brought to you by CORE



UNIVERSITI PUTRA MALAYSIA

EFFECTS OF LACTOBACILLUS STRAINS AS A PROBIOTIC AND A HYPOLIPIDAEMIC AGENT FOR CHICKENS

KALAVATHY RAMASAMY

IB 2003 2



EFFECTS OF *LACTOBACILLUS* STRAINS AS A PROBIOTIC AND A HYPOLIPIDAEMIC AGENT FOR CHICKENS

 $\mathbf{B}\mathbf{y}$

KALAVATHY RAMASAMY

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirement for the Degree of Doctor of Philosophy

UPM

Abstract of the thesis submitted to the senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

EFFECTS OF LACTOBACILLUS STRAINS AS A PROBIOTIC AND A HYPOLIPIDAEMIC AGENT FOR CHICKENS

By

KALAVATHY RAMASAMY

August 2003

Chairman: Professor Dr. Ho Yin Wan

Institute : Bioscience

In recent years, there has been considerable interest in the beneficial effects of probiotics (direct-fed microbials, which include Lactobacillus) to modulate the lipid metabolism. However, the mechanism(s) involved remains unclear. A series of experiments was carried out to investigate the ability of 12 Lactobacillus strains to deconjugate bile salts and to remove cholesterol in vitro, and to assess their potential as a probiotic and as a hypolipidaemic agent for broilers and laying hens. Bile salt hydrolase (BSH) activity (resulting in bile salt deconjugation) of intestinal bacteria is closely linked to the lowering of cholesterol. The results of the *in vitro* studies showed that all the 12 Lactobacillus strains could deconjugate sodium glychocholate (GCA) and sodium taurocholate (TCA) bile salts, and all the strains, except L. fermentum I 24, had a higher affinity for GCA. However, only eight strains could deconjugate sodium taurodeoxycholate (TDCA). This indicates that the BSH of the Lactobacillus strains is substrate specific. The 12 Lactobacillus strains showed significant differences in their ability to reduce cholesterol from the growth medium (27 to 85 %) with or without bile salt, indicating that bile salt is not a prerequisite for the removal of cholesterol. Lactobacillus acidophilus I 16, L. crispatus I 12, L.



brevis C 17 and I 211, and L. fermentum I 24 and I 25 removed cholesterol from the growth medium mainly through assimilation of cholesterol into the cells. On the other hand, L. brevis C 1, C 10, I 23 and I 218, and L. fermentum C 16 removed cholesterol through both assimilation and co-precipitation of deconjugated bile salt with cholesterol at low pH. The Lactobacillus strains assimilated more esterified than non-esterified cholesterol and the assimilated cholesterol was tightly bound to the cells. Cells grown in the presence of cholesterol were more resistant to lysis by sonication than when grown in its absence, suggesting a possible alteration of the cell wall or membrane by the assimilated cholesterol. Cholesterol removal by the Lactobacillus strains was also affected by Tween 80.

The feeding trials showed that the supplementation of a mixture of the 12 Lactobacillus cultures (LC), as a probiotic for broilers, significantly improved growth equivalent to that provided by the antibiotic, oxytetracycline, but the feed conversion ratio was better in LC-fed broilers. The supplementation of LC also significantly lowered the total cholesterol, low density lipoprotein cholesterol and triglycerides of the serum; the cholesterol of the carcass and liver; abdominal fat deposition; and fat contents of the liver, muscle and carcass of broilers; but there was little effect on the fatty acid compositions of the liver, muscle and carcass.

