

UNIVERSITI PUTRA MALAYSIA

SHADING RESPONSES OF THE SEAGRASS

HALOPHILA OVALIS (R. BR.) HOOK. F. FROM

TELUK KEMANG, NEGRI SEMBILAN, MALAYSIA

MOHAMMAD ROZAIMI B JAMALUDIN

FS 2008 31



SHADING RESPONSES OF THE SEAGRASS HALOPHILA OVALIS (R. BR.) HOOK. F. FROM TELUK KEMANG, NEGRI SEMBILAN, MALAYSIA

MOHAMMAD ROZAIMI B JAMALUDIN

MASTER OF SCIENCE UNIVERSITI PUTRA MALAYSIA 2008



SHADING RESPONSES OF THE SEAGRASS HALOPHILA OVALIS (R. BR.) HOOK. F. FROM TELUK KEMANG, NEGRI SEMBILAN, MALAYSIA

Ву

MOHAMMAD ROZAIMI B JAMALUDIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Masters of Science

June 2008



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science

> SHADING RESPONSES OF THE SEAGRASS HALOPHILA OVALIS (R. BR.) HOOK. F. FROM TELUK KEMANG, NEGRI SEMBILAN, MALAYSIA

> > By

MOHAMMAD ROZAIMI B JAMALUDIN

June 2008

Chairman: Japar Sidik Bujang, PhD

Faculty: Science

The seagrass Halophila ovalis from Teluk Kemang coast (2 ° 30'N, 101 ° 45'E) in Port Dickson, Negeri Sembilan was studied to elucidate its responses towards artificial shading. Responses were firstly based on autotrophic productivity of H. ovalis through photosynthesis experiments to determine the effects of prior acclimation to the condition of either in the field (naturally growing) or in cultures (light reduced to 85-90% of ambient conditions). Results showed that the light compensation values in field and cultured leaves (8-13 µmol m⁻² s⁻¹) were similar while saturation point was in the range of 268-275 µmol m⁻² s⁻¹ for field leaves and increased to 290-293 μmol m⁻² s⁻¹ for cultured leaves. A one-month long artificially imposed shading was then performed to plants in the field (50%, 65%, 80% and 95%) shading relative to field light intensity) and in cultures (92% shading – Tank 1, and 96% shading - Tank 2, relative to field light intensity) and compared to unshaded plants as a control showed the following Photosynthetic rates of field *H. ovalis* at two tide levels as determined using



the Biological Oxygen Demand bottle method was up to six times higher when compared to the oxygen electrode method. Leaf chlorophyll content was significantly higher from plants under shading for both field and cultured leaves compared to control where leaves from cultures (Tank 2) showed the highest value in leaf chlorophyll content (1353.40 ± 74.00 µg chlorophyll $a g^{-1}$, p < 0.01, and 11.92 $\pm 0.59 \mu g$ chlorophyll $a cm^{-2}$, p < 0.01, by leaf fresh weight and leaf surface area respectively, and 744.30 ± 46.55 chlorophyll b g^{-1} , p < 0.01 and 6.56 ± 0.39 µg chlorophyll $b \text{ cm}^{-2}$, p < 0.01, by leaf fresh weight and leaf surface area respectively). For carbohydrates, starch and the reducing sugars of glucose, sucrose, fructose and maltose were tested for in the below-ground portions of field plants, and above-ground and belowground portions of cultured plants. Starch was not detected in both aboveground and below-ground plant portions of both field and culture studies. Glucose content was highest among the four sugars, in both field and culture plants but not significantly different compared to the control. Changes in growth rates were the most discernible where increased shading results in decreased growth rates (3.72 ± 0.51 mm apex⁻¹ day⁻¹ from control plants, to the significantly lowest recorded growth rate value of 0.746 \pm 0.205 mm apex⁻¹ day⁻¹, p < 0.01, from Tank 1 plants). Leaf morphology based on leaf length, leaf width, leaf petiole length, number of cross veins per leaf, leaf fresh weight and leaf surface area were significantly higher for leaves under shading in culture condition compared to field-shaded leaves and the control. This is substantiated by the data from Tank 2 where leaf length is 24.73 ± 0.54 mm, leaf width -9.38 ± 0.23 , leaf length-width ratio -2.80 ± 0.030 , leaf petiole length - 28.48 ± 1.03, leaf cross vein number - 14.47 ± 0.27, leaf



