



**UNIVERSITI PUTRA MALAYSIA**

**ISOLATION, SCREENING FOR BIOACTIVITIES AND  
IDENTIFICATION OF SELECTED ENDOPHYTE FUNGI BY  
SEQUENCING OF 18S rRNA/ITS GENES**

**CHEAH YOKE KQUEEN**

**FSMB 2001 10**

**ISOLATION, SCREENING FOR BIOACTIVITIES AND IDENTIFICATION  
OF SELECTED ENDOPHYTE FUNGI BY SEQUENCING  
OF 18S rRNA/ITS GENES**

**CHEAH YOKE KQUEEN**

**MASTER OF SCIENCE  
UNIVERSITI PUTRA MALAYSIA**

**2001**



**ISOLATION, SCREENING FOR BIOACTIVITIES AND IDENTIFICATION  
OF SELECTED ENDOPHYTE FUNGI BY SEQUENCING  
OF 18S rRNA/ITS GENES**

**By**

**CHEAH YOKE KQUEEN**

**Thesis Submitted in Fulfilment of the Requirement for the  
Degree of Master of Science in the Faculty  
of Food Science and Biotechnology  
Universiti Putra Malaysia**

**August 2001**



## DEDICATION

To my mother, Sheon Yoke Chun and  
my father, Cheah Seak Yuen for their help  
and prayers.....

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement for the degree of Master of Science

**ISOLATION, SCREENING FOR BIOACTIVITIES AND IDENTIFICATION  
OF SELECTED ENDOPHYTE FUNGI BY SEQUENCING  
OF 18S rRNA/ITS GENES**

By

**CHEAH YOKE KQUEEN**

**August 2001**

**Chairman : Associate Professor Son Radu, Ph. D.**

**Faculty : Food Science and Biotechnology**

Endophytic fungi occur within plant tissues such as leaves and stems of healthy plants without producing any apparent infections and symptoms. Experiments were conducted to isolate endophytic fungi from healthy medicinal plants as well as detecting any reoccurrence of any particular endophytic fungi predominantly in selected local medicinal plants. Endophytic fungi have also been recognised as repository of novel secondary metabolites, which have beneficial biological activities, biocontrol of insects and oligosaccharides degrading enzymes. Thus, isolated endophytic fungi were screened for their bioactive properties by using Thin Layer Chromatography and Agar Diffusion Assay. Specially engineered yeast strains i.e. UCS and UCK from Kyowa Hakko Company, Japan were used to screened for anti-tumour activity. 18S ribosomal Ribonucleic Acid (18S



rRNA) / Internal Transcribed Spacer (ITS) gene sequencing were conducted to identified certain isolates.

Isolation of the endophytic fungi from healthy local medicinal plants showed that 61 out of 72 (84.7%) yielded endophytic fungi. Most of the endophytes were obtained from the leaves and very few from the stems. The reoccurrence rate of the endophytic fungi was 1.39% (1 in 72). Nevertheless, no predominant endophytic fungi association with types of medicinal plant was observed.

All the isolated endophytic fungi were able to degrade starch, xylan, mannan and inulin. 98.33% of the endophytic tested were able to degrade sago starch and rice starch. About 96.67% of the isolates were detected producing potato starch and starch wheat unmodified degrading enzyme. However, 95% of the endophytes produced tapioca starch and corn starch degrading enzyme.

Among the isolated endophytic fungi, 16.9% of them were considered important with regard to the bioactive screening results. Thus, 22 isolates from 130 isolated fungi gave satisfactory results in bioactive screening. There were 16 isolates that gave positive results in bio-activity test against UCS/UCK yeast strain. This showed that 16 out of 130 isolates produced potential bioactive compound for anti-tumour.

