



UNIVERSITI PUTRA MALAYSIA

**KINETICS, MODELLING AND SCALING-UP OF KOJIC ACID
FERMENTATION BY ASPERGILLUS FLAVUS 44-1
USING DIFFERENT CARBON SOURCES**

ROSFARIZAN MOHAMAD

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By

ROSFARIZAN MOHAMAD

**Thesis Submitted in Fulfilment of the Requirement for the Degree of Doctor of
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December 2000



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ROSFARIZAN MOHAMAD

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Chairman : Assoc. Prof. Dr. Arbakariya Ariff

Faculty : Food Science and Biotechnology

Kojic acid production by *Aspergillus flavus* strain 44-1 using different types of carbon source (glucose, starch hydrolysate from enzymatic hydrolysis of sago starch, sucrose, fructose and gelatinized sago starch) was carried out in 250 mL shake flask, 2 L, 8 L and 50 L stirred tank fermenters. The experimental data from batch fermentation and resuspended cell system were analysed in order to form the basis for a kinetic model of the process. Unstructured model based on logistic and Luedeking-Piret equations was found suitable to describe growth, substrate consumption and kojic acid production by *Aspergillus flavus* in batch and also resuspended cell system using either glucose or sucrose. From the modelling, it was found that kojic acid production by *A. flavus* was non-growth associated process. The kinetic parameter values for each fermenter were calculated from the modelling



and they can be used to verify the experimental data using various types and concentration of carbon source.

Kojic acid production (23.5 g/L) using 100 g/L sago starch in a shake flask was comparable to fermentation of glucose (32.5 g/L) and starch hydrolysate (27.9 g/L) but in the 8 L and 50 L fermenter kojic acid production was greatly reduced due to non-optimal aeration conditions. Fed-batch fermentation with intermittent feeding of concentrated sago starch (140 g/L) can be employed to improve direct fermentation of sago starch to kojic acid by about 4 times higher as compared to batch fermentation. *A. flavus* was also capable to utilise sucrose for kojic acid fermentation where the highest production (40.23 g/L) in 2 L fermenter was obtained at 150 g/L sucrose. Kojic acid production (10.25 g/L) was greatly reduced in fermentation using fructose as the sole carbon source. Scaling-up based on a constant impeller tip speed (1.65 m/s) together with optimal DOT and pH control strategies was successfully used for kojic acid fermentation in 50 L fermenter using glucose and sucrose as carbon sources.

Kojic acid fermentation by *A. flavus* can be divided into two phases; growth and production phase. The culture pH during growth phase influenced the performance of kojic acid fermentation to a further extent than did the pH during the production phase. The fermentation without pH controlled (started with an initial culture pH 3) showed higher kojic acid production than single-phase pH controlled fermentation at a range of pH 2.2 – 4.0. Comparable kojic acid production to fermentation without pH controlled was obtained in two-phase pH

controlled fermentation (started with initial culture pH, without control during growth phase and switched to 3 during production phase).

Efficient conversion of glucose to kojic acid was achieved in a resuspended cell system, in a solution containing only glucose with citrate buffer at pH 3.5 and 30°C. The rate of glucose conversion to kojic acid was increased with increasing glucose concentration up to 100 g/L, suggesting that the biotransformation of glucose to kojic acid by the cell-bound enzymes followed the Michaelis-Menten enzyme kinetic models. The value of K_m and V_{max} for the reaction, as determined by using Langmuir plot, was 10.042 g/L glucose and 0.076 g/L.h, respectively.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk bergraduasi Ijazah Doktor Falsafah

**KINETIK, PERMODELAN DAN PENINGKATAN SKALA FERMENTASI
ASID KOJIK OLEH *ASPERGILLUS FLAVUS* 44-1 MENGGUNAKAN
SUMBER-SUMBER KARBON YANG BERBEZA**

oleh

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Pengerusi : Assoc. Prof. Dr. Arbakariya Ariff

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Penghasilan asid kojik oleh *Aspergillus flavus* strain 44-1 menggunakan pelbagai sumber karbon (glukosa, hidrolisat kanji daripada hidrolisis berenzim kanji sagu, sukrosa, fruktosa dan kanji sagu) telah dijalankan menggunakan kelalang bergoncang 250 mL dan pelbagai saiz fermenter berpengaduk (2 L, 8 L dan 50 L). Data-data eksperimen daripada proses fermentasi sesekelompok dan sistem sel terampai telah dianalisa untuk membentuk asas bagi model proses kinetik. Model tidak berstruktur berdasarkan persamaan-persamaan logistik dan Luedeking-Piret didapati sesuai untuk menerangkan pertumbuhan *Aspergillus flavus*, penggunaan substrat dan penghasilan asid kojik dalam sistem sesekelompok dan sel terampai menggunakan glukosa ataupun sukrosa. Daripada permodelan, penghasilan asid

kojik oleh *A. flavus* telah ditunjukkan sebagai proses pertumbuhan tidak berkait. Nilai-nilai parameter kinetik bagi setiap proses fermentasi telah dikira daripada permodelan dan ianya boleh digunakan untuk menguji benar tidaknya data eksperimen menggunakan pelbagai jenis dan kepekatan sumber-sumber karbon.