fresh weight -0.0179 ± 0.00134 and leaf surface area -2.011 ± 0.126) compared to the unshaded control (leaf length: 13.20 + 0.54 mm; leaf width: 6.81 ± 0.29; leaf length-width ratio: 1.93 ± 0.037; leaf petiole length: 11.20 ± 1.43; leaf cross vein number: 11.40 ± 0.35; leaf fresh weight: 0.00680 ± 0.000548; and leaf surface area: 0.796 \pm 0.0744). For field biomass values, there were no significant differences between shaded plants and the control. Comparatively, culture biomass values of Tank 1 were significantly higher for both above-ground biomass (0.0127 \pm 0.00238 g DW rhizome⁻¹, p < 0.01) and below-ground biomass (0.0282 \pm 0.00245 g DW rhizome⁻¹, p < 0.01) compared to the unshaded control (0.0107 ± 0.000914 g DW rhizome⁻¹ and 0.0192 ± 0.00109 g DW rhizome⁻¹ for above-ground and below-ground biomass respectively). All the observations and results collated showed H. ovalis tolerates extreme low light conditions as low as 96% shading (80 μmol m⁻² s⁻¹) by modifying its various physical and biochemical characteristics accordingly with its light environment. This is also evident that the plant survives continues to maintain productivity with and respect to photosynthesis and carbohydrate production even under the highest shading levels imposed in both field (95% shading) and cultures (Tank 2 - 96% shading). Furthermore, it is possible to culture *H. ovalis*, although maximum growth densities equivalent to those observed in the field were not achieved. The findings suggest that lowered light availability may not be the sole causal factor for H. ovalis loss in a particular area. Other aspects such as epiphytic fouling and available nutrients could be more important in the loss of H. ovalis vegetation, although an interaction of the factor of reduced light and these other factors should not be discounted.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master of Science

SHADING RESPONSES OF THE SEAGRASS HALOPHILA OVALIS (R. BR.) HOOK. F.FROM TELUK KEMANG, NEGRI SEMBILAN, MALAYSIA

Oleh

MOHAMMAD ROZAIMI B JAMALUDIN

Jun 2008

Pengerusi: Japar Sidik Bujang, PhD

Fakulti: Sains

Kajian terhadap Halophila ovalis dari Teluk Kemang (2 ° 30'N, 101 ° 45'E), Port Dickson, Negeri Sembilan telah dibuat untuk melihat tindakbalas rumput laut ini kepada keredupan tiruan. Tindakbalas berdasarkan produktiviti autotrofik H. ovalis melalui beberapa eksperimen fotosintesis adalah untuk mengenalpasti kesan adaptasi tumbuhan kepada di lapangan (pertumbuhan semulajadi) atau di dalam kultur (cahaya dikurangkan ke 85-90% dari cahaya semulajadi). Hasil pemerhatian keamatan mendapati kepampasan cahaya adalah tidak berbeza di antara daun dari lapangan atau daun dari kultur (8-13 µmol m⁻² s⁻¹). Manakala titik ketepuan cahaya adalah berada dalam linkungan 268-275 umol m⁻² s⁻¹ bagi daun dari lapangan dan nilai titik ketepuan cahaya bagi daun dari kultur meningkat ke linkungan 290-293 µmol m⁻² s⁻¹. Kajian selama satu bulan telah dibuat terhadap tumbuhan di lapangan (tahap 50%, 65%, 80% dan 95% daripada intensiti cahaya lapangan) dan di dalam kultur (keredupan 92% pada Tangki 1 dan 96% keredupan pada Tangki 2) berbanding dengan kawalan tanpa keredupan



