

Provided by Universiti Putra Malaysia Institutional Repository

UNIVERSITI PUTRA MALAYSIA

USE OF OIL PALM EMPTY FRUIT BUNCH FOR LIGNOPHENOL PRODUCTION

SHARIFAH SOPLAH BINTI SYED ABDULLAH

FK 2008 10

USE OF OIL PALM EMPTY FRUIT BUNCH FOR LIGNOPHENOL PRODUCTION

By

SHARIFAH SOPLAH BINTI SYED ABDULLAH

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science Universiti Putra Malaysia

February 2008

Dedicated to my husband, daughter and my extended family who have brought a new level of love, patience and understanding into my life.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

USE OF OIL PALM EMPTY FRUIT BUNCH FOR LIGNOPHENOL PRODUCTION

By

SHARIFAH SOPLAH BINTI SYED ABDULLAH

February 2008

Chairman: Professor Mohd. Ali Hassan, PhD

Faculty : Engineering

Oil palm empty fruit bunch (OPEFB) is a by-product in palm oil industry and represents an abundant, inexpensive and renewable resource which has not been utilized satisfactorily. It can be categorized as lignocellulosic material due to its cellulose, hemicellulose and lignin content. Due to the reasons, this research was done to evaluate the potential of OPEFB as starting material for lignophenol production by using one step and two-step processes to compare the efficiency of the methods. The best method was further applied for the next experiment to produce lignophenols from different type of OPEFB. The separation of lignin from OPEFB is based on the phase separation reaction system at room temperature (~ 28°C) to produce lignophenol. This process composed of phenol derivative (*p*-cresol) and concentrated acid (72% sulfuric acid) where lignin is present in organic phase and carbohydrates in aqueous phase after 1 hour of stirring. From the results obtained, the two step process is the best method to

produce lignophenol due to competitive yield obtained and less chemicals used compared to one step process. The yield of lignophenol produced by one step and two step processes were 68% and 61%, respectively. Three types of OPEFB have been used, i.e. extractives free OPEFB (LP1), non-extractives free OPEFB (LP2) and OPEFB powder obtained from Sabutek Mill, Perak (LP3). The chemical composition of each OPEFB was determined prior to production of lignophenol by two step process. Basically, the lignin content in LP1 and LP2 were not so much different as compared to LP3 which has lower lignin and cellulose content. There were no difference in yield of lignophenol being produced from LP1 and LP2 which gave 61±1% respectively, whereas LP3 gave 56±1% yield based on lignin content. Lignophenol from LP1 and LP2 also appeared in white pinkish color which is comparable to the previous work. However, lignophenol from LP3 appeared in dark brown color. The sugars hydrolyzed in the phase separation system in terms of percent conversion were 89%, 99% and 97% in LP1, LP2 and LP3, respectively. The lignophenol sample from LP1, LP2 and LP3 were further analyzed and characterized by Proton Nuclear Magnetic Resonance (¹H-NMR). Fourier Transform Infrared Spectroscopy (FTIR), Permeation Gel Chromatography (GPC), Ultraviolet spectroscopy (UV) and thermomechanical analysis (TMA). However, LP3 could not be analyzed due to insolubility in the solvents used. From the results obtained, lignophenol from LP1 and LP2 showed similar results in structure and physical properties. The molecular weight of lignophenols from LP1 and LP2 were 5759 g mol⁻¹ and 5866 g mol⁻¹, respectively. There were no significant difference in the amount of attached cresol between LP1 and LP2 which gave 26±1%. In summary, it can be concluded that OPEFB has a good potential as a starting material

in lignophenol production by using two step process. Furthermore, it does not require ethanol/benzene extraction to treat the material.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

