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Surface Enhanced Raman Scattering of Crystal Violet

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ABSTRACT

Despite the ubiquity of Raman spectroscopy, fluorescence, poor signal strength and photobleaching pose a significant challenge to researchers in the biomedical field. Here, we demonstrate a 17-fold signal enhancement in Raman spectra of crystal violet via surface-enhanced Raman scattering (SERS). The SERS substrate was fabricated by electron beam lithography (EBL); the nanostructured surface was an array of G-shaped elements made of Au on SiO₂/Si. In addition to the SERS spectra, finite-difference time-domain simulations were performed to illustrate the distribution of electric-field hot-spots on the SERS substrate. The electric-field hot-spots were prominent at the vertices and edges of the nanostructured G-shaped motifs. The results presented here demonstrate that EBL is a high-end choice for SERS substrate fabrication that opens the way for more complex Raman spectroscopies, for instance involving nonlinear optics or chiral analytes.

Keywords: Surface enhanced Raman scattering, crystal violet, finite-difference time-domain simulation, electron beam lithography

1. INTRODUCTION

Raman scattering spectroscopy is a versatile technique that exploits the inelastic scattering of light to characterize and identify chemicals and materials based on their quantized vibrational energy levels¹⁻⁴. When light inelastically scatters via a light-matter interaction, the light can lose energy (Stokes scattering) by increasing the energy of vibration (or rotational-vibration) of the scattering analyte. If, however, the analyte is initially in an excited state of vibrational energy, the light can gain energy via the scattering event leaving the analyte in a lower energy state. Most instruments observe the Stokes Raman shift in wavelength of the scattered light meaning that the observed wavelength is larger than the incident light. The observed Raman shifts are typically quantized according to the quantization of vibrational energy levels in the scattering material and are subject to selection rules⁵. Hence, crystals and molecules can be identified by their unique Raman spectrum of distinct peaks.

Raman spectroscopy is particularly attractive to biologists as it is a chemically selective hyperspectral imaging technique that seldom requires pretreatment of the specimen with molecular tags. These pretreatment techniques of tissues invariably alter the metabolic processes of the cells^{6,7}. Despite these advantages, Raman spectroscopy can be challenging with certain types of samples due to fluorescence or photobleaching. Sometimes, these effects can be mitigated by increasing the wavelength of the incident light or by algorithmically removing the background fluorescence⁸. However, Raman scattering is an inherently weak effect with only 1 in 10⁸ photons undergoing spontaneous Raman scattering⁹. Hence, researchers often employ plasmonic effects to enhance the Raman signal.

Raman spectroscopy has become a useful tool for characterizing a variety of materials, such as 2D materials (including graphene¹⁰⁻¹² and transition metal dichalcogenides¹³⁻¹⁵). Raman spectroscopy can help establish the number of monolayers^{11,14,16,17}, interlayer breathing and shear modes¹⁸, in-plane anisotropy¹⁹, doping²⁰⁻²², disorder^{12,23-25}, thermal conductivity¹³, strain²⁶ and phonon modes²⁷⁻²⁹.

Surface enhanced Raman scattering (SERS) was first observed in 1974 when Fleischmann et al. noticed a significant increase in the Raman signal of pyridine on a roughened silver electrode³⁰. SERS can be achieved using various forms of nanostructured materials, such as plasmonic nanoparticles. Surface plasmons are coherent oscillations of the surface electrons in metals that can be driven by visible light³¹. Numerous theoretical models have been proposed in order to achieve such tuning^{32,33}. Nanoparticle aggregates are a common method for SERS but the consistency of aggregation, and hence the homogeneity of enhancement, can be challenging^{34–38}. Electron beam lithography is an exquisitely precise method for producing SERS substrates with homogeneous hot-spots of SERS^{39,40}. The G-shaped nanoarrays have been the subject of numerous experimental studies, especially using second harmonic generation^{41,42} – an interface-sensitive technique^{43,44} whose efficiency scales as the fourth power of the optical near-fields, similarly to the enhancement factor of SERS^{45,46}.

Here, we demonstrate SERS using plasmonic G-shaped nanostructures fabricated from Au on a SiO₂/Si substrate using electron beam lithography as detailed in refs⁴⁷. Crystal violet (CV) is used as the analyte as it constitutes an excellent reference, with a distinct and well known Raman spectrum⁴⁸.