cahaya. Kadar fotosintesis H. ovalis di lapangan pada aras air surut dan pasang sederhana dan juga daripada kultur berdasarkan kaedah botol 'Biological Oxygen Demand' adalah sehingga enam kali lebih tinggi dari nilai yang didapati melalui kaedah elektrod oksigen. Kandungan klorofil pada daun tumbuhan di lapangan dan kultur yang diredupkan adalah lebih tinggi berbanding dengan kawalan di mana daun dari kultur (Tangki 2) menunjukkan nilai kandungan klorofil tertinggi (1353.40 ± 74.00 µg klorofil a g^{-1} , p < 0.01 bagi berat daun segar, dan 11.92 \pm 0.59 μ g klorofil a cm⁻², p <0.01, bagi kawasan permukaan daun, serta 744.30 \pm 46.55 klorofil b g⁻¹, p < 0.01 bagi berat daun segar dan 6.56 \pm 0.39 μ g klorofil b cm⁻², p < 0.01, bagi kawasan permukaan daun). Untuk kandungan karbohidrat, kanji dan empat jenis gula – glukos, sukros, fruktos dan maltos telah diuji pada bahagian tumbuhan yang di atas permukaan substrat ("above-ground") dan di bawah substrat ("below-ground") untuk di lapangan dan kultur. Kanji tidak dikesan pada kedua-dua bahagian tumbuhan "above-ground" dan "below-ground" untuk tumbuhan di lapangan dan kultur. Kandungan glukos adalah yang tertinggi berbanding gula yang lain tetapi nilainya tidak jauh berbeza dengan tumbuhan kawalan. Analisis kadar pertumbuhan telah menunjukkan nilai perbezaan yang paling ketara di mana didapati peningkatan kadar keredupan menyebabkan penurunan kadar pertumbuhan (pertumbuhan sebanyak 3.72 + 0.51 mm apex⁻¹ hari⁻¹ bagi tumbuhan kawalan berbanding dengan tumbuhan pada Tangki 1 yang menunjukkan rekod nilai pertumbuhan yang paling rendah iaitu pada 0.746 ± 0.205 mm apex⁻¹ hari⁻¹, p < 0.01). Morfologi daun berdasarkan parameter kepanjangan daun, kelebaran daun, nisbah panjang-kelebaran daun, kepanjangan 'petiole' daun,



jumlah 'cross veins' untuk sehelai daun, berat daun segar, dan luas permukaan daun di dalam keadaan keredupan di lapangan dan kultur menunjukkan nilai kesemua parameter-parameter ini adalah lebih tinggi berbanding tumbuhan kawalan. Ini disokong oleh data dari Tangki 2 di mana panjang daun adalah 24.73 ± 0.54 mm, kelebaran daun – 9.38 ± 0.23, nisbah panjang-kelebaran daun - 2.80 ± 0.030, kepanjangan 'petiole' daun - 28.48 \pm 1.03, jumlah 'cross vein' daun - 14.47 \pm 0.27, berat daun segar - 0.0179 \pm 0.00134 dan kawasan permukaan daun - 2.011 ± 0.126 jika dibandingkan dengan tumbuhan kawalan (panjang daun: 13.20 ± 0.54 mm; kelebaran daun: 6.81 ± 0.29; nisbah panjang-kelebaran daun: 1.20 ± 1.43; kepanjangan 'petiole' daun: 11.40 ± 0.35; jumlah 'cross vein' daun: 14.47 ± 0.27; berat daun segar: 0.00680 ± 0.000548; dan kawasan permukaan daun: 0.796 ± 0.0744). Bagi nilai biojisim, tiada perbezaan ketara antara tumbuhan yang diredup di lapangan dan tumbuhan kawalan. Secara bandingan, nilai biojisim bagi tumbuhan dari Tangki 1 adalah lebih tinggi (0.0127 ± 0.00238 g DW rhizome⁻¹, p < 0.01, bagi bahagian di atas permukaan substrat dan $0.0282 \pm$ 0.00245 g DW rhizome⁻¹, p < 0.01, bagi bahagian di bawah substrat) berbanding tumbuhan kawalan (0.0107 ± 0.000914 g DW rhizome⁻¹ bagi bahagian di atas permukaan substrat dan 0.0192 ± 0.00109 g DW rhizome⁻¹ bagi bahagian di bawah substrat). Berdasarkan kesemua pemerhatian dan hasil tinjauan yang telah dijalankan, didapati H. ovalis adalah toleran kepada keadaan keamatan cahaya yang rendah di mana tumbuhan ini melalui perubahan secara fizikal dan biokimia, mengikut kedapatan cahaya di persekitarannya. Ini juga terbukti bahawa tumbuhan ini mampu hidup dan mengekalkan produktiviti walaupun pada tahap keredupan yang tinggi, iaitu