KEGUNAAN TANDAN KOSONG KELAPA SAWIT UNTUK PENGHASILAN LIGNOPHENOL

Oleh

SHARIFAH SOPLAH BINTI SYED ABDULLAH

Februari 2008

Pengerusi: Professor Mohd. Ali Hassan, PhD

Fakulti : Kejuruteraan

Tandan kosong kelapa sawit (OPEFB) adalah produk sampingan di dalam industri minyak kelapa sawit dan ia boleh didapati dengan banyak, murah dan sentiasa diperbaharui tetapi tidak digunakan dengan sebaiknya. Bahan ini dikategorikan sebagai bahan lignoselulosa kerana kandungan selulosa, hemiselulosa dan lignin di dalamnya. Berdasarkan sebab-sebab di atas, penyelidikan ini telah dilakukan untuk mengkaji potensi OPEFB sebagai bahan pemula bagi penghasilan lignophenol menggunakan dua kaedah iaitu satu langkah pertama dan dua langkah bagi membandingkan kecekapan kedua-dua kaedah. Kaedah terbaik telah diaplikasikan di dalam eksperimen yang seterusnya untuk menghasilkan lignophenol daripada pelbagai jenis OPEFB. Pengasingan lignin daripada OPEFB adalah berdasarkan sistem fasa tindakbalas pengasingan pada suhu bilik (~ 28°C) untuk menghasilkan lignophenol. Proses ini terdiri daripada phenol (*p*-cresol) dan asid pekat (72% sulfurik asid) di mana lignin akan

terpisah ke dalam fasa organik dan karbohidrat di dalam fasa akues selepas pengacauan selama satu jam. Daripada keputusan yang diperolehi, kaedah dua langkah adalah kaedah terbaik untuk menghasilkan lignophenol berdasarkan angkali hasil yang kompetitif dan penggunaan amaun bahan kimia yang sedikit berbanding kaedah satu langkah. Angkali hasil perolehan lignophenol bagi kaedah satu langkah dan dua langkah adalah 68% dan 61% masing-masing. Tiga jenis OPEFB telah digunakan iaitu OPEFB bebas ekstraktif (LP1), OPEFB tidak bebas ekstraktif (LP2) dan serbuk OPEFB daripada kilang minyak sawit Sabutek, Perak (LP3). Komposisi kimia bagi setiap sampel telah ditentukan sebelum penghasilan lignophenol menggunakan kaedah dua langkah. Secara asasnya, kandungan lignin dalam LP1 dan LP2 tidak banyak berbeza berbanding LP3 yang mempunyai kandungan lignin dan selulosa yang yeng lebih rendah. Tiada perbezaan bagi angkali hasil lignophenol yang dihasilkan daripada LP1 dan LP2 dengan nilai 61±1% masing-masing, manakala LP3 menunjukkan 56±1% angkali hasil berdasarkan kandungan lignin. Lignophenol daripada LP1 dan LP2 juga berwarna putih merah jambu sebagaimana dengan hasil daripada kajian sebelum ini. Bagamanapun, lignophenol daripada LP3 berwarna coklat gelap. Gula yang dihidrolisis dalam sistem fasa pengasingan dalam peratus penukaran adalah 89%, 99% dan 97% bagi LP1, LP2 dan LP3 masing-masing. Ketiga-tiga lignophenol sampel kemudiannya dianalisis dan ditentukan ciri-cirinya menggunakan Proton Nuclear Magnetic Resonance (¹H-NMR), Fourier Transform Infrared Spectroscopy (FTIR), Gel Permeation Chromatography (GPC), Ultraviolet spectroscopy (UV)and thermomechanical analysis (TMA). Walaubagaimanapun, LP3 tidak dapat dianalisis kerana ketidaklarutannya di dalam pelarut yang digunakan. Daripada keputusan yang

diperolehi, lignophenol daripada LP1 dan LP2 menunjukkan keputusan yang sama dari segi struktur dan ciri-ciri fizikal. Berat molekul lignophenol daripada LP1 dan LP2 adalah 5759 g mol⁻¹ dan 5866 g mol⁻¹ masing-masing. Tiada perbezaan ketara dari segi amaun cresol yang melekat pada lignin di antara LP1 dan LP2 dengan nilai 26±1%. Secara ringkasnya, dapat disimpulkan bahawa OPEFB mempunyai potensi yang baik sebagai bahan pemula dalam penghasilan lignophenol menggunakan kaedah langkah kedua. Selain itu, ia tidak memerlukan ekstraksi ethanol/benzene untuk merawatnya.

