The therapeutic effect of dietary nitrate supplementation in healthy adults, individuals with type 2 diabetes mellitus and chronic obstructive pulmonary disease.

Submitted by Anthony Ian Shepherd to the University of Exeter
as a thesis for the degree of
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#### **Abstract**

#### Introduction and aim

Increases in the bioavailability of nitric oxide have been shown to reduce the oxygen (O<sub>2</sub>) cost of exercise, improve exercise performance, alter gastric blood flow and mediate glucose uptake in healthy individuals. Aim; does dietary nitrate reduce the O<sub>2</sub> cost of exercise, improve walking performance in individuals with type 2 diabetes mellitus (T2DM) and chronic obstructive pulmonary disease (COPD) or alter hepatic diffusion and positively affect glucose homeostasis in healthy adults?

#### Methods

Experimental chapters utilised a double-blind, placebo-controlled, randomised, experimental design. Breath by breath pulmonary gas analysis was utilised to assess the O<sub>2</sub> cost of exercise in 48 individuals with T2DM and 13 with COPD. Walking performance was assessed via the six minute walk test (6MWT) in cohorts 1 and 2. Magnetic resonance imaging was used to assess portal vein flux, velocity and the apparent diffusion coefficient, in order to assess hepatic microvascular diffusion (apparent diffusion coefficient (ADC)). Blood pressure (BP) was measured in all trials.

## Results

Relative to placebo, beetroot juice resulted in a significant increase in plasma nitrate and nitrite. There were no differences between placebo vs. beetroot juice for the  $O_2$  cost of walking (T2DM: placebo; 946 ± 221 vs. beetroot juice; 939 ± 223 ml·min<sup>-1</sup>; P=0.59) or cycling (COPD: placebo; 933 ± 323 vs. beetroot juice; 939 ± 302 ml: min<sup>-1</sup>; P=0.88), distance covered in the 6MWT (T2DM: placebo;

550 ± 83 vs. beetroot juice; 554 ± 90m; P=0.17 or COPD: placebo; 456 ± 86 vs. beetroot juice; 449 ± 79 m; P=0.37) or BP (T2DM: systolic: placebo; 134 ± 10 vs. beetroot juice; 132 ± 12 mmHg, P=0.17; diastolic: placebo; 77 ± 7: vs. beetroot juice; 76 ± 11 mmHg, P=0.27. COPD: systolic: placebo; 123 ± 14 vs. beetroot juice; 123 ± 14 mmHg; P=0.91; diastolic: placebo; 77 ± 9 vs. beetroot juice; 79 ± 9 mmHg; P=0.27). No differences were seen between placebo and beetroot juice for ADC (young adults: F(3, 45) = 0.25, P = 0.74; older adults; F(3, 42) = 1.3, P = 0.28), portal vein flux (young adults: F(3, 45) = 0.339, P = 0.79; older adults; F(3, 42) = 1.65, P = 0.19) however, there was an interaction effect in the young adults: (F(3, 45) = 2.9, P = 0.04) but not in the older adults; F(3, 42) = 1.8, P = 0.16) between visits for portal vein velocity.

Nitrate supplementation did not reduce plasma glucose concentrations (young adults:  $F_{(3, 45)} = 0.96$ , P = 0.42; older adults;  $F_{(3, 42)} = 0.04$ , P = 0.99). Nitrate supplementation did not reduce systolic blood pressure (young adults:  $F_{(3, 45)} = 0.20$ , P = 0.89; older adults;  $F_{(3, 42)} = 1.7$ , P = 0.18) or diastolic blood pressure (young adults:  $F_{(3, 45)} = 0.25$ , P = 0.86; older adults;  $F_{(3, 42)} = 0.45$ , P = 0.72).

# Conclusion

Dietary nitrate supplementation does not alter the  $O_2$  cost of exercise, improve walking performance or reduce BP in individuals with T2DM or COPD. Nitrate supplementation does not alter hepatic diffusion, glucose homeostasis or BP.

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#### **Authors declaration**

In experimental chapter 1; the high-performance liquid chromatography analysis of the antioxidant content of the beetroot juice was performed by Ewa Rembialkowska group in Warsaw, Poland.

#### **Abbreviations**

Nitric oxide; NO-

Nitric oxide synthase; NOS

Oxygen; O<sub>2</sub>

Neuronal nitric oxide synthase; nNOS

Inducible nitric oxide synthase; iNOS

Endothelial nitric oxide synthase; eNOS

Nicotinamide adenine dinucleotide phosphate hydrogen; NADPH

Flavin adenine dinucleotide; FAD

Flavin mononucleotide; FMN

Calmodulin; CaM

Tetrahydrobiopterin; BH4

Nitrate; NO<sub>3</sub>

Nitrite; NO<sub>2</sub>

Type 2 diabetes mellitus; T2DM

Cyclic guanosine monophosphate; cGMP

Blood pressure; BP

Oxygen uptake;  $\dot{V}O_2$ 

Peak oxygen uptake;  $\dot{V}O_{2peak}$ 

Work rate; WR

Gross efficiency; GE

Delta efficiency; DE

Systolic blood pressure; SBP

Diastolic blood pressure; DBP

Mean arterial blood pressure; MAP

Ischemic reperfusion; IR

Flow mediated dilation; FMD

Moderate intensity; MI

Rating of perceived exertion; RPE

Severe intensity; SI

Maximal oxygen uptake  $\dot{V}O_2$ 

Low intensity; LI

High intensity; HI

Phosphocreatine; PCr

Inorganic phosphate; Pi

Adenosine diphosphate; ADP

Adenosine tri phosphate; ATP

Gas exchange threshold; GET

Peripheral artery disease; PAD

Arterial oxygen saturation; S<sub>a</sub>o<sub>2</sub>

Limit of tolerance; Tlim

Phosphorylation efficiency; P/O ratio

Power output; PO

Time trial; TT

Time to exhaustion; TTE

Low nitrate food; LNF

High nitrate food HNF

Partial pressure of oxygen; PO<sub>2</sub>

Dimethoxybenzaldehyde; DMBA

Near infrared spectrometry; NIRS

Soluble guanylate cyclase; sGC

Respiratory quotation; RQ

Maximal voluntary contraction; MVC

Red blood cells; RBC

Chronic obstructive pulmonary disease; COPD

Glycated haemoglobin; HbA1c

Asymmetric dimethylarginine; ADMA

Glucagon like peptide 1; GLP-1

Glucose dependant insulintropic peptide; GIP

Glucagon like peptide 1 receptor; GLP-1R

Glucagon like peptide 2; GLP-2

Guanosine triphosphate; GTP

GTP cyclohydrolase; GTPCH1

Peroxynitrite; OONO<sub>2</sub>

Dihydropterin; BH<sub>2</sub>

Dihydrofolate reductase; DHFR

Asymmetric dimethylarginine; ADMA

Symmetrical dimethylarginine; SDMA

Monomethyl-L-arginine; L-NMMA

Protein-arginine methyltransferases; PMRTs

Superoxide dismutase; SOD

Angiotensin-converting enzyme inhibitor; ACEi

Angiotensin receptor blockers; ARBs

Cardiovascular disease; CVD

Diabetes related nephropathy; DRN

Forced expiratory volume in the first second; FEV<sub>1</sub>

Quality of life; QoL

Reactive oxygen species; ROS

Mean pulmonary artery pressure; mPAP

Six minute walk test; 6MWT

Cyclic guanosine 3' 5' monophosphate; cGMP

Pulmonary artery hypertension; PAH

Phosphodiesterase type 5; PDE-5

Sodium hydroxide; NaOH

Zinc sulphate; ZnSO<sub>4</sub>

Vanadium chloride; VCI

Spike recovery; SR

Sodium Iodide; Nal

Amplitude; Ap

Time delay; TDp

Time constant;  $\tau_p$ 

Mean response time; MRT

Region of interest; ROI

Signal intensity in low flow sensitivity image; S<sub>0</sub>

Signal intensity in flow sensitive image; S<sub>1</sub>

Magnetic field gradient used in low flow sensitivity image=250 s/mm<sup>2</sup>; b<sub>0</sub>

Magnetic field gradient used in flow sensitive image=750 s/mm<sup>2</sup>; b<sub>1</sub>

Standard deviation; SD

Interquartile range; IQR

Gas exchange threshold; GET

#### Publications and awards arising from work contained in this thesis

#### Journal articles:

- Anthony I Shepherd, Daryl P Wilkerson, Lee Dobson, James Kelly, Paul G Winyard, Andrew M Jones, Nigel Benjamin, Angela C Shore, Mark Gilchrist (2015). The effect of dietary nitrate supplementation on the oxygen cost of cycling, walking performance and resting blood pressure in individuals with chronic obstructive pulmonary disease: A double blind placebo controlled, randomised control trial. *Nitric Oxide*. doi:10.1016/j.niox.2015.01.002.
- 2. Anthony I Shepherd, Mark Gilchrist, Paul G Winyard, Andrew M Jones, Ewelina Hallmann, Renata Kazimierczak, Ewa Rembialkowska, Nigel Benjamin, Angela C Shore, Daryl P Wilkerson (2015). Effects of dietary nitrate supplementation on the oxygen cost of exercise and walking performance in individuals with type 2 diabetes: a randomised, double blind, placebo-controlled cross-over trial. Free Radical Biology & Medicine. doi:10.1016/j.freeradbiomed.2015.05.014

#### Abstracts;

- Anthony I Shepherd, Mark Gilchrist, Paul G Winyard, Andrew M Jones, Nigel Benjamin, Angela C Shore, Daryl P Wilkerson. (2014). The effect of dietary nitrate supplementation on the ability of individuals with Type 2 diabetes to perform walking exercise: A randomised control trial. Diabetes UK Professional Conference. Liverpool.
- Anthony I Shepherd, Mark Gilchrist, Paul G Winyard, Andrew M Jones,
   Nigel Benjamin, Angela C Shore, Daryl P Wilkerson. (2014). Effects of
   dietary nitrate supplementation on blood pressure, the O<sub>2</sub> cost of

exercise, and walking performance in individuals with Type 2 diabetes: A randomised control trial. The British Association of Sport and Exercise Sciences. Student Conference. Portsmouth.

Award: Best postgraduate oral presentation.

3. Anthony I Shepherd, Mark Gilchrist, Paul G Winyard, Andrew M Jones, Nigel Benjamin, Angela C Shore, Daryl P Wilkerson. (2014). Effects of dietary nitrate supplementation on blood pressure, the O<sub>2</sub> cost of exercise, and walking performance in individuals with Type 2 diabetes. The European College of Sport Science. Amsterdam.

Shortlisted for the young investigator award.

#### **Chapter 1: Introduction**

Nitric oxide (NO·) was first found to be an endothelium derived relaxation factor in 1987 by Ignarro et al. (1987). The research questions surrounding its functions and biochemical pathways have expanded vastly since the identification of a stepwise reduction on nitrate (NO<sub>3</sub><sup>-</sup>) to nitrite (NO<sub>2</sub><sup>-</sup>) via facultative anaerobic bacteria and subsequently a conversion to NO· within the stomach (Lundberg et al., 1994, Benjamin et al., 1994).

More recently, trials investigating the impact of supplementation of inorganic NO<sub>3</sub><sup>-</sup> have shown some remarkable findings, predominantly in young healthy individuals. These include: improvements in mitochondrial efficiency (Larsen et al., 2011); reduction in the metabolic rate (Larsen et al., 2014), increases in blood flow (matching to skeletal muscle requirements and delivery to the brain) (Presley et al., 2011); reductions in systolic and diastolic blood pressure (Webb et al., 2008a, Vanhatalo et al., 2010, Vanhatalo et al., 2011, Lansley et al., 2010, Lansley et al., 2011a, Engan et al., 2012, Wylie et al., 2013b, Thompson et al., 2014, Muggeridge et al., 2014, Keen et al., 2014, Levitt et al., 2015, Treichler et al., 2015, Kocoloski and Crecelius, 2015, Bond Jr et al., 2013, Bond et al., 2014) and increase glucose uptake (Merry et al., 2010). In this introduction the biochemical pathways of NO· synthesis will be reviewed along with exploration of what is known surrounding NO<sub>3</sub><sup>-</sup> supplementation and the potential clinical applications of these beneficial effects.

#### 1.1: Inorganic nitrate (NO<sub>3</sub>), nitrite (NO<sub>2</sub>) & NO-

#### 1.1.1: NO synthesis in man:

NO· is a gaseous signalling molecule involved in many physiological processes such as vasodilation (Ignarro et al., 1987), neuronal signalling (Garthwaite, 2008) calcium handling (Hart and Dulhunty, 2000) and protection against bacterial pathogens (Chakravortty and Hensel, 2003). There are two major pathways in which NO is produced in humans, firstly; the nitric oxide synthase (NOS) pathway, which is oxygen (O<sub>2</sub>) dependent and creates endogenously derived NO. There are three NOS isoforms which are bi-domain in structure including; (i) neuronal NOS (nNOS), (ii) inducible NOS (iNOS) and (iii) endothelial NOS (eNOS), all of which produce NO in different locations of the body (see figure 1). The families of NOS's catalyse a five stage oxidation of Larginine to produce the biologically active molecule NO and also L-citrulline (Stamler and Meissner, 2001). This reaction relies on a constant substrate availability of six key compounds in order to maintain NO production via the NOS pathway (Crabtree et al., 2009), these are: nicotinamide adenine dinucleotide phosphate hydrogen (NADPH), flavin adenine dinucleotide (FAD), flavin mononucleotide (FMN), haem, calmodulin (CaM) and tetrahydrobiopterin (BH4) (Alderton et al., 2001). If any of these substrates are not present the production of NO via NOS enzymes is diminished. For eNOS activation to occur, the release of its inhibition is required by two CaM attachments which transfer electrons from the reductase domain of the NOS to the oxygenase domian (Andrew and Mayer, 1999).

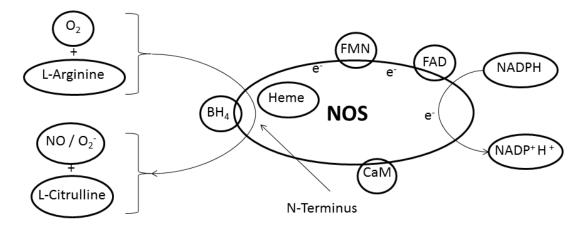


Figure 1: NO· production via NOS.

This graphical representation shows the movement of electrons through the NOS enzyme with its key co-factors. The net outcome produces NO·.

The second is the entero-salivary pathway (NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup> & NO· conversion); this pathway can act as a pool to provide additional NO·. This pathway functions with the body absorbing inorganic NO<sub>3</sub><sup>-</sup> from food and from NO<sub>3</sub><sup>-</sup> produced endogenously within the body. 60-80% of a western dietary intake of NO<sub>3</sub><sup>-</sup> is derived from vegetables (Ysart et al., 1999), with beetroot, leafy greens and cruciferous vegetables containing the highest concentrations of dietary NO<sub>3</sub><sup>-</sup>. The conditions in which plants is grown can have an impact on where the NO<sub>3</sub><sup>-</sup> is stored within the plant (Gilchrist et al., 2010). If the plant is grown in conditions with small amounts of light it will store the NO<sub>3</sub><sup>-</sup> as it cannot be converted to amino acids during photosynthesis. Beetroot store copious amounts of NO<sub>3</sub><sup>-</sup> in their roots as its main anion (Gilchrist et al., 2010).

NO<sub>3</sub><sup>-</sup> consumption through normal habitual dietary means has been described as part of a healthy balanced diet (Bryan and Hord, 2010). The average consumption of NO<sub>3</sub><sup>-</sup> per day is ~1-2 mmols in a typical western diet (ECETOC., 1988), of which the majority of the available NO<sub>3</sub><sup>-</sup> is quickly absorbed in the upper gastrointestinal tract (Florin et al., 1990, van Velzen et al., 2008). Approximately 60% of the NO<sub>3</sub><sup>-</sup> from our diets is excreted in our urine (Lundberg and Govoni, 2004) within 48 hours (Wagner et al., 1983). Of the available NO<sub>3</sub><sup>-</sup>

either produced endogenously or taken in from the diet, 25% passes into the entero-salivary circulation where it is concentrated by a magnitude of 10 fold in salivary glands (Lundberg and Weitzberg, 2010).

The entero-salivary pathway is reliant on a number of factors, principally the symbiotic relationship that humans share with facultative anaerobic bacteria that live on the surface of the tongue (Duncan et al., 1995). The salivary glands secrete high concentrations of NO<sub>3</sub><sup>-</sup> in our oral cavity. NO<sub>3</sub>- acts as an alternative electron acceptor (Duncan et al., 1995) where bacteria on the tongue reduces the NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup>. The NO<sub>2</sub><sup>-</sup> is swallowed and some of the NO<sub>2</sub><sup>-</sup> is further reduced to NO· in the acid environment of the stomach (Benjamin et al., 1994). Not all of the NO<sub>2</sub><sup>-</sup> is reduced to NO· in the stomach, some is absorbed into the circulating plasma where it can act as a storage pool for later reduction to NO· (Lundberg and Govoni, 2004).

This NO<sub>2</sub> to NO· reduction can also be catalysed by deoxyhaemoglobin, deoxymyoglobin, xanthine oxide, NOS, aldehyde oxidase, cytochrome P-450 and the mitochondrial electron transfer complexes (Cosby et al., 2003, Shiva et al., 2007, Zhang et al., 1998, Vanin et al., 2007, Li et al., 2008b, Kozlov et al., 2003, 1999). Other specific conditions can also expedite the conversion of NO<sub>2</sub> to NO· such as hypoxia (Castello et al., 2006) and acidic environments (Modin et al., 2001), which can occur during exercise (Bailey et al., 2009). This may be important for clinical populations, such as those with T2DM, who find day to day activities difficult, and have a reduced NOS derived NO· (Wu and Meininger, 2009). Lauer et al. (2008) used plasma NO<sub>2</sub> concentration change as a marker of NO· production. Lauer et al. (2008) suggest that a reduced concentration of NO<sub>2</sub> is associated with a reduced exercise tolerance.

# 1.1.2: The beneficial effects of NO<sub>3</sub>, NO<sub>2</sub> and NO:

Consuming a diet that is high in fruits and vegetables has been shown to lower blood pressure (Appel et al., 1997, Rouse et al., 1983). These diets have the ability to lower the risk of morbidity and mortality from cardiovascular disease (Bazzano et al., 2001, Joshipura et al., 1999, Joshipura et al., 2001). Due to large variation in the diet between individuals, it is difficult to elucidate which components decrease the risk of mortality and have cardio protective effects. Individuals who consume greater quantities of vegetables, such as those following the DASH diet (Appel et al., 1997), may take in up to 20 mmol of NO<sub>3</sub><sup>-1</sup> per day (Hord et al., 2009). Individuals who consume the greatest amounts of green leafy and cruciferous vegetables, which typically have high NO<sub>3</sub><sup>-1</sup> contents, have the largest protective effect from cardiovascular disease (Joshipura et al., 1999, Joshipura et al., 2001). More recently it has been shown that diets rich in green leafy vegetables may also protect against the risk of developing type 2 diabetes (T2DM) (Carter et al., 2010) and COPD (Hirayama et al., 2009, Watson et al., 2002).

## 1.1.3: Exercise efficiency & performance:

A basic premise of exercise physiology was that an individual's  $O_2$  requirement for an exercise bout could be accurately predicted if the work load was known (Jones and Poole, 2005).

The  $O_2$  cost of moderate intensity exercise is unaffected by having a disease such as T2DM, ageing, having a sedentary lifestyle and poor health status (Jones and Poole, 2005). Exercise training (in the short term) will not modulate the  $O_2$  cost of moderate intensity exercise (see Jones and Poole, 2005 for

review). It is, therefore, surprising that dietary supplementation with inorganic  $NO_3^-$  can elicit a reduction in the  $O_2$  cost of exercise.

A 3-5% reduction in the O<sub>2</sub> cost of exercise has been shown following NO<sub>3</sub><sup>-</sup> supplementation with beetroot juice (Bailey et al., 2009, Larsen et al., 2007b) and pharmacological sodium NO<sub>3</sub><sup>-</sup> (Larsen et al., 2010). Subsequent to these studies, a reduction in the O<sub>2</sub> cost of exercise following submaximal exercise and/or improvements in exercise performance following supplementation of beetroot juice or sodium NO<sub>3</sub><sup>-</sup> in recreationally active individuals have been shown in multiple studies (Bailey et al., 2009, Lansley et al., 2010, Larsen et al., 2011, Larsen et al., 2007b, Larsen et al., 2010, Vanhatalo et al., 2010, Bescos et al., 2011, Cermak et al., 2012b, Wylie et al., 2013a, Wylie et al., 2013b, Breese et al., 2013). It should be noted that some studies have not reported such positive effects (Christensen et al., 2013, Murphy et al., 2014, Martin et al., 2014, Fulford et al., 2013) (for more examples see review table 1 and 13 a,b and c).

The question that remains in the clinical context is: can this reduction in the O<sub>2</sub> cost of exercise be seen in populations that may benefit from improved exercise efficiency and elicit improvements in quality of life and activities of daily living? NO<sub>3</sub><sup>-</sup> supplementation in clinical populations is a novel concept and there is limited research published. At the commencement of this body of work only one study has been conducted with NO<sub>3</sub><sup>-</sup> supplementation in a clinical population. It demonstrated an increase in walking time, time to onset of claudication pain and improved muscle oxygenation in individuals with peripheral arterial disease (Kenjale et al., 2011). However, this study is very small (n=8) and was not

double blinded. Thus the robustness of the conclusions should be interpreted with care.

A recent review (Bailey et al., 2012) suggests that as athletes have elevated plasma NO<sub>3</sub> and NO<sub>2</sub> (Jungersten et al., 1997, Cuzzolin et al., 2002) concentrations compared to non-athletic controls, well-trained athletes may not gain the same benefits as recreationally active individuals following NO<sub>3</sub><sup>-</sup> supplementation. This perhaps indicates the existence of a ceiling limit, whereby further increases in the bioavailability of NO<sub>2</sub> may not have an additive effect on exercise performance. Other evidence exists to suggest that it is not a ceiling limit per se but it is more likely to have diminishing effects as the aerobic capacity of the individual increases. A recent study demonstrated that a cohort of well-trained cyclists over a 50 mile time trial did not cover the distance in a shorter time (Wilkerson et al., 2012). Interestingly, there was a significant correlation between a rise in plasma NO<sub>2</sub> concentration and time trial performance. Elite athletes tend to have a higher plasma NO<sub>2</sub> concentration (Wilkerson et al., 2012). This may diminish the ability of athletes to gain ergogenic effects from NO<sub>3</sub> supplementation. If this hypothesis is correct then individuals with reduced plasma NO<sub>3</sub> and NO<sub>2</sub> concentration such as individuals with T2DM (Vanhoutte, 2009) may stand to get the greatest benefits from NO<sub>3</sub> supplementation. Although Vanhoutte presents data suggesting individuals with T2DM have a reduced plasma NO<sub>3</sub> and NO<sub>2</sub> concentration the method used to assess this was the greiss method and is less sensitive at lower concentrations.

NO increases following exercise have been positively correlated to increases in cardiovascular health (Jungersten et al., 1997). There is an association

between the increase of endogenously produced NO<sub>2</sub> and an improvement in exercise tolerance (Dreißigacker et al., 2010) whilst an increase in plasma NO<sub>2</sub> concentration following beetroot juice has been correlated with improvements in time trial performance (Wilkerson et al., 2012). If an individual's tolerance to exercise improves then they have the potential to perform the same amount of exercise whilst feeling less physiological stress or do more work while feeling the same physiological stress as incurred previously at a lower level of exercise. Enabling people with chronic disease to exercise more, via reducing early onset fatigue, may lead to significant health and quality of life benefits. Exercise performance could improve in individuals with reduced plasma NO<sub>3</sub> and NO<sub>2</sub> by doing regular exercise via increases in endogenously produced NO. Whilst this is unlikely to have a large effect it could occur due to increases in vascular production of NO due to shear stress (Cooke et al., 1991, Miller and Burnett, 1992). As the individual exercises the subsequent increase in cardiac output will create shear stress producing more NO· (van Citters and Franklin, 1969). Given that individuals with reduced NO· bioavailability such as people with T2DM, and they have a lower  $\dot{V}O_2$  max (Regensteiner et al., 1995b, Regensteiner et al., 2009), NO<sub>3</sub> supplementation could enable individuals with disease to begin exercising at a lower VO<sub>2</sub>, thus exercise feels relatively easier and have the potentially to yield more endogenously derived NO. Another potential means of improving exercise tolerance would be to improve efficiency in a nonhaemodynamic process such as an improved mitochondrial efficiency or calcium handling.

# 1.1.4: NO· mechanisms for the reductions in the O<sub>2</sub> cost of exercise:

Multiple potential mechanisms for the reduction in O<sub>2</sub> cost of exercise following supplementation with inorganic NO<sub>3</sub><sup>-</sup> have been hypothesised. These include a reduced O<sub>2</sub> cost of mitochondrial ATP re-synthesis (Clerc et al., 2007, Basu et al., 2008) and or a reduction in the ATP turnover (Bailey et al., 2010b). A vasodilatory effect has also been postulated, allowing for a better coupling of blood flow to metabolic demand and finally an increase in the anaerobic energy production (Larsen et al., 2007b, Bailey et al., 2009). These mechanisms will be discussed below.

The reduction in the  $O_2$  cost of exercise has been shown in numerous studies with athletes, recreationally active individuals and groups of individuals with disease (See table 1 and 13). The reduction in the  $O_2$  cost of mitochondrial ATP synthesis may be brought about via two different mechanisms (Larsen et al., 2011). The first is that  $NO_2$  could act as an alternative electron acceptor via the inhibition of cytochrome-c (Basu et al., 2008). Cytochrome-c is the last enzyme in the electron transport chain. NO could become a surrogate for  $O_2$  at cytochrome-c's binding site, replacing the need for  $O_2$  during ATP production (Brown and Cooper, 1994). This, *in vitro* study appears to show a near linear relationship between increasing NO concentration (via light releasing nitroprusside) and decreasing  $O_2$  consumption of brain synaptosomes. At 60nM NO concentration and  $30\mu M$   $O_2$  concentration half of the cytochrome-c binding sites were inhibited (Brown and Cooper, 1994). This is of great importance given the difficulties of  $O_2$  delivery in many clinical populations.

The second proposed mechanism involves an increase in the phosphate -  $O_2$  (P/O) ratio. The term P/O ratio was coined by Hinkle (2005). The P/O ratio is

used as a measure of mitochondrial oxidative phosphorylation efficiency. The P/O ratio is the term given for the number of ATP molecules that can be produced for an molecule of  $O_2$ . If the ratio is relatively higher, then more ATP can be produced for the same amount of  $O_2$ . The reduction in the  $O_2$  cost is thought to have been brought about via a reduced slippage of protons across the mitochondrial proton pumps essentially reducing energy waste and increasing oxidative phosphorylation. This is thought to occur due to one of two reasons; firstly there is an increase in the efficiency of the electron transport chain and secondly a decrease in the proton gradient consuming process (Clerc et al., 2007). P-magnetic resonance spectrometry has been used to access the turnover rate of ATP. Bailey et al., measured phosphocreatine, ADP and inorganic phosphate in addition to pH. Bailey et al., showed that with  $NO_3$  supplementation you can significantly reduce the estimated turnover rate of ATP (essentially reduce the  $O_2$  cost of exercise) (Bailey et al., 2010b).

NO· is a potent vasodilator. The potential for NO· to vasodilate hypoxic and anoxic areas to improve oxygenation is a theory often referred to in NO· related blood pressure lowering effects (Webb et al., 2008b, Gautier et al., 2006). NO· production via eNOS is important for maintaining vascular tone. If the microvasculature has a greater flux of oxygenated blood, it is possible that a greater matching of O₂ delivery to O₂ demand within the working muscles may occur. However, this may be impaired in individuals with O₂ delivery deficiencies such as individuals with T2DM and chronic obstructive pulmonary disease (COPD). Moreover, if there is a greater matching of O₂ to the working muscles then uptake and utilisation may increase. Therefore, O₂ delivery is an unlikely source of reduction in steady state O₂ consumption but may mediate increased performance.

Given the reduced O<sub>2</sub> cost of exercise another potential mechanism for this improvement in efficiency could be an increase, proportionally, by the anaerobic contribution. This is unlikely based on two studies showing no increase in blood lactate levels measured via capillary blood sampling (Larsen et al., 2007b, Bailey et al., 2009) post NO<sub>3</sub><sup>-</sup> supplementation. Bailey et al., (2012) showed via p-magnetic resonance spectrometry that the pH was unchanged during moderate and severe exercise, further suggesting that the reduction in the O2 cost of exercise is not due to an increase in anaerobic contribution to ATP production.

With regards to the reduction in the O<sub>2</sub> cost of exercise, the evidence suggests that NO acts as a surrogate for O<sub>2</sub> in the electron transport chain (Basu et al., 2008, Brown and Cooper, 1994) in conjunction with an increase in the P/O ratio via mitochondrial protein changes (Larsen et al., 2011, Clerc et al., 2007). Larsen's 2011 protocol utilised muscle biopsies harvested from healthy young individuals. The primary skeletal muscle cell cultures were conducted in ambient conditions. This would place the cells in an artificially elevated O<sub>2</sub> rich environment which would cause oxidative stress. In order to assess the O2 consumption relative to the muscle the O2 concentration could be lowered to ~3% which would match the pO<sub>2</sub> at the muscle. This makes the findings within this study questionable as the utilisation of NO· in different redox balances is likely to be vastly different. The limitations of these data supporting a reduction of proton slippage at the proton pumps and improvements of efficiency via the P/O must be borne in mind when seeking to understand the mechanism behind the reduction in O<sub>2</sub> cost of exercise. However, the data shows the potential for improvements in efficiency and more research is warranted in this area.

#### 1.1.5: NO., cGMP and sGC in vascular function:

NO· is a small size molecule that carries no charge. Due to this, it can diffuse easily across membranes without the need for a transport system, removing a rate limiting step (Lancaster, 1996, Lancaster, 1994, Denicola et al., 1996). NO· is relatively 'unreactive' and moves freely from the cell in which it was created to a target cell prior to being utilised (Denninger and Marletta, 1999). Within plasma the half-life of NO· is thought to be very short ~ 3 seconds (Yoshizumi et al., 1993, Lundberg and Govoni, 2004); NO₂⁻ is ~ 110 seconds (Kelm, 1999) and NO₃⁻ is ~ 5 hours (McKnight et al., 1997).

The bioavailability of NO· is paramount for normal cellular function and in particular, smooth muscle relaxation and signalling transduction. Many actions of NO· mediated vascular function are produced via cyclic guanosine monophosphate (cGMP) (Alzawahra et al., 2008). cGMP is created by the reduction of guanosine triphosphate (GTP) which is catalysed by guanylate cyclase (GC) (Denninger and Marletta, 1999). GC can be soluble (sGC) or attached to a receptor on the cell membrane. NO· increases GC activity (Schultz et al., 1977, Arnold et al., 1977) and subsequently cGMP concentrations (Denninger and Marletta, 1999). The elevations in cGMP lead to the phosphorylation of protein kinase C activity which phosphorylates proteins and decreases intracellular levels of Ca<sup>2+</sup> via compartmentalisation or inhibits its release (Lohmann et al., 1997). This reduction leads to relaxation of vascular smooth muscle which ultimately lowers blood pressure (Lohmann et al., 1997).

#### 1.1.6: Entero-salivary pathway and its role in blood pressure regulation:

The importance of reduction of NO<sub>3</sub> to NO<sub>2</sub> in the entero-salivary pathway was initially tested by having participants refrain from swallowing their own saliva

(Webb et al., 2008a) or by spitting post NO<sub>3</sub> supplementation (Lundberg and Govoni, 2004). These studies suggest that by preventing the swallowing of NO<sub>2</sub>, the beneficial effects of NO<sub>3</sub> supplementation are removed due to interruption of the pathway. Recently, it has been shown by limiting the amount of microflora in the oral cavity (via the use of antibacterial mouthwash) the circulating plasma NO<sub>2</sub> concentration is reduced without any decrease in the plasma NO<sub>3</sub> concentration (Kapil et al., 2012; Petersson et al., 2009) again providing evidence that oral microflora are a key step in the entero-salivary pathway (Spiegelhalder et al., 1976, Lundberg et al., 2004, Duncan et al., 1995). Consequently, the use of antibacterial mouthwash has also been shown to increase the blood pressure of the participants involved (Kapil et al., 2012, Petersson et al., 2009) (shown in human and animal models), therefore, suggesting that antibacterial mouthwash may attenuate the NO stimulated blood pressure reducing effects. Individuals with elevated blood pressure could potentially have raised systolic and diastolic pressures due to a depletion in NO. (either due to increased scavenging, reduced NOS production, poor diet and or other failures in the entero-salivary pathway), as opposed to earlier hypotheses that a high blood pressure causes a reduced NO level (Rockey and Chung, 1998).

The role of NO<sub>3</sub><sup>-</sup> and its subsequent conversion to NO· has many beneficial effects. Previously, Lundberg et al., (2006) hypothesised that NO<sub>3</sub><sup>-</sup> rich vegetables may play a key role in blood pressure reduction. Subsequently, there have been numerous reports of a reduction in blood pressure, either with sodium NO<sub>3</sub><sup>-</sup> (Larsen et al., 2006) or beetroot juice (Bailey et al., 2010b, Lansley et al., 2010).

At the start of this project the only study to examine a clinical population with NO<sub>3</sub><sup>-</sup> supplementation was in individuals with peripheral artery disease which found a statistically significant reduction in diastolic blood pressure (BP) (7 mmHg) but no change in systolic BP (Kenjale et al., 2011). For more details on blood pressure changes in NO<sub>3</sub><sup>-</sup> supplementation studies see table 1 and 13.

Much of the early literature (and a number of more recent studies) used a placebo that is not a 'true' placebo, i.e. blackcurrant juice or prune/orange juice. Whilst the increase in exercise time or reductions in the O<sub>2</sub> cost of exercise in these studies are of interest, there are significant limitations. By utilising fruit juice they are likely to have a substantially different antioxidant content which could alter NO· bioavailability (Bondonno et al., 2012). This could therefore explain some of the variability in these studies. Moreover, related to the lack of a 'true' placebo where the participant did not know whether active or placebo juice was being taken, the widely reported beneficial effects of beetroot juice [in the lay press] on exercise performance/tolerance may have given rise to a "placebo" effect in those informed volunteers taking beetroot juice. For example the Daily Mail online published an article stating beetroot juice 'helps lower blood pressure by 7%'. On the 29<sup>th</sup> of June 2010 the Daily Mail stated that beetroot juice 'could save your life' (DailyMail, 2013).

Beetroot is known to contain a number of antioxidants within the juice and it was an important stage in  $NO_3^-$  research to determine if antioxidants could reduce the  $O_2$  cost of exercise. Bailey (2011) found that supplementing with N-acetylcysteine (an antioxidant) had no changes in the  $O_2$  cost of sub maximal exercise. There is, however, some evidence to suggest that antioxidants and polyphenols can reduce  $NO_2^-$  to  $NO_1^-$  (Gago et al., 2007, Carlsson et al., 2001)

albeit in the stomach and urine. Beetroot juice has a variety of antioxidants (see experimental chapter 1 for antioxidant content of beetroot juice). Lansley et al. (2010) removed the  $NO_3^-$  content from the beetroot juice to act as a placebo. This was performed via an ion exchange resin. They subsequently supplemented their participants with the active and placebo juices. These results provided evidence to suggest that in young healthy individuals it is  $NO_3^-$  that is the active ingredient as there was no reduction in the  $O_2$  cost of exercise with the  $NO_3^-$  depleted juice.

Table 1. A summary of dietary  $NO_3^-$  supplementation and its effects on plasma  $NO_3^-/NO_2^-$  concentrations, BP and exercise (2007 - 2012).

Author	Participants & Protocol	NO <sub>3</sub> <sup>-</sup> supplement	Plasma NO <sub>3</sub> & NO <sub>2</sub> concentration (BR vs. PL respectively)	BP changes	Exercise efficiency or performance and/or clinical relevance	Primary outcome successful?
Larsen et al. (2007b)	9 healthy well-trained males. Min 10 day washout. 5 mins cycling at 45, 60, 70, 80, 85 and 100% of VO <sub>2</sub> max	3 days supplementation of NaNO <sub>3</sub> <sup>-</sup> (0.1 mmol Kg <sup>-1</sup> day <sup>-1</sup> )	$NO_3^- = 182 \pm 55$ and $27 \pm 6.9 \mu M$ $NO_2^- = 226 \pm 87$ and $124 \pm 28 n M$ .	↓SBP ↓DBP	↓VO <sub>2</sub> over 4 lowest WR  ↑ average GE over 4 lowest WR  ↑ average DE over 4 lowest WR	Yes
Webb et al. (2008a)	14 healthy subjects (10 in phase two), min 7 day washout period. 3BP measurements taken every 15mins for 1 hour.	500ml BR juice (NO <sub>3</sub> = 45 mmol/L) or water	16 fold ↑ in plasma NO₃⁻  2 fold ↑ in plasma NO₂⁻  (Values not given)	2.5h peak ↓SBP 3h peak ↓ DBP and MAP	IR of the forearm slowed the FMD and protected against IR injury (phase two).	Yes
Bailey et al. (2009)	8 healthy males. With a min of a 10 day washout period. 3BP measurements taken at the 6 <sup>th</sup> supplementation point. 4 MI and 2 SI bouts on days 4, 5& 6.	500ml/day of BR juice (NO <sub>3</sub> <sup>-</sup> =5.5mmol/day) or black current cordial for 6 days	NO <sub>2</sub> = 273 ± 44 and 140 ± 50 nM	↓ SBP	<ul> <li>↓ VO₂ amplitude in MI</li> <li>↓ VO₂ slow component in SI</li> <li>↑ time to exhaustion in SI</li> <li>↓ deoxyhaemoglobin</li> <li>↑ haemoglobin</li> <li>↑ total haemoglobin</li> </ul>	Yes

Bailey et al. (2010b)	7 healthy makes. With a min of a 10 day washout period. 3BP measurements taken at the 6 <sup>th</sup> supplementation point. Metabolic milieu measured via P-MRS.	500ml/day of BR juice (NO <sub>3</sub> <sup>-</sup> =5.1mmol/day) or black current cordial for 6 days	NO <sub>2</sub> <sup>-</sup> = 643 ± 110 and 206 ± 93nM	↓SBP ↓DBP ↓MAP	↓ VO₂ amplitude in LI  ↓ VO₂ slow component in HI  ↑ time to exhaustion in SI  ↓ muscle ATP turnover rate  ↓ muscle ADP accumulation  ↓ muscle Pi accumulation  ↓ muscle PCr depletion	Yes
Vanhatalo et al. (2010)	8 healthy physically active subjects. With a min of a 10 day washout period. 4BP measurements were taken on each visit. Protocol consisted of a ramp and MI exercise.	500ml/day of BR juice (NO <sub>3</sub> =5.2mmol) or juice cordial for an acute dose (2.5h) and chronic exposure (5 and 15 days)	NO <sub>2</sub> = 454 ± 81 nM at baseline and increased by 39% (post 2.5h), 25% (after 5 days) and 46% after 15days.	↓SBP ↓MAP For all conditions	For acute and chronic supplementations.  ↓ $\dot{V}O_2$ amplitude in MI  After 15 days only:  † $\dot{V}O_2$ max  †peak work rate during the ramp test  †GET & WR	Yes

Larsen et al. (2010)	Nine (7 male) healthy subjects. With a min of a 7 day washout period. 2 BP measurements were taken on the final visit.	Acute (1h prior to exercise) & Chronic exposure; 2 days NaNO <sub>3</sub> <sup>-</sup> supplementation (0.1 mmol Kg <sup>-1</sup> day <sup>-1</sup> ) or placebo which was equally dosed NaCl.	$NO_2^- = 142 \pm 35$ and $61 \pm 11$ nM $NO_3^- = 230 \pm 31$ and $17.3 \pm 3$ µM	No change in resting SBP & DBP. ↓DBP post ex.	Acute ingestion;  VO₂ during LI cycle exercise  Chronic ingestion;  ↑ cGMP  ↑VO₂max	Yes
Vanhatalo et al. (2011)	9 healthy (7 male) physically active participants. BP was measured prior to exercise commencement after breathing normoxic or hypoxic air for 15 minutes.	750ml/day of BR juice (NO <sub>3</sub> =9.3mmol) or PL (NO <sub>3</sub> =0.006mmol). Juice was consumed 24, 12 and 2.5h prior to exercise.	NO <sub>2</sub> <sup>-</sup> = 194 ± 51 and 129 ± 23 nM	↓SBP ↓DBP ↓MAP	↑PCr recovery $\downarrow S_{aO2}$ in BR and PL compared to control. $\downarrow T_{lim}$ PL compared to control $\leftrightarrow T_{lim}$ BR to control	No
Larsen et al. (2011)	Mitochondria harvested from healthy volunteers post supplementation.	3 days NaNO <sub>3</sub> <sup>-</sup> supplementation (0.1 mmol Kg <sup>-1</sup> day <sup>-1</sup> )	$NO_2^- = 163 \pm 29$ and $35 \pm 7$ nM $NO_3^- = 169 \pm 18$ and $27 \pm 2.6$ µM	N/A	↑ Mitochondrial P/O ratio by 19%, suggesting a higher max ATP turnover which increased by 23% ↓ vO₂ in LI exercise	Yes

Lansley et al. (2011b)	9 healthy active males. With a min of a 10 day washout period. 4 BP measurements were taken on each visit to the lab (10 occasions)	500ml/day of BR juice (NO <sub>3</sub> =6.2mmol/day) or PL(NO <sub>3</sub> =0.003mmol/day) for 6 days	$NO_2^- = 373 \pm 211$ and $183 \pm 119$ nM	↓SBP	<ul> <li>↓ VO₂ whilst walking</li> <li>↓ VO₂ amplitude in MI</li> <li>↓ end exercise VO₂ in SI</li> <li>↑ time to exhaustion in SI</li> </ul>	Yes
Lansley et al. (2011a)	9 healthy male club level cyclists. With a min of a 48h washout period. Exercise involved a 4 & 16.1km TT.	500ml/day of BR juice (NO <sub>3</sub> <sup>-</sup> =6.2mmol) or PL(NO <sub>3</sub> <sup>-</sup> =0.005mmol/day) 2.5h prior to exercise	$NO_2^- = 575 \pm 199$ and 241 ± 125 nM	↓SBP	↑ TT performance (time=↓) ↑PO in TT	Yes
Kenjale et al. (2011)	8 (4 males) PAD patients. With a 7 – 14 day washout period. 1 BP measurement was taken prior to the intervention and 2h post juice. ABI and endothelium function were assessed.	500ml/day of BR juice (NO <sub>3</sub> <sup>-</sup> =18,181 umol/l) or PL (orange juice) 3h prior to exercise.	NO <sub>2</sub> = 943 ± 826 and 152 ± 72 nM	↓DBP	<ul> <li>↓ time to onset of claudication</li> <li>↑ walking distance</li> <li>↓ deoxyhaemoglobin</li> <li>↑ haemoglobin</li> <li>↑ total haemoglobin</li> </ul>	Yes
Bescós et al. (2011)	11 healthy participants. With a 7 day washout period.	NaNO <sub>3</sub> supplementation (10 mg.kg <sup>-1</sup> ) or equivalent dosed PL (NaCl).	NO <sub>2</sub> <sup>-</sup> = 2.3 $\mu$ M ± 157 nM and 2 $\mu$ M ± 206 nM NO <sub>3</sub> <sup>-</sup> = 250 ± 80 and 29 ± 8 $\mu$ M	N/A	↓VO₂peak without affected TTE or max power.	Yes

Cermak et al. (2012a)	12 active male cyclists. With a 14 day washout period. Exercise involved a 60min sub max cycling followed by a10-km TT.	500ml/day of BR juice (NO <sub>3</sub> =8mmol/day) or PL (NO <sub>3</sub> depleted juice) for 6 days.	Abstract only, figures not presented.	N/A	↓ TT time  ↑Power output  ↓ MI ऐO₂	Yes
Cermak et al. (2012b)	20 well-trained males. Exercise included a ~1h TT.	140ml BR 2.5h prior to exercise (NO <sub>3</sub> = 8.7 mmol).	$NO_2^- = 532 \pm 32$ and 271 ± 13 nM	N/A	<ul><li>→ TT time</li><li>→ Power output</li></ul>	No

↑ = statistical significant increase;  $\leftrightarrow$  = no statistical reduction;  $\downarrow$  = statistical significant reduction;  $\dot{V}O_2$  = oxygen uptake;  $\dot{V}O_{2peak}$  = peak oxygen uptake; WR = work rate; GE = gross efficiency; DE = delta efficiency; SBP = systolic blood pressure, DBP = diastolic blood pressure; MAP = mean arterial blood pressure; IR = ischemic reperfusion, FMD = flow-mediated dilation; MI = moderate intensity; RPE = rating of perceived exertion; SI = severe intensity;  $\dot{V}O_{2max}$  = maximal oxygen uptake; LI = low intensity; HI = high intensity; PCr = phosphocreatine; Pi = inorganic phosphate; ADP = adenosine diphosphate; ATP = adenosine triphosphate; GET = gas exchange threshold; PAD = peripheral arterial disease; cGMP = cyclic guanosine monophosphate;  $S_{aO2}$  = arterial  $O_2$  saturation; TIm = limit of tolerance; P/O ratio = phosphorylation efficiency; PO = power output; TT = time trial; TTE = time to exhaustion; LNF = low nitrate food; HNF = high nitrate food; PO<sub>2</sub> = partial pressure of oxygen; DMBA = dimethoxybenzaldehyde (a powerful organ-specific laboratory carcinogen); NIRS = near infrared spectrometry and sGC = Soluble guanylate cyclase; RQ = respiratory quotation; MVC = maximal voluntary contraction. Where NO<sub>3</sub> & NO<sub>2</sub> values are given it is from the final supplementation period if multiple doses given.

From 2008 until the start of this body of work, 20 papers were published in the field of NO<sub>3</sub> supplementation (regardless of the primary outcome). Variability of NO<sub>3</sub> dose between studies must be considered when comparing results from different trials. Webb et al., (2008a) administered ~22.5 mmol of NO<sub>3</sub><sup>-</sup> in a single 500ml bolus. Many studies have supplementation regimens between ~4.5 and 6.5 mmol (see table 1 above for published values). These differences are also seen in plasma NO<sub>3</sub><sup>-</sup> concentration values, which can range from 17.3 to  $27\mu M$  (23.8  $\pm$  5.6  $\mu M$ ) after placebo juice supplementation compared with the active juice ranging from 169 to 230 $\mu$ M (194 ± 32  $\mu$ M). Plasma NO<sub>2</sub> concentrations have greater variability between studies with concentrations following placebo juice supplementation from 29nM to 2μM (326 ± 524 nM) and concentrations following active juice supplementation ranging from 142nM to 2.3µM (543 ± 575 nM). A minority of published papers have reported plasma NO<sub>2</sub> concentration in the µM range with a minimal increase post supplementation (Bescos et al., 2011). The majority of the inorganic NO<sub>3</sub> supplementation studies have used chemiluminescence to determine NO<sub>3</sub> and NO<sub>2</sub> concentrations. Unsupplemented or placebo plasma NO<sub>2</sub> concentrations are predominantly in the nM range. The measurement of NO<sub>2</sub> poses significant challenges with substantial environmental contamination and plasma values that are often approaching the limits of detection of most assays. Furthermore there are a range of assay techniques available, with varying sensitivities. When this study is removed (Bescos et al., 2011) from the NO<sub>2</sub> and NO<sub>3</sub> data there is still a ~10 fold difference between the largest and the smallest baseline plasma NO2 values which some researchers would argue is due to training status. It is unlikely that the variation in reported plasma NO<sub>2</sub> values is entirely due to genuine biological variation and dietary intake. Dietary intake accounts for ~1 - 2 mmol of NO<sub>3</sub> per day

(ECETOC., 1988) and would therefore not elicit these variations with much less than this being derived from the L-arginine pathway. Assay variation remains a likely contributor to these large variations. A consensus within and between groups would help to establish norms within populations.

Variation in length of supplementation of NO<sub>3</sub><sup>-</sup> ranges from 2.5 hours to 15 days with variation in washout periods for acute and chronic supplementation ranging from 2.5 days to 28 days. For supplementation periods that do not have time to elicit protein expression within the mitochondria (i.e. < 3 days) (Larsen et al., 2011) the washout period is less important. Conversely, longer washout periods are needed for longer supplementation periods. Research examining the length of RNA expression changes following supplementation withdrawal would elucidate how long it would take to lose these mitochondrial adaptations.

### 1.1.7: Endothelial dysfunction and red blood cells:

Endothelial function and the blood pressure lowering effects of NO· have been closely linked in previous research studies. It has been hypothesised that eNOS could act as another NO<sub>2</sub><sup>-</sup> reductase in areas where it may be needed, such as hypoxia (Webb et al., 2008b) and anoxia (Gautier et al., 2006). Plasma NO<sub>2</sub><sup>-</sup> concentrations have been shown to mirror endothelial dysfunction in humans (Kleinbongard et al., 2006), with dysfunction in endothelial function and reduced bioavailability of NO· being closely linked to cardiovascular disease (Forte et al., 1997, Vanhoutte, 2009). This potentially could be the reason for poor control of vascular tone (Forte et al., 1997).

Endothelial dysfunction is a systemic disease of the vascular endothelium which is brought about via a disparity between vasodilation and vasoconstricting substances. This endothelial dysfunction is associated with T2DM and has been strongly linked to a reduced bioavailability of NO· (Calver et al., 1992, Cohen, 1993, Cockcroft et al., 2000). This is likely to be underpinned by multiple factors; firstly, an impairment of NOS (Ito et al., 2002, Huynh and Tayek, 2002) to create NO· and secondly, increased inactivation of NO· by oxidants or reactive O<sub>2</sub> species (Vallejo et al., 2000, Angulo et al., 1996). A third possibility includes scavenging of NO· in the vascular lumen which can be expedited by four key proteins such as, oxyhaemoglobin (to form methemoglobin and NO<sub>3</sub><sup>-</sup>), deoxyhaemoglobin (to create nitrosyl haemoglobin), O<sub>2</sub> and thiols (creating *S*-nitrosohaemoglobin) (Kharitonov et al., 1995, Gow et al., 1999, Gow and Stamler, 1998, Lancaster, 1996).

Nitrosyl-haemoglobin levels are higher in individuals with type 1 diabetes compared to healthy controls. It would appear that higher concentrations of glycated haemoglobin will reduce the bioavailability of NO· (Milsom et al., 2002). S-nitrosohaemoglobin related dilation is inversely related to oxygenation status (James et al., 2004). However, the deoxygenation of haemoglobin has been proposed as a mechanism of NO· delivery (Jia et al., 1996). NO· delivered via red blood cells (RBC) has been suggested to play a role in vasodilation in healthy participants under hypoxic conditions and contributes to a role in healthy physiology (McMahon and Stamler, 1999). If these metabolites do act as a NO· storage pool then they could be key for NO· mediated vasodilation in conditions such as acidosis or hypoxia (James et al., 2004) which are particularly prevalent conditions in skeletal muscle of individuals with T2DM and the respiratory muscles of individuals with chronic obstructive pulmonary disease (COPD). However, more NO· is bound to glycated haemoglobin (HbA1c) which appears to reduce the vasodilatory functions of red blood cells (James et al., 2004) and thus individuals with T2DM will retain more NO·

than non-diabetic controls. This RBC derived NO· delivery appears to be expedited under pathological conditions such as ischemia reperfusion injury (Duranski et al., 2005), pulmonary hypertension (Hsu et al., 2007), atherosclerosis (Kim-Shapiro et al., 2006) and cerebral vasospasm (Pluta et al., 2005).

Blood vessel relaxation can be induced by insulin and is mediated at least in part via nitric oxide (Scherrer et al., 1994). Vascular function is impaired in T2DM (Steinberg et al., 1996). However, the perturbations in nitric oxide production and metabolism in T2DM and consequent modulation of vascular health are currently not clear. Some investigators suggest basal nitric oxide production is reduced in individuals with T2DM compared with healthy controls (Woodman et al., 2006, Scherrer and Sartori, 2000, Tessari et al., 2010). This could be a result of the increased expression of the intracellular inhibitor of endothelial nitric oxide synthase (García-Cardeña et al., 1997), caveolin-1 in the adipose tissue of individuals with T2DM (Catalán et al., 2008), elevated levels of NOS inhibitors such as asymmetric dimethylarginine (ADMA) (Abbasi et al., 2001) or BH<sub>4</sub> oxidation. Other groups report elevated levels of oxidised products of nitric oxide (Maejima et al., 2001, Chien et al., 2005, Ghasemi et al., 2010) with associations with specific genetic polymorphisms of eNOS noted (Monti et al., 2003). Together these reports would suggest T2DM is accompanied by either impaired production of NO, rapid scavenging of nitric oxide before it can cause downstream changes or a failure of activation of secondary messengers such as cGMP which are known to mediate NO's vascular effects (Young and Leighton, 1998).

### 1.1.8: NO· hepatic glucose uptake and T2DM:

Following a meal, glucose along with other carbohydrates and dietary constituents gets absorbed from the gastrointestinal tract. These nutrients are taken directly to the liver by the portal blood vessels allowing the liver to control the levels of nutrients like glucose in the blood. Glucose is the predominant energy source for mammalian cells and these cells require a steady supply of glucose to maintain normal function. Blood glucose regulation is maintained by numerous actions including: the rate of glucose consumption, carbohydrate uptake from the intestine, skeletal muscle glucose uptake and utilisation, and the rate of uptake/release via the kidney (Nordlie and Arion, 1964) and via the generation of endogenous glucose when required. Under normal physiological conditions glucose does not accumulate in the plasma. The surplus glucose gets metabolised, stored as glycogen (glycogenesis) predominantly in the liver) or converted to fat. Following ingestion of food, portal vein diameter widens increasing blood flow to the liver (Pazahr et al., 2014). This process may expedite glucose disposal.

Incretins, such as glucagon like peptide (GLP-1) and Glucose dependent insulinotropic peptide (GIP), are a group of gastrointestinal hormones that are released into the blood stream from cells in the small intestine in response to ingestion of food (Holst, 2007). They are predominantly involved in the regulation of blood glucose levels via their ability to have an insulinotropic (i.e. they promote insulin secretion) effect essentially augmenting the glucose-induced insulin secretion (known as the incretin effect). Other functions of incretins include regulation of gut motility, gall bladder constriction, nutrient absorption and secretion of gastric acids and pancreatic enzymes (Drucker, 2006). Glucagon-like peptide receptors (GLP-1Rs) have been found in many cardiovascular tissues including vascular and

coronary endothelium smooth muscle, cardiomyocytes and endocardium (Wei and Mojsov, 1995, Bullock et al., 1996). A diverse range of beneficial cardiovascular effects have been shown by multiple groups which include: regulation of blood pressure and heart rate, vascular tone and myocardial contractility (Barragán et al., 1996, Vila Petroff et al., 2001, Yamamoto et al., 2002, Green et al., 2008). Importantly these beneficial effects have been shown in healthy adults and also individuals with T2DM (Nikolaidis et al., 2004a, Nystrom et al., 2004, Özyazgan et al., 2005, Sokos et al., 2007).

L-cells present in the intestines secrete the biologically active GLP-1 (7-37), GLP-1 (7-36 amide) and GLP-2 (Baggio and Drucker, 2007) into the microcirculation to which the cells are closely associated. GLP-1 is known to slow gastric emptying (Meier et al., 2006) which reduces feelings of hunger (Zander et al., 2002) and in combination with receptors within the brain, GLP-1 creates a feeling of satiety (Goke et al., 1995) which have been shown to improve weight loss in individuals with T2DM. Other beneficial effects have been shown such as increased myocardial and endothelial function (note, in cardiac disease patients) (Basu et al., 2007, Nikolaidis et al., 2004a, Nikolaidis et al., 2004b).

A consensus within the literature has emerged that individuals with T2DM have an impaired incretin response compared to healthy controls (Nauck et al., 2011). This appears to be due to both reduced GLP-1 secretion and a resistance to GIP. The impaired incretin response increases as T2DM progresses (Nauck et al., 2011). GLP-1 has been shown to elicit myocardial glucose uptake (Bhashyam et al., 2010) and increases skeletal muscle microvascular blood flow, glucose uptake, nitric oxide production and insulin clearance mediated via nitric oxide-dependant mechanisms

(Chai et al., 2012). Specifically, GLP-1 analogues in animal models (dog) have also been shown to up-regulate the production of the vasodilator NO· (Post et al., 2010) whilst GIP has been shown to stimulate NO· production from eNOS in the portal vein but not in the hepatic artery (Ding et al., 2004). The portal vein is of paramount importance for glucose disposal; it collates tributaries from the stomach, proximal small intestines and the spleen and directs nutrient rich blood towards the hepatic vasculature / parenchyma for processing.

NO mediated vasodilation facilitates insulin stimulated glucose uptake into skeletal muscle (measured via an L-NAME NO· blockade) (Roy et al., 1998). In endothelial cells insulin promotes nitric oxide synthesis (via eNOS) leading to activation of GLUT4 translocation and increased glucose uptake into the cell (Jiang et al., 2014). Therefore, increasing NO concentration may increase glucose uptake into cells via both activation of GLUT's translocation and/or via vasodilation. As described above dietary NO<sub>3</sub> is converted to NO<sub>2</sub> which is rapidly absorbed in the stomach and proximal small intestine. NO2, absorbed by the stomach and the proximal small intestine will be delivered via the portal vein along with other micro and macronutrients to the stomach. Therefore the highest plasma concentrations of NO<sub>2</sub><sup>-</sup> in the body will occur in the portal circulation. Nitrite may play a vital role in glucose homeostasis following a meal either by direct signalling as a vasodilator or following conversion to NO· (Petersson et al., 2007). The portal vein is partially deoxygenated. Deoxyhaemoglobin is a NO<sub>2</sub> reductase (Cosby et al., 2003) and thus there may be an increase in the bioavailability of NO. As the elevated plasma NO2 concentrations reach the liver, many more NO2 reductases are present such as neuroglobin, cytoglobin, xanthine oxide and aldehyde oxidase which may increase the bioavailability of NO·(Martin et al., 2004, Li et al., 2008b).

Over recent years experimental evidence has shown that NO· may be key for glucose uptake (particularly in animal models) with skeletal muscle and in the intestines. In 2002 Kingwell et al, used a NOS inhibitor to assess the role of glucose uptake with skeletal muscle of an individuals with T2DM. They present a 13% drop in glucose uptake (Kingwell et al., 2002). One year later Guan et al., infused GLP-2 into neonatal piglets with an without a NOS inhibitor and found a marked reduction in glucose uptake with L-NAME (Guan et al., 2003). L-NMMA has also be shown to reduce glucose uptake in skeletal muscle in a mouse model (Merry et al., 2010). Another animal model used eNOS deficient mice, supplemented half the mice with nitrate rich water for 10 weeks. Carlstrom et al., demonstrate reduction in HbA1c, baseline plasma glucose concentrations and post prandial glucose concentrations and reduction in weight and circulating triglycerides (Carlström et al., 2010). Together these animal models suggest that NO may be a mediator of glucose uptake. Potentially via an increase in GLUT-4 translocation (Bedard et al., 1997) or increased perfusion with the parenchyma. One study has also shown that nitrate supplementation in healthy young individuals increases glucose uptake compared to placebo during exercise (Wylie et al., 2013b).

An intervention that can increase the bioavailability of NO· within the liver (a major site for glucose disposal) may help expedite reductions in postprandial plasma glucose concentrations.

# 1.1.9: eNOS coupling

Uncoupling is a term used to describe the physiological process where eNOS no longer produces NO but creates  $O_2$  (Li and Forstermann, 2013). Coupled and uncoupled NOS are commonly present in the same cell (Li and Forstermann, 2013).

As the coupled NOS produces NO $\cdot$  and the uncoupled NOS produces  $O_2$ , the potential for an increase in peroxynitrite concentration and subsequently a change in redox balance increases (Zou et al., 2004).

eNOS is mainly found in the endothelium but also in the cardiomyocytes (Wei et al., 1996), airway endothelium (Giaid and Saleh, 1995), and the tubular cells of the kidneys (Tracey et al., 1994). The eNOS N-terminus (i.e. the start of a polypeptide) contains the oxygenase aspect of the NOS, BH<sub>4</sub>, heam and also the L-arginine binding sites.

L-arginine and BH<sub>4</sub> stabilise the eNOS protein when in its dimeric form (non-covalent bonded macromolecule of two similar sub-units (Reif et al., 1999, Venema et al., 1997)). Zinc thiolate is key in maintaining the BH<sub>4</sub> binding site and maintains its structure in a dimeric form (Raman et al., 1998). Therefore maintaining BH<sub>4</sub> and zinc thiolate is key to prevent uncoupling and an increase in oxidative stress. Guanosine triphosphate (GTP) is converted to BH<sub>4</sub> via GTP cyclohydrolase (GTPCH1). Under elevated conditions of oxidative stress BH<sub>4</sub> is converted to dihydropterin (BH<sub>2</sub>) which ultimately leads to a perpetual increase in NOS uncoupling. O<sub>2</sub><sup>-</sup> and NO· combine to produce peroxynitrite (OONO<sup>-</sup>), which is a powerful free radical. This is a reversible reaction which can be catalysed by dihydrofolate reductase (DHFR) (Crabtree et al., 2011). Individuals with T2DM are in a pro-oxidant state, which is likely to lead to uncoupled NOS and in particular eNOS (Siasos et al., 2007) which will generate O<sub>2</sub><sup>-</sup>. NO· can react with O<sub>2</sub><sup>-</sup> which will create OONO<sub>2</sub><sup>-</sup>. This will reduce the bioavailability of NO· (Beckman and Koppenol, 1996).

