Provided by Universiti Putra Malaysia Institutional Repositor



## **UNIVERSITI PUTRA MALAYSIA**

# THE EFFECT OF CARBOFURAN AND ENDOSULFAN ON THE AFRICAN CATFISH, CLARIAS GARIEPINUS

ISAM ELDIN MOHAMED ELAMIN ABU ZEID

FSAS 2001 58



## THE EFFECT OF CARBOFURAN AND ENDOSULFAN ON THE AFRICAN CATFISH, CLARIAS GARIEPINUS

### By

#### ISAM ELDIN MOHAMED ELAMIN ABU ZEID

Thesis Submitted in Fulfilment of the Requirements for the Degree of Doctor of Philosophy in the Faculty of Science and Environmental Studies
Universiti Putra Malaysia

February 2001



#### **DEDICATION**

To the memory of my late father, MOHAMED ELAMIN who left us alone while I am conducting this study, and to my beloved wife GHADA and sons MOHAMED and RAZI who were the source of inspiration and encouragement throughout the period of this study.



Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

THE EFFECT OF CARBOFURAN AND ENDOSULFAN ON THE AFRICAN CATFISH, CLARIAS GARIEPINUS

By

ISAM ELDIN MOHAMED ELAMIN ABU ZEID

February 2001

Chairman: Assoc. Prof. Dr. Mohd. Arif Syed

Faculty: Science and Environmental Studies

This study was undertaken to determine the toxic effects of sub-lethal concentrations of carbofuran and endosulfan on some behavioural.

morphological, biaccumulation, biochemical, histopathological, and molecular

aspects of the freshwater African catfish, Clarias gariepinus. The toxicity of

carbofuran and endsulfan was ascertained by estimating the LC<sub>50</sub>.

The calculated 96-h LC<sub>50</sub> values of carbofuran and endosulfan for

juveniles of Clarias gariepinus were found to be 10.4 p.p.m and 21.6 p.p.b

respectively, under tropical condition. The test fish swam erratically, struggled to

breath, often swam to the surface, followed by loss of equilibrium. The color of

the skin became progressively pale during the period of the test, The liver

glutamate oxaloacetate transaminase (GOT), glutamate pyruvate transaminase

(GPT) and acetycholinesterase (AchE) were determined photometrically. Within

16 days of treatment, the activity levels of GOT and GPT were significantly

(p<0.05, p<0.01, p<0.001) increased by 203% and 121% for carbofuran and 167% and 195% for endosulfan respectively, whereas, AchE activity levels were inhibited following exposure to the test pesticides. With no exceptions, the *in vivo* and *in vitro* effects of carbofuran and enosulfan on GOT, GPT and AchE were qualitatively similar.

The pesticide concentration in the tissues was in the order of liver> intestine> gill> brain> muscle. The highest concentration of the test pesticides (353.47 µg/g carbofuran and 1409.35 ng/g endosulfan) were found in the liver 24 hours after treatment. Liver lesions were observed following exposure to both pesticides. The evidence of pesticide accumulation was elucidated by necrosis and damaged fish liver.

The depletion in the protein content was observed following exposure to carbofuran and endosulfan. The RNA concentrations were significantly decreased in *Clarias gariepinus* exposed to both pesticides, whereas, DNA concentrations tend to remain constant. The DNA molecular weight of the control fish was found to be about 16832 bp, whereas, the DNA molecular weights of carbofuran and endosulfan exposed fish were 14505 and 14505 bp respectively. The liversomatic index decreased to 2.11 for carbofuran and 1.59 for endosulfan in comparison to control. The biochemical, molecular, bioaccumulation, histological, behavioural and morphological techniques employed in this study may be used to detect and assess any pesticidal pollution in the aquatic environment at an early stage of pollution.



Abstrak tesis dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Doktor Falsafah

KESAN KETOKSIKAN OLEH ENDOSULFAN DAN KARBOFURAN KE ATAS IKAN KELI AFRIKA. CLARIAS GARIEPINUS

Oleh

ISAM ELDIN MOHAMED ELAMIN ABU ZEID

Februari 2001

Pengerusi: Prof. Madya Dr. Mohd. Arif Syed

Fakulti: Sains dan Pengajian Alam Sekitar

Kesan ketoksikan endosulfan dan karbofuran ke atas perlakuan,

morfologi, bioakumulasi, biokimia, histopathologi dan aspek molekul telah dikaji

pada ikan keli Afrika dengan memberikan rawatan sublethal. Ketoksikan

endosulfan dan karbofuran telah ditentukan nilainya dengan LC<sub>50</sub>.