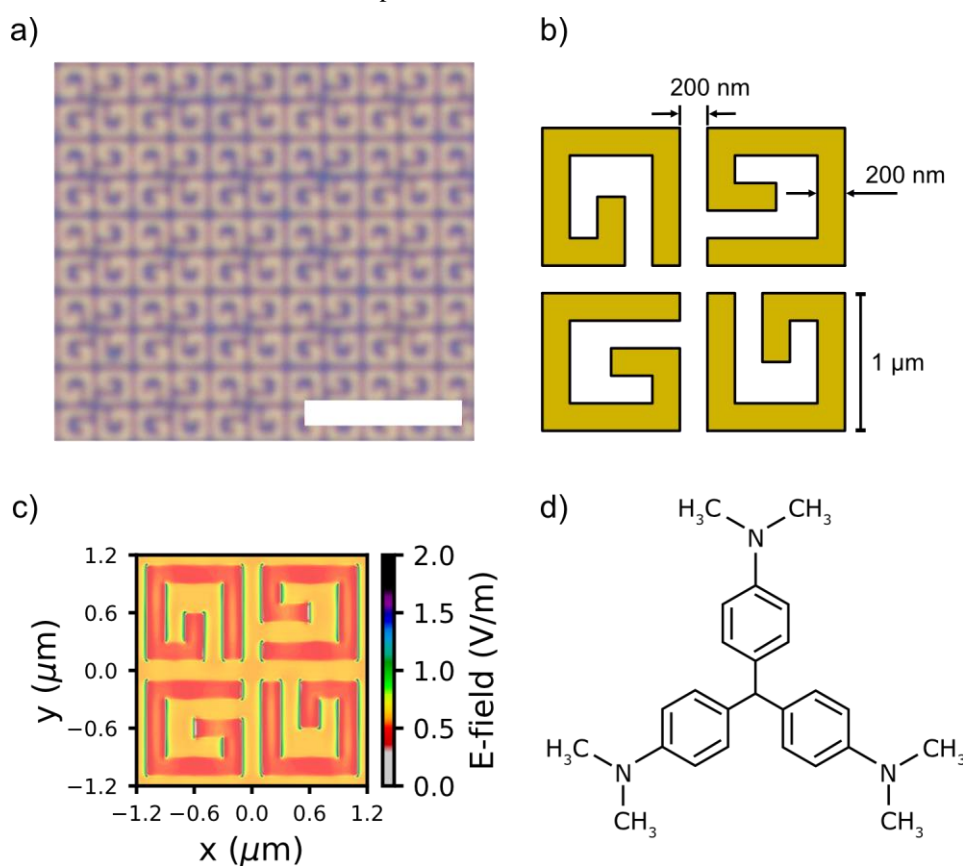


Figure 1. a) bright field microscope image of the G-shaped SERS substrate; scale bar 5 μm . b) schematic of the geometry of the nanostructured SERS substrate fabricated as per refs⁴⁷ c) simulation results of the electric-field distribution at 532 nm taken 1 nm above the SERS substrate. d) molecular structure of crystal violet.

2. EXPERIMENTAL PROTOCOL

Raman spectra of CV were acquired using a Renishaw InVia Raman microscope. A continuous wave laser (50 mW Cobolt RL532-08) was used to provide a narrow bandwidth light source at 532 nm on the sample. The spectrometer used a 1800 lines/mm grating blazed for 532 nm. The incident radiation and epi-scattered light were focused through an N-plan 50 \times objective with a numerical aperture of 0.75. Unless otherwise stated, spectra were averaged from a square grid of 25 uniformly distributed points (1 μm apart for CV on SERS and 5 μm apart for CV on Si to account for the increased inhomogeneity in CV distribution on bare Si). At each point on the grid, the spectrum acquisition was 10 seconds with

the spectrometer slit width set to 60 μm wide. The spectral resolution of the spectrometer is 0.3 cm^{-1} . The fluorescence background was removed using a polynomial fit algorithm in the Renishaw WiRE software.