Methylation of L-arginine naturally occurs in the cytoplasm of any normal functioning cell. Methylation of L-arginine can create one of three distinct derivatives of this amino acid (i), Asymmetric dimethylarginine (ADMA), (ii) symmetrical dimethylarginine (SDMA) and (iii) monomethyl-L-arginine (L-NMMA). Two distinct enzymes are required to produce ADMA and SDMA: they are type 1 (forms ADMA) and type 2 (forms SDMA) protein-arginine methyltransferases (PMRTs). ADMA can inhibit all three NOS isoforms and has a greater affinity for eNOS (Tousoulis et al., 2007). ADMA is elevated in individuals with T2DM (Abbasi et al., 2001, Lu et al., 2011, Schulze et al., 2006), and hypertension (Surdacki et al., 1999). Elevated ADMA has been shown to predict cardiovascular disease (Ravani et al., 2005, Lajer et al., 2008, Lu et al., 2011, Schnabel et al., 2005). Moreover, plasma ADMA concentrations have been shown to independently predict not only non-fatal myocardial infarctions but also death from cardiovascular causes (Schnabel et al., 2005, Meinitzer et al., 2007) in patients with coronary artery disease. Similar findings have been shown in haemodialysis patients (Zoccali et al., 2001) and angina (Lu et al., 2003a). Hyperglycaemia impairs dimethylargininase in the endothelium and smooth muscles which results in raised ADMA levels in individuals with T2DM (Lin et al., 2002). ADMA causes atherosclerosis lesions and renal damage which impairs NO· output in eNOS deficient mice (Suda et al., 2004). These deleterious effects are caused by upregulation of angiotensin-converting enzyme and elevated levels of O<sub>2</sub><sup>-</sup> (Suda et al., 2004).

# 1.1.10: Preventing eNOS uncoupling via pharmaceutical methods:

Uncoupling of eNOS decreases O<sub>2</sub> dependent NO· production and elevates oxidative stress (Li and Forstermann, 2013). Strategies that increase NO· bioavailability either through pharmacological methods or through dietary sources could alter the redox balance to prevent NO· and BH<sub>4</sub> oxidation. This may lead to reduced superoxide production from uncoupled NOS (Förstermann and Sessa,

2012). Below is a summary of supplements/drugs that can help prevent the uncoupling of eNOS and therefore elevate NO· bioavailability.

 $BH_4$ 

BH<sub>4</sub> supplementation has been shown to vasodilate blood vessels after arterial-infusion in patients with coronary heart disease (Cunnington et al., 2012). Its effectiveness as an anti-hypertensive is limited by its rapid oxidation. When BH<sub>4</sub> is administered as an oral treatment it has no effect on endothelial function (Cunnington et al., 2012). There is some evidence that BH<sub>4</sub> elevates iNOS activity (McNeill and Channon, 2012). Sepiapterin is a molecule that is converted to BH<sub>4</sub> via Sepiapterin-reductase and DHFR. These reductases have been shown to be downregulated in hypertension (Youn et al., 2012). Therefore, given the rapid oxidation of BH<sub>4</sub> as an oral agent (Cunnington et al., 2012), its use as a supplement is unlikely to be beneficial. Maintaining redox balance however, will prevent the uncoupling of NOS (Zou et al., 2004).

### **Antioxidants**

There are data to suggest that beetroot juice will not only increase the bioavailability of NO· by the entero-salivary pathway involving inorganic NO<sub>3</sub><sup>-</sup> but also via its antioxidant content (see experimental chapter 1 for table of antioxidant content) (Kanner et al., 2001). Antioxidant supplementation *per se* has the potential to alleviate oxidative stress via reductions in cell damage (de Boer et al., 2005), anti-inflammatory effects (Kelley et al., 2006) an inhibition of uric acid production (Jacob et al., 2003) and potentially prevent NOS uncoupling.

### Statins

Statins are a group of drugs that are utilised to lower plasma cholesterol levels via the inhibition of 3-hydroxy-3-methyl-glutaryl-CoA reductase. Statins enhance the availability of superoxide dismutase (SOD) and GPx isoforms (Carrepeiro et al., 2011, Landmesser et al., 2005). Statins also have an anti-atherosclerotic effect which is may in part be due to their impact on NO· bioavailability (Antonopoulos et al., 2012). *In vivo*, statins have been shown to increase the expression of BH<sub>4</sub> and also GTPCH1 in humans (Antoniades et al., 2011). This increase in GTPCH1 is likely to be responsible for the increase in BH<sub>4</sub> and ultimately reducing eNOS uncoupling (Hattori et al., 2003, Antoniades et al., 2011).

### Organic nitrates

Organic NO<sub>3</sub><sup>-</sup> act as NO· donors (such as isosorbide mononitrate). Pentaerithrityl tetranitrate has been shown to prevent eNOS uncoupling (again via elevated BH<sub>4</sub> concentrations (Schuhmacher et al., 2011)), induce SOD isoforms (Oppermann et al., 2009), HO-1, reduce NADPH oxidase and xanthine oxidase activity.

### ARB, ACEi and Beta Blockers

Angiotensin-converting enzyme inhibitor (ACEi) and angiotensin receptor blockers (ARBs) impact upon the renin-angiotensin-aldosterone system. ACEi have been shown to increase NO· production via reductions in NAPDH oxidase, O<sub>2</sub><sup>-</sup> and OONO whilst elevating BH<sub>4</sub> concentrations in an the wantanabe heritable hyperlipidemic rabbits (Imanishi et al., 2008). ARBs have antioxidant properties (Warnholtz et al., 1999) and long term exposure leads to re-coupling of eNOS (Warnholtz et al., 1999). This appears to be via the prevention of BH<sub>4</sub> oxidation (Wenzel et al., 2008). GTPCH1 mediates BH<sub>4</sub> synthesis (Wenzel et al., 2008, Satoh et al., 2008) and by up-regulation of dihydrofolate reductase mediates reduction of BH<sub>2</sub> to BH<sub>4</sub> (Oak and

Cai, 2007). Nebivol, a beta blocker, not only stimulates eNOS to produce NO-(Münzel and Gori, 2009) but has the ability to reverse its uncoupling (Mollnau et al., 2003) by inhibiting NADPH oxidase.

eNOS transcriptional enhancers (AVE3085)

AVE3085 is an eNOS transcription factor which is involved in the regulation of eNOS protein expression (Wohlfart et al., 2008). AVE3085 has been shown to elevate vascular BH<sub>4</sub> concentrations and thus reduce eNOS uncoupling (Wohlfart et al., 2008). AVE3085 also appears to have antioxidant effects and reduces the expression of NADPH oxidase (Westermann et al., 2009) which may contribute to elevated NO· bioavailability.

### 1.1.11 Ageing and NO.

Two key co-factors in the production of NO· are reduced in concentration as we age. These are BH₄ and L-arginine (Delp et al., 2008). This may mean that as we age we get a greater increase in uncoupled NOS, creating more superoxide and subsequently peroxynitrite (Kang et al., 2009). Moreover, plasma nitrite concentrations have also been shown to be reduced in older adults (Sindler et al., 2011). Given the putative benefits of NO· in healthy young adults, using a dietary source of nitrate may increase NO bioavailability in healthy older adults and enable them to benefit from the positive association that has been shown between NO· and vascular health (Ignarro et al., 1999).

### 1.1.12: Toxicology:

Methemoglobinemia occurs when the heme group in a haemoglobin is oxidised to its ferric state ( $Fe^{3+}$ ) and is therefore incapable of transporting  $O_2$  (Avery, 1999). As far back as 1868 the combination of  $NO_3^-$  and blood has been shown to produce

methemoglobinemia (Eusterman and Keith, 1929, Gamgee, 1868, Reichert, 1880). This was later corroborated in animal models as large doses of NO<sub>3</sub><sup>-</sup> caused the formation of methemoglobinemia (Binz and Geringer, 1901). In 1945 concerns about nitrates' potential toxicity emerged again as fears over methemoglobinemia or blue baby syndrome arose. Blue baby syndrome was first reported by Comly (1945) with the observation that the well-water had large contamination of NO<sub>3</sub><sup>-</sup>. Occurrences of methemoglobinemia were shown in (Walton) 1951 to be rare in water that contained < 44mg/l of NO<sub>3</sub><sup>-</sup>. Given this knowledge, the level at which the US (44 mg/l) and the European union (50 mg/l) set the NO<sub>3</sub><sup>-</sup> levels in drinking water was addressed. The notion that NO<sub>3</sub><sup>-</sup> by itself can cause methemoglobinemia has been strongly argued against by Avery (1999). Avery proposed a more likely scenario that NO<sub>3</sub><sup>-</sup> in the well water contributed to methemoglobinemia. Bacterial contamination within the well was converting NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup>. Furthermore, given the beneficial effects of NO<sub>3</sub><sup>-</sup> that are described later in this chapter, the removal of NO<sub>3</sub><sup>-</sup> from water supplies could be detrimental to human health by reducing the daily intake of NO<sub>3</sub><sup>-</sup>.

N-nitrosamines were shown to cause hepatic tumours in rats (Magee and Barnes, 1956). In 1976 two independent groups from Germany and the USA introduced the concept that NO<sub>3</sub><sup>-</sup> could be transformed into N-nitrosamines under acidic environments (Tannenbaum et al., 1976, Spiegelhalder et al., 1976). Given that NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> have been used as a preservative in food for centuries and that NO<sub>3</sub><sup>-</sup> is abundant in many foods which epidemiological evidence has suggested to have cardio protective effects such as vegetables (Joshipura et al., 1999, Joshipura et al., 2001), the publication by (Newberne, 1979) stating that NO<sub>2</sub><sup>-</sup> caused lymphomas in rats was surprising. The food and agriculture organisation and the world health organisation expert committee on food additives jointly released a statement which

stated "Overall, the epidemiological studies showed no consistently increased risk for cancer with increasing consumption of NO<sub>3</sub><sup>-</sup>. These data, combined with the results of the epidemiological studies considered by the Committee at its forty-fourth meeting, do not provide evidence that NO<sub>3</sub><sup>-</sup> is carcinogenic to humans" (Speijers and Brandt, 2003).

# 1.2: Introduction to Type 2 Diabetes Mellitus:

T2DM is a chronic disease and progressive in nature. The aetiology of the disease is chronic hyperglycaemia caused via insulin resistance and diminishing pancreatic beta cell insulin production (Stumvoll et al., 2007). These characteristics cause micro and macrovascular complications (such as cardiovascular disease) which ultimately result in elevated morbidity and mortality (Jaffe et al., 1984, Sprafka et al., 1991). T2DM is emerging as an epidemic in both the developed world as well as the developing world. Life expectancy has risen sharply over recent years (Oeppen and Vaupel, 2002) whilst obesity is rising due to a sedentary lifestyle and excessive intake of calories. This review will aim to explore the demographics of T2DM, insulin resistance and the development of T2DM, measurement of T2DM, genetics, response to exercise, macrovascula,r microvascular disease and metabolic complications and NO·.

### 1.2.1: T2DM demographics and costs:

T2DM accounts for 90 - 95% of all cases of diabetes worldwide (Harris, 1998, Hariri et al., 2006). Physical inactivity and obesity increases the prevalence of this metabolic disease (Wild et al., 2004). As we age (on a population level) we do less and less activity (Sallis, 2000) which may explain some of the increased risk with aging. As the global population expands the number of people with T2DM is increasing (Wild et al., 2004). The chances of developing diabetes increases from 0.3 - 0.5 % per year for 50-60 year olds to 0.5 - 1% for 60-70 year olds and greater than 1% a year for over 70 year olds (Berger et al., 1999). Wild et al., (2004) projected that by 2030 this figure of 171 million will have doubled to over 340 million worldwide. However, this may be conservative given these calculations were based on obesity levels remaining stable and the observation that obesity levels are in fact

increasing. More recent estimates of 439 million by 2030 have been suggested after extrapolation from the 2010 figure of 285 million individuals (Shaw et al., 2010). In the UK 3.1 million people (diagnosed and undiagnosed) have T2DM, which equates to 7.4% of the population (PHO, 2010). With increasing numbers of individuals developing T2DM the associated costs increase (Caro et al., 2002). T2DM is responsible for 8% of the NHS annual budget equating to £8.8 billion per annum. The total cost of direct and indirect care comes to a total of £20.5 billion (Hex et al., 2012).

# 1.2.2: Insulin resistance, beta cell dysfunction, and type 2 diabetes mellitus:

Reduction in sensitivity to the hormone insulin causes three major abnormalities to occur in the body: firstly, a reduced insulin mediated uptake of glucose by skeletal muscle, secondly, an increase in free fatty acid metabolism via adipose tissues and finally over production/secretion of glucose by the liver (Kabir et al., 2005). As insulin resistance progresses hyperglycaemia will begin to manifest itself in the fasted state (Lebovitz, 1999). Insulin resistance is a precursor to the development of T2DM in which the cells within the bodies fail to respond to the hormone correctly. The body produces adequate concentrations of insulin, however, changes in the cell surface receptors, (Kolterman et al., 1980), mitochondrial dysfunction (Holloszy, 2009), endoplasmic reticulum stress (Cnop et al., 2012), fat in skeletal muscle and glucose modification (Hart et al., 2011) (via the hexosamine biosynthesis pathway) prevent the correct response (i.e. the uptake of glucose into the cell). As peripheral insulin resistance progresses in skeletal muscle, liver, and fat tissues circulating plasma glucose concentrations will rise. Rising plasma glucose concentrations stimulate the pancreatic ß-cells to secrete more insulin (hyperinsulinanemia) in an attempt to

maintain metabolic homeostasis. This will continue until total ß-cells' death (Stumvoll et al., 2005) (See figure 2 below).

The aetiology of T2DM and the subsequent insulin resistance and beta cell dysfunction is multifactorial. Of primary importance are the following: relative excessive calorie intake (Bloomgarden, 2000), aging (Harris et al., 1998), and a lack of exercise (Morgan et al., 2002, Shaw et al., 2010). This means that insulin resistant individuals are in a chronically elevated energy balance (Stumvoll et al., 2005, Eriksson, 2007, Kahn et al., 2006). If an individual has a persistent positive energy balance (i.e. over and above the requirements for daily living) adipocytes will uptake triglycerides until they become saturated, creating adiposopathy (Bays, 2011). This mainly occurs in the visceral adipocytes. If adipocytes are at maximum capacity the ability to reduce circulating fats is diminished (Tushuizen et al., 2007). Hyperglycaemic mediated insulin production has been shown to be stunted by exposure of the ß-cells to fatty acid deposits, which are present in higher concentrations due to reduced clearance (Lee et al., 1994, Tushuizen et al., 2007). Furthermore, persistent chronically elevated fatty acid and triglyceride levels lead to abnormal amounts of these fats being stored in skeletal muscle (Boden, 1997, Boden and Chen, 1995) and the liver (Kabir et al., 2005). Storage of fats in skeletal muscle and the liver expedites the insulin resistance as skeletal muscle is the predominate source of glucose uptake (Berg et al., 2001, Kelley et al., 1993).

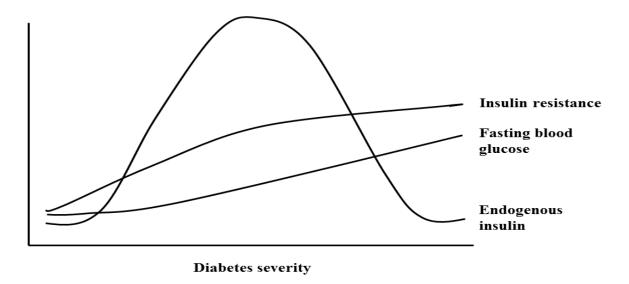


Figure 2: Change in insulin resistance, insulin production and fasting blood glucose as diabetes severity increases.

#### 1.2.3: Measurement of insulin resistance and T2DM

HbA1c is haemoglobin that has been glycated. HbA1c is created via an Amadori reaction which causes an irreversible binding of glucose molecules to the  $\beta$  -N-terminal valine residues on the globin chains. During the reaction the haemoglobin is rearranged to 1-deoxy-1-N-valyl-fructose (Bookchin and Gallop, 1968, Dixon, 1972, Bunn et al., 1975). Although exposure of haemoglobin to glucose during the 120 day life time is stable in individuals with controlled diabetes, the HbA1c concentration is higher in older erythrocytes (Huisman and Dozy, 1962, Fitzgibbons et al., 1976). When circulating glucose concentrations rise the proportion of HbA1c rises as well, therefore the higher the plasma glucose the higher the HbA1c (Rohlfing et al., 2002). HbA1c is used as a measure of glycaemic control over a prolonged period as it is stable over time, typically accurate over a 3 month period due to the replacement of new red blood cells. The WHO have published a report on the use of HbA1c as a diagnostic tool for diabetes (WHO, 2011).

#### 1.2.4: Genetics

T2DM appears to have a heritable component with heritability of T2DM being suggested to be greater than 50% in studies of monozygotic twins (Herder and Roden, 2011, Poulsen et al., 2009), and first degree relatives of individuals with T2DM. (Hemminki et al., 2010, Poulsen et al., 2009, Tattersall and Fajans, 1975, Saxena et al., 2007). Genetic mapping over recent years has shown more than 40 loci that are thought to be partly responsible for the development of T2DM (Herder and Roden, 2011, Voight et al., 2010, Dupuis et al., 2010, Zeggini et al., 2008, Saxena et al., 2010). Most of these loci are associated with the dysfunction of pancreatic β-cells with the most common named 'TCF7L2' (Dupuis et al., 2010). The identified loci can only explain 10% of the heritability in T2DM (Herder and Roden, 2011). The remaining cause of heritability is unknown. However, Nolan and colleagues (2011) suggest that variants in allele frequency maybe involved. Genetics also have a role to play in the development of microvascular disease, there appears to be an abnormality in the loci within chromosomes 7 and 20 which increases susceptibility to nephropathy (Imperatore et al., 1998).

Ethnicity and environment can affect the risk of developing T2DM and it can also affect the likelihood of complications. Fasting plasma glucose levels were measured in Pima Indians (Pettitt et al., 1980, McCance et al., 1994), Egyptians (Engelgau et al., 1997), sections of the US population (Alberti and Zimmet, 1998) and Caucasians (Jarrett et al., 1984) independently. Measurements of the associated risk diabetes related complications and fasting plasma glucose concentrations were correlated with the mg/dL ranging from 108-140. African American and Mexican Americans have been shown to be at greater risk of diabetes related retinopathy compared to Caucasian Americans. In African American individuals this could be explained by a

larger % of the population having high blood pressure and a higher HbA1c. However, in Mexican Americans this is unexplained (Harris et al., 1998).

# 1.2.5: T2DM and response to exercise:

#### Fatique:

During exercise T2DM can lead to increased discomfort and early fatigue (Regensteiner et al., 1995a, Regensteiner et al., 1998) which may impair people with T2DM's ability to achieve the recommended amounts of exercise that are a key part of the management of T2DM (NICE, 2008). This premature fatigue has been attributed to abnormalities occurring at multiple points during the transport of O<sub>2</sub> from the lungs to the muscles where it is utilised during exercise (Scognamiglio et al., 1998). Possible explanations for this impairment include compromised cardiac function which can manifest with reduced stroke volume (Gusso et al., 2008), reduced cardiac innervation (Scognamiglio et al., 1998) reduced left ventricular diastolic function (Poirier et al., 2000) and impaired cardiac autonomic function (Gerritsen et al., 2001). Other explanations of exercise impairment include diminished skeletal muscle O2 diffusion and a reduced mitochondrial O2 utilisation (Regensteiner et al., 1995a, Regensteiner et al., 1998, Baldi et al., 2002). These effects are likely underpinned by reductions in mitochondrial density (MÅrin et al., 1994, Mathieu-Costello et al., 2005, Ritov et al., 2005) and amplified levels of mitochondrial dysfunction (Padilla et al., 2007). This dysfunction is characterised by an impaired bioenergetic capacity (specifically diminished NADH and citrate synthase (Kelley et al., 2002) and a reduced ability to uptake glucose (Mathieu-Costello et al., 2003).

### Maximal O<sub>2</sub> uptake:

Individuals with T2DM have an elevated resting O2 consumption coupled with a profound reduction in maximal O<sub>2</sub> uptake (Baldi et al., 2002, Regensteiner et al., 1998, Regensteiner et al., 1995a, Wilkerson et al., 2011, O'Connor et al., 2015). Maximal O<sub>2</sub> uptake is an important predictor of all-cause mortality (Myers et al., 2002, Galant et al., 2012). Therefore, individuals with T2DM are likely to be closer to their maximum exercise capacity than individuals without diabetes for any given intensity of exercise (Regensteiner et al., 1998). The consequence of being closer to the maximal exercise capacity is that a greater physiological stress and thus greater discomfort will be experienced by the individual (Regensteiner et al., 1998), resulting in a reduction in exercise tolerance. If two individuals exercise at the same work load, the one with higher O<sub>2</sub> consumption will typically find the exercise more difficult. This may translate with individuals with T2DM having greater perceived exertion at the same workload. Difficulties completing submaximal exercise tasks may manifest as a barrier to attain the recommended amounts of exercise recommended by the WHO (Huebschmann et al., 2009). The reduction in maximal O<sub>2</sub> uptake is ultimately caused by an irregularity in O<sub>2</sub> delivery (Regensteiner et al., 1995a, Regensteiner et al., 1998, Baldi et al., 2002, Wilkerson et al., 2011).

### O<sub>2</sub> delivery:

Many of the abnormalities that are related to premature fatigue in individuals with T2DM are likely caused via  $O_2$  delivery perturbations. These include  $O_2$  transportation form the lungs to the muscle during exercise (Scognamiglio et al., 1998), along with reduced stroke volume (Gusso et al., 2008), cardiac innervation (Scognamiglio et al., 1998), cardiac autonomic function (Gerritsen et al., 2001) and left ventricular diastolic function (Poirier et al., 2000).  $O_2$  delivery may also be limited

by a reduced total and fractional O<sub>2</sub> extraction into the mitochondria which is caused via reduction in mitochondria density, size and function (Regensteiner et al., 1995a, Regensteiner et al., 1998, Baldi et al., 2002, Padilla et al., 2007, Shore et al., 1994, Mårin et al., 1994, Mathieu-Costello et al., 2005). Individuals with T2DM also have elevated plasma endothelin-1 (endothelin-1 are a group of peptides that modulate vascular tone) (Schneider et al., 2002), reduced capillary density (Mårin et al., 1994, Mathieu-Costello et al., 2003), impaired haemodynamics (Padilla et al., 2007, Shore et al., 1994) and mitochondrial function (Kelley et al., 2002) in the skeletal muscle.

# $\dot{V}$ $O_2$ kinetics and exercise tolerance:

The rate at which  $\dot{V}O_2$  projects towards the steady state is a major determinant in the ability of an individual to tolerate exercise. The quicker an individual can attain a steady state following the onset of exercise, the greater the contribution of oxidative compared to non-oxidative metabolism (Jones and Poole, 2005). By increasing the speed of the O<sub>2</sub> uptake response, individuals can reduce the intracellular changes in metabolic milieu that have been linked to the fatigue process. By using a larger proportion of oxidative metabolism, the speed at which lactic acid concentrations build up are slowed. Lactic acid disassociates into lactate and H<sup>+</sup>, the H<sup>+</sup> causes acidosis and thus is an important determinant of fatigue (Bruton et al., 1998, Pate et al., 1995, Westerblad et al., 1997, Wiseman et al., 1996). Similarly, in anaerobic respiration, phosphocreatine is broken down into creatine and inorganic phosphate (Pi). Pi concentrations appear to have a larger contribution to fatigue than reduced pH levels (at least at physiological temperatures) (Dahlstedt et al., 2001, Dahlstedt et al., 2000, Dahlstedt and Westerblad, 2001, Fryer et al., 1995, Kabbara and Allen, 1999, Kabbara and Allen, 2001). Fatigue may however be expedited in low pH levels by inhibition of key enzymes involved in glycolysis and glycogenolysis. This may

slow the adenosine diphosphate to adenosine triphosphate turnover rate (Westerblad, Allen, & Lännergren, 2002).

The  $\dot{v}O_2$  kinetics of individuals with T2DM have been shown to be slowed (Bauer et al., 2007, Regensteiner et al., 1998, Regensteiner et al., 1995a, Brandenburg et al., 1999). Conversely, Wilkerson et al. (2011) showed no difference in the  $\dot{v}O_2$  kinetics of older individuals with T2DM compared to healthy age matched controls. More recently, the putative slowed  $\dot{v}O_2$  kinetics shown by some studies in older individuals with T2DM has been revisited by O'Connor et al. (2015) to assess whether the lack of effect shown in some studies in the kinetics of older individuals with T2DM was due to disease duration or age. Disease duration was matched between a younger group and an older group of individuals with T2DM. Their findings indicate that middle aged men have impaired  $\dot{v}O_2$  kinetics (i.e. a longer time constant) compared to healthy matched controls. However, older (65 years old) individuals with T2DM were similar to their healthy matched controls, replicating Wilkerson et al's findings in 2011. This suggests that aging is the predominant cause for slowed  $\dot{v}O_2$  kinetics.

### Medication and exercise performance:

T2DM is a disease that affects multiple systems. Many of the medications that are used to treat one aspect of their condition can also have many other beneficial effects. Drug treatment used to reduce hyperglycaemia such as sulfonylureas, insulin and metformin have all been shown to help reduce microvascular complications (Turner et al., 1998, UKPDS, 1998, Ohkubo et al., 1995, Patel et al., 2008) and improve exercise performance. For instance insulin when used to treat patients with T2DM and heart failure as a comorbidity have been shown to improve ventilatory efficiency and  $\dot{V}O_2$  (Guazzi et al., 2003). Moreover, ramipril has been

shown to increase treadmill walking time compared with a placebo in individuals with peripheral artery disease (Ahimastos et al., 2013).

# 1.2.6: Micro & macrovascular Complications:

Between 1950 and 1995 the absolute risk of myocardial infarction, strokes and intermittent claudication has fallen by ~50% in individuals with diabetes (Fox et al., 2004). However, individuals who have T2DM still have a two to four fold increased chance of developing coronary heart disease compared to individuals without T2DM (Stamler et al., 1993, Hanefeld et al., 1991, Wei et al., 1998, Wilson et al., 1991). Although the overall risk of CVD has fallen in individuals with T2DM over resent decades, more of the risk is attributable to T2DM (Fox et al., 2007) (the Framingham heart study). Having diabetes and CVD increases the risk of mortality compared to non-diabetic controls following a myocardial infarct by 50% (Sprafka et al., 1991). Jaffe et al., also showed an increased mortality in individuals with T2DM compared with controls. However when both groups were stratified for congestive heart failure the survival rates were similar (Jaffe et al., 1984).

Cholesterol is one of the major components involved in the pathogenesis of CVD (Lu et al., 2003b). Superficially, individuals with T2DM appear to have a similar lipid profile to age matched controls, for instance they have total cholesterol and LDL-cholesterol that are very similar (Laakso et al., 1985). Individuals with T2DM have a lower HDL-cholesterol and higher triglycerides (Wilson et al., 1985, Taskinen, 2003) which in turn are known risk factors for the development of CVD (Fontbonne et al., 1989, Assmann and Schulte, 1991).

Poor glycaemic control is detrimental to microvascular function and has been strongly associated with elevated microvascular dysfunction (Stratton et al., 2000).

The risk for microvascular disease is thought to be determined by the level of glycaemia, with extremely high glucose concentrations creating the greatest risk (Krolewski et al., 1995). Every percentage point decrease in HbA<sub>1c</sub> (and thus improving control) results in a 35% reduction in microvascular complications (UKPDS, 1998). This group also suggested that by improving glycaemic control via diet and metformin as opposed to insulin therapy, microvascular complications can be reduced by 32%. Some of these findings have since been replicated in a Japanese cohort (Shichiri et al., 2000) when comparing 3 or more insulin injections compared with 1-2 injections. When more injections were used a greater level of glycaemic control was delivered which correlated with a reduced risk of microvascular complications. Not all of the experimental and epidemiological evidence supports this notion. In 2009 evidence was published to suggested that intensive glycaemic control in individuals with T2DM had no effect in controlling for microvascular complications or even major cardiovascular events (ADVANCE trial 2009) (Duckworth et al., 2009). Another study in 2008 showed no effect where aggressive treatment of HbA1c with intensive therapy (increased medications with monthly visits to assess glycaemic control) compared with conventional therapy (as prescribed by their clinician) actually increases mortality (ACCORD (Hertzel C. Gerstein et al., 2008)).

### 1.2.7: NO· and related microvascular diseases:

The availability of NO· is important in the pathophysiology of diabetes related vascular disease, with reduced availability leading to reduced endothelial relaxation (Creager et al., 2003). This impairment ultimately contributes to diabetes related microvascular angiopathy. This microvascular disease is a hallmark of diabetes related retinopathy and nephropathy (Santilli et al., 2004). Polymorphisms of eNOS

have been shown to increase the severity of retinopathy and nephropathy in individuals with T2DM (Awata et al., 2004). By contrast a recent study has shown that an eNOS genotype is not associated with the development of diabetes related retinopathy (de Syllos et al., 2006, Neugebauer et al., 2000). However, eNOS polymorphisms are associated with nephropathy progression (Neugebauer et al., 2000). de Syllos and Awata et al's cohorts have ~50 participants with similar methodologies. The differences in these trials may be explained by differences in the cohorts. de Syllos et al's cohort had diabetic retinopathy for ~4 years more than Awata cohort. Awata et al's cohort also appear to have an abnormally low BMI for individuals with T2DM, at ~ 24 kg/m².

### 1.2.8: Retinopathy:

Awata et al., suggest that the down-regulation of basal eNOS expression resulted in the macular oedema (caused via protein deposits and fluid build-up under the macula) which leads to a degradation of the blood-retina barrier. Larger cohorts are required to definitively explore the effects of eNOS genetic variation on retinal status. If the retardation of eNOS is limiting NO· availability through the traditional NOS pathway, this may reduce its vasodilatory effect in the microvasculature. Habitual supplementation of NO₃ could alleviate the damage by providing a pool of NO₂ for later conversion to NO· in the microvasculature of the retina along with other microvessels. Retinopathy is the changes in the microvasculature of the retina. Initially the changes are characterised by increased vascular permeability which can progress to proliferative diabetic retinopathy (PDR) and eventually blindness. PDR is characterised by growth of new blood vessels on the retina and vitreous with an increase in blood flow (Patel et al., 1992). Macular oedema caused by retinal thickening is a common development with this disease (Fong et al., 2004). Within the

first two decades of diagnosis > 60% of individuals with T2DM have retinopathy (Fong et al., 2004). The greatest predictor for diabetes related retinopathy and progression is the duration of diabetes. As the years of disease increase so does the incidence of retinopathy, after 3 years there is a 8% prevalence which increased to 25% after 5 years and 60% at 10 years (Klein et al., 1984).

#### 1.2.9: Nephropathy:

Diabetes related nephropathy (DRN) is a progressive disease that affects the kidneys. It is caused by angiopathy of the capillaries within and surrounding the glomerulus (Christensen, 1971). DRN affects approximately 40% of all individuals with T2DM (Parving et al., 2001) with 5-10% of individuals with T2DM newly diagnosed with microalbuminuria each year (Ravid et al., 1993, Nelson et al., 1996, Gæde et al., 1999, HOPE, 2000). Approximately 40% of all end stage renal failure occurs due to diabetes related nephropathy (USRDS, 2012). Patients that have microalbuminuria have a 10-20 fold increased risk of developing nephropathy (Parving, 1996). Glomerular filtration rates may be elevated at diagnosis with T2DM due to hyperfiltration with or without microalbuminuria. Once macroalbuminuria develops the glomerular filtration rate progressively declines (Nelson et al., 1996). Ethnicity has a large impact upon nephropathy rates. However, age doesn't appear to have an impact. African American individuals have the highest risk followed by Asian and Hispanic with Caucasian individuals having the lowest risk factor (USRDS, 2012) with costs rising sharply to treat nephropathy with estimates ranging from \$68,000 to \$88,000 (USRDS, 2012, Foley and Collins, 2007).

# 1.2.10: Neuropathy:

Neuropathy is a microvascular complication thought to be brought about by damage to blood vessels that supply blood to nerves. Approximately 10% of individuals with

T2DM have neuropathy (Said, 2007). The prevalence of neuropathy is believed to increase with poor glycaemic control and the individual's duration of T2DM (Pirart, 1978, TDCCTRG, 1993, Martin et al., 2006). Neuropathy is usually a distal symmetric disease accounting for approximately 80% of all cases (Palumbo et al., 1978, Freitas et al., 1992). Progression of polyneuropathy can also be triggered by amyloid polyneuropathies (Said, 1981). Competitive inhibition of the breakdown of amyloid-β in individuals with T2DM, by insulin degrading enzyme, occurs predominantly due to hyperinsulinanemia. In individuals with an impaired insulin response (Qiu and Folstein, 2006), this process may lead to increases in amyloid related polyneuropathies that are diagnosed as diabetic neuropathy in individuals with T2DM.

#### 1.2.11: Conclusion:

T2DM metabolic disease with a rising prevalence that affects hundreds of millions of people worldwide. Attenuating sedentary time in individuals with T2DM appears to improve markers of metabolic health (Falconer et al., 2015). Therefore, strategies aimed at making exercise easier (thus removing barriers to exercise and fatigue) or reducing hyperglycaemia are needed to help combat the economic burden and improve quality of life in individuals with T2DM.

### 1.3: Introduction to chronic obstructive pulmonary disease

## 1.3.1: Background:

Chronic obstructive pulmonary disease (COPD) is an umbrella term given to group of diseases; emphysema, chronic bronchitis and chronic obstructive airways disease (Sin and Vestbo, 2009). Chronic obstructive pulmonary disease (COPD) is characterised by constricted airflow that is treatable but not entirely reversible, with a gradual decline in function (Kirkham and Rahman, 2006). Individuals with COPD typically report: exertional breathlessness, a chronic cough, regular sputum production, bronchitis and wheezing (GOLD., 2011). Multiple factors lead to relative hypoxemia in individuals with COPD such as reduced cardiac function (Jardin et al., 1984), muscle deconditioning (Maltais et al., 2014), irregular pulmonary blood flow distribution leading to incomplete gas exchange (Marshall et al., 1994) and cor pulmonale. These manifestations often lead to a significant reduction in quality of life (DiBonaventura et al., 2012) and feelings of breathlessness and fatigue (Nici et al., 2006). This review will consist of a brief introduction, demographics and economic burden, causes of the disease, COPD and exercise, pulmonary hypertension, dynamic hyperinflation and treatments.

### 1.3.2: Demographics and economic burden:

It is projected that by 2020 COPD will be the third most common cause of death worldwide (GOLD., 2011), due to an ageing population which have more exposure to COPD risk factors (Mathers and Loncar, 2006). There are approximately 3 million people living in the UK with COPD. However, 2 million of these remain undiagnosed (Healthcare-Commission, 2006). COPD accounts for 30,000 deaths a year in the UK with ~90% of these deaths occurring in those over 65 year olds (DoH, 2009). COPD accounts for 23% of all respiratory deaths in the UK (Society, 2006). There is a

substantial economic burden to society as a result of COPD. The Chief Medical Officer for England and the United Kingdom's governmental Chief Medical Advisor reported that COPD accounts for ~£800 million in direct health care costs (DoH, 2005), with other reports estimating the cost to be as high as £930 million (Society, 2006). The healthcare commission also states that more than 50% of these costs are related to direct provision of care, and as such it is the single most costly provision of care within hospitals (Healthcare-Commission, 2006). The total economic cost of COPD is much larger given that it accounts for the loss of 24 million working days in the UK each year (Healthcare-Commission, 2006).

# 1.3.3: Chronic bronchitis & emphysema:

Chronic bronchitis is a disease defined by bronchial hypersecretion and a cough which is present for a minimum of 3 months at a time in at least two consecutive years (MRC, 1965). However, not all patients with chronic bronchitis will develop airflow limitations (Vestbo and Lange, 2002). Chronic bronchitis is associated with a chronic secretion of mucus combined with a progressive decline in lung function. Chronic bronchitis increases incidences of hospitalisation and mortality (Vestbo et al., 1996). Chronic bronchitis is predominantly caused by smoking. Evidence following on from the clean air act of 1956 which improved urban air quality dramatically has shown an associated reduction in the incidence of COPD and its associated diseases.

As the disease progresses the alveoli are broken down which leads to large voids within the lungs, drastically reducing surface area for gas exchange (Snider, 1985). The pathology of emphysema involves the infiltration of alveoli by neutrophils, macrophages and lymphocytes (Taraseviciene-Stewart and Voelkel, 2008). Emphysema is also known to create a loss of elastic recoil within the lungs which is

caused by activated macrophages which release MMP-12 and neutrophil elastase which breaks down the elastin matrix between the alveoli (Mercer et al., 2004). Other cells such as epithelial and endothelial cells can also produce and secrete proteases (Mercer et al., 2004, Taraseviciene-Stewart et al., 2005). Subsequent scaring to lung tissue and a reduction in elastic recoil causes hyperinflation which causes major limitation to gas exchange (O'Donnell and Laveneziana, 2007). As the physiological drive to expire CO<sub>2</sub> rises and demand for O<sub>2</sub> increases, ventilation will increase (Fishman et al., 1955). The air enters the lung as the rib cage expands and pulls the scarred connective tissues. When the individual breathes out the air becomes trapped as the proximal and distal airways collapse. When the air becomes trapped the ability to breathe off CO<sub>2</sub> decreases which in turn will increase ventilation and create a vicious cycle of air trapping (Fishman et al., 1955).

#### 1.3.4: COPD assessment:

Severity or grade of COPD is accessed via the measurement of the forced expiratory volume in the first second (FEV<sub>1</sub> (GOLD., 2011)). The speed at which FEV<sub>1</sub> declines is a good indicator of survival (Anthonisen et al., 1986). The authors also showed that those individuals with a lower FEV<sub>1</sub> at the beginning of the trial had the smallest reduction in FEV<sub>1</sub> relative to baseline. Moreover, with individuals with relatively preserved FEV<sub>1</sub> their speed of reduction correlated well with their response to a bronchodilator (Anthonisen et al., 1986). This landmark study also demonstrated an average reduction in FEV<sub>1</sub> of 44 ml/year over three years (n = 985). The severity of dyspnoea correlated significantly with Quality of Life (QoL) as measured via a health related questionnaire (SF-36) but not significantly with FEV<sub>1</sub> (Mahler and Mackowiak, 1995). This suggests that as FEV<sub>1</sub> does not correlate well with the QoL of an individual (Mahler and Mackowiak, 1995), a qualitative approach should be

considered when assessing QoL in individuals with COPD. This may be due to COPD being a multisystem disease.

# 1.3.5: Smoking and oxidative stress:

The recent report (GOLD., 2011) looking at the effects and causes of COPD has emphasised the impact of 'noxious particles and gases' and recently defined COPD as "a disease state characterized by airflow limitation that is not fully reversible. The airflow limitation is usually both progressive and associated with an abnormal inflammatory response of the lungs to noxious particles or gases" (GOLD., 2011). COPD has been linked to elevations in oxidative stress within the lungs (Roy et al., 2007, Langen et al., 2003). Experimental research provides evidence that oxidants play a critical role in the pathogenesis of COPD. The principal contributory factor to the increase in oxidants is widely accepted to be cigarette smoke (Ciencewicki et al., 2008). Of all COPD related mortality, 85% is attributed to smoking (Healthcare-Commission, 2006). Cigarette smoke and smoke in general is known to have high concentrations of reactive oxygen species (ROS) (Chen et al., 2015). These ROS are thought to upregulate a transcription factor, NF-kB, known to be involved in inflammation (Li et al., 2009). Increases in ROS will therefore increase the inflammatory response and lead to alveoli breakdown. However, cigarette smoke is not the only cause of COPD. COPD can be caused via exposure to occupational air pollution for example, from burning wood or other biofuels (Salvi and Barnes, 2009). This is important because there is a delicate balance within the body between ROS and antioxidants termed as a redox balance. When this balance is shifted past the antioxidant capacity, ROS cause damage to proteins, lipids and DNA (Ciencewicki et al., 2008). ROS are also endogenously produced in epithelial and inflammatory cells

in response to cellular damage/injury and metabolism of O<sub>2</sub> (Ciencewicki et al., 2008, Thannickal and Fanburg, 2000).

### 1.3.6: Smoking and exercise:

Reduced exercise tolerance is often found in individuals with COPD. The reduction in exercise tolerance is not a linear relationship dependent upon the number of cigarettes smoked (Wüst et al., 2008). This suggests that it is either an acute effect or, that after a certain number of cigarettes, the amount of intolerance to exercise that an individual experiences will reach a plateau (Wüst et al., 2008). This reduction in tolerance to exercise could simply be due to a sedentary lifestyle which is often linked to smoking (Larsson et al., 1998). Approximately 80% of individuals diagnosed with COPD are current or were former smokers. The only known way to slow the decline in pulmonary function is smoking cessation. Aggressive smoking interventions can significantly slow the decline in FEV<sub>1</sub> (Anthonisen et al., 1994). The majority of the reduction in FEV<sub>1</sub> decline was shown within the first year and the individuals who stopped smoking for the longest periods (within the year) experienced the greatest pulmonary function protective effects. Other medications which are usually utilised for asthmatic relief are used to ease the symptoms of the disease such as mucolytic agents, corticosteroids or bronchodilators (GOLD., 2011).

# 1.3.7: Impact of pulmonary hypertension, and dynamic hyperinflation on exercise:

The formation of abnormal pulmonary blood vessels in individuals with COPD creates hypoxia via reduced gas exchange (O'Donnell and Laveneziana, 2007). Subsequent to these deleterious effects a diminished O<sub>2</sub> saturation occurs which contributes to pulmonary hypertension via pulmonary vasoconstriction (Hopkins & McLoughlin, 2002. Pulmonary hypertension is defined by a resting mean pulmonary artery pressure (mPAP) of 20-25 mmHg. The mPAP can be used as a prognostic

tool. For mPAP > 25, the 5 year survival rate is 36% compared to < 25 mmHg is 62% (Oswald-Mammosser et al., 1995). The mPAP has been treated effectively via O<sub>2</sub> therapy (Zieliński et al., 1998) and NO· inhalation (in rats) (Kouyoumdjian et al., 1994). Over time pulmonary hypertension leads to increasing vascular damage and further decreasing lung function (Hopkins and McLoughlin, 2002). Pulmonary hypertension is strongly associated with COPD, especially in the advanced stages of the disease (Barberà and Blanco, 2009, Barberà et al., 2003). Pulmonary hypertension is also a key predictor of mortality in COPD (Burrows et al., 1972), and can have significant implications for morbidity (Chaouat et al., 2005). There are conflicting views as to the prevalence of pulmonary hypertension depending on which of these values are used. The occurrence of pulmonary hypertension can range from 20% to 91% of the COPD cohort (Oswald-Mammosser et al., 1991, Weitzenblum et al., 1984, Scharf et al., 2002, Thabut et al., 2005).

Tibetans appear to be able to utilise NO· to control hypoxia (due to altitude) via an elevated blood flow without the negative effects of pulmonary hypertension or vascular resistance (Erzurum et al., 2007). Non-indigenous people who go to high altitude have higher mPAP which impedes NO· bioavailability (Bailey et al., 2010a). Other research groups have suggested that high endogenous NO· levels are correlated with less severe injury within the lungs and may have a protective effect for pulmonary and systemic endothelium and may act as a protective function for acute lung injury (McClintock et al., 2007).

The significance of pulmonary hypertension to individuals with COPD can be stark. Pulmonary hypertension is usually stable when patients are assessed at rest. However, pulmonary hypertension is exacerbated during exercise (Thabut et al., 2005). Blood pressure rises during exercise which can increase the pressure of

blood moving through the lungs. The severity of pulmonary hypertension has a direct effect on exercise performance in the 6 minute walk test (6MWT) (Cuttica et al., 2010, Sims et al., 2009). Individuals with COPD, with or without pulmonary hypertension, may have a significant overlap within measures of lung function such as:  $S_pO_2$ , peak  $O_2$  uptake, ventilatory efficiency, maximal ventilation and spirometry levels has been previously demonstrated (Holverda et al., 2008). Reduction in  $S_pO_2$  at rest which exacerbate when exercising have a clear correlation with pulmonary hypertension (Holverda et al., 2008).

When individuals with COPD exercise they can have a further hyperinflation known as dynamic hyperinflation. End-expiratory lung volume increases due to a superimposed increase above the hyperinflation at rest (Marin et al., 2001, O'Donnell et al., 1998, Gallagher, 1994).

### 1.3.8: COPD, exercise and medicines:

COPD is not only a pulmonary disease, but also a systemic disease. Impairments in  $FEV_1$  do not account for all of the limitations associated with COPD (Roy et al., 2009). Diminished functional capacity is brought about via a higher resting  $O_2$  requirement and lower peak  $\dot{V}O_2$  (Gosker et al., 2003). The elevated energy requirement at rest represents a much higher fraction of their maximal exercise capacity (Gosker et al., 2003) and thus patients experience greater physiological stress during exercise. Other reasons for a reduced exercise tolerance are muscular atrophy, fibre type plasticity (changes from slow to fast) (Wüst and Degens, 2007), changes in pulmonary blood flow distribution (Marshall et al., 1994) and cardiac deconditioning (Maltais et al., 2014).

Some improvements in exercise capacity have been shown following structured exercise programs (Griffiths et al., 2000, Cockcroft et al., 1981). To date, three processes have been shown to aid in this improvement: physical reconditioning (Casaburi et al., 1991), improved ventilatory efficiency (Casaburi et al., 1997) and desensitisation to dyspnoea (Cooper, 2009). Some researchers have also suggested that improvements in ventilatory muscle function could also aid these improvements (Cooper, 2009).

Individuals with COPD can have numerous exercise limiting cardiovascular defects.

These consist of:-

- (i) An impaired left ventricular function as a direct result of hypoxemia. Hypoxemia creates a reduced pulmonary blood flow (Cooper, 1995) which exacerbates the pulmonary hypertension causing a negative feedback loop.
- (ii) There is an increasing amount of experimental evidence that suggests that deconditioning due to a sedentary lifestyle in individuals with COPD brings about markers of fatigue such as elevated blood lactate levels (Cooper, 1995).
- (iii) A shift from aerobic to anaerobic metabolism due to a reduced delivery of  $O_2$  to the muscle causes lactate production (Cooper, 1995). This process stimulates increased ventilation which in turn further reduces the time available for exhalations, creating a vicious cycle of air trapping (Cooper, 2001).

Reductions in exercise tolerance are also brought about via feelings of dyspnoea and early onset fatigue. Cooper et al., (2009) showed that a general physical training program elicited an improved feeling of dyspnoea as determined via a visual analogue scale following a 6MWT, whereas inspiratory muscle training had no effect.

However, participants with COPD can become 'acclimatised' to exercise after being deconditioned for a prolonged period which leads to an alleviation of the fear dyspnoea (Agle et al., 1973).

# 1.3.9: Nitric oxide (NO·) and its effects as a bronchodilator:

NO· is an endothelium derived relaxant factor in the vasculature. NO· and the ROS that develop from this molecule play a critical role in COPD pathophysiology (Bove and van der Vliet, 2006). NO· acts as a bronchodilator (Dupuy et al., 1992) via mediation of the actions of nitrovasodilator drugs such as glyceryl trinitrate and nitroprusside. These vasodilators relax smooth muscle in the airways which subsequently increases production of soluble guanylyl cyclase (sGC) and cyclic guanosine 3' 5' monophosphate (cGMP) (Dupuy et al., 1992, Gruetter et al., 1989). In animal models, inhaled NO· diffuses across the alveolar membrane into the vascular smooth muscle due to its highly lipophilic properties (Shaw and Vosper, 1977) and induces vasodilation, which reduces hypoxia induced pulmonary vasoconstriction (Frostell et al., 1991, Fratacci et al., 1991). However, when NO· comes into contact with haemoglobin a rapid oxidation (Gruetter et al., 1981) occurs which essentially makes the NO· inert due to its oxidation to NO₂¹, diminishing its potent vasodilatory effects in the systemic circulation. COPD can cause an increase in expired NO· (Clini et al., 2001) compared to healthy controls.

# 1.3.10: COPD therapies and treatments:

There are numerous therapies used in the treatment of individuals with COPD and its associated co-morbidities, some of which are briefly reviewed below.

# Smoking cessation

Smoking cessation is the only known intervention that has been shown to slow the progression of the disease (Barnes, 2003) and therefore is the most important intervention (Pauwels, 2000). An aggressive smoking cessation program can significantly reduce the progressive decline of FEV<sub>1</sub> shown in smokers (Anthonisen et al., 1994), in addition to improving long term prognosis in individuals with mild-moderate COPD (Scanlon et al., 2000).

#### **Broncodilators**

COPD is characterised by airway obstruction caused via airway narrowing in combination with cholinergic vagal bronchoconstriction (GOLD., 2011). The ability to use a drug in order to vasodilate the airways is key for day to day activities of individuals with COPD but also to reduce symptoms during exacerbations. There are currently 3 classes of broncodilators in clinical use: anticholinergics, \$\mathcal{B}\_2\$-sympathomimetic agonists and methylxanthines. All these classes of drugs when administered work via smooth vessel relaxation. The mechanism of action however, is different. Anticholinergics cause antagonism of acetylcholine within the airway smooth muscle (Barnes, 1995) and \$\mathcal{B}\_2\$-sympathomimetic agonists stimulate \$\mathcal{B}\_2\$-recptors inducing elevations in cAMP (Lulich et al., 1988), methylxanthines work by inhibiting phosphodiesterase which breaks down cAMP.

#### Simvastatin & Beta-blockers

Simvastatin, a drug usually used to lower cholesterol, has been shown to reduce pulmonary arterial hypertension (PAH (Sun and Ku, 2008, Girgis et al., 2003)) in animal models (male adult Sprague-Dawley rats) perhaps due to their anti-inflammatory, antioxidant and antithrombotic effects (Wang et al., 2008a). However,

Wilkins et al. (2010) provide evidence that simvastatin does also enhance exercise tolerance, as measured via the 6MWT (n = 42 in a parallel group design).

Beta-blockers are often prescribed for COPD patients with cardiovascular comorbidity. Approximately 27% of individuals with COPD have coronary heart disease (Karoli and Rebrov, 2005). Beta-blockers have been shown to decrease mortality in individuals with severe COPD (Chen et al., 2001) and improve outcomes following a myocardial infarction (Quint et al., 2013). Their use in some patients is limited by bronchoconstriction (Albouaini et al., 2007).

#### Sildenafil

Sildenafil is used to treat/reduce pulmonary hypertension and secondly it is used to treat erectile dysfunction. Both of these effects are mediated by an inhibition of phosphodiesterase type 5 (PDE-5) enzyme which lowers the activity of PDE-5 via competitive inhibition of cGMP (Corbin, 2004). Therefore, cGMP concentrations rise. Intracellular cGMP levels are key as they play a critical role in the activation of protein kinases, protein phosphorylation and subsequent smooth muscle cell (SMC) relaxation via calcium modulation (Lucas et al., 2000, Wang et al., 2008b). There are a total of 11 known PDE isoenzymes which are all thought to act on specific tissue targets and substrates (Ramani and Park, 2010). Critically, PDE-5 has been a focus for PAH therapeutics due to its high expression with lung tissues, vascular beds and systemic circulation (Francis and Corbin, 2005). Sildenafil inhibits PDE-5 which extends the half-life of cGMP (the intracellular mediator for NO· vasodilatory pathways) and enhances SMC relaxation creating an increase in pulmonary vasodilation (Steiner et al., 2005).

# Nitric Oxide gas

NO· inhalation has been shown to have both positive and negative effects in individuals with COPD. In individuals with COPD and CVD, it can reduce peripheral vascular resistance (Kieler-Jensen et al., 1994) and it has been shown to increase the exercise capacity of heart failure patients with pulmonary hypertension (Koelling et al., 1998). However, there is evidence to suggest that in some individuals the NO-gas can lead to increased venous return to the lungs which can cause elevated pulmonary capillary pressure and exacerbate pulmonary hypertension in heart failure patients (Ichinose et al., 2004). Furthermore, to gain the greatest beneficial effects, the NO· needs to be administered with a small amount of O<sub>2</sub> (Yoshida et al., 1997). This has been shown to enable a greater PaO<sub>2</sub> in comparison to breathing O<sub>2</sub> alone or when mixed with air (Barberà et al., 1996). O<sub>2</sub> and NO· inhalation has subsequently been shown to work over a 3 month period with decreases in pulmonary arterial pressure and pulmonary vascular resistance with increases in cardiac output (Vonbank et al., 2003).

## 1.4: Aims, objective and hypothesis

Dietary NO<sub>3</sub> supplementation appears to offer a low cost therapeutic supplement that may alter the O<sub>2</sub> cost of exercise, improve exercise performance, lower blood pressure (in individuals with T2DM and COPD) alter hepatic diffusion and improve glucose homeostasis in healthy young and older adults.

The global aim of this thesis is to assess the therapeutic effects of inorganic dietary NO<sub>3</sub><sup>-</sup> supplementation in individuals with type 2 diabetes (T2DM), chronic obstructive pulmonary disease (COPD) and healthy adults.

Specifically, in individuals with T2DM and COPD, will dietary NO<sub>3</sub>:

- 1. Lower the O<sub>2</sub> cost of exercise?
- 2. Improve walking performance?

Additionally, in healthy young and older adults, can an acute NO<sub>3</sub> rich meal alter:

- 1. Hepatic diffusion?
- 2. Affect glucose homeostasis?

## Secondary objectives;

1. To assess the effects of dietary NO<sub>3</sub> on blood pressure in all the above cohorts.

## Hypothesis

- 1. Dietary nitrate supplementation will reduce the oxygen cost of exercise in individuals with T2DM and COPD.2. Dietary nitrate supplementation will improve walking performance in individuals with T2DM and COPD.
- 3. Dietary nitrate supplementation will reduce blood pressure in individuals with T2DM and COPD.
- 4. Dietary nitrate supplementation will increase portal vein velocity, flux and microvascular diffusion in healthy young and older adults.
- 5, Dietary nitrate supplementation will reduce postprandial plasma glucose concentrations in healthy young and older adults.
- 6. Dietary nitrate supplementation will reduce blood pressure in healthy young and older adults.

#### 2.0: Methods

#### 2.1: Chemiluminescence

# 2.1.1: Blood collection and sampling

Blood sampling and processing were identical for all three experimental chapters. Free flowing blood samples for NO<sub>3</sub> and NO<sub>2</sub> analysis were drawn into room temperature lithium heparin tubes (Sarstedt S-Monovette, Nümbrecht, Germany). Samples were immediately centrifuged at 3600 rpm for 10 minutes at 4°C. The plasma was then aliquoted into (barcoded eppendorf's for experimental chapter 1) eppendorf's and flash frozen in liquid nitrogen prior to storing in -80°C freezers. Plasma samples were stored with minimal headspace between the sample and lid to minimise NO· / NO<sub>2</sub> contamination. Eppendorf's were triple washed and left to dry overnight to remove NO<sub>2</sub> contamination. Determination of plasma NO<sub>3</sub> and NO<sub>2</sub> were performed nitric oxide concentrations on а analyser via ozone chemiluminescence (Sievers NOA 280; Analytix Ltd, Durham, UK).

## 2.12: Deproteinisation

Samples were thawed in batches at room temperature on the laboratory bench. Samples were only used for a single freeze thaw cycle. Samples were completely defrosted prior to analysis (~1 hour) and deproteinised as soon as thawing had occurred. 200 µl of sample was added to the triple washed eppendorf's. 400 µl of 1M sodium hydroxide (NaOH) was added to the eppendorf, vortexed for 1 minute and incubated for 10 minutes (at room temperature). 400µl of zinc sulphate (ZnSO<sub>4</sub> (10% w/v)) was added and vortexed once more and left to stand for a further 10 minutes. Samples were centrifuged for 5 minutes and decanted into triple washed eppendorfs.

Fresh reagents used in the deproteinisation process were created each day to prevent the accumulation of NO· from the ambient air (Pelletier et al., 2006).

# 2.1.3: $NO_3^-$ analysis

For both NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> assays, following their respective reduction of NOx content to NO·, the NO· reacted with ozone and created nitrogen dioxide, which on creation emits light (via the release of a photon). This luminescence is detected by a thermoelectrically cooled, red-sensitive photomultiplier tube. This tube is held within the chemiluminescence nitric oxide analyser (Sievers NOA 280i, Analytix Ltd, Durham, UK) (see figure 3).

$$NO \cdot + O_3 \rightarrow NO_2^- + O_2$$

$$NO_2^- \rightarrow NO_2 + \lambda v$$

 $\Lambda v = poton$ 

 $O_3 = Ozone$ 

The analysis of NO<sub>3</sub> was performed with an injection of 100µl of deproteinised plasma and performed in duplicate. To analyse plasma NO<sub>3</sub> concentration it was reduced in a stepwise order to NO· via a 5ml solution of vanadium (III) chloride (VCI) in 1M hydrochloric acid (0.8% w/v) at 95°C. To quantify NO· concentration within the plasma samples it was bubbled through the purge vessel (where the VCI reduced NO<sub>3</sub> to NO·), condenser and NaOH trap with an inert gas (nitrogen). The NaOH trap was required in order to neutralise any corrosive gases that had bubbled through with the NO· and nitrogen prior to reacting the analyser.

The effectiveness of the reagents diminished over the course of the assay. For NO<sub>3</sub> analysis the VCI remained stable for ~12 injections. The reagents reduced NO<sub>3</sub> at a slower speed. This manifested as an area under the curve that was stable but the peak had lowered. In order to account for this a biological standard was used at the start and end of each assay and the known concentration checked. The assay was accepted if the biological standard was within 10% of the previously determined inter assay mean. Reagents were changed in a timely manner, usually for each assay.

Equation 1: Plasma NO<sub>3</sub> reduction

$$2NO_3^- + 3V_3^+ + 2H_2O \rightarrow 2NO + 3\dot{V}O_2^+ + 4H^+$$

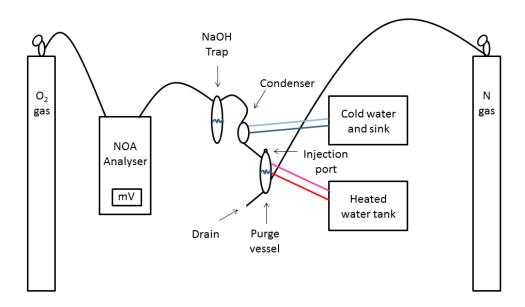


Figure 3. NOA analyser and associated equipment for nitrate reduction.

Depicts the NOA analyser and associated equipment for nitrate reduction to NO $\cdot$ . 100µl of deproteinised plasma was injected in the injection port. The VCl reduced the nitrate and nitrite to NO $\cdot$ , where nitrogen bubbles the NO $\cdot$  through the purge vessel, condenser, NaOH trap and into the analyser.

# 2.1.4: NO<sub>3</sub> assay validation

To ensure that  $NO_3^-$  was being accurately measured  $NO_3^-$  spiked samples were used. A plasma standard was created by following the blood collection procedures above and subsequently pooling 300 ml of plasma from 3 individuals in a  $NO_3^-$  and  $NO_2^-$  free sterile beaker prior to aliquoting. Post deproteinisation spiked recovery was used to quantify the concentration of the  $NO_3^-$  and  $NO_2^-$  concentration in the plasma samples. Two separate, known concentrations of (100 $\mu$ l) standards (typically 1 $\mu$ M and 5 $\mu$ M) were added to separately to different eppendorf's with 100 $\mu$ l of biological standard prior to deproteinisation. These were vortexed for ~1 minute and deproteinised. 4x 200 $\mu$ l of plasma was also deproteinised and analysed to compared with the spike recovery concentration and repeated on numerous days. An average of the 4x 200 $\mu$ l of plasma was taken for the equation.

(Average + standard concentration / 2) / (concentration of the spiked plasma x 100)

Equation 2: A working example (spike recovery):

$$((44.7 \mu M + 20 \mu M) / 2 = 32.35 \mu M) / (34.55 \mu M \times 100 = 93.6\%)$$

Initial attempts to perform a spike recovery revealed that the method was either not being reliably executed or that there was contamination. The initial spike recoveries ranged between 140 – 210% of predicted concentration. Standard operating procedure was being strictly followed and therefore it was more likely to be explained by contamination. To get a reliable spike recovery (SR) for the plasma NO<sub>3</sub><sup>-</sup> concentration distinct methodological changes were required. Numerous

amendments to the protocol were required in order to establish where the poor repeatability was emanating from. Firstly, the vortex was used for a longer period to ensure the plasma was mixed well with the ZnSO<sub>4</sub> and NaoH. This was done for ~60 seconds and the mixing was confirmed by eye. A visible swirl of the mixture can be seen moving down the Eppendorf. When it reached the bottom of the Eppendorf it was mixed. This maximised the removal of as much protein in the precipitate.

