

UNIVERSITI PUTRA MALAYSIA

AETIOLOGIC AGENTS OF FRY MORTALITY SYNDROME IN THE RAINBOW TROUT (Oncorhynchus mykiss) IN IRAN

SEYED MOHAMMAD EBRAHIM JALIL ZORRIEHZAHRA

FPSK(P) 2008 5



AETIOLOGIC AGENTS OF FRY MORTALITY SYNDROME IN THE RAINBOW TROUT (Oncorhynchus mykiss) IN IRAN

By

SEYED MOHAMMAD EBRAHIM JALIL ZORRIEHZAHRA

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in the Fulfilment of Requirements for the Degree of Doctor of Philosophy

30 October 2008



DEDICATION

WITH LOVE AND APPRECIATION TO:

My dear wife: Masoumeh Kohinejad

My dear son: Seyed Mohammad Ehsan

My dear brothers: Seyed Kamal, Seyed Jalal, Seyed Amir Ahmad and

Seyed Mohammad Mehdi

and

My dear sister: Eftekhar Sadat



Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in the fulfilment of the requirements for the Degree of Doctor of Philosophy

AETIOLOGIC AGENTS OF FRY MORTALITY SYNDROME IN RAINBOW

TROUT (Oncorhynchus mykiss) IN IRAN

By

SEYED MOHAMMAD EBRAHIM JALIL ZORRIEHZAHRA

April 2008

Chairman: Associate Professor Dr Hassan Hj. Mohd Daud, Ph.D.

Faculty: Veterinary Medicine

An investigation was conducted in order to find out the etiological factors of Fry

Mortality Syndrome (FMS) that causes serious economical loss in rainbow trout

farms in Iran. In recent years obscure fry mortalities have been observed in many

hatchery farms in Iran. It was reported that the rate of fry and juvenile mortality

increased dramatically in some provinces e.g. 23 million fry were produced in

hatchery centers of Chahar Mohal Bakhtiary province in 2002 but nearly 21 million

fry (91.3%) in different stages of growth died before distribution to farmers. Also

close to 23 million fry were produced in Mazandaran province, but 12 million fry

equivalent to 52.12% of total fry production died mysteriously. This investigation

was carried out with objectives of detecting and confirming the main causative agent

that contribute to the occurrence of Fry Mortality Syndrome in Iran. During 32

months, from October of 2001 until May of 2004, 52 different hatchery centers and

rearing farms of rainbow trout (Oncorhynchus mykiss) which were located in Tehran,

Mazandaran, Guilan, Fras, Markazi, Kerman and Kohkiloyeh Boyerahmad provinces, were visited and various samples from affected farms were collected. Collected samples consisted of ovarian fluid, milts, eggs, eyed-eggs, larvae, fry < 1 g and 1-3 g as well as internal organs from adult fishes. A total of 2,107 samples were collected from farms in six provinces and were examined by five methods such as virology (410 samples), bacteriology (899 samples), serology (consisted of IFAT: 392 samples and ELISA: 44 samples), histopathology (160 samples) and hematology (202 samples). Some of the mentioned approaches such as fish cell culture, ELISA and IFAT techniques were set-up and optimized for the first time in Iran.

The clinical signs of suspected fishes were darkening, exophthalmia, ascites, abnormal swimming and whirling. From 410 samples that of tissues inoculated on to cell cultures two samples showed CPE in EPC and BF-2 cell lines which were inoculated with ovarian fluid from broodstock obtained from hatchery farms in Mazandaran province. The CPE was similar to IHN virus induced. The CPE foci revealed dying cells congegrated as grape-like clusters (ballony performance with cytolysis). TEM findings in infected cells showed bullet-shaped particles having sizes of 130-180 nm in length and 65-70 nm in diameter. From the virion morphology it was suggested that observed particles were similar to Rhabdovirus. FAT examination revealed that all samples were examined with MAbs and PAbs against IPNV and VHSV were negative. On the other hand, two samples were positive when examined with MAbs and PAbs against IHNV. These smears were originated from samples that had showed CPE in EPC and BF-2 cell lines and bullet shaped particles in electron microscopy. ELISA findings (cut-off value, optical density and detection-level percentage) showed that IHNV had higher percentage of detection with 23.25%