Four out of the twenty two important isolates were identified through microscopic examination and 18S/ITS gene sequencing. Isolate 12L was

identified as *Penicillium* spp. through microscopic morphology observation. Those isolates identified by 18S/ITS gene sequencing included 1B, *Endothia gyrosa*; 19L, *Colletotrichum gloeosporioides* and 22L, *Botryosphaeria ribis* . BLAST programme was used as a tool to determine the homology of the sequence obtained with the database sequence.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi syarat keperluan memperoleh ijazah Master Sains

**PEMENCILAN, PENSKRINAN BIOAKTIVITI DAN IDENTIFIKASI  
"ENDOPHYTE FUNGI" YANG TERPILIH DENGAN  
PENJUJUKKAN GEN '18S/ITS'NYA**

Oleh

**CHEAH YOKE KQUEEN**

**Ogos 2001**

**Pengerusi : Profesor Madya Son Radu, Ph. D.**

**Fakulti : Sains Makanan and Bioteknologi**

"Endophytic fungi" terdapat di antara tisu tumbuhan umpamanya tisu, daun dan batang tumbuhan yang sihat tanpa menyebabkan sebarang infeksi dan simptom-simptom yang ketara. Eksperimen dijalankan untuk mengisolatkan "endopytic fungi" daripada tumbuhan berubat yang sihat dan mengenalpasti sebarang kehadiran semula "endophytic fungi" secara pradominan dalam tumbuhan berubat tempatan yang terpilih. "Endophytic fungi" juga di kenali sebagai tempat penyimpanan metabolit sekunder yang bermanfaat dari segi aktiviti biologi, kawalan biologi terhadap serangga dan enzim pencernaan oligosakarid. Dengan demikian, "endophytic fungi" yang dipencilkan itu diskrinkan terhadap unsur bioaktif dengan "Thin Layer Chromatography" dan "Agar Diffusion Assay". Yeast yang khas dijurutera i.e. UCS dan UCK dari syarikat Kyowa Hakko digunakan untuk penskrinan unsur bioaktif antitumor.



Penjukkan gen "18S ribosomal ribonucleic acid" (18S rRNA) / "Internal Transcribed Spacer" (ITS) dijalankan untuk mengenalpasti pencilan-pencilan tertentu.

Pemencilan "endophytic fungi" daripada tumbuhan berubat yang sihat menunjukkan 61 daripada 72 (84.7%) terdapatnya "endophytic fungi". Kebanyakan "endophytes" yang diperolehi adalah daripada daun dan sangat jarang daripada batang ataupun tangkai. Kadar kehadiran semula "endophytic fungi" adalah 1.39% (1 in 72). Walaubagaimanapun, tiada "endophytic fungi" yang pradominan berkait dengan sebarang jenis tumbuhan yang dikaji kelihatan.

Semua pencilan "endophytic fungi" berupaya menurunkan kanji, xylan, mannan dan inulin. 98.33% daripada "endophytic fungi" yang dikaji berupaya menurunkan kanji sago dan kanji beras. Lebih kurang 96.67% daripada pencilan dikesan menghasilkan enzim penurunan kanji kentang dan "starch wheat unmodified". Namun demikian, 95% daripada "endophytes" menghasilkan enzim penurunan kanji ubi dan kanji jagung.

Di kalangan pencilan-pencilan "endophytic fungi", 16.9% daripadanya di katakan mustahak berhubungan dengan penskrinan bioaktif. Oleh itu, 22 pencilan daripada 130 "endophytic fungi" memberi keputusan yang memuaskan dalam penskrinan bioaktif. Terdapat 16 pencilan mempamerkan keputusan positif dalam bio-aktiviti terhadap UCS/UCK. Ini menunjukkan 16

daripada 130 pencilan berpotensi menghasilkan kompaun bioaktif terhadap anti-tumor.

Empat daripada dua puluh dua pencilan dikenalpasti melalui pemeriksaan mikroskopik dan penjujukan gen 18S/ITS. Pencilan 12L dikenalpasti sebagai *Penicillium* spp. melalui penelitian morfologi secara mikroskopik. Pencilan-pencilan yang dikenalpastikan dengan penjujukan 18S/ITS termasuk 1B, *Endothia gyrosa*; 19L, *Colletotrichum gloeosporioides* dan 22L, *Botryosphaeria ribis*. Program BLAST digunakan sebagai alat untuk memastikan homologi jujukan yang diperolehi dengan database jujukan.

## ACKNOWLEDGEMENTS

My most sincere grateful thanks is due to Associate Professor Dr. Son Radu, the Chairman of the Supervisory Committee, for his patience, encouragement, persuasion and guidance in preparation of this thesis and his helpful counsel during the course of study.