Nilai LC<sub>50</sub> bagi rawatan 96 jam karbofuran dan endosulfan bagi Clarias

gariepinus didapati masing-masing adalah 10.4 ppm dan 21.6 ppb. Ikan didapati

menyelam tidak tentu hala, menghadapi masalah pernafasan, kerap muncul ke

permukaan dan hilang daya imbangan. Semasa ujikaji, warna kulit dilihat

menjadi semakin pucat. Pada hati, enzim glutamat oxaloacetat transaminase

(GOT), glutamat piruvat transaminase (GPT) dan asetilkolinesterase (AchE)

ditentukan secara fotometrik. Semasa 16 hari rawatan, paras aktiviti GOT dan

GPT adalah signifikan (p<0.05, p<0.01, p<0.001). Paras masing-masing telah

meningkat sebanyak 203% dan 121% untuk karbofuran, 167% dan 195% untuk

endosulfan. Aktiviti AchE pula didapati terencat akibat pendedahan kepada racun

v

perosak tersebut. Dari permerhatian in vivo dan in vitro, kesan karbofuran dan endosulfan ke atas GOT, GPT dan AchE adalah sama secara kualilitatif.

Kandungan racun perosak pada tisu adalah tinggi menurut turutan hati>usus>insang>otak>otot. Kandungan tertinggi racun perosak (353.47μg/g karbofuran dan 1409.35ng/g endosulfan) di dalam hati, selepas 24 jam rawatan. Pemerhatian kerosakan hati telah dilakukan selepas dirawat racun perosak. Kesan pengumpulan racun perosak telah berjaya dibuktikan melalui kerosakan dan nekrosis pada hati tersebut.

Kandungan protein didapati berkurang pada ikan yang telah dirawat dengan carbofuran dan endosulfan. Jumlah RNA juga didapati menurun pada Clarias gariepinus yang terdedah kepada kedua-dua racun perosak ini manakala kepekatan DNA didapati tidak ada sebarang perubahan. Berat molekul DNA bagi kawalan adalah 16832 bp dan ikan terdedah kepada endosulfan dan carbofuran masing-masing berat molekulnya adalah 14505bp. Nisbah berat badan ikan kepada berat hati jika dibandingkan dengan kawalan didapati telah menurun kepada 2.11 bagi carbofuran dan 1.59 bagi endosulfan.

Kesimpulannya, teknik biokimia, molekul, bioakumulasi, histologikal, perlakuan dan morfologikal dapat digunakan bagi mengesan dan mengenalpasti pencemaran racun perosak di dalam persekitaran akuatik terutama di peringkat awal pemcemaran berlaku.



#### **ACKNOWLEDGEMENTS**

First and foremost, I would like to express my most sincere gratitude and deep appreciation to the chairperson of my supervisory committee, Assoc. Prof. Dr. Mohd. Arif Syed for his guidance, encouragement, objective and enthusiastic discussion, keen interest, and support throughout the course of the study.

Words are not enough to express my heartfelt thanks to the members of my supervisory committee, Assoc. Prof. Dr. Nor Aripin Shamaan, Assoc. Prof. Dr. Johari Ramli and Assoc. Prof. Dr. Juzu Hayati Arshad for their careful supervision, invaluable advice, constructive suggestions, review of my work during the period of this study, and for extending their time and effort to held a weekly research group meeting with us.

