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2 **Ecology of testate amoebae in an Amazonian peatland and**  
3 **development of a transfer function for palaeohydrological**  
4 **reconstruction**

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7 *Manuscript for MICROBIAL ECOLOGY*

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30

31 Tropical peatlands represent globally important carbon sinks with a unique biodiversity and are cur-  
32 rently threatened by climate change and human activities. It is now imperative that proxy methods are  
33 developed to understand the natural ecohydrological dynamics of these systems and for testing peat-  
34 land development models. Testate amoebae have been used as environmental indicators in ecological  
35 and palaeoecological studies of peatlands, primarily in ombrotrophic *Sphagnum*-dominated peatlands  
36 in the mid- and high latitudes. We present the first ecological analysis of testate amoebae in a tropical  
37 peatland, a nutrient-poor domed bog in western (Peruvian) Amazonia. Litter samples were collected  
38 from different hydrological microforms (hummock to pool) along a transect from the edge to the interior  
39 of the peatland. We recorded 47 taxa from 21 genera. The most common taxa are *Cryptodiffugia ovi-*  
40 *formis*, *Euglypha rotunda* type, *Phryganella acropodia*, *Pseudodiffugia fulva* type and *Trinema lineare*.  
41 One species found only in the southern hemisphere, *Argygnia spicata*, is present. *Arcella* spp., *Cent-*  
42 *tropyxis aculeata* and *Lesqueresia spiralis* are indicators of pools containing standing water. Canonical  
43 Correspondence Analysis and Non-Metric Multidimensional Scaling illustrate that water table depth  
44 is a significant control on the distribution of testate amoebae, similar to the results from mid- and  
45 high latitude peatlands. A transfer function model for water table based on weighted averaging partial  
46 least-squares (WAPLS) regression is presented and performs well under cross-validation ( $r_{\text{apparent}}^2 =$   
47  $0.76$ ,  $\text{RMSE} = 4.29$ ;  $r_{\text{jack}}^2 = 0.68$ ,  $\text{RMSEP} = 5.18$ ). The transfer function was applied to a 1-m peat  
48 core and sample-specific reconstruction errors were generated using bootstrapping. The reconstruction  
49 generally suggests near-surface water tables over the last 3,000 years, with a shift to drier conditions  
50 at c. cal. AD 1218-1273.

# 51 1 Introduction

52 Tropical peatlands are thought to contain approximately 88.6 Gt of carbon, equivalent to 15-19 %  
53 of the global peatland carbon pool [1, 2]. They support important ecosystems and are found in both  
54 lowland and upland areas in SE Asia, Africa and Central and South America [3, 4, 5, 6]. A wide variety  
55 of peatlands have recently been discovered in the subsiding Pastaza-Marañón basin in Peruvian Ama-  
56 zonia including minerotrophic palm swamps and ombrotrophic domed bogs, classified by differences in  
57 surface nutrient status, topography and vegetation communities [5, 7]. Peat thickness is also variable  
58 (from <1 to 7.5 m) [5, 7]. These peatlands are different to those in SE Asia as they have not been  
59 heavily disturbed by human activity and the domed bogs may therefore be the best remaining exam-  
60 ples in the world [7, 8]. Peatlands have also been recently reported from Central Amazonia (Brazil),  
61 although peat thicknesses are not as great as in the west [9].

62  
63 Peatlands in the tropics are vulnerable to destabilisation through climate-induced changes and human  
64 activities including deforestation, drainage and burning [2, 10, 11]. To fully understand how tropical  
65 peatlands may respond to such drivers of change, knowledge of their developmental history and past  
66 ecohydrological dynamics is needed. The use of testate amoebae for palaeohydrological reconstruction  
67 is well established for mid-latitude peatlands [12, 13, 14, 15, 16, 17] and subarctic/boreal peatlands  
68 to a lesser extent [18]. However, their potential as hydrological indicators in tropical peatlands has  
69 not yet been assessed despite several ecological studies of testate amoebae in the tropics [19, 20, 21].  
70 Hydrological reconstructions from tropical peatlands may prove particularly useful as relatively little  
71 hydrological monitoring data exists, especially from sites in Africa and S. America.