sebanyak 95% keredupan di lapangan dan sebanyak 96% keredupan di dalam kultur (Tangki 2). Adalah tidak mustahil untuk mengkulturkan *H. ovalis*, walaupun kadar maksimum bagi kepadatan pertumbuhan seperti tumbuhan di lapangan tidak tercapai. Hasil kajian ini memperlihatkan bahawa kerendahan terdapatan cahawa bukan hanya faktor yang menyebabkan kehilangan *H. ovalis* di sesuatu kawasan. Aspek-aspek lain seperti "epiphytic fouling" dan kedapatan nutrien berinteraksi dengan faktor kurangnya terdapatan cahaya perlu diambil kira juga.



ACKNOWLEDGEMENTS

It is with the utmost and foremost humility that I owe my thanks to the One Great God, Allah Almighty, for the success of this thesis and study. I am in gratitude to my mentor and teacher, Dr Japar Sidik Bujang for accepting me as his student, for guiding me throughout my tenure as a post-graduate candidate, and for being patient with me in my haste to graduate. My gratitude goes towards my co-supervisors, Dr Misri Kusnan and Dr Hishammudin Omar as well, for their guidance in my study.

I would also like to thank my parents, Jamaludin and Jumiah, my wife, Raja Yana, my brothers, Mohammad Roslan and Mohammad Rozmand, for their continued inspiration, support and belief in me to succeed in this endeavour. Also not forgetting are friends like Mahathir and Efrizal who ever so often had been there for me in so many ways in this journey. Many thanks are also due to lab-mates and the staff of Universiti Putra Malaysia Research Station for helping me in the whole study. Lastly, I would also like to thank anyone else not mentioned here who have helped complete this study in one way or another.

This research is made possible through the grant funded by the Ministry of Science, Technology and Environment Malaysia, under the 'Intensification of Research in Priority Areas' programme entitled "Seagrass taxonomy, biology and habitat characteristics: EA-001-09-02-04-0679". Some financial and travel supports from Japan Society for the Promotion of Science (JSPS) are also acknowledged.



I certify than an Examination committee has met on the 12th of June, 2008 to conduct the final examination of Mohammad Rozaimi b Jamaludin on his Master of Science thesis entitled "Shading responses of the seagrass *Halophila ovalis* (R. Br.) Hook. f. from Port Dickson, Negri Sembilan, Malaysia" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and the Universiti Pertanian Malaysia (Higher Degree) Regulations1981. The Committee recommends that the student be awarded the Master of Science.

Members of the Examination Committee were as follows:

Aziz Arshad, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Umi Kalsom Yusuf, PhD

Professor Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Abdul Rahim Ismail, PhD

Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Phang Siew Moi, PhD

Professor Faculty of Science University of Malaya Malaysia (External Examiner)

HASANAH MOHD. GHAZALI, PhD

Pofessor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of **Master of Science**. The members of the Supervisory committee were as follows:

Japar Sidik Bujang, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Chairman)

Misri Kusnan, PhD

Faculty of Science Universiti Putra Malaysia (Member)

Hishamuddin Omar, PhD

Faculty of Science Universiti Putra Malaysia (Chairman)