The SERS substrate was fabricated on a Silicon substrate with a thermally grown 200 nm SiO_2 surface layer. A double polymethyl methacrylate-methyl methacrylate resist layer was used to cover the substrate. The G-shaped nanostructures were then created by electron beam lithography. A 25 nm layer of Au was deposited by evaporation using a DC sputtering system⁴⁷. The resist was then removed by a ‘lift-off’ procedure. Figure 2 a) shows a bright field microscope image of the G-shaped nanostructured SERS substrate taken using a 100 \times objective. The geometry of the unit cell with the associated dimensions is shown in Figure 3 b); the unit cell comprised of four such motifs in a four-fold tetrad. The array of Au nanostructures covered an area of 2.5 mm \times 2.5 mm on the Si substrate. Hence, the enhancement of the Raman signal from CV on the SERS part of the sample was compared to measurements of CV on the clean Si area of the sample.

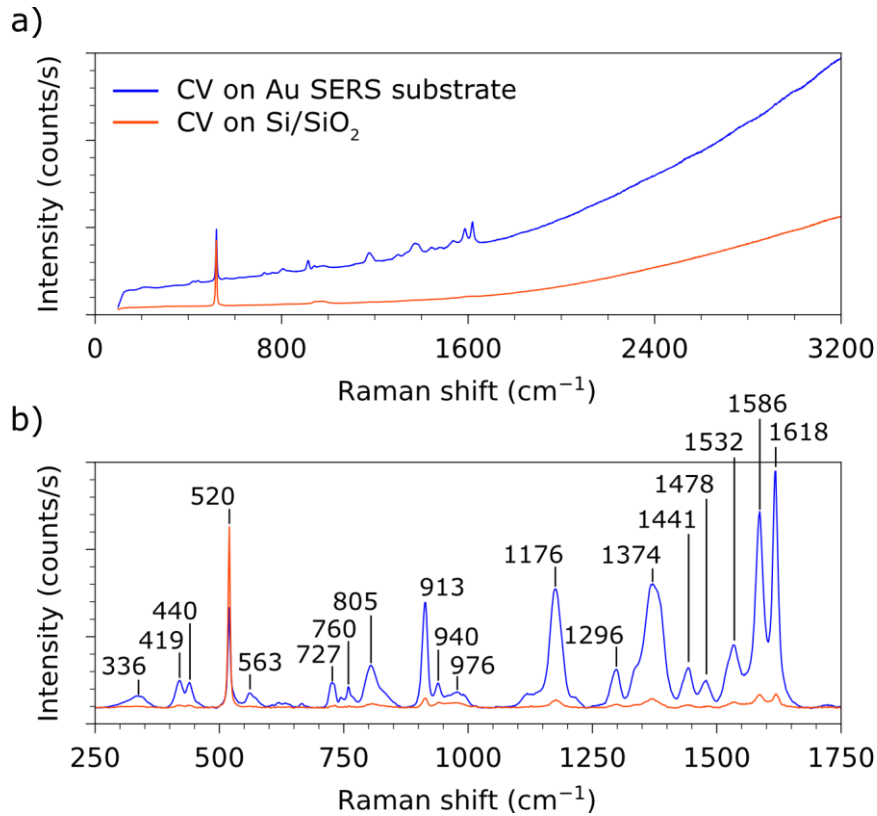


Figure 4. Raman spectra of crystal violet on the SERS substrate and on silicon using 532 nm incident excitation. a) full range of Raman shifts taken at a single point with 100% of the available laser power for 10 seconds. b) spectral region of interest with fluorescence background removed taken from a uniform 5 \times 5-point grid (25 points in total) with a 10 second acquisition per point and averaged; the laser power was attenuated to 10% using neutral density filters. The sample points in the map had a granularity of 1 μm on the SERS substrate and 5 μm on the silicon substrate to account for increased inhomogeneity in the distribution of CV on bare Si.

