



UNIVERSITI PUTRA MALAYSIA

**EXPRESSION OF HAEMAGGLUTININ-NEURAMINIDASE ENVELOPE
PROTEIN OF NEWCASTLE DISEASE VIRUS STRAIN AF2240 IN
CENTELLA ASIATICA (PEGAGA) EMBRYOGENIC CALLI THROUGH
OPTIMIZED PARTICLE BOMBARDMENT METHOD**

LAI KOK SONG

FBSB 2008 18

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**MASTER OF SCIENCE
UNIVERSITI PUTRA MALAYSIA**

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By

LAI KOK SONG

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirement for the Degree of Master of Science**

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Abstract of thesis presented to the Senate of University Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EXPRESSION OF HAEMAGGLUTININ-NEURAMINIDASE ENVELOPE PROTEIN OF NEWCASTLE DISEASE VIRUS STRAIN AF2240 IN *CENTELLA ASIATICA* (PEGAGA) EMBRYOGENIC CALLI THROUGH OPTIMIZED PARTICLE BOMBARDMENT METHOD

By

LAI KOK SONG

June 2008

Chairman: Professor Maziah Mahmood, PhD

Faculty: Biotechnology and Biomolecular Sciences

Centella asiatica is a locally important medicinal plant. It is non-toxic, high in medicinal values, and can serve as a good candidate for genetic manipulation. However, to date no transformation protocol has been developed to fully utilize the potential of this plant. Therefore, this research is to establish an efficient particle bombardment transformation protocol for *C. asiatica* embryogenic calli. In addition, an attempt to express the haemagglutinin-neuraminidase (HN) protein from Newcastle disease virus (NDV) strain AF2240 in *C. asiatica* embryogenic calli were carried out using the developed transformation system. The HN protein can serve as a potential vaccine candidate for Newcastle disease (ND) in poultry. The induced embryogenic calli revealed the presence of extracellular matrix layer (ECM) during the microscopy studies. Particle bombardment transformation protocol was developed using the green fluorescent protein (GFP) as reporter. A total of eight parameters mainly different target distance, helium pressure, gold particles size, chamber vacuum



pressure, number of bombardment, precipitation agents, post-bombardment incubation time, and plasmid DNA concentration were identified and successfully optimized. Based on the established protocol, transformations of *C. asiatica* embryogenic calli were performed using the constructed recombinant pMDC32'HN and HBT95:sGFP(S65T)-NOS'HN plasmids. Genomic PCR analysis revealed the presence of HN transgene in the transformed lines. Unfortunately no protein bands were detected during SDS-PAGE and western blotting, indicating low or no HN protein expression. Transformation using recombinant HBT95:sGFP(S65T)-NOS'HN plasmid resulted in very low GFP expression as compared to the positive control. Nonetheless, the mRNA transcripts were detected in the RT-PCR analysis. Positive signal from the dot blot assay further confirmed the presence of the HN protein expression in the transformed lines.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**EXPRESI PROTEIN HAEMAGGLUTININ-NEURAMINIDASE DARI
VIRUS PENYAKIT NEWCASTLE STRAIN AF2240 UNTUK
KALUS EMBRYOGENIK *CENTELLA ASIATICA* (PEGAGA) MELALUI
KAEDAH MIKROPROJEKTIL BEDILAN YANG DIOPTIMASIKAN**

Oleh

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Centella asiatica merupakan tumbuhan ubatan tempatan yang penting. Ia tidak bertoksik, kaya dengan nilai ubatan, dan boleh bertindak sebagai calon manipulasi genetik yang baik. Malangnya, hingga kini tiada protokol transformasi yang dihasilkan dalam memaksimumkan potensi tumbuhan ini. Justeru itu, penyelidikan ini bermatlamat untuk menghasilkan protokol mikroprojektil bedilan transformasi yang cekap untuk kallus embryogenik *C. asiatica*. Tambahan pula, cubaan untuk mengekspreskan protein haemagglutinin-neuraminidase (HN) strain virus penyakit Newcastle (NDV) AF2240 dalam kallus embryogenik *C. asiatica* juga dijalankan berdasarkan sistem protokol yang dihasilkan. Protein HN boleh digunakan sebagai calon vaksin yang berpotensi untuk penyakit Newcastle (ND) dalam perternakan. Kallus embryogenik yang diinduksikan menunjukkan kehadiran lapisan matrik luaran (ECM) semasa kajian mikroskop. Protokol mikroprojektil bedilan transformasi telah dihasilkan dengan menggunakan



