



UNIVERSITI PUTRA MALAYSIA

**ANTIOXIDANT AND VASCULAR RELAXATION ACTIVITIES OF TROPICAL
PLANT EXTRACTS**

IRINE RUNNIE ANAK HENRY GINJOM

FSMB 2003 17

**ANTIOXIDANT AND VASCULAR RELAXATION ACTIVITIES OF TROPICAL
PLANT EXTRACTS**

IRINE RUNNIE ANAK HENRY GINJOM

**MASTER OF SCIENCE
UNIVERSITI PUTRA MALAYSIA**

**ANTIOXIDANT AND VASCULAR RELAXATION ACTIVITIES OF TROPICAL
PLANT EXTRACTS**

By

IRINE RUNNIE ANAK HENRY GINJOM

**Thesis Submitted in Fulfillment of the Requirement for the Degree of Master of
Science in the Faculty of Food Science and Biotechnology
Universiti Putra Malaysia**

April 2003

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science.

ANTIOXIDANT AND VASCULAR RELAXATION ACTIVITIES OF TROPICAL PLANT EXTRACTS

By

IRINE RUNNIE ANAK HENRY GINJOM

April 2003

Chairman : Professor Suhaila Mohamed, Ph.D.

Faculty : Food Science and Biotechnology

In this study, the methanolic extracts of twelve plants which are the leaves of noni (*Morinda citrifolia* L.), mamon (*Gynandropsis gynandra* L.), cashew (*Anacardium occidentale* L.), sweet potato (*Ipomoea batatas* L.), papaya (*Carica papaya* L.), mint (*Mentha arvensis* L.), semambu (*Azadirachta indica*), betel (*Piper betle* L.); lemongrass stalk (*Cymbopogon citrates* Stapf.), bird chili fruit (*Capsicum frutescens* L.), roselle calyx (*Hibiscus sabdariffa*), and oil palm frond (*Elaeis guineensis* Jacq.) were analyzed for *in vitro* antioxidant and vascular activities. Their total phenolic content was also estimated. Plant extract showing high antioxidant and vascular relaxing activities were further tested for their *in vivo* antioxidant and cholesterol lowering effects in hypercholesterolemic New Zealand White rabbits.

In vivo antioxidant test (ferric reducing/antioxidant potential [FRAP] assay) showed the highest activity in cashew leaf extract (4.3 mmol FRAP/g d.w.) and the lowest in lemongrass extract (0.2 mmol FRAP/g d.w.). The antioxidant activity of the cashew leaf extract was equivalent to those of green tea extract while lemongrass extract's antioxidant activity was equivalent to those of purified apigenin. The total phenols ranged from 55-256 mg gallic acid equivalent / g dry weight (d.w.) extracted samples with the highest concentration in oil palm frond extract. In this study, the antioxidant activity of the extracts did not correlate with the total phenolic contents, mainly due to the heterogeneity of the samples tested.

For vascular activity, sweet potato, betel, cashew, mamon, mint, oil palm frond, semambu, bird chili, and papaya extracts showed more than 50 % relaxing effects in Wistar Kyoto rat's isolated aortic ring preparation. Most of the relaxing effects were endothelium-dependent with nitric oxide as the main mediator. Prostacyclin and endothelium-derived hyperpolarizing factors (EDHF) effects were also observed in small amounts. For the smaller vessels, mesenteric arteries, the extent of relaxation is reduced, where only oil palm frond, betel and lemongrass extracts showed more than 50 % relaxing effect. It is observed that in the smaller vessel, the EDHFs effects are more prominent, especially in the betel and lemongrass extracts.

Oil palm frond was selected for the *in vivo* study for its high phenolic content and high vascular relaxing activities in both blood vessel preparations. This study showed that oil palm frond supplementation in the diets of hypercholesterolemic rabbits led to a delayed

increment in serum total cholesterol levels. There was no significant antioxidant and toxicity effect on liver, kidney and muscle observed.