I am also indebted to Assoc. Prof. Dr. Nashriyah Mat, Malaysian Institute of Nuclear Technology (MINT) for providing some of the facilities for the bioaccumulation study. Thanks are also extended to the technical staff of the Laboratory of Aquatic Resources Technology, Institute of Bioscience, UPM, particularly, Muhamed, Jasni, Krishnan, Zaidi and Nordin for their valuable assistance

Furthermore, I would like to acknowledge the staff of the Electron Microscopy Unit, for their help especially Dr. Fawziah Othman and Assoc. Prof. Dr. Mohd. Hair Bejo of the Faculty Veterinary Medicine, UPM for their valuable



comments on the histopathological work. I also appreciate the assistance of the staff of Genome Centre, Institute of Bioscience particularly, Dr. Tan Siang Hee and Musliyana Mansor. Special thanks also to the technical staff of the Department of Biochemistry and Microbiology, UPM; Khalid, Ibrahim, Mohd. Zamros, Onn and Husain. The financial supports given by the Malaysian Government through Intensification of Research in Priority Areas (IRPA), grant no. 01-02-04-078 is gratefully acknowledged.

Thanks are extended to my fellow graduate students for their moral support and hospitality. Special words of thanks to my friends, Yunus, Jeeven, Parichehr and Arif, for their help and assistance. Thanks are due to all who have helped or contributed in one way or other towards the completion of this study.

Last but definitely not the least, I wish to express my deepest and heartfelt feelings to my mother, brothers, and sisters, for providing me with guidance and support since my childhood and understanding of my ambition. My heartfelt appreciation to my wife Ghada for being supportive, enduring with patience, and understanding, the many years we spent away from home. I am very grateful to my sons, Mohamed and Razi for the incredible joy, and privilege, they give me while I am doing this study

And above all to Almighty ALLAH, the Merciful and Benevolent.

I certify that an Examination Committee met on 2<sup>nd</sup> February 2001 to conduct the final examination of Isam El Din Mohamed El Amin Abu Zeid on his Doctor of Philosophy thesis entitled "The Effect of Carbofuran and Endosulfan on the African Catfish, *Clarias gariepinus*" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

#### ABU BAKAR SALLEH, Ph.D,

Professor Faculty of Science and Environmental Studies, Universiti Putra Malaysia (Chairman)

#### MOHD. ARIF SYED, Ph.D,

Associate Professor, Faculty of Science and Environmental Studies, Universiti Putra Malaysia. (Member)

#### NOR ARIPIN SHAMAAN, Ph.D,

Associate Professor, Faculty of Science and Environmental Studies, Universiti Putra Malaysia. (Member)

#### JOHARI RAMLI, Ph.D,

Associate Professor/ Deputy Dean, Faculty of Science and Environmental Studies, Universiti Putra Malaysia. (Member)

#### JUZU HAYATI ARSHAD, Ph.D,

Associate Professor, Faculty of Science and Environmental Studies, Universiti Putra Malaysia. (Member)

#### THOMAS WILLIAM JORDAN, Ph.D,

Professor School of Biological Sciences Victoria University of Wellington New Zealand (Independent Examiner)

> MOHD GHAZALI MOHAYIDIN, Ph.D Professor/Deputy Dean of Graduate School

Date: 0 8 FEB 2001

This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirement for degree of Doctor of Philosophy.

MOHD GHAZALI MOHAYIDIN, Ph.D.

Professor

Deputy Dean of Graduate School Universiti Putra Malaysia

Date: - 12 APR 2001.



#### **DECLARATION**

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

Isam El Din Mohamed El Amin

Date:



## TABLE OF CONTENTS

			Page
		ION	
		T	
		LEDGEMENTS	
		AL SHEETS	
		ATION FORM	
		ABLES	
		IGURES	
LIST	OF A	BBREVIATIONS	XX
CHAF	PTER		
	I	INTRODUCTION	1
	TT	LITED ATTIDE DEVIEW	4
	II	LITERATURE REVIEW	
		Pesticides Problems in Malaysia	
		Pesticide Usage in Sudan	
		Toxicity of Pesticides to Fish	
		Behavioural Indicators of Pesticide Stress	
		Morphological Changes	
		Biochemical Changes	
		Effect of Pesticides on Fish Enzymes	9
		Effect of Pesticides on other Biochemical	10
		Parameters in Fish	
		Histological Changes induced by Pesticides in Fish	
		Molecular Changes.	
		Bioaccumulation of Pesticides in Fish	10
		Techniques for Assessment of Stress on Aquatic	17
		Organisms	
		Life Cycle Tests.	
		Toxicity Tests	18
		Biological Markers and their Advantages	0.1
		in Environmental Monitoring	
		Effect of Pesticides on Catfish.	23
		Effect of Environmental Pollutants on African Catfish,	0.7
		Claris gariepinus	
		Clarias gariepimus as Test Organism	
		Carbofuran as Test Chemical and Fish Toxicity	30
		The Toxic Effects of Endosulfan on Fish	
		Acetylcholinesterase (AchE) Inhibition	37
		Mechanism of Inhibition of AchE by Carbamate and	