green fluorescent protein (GFP) sebagai sistem pelapor. Sejumlah lapan parameter yang terdiri dari jarak tisu sasaran, tekanan helium, saiz partikel emas, tekanan ruang vakum, bilangan bedilan, bahan pengikatan, masa pos-bedilan, dan kepekatan DNA plasmid telah dikenalpasti dan berjaya dioptimasikan. Berdasarkan protokol yang dihasilkan, transformasi dijalankan pada kalus embryogenik *C. asiatica* dengan menggunakan plasmid pMDC32'HN dan HBT95:sGFP(S65T)-HN. Analisis PCR genomik menunjukkan kehadiran transgen HN dalam transforman. Malangnya, tiada produk protein yang dikesan semasa SDS-PAGE dan blot western menunjukkan rendah atau ketiadaan ekspresi protein HN. Transformasi menggunakan plasmid HBT95:sGFP(S65T)-HN memberi ekspresi GFP yang rendah berbanding pada kawalan positif. Walaubagaimanapun, transkrip mRNA berjaya dikesan dalam analisis RT-PCR. Keputusan positif dari pemblotan titikan memastikan kehadiran ekspresi protein HN pada transforman.

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I certify that an Examination Committee has met on 25 September 2008 to conduct the final examination the of Lai Kok Song on his Master of Science thesis entitled “Expression of Haemagglutinin-neuraminidase Envelope Protein from Newcastle Disease Virus Strain AF2240 in *Centella asiatica* (Pegaga) Embryonic Calli Through Optimized Particle Bombardment Method” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the student be awarded the Master of Science.

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DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledge. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institutions.

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TABLE OF CONTENTS

| | Page |
|---|-------------|
| ABSTRACT | iii |
| ABSTRAK | v |
| ACKNOWLEDGEMENTS | vii |
| APPROVAL | viii |
| DECLARATION | ix |
| LIST OF TABLES | xiv |
| LIST OF FIGURES | xv |
| LIST OF ABBREVIATIONS | xvii |
| | |
| CHAPTER | |
| | |
| 1 INTRODUCTION | 1 |
| | |
| 2 LITERATURE REVIEW | 4 |
| 2.1 Newcastle Disease | 4 |
| 2.2 Newcastle Disease Virus | 5 |
| 2.2.1 Classification | 5 |
| 2.2.2 Virion Structure | 6 |
| 2.2.3 Viral Genome Organization | 9 |
| 2.2.4 Haemagglutinin-neuraminidase Protein | 11 |
| 2.3 Plant Tissue Culture | 14 |
| 2.3.1 <i>Centella asiatica</i> | 14 |
| 2.3.2 Embryogenic Tissue | 15 |
| 2.3.3 Organogenesis | 17 |
| 2.4 Plant Genetic Transformation | 19 |
| 2.4.1 DNA Delivery Methods | 19 |
| 2.4.2 Particle Bombardment | 21 |
| 2.5 Plant protein Expression System | 25 |
| 2.5.1 Strategies for Protein Expression | 25 |
| 2.5.2 Why Plant the Better Expression System? | 28 |
| | |
| 3 MATERIALS & METHODS | 32 |
| 3.1 Materials | 32 |
| 3.1.1 Chemicals, Biological Reagents, Kits, Enzymes, and Markers | 32 |
| 3.1.2 Virus | 32 |
| 3.1.3 Bacteria | 32 |
| 3.1.4 Plant Material | 33 |
| 3.1.5 Plasmid Constructs | 33 |
| 3.2 Methods | 34 |
| 3.2.1 General Procedures | 34 |
| 3.3 Plant Tissue Culture and Microscopy Studies | 35 |
| 3.3.1 Preparation of Tissue Culture Media | 35 |
| 3.3.2 Leaf Explant Sterilization | 35 |
| 3.3.3 Calli Initiation and Maintenance | 36 |
| 3.3.4 Growth Measurement of the Calli Culture | 36 |