In laying hens, the supplementation of LC improved the feed efficiency and hen-day egg production during the early stage of the laying cycle, and increased egg weight and influenced a shift from small and medium to large and extra large eggs throughout the laying cycle. However, LC had very little effect on improving the fatty acid composition, and the cholesterol and total fat contents of eggs.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

KESAN PELBAGAI STRAIN *LACTOBACILLUS* SEBAGAI PROBIOTIK DAN AGEN HIPOLIPIDIMIK UNTUK AYAM

Oleh

KALAVATHY RAMASAMY

Ogos 2003

Pengerusi: Profesor Dr. Ho Yin Wan

Institut : Biosains

Sejak kebelakangan ini, kecenderungan untuk menggunakan probiotik (mikrob makanan, termasuk Lactobacillus) dalam mengawal atur metabolisma lipid semakin berkembang. Namun demikian, mekanisma yang terlibat masih tidak jelas. Satu siri eksperimen telah dijalankan untuk mengkaji keupayaan 12 strain Lactobacillus untuk melakukan dikonjugasi garam hempedu (garam konjugat) dan mengurangkan kolesterol secara in vitro, serta kesannya sebagai probiotik dan agen hypolipidimik terhadap ayam pedaging dan ayam penelur. Aktiviti enzim "bile salt hydrolase (BSH)" (yang menyebabkan dikonjugasi garam hempedu) usus berkait rapat dengan pengurangan kolesterol. Hasil kajian in vitro menunjukkan bahawa kesemua 12 strain *Lactobacillus* berupaya melakukan dikonjugasi garam "glychocholate" (GCA) dan garam "taurocholate" (TCA), dan kesemua strain, kecuali L. fermentum I 24, menunjukkan afiniti yang lebih tinggi terhadap GCA. Tetapi hanya lapan strain berupaya melakukan dikonjugasi "taurodeoxycholate". Ini menunjukkan bahawa aktiviti BSH Lactobacillus adalah spesifik substrat. Duabelas strain Lactobacillus ini juga menunjukkan keupayaan untuk mengurangkan kolesterol dari media kultur (25 hingga 85 %) yang ada atau tiada garam hempedu. Pengurangan kolesterol dari media kultur oleh L. acidophilus



I 16, *L. crispatus* I 12, *L. brevis* C 17 dan I 211, dan *L. fermentum* I 24 dan I 25 adalah terutamanya melalui asimilasi kolesterol oleh sel. Pengurangan kolesterol oleh *L. brevis* C 1, C 10, I 23 dan I 218, dan *L. fermentum* C 16 pula, adalah melalui asimilasi dan juga ko-mendakan garam hempedu tak berkonjugat bersama kolesterol pada pH yang rendah. Strain *Lactobacillus* mengasimilasi lebih banyak kolesterol ester berbanding dengan kolesterol bebas dan kolesterol yang diasimilasi didapati terikat dengan kuat pada sel. Sel yang ditumbuhkan bersama kolesterol juga lebih resistan kepada sonikasi, mencadangkan bahawa pengubahsuaian pada dinding atau membran sel berlaku setelah mengasimilasi kolesterol. Pengurangan kolesterol oleh strain *Lactobacillus* juga bergantung pada Tween 80.

Hasil kajian *in vivo* menunjukkan bahawa campuran 12 strain *Lactobacillus* (LC), sebagai probiotik pada ayam pedaging dapat meningkatkan berat badan sama seperti antibiotik "oxytetracycline", tetapi kadar penukaran makanan ayam adalah lebih baik pada ayam yang di beri LC. Penambahan LC pada ayam juga dapat menurunkan paras "total" kolesterol, "low density lipoprotein" kolesterol dan trigliserida di serum; kandungan kolesterol pada karkas dan hati; lemak berlebihan pada bahagian abdomen; dan kandungan lemak pada hati, otot dan karkas; tetapi tidak berupaya mengubah profil asid lemak pada hati, otot dan karkas.

Ayam penelur yang di beri LC dapat meningkatkan kadar penukaran makanan dan produksi telur pada peringkat awal peneluran serta dapat menghasilkan telur yang lebih berat dan saiz yang lebih besar sepanjang proses peneluran. Namun demikian, LC kurang berkesan untuk mengubah profil asid lemak, atau menurunkan paras kolesterol dan lemak di telur.



