

Modeling green microalgal growth, nutrient uptake and storage in the ASM framework

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Publication date:
2015

Document Version
Peer reviewed version

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Citation (APA):

Wágner, D. S., Valverde Perez, B., Sæbø, M., Bregua de la Sotilla, M., Van Wageningen, J., Smets, B. F., & Plósz, B. G. (2015). Modeling green microalgal growth, nutrient uptake and storage in the ASM framework [Sound/Visual production (digital)]. 9th IWA Symposium on Systems Analysis and Integrated Assessment (Watermatex 2015), Gold Coast, Queensland, Australia, 14/06/2015

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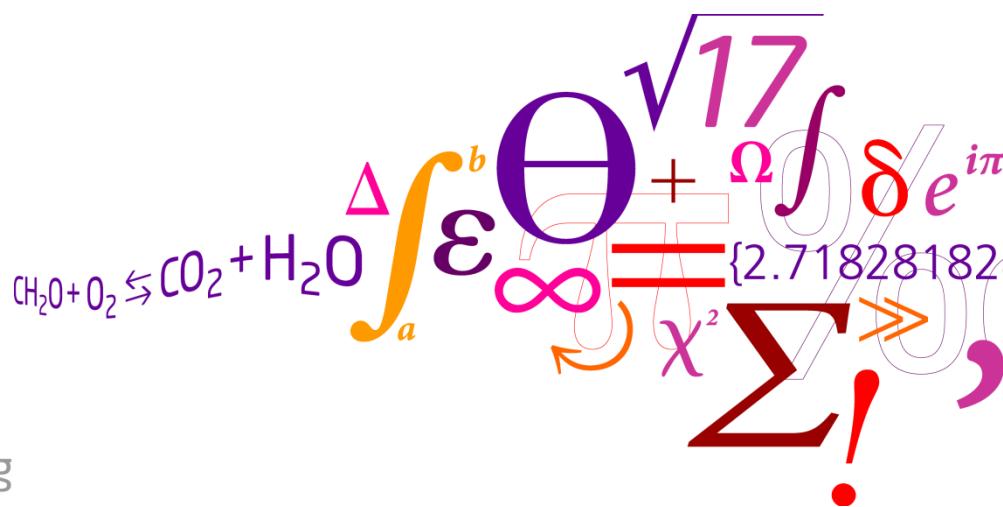
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Modeling green microalgal growth, nutrient uptake and storage in the ASM framework

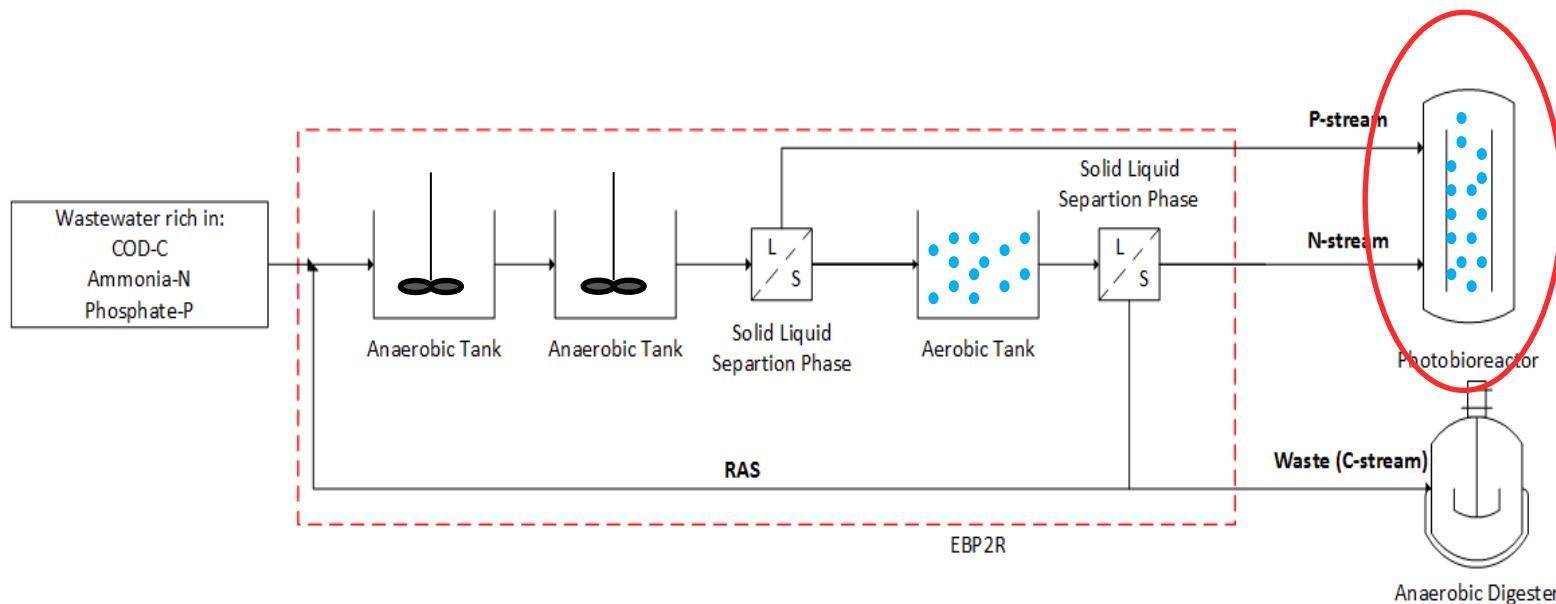
Dorottya S. Wágner, Borja Valverde-Pérez, Mariann Sæbø, Marta Bregua de la Sotilla, Jonathan van Wagenen, Barth F. Smets, Benedek Gy. Plósz

$$\text{CH}_2\text{O} + \text{O}_2 \xrightarrow{\Delta} \text{CO}_2 + \text{H}_2\text{O}$$


A collage of mathematical symbols including integrals, summations, and Greek letters like theta, omega, and sigma, along with a chemical reaction equation.

Motivation

- Microalgae photobioreactors can be used for wastewater treatment as:
 - Tertiary treatment step for nutrient removal
 - Nutrient recovery technology due to the phosphorus and nitrogen internal storage



Valverde-Pérez et al. (2015)

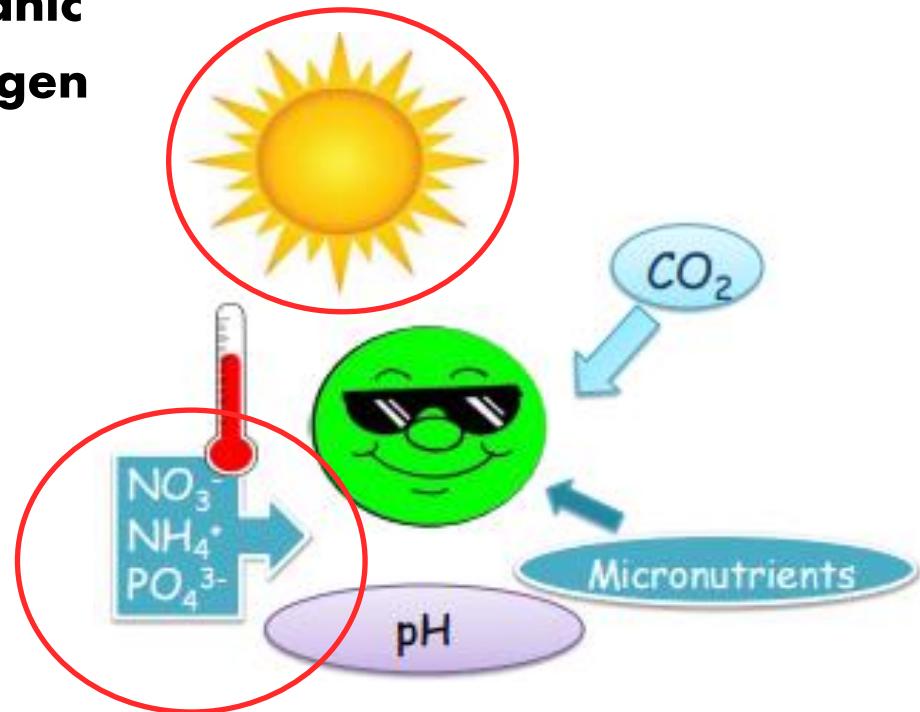
Motivation

- Available models are good, however the applicability may be limited:

- By the number of considered variables
- By the model structure

- Factors affecting microalgal growth:

- Carbon, both organic and inorganic
- Nutrients: phosphorus and nitrogen
- pH
- Light
- Micronutrients (e.g. iron)



Agenda

- **Model development**
- **Targeted experiments for parameter estimation:**
 - **Green microalgae identification and equipment**
 - **Microbatch and 1-L batch experiments**
 - **Open pond experiments**
- **Model evaluation**

Model development (ASM-A)

- Mechanistic description of **biokinetic** processes: multiple-substrate kinetic process rate equations based on literature