This study shows that most of the selected plants possess a high antioxidant and vascular relaxing activities. Antioxidants in food are important in minimizing food deterioration i.e. rancidity which affect the taste and textures of food containing lipids. In human, dietary antioxidants help to reduce oxidative stress, which is associated with the etiology of several chronic diseases such as hypertension, cancer, aging diseases, atherosclerosis and coronary artery diseases. Similarly, consumption of plants possessing vascular relaxing activities may restore and/or improve vascular functions and hence protect against vascular diseases such as hypertension.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Master Sains

**AKTIVITI ANTIOSIDAN DAN VASODILATOR EKSTRAK TUMBUHAN
TROPICAL**

Oleh

IRINE RUNNIE ANAK HENRY GINJOM

April 2003

Pengerusi : Profesor Suhaila Mohamed, Ph.D.

Fakulti : Sains Makanan dan Bioteknologi

Dalam kajian ini, ekstrak metanol dua-belas tumbuhan iaitu daun noni (*Morinda citrifolia* L.), mamon (*Gynandropsis gynandra* L.), gajus (*Anacardium occidentale* L.), keledak (*Ipomoea batatas* L.), betik (*Carica papaya* L.), pudina (*Mentha arvensis* L.), semambu (*Azadirachta indica*) dan sirih (*Piper betle* L.); batang serai (*Cymbopogon citrates* Stapf.), buah cili burung (*Capsicum frutescens* L.) kelopak roselle (*Hibiscus sabdariffa*) dan daun kelapa sawit (*Elaeis guineensis* Jacq.) telah diuji untuk aktiviti-aktiviti antioksidan dan vaskular *in vitro*. Jumlah kandungan fenol juga dianggarkan. Ekstrak tumbuhan yang menunjukkan aktiviti antioksidan dan vaskular yang tinggi selanjutnya diuji untuk kesan antioksidan dan penurunan kolesterol *in vivo* dalam arnab New Zealand White.

Ujian antioksidasi *in vivo* (kajian ferric reducing/antioxidant potential [FRAP]) menunjukkan aktiviti yang tertinggi dalam ekstrak daun gajus (4.3 mmol FRAP/g berat kering [b.k]). Aktiviti antioksidasi ekstrak daun gajus adalah setara dengan aktiviti yang ditunjukkan oleh ekstrak teh hijau manakala aktiviti antioksidasi ekstrak serai adalah setara dengan yang terdapat pada apigenin. Jumlah kandungan fenol pula mempunyai julat di antara 55-256 mg GAE (gallic acid equivalent) / g b.k. dengan kandungan yang tertinggi dalam ekstrak daun kelapa sawit. Dalam kajian ini, aktiviti antioksidasi ekstrak-ekstrak dengan kandungan fenol tidak menunjukkan sebarang korelasi, yang berkemungkinan besar disebabkan oleh kepelbagaian sampel tumbuhan.

Untuk aktiviti vaskular, ekstrak daun keledak, sirih, gajus, pudina, kelapa sawit, semambu, buah cili dan daun betik menunjukkan kesan relaksasi yang melebihi 50 % dalam penyediaan gelung aorta tikus Wistar Kyoto. Kebanyakan kesan relaksasi adalah bergantung kepada kehadiran endotelium yang mana nitrik oksida merupakan pengantara utama. Sebilangan kecil daripada kesan relaksasi adalah disebabkan oleh prostasiklin dan faktor hiperpolarisasi endotelium (EDHF). Untuk arteri mesenterik, iaitu salur darah yang lebih kecil, kesan relaksasi berkurangan jika dibandingkan dengan gelung aorta, dengan hanya ekstrak daun kelapa sawit, sirih dan serai yang menunjukkan kesan relaksasi melebihi 50 %. Pemerhatian menunjukkan bahawa dalam salur darah yang lebih kecil, kesan EDHF adalah lebih ketara, terutamanya bagi ekstrak sirih dan serai.

Daun kelapa sawit telah dipilih untuk kajian *in vivo* kerana ia mempunyai kandungan fenol dan kesan relaksasi vaskular dalam kedua-dua penyediaan salur darah. Dalam kajian ini, penambahan daun kelapa sawit ke dalam makanan amab yang

hiperkolesterolemik menjurus kepada penangguhan dalam penambahan paras kolesterol di dalam serum. Tiada kesan antioksidan dan ketoksikan yang signifikan ke atas hati, buah pinggang, dan otot ditunjukkan.