ACKNOWLEDGEMENTS

I wish to express my deep appreciation and most sincere gratitude to the chairman of the supervisory committee, Professor Dr. Ho Yin Wan, for her invaluable guidance and advice, endless support, patience, and encouragement throughout the duration of this study and for her critical analysis, constructive criticism and helpful suggestions during the preparation of my thesis.

I am deeply grateful and indebted to Associate Professor Dr. Norhani Abdullah and Dr. Clemente Michael Wong, who are members of the supervisory committee, for their kind assistance, advice and guidance throughout the course of my work and in the preparation of the thesis.

Special appreciation goes to Tan Sri Dato Dr. Syed Jalaludin Syed Salim (who was a member of the supervisory committee till his retirement in 2001) for his wise counsel, support and constant encouragement.

My heartfelt appreciations are extended to Madam Haw Ah Kam, Mr. Khairul Kamar Bakri, Mr. Nagayah Muniandy, Mr. Jivanathan Arumugam and Mr. Paimon Lugiman, staff of the Enzyme and Microbial Technology Laboratory, and Mr. Saparin Denim and Mr. Ibrahim Mohsin, staff of Animal Nutrition Laboratory, for their technical support and kind assistance. Thanks are also due to Dr. Goh Yong Meng for his assistance on the preparation of samples for the fatty acid determination using Gas Chromatography.



I wish to extend my sincere thanks to my post graduate friends Chin Chin, Latiffah, Lan, Darlis, Vicky, Wan, Thongsuk, Lee, Pit Kang and Sidieg for their friendship, support, encouragement and their sense of humor that made the many hours in the laboratory very pleasant, which contributed to the successful completion of this work.

Finally, very special thanks are due to my family for their unconditional love, untiring patience, support and encouragement, which inspired and motivated me throughout the course of this study.



I certify that an Examination Committee met on 6th August 2003 to conduct the final examination of Kalavathy Ramasamy on her Doctor of Philosophy thesis entitled "Effects of *Lactobacillus* Strains as a Probiotic and a Hypolipidaemic Agent for Chickens" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulation 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

ABDUL RAZAK ALIMON, Ph.D.

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

HO YIN WAN, Ph.D.

Professor Faculty of Science and Environmental Studies Universiti Putra Malaysia (Member)

NORHANI ABDULLAH, Ph.D.

Associate Professor Faculty of Science and Environmental Studies Universiti Putra Malaysia (Member)

CLEMENTE MICHAEL WONG, Ph.D.

Faculty of Food Science and Biotechnology Universiti Putra Malaysia (Member)

HYUNG TAI SHIN, Ph.D.

Professor
Department of Food and Bioresources
Faculty of Life Science and Technology
Sung Kyun Kwan University
300 Chunchun-Dong, Jangan-Ku
Suwon 440-746, Republic of Korea
(Independent Examiner)

GULAM RUSUL RAHMAT ALI, Ph.D.

Professor/Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 4 SEP 2003



This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee are as follows:

HO YIN WAN, Ph.D.

Professor Faculty of Science and Environmental Studies Universiti Putra Malaysia (Chairman)

NORHANI ABDULLAH, Ph.D.

Associate Professor Faculty of Science and Environmental Studies Universiti Putra Malaysia (Member)

CLEMENTE MICHAEL WONG, Ph.D.

Faculty of Food Science and Biotechnology Universiti Putra Malaysia (Member)

AINI IDERIS, Ph.D.