Model	Autotrophic algal growth										Heterotrophic algal growth	
	Nutrients			DIC	Light							
	N	P		Monod	Haldane	Steele	P&E	P&J	Smith	Poisson		
Moya et al. ⁴⁶											H	
Bouterfas et al. ⁴⁸												
Ambrose et al. ³¹	D	D										
Wolf et al. ³²	M											
Bougaran et al. ⁴²	D	D										
Quinn et al. ³³	D											
Broekhuizen et al. ³⁴	M											
Skjelbred et al. ⁴⁹												
Guest et al. ³⁵	D											
Decostere et al. ³⁶												
Coppens et al. ³⁸	M											
Van Wagenen et al. ⁵²												
ASM-A (This study)	D										M	

D: Droop

M: Monod

H: Haldane

Model development (ASM-A)

- Developed as an **extension of ASM-2d**, so compatible with activated sludge models
- Units and nomenclature expressed according to the **ASM** (activated sludge modelling) **framework**

Component	NH_4	NO_3	Internal quota N	PO_4	Internal quota P	Inorg. carbon	Acetate	O_2	Algal Biomass	Inert Particulates
Symbol	S_{NH_4}	S_{NO}	$X_{\text{Alg},N}$	S_{PO_4}	$X_{\text{Alg},PP}$	S_{Alk}	S_A	S_{O_2}	X_{Alg}	X_I
Unit	gN/m^3	gN/m^3	gN/m^3	gP/m^3	gP/m^3	gC/m^3	gCOD/m^3	gCOD/m^3	gCOD/m^3	gCOD/m^3
Process										
Uptake and storage of nitrogen from NH_4	-1		1							
Uptake and storage of nitrogen from NO_3		-1	1							
Uptake and Storage of PO_4				-1	1					
Autotrophic growth			$-i_{\text{NX}_{\text{alg}}}$		$-i_{\text{PX}_{\text{alg}}}$	$-1/Y_{\text{X}_{\text{alg}}, \text{S}_{\text{Alk}}}$		$1/(2.67 * Y_{\text{X}_{\text{alg}}, \text{S}_{\text{Alk}}})$	1	
Heterotrophic growth			$-i_{\text{NX}_{\text{alg}}}$		$-i_{\text{PX}_{\text{alg}}}$	$1/(0.4 * Y_{\text{X}_{\text{alg}}, \text{S}_A})$	$-1/(1.067 * Y_{\text{X}_{\text{alg}}, \text{S}_A})$	$-1/(1.067 * Y_{\text{X}_{\text{alg}}, \text{S}_A})$	1	
Decay	$i_{\text{NX}_{\text{alg}}} - f_{\text{Xi}} * i_{\text{NX}_{\text{Alg},D}}$			$i_{\text{PX}_{\text{alg}}} - f_{\text{Xi}} * i_{\text{PX}_{\text{Alg},D}}$				$-(1-f_{\text{Xi}})$	-1	f_{Xi}

Microalgal nutrient uptake and storage



- Processes 1&2: uptake and storage of nitrogen using ammonia and nitrate as nitrogen source

$$k_{NH4} \cdot \frac{S_{NH4}}{S_{NH4} + K_{NH4,Alg}} \cdot \frac{X_{Alg,Nmax} \cdot X_{Alg} - X_{Alg,N}}{X_{Alg,Nmax} \cdot X_{Alg}} \cdot X_{Alg}$$

$$k_{NO} \cdot \frac{S_{NO}}{S_{NO} + K_{NO,Alg}} \cdot \frac{K_{NH4,Alg}}{K_{NH4,Alg} + S_{NH4}} \cdot \frac{X_{Alg,Nmax} \cdot X_{Alg} - X_{Alg,N}}{X_{Alg,Nmax} \cdot X_{Alg}} \cdot X_{Alg}$$

- Process 3: uptake and storage of phosphate

$$k_{PO4} \cdot \frac{S_{PO4}}{S_{PO4} + K_{PO4,Alg}} \cdot \frac{X_{Alg,PPmax} \cdot X_{Alg} - X_{Alg,PP}}{X_{Alg,PPmax} \cdot X_{Alg}} \cdot X_{Alg}$$

Rieger et al. (2001)

Rhee (1973)

Ambrose et al. (2006)

Microalgal growth and decay

• Processes 4: autotrophic growth rate

$$\mu_{A,max} \cdot \left(1 - \frac{X_{Alg,Nmin} X_{Alg}}{X_{Alg,N}}\right) \cdot \left(1 - \frac{X_{Alg,PPmin} X_{Alg}}{X_{Alg,PP}}\right) \cdot \frac{S_{Alk}}{S_{Alk} + K_{Alk}} \cdot \frac{I_{Av}}{I_S} \cdot e^{1 - \frac{I_{Av}}{I_S}} \cdot X_{Alg}$$

• Process 5: heterotrophic growth rate

$$\mu_{H,max} \cdot \left(1 - \frac{X_{Alg,Nmin} X_{Alg}}{X_{Alg,N}}\right) \cdot \left(1 - \frac{X_{Alg,PPmin} X_{Alg}}{X_{Alg,PP}}\right) \cdot \frac{S_A}{S_A + K_A} \cdot \frac{S_{O2}}{S_{O2} + K_{O2}} \cdot \frac{K_I}{K_I + I_{Av}} \cdot X_{Alg}$$

• Process 6: decay

$$b_{X_{Alg}} \cdot X_{Alg}$$

Droop (1974)

Broekhuizen et al. (2012)

Rieger et al. (2001)

Ambrose et al. (2006)

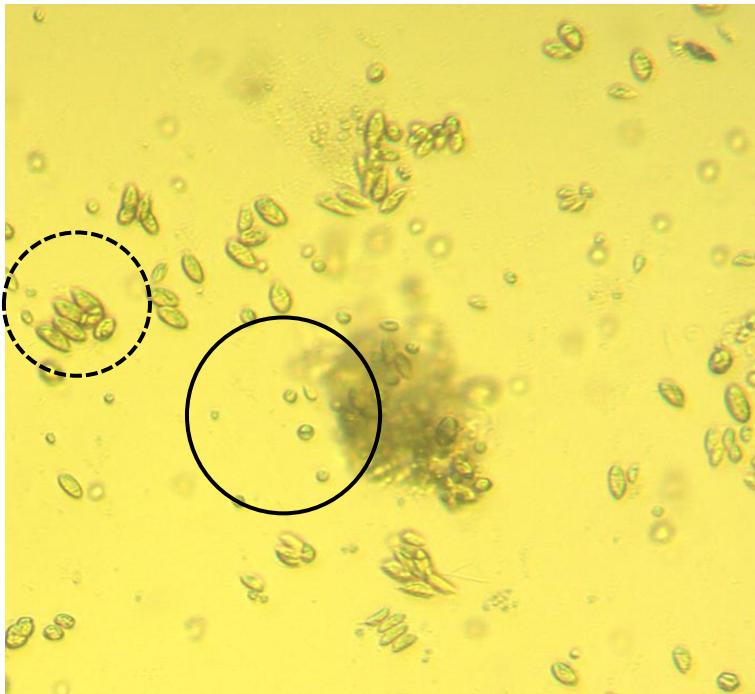
Chen and Johns (1994)

Agenda

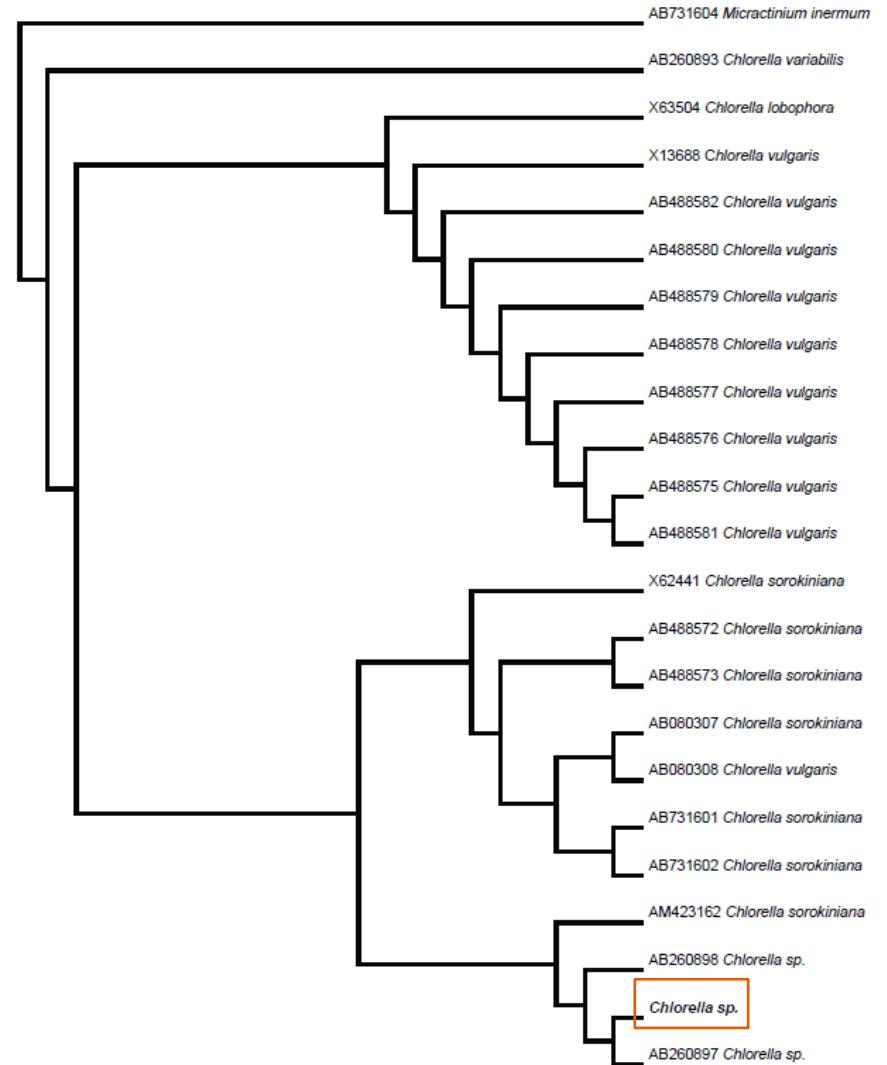
- Model development
- Targeted experiments for parameter estimation:
 - Green microalgae identification and equipment
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Microalgal culture