	Organophosphorus Insecticides	
	Mechanism of Inhibition of AchE by Organochlorine	
	Pesticides	4
	Alteration in Transaminase Enzymes as	
	Indicators of Pesticides Stress.	
	Statement of the Problem	
	Significance of the Study	
	Objectives of the Study	50
Ш	GENERAL MATERIALS AND METHODS	<b>5</b> 1
	Experimental Design	
	Acclimatisation of the Test Organism	52
	Water Quality Monitoring	52
	Experimental Set up	
	Analysis of Different Parameters	
	Data Collection.	55
	Statistical Analysis	56
ΙV	COMPARATIVE TOXICITY, BEHAVIOURAL	
	AND MORPHOLOGICAL CHANGES INDUCED	
	BY CARBOFURAN AND ENDOSULFAN IN THE	
	AFRICAN CATFISH, CLARIAS GARIEPINUS	57
	Introduction	
	Materials and Methods	
	Test Animal and Water.	
	Preliminary Toxicity Experiments	
	Test Procedures.	
	Calculation of the LC <sub>50</sub> Values	
	Results.	
	Discussion.	
	Conclusion.	
V	BIOACCUMULATION OF RADIOLABELLED	
•	CARBOFURAN AND ENDOSULFAN IN DIFFERENT	
	TISSUES OF AFRICAN CATFIS, CLARIAS	
	GARIEPINUS	72
	Introduction.	
	Materials and Methods	
	Chemicals	
	Test Organisms	
	Test System and Sampling.	
	Determination of Pesticide Residues in Fish Tissues.	
	Statistical Analysis	
	Results	
		/O Q1



	Conclusion	86
VI	ENZYME MODULATION BY CARBOFURAN	
	AND ENDOSULFAN IN THE LIVER OF AFRICAN	
	CATFISH, CLARIAS GARIEPINUS	87
	Introduction.	
	Materials and Methods.	
	Test Fish.	
	Chemicals	
	Test Procedures	
	Preparation of Tissue Homogenate	
	Assay of the Test Enzymes	
	Statistical Analysis	
	Results	92
	GOT, GPT and AchE Activity Levels	
	of Control Fish	
	GOT Activity Levels in Fish Exposed to Pesticides	
	GPT Activity Levels in Fish Exposed to Pesticides	
	Effect of Pesticides on AchE Activity	102
	Discussion	107
	Conclusion	114
VII	HISTOLOGICAL CHANGES INDUCED BY	
	CARBOFURAN AND ENDOSULFAN IN THE	
	LIVER OF AFRICAN CATFISH, CLARIAS	
	GARIEPINUS	115
	Introduction	
	Materials and Methods.	
	Test Fish	
	Light Microscopy	
	Transmission Electron Microscopy	
	Results	
	Light Microscopy	
	Transmission Electron Microscopy	
	Discussion	
	Conclusion.	126
VIII	MOLECULAR CHANGES INDUCED BY	
	CARBOFURAN AND ENDOSULFAN IN THE	
	LIVER OF AFRICAN CATFISH, CLARIAS	
	GARIEPINUS	127
	Introduction	
	Materials and Methods	
	Analysis of Different Parameters	
	Statistical Analysis.	
	Results	130



	Discussion	. 134
	Conclusion	. 136
IX	GENERAL DISCUSSION AND CONCLUSION	. 137
	BIBLIOGRAPHY	. 147
	APPENDICES	. 187
	Tissue Processor	188
	BIODATA OF THE AUTHOR	189