Professor/Dean School of Graduate Studies Universiti Putra Malaysia

Date: 1 6 SEP 2003



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

KALAVATHY RAMASAMY

Date: 22 /9/03



TABLE OF CONTENTS

		Page
ABSTRAK ACKNOWLEI APPROVAL DECLARATIO LIST OF TABI	DGEMENTS DN LES JRES REVIATIONS	ii iv vi viii x xiv xvii xx
CHAPTER		
1	INTRODUCTION	1
2	2.1 Poultry Industry 2.2 Global Challenges in the Modern Poultry Industry 2.3 Lipids and Human Health 2.4 Lipids in Broiler Meat and Eggs 2.4.1 Lipid 2.4.2 Fat deposition, Cholesterol and Fatty Acids 2.4.3 Strategies in Improving the Lipid Content in Broiler Meat 2.4.4 Strategies in Improving the Lipid Content in Eggs 2.4.5 Biological Methods in Improving Lipids in Broiler Meat and Eggs	5 6 8 10 10 12 14 16
	 2.5 Antibiotics in Poultry Production: Benefits and Risks 2.6 Probiotics 2.6.1 Contributions of the Intestinal Microflora 2.6.2 Definition 2.6.3 Probiotics Currently in Use 2.6.4 Mode of Action of Probiotics 2.6.5 Selection Criteria for Probiotics 2.6.6 Benefits of Probiotics on Poultry Performance 2.7 Hypocholesterolaemic Effect of Lactic Acid Bacteria 2.8 Bile Salt Deconjugation of Lactic Acid Bacteria 2.8.1 Enterohepatic Circulation of Bile Acids 2.8.2 Significance of Bile Salt Deconjugation by the Lactic Acid Bacteria 	18 22 22 23 24 25 28 28 33 36 36
3	BILE SALT HYDROLASE ACTIVITY OF LACTOBACILLUS CULTURES FROM CHICKEN 3.1 Introduction	40 40



		 3.2.1 Source and Maintenance of <i>Lactobacillus</i> Strains 3.2.2 Bile Salt Deconjugation by <i>Lactobacillus</i> Strains 3.2.3 Kinetics of Bile Salt Deconjugation
		3.2.4 Bile Tolerance Test
	3.3	Results
	3.3	3.3.1 Morphological Characteristics
		3.3.2 Bile Salt Deconjugation by <i>Lactobacillus</i> Strains 3.3.3 Kinetic Parameters of Bile Salt Deconjugation
		3.3.4 Bile Tolerance
	3.4	
4	СН	OLESTEROL-REDUCING ABILITY OF <i>LACTOBACILLUS</i>
•		RAINS IN VITRO AND THE MECHANISM (S)
		OLVED
		Introduction
	4.2	Materials and Methods
	4.2	
		4.2.1 Preliminary Study of Cholesterol Reduction by 12
		Lactobacillus Strains
		4.2.2 Effects of Various Bile Salt Concentrations on the
		Reduction of Cholesterol
		4.2.3 Effects of Various Concentrations of Tween 80 on
		the Reduction of Cholesterol
		4.2.4 Effect of Cholesterol on Growth of Lactobacillus
		Strains
		4.2.5 Quantitative Analysis of Cholesterol in the Culture
		Supernatant and Bacterial Cell Pellet of Three
		Lactobacillus Strains
		4.2.6 Qualitative Analysis on the Assimilation of
		Cholesterol by Lactobacillus Strains
		4.2.7 Effects of Cholesterol and Bile Salts on Lysis of
		Lactobacillus by Sonication
		4.2.8 Effects of pH and Bile Salts on Solubility of Cholesterol
	4.3	Results
		4.3.1 Reduction of Cholesterol in Growth Media by 12 Lactobacillus Strains
		4.3.2 Effects of Bile Salt on Cholesterol Reduction
		4.3.3 Effects of Concentrations of Tween 80 on Cholesterol
		Reduction
		4.3.4 Effect of Cholesterol on Growth of Lactobacillus
		Strains
		4.3.5 Quantitative Analysis of Cholesterol in the Culture
		Supernatant and Cell Pellet of Lactobacillus Strains
		4.3.6 Qualitative Analysis of Cholesterol in Cell Pellets of
		Lactobacillus Strains
		4.3.7 Effect of Cholesterol and Bile Salts on Lysis of
		Lactobacillus by Sonication
		4.3.8 Influence of pH and Bile Salts on Solubility of
	, .	Cholesterol
	44	Discussion