Kajian ini menunjukkan yang kebanyakan tumbuhan yang dipilih mempunyai aktiviti-aktiviti antioksidan dan relaksasi vaskular yang tinggi. Antioksidan di dalam makanan adalah penting untuk meminimalkan kerosakan makanan iaitu ketengikan yang mempengaruhi rasa dan tekstur makanan yang mengandungi lemak. Pada manusia, antioksidan dari makanan membantu mengurangkan tekanan oksida, yang dikaitkan dengan pembentukan beberapa jenis penyakit kronik seperti hipertensi, barah, penyakit yang berkaitan dengan penuaan, aterosklerosis dan penyakit koronari arteri. Sehubungan dengan itu, pemakanan yang mengandungi tumbuhan yang mempunyai aktiviti relaksasi vaskular yang tinggi mungkin dapat mengekalkan dan/atau memperbaiki fungsi vaskular dan seterusnya memberi perlindungan daripada penyakit vaskular seperti hipertensi.

ACKNOWLEDGEMENTS

This work could never been completed without these people. First and foremost, I would like to thank Professor Dr. Suhaila Mohamed for her excellent supervisory. I was also privileged for having such a supportive team of co-supervisors namely Associate Professor Dr. Noordin Mohamed Mustapha, Associate Professor Dr. Radzali Muse, Associate Professor Dr. Azizah Hamid, Associate Professor Dr. Hapizah Nawawi and Dr. Mahinda Abeywardena. I am thankful for all of them.

I would also like to acknowledge UPM for without their financial supports, this study would never come to realization. To all the laboratory assistants and staffs in the Food Science Biotechnology, UPM, thank you very much for your assistance. I am also especially grateful to CSIRO Health Sciences & Nutrition, Adelaide which had offered me a place for the 18-month technical training in the first place. The technical, research, academic, moral and financial supports provided had been very much appreciated. Not only that, I am also indebted by Dr. Mahinda Abeywardena and his co-researchers' kindness and helpfulness. To all my colleagues, thank you for your support and friendship. To all my friends, thank you for your understanding and help and also for bearing with me for all these times. A big thank you goes to my parents, brothers and relatives that has supported me all the time. To others that is too many to mention, thank you for being there.

Finally, I thank God for restoring me physically, mentally and spiritually during this long yet adventurous road to obtain the scroll. There is no doubt I have learned a lot from it.

I certify that an Examination Committee has met on 15th April 2003 to conduct the final examination of Irine Runnie ak Henry Ginjom on her Master of Science thesis entitled “Antioxidant and Vascular Relaxation Activities of Tropical Plant Extracts” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Nazamid Saari, Ph.D.

Associate Professor
Representative from Graduate School Office
(Chairman)

Suhaila Mohamed, Ph.D.

Professor
Faculty of Food Science and Biotechnology
Universiti Putra Malaysia
(Member)

Azizah Hamid, Ph.D.

Associate Professor
Faculty of Food Science and Biotechnology
Universiti Putra Malaysia
(Member)

Radzali Muse, Ph.D.

Associate Professor
Faculty of Sciences and Environmental Studies
Universiti Putra Malaysia
(Member)

Hapizah Mohd. Nawawi, MRCPATH.

Associate Professor
Faculty of Medicine
Universiti Kebangsaan Malaysia
(Member)



.....
GULAM RUSUL RAHMAT ALI, Ph.D.,
Professor / Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia.

Date: 17 JUL 2003

This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfillment of the requirements for the degree of Master of Science. The members of the Supervisory Committee are as follows:

Suhaila Mohamed, Ph.D.

Professor
Faculty of Food Science and Biotechnology
Universiti Putra Malaysia
(Chairman)

Radzali Muse, Ph.D.

Associate Professor
Faculty of Science and Environmental Studies
Universiti Putra Malaysia
(Member)

Azizah Hamid, Ph.D.

Associate Professor
Faculty of Food Science and Biotechnology
Universiti Putra Malaysia
(Member)

Noordin Mohamed Mustapha, Ph.D.