72  
73 This paper has three aims:

- 74 (1) To describe the testate amoebae communities in an Amazonian peatland;
- 75 (2) To determine the most important environmental parameters that influence the testate amoeba  
76 communities;
- 77 (3) To elucidate if testate amoeba transfer functions for the reconstruction of hydrological change can  
78 be developed in these environments.

79 We test the hypothesis that water table depth is the strongest environmental control on the distribution  
80 of testate amoebae in an Amazonian peatland.

## 81 **2 Study site**

82 Aucayacu is a nutrient-poor domed peatland in Peruvian Amazonia which now operates as an om-  
83 brotrophic 'raised bog' system [7]. It is situated on alluvial fan sediments between a stream of the  
84 Pastaza fan and the Tigre River (Figure 1). The peatland was initially a nutrient rich minerotrophic  
85 system that gradually became an ombrotrophic raised bog [8]. Aucayacu represents the deepest peat-  
86 land that has been discovered in the Amazon basin (up to 7.5 m thick) and peat initiation at the site  
87 has been dated to c. 8870 cal. BP [8]. The vegetation of Aucayacu is characterised by 'pole' and  
88 'dwarf' forest communities.

89  
90 At the nearby city of Iquitos (Figure 1), average annual rainfall of up to 3000 mm is typical, with  
91 the wet season spanning the months November to March when the ITCZ has migrated just south of  
92 the equator and is positioned over northern Peru [22]. Even the dry season from June to September  
93 can experience monthly rainfall totals of up to 170 mm [23]. Owing to its equatorial position, the  
94 altitude of the midday sun is always close to vertical, leading to nearly constant monthly temperatures  
95 throughout the year. The average annual temperature at Iquitos is 26°C, with a diurnal range of  
96 approximately 10°C (30-32°C daytime temperatures and 21-22°C at night) [23]. The climate of this  
97 region is classed as equatorial under the Köppen climate classification (*Af*).

## 98 **3 Methods**

99 Linear transects from the edge to the interior of the Aucayacu peatland were established and 100 sur-  
100 face sampling points were selected to cover the entire microtopographical/hydrological gradient. The  
101 transect was surveyed using a Leica level and staff and the locations of the sample points recorded  
102 using a hand-held GPS. Litter samples of approximately 5 cm<sup>3</sup> were sampled from each point and  
103 placed into ziplock bags. The size and shape of each microform along with the vegetation composition  
104 (within an area of 5 m<sup>2</sup>), % litter and vegetation cover was recorded at each location (Supplementary  
105 file 1, 2). A hole was augered at each sampling point and the water table depth measured at regular  
106 intervals until it equalised before being measured with a metal ruler (Supplementary file 3). The water  
107 table measurements were carried out over a three day period to ensure they were internally consistent.  
108 pH and conductivity measurements were carried out on water samples extracted from the boreholes

109 using calibrated field meters. A 1-m core was extracted from the interior of the Aucayacu peatland  
110 using a Russian corer following the parallel hole method [24, 25].

111  
112 Following courier transport, all samples were returned to the laboratory and stored in refrigeration at  
113 4°C prior to further analysis. Approximately one half of each litter sample was weighed, oven dried  
114 and re-weighed to determine moisture content. The samples were subsequently burnt in a muffle fur-  
115 nace at 450°C for 8 hours to determine loss-on-ignition [26]. Testate amoebae were prepared using a  
116 modified version of the standard method [27]. Samples of known volume were sieved through a 300 µm  
117 sieve and no fine-sieving was carried out following [28]. The samples were stored in deionised water.  
118 Testate amoebae were counted under transmitted light at 200-400× and identified using morphology,  
119 composition, size and colour to distinguish taxa. At least 100 specimens were counted per sample  
120 [29]. The taxonomy uses a morphospecies approach in certain circumstances, where a designation that  
121 includes other species has been classed as a "type". Testate amoebae were identified using several  
122 standard keys [30, 31, 32, 33, 34]. Scanning electron microscope images were taken using a Hitachi  
123 S-3700N scanning electron microscope (<http://www.sem-eds.amu.edu.pl/>). The core was sub-sampled  
124 in the laboratory and samples were prepared for testate amoebae analysis as outlined above.