in comparison with other relevant viral diseases i.e. IPNV with 7.31% and VHSV with 14.29%. Results of histopathological study on the sampled fry revealed that the target tissues in the kidney, liver, spleen, hepatopancreas, intestine and gills showed different degree of tissue changes beginning from cell degeneration to complete necrosis. There were also renal blood vessels congestion, marked degenerative changes in posterior kidney with tubular necrosis and interstitial hematopoeitic tissue degeneration. In addition, interstitial degeneration and oedema in anterior portion of kidney, focal necrosis in the tubular area and several stages of cell necrosis in the hematopoeitic tissue were the most important histopathological changes seen in kidney tissues examined. Hepatopancreatic tissues also revealed marked changes such as congestion, atrophy and necrosis of pancreatic acinar cells and Islets of Langerhans. Spleen samples revealed spleenic congestion, severe necrosis, hemosiderosis and increased presence of melanomacrophage centers (MMC). Gills tissue in sampled fry showed hyperplasia, clubbing and fusion of lamellae. Hematological findings revealed that total white blood cell count, i.e. lymphocyte and neutrophil in investigated fish showed significant increased compared with the

Hematological findings revealed that total white blood cell count, i.e. lymphocyte and neutrophil in investigated fish showed significant increased compared with the control fish (p< 0.05). On the contrary, all the samples showed a decreased in RBC, Hb and HCT values. In addition, MCHC and total protein plasma showed a marked decreased (p<0.05). In the blood serum components analysis, similarly it was revealed LDH and AST showed a significant decreased (p<0.05).

In conclusion, with marked clinical signs, cell culture observation and TEM findings, ELISA and IFAT results, histopathology and hematological findings (blood and biochemical parameters) seen in the current investigation lead to possibility of a viral disease agent infection as the cause of fry mortality syndrome in the hatchery



and rearing trout farms in Iran. From findings of the current study, it is concluded that IHN-like virus could be most probable etiologic of fry mortality syndrome in Iran.

Key words: Fry Mortality Syndrome, Rainbow trout, Cell culture, TEM, ELISA, IFAT, Histopathology, Hematology, IHNV, IPNV, VHSV, Iran



Oleh

SEYED MOHAMMAD EBRAHIM JALIL ZORRIEHZAHRA

April 2008

Pengerusi: Profesor Madya Dr. Hassan Hj. Mohd Daud, Ph.D.

Fakulti: Perubatan Veterinar

Satu penyiasatan telah dijalankan untuk menentukan faktor etiologik Sindrom

Kematian Anak Ikan yang telah menyebabkan kehilangan ekonomi yang serius

dalam ladang ikan trout pelangi di Iran. Dalam tahun kebelakangan ini, kematian

anak ikan yang tidak di ketahui punca telah dilihat di banyak ladang penetasan di

Iran. Di lapurkan bahawa kadar kematian anak ikan dan ikan juvenil telah

meningkat secara mendadak di beberapa daerah, contohnya 23 juta anak ikan di

keluarkan di pusat penetasan daerah Chahar Mohal Bakhtiary dalam tahun 2002,

hampir 21 juta anak ikan (91.3%) dalam pelbagai peringkat pertumbuhan mati

sebelum dapat diedarkan kepada penternak. Juga hampir 23 juta anak ikan yang

dihasilkan di daerah Mazandaran, 12 juta iaitu 52.12% dalam jumlah keseluruhan

anak ikan mati secara misteri.

Kajian ini dijalankan dengan objektif untuk mengesan dan mengesahkan agen utama

penyebab kejadian sindrom kematian anak ikan di Iran. Dalam masa 32 bulan iaitu

dari Oktober 2001 hingga Mei 2004, sejumlah 52 pusat penetasan dan ladang

peliharaan ikan trout pelangi (Oncorhynchus mykiss) terletak di daerah Tehran,

vii

Mazandaran, Guilan, Fras, Markazi, Kerman dan Kohkiloyeh Boyerahmad dilawati dan pelbagai sampel dikumpulkan. Sampel yang diambil termasuk cecair obari, cecair sperma, telur, telur bermata, larvae, ikan fri bersaiz < 1 gm dan bersaiz 1-3 gm dan juga organ dalaman dari ikan dewasa. Sebanyak 2,107 sampel telah dikumpul dari ladang di enam daerah dan disiasat menggunakan lima tatacara iaitu virologi (410 sampel), bakteriologi (899 sampel), serologi (terdiri dari IFAT: 392 sampel dan ELISA: 44 sampel), histopatologi (160 sampel) dan hematologi (202 sampel). Sesetengah dari prosedur seperti kultur sel ikan, ELISA dan IFAT teknik adalah pertama dibangunkan dan optimakan di Iran.