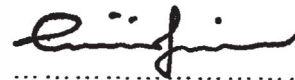
Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Hapizah Mohd. Nawawi, MRCPPath.

Associate Professor
Faculty of Medicine
Universiti Kebangsaan Malaysia
(Member)

Mahinda Abeywardena, Ph.D.

CSIRO Health Sciences and Nutrition
Adelaide, Australia
(Member)



.....
AINI IDERIS, Ph.D.
Professor / Dean
School of Graduate Studies
Universiti Putra Malaysia.

Date: **15** AUG 2003

DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.


.....

IRINE RUNNIE AK HENRY GINJOM

Date: 14/8/2003

TABLE OF CONTENTS

	Page
ABSTRACT.....	ii
ABSTRAK	v
ACKNOWLEDGEMENTS	viii
APPROVAL	ix
DECLARATION.....	xi
TABLE OF CONTENTS.....	xii
LIST OF TABLES.....	xv
LIST OF FIGURES.....	xix
LIST OF ABBREVIATIONS.....	xxi

CHAPTER

I	INTRODUCTION.....	1
	Background.....	1
	Research Objectives.....	3
II	LITERATURE REVIEW.....	5
	Introduction.....	5
	Free Radicals in Health and Diseases.....	6
	Endogenous Sources of Free Radicals.....	7
	Exogenous Sources of Free Radicals.....	9
	Damaging Reactions of Free Radicals.....	11
	Antioxidant Defense System.....	13
	Enzymatic Antioxidants.....	14
	Non-enzymatic Antioxidants.....	15
	Other Antioxidants.....	17
	Flavonoids.....	19
	Structure and Distribution.....	19
	The Antioxidant and Cardio-protective Effects of Flavonoids..	20
	Endothelial Function and Vascular Health.....	23
	Vascular Endothelial Cells.....	23
	Endothelium-derived Vasoactive Factors.....	25
	Endothelial Dysfunction in Diseased Conditions.....	29
	Therapeutic Strategies.....	34
	Health Claims on Some of the Plant Sample Selected.....	35
	Cashew (<i>Anacardium occidentale</i> L.).....	35
	Mint (<i>Mentha arvensis</i> L.).....	35
	Oil palm (<i>Elaeis guineensis</i> Jacq.).....	36
	Roselle (<i>Hibiscus sabdariffa</i>).....	36

	Maman (<i>Gynandropsis gyantra</i> L. Brig.).....	36
	Papaya (<i>Carica papaya</i> Linn.)	36
	Noni (<i>Morinda citrifolia</i> L.)	37
	Lemongrass (<i>Cymbopogon citrates</i> Stapf.)	37
	Sweet potato (<i>Ipomoea batatas</i> L.)	38
III	ANTIOXIDANT ACTIVITY, TOTAL PHENOLIC CONTENT AND FLAVONOID CONTENT OF SELECTED PLANT EXTRACTS.....	39
	Introduction.....	39
	Material and Methods.....	40
	Plants Sample.....	40
	Chemicals and Solvents.....	41
	Preparation of Plant Extract.....	41
	FRAP Assay – <i>In Vitro</i> Measurement of Antioxidant Activity.....	42
	Determination of Total Polyphenolic Compounds.....	43
	Determination of Flavonoids by RP-HPLC with DAD.....	43
	Statistics.....	45
	Results.....	45
	Antioxidant Activity.....	45
	Amount of Total Phenolics.....	47
	Identification and Quantification of Selected Flavonoids.....	48
	Relationship Among Antioxidant Activities, Total Phenol Compounds and Individual Flavonoid Contents.....	50
	Discussions.....	50
IV	VASCULAR RELAXATION EFFECTS OF SELECTED PLANT EXTRACTS	56
	Introduction.....	56
	Material and Methods.....	57
	Preparation of Plant Extracts.....	57
	Drugs and Chemicals	57
	Vascular Studies.....	58
	Statistics.....	59
	Results.....	60
	Relaxing Effects on Aortic Ring Preparations.....	60
	Relaxing Effects on Mesenteric Artery Bed Preparations.....	64
	Comparison of Relaxation Responses in Different Vessel Preparations and to Different Antagonists	66
	Discussions.....	68
V	EFFECTS OF OIL PALM FROND SUPPLEMENTATION ON LIPID AND ANTIOXIDANT STATUS OF HYPERCHOLESTEROLEMIC RABBITS.....	76
VI	Introduction.....	76