Another key aspect was to use VCI as the purge vessel was warming up. Previously, the vessel was washed with deionised water. The high temperature and the VCI cleaned the vessel of all detergent and debris. Prior to the analysis fresh reagent was added to the reaction chamber. This step allowed for a repeatable standard curve. When these errors were corrected the spike recovery was much more reliable. The final five plasma samples averaged  $45.68 \pm 2.91 \,\mu\text{M}$ , the spike recovery with  $10\mu\text{M}$  sodium  $NO_3^-$  concentration averaged  $95.45 \pm 4.1 \,\mu\text{M}$  % and the  $20\mu\text{M}$  sodium  $NO_3^-$  concentration averaged  $97.23 \pm 4.6 \,\mu\text{M}$  %.

Table 2. Nitrate plasma standards and spike recovery reproducibility.

This table depicts the raw values, means, SD and CV for plasma standards, and two different spike recovery concentrations (10 and 20 µM).

	Plasma standard		
Nitrate	μМ	SR10	SR20
	48.4	97.3	93
	47.7	95.7	100
	41.9	99.8	102.3
	44.7	89	93.6
	46.7	93.5	N/A
	50.1	N/A	N/A
Mean	46.58	95.06	97.23
SD	2.91	4.09	4.63
CV %	6.252	4.307	4.767

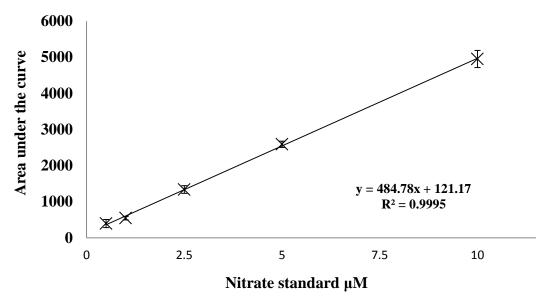


Figure 4. NO<sub>3</sub>, representative standard curves.

This figure depicts the average  $\pm$  SD of 10 representative NO $_3$  standard curves

# 2.1.5: $NO_2^-$ analysis

To analyse plasma  $NO_2^-$  concentration it was reduced to  $NO_2^-$  via 5ml glacial acetic acid and 1ml sodium lodide (NaI) (0.5% w/v) solution at 30°C. To quantify  $NO_2^-$  (post reagent reduction) within the plasma samples it was carried through the reduction agents with nitrogen. Note the NaOH trap was not required with this reaction. The analysis of  $NO_2^-$  was performed with an injection of 100µl of deproteinised plasma and performed in duplicate. For  $NO_2^-$ , the acetic acid and NaI was able to reduce the  $NO_2^-$  to  $NO_2^-$  for longer (~20 injections) with consistent peaks.

Equation 3: Plasma NO<sub>2</sub> reduction

$$2I^{-} + 2NO_{2}^{-} + 4H^{+} \rightarrow 2NO + I_{2} + 2H_{2}O$$



Figure 5. NOA analyser.

Depicts the NOA analyser and equipment used to reduce the NO<sub>2</sub> to NO·.

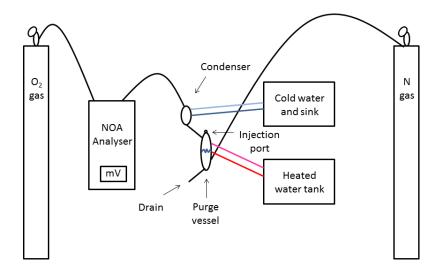


Figure 6. NOA analyser and associated equipment.

Shows the NOA analyser and associated equipment for nitrite reduction to NO·. 100µl of deproteinised plasma was injected in the injection port. The Nal reduced the nitrite to NO·, where nitrogen bubbles the NO· through the purge vessel, condenser and into the analyser.

To quantify the concentrations of  $NO_3^-$  and  $NO_2^-$  standard curves were created daily using sodium  $NO_3^-$  at known concentrations of 500nM to 10 $\mu$ M and sodium  $NO_2^-$  from 10nM to 250nM and the luminescence was plotted via signal area in mV. At least 5 points were used to create each curve.

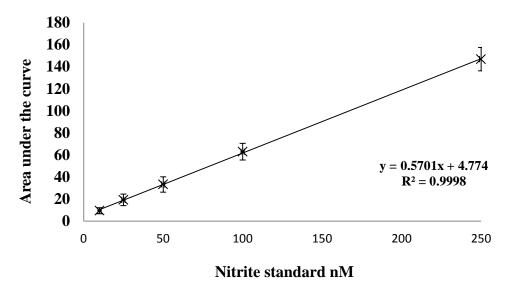


Figure 7. Nitrite, representative standard curves.

This figure depicts the average ± SD of 10 representative NO<sub>2</sub> standard curves.

Table 3. Nitrite plasma standards and spike recovery reproducibility.

Table depicts the raw values, means, SD and CV for plasma standards, and two different spike recovery concentrations (100 and 250 nM).

Nitrite	Plasma standard nM	SR100	SR250
	508	118	93
	502.5	127	102
	518.5	119	94
	571	117	100
	547	132	Missing value
	493	111	118
Mean	523.33	120.67	101.40
SD	28.90	6.58	4.43
CV %	5.522	5.453	4.364

# 2.2: O<sub>2</sub> uptake measurement and analysis

For experimental chapter 3 and 4, breath-by-breath pulmonary gas exchange and ventilation were measured during walking (MedGraphics, CardiO<sub>2</sub> Cardiopulmonary Diagnostic Systems, St. Paul, MN, USA) and cycling exercise (Vmax<sup>TM</sup> Encore, Yorba, Linda, CA) respectively. Before each session the O<sub>2</sub> and CO<sub>2</sub> analysers were calibrated using known concentration gases. The volume transducer was calibrated using a 3-litre syringe (Hans Rudolph, Kansas City, MO, USA).

Participant's breath-by-breath  $\dot{V}O_2$  data were initially checked for 'erroneous' breaths. These breaths can be caused by coughing and swallowing and were

removed (Lamarra et al., 1987). Breaths over 4 standard deviations from the local mean were removed. For each individual breath-by-breath data were time aligned and interpolated for each transition to give second by second values (chapter 3 had 3 transitions and chapter 4 had two). A nonlinear least squares algorithm was then used to fit the ensemble-averaged data, as described in the following equation.

Equation 4: Nonlinear least square algorithm:

$$\dot{V}O_2$$
 (t) =  $\dot{V}O_2$  baseline +  $A_p(1-e^{-(t-TDp)/\tau p})$ 

where  $\dot{V}O_2$  (*t*) represents the absolute  $\dot{V}O_2$  at a given time *t*;  $\dot{V}O_{2\text{baseline}}$  represents the mean  $\dot{V}O_2$  in the baseline period (when participants were stood still on the treadmill or when cycling at 0 watts);  $A_p$ ,  $TD_p$ , and  $\tau_p$  represent the amplitude, time delay, and time constant, respectively, describing the increase in  $\dot{V}O_2$  above baseline when exercise had commenced. The end-exercise  $\dot{V}O_2$  was defined as the mean  $\dot{V}O_2$  measured over the final 30 seconds of exercise. The mean response time (MRT) was calculated by fitting a single exponential curve to the data from the onset to the end of exercise.

Equation 5 Nonlinear least square algorithm (without TD):

$$\dot{V}O_2(t) = \dot{V}O_2$$
 baseline + Ap(1-e<sup>-(t)/Tp</sup>)

The  $O_2$  deficit was calculated as the product of the  $\dot{V}O_2$  response amplitude (i.e. the baseline to the point that a steady state was attained) and the MRT. The coefficient of variation for day to day variability of  $O_2$  uptake for treadmill walking using the MedGraphics equipment can be seen in table 4.

Table 4.  $\dot{\mathbf{V}}\mathbf{O}_2$  repeatability data.

This table shows day to day  $\dot{V}O_2$  repeatability. Data from 1 individual after interpolation and modelled as described above for 3 transitions, with the mean, SD and coefficient of variation calculated over 3 separate days.

	Time constant	MRT	O <sub>2</sub> deficit	Baseline	Amplitude	End <b>V</b> O <sub>2</sub>
Day 1	42.35	47.62	0.86	296.15	1488.17	1744.62
Day 2	42.18	48.40	0.84	306.88	1467.04	1737.20
Day 3	41.82	48.92	0.78	327.62	1357.33	1641.72
Mean	42.12	48.31	0.83	310.21	1437.51	1707.85
SD	0.22	0.54	0.03	13.06	57.35	46.86
CV %	0.52	1.11	3.99	4.21	3.99	2.74

For the experimental overview, exercise modality and transitions, please see individual experimental chapters.

# 2.3: The apparent diffusion co-efficient of the liver, portal vein flux and velocity For experimental chapter 3 a 1.5 T Philips (Amsterdam, The Netherlands) magnetic resonance imaging (MRI) scanner was used in order to examine modifications in velocity and flow to the liver (portal vein) and estimate capillary diffusion in the

posterior right lobe of the liver following NO<sub>3</sub> or placebo supplementation.

The human body is comprised predominantly of water. An MRI scanner creates a homogenous magnetic field which allows protons within hydrogen nuclei to occupy one of two distinct energy states. If radiofrequency radiation is then transmitted at the appropriate frequency it is possible to cause transitions of protons between these

two energy levels. The presence of protons in the excited energy state can be detected via the use of appropriate detector devices and it is this mechanism that gives rise to the means of generating MRI images.

## 2.3.1: Portal vein (flow & velocity);

Initially, structural scans were obtained in different planes to identify the location and orientation of the portal vein. Subsequently, an 8mm thick imaging slice was selected that lay perpendicular to the long axis of the vein. Landmarks (such as the inferior vena cava) were used to ensure replication of slice positioning between each repeat measurement and for visits 1 and 2. The circumference of the portal vein was checked between all scans from V1 and V2. To determine flow and velocity a cardiac triggered velocity sensitive phase encoding imaging sequence (Gatehouse et al., 2005) was employed which obtained image data at 20 time points throughout the cardiac cycle. Analysis was subsequently undertaken using a package supplied as part of the general philips scanner software. For each separate measurement the circumference of the vessel was manually drawn and recorded for each of the 20 time points to establish a defined region of interest (ROI). Within this ROI, flow and velocity were automatically calculated to give profiles throughout the cardiac cycle.

# 2.3.2: Apparent diffusion coefficient (ADC);

When examined as a group of voxels, capillary flow represents a pseudo-diffusion process, such that fluid flow is taking place in a 3D isotropic fashion. To examine this, a magnetic resonance (MR) sequence sensitive to flow, via the application of magnetic field gradients in three orthogonal directions, was employed. From this a parameter known as the apparent diffusion coefficient (ADC), dependent upon capillary flow averaged over all directions, could be determined. To calculate ADC within the posterior left lobe of the liver, a ROI (typical dimensions = 2500mm<sup>3</sup>) was

manually drawn using the scanner software and the signal intensity within determined. ADC was subsequently calculated based upon the ratio of signal intensity from the two images generated from the MR sequence employed, one of which was sensitive to flow, whereas the other had a low sensitivity to flow, where:

Equation 6: ADC:

ADC=  $1/(b_1-b_0)$  Ln  $(S_0/S_1)$ 

S<sub>0</sub> signal intensity in low flow sensitivity image

S<sub>1</sub> signal intensity in flow sensitive image

b<sub>0</sub> magnetic field gradient used in low flow sensitivity image=250 s/mm<sup>2</sup>

b<sub>1</sub> magnetic field gradient used in flow sensitive image=750 s/mm<sup>2</sup>

# 2.3.3: ADC and portal vein repeatability

To establish the repeatability of the portal vein velocity (see table 5 below) and ADC measurement (see table 6 below) 6 different, fasted individuals were assessed over two separate mornings. A CV was created between each of the ADC indices and then averaged.

Table 5. MRI reproducibility.

This table describes the CV for the portal vein flux and velocity for 6 fasted individuals on two separate mornings.

	Portal vein flux	Portal vein velocity	ADC
Mean	16.64	13.61	1.15
SD	2.28	2.18	0.12
CV %	13.36	16.366	9.77

Table 6. Description of multiple ROI and its effect on reproducibility.

This table describes the mean, SD and CV for the ADC for 6 fasted individuals on two separate mornings. Different numbers of regions of interests (ROI) were used to find the optimum number to increase repeatability.

	ADC	SD	CV
6 ROI	1.09	0.33	30.32
3 ROI	1.10	0.23	21.34
2 ROI	1.11	0.21	15.29
1 ROI	1.15	0.12	9.77

For study analysis the ADC and the portal vein assessment (velocity and ADC), the baseline scans of each day are compared with 1, 2 and 3 hours post supplementation. See experimental chapter 3 for a description for detailed description of experimental procedures.

#### 2.4: Standardised breakfast

For chapter 5, (experimental chapter 3 / DiMPLe) the participants came to the MRI research centre at St Luke's Campus in a fasted state. Participants were asked to fast from 10pm the night before however water was admissible. Approximately 1 hour after arrival participants were provided with breakfast. Each participant was supplemented (breakfast) with 76g of carbohydrates which is comparable to the glucose dose given in an OGTT (75g(Bartoli et al., 2011b)). The carbohydrate supplement was in the form of 2 beetroot juice sports shots (32g), 2 slices of brown bread (25g), butter (~0g) and jam (19g).

An OGTT test begins with an overnight fast and an acclimatisation period with the patient seated throughout the testing period. A baseline plasma glucose

measurement is then taken. Two hours after the carbohydrate bolus another plasma glucose measurement is taken. In 1997 the American Diabetes Association(Gavin et al., 1997) lowered the fasting plasma glucose measurement cut criteria for impaired fasting glycaemia from 140 mg/dl (7.7) to 126 (6.99mmol/L). In 1979 the national diabetes data group set a distinct cut of value for impaired glucose tolerance which was given following the 2 hour post plasma glucose concentrations for impaired glucose tolerance as 140 (7.7) - 199 mg/dl (11mmol/L) (NDDG, 1979). In the present study (experimental chapter 3), blood glucose concentrations were measured every hour following the standard breakfast for 3 subsequent hours.

# 2.5: General data handling and methods

Prior to data collection for any of the experimental chapters Microsoft Access (Redmond, WA, USA) databases were set up with all appropriate variables encoded. Data was double entered by an independent researcher. When both data sets were entered and the study complete a macro was run between the files to check for anomalies. All discrepancies were checked within the clinical records folder and amended into one database. Patient identifiable data were entered into a separate database and saved on a non-networked password encrypted hard drive in the diabetes and vascular research centre.

Variables were transposed into SPSS software version 21.0 (Chicago, IL, USA). All data were tested for normality prior to determination of statistical processing (i.e. parametric or non-parametric). Statistical difference was accepted when P < 0.05. For specific randomisation blinding and statistical tests please see experimental chapters.

## 3.0: Experimental chapters

# 3.1: Experimental chapter 1(Dietary NO<sub>3</sub><sup>-</sup> supplementation in T2DM)

Title: Effects of dietary nitrate supplementation on the oxygen cost of exercise and walking performance in individuals with type 2 diabetes: a randomised, double blind, placebo-controlled cross-over trial.

3.1.1: Authors: Anthony I Shepherd<sup>1,2</sup>, Mark Gilchrist<sup>2</sup>, Paul G Winyard<sup>2</sup>, Andrew M Jones<sup>1</sup>, Ewelina Hallmann<sup>3</sup>, Renata Kazimierczak<sup>3</sup>, Ewa Rembialkowska<sup>3</sup>, Nigel Benjamin<sup>2</sup>, Angela C Shore<sup>2</sup>, Daryl P Wilkerson<sup>1</sup>

3.1.2: Affiliations: College of Life and Environmental Sciences, Sport and Health Sciences, University of Exeter, Devon, UK, <sup>2</sup> University of Exeter Medical School and NIHR Exeter Clinical Research Facility, Royal Devon and Exeter Hospital, Exeter, Devon, UK, <sup>3</sup> Warsaw University of Life Sciences, Warsaw, Poland.

## 3.1.3: Abstract

Background: Dietary NO<sub>3</sub><sup>-</sup> supplementation has been shown to reduce the O<sub>2</sub> cost of exercise and enhance exercise tolerance in healthy individuals. This study assessed whether similar effects could be observed in individuals with type 2 diabetes (T2DM).

Methods: In a randomised, double blind, placebo-controlled cross-over study, 48 participants with T2DM supplemented their diet for four days with either NO<sub>3</sub><sup>-</sup> rich beetroot juice (70 ml/day, 6.43 mmol NO<sub>3</sub><sup>-</sup>/day) or NO<sub>3</sub><sup>-</sup> depleted beetroot juice as placebo (70 ml/day, 0.07 mmol NO<sub>3</sub><sup>-</sup>/day). After each intervention period, resting plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations were measured subsequent to participants completing moderate-paced walking. Pulmonary gas exchange was measured to

assess the O<sub>2</sub> cost of walking. Following a rest period, participants performed the six-minute walk test (6MWT).

Results: Relative to placebo, beetroot juice resulted in a significant increase in plasma NO<sub>3</sub><sup>-</sup> (placebo; 57 ± 66 vs. beetroot juice; 319 ± 110  $\mu$ M; P <0.001) and plasma nitrite concentration (placebo; 680 ± 256 vs. beetroot juice; 1065 ± 607 nM; P <0.001). There were no differences between placebo juice vs. beetroot juice for the O<sub>2</sub> cost of walking (placebo; 946 ± 221 vs. beetroot juice; 939 ± 223 ml·min<sup>-1</sup>, respectively; P =0.59), and distance covered in the 6MWT (placebo; 550 ± 83 vs. beetroot juice; 554 ± 90m, respectively; P =0.17).

Conclusion: Nitrate supplementation did not affect the O<sub>2</sub> cost of moderate-paced walking or improve performance in the 6MWT. These findings indicate that dietary NO<sub>3</sub><sup>-</sup> supplementation does not modulate the response to exercise in individuals with T2DM.

## 3.1.4: Introduction

Individuals with type 2 diabetes mellitus (T2DM) have profound reductions in their tolerance to exercise compared to healthy individuals (Regensteiner et al., 1995b). The discomfort experienced by individuals with T2DM during exercise may impact upon their ability or willingness to attain their recommended level of exercise, which is a key aspect of disease management (NICE, 2008). Exercise intolerance in this population has been attributed to abnormalities at multiple points during the transport of O<sub>2</sub> from the lungs to its site of utilisation in the muscles (Scognamiglio et al., 1998, Regensteiner et al., 1995a, Regensteiner et al., 1998). There is evidence that individuals with T2DM have reductions in the biosynthesis and bioavailability of the biological messenger nitric oxide (NO·) (Xu et al., 2007, Meininger et al., 2000). NO·

is known to play an important role in muscle contractility, skeletal muscle glucose uptake, calcium handling (Stamler and Meissner, 2001), vascular function, blood flow regulation (Dejam et al., 2007), and mitochondrial respiration and biogenesis (Cooper and Giulivi, 2007).

There are two known pathways by which NO· is synthesised, the L-arginine pathway and the entero-salivary pathway. The L-arginine pathway involves the synthesis of NO· from L-arginine by the nitric oxide synthase (NOS) family of enzymes. The entero-salivary pathway involves a stepwise conversion of NO₃⁻ to NO₂⁻, and subsequently, NO₂⁻ to NO·. Briefly, NO₃⁻ from the diet (or from NO·/ NO₂⁻ oxidation) is absorbed into the circulation where it is concentrated in the salivary glands. NO₃⁻ is then reduced to NO₂⁻ via facultative anaerobic bacteria on the surface of the tongue (Duncan et al., 1995) and subsequently swallowed. Some of the swallowed NO₂⁻ is reduced to NO· in the acidic environment of the stomach, with important local effects on gastric function and protection against enteric pathogens (Gilchrist et al., 2010) and the remainder enters the circulation. Circulating NO₂⁻ acts as a reservoir for NO·, with its reduction to NO· potentiated in acidic or hypoxic areas, such as contracting skeletal muscle (Bryan, 2006). It has been suggested that the enterosalivary pathway is a complementary system for NO· synthesis (Lundberg et al., 2008).

There is extensive evidence in healthy individuals that NO<sub>3</sub><sup>-</sup> supplementation, via either sodium NO<sub>3</sub><sup>-</sup> or NO<sub>3</sub><sup>-</sup> rich beetroot juice, increases plasma NO<sub>2</sub><sup>-</sup> concentration (Bailey et al., 2009, Lundberg and Govoni, 2004), lowers blood pressure (BP) (Bailey et al., 2010b, Larsen et al., 2006), and reduces the O<sub>2</sub> cost of exercise (Bailey et al., 2009, Larsen et al., 2007a, Wylie et al., 2013a). The reduction in the O<sub>2</sub> cost of exercise has been proposed to be related to a reduction in the ATP cost of muscle

force production (Bailey et al., 2010b), an improvement in mitochondrial efficiency (Larsen et al., 2011), or a combination of both (Bailey et al., 2012).

It has been reported that the bioavailability of NO is reduced in individuals with T2DM due to increased scavenging and/or reduced synthesis. Reactive oxygen species (ROS) (Crabtree et al., 2011) and hyperglycaemia (Ding et al., 2004) may result in lower L-arginine derived NO. Further, there is evidence that NO. production is suppressed by NOS inhibitors such as asymmetric dimethylarginine (ADMA) (Abbasi et al., 2001) and caveolin-1 (Catalán et al., 2008), which are elevated in T2DM. Conversely, others have reported increased plasma NO<sub>3</sub> and NO<sub>2</sub> concentrations in individuals with T2DM (Ghasemi et al., 2010, Chien et al., 2005), which may indicate a quenching of NO activity or upregulation of NO synthesis to counteract resistance. It is possible that dietary NO<sub>3</sub> supplementation in this population could ameliorate the impact of T2DM on NOS dependent NO· synthesis by increasing the amount of NO produced via the entero-salivary pathway (Lundberg et al., 2008). We recently reported no effect of dietary NO<sub>3</sub><sup>-</sup> supplementation on the 24h ambulatory BP in individuals with T2DM (Gilchrist et al., 2013), but the impact on the O2 cost of exercise, and exercise tolerance in individuals with T2DM remains unknown.

The aim of this study was, therefore, to examine the effects of dietary NO<sub>3</sub><sup>-</sup> supplementation (NO<sub>3</sub><sup>-</sup> rich beetroot juice) on the O<sub>2</sub> cost of exercise and walking performance, and to confirm our previous finding regarding resting BP, in individuals with T2DM. It was hypothesised that, compared to NO<sub>3</sub><sup>-</sup> depleted beetroot juice, NO<sub>3</sub><sup>-</sup> rich beetroot juice would reduce the amount of O<sub>2</sub> required per unit of time to perform moderate-paced walking, and increase the distance covered in the 6-minute walk test.

## 3.1.5: Methods

#### **Patients**

Forty eight patients (35 males) with T2DM (see Table 7 for characteristics) volunteered to participate in this randomised, double blind, placebo-controlled crossover study. Participants were recruited from the NIHR Exeter Clinical Research Facility, Exeter 10,000 cohort, a database of individuals who have consented to be contacted for research. Ethical approval was granted by the Cornwall and Plymouth NRES Committee 12/SW/0118 and the study was registered as a clinical trial on the ClinicalTrials.gov website, ID # NCT02206074. All participants provided written informed consent. Patients were recruited if they had been diagnosed with T2DM (as defined by the WHO) at least five years prior to enrolment in the study and were 35-75 years of age. Patients were excluded from the study if they had significant renal impairment (eGFR <30), uncontrolled hypertension, a BMI <25 or >35 (kg/m<sup>2</sup>), a history of myocardial infarction or cerebro-vascular event, were taking regular organic NO<sub>3</sub> or nicorandil, or were smokers (or who had stopped smoking within the previous three months). Patients taking phosphodiesterase inhibitors were asked to refrain from doing so for the duration of the study. On experimental days, participants were asked to arrive at the laboratory in a rested and fully hydrated state, at least 3 hours postprandial, and having avoided caffeine and alcohol for 6 and 24 hours, respectively. Participants were also asked to avoid exercise for 24 hours prior to each testing session. Participants were asked to record their diet for 24 hours prior to each experimental visit. Following the crossover the participant were asked to replicate their previous diet and this was verbally confirmed at the second visit.

Table 7. Characteristics of patients included in the final analysis. Data are mean  $\pm$  SD, or as a % of the cohort on a medication.

Age (years)	63.35 ± 7.27
Diabetes Duration (years)	10.31 ± 5.26
HbA1c (mmol/mol)	60.83 ± 13.37
Body Mass Index (kg/m²)	$30.17 \pm 2.93$
Baseline systolic BP (mmHg)	142.13 ± 12.84
Baseline diastolic BP (mmHg)	$80.85 \pm 7.36$
No. of 30+ minutes of exercise per week	3.83 ± 3.16
Portions of fruit and vegetables per day	$3.83 \pm 2.00$
Metformin %	83.33
Insulin %	25.00
ARB & ACEi %	47.92
Statins %	83.33

Participants attended the NIHR Exeter Clinical Research Facility where we completed informed consent, a medical history, anthropometric measures, and a resting ECG. A familiarisation session was performed for the treadmill exercise, allowing the determination of the walking speed for each participant that would be used in the experimental visits. The target speed was a pace equivalent to the participants' usual walking speed that could comfortably be maintained for 6 minutes.

# **Experimental Overview**

Following the pre-experimental visit, participants were assigned in a double-blind, randomised, crossover design to consume 70 ml/day of  $NO_3^-$  rich beetroot juice (beetroot juice; containing 6.43 mmol of  $NO_3^-$ ; Beet it, James White Drinks Ltd., lpswich) or  $NO_3^-$  depleted beetroot juice (placebo;  $NO_3^-$  depleted beetroot juice

containing 0.07 mmol of NO<sub>3</sub><sup>-</sup>; Beet it, James White Drinks Ltd., Ipswich) for four days. The dose of 6.43 mmol of NO<sub>3</sub><sup>-</sup> equated to 0.072 mmol.kg<sup>-1</sup> per day for 3.5 days. Similar dosing regimens (0.068 (Bailey et al., 2010b); 0.064 (Muggeridge et al., 2013b); 0.086 (Berry et al., 2014)mmol.kg<sup>-1</sup>) to ours have been shown to be effective at this relative dose. Production of the placebo juice has been detailed previously (Lansley et al., 2010), with the final product being indistinguishable in taste, colour, texture, appearance and odour to the NO<sub>3</sub><sup>-</sup> rich beetroot juice (Gilchrist et al., 2014). The final 70 ml of juice was consumed ~3 hours before the commencement of exercise on the morning of testing. Participants were required to abstain from using antibacterial mouthwash and chewing gum throughout the study, as this has been shown to reduce the concentration of oral bacteria responsible for the reduction of NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup> (Govoni et al., 2008).

During the two experimental visits, resting BP was measured and venous blood samples were drawn into lithium-heparin tubes (Sarstedt S-Monovette, Nümbrecht, Germany) and prepared for NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> analysis as previously described (Gilchrist et al., 2013). Subsequently, participants performed 3 bouts of walking on a motorised treadmill (NordicTrack T14.0, Chaska, MN, USA) at the target speed that was determined in the pre-experimental visit. Each bout was preceded by a resting baseline period of three minutes, with the three bouts separated by 15 minutes of passive recovery. Following a further 15 minutes of seated rest, participants performed a 6 minute walk test (6MWT) to assess functional capacity. This test was conducted indoors on a straight, flat course between 2 cones which were 32.2 meters apart, with patients instructed to cover as much distance as possible in the allotted time. Standardised verbal encouragement was given throughout. Following

their first experimental visit, participants began a washout period (10-14 days) before entering the opposing arm of the study.

#### Measurements

Prior to any exercise testing during the experimental visits, resting BP of the brachial artery was measured using an automated sphygmomanometer (Omron M6, Kyoto, Japan). Five measurements were taken in total, with the mean of the final three measurements being recorded.

For the determination of plasma NO<sub>2</sub> concentration, venous blood samples (~4 ml) were drawn into lithium-heparin tubes (Sarstedt S-Monovette, Nümbrecht, Germany), centrifuged, aliquoted and immediately frozen in liquid nitrogen and stored at -80°C. Prior to analysis all samples were deproteinized using a variant of the protocol used by Higuchi and Motomizu (1999). Fresh reagents were created each day and checked for contamination. No contamination was detectable for water, sodium hydroxide, zinc sulphate or sodium iodide. Determination of plasma NO<sub>3</sub> and NO<sub>2</sub> concentrations was performed on a nitric oxide analyser via ozone chemiluminescence (Sievers NOA 280; Analytix Ltd, Durham, UK) using the protocol described by Bateman et al. (2002).

Pulmonary gas exchange was measured breath by breath during all treadmill walking exercise (MedGraphics CardiO<sub>2</sub> Cardiopulmonary Diagnostic Systems, St. Paul, MN, USA). The volume transducer was calibrated before each test with a 3-liter calibration syringe (Hans Rudolph, Kansas City, MO, USA) and the O<sub>2</sub> and CO<sub>2</sub> analysers were calibrated using gases of known concentration. During treadmill walking exercise, heart rate (HR) was measured every 5 seconds via telemetry (Polar R5400sd, Kempele, Finland).

## Data Analysis

The breath by breath  $O_2$  uptake ( $\dot{V}O_2$ ) data were initially inspected for errant breaths (e.g. associated with coughing and swallowing), with values lying more than four SDs from the local mean being removed. The breath-by-breath data were linearly interpolated to provide second-by-second values and, for each individual, the three bouts were time-aligned to the start of exercise and ensemble-averaged. A nonlinear least-square algorithm was used to fit the data, as described in the following equation:

$$\dot{V}O_2$$
 (t) =  $\dot{V}O_2$  baseline + A<sub>p</sub> (1-e<sup>-(t-TDp)/\taup</sup>)

where  $\dot{V}O_2$  (t) represents the absolute  $\dot{V}O_2$  at a given time t;  $\dot{V}O_{2baseline}$  represents the mean  $\dot{V}O_2$  in the baseline period (when participants were stood still on the treadmill);  $A_p$ ,  $TD_p$ , and  $\tau_p$  represent the amplitude, time delay, and time constant, respectively, describing the increase in  $\dot{V}O_2$  above baseline when exercise had commenced. The end-exercise  $\dot{V}O_2$  was defined as the mean  $\dot{V}O_2$  measured over the final 30 seconds of exercise. The mean response time (MRT) was calculated by fitting a single exponential curve to the data with no time delay from the onset to the end of exercise. The  $O_2$  deficit was calculated as the product of the  $\dot{V}O_2$  response amplitude (i.e. the baseline to the point that a steady state was attained) and the MRT. For a schematic representation of the  $\dot{V}O_2$  kinetics parameters, see Figure 1.

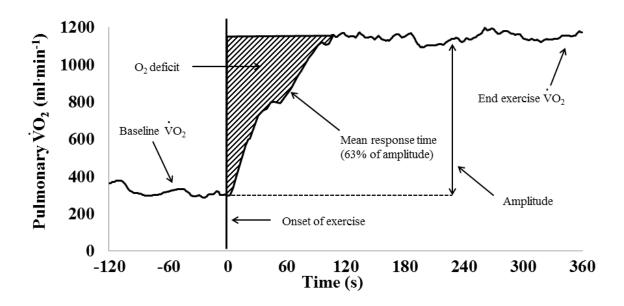


Figure 1. Schematic showing the parameters of the  ${\rm O_2}$  uptake kinetics on a representative participant's data.

## Juice analysis

It has previously been suggested that the antioxidant content of beetroot juice may have beneficial effects (Lansley et al., 2011a). In order to establish whether the concentrations of known antioxidants in beetroot juice (polyphenols and betacyanins) were different in the active and placebo juice, both were analysed using the high-performance liquid chromatography technique according to our previously described methods (Kazimierczak et al., 2014).

## Sample size and randomisation

An *a priori* sample size calculation was performed by a statistician from the University of Exeter Medical School. A pilot study (n=6) in individuals who have T2DM revealed a mean difference between beetroot juice and placebo juice of 45 ml.min<sup>-1</sup> for end exercise  $\dot{V}O_2$  (1SD) during cycling exercise. For 90% power and an  $\alpha$ -level set at P=0.05 (two tailed), to detect a 0.5 SD difference 44 patients were

required. In order to account for dropout, we anticipated that 48 patients would begin the study. Each participant was randomised by a research nurse, who also supplied them with the appropriate juice.

## Data and statistical analysis

All data were tested for normality and are presented as means  $\pm$  standard deviation (*SD*) unless otherwise stated. Differences in plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations, BP,  $\dot{V}$ O<sub>2</sub>, and distance covered in the 6-minute walk test between the conditions were analysed using two-tailed, paired-samples *t*-tests. Where normality of data was not met, differences between the aforementioned data were tested using a non-parametric test (Wilcoxon rank-sum test). When the sample was split for patients taking different classes of medications, independent samples *t*-tests were performed. Pearson product–moment correlation coefficients were used to assess the relationships between variables. Statistical analyses were performed on SPSS software version 21.0 (Chicago, IL, USA), with statistical difference accepted when *P* < 0.05.

#### 3.1.6: Results

48 subjects completed the study. For detailed information regarding participant recruitment and withdrawal, see Figure 8. Participants' self-reported adherence to beetroot juice and avoidance of mouth wash was 100% for both arms of the study. All participants reported similar physical activity and dietary patterns for both supplementation periods. The ingestion of both juices was well tolerated with no adverse side effects. As in previous studies using beetroot juice supplementation, participants reported beeturia (red urine) and red stools (Bailey et al., 2009).

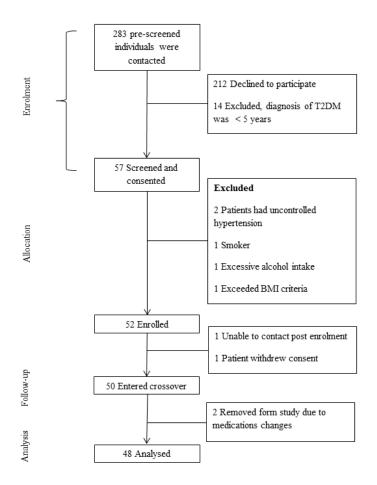


Figure 8. Flow diagram of trial

*Plasma NO*<sub>3</sub> and *NO*<sub>2</sub> concentrations: Relative to placebo, beetroot juice resulted in a significant increase in plasma NO<sub>3</sub> concentration (placebo: 57 ± 66 vs. beetroot juice 319 ± 90μM, P < 0.001, 95% CI -220, 302; Figure 9A). Plasma NO<sub>2</sub> concentration also increased for beetroot juice in comparison to placebo (placebo: 680 ± 256 vs. beetroot juice 1065 ± 607 nM, P < 0.001, 95% CI -220, 548; Figure 9B). There were no differences in baseline (placebo) plasma NO<sub>2</sub> when subjects were split into groups based on drug classes; ACEi and ARB (n= 25; difference *vs.* those not on these drugs -68 ± 75 nM, P = 0.37, 95% CI -218, 83), metformin (n = 40; difference -18 ± 105 nM, P = 0.87, 95% CI -231, 196), insulin (n= 12; difference 3 ± 86 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177) statins (n = 40; difference 164 ± 97 nM, P = 0.97, 95% CI -171, 177)

0.09, 95% CI -31, 360) and sulphonylurea (n = 15; difference 35  $\pm$  134 nM, P = 0.79, 95% CI -237, 306). Other classifications of drugs were prescribed, however the sample size was too small to make inferences; gliptins (n = 2), exenatide (n = 2), sulfazalazine (n = 1), alpha blocker (n = 3), beta blocker (n = 1), calcium channel blocker (n = 6), loop diuretic (n = 1), thiazide (n = 2) P > 0.05).

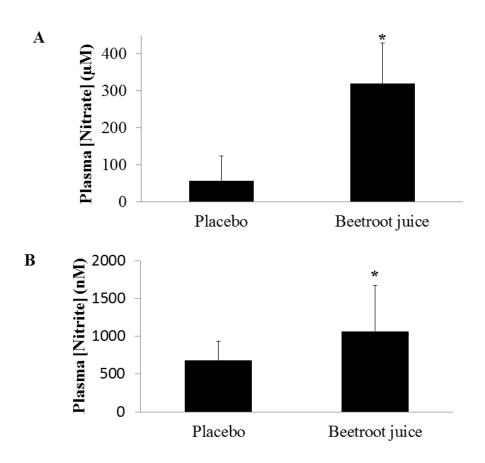


Figure 9. Plasma NO<sub>3</sub> and NO<sub>2</sub> concentration in individuals with T2DM.

A depicts plasma  $NO_3^-$  and B shows plasma  $NO_2^-$  group mean concentrations analysed via chemiluminescence. Values are means  $\pm$  SD. \*significantly different from the placebo P < 0.001.

Treadmill walking,  $\dot{V}O_2$  kinetics, and heart rate: Relative to placebo, beetroot juice supplementation had no effect on baseline (placebo: 282 ± 50 vs. beetroot juice: 281 ± 46 ml.min<sup>-1</sup>; P = 0.82, 95% Cl -12.9, 10.21) or end-exercise  $\dot{V}O_2$  (placebo: 939 ± 223 vs. beetroot juice: 946 ± 221 ml.min<sup>-1</sup>; P = 0.60, 95% Cl -20, 34.3; Figure 10). The time constant and MRT of  $\dot{V}O_2$  response, and the  $O_2$  deficit were also not different between conditions (see Table 8). End-exercise heart rate was not different between conditions (placebo: 96 ± 12 vs. beetroot juice: 94 ± 11 BPM, P = 0.36, 95% Cl -4.7, 1. 7). No significant correlation was observed between the change in plasma  $NO_2$  concentration and end-exercise  $\dot{V}O_2$  (r = 0.04, P = 0.77).

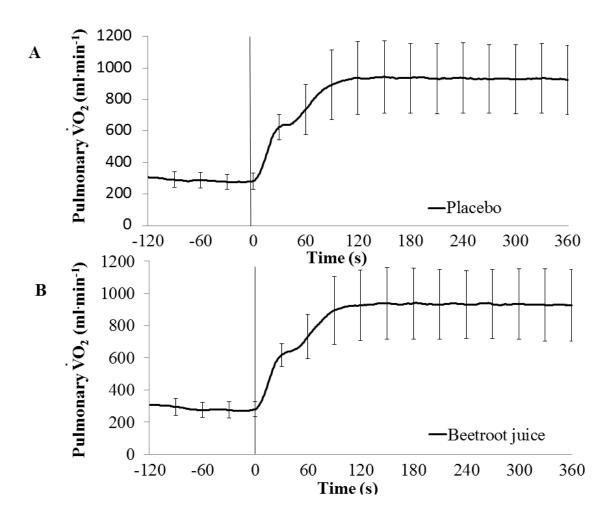


Figure 10. Pulmonary O<sub>2</sub> uptake response to beetroot juice in individuals with T2DM.

The group mean pulmonary  $O_2$  uptake response for placebo (A) and  $NO_3^-$  rich beetroot (B). The vertical line represents the initiation of walking exercise from the standing baseline. Group mean responses are shown with error bars every 30s.

Table 8. O<sub>2</sub> uptake kinetics during walking exercise with placebo and beetroot juice supplementation.

	Placebo	Beetroot juice
Baseline (ml·min <sup>-1</sup> )	282 ± 50	281 ± 46
Primary amplitude (ml·min-1)	656 ± 205	665 ± 199
End-exercise (ml <sup>-1</sup> )	938 ± 223	946 ± 221
O <sub>2</sub> deficit (L)	$0.48 \pm 0.17$	$0.50 \pm 0.19$
Mean response time (s)	45 ± 10	45 ± 10

Six minute walk test: No difference was found between placebo and beetroot juice for distance covered during the six minute walk test (placebo:  $554 \pm 90$  vs. beetroot juice:  $550 \pm 83$  m, P = 0.17, 95% CI -11, 2.04). No difference was observed for distance covered during the six minute walk test when the group was split for ACEi and ARB compared with individuals not on this classification of drug (change -7.6  $\pm$  44.6 m, P = 0.25, 95% CI -20.6, 5.4).

*Blood pressure:* Comparisons between placebo and beetroot juice revealed no statistically significant effect on systolic BP (placebo:  $134 \pm 10 \ vs.$  beetroot juice:  $132 \pm 12 \ mmHg$ , P = 0.17, 95% CI -4.09, 1.17) or diastolic BP (placebo:  $77 \pm 7 \ vs.$  beetroot juice:  $76 \pm 11 \ mmHg$ , P = 0.27, 95% CI -4.04, 0.74; Figure 11 A and B).

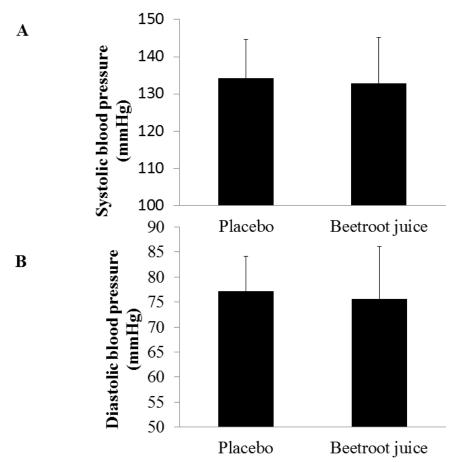


Figure 11. Systolic and diastolic blood pressure response to beetroot juice in individuals with T2DM.

Effects of  $NO_3^-$  on blood pressure, A: Groups means for systolic blood pressure and B depicts the group mean for diastolic blood pressure. Values are means  $\pm$  SD.

Antioxidants in beetroot juice: Antioxidant concentrations in placebo and active beetroot juice are displayed in Table 9. Betacyanins and polyphenol compounds in beetroot juice.

The content of total betacyanins and polyphenol compounds in two different beetroot juices. Table A, depicts all identified compounds and figure B shows the compound groups. Mean value ± standard deviation (in mg/100 ml of fresh juice).

Α

Identified compound	Beetroot juice	Placebo
Betanin-3-O-glucoside	452.71 ± 19.56	519.31 ± 11.43
Betanidine	$6.03 \pm 0.40$	3.61 ± 0.19
Gallic acid	1.80 ± 0.12	1.43 ± 0.14
Chlorogenic acid	$3.03 \pm 0.55$	2.10 ± 0.22
Caffeic acid	$0.90 \pm 0.20$	0.58 ± 0.11
Ferulic acid	$0.56 \pm 0.00$	$0.53 \pm 0.01$
Rutinoside-3-O-quercetin	$1.97 \pm 0.08$	1.81 ± 0.04
Glucoside-3-O-quercetin	$0.89 \pm 0.04$	$0.83 \pm 0.02$
Myrycetin	$0.44 \pm 0.01$	$0.47 \pm 0.02$
Luteolin	$0.23 \pm 0.01$	0.16 ± 0.01
Quercetin	0.25 ± 0.01	$0.19 \pm 0.00$
Kaempferol	$0.22 \pm 0.01$	$0.17 \pm 0.00$

В

Group of compounds	Beetroot juice	Placebo
Total betacyanins	458.73 ±1 9.73	522.92 ± 11.43
Total polyphenols	$10.29 \pm 0.72$	$8.30 \pm 0.34$
Total phenolic acids	$6.30 \pm 0.67$	$4.65 \pm 0.34$
Total flavonoids	$3.99 \pm 0.10$	$3.65 \pm 0.05$

### 3.1.7 : Discussion

This is the first study to investigate the effects of dietary NO<sub>3</sub><sup>-</sup> supplementation on the exercise responses of individuals with T2DM. The principal findings of this investigation were that short-term dietary NO<sub>3</sub><sup>-</sup> supplementation did not reduce the O<sub>2</sub> cost of walking, or increase the distance covered in the six minute walk test in this population. We also confirmed our previous finding that NO<sub>3</sub><sup>-</sup> supplementation had no effect on resting blood pressure in individuals with T2DM. These findings may be considered surprising given the compelling effects of dietary NO<sub>3</sub><sup>-</sup> supplementation reported in other populations. Possible explanations for the lack of effects in the present study relate to an elevated baseline plasma NO<sub>2</sub><sup>-</sup> concentration in the control condition, and/or reductions in the bioavailability of NO· in individuals with T2DM.

Nitrate supplementation and plasma NO<sub>3</sub> and plasma NO<sub>2</sub> concentration.

Plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations were significantly elevated following NO<sub>3</sub><sup>-</sup> supplementation, which is consistent with previous studies examining young (Bailey et al., 2009, Lundberg and Govoni, 2004) and older healthy participants (Kelly et al., 2013a), and individuals with peripheral arterial disease (Kenjale et al., 2011). The post placebo plasma NO<sub>2</sub><sup>-</sup> concentration was approximately two to six-times higher in the present study than those reported in the aforementioned studies. The elevated plasma NO<sub>2</sub><sup>-</sup> concentration in individuals with T2DM in the placebo condition may be associated with the habitual up-regulation of iNOS which is endemic in this population (Krause et al., 2012). It is possible that the elevated baseline (placebo) plasma NO<sub>2</sub><sup>-</sup> concentration is indicative of disease pathology and mitigates the attainment of the benefits of dietary NO<sub>3</sub><sup>-</sup> supplementation that have been reported in other populations.

The plasma NO<sub>2</sub> concentration was 680 ± 256 nM in the placebo arm in the present study. In the only other study which could be directly comparable, the plasma NO<sub>2</sub><sup>-</sup> concentration was 232 nmol/L (200, 265), median (IQR) in the placebo arm (Gilchrist et al., 2013). The control plasma from Gilchrist et al. (2013) was re-measured during the current study to establish agreement between the NO2 values measured in the two studies. It is therefore unlikely that analytical error is the reason for our elevated 'baseline' plasma NO<sub>2</sub> concentrations. There are notable differences between the two studies with respect to the timing of beetroot juice doses and plasma sampling. In the present work, studies were conducted at midday, with subjects having had breakfast including the beetroot juice and their usual morning hypoglycaemic and antihypertensive medications 3 hours previously. In our previous study in subjects with T2DM (Gilchrist et al., 2013), plasma sampling occurred after an overnight fast, with subjects having omitted their usual morning hypoglycaemic and antihypertensive medications, 16 hours after their last beetroot juice.

Multiple agents within these broad classes of medication have been shown to upregulate eNOS activity (Davis et al., 2006, Ceconi et al., 2007, Andrade et al., 2013). When the current cohort was split for drug classifications, no differences in baseline plasma NO<sub>2</sub><sup>-</sup> were detectable. It is therefore unlikely that drug classification affected plasma NO<sub>2</sub><sup>-</sup> concentrations, though the study was not powered to detect any such difference. Furthermore, there are data to suggest circadian variation in eNOS activity, with the lowest levels in the morning, rising through the day before falling again at night (Elherik et al., 2002). This could further influence the difference between plasma NO<sub>2</sub><sup>-</sup> concentrations in the two groups. With a growing number of studies in patient groups, understanding the effect of concomitant medication on plasma NO<sub>2</sub><sup>-</sup> and NO<sub>3</sub><sup>-</sup> concentrations is becoming increasingly important.

Accordingly, sufficiently powered research is required to examine the pharmacokinetics and dynamics of medications and NOx concentrations in order to elucidate the possibility of NO<sub>3</sub> related therapeutic effects on exercise performance.

# Nitrate supplementation and pulmonary O<sub>2</sub> uptake

There was no difference between active and placebo juice in the  $\dot{V}O_2$  responses to low intensity walking exercise in the present study. A reduction in the baseline (Lansley et al., 2010) and steady state (Bailey et al., 2009, Vanhatalo et al., 2010, Lansley et al., 2010) O<sub>2</sub> cost of low intensity exercise has previously been reported in young healthy individuals following similar NO<sub>3</sub> dosage regimens to that which was implemented in the present study. No change in the O2 cost of exercise was reported in healthy older individuals following NO<sub>3</sub> supplementation (Kelly et al., 2013a). However, these authors reported a significant speeding of the VO<sub>2</sub> kinetics, something that was not observed in the present study. It is therefore unlikely that aging per se explains the lack of effect on the  $\dot{V}O_2$  kinetics in the present study. Other studies have reported no significant difference in the VO<sub>2</sub> response to exercise subsequent to NO<sub>3</sub> supplementation in healthy, well-trained athletes (Wilkerson et al., 2012, Bescos et al., 2011). This lack of effect has also been reported in individuals with COPD (Shepherd et al., 2015, Berry et al., 2014, Kerley et al., 2015). These studies all had relatively small sample sizes (n = 8 (Wilkerson et al., 2012), 11 (Bescos et al., 2011, Kerley et al., 2015), 13 (Shepherd et al., 2015), and 15 (Berry et al., 2014)).

One possible explanation for the lack of effect on the  $O_2$  cost of exercise may relate to the elevated plasma  $NO_2^-$  concentration in the placebo condition (baseline) of this study which might reduce the scope for the further increases in plasma  $NO_2^-$ 

concentration that have been realised via NO<sub>3</sub> supplementation in other populations. Thus the lack of effect of dietary NO<sub>3</sub> supplementation on the O<sub>2</sub> cost of exercise noted in the present study may indicate the existence of an upper limit for the baseline plasma NO<sub>2</sub> concentration, beyond which the scope for positive effects from further increasing plasma NO<sub>2</sub> concentration is reduced. In support of this suggestion, a significant negative correlation has been reported between the changes in plasma NO<sub>2</sub> concentration with beetroot juice vs. placebo juice and the change in performance during cycling exercise (i.e. participants whose plasma NO<sub>2</sub> increased more following NO<sub>3</sub> supplementation experienced a greater improvement in exercise performance, and *vice versa*) (Wilkerson et al., 2012). However, in the current study there was no correlation between either placebo plasma NO<sub>2</sub> concentration, or change in plasma NO<sub>2</sub> concentration with NO<sub>3</sub> rich vs. placebo juice, and the O<sub>2</sub> cost of exercise.

An alternative suggestion for the lack of effect of NO<sub>3</sub> supplementation on the O<sub>2</sub> cost of exercise may be related to the reduction in the bioavailability of NO· which has been reported in individuals with T2DM (Calver et al., 1992, Cockcroft et al., 2000, Cohen, 1993). This reduced bioavailability has been linked with the concentration of plasma ADMA (an analogue of L-arginine), which is elevated in individuals with T2DM (Devangelio et al., 2007). ADMA is known to inhibit all three NOS isoforms, particularly eNOS (Siasos et al., 2007), leading to eNOS uncoupling and the generation of superoxide radicals (Tousoulis et al., 2007), ultimately resulting in increased oxidative stress. When NO· and superoxide react, peroxynitrite is generated. This is known to result in a quenching of NO· activity and thus a reduction in its bioavailability, which may serve to diminish any positive impact of NO<sub>3</sub> supplementation (Beckman and Koppenol, 1996, Coppey et al., 2001).

Furthermore, sustained exposure to oxidative and nitrative stress could result in damage to mitochondrial membranes (Kowaltowski and Vercesi, 1999) and thus reduce the P/O ratio (i.e more  $O_2$  would be required to produce a given amount of ATP). It is possible that damaged mitochondria associated with T2DM (Kowaltowski and Vercesi, 1999) means that this population are less likely to benefit from the improved mitochondrial function subsequent to  $NO_3^-$  supplementation, which may underpin, in part, the reduced  $O_2$  cost of exercise in healthy individuals (Cooper and Giulivi, 2007).

A further possible explanation for the lack of effect of NO<sub>3</sub> supplementation on the exercise response in individuals with T2DM may relate to pathological consumption of NO<sub>2</sub> during exercise. Post-exercise plasma NO<sub>2</sub> concentration has been shown to fall markedly in individuals with cardiovascular risk factors (Rassaf et al., 2010) compared with the typical rise in plasma NO<sub>2</sub> reported in healthy individuals (Cuzzolin et al., 2000). This may suggest that either the production of NO· cannot keep up with metabolic demand, or the NO· produced is scavenged more rapidly in individuals with cardiovascular risk factors compared with healthy individuals. It is plausible that a similar effect is seen in individuals with T2DM, with a marked net consumption of plasma NO<sub>2</sub> during exercise. It should be noted, however, that this would not explain the lack of effect of NO<sub>3</sub> supplementation on BP in our study as these measurements were conducted at rest.

It should be noted that although we have postulated a pro-oxidant state as an explanation for the lack of effect of  $NO_3^-$  supplementation in individuals with T2DM, beneficial effects of  $NO_3^-$  have been seen in other patient groups who are likely to be in a pro-oxidant state. Zamani et al. (2015) reported an improvement in exercise capacity (but no reduction in the  $O_2$  cost of exercise) following beetroot juice

supplementation in individuals with heart failure. Similar findings have been reported in individuals with peripheral artery disease (Kenjale et al., 2011) and chronic obstructive pulmonary disease (Shepherd et al. (2015), Berry et al., 2014, Kerley et al., 2015). It should be noted, however, that to date no study has shown a reduction in the O<sub>2</sub> cost of exercise in a patient group, despite reporting improvements in exercise tolerance. It is feasible that the improvement in exercise tolerance in the aforementioned studies is associated with elevated muscle blood flow consequent to an elevated plasma NO<sub>2</sub><sup>-</sup> concentration (Cosby et al., 2003), although this requires further investigation to confirm.

Nitrate supplementation and functional Capacity.

There was no difference between the active and placebo juice conditions for the distance covered in the 6MWT. This is consistent with the only other study to have investigated the impact of  $NO_3^-$  supplementation on exercise performance using the 6MWT (in healthy older individuals) (Kelly et al., 2013a). Since any improvement in walking performance could reasonably be assumed to be underpinned by alterations in the  $\dot{V}O_2$  response to exercise, and considering that this was not modulated by  $NO_3^-$  supplementation, it is perhaps not surprising that functional capacity was also not different between conditions. It is likely that the explanation for the lack of effect of  $NO_3^-$  supplementation on walking performance is synonymous with the potential explanations for the lack of effect on the  $O_2$  cost of exercise (see previous section).

Walking performance has been shown to be enhanced in individuals who are taking ACEi (Ahimastos et al., 2013). Our cohort included 48% of individuals on ACEi and/or ARBs, thus any potential improvements in walking performance subsequent to NO<sub>3</sub><sup>-</sup> supplementation may have been masked. When we separated the group into

those prescribed ACEi or ARBs compared to the remaining individuals, no difference was seen between groups for walking performance. The groups were split 48% (n = 23) with prescribed ACEi or ARBs and 52% (n = 25) not taking ACEi/ARB. One possible explanation for the finding is that the 6MWT was the last of the experimental procedures to be conducted and followed 18 minutes of treadmill walking. The prior exercise (and time since last  $NO_3^-$  supplementation) may have meant that some of the additional  $NO_2^-$  which was available following  $NO_3^-$  supplementation had been utilised, especially given our cohort's elevated cardiovascular risk (Rassaf et al., 2010).

Nitrate supplementation and resting blood pressure.

In agreement with our previous study examining the effect of dietary NO<sub>3</sub><sup>-</sup> supplementation on blood pressure in individuals with T2DM (Gilchrist et al., 2013), we found no difference in BP between conditions. NO<sub>3</sub><sup>-</sup> doses similar to that which was administered in the present study (6.43 mmol per day) have elicited reductions systolic BP in healthy young (Bailey et al., 2009, Bailey et al., 2010b, Lansley et al., 2010) and old (Kelly et al., 2013a) individuals and reduced diastolic BP in healthy young individuals (Bailey et al., 2009) and those with peripheral arterial disease (Kenjale et al., 2011).

There are a number of possible explanations for the lack of effect of NO<sub>3</sub> supplementation on the resting BP of individuals with T2DM. A reduced NO-responsiveness has been linked to vascular stiffening in older individuals (Lyons et al., 1997). However, aging *per se* is unlikely to explain the lack of effect in the present study as BP was significantly reduced subsequent to NO<sub>3</sub> supplementation in healthy older adults (on no medication) (Kelly et al., 2013a). Secondly, the

elevated oxidative stress prevalent in individuals with T2DM would be expected to result in an increase in the scavenging of NO·, potentially diminishing any hypotensive effects from NO<sub>3</sub> supplementation.

## Antioxidants in beetroot juice

It has been demonstrated that the beneficial effects of beetroot juice are largely explained by its  $NO_3^-$  content (Lansley et al., 2010). However, it has also been suggested that an antioxidant effect may also be occurring, such as is observed under in vitro experiments with polyphenols (Kanner et al., 2001). Though there are small differences in antioxidants between the two juices used (placebo and active), the magnitude of this difference is unlikely to be physiologically relevant. Typical total daily polyphenol intake across multiple populations has been estimated to be in the region of 1g daily (Bohn, 2014); the effective dose from either juice in the current study therefore represents less than 1% of average total intake. Betacyanins were present in much higher quantities. The bioavailability of these compounds is however uncertain, but it appears to be very low with typical estimates from <1% to 4% of an oral dose, with some individuals having far greater absorption (Watts et al., 1993, Tesoriere et al., 2004). Furthermore, the removal of betanin from plasma is rapid, with a  $t_{1/2}$  of 0.94±0.07 hours. These factors suggest the betacyanin content of beetroot juice is unlikely to have a clinically meaningful effect.

## Strengths and limitations.

To date this is the largest trial, implementing robust methods, to examine the effect of inorganic  $NO_3^-$  or pharmacological  $NO_3^-$  supplementation on the  $O_2$  cost of exercise. The *a priori* sample size calculation was designed to enable the study to detect an 8% reduction in the  $O_2$  cost of low intensity exercise with a 0.05 alpha level

and an 80% power. From the 95% CI it is likely that the actual difference for the O<sub>2</sub> cost of low intensity exercise following NO<sub>3</sub><sup>-</sup> supplementation in individuals with T2DM was between -2.1% and 3.6%. Therefore the minimum detectable reduction in the O<sub>2</sub> cost of exercise in our study is 2.1%. This is very similar to the day to day variability of  $\dot{V}O_2$  measurement, thus it is unlikely that a larger sample size would elucidate a clinically significant difference in the O<sub>2</sub> cost of walking exercise in this patient group. A potential limitation of the present study is that the 6MWT was completed up to 4 hours subsequent to the consumption of the final beverage, and after 18 minutes of walking when the increments in NO<sub>2</sub><sup>-</sup> from supplementation may have been utilised already. As plasma NO<sub>2</sub><sup>-</sup> concentration was not determined prior to the 6MWT, we cannot be certain that this remained elevated following NO<sub>3</sub><sup>-</sup> supplementation compared to the placebo condition. However, this does mean that the study provides knowledge of the level of improvement (or lack of it) that a patient with type 2 diabetes could expect in the early afternoon of normal daily living.

## 3.1.8: Conclusion

In contrast to much of the literature in young healthy individuals and despite a statistically significant and physiologically meaningful increase in plasma NO<sub>2</sub><sup>-</sup> concentration, four days of beetroot juice supplementation with 6.43 mmol of NO<sub>3</sub><sup>-</sup> did not reduce the O<sub>2</sub> cost of walking, improve functional capacity as determined by the 6MWT, or reduce resting BP in individuals with T2DM. The lack of effects of dietary NO<sub>3</sub><sup>-</sup> supplementation in individuals with T2DM may be explained by increased oxidative stress and its impact on the bioavailability of NO·, or an elevated 'baseline' plasma NO<sub>2</sub><sup>-</sup> concentration which reduces the scope for the beneficial effects reported in other populations.

# 3.2: Experimental chapter 2 (Dietary nitrate supplementation in COPD)

Title: The effect of dietary nitrate supplementation on the oxygen cost of cycling, walking performance and resting blood pressure in individuals with chronic obstructive pulmonary disease: A double blind placebo controlled, randomised control trial.

3.2.1: Authors: Anthony I Shepherd<sup>1,2</sup>, Daryl P Wilkerson<sup>1</sup>, Lee Dobson<sup>3</sup> James Kelly<sup>1</sup>, Paul G Winyard<sup>2</sup>, Andrew M Jones<sup>1</sup>, Nigel Benjamin<sup>2,3</sup>, Angela C Shore<sup>2</sup>, Mark Gilchrist<sup>2</sup>

3.2.2: Affiliation: <sup>1</sup>College of Life and Environmental Sciences, Sport and Health Sciences, University of Exeter, Devon, UK, <sup>2</sup> University of Exeter Medical School and NIHR Exeter Clinical Research Facility, Royal Devon and Exeter Hospital, Exeter, Devon, UK, <sup>3</sup> Torbay Hospital, Heart and Lung Unit, Torquay, Devon, UK.

### 3.2.3: Abstract

### Background

Chronic obstructive pulmonary disease (COPD) results in exercise intolerance. Dietary NO<sub>3</sub> supplementation has been shown to lower blood pressure (BP), reduce the O<sub>2</sub> cost of exercise, and enhance exercise tolerance in healthy volunteers. This study assessed the effects of dietary NO<sub>3</sub> on the O<sub>2</sub> cost of cycling, walking performance and BP in individuals with mild-moderate COPD.

### Methods

Thirteen patients with mild-moderate COPD were recruited. Participants consumed 70 ml of either NO<sub>3</sub> rich (6.77 mmol NO<sub>3</sub>; beetroot juice) or NO<sub>3</sub> depleted beetroot juice (0.002 mmol NO<sub>3</sub>; placebo) twice a day for 2.5 days, with the final supplement ~3 hours before testing. BP was measured before completing two bouts of moderate-intensity cycling, where pulmonary gas exchange was measured throughout. The six-minute walk test (6MWT) was completed 30 minutes subsequent to the second cycling bout.

