

UNIVERSITI PUTRA MALAYSIA

MOLECULAR GENETIC CHARACTERIZATION OF DIFFERENT ACCESSIONS OF CENTELLA ASIATICA

WONG SOOK MUN

FSAS 2003 12



MOLECULAR GENETIC CHARACTERIZATION OF DIFFERENT ACCESSIONS OF CENTELLA ASIATICA

WONG SOOK MUN

DEGREE OF MASTER OF SCIENCE UNIVERSITI PUTRA MALAYSIA

2003



MOLECULAR GENETIC CHARACTERIZATION OF DIFFERENT ACCESSIONS OF CENTELLA ASIATICA

By

WONG SOOK MUN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

January 2003



Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Master of Science

MOLECULAR GENETIC CHARACTERIZATION OF DIFFERENT ACCESSIONS OF CENTELLA ASIATICA

By

WONG SOOK MUN

January 2003

Chairperson: Assoc. Prof. Siti Khalijah Daud, Ph.D.

Faculty: Science and Environmental Studies

Centella asiatica or locally known as "pegaga" belongs to the Apiceae family. This medicinal plant is one of the most important medicinal herbs, and widely used in health foods, pharmaceutical and cosmetic industries. Twelve accessions of C. asiatica planted in MARDI, originated from different locations in Peninsular Malaysia, were used for this study. Phenotypic differences among these accessions are not very distinct, thus this study is undertaken to determine whether there are any genetic differences.

DNA markers, unaffected by environmental or physiological factors, have potential utility in the characterization of plant species. High discriminating power of this class of markers demonstrated uniformity and stability within genetically complex cultivars. Good quality DNA was extracted from leaf samples using conventional hexadecyltrimethylammonium bromide (CTAB) method. Two PCR-based DNA markers

UPM N

system, namely Amplified Fragment Length Polymorphisms (AFLPs) and Long Primer Randomly Amplified Polymorphic DNA (LP-RAPDs), were employed. This study has successfully analyzed the genetic relationships among the accessions of *C. asiatica*. Two phylogenetic trees had been constructed from the unweighted pair group method with arithmatical average (UPGMA) pairwise analyses. Genetic distances was calculated based on the Dice similarity index. From both analyses, the CA01 and CA02 as well as CA05 and CA06 were closer (D=0.119) within the same cluster indicating that they are closely related. Based on the genetic distances, CA10 represented as highest distant group in the LP-RAPDs analyses whereas CA03 represented as highest distant group in the AFLPs analyses. Furthermore, diagnostic band with highest molecular weight (3000 bp) was found in CA10 by using long primer PEH A3. The amplification of CA03 genotypes with AFLP primer pair ACG/CTA has shown a unique DNA profile. In addition, CA03 is easily distinguish from other accessions morphologically due to its wavy shape of the plant leaf.

Different levels of genetic diversity among the accessions suggested that all the accessions are genetically non identical. CA05 showed the lowest percentage of polymorphism within the accession, approximately 6.18 % (LP-RAPDs) and 8.15% (AFLPs). Both techniques employed 6 primers and adequate DNA markers were obtained. However, the AFLP technology produced relatively greater amount of DNA markers, 3616 polymorphic bands, compared to LP-RAPD which had only 773 polymorphic bands. Both marker systems have successfully described genetic diversity of *C. asiatica* although both the AFLPs and LP-RAPDs are dominance inherited markers.



Abstrak tesis yang dikemukakan kepada Senat Universit Putra Malaysia sebagai Memenuhi keperluan untuk ijazah Master Sains.

PENENTUAN CIRI-CIRI GENETIK MOLEKUL ANTARA ASESI CENTELLA ASIATICA YANG BERLAINAN

Oleh

WONG SOOK MUN

Januari 2003

Pengerusi : Prof. Madya Siti Khalijah Daud, Ph.D.