| | | |
|--------|--|----|
| 3.3.5 | Organogenesis of <i>Centella asiatica</i> Calli Culture | 37 |
| 3.3.6 | Histology: Light Microscopy | 37 |
| 3.3.7 | Scanning Electron Microscopy (SEM) | 38 |
| 3.3.8 | Transmission Electron Microscopy (TEM) | 39 |
| 3.4 | Molecular Cloning Methods | 40 |
| 3.4.1 | Virus Cultivation and Purification | 40 |
| 3.4.2 | Haemagglutinin (HA) Test | 41 |
| 3.4.3 | Viral RNA Extraction | 42 |
| 3.4.4 | Primer Design | 43 |
| 3.4.5 | Reverse Transcription-Polymerase Chain Reaction (RT-PCR) | 43 |
| 3.4.6 | DNA Agarose Gel Purification | 45 |
| 3.4.7 | Isolation of DNA from Agarose Gel | 45 |
| 3.4.8 | Preparation of <i>Escherichia Coli</i> Competent Cells | 45 |
| 3.4.9 | Cloning of HN Gene into pCR [®] 8/GW/TOPO Vector | 46 |
| 3.4.10 | Subcloning of HN Gene into pMDC32'HN | 46 |
| 3.4.11 | Cloning of HN Gene into pGEM [®] -T Easy Vector | 48 |
| 3.4.12 | Subcloning of HN Gene into HBT95:sGFP(S65T)-NOS | 48 |
| 3.4.13 | Transformation of <i>Escherichia Coli</i> Competent Cells | 49 |
| 3.4.14 | Small Scale Plasmid Extraction | 49 |
| 3.4.15 | Restriction Enzyme Digestion | 51 |
| 3.4.16 | Polymerase Chain Reaction (PCR) | 51 |
| 3.4.17 | DNA Sequencing | 52 |
| 3.5 | Particle Bombardment Transformation Parameters Optimization | 52 |
| 3.5.1 | Consumables for Particle Bombardment (PDS/He 1000 Bio-Rad System) | 52 |
| 3.5.2 | Target Tissue Preparation for Transformation | 53 |
| 3.5.3 | Large Scale Plasmid Extraction | 53 |
| 3.5.4 | Preparation of Gold Microcarriers and Coating of Plasmid DNA onto Gold Microcarriers | 53 |
| 3.5.5 | Particle Bombardment | 54 |
| 3.5.6 | GFP Monitoring | 56 |
| 3.6 | Transformation of pMDC32'HN and HBT95:sGFP(S65T)-NOS'HN | 56 |
| 3.7 | Selection and Molecular Analyses of the Transformants | 57 |
| 3.7.1 | Minimal Hygromycin Concentration Determination | 57 |
| 3.7.2 | Selection of Putative Transformants | 57 |
| 3.7.3 | Plant Genomic DNA Extraction | 58 |
| 3.7.4 | Plant Genomic PCR Analysis | 58 |
| 3.7.5 | Total Plant RNA Extraction | 59 |
| 3.7.6 | Total Soluble Protein Extraction | 60 |
| 3.7.7 | Bradford Assay | 60 |
| 3.7.8 | SDS-Polyacrylamide Gel Electrophoresis (SDS-PAGE) | 61 |
| 3.7.9 | Western Blot Analysis | 62 |