### Results

Plasma NO<sub>3</sub><sup>-</sup> concentration was significantly elevated following beetroot juice vs. placebo (placebo;  $48 \pm 86$  vs. beetroot juice;  $215 \pm 84$  µM, P=0.002). No significant differences were observed between placebo vs. beetroot juice for O<sub>2</sub> cost of exercise (933 ± 323 vs. 939 ± 302 ml: min<sup>-1</sup>; P=0.88), distance covered in the 6MWT (456 ± 86 vs. 449 ± 79 m; P=0.37), systolic BP (123 ± 14 vs. 123 ± 14 mmHg; P=0.91), or diastolic BP (77 ± 9 vs. 79 ± 9 mmHg; P=0.27).

### Conclusion

Despite a large rise in plasma NO<sub>3</sub><sup>-</sup> concentration, two days of NO<sub>3</sub><sup>-</sup> supplementation did not reduce the O<sub>2</sub> cost of moderate intensity cycling, increase distance covered in the 6MWT, or lower BP.

## 3.2.4: Introduction

Exercise in individuals with COPD is limited by multiple factors which can result in hypoxemia. These include loss of normal lung architecture, impaired cardiac function (Jardin et al., 1984), abnormal pulmonary blood flow distribution (Marshall et al., 1994) and peripheral muscle de-conditioning (Maltais et al., 2014). O<sub>2</sub> uptake in the lungs and delivery of O<sub>2</sub> to working muscle is impaired by increases in pulmonary blood flow which increase shunting through blood vessels resulting in incomplete gas exchange (Cooper, 2009) and cor pulmonale later in the disease course. These abnormalities result in feelings of breathlessness and fatigue (Nici et al., 2006), with individuals often finding that activities of daily living are physically challenging.

The beneficial effects of a diet rich in vegetables upon cardiovascular health (Gilchrist et al., 2010), risk of morbidity and mortality (Joshipura et al., 2001), and

COPD development (Hirayama et al., 2009, Watson et al., 2002) have been well described. These positive effects have, in part, been attributed to inorganic NO<sub>3</sub><sup>-</sup> (NO<sub>3</sub><sup>-</sup>) which is found in particularly high quantities in leafy green vegetables and some root vegetables such as beetroot (Bryan and Hord, 2010). Nitrate supplementation in the form of sodium NO<sub>3</sub><sup>-</sup> or NO<sub>3</sub><sup>-</sup> rich beetroot juice has been shown to have remarkable effects in healthy young individuals and athletes, including reductions in the O<sub>2</sub> cost of exercise (Bailey et al., 2009), enhanced exercise tolerance/performance and reduced blood pressure (BP) (Bailey et al., 2009, Bailey et al., 2010b). Some of these effects have subsequently been observed in individuals with peripheral artery disease following dietary NO<sub>3</sub><sup>-</sup> supplementation (Kenjale et al., 2011). These findings have been attributed to an increase in the bioavailability of nitric oxide (NO·).

NO· is a signalling molecule with multiple functions including regulation of vascular tone, mitochondrial respiration and skeletal muscle function (Cosby et al., 2003, Merry et al., 2010, Larsen et al., 2011). These factors are important in the physiological response to exercise. NO· is produced in two distinct ways in man. The best known is the classical L-arginine nitric oxide synthase (NOS) pathway which is O₂ dependent (Alderton et al., 2001). The second is the entero-salivary pathway and is O₂ independent. Briefly, NO₃ from the diet is rapidly and extensively absorbed in the stomach and proximal small intestine with bioavailability approaching 100% (Florin et al., 1990). Nitrate is then concentrated in the salivary glands, with concentrations 10 fold greater in saliva than in plasma. NO₃ secreted in saliva is reduced to NO₂ by facultative anaerobic bacteria on the dorsum of the tongue (Duncan et al., 1995). On swallowing, the acidic environment of the stomach results in NO· formation with important local effects on gastric function and host defence

(Gilchrist et al., 2010, Benjamin et al., 1994). Some NO<sub>2</sub> is absorbed into the circulation where it acts as a storage pool for subsequent NO· production (Cosby et al., 2003). The conversion of NO<sub>2</sub> to NO· is expedited in conditions of acidosis (Modin et al., 2001) or hypoxemia (Cosby et al., 2003) which often occur in the exercising muscle of individuals with COPD (Fiaccadori et al., 1987).

In many individuals with COPD, functional capacity is reduced to a level where activities of daily living may impose a challenge due to an energy requirement representing a high fraction of their maximal O<sub>2</sub> uptake. While a number of cardiovascular and physiological benefits have been shown as a result of dietary NO<sub>3</sub><sup>-</sup> supplementation in healthy populations, little is known about possible effects in clinical populations. We aimed to determine whether dietary NO<sub>3</sub><sup>-</sup> supplementation has a beneficial impact upon the O<sub>2</sub> cost of sub-maximal cycling exercise, walking performance and BP in individuals with COPD.

## **Purpose**

The aim of this study was to assess the effects of 2.5 days of dietary  $NO_3$  supplementation on the  $O_2$  cost of sub-maximal cycling, walking performance, and resting BP in individuals with mild-moderate COPD.

## 3.2.5: Methods

### **Patients**

Fourteen individuals with mild-moderate COPD (see table 10 for patient characteristics) gave written informed consent to participate in this double-blind, placebo-controlled, cross-over design study between April 2013 and January 2014. The study was registered as a clinical trial at ClinicalTrials.gov (NCT01712386). The

Exeter NRES Committee gave ethical approval (12//SW//0327). Patients were recruited if lung function was between 30-80% of predicted FEV<sub>1</sub> values, aged 40-75 years old and able to give informed consent. Participants were excluded if they had chronic kidney disease (estimated glomerular filtration rate <30 ml/min/1.73 m<sup>2</sup>), uncontrolled hypertension (systolic BP> 160 mmHg or diastolic >100 mmHg), were smokers (smoked within past 3 months), consumed regular organic NO<sub>3</sub><sup>-</sup> or nicorandil. Patients taking phosphodiesterase inhibitors were asked to refrain from doing so for the duration of the study.

Table 9. Characteristics of patients included in the final analyses.

Data are mean ± SD or as % of the cohort on a medication.

Age (years)	64.7 ± 7.7
FEV <sub>1</sub> (%)	57 ± 9
FVC (I)	$3.6 \pm 0.9$
FEV <sub>1</sub> /FVC	41 ± 16
Body Mass Index (kg/m²)	29 ± 8
Baseline SBP (mmHg)	132 ± 15
Baseline DBP (mmHg)	85 ± 10
No. of 30+ minutes of exercise per week	$4.7 \pm 3.6$
ARB /ACEi (%)	23
Calcium channel blocker (%)	15
Short-acting β2-agonists (%)	77
Long-acting β2-agonists (%)	77

# **Pre-experimental tests**

Participants arrived at the Heart and Lung unit at Torbay hospital where informed consent, medical history, anthropometric measures, BP, lung function and an ECG were performed. Participants completed a ramp incremental cycle ergometer test (10 W·min<sup>-1</sup>) to determine their gas exchange threshold (GET). Breath-by-breath pulmonary gas exchange was measured throughout and the GET was determined using the V-slope method as described previously (Beaver et al., 1986).

# **Experimental Overview**

Participants consumed 70 ml of NO<sub>3</sub> rich beetroot juice (beetroot juice; 6.77 mmol NO<sub>3</sub>; Beet it, James White Drinks Ltd., Ashbocking, UK) or NO<sub>3</sub> depleted beetroot juice as a placebo (placebo; 0.002 mmol NO<sub>3</sub>; Beet it, James White Drinks Ltd., Ashbocking, UK), with one beverage in the morning and one in the evening for two days preceding testing. On study days, participants consumed a final 70 ml beetroot juice drink ~3 hours prior to exercising. Participants self-reported concordance with the supplementation regime which was confirmed by measurement of plasma NO<sub>3</sub> concentration. After exercise testing the participants began a washout period (7 days) before entering the opposing arm of the study. The placebo was indistinguishable from the NO<sub>3</sub> rich juice in taste, colour, texture, appearance and odour as described previously (Gilchrist et al., 2014).

Participants arrived at the laboratory in a fully hydrated state, having avoided consumption of caffeine, alcohol, cruciferous vegetables, leafy greens, beetroot, and completion of strenuous exercise 24 hours prior to testing. Participants were asked to record their food intake for 24 hours prior to testing and to replicate this after the crossover and this was verbally confirmed on the second exercise visit. Participants

avoided antibacterial mouthwash for 7 days prior to testing. Participants arrived 45 minutes before the initiation of exercise following ingestion of the randomised juice with their morning meal. Brachial artery BP was taken, after a 10 minute resting period whilst supine, with an automated sphygmomanometer (Omron M6, Kyoto, Japan). Five measurements were performed and the mean of the last three was recorded. Venous blood was drawn and processed for plasma NO<sub>3</sub> concentration as per our previously described chemiluminescence technique (Gilchrist et al., 2013). Participants completed two bouts of cycling at 80% of their GET on a cycle ergometer (Ergoselect 100, Bitz, Germany) with 30 minutes recovery between bouts. Following 30 minutes rest, participants performed a six-minute walk test to assess functional capacity. Participants walked around a clear rectangular corridor (14 x 12m) for a total of 52m per lap, covering as much distance as possible. Standardised verbal encouragement was given throughout.

#### Measurements

Pulmonary gas exchange and ventilation were measured during the cycling exercise (Vmax<sup>TM</sup> Encore, Yorba, Linda, CA). Before each session the analysers were calibrated using gases of known concentration. The volume transducer was calibrated using a 3-litre syringe (Hans Rudolph, Kansas City, MO, USA).

#### **Outcome measures**

Primary outcome measure: does 2.5 days of NO<sub>3</sub><sup>-</sup> supplementation reduce the O<sub>2</sub> cost of moderate intensity cycling? Secondary outcome measure (i): does 2.5 days of NO<sub>3</sub><sup>-</sup> supplementation improve functional capacity as measured via the six-minute walk test? Secondary outcome measure (ii): does 2.5 days of NO<sub>3</sub><sup>-</sup> supplementation reduce resting BP?

# Sample size and randomisation

An *a priori* sample size calculation was performed. Previous literature in healthy young volunteers has shown a mean change between beetroot juice and control of 69 ml for end exercise pulmonary  $O_2$  uptake  $(\dot{V}O_2)$  (1SD) and 121 ml for  $\dot{V}O_2$  amplitude (2SD) (Bailey et al., 2009). For 90% power and an  $\alpha$ -level set at P=0.05 (two tailed), to detect a 1 SD difference 13 patients were required. The reproducibility of these measures in patients with COPD are similar to healthy controls (Covey et al., 1999). An unrestricted computer generated sequence was used by a research nurse to assign each participant a randomisation number and supply them with the requisite juice.

# Data and statistical analysis

Participant's breath by breath  $\dot{V}O_2$  data were initially checked for erroneous breaths (caused by coughing and swallowing). Breaths > 4 SDs away from the local mean were removed prior to interpolation. Breath by breath data for each cycling bout were time aligned and interpolated to provide second by second values. A nonlinear least squares algorithm was then used to fit the ensemble-averaged data. The overall  $\dot{V}O_2$  kinetics were described using the mean response time (MRT), which was calculated by fitting a single exponential curve to the data with no time delay from the onset to the end of exercise. The  $O_2$  deficit was calculated as the product of the  $\dot{V}O_2$  response amplitude (i.e. the baseline to the point that a steady state was attained) and the MRT. For a schematic representation of the kinetics parameters, please see figure 12.

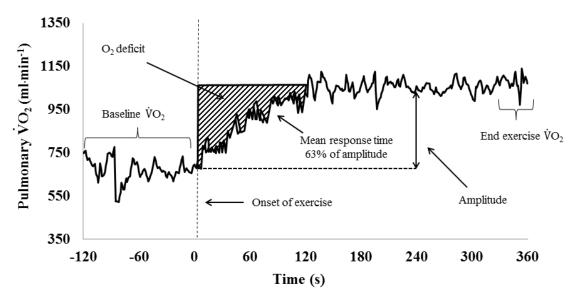


Figure 12. Parameters of O<sub>2</sub> uptake kinetics.

All data were tested for normality. Statistical differences were assessed using paired t-tests for normally distributed data and Wilcoxon rank-sum test for non-normally distributed data. All data are presented as means  $\pm$  standard deviation (*SD*). Statistical analysis was performed on SPSS software version 21.0 (Chicago, IL, USA). Statistical difference was accepted when P < 0.05.

## 3.2.6: Results

14 individuals with COPD provided written informed consent. Following screening, one individual was withdrawn due to FEV<sub>1</sub> < 30%. 13 participants were randomised to start in either the beetroot juice or placebo condition of the study. All participants reported 100% adherence to the supplementation regime. Participants reported similar dietary patterns and physical activity during both study arms. Dietary NO<sub>3</sub> supplementation was well tolerated with no adverse events apart from red stools and beeturia, as in previous studies (Bailey et al., 2009).

*Plasma*  $NO_3^-$  *concentration*: Relative to placebo, beetroot juice significantly increased plasma  $NO_3^-$  concentration (placebo; 48 ± 85 vs. beetroot juice; 215 ± 84 $\mu$ M, P = 0.002, 95% CI 75, 260; Figure 13).



Figure 13. Plasma NO<sub>3</sub> concentration in individuals with COPD.

Plasma  $NO_3^-$  concentration following placebo and beetroot juice supplementation. The open bar represents placebo and the closed bar, beetroot juice. \*Significantly different from placebo P < 0.01.

Effects on the  $O_2$  cost of cycling exercise: The group mean pulmonary  $\dot{V}O_2$  response to exercise for both placebo and beetroot juice conditions can be seen in figure 14, with the  $\dot{V}O_2$  kinetics resulting from the model fits displayed in Table 11. Relative to placebo, beetroot juice supplementation had no effect on baseline  $\dot{V}O_2$  (placebo; 634  $\pm$  233 vs. beetroot juice; 622  $\pm$  253 ml·min<sup>-1</sup>, P = 0.56, 95% CI -57, 32) or end exercise  $\dot{V}O_2$  (placebo; 933  $\pm$  323 vs. beetroot juice; 939  $\pm$  302 ml·min<sup>-1</sup>, P = 0.88,

95% CI -68, 78). There were no differences between conditions for the MRT (P = 0.90, CI -25, 28) or the O<sub>2</sub> deficit (P = 0.71, CI -.2, .3).

Table 10. Pulmonary gas exchange during moderate intensity cycling following placebo and beetroot juice supplementation.

	Dlasaha	Dootroot inico
	Placebo	Beetroot juice
	004 000	000 050
Baseline VO <sub>2</sub> (ml/min)	634 ± 233	622 ± 253
VO₂ Amplitude (ml/min)	$292 \pm 80$	$308 \pm 85$
End-exercise $\dot{V}O_2$ (ml/min)	933 ± 323	939 ± 302
O <sub>2</sub> deficit (L)	$0.39 \pm 0.19$	$0.43 \pm 0.29$
MRT (s)	80 ± 19	81 ± 35

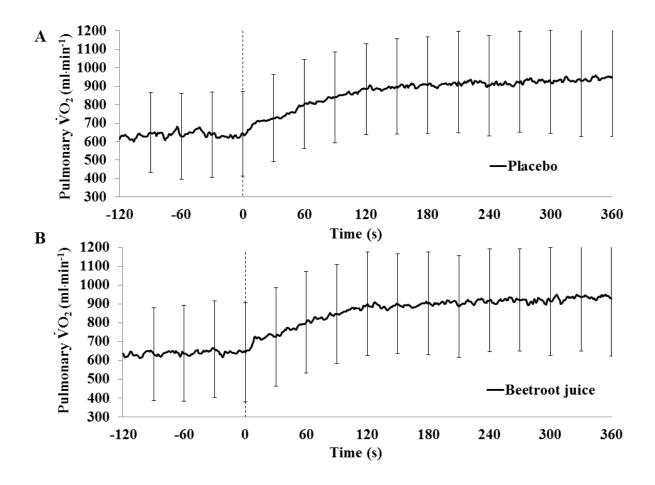


Figure 14. Pulmonary O<sub>2</sub> uptake response to beetroot juice in individuals with COPD.

The pulmonary  $O_2$  uptake response during the transition from unloaded cycling to cycling at 80% of the GET for 6 minutes following placebo (A) and beetroot juice supplementation (B). The vertical line denotes the transition from baseline to moderate intensity cycling.

Effects on functional capacity: There was no difference between conditions for distance covered during the six-minute walk test (placebo;  $456 \pm 86 \text{ vs.}$  beetroot juice;  $449 \pm 79 \text{ m}$ , P = 0.17, 95% CI -22, 9).

Effects on resting blood pressure: Compared to the placebo juice, beetroot juice did not significantly reduce systolic BP (placebo;  $123 \pm 14 \ vs.$  beetroot juice;  $123 \pm 14 \ mmHg$ , P = 0.91, 95% CI -5, 4) or diastolic BP (placebo;  $78 \pm 9 \ vs.$  beetroot juice;  $79 \pm 9 \ mmHg$ , P = 0.25, 95% CI -2, 5; Figure 15).

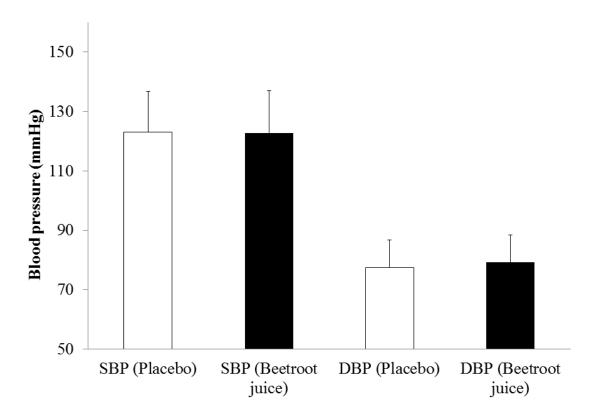


Figure 15. Systolic (SBP) and diastolic (DBP) blood pressure following placebo and beetroot juice supplementation.

The open bar represents placebo and the closed box, beetroot juice.

### 3.2.7: Discussion

Beetroot juice supplementation, ( $NO_3^-$ ; 6.77 mmol) twice daily, for 2.5 days did not reduce the  $O_2$  cost of cycle ergometer exercise, improve functional capacity or reduce resting BP in individuals with COPD. There was no difference between conditions for these variables despite a statistically significant and physiologically meaningful rise in plasma  $NO_3^-$  concentration following  $NO_3^-$  supplementation.

Possible explanations for the lack of effect in this study include NO<sub>3</sub><sup>-</sup> dosage, efficacy of NO<sub>3</sub><sup>-</sup> reduction to NO<sub>2</sub><sup>-</sup>, oxidative stress, and the age of the participants.

Nitrate supplementation and effects on plasma NO<sub>3</sub> concentration.

Plasma NO<sub>3</sub> concentration was 48µM post placebo and 215µM following NO<sub>3</sub> rich beetroot juice, which is consistent with much of the literature in healthy young individuals (Larsen et al., 2010, Bescos et al., 2011) and individuals with type 2 diabetes (Gilchrist et al., 2014). Similar changes in plasma NO<sub>3</sub> concentrations have been shown to elicit reductions in the O2 cost of exercise, improved exercise tolerance/performance and reductions in BP (Larsen et al., 2007a, Muggeridge et al., 2013b, Wylie et al., 2013a). Due to logistical constraints, plasma NO<sub>2</sub> concentration was not assessed in this study. In all previous studies involving dietary NO<sub>3</sub><sup>-</sup> supplementation where plasma NO2 concentration has been determined, a rise in plasma NO<sub>3</sub> concentration similar to the magnitude observed in the present study has been accompanied by a physiologically meaningful and statistically significant rise in plasma NO<sub>2</sub> concentration (Muggeridge et al., 2013b, Larsen et al., 2007a, Larsen et al., 2011). However, we cannot exclude the possibility that there is an impaired capacity for reduction of NO<sub>3</sub> to NO<sub>2</sub> in individuals with COPD. Such an impairment could potentially be related to differences between individuals with COPD and healthy individuals in oral microflora due to oral steroids and repeated exposure to courses of antibiotics (Mobbs et al., 1999).

Nitrate supplementation and effects on the  $O_2$  cost of cycling exercise.

We found no reduction in the  $O_2$  cost of cycling exercise at baseline or end-exercise in individuals with COPD following  $NO_3^-$  rich beetroot juice supplementation compared to placebo. Nitrate supplementation in healthy young individuals has

previously resulted in reductions in the  $O_2$  cost of exercise (Bailey et al., 2009). However, we recently reported that the  $O_2$  cost of exercise was not altered by dietary  $NO_3^-$  supplementation in a group of healthy older adults (Kelly et al., 2013a). The current study is the first to examine the effects of  $NO_3^-$  supplementation on the  $O_2$  cost of exercise in any clinical population. The supplementation regime used in this study, consisting of 6.77 mmol twice a day for 2.5 days, has previously been shown to increase plasma  $NO_2^-$  concentrations (Bailey et al., 2009, Vanhatalo et al., 2010, Lansley et al., 2010) and elicit reductions in the  $O_2$  cost of exercise (Muggeridge et al., 2013b). It is therefore unlikely that the dosage and the timing of  $NO_3^-$  supplementation explain why no effect on the  $O_2$  cost of exercise was observed.

One possible explanation for the reduction in O<sub>2</sub> cost following dietary NO<sub>3</sub> supplementation in other populations is an increase in the P/O ratio (i.e. less O<sub>2</sub> being consumed to produce a given amount of ATP). Larsen et al (Larsen et al., 2011) reported an increase in the P/O ratio of harvested mitochondria following three days NO<sub>3</sub> supplementation. However, we did not observe a reduction in the O<sub>2</sub> cost of exercise, which may be related to the impact of oxidative stress, which is reported to damage mitochondrial membranes (Kowaltowski and Vercesi, 1999), potentially resulting in a reduction in the P/O ratio. COPD is associated with increased oxidative stress, with reactive oxygen species (ROS) being produced within the inflammatory cells and epithelial cells of the airways in conjunction with increased systemic generation of ROS (Ciencewicki et al., 2008). Oxidative stress leads to uncoupling of the NO· synthase enzymes (Li and Forstermann, 2013), thus reducing NO-bioavailability and creating a negative feedback loop of diminishing NO· production and elevated NO· scavenging. This may be a substantial barrier to NO· based therapeutics in COPD.

Nitrate supplementation and effects on functional capacity.

No statistical difference in distance covered for the six-minute walk test was observed between conditions. Considering that the  $O_2$  cost of exercise and rate of adaptation of  $\dot{V}O_2$  were not altered following  $NO_3^-$  supplementation, it is perhaps not surprising that functional capacity was also not different between conditions. It is likely that these lack of effects share a common explanation, which may be related to the impact of oxidative stress on the bioavailability of  $NO_2$  (Li and Forstermann, 2013) (see previous section).

The only other studies that have examined the impact of dietary NO<sub>3</sub><sup>-</sup> supplementation on walking performance have reported both positive and neutral effects. Kenjale et al. (2011) reported an increased walking time to exhaustion (17%) in a cohort of peripheral artery disease patients. However, dietary NO<sub>3</sub><sup>-</sup> supplementation had no effect on the distance covered in a six-minute walk test in healthy older individuals (Kelly et al., 2013a). Since plasma NO<sub>3</sub> (and NO<sub>2</sub> in Kenjale et al. 2011 and Kelly et al. 2013) concentrations were similar for the present study and two previous studies, the differences in walking performance post NO<sub>3</sub><sup>-</sup> supplementation are likely related to methodological differences. Kenjale et al. (2011) assessed walking performance via an incremental test to exhaustion on a treadmill, whereas in the present study and that of Kelly et al. (2013a) walking performance was assessed via completion of a (submaximal) six-minute walk test. It is likely that the higher exercise intensity encountered by the participants in Kenjale et al. (2011) resulted in the development of a hypoxic and acidic cellular environment that is known to be conducive for the reduction of NO<sub>3</sub> to NO<sub>2</sub> (Modin et al., 2001). Such an environment would have been less likely to occur during the lower exercise intensity in the present and Kelly et al's (2013a) study. Finally, Kenjale et al. (2011) did not use a placebo that was indistinguishable from their active juice, thus a 'placebo effect' cannot be ruled out.

Nitrate supplementation and effect on resting BP.

There was no difference in systolic or diastolic resting BP following NO<sub>3</sub> rich beetroot juice compared to placebo. This may be related to a factor specific to COPD such as the elevated oxidative stress (Ciencewicki et al., 2008) in this population would be expected to increase the scavenging of NO, thus reducing its effectiveness. Alternatively there are multiple other factors which may modify the BP effect. Studies investigating the effects of dietary NO<sub>3</sub> supplementation in older subjects with and without pathology have reported inconsistent BP effects. Gilchrist et al (2013) examined the impact of dietary NO<sub>3</sub> supplementation in individuals with type 2 diabetes, and found no statistical difference in mean 24h ambulatory BP. In subjects with peripheral artery disease Kenjale et al. (2011) reported a statistically significant reduction in diastolic BP (7 mmHg) but no change in systolic BP. It is possible that ageing per se may attenuate NO· mediated BP reduction, however in Kelly et al. (2013a) study of healthy older adults dietary NO<sub>3</sub> supplementation resulted in reductions in systolic and diastolic BP of 5 and 3 mmHg, respectively. In contrast, more recently, a larger study by Bondonno et al. (2014a) used a vegetable based, NO<sub>3</sub> rich diet for 7 days. Ten hour ambulatory BP along with home and office based measurements were used to assess BP. They found no reductions in BP or arterial stiffness. There are key differences around the supplementation protocol and timing and method of blood pressure measurement. In Kelly et al's study the office based blood pressure measurement was timed to coincide with the plasma NO<sub>2</sub> peak post NO<sub>3</sub> ingestion. In the Bondonno et al study measurements took place outside this window.

It is also worth noting the differing BMI's in these studies and our present manuscript. Kelly et al's cohort of older adults are the only group in the normal range (24±3 kg/m²). Our present cohort had a mean BMI of 29 ± 8kg/m², Bondonno et al's cohort were overweight 27±4 kg/m², and in our previous study of subjects with type 2 diabetes the group mean BMI was 30.8±3.2 kg/m². This raises the possibility that adiposity may attenuate the response to inorganic NO<sub>3</sub> by an as yet unknown mechanism.

One factor which may have had an impact is that subjects in the present study were taking multiple classes of drugs including antihypertensives. It is possible that the scope for reductions in BP subsequent to NO<sub>3</sub><sup>-</sup> supplementation is significantly reduced when individuals are already taking antihypertensive medication. It is noteworthy that in studies where subjects were taking antihypertensives (current study - 38% prescribed antihypertensives; Gilchrist et al. (2013) - 98% prescribed antihypertensives), no reductions in BP have been reported (see table 1 for drug classifications). Alternatively, the healthy older adults, on no medications, studied by Kelly et al. (2013a) showed a significant reduction in BP following NO<sub>3</sub><sup>-</sup> supplementation. It is possible that antihypertensive agents mitigate the NO-mediated reduction in BP.

There is conflicting evidence to suggest that ACEi/ARBs can alter the bioavailability of NOx with some studies showing reduction (Cacciatore et al., 2011) and others proposing increases (Ceconi et al., 2007, Jacoby and Rader, 2003). Therefore the direction in which ACEi/ARBs may alter the bioavailability of NO· remains unclear. \$2-adrenergic receptor agonists are known to increase endothelial NO· production and are at least, in part, responsible for their vasodilatory effects (Yong-Xiang et al., 1993). \$2-agonists are the most common treatment for individuals with COPD and

thus we could not reasonably exclude individuals who were prescribed this medication. We cannot exclude the possibility that prescribed medications which modulate NO· bioavailability may attenuate a beneficial effect from dietary NO<sub>3</sub><sup>-</sup> supplementation. This study is a crossover design and therefore both treatment arms will be equally affected. Further study is required to better understand the possible interaction of different medications and inorganic NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup>.

Berry et al. (2014) recently examined the effect of NO<sub>3</sub> rich beetroot juice vs. prune juice (as a placebo) in individuals with COPD. Plasma NO<sub>2</sub> concentration was significantly higher post beetroot juice compared to post prune juice, which indicates that the entero-salivary pathway is operational in people with COPD. The authors reported reductions in resting systolic BP, iso-time (defined as: last 60s of the shortest exercise time during either active or placebo visits compared with the same time point from the longer exercise time) BP, end exercise diastolic BP and an improvement in exercise tolerance (i.e. lengthened time to exhaustion during submaximal constant rate). Whilst the increase in exercise time is of interest, there are significant limitations in this study. Firstly, the design utilises prune juice as the placebo, which is likely to have a substantially different antioxidant content which could alter NO bioavailability (Bondonno et al., 2012). Secondly, and related to the lack of a 'true' placebo where the participant did not know whether active or placebo juice was being taken (as used in the present study), the widely reported (in the national press as well as in scientific literature) beneficial effects of beetroot juice on exercise performance/tolerance may have given rise to a placebo effect in informed volunteers. The authors do not show a reduction in the O<sub>2</sub> cost of exercise which is consistent with the present study. However, with no reduction in the O2 cost of exercise, it is not immediately clear what mechanism underpins the improved time to exhaustion reported by Berry et al. (2014).

This is the first double blind, randomised, placebo, controlled, crossover design study to examine the effects of NO<sub>3</sub><sup>-</sup> supplementation on the O<sub>2</sub> cost of exercise, walking performance and BP in individuals with COPD. The study had a robust experimental design (double-blind, placebo-controlled, randomised, cross-over study). A limitation is that we were not able to ascertain whether or not the increase in plasma NO<sub>3</sub><sup>-</sup> concentration lead to an increase in plasma NO<sub>2</sub><sup>-</sup> concentration, as we were not able to measure the latter due to logistical constraints. However, a recent study examining NO<sub>2</sub><sup>-</sup> levels in individuals with COPD did show elevated plasma NO<sub>2</sub><sup>-</sup> concentrations (Berry et al., 2014) which suggests the entero-salivary pathway is operational.

# 3.2.8: Conclusion

In contrast to findings in healthy young individuals, and despite a statistically significant and physiologically meaningful rise in plasma  $NO_3^-$  concentration, 2.5 days of beetroot supplementation with 6.77 mmol of  $NO_3^-$  twice daily did not reduce the  $O_2$  cost of cycling exercise, improve functional capacity or reduce resting blood pressure. Potential explanations for the lack of effect include a reduced P/O ratio due to systemic ROS generation associated with oxidative stress, or a reduced conversion of  $NO_3^-$  to  $NO_2^-$  in this population.

3.3: Experimental chapter 3 (Nitrate supplementation, hepatic diffusion and glucose homeostasis)

Title: Nitrate supplementation, hepatic diffusion and glucose homeostasis: A double-blind, placebo controlled, randomised control trial.

3.3.1: Authors: Anthony I Shepherd<sup>1,2</sup>, Daryl P Wilkerson<sup>1</sup>, Jon Fulford<sup>2</sup>, Paul G Winyard<sup>2</sup>, Nigel Benjamin<sup>2,3</sup>, Angela C Shore<sup>2</sup>, Mark Gilchrist<sup>2</sup>

3.3.2: Affiliation: <sup>1</sup>College of Life and Environmental Sciences, Sport and Health Sciences, University of Exeter, Devon, UK, <sup>2</sup> University of Exeter Medical School and NIHR Exeter Clinical Research Facility, Royal Devon and Exeter Hospital, Exeter, Devon, UK, <sup>3</sup> Torbay Hospital, Heart and Lung Unit, Torquay, Devon, UK.

# 3.3.3: Abstract

# Background:

Type 2 diabetes mellitus is a growing burden on the NHS budget predominantly due to an increased cardiovascular and diabetes risk factors. Nitrates derivatives (nitrite and nitric oxide) have been shown to alter gastric blood flow, improve vascular function and mediate glucose uptake within the intestines and skeletal muscle. Dietary NO<sub>3</sub><sup>-</sup> appears to be a potential low cost therapy that may help maintain glucose homeostasis.

### Methods:

In a randomised, double blind, placebo-controlled cross-over study, 31 participants (16 young adults and 15 healthy older adults) arrived fasted overnight and had a standardised breakfast, supplemented with either  $NO_3^-$  rich beetroot juice (11.91 mmol  $NO_3^-$ ) or  $NO_3^-$  depleted beetroot juice as placebo (0.01 mmol  $NO_3^-$ ). A

minimum 7 day washout period was employed. Magnetic resonance imaging was used to assess apparent diffusion coefficient (ADC), portal vein flux and velocity. Plasma glucose concentrations and blood pressure were also assessed. On each visit outcome variables were measured at baseline and again every hour for 3 subsequent hours. Repeated measures ANOVA were used to assess the interaction effect of time and supplement.

### Results:

Compared with a placebo, beetroot juice resulted in a physiologically meaningful and significant elevation in plasma NO<sub>3</sub> and plasma nitrite concentration in both cohorts. No differences were seen between placebo and beetroot juice for ADC (young adults:  $F_{(3, 45)} = 0.25$ , P = 0.74; older adults;  $F_{(3, 42)} = 1.3$ , P = 0.28), portal vein flux (young adults:  $F_{(3, 45)} = 0.339$ , P = 0.79; older adults;  $F_{(3, 42)} = 1.65$ , P = 0.19), however, there was an interaction effect in the young adults: ( $F_{(3, 45)} = 2.9$ , P = 0.04) but not in the older adults;  $F_{(3, 42)} = 1.8$ , P = 0.16) between visits for portal vein velocity. Nitrate supplementation did not reduce plasma glucose concentrations (young adults:  $F_{(3, 45)} = 0.96$ , P = 0.42; older adults;  $F_{(3, 42)} = 0.04$ , P = 0.99). Nitrate supplementation did not reduce systolic blood pressure (young adults:  $F_{(3, 45)} = 0.20$ , P = 0.89; older adults;  $F_{(3, 42)} = 1.7$ , P = 0.18) or diastolic blood pressure (young adults:  $F_{(3, 45)} = 0.25$ , P = 0.86; older adults;  $F_{(3, 42)} = 0.45$ , P = 0.72).

## Conclusion:

This was the first study to examine the hepatic blood flow response to  $NO_3^-$  supplementation. Despite a large and physiologically meaningful elevation in plasma  $NO_2^-$  concentration following an acute dose of 11.91 mmol of  $NO_3^-$ , there was no

effect on hepatic blood flow, plasma glucose concentrations or systolic blood pressure.

### 3.3.4: Introduction

The total cost of direct and indirect care of individuals with T2DM is in the UK is £20.5 billion and equates to 8% of the annual NHS budget (Hex et al., 2012). It is anticipated that by 2030, 439 million individuals will have type 2 diabetes (T2DM) worldwide (Shaw et al., 2010). This represents a serious challenge to global healthcare systems (Wild et al., 2004). T2DM is a disease of progressive hyperglycemia. Hyperglycemia is strongly associated with cardiovascular disease and microvascular complications such as retinopathy, neuropathy, nephropathy (Stratton et al., 2000). Reductions in glycated hemoglobin (HbA<sub>1c</sub>) have been associated with reduced risk of both micro and macrovascular complications (Stratton et al., 2000) and events (Patel et al., 2008).

Diets rich in vegetables have been shown to have beneficial effects on cardiovascular health (Gilchrist et al., 2010), morbidity and mortality (Joshipura et al., 2001) and reduces the risk of developing T2DM (Carter et al., 2010). These diets are rich in inorganic NO<sub>3</sub><sup>-</sup>. There is growing evidence to suggest NO<sub>3</sub><sup>-</sup> is at least in part responsible for these beneficial effects (Bryan and Hord, 2010). Recent reports have shown NO<sub>3</sub><sup>-</sup> supplementation reduces systolic and diastolic blood pressure in healthy older adults (Jajja et al., 2014, Kapil et al., 2015, Kelly et al., 2013a) and in clinical cohorts with elevated cardiovascular risk factors (Berry et al., 2014, Kerley et al., 2015, Kenjale et al., 2011). This may in part be due to its effects on vascular tone and prevention of vasospasm (Cosby et al., 2003, Pluta et al., 2005).

One of NO<sub>3</sub><sup>-</sup> derivatives, nitric oxide (NO) has been shown to mediate glucose uptake from the intestines (Guan et al., 2003) and facilitate its disposal into skeletal muscle in animal models (Merry et al., 2010) and in individuals with T2DM (Kingwell et al., 2002). Whilst another derivative of NO<sub>3</sub><sup>-</sup>, nitrite (NO<sub>2</sub><sup>-</sup>) when supplemented in eNOS deficient mice for 10 weeks has shown to reduce glycated hemoglobin concentrations, lower baseline and postprandial glucose concentrations (Carlström et al., 2010). An vivo study in young adults has shown a reduction in plasma glucose concentrations following nitrate supplementation following exercise (Wylie et al., 2013b). Recently, another study has described reduction in baseline plasma glucose concentrations 2.5h after supplementation with pharmacological sodium NO<sub>3</sub><sup>-</sup> in individuals with T2DM whilst there was no effect on post prandial glucose concentrations following an oral glucose tolerance test (Cermak et al., 2015).

Incretins are a group of gastrointestinal hormones which stimulate NO· production (Hattori et al., 2010, Ding et al., 2004). Upon ingestion of food, plasma incretin concentrations rise following release from the small intestines (Holst, 2007) and concurrently, NO· increases gastric blood flow (Björne et al., 2004, Petersson et al., 2007). Incretins are intrinsically linked with insulin and are a key component in glucose homeostasis via their insulinotropic effect. Incretins mediate the uptake of glucose in the intestines in a NO· dependent fashion (Guan et al., 2003). Incretins have also been shown to expedite the production of NO· within the portal vein (Ding et al., 2004).

NO· is produced within the body via two independent pathways. The first pathway in which NO· is produced is via the amino acid L-arginine combining with an enzyme, nitric oxide synthase (NOS) and is O<sub>2</sub> dependent (Alderton et al., 2001). The second pathway, known as the entero-salivary pathway is O<sub>2</sub> independent. Nitrate from the

diet is ingested, absorbed through the stomach wall and proximal small intestine (Bartholomew and Hill, 1984, Florin et al., 1990) into the circulation where it is concentrated in the salivary glands. A reduction of NO<sub>3</sub> to NO<sub>2</sub> via facultative anaerobic bacteria occurs within the mouth (Duncan et al., 1995). Nitrite is then swallowed where some NO<sub>2</sub> is converted to NO· in the acidic environment of the stomach (Benjamin et al., 1994) whilst some of the NO<sub>2</sub> is absorbed into the circulation and acts as a storage pool for subsequently conversion to NO. (Lundberg et al., 2008). Another possible mechanism exists for the conversion of NO<sub>3</sub> to NO<sub>2</sub>, whereby hepatic xanthine oxidoreductase reduces NO<sub>3</sub> to NO<sub>2</sub> (Jansson et al., 2008). This mechanism in conjunction with the uptake of NO<sub>2</sub> into the portal circulation may explain why one of the highest concentrations of nitrite in any organ are found within the liver (Totzeck et al., 2012, Bryan et al., 2005). Subsequent increases in the bioavailability of NO· within the liver may be expedited by a number of nitrite reductases which are in relatively high concentrations such as; xanthine oxidoreductase (Li et al., 2008b, Martin et al., 2004), aldehyde oxidase (Li et al., 2008a, Kundu et al., 2012), neuroglobin (Fordel et al., 2007, Tiso et al., 2011), cytoglobin (Burmester et al., 2002, Fordel et al., 2007, Li et al., 2012), deoxyhaemoglobin (Cosby et al., 2003) and myoglobin (Cossins et al., 2009). The potential increase of NO· within the hepatic vasculature may lead to vasodilation of the parenchyma and lead to greater surface area for glucose uptake to occur. Increases in the bioavailability of NO has also bene shown to stimulate insulin secretion (Nystrom et al., 2012) and increase GLUT4 translocation (Li et al., 2004).

# Purpose / hypothesis

By reducing post-prandial hyperglycaemia it may be possible to reduce the associated micro and macrovascular complications (Stratton et al., 2000). A

reduction in hyperglycaemic exertions is a long term goal in the treatment of T2DM in order to protect against cardiovascular events and microvascular damage (Shamoon et al., 1993, Del Prato et al., 2005) and even the risk of morbidity and mortality. Therefore, dietary NO<sub>3</sub><sup>-</sup> may offer a simple, low cost means of modifying diabetes and cardiovascular risk factors.

The aim of this study was to assess if inorganic NO<sub>3</sub><sup>-</sup> modulates portal vein flux and velocity and hepatic microvascular diffusion and secondly; does NO<sub>3</sub><sup>-</sup> lower post-prandial plasma glucose concentrations and blood pressure in healthy young and older adults. We hypothesised that supplementation of the diet with dietary NO<sub>3</sub><sup>-</sup> would increase blood flow to the liver and vasodilate the microvasculature causing improved postprandial glucose uptake. We also hypothesise that dietary nitrate supplementation would reduce blood pressure.

### 3.3.5: Methods

#### Volunteers

37 individuals (17 healthy young individuals and 20 healthy older adults) provided written informed consent to participate in this double-blind, placebo-controlled, crossover design study (see table 12 for subject characteristic). The healthy young individuals were recruited via word of mouth. The older adults were recruited via the NIHR Exeter Clinical Research Facility, Exeter 10,000 cohort. This is a database of individuals who have been pre-screened and consented to be contacted as research volunteers. The trial commenced in July 2014 and ended in April 2015. Ethical approval was obtained from the Exeter NRES Committee (14/SW/0092). This trial was registered on the ClinicalTrials.gov website (NCT02195856). Healthy young individuals were recruited if they were aged between 18 and 35. Older adults were

recruited if they were aged between 50 and 75. Exclusion criteria were incapacity to consent, on any regular vasoactive medications, uncontrolled hypertension, antibiotic therapy within the preceding two weeks, on regular organic NO<sub>3</sub>-, thiazolidinidiones or nicorandil, severe claustrophobia, were smokers (smoked within past 3 months), presence of chronic kidney disease (estimated glomerular filtration rate (eGFR) <30 ml/min/1.73 m<sup>2</sup>), myocardial infarction or cerebro-vascular event within the preceding 3 months, previous brain surgery, cardiac pacemaker, metal fragments in the eye or larger metal objects that would interfere with data collection or analysis. Volunteers, who had medical interventions where metal implants (including some plates, clips, staples or sutures) were inserted, were thoroughly assessed to determine safety in the scanner.

Table 11. Participant characteristics included in the final analysis.

	Young adults	Older adults
n	16	15
Age (years)	26.6	59.2
Male (%)	68.8	53.3
Height (m)	1.75	1.69
Weight (kg)	76.2	75.3
BMI (kg/m²)	24.6	26.3
Baseline SBP (mmHg)	121.0	130.0
Baseline DBP (mmHg)	70.0	79.0

# **Experimental Overview**

Screening and consent took place at the Diabetes and Vascular Research Centre at the NIHR Exeter Clinical Research Facility. Following arrival, volunteers completed informed consent followed by a medical history, anthropometric measures and a resting ECG were taken. Venous blood samples were taken and checked for markers of undiagnosed liver disease, lipid profile, eGFR and glycated haemoglobin. Following the screening checks the volunteers were randomly assigned to a double-blind crossover experimental design to consume 140 ml of NO<sub>3</sub><sup>-</sup> rich beetroot juice (beetroot juice; containing 11.91 mmol of NO<sub>3</sub><sup>-</sup>; Beet it, James White Drinks Ltd., lpswich) or NO<sub>3</sub><sup>-</sup> depleted beetroot juice (placebo; NO<sub>3</sub><sup>-</sup> depleted beetroot juice containing 0.01 mmol of NO<sub>3</sub><sup>-</sup>; Beet it, James White Drinks Ltd., lpswich). The placebo production has been detailed previously (Gilchrist et al., 2014); the final product is identical in appearance, odour taste, colour and texture.

Volunteers were asked to fast overnight (from 10pm) although water consumption was allowed to ensure they arrived in a hydrated state. Volunteers were asked to refrain from antibacterial mouthwash throughout the study and for at least 7 days prior to the experimental visits. Antibacterial mouthwash has been demonstrated to reduce the concentration of oral bacterial anaerobes responsible for the reduction of NO<sub>3</sub><sup>-</sup> in the entero-salivary pathway (Govoni et al., 2008). Volunteers were also asked to avoid caffeine for 12 hours, alcohol and strenuous activity for 24 hours and NO<sub>3</sub><sup>-</sup> rich foods on the day prior to their visits..

Volunteers arrived at the Peninsula Magnetic Resonance Research Centre at the University of Exeter. A 30 minute acclimatisation period was implemented prior to the magnetic resonance imaging (MRI) scans. During this acclimatisation period a cannula was inserted in order to take baseline plasma concentrations for glucose (fluoride & EDTA tubes; Sarstedt, S-Monovette, Nümbrecht, Germany). Plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> were collected (lithium heparin tubes; Sarstedt, S-Monovette, Nümbrecht, Germany) and analysis was performed as previously described (Gilchrist et al., 2013). Immediately prior to the baseline MRI scan, 5 resting blood pressure (BP)

measurements were taken (Schiller Medical, Wissembourg, France) and an average of the final 3 recorded.

Following the baseline MRI scans the volunteers were provided with either the NO<sub>3</sub>-rich beetroot juice or the placebo with two slices of toast, butter and jam. The combined quantity of carbohydrate equated to 76 grams and is equivalent to that that would be consumed during an oral glucose tolerance test (Bartoli et al., 2011a). Every hour, for three subsequent hours, from the consumption of the beetroot juice another set of scans were performed. Immediately prior to each scan brachial artery blood pressure and venous blood samples were taken and processed as previously described. A minimum 7 day washout period between the crossover was employed.

### **MRI** scans

A 1.5 T (Philips, Amsterdam, The Netherlands) magnetic resonance imaging (MRI) scanner was used in order to examine changes in velocity and flux in the portal vein and microvascular diffusion in the posterior right lobe of the liver.

Initial structural images were obtained to orientate the portal vein and an 8mm slice was selected perpendicular to the long axis of the vein. To determine flux and velocity a cardiac triggered velocity sensitive phase encoding imaging sequence (Gatehouse et al., 2005) was employed which obtained image data at 20 time points throughout the cardiac cycle. Analysis of the portal vein was subsequently undertaken using a package supplied as part of the general scanner software. For each separate measurement the circumference of the vessel was manually drawn and recorded for each of the 20 time points to establish a defined region of interest (ROI). Within this ROI, flow and velocity were automatically calculated to give

profiles throughout the cardiac cycle. A mean across the cardiac cycle for flux and velocity was created.

To examine the microvascular diffusion in the posterior right lobe of the liver, a magnetic resonance sequence sensitive to flow was employed via the application of magnetic field gradients in three orthogonal directions. Microvascular diffusion was averaged over all directions within the region of interest and is known as the apparent diffusion coefficient (ADC). Day to day repeatability was assessed in 6 individuals for ADC (1 ROI: ADC =  $1.15 \pm 0.12$ , CV = 9.77) and with multiple ROI. One ROI away from any major vessels had greater repeatability that 2, 3 and 6 sites (2 ROI: ADC =  $1.11 \pm 0.21$ , CV = 15.29; 3 ROI: ADC =  $1.10 \pm 0.23$ , CV = 21.34; 6 ROI: ADC =  $1.09 \pm 0.33$ , CV = 30.32). To calculate ADC within the posterior left lobe of the liver, a ROI (typically  $2500 \text{mm}^3$ ) was manually drawn using the scanner software and the signal intensity within determined. ADC was subsequently calculated based upon the ratio of signal intensity from the two images generated from the MR sequence employed, one of which was sensitive to flow, whereas the other had a low sensitivity to flow, where:

$$ADC = -1/(b_1-b_0) Ln (S_0/S_1)$$

S<sub>0</sub> signal intensity in low flow sensitivity image

S<sub>1</sub> signal intensity in flow sensitive image

b<sub>0</sub> magnetic field gradient used in low flow sensitivity image=250 s/mm<sup>2</sup>

b<sub>1</sub> magnetic field gradient used in flow sensitive image=750 s/mm<sup>2</sup>.

## Sample size and randomisation

For 90% power with an  $\alpha$ -level set at P=0.05 (two tailed), to detect a 1 SD difference, 13 volunteers were required to compare between placebo and active conditions. A computer program was used to generate a randomisation schedule for both groups.

### Data and statistical analysis

All data were tested for normality. Data are presented as means  $\pm$  standard deviation (*SD*). Statistical analyses were performed on SPSS software version 21.0 (Chicago, IL, USA). Statistical difference was accepted when P < 0.05. Statistical differences were assessed using paired samples t-tests.

### 3.3.6: Results

37 individuals (17 healthy young individuals and 20 healthy older adults) gave written informed consent to participate. Post screening and consent 6 individuals were excluded from the trial. 1 individual had abnormal liver function, 1 had a metal pin (in an area which would interfere with data collection) and 4 had previously undiagnosed claustrophobia. 31 individuals (16 healthy young individuals and 15 healthy older adults) were randomised to start in either the NO<sub>3</sub><sup>-</sup> rich beetroot arm or the placebo arm. No differences between dietary intake or exercise patterns were recorded prior to both study visits. The beetroot juice was well tolerated and no adverse events were reported.

### Plasma NO<sub>3</sub><sup>-</sup> concentration:

Repeated measures ANOVA revealed a significant main effect for time, supplementation and an interaction for plasma  $NO_3^-$  concentration in the young adults and the older adults; P < 0.001. Post hoc analysis revealed no significant

differences at baseline (prior to any supplementation) for plasma NO $_3$ <sup>-</sup> concentration (young adults: mean difference; 2 ± 4.4  $\mu$ M, P = 0.64, 95% CI -7.4, 11.5; older adults: mean difference; 1.5 ± 4.1  $\mu$ M, P = 0.72, 95% CI -10.4, 7.4) Post hoc analysis revealed a significant increase when beetroot juice was compared with placebo at 1 hour post supplementation (young adult: mean difference; 543 ± 37  $\mu$ M, P < 0.001, 95% CI 463, 624; older adult: mean difference; 505 ± 39  $\mu$ M, P < 0.001, 95% CI 420, 590), 2 hours post (young adult: mean difference; 645 ± 29  $\mu$ M, P < 0.001, 95% CI 581, 707; older adult: mean difference; 632 ± 35  $\mu$ M, P < 0.001, 95% CI 556, 710) and 3 hours post (young adult: mean difference; 598 ± 31  $\mu$ M, P < 0.001, 95% CI 530, 665; older adult: mean difference; 616 ± 26  $\mu$ M, P < 0.001, 95% CI 559, 673).

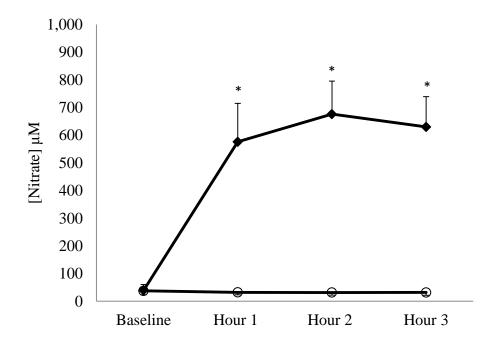


Figure 16. Plasma NO<sub>3</sub> concentration in a young adult cohort.

This figure shows plasma  $NO_3^-$  concentration for the young adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot juice. The \* represents a significant difference when beetroot juice is compared with placebo < 0.001.

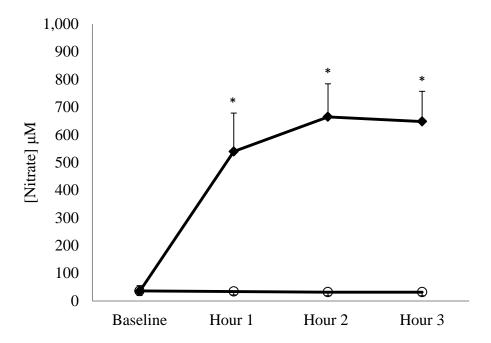


Figure 17. Plasma NO<sub>3</sub> concentration in a older adult cohort.

This figure shows plasma  $NO_3^-$  concentration for the older adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot juice. The \* represents a significant difference when beetroot juice is compared with placebo < 0.001.

### Plasma NO<sub>2</sub> concentration:

Repeated measures ANOVA revealed a significant main effect for time, supplementation and an interaction for plasma  $NO_2^-$  concentration in the young adults and the older adults; P < 0.001. Post hoc analysis revealed no significant differences at baseline (prior to any supplementation) for plasma  $NO_2^-$  concentration (young adults: mean difference; -3.3 ± 74 nM, P = 0.86, 95% CI -43, 36; older adults: mean difference; 26.5 ± 78 nM, P = 0.21, 95% CI -17, 69) Post hoc analysis revealed a significant increase when beetroot juice was compared with placebo at 1 hour post supplementation (young adults: mean difference; 283 ± 201 nM, P < 0.001, 95% CI 176, 391: older adults: mean difference; 471 ± 381 nM, P < 0.001, 95% CI 260, 682), 2 hours post (young adults: mean difference; 497 ± 259 nM, P < 0.001,

95% CI 353, 640: older adults: mean difference; 545  $\pm$  325 nM, P < 0.001, 95% CI 364, 325) and 3 hours post (young adults: mean difference; 559  $\pm$  201 nM, P < 0.001, 95% CI 442, 675: older adults: mean difference; 797  $\pm$  525 nM, P < 0.001, 95% CI 493, 1100). There was also a significant increase at peak plasma nitrite concentration (hour 3), compared with hour 2 (mean difference; 201  $\pm$  344 nM, P = 0.039, 95% CI 11, 392).

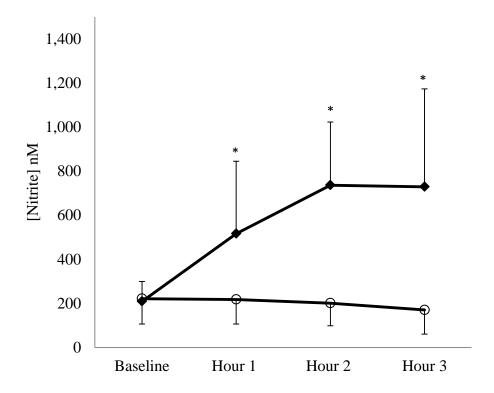


Figure 18. Plasma NO<sub>2</sub> concentration in a young adult cohort.

This figure depicts plasma NO<sub>2</sub> concentration for the young adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot juice. The \*represents a significant difference when beetroot juice is compared with placebo < 0.001.

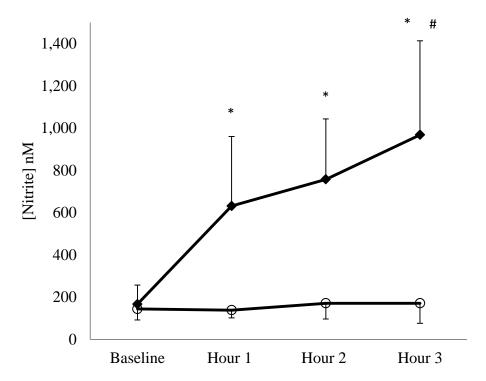


Figure 19. Plasma NO<sub>2</sub> concentration in a older adult cohort.

This figure depicts plasma  $NO_2^-$  concentration for the older adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot juice. The \* represents a significant difference when beetroot juice is compared with placebo < 0.001. The # shows a significant difference in the beetroot condition when hour 2 is compared to hour 3.

### ADC:

Repeated measures ANOVA revealed no significant main effect for time (young adults:  $F_{(3, 45)} = 0.96$ , P = 0.42; older adults;  $F_{(3, 42)} = 0.27$ , P = 0.85), supplementation (young adults:  $F_{(1, 15)} = .314$ , P = 0.58; older adults;  $F_{(1, 14)} = 1.65$ , P = 0.22) or an interaction (young adults:  $F_{(3, 45)} = 0.25$ , P = 0.74; older adults;  $F_{(3, 42)} = 1.3$ , P = 0.28) between visits for ADC index.

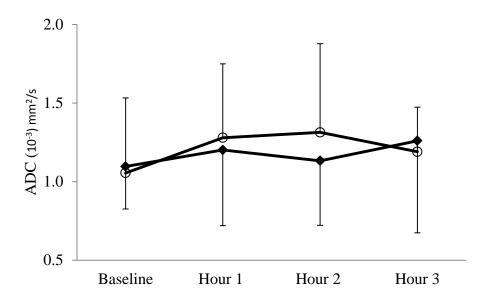


Figure 20. ADC in a young adult cohort across time for beetroot and placebo juice.

The figure shows ADC (microvascular diffusion) for the young adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot juice.

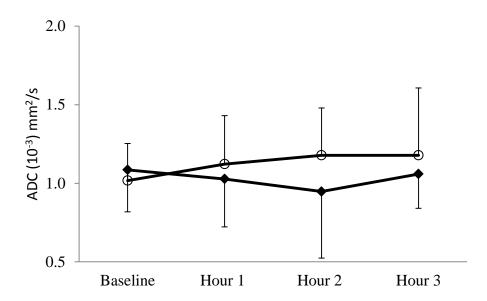


Figure 21. ADC in a older adult cohort across time for beetroot and placebo juice.

This table shows ADC (microvascular diffusion) for the older adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot.

#### Portal Vein Flux

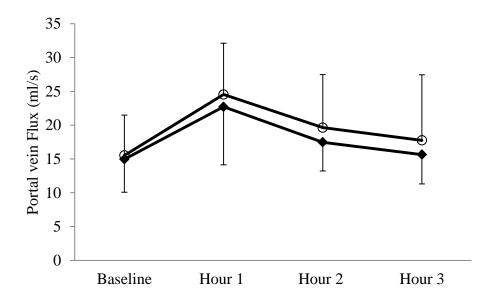


Figure 22. Portal vein flux in a young adult cohort across time for beetroot and placebo juice.

This figure shows portal vein flux for the young adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot.

Repeated measures ANOVA revealed a significant main effect for time (young adults:  $F_{(3, 45)} = 14.07$ , P < 0.001; older adults;  $F_{(3, 42)} = 32.84$ , P < 0.001), however, no significant effect for supplementation (young adults:  $F_{(1, 15)} = 1.00$ , P = 0.33; older adults;  $F_{(1, 14)} = 3.04$ , P = 0.10) or interaction (young adults:  $F_{(3, 45)} = 0.339$ , P = 0.79; older adults;  $F_{(3, 42)} = 1.65$ , P = 0.19) between visits for portal vein flux. However, there was a baseline difference for older adults (older adults: placebo; 14.6 ± 4.3 vs. beetroot juice; 11.7 ± 2.9 ml/s, P = 0.04, 95% CI -5.67, -0.13).

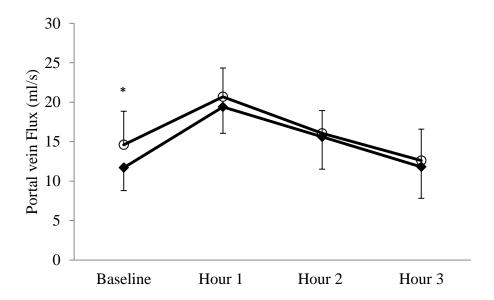


Figure 23. Portal vein flux in an older adult cohort across time for beetroot and placebo juice.

This table shows portal vein flux for the older adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot. The \* represents a statistically different baseline portal vein flux between conditions.

### Portal Vein Velocity

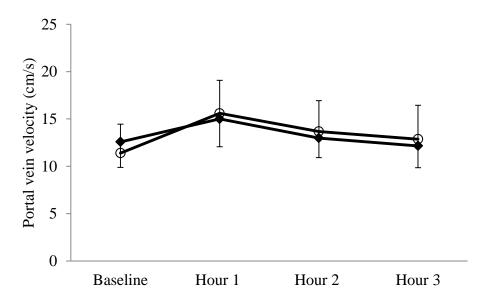


Figure 24. Portal vein velocity in a young adult cohort across time for beetroot and placebo juice.

This figure shows portal vein velocity for the young adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot. The \* represents a statistically different baseline portal vein velocity between conditions.

Repeated measures ANOVA revealed a significant main effect for time (young adults:  $F_{(3, 45)} = 10.7$ , P < 0.001; older adults;  $F_{(3, 42)} = 17.7$ , P < 0.001), however, no significant effect for supplementation (young adults:  $F_{(1, 15)} = 0.1$ , P = 0.75; older adults;  $F_{(1, 14)} = 2.3$ , P = 0.15) however, there was an interaction effect in the young adults: ( $F_{(3, 45)} = 2.9$ , P = 0.04) but not in the older adults;  $F_{(3, 42)} = 1.8$ , P = 0.16) between visits for portal vein velocity. Post hoc analysis of the interaction revealed no statistical differences. However, there was a baseline difference for older adults (older adults: placebo;  $13 \pm 3.4$  vs. beetroot;  $11.1 \pm 3$  cm/s, P = 0.04, 95% CI -3.7, -1.1).

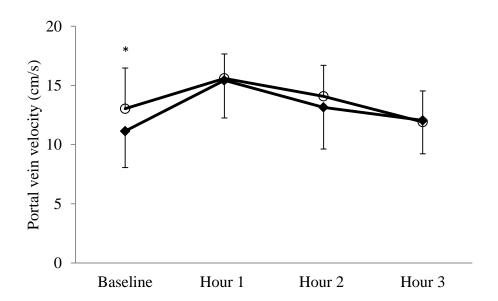


Figure 25. Portal vein velocity in a older adult cohort across time for beetroot and placebo juice.

This figure shows portal vein velocity for the older adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot. The \* represents a statistically different baseline portal vein velocity between conditions.

