



UNIVERSITI PUTRA MALAYSIA

BIOLOGICAL ACTIVITIES AND MOLECULAR ANALYSIS OF NOVEL DITHIOCARBAZATE COMPLEX COMPOUNDS ON GLIOMA CELL LINES

SHABAN A. KH. AWIDAT

FPSK(P) 2005 1



BIOLOGICAL ACTIVITIES AND MOLECULAR ANALYSIS OF NOVEL DITHIOCARBAZATE COMPLEX COMPOUNDS ON GLIOMA CELL LINES

Ву

SHABAN A. KH. AWIDAT

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

December 2005



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

BIOLOGICAL ACTIVITIES AND MOLECULAR ANALYSIS OF NOVEL DITHIOCARBAZATE COMPLEX COMPOUNDS ON GLIOMA CELL LINES

By

SHABAN A. KH. AWIDAT

December 2005

Chairman:

Associate Professor. Rozita Rosli, PhD

Faculty:

Medicine and Health Sciences

The object of research in the exploration of new chemotherapy agents is to kill cancerous cells and not harm the healthy cells. In addition, an effective dose of these agents is essential in conducting clinical studies in the treatment of cancer. In this study, an investigation of the anticancer effects of a group of synthetic compounds on human glioma cell lines was carried out. Initially, 11 compounds were screened using cytotoxicity assays. The most active compounds were found to be derived from bis (S-methyl-β-N-(2-acetylfuran) dithiocarbazate) (SMDB) and bis (S-benzyl-β-N-(2-acetylfuran) dithiocarbazate) (SBD4) complexed with zinc, cadmium and platinum ions. The glioma cell lines, A172, U87MG and T98G and normal brain cell line HCN-2, were used in this study. The IC₅₀ values of the cell lines treated with the compounds were determined by using (3-4,5-dimethylthiazol-2-yl)-2-5-diphenyltetrazolium

bromide (MTT) assay. Tamoxifen was used as a control as it is the current drug of choice in the treatment of brain cancer.

From the cytotoxicity assays, it was found that the compounds which showed the most potential are SMDB-Cd and SMDB-Zn. The IC₅₀ values for SMDB-Cd on A172, U87MG, T98G and HCN-2 were $0.65\mu g/ml$, $0.29\mu g/ml$, $0.4\mu g/ml$, and $1.4\mu g/ml$, while that for SMDB-Zn were at $3.7\mu g/ml$, $1.76\mu g/ml$, $2.7\mu g/ml$ and $7\mu g/ml$, respectively. The IC₅₀ values for tamoxifen for the same cell lines were $6.7\mu g/ml$, $5.3\mu g/ml$, $6.3\mu g/ml$ and $6\mu g/ml$ respectively.

Several methods were employed towards understanding the mechanism of action at the molecular level for SMDB-Cd and SMDB-Zn on glioma cell lines. Tunel assay displayed the typical morphological features of apoptosis cells with condensed and fragmented nuclei at 48 hours. The percentage of apoptotic cells in all treated cells with tamoxifen, SMDB-Zn and SMDB-Cd were significantly (p<0.05) increased.

Reverse Transcription-Polymerase Chain Reaction (RT-PCR) was used in monitoring the gene expression level of two key genes, Epidermal Growth Factor Receptor (EGFR) and Mouse Double Minute 2 (MDM2). The expression of EGFR gene was suppressed in all three-cell lines. However, MDM2 gene was suppressed only in A172 and T98G. Therefore, the suppression of EGFR and MDM2 by the compounds was one of the pathways to apoptosis in the glioma cells.

In the flow cytometry analysis, the effect of SMDB-Cd and tamoxifen on the cell cycle after 3, 6, 12 and 24 hr treatment showed glioma cells A172, U87MG and T98G were arrested in G₁ phase and the SMDB-Zn arrested glioma cell lines U87MG, T98G and A172 in, G2/M, S phase and G₁ phase, respectively.

The SMDB-Cd and tamoxifen arrested the cell cycle by preventing replication (phase specific G₁) whereas SMDB-Zn was not phase specific which can arrest the cell at any point in the cell cycle.