AINI IDERIS, PhD

Pofessor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 14 August 2008



DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

MOHAMMAD ROZAIMI B JAMALUDIN

Date: 8th July 2008



TABLE OF CONTENTS

ABSTRACT ABSTRAK ACKNOWLEDGEMENTS APPROVAL DECLARATION LIST OF TABLES LIST OF FIGURES LIST OF ABBREVIATIONS		Page iii vi x xi xiii xvii xxvii xx
CHAPTER		
1	GENERAL INTRODUCTION	1
2	LITERATURE REVIEW Seagrasses and their distributions Seagrasses from Malaysia Importance of seagrasses and the threats to its	5 5 7
	existence Light attenuation in the sea and its effects on seagrasses Physiological responses to light reduction by	10 19
	shading <i>Halophila ovalis</i>	28 33
3	PHOTOSYNTHETIC LIGHT RESPONSES OF NATURALLY GROWING AND CULTURED HALOPHILA OVALIS Abstract Introduction Materials and methods Plant material Experimental mechanism Leaf parameters Graphical analyses Statistical analyses Results Discussions	38 39 42 42 43 44 45 46 50
4	IN SITU RESPONSES OF HALOPHILA OVALIS TOWARDS SHADING Abstract Introduction Materials and Methods Study site Shading apparatus Analyses of plant material Photosynthetic rates Chlorophyll content Carbohydrate content Plant growth rates	57 59 63 63 65 67 68 69 69



	Leaf morphological	
	measurements	70
	Plant biomass	71
	Statistical analyses	71
	Results	72
	Photosynthetic rates	72
	Mean photosynthetic rates based	· -
	on leaf fresh weight	73
	Mean photosynthetic rates based	70
	on leaf surface area	76
		70
	Mean photosynthetic rates based	77
	on leaf chlorophyll content	77 70
	Chlorophyll content	79
	Mean chlorophyll <i>a</i> content	81
	Mean chlorophyll b content	82
	Mean ratio of chlorophyll a to	
	chlorophyll <i>b</i>	84
	Carbohydrate content	84
	Mean glucose content	87
	Mean sucrose content	87
	Mean fructose content	88
	Mean maltose content	88
	Plant growth rates	89
	Leaf morphological measurements	89
	Mean leaf length	91
	Mean leaf width	91
	Mean leaf length to width ratio	93
	Mean leaf petiole length	93
	Mean number of leaf cross-veins	95
	Mean leaf fresh weight	95
	Mean leaf surface area	97
	Plant biomass	97
	Above-ground biomass	99
	Below-ground biomass	99
	Above-ground to below-ground	
	biomass ratio	101
	Discussions	101
5	RESPONSES OF <i>HALOPHILA OVALIS</i> TOWARDS	
	SHADING IN CULTURES	120
	Abstract	120
	Introduction	122
	Materials and Methods	126
	Plant source	136
	Plant material and shading setup	127
	Analyses of plant material	132
	Leaf chlorophyll content and leaf	
	morphology	132
		102
	9	100
	ground biomass and ratios	133
	Carbohydrate content	133
	Statistical analyses	135