125  
126 Above ground plant material (e.g. leaf fragments, wood and seeds) were extracted from the peat  
127 samples and AMS <sup>14</sup>C dates at <sup>14</sup>Chrono (Queen's University Belfast) and the SUERC Accelerator  
128 Mass Spectrometer Laboratory (East Kilbride, Scotland). All samples were acid-alkali-acid washed  
129 prior to analysis. We looked for Spheroidal Carbonaceous Particles (SCPs) in the top 50 cm of the  
130 peat core in an attempt to date the recent century [35]; however, none were present.

131  
132 The gradient length of the contemporary data was determined using Detrended Correspondence Analy-  
133 sis (DCA). As the data are characterised by a long gradient, Canonical Correspondence Analysis (CCA)  
134 was used to explore the relationships between testate amoebae taxa and environmental variables. The  
135 relative contributions of the environmental variables were investigated using a series of partial CCAs  
136 [36], enabling an estimation of how the total variance is partitioned and the intercorrelations between  
137 variables. Monte-Carlo permutation tests were used to determine the statistical significance of these  
138 analyses (e.g. Dale and Dale 2002). Our use of CCA enables direct comparisons with previous studies

139 of peatland testate amoebae that have relied on this technique. A number of environmental variables  
140 (plant functional types, distances and heights, peat thickness, % litter and vegetation cover, number  
141 of plant taxa, microform area) were considered as indirect factors and were included as passive (sup-  
142 plementary) variables in the analysis. As there have been some criticisms of the use of the  $\chi^2$  distance  
143 in CCA [38, 39], Nonmetric Multidimensional Scaling (NMDS), [40, 41] was also used to examine the  
144 relationship between testate amoebae and environmental variables. In contrast to CCA, NMDS does  
145 not make assumptions about species distributions over environmental gradients. Species data were  
146 square root transformed prior to NMDS ordination and Sorensen distance measure was used. The  
147 optimum solution was identified through comparison of final stress values. The analysis was carried  
148 out using the Vegan package in R version 2.15.1 [42, 43]. The Shannon Diversity Index (SDI) was used  
149 to examine the community diversity [44]. The SDI is defined as:

$$SDI = \sum_{i=1}^s \left( \frac{X_i}{N_i} \right) \ln \left( \frac{X_i}{N_i} \right)$$

155 where  $X_i$  is the abundance of each taxon in a sample,  $N_i$  is the total abundance of the sample,  
156 and  $s$  is equal to the species richness of the sample. Environments are considered to be healthy if  
157 the SDI falls between 2.5 and 3.5, in transition between 1.5 and 2.5, and stressed between 0.1 and 1.5  
158 [45, 46].

159  
160 Detrended Canonical Correspondence Analysis (DCCA) was performed on the dataset to determine  
161 the gradient length and therefore to deduce whether linear or unimodal-based regression methods  
162 would best represent the taxon-environment relationships [47]. Transfer functions were constructed  
163 using several regression models - Weighted averaging (WA), tolerance-downweighted weighted averag-  
164 ing (WA-Tol), locally-weighted weighted averaging (LWWA), weighted averaging partial least-squares  
165 (WA-PLS) and maximum likelihood (ML). Models were also developed using the modern analogue  
166 technique (MAT) and weighted modern analogue technique (WMAT). The models were built using C2  
167 [48]. The performance of the models was assessed using  $r^2$  and the root mean square error of prediction  
168 (RMSEP) with leave-one-out cross validation (jack-knifing) and bootstrapping. The transfer function

169 models were improved through removal of 19 samples with high residual values (>10 cm). A further 11  
170 samples were screened out based on influence of other (non-hydrological) factors. The best performing  
171 model was the one based on WA-PLS (component 2). The water table transfer function was applied to  
172 subfossil data and sample-specific errors of prediction were generated by 999 bootstrap cycles [49, 50].