### Plasma glucose concentration:

Repeated measures ANOVA revealed a significant main effect for time (young adults:  $F_{(3, 45)} = 7.3$ , P < 0.001; older adults;  $F_{(3, 42)} = 17.7$ , P < 0.001), however, no significant main effect on supplementation (young adults:  $F_{(1, 15)} = 0.96$ , P = 0.35; older adults;  $F_{(1, 14)} = 1.4$ , P = 0.26) or an interaction (young adults:  $F_{(3, 45)} = 0.96$ , P = 0.42; older adults;  $F_{(3, 42)} = 0.04$ , P = 0.99) between visits for plasma glucose concentration.

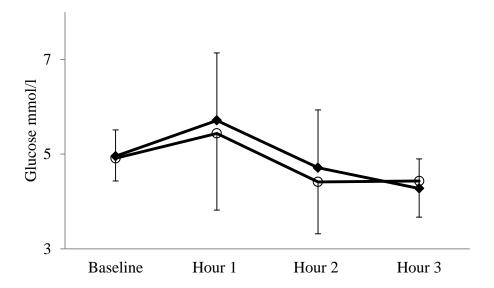


Figure 26. Plasma glucose concentrations in a young adult cohort across time for beetroot and placebo juice.

Figure shows plasma glucose concentration for the young adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot.

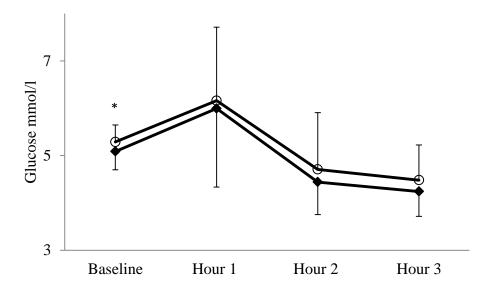


Figure 27. Plasma glucose concentrations in a older adult cohort across time for beetroot and placebo juice.

This figure shows plasma glucose concentration for the older adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot. The \*represents a statistically different baseline glucose concentration between conditions.

### Effects on resting blood pressure:

Repeated measures ANOVA revealed a significant main effect for time (young adults:  $F_{(3, 45)} = 4.5$ , P = 0.008; older adults;  $F_{(3, 42)} = 13.3$ , P < 0.001), however, no significant main effect on supplementation (young adults:  $F_{(1, 15)} = 1.2$ , P = 0.28; older adults;  $F_{(1, 14)} = 1.7$ , P = 0.20) or an interaction (young adults:  $F_{(3, 45)} = 0.20$ , P = 0.89; older adults;  $F_{(3, 42)} = 1.7$ , P = 0.18) between visits for systolic blood pressure.

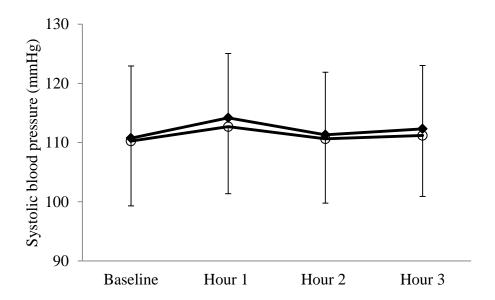


Figure 28. Systolic blood pressure in a young adult cohort across time for beetroot and placebo juice.

Depicts systolic blood pressure for the young adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot.

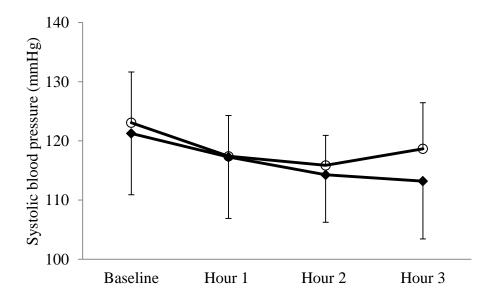


Figure 29. Systolic blood pressure in a older adult cohort across time for beetroot and placebo juice.

Depicts systolic blood pressure for the older adult cohort with changes across time for beetroot and placebo juice.

Repeated measures ANOVA revealed a non-significant main effect on diastolic blood pressure for time in young adults ( $F_{(3, 45)} = 2.4$ , P = 0.08) and a significant main effect of time in older adults ( $F_{(3, 42)} = 18.8$ , P < 0.001), however, no significant main effect on supplementation in young adults ( $F_{(1, 15)} = 2.6$ , P = 0.13). There was however, a trend for a reduction as a main effect of supplementation in older adults ( $F_{(1, 14)} = 4.0$ , P = 0.06). No effect on interaction (young adults:  $F_{(3, 45)} = 0.25$ , P = 0.86; older adults;  $F_{(3, 42)} = 0.45$ , P = 0.72) was found between visits for diastolic blood pressure.

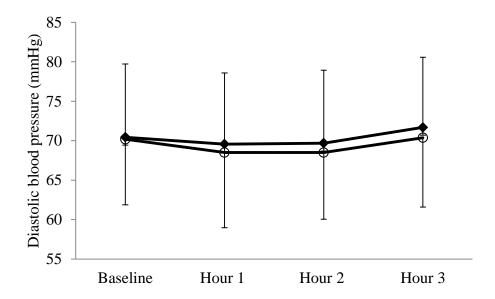


Figure 30. Diastolic blood pressure in a young adult cohort across time for beetroot and placebo juice.

Depicts diastolic blood pressure for the young adult cohort with changes across time for beetroot and

placebo juice. The open circles are placebo and the closed circles beetroot.

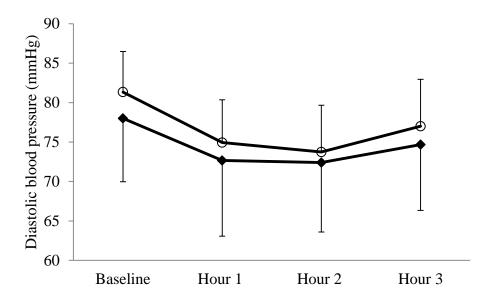


Figure 31. Diastolic blood pressure in a older adult cohort across time for beetroot and placebo juice.

Depicts diastolic blood pressure for the older adult cohort with changes across time for beetroot and placebo juice. The open circles are placebo and the closed circles beetroot.

### 3.3.7: Discussion

This is the first study to investigate the effects of dietary NO<sub>3</sub> supplementation on hepatic blood flow and glucose homeostasis in a healthy older adult population. The primary outcomes were to assess changes in microvascular diffusion (ADC), portal vein flux and velocity. Nitrate supplementation did not alter portal vein flux, velocity or affect ADC. Secondary outcomes were to assess plasma glucose concentration and blood pressure changes. Nitrate supplementation did not alter plasma glucose concentration. Nitrate supplementation did not lower systolic or diastolic blood pressure.

Nitrate supplementation and plasma NO<sub>3</sub> and plasma NO<sub>2</sub> concentration.

Plasma NO<sub>3</sub> concentration peaked post NO<sub>3</sub> rich beetroot juice compared with placebo for both the young (30.9 vs. 664.3 μM) and older adult (32.1 vs. 633.8 μM)

groups at 2 hours by ~2000%. Similarly, plasma NO<sub>2</sub><sup>-</sup> concentration rose following NO<sub>3</sub><sup>-</sup> rich beetroot juice compared to the placebo and peaked at 2 hours for the young group (218.3 vs. 715.5 nM) and 3 hours for the older adult group (171 vs. 967.9 nM) representing a ~230 and 460% increase, respectively. The pharmacokinetic response for both plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations of both cohorts are similar to a previously reported does response study in young healthy individuals (Wylie et al., 2013a). However, the pharmacokinetic response of plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations have not previously been reported in a group of healthy older adults. Kelly et al. (2013a) recently reported a rise in plasma NO<sub>2</sub><sup>-</sup> concentration of a similar magnitude by 418% (248 to 1037 nM) in a group of healthy older adults. The pharmacokinetic response in the healthy older adult group shows a peak at 3 hours. This is much higher than at hour 2 (757 nM) and therefore may still have been rising at 3 hours.

The lengthened time to peak plasma NO<sub>2</sub><sup>-</sup> concentration in the older adult cohort may be associated with the commensal oral bacteria which play a fundamental role in the entero-salivary pathway. Serum antibodies to these oral bacteria decrease in concentration as we age (Percival et al., 1996, Challacombe et al., 1995, Percival et al., 1997). These reductions can lead to increased risk of infection and disease from pathogens (Marsh and Percival, 2006). However, this increase in oral microflora may lead to a greater increase in the conversion of NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup>, potentially changing the bacteria's' contribution as a rate limiting factor which may explain why plasma NO<sub>2</sub><sup>-</sup> concentrations were still rising 3 hours after NO<sub>3</sub><sup>-</sup> supplementation. Resting salivary flow rates are not different between young and older healthy adults. Therefore salivation is unlikely to be a contributory factor leading to the delayed peak in plasma

 $NO_2^-$  concentration (Ship and Fischer, 1997). This suggests that more  $NO_3^-$  will be converted to  $NO_2^-$ , however, due to the flow rate of saliva the peak is delayed.

Nitrate supplementation and hepatic blood flow

Despite a statistically significant and physiologically meaningful rise in plasma NO<sub>2</sub> concentration, this did not lead to an increase in portal vein flux, velocity or microvascular diffusion (ADC). The portal vein supplies 75% of inflow, with the remainder supplied by the hepatic artery (Rappaport, 1979) and even though the blood it supplies is partially deoxygenated, the portal vein supplies ~50% of the O<sub>2</sub> delivery (See Hwang., 2011 and Vollmar and Menger., 2009 for reviews). A higher proportion of deoxyhaemoglobin may lead to a faster conversion of NO<sub>2</sub> to NO· (Cosby et al., 2003) in the portal vein. It is therefore likely that the highest concentrations of NO· and plasma glucose concentrations would be found in the liver.

The hepatic artery has the capacity to dilate in response to changes in portal vein flux however, the portal vein cannot dilate in response to changes in hepatic arterial flow (Vollmar and Menger, 2009, Legare and Lautt, 1987). Thus if the portal vein flow decreases the hepatic artery will dilate to increase, flow and if the portal vein increases flow the hepatic artery will constrict to reduce flow (Lautt et al., 1990). This is known as the known as the 'hepatic arterial buffer response' (Hwang, 2011). This buffering response is limited and may be able to compensate for 25 - 60% of change in flow from the portal vein (Lautt, 1980, Lautt, 1977). The underlying reason behind this is to enable a steady perfusion rate in order to manage changes in blood flow from splanchnic organs and the associated hormones and nutrients (Lautt, 1980, Lautt, 1977). This could, in part explain the lack of effect of an increased plasma

NO<sub>2</sub><sup>-</sup> concentration on portal vein blood flow. As expected, there was an increase in portal vein flux and velocity from baseline to the liver regardless of condition.

There was, however, a statistical difference in baseline portal vein flux (placebo: 14.6 and active 11.7 cm/s; mean difference of 2.9 cm/s) and velocity (placebo: 13 and active 11.1 cm/s; mean difference of 1.9 ml/s) on the two visits. The day to day coefficient of variation for portal vein flux and velocity was 13% and 16% respectively. The mean differences are outside our repeatability and are likely to be genuine differences between visits. This suggests that portal vein flux and velocity were faster on the placebo visit at baseline. This may cloud any differences between the active condition and placebo later in the time course. Participants were fasted overnight, having avoided caffeine, strenuous exercise and were in a hydrated state. Therefore it is unclear why there is a baseline difference. It may be due to a type 2 error.

### Nitrate supplementation and plasma glucose concentrations

There was no significant difference between active or placebo juice for plasma glucose concentration. It should be noted that in the older adult cohort there was statistically significant baseline difference in plasma glucose concentration (mean difference = -0.2 mmol/l or 3.8%). On the active testing days plasma glucose concentration was already ~4% lower than placebo. Mechanisms for the uptake of glucose have been partially elucidated. NO· is known to mediate glucose uptake from the intestines and skeletal muscle (Merry et al., 2010, Guan et al., 2003, Kingwell et al., 2002). Potentially via elevated NO· bioavailability which has been shown to stimulate insulin secretion (Nystrom et al., 2012) and increase GLUT4 translocation (Li et al., 2004). The potential to modifying this diabetes risk factor via

reductions in plasma glucose concentrations would likely reduce micro and macrovascular complications which are a severe cause of reduced quality of life, morbidity and mortality (Sprafka et al., 1991, Jaffe et al., 1984). Larger samples sizes that are appropriately powered are required to assess the effect of nitrate supplementation on plasma glucose concentrations.

Nitrate supplementation and resting blood pressure.

There was no significant difference in systolic or diastolic BP following NO<sub>3</sub> rich beetroot juice supplementation (at peak plasma NO<sub>2</sub> concentration) compared to placebo in the healthy young adult cohort. This is similar to some acute NO<sub>3</sub><sup>-</sup> supplementation studies (Wilkerson et al., 2012, Coles and Clifton, 2012, Sandbakk et al., 2014). However, most studies have shown hypotensive effects with acute supplementation regimens (Webb et al., 2008a, Vanhatalo et al., 2010, Vanhatalo et al., 2011, Lansley et al., 2010, Lansley et al., 2011a). Larger sufficiently powered trials are required to determine the effect of NO<sub>3</sub> as a hypotensive agent in young individuals. There was no significant reduction in systolic BP following NO<sub>3</sub> rich beetroot juice compared to placebo in the healthy older adult cohort. We report a non significant 5 mm Hg drop in systolic BP. Kelly et al. (2013a) report a statistically significant 5 mmHg drop in systolic BP, whilst another groups reported larger reductions (Jajja et al., 2014). Another study in healthy older adults reported no effect of nitrate supplementation on systolic BP (Bondonno et al., 2014b). Kelly et al. reported diastolic BP reductions of 3 mmHg. Whereas we report a non-significant change in diastolic BP of2 mmHg. A detailed pharmacokinetic study in healthy older adults over a prolonged period may elucidate at what time peak plasma NO<sub>2</sub>- $NO_2^$ concentrations occur. If the pharmacokinetic response of plasma

concentrations in the healthy older adult group continued to rise past 3 hours, a greater hypotensive effect than measured here may have occurred.

Strengths and limitations.

This is the first study to examine the effect of NO<sub>3</sub><sup>-</sup> supplementation on hepatic blood flow. This study has a robust experimental design as a double-blind, placebo controlled, crossover trial. A limitation to this study is that we did not measure plasma NO<sub>2</sub><sup>-</sup> concentration beyond 3 hours supplementation. Future research should aim to elucidate if plasma NO<sub>2</sub><sup>-</sup> concentration rises beyond 3 hours in healthy older adults.

### 3.3.7: Conclusion

This was the first study to examine the hepatic blood flow response to  $NO_3^-$  supplementation. Despite a large and physiologically meaningful elevation in plasma  $NO_2^-$  concentration following an acute dose of 11.91 mmol of  $NO_3^-$ , there was no effect on hepatic blood flow, plasma glucose concentrations or systolic blood pressure.

#### 4.0: General discussion

This body of work has made a number of novel contributions in this area of research; it was the first study to investigate the effects of dietary  $NO_3^-$  supplementation on the exercise responses of individuals with T2DM. Dietary  $NO_3^-$  supplementation did not reduce the  $O_2$  cost of walking in individuals with T2DM or increase the distance covered in the six minute walk test. In addition this work confirms previous findings that  $NO_3^-$  supplementation has no effect on resting blood pressure in individuals with T2DM.

This was the third peer reviewed article to assess the effects on individuals with COPD (all published in *Nitric Oxide* within 6 weeks of each other). Dietary NO<sub>3</sub> supplementation did not reduce the O<sub>2</sub> cost of cycling in individuals with COPD, or increase the distance covered in the six minute walk test. In contrast to the other reports in individuals with COPD, no hypotensive effect was found.

This is the first study to examine the liver diffusion or delivery. The data shows no increase in liver diffusion or delivery. The data shows no effect of nitrate in blood glucose concentrations or blood pressure.

Dietary NO<sub>3</sub> supplementation in the form of beetroot juice (in an older adult population) is unlikely to reduce the O<sub>2</sub> cost of exercise, improve walking performance or reduce BP in individuals with T2DM and COPD. Moreover, it does not alter glucose homeostasis or affect blood pressure in healthy adults.

I will begin the discussion with an update of the studies published since the beginning of this body of work. Table 13 describes each trial published and each of the key outcome measures assessed. I will then describe and critically appraise my findings in the context of recent literature.

Table 12: A, B and C. Publications on NO<sub>3</sub> supplementation from 2012 – April 2015 (split via age and disease status).

A, Healthy young individuals

B, Healthy older individuals

C, Clinical populations

Studies in table 13, B and C (that are not abstracts), will be critically reviewed in the discussion below. Please see marked with a \*.

Table A: Nitrate supplementation studies in healthy young individuals

Author	Participants & Protocol	NO <sub>3</sub> supplement	Plasma NO <sub>3</sub> <sup>-</sup> & NO <sub>2</sub> <sup>-</sup> concentration (BR vs. PL respectively)	BP changes	Exercise efficiency or performance and/or clinical relevance	Primary outcome successful?
Wilkerson et al. (2012)	8 well-trained male cyclists. A minimum of a 7 day washout was given. 4BP measurements were taken on each visit. Exercise involved a 50 mile TT.	500ml of BR juice (NO <sub>3</sub> = 6.2mmol) or PL (NO <sub>3</sub> = 0.005mmol) 2.5 hours prior to exercise.	NO <sub>2</sub> = 379 ± 94 and 472 ± 96 nM	⇔SBP, DBP or MAP	↔ TT performance	No
Masschelein et al. (2012)	15 healthy volunteers. With a 14 day washout period. Exercise included cycling in normoxia and hypoxia.	500ml of BR (0.07 Kg <sup>-1</sup> day <sup>-1</sup> of body weight) for 6 days or PL (apple and blackcurrant).	$NO_2^- = 3.8 \pm 0.1$ and $5.3 \pm 0.3 \mu M$ $NO_3^- = 41 \pm 3$ and $147 \pm 7 \mu M$	N/A	BR negates some negative effects of hypoxia.  ↑Arterial and muscle oxygenation.	Mixed

Engan et al. (2012)	12 healthy subjects. Tests; vital capacity, apnoeic duration and arterial O <sub>2</sub> saturation. Washout period not stated.	70ml of BR juice (NO <sub>3</sub> <sup>-</sup> =5mmol) or PL (NO <sub>3</sub> <sup>-</sup> =0.003mmol) 2.5h prior to each test.	Figures not given.	↓MAP	↓S <sub>aO2</sub> ↑ Max apnoeic duration by 11%	Yes
Hobbs et al. (2012).	14 subjects (sex and disease status unknown). 24h ambulatory BP was measured.	Subjects consumed BR juice (0, 100, 250 or 500g) or water (PL) and bread (PL) or BR bread (white and red beets).	Abstract only, figures not presented.	N/A	↓BP in a ~ dose dependant manor.  ↔ BP for white BR bread	Yes
Bond et al. (2012). (Abstract only)	14 well-trained junior male rowers. A 7 day washout period was employed. Exercise included 6 maximal 500m ergometer reps.	500ml of BR or PL daily for 6 days.	Abstract only, figures not presented.	N/A	↑ average repetition time	Yes
Coles and Clifton (2012)	30 healthy subjects (15 males). 14 day washout was employed. 24h ambulatory blood pressure was taken	500ml of BR juice (NO <sub>3</sub> <sup>-</sup> =15 mmol) or PL (apple juice matched for colour and sweetness)	Figures not given.	Trend for ↓SBP	↓SBP in males at 6h post supplement	Yes
Krajka- Kuzniak et al. (2012).	24 male 6 week old rats. Split into 4 independent groups. DNA damage and liver injury induced by hepatocarcinogenic N-nitrosodiethylamine (NDEA).	Two groups fed beetroot juice and one of two different enzymes. Groups ¾ had PL and water.	Figures not published.	N/A	Protects against oxidative stress Protects against liver damage	Yes

Klewicka et al. (2012)	Caco-2 cell line (Rat cells/colons acquired from 32 rats, 4 groups of 8).	Group 1 had a basal diet. Group 2 was fed a basal diet and BR juice for 30 days. Group 3 was fed BR and a carcinogen. Group 4 had BR and a different carcinogen.	N/A	N/A	Protect against precancerous aberrant crypt formation.  ↓Oxidative stress	Yes
Ferguson et al. (2013)	Rats were fed BR or PL (H <sub>2</sub> O) for 5 days. Microvascular PO <sub>2</sub> was measured during 180s of electorally induced muscle contractions.	BR ( $NO_3^- = \sim 0.8$ mmol/kg/day) or PL ( $H_2O$ )	↑ NO₂ ↑ NO₃ ↑ Values not provided	N/A		Mixed
Christensen et al. (2013)	10 highly trained male cyclists. An approximate 14 day washout was given. The exercise involved a 120 min preload followed by a 400-kcal TT.	500ml/day of BR juice (~0.5g of NO <sub>3</sub> ) per day for 6 days. PL was a blackcurrant cordial. Day 6 was consumed 3h prior to exercise.	$NO_x = 159 \pm 103$ and $40 \pm 7 \mu M$	N/A	↔VO <sub>2</sub> kinetics ↔ TT performance	No
Fulford et al. (2013)	8 healthy male subjects. A minimum of a 14 day washout. Exercise involved two tasks. Repeated MVC and short bouts of continuous exercise.	500ml of BR juice (NO <sub>3</sub> <sup>-</sup> =10.2 mmol) or PL (NO <sub>3</sub> <sup>-</sup> =0.17 mmol) per day for 15 days. Participants arrived 2.5h, 5 and 15 days post supplementation.	NO <sub>2</sub> <sup>-</sup> = 408 ± 131 and 232 ± 49 nM	N/A		Mixed

Wylie et al. (2013b)	14 healthy male subjects. A minimum 72h washout period was used. Exercise involved a Yo-Yo IR1.	BR juice (NO <sub>3</sub> <sup>-</sup> =4.1 mmol) or PL (NO <sub>3</sub> <sup>-</sup> =0.04 mmol). Participants consumed two shots in the morning and two in the evening the day before testing. On the day two shots were consumed 2.5h and 1 shot 1.5h prior to exercise.	$NO_{2}^{-} = 584 \pm 343$ and $118 \pm 44$ nM $NO_{3}^{-} = 768 \pm 180$ and $25 \pm 9$ µM. $NO_{2}^{-}$ values declined by 20% in PL and 54% in BR. $NO_{3}^{-}$ rose by 17% in PL and 10% in BR.	N/A	↑ Yo-Yo performance (4.2%) ↑ Plasma potassium ↓Blood glucose levels	Yes
Wylie et al. (2013a)	10 healthy male subjects. A minimum of 3 days washout was given. Exercise involved a 2 MI and 1 SI bout on each of 6 visits (1 visit per condition).	Participants consumed 70ml, 140ml and 280ml BR juice (NO <sub>3</sub> <sup>-</sup> = 4.2 mmol) or PL ((NO <sub>3</sub> <sup>-</sup> =0.04 mmol) per shot).	$NO_2^- = 425 \pm 225$ and $150 \pm 73$ nM $NO_3^- = 580 \pm 89$ and $130 \pm 17 \mu$ M. $\uparrow$ in a dose dependant manor.	↓SBP (peak at 4h) ↓DBP (peak at 4h) ↓MAP (peak at 1h)	140 & 280 ↓ steady state O₂ and time to task failure.	Yes
Kelly et al. (2013b)	9 healthy active males. Exercise involved 4 SI bouts at different % of peak power to exhaustion.	500ml of BR juice (NO <sub>3</sub> = 8.2 mmol) or PL (NO <sub>3</sub> = 0.006 mmol)	$NO_2^- = 297 \pm 98 \text{ nM}$ and $98 \pm 41 \text{ nM}$	N/A	↑ at 60%, 70% and 80% peak power but not 100%. ↔CP or W	Yes
Breese et al. (2013)	9 (4 males) healthy subjects. A 7 day washout period was utilised. NIRS, muscle oxygenation and VO <sub>2</sub> kinetics were assessed.	Participants consumed 70ml, BR juice (NO <sub>3</sub> = 4 mmol) or PL ((NO <sub>3</sub> = 0.002 mmol) per shot) in the morning and the evening for 6 days.	NO <sub>2</sub> = 348 ± 170 nM and 65 ± 32 nM	N/A	↑ speed of VO₂ kinetics ↑ deoxyhaemoglobin kinetics (derived from NIRS) ↑time to task failure	Yes

Hoon et al. (2013)	10 highly trained male rowers. Exercise involved a 2000m TT.	Participants consumed 70ml, BR juice (NO <sub>3</sub> = 4.2 mmol or 8.4 mmol) or PL (NO <sub>3</sub> = 0 mmol).	Abstract only, figures not presented.	N/A		No
Muggeridge et al. (2013a)	8 healthy male kayakers. Exercise included 15m paddling at 60% max WR and 5x10s all out sprints and a 1km TT.	Participants consumed 70ml, BR juice or PL.	NO <sub>2</sub> = $687.9 \pm 20$ and $519.8 \pm 25.8$ nM NO <sub>3</sub> = $152 \pm 3.5$ and $33.8 \pm 1.9$ µM.	N/A	<ul> <li>→ TT performance</li> <li>→ Peak power</li> <li>↓ steady state VO₂</li> </ul>	No
Muggeridge et al. (2013b)	9 male trained cyclists. Exercise included a 15 ride at 60% max WR and a 16.1 km TT at 2500m simulated altitude.	Participants consumed 70ml, BR juice or PL 3h prior to exercise.	NO <sub>2</sub> <sup>-</sup> = 678.1 ± 103.5 and 289.8 ± 27.9 nM NO <sub>3</sub> <sup>-</sup> = 150.5 ± 9.3 and 39.1 ± 3.5 $\mu$ M.	N/A	↓ steady state VO <sub>2</sub> ↑TT performance	Yes
Velmurugan et al. (2013)	24 healthy subjects (12 males). With a 1-4 week washout period. In vivo and in vitro methods were used.	250ml BR juice or K $NO_3$ ( $NO_3$ = 8 mmol) vs. PL.	Figures not published.	N/A	↓Platelet reactivity in males but not in females. Males have a greater dependence on the NO- sGC pathway.	Yes
Lane et al. (2013)	24, Trained cyclists completed 4 experimental conditions. BR, caffeine, BR & caffeine and control. Ergometer TT.	Caffeine = 3 Kg <sup>-1</sup> day <sup>-1</sup> of body weight and BR (8.4 mmol) supplementation 2h prior to testing.	Abstract only, figures not presented.	N/A		No
Bond Jr et al. (2013) (Abstract only)	12 healthy young african americans. Protocol, rest and 40, 60 and 80% peak $\dot{V}O_2$ .	BR and orange juice as placebo.	Abstract only, figures not presented.	↓SBP ↔DBP	↓ VO <sub>2</sub> ↔ in RQ, minute ventilation and HR	Yes

Schiffer et al. (2013)	15 healthy young males. Indirect calorimetry.	Three days (NO <sub>3</sub> = 0.1mmol Kg <sup>-1</sup> day <sup>-1</sup> ) or PL (NaCl)	Published abstract, not data presented.	N/A	↓ metabolic rate     Plasma NO₂     correlates (r 0.7) with     BMR.	Yes
Murphy et al. (2014) (Abstract only)	13 recreationally active males. 10 day washout. Protocol, 10 x 6s repeated sprints with 30s recovery.	BR (5 mmol) supplementation vs. PL 2.5h prior to exercise.	Abstract only, figures not presented.	N/A	↔ VO <sub>2</sub> or fatigue	No
Hoon et al. (2014)	26 well-trained cyclists. 2 x 4min TT, separated by 75 mins. 6 day washout.	70ml BR 75 or 150 mins prior to exercise with 35ml top up following first bout.	Figures not published, but peaked at 75 mins.	N/A	<ul> <li>         ← in TT performance following BR vs. PL. Potential negative effects.     </li> </ul>	No
Boorsma et al. (2014)	8 elite male 1500m runners. 7+ day washout. Protocol incudes a submaximal run and 1500 m TT run.	210ml BR juice for 8 days (NO <sub>3</sub> = 19.5 mmol) vs. PL.	$NO_3^- = 870 \pm 259$ and $37 \pm 15 \mu M$ .	N/A	$\leftrightarrow$ $\dot{V}O_2$ or improvement in TT performance.	No
Peeling et al. (2014)	Study A: 6 male national level kayakers completed a 7 x 4 min lab based step test TT.  Study B: 5 Females national level kayakers completed a 500 m 'on water' TT.	Study A: 70 ml BR (4.8 mmol) or PL (not a true PL) 2.5h prior to exercise.  Study B: 140 ml BR (9.6 mmol) or PL (not a true PL). 2 h prior to exercise.	Abstract only, figures not presented.	N/A	↓ VO₂ (mod)  ↔ in TT performance in the laboratory but  ↑ TT in the field.	Primary outcome not stated.
Richards et al. (2014)	7 young adults performed hand grip strength at 5, 15 and 25% of MVC. Doppler ultrasound was used to assess changes in blood flow.	12.6 mmol BR post baseline. Exercise 2 hours post ingestion.	Abstract only, figures not presented.	N/A	↑ forearm blood flow and sig more as % MVC went up and also in hypoxia	Yes

Thompson et al. (2014)	16 recreationally active males. 20 min stages at 50%, 70% $\dot{V}O_{2 \text{ peak}}$ and exercise to volitional exhaustion at 90% $\dot{V}O_{2 \text{ peak}}$ , 7 day washout.	450ml BR (5 mmol) or blackcurrant PL.	$NO_2^- = 222 \pm 61$ and 124 ± 10 nM	↓ SBP ↔ DBP	↑ exercise tolerance at 90% VO <sub>2peak.</sub> ↔ in RPE, energy levels or cognitive function.	Yes
Martin et al. (2014)	16 recreationally active male (9) and females (7). Repeated 8s sprints with 30s recovery until volitional exhaustion.	70 ml (0.3 g of NO <sub>3</sub> ) or PL 2h prior to exercise.	NOx not measured.	N/A	<ul><li>             ← repeated sprint performance.             ↓ total work         </li></ul>	No
Sandbakk et al. (2014)	9 elite male cross-country skiers. Sub maximal running economy and 5km TT.	A; L-arginine (6g) & NO <sub>3</sub> <sup>-</sup> (614 mg). B; NO <sub>3</sub> <sup>-</sup> (614 mg) & PL. C; L-arginine and PL.	NO <sub>2</sub> = 328 ± 107 nM and 149 ± 64 nM	↔ MAP -	L-arginine did not increase NO₂ concentration.	No
Bond et al. (2014)	13 female African Americans. Washout not specified. Measures were taken at baseline, 40 & 80% VO <sub>2 peak</sub> . HR variability measured via SD of RR intervals of an ECG. Collofello et al. (2014)	500 ml BR (750 mg), PL = orange juice. 2h 20min prior to exercise.	NO = 21.1 ± 4.9 and 4.4 ± 1 nM	↓SBP	↑ Heart rate variability during exercise.	Yes
Collofello et al. (2014)	12 healthy females. 20s submaximal static apnoea with 90s rest followed by a max effort apnoea.	70ml BR juice for 8 days $(NO_3^- = 5 \text{ mmol}) \text{ vs. PL}$ $(NO_3^- = 0.003 \text{ mmol}). 2.5$ post ingestion the measurement begun.	NOx not measured / reported.	↓ SBP ↔ DBP ↔ MAP	↔ maximal static apnoea	No
Pinna et al. (2014)	14 moderately trained male masters swimmers performed two incremental swimming tests.	500ml BR juice for 6 days (NO <sub>3</sub> = 5.5 mmol) vs. no supplementation.	NOx not measured / reported.	N/A	↓ VO₂ during swimming. ↑ anaerobic threshold	Yes

Bentley et al. (2014)	9 healthy males. fMRI was performed at 15% & 25% MVC.	700 ml BR 2.5 h prior to exercise scan vs. no juice.	NOx not measured / reported.	N/A	tissue oxygenation     maximal change (more     effective at lower     intensities)     ↔ recovery rate	Yes
Puype et al. (2014)	22 healthy males in independent groups (n=11). 6 week training program. 5 x 30mins a week sustaining a BLa of 4-6 mmol.	NO <sub>3</sub> = 0.07 mmol Kg <sup>-1</sup> day <sup>-1</sup> or PL (apple-blackcurrant juice) 2.5 h prior to each training session.	NO <sub>3</sub> concentrations not given. Estimate from figure = 165 and 35 μM.	N/A	16% ↑ PO & TT. ↔ activated AMP, glycogen breakdown or mRNA variables.	Yes
Ferguson et al. (2014)	20 rats were split into independent groups. Microvascular PO <sub>2</sub> was measured during 180s of electorally induced twitches	BR juice (NO <sub>3</sub> <sup>-</sup> = 0.08 mmol Kg <sup>-1</sup> day <sup>-1</sup> ) or water day <sup>-1</sup> )	NO <sub>3</sub> <sup>-</sup> & NO <sub>2</sub> <sup>-</sup> concentrations but given. Estimate from figure = NO <sub>3</sub> <sup>-</sup> = 140 and 22 $\mu$ M. NO <sub>2</sub> <sup>-</sup> = 640 and 200 nM	N/A	<ul> <li>→ Microvascular PO<sub>2</sub></li> <li>↓ O<sub>2</sub> driving pressure</li> </ul>	No
Ashmore et al. (2014)	40 male wister rats. Exposed to chronic hypoxia (13%).	0.7 mmol I-1 NaCl (as control) or 0.7 mmol I-1 NaNO3,	Elevated plasma NO <sub>3</sub> levels by 80%. Figures not given but graphical representation is.	N/A	Nitrate maintains redox state (compared to hypoxia) ↑ ETC protein levels ↓ Respiration rate, ↑ ATP ↑ L-arginine	Yes
Muggeridge et al. (2014)	9 trained cyclists. 10 mins sub max followed by a 10 mile TT. 4 conditions, NO <sub>3</sub> GEL and UV-A, UV-A and PL, NO <sub>3</sub> GEL and sham light and PL and sham light	2 gels (=16.2 mmol), 2.5 h prior to exercise	Plasma NO <sub>2</sub> was higher for GEL + SHAM 332; and NIT + UV-A 456 nM; PL + SHAM 215. PL 282 nM	↓ SBP ↔ DBP ↓ MAP	UV-A light and NO <sub>3</sub> alters response to exercise. UV-A does not improve exercise performance alone.	No

Kim et al. (2014) abstract only	12 young healthy adults. 3h post supplementation, brachial artery diameter, flow, and blood velocity were measured (Doppler ultrasound) at rest and during 6 exercise intensities.	140 ml of BR (= 12.9 mmol) or PL.	NO <sub>3</sub> supplementation raised plasma NO <sub>3</sub> (19.5-fold) and NO <sub>2</sub> (1.6-fold) concentrations. Values not given.	N/A		Yes
Carpentier et al. (2014)	13 recreationally active males. 20 min at 85% $\dot{V}O_2$ max	450 mg potassium NO <sub>3</sub> or PL for 7 days.	Not measured.	N/A	↔ kidney function	No
Porcelli et al. (2014)	21 young male individuals (28.2 to 81.7 ml. kg-1. min-1). Sub max running and a incremental running test. + 3k TT	5.5 mmol of NO <sub>3</sub> per day for 6 days.	NO <sub>2</sub> <sup>-</sup> = 462.3 ± 249.5 and 265.7 ± 163.7 nM NO <sub>3</sub> <sup>-</sup> = 181 ± 32.7 and 21.3 ± 3.4 $\mu$ M.	N/A	A strong negative correlation (-0.8) between reduction of O2 cost of submaximal exercise and individual aerobic fitness level was observed. Strong positive correlation (0.75) for aerobic fitness and increase in NO <sub>2</sub> - concentration.	Yes
Keen et al. (2014)	6 healthy adults. Microdialysis fibers on ventral forearm. L-NAME inhibitor. Laser-Doppler	70 ml (5 mmol) one a day for 3 days	Not measured.	↓ MAP ↓ DBP	↑ NOS independent vasodilation to local heating	Yes
Casey et al. (2015)	13 young and 12 older adults. 20% MVC during normoxia and hypoxia. Forearm vascular conductance was measured	140 ml BR or PL (topped up with water to 500ml). NO <sub>3</sub> content not assessed.	Precise figures not given. State that it increased significantly P < 0.0001.	N/A	↑ compensatory vasodilatory response to hypoxia in older but not young adults	Yes

Levitt et al. (2015)	7 healthy males. Microdialysis fibers on ventral forearm. L-NAME inhibitor. Whole body heating	70 ml (5 mmol) one a day for 3 days	Not measured.	↓ MAP     ↓ SBP     ↔ DBP     (during heat stress)	⇔ skin blood flow     ↑ vasodilation in the     skin	No
Rocha et al. (2015)	2 healthy adults and male Wister rats (n not specified). Gastric head space was sampled for ethyl NO <sub>2</sub> and smooth muscle relaxation in the rats	50g of lettice and wine or whiskey	NO particles per billion. Figures not given but in graphical representation.	N/A	Nitrosation of ethanol produced ethyl NO <sub>2</sub> in the stomach after wine and whiskey when consumed with NO <sub>3</sub> rich foods.	Yes
Kocoloski and Crecelius (2015)	6 young, moderate active males. Performed a ramp test and 45 min moderate exercise (38% max).	70 ml 0.4g NO <sub>3</sub> 2 hours prior to exercise. Placebo was created via the use of mouthwash.	Abstract only, figures not presented.	↓ MAP	↔ O <sub>2</sub> cost	No

Table 13, B. Nitrate supplementation studies in healthy older adults.

Author	Participants & Protocol	NO₃⁻ supplement	Plasma NO <sub>3</sub> <sup>-</sup> & NO <sub>2</sub> <sup>-</sup> concentration (BR vs. PL respectively)	BP changes	Exercise efficiency or performance and/or clinical relevance	Primary outcome successful?
Treichler et al. (2015) abstract only	13 young and 12 older normotensive adults. Aortic BP and wave reflection were assessed.	Data not available. Looks like the same cohort as Casey et al., 2015.	Precise figures not given. State that it increased significantly P < 0.0001	↓ SBP ↓ DBP	↓ central aortic pressure	Yes
Jajja et al. (2014)*	21 older healthy participants had BP measured over a 4 week period with 3 different measures. Resting clinic BP, 24 hour ambulatory BP and home monitoring.	70 ml (300 mg of NO <sub>3</sub> ) or PL (blackcurrant juice) 2h prior to exercise.	NO <sub>3</sub> concentrations not given. Estimate from salivary figure = 6 mmol/l and 1.2 mmol/l.			Yes
Kelly et al. (2013a)*	12 healthy older adults (64 ± 4 years). Minimum of a 72h washout. Exercise involved a 6MWT, cognitive function tests and a symptom limited ramp test.	70ml BR juice (NO <sub>3</sub> =4.8 mmol) or PL (NO <sub>3</sub> =0.01 mmol) twice a day for 2.5 days.	NO <sub>2</sub> = 1,037 ± 627 and 248 ± 182 nM	↓SBP ↓DBP ↓MAP	⇔ O₂ cost of exercise     ⇔Cognitive function     ⇔ 6MWT     ⇔Brain metabolite concentrations     ↑VO₂ kinetics	No
Miller et al. (2012). (Abstract only)	13 older (72.5 ± 4.7 years) healthy adults (8 males & 5 females). BP and NO <sub>3</sub> & NO <sub>2</sub> were measured before each meal and hourly for 3h post.	Supplementation involved 3 days of LNF and BR juice, HNF and HNF & BR juice.	↑ NO <sub>3</sub> ⁻ & NO₂⁻ with foods and supplement	↓SBP	All treatment arms ↓SBP in older adults.	Yes

Bondonno et al. (2014b)*	38 (60.6 ± 7 years) healthy older adults. Ambulatory, home and office BP measured. Arterial stiffness	Dietary intervention. Low and high NO <sub>3</sub> diets. High diets = > 300mg/day. Low diet < 100 mg/day.	_	↔ SBP ↔ DBP	Ambulatory BP ↔ Home BP ↔ office BP ↔	No
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Table 13, C. Nitrate supplementation studies in clinical populations.

Author	Participants & Protocol	NO <sub>3</sub> <sup>-</sup> supplement	Plasma NO <sub>3</sub> <sup>-</sup> & NO <sub>2</sub> <sup>-</sup> concentration (BR vs. PL respectively)	BP changes	Exercise efficiency or performance and/or clinical relevance	Primary outcome successful?
Kapil et al. (2015)*	68 hypertensive adults. 2 week washout. BP measured at clinic, ambulatory and home (34 in independent groups, BR and PL).	250ml BR or PL. 6.4 mmol. Once per day for 4 weeks.	Figures not given.	↓SBP 24h ambulatory and home.	Improved endothelial function. Measured via FMD. PWV not altered.	Yes
Gilchrist et al. (2014)*	27 individuals with T2DM. Protocol includes a battery of 5 cognitive function tests.	250 ml (7.5 mmol) vs. PL (0.002 mmol) daily for 2 weeks.	$NO_2^- = (IQR) 390$ and 232 nM $NO_3^- = 150$ and 31 $\mu$ M.	N/A	↓ simple reaction time	No
Zamani et al. (2015)*	17 individuals with heart failure. Supine $\dot{V}O_2$ max with measures of cardiac output and skeletal muscle oxygenation.(5 day washout period)	A single dose of NO <sub>3</sub> 3 hours prior to exercise (12.9 mmol).	Data not given.	N/A	↑ ejection fractions  ↔ O <sub>2</sub> cost of exercise  ↓ arterial wave reflections  Tendency to improve mitochondrial efficiency.	

Gilchrist et al. (2013)*	27 (18 males). T2DM subjects. A four week washout period was utilised. 24h ambulatory BP was taken. Macro & Microvascular endothelial fiction measures were assed	250ml of BR juice (NO <sub>3</sub> <sup>-</sup> =7.5 mmol) or PL (NO <sub>3</sub> <sup>-</sup> =0.002 mmol) a day for 14 days. BR & PL were consumed with their evening meal to prevent hyperglycaemic events.	$NO_2^{-} = 537$ and 390 nM $NO_3^{-} = 150$ and 31 $\mu$ M.	↔SBP ↔DBP ↔MAP		No
Shepherd et al. (2015)* Experimental chapter 2.	13 individuals with COPD. 80% GET. BP and 6MWT	70 ml of 6.77 mmol twice daily for 2 days. Plus 70 ml ~3h prior to exercise.	NO <sub>3</sub> <sup>-</sup> 215 and 48 μM	↔ SBP ↔ DBP	↔ O <sub>2</sub> cost ↔ 6MWT	No
Berry et al. (2014)*	15 individuals with COPD. Sub-maximal 75% max work rate.	140ml BR juice (NO <sub>3</sub> = 7.58 mmol) vs. PL (prune juice) 2.5h prior to exercise.	NO <sub>2</sub> increased by 379% accurate values not presented. 375nM and 175 nM (estimates from fig)	↓ SBP ↔ DBP	↑ exercise isotime  ↔ sub-max VO₂  ↔ dyspnoea  ↔ dynamic hyperinflation	Yes
Kerley et al. (2015)*	11 individuals with COPD. Incremental shuttle walk test and BP	140 ml (12.9 mmol) 3h prior to test plus blackcurrant juice. Vs PL = water and blackcurrant juice	$NO_2^- = 751$ and 139 nM $NO_3^- = 508$ and 56 $\mu$ M.	↓ SBP ↓ DBP ↓ MAP	† Incremental shuttle walk test	Yes

<sup>↑ =</sup> statistical significant increase;  $\leftrightarrow$  = no statistical reduction; ↓ = statistical significant reduction;  $\dot{V}O_{2peak}$  = peak oxygen uptake;  $\dot{V}O_{2peak}$  = peak oxygen;  $\dot{V}O_{2peak}$  =

#### General issues in the NO<sub>3</sub> literature

A major limitation of the majority of the inorganic NO<sub>3</sub><sup>-</sup> supplementation literature (*n*=53) regardless of the outcome measure is that they have small sample sizes (mean of 14.2 ± 9.4, with a range of 7 - 68 as of 01/12/14). Moreover, Kapil et al. (2015) who had the largest n (68) used a parallel group design. The largest crossover design (apart from experimental chapter 1; n = 48) examined blood pressure changes following NO<sub>3</sub><sup>-</sup> in older adults. The cohort size was 38 (Bondonno et al., 2014b). Reporting of power calculations and confidence intervals are rare. This makes determination of effect sizes difficult. The lack of power in assessing BP changes following NO<sub>3</sub><sup>-</sup> supplementation has been somewhat explored by meta-analysis by Siervo et al. (2013). His findings showed an association for a significant reduction in systolic blood pressure of 4.4 mmHg. However, larger sample sizes would elucidate more definitive conclusions.

Assay variation is still an issue within the NO<sub>3</sub> research community. Kapil et al., (2011) was the first study to report plasma NO<sub>2</sub> concentration in the μM range in an unsupplemented or placebo condition. Since the beginning of this body of work two further studies have reported plasma NO<sub>2</sub> concentration in the μM range (Bondonno et al., 2014b, Masschelein et al., 2012). The values reported in these studies (Bondonno et al., 2014b, Masschelein et al., 2012, Bescos et al., 2011) are unlikely to be accurate given that dietary intake accounts for ~1 - 2 mmol of NO<sub>3</sub> per day (ECETOC., 1988). Moreover, these studies gave a much lower dose of NO<sub>3</sub> than Webb et al. (2008a) Wylie et al. (2013b) Kapil et al. (2011) who gave the largest doses of NO<sub>3</sub> (~20 mmol) to date and did not reach the μM range post NO<sub>3</sub> rich beetroot juice supplementation.

Nitrate supplementation and effects on plasma NO<sub>2</sub>- concentration

Baseline plasma NO<sub>2</sub> concentration varied markedly between cohorts in this body of work. The healthy young individuals had a mean baseline plasma NO<sub>2</sub> concentration of 218 nM while the healthy older adults had a mean of 171 nM. These concentrations rose following NO<sub>3</sub> rich beetroot juice to 715 and 968 nM, respectively. These concentrations are within the expected range given the peer reviewed literature (Kelly et al., 2013a, Sandbakk et al., 2014, Muggeridge et al., 2013b). Moreover, the pharmacokinetic responses and peak plasma NO<sub>2</sub><sup>-</sup> concentration were similar to a profile from a cohort of young healthy individuals previously published (Wylie et al., 2013a). Interestingly, the baseline plasma NO<sub>2</sub><sup>-</sup> concentrations of individuals with T2DM were significantly elevated (see figure 32). A detailed discussion of the possible ramifications and explanations of an elevated baseline are explored in the discussion of experimental chapter 1. However, previous reports reviewed by Vanhoutte (2009) suggest that NO concentration is reduced in individuals with T2DM. This is brought about via a reduced production of NO· via eNOS (Vanhoutte, 2009). The reduction in eNOS activity is likely due to endopenetration by macrophages, cellular growth and atherosclerosis (Voetsch et al., 2004, Li et al., 2002). If eNOS activity is reduced, explanations for the high concentrations of plasma NO<sub>2</sub> found in experimental chapter 1 consist of; an habitual up-regulation of iNOS activity in this population (Krause et al., 2012), the elevated oxidative stress prevalent in individuals with T2DM (Stadler, 2013) which would be expected to result in an increase in the scavenging of NO, leading to higher NO<sub>2</sub> concentrations or medications which increase NOS activity (Davis et al., 2006, Ceconi et al., 2007, Andrade et al., 2013).

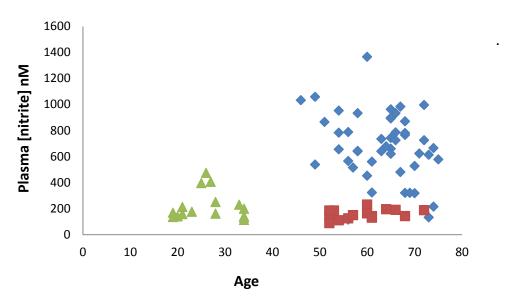


Figure 32. Baseline plasma NO<sub>2</sub> concentration for young adults, older adults and individuals with T2DM.

Shows the baseline plasma  $NO_2^-$  concentration for all samples collected from this body of work for young adults (green triangles), older adults (red squares) and individuals with T2DM (blue diamonds).

# NO<sub>3</sub> supplementation and its effect on exercise in individuals with T2DM.

The O<sub>2</sub> requirement of exercise is a key determinant of an individual's ability to tolerate exercise (Whipp et al., 1981). By reducing the O<sub>2</sub> cost of exercise the individual will be further away from the maximal O<sub>2</sub> uptake and thus, will feel less physiological stress and will fatigue slower (Regensteiner et al., 1998, Regensteiner et al., 1995a). The aetiology of fatigue in individuals with T2DM is predominantly due to deficiencies in O<sub>2</sub> delivery (Regensteiner et al., 1998, Regensteiner et al., 1995b, Scognamiglio et al., 1998). These deficiencies in delivery of O<sub>2</sub> include; transportation from the lungs (Scognamiglio et al., 1998), reduced stroke volume (Gusso et al., 2008), cardiac autonomic function (Gerritsen et al., 2001) and reduced O<sub>2</sub> extraction in the mitochondria (Baldi et al., 2002; MÅrin et al., 1994; O. Mathieu-Costello et al., 2005; Padilla et al., 2007; J. Regensteiner, G et al., 1998; J. Regensteiner, G et al., 1995).

Reducing the  $O_2$  cost of exercise or improving the kinetics of  $O_2$  uptake in any clinical population would likely have important implications. By reducing the  $O_2$  cost of exercise it may be possible to improve the quality of life in these patients. An individual's  $\dot{V}O_2$  kinetics and the  $O_2$  deficit are key determinants to the individual's ability to tolerate exercise (Jones and Poole, 2005). For example, by speeding the uptake of  $O_2$  so the individual attains a steady state  $\dot{V}O_2$ , the greater the contribution of a oxidative metabolism compared to an anaerobic contribution (Jones and Poole, 2005). By using oxidative metabolism compared to an anaerobic metabolism the speed that lactic acid concentrations build up is slowed (Bruton et al., 1998, Pate et al., 1995, Westerblad et al., 1997, Wiseman et al., 1996).

We reported no difference between the active and placebo conditions for the  $\dot{V}O_2$  responses (kinetics,  $O_2$  deficit or the  $O_2$  cost of exercise) to moderate intensity walking exercise in experimental chapter 1. This is in contrast to much of the literature which has described a reduction in steady state  $O_2$  cost of moderate intensity exercise (Bailey et al., 2009, Vanhatalo et al., 2010, Lansley et al., 2010). No reduction in the  $O_2$  cost of exercise has been reported in any clinical population to date (Berry et al., 2014, Zamani et al., 2015) (including experimental chapters 1 and 2) whilst other studies examining the effect of  $NO_3^-$  supplementation in clinical populations have not looked at the  $O_2$  cost of exercise and have focused on exercise capacity in terms of walking performance (Kenjale et al., 2011, Kerley et al., 2015). Nitrate supplementation has also been shown to improve cardiac output and stroke volume in heart failure patients (Zamani et al., 2015) but not reduce the  $O_2$  cost of exercise.

There was no difference between the active and placebo juice conditions for the distance covered in the 6MWT. Walking performance, however, has been improved following NO<sub>3</sub><sup>-</sup> supplementation in other clinical populations such as peripheral artery disease and COPD (Kenjale et al., 2011, Kerley et al., 2015, Berry et al., 2014). Potential reasons for the lack of effect in individuals with T2DM include: an elevated baseline plasma NO<sub>2</sub><sup>-</sup> concentrations, reduced NO· bioavailability due to higher levels of oxidative stress or pathological consumption of NO<sub>2</sub><sup>-</sup> during exercise and medications.

This cohort was split for drug classification (ACEi/ARB, metformin, insulin and statins; other drug sample sizes too small), no effect on plasma NO<sub>2</sub><sup>-</sup> concentration, walking performance, O<sub>2</sub> cost of exercise or blood pressure were found. It is therefore unlikely that medication is having an effect on these outcomes (note: we were not powered to find these effects). Unfortunately, in experimental chapter 1, plasma samples were only taken prior to exercise. A detailed pharmacokinetic study of NO<sub>2</sub><sup>-</sup> concentration levels during exercise (Rassaf et al., 2010) may elucidate the speed of change plasma NO<sub>2</sub><sup>-</sup> in this cohort. Diminished NO- bioavailability due to oxidative stress maybe the reason for the lack of effects following NO<sub>3</sub><sup>-</sup> supplementation in individuals with T2DM. Restoring balance to the redox state in individuals with T2DM (prior to nitrate supplementation) may prolong the half-life and thus, bioavailability of NO·.

Nitrate supplementation in individuals with COPD and A critical appraisal of the literature.

Recently Berry et al. (2014) and Kerley et al. (2015) examined the effect of NO<sub>3</sub> rich beetroot juice supplementation vs. prune juice or water (respectively: as a placebo) in individuals with COPD. In experimental chapter 2 (NO<sub>3</sub> and COPD) logistical

restraints prevented the measurement of plasma NO<sub>2</sub> concentration. Berry et al. (2014) and Kerley et al. (2015) reported significantly elevated plasma NO<sub>3</sub> and NO<sub>2</sub> concentrations compared with placebo. This indicates that the entero-salivary pathway is operational in people with COPD.

Berry et al., reported reductions in resting systolic BP, iso-time BP, end exercise diastolic BP. Whilst Kerley et al., report reductions in systolic, diastolic blood pressure and MAP following NO<sub>3</sub><sup>-</sup> supplementation compared to placebo. Following placebo (water) Kerley et al., showed a statistically significant increase in systolic (3 mmHg) diastolic (9 mmHg) blood pressure and MAP (7 mmHg). Berry et al., reported an improvement in exercise tolerance (i.e. lengthened time to exhaustion during submaximal constant rate).

Kerley et al., report improved distance covered in an incremental shuttle walk test following NO<sub>3</sub><sup>-</sup> rich beetroot juice compared to placebo. The placebo condition significantly decreased walking performance by 7.6% from baseline. The significant change in outcome measures (blood pressure and exercise performance) following placebo in the Kerley et al cohort is difficult to explain given the 'double-blind' nature of the trial. The O<sub>2</sub> cost of exercise was not assessed in this study. Berry et al., do not show a reduction in the O<sub>2</sub> cost of exercise which is consistent with experimental chapter 2. However, with no reduction in the O<sub>2</sub> cost of exercise, it is not clear what mechanism underpins the improved time to exhaustion reported by Berry et al. (2014) and Kerley et al. (2015).

A potential reason for the improvement seen in walking performance that does not reflect a reduction in the O<sub>2</sub> cost of exercise maybe an increase in the delivery of O<sub>2</sub> to the working muscle (Ferguson et al., 2014, Ferguson et al., 2013). This would

mask any reduction in the  $O_2$  cost of exercise and help to improve exercise performance. However, given the lack of evidence in the experimental chapters 1 and 2, we were either underpowered to detect this or this is not the mechanism for the improvement in walking performance.

Although these studies in the literature highlight the potential for NO<sub>3</sub> supplementation to be beneficial to individuals with COPD in terms of blood pressure reduction and improved walking performance these studies have major limitations. Of paramount importance with both studies is a lack of a true placebo as used in experimental chapter 2. Given the substantial media attention given in the scientific literature, written press and on the television in recent years describing the beneficial effects of beetroot juice for exercise performance/economy and blood pressure it is impossible to rule out a placebo effect for knowledgeable and enthused volunteers. In informal discussions with the cohort of volunteers in the present trials, many of the participants in this body of work were aware of the putative beneficial effects of beetroot prior to receiving the participant information sheets. An example published on the 29/06/2010 in the daily mail online (DailyMail, 2013) had a headline which read: beetroot juice 'could save your life'. Secondly, the antioxidant content of prune juice and water are likely to be significantly different to beetroot juice (Donovan et al., 1998). This change in antioxidant content may affect NO bioavailability (Bondonno et al., 2012) due to reduced scavenging of NO. This may increase the half-life on NO· in diseased populations. Berry et al., matched the calorie content of the different juice supplementation to control for the performance benefits of carbohydrate availability. Kerley et al., did not match this and had a 120 kcal difference between the conditions. However, this is unlikely to be the reason for the improvement given

evidence from Vermeeren et al. (2001) who demonstrated a 250 kcal supplement did not improve submaximal cycling performance in individuals with COPD.

Berry et al., used an iso-time to established the  $O_2$  cost of exercise. This method takes an average of the last minute of  $O_2$  consumption of the shorter exercise period and compares to the longer exercise period. These averages were then compared. A methodological weakness exists in Berry et al's assessment of the  $O_2$  cost of exercise. Using one minute of data is not a robust measure of the  $O_2$  cost of exercise. Towards the end of an exercise session coughing and spluttering as you fatigue is common (Lamarra et al., 1987). Typically, 6 minutes of steady state  $\dot{V}O_2$  is used with multiple transitions to improve repeatability of the measure. This cohort of individuals have severe respiratory dysfunction as described in the introduction. This will increase the variability of data and given the short period of assessment this will magnify the variability. Given this, it is unlikely that this method of assessment of iso-time assessment of  $O_2$  uptake is appropriate in this cohort of participants.

While the findings from Berry and Kerley were of interest, they were not replicated in a truly double blind trial and, in part, do not utilise the most robust measurement for O<sub>2</sub> uptake dynamics.

Although many mechanisms were explored for the lack of effect on the  $O_2$  cost of exercise in the discussion of experimental chapter 2, a further possible explanation exists. In individuals with COPD, increasing the bioavailability of NO· (NO· gas in this study) may increase venous return and caused elevated pulmonary artery pressure in heart failure patients (Ichinose et al., 2004). By elevating pulmonary artery pressure, gas exchange may be slowed which may prevent the reduction in the  $O_2$  cost of exercise that was hypothesised. However, the inorganic molecule GTN (in

dogs), has been shown to reduce pulmonary artery hypertension (Kim et al., 2010). More research is needed in this area to establish whether increasing NO-bioavailability increases or decreases pulmonary artery hypertension. Conversely, it may be due to the differing effects of the pharmacokinetic and dynamic responses of inorganic vs. organic NO<sub>3</sub><sup>-</sup>/NO<sub>2</sub><sup>-</sup> and not the bioavailability of NO· (Omar et al., 2012).

Nitrate supplementation and blood pressure in other clinical populations.

The gold standard measurement of blood pressure is 24h ambulatory blood pressure monitoring. For ease of measurement repeated rested brachial artery BP was used for all experimental chapters. This method can be susceptible to the white coat syndrome (Pickering, 1996). Given the timing of blood pressure measurement (described in the experimental chapters) it coincided with peak plasma NO<sub>2</sub><sup>-</sup> (Wylie et al., 2013a) which would give the optimal chance for observing any blood pressure reductions.

Recently, the results from a 24h ambulatory blood pressure monitoring protocol following NO<sub>3</sub><sup>-</sup> supplementation or placebo were reported in a group of individuals with T2DM. The authors reported no reductions in blood pressure (Gilchrist et al., 2013). The mean change in systolic blood pressure was 0.5 mmHg. This is similar to the effect shown in systolic blood pressure in experimental chapter 1 (-2 mmHg). A recently reported phase II trial in hypertensive adults following NO<sub>3</sub><sup>-</sup> supplementation was published. The trial by Kapil et al. (2015) is the largest NO<sub>3</sub><sup>-</sup> supplementation study, in any population to date (apart from experimental chapter 1). Clinic BP, home BP and 24h ambulatory blood pressure monitoring was employed. They showed in a phase 2, randomised, parallel group design that 4 weeks of NO<sub>3</sub><sup>-</sup> supplementation causes a reduction in diastolic (2.4 – 3.8 mmHg depending on method) and systolic

blood pressure (7.7 - 8.1 mmHg depending on method). Following withdrawal of  $NO_3^-$ , BP returned to baseline levels. One study has looked at the effect of  $NO_3^-$  supplementation in 8 individuals with peripheral artery disease. Kenjale et al. (2011) reported reductions in diastolic blood pressure at rest and during exercise. No reductions in systolic blood pressure or MAP were present.

Nitrate supplementation and blood pressure in healthy individuals.

Varying reports of the blood pressure lowering effects of NO<sub>3</sub><sup>-</sup> supplementation have been reported in young healthy individuals. In experimental chapter 3 we report no reductions in systolic or diastolic blood pressure following acute supplementation of NO<sub>3</sub><sup>-</sup> rich beetroot juice in young healthy individuals. This is similar to some of the studies (Wilkerson et al., 2012, Coles and Clifton, 2012, Sandbakk et al., 2014) whilst most have reported reductions (Webb et al., 2008a, Vanhatalo et al., 2010, Vanhatalo et al., 2011, Lansley et al., 2010, Lansley et al., 2011a, Engan et al., 2012, Wylie et al., 2013b, Thompson et al., 2014, Muggeridge et al., 2014, Keen et al., 2014, Levitt et al., 2015, Treichler et al., 2015, Kocoloski and Crecelius, 2015, Bond Jr et al., 2013, Bond et al., 2014). There does not appear to be obvious differences in age, BMI, disease status, ethnicity or physical activity level between the studies who found a reduction and those that did not.