Results, of caspase-8/9 activity assay of tamoxifen, SMDB-Cd and SMDB-Zn on glioma cells showed that caspase-8 activity was significantly induced but no significant activity for caspase-9 was observed. Therefore, the activation of caspase-8 may be the mechanism through which tamoxifen, SMDB-Cd and SMDB-Zn induces apoptosis.

The comet assay used to study the genotoxic activity of SMDB-Cd and SMDB-Zn in CHO cell line showed no genotoxic activity in both compounds. In conclusion, the two compounds have the potential to be developed as chemotherapeutic agents.

Nilai IC₅₀ untuk sel-sel tersebut yang telah dirawat dengan sebatian-sebatian di-atas dipastikan dengan menggunakan kaedah (3-4, 5-dimethylthiazol-2-yl)-2-5-diphenyltetrazolium bromide (MTT). Tamoksifen telah digunakan sebagai kawalan memandangkan ia adalah dadah pilihan semasa dalam rawatan kanser otak.

Dari kaedah sitotoksik itu sebatian-sebatian yang ditemui menunjukkan potensi adalah SMDB-Cd dan SMDB-Zn. Nilai IC₅₀ untuk SMDB-Cd pada A172, U87MG, T98G dan HCN-2 adalah 0.65µg/ml, 0.29µg/ml, 0.4µg/ml, dan 1.4µg/ml, sementara itu bagi SMDB-Zn adalah 3.7µg/ml, 1.76µg/ml, 2.7µg/ml and 7µg/ml. Nilai IC₅₀ bagi tamoksifen pula untuk sel-sel yang sama tersebut adalah 6.7µg/ml, 5.3µg/ml, 6.3µg/ml and 6µg/ml.

Beberapa kaedah telah dijalankan ke arah memahami mekanisme tindakan SMDB-Cd dan SMDB-Zn tersebut dalam sel-sel glioma pada peringkat molekul. Kaedah Tunel telah menunjukkan ciri-ciri morfologi yang tipikal bagi sel-sel apoptotik dengan nukleusnya yang menjadi padat dan pecah pada 48 jam, peratus sel-sel yang apoptotik dalam semua sel-sel yang dirawat bersama tamoxifen, SMDB-Zn dan SMDB-Cd adalah sangat bermakna (p<0.05).

"Reverse Transcription-Polymerase Chain Reaction" (RT-PCR) telah digunakan dalam pemerhatian paras ekspresi gen terhadap dua gen ini, "Epidermal Growth Factor Receptor" (EGFR) dan Mouse Double Minute 2 (MDM2). Ekspresi gen EGFR telah dihalang didalam ketiga-tiga sel yang digunakan.

ACKNOWLEDGEMENTS

My utmost appreciation goes to Associate Prof Dr. Rozita Rosli, without her continuous support, help, limitless patience, encouragement and advice I would not have been able to continue and complete this project. I wish to express my deepest thanks to Professor Karen Crouse and Professor Peter Pook for their guidance and support. I also would like to thank my fellow lab members.

I am very grateful to all my family members for their continued moral support of me in pursuing my dream. This work may not have masterialised without the prayers and love from my wife Rabiea Amor. I would like to dedicate this Ph.D, thesis to them and my committee. I certify that an Examination Committee met on 13th December 2005 to conduct the final examination of Shaban A. Kh. Awidat on his Doctor of Philosophy thesis entitled "Biological Activities and Molecular Analysis of Novel Dithiocarbazate Complex Compounds of Glioma Cell Lines" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Dato' Abdul Salam Abdullah, PhD

Professor Faculty of Medicine and Health Science Universiti Putra Malaysia (Chairman)

Fauziah Othman, PhD

Associate Professor Faculty of Medicine and Health Science Universiti Putra Malaysia (Internal Examiner)

Chong Pei Pei, PhD

Lecturer
Faculty of Medicine and Health Science
Universiti Putra Malaysia
(Internal Examiner)