- Mixed green microalgal culture consists mainly of ***Chlorella sp.*** (*C. sorokiniana*) and ***Scenedesmus sp.***



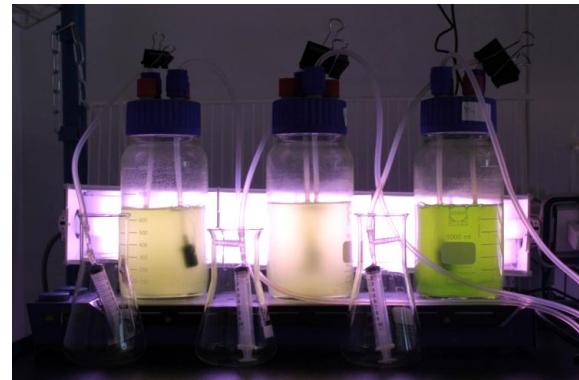
Microscope image of the mixed green microalgae culture in the batch experiment.



Reactors

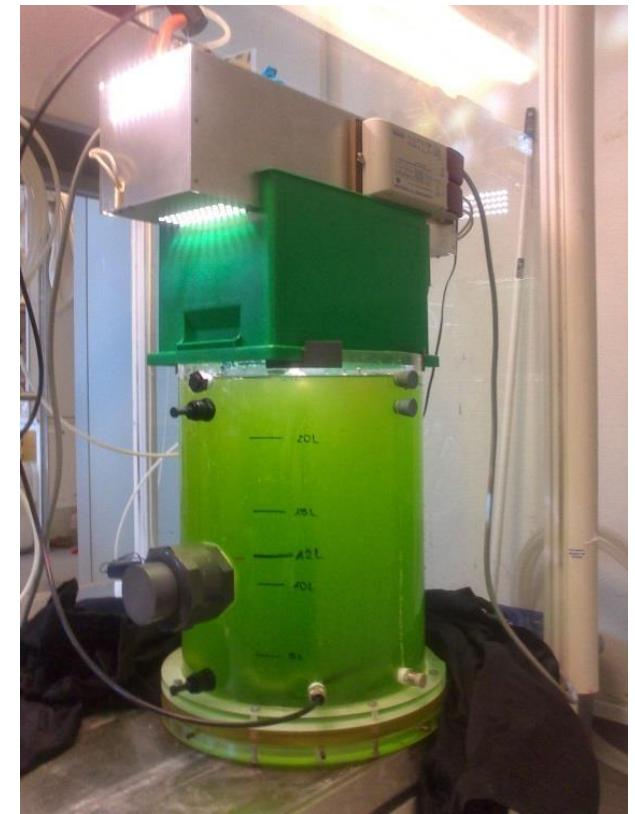


2 mL microbatches



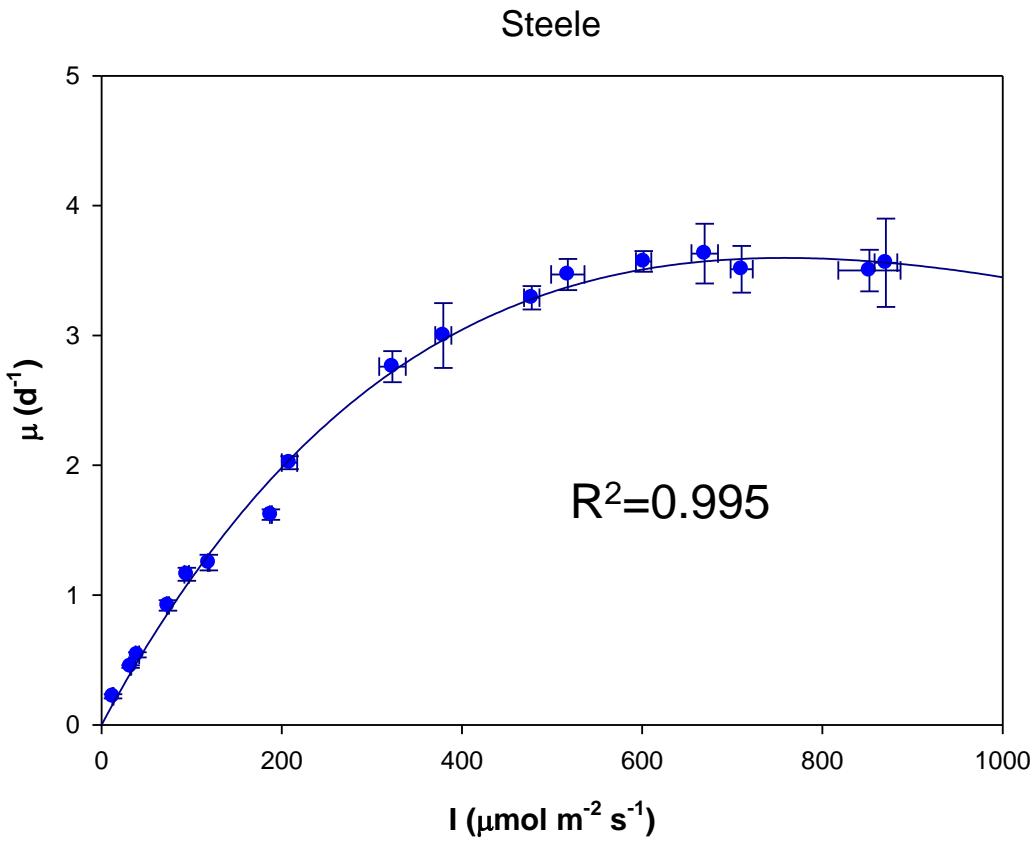
1-L batches

24 L open airlift PhBR



Microbath experiments: light intensity effect

- Assessing the specific growth rate under different light intensities



Steele equation:

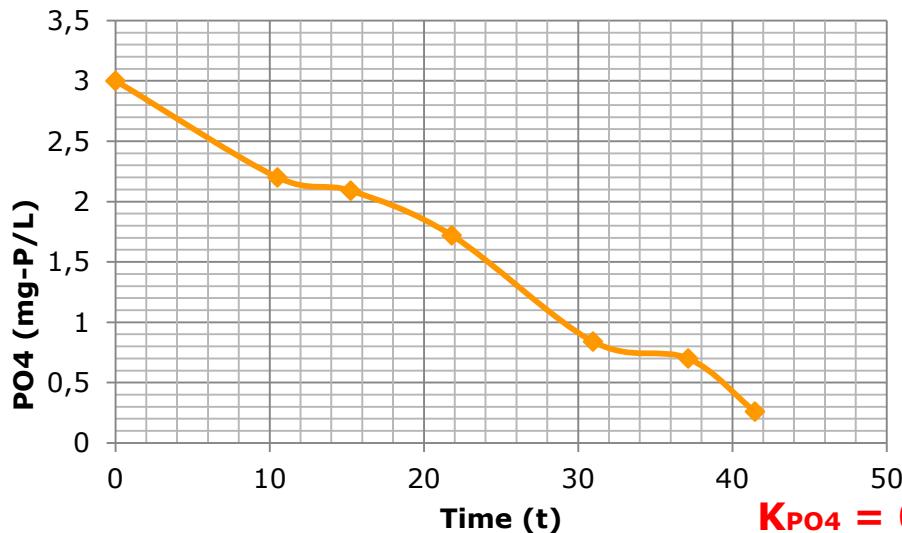
$$\mu = \frac{\mu_{max} * I}{I_s} \exp^{1-I/I_s}$$