In experimental chapter 3 systolic and diastolic blood pressure were assessed as a secondary outcome measure. These data confirms the findings described by Bondonno et al. (2014b) who found no effect of NO<sub>3</sub><sup>-</sup> supplementation on systolic blood pressure. Kelly et al. (2013a), Jajja et al. (2014) and two other studies have shown reductions in systolic blood pressure in an healthy older adult population however, only abstracts are available online (Miller et al., 2012, Treichler et al., 2015). However, these studies are the only other studies to examine the effects of

NO<sub>3</sub> in an older adult cohort. Kelly et al., and the data from experimental chapter 3 report a 5 mmHg drop whilst Jajja et al. (2014) report a 7 mmHg drop in systolic blood pressure compared to baseline. It should be noted that Jajja et al., did not use a true placebo and did not find reduction using 24 hour blood pressure monitoring or clinical blood pressure assessment. Bondonno et al., cohort had a mean age of ~60 but there range was from 38-70 years old. Moreover some participants were on antihypertensives. A discussion on drug therapy and its effects on NO<sub>3</sub><sup>-</sup> supplementation (and study outcomes) can be found in the discussion sections of experimental chapters 1 and 2. Bondonno et al., also used (predominantly) green leafy vegetables to increase the NO<sub>3</sub> dose in each individual's diet. Although it is relatively easy to estimate the NO<sub>3</sub> content of foods, it is much harder to standardise the dose. NO<sub>3</sub> content of different crops could have different concentrations depending on growth rate (Rodgers and Barneix, 1988) although this is not always the case (Mattsson et al., 1991). Kelly et al., report a statistically significant 3 mmHg drop in diastolic blood pressure whilst we report a non-significant 2 mmHg drop. Jajja et al. (2014) report no change in diastolic blood pressure. A larger trial examining the effect in older adults is required to examine the efficacy of NO<sub>3</sub> supplementation in normotensive older adults. If NO<sub>3</sub> lowers blood pressure it may prevent/delay the onset of hypertension.

Nitrate supplementation, portal vein flux, velocity and hepatic diffusion and glucose concentration

Camilleri et al. (1985) described the uptake of solids and liquid foods from the stomach (see figure 33). At 1 hour the food has started to be absorbed with ~ 60% of the fluids taken up and 20% of the solids. At 2 hours 65% of the solids have been taken up and 90% of the fluids. At 3 hours both the fluids and solids have been

absorbed. Thus it is likely that hepatic blood flow and importantly, peak glucose concentration would occur at 2 hours.

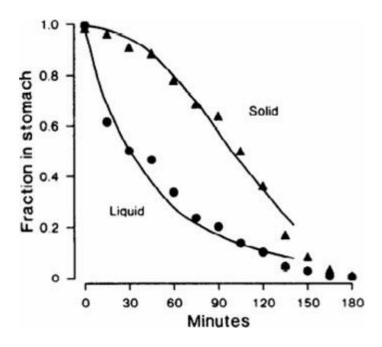


Figure 33. Uptake of liquids and solids from the stomach.

This figure is from Camilleri et al. (1985) with the appropriate rights from the publishers.

As glucose and other micro nutrients are absorbed through the stomach and proximal small intestine, nitrite is simultaneously absorbed. Peak plasma nitrite concentrations occur at 2 – 2.5 hours (Wylie et al., 2013a, Webb et al., 2008a). The nitrite along with the nutrients will meet within the portal vein from tributaries of the stomach, proximal small intestine and the spleen. Blood from the portal vein, having already been through vascular beds will be deoxygenated. Deoxyhaemoglobin is a nitrite reductase (Cosby et al., 2003). Therefore, NO· bioavailability would be expected to increase and thus vasodilate the portal vein leading to an increase in flux.

The liver has many nitrite reductases. Neuroglobin and cytoglobin are in small concentrations and therefore may not have a meaningful effect (Reeder, 2010, Omar and Webb, 2014). However, xanthine oxide and aldehyde oxidase are in much higher concentrations (Martin et al., 2004, Li et al., 2008b). Increase in the bioavailability of NO· has the potential to have significant biological effects within the liver. NO has previously been shown to mediate the uptake of glucose into skeletal muscle (Merry et al., 2010) and mediate its uptake from the intestines via incretins (Guan et al., 2003) into the circulation. Moreover, a study by Carlstrom et al., showed that in eNOS deficient mice that 10 weeks of nitrite supplementation in their drinking water, redcued HbA1c, baseline glucose concentrations and post prandial glucose concentrations (Carlström et al., 2010). In healthy humans and in individuals with T2DM, a NOS inhibitor has shown a reduction in the uptake of glcusoe within the exercising skeletal muscle (Kingwell et al., 2002). It is therefore possible or even probable (given the elevated NO and glucose concentrations) that it will help mediate the uptake of glucose in to hepatic tissue along with skeletal muscle. In vitro work on liver cells may help to elucidate this mechanism. GLP-1 and incretins in general appear to be interlinked in many biological processes such as increasing glucose utilisation (in an NO dependant mechanism) (Chai et al., 2012) and production of NO· in the portal vein (Ding, Zhong, Xu, & Isales, 2004). By elevating the NO· bioavailability we may increase GLP-1 and vice versa (Post et al., 2010) and thus facilitate uptake of glucose (Bhashyam et al., 2010) into cells leading to lower blood glucose concentrations.

Experimental chapter 3 shows that dietary NO<sub>3</sub><sup>-</sup> supplementation did not alter portal vein flux, velocity or hepatic diffusion. The hepatic arterial buffer response described by Hwang (2011) suggests that the hepatic artery can vasodilate in response to the

portal vein. Therefore if the portal vein changes in flow the hepatic artery responds with constriction of relaxation (Hwang, 2011, Lautt et al., 1990). The buffer response can only compensate for 25-60% of alterations in blood flow from the portal vein (Lautt, 1980, Lautt, 1977). The primary reason behind this mechanism is to maintain a constant perfusion rate from organs such as the stomach, spleen and intestines (Lautt, 1980, Lautt, 1977). Therefore dietary NO<sub>3</sub><sup>-</sup> has the capacity to affect up to 75% of flow to the liver. This may be a potential reason why we saw no effect in this study.

Methodological issues / limitations with experimental chapters.

The link between NO·, glucose uptake, insulin and incretin release has been discussed in depth with in experimental chapter 3 and the general discussion. Samples for incretin and insulin quantification for experimental chapter 3 have been stored. Unfortunately, we did not secure the funding to analyse these samples and this should be recognised as a limitation of the study and thesis in general.

### Hepatic ADC analysis

From repeatability analysis involving multiple regions of interest it appears that to gain an accurate and repeatable ADC, the region of interest needs to be away from large vessels such as the right branch of the hepatic vein and the anteroinferior branch of the portal vein. The closer the ROI to the vessel the more unpredictable the ADC becomes. Potentially due to a higher variability of flow in larger blood vessels compared with a constant flow in the liver parenchyma.

The area in red and its close surrounding allow for reproducibility. The closer you get to the black box the more variable the data becomes.

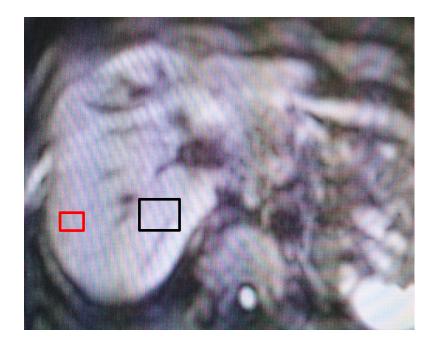


Figure 34. Depicts a slice of a liver from a representative participant. The highlighted boxes shows the regions of interest with the best and worst repeatability.

#### **Future work**

The results from this body of work have helped to answer many questions and has raised others. Notably, the pharmacokinetic response of older adults, the potential for blood pressure reductions in older adults, antioxidants in individuals with T2DM and critically the effect of dietary NO<sub>3</sub> on plasma glucose concentration.

#### Antioxidants in clinical populations.

Previously it has been demonstrated that the beneficial effects of beetroot juice can be predominantly explained by its large NO<sub>3</sub><sup>-</sup> content (Lansley et al., 2010). It has been suggested that polyphenols (in vitro) may have a beneficial antioxidant effects (Kanner et al., 2001). Given the pro-oxidant state of many clinical populations a supplement that could help to re-balance the redox state could help to prolong the NO· half-life. If this is possible then the hypothesis in experimental chapter one with improved efficiency in individuals with T2DM could be revisited. Thus enabling individuals with T2DM to gain from the mitochondrial adaptations that healthy young adults appear to benefit from.

Pharmacokinetic response in older adults and a phase II trial.

In experimental chapter 3 the data show that plasma NO<sub>2</sub> concentrations are continuing to rise after 2 hours and peak at 3. This suggests that plasma NO<sub>2</sub> concentration may rise beyond 3 hours which has only been shown once before (Kapil et al., 2011) with a peak at 6 hours. Future work should try to elucidate the precise pharmacokinetic response to dietary NO<sub>3</sub>. In experimental chapter 3, systolic blood pressure was reduced. This coincided with peak plasma NO<sub>2</sub> concentration. If the conversion of NO<sub>3</sub> to NO<sub>2</sub> continues past 3 hours this hypotensive effect may continue.

Three small studies to date have consistently shown a reduction in blood pressure of normotensive, drug naive older adults. A larger phase II trial in normotensive adults would help to characterise the blood pressure reductions shown in this work and by Kelly et al. (2013a) and Jajja et al. (2014). Dietary NO<sub>3</sub> supplementation could slow the onset of hypertension in healthy older adults. The potential to save the NHS money on treatment of hypertension and the associated diseases caused via hypertension is vast. A health economist should also be used to assess any potential savings.

#### Plasma glucose concentration

The potential to reduce the plasma glucose concentration following a NO<sub>3</sub><sup>-</sup> rich meal compared to baseline has been shown in experimental chapter 3. Nitrate-rich beetroot juice reduced plasma glucose concentration levels where placebo did not. However, this experiment did not show a reduction at two hours post supplementation and a standardised meal between conditions. Although the data are not unequivocal it does raise the possibility of a reduction in postprandial glucose concentrations and should be assessed again in larger numbers of healthy older

adults. This could help to prevent delay the onset of insulin resistance and even T2DM. If the findings are replicated then a trial in individuals with T2DM would be of interest. The potential to reduce plasma glucose concentrations in this cohort could help to manage the disease and has the potential to save the NHS vast amounts of money in treatment costs. Although this would require a significant increase in the nationwide uptake of NO<sub>3</sub><sup>-</sup> supplementation. A phase II trial would be the next step in this process. This may be easier to achieve via pharmacological means i.e. sodium or potassium NO<sub>3</sub><sup>-</sup> than NO<sub>3</sub><sup>-</sup> rich beetroot juice or increasing the intake of NO<sub>3</sub><sup>-</sup> rich vegetables.

# Summary and conclusion of findings in this body of work

This is the first body of work to assess the effect of dietary NO<sub>3</sub> supplementation on the O<sub>2</sub> cost of exercise in individuals with T2DM and COPD with a true placebo. Supplementation of the diet with dietary NO<sub>3</sub> does not reduce the O<sub>2</sub> cost of walking (in individuals with T2DM) or cycling (in individuals with COPD), improve distance covered in the 6MWT or reduce BP in individuals with T2DM or COPD. This body of work is the first to assess the effect NO<sub>3</sub> rich beetroot juice on hepatic blood flow. Dietary NO<sub>3</sub> does not affect liver diffusion, portal vein flux or velocity, reduce systolic, diastolic BP or plasma glucose concentrations in healthy adults. By modifying an individual's diet with NO<sub>3</sub> supplementation it is unlikely to reduce the O<sub>2</sub> cost of exercise, improve walking performance or reduce BP. Potential reasons for this are; an elevated baseline plasma NO<sub>2</sub> concentration in the placebo condition, and/or reductions in the bioavailability of NO· in individuals with T2DM. In individuals with COPD likely reasons for the lack of effect include; NO<sub>3</sub> dosage, oxidative stress, ageing and efficacy of NO<sub>3</sub> reduction to NO<sub>2</sub>. A hypothesis for the lack of effect in microvascular diffusion is the hepatic arterial buffer response.

Modifying an individual diet with supplementary NO<sub>3</sub> does not appear to reduce the O<sub>2</sub> cost of exercise, improve walking performance or reduce BP in individuals with T2DM or COPD. Nitrate supplementation does not affect systolic, diastolic blood pressure or plasma glucose concentrations in healthy young and older adults.

## References

ABBASI, F., ASAGMI, T., COOKE, J. P., LAMENDOLA, C., MCLAUGHLIN, T., REAVEN, G. M., STUEHLINGER, M. & TSAO, P. S. 2001. Plasma concentrations of asymmetric

- dimethylarginine are increased in patients with type 2 diabetes mellitus. *The American Journal of Cardiology,* 88, 1201-1203.
- AGLE, D. P., BAUM, G. L., CHESTER, E. H. & WENDT, M. 1973. Multidiscipline Treatment of Chronic Pulmonary Insufficiency. *Psychosomatic Medicine*, 35, 41-49.
- AHIMASTOS, A. A., WALKER, P. J., ASKEW, C. & ET AL. 2013. Effect of ramipril on walking times and quality of life among patients with peripheral artery disease and intermittent claudication: A randomized controlled trial. *JAMA*, 309, 453-460.
- ALBERTI, K. G. M. M. & ZIMMET, P. Z. 1998. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus. Provisional report of a WHO Consultation. *Diabetic Medicine*, 15, 539-553.
- ALBOUAINI, K., ANDRON, M., ALAHMAR, A. & EGRED, M. 2007. Beta-blockers use in patients with chronic obstructive pulmonary disease and concomitant cardiovascular conditions. *International journal of chronic obstructive pulmonary disease*, 2, 535-540.
- ALDERTON, W. K., COOPER, C. E. & KNOWLES, R. G. 2001. Nitric oxide synthases: structure, function and inhibition.
- ALZAWAHRA, W. F., TALUKDER, M. A., LIU, X., SAMOUILOV, A. & ZWEIER, J. L. 2008. Heme proteins mediate the conversion of nitrite to nitric oxide in the vascular wall. *Am J Physiol Heart Circ Physiol*, 295, H499-508.
- ANDRADE, V. L., SERTÓRIO, J. T. C., ELEUTERIO, N. M., TANUS-SANTOS, J. E., FERNANDES, K. S. & SANDRIM, V. C. 2013. Simvastatin treatment increases nitrite levels in obese women: Modulation by T–786C polymorphism of eNOS. *Nitric Oxide*.
- ANDREW, P. J. & MAYER, B. 1999. Enzymatic function of nitric oxide synthases. *Cardiovascular Research*, 43, 521-531.
- ANGULO, J., SANCHEZ-FERRER, C. F., PEIRO, C., MARIN, J. & RODRIGUEZ-MANAS, L. 1996. Impairment of endothelium-dependent relaxation by increasing percentages of glycosylated human hemoglobin. Possible mechanisms involved. *Hypertension*, 28, 583-92.
- ANTHONISEN, N., R, CONNETT, J., E & KILEY, J., P 1994. Effects of smoking intervention and the use of an inhaled anticholinergic bronchodilator on the rate of decline of fev1: The lung health study. *JAMA: The Journal of the American Medical Association*, 272, 1497-1505.
- ANTHONISEN, N., WRIGHT, E. & HODGKIN, J. 1986. Prognosis in chronic obstructive pulmonary disease. *The American review of respiratory disease*, 133, 14-20.
- ANTONIADES, C., BAKOGIANNIS, C., LEESON, P., GUZIK, T. J., ZHANG, M.-H., TOUSOULIS, D., ANTONOPOULOS, A. S., DEMOSTHENOUS, M., MARINOU, K. & HALE, A. 2011. Rapid, direct effects of statin treatment on arterial redox state and nitric oxide bioavailability in human atherosclerosis via tetrahydrobiopterin-mediated endothelial nitric oxide synthase coupling. *Circulation*, 124, 335-345.
- ANTONOPOULOS, A. S., MARGARITIS, M., LEE, R., CHANNON, K. & ANTONIADES, C. 2012. Statins as anti-inflammatory agents in atherogenesis: molecular mechanisms and lessons from the recent clinical trials. *Current Pharmaceutical Design*, 18, 1519.
- APPEL, L., J, MOORE, T., J, OBARZENEK, E., VOLLMER, W., M, SVETKEY, L., P, SACKS, F., M, BRAY, G., A, VOGT, T., M, CUTLER, J., A, WINDHAUSER, M., M, LIN, P. & KARANJA, N. 1997. A clinical trial of the effects of dietary patterns on blood pressure. *The New England Journal of Medicine*, 336, 1117-1124.
- ARNOLD, W. P., MITTAL, C. K., KATSUKI, S. & MURAD, F. 1977. Nitric oxide activates guanylate cyclase and increases guanosine 3': 5'-cyclic monophosphate levels in various tissue preparations. *Proceedings of the National Academy of Sciences*, 74, 3203-3207.
- ASHMORE, T., FERNANDEZ, B. O., BRANCO-PRICE, C., WEST, J. A., COWBURN, A. S., HEATHER, L. C., GRIFFIN, J. L., JOHNSON, R. S., FEELISCH, M. & MURRAY, A. J. 2014. Dietary nitrate increases arginine availability and protects mitochondrial complex I and energetics in the hypoxic rat heart. *The Journal of physiology*, 592, 4715-4731.

- ASSMANN, G. & SCHULTE, H. 1991. Triglycerides and atherosclerosis; results from the Prospective Cardiovascular Munster Study. *Arterioscler Rev,* 22, 51-57.
- AVERY, A. A. 1999. Infantile methemoglobinemia: Reexamining the role of drinking water nitrates. *Environmental Health Perspectives*, 107, 583-586.
- AWATA, T., NEDA, T., IIZUKA, H., KURIHARA, S., OHKUBO, T., TAKATA, N., OSAKI, M., WATANABE, M., NAKASHIMA, Y., SAWA, T., INUKAI, K., INOUE, I., SHIBUYA, M., MORI, K., YONEYA, S. & KATAYAMA, S. 2004. Endothelial Nitric Oxide Synthase Gene Is Associated With Diabetic Macular Edema in Type 2 Diabetes. *Diabetes Care*, 27, 2184-2190.
- BAGGIO, L. L. & DRUCKER, D. J. 2007. Biology of incretins: GLP-1 and GIP. *Gastroenterology*, 132, 2131-57.
- BAILEY, D. M., DEHNERT, C., LUKS, A. M., MENOLD, E., CASTELL, C., SCHENDLER, G., FAORO, V., GUTOWSKI, M., EVANS, K. A., TAUDORF, S., JAMES, P. E., MCENENY, J., YOUNG, I. S., SWENSON, E. R., MAIRBÄURL, H., BÄRTSCH, P. & BERGER, M. M. 2010a. High-altitude pulmonary hypertension is associated with a free radical-mediated reduction in pulmonary nitric oxide bioavailability. *The Journal of Physiology*, 588, 4837-4847.
- BAILEY, S., J , FULFORD, J., VANHATALO, A., WINYARD, P., BLACKWELL, J., R, DIMENNA, F., J, WILKERSON, D., P, BENJAMIN, N. & JONES, A., M 2010b. Dietary nitrate supplementation enhances muscle contractile efficiency during knee-extensor exercise in humans. *Journal of Applied Physiology*, 109, 135-148.
- BAILEY, S., J, WINYARD, P., VANHATALO, A., BLACKWELL, J., R, DIMENNA, F., J, WILKERSON, D., P, TARR, J., BENJAMIN, N. & JONES, A., M 2009. Dietary nitrate supplementation reduces the O<sub>2</sub> cost of low-intensity exercise and enhances tolerance to high-intensity exercise in humans. *Journal of Applied Physiology*, 107, 1144-1155.
- BAILEY, S. J., VANHATALO, A., WINYARD, P. G. & JONES, A. M. 2012. The nitrate-nitrite-nitric oxide pathway: Its role in human exercise physiology. *European Journal of Sport Science*, 12, 309-320.
- BAILEY, S. J., WINYARD, P. G., BLACKWELL, J. R., VANHATALO, A., LANSLEY, K. E., DIMENNA, F. J., WILKERSON, D. P., CAMPBELL, I. T. & JONES, A. M. 2011. Influence of N-acetylcysteine administration on pulmonary O2 uptake kinetics and exercise tolerance in humans. *Respiratory physiology & neurobiology*, 175, 121-129.
- BALDI, J., C, AOINA, J., L, OXENHAM, H., C, BAGG, W. & DOUGHTY, R., N 2002. Reduces exercise arteriovenous O<sub>2</sub> differences in Type 2 diabetes. *Journal of Applied Physiology*, 94, 1033-1038
- BARBERÀ, J. A. & BLANCO, I. 2009. Pulmonary Hypertension in Patients with Chronic Obstructive Pulmonary Disease: Advances in Pathophysiology and Management. *Drugs*, 69, 1153-1171 10.2165/00003495-200969090-00002.
- BARBERÀ, J. A., PEINADO, V. I. & SANTOS, S. 2003. Pulmonary hypertension in chronic obstructive pulmonary disease. *European Respiratory Journal*, 21, 892-905.
- BARBERÀ, J. A., ROGER, N., ROCA, J., RODRIGUEZ-ROISIN, R., ROVIRA, I. & HIGENBOTTAM, T. W. 1996. Worsening of pulmonary gas exchange with nitric oxide inhalation in chronic obstructive pulmonary disease. *The Lancet*, 347, 436-440.
- BARNES, P. 1995. Bronchodilators: basic pharmacology. *Chronic obstructive pulmonary disease*. Springer.
- BARNES, P. J. 2003. Chronic obstructive pulmonary disease \* 12: New treatments for COPD. *Thorax,* 58, 803-808.
- BARRAGÁN, J. M., RODRÍGUEZ, R. E., ENG, J. & BLÁZQUEZ, E. 1996. Interactions of exendin-(9–39) with the effects of glucagon-like peptide-1-(7–36) amide and of exendin-4 on arterial blood pressure and heart rate in rats. *Regulatory Peptides*, 67, 63-68.
- BARTHOLOMEW, B. & HILL, M. 1984. The pharmacology of dietary nitrate and the origin of urinary nitrate. *Food and Chemical Toxicology*, 22, 789-795.

- BARTOLI, E., FRA, G. & SCHIANCA, G. C. 2011a. The oral glucose tolerance test (OGTT) revisited. *European journal of internal medicine*, 22, 8-12.
- BARTOLI, E., FRA, G. P. & SCHIANCA, G. P. C. 2011b. The oral glucose tolerance test (OGTT) revisited. *European Journal of Internal Medicine*, 22, 8-12.
- BASU, A., CHARKOUDIAN, N., SCHRAGE, W., RIZZA, R. A., BASU, R. & JOYNER, M. J. 2007. Beneficial effects of GLP-1 on endothelial function in humans: dampening by glyburide but not by glimepiride. *American Journal of Physiology-Endocrinology and Metabolism*, 293, E1289-E1295.
- BASU, S., AZAROVA, N. A., FONT, M. D., KING, S. B., HOGG, N., GLADWIN, M. T., SHIVA, S. & KIM-SHAPIRO, D. B. 2008. Nitrite Reductase Activity of Cytochrome c. *Journal of Biological Chemistry*, 283, 32590-32597.
- BATEMAN, R. M., ELLIS, C. G. & FREEMAN, D. J. 2002. Optimization of Nitric Oxide Chemiluminescence Operating Conditions for Measurement of Plasma Nitrite and Nitrate. *Clinical Chemistry*, 48, 570-573.
- BAUER, T. A., REUSCH, J. E. B., LEVI, M. & REGENSTEINER, J. G. 2007. Skeletal Muscle Deoxygenation After the Onset of Moderate Exercise Suggests Slowed Microvascular Blood Flow Kinetics in Type 2 Diabetes. *Diabetes Care*, 30, 2880-2885.
- BAYS, H. E. 2011. Adiposopathy: is "sick fat" a cardiovascular disease? *Journal of the American College of Cardiology*, 57, 2461-2473.
- BAZZANO, L., A, HE, J., OGDEN, L., G, LORIA, C., M, VUPPUTURI, S., MYERS, L. & WHELTON, P., K 2001. Fruit and vegetable intake and risk of cardiovascular disease in US adults: the first National Health and Nutrition Examination Survey Epidemiologic Follow-up Study<sup>1-3</sup>. *American Society for Clinical Nutrition* 76, 93-99.
- BEAVER, W. L., WASSERMAN, K. & WHIPP, B. J. 1986. A new method for detecting anaerobic threshold by gas exchange. *J Appl Physiol*, 60, 2020-2027.
- BECKMAN, J. S. & KOPPENOL, W. H. 1996. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and the ugly. *American Journal of Physiology-Cell Physiology*, 40, C1424.
- BEDARD, S., MARCOTTE, B. & MARETTE, A. 1997. Cytokines modulate glucose transport in skeletal muscle by inducing the expression of inducible nitric oxide synthase. *Biochem J*, 325, 487-93.
- BENJAMIN, N., O'DRISCOLL, F., DOUGALL, H., DUNCAN, C., SMITH, S., GOLDEN, M. & MCKENZIE, H. 1994. Stomach NO synthesis. *Nature*, 368, 502.
- BENTLEY, R., GRAY, S. R., SCHWARZBAUER, C., DAWSON, D., FRENNEAUX, M. & HE, J. 2014. Dietary nitrate reduces skeletal muscle oxygenation response to physical exercise: a quantitative muscle functional MRI study. *Physiological reports*, 2, e12089.
- BERG, A. H., COMBS, T. P., DU, X., BROWNLEE, M. & SCHERER, P. E. 2001. The adipocyte-secreted protein Acrp30 enhances hepatic insulin action. *Nature medicine*, **7**, 947-953.
- BERGER, B., STENSTRÖM, G. & SUNDKVIST, G. 1999. Incidence, prevalence, and mortality of diabetes in a large population. A report from the Skaraborg Diabetes Registry. *Diabetes Care*, 22, 773-778.
- BERRY, M. J., JUSTUS, N. W., HAUSER, J. I., CASE, A. H., HELMS, C. C., BASU, S., ROGERS, Z., LEWIS, M. T. & MILLER, G. D. 2014. Dietary nitrate supplementation improves exercise performance and decreases blood pressure in COPD patients. *Nitric Oxide*.
- BESCOS, R., RODRIGUEZ, F. A., IGLESIAS, X., FERRER, M. D., IBORRA, E. & PONS, A. 2011. Acute administration of inorganic nitrate reduces VO(2peak) in endurance athletes. *Med Sci Sports Exerc*, 43, 1979-86.
- BESCÓS, R., RODRÍGUEZ, F. A., IGLESIAS, X., FERRER, M. D., IBORRA, E. & PONS, A. 2011. Acute administration of inorganic nitrate reduces VO (2peak) in endurance athletes. *Medicine and science in sports and exercise*, 43, 1979.
- BHASHYAM, S., FIELDS, A. V., PATTERSON, B., TESTANI, J. M., CHEN, L., SHEN, Y.-T. & SHANNON, R. P. 2010. Glucagon-Like Peptide-1 Increases Myocardial Glucose Uptake via p38α MAP Kinase–

- Mediated, Nitric Oxide—Dependent Mechanisms in Conscious Dogs With Dilated Cardiomyopathy. *Circulation: Heart Failure*, 3, 512-521.
- BINZ, C. & GERINGER, P. 1901. Die Reduktion des Natrimnitrats in Tierkorper. *Archive of Internation Pharmacodynamics*, 9, 441-450.
- BJÖRNE, H., PETERSSON, J., PHILLIPSON, M., WEITZBERG, E., HOLM, L. & LUNDBERG, J. O. 2004. Nitrite in saliva increases gastric mucosal blood flow and mucus thickness. *The Journal of clinical investigation*, 113, 106-114.
- BLOOMGARDEN, Z. T. 2000. Obesity and diabetes. Diabetes care, 23, 1584-1590.
- BODEN, G. 1997. Role of fatty acids in the pathogenesis of insulin resistance and NIDDM. *Diabetes*, 46, 3-10.
- BODEN, G. & CHEN, X. 1995. Effects of fat on glucose uptake and utilization in patients with non-insulin-dependent diabetes. *Journal of Clinical Investigation*, 96, 1261.
- BOHN, T. 2014. Dietary factors affecting polyphenol bioavailability. *Nutrition Reviews*, 72, 429-452.
- BOND, H., MORTON, L. & BRAAKHUIS, A. J. 2012. Dietary nitrate supplementation improves rowing performance in well-trained rowers. *Int J Sport Nutr Exerc Metab*, 22, 251-6.
- BOND JR, V., CURRY, B. H., ADAMS, R. G., MILLIS, R. M. & HADDAD, G. E. 2013. Cardiorespiratory function associated with dietary nitrate supplementation. *Applied Physiology, Nutrition, and Metabolism,* 39, 168-172.
- BOND, V., CURRY, B. H., ADAMS, R. G., ASADI, M. S., STANCIL, K. A., MILLIS, R. M. & HADDAD, G. E. 2014. Effects of Nitrate Supplementation on Cardiovascular and Autonomic Reactivity in African-American Females. *International Scholarly Research Notices*, 2014.
- BONDONNO, C. P., LIU, A. H., CROFT, K. D., WARD, N. C., YANG, X., CONSIDINE, M. J., PUDDEY, I. B., WOODMAN, R. J. & HODGSON, J. M. 2014a. Short-term effects of nitrate-rich green leafy vegetables on blood pressure and arterial stiffness in individuals with high-normal blood pressure. *Free Radic Biol Med*.
- BONDONNO, C. P., LIU, A. H., CROFT, K. D., WARD, N. C., YANG, X., CONSIDINE, M. J., PUDDEY, I. B., WOODMAN, R. J. & HODGSON, J. M. 2014b. Short-term effects of nitrate-rich green leafy vegetables on blood pressure and arterial stiffness in individuals with high-normal blood pressure. *Free Radic Biol Med*, 77, 353-62.
- BONDONNO, C. P., YANG, X., CROFT, K. D., CONSIDINE, M. J., WARD, N. C., RICH, L., PUDDEY, I. B., SWINNY, E., MUBARAK, A. & HODGSON, J. M. 2012. Flavonoid-rich apples and nitrate-rich spinach augment nitric oxide status and improve endothelial function in healthy men and women: a randomized controlled trial. *Free Radical Biology and Medicine*, 52, 95-102.
- BOOKCHIN, R. M. & GALLOP, P. M. 1968. Structure of hemoglobin Alc: nature of the N-terminal beta chain blocking group. *Biochem Biophys Res Commun*, 32, 86-93.
- BOORSMA, R. K., WHITFIELD, J. & SPRIET, L. L. 2014. Beetroot Juice Supplementation Does Not Improve Performance in Elite 1500-m Runners. *Medicine and science in sports and exercise*.
- BOVE, P. F. & VAN DER VLIET, A. 2006. Nitric oxide and reactive nitrogen species in airway epithelial signaling and inflammation. *Free Radical Biology & Medicine*, 41, 515-527.
- BRANDENBURG, S. L., REUSCH, J., BAUER, T. A., JEFFERS, B. W., HIATT, W. R. & REGENSTEINER, J. G. 1999. Effects of exercise training on oxygen uptake kinetic responses in women with type 2 diabetes. *Diabetes care*, 22, 1640-1646.
- BREESE, B. C., MCNARRY, M. A., MARWOOD, S., BLACKWELL, J. R., BAILEY, S. J. & JONES, A. M. 2013. Beetroot juice supplementation speeds O2 uptake kinetics and improves exercise tolerance during severe-intensity exercise initiated from an elevated baseline. *Am J Physiol Regul Integr Comp Physiol*, 2, 2.
- BROWN, G. C. & COOPER, C. E. 1994. Nanomolar concentrations of nitric oxide reversibly inhibit synaptosomal respiration by competing with oxygen at cytochrome oxidase. *FEBS Letters*, 356, 295-298.

- BRUTON, J. D., LÄNNERGREN, J. & WESTERBLAD, H. 1998. Effects of CO2-induced acidification on the fatigue resistance of single mouse muscle fibers at 28 C. *Journal of Applied Physiology*, 85, 478-483.
- BRYAN, N., S 2006. Nitrite in nitric oxide biology: Cause or consequence? A systems based review. *Free Radical Biology & Medicine* 41, 691-701.
- BRYAN, N., S & HORD, N., G 2010. *Dietary nitrate and nitrites: The physiological context for potential health benefits*/ Food, nutrition and the nitric oxide pathway, DEStech Publications.
- BRYAN, N. S., FERNANDEZ, B. O., BAUER, S. M., GARCIA-SAURA, M. F., MILSOM, A. B., RASSAF, T., MALONEY, R. E., BHARTI, A., RODRIGUEZ, J. & FEELISCH, M. 2005. Nitrite is a signaling molecule and regulator of gene expression in mammalian tissues. *Nat Chem Biol*, 1, 290-297.
- BULLOCK, B. P., HELLER, R. S. & HABENER, J. F. 1996. Tissue distribution of messenger ribonucleic acid encoding the rat glucagon-like peptide-1 receptor. *Endocrinology*, 137, 2968-2978.
- BUNN, H. F., HANEY, D. N., GABBAY, K. H. & GALLOP, P. M. 1975. Further identification of the nature and linkage of the carbohydrate in hemoglobin A1c. *Biochem Biophys Res Commun*, 67, 103-9.
- BURMESTER, T., EBNER, B., WEICH, B. & HANKELN, T. 2002. Cytoglobin: A Novel Globin Type Ubiquitously Expressed inVertebrate Tissues. *Molecular Biology and Evolution*, 19, 416-421.
- BURROWS, B., KETTEL, L. J., NIDEN, A. H., RABINOWITZ, M. & DIENER, C. F. 1972. Patterns of Cardiovascular Dysfunction in Chronic Obstructive Lung Disease. *New England Journal of Medicine*, 286, 912-918.
- CACCIATORE, F., BRUZZESE, G., VITALE, D. F., LIGUORI, A., DE NIGRIS, F., FIORITO, C., INFANTE, T., DONATELLI, F., MINUCCI, P. B. & IGNARRO, L. J. 2011. Effects of ACE inhibition on circulating endothelial progenitor cells, vascular damage, and oxidative stress in hypertensive patients. *European journal of clinical pharmacology*, 67, 877-883.
- CALVER, A., COLLIER, J. & VALLANCE, P. 1992. Inhibition and stimulation of nitric oxide synthesis in the human forearm arterial bed of patients with insulin-dependent diabetes. *Journal of Clinical Investigation*, 90, 2548.
- CAMILLERI, M., MALAGELADA, J., BROWN, M., BECKER, G. & ZINSMEISTER, A. 1985. Relation between antral motility and gastric emptying of solids and liquids in humans. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 249, G580-G585.
- CARLSSON, S., WIKLUND, N. P., ENGSTRAND, L., WEITZBERG, E. & LUNDBERG, J. O. N. 2001. Effects of pH, Nitrite, and Ascorbic Acid on Nonenzymatic Nitric Oxide Generation and Bacterial Growth in Urine. *Nitric Oxide*, 5, 580-586.
- CARLSTRÖM, M., LARSEN, F. J., NYSTRÖM, T., HEZEL, M., BORNIQUEL, S., WEITZBERG, E. & LUNDBERG, J. O. 2010. Dietary inorganic nitrate reverses features of metabolic syndrome in endothelial nitric oxide synthase-deficient mice. *Proceedings of the National Academy of Sciences*, 107, 17716-17720.
- CARO, J. J., WARD, A. J. & O'BRIEN, J. A. 2002. Lifetime Costs of Complications Resulting From Type 2 Diabetes in the U.S. *Diabetes Care*, 25, 476-481.
- CARPENTIER, A., STRAGIER, S., BRÉJEON, C. & POORTMANS, J. R. 2014. Nitrate Supplementation, Exercise, and Kidney Function: Are There Detrimental Effects? *Medicine and science in sports and exercise*.
- CARREPEIRO, M. M., ROGERO, M. M., BERTOLAMI, M. C., BOTELHO, P. B., CASTRO, N. & CASTRO, I. A. 2011. Effect of n-3 fatty acids and statins on oxidative stress in statin-treated hypercholestorelemic and normocholesterolemic women. *Atherosclerosis*, 217, 171-178.
- CARTER, P., GRAY, L. J., TROUGHTON, J., KHUNTI, K. & DAVIES, M. J. 2010. Fruit and vegetable intake and incidence of type 2 diabetes mellitus: systematic review and meta-analysis. *BMJ*, 341.
- CASABURI, R., PATESSIO, A., IOLI, F., ZANABONI, S., DONNER, C. F. & WASSERMAN, K. 1991.

  Reductions in Exercise Lactic Acidosis and Ventilation as a Result of Exercise Training in Patients with Obstructive Lung Disease. *American Journal of Respiratory and Critical Care Medicine*, 143, 9-18.

- CASABURI, R., PORSZASZ, J., BURNS, M. R., CARITHERS, E. R., CHANG, R. S. & COOPER, C. B. 1997. Physiologic benefits of exercise training in rehabilitation of patients with severe chronic obstructive pulmonary disease. *American Journal of Respiratory and Critical Care Medicine*, 155, 1541-51.
- CASEY, D. P., TREICHLER, D. P., GANGER, C. T., SCHNEIDER, A. C. & UEDA, K. 2015. Acute dietary nitrate supplementation enhances compensatory vasodilation during hypoxic exercise in older adults. *Journal of Applied Physiology*, 118, 178-186.
- CASTELLO, P. R., DAVID, P. S., MCCLURE, T., CROOK, Z. & POYTON, R. O. 2006. Mitochondrial cytochrome oxidase produces nitric oxide under hypoxic conditions: Implications for oxygen sensing and hypoxic signaling in eukaryotes. *Cell Metabolism*, 3, 277-287.
- CATALÁN, V., GÓMEZ-AMBROSI, J., RODRÍGUEZ, A., SILVA, C., ROTELLAR, F., GIL, M. J., CIENFUEGOS, J. A., SALVADOR, J. & FRÜHBECK, G. 2008. Expression of caveolin-1 in human adipose tissue is upregulated in obesity and obesity-associated type 2 diabetes mellitus and related to inflammation. *Clinical Endocrinology*, 68, 213-219.
- CECONI, C., FOX, K. M., REMME, W. J., SIMOONS, M. L., BERTRAND, M., PARRINELLO, G., KLUFT, C., BLANN, A., COKKINOS, D. & FERRARI, R. 2007. ACE inhibition with perindopril and endothelial function. Results of a substudy of the EUROPA study: PERTINENT. *Cardiovascular Research*, 73, 237-246.
- CERMAK, N. M., GIBALA, M. J. & VAN LOON, L. J. 2012a. Nitrate supplementation's improvement of 10-km time-trial performance in trained cyclists. *Int J Sport Nutr Exerc Metab*, 22, 64-71.
- CERMAK, N. M., HANSEN, D., KOUW, I. W., VAN DIJK, J.-W., BLACKWELL, J. R., JONES, A. M., GIBALA, M. J. & VAN LOON, L. J. 2015. A single dose of sodium nitrate does not improve oral glucose tolerance in patients with type 2 diabetes mellitus. *Nutrition Research*.
- CERMAK, N. M., RES, P., STINKENS, R., LUNDBERG, J. O., GIBALA, M. J. & VAN LOON, L. J. 2012b. No improvement in endurance performance after a single dose of beetroot juice. *Int J Sport Nutr Exerc Metab*, 22, 470-8.
- CHAI, W., DONG, Z., WANG, N., WANG, W., TAO, L., CAO, W. & LIU, Z. 2012. Glucagon-Like Peptide 1 Recruits Microvasculature and Increases Glucose Use in Muscle via a Nitric Oxide—Dependent Mechanism. *Diabetes*, 61, 888-896.
- CHAKRAVORTTY, D. & HENSEL, M. 2003. Inducible nitric oxide synthase and control of intracellular bacterial pathogens. *Microbes and Infection*, 5, 621-627.
- CHALLACOMBE, S., PERCIVAL, R. & MARSH, P. 1995. Age-related changes in immunoglobulin isotypes in whole and parotid saliva and serum in healthy individuals. *Oral microbiology and immunology*, 10, 202-207.
- CHAOUAT, A., BUGNET, A.-S., KADAOUI, N., SCHOTT, R., ENACHE, I., DUCOLONÉ, A., EHRHART, M., KESSLER, R. & WEITZENBLUM, E. 2005. Severe Pulmonary Hypertension and Chronic Obstructive Pulmonary Disease. *American Journal of Respiratory and Critical Care Medicine*, 172, 189-194.
- CHEN, J., RADFORD, M. J., WANG, Y., MARCINIAK, T. A. & KRUMHOLZ, H. M. 2001. Effectiveness of beta-blocker therapy after acute myocardial infarction in elderly patients with chronic obstructive pulmonary disease or asthma. *Journal of the American College of Cardiology*, 37, 1950-1956.
- CHEN, M., YANG, T., MENG, X. & SUN, T. 2015. Azithromycin attenuates cigarette smoke extract-induced oxidative stress injury in human alveolar epithelial cells. *Mol Med Rep*.
- CHIEN, W. L., LIANG, K. C., TENG, C. M., KUO, S. C., LEE, F. Y. & FU, W. M. 2005. Enhancement of learning behaviour by a potent nitric oxide-guanylate cyclase activator YC-1. *European Journal of Neuroscience*, 21, 1679-1688.
- CHRISTENSEN, N. J. 1971. Diabetic angiopathy and neuropathy. A review with special reference to circulation in the extremities, the effect of hypophysectomy on capillary resistance and capillary permeability, functional abnormalities in early diabetes. *Acta medica Scandinavica*. *Supplementum*, 541, 3-60.

- CHRISTENSEN, P. M., NYBERG, M. & BANGSBO, J. 2013. Influence of nitrate supplementation on VO(2) kinetics and endurance of elite cyclists. *Scand J Med Sci Sports*, 23, 1.
- CIENCEWICKI, J., TRIVEDI, S. & KLEEBERGER, S. R. 2008. Oxidants and the pathogenesis of lung diseases. *The Journal of allergy and clinical immunology,* **122,** 456-68; quiz 469-70.
- CLERC, P., RIGOULET, M., LEVERVE, X. & FONTAINE, E. 2007. Nitric oxide increases oxidative phosphorylation efficiency. *Journal of bioenergetics and biomembranes*, 39, 158-166.
- CLINI, E., BIANCHI, L., FOGLIO, K., PORTA, R., VITACCA, M. & AMBROSINO, N. 2001. Effect of pulmonary rehabilitation on exhaled nitric oxide in patients with chronic obstructive pulmonary disease. *Thorax*, 56, 519-523.
- CNOP, M., FOUFELLE, F. & VELLOSO, L. A. 2012. Endoplasmic reticulum stress, obesity and diabetes. *Trends in molecular medicine*, 18, 59-68.
- COCKCROFT, A. E., SAUNDERS, M. J. & BERRY, G. 1981. Randomised controlled trial of rehabilitation in chronic respiratory disability. *Thorax*, 36, 200-203.
- COCKCROFT, J. R., WEBB, D. J. & WILKINSON, I. B. 2000. Arterial stiffness, hypertension and diabetes mellitus. *J Hum Hypertens*, 14, 377-80.
- COHEN, R. A. 1993. Dysfunction of vascular endothelium in diabetes mellitus. *Monograph-American Heart Association*, 87, V67-V76.
- COLES, L. T. & CLIFTON, P. M. 2012. Effect of beetroot juice on lowering blood pressure in free-living, disease-free adults: a randomized, placebo-controlled trial. *Nutr J*, 11, 1475-2891.
- COLLOFELLO, B., MOSKALIK, B. & ESSICK, E. 2014. Acute Dietary Nitrate Supplementation Decreases Systolic Blood Pressure and Increases Dry Static Apnea Performance in Females. *Journal of Exercise Physiology Online*, 17.
- COMLY, H., H 1945. Landmark article Sept 8, 1945: Cyanosis in infants caused by nitrates in well-water. . *Journal of American Medical Association* 257, 2788-2792.
- COOKE, J., ROSSITCH JR, E., ANDON, N., LOSCALZO, J. & DZAU, V. 1991. Flow activates an endothelial potassium channel to release an endogenous nitrovasodilator. *J. Clin. Invest*, 88, 1663-1671.
- COOPER, C. B. 1995. Determining the role of exercise in patients with chronic pulmonary disease. *Medicine & Science in Sports & Exercise*, 27, 147-157.
- COOPER, C. B. 2001. Exercise in chronic pulmonary disease: limitations and rehabilitation. *Medicine* and science in sports and exercise, 33, S643-6.
- COOPER, C. B. 2009. Desensitization to dyspnea in COPD with specificity for exercise training mode. *International journal of chronic obstructive pulmonary disease*, **4**, 33-43.
- COOPER, C. E. & GIULIVI, C. 2007. Nitric oxide regulation of mitochondrial oxygen consumption II: molecular mechanism and tissue physiology. *American Journal of Physiology-Cell Physiology*, 292, C1993-C2003.
- COPPEY, L. J., GELLETT, J. S., DAVIDSON, E. P., DUNLAP, J. A., LUND, D. D. & YOREK, M. A. 2001. Effect of Antioxidant Treatment of Streptozotocin-Induced Diabetic Rats on Endoneurial Blood Flow, Motor Nerve Conduction Velocity, and Vascular Reactivity of Epineurial Arterioles of the Sciatic Nerve. *Diabetes*, 50, 1927-1937.
- CORBIN, J. 2004. Mechanisms of action of PDE5 inhibition in erectile dysfunction. *International journal of impotence research*, 16, S4-S7.
- COSBY, K., PARTOVI, K., CRAWFORD, J., H, PATEL, R., P, REITER, C., D, MARTYR, S., YANG, B., K, WACLAWIW, M., A, ZALOS, G., XU, X., HUANG, K., T, SHIELDS, H., KIM-SHAPIRO, D., B, SCHECHTER, A., N, CANNON III, R., O & GLADWIN, M., T 2003. Nitrite reduction to nitric oxide by deoxyhemoglobin vasodiltaes the human circulation *Nature Medicine* 9, 1498-1505.
- COSSINS, A. R., WILLIAMS, D. R., FOULKES, N. S., BERENBRINK, M. & KIPAR, A. 2009. Diverse cell-specific expression of myoglobin isoforms in brain, kidney, gill and liver of the hypoxiatolerant carp and zebrafish. *Journal of Experimental Biology*, 212, 627-638.
- COVEY, M. K., LARSON, J. L. & WIRTZ, S. 1999. Reliability of submaximal exercise tests in patients with COPD. Chronic obstructive pulmonary disease. *Medicine and science in sports and exercise*, 31, 1257-1264.

- CRABTREE, M. J., HALE, A. B. & CHANNON, K. M. 2011. Dihydrofolate reductase protects endothelial nitric oxide synthase from uncoupling in tetrahydrobiopterin deficiency. *Free Radic Biol Med*, 50, 1639-46.
- CRABTREE, M. J., TATHAM, A. L., HALE, A. B., ALP, N. J. & CHANNON, K. M. 2009. Critical Role for Tetrahydrobiopterin Recycling by Dihydrofolate Reductase in Regulation of Endothelial Nitric-oxide Synthase Coupling: RELATIVE IMPORTANCE OF THE DE NOVO BIOPTERIN SYNTHESIS VERSUS SALVAGE PATHWAYS. *Journal of Biological Chemistry*, 284, 28128-28136.
- CREAGER, M. A., LÜSCHER, T. F., OF, P. W. T. A., COSENTINO, F. & BECKMAN, J. A. 2003. Diabetes and Vascular Disease: Pathophysiology, Clinical Consequences, and Medical Therapy: Part I. *Circulation*, 108, 1527-1532.
- CUNNINGTON, C., VAN ASSCHE, T., SHIRODARIA, C., KYLINTIREAS, I., LINDSAY, A. C., LEE, J. M., ANTONIADES, C., MARGARITIS, M., LEE, R. & CERRATO, R. 2012. Systemic and vascular oxidation limits the efficacy of oral tetrahydrobiopterin treatment in patients with coronary artery disease. *Circulation*, 125, 1356-1366.
- CUTTICA, M. J., KALHAN, R., SHLOBIN, O. A., AHMAD, S., GLADWIN, M., MACHADO, R. F., BARNETT, S. D. & NATHAN, S. D. 2010. Categorization and impact of pulmonary hypertension in patients with advanced COPD. *Respiratory Medicine*, 104, 1877-1882.
- CUZZOLIN, F. S. L., ROSSI, L., PASETTO, M. & BENONI, G. 2002. Plasma nitrite/nitrate and erythropoietin levels in cross-country skiers during altitude training. *Journal of Sports Medicine and Physical Fitness*, 42, 129-34.
- CUZZOLIN, L., LUSSIGNOLI, S., CRIVELLENTE, F., ADAMI, A., SCHENA, F., BELLAVITE, P., BROCCO, G. & BENONI, G. 2000. Influence of an acute exercise on neutrophil and platelet adhesion, nitric oxide plasma metabolites in inactive and active subjects. *International journal of sports medicine*, 21, 289-293.
- DAHLSTEDT, A. J., KATZ, A. & WESTERBLAD, H. 2001. Role of myoplasmic phosphate in contractile function of skeletal muscle: studies on creatine kinase-deficient mice. *J Physiol*, 533, 379-88.
- DAHLSTEDT, A. J., KATZ, A., WIERINGA, B. & WESTERBLAD, H. 2000. Is creatine kinase responsible for fatigue? Studies of isolated skeletal muscle deficient in creatine kinase. *Faseb j*, 14, 982-90.
- DAHLSTEDT, A. J. & WESTERBLAD, H. 2001. Inhibition of creatine kinase reduces the rate of fatigue-induced decrease in tetanic [Ca(2+)](i) in mouse skeletal muscle. *J Physiol*, 533, 639-49. DAILYMAIL. 2013. *Daily Mail*, 29/6/2010.
- DAVIS, B. J., XIE, Z., VIOLLET, B. & ZOU, M.-H. 2006. Activation of the AMP-Activated Kinase by Antidiabetes Drug Metformin Stimulates Nitric Oxide Synthesis In Vivo by Promoting the Association of Heat Shock Protein 90 and Endothelial Nitric Oxide Synthase. *Diabetes*, 55, 496-505.
- DE BOER, V. C., DIHAL, A. A., VAN DER WOUDE, H., ARTS, I. C., WOLFFRAM, S., ALINK, G. M., RIETJENS, I. M., KEIJER, J. & HOLLMAN, P. C. 2005. Tissue distribution of quercetin in rats and pigs. *The Journal of nutrition*, 135, 1718-1725.
- DE SYLLOS, R. W. C., SANDRIM, V. C., LISBOA, H. R. K., TRES, G. S. & TANUS-SANTOS, J. E. 2006. Endothelial nitric oxide synthase genotype and haplotype are not associated with diabetic retinopathy in diabetes type 2 patients. *Nitric Oxide*, 15, 417-422.
- DEJAM, A., HUNTER, C., J, TREMONTI, C., PLUTA, R., M, HON, Y., Y, GRIMES, G., PARTOVI, K., PELLETIER, M., M, OLDFIELD, E., H, CANNON III, R., O, SCHECHTER, A., N & GLADWIN, M., T 2007. Nitrate Infusion of Humans and Nonhuman Primates: Endocrine Effects, Pharmacokinetics, and Tolerance Formation. *Circulation*, 116, 1821-1831.
- DEL PRATO, S., FELTON, A. M., MUNRO, N., NESTO, R., ZIMMET, P. & ZINMAN, B. 2005. Improving glucose management: ten steps to get more patients with type 2 diabetes to glycaemic goal. *International journal of clinical practice*, 59, 1345-1355.
- DELP, M. D., BEHNKE, B. J., SPIER, S. A., WU, G. & MULLER-DELP, J. M. 2008. Ageing diminishes endothelium-dependent vasodilatation and tetrahydrobiopterin content in rat skeletal muscle arterioles. *J Physiol*, 586, 1161-8.

- DENICOLA, A., SOUZA, J. M., RADI, R. & LISSI, E. 1996. Nitric oxide diffusion in membranes determined by fluorescence quenching. *Archives of Biochemistry and Biophysics*, 328, 208-212.
- DENNINGER, J. W. & MARLETTA, M. A. 1999. Guanylate cyclase and the sup NO/cGMP signaling pathway. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1411, 334-350.
- DEVANGELIO, E., SANTILLI, F., FORMOSO, G., FERRONI, P., BUCCIARELLI, L., MICHETTI, N., CLISSA, C., CIABATTONI, G., CONSOLI, A. & DAVÌ, G. 2007. Soluble RAGE in type 2 diabetes: association with oxidative stress. *Free Radical Biology and Medicine*, 43, 511-518.
- DIBONAVENTURA, M., PAULOSE-RAM, R., SU, J., MCDONALD, M., ZOU, K. H., WAGNER, J. S. & SHAH, H. 2012. The impact of COPD on quality of life, productivity loss, and resource use among the elderly United States workforce. *Copd*, 9, 46-57.
- DING, K.-H., ZHONG, Q., XU, J. & ISALES, C. M. 2004. Glucose-dependent insulinotropic peptide: differential effects on hepatic artery vs. portal vein endothelial cells. *American Journal of Physiology Endocrinology and Metabolism*, 286, E773-E779.
- DIXON, H. B. 1972. A reaction of glucose with peptides. *Biochem J*, 129, 203-8.
- DOH, D. O. H. 2005. On the state of the public health: Annual Report of the Chief Medical Officer 2004.
- DOH, D. O. H. 2009. Chronic Obstructive Pulmonary Disease (COPD) Indicator Set: Dataset and Business rules: new GMS Contract QOF Implementation. (Version 14.0). *London: Department of Health*.
- DONOVAN, J. L., MEYER, A. S. & WATERHOUSE, A. L. 1998. Phenolic composition and antioxidant activity of prunes and prune juice (Prunus domestica). *Journal of agricultural and food chemistry*, 46, 1247-1252.
- DREIßIGACKER, U., WENDT, M., WITTKE, T., TSIKAS, D. & MAASSEN, N. 2010. Positive correlation between plasma nitrite and performance during high-intensive exercise but not oxidative stress in healthy men. *Nitric Oxide*, 23, 128-135.
- DRUCKER, D. J. 2006. The biology of incretin hormones. Cell Metabolism, 3, 153-165.
- DUCKWORTH, W., ABRAIRA, C., MORITZ, T., REDA, D., EMANUELE, N., REAVEN, P. D., ZIEVE, F. J., MARKS, J., DAVIS, S. N., HAYWARD, R., WARREN, S. R., GOLDMAN, S., MCCARREN, M., VITEK, M. E., HENDERSON, W. G. & HUANG, G. D. 2009. Glucose Control and Vascular Complications in Veterans with Type 2 Diabetes. *New England Journal of Medicine*, 360, 129-139.
- DUNCAN, C., DOUGALL, H., JOHNSTON, P., GREEN, S., BROGAN, R., SMITH, L., GOLDEN, M. & BENJAMIN, N. 1995. Chemical generation of nitric oxide in the mouth from the entrosalivery circulation of dietary nitrate. *Nature Medicine*, 1, 546-551.
- DUPUIS, J., LANGENBERG, C., PROKOPENKO, I., SAXENA, R., SORANZO, N., JACKSON, A. U., WHEELER, E., GLAZER, N. L., BOUATIA-NAJI, N. & GLOYN, A. L. 2010. New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. *Nature genetics*, 42, 105-116.
- DUPUY, P. M., SHORE, S. A., DRAZEN, J. M., FROSTELL, C., HILL, W. A. & ZAPOL, W. M. 1992. Bronchodilator action of inhaled nitric oxide in guinea pigs. *The Journal of clinical investigation*, 90, 421-428.
- DURANSKI, M. R., GREER, J. J., DEJAM, A., JAGANMOHAN, S., HOGG, N., LANGSTON, W., PATEL, R. P., YET, S.-F., WANG, X. & KEVIL, C. G. 2005. Cytoprotective effects of nitrite during in vivo ischemia-reperfusion of the heart and liver. *Journal of Clinical Investigation*, 115, 1232-1240.
- ECETOC. 1988. Technical report No. 27, Nitrate and Drinking Water, European Chemical Industry Ecology and Toxicology Centre, ISSN: 0773-8072-27.: Brussels: European Chemical Industry Ecology and Toxicology Centre.
- ELHERIK, K., KHAN, F., MCLAREN, M., KENNEDY, G. & BELCH, J. J. F. 2002. Circadian variation in vascular tone and endothelial cell function in normal males. *Clinical Science*, 102, 547-552.

- ENGAN, H. K., JONES, A. M., EHRENBERG, F. & SCHAGATAY, E. 2012. Acute dietary nitrate supplementation improves dry static apnea performance. *Respir Physiol Neurobiol*, 182, 53-9.
- ENGELGAU, M. M., THOMPSON, T. J., HERMAN, W. H., BOYLE, J. P., AUBERT, R. E., KENNY, S. J., BADRAN, A., SOUS, E. S. & ALI, M. A. 1997. Comparison of Fasting and 2-Hour Glucose and HbA1c Levels for Diagnosing Diabetes: Diagnostic criteria and performance revisited. *Diabetes Care*, 20, 785-791.
- ERIKSSON, J. W. 2007. Metabolic stress in insulin's target cells leads to ROS accumulation—a hypothetical common pathway causing insulin resistance. *FEBS letters*, 581, 3734-3742.
- ERZURUM, S. C., GHOSH, S., JANOCHA, A. J., XU, W., BAUER, S., BRYAN, N. S., TEJERO, J., HEMANN, C., HILLE, R., STUEHR, D. J., FEELISCH, M. & BEALL, C. M. 2007. Higher blood flow and circulating NO products offset high-altitude hypoxia among Tibetans. *Proceedings of the National Academy of Sciences of the United States of America*, 104, 17593-17598.
- EUSTERMAN, G., B & KEITH, N., M 1929. Transient methemoglobinemia following administration of ammonium nitrate. *M Clin*, 12.
- FALCONER, C. L., PAGE, A. S., ANDREWS, R. C. & COOPER, A. R. 2015. The Potential Impact of Displacing Sedentary Time in Adults with Type 2 Diabetes. *Medicine & Science in Sports & Exercise*.
- FERGUSON, S., HOLDSWORTH, C., WRIGHT, J., FEES, A., MUSCH, T. & POOLE, D. 2014. Impact of nitrate supplementation via beetroot juice on capillary hemodynamics in skeletal muscle of rats in chronic heart failure (1106.16). *The FASEB Journal*, 28, 1106.16.
- FERGUSON, S. K., HIRAI, D. M., COPP, S. W., HOLDSWORTH, C. T., ALLEN, J. D., JONES, A. M., MUSCH, T. I. & POOLE, D. C. 2013. Effects of nitrate supplementation via beetroot juice on contracting rat skeletal muscle microvascular oxygen pressure dynamics. *Respir Physiol Neurobiol*, 187, 250-5.
- FIACCADORI, E., DEL CANALE, S., VITALI, P., COFFRINI, E., RONDA, N. & GUARIGLIA, A. 1987. SKeletal muscle energetics, acid-base equilibrium and lactate metabolism in patients with severe hypercapnia and hypoxemia. *Chest*, 92, 883-887.
- FISHMAN, A. P., SAMET, P. & COURNAND, A. 1955. Ventilatory drive in chronic pulmonary emphysema. *The American Journal of Medicine*, 19, 533-548.
- FITZGIBBONS, J. F., KOLER, R. D. & JONES, R. T. 1976. Red cell age-related changes of hemoglobins Ala+b and Alc in normal and diabetic subjects. *J Clin Invest*, 58, 820-4.
- FLORIN, T. H. J., NEALE, G. & CUMMINGS, J. H. 1990. The effect of dietary nitrate on nitrate and nitrite excretion in man. *British Journal of Nutrition*, 64, 387-397.
- FOLEY, R. N. & COLLINS, A. J. 2007. End-Stage Renal Disease in the United States: An Update from the United States Renal Data System. *Journal of the American Society of Nephrology,* 18, 2644-2648.
- FONG, D. S., AIELLO, L., GARDNER, T. W., KING, G. L., BLANKENSHIP, G., CAVALLERANO, J. D., FERRIS, F. L. & KLEIN, R. 2004. Retinopathy in Diabetes. *Diabetes Care*, 27, s84-s87.
- FONTBONNE, A., ESCHWEGE, E., CAMBIEN, F., RICHARD, J.-L., DUCIMETIERE, P., THIBULT, N., WARNET, J.-M., CLAUDE, J.-R. & ROSSELIN, G.-E. 1989. Hypertriglyceridaemia as a risk factor of coronary heart disease mortality in subjects with impaired glucose tolerance or diabetes. *Diabetologia*, 32, 300-304.
- FORDEL, E., THIJS, L., MOENS, L. & DEWILDE, S. 2007. Neuroglobin and cytoglobin expression in mice. *FEBS Journal*, 274, 1312-1317.
- FÖRSTERMANN, U. & SESSA, W. C. 2012. Nitric oxide synthases: regulation and function. *European Heart Journal*, 33, 829-837.
- FORTE, P., COPLAND, M., SMITH, L. M., MILNE, E., SUTHERLAND, J. & BENJAMIN, N. 1997. Basal nitric oxide synthesis in essential hypertension. *The Lancet*, 349, 837-842.