| | | |
|----------|---|------------|
| 3.7.10 | Dot Blot Analysis | 63 |
| 4 | RESULTS AND DISCUSSION | 65 |
| 4.1 | Plant Tissue Culture | 65 |
| 4.1.1 | <i>Centella asiatica</i> Callus Induction | 65 |
| 4.1.2 | The Growth Curve of Callus | 66 |
| 4.1.3 | Organogenesis of <i>Centella asiatica</i> Callus | 68 |
| 4.2 | Microscopy Studies | 70 |
| 4.2.1 | Histology: Light Microscopy | 70 |
| 4.2.2 | Scanning Electron Microscopy (SEM) | 73 |
| 4.2.3 | Transmission Electron Microscopy (TEM) | 76 |
| 4.3 | Minimal Hygromycin Concentration Determinations | 81 |
| 4.4 | Particle Bombardment Parameters Optimisation | 83 |
| 4.4.1 | Effect of Different Target Distance | 83 |
| 4.4.2 | Effect of Different Helium Pressure (psi) | 84 |
| 4.4.3 | Effect of Gold Particle Size | 86 |
| 4.4.4 | Effect of Chamber Vacuum Pressure | 87 |
| 4.4.5 | Effect of Bombardment Number | 89 |
| 4.4.6 | Effect of Spermidine and Calcium Chloride Precipitation Agents | 90 |
| 4.4.7 | Effect of Post-Bombardment Incubation Time | 92 |
| 4.4.8 | Effect of Total Plasmid DNA Concentration | 93 |
| 4.5 | RT-PCR and Construction of pMDC32'HN | 96 |
| 4.6 | Selection of the Putative Transformants | 100 |
| 4.7 | Genomic PCR Analyses of the Transformants | 101 |
| 4.8 | Protein Analysis with SDS-PAGE and Western Blotting | 103 |
| 4.9 | PCR and Construction of HBT:sGFP(S65T)-NOS'HN | 107 |
| 4.10 | Transient GFP Expression Monitoring | 111 |
| 4.11 | RT-PCR and Dot Blot Analysis | 111 |
| 5 | CONCLUSIONS | 114 |
| | REFERENCES | 118 |
| | APPENDICES | 135 |
| | LIST OF PUBLICATIONS | 143 |
| | BIODATA OF THE AUTHOR | 144 |



LIST OF TABLES

| Table | | Page |
|--------------|---|-------------|
| 3.1 | Oligonucleotide primers used in amplification of the HN gene | 44 |
| 3.2 | The bombardment parameters were studied independently in the co-bombardment event | 56 |

LIST OF FIGURES

| Figure | Page | |
|--------|---|----|
| 2.1 | Virion structure of NDV | 8 |
| 2.2 | TEM of NDV particles purified from allantoic fluid | 8 |
| 2.3 | NDV genome organization and viral transcripts | 10 |
| 2.4 | Schematic diagram of important features and domains of HN protein from NDV | 13 |
| 3.1 | Flow chart of the construction of the recombinant pMDC32'HN plasmid | 47 |
| 4.1 | <i>C. asiatica</i> calli derived from leaf explants | 67 |
| 4.2 | Growth curve measurement of <i>C. asiatica</i> | 67 |
| 4.3 | Organogenesis of <i>C. asiatica</i> callus | 69 |
| 4.4 | Light micrographs of embryogenic and non-embryogenic callus | 71 |
| 4.5 | SEM of embryogenic and non-embryogenic calli | 74 |
| 4.6 | SEM of embryogenic calli | 75 |
| 4.7 | TEM of embryogenic and non-embryogenic cells | 77 |
| 4.8 | The ultrastructure of ECM layer of the embryogenic cells | 79 |
| 4.9 | The hygromycin killing level for <i>C. asiatica</i> embryogenic calli | 82 |
| 4.10 | Effect of different target distance on transient GFP expression in <i>C. asiatica</i> embryogenic calli | 85 |
| 4.11 | Effect of different helium pressure (psi) on transient GFP expression in <i>C. asiatica</i> embryogenic calli | 85 |
| 4.12 | Effect of different gold particle size (μm) on transient GFP expression <i>C. asiatica</i> embryogenic calli | 88 |
| 4.13 | Effect of different chamber vacuum pressure (mm Hg) on transient GFP expression in <i>C. asiatica</i> embryogenic calli | 88 |
| 4.14 | Effect of different number of bombardments on transient GFP expression in <i>C. asiatica</i> embryogenic calli | 91 |

| | | |
|------|---|-----|
| 4.15 | Effect of different treatments on transient GFP expression in <i>C. asiatica</i> embryogenic calli. | 91 |
| 4.16 | Effect of different post bombardment time on transient GFP expression in <i>C. asiatica</i> embryogenic calli | 94 |
| 4.17 | Effect of different plasmid DNA concentration (μg) on transient GFP expression in <i>C. asiatica</i> embryogenic calli | 94 |
| 4.18 | Transient GFP expression on <i>C. asiatica</i> embryogenic calli | 95 |
| 4.19 | Construction of recombinant pMDC32'HN plasmid | 98 |
| 4.20 | Morphology of the survive transformants after 2 months subculturing in 15 mg/L hygromycin medium | 102 |
| 4.21 | Genomic PCR analyses of the transformants | 102 |
| 4.22 | SDS-12% PAGE and coomassie blue staining of total soluble protein from transformed and non-transformed lines | 106 |
| 4.23 | Western blot analysis of total soluble protein from transformed and non-transformed lines | 106 |
| 4.24 | Construction of recombinant HBT95:sGFP(S65T)-NOS'HN plasmid | 109 |
| 4.25 | Transient GFP expression analysis | 113 |
| 4.26 | RT-PCR and dot blot analysis of transformants | 113 |