Material and Methods.....	77
Plant Material.....	77
Chemicals and Solvents.....	78
Preparation of Animal Feed.....	78
Animal Study.....	80
Determination of Serum Lipids.....	80
Determination of Blood Antioxidant Enzymes.....	81
Determination of Serum and Liver MDA.....	83
Determination of Liver and Kidney Damage Biomarkers.....	85
Statistics.....	85
Results.....	86
Lipid Levels in Serum.....	86
Antioxidant Status.....	92
Body and Internal Organ Weights.....	97
Biochemical Markers for Liver, Kidney and Muscle Damages	99
Discussions.....	106
 GENERAL CONCLUSION.....	 113
 REFERENCES.....	 118
 APPENDICES.....	 135
A1 Ferric Reducing/Antioxidant Potential (FRAP) Assay.....	135
A2 Determination of Total Polyphenols Using Folin-Ciocalteu Method....	140
A3 Reversed-phase HPLC with Photo Diode Array Detection (DAD)..	142
A4 Aortic Rings Preparation.....	148
A5 Mesenteric Arterial Bed Preparation.....	152
A6 Pictures of Twelve Tropical Plants Used in This Study.....	158
B Original Data.....	159
 BIODATA OF THE AUTHOR.....	 162

LIST OF TABLES

Table	Page
2.1 Typical ROS formations <i>in vivo</i>	7
2.2 Structure of common flavonoid aglycones and glycosides.....	20
2.3 Antithrombotic properties of endothelial cells.....	29
3.1 The thirteen plants (common, generic and family) used in this study...	42
3.2 Antioxidant activity of selected plant extracts and flavonoids; and total phenolic content in plant extracts.....	46
3.3 Flavonoid content of green tea, oil palm frond and betel extracts as measured by the reversed-phase HPLC-DAD method.....	49
4.1 Maximum relaxation (%) of Ach, plant extracts and flavonoids in aortic rings pre-contracted with NA, NA+NOLA and NA without endothelium	61
4.2 Maximum relaxation (%) of flavonoids in aortic rings pre-contracted with NA, NA+NOLA and NA without endothelium.....	62
4.3 Loss of vascular relaxation activity (%) as compared to responses in preparations with intact endothelium, pre-contracted with NA.....	63
4.4 Maximum relaxation (%) of Ach, Hist, plant extracts and flavonoids in mesenteric arterial bed pre-contracted with NA and NA+NOLA+INDO	65
4.5 Maximum relaxation (%) of flavonoids in mesenteric arterial bed pre-contracted with NA and NA+NOLA+INDO	66
4.6 Loss of activity of cashew, betel and lemongrass extracts and the most likely relaxation mediator in aortic ring preparations.....	70
4.7 Loss of activity of cashew, betel and lemongrass extracts and the most likely relaxation mediator in MAB preparations.....	70
5.1 Ingredients and compositions of four different diets fed to rabbits in this study.....	79

5.2	Comparison of serum total cholesterol (TC) levels of rabbits before and after being fed with experimental diets for each group	87
5.3	Comparison of % delta TC of rabbits at 8 and 16 weeks between groups.....	87
5.4	Comparison of serum high-density lipoprotein cholesterol (HDL-C) levels of rabbits before and after being fed with experimental diets for each group.....	88
5.5	Comparison of % delta HDL-C of rabbits at 8 and 16 weeks between groups.....	89
5.6	Comparison of serum triglycerides (TG) levels of rabbits before and after being fed with experimental diets for each group	90
5.7	Comparison of % delta TG of rabbits at 8 and 16 weeks between groups.....	90
5.8	Comparison of TC/HDL-C ratio of rabbits before and after being fed with experimental diets for each group	91
5.9	Comparison of % delta TC/HDL-C of rabbits at 8 and 16 weeks between groups.....	92
5.10	Comparison of erythrocyte catalase activity of rabbits before and after being fed with experimental diets for each group.....	93
5.11	Comparison of % delta catalase activities of rabbits at 8 and 16 weeks between groups.....	93
5.12	Comparison of blood glutathione peroxidase activities (GSH-Px) of rabbits before and after being fed with experimental diets for each group.....	94
5.13	Comparison of % delta GSH-Px of rabbits at 8 and 16 weeks between groups.....	95
5.14	Comparison of liver and serum malonaldehyde (MDA) levels of rabbits before and after being fed with experimental diets for each group	96