A word of thanks is to Professor Dr. Mohamed Ismail Abdul Karim and Dr. Abdul Reezal Abdul Latif, members of the Supervisory Committee, for their invaluable guidance and advice throughout in the thesis preparation. My sincere thanks also to Associate Professor Dr. Mohd Said Saad, Head of the Plant Genetic Resource Centre University Putra Malaysia for providing me the plant samples. I also wish to acknowledge Japan Society for the Promotion of Science (JSPS) for its continuous support of media and apparatus.

I am indebted to Ms. Ooi Wai Ling for the technical assistances while conducting my laboratory experiments. Thanks are also due to fellow graduate students, especially Puan Hajjah Endang Purwati, Mr. Samuel Lihan, Mr. Ahmad Zainuri, Mr. Yuherman, Puan Noor Zaleha binti Awang Salleh, Ms. Woo Kwan Kit, Ms. Lee Yock Ann, Ms. Haryanti Toosa, Ms. Noorlis Ahmad, Ms. Tengku Ahbrizal Farizal binti Tengku Ahmad, Ms. Apinya Vanichpun and many others, for their help and encouragement during my study.



This manuscript and the work it represents could not have been achieved without the sacrifices, patience, understanding and moral support of my father, mother and sister whose encouragement and moral support are boundless.



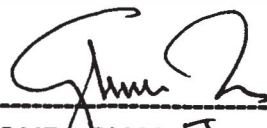
I certify that an Examination Committee met on 10<sup>th</sup> August 2001 to conduct the final examination of Cheah Yoke Kqueen on his Master of Science thesis entitled "Isolation, Screening for Bioactivities and Identification of Selected Endophyte Fungi by Sequencing of 18s rRNA/ITS Genes" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

**ABDUL MANAF ALI, Ph.D.**  
Professor  
Faculty of Food Science and Biotechnology,  
Universiti Putra Malaysia.  
(Chairman)

**SON RADU, Ph.D.**  
Associate Professor  
Faculty of Food Science and Biotechnology,  
Universiti Putra Malaysia.  
(Member)

**MOHAMED ISMAIL ABDUL KARIM, Ph.D.**  
Professor  
Faculty of Food Science and Biotechnology,  
Universiti Putra Malaysia.  
(Member)

**ABDUL REEZAL ABDUL LATIF, Ph.D.**  
Faculty of Food Science and Biotechnology,  
Universiti Putra Malaysia.  
(Member)



---

**MOHD. GHAZALI MOHAYIDIN, Ph.D.,**  
Professor  
Deputy Dean of Graduate School,  
Universiti Putra Malaysia

Date: 21 AUG 2001

This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirement for the degree of Master of Science.



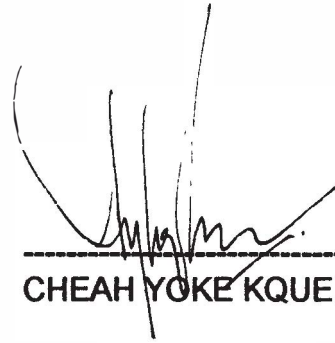
---

AINI IDERIS, Ph.D.  
Professor  
Dean of Graduate School,  
Universiti Putra Malaysia

Date: 08 NOV 2009

## DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations, which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.