Rahmah Mohamed, PhD

Professor Faculty of Science and Technology Universiti Kebangsaan Malaysia (External Examiner)

HASANAH MOHD. GHAZALI, PhD

Professor/Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:

19 JAN 2006



This thesis submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee are as follows:

Rozita Rosli, PhD Associate Professor` Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Karen Ann Crouse, PhD Professor Faculty of Science and Environmental Studies Universiti Putra Malaysia (Member)

Peter Pook Chuen Keat, PhD Professor Faculty of Medicine International Medical Universiti (Member)

> AINI IDERIS, PhD Professor/Dean School of Graduate Studies

Universiti Putra Malaysia

Date;

07 FEB 2006

DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations, which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

SHABAN A. KH. AWIDAT

Date: 16/1/06

TABLE OF CONTENTS

| | | Page |
|--|---|---|
| ABS ACK APP DEC LIST LIST | STRACT STRAK KNOWLEDGEMENT PROVAL CLARATION T OF FIGURES T OF TABLES T OF ABBREVIATIONS | ii v viii ix xi xv xviii xix |
| СНА | APTER | |
| 1 | INTRODCTION 1.1 General Background 1.2 Brain tumor 1.3 Nitrogen-sulphur Donor Ligands 1.4 Significance of the present study | 1 1 1 3 5 |
| 2 | LITERATURE REVIEW 2.1. Cancer 2.2. Brain tumor | 7 10 10 10 11 11 12 12 13 13 |
| | 2.3.2.1. A172 cell line 2.3.2.2. U87MG cell line 2.3.2.3.T98G cell line 2.3.3. Pathophysiology of glioma 2.4. Cancer as a Cell Cycle Disease 2.4.1. Deregulation of cell cycle entry 2.4.2. Cell cycle check points 2.4.3. G1–S checkpoint 2.4.4. G2 checkpoint | 13 13 14 14 17 18 22 23 25 |
| | 2.4.5. Mitotic spindle checkpoint 2.5. Current treatment strategies for brain gliomas 2.5.1. Surgery 2.5.2. Radiation therapy | 26 28 28 29 |

| | | 2.5.3. Chemotherapy | 31 |
|---|---|---|------------|
| | | 2.5.3.1. Tamoxifen | 36 |
| | | 2.5.3.2. Cisplatin | 38 |
| | | 2.5.3.3. The synthetic compounds | 39 |
| | | 2.5.3.4. Structure and chemical properties of synthetic | 39 |
| | | compounds | |
| | | 2.5.3.5. Nitrogen-sulphur donor ligand and their metal complexes | 39 |
| | | 2.5.3.6. Bioactive Properties of the metal complexes containing nitrogen- sulfur Schiff bases | 4 1 |
| | | 2.5.3.7. Cytotoxic of the metal complexes containing nitrogen-sulfur Schiff base | 45 |
| | 2.6. | Drug delivery | 47 |
| | 2.7. | Program of cell death | 49 |
| | | 4.7.1. Necrosis | 50 |
| | | 2.7.2. Apoptosis | 52 |
| | 2.8. | Caspases and their activation during apoptosis | 53 |
| | 2.9 | Genotoxicity | 55 |
| 3 | ΜΔΤΙ | ERIAL AND METHODS | |
| • | 3.1. | Materials | 57 |
| | 3.2. | Cell culture | 61 |
| | 0.2. | 3.2.1. Cell viability | 61 |
| | 3.3. | Cytotoxicity Determination | 62 |
| | • | 3.3.1. Treatment | 63 |
| | | 3.3.2. MTT assay | 63 |
| | | 3.3.3. Statistical analysis | 64 |
| | 3.4. | Reverse transcription-polymerase chain reaction (RT-PCR) | 65 |
| | | 3.4.1. Preparation of Total Ribonucleic Acid (RNA) | 65 |
| | | 3.4.2. Complementary deoxyribonucleic acid (cDNA) synthesis | 67 |
| | | 3.4.3. PCR amplification | 67 |
| | | 3.4.4. Agarose gel electrophoresis | 69 |
| | 3.5. | Apoptosis assay | 70 |
| | | 3.5.1. Tunel system | 70 |
| | | 3.5.2. Tunel assay | 70 |
| | | 3.5.3. Data analysis | 72 |
| | 3.6. | Cell cycle analysis | 73 |
| | | 3.6.1. Flow cytometry. | 73 |
| | | 3.6.2. Flow cytometry analysis using RNase A/Propidium lodide | 73 |
| | | 3.6.3. Data analysis | 74 |
| | 3.7. | Caspase activity | 75 |
| | | 3.7.1. Caspase assay | 75 |
| | | 3.7.2. Data analysis | 76 |