## LIST OF TABLES

Γable		Page
1	LC50 of some pesticides for different species of catfish.	. 24
2	Carbofuran acute toxicity (LC50) for some fish species.	32
3	Endosulfan acute toxicity (LC50) for some fish species.	36
4	Percentage of dead fish, empirical probits, and approximate expected probits for the estimation of LC <sub>50</sub> of carbofuran for C. gariepinus	64
5	Percentage of dead fish, empirical probits, and approximate expected probits for the estimation of LC <sub>50</sub> of endosulfan for <i>C. gariepinus</i>	64
6	Bioaccumulation (μg/g dry wt.) of radiolabelled carbofuran in different tissues of C. griepinus	. 78
7	Bioaccumulation (ng/g dry wt.) of radiolabelled endosulfan in different tissues of C. gariepinus	78
8	In vivo effect of sub-lethal concentrations of carbofuran and endosulfan on the activity levels (U/l) of GOT in the liver of C. gariepinus.	94
9	In vivo effect of sub-lethal concentrations of carbofuran and endosulfan on the activity levels (U/l) of GPT in the liver of C. gariepinus.	96
10	In vitro effect of sub-lethal concentrations of carbofuran and endosulfan on the activity levels (U/l) of GOT in the liver of C. gariepinus at different incubation time.	98
11	In vitro effect of sub-lethal concentrations of carbofuran and endosulfan on the activity levels (U/I) of GPT in the liver of C. gariepimus at different incubation time.	100
12	In vivo effect of sub-lethal concentrations of carbofuran	



	and endosultan on the activity levels (µmole/min/mg protein) of AchE in the liver of C. gariepinus
13	In vitro effect of sub-lethal concentrations of carbofuran and endosulfan on the activity levels (µmole/min/mg protein) of AchE in the liver of C. gariepinus at
	different incubation time
14	Effect of carbofuran and endosulfan on total protein (mg/g wet wt.), total RNA and DNA (μg/g wet wt.), RNA/DNA ratio and liver somatic index in the liver
	of C. gariepinus at 16 days exposure132
15	Transformation of percentage to probits
16	Tissue processing by an automatic tissue processor



## LIST OF FIGURES

Figur	e	Page
1	Schematic diagram of the possible directions of movement and fates of a pollutant after it has been absorbed into the bloodstream of fish.	17
2	Chemical formula of carbofuran	31
3	Chemical formula of endosulfan	35
4	Diagrammatic representation of the reaction of acetylcholinesterase with its normal substrate and organophospahte or carbamate inhibitors	42
5	Determination of the LC50 of carbofuran for C. gariepinus	65
6	Determination of the LC50 of endosulfan for C. gariepinus.	66
7	Morphology of the African catfish, C. gariepinus (control)	67
8	Morphological changes in the African catfish, C. exposed to carbofuran	67
9	Morphological changes in the African catfish,  C. gariepinus exposed to endosulfan	67
10	Biaccumulation of radiolabelled carbofuran in different tissues of C. gariepimus.	79
11	Biaccumulation of radiolabelled endosulfan in different tissues of <i>C. gariepinus</i>	80
12	In vivo effects of carbofuran and endosulfan on the GOT activity levels in the liver of C. gariepinus.	95
13	In vivo effects of carbofuran and endosulfan on the GPT activity levels in the liver of C. gariepinus.	97
14	In vitro effects of carbofuran and endosulfan on the GOT	



	activity levels in the liver of C. gariepinus	99
15	In vitro effects of carbofuran and endosulfan on the GPT activity levels in the liver of C. gariepinus	10
16	In vivo effects of carbofuran and endosulfan on the AchE activity levels in the liver of C. gariepinus	104
17	In vitro effects of carbofuran and endosulfan on the AchE activity levels in the liver of C. gariepinus	106
18	Light micrograph of control C. gariepinus liver	121
19	Light micrograph of carbofuran exposed C. gariepinus liver	121
20	Light micrograph of endosulfan exposed C. gariepinus liver	. 122
21	Electron micrograph of control C. gariepimus liver	. 123
22	Ultrastructural changes in the liver of C. gariepinus exposed to carbofuran	123
23	Ultrastructural changes in the liver of C. gariepinus exposed to endosulfan	124
24	Effect of pesticides on DNA pattern in the liver	133



#### LIST OF ABBREVIATIONS

AchE Acetylcholinesterase

ANOVA Analysis of variance

Bp Base-pair

DDVP Dichlorvos

DO Dissolved oxygen

EDB Ethylene dibromide

EROD Ethoxyresorufin O-deethylase

GOT Glutamate oxaloacetate transaminase

GPT Glutamate pyruvate transaminase

H & E Hematoxylin and Eosin

H Hepatic cell

HCH Hexachlorocyclohexane

IAEA International Atomic Energy Agency

LC<sub>50</sub> Lethal concentration that results in 50% death.