Fakulti : Sains dan Pengajian Alam Sekitar

Centella asiatica atau dikenali sebagai pegaga oleh penduduk tempatan tergolong dalam Famili Apiceae. Ia merupakan herba yang penting dalam industri perubatan dan kosmetik. Dua belas asesi pegaga yang ditanam di MARDI diperolehi dari lokasi yang berlainan di Semenanjung Malaysia. Perbezaan fenotip antara asesi ini tidak berapa ketara. Oleh itu, kajian ini dijalankan untuk menentukan perbezaan dari segi genetiknya.

Penentu DNA yang tidak dipengaruhi oleh faktor-faktor sekitaran atau fisiologi mempunyai potensi untuk digunakan dalam mencirikan spesies tumbuhan. Kuasa diskriminasi yang tinggi bagi kelas penanda DNA ini dapat menunjukkan keseragaman dan kestabilan dalam tanaman yang mempunyai genetic yang kompleks.

UPM

DNA yang berkualiti diekstrak daripada semua sampel daun dengan kaedah CTAB yang konvensional. Dua kaedah berasaskan PCR iaitu AFLP dan LP-RAPD telah digunakan. Kajian ini berjaya menganalisis perhubungan genetik antara asesi-asesi pegaga. Dua pokok filogenetik telah dibina berdasarkan analisis UPGMA yang berasaskan Indeks Keserupaan Dice. Keputusan menunjukkan antara asesi CA01 dan CA02 serta antara asesi CA05 dan CA06 adalah berkait rapat antara satu sama lain dengan jarak genetik bernilai 0.119. Berdasarkan jarak genetik, CA10 merupakan asesi yang mempunyai hubungan genetik yang paling jauh daripada asesi yang lain berdasarkan LP-RAPDs, manakala CA03 merupakan kumpulan yang paling jauh jarak genetiknya dengan asesi yang lain berdasarkan penanda AFLPs. Selain itu, kewujudan jalur penentu yang paling berat jisim molekulnya (3000 bp) bagi CA10 dengan primer panjang PEH A3 dan juga keunikan corak DNA bagi CA03 dengan pasangan primer ACG/CTA telah menyokong keputusan tersebut. Selain itu, perbezaan CA03 amat ketara berbanding dengan asesi lain dari segi morfologi iaitu mempunyai daun berbentuk ombak.

Perbezaan aras diversiti genetik yang berlainan telah ditunjukkan di antara asesi dan ini menunjukkan tidak wujud kesamaan genetik di antara dua belas asesi pegaga tersebut. Asesi CA05 telah menunjukkan peratus polimorfisme terendah dalam asesi, iaitu 6.18 % (LP-RAPD) dan 8.15 % (AFLP). Kedua-dua teknik menggunakan 6 primers dan penanda DNA yang mencukupi telah diperolehi. Walau bagaimanapun, teknologi AFLP telah menghasilkan jumlah DNA yang agak banyak, berjumlah 3616 jalur polimorfisme iaitu jauh lebih banyak berbanding dengan 773 jalur polimorfisme yang dihasilkan oleh LP-RAPD.



Kedua-dua sistem penanda ini berjaya menggambarkan diversiti genetik pegaga walaupun kedua-dua AFLP dan LP-RAPD adalah penanda yang bersifat dominans.



ACKNOWLEDGEMENTS

The author first and foremost wish to acknowledge with gratitude her supervisors, Associate Professor Dr. Siti Khalijah Daud, Professor Dr. Marziah Mahmood and Dr. Nor'Aini bt. Fadzillah for their kind words of wisdom, guidance and encouragement throughout this project.

The author would like to extend her sincere thanks to Professor Tan Soon Gan for his concern in accessibility of the equipments and radioactive room. The author would like to thank the staff of Department of Biochemistry and Microbiology, Universiti Putra Malaysia (UPM) for the facilities of dark room.