ACKNOWLEDGEMENTS

Alhamdulillah. Thanks to Almighty God for the opportunity and grace to have accomplished this study entitled, "Use of oil palm empty fruit bunch for lignophenol production".

My most gratitude to Majlis Amanah Rakyat (MARA) and Universiti Kuala Lumpur (UniKL) as a sponsorship throughout my studies. My sincere thanks and appreciation to the chairman of my supervisory committee, Prof. Dr. Mohd Ali Hassan and members of the supervisory committee, Prof. Ir. Dr. Azni Hj. Idris and Prof. Dr. Yoshihito Shirai for providing me continue supervision, guidance and sacrificing their invaluable time to the presentation of this thesis. I would like to send a special thanks to UPM, Prof. Dr. Mohd Ali Hassan and Prof. Dr. Yoshihito Shirai for giving me opportunity to do research work in Japan. I also wish to extend my gratitude to Prof. Funaoka, Mr. Shinano and other Lignophenol laboratory members in Mie University, Japan and also Kuroda and other laboratory members in Kysuhu Institute of Technology (KIT) for their guidance, useful discussions, helps and advice throughout the student exchange program.

I would like to express my sincere gratitude to my colleagues in Environmental Biotechnology group; Dr. Baharudin, Dr. Nor Aini, Dr. Suriani, Dr. Nazlin, Huzairi, Hidayah, Majd, Asma', Chong Mei Ling, Zulkhairi, Asma, Azhari, Firwance, Rafein, Alawi, Tengku Elida, Zatil, Sam, Mumtaz, Zainuri, Munir and others for their most pleasant collaboration and encouragement; Mr. Rosli Aslim, Madam Renuga, Madam Aluyah and all staff at Bioprocess Laboratory, Faculty of Biotechnology and Biomolecular Sciences for the skill and flexible technical assistance. My special thanks to Mr. Zulkifli and Mr. Saparin in Animal Science Department, Faculty of Agriculture.

My thanks are due to my dearest ones, family and friends for giving me so much joy and happiness outside the lab. My beloved husband Ahmad Kamal Abdullah, my child Nur Syamim Fatihah, my mother Esah Bt. Yusof and other family members who have given me enormous support, love and understanding in all my life throughout these years. I certify that an Examination Committee has met on 13th February 2008 to conduct the final examination of Sharifah Soplah Binti Syed Abdullah on her Master of Science thesis entitled "Use of Oil Palm Empty Fruit Bunch for Lignophenol Production" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommends that the candidate be awarded the relevant degree.

Members of the examination Committee are as follows:

Robiah Yunus, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Chaiperson)

Ling Tau Chuan, PhD

Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Norhafizah Abdullah, PhD

Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Abdul Latif Bin Ahmad, PhD

Professor School of Chemical Engineering Universiti Sains Malaysia (External Examiner)

HASANAH MOHD GHAZALI, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of Supervisory Committee were as follows:

Mohd. Ali Hassan, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Azni Hj. Idris, PhD

Professor, Ir. Faculty of Engineering Universiti Putra Malaysia (Member)

Yoshihito Shirai, PhD

Professor Graduate School of Life Science and System Engineering Kyushu Institute of Technology, Japan (Member)

AINI IDERIS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 8 May 2008

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

SHARIFAH SOPLAH SYED ABDULLAH

Date: 27 April 2008

TABLE OF CONTENTS

Page

DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL SHEETS	xi
DECLARATION FORM	xiii
LIST OF TABLES	xvii
LIST OF FIGURES	xviii
LIST OF APPENDICES	XX
LIST OF ABBREVIATIONS	xxi