## 173 4 Results

### 174 4.1 Site characteristics

175  
176 The topographic survey illustrates that Aucayacu is a domed peatland with a steep rand (Figure  
177 2). Environmental parameters vary within the peatland and there is clear evidence of river influence  
178 at the edge of the site causing higher pH and reduced loss-on-ignition (through delivery of minerogenic  
179 material). A silty clay deposit underlies the peatland and there is a small natural levee at the peatland  
180 edge (Figure 2). The vegetation survey suggests the presence of at least 87 plant taxa (Figure 3;  
181 Supplementary file 1), the most commonly encountered trees and 'small trees' include *Alibertia* sp. 1,  
182 *Iryanthera ulei*, *Virola pavonis*, *Zygia* sp. 1 and *Oxandra euneura*. The most commonly occurring palm  
183 trees are *Mauritia flexuosa* and *Oenocarpus mapora*. Understory herbs include *Trichomanes pinnatum*  
184 and *Pariana* sp. 1. There are variations in the distribution of plant functional types (PFTs) and the  
185 relative contribution of different plant families across the site (Figure 3). For example, individuals from  
186 the families Myristicaceae and Arecaceae become more abundant with distance from the river. Plants  
187 from the Euphorbiaceae and Annonaceae families are well represented in both the peatland margins  
188 and interior, whereas there is a zone of plants from the Rubiaceae family approximately 500-800 m  
189 from the river.

190  
191 A series of microforms were encountered in the Aucayacu peatland including hollows and pools (the  
192 latter characterised by standing water), flat areas ('litter flats'), raised ridges and mounds of accu-  
193 mulated litter ('litter hummocks' and 'ridges') and raised areas caused by litter accumulation around  
194 the roots of large trees ('tree hummocks'). The pools and hollows become larger in the interior of the  
195 peatland and more aligned to the contours, similar to Northern peatlands (Figure 2, Supplementary  
196 file 2). The microforms are characterised by contrasting water table depths (Figure 4, Supplementary  
197 file 3).

199 *4.2 Ecology of testate amoebae*

200

201 A total of 47 testate amoebae taxa from 21 genera were identified at Aucayacu (Figures 5, 6ab,  
 202 Table 1). The most common taxa are *Cryptodiffugia oviformis*, *Euglypha rotunda* type, *Phryganella*  
 203 *acropodia*, *Pseudodiffugia fulva* type and *Trinema lineare*. One species found only in the southern  
 204 hemisphere, *Argygnia spicata*, is present [51]. *Arcella* spp., *Centropyxis aculeata* and *Lesqueresia spi-*  
 205 *ralis* are indicators of pools with standing water. We recorded one potentially new species of *Arcella* -  
 206 details of which will be published elsewhere. The Shannon Diversity Index values of the samples range  
 207 between 0.8-2.8.

208

209 CCA axes one (eigenvalue = 0.149) and two (eigenvalue = 0.063) explain 11.7% of the variance in  
 210 the testate amoebae data (Figure 7). The hydrological variables (moisture content and water table)  
 211 and loss-on-ignition are strongly correlated to axis one. The associated Monte Carlo permutation test  
 212 shows that CCA axis one is highly significant ( $p < 0.001$ , 999 random permutations), pH and conductiv-  
 213 ity are correlated with axis two. A series of partial CCAs show that water table depth explains 15.3%  
 214 of the variance in the data ( $p < 0.002$ ). pH explains 12.9% ( $p < 0.002$ ), Moisture content explains 8.9%  
 215 ( $p < 0.025$ ), conductivity explains 11.8% ( $p < 0.030$ ) and loss-on-ignition explains 10.0% ( $p < 0.0470$ ).  
 216 The strong influence of hydrological variables is also illustrated by the NMDS ordination as water  
 217 table and moisture content are correlated with NMDS coordinate 1 (Figure 7).