5	CHICKENS	117
	CHICKENS	117
	5.2 Materials and Methods	117
		119
	5.2.1 Animals and Rearing Management	
	5.2.3 Experiment II	122
	5.2.4 Statistical Analysis	129
	5.3 Results	129
	5.3.1 Experiment I	129
	5.3.2 Experiment II	132
	5.4 Discussion	142
6	EFFECTS OF LACTOBACILLUS CULTURES ON LAYING	
	HENS	157
	6.1 Introduction	157
	6.2 Materials and Methods	158
	6.2.1 Animals and Rearing Management	158
	6.2.2 Dietary Treatment	159
	6.2.3 Layer Performance and Production	159
	6.2.4 Egg Quality and Egg Storage Test	161
	6.2.5 Yolk Total Lipids, Fatty Acid Composition and	101
	Cholesterol	163
	6.2.6 Statistical Analysis	164
	6.3 Results	164
	6.3.1 Ambient Temperature and Relative Humidity	164
	6.3.2 Layer Performance and Production	165
	· · · · · · · · · · · · · · · · · · ·	175
	6.3.3 Egg Quality and Storage Test	1/3
	6.3.4 Egg Yolk Cholesterol, Total Lipids and Fatty Acid	100
	Composition	180
	6.4 Discussion	185
7	GENERAL DISCUSSION AND CONCLUSIONS	194
	7.1 General Discussion	194
	7.2 Conclusions	206
	BIBLIOGRAPHY	208
	VITA	234



LIST OF TABLES

Table		Page
1	Possible modes of actions of probiotics	27
2	Criteria for an effective probiotic strain	29
3	Hypocholesterolaemic effects of lactic acid bacteria on various hosts	34
4	Lactobacillus strains (from chicken) used in the study	42
5	Bile salt hydrolase (BSH) activity of <i>Lactobacillus</i> strains on MRS + sodium taurodeoxycholate (MRS + TDCA) agar plates	51
6	Comparison of the deconjugation of sodium taurocholate and sodium glychocholate by 12 <i>Lactobacillus</i> strains	59
7	Kinetics of bile salt deconjugation by <i>L. brevis</i> C 10 from 2 to 24 h of incubation	63
8	Kinetics of bile salt deconjugation by <i>L. fermentum</i> C16 from 2 to 24 h of incubation	65
9	Kinetics of bile salt deconjugation by <i>L. acidophilus</i> I 26 from 2 to 24 h of incubation	67
10	Kinetics of bile salt deconjugation by <i>L. acidophilus</i> I 16 from 2 to 24 h of incubation	69
11	Growth of <i>Lactobacillus</i> strains in MRS broth and MRS with 0.3 % bile salt at 4 h	71
12	Reduction of cholesterol in growth media by 12 Lactobacillus strains	89
13	Effects of bile salt concentrations on cholesterol reduction by Lactobacillus strains	91
14	Effects of Tween 80 concentrations on cholesterol reduction by Lactobacillus strains	92
15	Comparison of growth of three <i>Lactobacillus</i> strains in various growth media	94
16	Percentages of cholesterol reduced in the MRSC and MRSBC supernatants and percentages of cholesterol assimilated in the cell pellets of three <i>Lactobacillus</i> strains	97