	Results	135
	Photosynthesis rates	136
	Mean photosynthetic rates	based
	on leaf fresh weight	136
	Mean photosynthetic rates	based
	on leaf surface area	138
	Mean photosynthetic rates	based
	on leaf chlorophyll content	138
	Chlorophyll content	140
	Mean chlorophyll <i>a</i> conten	t 140
	Mean chlorophyll <i>b</i> conten	t 144
	Mean ratio of chlorophyll a	
	chlorophyll <i>b</i>	146
	Carbohydrate content	148
	Mean glucose content	150
	Mean sucrose content	150
	Mean fructose content	151
	Mean maltose content	152
	Plant growth rates	152
	Leaf morphological measurement	
	Mean leaf length	153
	Mean leaf width	155
	Mean leaf length to width r	
	Mean leaf petiole length	157
	Mean number of leaf cross	
	Mean leaf fresh weight	160
	Mean leaf surface area	162
	Above-ground and below ground biomass	163
	Mean above-ground and b	
	ground biomass	163
	Mean ratio of above-groun	
	below-ground biomass	165
	Discussions	167
	D13003310113	107
6	GENERAL DISCUSSIONS	180
· ·	Basics of seagrass shading	180
	Study descriptions	181
	Various responses of <i>H. ovalis</i> to shading	
	•	.
7	CONCLUSION	216
REFERENC	SEC.	218
_	1 – EXPERIMENTAL METHODS	237
	2 – DATA VALUES	244
	3 – REGRESSION ANALYSIS OF FIELD	244
I LIIDIX	EXPERIMENTS	272
BIODATA C	OF THE AUTHOR	282
	JBLICATIONS PRODUCED	283
	· · · · · · · · · · · · · · · · · · ·	_30



LIST OF TABLES

No.	Table	Page
1	Functions and values of seagrass from the wider ecosystem perspective.	13
2	Characteristic differences between plants adapted or acclimated to sunny versus shady extremes in irradiance level.	32
3	Summary of the photosynthetic rates (R_{dark} , I_c , I_k and P_{max} values) inferred from their respective curves.	51
4	Photosynthetic irradiance values (I_c and I_k) and its corresponding plant part used from selected <i>Halophila</i> by exposure to graded light regimes.	52
5	Comparisons of the photosynthetic rates between the method used in Chapter 3 (oxygen electrode method) and that used in this chapter (BOD incubations).	104
6	Comparisons of the photosynthetic rates between the method used in Chapter 3 (oxygen electrode method) and that used in this chapter (BOD incubations).	169
7a	Summary of the photosynthetic rates of field and cultured <i>Halophila ovalis</i> as recorded through oxygen electrode incubation.	195
7b	Summary of the photosynthetic rates of field and cultured <i>Halophila ovalis</i> as recorded through biological oxygen demand (BOD) bottle incubation method.	196
8	Summary of the chlorophyll content of field and cultured Halophila ovalis.	197
9a	Summary of starch content of field and cultured Halophila ovalis.	198
9b	Summary of sugar content of field and cultured Halophila ovalis.	199
10	Summary of growth rates of field and cultured Halophila ovalis.	200
11a	Summary of the morphology (leaf length, leaf width, leaf length to width ratio and leaf petiole length) of field and cultured <i>Halophila ovalis</i> .	201
11b	Summary of the morphology (leaf cross-vein number, leaf fresh weight and leaf surface area) of field and cultured	202



Halophila ovalis.

12a	Summary of the biomass of field and cultured <i>Halophila</i> ovalis.	203
12b	Summary of above-ground to below-ground biomass ratio.	204
13	Photosynthetic rates ($\bar{x} \pm S$. E.) of <i>Halophila ovalis</i> based on leaf fresh weight (13a), leaf surface area (13b) and leaf chlorophyll content (13c).	244
14a	Values of mean photosynthesis rates at low tide level.	245
14b	Values of mean photosynthesis rates at moderate tide level.	246
15a	Values of mean chlorophyll <i>a</i> content relative to the respective parameters.	247
15b	Values of mean chlorophyll \boldsymbol{b} content relative to the respective parameters.	247
15c	Table 15c. Mean ratio value of chlorophyll a to b content.	248
16	Values of mean sugar content (glucose, sucrose, fructose and maltose).	249
17	Mean values of the growth rates of <i>Halophila ovalis</i> rhizomes from the four shading grades and control.	250
18	Mean values of the morphological measurements from the parameters of leaf length, leaf width, leaf length to width ratio, leaf petiole length, number of leaf cross-veins, leaf fresh weight and leaf surface area.	251
19a	Mean values of above-ground biomass.	253
19b	Mean values of below-ground biomass	253
19c	Mean value of the ratio of above-ground to below-ground biomass.	253
20	Photosynthesis rates from the parameters of leaf fresh weight (20a), leaf surface area (20b) and leaf chlorophyll amount (20c).	254
21	Chlorophyll a content (21a-i, ii), chlorophyll b content (21b-i, ii) and chlorophyll a to b ratios (21c) from culture shadings.	257
22	Values of mean glucose (Table 22a), sucrose (Table 22b), fructose (Table 22c) and maltose (Table 22d) content	262