218

219 *4.3 Transfer function and application to core*

220

221 The performance statistics for the transfer function models are shown in Table 2. The best perform-  
 222 ing transfer function model (Figure 8) is based on weighted averaging partial least-squares (WAPLS)  
 223 component 2 ( $r_{\text{apparent}}^2 = 0.53$ , RMSE = 7.70,  $r_{\text{jack}}^2 = 0.40$ , RMSEP = 9.13). After the screening  
 224 of samples (Supplementary file 5) the model performance greatly improved ( $r_{\text{apparent}}^2 = 0.76$ , RMSE  
 225 = 4.29;  $r_{\text{jack}}^2 = 0.68$ , RMSEP = 5.18). The most common subfossil testate amoebae present in the  
 226 core from Aucayacu include *Hyalosphenia subflava* "major" ( $> 60\mu\text{m}$ ), *Hyalosphenia subflava* "minor"  
 227 ( $< 60\mu\text{m}$ ), *Phryganella acropodia*, *Trigonopyxis arcula* "polygon aperture", *Centropyxis aculeata* and



228 *Cryptodiffugia oviformis*. The transfer function was applied to the subfossil data and there were no  
229 missing modern analogues. The directional changes in the water table reconstruction are mirrored by  
230 principal components analysis (PCA) axis one scores, suggesting that the transfer function is correctly  
231 representing the structure in the subfossil data (Supplementary file 6). The reconstruction suggests  
232 near-surface water tables over the last 3,000 years (Figure 9; Supplementary file 7) with a marked  
233 shift to drier conditions at c. 50 cm (c. cal. AD 1218-1273).

## 234 5 Discussion

235 To our knowledge this is the first study examining the ecology of testate amoebae in a tropical peat-  
236 land. We have demonstrated that testate amoebae are sensitive hydrological indicators in Amazo-  
237 nian peatlands, suggesting they have the potential to be used more widely in tropical peatland re-  
238 search. The multivariate statistical analysis illustrates the strong hydrological controls on the dis-  
239 tribution of testate amoebae, similar to the research findings from mid- and high latitude peatlands  
240 [12, 13, 14, 15, 16, 17, 18]. pCCA also shows that pH is an important control on testate amoebae in the  
241 Aucayacu peatland - two species (*Trinema grandis*, *Pyxidicula operculata*) are indicators of higher pH  
242 conditions. However, the statistical analysis also demonstrates that a large proportion of variance in  
243 the testate amoeba data remains unexplained. This may be due to a combination of inter-correlations  
244 between environmental variables and unmeasured environmental (edaphic/abiotic) factors. Such fac-  
245 tors may include the characteristics of the canopy (determining the amount of moisture reaching the  
246 peatland surface), litter quality, diversity and decomposition [21], variations in nutrient status, and  
247 other unmeasured geochemical factors. It has also been suggested that short-term environmental vari-  
248 ability may be an important factor in the community dynamics of testate amoebae [52]. There is evidence  
249 that the Aucayacu stream has an influence on the SW margin of the peatland (increased pH and  
250 loss-on-ignition - Figure 2) which may affect the testate amoebae communities there. There is also a  
251 possibility that occasional high-magnitude river flooding events affects the peatland interior, although  
252 there is no evidence for this.

253  
254 There appears to be differences in the ecology of certain species compared to the findings from mid- and  
255 high latitude peatlands. For example, *Hyalosphenia subflava*, *Diffugia pulex* and *Trigonopyxis arcuata*  
256 are not unambiguous dry indicators as reported from mid-latitude *Sphagnum* peatlands [15, 17, 53].

257 However, the ecology of these taxa may be complex as they have been observed in wet fen environ-  
258 ments as well as dry bog hummocks in subarctic and boreal peatlands [18]. The large abundance of  
259 *Diffugia pulex* and *Hyalosphenia subflava* in this tropical peatland is particularly interesting as there  
260 have been some problems finding modern analogues for these taxa in temperate peatlands [12, 14, 54]  
261 and it has also been suggested that these two taxa are characteristic of highly variable conditions [52].  
262 However, *Centropyxis aculeata* is consistently a wet indicator in both tropical and temperate peatlands  
263 [55]. There is morphological variability of certain taxa reported here such as the marked differences in  
264 the aperture of *Trigonopyxis arcuata*. Ogden and Hedley (1980) describe the highly variable aperture  
265 shape of *T. arcuata* [31], which was also noted by Bobrov et al. (1995) in populations from Russia and  
266 Canada including the occurrence of 3-point, 4-point and almost polygonal/circular apertures [56].