1 /	in various media and stained with filipin	98
18	Fluorescence intensity of cell pellets of <i>Lactobacillus</i> strains grown in various media and stained with Nile Red	101
19	Effects of cholesterol and bile salts on lysis of <i>Lactobacillus</i> by sonication	105
20	Composition of the basal diets	121
21	Effects of <i>Lactobacillus</i> cultures (LC) or oxytetracycline on body weight, weight gain, feed intake and feed to gain ratio of broiler chickens for 42 days	131
22	Effects of <i>Lactobacillus</i> cultures (LC) on body weight, weight gain and feed to gain ratio of broiler chickens for 42 days	133
23	Percentage by weight of organs from broiler chickens fed diets with or without <i>Lactobacillus</i> cultures (LC) from 21 to 42 days of age	134
24	Abdominal fat deposition of broiler chickens fed with or without Lactobacillus cultures (LC) from 21 to 42 days of age	135
25	Serum lipid concentrations in broiler chickens fed with or without Lactobacillus cultures (LC) from 21 to 42 days of age	140
26	Effects of <i>Lactobacillus</i> cultures (LC) on cholesterol contents of carcass, liver and muscle of broiler chickens at 42 days of age	141
27	Effects of <i>Lactobacillus</i> cultures (LC) on fat contents of carcass, liver and muscle of broiler chickens at 42 days of age	141
28	Fatty acid composition of carcass from broilers supplemented with or without <i>Lactobacillus</i> cultures (LC) at 42 days of age	143
29	Fatty acid composition of liver from broilers supplemented with or without <i>Lactobacillus</i> cultures (LC) at 42 days of age	144
30	Fatty acid composition of muscle from broilers supplemented with or without <i>Lactobacillus</i> cultures (LC) at 42 days of age	145
31	Composition of the basal diet	160
32	Egg size distribution	161
33	Effects of <i>Lactobacillus</i> cultures (LC) on feed intake, feed efficiency, hen-day egg production and mortality of laying hens from 20 to 68 weeks of age	166



34	Effects of <i>Lactobacillus</i> cultures (LC) on egg weight and egg mass of laying hens from 20 to 68 weeks of age	170
35	Effects of <i>Lactobacillus</i> cultures (LC) on egg size of laying hens from 20 to 68 weeks of age	173
36	Effects of <i>Lactobacillus</i> cultures (LC) on egg quality of hens from 20 to 35 weeks of age	176
37	Cholesterol contents of eggs from hens supplemented with or without <i>Lactobacillus</i> cultures (LC) at 24, 28, 32 and 68 weeks of age	181
38	Total lipid contents of eggs from hens supplemented with or without <i>Lactobacillus</i> cultures (LC) at 24, 28 and 32 weeks of age	181
39	Fatty acid composition of eggs from hens supplemented with or without <i>Lactobacillus</i> cultures (LC) at 24 weeks of age	182
40	Fatty acid composition of eggs from hens supplemented with or without <i>Lactobacillus</i> cultures (LC) at 28 weeks of age	183
41	Fatty acid composition of eggs from hens supplemented with or without <i>Lactobacillus</i> cultures (LC) at 32 weeks of age	184



LIST OF FIGURES

Figure		Page
1	Cell morphology of <i>Lactobacillus</i> strains observed using light microscopy	48
2	Colonies of Lactobacillus strains on MRS agar	49
3	Plate assay showing high bile salt hydrolase (BSH) activity of L. fermentum C 16	52
4	Plate assay showing high bile salt hydrolase (BSH) activity of L. brevis C 1	53
5	Plate assay showing high bile salt hydrolase (BSH) activity of <i>L. brevis</i> C 10	54
6	Plate assay showing no bile salt hydrolase (BSH) activity of <i>L. brevis</i> C 17	55
7	Plate assay showing low bile salt hydrolase (BSH) activity of L. crispatus I 12 and L. brevis I 23	56
8	Plate assay showing absence of bile salt hydrolase (BSH) activity in L. fermentum I 24 and L. acidophilus I 26	57
9	Precipitates due to bile salt hydrolase (BSH) activity as observed under the light microscope	58
10	Deconjugation of sodium glychocholate (GCA) and sodium taurocholate (TCA) by <i>Lactobacillus</i> strains	60
11	Growth and changes in pH, and dissappearance of conjugated bile salt in MRS broth supplemented with sodium taurocholate (TCA) and sodium glycocholate (GCA) of <i>L. brevis</i> C 10 from 0 to 24 h of incubation	64
12	Growth and changes in pH, and dissappearance of conjugated bile salt in MRS broth supplemented with sodium taurocholate (TCA) and sodium glycocholate (GCA) of <i>L. fermentum</i> C 16 from 0 to 24 h of incubation	66
13	Growth and changes in pH, and dissappearance of conjugated bile salt in MRS broth supplemented with sodium taurocholate (TCA) and sodium glycocholate (GCA) of <i>L. acidophilus</i> I 26 from 0 to 24 h of incubation	68