LSC Liquid scintillation counter

LSI Liver-somatic index

MATC Maximum acceptable toxicant concentration

MFO Mixed function oxygenase

N Nucleus

NC Necrotic cell



NRC National Research Council

NRCC National Research Council of Canada

NU Nucleolus

PCB Polychlorinated biphenyl

PCP Pentachlorophenol

SAC Safe application concentration

SD Standard deviation

SDS Sodium dodecylsulfate

SEM Scanning electron microscope

TCDD Tetrachlorodibenzo-p-dioxin

TEM Transmission electron microscope

UPM Universiti Putra Malaysia



#### CHAPTER I

#### INTRODUCTION

The growing demand for increased food productivity to meet the needs of the global population has led farmers to use sophisticated agricultural technology in which pesticides play a crucial role. Pesticide use has a positive and dramatic impact on agricultural production through protection of crops against insects, pests and diseases. The extensive use of pesticides in agriculture has given rise to criticisms in recent years, due to their persistent nature in the environment (Hernandez et al., 1993), and accumulation in different tissues of plants (Kaplan, 1999), animals (Miao et al., 2000) and human beings (Saleh et al., 1998). Therefore, the use of pesticides is a mixed blessing, while their benefits for preventing crop losses (Kacew et al., 1996) and saving human lives (Emerson et al., 1999) are well recognised, they very often result in unwanted side effects. Therefore, the identification of pesticides which are effective against pests and at the same time relatively safe to human and non-target organism is of considerable importance.

Among the numerous environmental impact of the application of pesticides in agriculture are their undesirable effect on the aquatic fauna of freshwater courses (Barlas, 1999). Pesticides have the potential to enter the aquatic environment by direct spraying or broadcast of granular formulations,



drift deposition of sprayable formulations, and in runoff water from treated field (Sharma, 1990). Redeposition from the atmosphere is another route of entry, as some studies have shown the presence of pesticides in rain water (Richards et al., 1987). The pollution of the aquatic environment by pesticides is known to pose a constant threat to fish by altering their habitat, behavioural pattern, growth, and reproduction (Jarvian et al., 1977). Fish is extremely sensitive to pollutants and exhibit a very high bioaccumulation rate of dissolved chemicals relative to their concentration (Al-Yousuf et al., 2000). Fish may accumulate pollutants and pass them to human beings through food causing chronic or acute diseases (Adeyeye et al., 1996). The importance of fish as one of the major source of cheap and available protein-rich food for human being is recognised (Begum and Vijayaraghavan, 1996). The nutritional value of different fishes depends on their biochemical composition; protein, amino acids, vitamins, and mineral contents (Ganeson et al., 1989). Pesticides affect these biochemical composition and may cause biochemical and physiological changes in different fish tissues (Ramaswamy et al., 1999).

Due to increasing pesticide applications, it has become necessary to evaluate their hazards and develop biological indicators of aquatic contamination. Various approaches have been used to evaluate or predict the effects of environmental stress on fish. The most common of these are laboratory tests of acute and chronic toxicity. Although these approaches are valuable for achieving such objectives as formal water quality criteria (Adams, 1990), however, they



lack ecological realism (Cairns, 1981). Recently, researchers used the biological markers approach to detect the effect of pollutants in the environment (Hoque, 1997). Biological markers permit the detection of stress-related factors that are biologically and ecologically variable. They provide early signals of adverse ecological effects as they use the lower levels of biological organisation, evaluate the specific response for each type of environmental stress as well as the overall integrated response, and can predict and evaluate the ecological significance and chronic stress (Adams, 1990). Changes in fish such as behavioural (Rice et al., 1997), morphological (Richmond and Dutta, 1992), biochemical (Juzu et al., 1998; Abu Zeid et al., 1997), histological (Dhanapaleiam and Premlatha, 1994), and molecular (Thomas, 1990) have been attributed to pesticides. These changes have the potential to be used as possible biological markers for the assessment of pollution in the aquatic environment.