Finite-difference time-domain simulations can be performed with a variety of Maxwell equations solvers, such as MAGMAS^{49,50}, Lumerical⁵¹, MEEP⁵², RSoft’s DiffractMOD⁵³, etc. We performed simulations in ANSYS™ Lumerical to illustrate the concentration of electric field ‘hot-spots’ at the vertices of the G-shaped structures. The geometry of the simulation was based on that shown in Figure 5 b). Periodic boundary conditions were imposed around the edges of the unit cell to simulate an infinite array of unit cells. The Si substrate model was a 500 nm thick slab of Si based on Palik material model⁵⁴. The G-shaped unit cell was modelled in Autodesk Inventor and imported into Lumerical with a CRC material model⁵⁵. A plane wave source was introduced from 1 μm above the surface with a wavelength range of 0.2 to 1.8 μm . The void above the plane wave source extended by an additional 1 μm . Perfectly

matched layer boundary conditions were applied to the top and bottom plane of the simulation domain. Hence, the simulation perceived the Si substrate to be infinite in extent. Figure 6 c) shows the electric field distribution at 532 nm taken in the plane 1 nm above the surface of the Au G-shaped nanostructures. The electric field hot-spots are clearly concentrated at the edges and vertices of the G-shaped motifs as expected.

A 0.3 mM solution of CV in ethanol was drop-casted onto the SERS substrate and allowed to evaporate in air. Because CV has three amino groups, it bonds to the Au surface and the effect of liquid tension during the evaporation of ethanol is minimized. Figure 7 d) shows a schematic of the organic molecule that is used for our demonstration of SERS; it has also been used in surface enhanced hyper-Raman scattering experiments⁵⁶.

3. RESULTS AND DISCUSSION

Figure 8 a) shows a spectrum of CV on the Au SERS substrate (blue) and on bare Si (red). These spectra (Fig. 9a) were acquired using 100% of the available laser power at a single point on the sample with an acquisition time of 10 seconds. The presence of fluorescence is evident and was subsequently removed using a polynomial fit algorithm in the Renishaw WiRE software. The distinct peak at 520 cm^{-1} corresponds to the Raman peak of Si.

Table 1. Raman peaks of crystal violet identified on the SERS substrates with associated band assignments measured using 532 nm excitation; band assignment taken from ref.⁴⁸.

Raman shift (cm^{-1})	Band assignment
727, 760, 805	C-H out of plane bending
913, 940, 976	Ring skeletal vibrations
1176	C-H in plane bending
1374	N-phenyl stretching
1532, 1586, 1618	C-C stretching

Figure 10 b) presents the spectra of CV measured on the SERS surface and on the Si surface in the finite range of Raman shifts of interest. The fluorescence background has been removed using the method stated above. Again, the Raman peak of Si at 520 cm^{-1} is observed. The prominent Raman peaks associated with CV are labelled and identified. The band assignments are presented in Table 1 and are in good agreement with refs^{48,57}. The enhancement of the signal on the SERS sample is approximately 17 \times that of the measurements taken from CV on the Si surface. Hence, we demonstrate that these samples, fabricated using electron beam lithography, are excellent SERS substrates with good homogeneity.

4. CONCLUSIONS

We have demonstrated a 17-fold signal enhancement of Raman scattering spectra in crystal violet using a G-shaped Au nanostructured SERS substrate relative to CV on Si wafer. Finite-difference time-domain simulations revealed that the electric field enhancement was most concentrated in the vicinity of the edges and vertices of the nanostructured motifs. EBL is a rather costly and time-consuming method, with limited potential for scaling up production, especially in the case of large area SERS substrates. However, the commercial availability of SERS substrates with homogeneous and reproducible enhancement in Raman scattering is also limited. EBL provides excellent nanoscale precision and design freedom that can be greatly beneficial for tailoring the SERS substrate to a particular analyte. The SERS enhancement can be further augmented by tuning the resonance wavelength of the nanoparticles to that of the incident laser. Nanostructures with a chiral design⁵⁸, such as the G-shaped nanoarrays, could next be used for chiral optical SERS investigations. Moreover, as they have well-known nonlinear optical conversion properties, these nanoarrays open the way for surface enhanced hyper-Raman scattering studies. Therefore, our work paves the way for SERS-based investigations of chiral organic molecules, such as pesticides and pharmaceuticals in the environment. Moreover, SERS find applications in the detection of explosives/chemical weapons⁵⁹, unmodified DNA⁶⁰, and pathogens⁶¹.

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SUPPORTING INFORMATION

All data supporting this study are openly available from the University of Bath data archive at <https://doi.org/10.15125/BATH-00991>.

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