- FOX, C. S., COADY, S., SORLIE, P. D., D'AGOSTINO, R. B., SR., PENCINA, M. J., VASAN, R. S., MEIGS, J. B., LEVY, D. & SAVAGE, P. J. 2007. Increasing cardiovascular disease burden due to diabetes mellitus: the Framingham Heart Study. *Circulation*, 115, 1544-50.
- FOX, C. S., COADY, S., SORLIE, P. D., LEVY, D., MEIGS, J. B., D'AGOSTINO, R. B., SR., WILSON, P. W. & SAVAGE, P. J. 2004. Trends in cardiovascular complications of diabetes. *Jama*, 292, 2495-9.
- FRANCIS, S. H. & CORBIN, J. D. 2005. Sildenafil: efficacy, safety, tolerability and mechanism of action in treating erectile dysfunction. *Expert Opinion on Drug Metabolism Toxicology*, 1, 283-293.
- FRATACCI, M. D., FROSTELL, C. G., CHEN, T. Y., WAIN, J. C., ROBINSON, D. R. & ZAPOL, W. M. 1991. Inhaled nitric oxide. A selective pulmonary vasodilator of heparin-protamine vasoconstriction in sheep. *Anesthesiology*, 75, 990-999.
- FREITAS, M. R. D., NASCIMENTO, O. J., CHIMELLI, L., ESCADA, T. & NEVARES, M. 1992. Neuropatia diabética: I. Conceito, epidemiologia, classificação, quadro clínico e eletroneuromiográfico: estudo de 210 casos; Diabetic neuropathy: I. Definition, epidemiology, classification, clinical features and electrophysiologic changes: study of 210 patients. *Rev. bras. neurol*, 28, 69-73.
- FROSTELL, C., FRATACCI, M. D., WAIN, J. C., JONES, R. & ZAPOL, W. M. 1991. Inhaled nitric oxide. A selective pulmonary vasodilator reversing hypoxic pulmonary vasoconstriction. *Circulation*, 83, 2038-47.
- FRYER, M. W., OWEN, V. J., LAMB, G. D. & STEPHENSON, D. G. 1995. Effects of creatine phosphate and P(i) on Ca2+ movements and tension development in rat skinned skeletal muscle fibres. *J Physiol*, 482 ( Pt 1), 123-40.
- FULFORD, J., WINYARD, P. G., VANHATALO, A., BAILEY, S. J., BLACKWELL, J. R. & JONES, A. M. 2013. Influence of dietary nitrate supplementation on human skeletal muscle metabolism and force production during maximum voluntary contractions. *Pflugers Arch*, 465, 517-28.
- GÆDE, P., VEDEL, P., PARVING, H.-H. & PEDERSEN, O. 1999. Intensified multifactorial intervention in patients with type 2 diabetes mellitus and microalbuminuria: the Steno type 2 randomised study. *The Lancet*, 353, 617-622.
- GAGO, B., LUNDBERG, J. O., BARBOSA, R. M. & LARANJINHA, J. 2007. Red wine-dependent reduction of nitrite to nitric oxide in the stomach. *Free Radical Biology and Medicine*, 43, 1233-1242.
- GALANT, L. H., FORGIARINI, J. L., DIAS, A. S. & MARRONI, C. A. 2012. Maximum oxygen consumption predicts mortality in patients with alcoholic cirrhosis. *Hepato-gastroenterology*, 60, 1127-1130.
- GALLAGHER, C., G 1994. Exercise limitation and clinical exercise testing in chronic obstructive pulmonary disease. *Clinics in Chest Medicine*, 15:, 305-326.
- GAMGEE, A. 1868. Researches on the blood; on the Action of Nitrates on the Blood. . PHILOSOPHICAL TRANSACTIONS OF THE ROYAL SOCIETY OF LONDON, 158, 589-625.
- GARCÍA-CARDEÑA, G., MARTASEK, P., MASTERS, B. S. S., SKIDD, P. M., COUET, J., LI, S., LISANTI, M. P. & SESSA, W. C. 1997. Dissecting the Interaction between Nitric Oxide Synthase (NOS) and Caveolin FUNCTIONAL SIGNIFICANCE OF THE NOS CAVEOLIN BINDING DOMAININ VIVO. *Journal of Biological Chemistry*, 272, 25437-25440.
- GARTHWAITE, J. 2008. Concepts of neural nitric oxide-mediated transmission. *European Journal of Neuroscience*, 27, 2783-2802.
- GATEHOUSE, P. D., KEEGAN, J., CROWE, L. A., MASOOD, S., MOHIADDIN, R. H., KREITNER, K.-F. & FIRMIN, D. N. 2005. Applications of phase-contrast flow and velocity imaging in cardiovascular MRI. *European radiology*, 15, 2172-2184.
- GAUTIER, C., VAN FAASSEN, E., MIKULA, I., MARTASEK, P. & SLAMA-SCHWOK, A. 2006. Endothelial nitric oxide synthase reduces nitrite anions to NO under anoxia. *Biochemical and Biophysical Research Communications*, 341, 816-821.
- GAVIN, J., ALBERTI, K., DAVIDSON, M., DEFRONZO, R., DRASH, A., GABBE, S., GENUTH, S., HARRIS, M., KAHN, R. & KEEN, H. 1997. Report of the expert committee on the diagnosis and classification of diabetes mellitus. *Diabetes care*, 20, 1183-1197.

- GERRITSEN, J., DEKKER, J. M., TENVOORDE, B. J., KOSTENSE, P. J., HEINE, R. J., BOUTER, L. M., HEETHAAR, R. M. & STEHOUWER, C. D. A. 2001. Impaired Autonomic Function Is Associated With Increased Mortality, Especially in Subjects With Diabetes, Hypertension, or a History of Cardiovascular Disease: The Hoorn Study. *Diabetes Care*, 24, 1793-1798.
- GHASEMI, A., ZAHEDIASL, S. & AZIZI, F. 2010. Nitric oxide and clustering of metabolic syndrome components in pediatrics. *European Journal of Epidemiology*, 25, 45-53.
- GIAID, A. & SALEH, D. 1995. Reduced expression of endothelial nitric oxide synthase in the lungs of patients with pulmonary hypertension. *New England Journal of Medicine*, 333, 214-221.
- GILCHRIST, M., WINYARD, P. & BENJAMIN, N. 2010. Dietary Nitrate Good or bad? *Nitric Oxide*, 22, 104-109.
- GILCHRIST, M., WINYARD, P. G., AIZAWA, K., ANNING, C., SHORE, A. & BENJAMIN, N. 2013. Effect of dietary nitrate on blood pressure, endothelial function, and insulin sensitivity in type 2 diabetes. *Free Radical Biology and Medicine*, 60, 89-97.
- GILCHRIST, M., WINYARD, P. G., FULFORD, J., ANNING, C., SHORE, A. & BENJAMIN, N. 2014. Dietary nitrate supplementation improves reaction time in type 2 diabetes: Development and application of a novel nitrate-depleted beetroot juice placebo. *Nitric Oxide*.
- GIRGIS, R. E., LI, D., ZHAN, X., GARCIA, J. G. N., TUDER, R. M., HASSOUN, P. M. & JOHNS, R. A. 2003. Attenuation of chronic hypoxic pulmonary hypertension by simvastatin. *American Journal of Physiology Heart and Circulatory Physiology*, 285, H938-H945.
- GOKE, R., LARSEN, P. J., MIKKELSEN, J. D. & SHEIKH, S. P. 1995. Distribution of GLP-1 binding sites in the rat brain: evidence that exendin-4 is a ligand of brain GLP-1 binding sites. *Eur J Neurosci*, 7, 2294-300.
- GOLD., G. I. F. C. O. L. D. 2011. *Guidlines: Global strategy for the diagnosis, managment and prevention of COPD.* [Online]. <a href="http://www.goldcopd.org/">http://www.goldcopd.org/</a>. [Accessed 16/10/2012 2012].
- GOSKER, H. R., LENCER, N. H. M. K., FRANSSEN, F. M. E., VAN DER VUSSE, G. J., WOUTERS, E. F. M. & SCHOLS, A. M. W. J. 2003. Striking Similarities in Systemic Factors Contributing to Decreased Exercise Capacity in Patients With Severe Chronic Heart Failure or COPD\*. *CHEST Journal*, 123, 1416-1424.
- GOVONI, M., JANSSON, E. Å., WEITZBERG, E. & LUNDBERG, J. O. 2008. The increase in plasma nitrite after a dietary nitrate load is markedly attenuated by an antibacterial mouthwash. *Nitric Oxide*, 19, 333-337.
- GOW, A. J., LUCHSINGER, B. P., PAWLOSKI, J. R., SINGEL, D. J. & STAMLER, J. S. 1999. The oxyhemoglobin reaction of nitric oxide. *Proc Natl Acad Sci U S A*, 96, 9027-32.
- GOW, A. J. & STAMLER, J. S. 1998. Reactions between nitric oxide and haemoglobin under physiological conditions. *Nature*, 391, 169-173.
- GREEN, B. D., HAND, K. V., DOUGAN, J. E., MCDONNELL, B. M., CASSIDY, R. S. & GRIEVE, D. J. 2008. GLP-1 and related peptides cause concentration-dependent relaxation of rat aorta through a pathway involving KATP and cAMP. *Archives of Biochemistry and Biophysics*, 478, 136-142.
- GRIFFITHS, T. L., BURR, M. L., CAMPBELL, I. A., LEWIS-JENKINS, V., MULLINS, J., SHIELS, K., TURNER-LAWLOR, P. J., PAYNE, N., NEWCOMBE, R. G., LONESCU, A. A., THOMAS, J. & TUNBRIDGE, J. 2000. Results at 1 year of outpatient multidisciplinary pulmonary rehabilitation: a randomised controlled trial. *The Lancet*, 355, 362-368.
- GRUETTER, C. A., CHILDERS, C. E., BOSSERMAN, M. K., LEMKE, S. M., BALL, J. G. & VALENTOVIC, M. A. 1989. Comparison of Relaxation Induced by Glyceryl Trinitrate, Isosorbide Dinitrate, and Sodium Nitroprusside in Bovine Airways. *American Journal of Respiratory and Critical Care Medicine*, 139, 1192-1197.
- GRUETTER, C. A., GRUETTER, D. Y., LYON, J. E., KADOWITZ, P. J. & IGNARRO, L. J. 1981. Relationship between cyclic guanosine 3':5'-monophosphate formation and relaxation of coronary arterial smooth muscle by glyceryl trinitrate, nitroprusside, nitrite and nitric oxide: effects of methylene blue and methemoglobin. *Journal of Pharmacology and Experimental Therapeutics*, 219, 181-186.

- GUAN, X., STOLL, B., LU, X., TAPPENDEN, K. A., HOLST, J. J., HARTMANN, B. & BURRIN, D. G. 2003. GLP-2-mediated up-regulation of intestinal blood flow and glucose uptake is nitric oxide-dependent in TPN-fed piglets. *Gastroenterology*, 125, 136-147.
- GUAZZI, M., TUMMINELLO, G., MATTURRI, M. & GUAZZI, M. D. 2003. Insulin ameliorates exercise ventilatory efficiency and oxygen uptake in patients with heart failure—type 2 diabetes comorbidity. *Journal of the American College of Cardiology*, 42, 1044-1050.
- GUSSO, S., HOFMAN, P., LALANDE, S., CUTFIELD, W., ROBINSON, E. & BALDI, J. 2008. Impaired stroke volume and aerobic capacity in female adolescents with type 1 and type 2 diabetes mellitus. *Diabetologia*, 51, 1317-1320.
- HANEFELD, M., FISCHER, S., SCHMECHEL, H., ROTHE, G., SCHULZE, J., DUDE, H., SCHWANEBECK, U. & JULIUS, U. 1991. Diabetes Intervention Study. Multi-intervention trial in newly diagnosed NIDDM. *Diabetes Care*, 14, 308-17.
- HARIRI, S., YOON, P. W., QURESHI, N., VALDEZ, R., SCHEUNER, M. T. & KHOURY, M. J. 2006. Family history of type 2 diabetes: a population-based screening tool for prevention? *Genetics in Medicine*, 8, 102-108.
- HARRIS, M. I. 1998. Diabetes in America: Epidemiology and Scope of the Problem. *Diabetes Care*, 21, C11-C14.
- HARRIS, M. I., KLEIN, R., COWIE, C. C., ROWLAND, M. & BYRD-HOLT, D. D. 1998. Is the Risk of Diabetic Retinopathy Greater in Non-Hispanic Blacks and Mexican Americans Than in Non-Hispanic Whites With Type 2 Diabetes?: A U.S. population study. *Diabetes Care*, 21, 1230-1235.
- HART, G. W., SLAWSON, C., RAMIREZ-CORREA, G. & LAGERLOF, O. 2011. Cross talk between O-GlcNAcylation and phosphorylation: roles in signaling, transcription, and chronic disease. *Annual review of biochemistry*, 80, 825.
- HART, J. D. & DULHUNTY, A. F. 2000. Nitric oxide activates or inhibits skeletal muscle ryanodine receptors depending on its concentration, membrane potential and ligand binding. *J Membr Biol*, 173, 227-36.
- HATTORI, Y., JOJIMA, T., TOMIZAWA, A., SATOH, H., HATTORI, S., KASAI, K. & HAYASHI, T. 2010. RETRACTED ARTICLE: A glucagon-like peptide-1 (GLP-1) analogue, liraglutide, upregulates nitric oxide production and exerts anti-inflammatory action in endothelial cells. *Diabetologia*, 53, 2256-2263.
- HATTORI, Y., NAKANISHI, N., AKIMOTO, K., YOSHIDA, M. & KASAI, K. 2003. HMG-CoA reductase inhibitor increases GTP cyclohydrolase I mRNA and tetrahydrobiopterin in vascular endothelial cells. *Arteriosclerosis, Thrombosis, and Vascular Biology,* 23, 176-182.
- HEALTHCARE-COMMISSION 2006. Clearing the air: A national study of chronic obstructive pulmonary disease. Healhcare Commission.
- HEMMINKI, K., LI, X., SUNDQUIST, K. & SUNDQUIST, J. 2010. Familial Risks for Type 2 Diabetes in Sweden. *Diabetes Care*, 33, 293-297.
- HERDER, C. & RODEN, M. 2011. Genetics of type 2 diabetes: pathophysiologic and clinical relevance. *European Journal of Clinical Investigation*, 41, 679-692.
- HERTZEL C. GERSTEIN, MCMASTER HAMILTON, MICHAEL E. MILLER, ROBERT P. BYINGTON, DAVID C. GOFF, WAKE FOREST, BIGGER, T., JOHN B. BUSE, CHAPEL HILL, WILLIAM C. CUSHMAN, SAUL GENUTH, RICHARD H. GRIMM, JEFFREY L. PROBSTFIELD, DENISE G. SIMONSMORTON & WILLIAM T. FRIEDEWALD 2008. Effects of Intensive Glucose Lowering in Type 2 Diabetes. New England Journal of Medicine, 358, 2545-2559.
- HEX, N., BARTLETT, C., WRIGHT, D., TAYLOR, M. & VARLEY, D. 2012. Estimating the current and future costs of Type 1 and Type 2 diabetes in the UK, including direct health costs and indirect societal and productivity costs. *Diabetic Medicine*, 29, 855-862.
- HIGUCHI, K. & MOTOMIZU, S. 1999. Flow-injection spectrophotometric determination of nitrite and nitrate in biological samples. *Analytical sciences*, **15**, **129-134**.

- HINKLE, P. C. 2005. P/O ratios of mitochondrial oxidative phosphorylation. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1706, 1-11.
- HIRAYAMA, F., LEE, A. H., BINNS, C. W., ZHAO, Y., HIRAMATSU, T., TANIKAWA, Y., NISHIMURA, K. & TANIGUCHI, H. 2009. Do vegetables and fruits reduce the risk of chronic obstructive pulmonary disease? A case—control study in Japan. *Preventive Medicine*, 49, 184-189.
- HOBBS, D. A., KAFFA, N., GEORGE, T. W., METHVEN, L. & LOVEGROVE, J. A. 2012. Blood pressure-lowering effects of beetroot juice and novel beetroot-enriched bread products in normotensive male subjects. *Br J Nutr*, 108, 2066-74.
- HOLLOSZY, J. O. 2009. Skeletal muscle "mitochondrial deficiency" does not mediate insulin resistance. *The American journal of clinical nutrition*, 89, 463S-466S.
- HOLST, J. J. 2007. The physiology of glucagon-like peptide 1. *Physiol Rev*, 87, 1409-39.
- HOLVERDA, S., BOGAARD, H. J., GROEPENHOFF, H., POSTMUS, P. E., BOONSTRA, A. & VONK-NOORDEGRAAF, A. 2008. Cardiopulmonary exercise test characteristics in patients with chronic obstructive pulmonary disease and associated pulmonary hypertension. *Respiration; international review of thoracic diseases*, 76, 160-167.
- HOON, M. W., HOPKINS, W. G., JONES, A. M., MARTIN, D. T., HALSON, S. L., WEST, N. P., JOHNSON, N. A. & BURKE, L. M. 2014. NITRATE SUPPLEMENTATION AND HIGH-INTENSITY PERFORMANCE IN COMPETITIVE CYCLISTS. *Applied Physiology, Nutrition, and Metabolism*.
- HOON, M. W., JONES, A. M., JOHNSON, N. A., BLACKWELL, J. R., BROAD, E. M., LUNDY, B., RICE, A. J. & BURKE, L. M. 2013. The Effect of Variable Doses of Inorganic Nitrate-Rich Beetroot Juice on Simulated 2,000 m Rowing Performance in Trained Athletes. *Int J Sports Physiol Perform*, 30, 30.
- HOPE 2000. Effects of ramipril on cardiovascular and microvascular outcomes in people with diabetes mellitus: results of the HOPE study and MICRO-HOPE substudy. *The Lancet*, 355, 253-259.
- HOPKINS, N. & MCLOUGHLIN, P. 2002. The structural basis of pulmonary hypertension in chronic lung disease: remodelling, rarefaction or angiogenesis? *Journal of Anatomy*, 201, 335-348.
- HORD, N., G, TANG, Y. & BRYAN, N., S 2009. Food sources of nitrates and nitrites: the physiologic context for potential health benefits<sup>1-3</sup>. *American Journal of Clinical Nutrition* 90, 1-10.
- HSU, L. L., CHAMPION, H. C., CAMPBELL-LEE, S. A., BIVALACQUA, T. J., MANCI, E. A., DIWAN, B. A., SCHIMEL, D. M., COCHARD, A. E., WANG, X. & SCHECHTER, A. N. 2007. Hemolysis in sickle cell mice causes pulmonary hypertension due to global impairment in nitric oxide bioavailability. *Blood*, 109, 3088-3098.
- HUEBSCHMANN, A. G., REIS, E. N., EMSERMANN, C., DICKINSON, L. M., REUSCH, J. E. B., BAUER, T. A. & REGENSTEINER, J. G. 2009. Women with type 2 diabetes perceive harder effort during exercise than nondiabetic women. *Applied Physiology, Nutrition, and Metabolism,* 34, 851-857.
- HUISMAN, T. H. & DOZY, A. M. 1962. Studies on the heterogeneity of hemoglobin. V. Binding of hemoglobin with oxidized glutathione. *J Lab Clin Med*, 60, 302-19.
- HUYNH, N. T. & TAYEK, J. A. 2002. Oral arginine reduces systemic blood pressure in type 2 diabetes: its potential role in nitric oxide generation. *Journal of the American College of Nutrition*, 21, 422-427.
- HWANG, S. 2011. Microcirculation of the Liver. Venous Embolization of the Liver. Springer.
- ICHINOSE, F., ROBERTS, J. D. & ZAPOL, W. M. 2004. Inhaled Nitric Oxide. Circulation, 109, 3106-3111.
- IGNARRO, L. J., BUGA, G. M., WOOD, K. S., BYRNS, R. E. & CHAUDHURI, G. 1987. Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. *Proceedings of the National Academy of Sciences*, 84, 9265-9269.
- IGNARRO, L. J., CIRINO, G., CASINI, A. & NAPOLI, C. 1999. Nitric oxide as a signaling molecule in the vascular system: an overview. *J Cardiovasc Pharmacol*, 34, 879-86.
- IMANISHI, T., IKEJIMA, H., TSUJIOKA, H., KUROI, A., KOBAYASHI, K., MURAGAKI, Y., MOCHIZUKI, S., GOTO, M., YOSHIDA, K. & AKASAKA, T. 2008. Addition of eplerenone to an angiotensin-

- converting enzyme inhibitor effectively improves nitric oxide bioavailability. *Hypertension*, 51, 734-741.
- IMPERATORE, G., HANSON, R. L., PETTITT, D. J., KOBES, S., BENNETT, P. H. & KNOWLER, W. C. 1998. Sib-pair linkage analysis for susceptibility genes for microvascular complications among Pima Indians with type 2 diabetes. Pima Diabetes Genes Group. *Diabetes*, 47, 821-830.
- ITO, A., EGASHIRA, K., NARISHIGE, T., MURAMATSU, K. & TAKESHITA, A. 2002. Angiotensin-converting enzyme activity is involved in the mechanism of increased endogenous nitric oxide synthase inhibitor in patients with type 2 diabetes mellitus. *Circulation journal: official journal of the Japanese Circulation Society*, 66, 811-815.
- JACOB, R. A., SPINOZZI, G. M., SIMON, V. A., KELLEY, D. S., PRIOR, R. L., HESS-PIERCE, B. & KADER, A. A. 2003. Consumption of cherries lowers plasma urate in healthy women. *The Journal of nutrition*, 133, 1826-1829.
- JACOBY, D. S. & RADER, D. J. 2003. Renin-angiotensin system and atherothrombotic disease: from genes to treatment. *Archives of internal medicine*, 163, 1155-1164.
- JAFFE, A. S., SPADARO, J. J., SCHECHTMAN, K., ROBERTS, R., GELTMAN, E. M. & SOBEL, B. E. 1984. Increased congestive heart failure after myocardial infarction of modest extent in patients with diabetes mellitus. *Am Heart J*, 108, 31-7.
- JAJJA, A., SUTYARJOKO, A., LARA, J., RENNIE, K., BRANDT, K., QADIR, O. & SIERVO, M. 2014. Beetroot supplementation lowers daily systolic blood pressure in older, overweight subjects. *Nutrition Research*, 34, 868-875.
- JAMES, P. E., LANG, D., TUFNELL-BARRET, T., MILSOM, A. B. & FRENNEAUX, M. P. 2004. Vasorelaxation by red blood cells and impairment in diabetes reduced nitric oxide and oxygen delivery by glycated hemoglobin. *Circulation Research*, 94, 976-983.
- JANSSON, E. A., HUANG, L., MALKEY, R., GOVONI, M., NIHLEN, C., OLSSON, A., STENSDOTTER, M., PETERSSON, J., HOLM, L., WEITZBERG, E. & LUNDBERG, J. O. 2008. A mammalian functional nitrate reductase that regulates nitrite and nitric oxide homeostasis. *Nat Chem Biol*, 4, 411-417.
- JARDIN, F., GUERET, P., PROST, J., FARCOT, J., OZIER, Y. & BOURDARIAS, J. 1984. Two-dimensional echocardiographic assessment of left ventricular function in chronic obstructive pulmonary disease. *The American review of respiratory disease*, 129, 135-142.
- JARRETT, R. J., KEEN, H. & MCCARTNEY, P. 1984. The Whitehall Study: Ten Year Follow-up Report on Men With Impaired Glucose Tolerance With Reference to Worsening to Diabetes and Predictors of Death. *Diabetic Medicine*, 1, 279-283.
- JIA, L., BONAVENTURA, C., BONAVENTURA, J. & STAMLER, J. S. 1996. S-nitrosohaemoglobin: a dynamic activity of blood involved in vascular control.
- JIANG, H., TORREGROSSA, A. C., POTTS, A., PIERINI, D., ARANKE, M., GARG, H. K. & BRYAN, N. S. 2014. Dietary nitrite improves insulin signaling through GLUT4 translocation. *Free Radical Biology and Medicine*, 67, 51-57.
- JONES, A., M & POOLE, D., C 2005. Towards an understanding of the mechanistic bases of oxygen uptake kinetics *Exercise and Medicine* 294-328.
- JOSHIPURA, K., J, ASCHERIO, A., MANSON, J., E, STAMPFER, M., J, RIMM, E., B, SPEIZER F, E., HENNEKENS, C., H, SPIEGELMANN, D. & WILLET, W., C 1999. Fruit and Vegetable Intake in Relation to Risk of Ischemic Stroke. *American Medical Association* 282, 1233-1239.
- JOSHIPURA, K., J, HU, F., B, MANSON, J., E, STAMPFER, M., J, RIMM, E., B, SPEIZER F, E., COLDITZ, G., ASCHERIO, A., ROSNER, B., SPIEGELMANN, D. & WILLET, W., C 2001. The Effect of Fruit and Vegetable Intake on Risk of Coronary Heart Disease. *Annals of Internal Medicine*, 134, 1106-1114.
- JUNGERSTEN, L., AMBRING, A., WALL, B. & WENNMALM, Å. 1997. Both physical fitness and acute exercise regulate nitric oxide formation in healthy humans. *Journal of Applied Physiology*, 82, 760-764.

- KABBARA, A. A. & ALLEN, D. G. 1999. The role of calcium stores in fatigue of isolated single muscle fibres from the cane toad. *J Physiol*, 519 Pt 1, 169-76.
- KABBARA, A. A. & ALLEN, D. G. 2001. The use of the indicator fluo-5N to measure sarcoplasmic reticulum calcium in single muscle fibres of the cane toad. *J Physiol*, 534, 87-97.
- KABIR, M., CATALANO, K. J., ANANTHNARAYAN, S., KIM, S. P., VAN CITTERS, G. W., DEA, M. K. & BERGMAN, R. N. 2005. Molecular evidence supporting the portal theory: a causative link between visceral adiposity and hepatic insulin resistance. *American Journal of Physiology-Endocrinology and Metabolism*, 288, E454-E461.
- KAHN, S. E., HULL, R. L. & UTZSCHNEIDER, K. M. 2006. Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature*, 444, 840-846.
- KANG, L. S., REYES, R. A. & MULLER-DELP, J. M. 2009. Aging impairs flow-induced dilation in coronary arterioles: role of NO and H(2)O(2). *Am J Physiol Heart Circ Physiol*, 297, H1087-95.
- KANNER, J., HAREL, S. & GRANIT, R. 2001. Betalains a new class of dietary cationized antioxidants. *Journal of Agricultural and Food Chemistry*, 49, 5178-5185.
- KAPIL, V., HAYDAR, S. M. A., PEARL, V., LUNDBERG, J. O., WEITZBERG, E. & AHLUWALIA, A. 2012. Physiological role for nitrate-reducing oral bacteria in blood pressure control. *Free Radical Biology and Medicine*.
- KAPIL, V., KHAMBATA, R. S., ROBERTSON, A., CAULFIELD, M. J. & AHLUWALIA, A. 2015. Dietary Nitrate Provides Sustained Blood Pressure Lowering in Hypertensive Patients: A Randomized, Phase 2, Double-Blind, Placebo-Controlled Study. *Hypertension*, 65, 320-327.
- KAPIL, V., MILSOM, A., B, OKORIE, M., MALEKI-TOYSERKANI, S., AKRAM, F., REHMAN, F., ARGHANDAWI, S., PEARL, V., BENJAMIN, N., LOUKOGEORGAKIS, S., MACALLISTER, R., HOBBS, A., J, WEBB, A., J & AHLUWALIA, A. 2011. Inorganic Nitrate Supplimentation Lowers Blood Pressure in Humans. Role for Nititre-Derived NO. *Hypertension*, 56, 274-281.
- KAROLI, N. A. & REBROV, A. P. 2005. Chronic obstructive lung disease and coronary heart disease. *Klinicheskaia meditsina*, 83, 72-76.
- KAZIMIERCZAK, R., HALLMANN, E., LIPOWSKI, J., DRELA, N., KOWALIK, A., PUSSA, T., MATT, D., LUIK, A., GOZDOWSKI, D. & REMBIALKOWSKA, E. 2014. Beetroot (Beta vulgaris L.) and naturally fermented beetroot juices from organic and conventional production: metabolomics, antioxidant levels and anticancer activity. *J Sci Food Agric*, 94, 2618-29.
- KEEN, J. T., LEVITT, E. L., HODGES, G. J. & WONG, B. J. 2014. Short-term dietary nitrate supplementation augments cutaneous vasodilatation and reduces mean arterial pressure in healthy humans. *Microvascular research*.
- KELLEY, D. E., HE, J., MENSHIKOVA, E. V. & RITOV, V. B. 2002. Dysfunction of Mitochondria in Human Skeletal Muscle in Type 2 Diabetes. *Diabetes*, 51, 2944-2950.
- KELLEY, D. E., MOKAN, M., SIMONEAU, J.-A. & MANDARINO, L. 1993. Interaction between glucose and free fatty acid metabolism in human skeletal muscle. *Journal of Clinical Investigation*, 92, 91.
- KELLEY, D. S., RASOOLY, R., JACOB, R. A., KADER, A. A. & MACKEY, B. E. 2006. Consumption of Bing sweet cherries lowers circulating concentrations of inflammation markers in healthy men and women. *The Journal of nutrition*, 136, 981-986.
- KELLY, J., FULFORD, J., VANHATALO, A., BLACKWELL, J. R., FRENCH, O., BAILEY, S. J., GILCHRIST, M., WINYARD, P. G. & JONES, A. M. 2013a. Effects of short-term dietary nitrate supplementation on blood pressure, O2 uptake kinetics, and muscle and cognitive function in older adults. *Am J Physiol Regul Integr Comp Physiol*, 304, 21.
- KELLY, J., VANHATALO, A., WILKERSON, D. P., WYLIE, L. J. & JONES, A. M. 2013b. Effects of nitrate on the power-duration relationship for severe-intensity exercise. *Med Sci Sports Exerc*, 45, 1798-806.
- KELM, M. 1999. Nitric oxide metabolism and breakdown. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1411, 273-289.

- KENJALE, A. A., HAM, K. L., STABLER, T., ROBBINS, J. L., JOHNSON, J. L., VANBRUGGEN, M., PRIVETTE, G., YIM, E., KRAUS, W. E. & ALLEN, J. D. 2011. Dietary nitrate supplementation enhances exercise performance in peripheral arterial disease. *Journal of Applied Physiology,* 110, 1582-1591.
- KERLEY, C. P., CAHILL, K., BOLGER, K., MCGOWAN, A., BURKE, C., FAUL, J. & CORMICAN, L. 2015. Dietary nitrate supplementation in COPD: An acute, double-blind, randomized, placebocontrolled, crossover trial ☆. *Nitric Oxide*, 44, 105-111.
- KHARITONOV, V. G., SUNDQUIST, A. R. & SHARMA, V. S. 1995. Kinetics of nitrosation of thiols by nitric oxide in the presence of oxygen. *Journal of Biological Chemistry*, 270, 28158-28164.
- KIELER-JENSEN, N., RICKSTEN, S. E., STENQVIST, O., BERGH, C. H., LINDELÖV, B., WENNMALM, A., WAAGSTEIN, F. & LUNDIN, S. 1994. Inhaled nitric oxide in the evaluation of heart transplant candidates with elevated pulmonary vascular resistance. *The Journal of heart and lung transplantation*: the official publication of the International Society for Heart *Transplantation*, 13, 366-375.
- KIM-SHAPIRO, D. B., SCHECHTER, A. N. & GLADWIN, M. T. 2006. Unraveling the reactions of nitric oxide, nitrite, and hemoglobin in physiology and therapeutics. *Arteriosclerosis, Thrombosis, and Vascular Biology,* 26, 697-705.
- KIM, J.-K., MOORE, D. J., MAURER, D. G., KIM-SHAPIRO, D. B., BASU, S., FLANAGAN, M. P., SKULAS-RAY, A. C., KRIS-ETHERTON, P. & PROCTOR, D. N. 2014. Acute dietary nitrate supplementation does not augment submaximal forearm exercise hyperemia in healthy young men. *Applied Physiology, Nutrition, and Metabolism*, 40, 1-7.
- KIM, J. S., NAM, M. H., DO, Y. S., LEE, C. J., KIM, C. S., SIM, W. S., PARK, K. B., LEE, S. H., HAN, J. S. & SHIN, B. S. 2010. Efficacy of milrinone versus nitroglycerin in controlling pulmonary arterial hypertension induced by intravenous injections of absolute ethanol in anesthetized dogs. *Journal of Vascular and Interventional Radiology*, 21, 882-887.
- KINGWELL, B., A, FORMOSA, M., MUHLMANN, M., BRADLEY, S., J & MCCONELL, G., K 2002. Nitric Oxide Synthase Inhibition Reduces Glucose Uptake During Exercise in Individuals With Type 2 Diabtes More Than in Control Subjects *Diabetes*, 51, 2572-2580.
- KIRKHAM, P. & RAHMAN, I. 2006. Oxidative stress in asthma and COPD: Antioxidants as a therapeutic strategy. *Pharmacology & Company*, *Therapeutics*, 111, 476-494.
- KLEIN, R., KLEIN, B. E., MOSS, S. E., DAVIS, M. D. & DEMETS, D. L. 1984. The Wisconsin epidemiologic study of diabetic retinopathy: III. Prevalence and risk of diabetic retinopathy when age at diagnosis is 30 or more years. *Archives of Ophthalmology*, 102, 527.
- KLEINBONGARD, P., DEJAM, A., LAUER, T., JAX, T., KERBER, S., GHARINI, P., BALZER, J., ZOTZ, R. B., SCHARF, R. E., WILLERS, R., SCHECHTER, A. N., FEELISCH, M. & KELM, M. 2006. Plasma nitrite concentrations reflect the degree of endothelial dysfunction in humans. *Free Radical Biology and Medicine*, 40, 295-302.
- KLEWICKA, E., NOWAK, A., ZDUNCZYK, Z., JUSKIEWICZ, J. & CUKROWSKA, B. 2012. Protective effect of lactofermented red beetroot juice against aberrant crypt foci formation, genotoxicity of fecal water and oxidative stress induced by 2-amino-1-methyl-6-phenylimidazo[4,5-b] pyridine in rats model. *Environ Toxicol Pharmacol*, 34, 895-904.
- KOCOLOSKI, G. & CRECELIUS, A. 2015. Effects of single-dose dietary nitrate on oxygen consumption during and after prolonged submaximal exercise in healthy humans. *The FASEB Journal*, 29, 675.15.
- KOELLING, T. M., KIRMSE, M., DI SALVO, T. G., DEC, G. W., ZAPOL, W. M. & SEMIGRAN, M. J. 1998. Inhaled nitric oxide improves exercise capacity in patients with severe heart failure and right ventricular dysfunction. *The American Journal of Cardiology*, 81, 1494-1497.
- KOLTERMAN, O. G., INSEL, J., SAEKOW, M. & OLEFSKY, J. M. 1980. Mechanisms of insulin resistance in human obesity: evidence for receptor and postreceptor defects. *Journal of Clinical Investigation*, 65, 1272-1284.

- KOUYOUMDJIAN, C., ADNOT, S., LEVAME, M., EDDAHIBI, S., BOUSBAA, H. & RAFFESTIN, B. 1994. Continuous inhalation of nitric oxide protects against development of pulmonary hypertension in chronically hypoxic rats. *The Journal of clinical investigation*, 94, 578-584.
- KOWALTOWSKI, A. J. & VERCESI, A. E. 1999. Mitochondrial damage induced by conditions of oxidative stress. *Free Radical Biology and Medicine*, 26, 463-471.
- KOZLOV, A. V., DIETRICH, B. & NOHL, H. 2003. Various intracellular compartments cooperate in the release of nitric oxide from glycerol trinitrate in liver. *British Journal of Pharmacology,* 139, 989-997.
- KOZLOV, A. V., STANIEK, K. & NOHL, H. 1999. Nitrite reductase activity is a novel function of mammalian mitochondria. *FEBS Letters*, 454, 127-130.
- KRAJKA-KUZNIAK, V., SZAEFER, H., IGNATOWICZ, E., ADAMSKA, T. & BAER-DUBOWSKA, W. 2012.

  Beetroot juice protects against N-nitrosodiethylamine-induced liver injury in rats. *Food Chem Toxicol*, 50, 2027-33.
- KRAUSE, M., RODRIGUES-KRAUSE, J., O'HAGAN, C., DE VITO, G., BOREHAM, C., SUSTA, D., NEWSHOLME, P. & MURPHY, C. 2012. Differential nitric oxide levels in the blood and skeletal muscle of type 2 diabetic subjects may be consequence of adiposity: a preliminary study. *Metabolism*, 61, 1528-1537.
- KROLEWSKI, A. S., LAFFEL, L. M. B., KROLEWSKI, M., QUINN, M. & WARRAM, J. H. 1995. Glycosylated Hemoglobin and the Risk of Microalbuminuria in Patients with Insulin-Dependent Diabetes Mellitus. *New England Journal of Medicine*, 332, 1251-1255.
- KUNDU, T. K., VELAYUTHAM, M. & ZWEIER, J. L. 2012. Aldehyde Oxidase Functions as a Superoxide Generating NADH Oxidase: An Important Redox Regulated Pathway of Cellular Oxygen Radical Formation. *Biochemistry*, 51, 2930-2939.
- LAAKSO, M., VOUTILAINEN, E., SARLUND, H., ARO, A., PYÖRÄLÄ, K. & PENTTILÄ, I. 1985. Serum lipids and lipoproteins in middle-aged non-insulin-dependent diabetics. *Atherosclerosis*, 56, 271-281.
- LAJER, M., TARNOW, L., JORSAL, A., TEERLINK, T., PARVING, H.-H. & ROSSING, P. 2008. Plasma concentration of asymmetric dimethylarginine (ADMA) predicts cardiovascular morbidity and mortality in type 1 diabetic patients with diabetic nephropathy. *Diabetes Care*, 31, 747-752
- LAMARRA, N., WHIPP, B. J., WARD, S. A. & WASSERMAN, K. 1987. Effect of interbreath fluctuations on characterizing exercise gas exchange kinetics. *J Appl Physiol (1985)*, 62, 2003-12.
- LANCASTER, J. R. 1994. Simulation of the diffusion and reaction of endogenously produced nitric oxide. *Proceedings of the National Academy of Sciences*, 91, 8137-8141.
- LANCASTER, J. R., JR. 1996. Diffusion of free nitric oxide. Methods Enzymol, 268, 31-50.
- LANDMESSER, U., BAHLMANN, F., MUELLER, M., SPIEKERMANN, S., KIRCHHOFF, N., SCHULZ, S., MANES, C., FISCHER, D., DE GROOT, K. & FLISER, D. 2005. Simvastatin versus ezetimibe pleiotropic and lipid-lowering effects on endothelial function in humans. *Circulation*, 111, 2356-2363.
- LANE, S. C., HAWLEY, J. A., DESBROW, B., JONES, A. M., BLACKWELL, J. R., ROSS, M. L., ZEMSKI, A. J. & BURKE, L. M. 2013. Single and combined effects of beetroot juice and caffeine supplementation on cycling time trial performance 1. *Applied Physiology, Nutrition, and Metabolism*, 39, 1-8.
- LANGEN, R. C. J., KORN, S. H. & WOUTERS, E. F. M. 2003. ROS in the local and systemic pathogenesis of COPD. *Free Radical Biology and Medicine*, 35, 226-235.
- LANSLEY, K., E, WINYARD, P., FULFORD, J., VANHATALO, A., BAILEY, J., B, BLACKWELL, J., R, DIMENNA, F., J, GILCHRIST, M., BENJAMIN, N. & JONES, A., M 2010. Dietary nitrate supplementation reduces the O<sub>2</sub> cost of walking and running: a placebo-controlled study. *Journal of Applied Physiology*, 110, 591-600.

- LANSLEY, K. E., WINYARD, P. G., BAILEY, S. J., VANHATALO, A., WILKERSON, D. P., BLACKWELL, J. R., GILCHRIST, M., BENJAMIN, N. & JONES, A. M. 2011a. Acute dietary nitrate supplementation improves cycling time trial performance. *Med Sci Sports Exerc*, 43, 1125-1131.
- LANSLEY, K. E., WINYARD, P. G., FULFORD, J., VANHATALO, A., BAILEY, S. J., BLACKWELL, J. R., DIMENNA, F. J., GILCHRIST, M., BENJAMIN, N. & JONES, A. M. 2011b. Dietary nitrate supplementation reduces the O2 cost of walking and running: a placebo-controlled study. *Journal of Applied Physiology*, 110, 591-600.
- LARSEN, F., WEITZBERG, E., LUNDBERG, J. & EKBLOM, B. 2007a. Effects of dietary nitrate on oxygen cost during exercise. *Acta Physiologica*, 191, 59-66.
- LARSEN, F. J., EKBLOM, B., SAHLIN, K., LUNDBERG, J. O. & WEITZBERG, E. 2006. Effects of Dietary Nitrate on Blood Pressure in Healthy Volunteers. *New England Journal of Medicine*, 355, 2792-2793.
- LARSEN, F. J., SCHIFFER, T. A., BORNIQUEL, S., SAHLIN, K., EKBLOM, B., LUNDBERG, J. O. & WEITZBERG, E. 2011. Dietary Inorganic Nitrate Improves Mitochondrial Efficiency in Humans. *Cell Metabolism*, 13, 149-159.
- LARSEN, F. J., SCHIFFER, T. A., EKBLOM, B., MATTSSON, M. P., CHECA, A., WHEELOCK, C. E., NYSTRÖM, T., LUNDBERG, J. O. & WEITZBERG, E. 2014. Dietary nitrate reduces resting metabolic rate: a randomized, crossover study in humans. *The American Journal of Clinical Nutrition*.
- LARSEN, F. J., WEITZBERG, E., LUNDBERG, J. O. & EKBLOM, B. 2007b. Effects of dietary nitrate on oxygen cost during exercise. *Acta Physiologica*, 191, 59-66.
- LARSEN, F. J., WEITZBERG, E., LUNDBERG, J. O. & EKBLOM, B. 2010. Dietary nitrate reduces maximal oxygen consumption while maintaining work performance in maximal exercise. *Free Radical Biology and Medicine*, 48, 342-347.
- LARSSON, L., ÖRLANDER, J., ANSVED, T. & EDSTRÖM, L. 1998. Effects of chronic nicotine exposure on contractile enzyme-histochemical and biochemical properties of fast- and slow-twitch skeletal muscles in the rat. *Acta Physiologica Scandinavica*, 134, 519-527.
- LAUER, T., HEISS, C., BALZER, J., KEHMEIER, E., MANGOLD, S., LEYENDECKER, T., ROTTLER, J., MEYER, C., MERX, M., KELM, M. & RASSAF, T. 2008. Age-dependent endothelial dysfunction is associated with failure to increase plasma nitrite in response to exercise. *Basic Research in Cardiology*, 103, 291-297.
- LAUTT, W. 1980. Control of hepatic arterial blood flow: independence from liver metabolic activity. *The American journal of physiology,* 239, H559-H564.
- LAUTT, W. W. 1977. The hepatic artery: subservient to hepatic metabolism or guardian of normal hepatic clearance rates of humoral substances. *General Pharmacology: The Vascular System,* 8, 73-78.
- LAUTT, W. W., LEGARE, D. J. & EZZAT, W. R. 1990. Quantitation of the hepatic arterial buffer response to graded changes in portal blood flow. *Gastroenterology*, 98, 1024-1028.
- LEBOVITZ, H. E. 1999. Type 2 Diabetes: An Overview. Clinical Chemistry, 45, 1339-1345.
- LEE, Y., HIROSE, H., OHNEDA, M., JOHNSON, J., MCGARRY, J. D. & UNGER, R. H. 1994. Beta-cell lipotoxicity in the pathogenesis of non-insulin-dependent diabetes mellitus of obese rats: impairment in adipocyte-beta-cell relationships. *Proceedings of the National Academy of Sciences*, 91, 10878-10882.
- LEGARE, D. J. & LAUTT, W. W. 1987. Hepatic venous resistance site in the dog: localization and validation of intrahepatic pressure measurements. *Canadian journal of physiology and pharmacology*, 65, 352-359.
- LEVITT, E. L., KEEN, J. T. & WONG, B. J. 2015. Augmented reflex cutaneous vasodilatation following short-term dietary nitrate supplementation in humans. *Experimental physiology*.
- LI, H., CUI, H., KUNDU, T. K., ALZAWAHRA, W. & ZWEIER, J. L. 2008a. Nitric Oxide Production from Nitrite Occurs Primarily in Tissues Not in the Blood CRITICAL ROLE OF XANTHINE OXIDASE AND ALDEHYDE OXIDASE. *Journal of Biological Chemistry*, 283, 17855-17863.

- LI, H., CUI, H., KUNDU, T. K., ALZAWAHRA, W. & ZWEIER, J. L. 2008b. Nitric Oxide Production from Nitrite Occurs Primarily in Tissues Not in the Blood: CRITICAL ROLE OF XANTHINE OXIDASE AND ALDEHYDE OXIDASE. *Journal of Biological Chemistry*, 283, 17855-17863.
- LI, H. & FORSTERMANN, U. 2013. Uncoupling of endothelial NO synthase in atherosclerosis and vascular disease. *Curr Opin Pharmacol*, 13, 161-7.
- LI, H., HEMANN, C., ABDELGHANY, T. M., EL-MAHDY, M. A. & ZWEIER, J. L. 2012. Characterization of the Mechanism and Magnitude of Cytoglobin-mediated Nitrite Reduction and Nitric Oxide Generation under Anaerobic Conditions. *Journal of Biological Chemistry*, 287, 36623-36633.
- LI, H., WALLERATH, T., MÜNZEL, T. & FÖRSTERMANN, U. 2002. Regulation of endothelial-type NO synthase expression in pathophysiology and in response to drugs. *Nitric Oxide*, **7**, 149-164.
- LI, J., HU, X., SELVAKUMAR, P., RUSSELL, R. R., CUSHMAN, S. W., HOLMAN, G. D. & YOUNG, L. H. 2004. Role of the nitric oxide pathway in AMPK-mediated glucose uptake and GLUT4 translocation in heart muscle. *American Journal of Physiology-Endocrinology and Metabolism*, 287, E834-E841.
- LI, Q., SPENCER, N. Y., OAKLEY, F. D., BUETTNER, G. R. & ENGELHARDT, J. F. 2009. Endosomal Nox2 facilitates redox-dependent induction of NF-kappaB by TNF-alpha. *Antioxidants & redox signaling*, 11, 1249-1263.
- LIN, K. Y., ITO, A., ASAGAMI, T., TSAO, P. S., ADIMOOLAM, S., KIMOTO, M., TSUJI, H., REAVEN, G. M. & COOKE, J. P. 2002. Impaired nitric oxide synthase pathway in diabetes mellitus role of asymmetric dimethylarginine and dimethylarginine dimethylaminohydrolase. *Circulation*, 106, 987-992.
- LOHMANN, S. M., VAANDRAGER, A. B., SMOLENSKI, A., WALTER, U. & DE JONGE, H. R. 1997. Distinct and specific functions of cGMP-dependent protein kinases. *Trends in biochemical sciences*, 22, 307-312.
- LU, T.-M., CHUNG, M.-Y., LIN, M.-W., HSU, C.-P. & LIN, S.-J. 2011. Plasma asymmetric dimethylarginine predicts death and major adverse cardiovascular events in individuals referred for coronary angiography. *International journal of cardiology*, 153, 135-140.
- LU, T.-M., DING, Y.-A., LIN, S.-J., LEE, W.-S. & TAI, H.-C. 2003a. Plasma levels of asymmetrical dimethylarginine and adverse cardiovascular events after percutaneous coronary intervention. *European Heart Journal*, 24, 1912-1919.
- LU, W., RESNICK, H. E., JABLONSKI, K. A., JONES, K. L., JAIN, A. K., HOWARD, W. J., ROBBINS, D. C. & HOWARD, B. V. 2003b. Non-HDL cholesterol as a predictor of cardiovascular disease in type 2 diabetes the strong heart study. *Diabetes care*, 26, 16-23.
- LUCAS, K. A., PITARI, G. M., KAZEROUNIAN, S., RUIZ-STEWART, I., PARK, J., SCHULZ, S., CHEPENIK, K. P. & WALDMAN, S. A. 2000. Guanylyl Cyclases and Signaling by Cyclic GMP. *Pharmacological Reviews*, 52, 375-414.
- LULICH, K. M., GOLDIE, R. G. & PATERSON, J. W. 1988. Beta-adrenoceptor function in asthmatic bronchial smooth muscle. *Gen Pharmacol*, 19, 307-11.
- LUNDBERG, J., O, FEELISCH, M., BJORNE, H., JANSSON, E., A & WEITZBERG, E. 2006. Cardioprotective effects of vegetables: Is nitrate the answer? . *Nitric Oxide*, 15, 359-362.
- LUNDBERG, J., O, WEITZBERG, E., LUNDBERG, J., M & ALVING, K. 1994. Intragastric nitric oxide production in humans: measurements in expelled air. *British Medical Journal* 35, 1543-1546.
- LUNDBERG, J., WEITZBERG, E. & GLADWIN, M., T 2008. The nitrate-nitrite-nitric oxide pathway in physiology and theraputics. *Nature Reviews Drug Discovery* **7**, 156-167.
- LUNDBERG, J. O. & GOVONI, M. 2004. Inorganic nitrate is a possible source for systemic generation of nitric oxide. *Free Radical Biology and Medicine*, 37, 395-400.
- LUNDBERG, J. O. & WEITZBERG, E. 2010. NO-synthase independent NO generation in mammals. *Biochemical and Biophysical Research Communications*, 396, 39-45.
- LUNDBERG, J. O., WEITZBERG, E., COLE, J. A. & BENJAMIN, N. 2004. Nitrate, bacteria and human health. *Nat Rev Microbiol*, 2, 593-602.

- LYONS, D., ROY, S., PATEL, M., BENJAMIN, N. & SWIFT, C. G. 1997. Impaired nitric oxide-mediated vasodilatation and total body nitric oxide production in healthy old age. *Clinical Science*, 93, 519-525.
- MAEJIMA, K., NAKANO, S., HIMENO, M., TSUDA, S.-I., MAKIISHI, H., ITO, T., NAKAGAWA, A., KIGOSHI, T., ISHIBASHI, T. & NISHIO, M. 2001. Increased basal levels of plasma nitric oxide in type 2 diabetic subjects: relationship to microvascular complications. *Journal of Diabetes and its Complications*, 15, 135-143.
- MAGEE, P. & BARNES, J. 1956. The Production of Malignant Primary Hepatic Tumours in the Rat by Feeding Dimethylnitrosamine. *Br J Cancer*, 10

## 114-122.

- MAHLER, D. A. & MACKOWIAK, J. I. 1995. Evaluation of the short-form 36-item questionnaire to measure health-related quality of life in patients with COPD. *CHEST Journal*, 107, 1585-1589.
- MALTAIS, F., DECRAMER, M., CASABURI, R., BARREIRO, E., BURELLE, Y., DEBIGARE, R., DEKHUIJZEN, P. N., FRANSSEN, F., GAYAN-RAMIREZ, G., GEA, J., GOSKER, H. R., GOSSELINK, R., HAYOT, M., HUSSAIN, S. N., JANSSENS, W., POLKEY, M. I., ROCA, J., SAEY, D., SCHOLS, A. M., SPRUIT, M. A., STEINER, M., TAIVASSALO, T., TROOSTERS, T., VOGIATZIS, I. & WAGNER, P. D. 2014. An official American Thoracic Society/European Respiratory Society statement: update on limb muscle dysfunction in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*, 189, 201402-0373ST.
- MARIN, J., CARRIZO, S., GASCON, M., SANCHEZ, A., GALLEGO, B. & CELLI, B. 2001. Inspiratory Capacity, Dynamic Hyperinflation, Breathlessness, and Exercise Performance during the 6-Minute-Walk Test in Chronic Obstructive Pulmonary Disease. *American Journal of Respiratory and Critical Care Medicine*, 163, 1395-1399.
- MÅRIN, P., ANDERSSON, B., KROTKIEWSKI, M. & BJÖRNTORP, P. 1994. Muscle Fiber Composition and Capillary Density in Women and Men With NIDDM. *Diabetes Care*, 17, 382-386.
- MARSH, P. D. & PERCIVAL, R. S. 2006. The oral microflora friend or foe? Can we decide? *International Dental Journal*, 56, 233-239.
- MARSHALL, B., HANSON, C., FRASCH, F. & MARSHALL, C. 1994. Role of hypoxic pulmonary vasoconstriction in pulmonary gas exchange and blood flow distribution. *Intensive care medicine*, 20, 379-389.
- MARTIN, C. L., ALBERS, J., HERMAN, W. H., CLEARY, P., WABERSKI, B., GREENE, D. A., STEVENS, M. J. & FELDMAN, E. L. 2006. Neuropathy among the diabetes control and complications trial cohort 8 years after trial completion. *Diabetes Care*, 29, 340-4.
- MARTIN, H. M., MOORE, K. P., BOSMANS, E., DAVIES, S., BURROUGHS, A. K., DHILLON, A. P., TOSH, D. & HARRISON, R. 2004. Xanthine oxidoreductase is present in bile ducts of normal and cirrhotic liver. *Free Radical Biology and Medicine*, 37, 1214-1223.
- MARTIN, K., SMEE, D., THOMPSON, K. G. & RATTRAY, B. 2014. No improvement of repeated-sprint performance with dietary nitrate. *IJSPP*, 9.
- MASSCHELEIN, E., VAN THIENEN, R., WANG, X., VAN SCHEPDAEL, A., THOMIS, M. & HESPEL, P. 2012. Dietary nitrate improves muscle but not cerebral oxygenation status during exercise in hypoxia. *J Appl Physiol*, 113, 736-45.
- MATHERS, C. D. & LONCAR, D. 2006. Projections of global mortality and burden of disease from 2002 to 2030. *PLoS medicine*, 3, e442.
- MATHIEU-COSTELLO, O., JU, Y., TREJO-MORALES, M. & CUI, L. 2005. Greater capillary-fiber interface per fiber mitochondrial volume in skeletal muscles of old rats. *Journal of Applied Physiology*, 99, 281-289.
- MATHIEU-COSTELLO, O., KONG, A., CIARALDI, T. P., CUI, L., JU, Y., CHU, N., KIM, D., MUDALIAR, S. & HENRY, R. R. 2003. Regulation of skeletal muscle morphology in type 2 diabetic subjects by troglitazone and metformin: relationship to glucose disposal. *Metabolism*, 52, 540-546.
- MATTSSON, M., JOHANSSON, E., LUNDBORG, T., LARSSON, M. & LARSSON, C. M. 1991. Nitrogen Utilization in N-limited Barley during Vegetative and Generative Growth: I. GROWTH AND

- NITRATE UPTAKE KINETICS IN VEGETATIVE CULTURES GROWN AT DIFFERENT RELATIVE ADDITION RATES OF NITRATE-N. *Journal of Experimental Botany*, 42, 197-205.
- MCCANCE, D. R., HANSON, R. L., CHARLES, M.-A., JACOBSSON, L., PETTITT, D. J., BENNETT, P. H. & KNOWLER, W. C. 1994. Comparison of tests for glycated haemoglobin and fasting and two hour plasma glucose concentrations as diagnostic methods for diabetes. *BMJ: British Medical Journal*, 308, 1323.
- MCCLINTOCK, D. E., WARE, L. B., EISNER, M. D., WICKERSHAM, N., THOMPSON, B. T., MATTHAY, M. A., THE NATIONAL HEART, L. & NETWORK, B. I. A. 2007. Higher Urine Nitric Oxide Is Associated with Improved Outcomes in Patients with Acute Lung Injury. *American Journal of Respiratory and Critical Care Medicine*, 175, 256-262.
- MCKNIGHT, G., SMITH, L., DRUMMOND, R., DUNCAN, C., GOLDEN, M. & BENJAMIN, N. 1997. Chemical synthesis of nitric oxide in the stomach from dietary nitrate in humans. *Gut*, 40, 211-214.
- MCMAHON, T. J. & STAMLER, J. S. 1999. Concerted nitric oxide/oxygen delivery by hemoglobin. *Methods Enzymol*, 301, 99-114.
- MCNEILL, E. & CHANNON, K. M. 2012. The role of tetrahydrobiopterin in inflammation and cardiovascular. *Thromb Haemost*, 108, 832-839.
- MEIER, J. J., GETHMANN, A., GOTZE, O., GALLWITZ, B., HOLST, J. J., SCHMIDT, W. E. & NAUCK, M. A. 2006. Glucagon-like peptide 1 abolishes the postprandial rise in triglyceride concentrations and lowers levels of non-esterified fatty acids in humans. *Diabetologia*, 49, 452-8.
- MEININGER, C., MARINOS, R., HATAKEYAMA, K., MARTINEZ-ZAGUILAN, R., ROJAS, J., KELLY, K. & WU, G. 2000. Impaired nitric oxide production in coronary endothelial cells of the spontaneously diabetic BB rat is due to tetrahydrobiopterin deficiency. *Biochem. J*, 349, 353-356.
- MEINITZER, A., SEELHORST, U., WELLNITZ, B., HALWACHS-BAUMANN, G., BOEHM, B. O., WINKELMANN, B. R. & MÄRZ, W. 2007. Asymmetrical dimethylarginine independently predicts total and cardiovascular mortality in individuals with angiographic coronary artery disease (the Ludwigshafen Risk and Cardiovascular Health study). *Clinical Chemistry*, 53, 273-283.
- MERCER, B. A., KOLESNIKOVA, N., SONETT, J. & D'ARMIENTO, J. 2004. Extracellular regulated kinase/mitogen activated protein kinase is up-regulated in pulmonary emphysema and mediates matrix metalloproteinase-1 induction by cigarette smoke. *Journal of Biological Chemistry*, 279, 17690-17696.
- MERRY, T. L., STEINBERG, G. R., LYNCH, G. S. & MCCONELL, G. K. 2010. Skeletal muscle glucose uptake during contraction is regulated by nitric oxide and ROS independently of AMPK. American Journal of Physiology-Endocrinology and Metabolism, 298, E577-E585.
- MILLER, G. D., MARSH, A. P., DOVE, R. W., BEAVERS, D., PRESLEY, T., HELMS, C., BECHTOLD, E., KING, S. B. & KIM-SHAPIRO, D. 2012. Plasma nitrate and nitrite are increased by a high-nitrate supplement but not by high-nitrate foods in older adults. *Nutr Res*, 32, 160-8.
- MILLER, V. M. & BURNETT, J. C. 1992. Modulation of NO and endothelin by chronic increases in blood flow in canine femoral arteries. *American Journal of Physiology Heart and Circulatory Physiology*, 263, H103-H108.
- MILSOM, A. B., JONES, C. J., GOODFELLOW, J., FRENNEAUX, M. P., PETERS, J. R. & JAMES, P. E. 2002. Abnormal metabolic fate of nitric oxide in Type I diabetes mellitus. *Diabetologia*, 45, 1515-22.
- MOBBS, K., VAN SAENE, H., SUNDERLAND, D. & DAVIES, P. 1999. Oropharyngeal gram-negative bacillary carriage in chronic obstructive pulmonary disease: relation to severity of disease. *Respiratory medicine*, 93, 540-545.
- MODIN, A., BJORNE, H., HERULF, M., ALVING, K., WEITZBERG, E. & LUNDBERG, J. 2001. Nitritederived nitric oxide: a possible mediator of 'acidic-metabolic' vasodilation. *Acta Physiologica Scandinavica*, 171, 9-16.

- MOLLNAU, H., SCHULZ, E., DAIBER, A., BALDUS, S., OELZE, M., AUGUST, M., WENDT, M., WALTER, U., GEIGER, C. & AGRAWAL, R. 2003. Nebivolol prevents vascular NOS III uncoupling in experimental hyperlipidemia and inhibits NADPH oxidase activity in inflammatory cells. *Arteriosclerosis, Thrombosis, and Vascular Biology,* 23, 615-621.
- MONTI, L. D., BARLASSINA, C., CITTERIO, L., GALLUCCIO, E., BERZUINI, C., SETOLA, E., VALSECCHI, G., LUCOTTI, P., POZZA, G. & BERNARDINELLI, L. 2003. Endothelial nitric oxide synthase polymorphisms are associated with type 2 diabetes and the insulin resistance syndrome. *Diabetes*, 52, 1270-1275.
- MORGAN, C., M, TANOFSKY-KRAFT, M., WILFREY, D., E & YANOVSKI, J., A 2002. Childhood Obesity. *Child and Adolescent Psychiatric Clinics of North America*, 11, 257.
- MRC 1965. Definition and classification of chronic bronchitis for clinical and epidemiological purposes. A report to the Medical Research Council by their Committee on the Aetiology of Chronic Bronchitis. *Lancet*, 1, 775-9.
- MUGGERIDGE, D. J., HOWE, C. C., SPENDIFF, O., PEDLAR, C., JAMES, P. E. & EASTON, C. 2013a. The Effects of a Single Dose of Concentrated Beetroot Juice on Performance in Trained Flatwater Kayakers. *Int J Sport Nutr Exerc Metab*, 9, 9.
- MUGGERIDGE, D. J., HOWE, C. C., SPENDIFF, O., PEDLAR, C., JAMES, P. E. & EASTON, C. 2013b. A Single Dose of Beetroot Juice Enhances Cycling Performance in Simulated Altitude. *Med Sci Sports Exerc*, 10, 10.
- MUGGERIDGE, D. J., SCULTHORPE, N., GRACE, F. M., WILLIS, G., THORNHILL, L., WELLER, R. B., JAMES, P. E. & EASTON, C. 2014. Acute whole body UVA irradiation combined with nitrate ingestion enhances time trial performance in trained cyclists. *Nitric Oxide*.
- MÜNZEL, T. & GORI, T. 2009. NebivololThe Somewhat-Different β-Adrenergic Receptor Blocker. *Journal of the American College of Cardiology,* 54, 1491-1499.
- MURPHY, S., WITMER, C. A., DAVIS, S. E. & SAUERS, E. J. Effects of Beetroot Juice Supplementation on Performance during a Repeated-Sprint Rest in Active Males. International Journal of Exercise Science: Conference Proceedings, 2014. 61.
- MYERS, J., PRAKASH, M., FROELICHER, V., DO, D., PARTINGTON, S. & ATWOOD, J. E. 2002. Exercise Capacity and Mortality among Men Referred for Exercise Testing. *New England Journal of Medicine*, 346, 793-801.
- NAUCK, M. A., VARDARLI, I., DEACON, C. F., HOLST, J. J. & MEIER, J. J. 2011. Secretion of glucagon-like peptide-1 (GLP-1) in type 2 diabetes: what is up, what is down? *Diabetologia*, 54, 10-8.
- NDDG, N. D. D. G. 1979. Classification and diagnosis of diabetes mellitus and other categories of glucose intolerance. *Diabetes*, 28, 1039-1057.
- NELSON, R. G., BENNETT, P. H., BECK, G. J., TAN, M., KNOWLER, W. C., MITCH, W. E., HIRSCHMAN, G. H. & MYERS, B. D. 1996. Development and Progression of Renal Disease in Pima Indians with Non-Insulin-Dependent Diabetes Mellitus. *New England Journal of Medicine*, 335, 1636-1642.
- NEUGEBAUER, S., BABA, T. & WATANABE, T. 2000. Association of the nitric oxide synthase gene polymorphism with an increased risk for progression to diabetic nephropathy in type 2 diabetes. *Diabetes*, 49, 500-503.
- NEWBERNE, P. M. 1979. Nitrite promotes lymphoma incidence in rats. *Science (New York, N.Y.)*, 204, 1079-1081.
- NICE 2008. Type 2 diabetes: The management of type 2 diabetes. NICE Clinical Guidlines 66.
- NICI, L., DONNER, C., WOUTERS, E., ZUWALLACK, R., AMBROSINO, N., BOURBEAU, J., CARONE, M., CELLI, B., ENGELEN, M., FAHY, B., GARVEY, C., GOLDSTEIN, R., GOSSELINK, R., LAREAU, S., MACINTYRE, N., MALTAIS, F., MORGAN, M., O'DONNELL, D., PREFAULT, C., REARDON, J., ROCHESTER, C., SCHOLS, A., SINGH, S. & TROOSTERS, T. 2006. American Thoracic Society/European Respiratory Society statement on pulmonary rehabilitation. *Am J Respir Crit Care Med*, 173, 1390-413.