Tanda klinikal ikan yang dijangkiti adalah kehitaman, eksoptalmia, asites, berenang tidak normal dan berputar. Daripada 410 sampel yang diinokulasikan dalam kultur sel, dua sampel menunjukkan kesan sitopatik dalam sel turutan EPC dan BF-2 yang mana diinokulasi dengan cairan ovum dari induk berasal pusat penetasan di daerah Mazandaran. Kesan sitopatik itu adalah sama seperti cetusan virus IHN. Kawasan kesan sitopatik menunjukkan sel-sel nazak berkumpul seperti buah anggur (berbentuk belon dan sitolisis). Keputusan TEM menunjukkan partikel berbentuk peluru bersaiz 130-180 nm panjang dan 65-70 nm diameter. Dari morfologi partikel virion yang dilihat ianya adalah serupa seperti Rhabdovirus. Pemeriksaan IFAT menunjukkan kesemua sampel yang diuji dengan MAbs dan PAbs terhadap IPNV dan VHSV adalah negatif. Walaubagaimana pun dua sampel adalah positif apabila diuji dengan MAbs dan PAbs terhadap IHNV. Smer ini adalah berasal dari sampel yang menunjukkan kesan sitopatik dalam sel turutan EPC dan BF-2 dan partikel berbentuk peluru dalam TEM. Keputusan ELISA (titik penggalan, ketumpatan optik dan peratus aras pengesanan) menunjukkan bahawa IHNV mempunyai peratus



pengesanan setinggi 23.25% berbanding dengan penyakit virus relevan yang lain seperti IPNV dengan 7.31% dan VHSV dengan 14.29%. Keputusan kajian hematologi pada anak ikan menunjukkan bahawa tisu tumpuan dalam ginjal, hepar, limfa, hepatopankreas, usus dan insang memperlihatkan pelbagai tahap perubahan bermula dengan degenerasi sel hingga nekrosis penuh. Terdapat kongesi saluran darah renal, perubahan degeneratif nyata di ginjal posterior dengan nekrosis tubular dan degenerasi tisu perantaraan hematopoeitik. Tambahan pula, degenersasi tisu perantaraan dan edema dalam ginjal anterior, nekrosis fokus dalam kawasan tubul dan beberapa peringkat nekrosis sel dalam tisu hematopoeitik adalah perubahan histopatologi yang penting dalam tisu ginjal yang diperiksa. Tisu hepatopankreas mempamerkan juga perubahan nyata seperti kongesi, atrofi dan nekrosis dalam sel asinar pankreas dan Islets of Langerhans. Sampel limfa menunjukkan kongesi, nekrosis teruk, hemosiderosis dan penambahan kehadiran pusat melanomakrofaj (MMC). Tisu insang dalam anak ikan yang disampel menunjukkan hiperplasia, berbentuk "club" dan percantuman lamella.

Keputusan hematologikal menunjukkan bahawa jumlah sel darah putih iaitu limfosit dan neutrofil dalam ikan yang disiasat menunjukkan keputusan yang bererti apabila dibandingkan dengan ikan kawalan (p<0.05). Ikan-ikan tersebut menunjukkan peningkatan dalam jumlah sel darah putih, limfosit dan neutrofil. Walaubagaimana pun kesemua sampel ikan menunjukkan penurunan nilai sel darah merah, hemoglobin dan hematokrit yang bererti (p<0.05). Tambahan lagi, MCHC dan jumlah protein plasma juga menunjukkan kekurangan yang nyata (p<0.05). Dalam analisis komponen serum, ia telah menunjukan bahawa LDH dan AST telah menurun dengan bererti (p<0.05).