CHEAH YOKE KQUEEN

Date: 22 AUGUST 2001

## TABLE OF CONTENTS

|                |   | Page  |
|----------------|---|-------|
|                | DEDICATION.....   | ii    |
|                | ABSTRACT.....   | iii   |
|                | ABSTRAK.....  | vi    |
|                | ACKNOWLEDGEMENTS.....                                       | ix    |
|                | APPROVAL SHEETS.....  | xi    |
|                | DECLARATION .....   | xiii  |
|                | LIST OF TABLES.....   | xvi   |
|                | LIST OF FIGURES.....  | xvii  |
|                | LIST OF PLATES.....   | xviii |
|                | LIST OF ABBREVIATIONS.....                                  | xx    |
| <br>           |   |       |
| <b>CHAPTER</b> |   |       |
| <br>           |   |       |
| I              | INTRODUCTION.....   | 1     |
| <br>           |   |       |
| II             | LITERATURE REVIEW.....                                      | 8     |
|                | Endophytes.....   | 8     |
|                | Isolation.....  | 10    |
|                | Ecological Diversity.....                                   | 10    |
|                | Bioactive Properties.....                                   | 11    |
|                | Oligosaccharides.....                                       | 12    |
|                | Secondary Metabolites and Antibiotics.....                  | 13    |
|                | Anti-microbial.....   | 14    |
|                | Anti-fungal.....  | 15    |
|                | Anti-tumour .....   | 15    |
|                | Thin Layer Chromatography.....                              | 16    |
|                | Polymerase Chain Reaction and 18S rRNA/ ITS Sequencing..... | 16    |
|                | Basic Local Alignment Search Tool (BLAST).....              | 18    |
| <br>           |   |       |
| III            | METHODOLOGY.....  | 20    |
|                | Isolation of Endophytic Fungi.....                          | 20    |
|                | Ecological Diversity of Endophytic Fungi.....               | 20    |
|                | Detection of Starch Degrading Endophytes.....               | 21    |
|                | Cultivation and Enzyme Reaction.....                        | 21    |
|                | Thin Layer Chromatography Methods.....                      | 22    |
|                | Anti-microbial Activity .....                               | 22    |
|                | Anti-tumour Activity.....                                   | 23    |
|                | Identification of Endophytic Fungi.....                     | 24    |
| <br>           |   |       |
| IV             | RESULTS.....  | 28    |
|                | Isolation of Endophytic Fungi.....                          | 28    |
|                | Ecological Diversity of Endophytic Fungi.....               | 29    |





|    |   |    |
|----|---|----|
|    | Detection of Starch Degrading Endophytes.....                   | 32 |
|    | Thin Layer Chromatography in Detection of Oligosaccharides..... | 33 |
|    | Antimicrobial Activity and Anti-tumour Activity.....            | 41 |
|    | Identification of Endophytic Fungi.....                         | 44 |
| V  | DISCUSSION.....   | 50 |
|    | Recommendation.....   | 58 |
| VI | CONCLUSION.....   | 59 |
|    | REFERENCES.....   | 61 |
|    | APPENDICES.....   | 75 |
| A  | Simplified Flowchart of Isolation Method.....                   | 76 |
| B  | List of Medicinal Plant with Its Medicinal Value.....           | 78 |
|    | CURRICULUM VITAE.....   | 82 |



## LIST OF TABLES

| Table |  | Page |
|-------|--|------|
| 1     | Primers for Amplification of Fungal Ribosomal RNA Genes.....                         | 27   |
| 2     | Isolated endophytic fungi from different medicinal herbs...                          | 30   |
| 3     | Oligosaccharide production according to isolates.....                                | 38   |
| 4     | Endophytic fungi isolates showing bio-activity against test organisms.....           | 42   |
| 5     | Sequence obtained with automated sequencer.....                                      | 48   |
| 6     | List of medicinal plants with its medicinal values that were used in this study..... | 79   |

## LIST OF FIGURES

| Figure |   | Page |
|--------|---|------|
| 1      | Simplified diagram of isolation method..... | 77   |



## LIST OF PLATES

| Plate |  | Page |
|-------|--|------|
| 1     | Top view of some isolated endophytic fungi.....  | 28   |
| 2     | Bottom view of some isolated endophytic fungi.....   | 29   |
| 3     | Starch degrading endophyte with clear zone.....  | 32   |
| 4     | Thin Layer Chromatography plate with xylanoligosaccharide production by endophytic fungi isolated from local medicinal plants.....                                       | 33   |
| 5     | Thin Layer Chromatography plate with manooligosaccharide production by endophytic fungi isolated from local medicinal plants.....  | 34   |
| 6     | Thin Layer Chromatography plate with inulooligosaccharide production by endophytic fungi isolated from local medicinal plants.....                                       | 34   |
| 7     | Thin Layer Chromatography plate with oligosaccharide production from degradation of sago starch by isolated endophytic fungi from local medicinal plants.....            | 35   |
| 8     | Thin Layer Chromatography plate with oligosaccharide production from degradation of rice starch by isolated endophytic fungi from local medicinal plants.....            | 35   |
| 9     | Thin Layer Chromatography plate with oligosaccharide production from degradation of potato starch by isolated endophytic fungi from local medicinal plants.....          | 36   |
| 10    | Thin Layer Chromatography plate with oligosaccharide production from degradation of tapioca starch by isolated endophytic fungi from local medicinal plants.....         | 36   |
| 11    | Thin Layer Chromatography plate with oligosaccharide production from degradation of corn starch by isolated endophytic fungi from local medicinal plants.....            | 37   |
| 12    | Thin Layer Chromatography plate with oligosaccharide production from degradation of starch wheat unmodified by isolated endophytic fungi from local medicinal plants.... | 37   |
| 13    | Results of crude extracts from some of the endophyte isolates on <i>Bacillus subtilis</i> with C as positive control.....  | 42   |