| | 3.8. | Single-Cell gel electrophoresis | 77 |
|-----------------------|--------|--|----------|
| | | 3.8.1. Concentration selection and cytotoxicity test | 77 |
| | | 3.8.2. Comet assay | 78 |
| | | 3.8.3. Data analysis | 79 |
| | | 3.8.3.1. Image analysis system | 79 |
| | | 3.8.3.2. Statistical analysis | 79 |
| 4 | RES | JLTS | 80 |
| | 4.1. | Growth curve | 81 |
| | 4.2. | Cytotoxicity | 82 |
| | | 4.2.1. Morphology of cells treated | 87 |
| | 4.3. | Apoptosis | 91 |
| | 4.4. | RT-PCR Gene expression | 96 |
| | 4.5. | Cell cycle analysis | 102 |
| | 4.6. | Mechanism of apoptosis | 107 |
| | | 4.6.1 Caspase activity | 107 |
| | 4.7. | Genotoxicity | 117 |
| | | 4.7.1. Comet assay | 117 |
| | 4.8 | Toxicity of SMDB-Cd, SMDB-Zn and tamoxifen on normal brain cell line | 124 |
| 5 | DISC | USSION | 126 |
| 6 | CON | CLUSION | 141 |
| REE | FRFN | CES . | 144 |
| APPENDICES | | | 160 |
| BIODATA OF THE AUTHOR | | | 180 |
| PUBLICATIONS | | | 180 |
| APF BIO | DATA (| ES OF THE AUTHOR | 16 18 |

LIST OF TABLES

| Table | Table | |
|-------|---|-----|
| 2.1 | Chemotherapeutic agents used for the treatment of brain tumors | 33 |
| 2.2 | Difference between apoptosis and necrosis | 51 |
| 4.1 | IC $_{50}$ values for synthetic compounds SMDB, SMDB-Pt, SMDB-Zn and SMDB-Cd ($\mu g/ml$) on glioma cell lines | 83 |
| 4.2 | IC_{50} values for synthetic compounds SBD4, SBD4-Pt, SBD4-Zn SBD4-Cd and tamoxifen ($\mu g/ml$) on glioma cell lines | 83 |
| 4.3 | IC ₅₀ values for cadmium, zinc and platinum on glioma cell lines | 84 |
| 4.4 | Doses of compounds tamoxifen were used to treatment glioma cell line for induce caspase | 107 |
| 4.5 | Statistical validity of comet assay results | 123 |