5.15	Comparison of % delta serum MDA level of rabbits at 8 and 16 weeks between groups.....	96
5.16	Comparison of body weight of rabbits before and after being fed with experimental diets for each group.....	97
5.17	Comparison of % delta body weight of rabbits at 8 and 16 weeks between groups.....	98
5.18	Comparison of internal organ weight/body weight of rabbits between groups after being fed with experimental diets.....	98
5.19	Comparison of serum alanine aminotransferase (ALT) levels of rabbits before and after being fed with experimental diets for each group.....	100
5.20	Comparison of % delta ALT of rabbits at 8 and 16 weeks between groups.....	100
5.21	Comparison of serum gamma glutamyl transpeptidase (GGT) levels of rabbits before and after being fed with experimental diets for each group.....	101
5.22	Comparison of % delta GGT of rabbits at 8 and 16 weeks between groups.....	101
5.23	Comparison of serum urea levels of rabbits before and after being fed with experimental diets for each group.....	102
5.24	Comparison of % delta urea of rabbits at 8 and 16 weeks between groups.....	103
5.25	Comparison of serum creatinine levels of rabbits before and after being fed with experimental diets for each group.....	103
5.26	Comparison of % delta creatinine level of rabbits at 8 and 16 weeks between groups.....	104
5.27	Comparison of serum total creatine kinase (TCK) levels of rabbits before and after being fed with experimental diets for each group.....	105
5.28	Comparison of % delta TCK of rabbits at 8 and 16 weeks between groups.....	105

A.1	Cobas-Bio printout.....	137
A.2	Preparation of 0-40 mg / mL gallic acid.....	140
A.3	Retention times and the correlation (equations, R ² values) between flavonoid concentrations and peak area.....	145
A.4	Chemicals for preparation of 10x Krebs Henseleit solution.....	149
A.5	Preparation of 10-160 mg / mL KCl.....	153
A.6	Preparation of 0.1-1000 µg / mL NA.....	154
A.7	Preparation of 10 ⁻¹⁰ – 10 ⁻³ M Histamine.....	154
A.8	Preparation of flavonoids stock (10 ⁻³ M).....	155
B.1	Maximum relaxation (%) of plant extracts in aortic rings pre-contracted with NA, NA+NOLA and NA without endothelium.....	159
B.2	Maximum relaxation (%) of plant extracts in mesenteric arterial bed pre-contracted with NA and NA+NOLA+INDO.....	160
B.3	Maximum relaxation (%) of flavonoids in aortic rings pre-contracted with NA, NA+NOLA and NA without endothelium (-E).....	161
B.4	Maximum relaxation (%) of flavonoids in mesenteric arterial bed pre-contracted with NA and NA+NOLA+INDO.....	161

LIST OF FIGURES

Figure	Page
2.1	The sequence of ROS generation and the various sites of action of the different antioxidants..... 15
2.2	The generic structure of flavonoids..... 19
2.3	Cross-section of a normal blood vessel..... 24
2.4	Common determinants of endothelial-derived vasomotor tone..... 25
2.5	Blood vessel in normal, hypertensive and atherosclerotic states..... 30
3.1	RP-HPLC chromatogram of flavonoid standards at 330 nm..... 48
3.2	RP-HPLC chromatogram of flavonoid standards at 370 nm..... 49
4.1	Representative curves for the relaxing effect of cashew, □; mamon, ●; and sweet potato, Δ; in aortic rings with endothelium. Extracts were added cumulatively at 5 min intervals to organ baths containing 10 ⁻⁸ M NA-contracted aortic ring preparations. Resulting curves were means (n=4) ± S.E.M..... 62
4.2	Relaxation effects of cashew, betel and lemongrass extracts on aortic rings and mesenteric artery bed preparation incubated in various antagonists (NOLA and INDO). Bars indicate means ± S.E.M..... 67
4.3	Correlations between various parameters of plant extracts..... 72
A.1	Plot of Fe ²⁺ standards reaction kinetics..... 138
A.2	Reaction kinetics plots of plant extracts. Abbreviation OPF represents oil palm frond..... 138
A.3	Plot of flavonoid reaction kinetics..... 139
A.4	A calibration plot of Fe ²⁺ absorbance change against Fe ²⁺ concentration..... 139
A.5	Linear relationship between absorbance reading at 750 nm and gallic acid concentration (mg / mL)..... 141