|    |  |    |
|----|--|----|
| 14 | Result of crude extract from endophyte isolates showing a clear zone of growth inhibition against <i>Saccharomyces cerevisiae</i> .....  | 43 |
| 15 | Result of crude extract from an endophyte isolate (E) showing a clear zone of growth inhibition against <i>Alternaria</i> spp.....   | 43 |
| 16 | Growth of the UCS/UCK yeast strain as positive result in anti-tumour activity screening.....   | 44 |
| 17 | Microscopic structure of endophytic fungi strain 12L with 600 times magnification.....   | 44 |
| 18 | Microscopic structure of endophytic fungi strain 12L with 1500 times magnification.....  | 45 |
| 19 | PCR amplification products with NS1 - NS6 primers (lanes with no. 2, lanes with no. 5, lanes with no. 8 and lanes with no. 11) and NS5 - NS8 primers (lanes with no. 3, lanes with no. 6, lanes with no. 9 and lanes with no. 12). Lane 0 is marker. Lanes with no. 2 and 3 are 1B isolate. Lanes with no. 5 and 6 are 12L isolate. Lanes with no. 8 and 9 are 19L isolate and Lanes with no. 11 and 12 are 22L isolate..... | 46 |
| 20 | PCR amplification with ITS primers (lane 1, lane 4, lane 7 and lane 10), NS1 - NS6 primers (lane 2, lane 5, lane 8 and lane 11) and NS5 - NS8 primers (lane 3, lane 6, lane 9 and lane 12). Lane 0 is marker. Lanes 1 - 3 are 1B isolate. Lanes 4 - 6 are 12L isolate. Lanes 7 - 9 are 19L isolate and Lanes 10 - 12 are 22L isolate.....  | 46 |
| 21 | PCR amplification products with NS1 - NS6 primers (lane with no. 2, lane with no. 5, lane with no. 8 and lane with no. 11) and NS5 - NS8 primers (lane with no. 3, lane with no. 6, lane with no. 9 and lane with no. 12). Lane 0 is marker. Lanes with no. 2 and 3 are 1B isolate. Lanes with no. 5 and 6 are 12L isolate. Lanes with no. 8 and 9 are 19L isolate and Lanes with no. 11 and 12 are 22L isolate..            | 47 |



## LIST OF ABBREVIATIONS

|                                 |                                     |
|---------------------------------|-------------------------------------|
| %                               | Percent                             |
| μl                              | Microlitre (s)                      |
| °C                              | Degree Celsius                      |
| BLAST                           | Basic Local Alignment Search Tool   |
| bp                              | Base pair                           |
| cm                              | Centimetre (s)                      |
| CMA                             | Corneal Agar                        |
| CO <sub>2</sub>                 | Carbon dioxide                      |
| DNA                             | Deoxyribonucleic Acid               |
| dNTP                            | Deoxynucleotide Triphosphate        |
| e.g.                            | <i>Exempli gratia</i> (for example) |
| FeSO <sub>4</sub>               | Ferum Sulphate                      |
| g                               | Gram (s)                            |
| H <sub>2</sub> O                | Water                               |
| i.e.                            | <i>Id est</i> (that is)             |
| IGS                             | Intergenic Spacer                   |
| ITS                             | Internal Transcribed Spacer         |
| KCl                             | Potassium Chloride                  |
| KH <sub>2</sub> PO <sub>4</sub> | Dihydrogen Potassium Phosphate      |
| LSU                             | Large Subunit                       |
| MEA                             | Malt Extract Agar                   |
| mg                              | Milligram (s)                       |
| mg/l                            | Milligram per Litre                 |