The author also not to forget to express special thanks to Puan Indubala (Malaysia Agricultural Research Development Institution, MARDI) for the supplied of *C. asistia*, Miss Chong Lee Kim (Veterinary Department, UPM), Miss Chong Wai Ling (Biotechnology Department, UPM), Mr. Vijay (Biology Department, UPM), Miss Subha (Biology Department, UPM), Miss Phang Chai Ching (Biology Department, UPM), Mr. Chan Soon Choy (Biology Department, UPM), Mr. Ho Boon Peng (Biology Department, UPM), Mr. Kok Lian (Biology Department, UPM), Mr. Wong Sing King (Biology Department, UPM) for their generous information about the various skills applied into the techniques used in this project.

The author would very much appreciate the understanding and supports from her friends

Cik Salina, Puan Hariyati, Puan Intan, Miss Lim Yuet Mee, Mr Lim Fang Loong, Mr. Loi

Ee Hui, Miss Mary Kho, Miss Anna Ling, Encik Sobri.



Financial support from the Government of Malaysia through IRPA (Intensification of Research in Priority Areas) fund account number 01-04-01-T0123 MMBP SUB PROGRAME ON NATURAL PRODUCT DISCOVERY.

Last but not least, my deepest appreciation to her beloved parents Mr. Wong Swee Sung and Madam Chan Choon Ping, sister Ms Wong Wai Mun, Brothers Mr. Wong Kai Kong and Mr. Wong Kai Kuen.



I certify that an Examination Committee met on 8th January 2003 to conduct the final examination of Wong Sook Mun on her Master of Science thesis entitled "Molecular Genetic Characterization of Different Accessions of *Centella asiatica*" in accordance with Universiti Pertanian Malaysia (Higher Degreee) Act 1980 and Universiti Pertanian Malaysia (Higher Degreee) Regulation 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are follows:

SITI SHAPOR SIRAJ, Ph. D.

Associate Professor Department of Biology Faculty of Science and Environment Studies Universiti Putra Malaysia (Chairman)

SITI KHALIJAH DAUD, Ph.D.

Associate Professor Department of Biology Faculty of Science and Environment Studies Universiti Putra Malaysia (Member)

MAZIAH MAHMOOD, Ph.D.

Professor Department of Microbiology and Biochemistry Faculty of Science and Environment Studies Universiti Putra Malaysia (Member)

NOR'AINI MOHD. FADZILLAH, Ph.D.

Department of Biology Faculty of Science and Environment Studies Universiti Putra Malaysia (Member)

GULAM RUSUL RAHMAT ALI, Ph,D.

Professor/Deputy Dean, School of Graduate Studies, Unversiti Putra Malaysia.

Date: 16 JUN 2003



This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfillment of the requirement of the degree of Master of Science. The members of the Supervisory Committee are as follows:

SITI KHALIJAH DAUD, Ph.D.

Associate Professor Department of Biology Faculty of Science and Environment Studies Universiti Putra Malaysia (Chairman)

MAZIAH MAHMOOD, Ph.D.

Professor Department of Microbiology and Biochemistry Faculty of Science and Environment Studies Universiti Putra Malaysia (Member)

NOR'AINI MOHD. FADZILLAH, Ph.D.

Department of Biology Faculty of Science and Environment Studies Universiti Putra Malaysia (Member)

AINI IDERIS, Ph.D.
Professor/Deputy Dean,
School of Graduate Studies,
Unversiti Putra Malaysia.

Date: 11 JUL 2003



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations, which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