Pada kesimpulannya, dengan tanda klinikal yang nyata, pemerhatian kultur sel, penemuan TEM, keputusan ELISA dan IFAT, serta penemuan histopatologi dan hematologi dalam kajian ini telah mengarah kepada kemungkinan bahawa jangkitan penyakit virus adalah penyebab sindrom kematian anak ikan dalam pusat penetasan dan ladang ternakan di Iran. Dari penemuan kajian sekarang, adalah disimpulkan bahawa virus serupa IHN adalah agen etiologik utama penyebab sindrom kematian anak ikan di Iran.

Perkataan kunci: Sindrom Kematian Anak Ikan, Trout Pelangi, Kultur sel, TEM, ELISA, IFAT, Histopatologi, Hematologi, IHNV, IPNV, VHSV, Iran



ACKNOWLEDGEMENTS

Praise is to the Almightly Allah. Lord of all creation, for his heavenly, luxurious blessings over me throughout my life and the period of this study.

I would like to express my heartfelt gratitude and appreciation to my main Supervisor, Associate Professor Dr. Hassan Hj. Mohd Daud, for his kindly and valuable guidance and constructive suggestions throughout the period of my study. I sincerely appreciate the innumerable hours he spent reading the draft and the suggestions made to improve the thesis.

I wish to express my deepest thanks to my co-supervisors: Professor Dr. Mohd Hair Bejo and Professor Dr. Mehdi Soltani, for their valuable suggestions and kind assistance throughout this study.

I wish to express my deepest thanks to my dear wife, lovely son Mr.Ehsan for their endless support, lovely encouragement and patience during my life and study. These words would be one drop in front of their ocean of kindness.

A very special acknowledgement is given to my dear brother Dr. Sohrab Rezvani and Dr. Abbas Ali Motallebi, Head of Iranian Fisheries Research Organization (IFRO), and Dr.Mostafa Shariff Rohani, Research Deputy of IFRO for their cooperation during the process of conducting the study.



Very special thanks go my dear friends Dr. Issa Sharifpour for his vision and constant encouragement, Dr.Mohammad Gholizadeh and Dr. Hamid Sanatnama for their kind assistance during my study in UPM and Dr.Ali Asghar Saiedi for his kind efforts in research program.

I would like to express my thanks to all my friends whom I obtained their assistance during this study especially Dr. Mohammad Reza Mehrabi, Dr. Rozbeh Fallahi and Dr. Abolfazle Sepahdari and Dr. Taheri, Mr.Bagheri, Mr.Saydanlo from Veterinary Faculty of Tehran University for his invaluble help and assistance.

I am also grateful to the staff of Iranian Fisheries Research Organization, Faculty of Veterinary Medicine, University of Tehran, for their cooperation.

Last but not least, I would like to record my gratitudes to many others whose name do not appear here, who have helped me during my study period.



I certify that an Examination Committee has met on 30 October 2008 to conduct the final examination of Seyed Mohammad Ebrahim Jalil Zorriehzahra on his Doctor of Philosophy thesis entitled "Aetiologic Agents of Fry Mortality Syndrome in the Rainbow Trout *(Oncorhynchus mykiss)* in Iran" in accordance with Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Examination Committee were as follows:

Mohamed Ali Rajion, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Tengku Azmi Tengku Ibrahim, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Abdul Rani Bahaman, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Faizah Mohd. Shaharom, Ph.D

Professor Institute of Tropical Aquaculture Universiti Malaysia Terengganu (External Examiner)

HASANAH MOHD.GHAZALI, PHD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 29 January 2009



This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee are as follows:

Hassan Hj Mohd Daud, Ph.D

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Mohd Hair Bejo, Ph.D

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

Mehdi Soltani, Ph.D

Professor Faculty of Veterinary Medicine Tehran University (Member)

HASANAH MOHD.GHAZALI, PHD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 12 February 2009



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

SEYED MOHAMMAD EBRAHIM JALIL ZORRIEHZAHRA

Date:



TABLE OF CONTENTS

A A D T L	PPROY ECLA ABLE IST OI IST OI	ACT AK WLEDGEMENTS	Page iii vii xi xiii xv xvi xxiii xx xxiii
	HAPT		
1	INTR	ODUCTION	1
	1.1	Importance of study	1
	1.2	Objectives of the study	3
	1.3	Hypothesis	3 3
	1.4	Research approach	3
2	LITE	RATURE REVIEW	6
	2.1	Characteristics and structure of the sector	6
		2.1.1 History and general overview	8
		2.1.2 Farming System	9
		Warm water fish culture	9
		Cold water fish culture (Rainbow trout) Inland based fisheries	10 11
		2.1.3 Sector performance	11
		Production	11
		Aquaculture contribution to Economy	13
		2.1.4 Trout farming	14
		2.1.5 New technology acquisition	14
		2.1.6 Geographical distribution of Coldwater Aquaculture areas in Iran	15
		2.1.7 Overview of the Fry Mortality Syndrome in Iran and the world	18
	2.2	Infectious agents	20
		2.2.1 Bacteria agent	20
		2.2.2 Viral agents2.2.3 Parasite Diseases	29 35
	2.3	Non- Infectious agents	36
	5	2.3.1 Nutritional factors	36
		2.3.1 Nutritional factors 2.3.2 Environmental factors	39
	2.4	The fish immune system	43
	, :	2.4.1 Lymphoid organs	44
		2.4.2 Innate Immunity	45
		2.4.3 Adaptive Immunity	46



	2.5	Diseases Status in Iran	48
		2.5.1 Bacterial Diseases	49
		2.5.2 Viral Diseases	50
		2.5.3 Non-Infectious Diseases	50
3	DETE	RMINATION OF THE PRESENCE OF VIRAL AGENTS IN	52
		CTED FISH WITH FRY MORTALITY SYNDROME USING CULTURE ISOLATION AND TRANSMISSON ELECTRON	
		COSCOPY	
	3.1	Introduction	52
	3.2	Research objectives	53
	3.3	Fish Sampling	53
		3.3.1 Samples collection methods	56
		Sampling from reproductive secretions of broodstock (ovarian fluid, ova and milt)	56
		Sampling of green egg, eyed-egg and yolk sac fry from hatchery	57
		Sampling from fry less than one gram and fry1-3 gm Sampling of internal organs	57 59
	3.4	Smear preparation from broodstock's gonadal secretion (Milt, ovarian	60
	5	fluid and ova)	00
	3.5	Samples preparation for inoculation on fish cell line	61
	3.6	Media preparation and Cell line cultivation	62
	3.7	Culture Media Preparation	65
	3.8	Cell line passage	66
	3.9	Samples inoculation onto cell lines	67
	3.10	Electron Microscopy	68
	3.11	Results	68
		3.11.1 Virus isolation	68
		3.11.2 Electron Microscopy Examination (TEM)	72
	3.12	Discussion	73
4		TIFICATION OF BACTERIA ISOLATED FROM INFECTED	76
		(FRY, BROODSTOCK) WITH FRY MORTALITY SYNDROME	
	4.1	Introduction	76
	4.2	Research objectives	78
	4.3	Materials and Methods	78
		4.3.1 Fish sampling	78 79
		Sampling from redult fish	79 79
		Sampling from adult fish Sampling of fish external surface	83
		Sampling of fish internal organs	83
		Small fish (10-15 g)	84
		Adult fish >15 g	84
	4.4	Results	84
		4.4.1 Clinical sign results	84
		4.4.2 Bacteriological examination results	86
		Fry and larvae	86