14	salt in MRS broth supplemented with sodium taurocholate (TCA) and sodium glycocholate (GCA) of <i>L. acidophilus</i> I 16 from 0 to 24 h of incubation	70
15	Growth of <i>L. brevis</i> C 10, <i>L. acidophilus</i> I 26 and <i>L. acidophilus</i> I 16 in four different media with or without cholesterol	95
16	Fluorescence micrographs of cell pellets of <i>L. acidophilus</i> I 26 stained with filipin	99
17	Fluorescence micrographs of cell pellets of <i>L. acidophilus</i> I 26 stained with Nile Red	102
18	Fluorescence micrographs of cell pellets of <i>L. brevis</i> C 10 stained with Nile Red	103
19	Influence of pH and bile salts on solubility of cholesterol	106
20	Abdominal fat depositions of broiler chickens at 42 days of age fed without or with <i>Lactobacillus</i> cultures	136
21	Fat depositions at different areas in broiler chickens at 42 days of age fed without or with <i>Lactobacillus</i> cultures	137
22	Fat deposited on the skin of broiler chickens at 42 days of age fed without or with <i>Lactobacillus</i> cultures	138
23	Effect of <i>Lactobacillus</i> cultures (LC) on feed efficiency of laying hens from 20 to 68 weeks of age	167
24	Effect of <i>Lactobacillus</i> cultures (LC) on egg production of laying hens from 20 to 68 weeks of age	168
25	Effect of <i>Lactobacillus</i> cultures (LC) on egg weight of laying hens from 20 to 68 weeks of age	171
26	Effect of Lactobacillus cultures (LC) on egg mass of laying hens from 20 to 68 weeks of age	172
27	Effect of <i>Lactobacillus</i> cultures (LC) on egg size of laying hens from 20 to 44 weeks of age	174
28	Effect of <i>Lactobacillus</i> cultures (LC) on egg size of laying hens from 45 to 68 weeks of age	174
29	Effect of <i>Lactobacillus</i> cultures (LC) and storage time on internal	177



30	Internal egg quality of a fresh egg and an egg that was stored for 7 days from a top view	178
31	Internal egg quality of a fresh egg (A) and an egg that was stored for 7 days (B) from a lateral view	179



LIST OF ABBREVIATIONS

AAP Aminoantipyrine
ADP Adenosine diphosphate
AFTA Asean Free Trade Centre

AOAC Association of Official Analytical Chemists

ATP Adenosine triphosphate
BSH Bile salt hydrolase
CFU Colony forming unit

cm centimetre
CP Cell pellet

d Day

FAME Fatty acid methyl ester

FAO Food and Agriculture Organisation FDA Food and Drug Administrations

g gram

GC Gas Chromatography
GCA Sodium glychocholate

GRAS Generally Recognized as Safe

h hour

H₂O₂ Hydrogen peroxide

HACCP Hazzard Analysis Critical Control Points

HBA Hydroxybenzoic acid HDL High density lipoprotein

HMG CoA Hydroxymethylglutaryl coenzyme A HPLC High Performance Liquid Chromatography

HU Haugh unit

IDL Intermediate density lipoprotein

IU International Unit

kg kilogram

KIC α-ketoisocaproic acid KOH Potassium hydroxide

l litre

LABIP International Platform for Lactic Acid Bacteria

LC A mixture of 12 *Lactobacillus* cultures

LDL Low density lipoprotein

M Molar
m metre
mg milligram
min minute
MJ megajoules

mRNA Messenger Ribonucleic Acid

MRS Man Rogoso Sharpe
MRSB MRS containing bile salt
MRSC MRS containing cholesterol