Penghasilan asid kojik (23.5 g/L) menggunakan 100 g/L kanji sagu di dalam kelalang bergoncang adalah setara dengan fermentasi menggunakan glukosa (32.5 g/L) dan hidrolisat kanji (27.9 g/L) tetapi di dalam fermenter 8 dan 50 L, penghasilan telah berkurang kerana keadaan pengudaraan yang tidak optima. Fermentasi suapan sesekelompok dengan penambahan bersela kanji sagu pekat (140 g/L) boleh digunakan untuk meningkatkan prestasi fermentasi asid kojik menggunakan kanji sagu sebanyak empat kali ganda lebih tinggi berbanding dengan fermentasi sesekelompok. *A. flavus* juga berupaya menggunakan sukrosa bagi fermentasi asid kojik di mana penghasilan tertinggi (40.23 g/L) di dalam fermenter 2 L diperolehi pada kepekatan sukrosa 150 g/L. Penghasilan asid kojik (10.25 g/L) menurun dengan banyaknya bagi fermentasi menggunakan fruktosa sebagai sumber karbon. Peningkatan skala berdasarkan halaju hujung pengaduk yang tetap (1.65 m/s) bersama-sama strategi kawalan kepekatan oksigen terlarut dan pH yang optima telah berjaya digunakan untuk fermentasi asid kojik dalam 50 L fermenter menggunakan glukosa dan sukrosa sebagai sumber karbon.

Fermentasi asid kojik boleh dibahagikan kepada dua fasa, fasa pertumbuhan dan fasa penghasilan. pH kultur semasa fasa pertumbuhan lebih banyak mempengaruhi proses fermentasi asid kojik berbanding pH semasa fasa

penghasilan. Fermentasi tanpa kawalan pH (dimulakan dengan pH 3) menunjukkan penghasilan asid kojik yang lebih tinggi berbanding fermentasi kawalan pH satu fasa pada julat pH 2.2 – 4.0. Penghasilan asid kojik yang setara dengan fermentasi tanpa kawalan pH telah diperolehi bagi fermentasi kawalan pH dua-fasa (dimulakan dengan pH 3, tanpa kawalan pH semasa fasa pertumbuhan dan ditukarkan kepada pH 3 semasa fasa penghasilan).

Penukaran glukosa kepada asid kojik yang berkesan telah diperolehi dalam sistem sel terampai, dalam larutan mengandungi hanya glukosa dengan penimbale sitrat pada pH 3.0 dan suhu 30°C. Kadar pertukaran glukosa kepada asid kojik adalah meningkat dengan peningkatan kepekatan glukosa sehingga 100 g/L, menunjukkan yang proses biotransformasi ini mengikuti model enzim kinetik Michaelis-Menten. Untuk tindakbalas ini, nilai K_m adalah 10.042 g/L glukosa dan nilai V_{max} adalah 0.076 g/L.j, seperti yang ditentukan menggunakan plot Langmuir.

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I certify that an Examination Committee met on 11 December 2000 to conduct the final examination of Rosfarizan Mohamad on her Doctor of Philosophy thesis entitled “Kinetics, Modelling and Scaling-up of Kojic Acid Fermentation by *Aspergillus flavus* 44-1 using Different Carbon Sources” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommends that the candidate be awarded the relevant degree. Members of the examination Committee are as follows:

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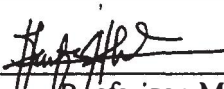
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DECLARATION FORM

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or currently submitted for any other degree at UPM or other institutions.


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LIST OF ABBREVIATIONS

α	Growth-associated rate constant for glucose consumption (g glucose/g cell)
β	Non-growth-associated rate constant for glucose consumption (g glucose/g cell.h)
C/N	Carbon to nitrogen ratio of medium in mM basis
D	Dilution rate
D_i	Impeller diameter
DOT	Dissolved oxygen tension
μ_{max}	Maximum or initial specific growth rate (h^{-1})
m	Growth associated rate constant for kojic acid production (g kojic acid/g cell)
n	Non-growth associated rate constant for kojic acid production (g kojic acid/g cell.h)
P_o	Initial kojic acid concentration (g/L)
P	Kojic acid concentration (g/L)
P_{max}	Maximum kojic acid concentration (g/L)
S_o	Initial substrate concentration (g/L)
S	Substrate concentration (g/L)
t	Time (h)
X	Cell concentration (g/L)
X_o	Initial cell concentration (g/L)
X_{max}	Maximum cell concentration (g/L)
$Y_{p/s}$	Yield of kojic acid based on glucose consumed (g/g)