		Adult Fish	88
	4.5	Discussion	88
5	(FRY	FIRMATION OF VIROLOGICAL FINDINGS IN INFECTED FISH , BROODSTOCK) WITH FRY MORTALITY SYNDROME USING AND ELISA	91
	5.1	Introduction	91
	5.2	Research objectives	92
	5.3	Serology examination	92
		5.3.1 Fluorescent Antibody Test	92
		Materials and Methods	93
		Procedure of Indirect Fluorescent Antibody Test (IFAT)	104
		5.3.2 Enzyme Linked Immunabsorbant Assay (ELISA)	105
		Materials and Methods	106
		Procedure of ELISA examination	106
		Main ELISA examination for antibody detection against IPNV and VHSV in broodstock serum	113
	5.4	Results	115
		5.4.1 Indirect Fluorescent Antibody Test	115
		5.4.2 ELISA	123
	5.5	Discussion	127
6	INFE	SSMENT OF HISTOPATHOLOGICAL CHANGES IN CTED RAIN BOW TROUT FRY AND FINGERLINGS WITH FRY TALITY SYNDROME	132
	6.1	Introduction	132
	6.2	Research objectives	132
	6.3	Materials and Methods	133
		6.3.1 Sample Collection	133
	6.4	Results	136
		6.4.1 Clinical Signs	136
		6.4.2 Histopathological Changes	
		Gills	136
		Intestine	136 137
			137 138
		Liver	137 138 141
		Hepatopancreas	137 138 141 145
		Hepatopancreas Kidney	137 138 141 145 147
		Hepatopancreas Kidney Spleen	137 138 141 145 147 151
	6.5	Hepatopancreas Kidney	137 138 141 145 147
7	ANAI PARA	Hepatopancreas Kidney Spleen Discussion and Conclusion LYSES AND COMPARISION OF HAEMATOLOGICAL AMETERS AND BLOOD ENZYMES BETWEEN INFECTED AND	137 138 141 145 147 151
7	ANAI PARA CONT	Hepatopancreas Kidney Spleen Discussion and Conclusion LYSES AND COMPARISION OF HAEMATOLOGICAL AMETERS AND BLOOD ENZYMES BETWEEN INFECTED AND TROL FRY SAMPLES	137 138 141 145 147 151 156
7	ANAI PARA CONT 7.1	Hepatopancreas Kidney Spleen Discussion and Conclusion LYSES AND COMPARISION OF HAEMATOLOGICAL AMETERS AND BLOOD ENZYMES BETWEEN INFECTED AND FROL FRY SAMPLES Introduction	137 138 141 145 147 151 156
7	ANAI PARA CONT 7.1 7.2	Hepatopancreas Kidney Spleen Discussion and Conclusion LYSES AND COMPARISION OF HAEMATOLOGICAL AMETERS AND BLOOD ENZYMES BETWEEN INFECTED AND FROL FRY SAMPLES Introduction Research objectives	137 138 141 145 147 151 156 161
7	ANAI PARA CONT 7.1	Hepatopancreas Kidney Spleen Discussion and Conclusion LYSES AND COMPARISION OF HAEMATOLOGICAL AMETERS AND BLOOD ENZYMES BETWEEN INFECTED AND FROL FRY SAMPLES Introduction	137 138 141 145 147 151 156



7.3.3 Blood enzymes measurement	169	
7.4 Results	170	
7.4.1 Statistic Data Analysis	178	
7.5 Discussion	178	
8 OVERALL DISCUSSION AND CONCLUSIONS	185	
REFERENCES	195	
APPENDIX		
A. Virology Examination	216	
B. Bacteriology Examination		
C. Serology Examination		
D. Histophatology Examination	233	
E. Hematology Examination	234	
RIODATA OF AUTHOR	235	



LIST OF TABLES

1	Aquaculture & Aquaculture based Fisheries trend in Iran (1997-2004)	12
2	Top 10 Farmed Trout-Producing Countries, 2005	12
3	Aquaculture Areas in Iran, 2003	13
4	Number of farm, pond area and annual production of Cold water fishes Source: Year book of Iran Fisheries Statistics, 2006	15
5	Total projection of Cold water fish production in different	
	provinces of Iran	16
6	Summary of vitamin deficiency signs (Halver, 1978)	39
7	Comparison between components of the immune system (Marh, 2008)	48
8	Samples numbers from hatchery and grow-out rainbow trout farms in various provinces of Iran used virological examination	55
9	Name of provinces and type of samples for virological examination	55
10	Total number and types of samples from six provinces	55
11	Technical information of the cell lines	63
12	Province distribution of eggs and larvae samples from some hatcheries and rearing farms for bacteriology examination	80
13	Province distribution of adult fish samples from some rearing farms for bacteriology assay	82
14	Isolated bacteria from fry Rainbow trout samples from some	87