LIST OF ABBREVIATIONS

| | |
|-------------------|-------------------------------------|
| 2'4 D | 2,4-dichlorophenoxy acetic acid |
| 5'-UTL | 5' untranslated leader |
| Ads | adenine sulfate |
| APS | ammonium persulfate |
| BA | 6-benzyladenine |
| BDMA | benzyl dimethylamine |
| BSA | bovine serum albumin |
| °C | Celsius |
| CaCl ₂ | calcium chloride |
| cDNA | complementary deoxyribonucleic acid |
| cm ² | centimetre square |
| DDSA | dodecenylsuccinic anhydride |
| DEPC | diethyl pyrocarbonate |
| dH ₂ O | distilled water |
| ECM | extracellular matrix |
| EDTA | ethylenediaminetetraacetic acid |
| ELISA | enzyme-linked immunosorbent assay |
| ER | endoplasmic reticulum |
| F | fusion (glycoprotein) |
| FAA | formaldehyde-acetic acid-ethanol |
| GFP | green fluorescent protein |
| GE | gene end |
| g | gram |
| GS | gene start |

| | |
|-------------------|---|
| h | hour |
| HN | haemagglutinin-neuraminidase (glycoprotein) |
| IBA | indole-3-butyric acid |
| Ig G | immunoglobulin G |
| kb | kilobase |
| kDa | kilo dalton |
| Kn | kinetin |
| kPa | kilopascal |
| L | litre |
| LB | Luria-Bertani |
| M | molar |
| mA | milliampere |
| MgCl ₂ | magnesium chloride |
| mg | milligram |
| min | minute |
| mL | millilitre |
| mm | millimeter |
| mM | millimolar |
| mm Hg | millimetre of mercury |
| MNA | 2-methyl-4-nitro-aniline |
| MS | Murashige and Skoog |
| mRNA | messenger ribonucleic acid |
| NAA | naphthalene acetic acid |
| NaOH | sodium hydroxide |
| NDV | Newcastle disease virus |

| | |
|----------|---|
| ng | nanogram |
| Nm | nanometer |
| NP | nucleoprotein |
| P | phosphoprotein |
| PBS | phosphate buffered saline |
| PCR | polymerase chain reaction |
| pmol | picomol |
| s | second (time) |
| scFv | single-chain variable fragment |
| SCW | silicon carbide whisker |
| SDS-PAGE | sodium dodecyl sulphate-PAGE |
| SEM | scanning electron microscope |
| RBC | red blood cell |
| RE | restriction enzyme |
| RNA | ribonucleic acid |
| rpm | rotations per minute |
| RT | room temperature |
| RT-PCR | reverse transcriptase polymerase chain reaction |
| TAE | 40 mM Tris-Cl(pH 7.4), 20mM sodium acetate, 1 mM EDTA |
| T-DNA | transfer deoxyribonucleic acid |
| TDZ | thidiazuran |
| TE | 10 mM Tris-Cl (pH 8.0), 1 mM EDTA |
| TEM | transmission electron microscope |
| TEMED | N,N,N',N'-tetramethylethylenediamine |
| TPBS | Tris phosphate buffer saline |



| | |
|---------------|----------------|
| μg | microgram |
| μL | microlitre |
| μm | micrometer |
| μM | micro Molar |
| v/v | volume /volume |
| w/v | weight/volume |

CHAPTER 1

INTRODUCTION

The term plant genetic engineering has long conveyed a highly efficient and precise process for the manipulation of plant genomes (Ow, 2007). This technology has become a versatile platform for cultivar improvement, expression and production of valuable proteins, and as well as studying gene function in plant. Recently, much effort has been channeled to develop the transformation system for medicinal and aromatic plants. In our country, *Centella asiatica* (*C. asiatica*) is an important medicinal plant that is grown for commercialization purposes. *C. asiatica* is non-toxic, easily grown, high in medicinal values, and can serve as a good platform for genetic engineering. Unfortunately, to date no efficient transformation protocol has been developed and genetic manipulation been carried out to fully exploit this essential medicinal herb. The aims of this research study are to develop an efficient particle bombardment transformation protocol for *C. asiatica* embryogenic calli and to express HN protein from Newcastle disease virus strain AF2240 based on the establish transformation protocol.