CHAPTER

1	INTI	RODUCTION	1
2	LITI	ERATURE REVIEW	
	2.1	Oil Palm Industry in Malaysia	4
	2.2	Oil Palm Biomass	5
		2.2.1 Oil Palm Empty Fruit Bunch (OPEFB) in Malaysia	5
		2.2.2 Chemical Characteristics of OPEFB	6
		2.2.3 Current Applications of OPEFB	7
	2.3	Lignocellulose	10
		2.3.1 Plant Cell Wall Constituents	11
		2.3.2 Lignin	13
		2.3.3 Physical Properties of Lignin	17
		2.3.4 Chemical Properties of Lignin	17
		2.3.5 Applications of Lignin	18
		2.3.6 Extractives	19
	2.4	Lignophenol	20
		2.4.1 Phase Separation System	21
		2.4.2 Process Design For Synthesizing Functional	
		Lignin Derivatives	26
		2.4.3 Characterization Of Lignophenol	28
		2.4.4 Material Balance in the Production of	
		Lignophenol from OPEFB	32
		2.4.5 Background of Lignophenol Applications	33
3	GEN	VERAL MATERIALS AND METHODS	

3.1 Chemical reagents	
-----------------------	--

3.2	Prepar	ration of Oil Palm Empty Fruit Bunch (OPEFB)	39
	3.2.1	Extractive-free OPEFB (LP1)	39

38

	3.2.2	Non extractives free OPEFB (LP2)	
	3.2.3	Powdered, non extractives-free OPEFB	
3.3	Experi	imental Overview	
3.4	Analy	tical methods	
	3.4.1	Determination of protein content in OPEFB	
	3.4.2	Determination of extractives in OPEFB	
	3.4.7	Determination of total sugar	
SYN	THESIS	OF LIGNOPHENOL FROM OIL PALM	
EMP	TY FRU	UIT BUNCH	
4.1	Introd	uction	
4.2	Mater	ials and methods	
	4.2.1	Oil Palm Empty Fruit Bunch (OPEFB)	
	4.2.2	Synthesis of lignophenol	
	4.2.3	Sugar hydrolyzed in synthesis of lignophenol	
	4.2.4	Material balance in lignophenol production	
4.3	Result	ts and discussion	
	4.3.1	Comparison on methods of lignophenol synthesis	
		from OPEFB	
	4.3.2	Material balance in lignophenol production	
	4.3.3	Effect of treatment of OPEFB on chemical	
		compositions	
	4.3.4	Effect of treatment of OPEFB on the yield of	
		1. 1 1 11 1 1	
		lignophenol and hydrolyzed sugar	

5 CHARACTERIZATION OF LIGNOPHENOL FROM OIL PALM EMPTY FRUIT BUNCH

Introduction			72	
Materials and methods				
Proton Nuclear Magnetic Resonance (1H-NMR)				
Analysis				74
Fourier Transform Infrared Spectroscopy				
(FTIR) Analysis				76
Thermal analysis	77			
Gel Permeation Chromatography (GPC) Analysis	5	78		
Ultraviolet (UV) Spectrophotometry Analysis	78			
Results and discussion				
¹ H-NMR Analysis		80		
FTIR Analysis		83		
Thermal analysis	86			
GPC Analysis		87		
Ultraviolet (UV) Spectrophotometry Analysis	88			
Conclusion			91	