LIST OF FIGURES

| Figur | res | page |
|-------|---|------|
| 2.1 | Cell cycle phases | 19 |
| 2.2 | Activation of the G1–S checkpoint | 21 |
| 2.3 | Activation of the G2-phase checkpoint | 24 |
| 2.4 | Mitotic spindle check point | 27 |
| 2.5 | Structure of tamoxifen | 36 |
| 2.4 | Structure of cispltatin | 38 |
| 3.1 | Structure of S-R-β- <i>N</i> -(2-acetylfuran) dithiocarbazate (SRDB) | 59 |
| 3.2 | Structure of [bis(S-R- β -N-(2-acetylfuran)-dithiocarbazato)](SMDB-M) | 59 |
| 4.1 | Cytotoxicity effects of SMDB-Cd on glioma cell lines | 84 |
| 4.2 | Effect of SMDB-Zn on glioma cell lines | 85 |
| 4.3 | Cytotoxicity effect of tamoxifen on glioma cell lines | 86 |
| 4.4 | Comparison of the IC ₅₀ of the SMDB-Cd, SMDB-Zn and tamoxifen on glioma cell lines. | 86 |
| 4.5 | Morphology of U-87MG before and after treatment | 88 |
| 4.6 | Morphology of A172 after being treated with two compounds and tamoxifen for 72h | 89 |
| 4.7 | Morphology of T98G cell line before and after treatment for 72h | 90 |
| 4.8 | Tunnel assay for A172 cell line treated with SMDB-Cd, SMDB-Zn and tamoxifen | 92 |
| 4.9 | Tunnel assay for U87MG cell line treated with SMDB-Cd, SMDB-Zn and tamoxifen. | 93 |
| 4.10 | Tunnel assay for T98G cell line treated with SMDB-Cd, SMDB-Zn and tamoxifen | 94 |
| 4.11 | Percentage of cells undergoing apoptosis in glioma cell treated with SMDB-Cd, SMDB-Zn and tamoxifen | 95 |

| 4.12 | The effect of SMDB-Cd on MDM2 gene expression in A172 and T98G cell lines | 97 |
|------|---|-----|
| 4.13 | The effect of SMDB-Zn on MDM2 gene expression in A172 and T98G cell lines | 98 |
| 4.14 | The effect of SMDB-Cd on EGFR gene expression in A172 and T98G cell lines | 99 |
| 4.15 | The effect of SMDB-Zn and SMDB-Cd on EGFR gene expression in U87MG cell line | 100 |
| 4.16 | The effect of synthetic compound SMDB-Zn on EGFR gene expression A172 and T98G cell lines | 101 |
| 4.17 | Effect of tamoxifen on glioma cell cycle | 104 |
| 4.18 | Effect of SMDB-Zn on glioma cell cycle | 105 |
| 4.19 | Effect of SMDB-Cd on glioma cell cycle | 106 |
| 4.20 | Activity of caspase-8/9 in U87MG treated with tamoxifen | 108 |
| 4.21 | Activity of caspase-8/9 in T98G treated with tamoxifen | 109 |
| 4.22 | Activity of caspase-8/9 in A172 treated with tamoxifen | 110 |
| 4.23 | Activity of caspase-8/9 in A172 treated with SMDB-Cd | 111 |
| 4.24 | Activity of caspase-8/9 in U87MG treated with SMDB-Cd | 112 |
| 4.25 | Activity of caspase-8/9 in T98G treated with SMDB-Cd | 113 |
| 4.26 | Activity of caspase-8/9 in T98G treated with SMDB-Zn | 114 |
| 4.27 | Activity of caspase-8/9 in U87MG treated with SMDB-Zn | 115 |
| 4.28 | Activity of caspase-8/9 in A172 treated with SMDB-Zn | 116 |

| 4.29 | Comet assay for positive and negative controls | 119 |
|------|--|-----|
| 4.30 | Genotoxic activity of SMDB-Zn on CHO cell line | 120 |
| 4.31 | Genotoxic activity of SMDB-Cd on CHO cell line | 121 |
| 4.32 | Effect of increasing concentration on tail length | 122 |
| 4.33 | Comparison of the IC ₅₀ values of SMDB-Cd, SMDB-Zn and tamoxifen on A172 and HCN-2 cell lines | 125 |

LIST OF ABBREVIATIONS

ATCC American Type Culture Collection

BBB Blood Brain Barrier

CDK Cyclin-dependent kinase

cDNA Complementary deoxyribonucleic acid

CMT Carrier-Mediated Transport

CFS Cerebral Spinal Fluid

CNS Central Nervous System

DNA Deoxyribonucleic acid

DMSO Dimethyl Sulfoxide

EGFR Epidermal growth factor receptor

EDTA Ethylenediamine tetraacetic acid

EMS Ethyl methenesulfonate

MDM2 Murine double minute 2

PDGF Platelet-Derived Growth Factor

pRB Retinoblastoma protein

RMT Receptor-Mediated Transport

RNA Ribose nucleic acid

RT-PCR Reverse transcription-polymerase

chain reaction

SBD4 (S-benzyl-β-*N*-(2-acetylfuran) dithiocarbazate)