MRSBC MRS containing bile salt and cholesterol

MRS-TDCA MRS agar supplemented with 0.5 % sodium taurodeoxycholate

MUFA Monounsaturated fatty acids

NaCl Sodium chloride NaOH Sodium hydroxide



ND No data

OD Optical density
OTC Oxytetracycline
PTA Phototungstic Acid

PPLO Pleuropneumonia-like organism
PUFA Polyunsaturated fatty acids
SAS Stastical Analysis Software
SCFA Short chain fatty acids
SFA Saturated fatty acids

ST Supernatant

TCA Sodium taurocholate
TDCA Sodium taurodeoxycholate
tRNA Transfer Ribonucleic Acid
UFA Unsaturated fatty acids

μg microgram μl microlitre

VLDL Very low density lipoprotein

W Watt

WHO World Health Organisation



CHAPTER 1

INTRODUCTION

The worldwide poultry industry provides a substantial proportion of the nutritional requirement of the human population. Poultry meat is perceived to be lean and low in cholesterol, so it may come as a surprise to learn that poultry scientists and producers are increasingly concerned about the amount of fat present in chicken meat. Chambers et al. (1981), Lin (1981) and Havenstein et al. (1994) reported that, as a result of selection strategy for body weight gain or growth rate, modern fast-growing broilers have been found to contain about four times higher amounts of abdominal fat than those in the 1960s. Eggs have also been viewed with suspicion today because of their high cholesterol content (Stadelman, 1999). In the US, egg consumption has declined from 256 eggs per capita per year in 1985 to 235 in 1995 (USDA, 1997). The lipid composition of animal products is a primary consumer concern as high fat and cholesterol intakes have been implicated to contribute to coronary heart disease, the most common chronic illness in developed countries. To the poultry producers, on the other hand, excess fat is an economic burden, as fat is lost during processing of the carcass or of the meat, resulting in lower meat yields and, furthermore, the discarded abdominal fat and visceral fat increases waste management problems. This has put the poultry production system under pressure and, therefore, much attention is now directed towards producing healthier meat and eggs such that the lipid fraction is improved (reduced cholesterol and fat and improvement of the fatty acid make-up). Animal feed strategies, genetic selections, and gene manipulation are some of the techniques that have been developed to alter the lipid composition in broilers (Jiménez-Colmenero, 2000) and



egg yolk (Hargis, 1988). However, very often these techniques are cost prohibitive or may impair performances and, therefore, not economically feasible to be applied at commercial scale. Animal welfare and environmental issues may also be linked in the application of these techniques.

Performance and economic returns are one of the main concerns of the commercial poultry industry. To achieve these goals, very often, intensive farming systems are adopted, subjecting broilers and laying hens to various stressful situations. Stress may lower the body's defense mechanism and create an imbalance in the intestinal microflora (Fuller, 1999), which in turn increases susceptibility to infectious diseases, resulting in poor performance. Efforts to prevent or reduce avian diseases include improved management practices, but inevitably at a cost, because this requires high quality feed manufacturing and feeding systems where the environment and the feed are relatively pathogen-free (Zhang-Barber et al., 1999). The benefits of incorporating antibiotic growth promoters in animal feeds are well substantiated (Bedford, 2000). These products have been used for many years by the poultry industry and have proved to be an effective way of enhancing animal status, uniformity and production efficiency. The Union of Concerned Scientists recently estimated that, each year, 11.2 million kg of antimicrobials are given to animals for non-therapeutic purposes, and 900,000 kg are given for theraphy, thus, it is fair to state that substantial amounts of antimicrobials are administered to food animals for growth promotion and feed efficiency in the absence of known disease (Gorbach, 2001). However, the use of antibiotics as growth promoters is severely restricted or totally banned in poultry production in many countries, largely because of concern on the development of resistant bacterial strains and residual toxicity in