WONG SOOK MUN

Date: 16 June 2003



TABLE OF CONTENTS

				Page
ABS				
ABS	TRAK			iv
ACF	KNOW	LEDEGI	EMENTS	vii
ABSTRACT ABSTRAK ACKNOWLEDEGEMENTS APPROVAL DECLARATION LIST OF TABLES LIST OF FIGURES LIST OF ABBREVIATION CHAPTER 1. INTRODUCTION 2. LITERATURE REVIEW 2.1 Centella asiatica 2.1.1 Geographical Distributions 2.1.2 Taxonomy of C. asiatica 2.1.3 Nomenclature 2.1.4 Plant Descriptions 2.1.5 Cytological Studies 2.1.6 Phytochemistry 2.1.7 Medicinal Uses 2.2 Plant Genome 2.3 Molecular Markers 2.4 Polymerase Chain Reaction 2.5 Randomly Amplified Polymporphic DNA Approach 2.6 Long Primer-Randomly Amplified Polymorpism Approach 2.7 Amplification Fragment Length Polymorpism Approach	ix			
DEC	CLARA	TION		xi
LIST	Γ OF Τ	ABLES		xv
LIST	r of f	IGURES		xvi
LIST	Γ OF A	BBREVI	IATION	XX
CHA	APTER			
1.				1
2.				6
	2.1	Centell		6
			U 1	6
			•	7
				7
			•	8
			• •	8
			· · · · · · · · · · · · · · · · · · ·	9
	0.0			10
				11
				12
		•		15
				17
		_	* * * * * * * * * * * * * * * * * * * *	20
	2.1	Amplif	ication Fragment Length Polymorpism Approach	22



			Page
3.	MA	TERIAL AND METHODS	29
	3.1	Plant Materials	29
 4. 	3.2	Total Leaf DNA Isolation	43
	3.3	DNAs Quantification and Qualification	44
	3.4	LP-RAPD Protocol	45
		3.4.1 Primer Screening	45
		3.4.2 Optimization of PCR Conditions	46
		3.4.3 Polymerase Chain Reaction	47
		3.4.4 2% Agarose Gel Electrophoresis	48
	3.5	AFLP Protocol	48
		3.5.1 Restriction Digestion of DNA	49
		3.5.2 Ligation of Adapters	49
		3.5.3 Dilution of 1:5	50
		3.5.4 Pre-amplification Reaction	50
		3.5.5 Dilution of 1:10	50
		3.5.6 EcoRI Primer Labeling	51
		Preparation of Primer Combination Mixtures (MIX 1)	51
		3.5.8 Preparation of PCR Master Mixtures (MIX 2)	51
		3.5.9 Selective AFLP Amplification	52
		3.5.10 40% Acrylamide Stock	52
		3.5.11 6% Denatured Polyacrylamide Sequencing Gel	52
		3.5.12 Denatured Sequencing Gel Analyses	53
	3.6	Data Analysis	54
4.	RES	RESULTS AND DISCUSSION	
	4.1	DNA Templates	56
	4.2	Long Primer RAPD Analyses	59
		4.2.1 Primers Screening	59
		4.2.2 Effects of MgCl ₂ Concentration	61
		4.2.3 Effects of DNA Concentration	62
		4.2.4 Effects of Annealing Temperature	63
		4.2.5 Effects of Various Number of PCR Cycles	64
	4.3	LP-RAPD Fingerprints	65
	4.4	AFLP Fingerprints	77
	4.5	Repeatability Test	87
	4.6	Phylogenetic Tree based on LP-RAPD Analyses	89
	4.7	Phylogenetic Tree based on AFLP Analyses	94
	4.8	Polymorphisms Data (%)	



		Page
5.	DISCUSSION	99
6.	CONCLUSION	102
REF	FRENCES	105
BIO	DDATA OF AUTHOR	123



LIST OF TABLES

Table		Page
1	Variables in common names of C. asiatica in various languages.	8
2	Several factors for choosing molecular techniques (Rashid, 1995).	13
3	List of Long Primers screened in the study (Gillings and Holley, 1997).	46
4	Different concentrations and conditions of PCR mixture in LP-RAPD analyses.	47
5	Various components with particular volume added in the restriction reaction mixture.	49
6	Amplified products (A.P) and Polymorphic bands (POL.B) scored from LP-RAPD fingerprints by using the 6 long primers	70
7	Amplified products (A.P) and Polymorphic bands (POL.B) scored from AFLP fingerprints by using the 6 selected AFLP primer pairs.	78
8	Genetic distance values calculated based on Dice similarity among and within the twelve accessions of <i>C. asiatica</i> using LP-RAPD analysis.	90
9	Genetic distance values calculated based on Dice similarity among and within the twelve accessions of <i>C. asiatica</i> using AFLP analysis.	95