Newcastle disease (ND) is a worldwide economic problem in poultry industry caused by NDV. The continued presence of ND causes economic losses to the industry in terms of production (such as in death, loss of body weight and impaired egg production) and control (such as quarantine, mass slaughter and disinfection) (Wong, 2004). The current protocol for NDV vaccination utilizes



both inactivated and live viruses for the control of ND in poultry. Although both types of vaccination produce excellent immunity among the flocks, the drawback of these vaccines still remain the major obstacle in the industry. Live vaccine will cause disease in the presence of the complicating infections, while it is often difficult to ensure complete inactivation of the virus which could remain as risk in inactivated vaccines (Wong, 2004). Studies had shown that one of the enveloped proteins of NDV known as haemagglutinin-neuraminidase (HN) protein is the potential candidate as immunogen for the development of NDV subunit vaccine. Passive immunization with either polyclonal or monoclonal antibodies to this protein is able to confer protection in chickens against NDV challenge (Reynolds and Maraqa, 2000). Thus many attempts had been applied to express the HN protein in different systems such as baculovirus and animal systems for the production of NDV subunit vaccine. Despite the successful expression and promising immunogenicity of the HN protein, they are still too costly for commercial production. As a result, a more feasible host protein expression system is needed to ensure the continuous supply of subunit vaccine for the control of NDV.

Extensive research over the past two decades has shown that plant can provide a new platform for the expression of recombinant proteins. Its protein synthesis pathway appears to be well conserved especially in glycosylation and post-translation modification enabling them for the production of various pharmacokinetics and biological active recombinant proteins. Moreover, plant is considered to be much safer than both animals and microbes because they generally lack human pathogens, endotoxin, and oncogenic sequences

(Commandeur, 2003). This system also provides lower cost of production and rapid scale up unlike the current available systems. Hence, plant can serve as a better and more feasible alternative system for the expression of recombinant proteins.

Thus the specific objectives of this study are:

1. to study the histochemical and morphological features of the target tissue, *C. asiatica* embryogenic calli;
2. to optimize transformation conditions for particle bombardment of *C. asiatica* embryogenic calli and to construct recombinant plasmids carrying the HN gene of NDV strain AF2240;
3. to bombard, select, verify the integration of transgene using PCR and to express the HN protein of NDV strain AF2240 in *C. asiatica* embryogenic calli.

CHAPTER 2

LITERATURE REVIEW

2.1 Newcastle Disease

Newcastle disease (ND) is a worldwide economic problem in poultry industry caused by Newcastle disease virus (NDV), a prototype *Avulavirus* in the subfamily *Paramyxovirinae* (De Leeuw and Peeters, 1999). Outbreaks of ND were first reported in poultry from Java, Indonesia and Newcastle-upon-Tyne in 1926 (Bruce et al., 2000). NDV has a wide host range with 27 of the 50 orders of birds reported to be infected by the virus (Kaleta and Baldauf, 1988). The NDV isolates can be divided into three main pathotypes: lentogenic, mesogenic, and velogenic depending on the severity of disease produced by the isolate.

Mildly virulent lentogenic strain usually causes mild respiratory infection in young chicks but not in adult birds. Mesogenic strain is more virulent and can induce mild disease with mortality accruing primarily in young chickens. Meanwhile the highly virulent velogenic strain induces severe diseases and mortality to birds of all ages. Differential diagnosis of NDV involves electron microscopic identification, hemagglutination inhibition with polyclonal NDV specific antisera, use of the ELISA, oligonucleotide probes, and viral genomic RNA fingerprint analysis (Bruce et al., 2000) The current protocol for NDV vaccination utilizes both inactivated and live viruses for the control of ND in poultry. Various routes of vaccination were applied such as injection, inhalation