$$\mu_{max} = 3.6 \text{ d}^{-1}$$

$$I_s = 758 \mu\text{mol m}^{-2} \text{s}^{-1}$$



Nutrient uptake and storage: P-uptake and storage



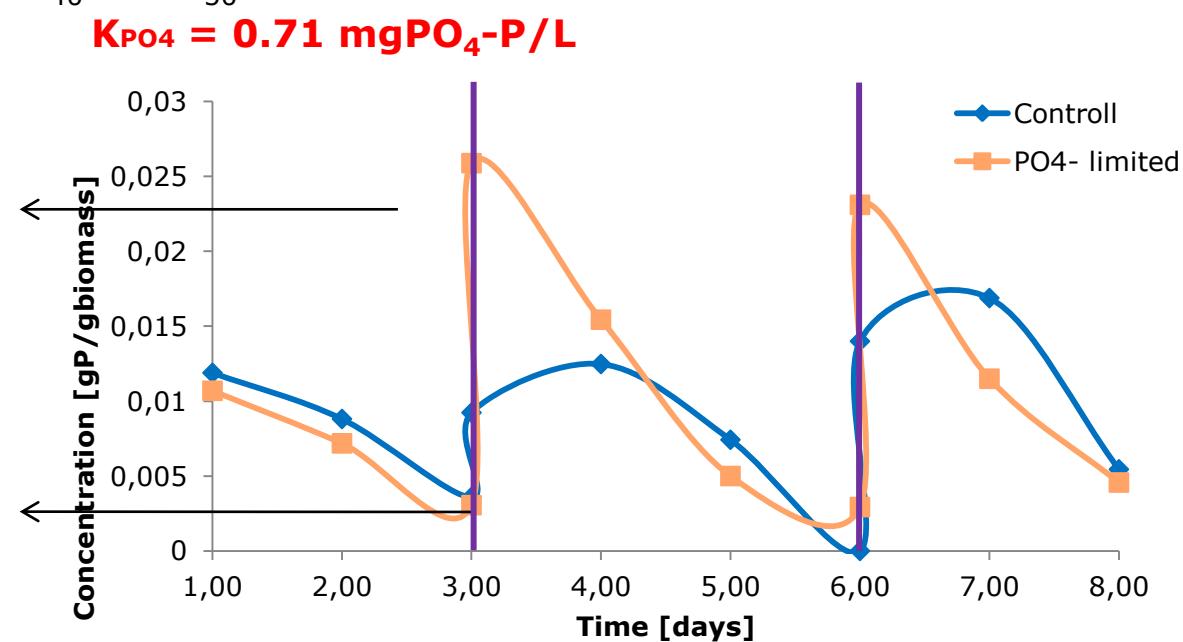
Parameter estimation using Simplex



$$I_{av} = 100 \text{ } \mu\text{mol m}^{-2}\text{s}^{-1}$$

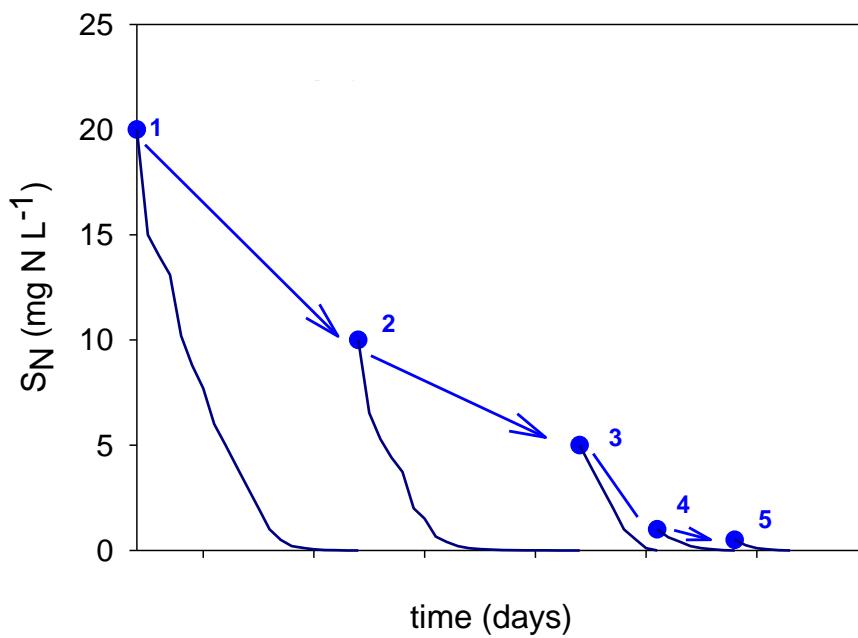
$$X_{Alg,PP_{max}} = 0.025 \text{ gP/gBiomass}$$

$$X_{Alg,PP_{min}} = 0.002 \text{ gP/gBiomass}$$



N-uptake and storage

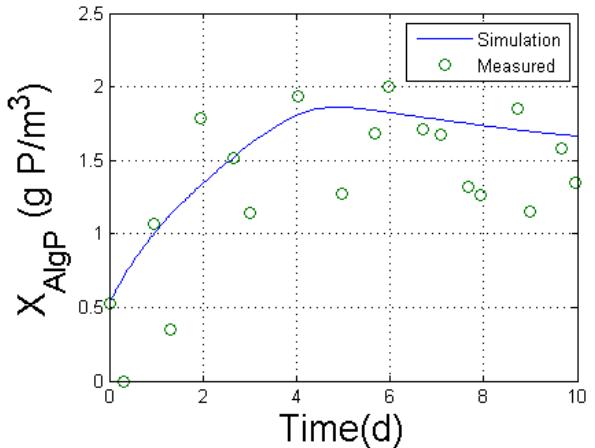
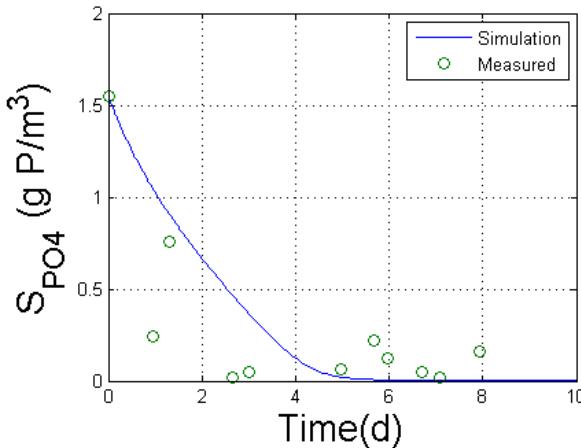
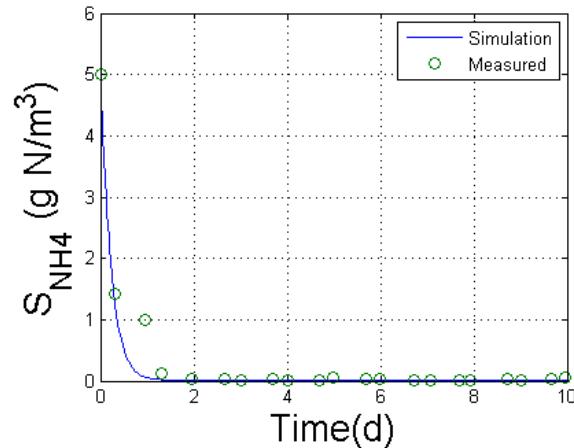
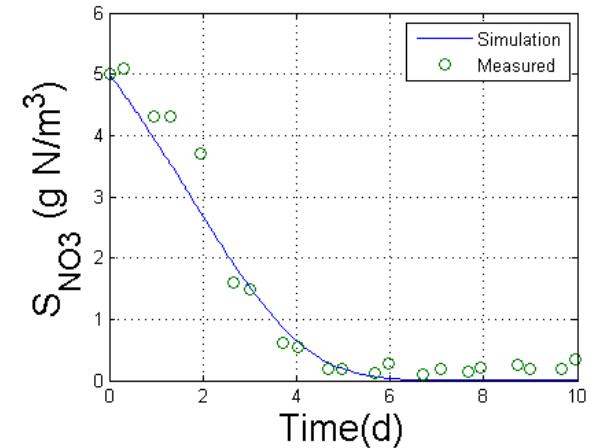
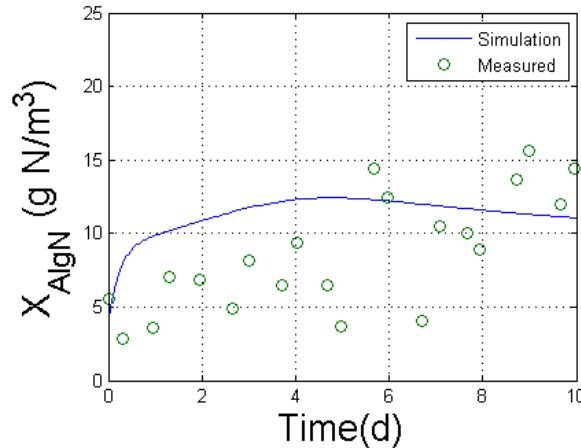
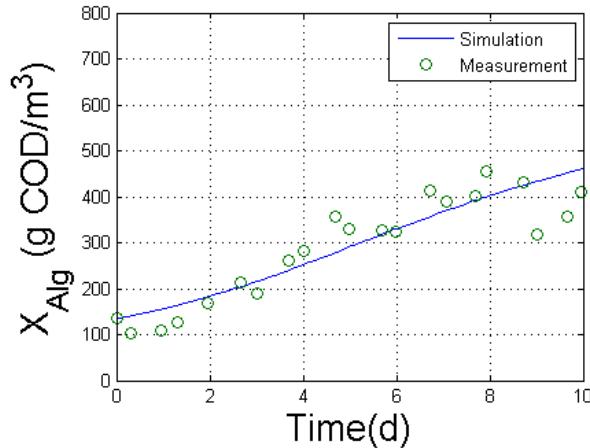
Cycle	Initial N conc. (g N/m ³)
1	20
2	10
3	5
4	1
5	0.5