LIST OF FIGURES

Figure		Page
1	The diagram showing an example of AFLP procedures using one primer pair (E-AAC/M-CAA).	24
2	A Peninsular Malaysia map showing the originated collection sites of the 12 accessions of <i>C</i> . asiatica (Jaganath, pers.comm.).	30
3	Centella asiatica accession collected from Brinchang, Cameron Highland, Pahang designated as CA01	31
4	Centella asiatica accession collected from Bukit Cahaya, Shah Alam, Selangor, designated as CA02.	32
5	Centella asiatica accession collected from Tapah, Perak, designated as CA03	33
6	Centella asiatica accession collected from Malacca designated as CA04	34
7	Centella asiatica accession collected from Tapah, Perak, designated as CA05	35
8	Centella asiatica accession collected from Serdang, Selangor, designated as CA06	36
9	Centella asiatica accession collected from Jalan Kebun, Kelang, Selangor, designated as CA07	37
10	Centella asiatica accession collected from Kundang, Muar, designated as CA08	38
11	Centella asiatica accession collected from Tanjung Karang, Perak designated as CA09	39
12	Centella asiatica accession collected from Serdang, Selangor, designated as CA10	40
13	Centella asiatica accession collected from Tanah Rata, Cameron Highland, Pahang, designated as CA11	41
14	Centella asiatica accession collected from Waterfall, Tapah, Perak, designated as CA12	42



15	Lane 1 to lane 16 showed the purified DNA isolated from C . asiatica leaves using CTAB conventional method with RNase treatment. Lane λ is the Lambda $\mathit{Hind III}$ DNA marker.	57
16	DNA extracts without RNase treatment. Lane 1 to lane 12 showed DNA samples with RNA contaminated.	58
17	Lane 1 to lane12 showed the DNA extracts without additional ethanol purification steps.	58
18	Lane 1 to 10 showed the amplification products by screening the 10 long primers on the CA01 accessions. Lane C was the control.	59
19	Lane 1 to 10 showed the amplification products by screening the 10 long primers on the CA03 accessions. Lane C was the control.	60
20	Lane 1 to 10 showed the amplification products by screening the 10 long primers on accessions CA 10. C was the control.	60
21	DNA banding patterns with different concentrations of MgCl ₂ . Lane 1 (2.0 mM), lane 2 (2.5 mM), lane 3 (3.0 mM), lane 4 (3.5 mM), lane 5 (4.0 mM) and lane C (control).	61
22	Effect of different DNA concentrations on DNA templates. Lane 1 (~25ng), lane 2 (~50ng), lane 3 (~100ng), lane 4 (~150ng) and lane 5 (~200ng).	62
23	LP-RAPD fingerprints in different annealing temperatures. Lane 1 (48°C), lane 2 (50°C), lane 3 (52°C), lane 4 (55°C), lane 5 (58°C), lane 6 (60°C) and lane C (Control).	63
24	DNA bands produced from different number of cycles during the amplification reactions. Lane 1 (30 cycles), lane2 (35cycles), lane 3 (40cycles) and lane 4 (45cycles).	64
25	LP-RAPD profile within the CA01 accession with primer PUC/13F. M was the 1kbp DNA ladder (Promega).	66
26	LP-RAPD profile within the CA02 accession with primer BOXAIR. M was the 1kbp DNA ladder (Promega).	67