6 CONCLUSION AND RECOMMENDA	ATIONS	
Conclusion		92
Recommendations	93	

REFERENCES	95
APPENDICES	101
BIODATA OF STUDENT	115

LIST OF TABLES

Table		Page
2.1	Frequently used thermal analysis techniques	31
4.1	Comparison on the methods of lignophenol production from extractives free OPEFB	60
4.2	Chemical composition of oil palm empty fruit bunch	67
4.3	Yield, physical appearance of lignophenol and percent conversion of hydrolyzed sugar	69
5.1	Amount of introduced cresol in lignophenol and yield of lignophenol	82
5.2	The weight average (Mw) and number average (Mn) molecular weights and polydispersity (Mw/Mn) of lignophenols from OPEFB	88
5.3	Lignin concentration in lignophenol	90
5.4	Molecular weight, amount of introduced cresol and yield of lignophenol from OPEFB	91

LIST OF FIGURES

Figure		Page
2.1	Different forms of OPEFB	8
2.2	Briquettes from oil palm empty fruit bunch	9
2.3	Complex polymers of cellulose, hemicellulose, and lignin	10
2.4	Chemical association in the plant cell wall	12
2.5	General scheme of the chemical wood components	13
2.6	General structural features of lignin	14
2.7	Structural model of spruce lignin	16
2.8	Model polymer structure of lignophenol and kraft lignin	20
2.9	Phase-separation system for the functionally control of lignocellulosics	23
2.10	A schematic model of phase separation system	24
2.11	Proposed major substructure of lignocatechol and lignocresol prepared by phase-separation system using phenols and sulfuric acid	24
2.12	Process for functionalizing native lignins in the phase-separative reaction system	25
2.13	Conversion of native lignin to diphenylmethane-type polymer by phenolation at $C\alpha$ -positions of side chain.	27
2.14	Schematic diagram of material balance	32
2.15	Project scheme on lignophenol production from OPEFB	33
2.16	Process flow in lignophenol production and continuously recycling molecular-level material through the ecosystem	34
3.1	Summary of processes involved in the preparation of OPEFB	40
3.2	Overall experimental overview	42

3.3	Soxhlet extraction apparatus	45
4.1a	Experimental layout for synthesis and analysis of lignophenol (one step process)	48
4.1b	Experimental layout for synthesis and analysis of lignophenol (two step process)	49
4.2	Lignophenol synthesis from OPEFB using two step process	53
4.3	Proposed process flow diagram of lignophenol production from oil palm empty fruit bunch (the process in the dotted line is studied further by another co-worker from the group)	62
4.4	Material balance in lignophenol production (two step process)	65
4.5	Lignophenol deposited to glassware during collection	66
4.6	Lignophenol from oil palm empty fruit bunch	69
5.1	Arrangement of aluminium disc and aluminium pan in thermal analysis	77
5.2a	¹ H-NMR spectra of lignophenol from treated OPEFB ethanol/benzene extraction)	81
5.2b	¹ H-NMR spectra of lignophenol from untreated OPEFB	81
5.3	IR spectra of lignophenol from oil palm empty fruit bunch (LP1 produced from ethanol-benzene extraction OPEFB, LP2 produced from without extraction OPEFB and LP3 produced from Sabutek OPEFB).	84
5.4	IR spectra of a ground lignin sample, a lignin sulfate sample and lignophenol from defatted wood powder	85
5.5	TMA profile of LP1 and LP2	87
5.6a	Ionization difference (Ei) spectra of LP1	89
5.6b	Ionization difference (Ei) spectra of LP2	89

LIST OF APPENDICES

Appendix		Page
A	Moisture content determination	102
В	Preparation of Neutral Detergent Fiber (NDF) in OPEFB	103
С	Preparation of Acid Detergent Fiber (ADF) in OPEFB	105
D	Preparation of Acid Detergent Lignin (ADL) in OPEFB	106
E	Standard curve for determination of total sugar concentration	108
F	Preparation of ethanol-benzene solution (1:2, v/v)	109
G	Preparation of 500 mL 72% sulfuric acid	110
Н	Preparation of 3mol/C ₉ <i>p</i> -cresol/acetone solution	111
Ι	Preparation of polystyrene standard	113
J	Preparation of 2N NaOH	114