Parameter
estimation using
Simplex

ASM-A calibration

Model calibration using data from cycle 2



Parameter Values

Parameter	Microplate	1-L batch	24-L batch	Literature Value	Unit
$\mu_{A,max}$	3.6 ± 0.04^a	3.3 ± 0.55^e	4.19 ± 1.19^g	$4.7^{30}; 0.1-11^{36};$ $1.4^{70}; 0.15^{71};$ $2.64^{67}; 1.47^{67};$ 3.26^{72}	d^{-1}
$\mu_{H,max}$	0.75 ± 0.1^b	3.23 ± 1.1^f	-	$14^{37}; 3.48^{73};$ $2.35-2.64^{67}; 0.96-$ 1^{67}	d^{-1}
I_s	758.2 ± 22.9^a	-	-	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$
$X_{Alg,PPmin}$	-	0.002 ± 0.0008^e	-	-	$\text{g P}\cdot\text{g}^{-1}\text{COD}$
$X_{Alg,Nmin}$	-	0.014 ± 0.0015^e	-	$0.023^{37}; 0.02^{72}$	$\text{g N}\cdot\text{g}^{-1}\text{COD}$
$X_{Alg,PPmax}$	-	0.02 ± 0.008^e	-	-	$\text{g P}\cdot\text{g}^{-1}\text{COD}$
$X_{Alg,Nmax}$	-	0.9 ± 0.5^e	-	$0.0683^{37}; 0.067^{72}$	$\text{g N}\cdot\text{g}^{-1}\text{COD}$
$K_{NO,Alg}$	-	3.39 ± 1.89^e	8.22 ± 0.97^g	$0.0602^{37}; 0.3^{71}$	$\text{g N}\cdot\text{m}^{-3}$
$K_{NH4,Alg}$	-	2.4 ± 0.42^e	3.91 ± 0.88^g	$5^{34}; 0.1-31.5^{71}$	$\text{g N}\cdot\text{m}^{-3}$
$K_{PO4,Alg}$	-	0.25 ± 0.18^e	0.36 ± 0.046^g	$0.046-10.5^{71}$	$\text{g P}\cdot\text{m}^{-3}$
$k_{NH4,Alg}$	-	2.97 ± 2.42	0.22 ± 0.01^g	-	$\text{g N}\cdot\text{g}^{-1}\text{COD}\cdot\text{d}^{-1}$
$k_{NO,Alg}$	-	2.97 ± 2.42	0.018 ± 0.012^g	$0.49^{37}; 0.43^{72}$	$\text{g N}\cdot\text{g}^{-1}\text{COD}\cdot\text{d}^{-1}$
$k_{PO4,Alg}$	-	0.066 ± 0.042	0.028 ± 0.021^g	-	$\text{g P}\cdot\text{g}^{-1}\text{COD}\cdot\text{d}^{-1}$
K_{Alk}	-	-	-	$3^{34}; 0.035-0.29^{71}$	$\text{g C}\cdot\text{m}^{-3}$
K_A	11.87 ± 5.2^b	89 ± 28^f	-	-	$\text{g COD}\cdot\text{m}^{-3}$
$K_{i,A}$	550.58 ± 232.1^b	-	-	-	$\text{g COD}\cdot\text{m}^{-3}$
K_I	878.6 ± 75^d	-	-	-	$\mu\text{mol m}^{-2}\text{s}^{-1}$
K_O				$2 \text{ (20\% of DO}_{\text{sat}}\text{)}_{56}$	g COD m^{-3}
b_{Xalg}	-	-	0.091 ± 0.0^g	$0.072 \text{ (2\% } * \mu_{A,max})_{33}; 3.7\% \mu_{A,max}^{30},$ $0.003-0.1^{71}$	d^{-1}
Y_{Alk}	-	-	-	$2.33; 1.06^{71}$	$\text{g COD}\cdot\text{g}^{-1}\text{C}$
Y_{Ac}	-	0.39^f	-	0.34^{45}	$\text{g COD}\cdot\text{g}^{-1}\text{COD}$
iP_{Xalg}	-	-		$0.00067^{31,34}$	$\text{g P}\cdot\text{g}^{-1}\text{COD}$
iN_{Xalg}	-	-		$0.0049^{31}; 0.0534^{34}$	$\text{g N}\cdot\text{g}^{-1}\text{COD}$
fX_I	-	-		0.1	$\text{g COD}\cdot\text{g}^{-1}\text{COD}$
N_{XAlgD}				0.00441	$\text{g N}\cdot\text{g}^{-1}\text{COD}$
P_{XAlgD}				0.000603	$\text{g P}\cdot\text{g}^{-1}\text{COD}$

Agenda

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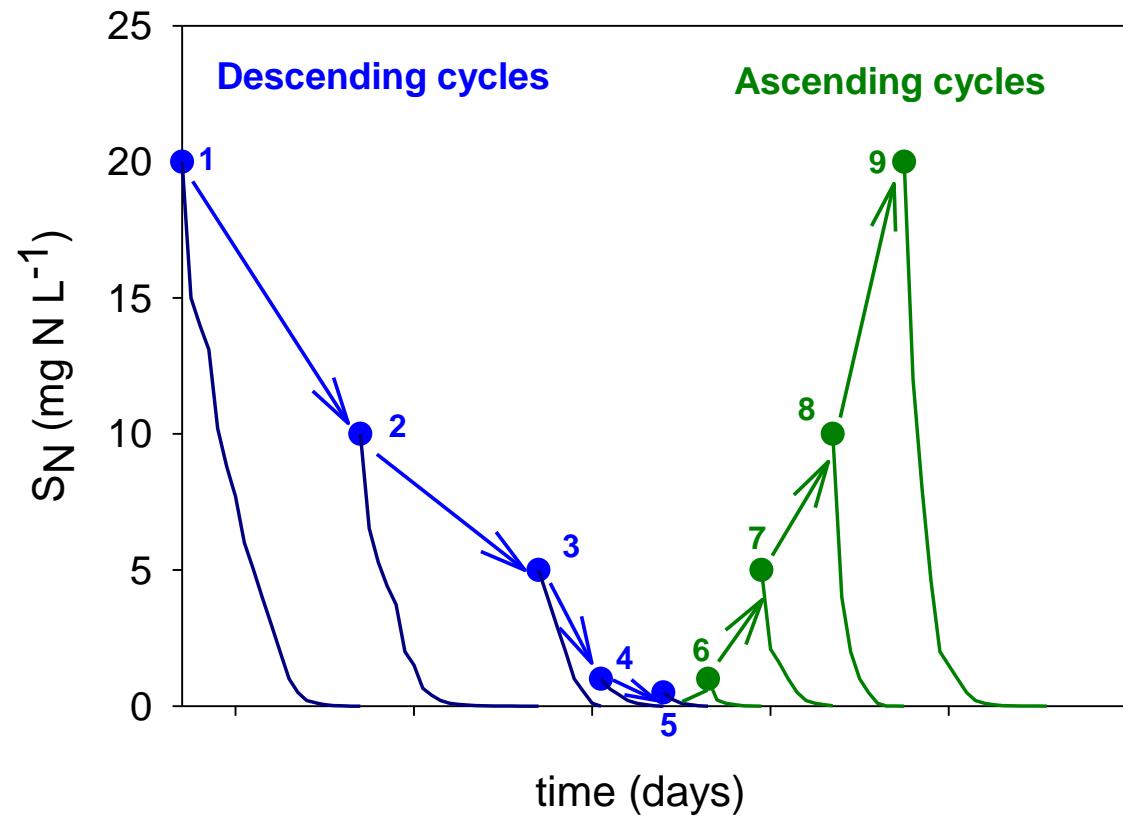
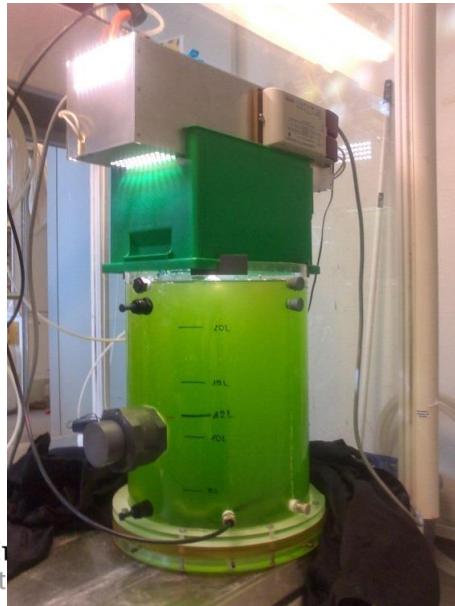
Model evaluation: assessment of parameter variability impact

Research questions:

- **What is the influence of culture history and/or substrate availability on parameter estimates?**
- **Can we use a default parameter set?**
- **Can we explain the discrepancy as a result of parameter variability?**