27	LP-RAPD profile within the CA03 accession with primer ERIC IR. M was the 1kbp DNA ladder (Promega).	68
28	LP-RAPD profile within the CA04 accession with primer ERIC IR. M was the 1kbp DNA ladder (Promega)	69
29	Monomorphic LP-RAPD profile within the CA05 accession with primer PUC/13F. M was the 1kb DNA ladder (Promega).	69
30	LP-RAPD profile within the CA06 accession with primer PEH A6. M was the 1kb DNA ladder (Promega).	70
31	LP-RAPD profile within the CA07 accession with primer BOX AIR. M was the 1kbp DNA ladder (Promega)	71
32	LP-RAPD profile within the CA08 accession with primer ERIC IR. M was the 1kbp DNA ladder (Promega)	72
33	LP-RAPD profile within the CA08 accession with primer ERIC 2. M was the 1kbp DNA ladder (Promega)	73
34	LP-RAPD profile within the CA09 accession with primer BOX AIR. M was the 1kbp DNA ladder (Promega)	73
35	LP-RAPD profile within the CA10 accession with primer PEH A3. M was the 1kbp DNA ladder (Promega)	74
36	LP-RAPD profile within the CA11 accession with primer PEH A3. M was the 1kbp DNA ladder (Promega).	75
37	LP-RAPD profile within the CA12 accession with primer PUC/M13F. M was the 100bp DNA ladder (Promega).	76
38	LP-RAPD profile from the CA01 to CA12 accession. (Lane 1 to lane 12) with primer ERIC IR. M was the 1kp DNA ladder (Promega). Ma was the 100bp DNA ladder (Promega).	77
39	AFLP fingerprints of the CA03 accession generated by primer E-AAG/M-CAG.	79
40	DNA banding pattern of the CA03 accession produced by AFLP primer pair, E-ACC/M-CTA.	80
41	A typical fingerprinting generated by using primer E-ACC/M-CTA on the	81



42	The CA05 accession AFLP fingerprints by using primer E-AAC/M-CAA.	83
43	Highest number of AFLP markers generated by using primer E-ACG/M-CTG on the CA12 accession.	84
44	AFLPs profile produced among the 12 accessions of <i>C. asiatica</i> by using AFLP primer pair E-ACG/M-CTG.	86
45	LP-RAPD reproducible test showing uniform DNA banding pattern. Lanes 1 - 3 are amplified products from 3 replicates of DNAs employed with the same PCR conditions.	88
46	AFLP repeatability test. Lanes 1 -5 are PCR products produced by the same DNAs and primer pairs. Lanes 4 and 5 showing DNA fingerprints using different models of thermal cycler.	88
47	A phylogenetic tree generated from genetic distance values calculated from the LP-RAPD markers for the 12 accessions f <i>C. asiatica</i> .	93
48	Phylogenetic tree constructed using UPGMA cluster analyses based on FLP data to reveal genetic relationships among the 12 accessions of <i>C. asiatica</i> .	97



ABBREVIATIONS AND SYMBOLS

% Percent

°C Degree centrigrade

λ Lamda Gamma γ bp Base pair kb Kilo base U Units ml Milliliter mm Micrometer μl Microliter μ M Micromolar Microgram (s) μg 1 X One time

dATP Deoxyadenosine triphosphate
dCTP Deoxycytidine triphosphate
dGTP Deoxyguanosine triphosphate
dTTP Deoxythymidine triphosphate
dNTPs Deoxynucleotide triphosphates

mer Oligomer

EDTA Ethylenediaminetetracetic acid
A₂₈₀ absorbance at wavelength 280 nm
A₂₆₀ absorbance at wavelength 260 nm

 $\begin{array}{cc} \text{nM} & \text{nanometer} \\ \text{mM} & \text{milimolar} \end{array}$

DNA Deoxyribonucleic acid MgCl₂ Magnesium chloride OD Optical density

RAPD Randomly Amplified Polymorphic DNA

LP Long primer

AFLP Amplified Fragment Length Polymorphisms

PCR Polymerase Chain Reaction SDS Sodium Dodecylsulphate

UV Ultra violet

TBE Tris-borate EDTA
RNA Ribonucleic acid
w/v Weight per volume

V Voltage

UPGMA Unweighted pair-group method with arithmetical averages



CHAPTER 1

INTRODUCTION

A medicinal plant is defined as any plant, one or more of its structures, containing substances that can be used for therapeutic semi-synthesis. Morphologically, medicinal plant species can be classified into trees, shrubs, herbs and ferns. There are approximately 500,000 plant species occupied the terrestrial habitat. About 7% to 14% (35,000 to 70,0000) of these species are used as medicinal plants worldwide. A large portion of these medicinal plants is found in the tropical rainforest biome (Allegra, 1984).