SBD4-Cd (S-benzyl-β-*N*-(2-acetylfuran) dithiocarbazate)

Cd (II)

SBD4-Pt (S-benzyl-β-*N*-(2-acetylfuran) dithiocarbazate)

Pt (II)

SBD4-Zn (S-benzyl-β-*N*-(2-acetylfuran) dithiocarbazate)

Zn (II)

SMDB S-methyl-β-*N*-(2-acetylfuran) dithiocarbazate

SMDB-Cd (S-methyl- β -N-(2-acetylfuran)-

dithiocarbazato) Cd (II).

SMDB-Pt (S-methyl- β -N-(2-acetylfuran)-

dithiocarbazato) Pt (II)

SMDB-Zn (S-methyl- β -N-(2-acetylfuran)-

dithiocarbazato) Zn (II)

CHAPTER 1

INTRODUCTION

1.1 General Background

Cancer is one of the three main causes of death among the economically active population. The two other main causes of mortality worldwide are accidents and cardiovascular diseases. Annually, there are more than 6 million deaths from a type of malignant neoplasia worldwide (Tovar-Guzman et al., 2001). The number of new cancer cases has been increasing over the past nine decades. The Malaysian Ministry of Health (1995) reported that malignant neoplasm is the major cause of death in government hospitals (45%), which is 2.8 times higher than heart diseases (16%). A total 26,089 of cancers were diagnosed among all residents in Peninsular Malaysia in the year 2002, comprising 11,815 males and 14,274 females. An estimated 10,000 cases were however, not registered. In terms of risk, 1 in 5.5 Malaysians can be expected to get cancer in his/her lifetime. Taking into accounts unregistered cases, the risk would be 1 in 4 Malaysians. The crude rate for all cancers in the year 2002 was 118.9 per 100,000 males and 148.4 per 100,000 females. (Lim et al., 2002).

1.2 Brain tumor

Primary tumors of the central nervous system account for less then 1.5% of all the cancer cases reported in the United States each year. According to the Malaysian national cancer registry in 2002, brain cancer incidence per 100,000 populations is 2.3 in males and 1.7 in female.

These infrequent tumors are the third leading cause of cancer-related deaths among men 15-54 years of age and fourth leading cause of death for women 15-34 years of age. Moreover, primary brain tumors are the most common solid tumor of childhood and the second leading cause of cancer death in children after leukemia (Kilic *et al.*, 2000).

Pathological approaches have shown that at the time of the initial diagnosis, most malignant glioma tumors have spread more then 15 mm over the area that can be identified by magnetic resonance imaging (MRI) scan (Kelly *et al.*, 1987, Greene *et al.*, 1989). Several molecular mechanisms are involved in the development of gliomas and their progression to more malignant tumors. For instance, the progression to higher grade glioma is associated with inactivation of the p53 tumor suppression gene on chromosome 17p, as well as over-expression of platelet-derived growth factor (PDGF) (Dunn *et al.*, 2000). Furthermore, progression to glioblastoma involves amplification of epidermal growth factor receptor (EGFR), murine double minute 2 (MDM2) genes and the expression of angiogenic factors such as vascular endothelial growth factor (VEGF) (Schlegel *et al.*, 1994, Biernat, *et al.*, 1997).

The prognosis for people affected by these rapidly growing tumors is currently very poor. Therapeutic modalities for malignant gliomas, including surgery, radiation and chemotherapy are of limited effectiveness and novel treatment modalities must be explored.



A direct targeting of cytotoxicity agents to cancer cells represents a modern approach to the treatment of malignant glioma and other cancers, because it should improve tumor inhibition and decrease toxicity.