Model evaluation: experimental design

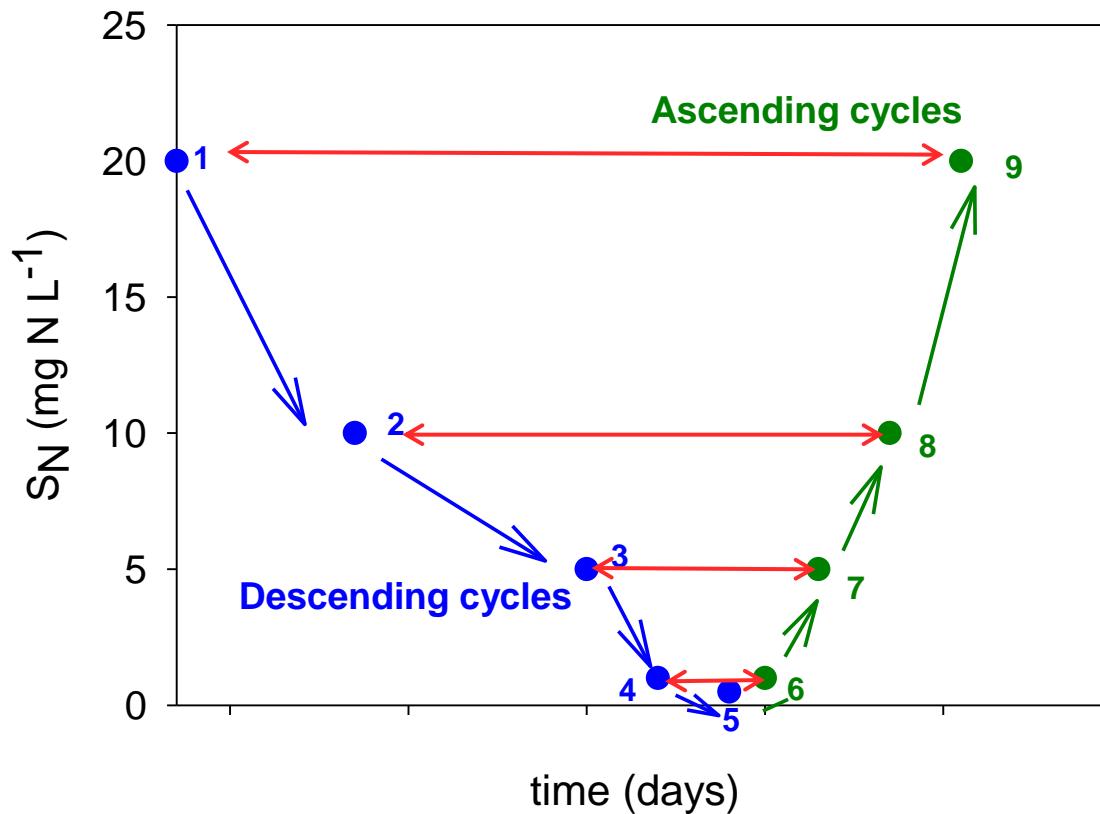
Cycle	Initial N conc. (g N/m ³)
1 and 9	20
2 and 8	10
3 and 7	5
4 and 6	1
5	0.5



Model evaluation: two evaluation steps

- Does culture history affect parameter values?

- Parameter sets obtained through the descending cycles confronted with data from ascending cycles
- Janus coefficient



Model evaluation: two evaluation steps



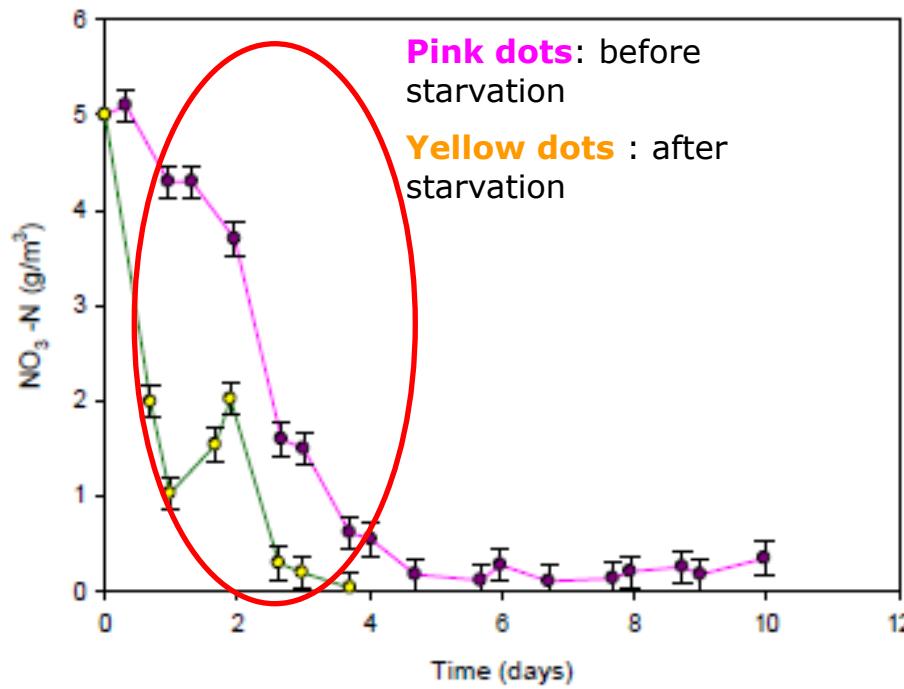
Cycle 2-8	RMSE calibration	RMSE evaluation	Janus coefficient
Ammonium in bulk liquid (S_{NH_4})	0.72	0.44	0.61
Nitrate in bulk liquid (S_{NO_3})	0.71	14.00	19.72
Phosphate in bulk liquid (S_{PO_4})	0.91	0.51	0.56
Algal biomass (X_{Alg})	0.19	0.1	0.53
Nitrogen quota (X_{AlgN})	1.27	0.70	0.55
Phosphorous quota (X_{AlgP})	0.91	0.14	0.15
Total	4.71	15.9	3.38

Nitrate prediction in the other cycles:

Cycle	RMSE calibration	RMSE evaluation	Janus coefficient
1-9	0.24	23.9	99.58
3-7	0.26	3.85	14.81
4-6	0.23	9.35	40.65

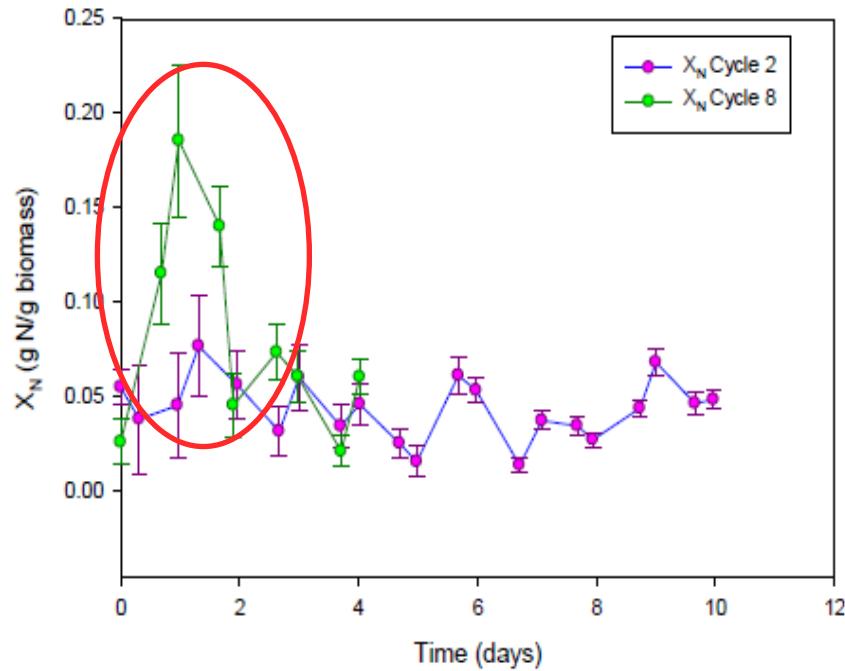
- $J \sim 1$ calibrated model prediction is good
- $J > >$ calibrated model prediction fails