267  
268 Palaeohydrological reconstructions based on testate amoebae may prove particularly useful for exam-  
269 ining the developmental history of tropical peatlands. Previous pollen and stratigraphic data suggest  
270 that Amazonian peatlands undergo major vegetation transitions in their developmental history [57].  
271 Testate amoebae may provide important information about the role of changing hydrology across such  
272 ecological transitions. However, poor preservation of tests may hinder this for older sections of the  
273 subfossil record [58, 59, 60].

274  
275 Relatively little is currently known about the ecohydrological dynamics of tropical peatlands. Peatland  
276 development models [61, 62] modified for tropical ecosystem PFTs, productivity and decomposition  
277 (e.g. [63]) may shed light on the long-term ecohydrological and C dynamics of these systems. Of  
278 particular interest is how peatlands respond to climatic shifts [64]. Testate amoebae-based reconstruc-  
279 tions may therefore prove useful for testing the hydrological outputs of such models and understanding  
280 peatland responses to changing climate. In the case of Amazonian peatlands, changing flooding regime  
281 through time and river channel migration may also affect the peatland development trajectories.

282  
283 Previous palaeoenvironmental studies of lakes and swamps in Amazonia have suggested distinct phases  
284 of climate changes during the Holocene. For example, there is compelling evidence for a period of in-  
285 creased precipitation from several areas of Amazonia at c. 700-1300 cal. BP [65, 66, 67, 68, 69].  
286 Further work is needed to examine the strength of the climatic signal preserved in Amazonian peat-

287 lands through i) the generation of modern and palaeoecological data from other suitable sites; ii) the  
288 development of robust core chronologies; iii) high-resolution sampling; iv) multiproxy approaches; iv)  
289 inter and intra-site comparison studies and v) the comparison of peat-based reconstructions to in-  
290 dependent palaeoclimatic data. Furthermore, hydrological monitoring data will help understand the  
291 sub-annual and inter-annual hydrological dynamics of Amazonian peatlands.

292  
293 There are major concerns about the effects of climate change and human activity on tropical peatlands  
294 as they are globally important carbon sinks [1, 2]. Future climate change scenarios for north-east Peru,  
295 derived from an average of 21 climate models and expressed as relative changes from the 1961-1990  
296 baseline climate to the year 2100, project an average annual increase in temperatures of 3.0 – 3.5 °C  
297 and a 5 – 10% increase in precipitation across the region [23]. Agreement between models is generally  
298 low (particularly for precipitation), but nonetheless a consistent pattern of warming and increasing  
299 precipitation is projected [23]. As the Amazonian rainforest is also of special interest as a biodiversity  
300 hotspot [70], further research is needed to examine the sensitivity of Amazonian peatlands to climate  
301 change and loss of biodiversity from human impacts. Testate amoebae may prove to be a particularly  
302 useful tool in this endeavour.

## 303 6 Conclusions

304 (1) We present the first analysis of testate amoebae from a tropical peatland (Aucayacu, Peruvian  
305 Amazonia). We recorded 47 testate amoebae taxa from 21 genera in surface litter samples. The most  
306 common taxa are *Cryptodiffugia oviformis*, *Euglypha rotunda* type, *Phryganella acropodia*, *Pseudodif-*  
307 *flugia fulva* type and *Trinema lineare*. *Arcella* spp., *Centropyxis aculeata* and *Lesqueresia spiralis* are  
308 indicators of pools with standing water.

309  
310 (2) Multivariate statistical analysis illustrates that water table depth is the most important control  
311 on the distribution of testate amoebae in the peatland explaining 15.3% of the variance in the data  
312 ( $p < 0.002$ ). pH is the next most important variable explaining 12.9% ( $p < 0.002$ ). A transfer function  
313 model for water table based on weighted averaging partial least-squares (WAPLS) regression is pre-  
314 sented and performs well under cross validation ( $r_{\text{apparent}}^2 = 0.76$ , RMSE = 4.29;  $r_{\text{jack}}^2 = 0.68$ , RMSEP  
315 = 5.18).