A.6	Calibration curve of five flavonoid standards.....	145
A.7 (a)	RP-HPLC chromatogram of extracts of green tea detected at 240, 280, 330 and 370 nm.....	146
A.7 (b)	RP-HPLC chromatogram of extracts of oil palm frond detected at 240, 280, 330 and 370 nm.....	147
A.8	Aortic rings set-up.....	148
A.9	Mesenteric arterial bed set-up.....	152
A.10	Aortic rings chart.....	156
A.11	Mesenteric artery chart.....	156
A.12	Picture of twelve tropical plants used in this study.....	158

LIST OF ABBREVIATIONS

ACE	Angiotensin converting enzyme
Ach	Acetylcholine
AIDS	Acquired Immune Deficiency Syndrome
ALT	Alanine aminotransferase
ATP	Adenosine triphosphate
BHA	Butylated hydroxyanisole
BHT	Butylated hydroxytoluene
BMI	Body Mass Index
C	Cholesterol control group
CAD	Coronary artery disease
CaM	Ca ²⁺ /calmodulin
cAMP	Cyclic adenosine monophosphate
cGMP	Cyclic guanosine monophosphate
CI	Confidence interval
CoQ10	Coenzyme Q10
CSIRO	Commonwealth Scientific and Industrial Research Organisation
Cu-Zn SOD	Copper-zinc superoxide dismutase
CVD	Cardiovascular diseases
DAD	Diode array detection
DNA	Deoxyribonucleic acid

DTNB	5,5'-Dithio-bis (2-Nitrobenzoic) Acid
EC-SOD	Extracellular superoxide dismutase
EDCF	Endothelium derived contraction factor
EDHF	Endothelium derived hyperpolarizing factor
EDRF	Endothelium-derived relaxing factor
EDTA	Ethylene diamine tetraacetic acid
eNOS	Endothelial nitric oxide synthase
FFA	Free fatty acid
FRAP	Ferric Reducing/Antioxidant Potential
GAE	Gallic acid equivalent
GGT	Gamma glutamyl transpeptidase
GSH	Glutathione
GSH-Px	Glutathione peroxidase
HCl	Hydrochloric acid
HDL	High-density lipoprotein
H ₂ O	Water
H ₂ O ₂	Hydrogen peroxide
His	Histamine
HMG-CoA	3-hydroxy-3-methylglutaryl-CoA
HOCl	Hypochlorous acid
HOCl ⁻	Hypochlorite
HPLC	High performance liquid chromatography

HPLC-MS	High performance liquid chromatography with mass spectrophotometer
INDO	Indomethacin
KCl	Potassium chloride
LC-MC	Liquid chromatography with mass spectrophotometer
LDL	Low-density lipoprotein
LDL-C	Low-density lipoprotein cholesterol
LO [•]	Lipid alkoxyl
LOO [•]	Lipid peroxy
M	Molar
MAB	Mesenteric arterial bed
MDA	Malonaldehyde
Mn-SOD	Manganese superoxide dismutase
N	Normal control group
NA	Noradrenaline
Na ₂ HPO ₄	Disodium phosphate
NaCl	Sodium chloride/saline
NADPH	Nicotinamide adenine dinucleotide phosphate, reduced form
NO	Nitric oxide
NO [•]	Nitric oxide radical
NOLA	N ω -Nitro-L-Arginine
NOS	Nitric oxide synthase