Model evaluation: experimental results



- The **nitrate uptake** after starvation conditions is **enhanced**

Model evaluation: experimental results

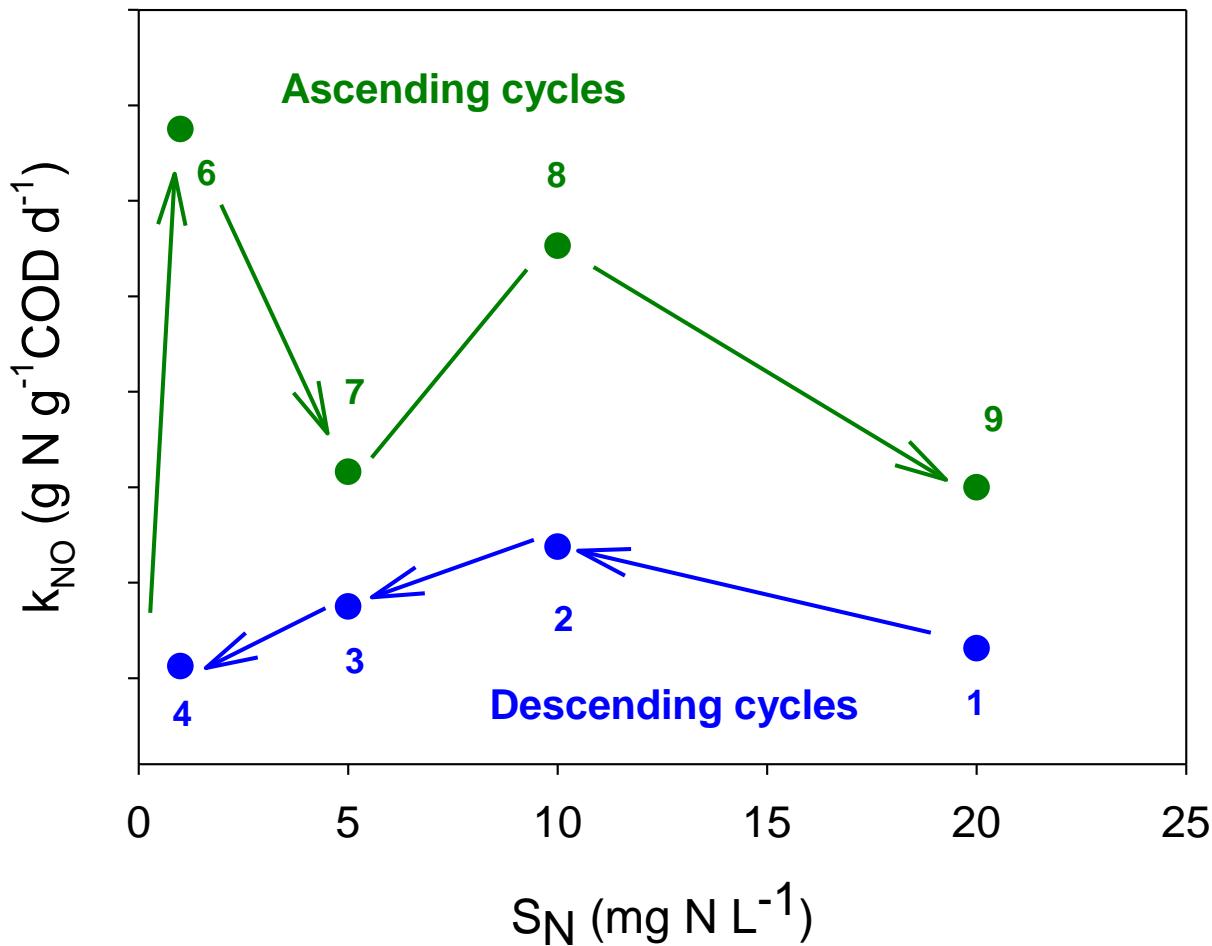


Pink dots: before starvation

Green dots: after starvation

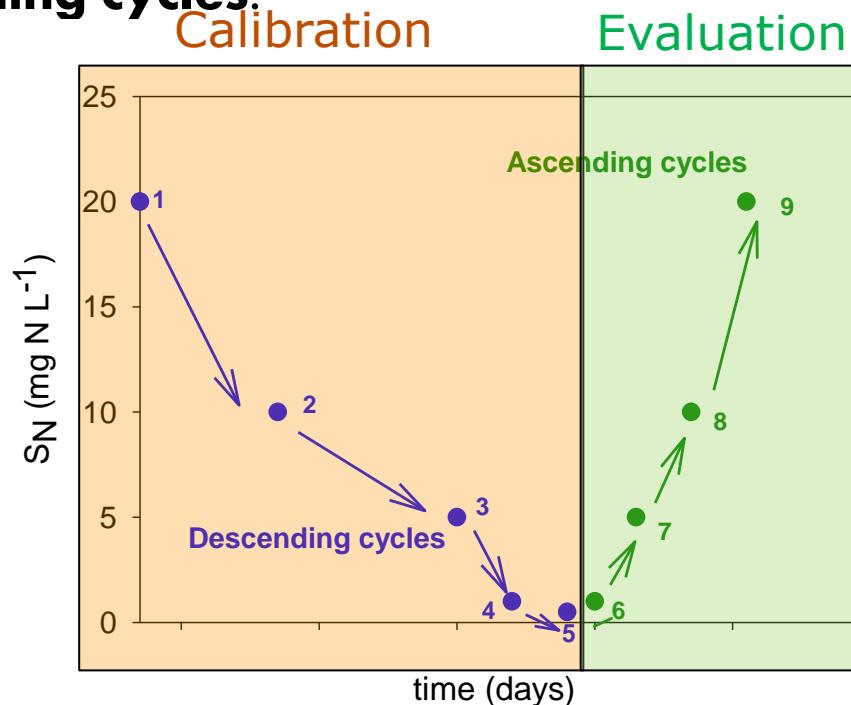
- After the **N quota** is replenished there is a **temporary enhanced** N storage

Model evaluation: parameter variability



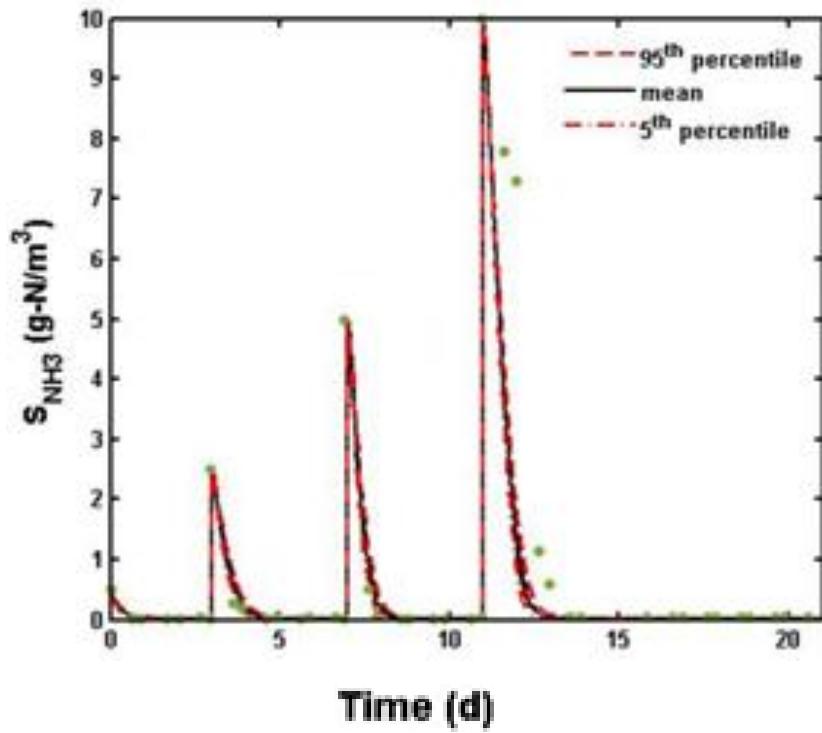
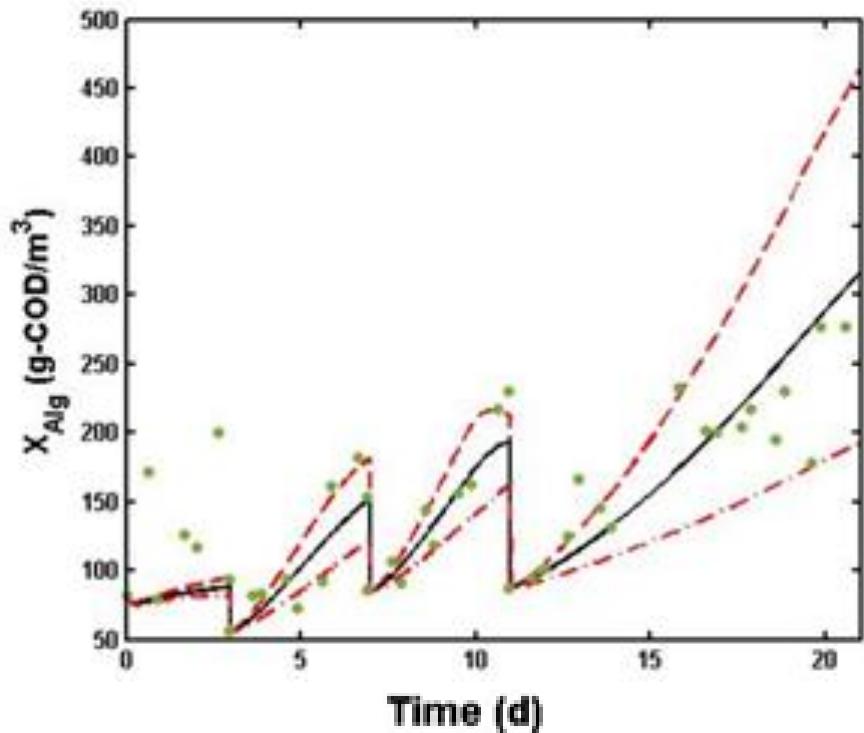
Model evaluation: two evaluation steps

- Can we use an average parameter set? Can we explain the discrepancy as a result of parameter variability?
 - Monte Carlo simulations run on the 4 ascending cycles
 - Parameter values: mean values of the estimated parameters through descending cycles.
 - Probability range: standard deviation of the mean values through descending cycles.