- NIKOLAIDIS, L. A., ELAHI, D., HENTOSZ, T., DOVERSPIKE, A., HUERBIN, R., ZOURELIAS, L., STOLARSKI, C., SHEN, Y. T. & SHANNON, R. P. 2004a. Recombinant glucagon-like peptide-1 increases myocardial glucose uptake and improves left ventricular performance in conscious dogs with pacing-induced dilated cardiomyopathy. *Circulation*, 110, 955-61.
- NIKOLAIDIS, L. A., MANKAD, S., SOKOS, G. G., MISKE, G., SHAH, A., ELAHI, D. & SHANNON, R. P. 2004b. Effects of glucagon-like peptide-1 in patients with acute myocardial infarction and left ventricular dysfunction after successful reperfusion. *Circulation*, 109, 962-965.
- NOLAN, C. J., DAMM, P. & PRENTKI, M. 2011. Type 2 diabetes across generations: from pathophysiology to prevention and management. *The Lancet*, 378, 169-181.
- NORDLIE, R. C. & ARION, W. J. 1964. Evidence for the Common Identity of Glucose 6-Phosphatase, Inorganic Pyrophosphatase, and Pyrophosphate- Glucose Phosphotransferase. *Journal of Biological Chemistry*, 239, 1680-1685.
- NYSTROM, T., GUTNIAK, M. K., ZHANG, Q., ZHANG, F., HOLST, J. J., AHREN, B. & SJOHOLM, A. 2004. Effects of glucagon-like peptide-1 on endothelial function in type 2 diabetes patients with stable coronary artery disease. *Am J Physiol Endocrinol Metab*, 287, 7.
- NYSTROM, T., ORTSATER, H., HUANG, Z., ZHANG, F., LARSEN, F. J., WEITZBERG, E., LUNDBERG, J. O. & SJOHOLM, A. 2012. Inorganic nitrite stimulates pancreatic islet blood flow and insulin secretion. *Free Radic Biol Med*, 53, 1017-23.
- O'CONNOR, E., GREEN, S., KIELY, C., O'SHEA, D. & EGAÑA, M. 2015. Differential effects of age and type 2 diabetes on dynamic versus peak response of pulmonary oxygen uptake during exercise.
- O'DONNELL, D., LAM, M. & WEBB, K. 1998. Measurement of Symptoms, Lung Hyperinflation, and Endurance during Exercise in Chronic Obstructive Pulmonary Disease. *American Journal of Respiratory and Critical Care Medicine*, 158, 1557-1565.
- O'DONNELL, D. E. & LAVENEZIANA, P. 2007. Dyspnea and activity limitation in COPD: mechanical factors. *COPD: Journal of Chronic Obstructive Pulmonary Disease*, **4**, 225-236.
- OAK, J.-H. & CAI, H. 2007. Attenuation of angiotensin II signaling recouples eNOS and inhibits nonendothelial NOX activity in diabetic mice. *Diabetes*, 56, 118-126.
- OEPPEN, J. & VAUPEL, J. W. 2002. Broken limits to life expectancy. Science, 296, 1029-1031.
- OHKUBO, Y., KISHIKAWA, H., ARAKI, E., MIYATA, T., ISAMI, S., MOTOYOSHI, S., KOJIMA, Y., FURUYOSHI, N. & SHICHIRI, M. 1995. Intensive insulin therapy prevents the progression of diabetic microvascular complications in Japanese patients with non-insulin-dependent diabetes mellitus: a randomized prospective 6-year study. *Diabetes Research and Clinical Practice*, 28, 103-117.
- OMAR, S. A., ARTIME, E. & WEBB, A. J. 2012. A comparison of organic and inorganic nitrates/nitrites. *Nitric Oxide*, 26, 229-240.
- OMAR, S. A. & WEBB, A. J. 2014. Nitrite reduction and cardiovascular protection. *Journal of Molecular and Cellular Cardiology*, 73, 57-69.
- OPPERMANN, M., BALZ, V., ADAMS, V., THAO-VI DAO, V., BAS, M., SUVORAVA, T. & KOJDA, G. 2009. Pharmacological induction of vascular extracellular superoxide dismutase expression in vivo. *Journal of Cellular and Molecular Medicine*, 13, 1271-1278.
- OSWALD-MAMMOSSER, M., APPRILL, M., BACHEZ, P., EHRHART, M. & WEITZENBLUM, E. 1991. Pulmonary hemodynamics in chronic obstructive pulmonary disease of the emphysematous type. *Respiration; international review of thoracic diseases*, 58, 304-310.
- OSWALD-MAMMOSSER, M., WEITZENBLUM, E., QUOIX, E., MOSER, G., CHAOUAT, A., CHARPENTIER, C. & KESSLER, R. 1995. Prognostic Factors in COPD Patients Receiving Long-term Oxygen Therapy Importance of Pulmonary Artery Pressure. *CHEST Journal*, 107, 1193-1198.
- ÖZYAZGAN, S., KUTLUATA, N., FER, U., AFŞAR, S., ÖZDAŞ, S., AKKAN, A. G. & KHAN, O. 2005. Effect of glucagon-like peptide-1 (7–36) and exendin-4 on the vascular reactivity in streptozotocin/nicotinamide-induced diabetic rats. *Pharmacology*, 74, 119-126.

- PADILLA, D. J., MCDONOUGH, P., BEHNKE, B. J., KANO, Y., HAGEMAN, K. S., MUSCH, T. I. & POOLE, D. C. 2007. Effects of Type II diabetes on muscle microvascular oxygen pressures. *Respiratory Physiology & Description*, 156, 187-195.
- PALUMBO, P. J., ELVEBACK, L. R. & WHISNANT, J. P. 1978. Neurologic complications of diabetes mellitus: transient ischemic attack, stroke, and peripheral neuropathy. *Adv Neurol*, 19, 593-601.
- PARVING, H.-H. 1996. Initiation and progression of diabetic nephropathy. *The New England Journal of Medicine*, 335, 1682-1683.
- PARVING, H., H, LEHNERT, H., BRÖCHNER-MORTENSEN, J., GOMIS, R., ANDERSEN, S. & ARNER, P. 2001. The Effect of Irbesartan on the Development of Diabetic Nephropathy in Patients with Type 2 Diabetes. *New England Journal of Medicine*, 345, 870-878.
- PATE, E., BHIMANI, M., FRANKS-SKIBA, K. & COOKE, R. 1995. Reduced effect of pH on skinned rabbit psoas muscle mechanics at high temperatures: implications for fatigue. *J Physiol*, 486 ( Pt 3), 689-94.
- PATEL, A., MACMAHON, S., CHALMERS, J., NEAL, B., BILLOT, L., WOODWARD, M., MARRE, M., COOPER, M., GLASZIOU, P. & GROBBEE, D. 2008. Intensive blood glucose control and vascular outcomes in patients with type 2 diabetes.
- PATEL, V., RASSAM, S., NEWSOM, R., WIEK, J. & KOHNER, E. 1992. Retinal blood flow in diabetic retinopathy. *Bmj*, 305, 678-683.
- PAUWELS, R. A. 2000. National and International Guidelines for COPD\*The Need for Evidence. *CHEST Journal*, 117, 20S-22S.
- PAZAHR, S., NANZ, D., ROSSI, C., CHUCK, N., STENGER, I., WURNIG, M. C., SCHICK, F. & BOSS, A. 2014. Magnetic resonance imaging of the liver: apparent diffusion coefficients from multiexponential analysis of b values greater than 50 s/mm2 do not respond to caloric intake despite increased portal-venous blood flow. *Invest Radiol*, 49, 138-46.
- PEELING, P., COX, G., BULLOCK, N. & BURKE, L. 2014. Beetroot Juice Improves on-Water 500 m Time-Trial Performance, and Laboratory-Based Paddling Economy in National and International-Level Kayak Athletes. *International journal of sport nutrition and exercise metabolism*.
- PELLETIER, M. M., KLEINBONGARD, P., RINGWOOD, L., HITO, R., HUNTER, C. J., SCHECHTER, A. N., GLADWIN, M. T. & DEJAM, A. 2006. The measurement of blood and plasma nitrite by chemiluminescence: pitfalls and solutions. *Free Radical Biology and Medicine*, 41, 541-548.
- PERCIVAL, R., MARSH, P. & CHALLACOMBE, S. 1997. Age-related changes in salivary antibodies to commensal oral and gut biota. *Oral microbiology and immunology,* 12, 57-63.
- PERCIVAL, R. S., MARSH, P. D. & CHALLACOMBE, S. J. 1996. Serum antibodies to commensal oral and gut bacteria vary with age. *FEMS Immunology & Medical Microbiology,* 15, 35-42.
- PETERSSON, J., CARLSTRÖM, M., SCHREIBER, O., PHILLIPSON, M., CHRISTOFFERSSON, G., JÄGARE, A., ROOS, S., JANSSON, E. Å., PERSSON, A. E. G., LUNDBERG, J. O. & HOLM, L. 2009. Gastroprotective and blood pressure lowering effects of dietary nitrate are abolished by an antiseptic mouthwash. *Free Radical Biology and Medicine*, 46, 1068-1075.
- PETERSSON, J., PHILLIPSON, M., JANSSON, E. Å., PATZAK, A., LUNDBERG, J. O. & HOLM, L. 2007. Dietary nitrate increases gastric mucosal blood flow and mucosal defense. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 292, G718-G724.
- PETTITT, D., LISSE, J., KNOWLER, W. & BENNETT, P. 1980. DEVELOPMENT OF RETINOPATHY AND PROTEINURIA IN RELATION TO PLASMA-GLUCOSE CONCENTRATIONS IN PIMA INDIANS. *The Lancet*, 316, 1050-1052.
- PHO, Y. A. H. 2010. APHO diabetes prevalence model for England [online].
- PICKERING, T. G. 1996. White coat hypertension. *Current opinion in nephrology and hypertension*, 5, 192-198.
- PINNA, M., ROBERTO, S., MILIA, R., MARONGIU, E., OLLA, S., LOI, A., MIGLIACCIO, G. M., PADULO, J., ORLANDI, C. & TOCCO, F. 2014. Effect of beetroot juice supplementation on aerobic response during swimming. *Nutrients*, 6, 605-615.

- PIRART, J. 1978. Diabetes Mellitus and Its Degenerative Complications: A Prospective Study of 4,400 Patients Observed Between 1947 and 1973. *Diabetes Care*, 1, 168-188.
- PLUTA, R. M., DEJAM, A., GRIMES, G., GLADWIN, M. T. & OLDFIELD, E. H. 2005. Nitrite infusions to prevent delayed cerebral vasospasm in a primate model of subarachnoid hemorrhage. *JAMA*, 293, 1477-1484.
- POIRIER, P., GARNEAU, C., BOGATY, P., NADEAU, A., MAROIS, L., BROCHU, C., GINGRAS, C., FORTIN, C., JOBIN, J. & DUMESNIL, J.-G. 2000. Impact of left ventricular diastolic dysfunction on maximal treadmill performance in normotensive subjects with well-controlled type 2 diabetes mellitus. *The American Journal of Cardiology*, 85, 473-477.
- PORCELLI, S., RAMAGLIA, M., BELLISTRI, G., PAVEI, G., PUGLIESE, L., MONTORSI, M., RASICA, L. & MARZORATI, M. 2014. Aerobic Fitness Affects the Exercise Performance Responses to Nitrate Supplementation. *Medicine and science in sports and exercise*.
- POST, S., SMITS, A. M., VAN DEN BROEK, A. J., SLUIJTER, J. P., HOEFER, I. E., JANSSEN, B. J., SNIJDER, R. J., MAGER, J. J., PASTERKAMP, G. & MUMMERY, C. L. 2010. Impaired recruitment of HHT-1 mononuclear cells to the ischaemic heart is due to an altered CXCR4/CD26 balance. *Cardiovascular Research*, 85, 494-502.
- POULSEN, P., GRUNNET, L. G., PILGAARD, K., STORGAARD, H., ALIBEGOVIC, A., SONNE, M. P., CARSTENSEN, B., BECK-NIELSEN, H. & VAAG, A. 2009. Increased Risk of Type 2 Diabetes in Elderly Twins. *Diabetes*, 58, 1350-1355.
- PRESLEY, T. D., MORGAN, A. R., BECHTOLD, E., CLODFELTER, W., DOVE, R. W., JENNINGS, J. M., KRAFT, R. A., BRUCE KING, S., LAURIENTI, P. J., JACK REJESKI, W., BURDETTE, J. H., KIM-SHAPIRO, D. B. & MILLER, G. D. 2011. Acute effect of a high nitrate diet on brain perfusion in older adults. *Nitric Oxide*, 24, 34-42.
- PUYPE, J., RAMAEKERS, M., VAN THIENEN, R., DELDICQUE, L. & HESPEL, P. 2014. No effect of dietary nitrate supplementation on endurance training in hypoxia. *Scandinavian journal of medicine & science in sports*.
- QUINT, J., HERRETT, E., BHASKARAN, K., TIMMIS, A., HEMINGWAY, H., WEDZICHA, J. & SMEETH, L. 2013. Effect of β blockers on mortality after myocardial infarction in adults with COPD: population based cohort study of UK electronic healthcare records. *BMJ: British Medical Journal*, 347.
- RAMAN, C., LI, H., MARTÁSEK, P., KRÁL, V., MASTERS, B. S. S. & POULOS, T. L. 1998. Crystal structure of constitutive endothelial nitric oxide synthase: a paradigm for pterin function involving a novel metal center. *Cell*, 95, 939-950.
- RAMANI, G. V. & PARK, M. H. 2010. Update on the clinical utility of sildenafil in the treatment of pulmonary arterial hypertension. *Drug design, development and therapy, 4*, 61-70.
- RAPPAPORT, A. 1979. Hepatic blood flow: morphologic aspects and physiologic regulation. International review of physiology, 21, 1-63.
- RASSAF, T., HEISS, C., MANGOLD, S., LEYENDECKER, T., KEHMEIER, E. S., KELM, M. & LAUER, T. 2010. Vascular Formation of Nitrite After Exercise Is Abolished in Patients With Cardiovascular Risk Factors and Coronary Artery Disease. *Journal of the American College of Cardiology,* 55, 1502-1503.
- RAVANI, P., TRIPEPI, G., MALBERTI, F., TESTA, S., MALLAMACI, F. & ZOCCALI, C. 2005. Asymmetrical dimethylarginine predicts progression to dialysis and death in patients with chronic kidney disease: a competing risks modeling approach. *Journal of the American Society of Nephrology,* 16, 2449-2455.
- RAVID, M., SAVIN, H., JUTRIN, I., BENTAL, T., KATZ, B. & LISHNER, M. 1993. Long-Term Stabilizing Effect of Angiotensin-Converting Enzyme Inhibition on Plasma Creatinine and on Proteinuria in Normotensive Type II Diabetic Patients. *Annals of Internal Medicine*, 118, 577-581.
- REEDER, B. J. 2010. The redox activity of hemoglobins: from physiologic functions to pathologic mechanisms. *Antioxidants & redox signaling*, 13, 1087-1123.

- REGENSTEINER, J., G, BAUER, T., A, REUSCH, J., E, B, BRANDENBURG, S., L, SIPPEL, J., M, VOGELSONG, A., M, SMITH, S., WOLFEL. E, E., ECKEL, R., H & HIATT. W, R. 1998. Abnormal, oxygen uptake kinetic responses in women with type II diabetes Mellitus. *American Physiology Society*, 85, 310-317.
- REGENSTEINER, J., G, BAUER, T., A, REUSCH, J., E, B, QUAIFE, R., A, CHEN, M., Y, SMITH, S., C, MILLER, T., M, GROVES, B., M & WOLFEL, E., E 2009. Cardiac dysfunction during exercise in uncomplicated type 2 diabetes. *Medicine and science in sports and exercise*, 41, 977.
- REGENSTEINER, J., G, SIPPEL, J., MCFARLING, E., T, WOLFEL, E., E & HIATT. W, R. 1995a. Effects of non-insulin-dependent diabetes on oxygen consumption during treadmill exercise. *Medicine & Science in Sports & Exercise*, 27, 661-667.
- REGENSTEINER, J. G., SIPPEL, J., MCFARLING, E. T., WOLFEL, E. E. & HIATT, W. R. 1995b. Effects of non-insulin-dependent diabetes on oxygen consumption during treadmill exercise. *Med Sci Sports Exerc*, 27, 661-7.
- REICHERT, E. T. 1880. On the Physiological Action of Potassium Nitrite. *The American Journal of the Medical Sciences*, 80, 158-180.
- REIF, A., FRÖHLICH, L. G., KOTSONIS, P., FREY, A., BÖMMEL, H. M., WINK, D. A., PFLEIDERER, W. & SCHMIDT, H. H. 1999. Tetrahydrobiopterin inhibits monomerization and is consumed during catalysis in neuronal NO synthase. *Journal of Biological Chemistry*, 274, 24921-24929.
- RICHARDS, J., RACINE, M., HEARON, C., PATEL, R. & DINENNO, F. 2014. Acute ingestion of dietary nitrate increases muscle blood flow via local vasodilation during exercise in young adults (705.9). *The FASEB Journal*, 28, 705.9.
- RITOV, V. B., MENSHIKOVA, E. V., HE, J., FERRELL, R. E., GOODPASTER, B. H. & KELLEY, D. E. 2005. Deficiency of Subsarcolemmal Mitochondria in Obesity and Type 2 Diabetes. *Diabetes*, 54, 8-14.
- ROCHA, B. S., GAGO, B., BARBOSA, R. M., CAVALEIRO, C. & LARANJINHA, J. 2015. Ethyl nitrite is produced in the human stomach from dietary nitrate and ethanol, releasing nitric oxide at physiological pH: potential impact on gastric motility. *Free Radical Biology and Medicine*, 82, 160-166.
- ROCKEY, D. C. & CHUNG, J. J. 1998. Reduced nitric oxide production by endothelial cells in cirrhotic rat liver: Endothelial dysfunction in portal hypertension. *Gastroenterology*, 114, 344-351.
- RODGERS, C. O. & BARNEIX, A. J. 1988. Cultivar differences in the rate of nitrate uptake by intact wheat plants as related to growth rate. *Physiologia Plantarum*, 72, 121-126.
- ROHLFING, C. L., WIEDMEYER, H.-M., LITTLE, R. R., ENGLAND, J. D., TENNILL, A. & GOLDSTEIN, D. E. 2002. Defining the relationship between plasma glucose and HbA1c analysis of glucose profiles and HbA1c in the Diabetes Control and Complications Trial. *Diabetes care*, 25, 275-278.
- ROUSE, I., L, BEILIN, L., J, ARMSTRONG, B., K & VANDONGEN, R. 1983. Blood-pressure-lowering effects of a vegetarian diet:: Controlled trial in normotensive subjects. *The Lancet* 1, 5-10.
- ROY, D., PERREAULT, M. & MARETTE, A. 1998. Insulin stimulation of glucose uptake in skeletal muscles and adipose tissues in vivo is NO dependent. *American Journal of Physiology-Endocrinology and Metabolism*, 274, E692-E699.
- ROY, K., BORRILL, Z. L., STARKEY, C., HAZEL, A. L., MORRIS, J., VESTBO, J. & SINGH, D. 2007. Use of different exhaled nitric oxide multiple flow rate models in COPD. *European Respiratory Journal*, 29, 651-659.
- ROY, K., SMITH, J., KOLSUM, U., BORRILL, Z., VESTBO, J. & SINGH, D. 2009. COPD phenotype description using principal components analysis. *Respiratory research*, 10, 41.
- SAID, G. 1981. [Progressive centripetal degeneration in polyneuropathies (author's transl)]. *Rev Neurol*, 137, 573-88.
- SAID, G. 2007. Diabetic neuropathy—a review. Nature Clinical Practice Neurology, 3, 331-340.
- SALLIS, J. F. 2000. Age-related decline in physical activity: a synthesis of human and animal studies.

- SALVI, S., S & BARNES, P., J 2009. Chronic obstructive pulmonary disease in non-smokers. *Lancet*, 29;374(9691), 733-43.
- SANDBAKK, S. B., SANDBAKK, Ø., PEACOCK, O., JAMES, P., WELDE, B., STOKES, K., BÖHLKE, N. & TJØNNA, A. E. 2014. Effects of Acute Supplementation of L-arginine and Nitrate on Endurance and Sprint Performance in Elite Athletes. *Nitric Oxide*.
- SANTILLI, F., CIPOLLONE, F., MEZZETTI, A. & CHIARELLI, F. 2004. The role of nitric oxide in the development of diabetic angiopathy. *Hormone and metabolic research*, 36, 319-335.
- SATOH, M., FUJIMOTO, S., ARAKAWA, S., YADA, T., NAMIKOSHI, T., HARUNA, Y., HORIKE, H., SASAKI, T. & KASHIHARA, N. 2008. Angiotensin II type 1 receptor blocker ameliorates uncoupled endothelial nitric oxide synthase in rats with experimental diabetic nephropathy. *Nephrology Dialysis Transplantation*, 23, 3806-3813.
- SAXENA, R., HIVERT, M.-F., LANGENBERG, C., TANAKA, T., PANKOW, J. S., VOLLENWEIDER, P., LYSSENKO, V., BOUATIA-NAJI, N., DUPUIS, J. & JACKSON, A. U. 2010. Genetic variation in GIPR influences the glucose and insulin responses to an oral glucose challenge. *Nature genetics*, 42, 142-148.
- SAXENA, R., VOIGHT, B. F., LYSSENKO, V., BURTT, N. P., DE BAKKER, P. I., CHEN, H., ROIX, J. J., KATHIRESAN, S., HIRSCHHORN, J. N., DALY, M. J., HUGHES, T. E., GROOP, L., ALTSHULER, D., ALMGREN, P., FLOREZ, J. C., MEYER, J., ARDLIE, K., BENGTSSON BOSTROM, K., ISOMAA, B., LETTRE, G., LINDBLAD, U., LYON, H. N., MELANDER, O., NEWTON-CHEH, C., NILSSON, P., ORHO-MELANDER, M., RASTAM, L., SPELIOTES, E. K., TASKINEN, M. R., TUOMI, T., GUIDUCCI, C., BERGLUND, A., CARLSON, J., GIANNINY, L., HACKETT, R., HALL, L., HOLMKVIST, J., LAURILA, E., SJOGREN, M., STERNER, M., SURTI, A., SVENSSON, M., TEWHEY, R., BLUMENSTIEL, B., PARKIN, M., DEFELICE, M., BARRY, R., BRODEUR, W., CAMARATA, J., CHIA, N., FAVA, M., GIBBONS, J., HANDSAKER, B., HEALY, C., NGUYEN, K., GATES, C., SOUGNEZ, C., GAGE, D., NIZZARI, M., GABRIEL, S. B., CHIRN, G. W., MA, Q., PARIKH, H., RICHARDSON, D., RICKE, D. & PURCELL, S. 2007. Genome-wide association analysis identifies loci for type 2 diabetes and triglyceride levels. *Science*, 316, 1331-6.
- SCANLON, P., D., CONNETT, J., E., WALLER, L., A., ALTOSE, M., D., BAILEY, W., C., SONIA, A. & TASHKIN, D. P. 2000. Smoking Cessation and Lung Function in Mild-to-Moderate Chronic Obstructive Pulmonary Disease. *American Journal of Respiratory and Critical Care Medicine*, 161, 381-390.
- SCHARF, S. M., IQBAL, M., KELLER, C., CRINER, G., LEE, S. & FESSLER, H. E. 2002. Hemodynamic Characterization of Patients with Severe Emphysema. *American Journal of Respiratory and Critical Care Medicine*, 166, 314-322.
- SCHERRER, U., RANDIN, D., VOLLENWEIDER, P., VOLLENWEIDER, L. & NICOD, P. 1994. Nitric oxide release accounts for insulin's vascular effects in humans. *Journal of Clinical Investigation*, 94, 2511.
- SCHERRER, U. & SARTORI, C. 2000. Defective nitric oxide synthesis: a link between metabolic insulin resistance, sympathetic overactivity and cardiovascular morbidity. *European journal of endocrinology*, 142, 315-323.
- SCHIFFER, T., WEITZBERG, E., LUNDBERG, J. O. & LARSEN, F. J. 2013. P75: Dietary inorganic nitrate reduces basal metabolic rate in man. *Nitric Oxide*, 31, S46.
- SCHNABEL, R., BLANKENBERG, S., LUBOS, E., LACKNER, K. J., RUPPRECHT, H. J., ESPINOLA-KLEIN, C., JACHMANN, N., POST, F., PEETZ, D. & BICKEL, C. 2005. Asymmetric Dimethylarginine and the Risk of Cardiovascular Events and Death in Patients With Coronary Artery Disease Results from the AtheroGene Study. *Circulation Research*, 97, e53-e59.
- SCHNEIDER, J. G., TILLY, N., HIERL, T., SOMMER, U., HAMANN, A., DUGI, K., LEIDIG-BRUCKNER, G. & KASPERK, C. 2002. Elevated plasma endothelin-1 levels in diabetes mellitus\*. *American journal of hypertension*, 15, 967-972.
- SCHUHMACHER, S., OELZE, M., BOLLMANN, F., KLEINERT, H., OTTO, C., HEEREN, T., STEVEN, S., HAUSDING, M., KNORR, M. & PAUTZ, A. 2011. Vascular dysfunction in experimental diabetes

- is improved by pentaerithrityl tetranitrate but not isosorbide-5-mononitrate therapy. *Diabetes*, 60, 2608-2616.
- SCHULTZ, K.-D., SCHULTZ, K. & SCHULTZ, G. 1977. Sodium nitroprusside and other smooth musclerelaxants increase cyclic GMP levels in rat ductus deferens.
- SCHULZE, F., LENZEN, H., HANEFELD, C., BARTLING, A., OSTERZIEL, K. J., GOUDEVA, L., SCHMIDT-LUCKE, C., KUSUS, M., MAAS, R. & SCHWEDHELM, E. 2006. Asymmetric dimethylarginine is an independent risk factor for coronary heart disease: results from the multicenter Coronary Artery Risk Determination investigating the Influence of ADMA Concentration (CARDIAC) study. *American Heart Journal*, 152, 493. e1-493. e8.
- SCOGNAMIGLIO, R., AVOGARO, A., CASARA, D., CREPALDI, C., MARIN, M., PALISI, M., MINGARDI, R., ERLE, G., FASOLI, G. & VOLTA, S., D 1998. Myocardial Dysfunction and Adrenergic Cardiac Innervation in Patients with Insulin-Dependent Diabetes Mellitus *American College of Cardiology* 31, 404-412.
- SHAMOON, H., DUFFY, H., FLEISCHER, N., ENGEL, S., SAENGER, P., STRELZYN, M., LITWAK, M., WYLIEROSETT, J., FARKASH, A. & GEIGER, D. 1993. The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulindependent diabetes-mellitus. *New England Journal of Medicine*, 329, 977-986.
- SHAW, A. W. & VOSPER, A. J. 1977. Solubility of nitric oxide in aqueous and nonaqueous solvents. Journal of the Chemical Society, Faraday Transactions 1: Physical Chemistry in Condensed Phases, 73, 1239-1244.
- SHAW, J. E., SICREE, R. A. & ZIMMET, P. Z. 2010. Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes Research and Clinical Practice*, 87, 4-14.
- SHEPHERD, A. I., WILKERSON, D. P., DOBSON, L., KELLY, J., WINYARD, P. G., JONES, A. M., BENJAMIN, N., SHORE, A. C. & GILCHRIST, M. 2015. The effect of dietary nitrate supplementation on the oxygen cost of cycling, walking performance and resting blood pressure in individuals with chronic obstructive pulmonary disease: a double blind placebo controlled, randomised control trial. *Nitric Oxide*.
- SHICHIRI, M., KISHIKAWA, H., OHKUBO, Y. & WAKE, N. 2000. Long-term results of the Kumamoto Study on optimal diabetes control in type 2 diabetic patients. *Diabetes Care*, 23 Suppl 2, B21-9.
- SHIP, J. A. & FISCHER, D. J. 1997. The Relationship Between Dehydration and Parotid Salivary Gland Function in Young and Older Healthy Adults. *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*, 52A, M310-M319.
- SHIVA, S., HUANG, Z., GRUBINA, R., SUN, J., RINGWOOD, L. A., MACARTHUR, P. H., XU, X., MURPHY, E., DARLEY-USMAR, V. M. & GLADWIN, M. T. 2007. Deoxymyoglobin Is a Nitrite Reductase That Generates Nitric Oxide and Regulates Mitochondrial Respiration. *Circulation Research*, 100, 654-661.
- SHORE, A. C., JAAP, A. J. & TOOKE, J. E. 1994. Capillary pressure in patients with NIDDM. *Diabetes*, 43, 1198-1202.
- SIASOS, G., TOUSOULIS, D., ANTONIADES, C., STEFANADI, E. & STEFANADIS, C. 2007. L-Arginine, the substrate for NO synthesis: An alternative treatment for premature atherosclerosis? *International journal of cardiology,* 116, 300-308.
- SIERVO, M., LARA, J., OGBONMWAN, I. & MATHERS, J. C. 2013. Inorganic nitrate and beetroot juice supplementation reduces blood pressure in adults: a systematic review and meta-analysis. *The Journal of nutrition,* 143, 818-826.
- SIMS, M. W., MARGOLIS, D. J., LOCALIO, A. R., PANETTIERI, R. A., KAWUT, S. M. & CHRISTIE, J. D. 2009. Impact of pulmonary artery pressure on exercise function in severe COPD. *Chest*, 136, 412-419.
- SIN, D. D. & VESTBO, J. 2009. Biomarkers in Chronic Obstructive Pulmonary Disease. *Proceedings of the American Thoracic Society, 6*, 543-545.

- SINDLER, A. L., FLEENOR, B. S., CALVERT, J. W., MARSHALL, K. D., ZIGLER, M. L., LEFER, D. J. & SEALS, D. R. 2011. Nitrite supplementation reverses vascular endothelial dysfunction and large elastic artery stiffness with aging. *Aging Cell*, 10, 429-37.
- SNIDER, G. 1985. The definition of emphysema; report of a National Heart, Lung and Blood Institute. Division of Lung Diseases. Workshop. *Am Rev Respir Dis*, 132, 182-185.
- SOCIETY, B. T. 2006. The Burden of Lung Disease. (2nd edition) British Thoracic Society
- SOKOS, G. G., BOLUKOGLU, H., GERMAN, J., HENTOSZ, T., MAGOVERN JR, G. J., MAHER, T. D., DEAN, D. A., BAILEY, S. H., MARRONE, G., BENCKART, D. H., ELAHI, D. & SHANNON, R. P. 2007. Effect of Glucagon-Like Peptide-1 (GLP-1) on Glycemic Control and Left Ventricular Function in Patients Undergoing Coronary Artery Bypass Grafting. *The American Journal of Cardiology*, 100, 824-829.
- SPEIJERS, G. & BRANDT, P., A 2003. Nitrate in: Food Additives Series, JointFAO/WHO Expert Committee on Food Additives. *Geneva*.
- SPIEGELHALDER, B., EISENBRAND, G. & PREUSSMANN, R. 1976. Influence of dietary nitrate on nitrite content of human saliva: Possible relevance to in vivo formation of N-nitroso compounds. *Food and Cosmetics Toxicology,* 14, 545-548.
- SPRAFKA, J. M., BURKE, G. L., FOLSOM, A. R., MCGOVERN, P. G. & HAHN, L. P. 1991. Trends in prevalence of diabetes mellitus in patients with myocardial infarction and effect of diabetes on survival. The Minnesota Heart Survey. *Diabetes Care*, 14, 537-43.
- STADLER, K. 2013. Oxidative stress in diabetes. Diabetes. Springer.
- STAMLER, J., S & MEISSNER, G. 2001. Physiology of Nitric Oxide in Skeletal Muscle. *American Physiology Society*, 81, 209-231.
- STAMLER, J., VACCARO, O., NEATON, J. D. & WENTWORTH, D. 1993. Diabetes, other risk factors, and 12-yr cardiovascular mortality for men screened in the Multiple Risk Factor Intervention Trial. *Diabetes Care*, 16, 434-444.
- STEINBERG, H. O., CHAKER, H., LEAMING, R., JOHNSON, A., BRECHTEL, G. & BARON, A. D. 1996. Obesity/insulin resistance is associated with endothelial dysfunction. Implications for the syndrome of insulin resistance. *Journal of Clinical Investigation*, 97, 2601.
- STEINER, M. K., PRESTON, I. R., KLINGER, J. R. & HILL, N. S. 2005. Pulmonary hypertension: inhaled nitric oxide, sildenafil and natriuretic peptides. *Current Opinion in Pharmacology*, 5, 245-250.
- STRATTON, I. M., ADLER, A. I., NEIL, H. A. W., MATTHEWS, D. R., MANLEY, S. E., CULL, C. A., HADDEN, D., TURNER, R. C. & HOLMAN, R. R. 2000. Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. *BMJ*, 321, 405-412.
- STUMVOLL, M., GOLDSTEIN, B. J. & VAN HAEFTEN, T. W. 2005. Type 2 diabetes: principles of pathogenesis and therapy. *The Lancet*, 365, 1333-1346.
- STUMVOLL, M., GOLDSTEIN, B. J. & VAN HAEFTEN, T. W. 2007. Pathogenesis of Type 2 Diabetes. *Endocrine Research*, 32, 19-37.
- SUDA, O., TSUTSUI, M., MORISHITA, T., TASAKI, H., UENO, S., NAKATA, S., TSUJIMOTO, T., TOYOHIRA, Y., HAYASHIDA, Y. & SASAGURI, Y. 2004. Asymmetric Dimethylarginine Produces Vascular Lesions in Endothelial Nitric Oxide Synthase–Deficient Mice Involvement of Renin-Angiotensin System and Oxidative Stress. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 24, 1682-1688.
- SUN, X. & KU, D. D. 2008. Rosuvastatin provides pleiotropic protection against pulmonary hypertension, right ventricular hypertrophy, and coronary endothelial dysfunction in rats. *American Journal of Physiology - Heart and Circulatory Physiology*, 294, H801-H809.
- SURDACKI, A., NOWICKI, M., SANDMANN, J., TSIKAS, D., BOEGER, R. H., BODE-BOEGER, S. M., KRUSZELNICKA-KWIATKOWSKA, O., KOKOT, F., DUBIEL, J. S. & FROELICH, J. C. 1999. Reduced urinary excretion of nitric oxide metabolites and increased plasma levels of asymmetric dimethylarginine in men with essential hypertension. *Journal of cardiovascular pharmacology*, 33, 652-658.

- TANNENBAUM, S., R, WEISMAN, M. & FETT, D. 1976. The effect of nitrate intake formulation in human saliva. *Food Cosmetic Toxicology* 14, 549-552.
- TARASEVICIENE-STEWART, L., SCERBAVICIUS, R., CHOE, K.-H., MOORE, M., SULLIVAN, A., NICOLLS, M. R., FONTENOT, A. P., TUDER, R. M. & VOELKEL, N. F. 2005. An animal model of autoimmune emphysema. *American journal of respiratory and critical care medicine*, 171, 734-742.
- TARASEVICIENE-STEWART, L. & VOELKEL, N. F. 2008. Molecular pathogenesis of emphysema. *The Journal of clinical investigation*, 118, 394.
- TASKINEN, M.-R. 2003. Diabetic dyslipidaemia: from basic research to clinical practice\*. *Diabetologia*, 46, 733-749.
- TATTERSALL, R. B. & FAJANS, S. S. 1975. Prevalence of Diabetes and Glucose Intolerance in 199 Offspring of Thirty-seven Conjugal Diabetic Parents. *Diabetes*, 24, 452-462.
- TDCCTRG, T. D. C. A. C. T. R. G. 1993. The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. . *N Engl J Med*, 329, 977-86.
- TESORIERE, L., ALLEGRA, M., BUTERA, D. & LIVREA, M. A. 2004. Absorption, excretion, and distribution of dietary antioxidant betalains in LDLs: potential health effects of betalains in humans. *The American journal of clinical nutrition*, 80, 941-945.
- TESSARI, P., CECCHET, D., COSMA, A., VETTORE, M., CORACINA, A., MILLIONI, R., IORI, E., PURICELLI, L., AVOGARO, A. & VEDOVATO, M. 2010. Nitric oxide synthesis is reduced in subjects with type 2 diabetes and nephropathy. *Diabetes*, 59, 2152-2159.
- THABUT, G., DAURIAT, G. L., STERN, J. B., LOGEART, D., LÉVY, A., MARRASH-CHAHLA, R. & MAL, H. 2005. Pulmonary Hemodynamics in Advanced COPD Candidates for Lung Volume Reduction Surgery or Lung Transplantation\*. *CHEST Journal*, 127, 1531-1536.
- THANNICKAL, V. J. & FANBURG, B. L. 2000. Reactive oxygen species in cell signaling. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 279, L1005-L1028.
- THOMPSON, K. G., TURNER, L., PRICHARD, J., DODD, F., KENNEDY, D. O., HASKELL, C., BLACKWELL, J. R. & JONES, A. M. 2014. Influence of dietary nitrate supplementation on physiological and cognitive responses to incremental cycle exercise. *Respiratory physiology & neurobiology*, 193, 11-20.
- TISO, M., TEJERO, J., BASU, S., AZAROV, I., WANG, X., SIMPLACEANU, V., FRIZZELL, S., JAYARAMAN, T., GEARY, L., SHAPIRO, C., HO, C., SHIVA, S., KIM-SHAPIRO, D. B. & GLADWIN, M. T. 2011. Human Neuroglobin Functions as a Redox-regulated Nitrite Reductase. *Journal of Biological Chemistry*, 286, 18277-18289.
- TOTZECK, M., HENDGEN-COTTA, U. B., LUEDIKE, P., BERENBRINK, M., KLARE, J. P., STEINHOFF, H.-J., SEMMLER, D., SHIVA, S., WILLIAMS, D., KIPAR, A., GLADWIN, M. T., SCHRADER, J., KELM, M., COSSINS, A. R. & RASSAF, T. 2012. Nitrite Regulates Hypoxic Vasodilation via Myoglobin-Dependent Nitric Oxide Generation. *Circulation*, 126, 325-334.
- TOUSOULIS, D., BÖGER, R. H., ANTONIADES, C., SIASOS, G., STEFANADI, E. & STEFANADIS, C. 2007. Mechanisms of disease: L-arginine in coronary atherosclerosis—a clinical perspective. *Nature Clinical Practice Cardiovascular Medicine*, 4, 274-283.
- TRACEY, W. R., POLLOCK, J. S., MURAD, F., NAKANE, M. & FORSTERMANN, U. 1994. Identification of an endothelial-like type III NO synthase in LLC-PK1 kidney epithelial cells. *American Journal of Physiology-Cell Physiology*, 266, C22-C28.
- TREICHLER, D., UEDA, K., GANGER, C., SCHNEIDER, A., HUGHES, W. & CASEY, D. 2015. Effects of Acute Dietary Nitrate Supplementation on Aortic Blood Pressure and Wave Reflection in Young and Older Adults. *The FASEB Journal*, 29, 950.8.
- TURNER, R., HOLMAN, R., CULL, C., STRATTON, I., MATTHEWS, D., FRIGHI, V., MANLEY, S., NEIL, A., MCELROY, K. & WRIGHT, D. 1998. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). *Lancet*, 352, 837-853.

- TUSHUIZEN, M. E., BUNCK, M. C., POUWELS, P. J., BONTEMPS, S., VAN WAESBERGHE, J. H. T., SCHINDHELM, R. K., MARI, A., HEINE, R. J. & DIAMANT, M. 2007. Pancreatic fat content and β-cell function in men with and without type 2 diabetes. *Diabetes Care*, 30, 2916-2921.
- UKPDS, U. P. D. S. G. 1998. Effect of intensive blood-glucose control with metformin on complications in overweight patients with type 2 diabetes (UKPDS 34). *Lancet*, 352, 854-865.
- USRDS, U. R. D. S. 2012. Annual Data Report: Atlas of Chronic Kidney Disease and End-Stage Renal Disease in the United States. *National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases, Bethesda*.
- VALLEJO, S., ANGULO, J., PEIRO, C., NEVADO, J., SÁNCHEZ-FERRER, A., PETIDIER, R., SANCHEZ-FERRER, C. & RODRÍGUEZ-MAÑAS, L. 2000. Highly glycated oxyhaemoglobin impairs nitric oxide relaxations in human mesenteric microvessels. *Diabetologia*, 43, 83-90.
- VAN CITTERS, R., L & FRANKLIN, D., L 1969. Cardiovascular Performance of Alaska Sled Dogs during Exercise. *Circulation Research*, 24, 33-42.
- VAN VELZEN, A. G., SIPS, A. J. A. M., SCHOTHORST, R. C., LAMBERS, A. C. & MEULENBELT, J. 2008. The oral bioavailability of nitrate from nitrate-rich vegetables in humans. *Toxicology Letters*, 181, 177-181.
- VANHATALO, A., BAILEY, J., B, BLACKWELL, J., R, DIMENNA, F., J, PAVEY, T., G, WILKERSON, D., P, BENJAMIN, N., WINYARD, P. & JONES, A., M 2010. Acute and chronic effects of dietary nitrate supplementation on blood pressure and the physiological repsonses to moderate-intensity and incremental exercise *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 299, 1121-1131.
- VANHATALO, A., FULFORD, J., BAILEY, S. J., BLACKWELL, J. R., WINYARD, P. G. & JONES, A. M. 2011. Dietary nitrate reduces muscle metabolic perturbation and improves exercise tolerance in hypoxia. *J Physiol*, 589, 5517-28.
- VANHOUTTE, P. M. 2009. Endothelial Dysfunction
- The First Step Toward Coronary Arteriosclerosis. Circulation Journal, 73, 595-601.
- VANIN, A. F., BEVERS, L. M., SLAMA-SCHWOK, A. & FAASSEN, E. E. 2007. Nitric oxide synthase reduces nitrite to NO under anoxia. *Cellular and Molecular Life Sciences*, 64, 96-103.
- VELMURUGAN, S., KAPIL, V., GHOSH, S. M., DAVIES, S., MCKNIGHT, A., ABOUD, Z., KHAMBATA, R. S., WEBB, A. J., POOLE, A. & AHLUWALIA, A. 2013. Antiplatelet effects of dietary nitrate in healthy volunteers: Involvement of cGMP and influence of sex. *Free Radic Biol Med*, 24, 00312-2.
- VENEMA, R. C., JU, H., ZOU, R., RYAN, J. W. & VENEMA, V. J. 1997. Subunit Interactions of Endothelial Nitric-oxide Synthase COMPARISONS TO THE NEURONAL AND INDUCIBLE NITRIC-OXIDE SYNTHASE ISOFORMS. *Journal of Biological Chemistry*, 272, 1276-1282.
- VERMEEREN, M. A., WOUTERS, E. F., NELISSEN, L. H., VAN LIER, A., HOFMAN, Z. & SCHOLS, A. M. 2001. Acute effects of different nutritional supplements on symptoms and functional capacity in patients with chronic obstructive pulmonary disease. *The American journal of clinical nutrition*, 73, 295-301.
- VESTBO, J. & LANGE, P. 2002. Can GOLD Stage 0 Provide Information of Prognostic Value in Chronic Obstructive Pulmonary Disease? *American Journal of Respiratory and Critical Care Medicine*, 166, 329-332.
- VESTBO, J., PRESCOTT, E. & LANGE, P. 1996. Association of chronic mucus hypersecretion with FEV1 decline and chronic obstructive pulmonary disease morbidity. Copenhagen City Heart Study Group. *American Journal of Respiratory and Critical Care Medicine*, 153, 1530-1535.
- VILA PETROFF, M. G., EGAN, J. M., WANG, X. & SOLLOTT, S. J. 2001. Glucagon-like peptide-1 increases cAMP but fails to augment contraction in adult rat cardiac myocytes. *Circ Res*, 89, 445-52.
- VOETSCH, B., JIN, R. C. & LOSCALZO, J. 2004. Nitric oxide insufficiency and atherothrombosis. *Histochemistry and cell biology,* 122, 353-367.

- VOIGHT, B. F., SCOTT, L. J., STEINTHORSDOTTIR, V., MORRIS, A. P., DINA, C., WELCH, R. P., ZEGGINI, E., HUTH, C., AULCHENKO, Y. S. & THORLEIFSSON, G. 2010. Twelve type 2 diabetes susceptibility loci identified through large-scale association analysis. *Nature genetics*, 42, 579-589.
- VOLLMAR, B. & MENGER, M. D. 2009. The hepatic microcirculation: mechanistic contributions and therapeutic targets in liver injury and repair. *Physiological reviews*, 89, 1269-1339.
- VONBANK, K., ZIESCHE, R., HIGENBOTTAM, T. W., STIEBELLEHNER, L., PETKOV, V., SCHENK, P., GERMANN, P. & BLOCK, L. H. 2003. Controlled prospective randomised trial on the effects on pulmonary haemodynamics of the ambulatory long term use of nitric oxide and oxygen in patients with severe COPD. *Thorax*, 58, 289-293.
- WAGNER, D. A., SCHULTZ, D. S., DEEN, W. M., YOUNG, V. R. & TANNENBAUM, S. R. 1983. Metabolic Fate of an Oral Dose of 15N-labeled Nitrate in Humans: Effect of Diet Supplementation with Ascorbic Acid. *Cancer Research*, 43, 1921-1925.
- WALTON, G. 1951. Survey of Literature Relating to Infant Methemoglobinemia Due to Nitrate-Contaminated Water. *American Journal of Public Health and the Nations Health*, 41, 986-996.
- WANG, C.-Y., LIU, P.-Y. & LIAO, J. K. 2008a. Pleiotropic effects of statin therapy: molecular mechanisms and clinical results. *Trends in Molecular Medicine*, 14, 37-44.
- WANG, C., WANG, J., ZHAO, L., WANG, Y., LIU, J., SHI, L., XU, M. & WANG, C. 2008b. Sildenafil Inhibits Human Pulmonary Artery Smooth Muscle Cell Proliferation by Decreasing Capacitative Ca<sup>2+</sup> Entry. *Journal of Pharmacological Sciences*, 108, 71-78.
- WARNHOLTZ, A., NICKENIG, G., SCHULZ, E., MACHARZINA, R., BRÄSEN, J. H., SKATCHKOV, M., HEITZER, T., STASCH, J. P., GRIENDLING, K. K. & HARRISON, D. G. 1999. Increased NADH-oxidase—mediated superoxide production in the early stages of atherosclerosis evidence for involvement of the renin-angiotensin system. *Circulation*, 99, 2027-2033.
- WATSON, L., MARGETTS, B., HOWARTH, P., DORWARD, M., THOMPSON, R. & LITTLE, P. 2002. The association between diet and chronic obstructive pulmonary disease in subjects selected from general practice. *European Respiratory Journal*, 20, 313-318.
- WATTS, A. R., LENNARD, M. S., MASON, S. L., TUCKER, G. T. & WOODS, H. F. 1993. Beeturia and the biological fate of beetroot pigments. *Pharmacogenetics and Genomics*, 3, 302-311.
- WEBB, A., J, PATEL, N., LOUKOGEORGAKIS, S., OKORIE, M., ABOUD, Z., MISRA, S., RASHID, R., MIALL, P., DEANFIELD, J., BENJAMIN, N., MACALLISTER, R., HOBBS, A., J & AHLUWALIA, A. 2008a. Acute Blood Pressure Lowering, Vasoprotective, and Antiplatelet Properties of Dietary Nitrate via Bioconversion to Nitrite. *Hypertension*, 51, 784-790.
- WEBB, A. J., MILSOM, A. B., RATHOD, K. S., CHU, W. L., QURESHI, S., LOVELL, M. J., LECOMTE, F. M. J., PERRETT, D., RAIMONDO, C., KHOSHBIN, E., AHMED, Z., UPPAL, R., BENJAMIN, N., HOBBS, A. J. & AHLUWALIA, A. 2008b. Mechanisms Underlying Erythrocyte and Endothelial Nitrite Reduction to Nitric Oxide in Hypoxia: Role for Xanthine Oxidoreductase and Endothelial Nitric Oxide Synthase. *Circulation Research*, 103, 957-964.
- WEI, C.-M., JIANG, S.-W., LUST, J. A., DALY, R. C. & MCGREGOR, C. G. Genetic expression of endothelial nitric oxide synthase in human atrial myocardium. Mayo Clinic Proceedings, 1996. Elsevier, 346-350.
- WEI, M., GASKILL, S. P., HAFFNER, S. M. & STERN, M. P. 1998. Effects of diabetes and level of glycemia on all-cause and cardiovascular mortality. The San Antonio Heart Study. *Diabetes Care*, 21, 1167-72.
- WEI, Y. & MOJSOV, S. 1995. Tissue-specific expression of the human receptor for glucagon-like peptide-I: brain, heart and pancreatic forms have the same deduced amino acid sequences. *FEBS Letters*, 358, 219-224.
- WEITZENBLUM, E., SAUTEGEAU, A., EHRHART, M., MAMMOSSER, M., HIRTH, C. & ROEGEL, E. 1984. Long-term course of pulmonary arterial pressure in chronic obstructive pulmonary disease. *The American review of respiratory disease*, 130, 993-998.

- WENZEL, P., SCHULZ, E., OELZE, M., MÜLLER, J., SCHUHMACHER, S., ALHAMDANI, M. S., DEBREZION, J., HORTMANN, M., REIFENBERG, K. & FLEMING, I. 2008. AT< sub> 1</sub>-receptor blockade by telmisartan upregulates GTP-cyclohydrolase I and protects eNOS in diabetic rats. *Free Radical Biology and Medicine*, 45, 619-626.
- WESTERBLAD, H., BRUTON, J. D. & LANNERGREN, J. 1997. The effect of intracellular pH on contractile function of intact, single fibres of mouse muscle declines with increasing temperature. *J Physiol*, 500 ( Pt 1), 193-204.
- WESTERMANN, D., RIAD, A., RICHTER, U., JÄGER, S., SAVVATIS, K., SCHUCHARDT, M., BERGMANN, N., TÖLLE, M., NAGORSEN, D. & GOTTHARDT, M. 2009. Enhancement of the endothelial NO synthase attenuates experimental diastolic heart failure. *Basic Research in Cardiology*, 104, 499-509.
- WHIPP, B., J, DAVIS, J., A, TORRES, F. & WASSERMAN, K. 1981. A test to determine parameters of aerobic function during exercise *The American Physiological Society*, 50, 217-221.
- WHO, W. H. O. 2011. Use of glycated haemoglobin (HbA1c) in the diagnosis of diabetes mellitus. *Geneva (Switzerland): The Organization*.
- WILD, S., ROGLIC, G., GREEN, A., SICREE, R. & KING, H. 2004. Global Prevalence of Diabetes: Estimates for the year 2000 and projections for 2030. *Diabetes Care*, 27, 1047-1053.
- WILKERSON, D. P., HAYWARD, G. M., BAILEY, S. J., VANHATALO, A., BLACKWELL, J. R. & JONES, A. M. 2012. Influence of acute dietary nitrate supplementation on 50 mile time trial performance in well-trained cyclists. *Eur J Appl Physiol*, 112, 4127-34.
- WILKERSON, D. P., POOLE, D. C., JONES, A. M., FULFORD, J., MAWSON, D. M., BALL, C. I. & SHORE, A. C. 2011. Older type 2 diabetic males do not exhibit abnormal pulmonary oxygen uptake and muscle oxygen utilization dynamics during submaximal cycling exercise. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 300, R685-R692.
- WILKINS, M. R., ALI, O., BRADLOW, W., WHARTON, J., TAEGTMEYER, A., RHODES, C. J., GHOFRANI, H. A., HOWARD, L., NIHOYANNOPOULOS, P., MOHIADDIN, R. H., GIBBS, J. S. & NULL 2010. Simvastatin as a treatment for pulmonary hypertension trial. *American Journal of Respiratory and Critical Care Medicine*, 181, 1106-1113.
- WILSON, P., KANNEL, W. & ANDERSON, K. 1985. Lipids, glucose intolerance and vascular disease: the Framingham Study. *Monographs on atherosclerosis*, 13, 1.
- WILSON, P. W., CUPPLES, L. A. & KANNEL, W. B. 1991. Is hyperglycemia associated with cardiovascular disease? The Framingham Study. *Am Heart J*, 121, 586-90.
- WISEMAN, R. W., BECK, T. W. & CHASE, P. B. 1996. Effect of intracellular pH on force development depends on temperature in intact skeletal muscle from mouse. *Am J Physiol*, 271, C878-86.
- WOHLFART, P., XU, H., ENDLICH, A., HABERMEIER, A., CLOSS, E. I., HÜBSCHLE, T., MANG, C., STROBEL, H., SUZUKI, T. & KLEINERT, H. 2008. Antiatherosclerotic effects of small-molecular-weight compounds enhancing endothelial nitric-oxide synthase (eNOS) expression and preventing eNOS uncoupling. *Journal of Pharmacology and Experimental Therapeutics*, 325, 370-379.
- WOODMAN, R. J., PLAYFORD, D. A. & WATTS, G. F. 2006. Basal production of nitric oxide (NO) and non-NO vasodilators in the forearm microcirculation in Type 2 diabetes: Associations with blood pressure and HDL cholesterol. *Diabetes Research and Clinical Practice*, 71, 59-67.
- WU, G. & MEININGER, C. J. 2009. Nitric oxide and vascular insulin resistance. BioFactors, 35, 21-27.
- WÜST, R., C & DEGENS, H. 2007. Factors contributing to muscle wasting and dysfunction in COPD patients. *Int J Chron Obstruct Pulmon Dis*, 2(3), 289-300.
- WÜST, R. C., MORSE, C. I., DE HAAN, A., RITTWEGER, J., JONES, D. A. & DEGENS, H. 2008. Skeletal muscle properties and fatigue resistance in relation to smoking history. *European journal of applied physiology*, 104, 103-110.
- WYLIE, L. J., KELLY, J., BAILEY, S. J., BLACKWELL, J. R., SKIBA, P. F., WINYARD, P. G., JEUKENDRUP, A. E., VANHATALO, A. & JONES, A. M. 2013a. Beetroot juice and exercise: pharmacodynamic and dose-response relationships. *J Appl Physiol*, 115, 325-36.

- WYLIE, L. J., MOHR, M., KRUSTRUP, P., JACKMAN, S. R., ERMIOTADIS, G., KELLY, J., BLACK, M. I., BAILEY, S. J., VANHATALO, A. & JONES, A. M. 2013b. Dietary nitrate supplementation improves team sport-specific intense intermittent exercise performance. *Eur J Appl Physiol*, 113, 1673-84.
- XU, J., WU, Y., SONG, P., ZHANG, M., WANG, S. & ZOU, M. H. 2007. Proteasome-dependent degradation of guanosine 5'-triphosphate cyclohydrolase I causes tetrahydrobiopterin deficiency in diabetes mellitus. *Circulation*, 116, 944-53.
- YAMAMOTO, H., LEE, C. E., MARCUS, J. N., WILLIAMS, T. D., OVERTON, J. M., LOPEZ, M. E., HOLLENBERG, A. N., BAGGIO, L., SAPER, C. B. & DRUCKER, D. J. 2002. Glucagon-like peptide-1 receptor stimulation increases blood pressure and heart rate and activates autonomic regulatory neurons. *Journal of Clinical Investigation*, 110, 43-52.
- YONG-XIANG, W., POON, K. S., RANDALL, D. J. & PANG, C. C. Y. 1993. Endothelium-derived nitric oxide partially mediates salbutamol-induced vasodilatations. *European Journal of Pharmacology*, 250, 335-340.
- YOSHIDA, M., TAGUCHI, O., GABAZZA, E. C., KOBAYASHI, T., YAMAKAMI, T., KOBAYASHI, H., MARUYAMA, K. & SHIMA, T. 1997. Combined inhalation of nitric oxide and oxygen in chronic obstructive pulmonary disease. *American Journal of Respiratory and Critical Care Medicine*, 155, 526-9.
- YOSHIZUMI, M., PERRELLA, M., BURNETT, J. & LEE, M.-E. 1993. Tumor necrosis factor downregulates an endothelial nitric oxide synthase mRNA by shortening its half-life. *Circulation research*, 73, 205-209.
- YOUN, J. Y., WANG, T., BLAIR, J., LAUDE, K. M., OAK, J.-H., MCCANN, L. A., HARRISON, D. G. & CAI, H. 2012. Endothelium-specific sepiapterin reductase deficiency in DOCA-salt hypertension. *American Journal of Physiology-Heart and Circulatory Physiology*, 302, H2243.
- YOUNG, M. & LEIGHTON, B. 1998. Evidence for altered sensitivity of the nitric oxide/cGMP signalling cascade in insulin-resistant skeletal muscle. *Biochem. J*, 329, 73-79.
- YSART, G., MILLER, P., BARRETT, G., FARRINGTON, D., LAWRANCE, P. & HARRISON, N. 1999. Dietary exposures to nitrate in the UK. *Food Additives & Contaminants*, 16, 521-532.
- ZAMANI, P., RAWAT, D., SHIVA-KUMAR, P., GERACI, S., BHUVA, R., KONDA, P., DOULIAS, P. T., ISCHIROPOULOS, H., TOWNSEND, R. R., MARGULIES, K. B., CAPPOLA, T. P., POOLE, D. C. & CHIRINOS, J. A. 2015. Effect of inorganic nitrate on exercise capacity in heart failure with preserved ejection fraction. *Circulation*, 131, 371-80.
- ZANDER, M., MADSBAD, S., MADSEN, J. L. & HOLST, J. J. 2002. Effect of 6-week course of glucagon-like peptide 1 on glycaemic control, insulin sensitivity, and beta-cell function in type 2 diabetes: a parallel-group study. *Lancet*, 359, 824-30.
- ZEGGINI, E., SCOTT, L. J., SAXENA, R., VOIGHT, B. F., MARCHINI, J. L., HU, T., DE BAKKER, P. I., ABECASIS, G. R., ALMGREN, P. & ANDERSEN, G. 2008. Meta-analysis of genome-wide association data and large-scale replication identifies additional susceptibility loci for type 2 diabetes. *Nature genetics*, 40, 638-645.
- ZHANG, Z., NAUGHTON, D., WINYARD, P. G., BENJAMIN, N., BLAKE, D. R. & SYMONS, M. C. R. 1998. Generation of Nitric Oxide by a Nitrite Reductase Activity of Xanthine Oxidase: A Potential Pathway for Nitric Oxide Formation in the Absence of Nitric Oxide Synthase Activity. *Biochemical and Biophysical Research Communications*, 249, 767-772.
- ZIELIŃSKI, J., TOBIASZ, M., HAWRYŁKIEWICZ, I., ŚLIWIŃSKI, P. & PAŁASIEWICZ, G. 1998. Effects of Long-term Oxygen Therapy on Pulmonary Hemodynamics in COPD Patients A 6-Year Prospective Study. *CHEST Journal*, 113, 65-70.
- ZOCCALI, C., BODE-BÖGER, S. M., MALLAMACI, F., BENEDETTO, F. A., TRIPEPI, G., MALATINO, L. S., CATALIOTTI, A., BELLANUOVA, I., FERMO, I. & FRÖLICH, J. C. 2001. Plasma concentration of asymmetrical dimethylarginine and mortality in patients with end-stage renal disease: a prospective study. *The Lancet*, 358, 2113-2117.

ZOU, M.-H., COHEN, R. A. & ULLRICH, V. 2004. Peroxynitrite and vascular endothelial dysfunction in diabetes mellitus. *Endothelium*, 11, 89-97.