1.3 Nitrogen-sulphur Donor Ligands

Coordination compounds are molecules containing coordinate bonding where ligands have excess electrons and can form coordinate linkages upon interaction with metal centers. The ligands may be neutral molecules or ions. Ligands are attached to the central atom by dative bonds or coordinate covalent bonds. In an ordinary covalent bond, each of the bonded atoms contributes one electron to the electron pair that forms the bond. The coordinating atom or ligand, called the donor, donates a pair of electrons to the central atom, called the acceptor. Proceeding from the donor atom to the acceptor atom often depicts the bond. Interaction between metal ions and ligands results in the formation of complexes. The entire aggregate of central atom and ligands is generally called a complex.

The study of nitrogen-sulfur donor ligands continues to be of great interest to researchers. Dithiocarbazate, NH₂NHCS₂ and its substituted derivatives have received considerable attention over the past few decades (Tarafder and Roy, 1988). The chemistry of such ligands warrants further study because dithiocarbazic acid and the Schiff bases derived from its S-alkyl esters form an interesting series of ligands whose properties can be greatly modified by introducing organic substituents into the ligand molecules thereby inducing different stereochemistry in the resultant metal complexes.



The synthesis of the ligands and metal complexes are of interest because of the intriguing observation that different ligands show different biological properties, although they differ only slightly in their molecular structures (Ali *et al.*, 1987; Majumder *et al.*, 1988).

Transition metal complexes of these ligands are widely studied because of their potential for therapeutic use (Tarafder and Roy 1988; Ali et al., 1996). base For example, the Schiff of 2-benzoylpyridine with Smethyldithiocarbazate (SMDTC) inhibits the growth of bacteria Escherichia coli and Staphylococcus aureus to some extent while that with Sbenzyldithiocarbazate (SBDTC) shows no effect at all on the two mentioned bacteria (Gou et al., 1990). The bioactivities of the ligands and the metal complexes such as cytotoxicity, antimicrobial and anticancer activities have not yet been widely studied. The mode of interaction of these compounds with the cancer cells and microbes are yet to be investigated.

In the present study, the anticancer effects of newly synthesized pure compounds on glioma cell lines, A172, U-87MG and T98G cell lines were investigated. Besides, screening these compounds as potential novel anticancer drugs for glioma, this study also investigates the mechanism involved, including the study on apoptosis, gene expression, cell cycle and genotoxic activity.



1.4 Significance of the present study

This project is of great importance due to the fact that some of the studied compounds are essentially new and no studies have been performed to investigate the action of these compounds toward cancer cells other than initial screening. It is anticipated that these compounds will open up many interesting avenues for further research. In other words, this project will give a new dimension to this field of brain cancer research and possibly by carrying out further studies on these new compounds, more information in this field will be gathered. This information may also enable further studies, for instance anticancer activities of related compounds. A number of compounds were tested (AR28, SB02 SPDRC, SMDCT, SBOTC, SMDB-Zn, SMDB-Pt, SBD4-Cd, SBD4-Pt, SBD4-Zn and SMDB-Cd) using cytotoxicity assay. SMDB-Cd and SMDB-Zn were found to be the most effective and hence were focused in this study. The Schiff bases and their metal complexes used in this study were synthesized and characterized by elemental analyses and various physico-chemical techniques (Chew et al., 2004).



Objectives:

The objectives of this study were:

- To study the cytotoxic effects of the pure synthetic compounds,
 (AR28, SB02 SPDRC, SMDCT, SBOTC, SMDB-Zn, SMDB-Pt, SBD4-Cd, SBD4-Pt, SBD4-Zn and SMDB-Cd) on glioma cell lines, A172,
 U-87MG and T98G.
- To study the effect of the most effective compounds on the expression of EGFR and MDM2 genes.
- To observe the morphology of the treated glioma cell lines by staining using fluorescence and light microscopy and to quantitatively determine apoptosis using tunnel assay.
- To study the effect of the pure synthetic compounds on the cell cycle by using flow cytometry.
- 5. To study the mechanism of apoptosis induction by the compounds in glioma cells by determining caspase 8 and 9 activation.
- To investigate the genotoxic effect of the compounds in CHO cells using comet assay and measuring the DNA damage using CASP software.