317 (3) The transfer function was applied to a 1-m peat core and sample-specific reconstruction errors  
318 were generated using bootstrapping. The reconstruction generally suggests near-surface water tables  
319 over the last 3,000 years, with a marked shift to drier conditions at c. cal. AD 1218-1273. Testate  
320 amoebae may prove very useful for reconstructing the hydrological dynamics of tropical peatlands in  
321 Amazonia and elsewhere.

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330 Sciences, Adam Mickiewicz University. We kindly thank Monika Lutynska for technical support.

## 331 **8 Figure captions**

332 Figure 1. Map showing the location of the Aucayacu peatland, Loreto region, Peruvian Amazonia.  
333 Location of the study site is shown on a Landsat TM RGB false color image (NASA Landsat Program,  
334 Orthorectified, WRS-2, Path 007, Row 063, downloaded from <http://earthexplorer.usgs.gov/>). Band  
335 4 was assigned to red, band 5 was assigned to green and band 7 was assigned to blue.

336

337 Figure 2. Topographic and stratigraphic profile of the Aucayacu peatland with environmental variables  
338 measured along the transects.

339

340 Figure 3. Plant families and plant functional types in the Aucayacu peatland (abundance plotted  
341 against distance from the river).

342

343 Figure 4. Boxplot of water table depths measured for each microform type.

344

345 Figure 5. Contemporary percentage testate amoebae data from Aucayacu peatland, ranked in or-  
346 der of water table depth. The total count and Shannon Diversity Index are also shown.

347

348 Figure 6a. Light microscope images of testate amoebae from Aucayacu: A. *Sphenoderia fissirostris*;  
349 B. *Centropyxis aerophila*; C. *Nebela penardiana*; D. *Lesqueresia spiralis*; E. *Cryptodiffugia oviformis*;  
350 F. *Diffugia pulex*; G. *Tracheleuglypha dentata*; H. *Centropyxis aculeata*; I. *Physochila griseola*; J.  
351 *Quadrullella symmetrica*; K. *Trigonopyxis arcuata* "3-point aperture"; L. *Centropyxis ecornis*.

352

353 Figure 6b. SEM images of testate amoebae from Aucayacu: A. *Argygnnia spicata*; B. *Hyalosphenia*  
354 *subflava* "major" ( $> 60\mu m$ ); C. *Euglypha rotunda* type; D. Aperture of *Euglypha rotunda* type; E.  
355 *Nebela barbata*; F. *Trinema lineare*; G. *Tracheleuglypha dentata*; H. *Physochila griseola*.

356

357 Figure 7. (a) CCA of testate amoebae from Aucayacu and environmental variables (water table  
358 depth, moisture content, pH, conductivity and loss-on-ignition. Abbreviated species codes and sample  
359 numbers are shown (see Table 1). (b) CCA showing environmental variables and other factors plotted  
360 as supplementary variables (plant functional types, number of plant taxa - S, % litter and vegetation,  
361 depth of peatland, distance from river, height above water level, microform area). (c) NMDS ordina-  
362 tion of the species and environmental data.

363

364 Figure 8. Graph of observed versus model estimated water table depth for (a) complete dataset;  
365 (b) screened dataset.

366

367 Figure 9. Percentage subfossil testate amoebae diagram from Aucayacu and water table reconstruc-  
368 tion. Radiocarbon dates are shown.

369

370 Table 1. Taxon codes.

371

372 Table 2. Transfer function model performance statistics.

373

374 Supplementary file 1. Vegetation survey data from Aucayacu.

375

376 Supplementary file 2. Photographs of peatland microforms at Aucayacu.

377

378 Supplementary file 3. Water table equalisation graphs.

379

380 Supplementary file 4. CCA results.

381

382 Supplementary file 5. Samples removed from the refined transfer function.

383

384 Supplementary file 6. Aucayacu water table reconstruction compared to PCA axis 1 scores.

385

386 Supplementary file 7.  $^{14}\text{C}$  dates.

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