In Malaysia, rainforest biome covers around 58.1% (19.12 million hectares) of the country's total land area. This area supports over 20,000 plant species, of which more than 2000 species were reported of having medicinal values. About 200 medicinal plants species used by different ethnic groups. The most popular local medicinal plant species, such as *Centella asiatica* (Pegaga), *Eurycoma longifolia* (Tongkat Ali), *Kaemferia galanga* (Cekur), *Zingiber officinale* (Halia), *Cymbopogan citratus* (Serai), *Curcums domestica* (Kunyit), *Andrographs paniculate* (Hempedu bumi), are widely consumed traditionally (Aziz, 1973).

Amongst these medicinal plants, *Centella asiatica* (Linn.) Urban belonging to *Apiceae* (also known as umbelliferae) family, was chosen in this study.



The study is part of the Malaysia-MIT Biotechnology Partnership Program (MMBPP). The twelve accessions of *C. asiatica* were coded as CA01, CA02, CA03, CA04, CA05, CA06, CA07, CA08, CA09, CA10, CA11, and CA12.

All the accessions were sampled from different places in Peninsular Malaysia. These accessions were collected and planted in nursery by a senior scientist, Madam Indu Bala Jaganath from the Strategic Environment and Natural Resource Center, MARDI.

Centella asiatica is indigenous to the Southern United States, but is widely distributed in Asia and South Africa (Grieve, 1974). This plant has increased its popularity as a vegetable crop due to its medicinal and nutritional values. It is commonly used as a source of raw material in health food, pharmaceutical and cosmetic industries. The World Health Organization (WHO) had recommended C. asiatica as one of the most important medicinal plant species to be conserved and cultivated (Belcaro et al., 1989).

For improving any reasonable gene conservation program, the precise understanding of the organization of the existing genetic diversity and its factors are emphasized. Thus, biotechnological approaches have a significant impact and play a major role in plant genetic resources conservation.

Traditional breeding procedures are mainly based on the evaluation of morphological characteristics on individual plants, in which environmental factors can influence the results. To overcome this drawback, molecular markers that unaffected by external environmental conditions can be used



effectively in evaluating genetic variation within plant species (Powell *et al.*, 1996).

According to Waugh and Powell (1996), genetic variation or polymorphisms revealed by molecular markers could help to select priority areas for conservation and provide vital information for the development of genetic sampling and improvement. Various types of molecular markers, such as allozymes, RFLPs (Restriction Fragment Length Polymorphisms), RAPDs (Randomly Amplified Polymorphic DNA), AFLPs (Amplified Fragment Length Polymorphisms), are widely used to distinguish between species and between varieties within a species. Recently, these technologies have received great attention in agriculture and horticulture (Cooke, 1995; Smith, 1995; Movell *et al.*, 1995; Cooke and Reeves, 1998).

A broad application of these markers has contributed to plant genetic resources management and also plant breeding programs (Bretting and WildrleChner, 1995). From various studies, DNA fingerprinting have proven to be more powerful tools than isozyme markers for assessing genetic diversity and polymorphisms among individuals in a particular species or even different populations of closely related species (Weising *et al.*, 1995).

The LP-RAPDs assay is a modified RAPD protocol using longer primer, 18 to 24 oligomers, to replace the normal RAPD primer, which having only 8 to 10 bases in length (Gillings and Holley, 1997). This technique is based on the use