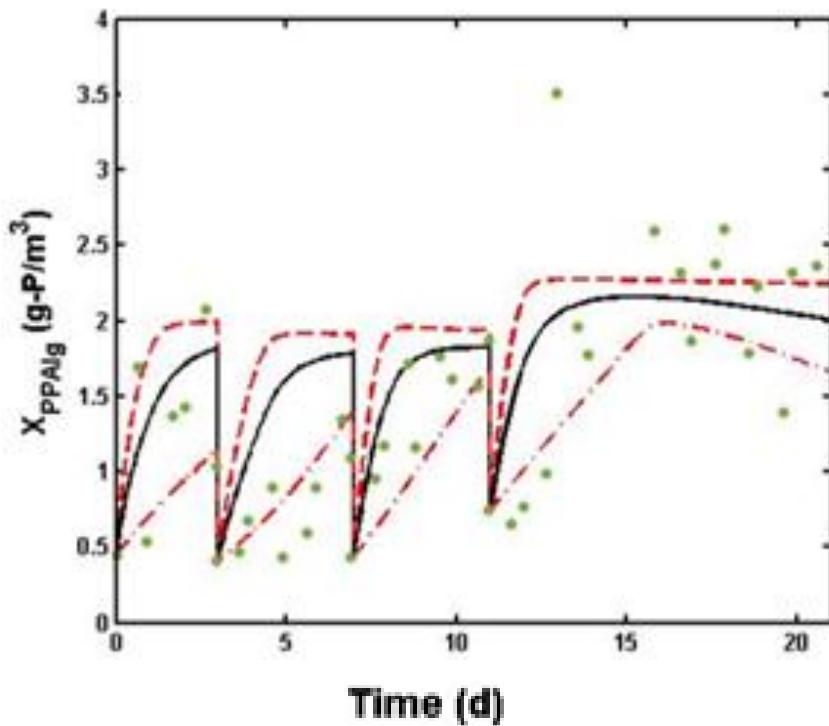
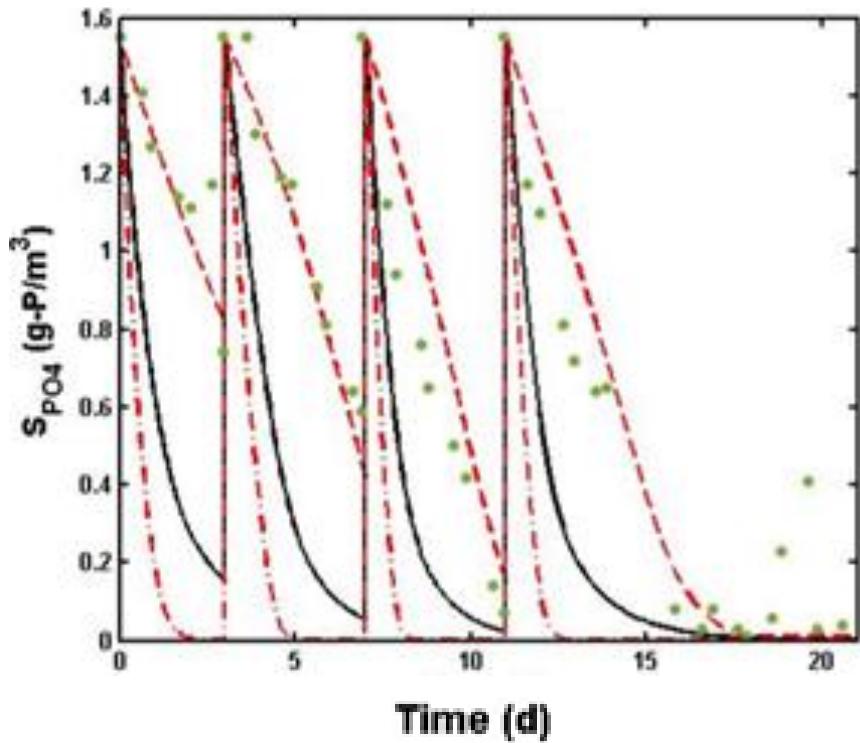
Model evaluation: second steps

- Algal biomass & ammonia: discrepancies can be explained by parameter variability



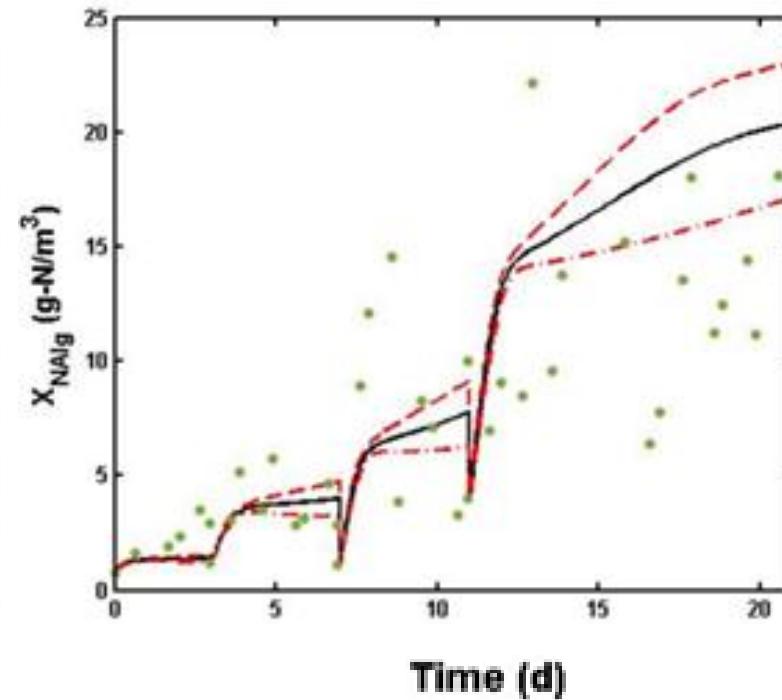
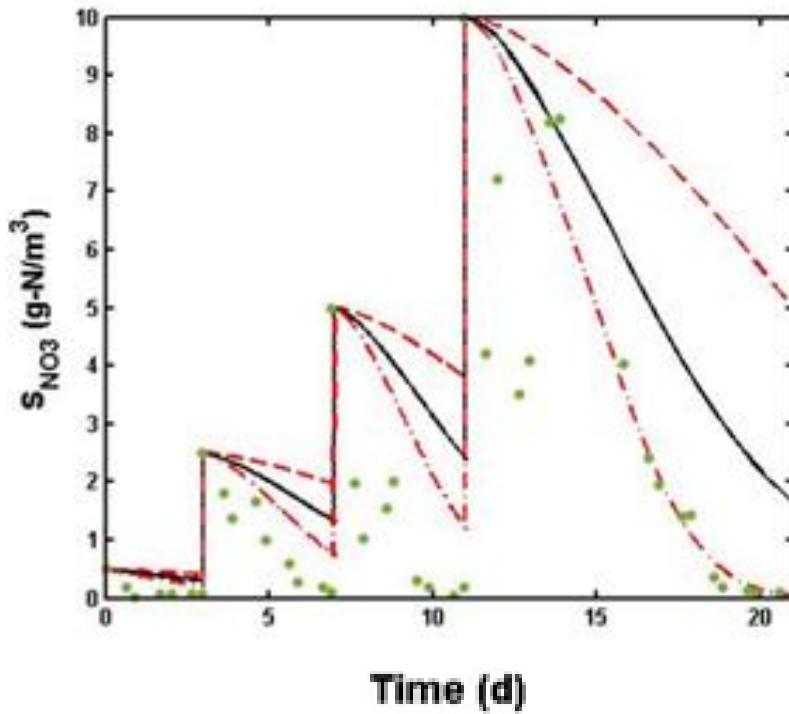
Model evaluation: two evaluation steps

- Soluble and stored phosphate: discrepancies can be explained by parameter variability



Model evaluation: two evaluation steps

- Soluble nitrate & stored nitrogen: the model prediction is compromised by
 - Culture history for nitrate
 - Substrate availability for nitrogen storage



Concluding remarks

- A novel process model in the ASM framework for predicting algal behavior in PBR has been identified, calibrated and critically evaluated
- Different scale lab experiments have been used to estimate different parameter sets.
- The model can predict algal biomass, ammonia, phosphate and internal PP quota using a mean parameter set
- Maximum nitrate uptake rate depends on the history of the culture

Future perspectives

- Model extensions including physic-chemical processes:
 - Mass transfer
 - pH
 - Light attenuation
 - Hydrodynamics
- Model extensions relevant to other end-uses: lipid accumulation for biodiesel
- Model calibration and evaluation using other microalgae species

Acknowledgement



Dorottya S. Wágner, Mariann Sæbø, Marta Bregua de la Sotilla,
Jonathan van Wageningen, Barth F. Smets, Benedek Gy. Plósz



Thank you for your attention!