hatchery and rearing farms for bacteriology examination

15	Group distribution of collected samples for IFAT examinations	97
16	Tissue distribution of collected samples for IFAT examinations	97
17	Score for flouresence colour reactions according to appearance in the dark-field fluorescence microscope	101
18	Specification of consumed viral antigens for polyclonal antibody production	103
19	Summary of optical density of negative control samples in ELISA examination and statistic analysis for Cut-off point measurement	124
20	Summary of ELISA final result for detection of probably important viral agents in fry rainbow trout (Oncorhynchus mykiss) mortality syndrome in Iran	127
21	Coldwater hatchery and grow-out farms in several Provinces where the samples were collected for histopathological study	135
22	Summarization of important histopathological findings seen in the collected samples	154
23	Normal hematological measurements in uninfected fry rainbow trout (<i>Oncorhynchus mykiss</i>) as control group (n=30) collected from three hatchery centers in Mazandaran province	171
24	Average Hematology parameters from rainbow trout fry obtained from hatchery centers in Mazandaran province	171
25	Analysis of leucocytes and differential count of rainbow trout fry from some hatchery centers in Mazandaran province	172
26	Comparison of hematological and biochemical indices between infected fry and uninfected fry as control group	173



27	Comparison of diagnostic methods used to detect the aetiologic agents of rainbow trout (O.mykiss) Fry Mortality Syndrome in Iran	193
28	Biochemical results of gram negative bacteria isolated from fry rainbow trout hatchery centers in some provinces of Iran	219
29	Biochemical results of gram negative bacteria isolated from adult rainbow trout rearing farms in some provinces of Iran	220
30	Time schedule of antigen injection to Rabbit for antibody production against $F.psychrophilum$	224
31	Time schedule of antigen injection to rabbit for polyclonal antibody production against IHN, IPN and VHS diseases	224
32	Specification of consumed viral antigens for polyclonal antibody production	226
33 34	Final results & Obtained (O.D) in ELISA examination for IHNV Final results & Obtained (O.D) in ELISA examination for IPNV	227 229
35	Final results & Obtained (O.D.) in FLISA evamination for VHSV	231



LIST OF FIGURES

1	Total fish production in Iran for 2004 (MT)	7
2	Fish production in Iran from 1993-2003	9
3	Aquaculture and Aquaculture based Fisheries in Iran	13
4	Contribution of Different Culture System in Coldwater production in 2004	14
5	Distribution of Coldwater Culture Sites in Iran	15
6	Increasing trend of Cold water fish production in last decade Iran (1995-2004)	17
7	Growth rate of Cold water production in forth of Iranian five years Social economical development plan	17
8	Clinical signs seen in affected fry such as darkening of the body, exophthalmia and lethargy	54
9	Collected samples consisted of milt, eggs and fry stored in EMEM media	54
10	Egg collection methods from broodstock	56
11	Eyed-egg in hatchery rainbow trout farm in Uremia city in West Azarbayejan province of Iran	57
12	An old raceway for fry production in Iranian hatchery rainbow trout farm in Shahryar city in Tehran province of Iran	58
13	Traditional raceway for fry production in Iranian hatchery rainbow trout farm in Haraz region in Mazandaran province of Iran	58



14	A new raceway for fry production in Iranian hatchery rainbow trout farm in Uremia city in West Azarbayejan province of Iran	58
15	A modern raceway and trough for fry production in Iranian hatchery rainbow trout farm in Faridan city in Isfahan province of Iran	59
16	Fish autopsy and internal organs sampling	59
17	Smear preparation for broodstock milt	60
18	Smear preparation for broodstock ova	61
19	Different kind of samples stored in bijou bottles and stored at -70°C before processing for inoculation on cell line	61
20	Disposable culture flasks for cell culturing	62
21	EPC cells monolayer grown at 25°C showing 100% confluency	63
22	FHM cells monolayer grown at 30°C showing 100% confluency	64
23	BF-2 cells monolayer grown at 25°C confluency	64
24	RTG-2 cells monolayer grown at 19°C confluency	65
25	CHSE-214 cells monolayer grown at 19°C confluency	65
26	Newly seeded cells showing pinkish color with good pH (1) in comparison to aged cells ready for passage (yellow color) (2)	66
27	CPE formed as faci of plagues in EPC cell line at 24 hour	70