LIST OF ABBREVIATIONS

¹ H-NMR	Proton Nuclear Magnetic Resonance Spectroscopy
ADF	acid detergent fiber
ADL	acid detergent lignin
СМС	carboxymethylcellulose
DSC	differential scanning calorimetry
DTA	differential thermal analysis
EFB	empty fruit bunches
FRIM	Forest Research Institute of Malaysia
FTIR	Fourier Transform Infrared Spectroscopy
GFC	gel filtration chromatography
GPC	Gel Permeation Chromatography
H_2SO_4	Sulfuric acid
IPN	semi-interpenetrating polymer network
IR	Infrared
KBr	potassium bromide
MRSM	Maktab Rendah Sains MARA
Mn	Number average molecular weight
Mw	Molecular weight
NaOH	Natrium/sodium hydroxide
NDF	neutral detergent fiber
OPEFB	Oil palm empty fruit bunch
OPF	oil palm fronds

OPT	oil palm trunks
PNB	p-nitrobenzaldehyde
SEC	size exclusion chromatography
TG	Thermogravimetry
THF	Tetrahydrofuran
TMA	thermomechanical analysis
UKM	Universiti Kebangsaan Malaysia
UPM	Universiti Putra Malaysia
UV	Ultraviolet
v/v	volume per volume
VOCs	volatile organic compounds
w/v	weight per volume

CHAPTER 1

INTRODUCTION

Cultivation of the oil palm (*Elaeis guineensis Jacq.*) has expanded tremendously in recent years such that it is now second only to soybean as a major source of the world supply of oils and fats. Presently, Southeast Asia is the dominant region of production with Malaysia being the leading producer and exporter of palm oil (Wahid et al., 2004). It is accounting for approximately 10% of the world's oil and fat production (1998). The total area of oil palm plantations is close to 3.2 million hectares, which account for almost 50% of the land under cultivation in Malaysia (Tanaka et al., 2004).

Due to its large economic scale of the industry, enormous amount of biomass is being generated daily. About 94% of agricultural residue comes from palm oil industry. In 2004, it was estimated that 26.7 million tonnes of solid biomass produced from 381 palm oil mills in Malaysia. The solid biomass is made up of 53% oil palm empty fruit bunch (OPEFB), 32% mesocarp and 15% fiber and palm kernel shell. Therefore, disposing large tonnages of biomass generated daily is a problem for the industry (Yacob, 2005). Full exploitation of this biomass can be done by maximizing the utilization of this biomass to form products of high value which not only comply to the zero waste strategy but also generate additional profit to the palm oil industry. More

than 14.4 million tonnes of OPEFB produced annually. About 65% of OPEFB is either incinerated for bunch ash or recycled back to the plantation as mulching or used as solid fuel in the boilers to generate steam and electricity for the mills (Mat Soom et al., 2004). This is only practice by the bigger plantation such as Golden Hope, Guthrie, Sime Darby and United Plantation. For old palm oil mills, the empty fruit bunch is burned in the incinerator to produce fertilizer. However, there are still plantation company that disposed the empty fruit bunches as landfill method particularly those mill without enough plantation or estates.

OPEFB, which is considered as lignocellulosics available in large quantities and has fairly high lignin content with an average of 20.5% based on an oven dried basis (Ramli et al., 2002), appears to be a potential material for lignophenol production. In recent years, lignocellulosic materials have shown interesting features to be used as raw materials for industrial material production. Lignocellulosics are renewable, available in abundance and inexpensive, offering great potential for transformation into chemical feed stocks or fuels. However, the lignocellulosics must be separated into individual components before use as chemical feedstocks (Mikame and Funaoka, 2006a).

Therefore, a new phase-separation system was originally designed by Funaoka and coworkers as a successive total utilization of lignocellulosics leading to sustainable development. Through the phase separation system, hydrophobic lignin and hydrophilic carbohydrates are subjected to selective structural conversion individually at different