|                                     |                                  |
|-------------------------------------|----------------------------------|
| <b>MgCl<sub>2</sub></b>             | <b>Magnesium Chloride</b>        |
| <b>MgSO<sub>4</sub></b>             | <b>Magnesium Sulphate</b>        |
| <b>min.</b>                         | <b>Minute (s)</b>                |
| <b>ml</b>                           | <b>Millilitre (s)</b>            |
| <b>mM</b>                           | <b>Millimolar</b>                |
| <b>MSP</b>                          | <b>Maximal Segment Pair</b>      |
| <b>nd</b>                           | <b>None detected</b>             |
| <b>ng</b>                           | <b>Nanogram (s)</b>              |
| <b>NH<sub>4</sub>NO<sub>3</sub></b> | <b>Ammonium Nitrate</b>          |
| <b>O<sub>2</sub></b>                | <b>Oxygen</b>                    |
| <b>PDA</b>                          | <b>Potato Dextrose Agar</b>      |
| <b>RNA</b>                          | <b>Ribonucleic Acid</b>          |
| <b>rpm</b>                          | <b>Rotation per minute</b>       |
| <b>SSU</b>                          | <b>Small Subunit</b>             |
| <b><i>Taq</i></b>                   | <b><i>Thermus aquaticus</i></b>  |
| <b>TLC</b>                          | <b>Thin Layer Chromatography</b> |
| <b>U</b>                            | <b>Unit</b>                      |
| <b>UV</b>                           | <b>Ultra violet</b>              |
| <b>v</b>                            | <b>Volume</b>                    |

## CHAPTER I

### INTRODUCTION

To microbes, land plants present a complex, spatially and temporally diverse ecological habitat. Symbiotic associations between microorganisms and plants are ancient and fundamental, and many examples of complex and highly specific symbioses between plants and microbes have been well known. Endophytic microbes are an intriguing group of organisms associated with various tissues and organs of terrestrial and some aquatic plants, and are the subject of increasing interest to mycologists, ecologists, and plant pathologist.

Biologically and ecologically, endophytes represent a diversity of nutritional modes from biotrophic parasites to interim or facultative saprotrophs, and associations with their hosts span the continuum from biotrophic mutualist and benign commensals to nectotrophic, antagonistic pathogens. Ecologically, interactions between host plants, pathogens, and herbivores are mediated by endophytes. The species composition and distribution of endophytic associations within and among hosts, and the reciprocal ecological effects of endophyte colonisation on host fitness and the composition of plant communities, are problems of common interest to endophytologists and plant ecologists. Research interests in endophytes are correspondingly diverse and include community analysis, anatomical-histological relationships, organismal biology, biodiversity studies, population



ecology and evolutionary biology, and ecological interactions among endophytes, hosts, and herbivores. Endophytism may be an important factor in microbial speciation and biodiversity. Accumulating evidence suggests that endophytes represent a large reservoir of genetic diversity and a rich source of heretofore undescribed species. Practical applications of endophytes include potential biological control agents, sources of novel metabolites for medicine, plant protection, and industrial use, and as research model systems for investigation of host-parasite interactions and evolution in natural systems.

Often the terms "endophyte" and "endophytic" are used with particular meaning by different workers and for particular groups of hosts and microbes. Among the definitions proposed for the term endophyte are "fungi colonising living plant tissue without causing any immediate, overt negative effects" (Hirsch and Braun, 1992). This definition includes virtually the entire spectrum of symbiotic interactions in which fungi and plants participate: parasitism, commensalism, and mutualism (Bills, 1996). This definition, however, fails to include prokaryotic microbes, such as bacteria and blue-green algae, or endophytic vascular plants (Fisher *et al.*, 1992; Mauseth *et al.*, 1985). A more inclusive definition of "endophyte" should stress the symptomless nature of the infection on the host without limiting the term to any particular group of organisms. Latent and quiescent pathogens are endophytes as are mutualistic microbes and benign commensals. Petrini (1991) considers the term endophyte to be purely topographical: "Endophytes colonise symptomlessly the living, internal tissues of their host,