23	Mean values of the growth rates of <i>Halophila ovalis</i> from cultures.	266
24	Mean morphological measurements from the parameters of leaf length, leaf width (Table 24a), leaf length to width ratio, leaf petiole length (Table 24b), number of leaf cross-veins, leaf fresh weight (Table 24c) and leaf surface area (Table 24d).	267
25	Mean values of above-ground and below-ground plant biomass (25a) and the mean ratio value of above-ground and below-ground biomass (25b).	271



LIST OF FIGURES

No.	Figure	Page
1	The major and important seagrass areas, associated habitats, utilization by coastal communities and other users in Peninsular Malaysia (A) and East Malaysia – Sabah (A) and Sarawak (C).	11
2	Depth limits compiled for 31 marine angiosperm species.	22
3	Halophila ovalis population in Teluk Kemang.	23
4	Ulva sp. canopy upon Halophila ovalis at Tanjung Chek Jawa, Singapore.	25
5	Halophila ovalis from Teluk Kemang covered with epiphytes.	25
6	Theoretical progression of a photosynthesis-irradiance (P-I) curve.	29
7	Comparisons of photosynthetic parameters of some studied seagrasses.	31
8	Botanical classification of Halophila ovalis.	35
9	Key to the species from section Halophila.	35
10	World geographical distribution of Halophila ovalis.	36
11a	Photosynthetic rates ($\bar{x} \pm S$. E.) based on leaf fresh weight by the oxygen electrode method.	47
11b	Photosynthetic rates ($\bar{x} \pm S$. E.) based on leaf surface area by the oxygen electrode method.	47
11c	Photosynthetic rates ($\bar{x} \pm S$. E.) based on leaf chlorophyll content by the oxygen electrode method	48
12	Location of the study site at Teluk Kemang (2 $^{\circ}$ 30 $^{\prime}$ N, 101 $^{\circ}$ 45 $^{\prime}$ E).	64
13	Shading frames staked upon the seabed in Teluk Kemang.	66
14	Some of the shading frames used for the field shading experiments at Teluk Kemang.	66
15a	Photosynthetic rate ($\bar{x} \pm S$. E.) of leaves from field by leaf	75
15b	fresh weight (FW). Photosynthetic rate ($\bar{x} \pm S$. E.) of leaves from field by leaf surface area (Area).	78



150	chlorophyll content (Chl).	80
16a	Mean of chlorophylls a and b ($\bar{x} \pm S$. E.) of leaves from field samples by leaf fresh weight (FW).	83
16b	Mean of chlorophylls a and b ($\bar{x} \pm S$. E.) of leaves from field samples by leaf surface area (Area).	83
16c	Mean of ratio ($\bar{x} \pm S$. E.) of chlorophyll a to b of leaves from field samples.	85
17	Mean content of reducing sugars ($\bar{x} \pm S$. E.) in below-ground plant portions per gram of dried field samples.	86
18	Mean values of the growth rate ($\bar{x} \pm S$. E.) of <i>Halophila ovalis</i> rhizomes from the four shading grades and control.	90
19a	Mean length of leaves ($\bar{x} \pm S$. E.) from field samples.	91
19b	Mean width of leaves ($\bar{x} \pm S$. E.) from field samples.	91
19c	Mean ratio ($\bar{x} \pm S$. E.) of leaf length to width of field samples.	94
19d	Mean petiole length of leaves ($\bar{x} \pm S$. E.) from field samples.	94
19e	Mean number of cross-veins ($\bar{x} \pm S$. E.) of leaves from field samples.	96
19f	Mean fresh weight of leaves ($\bar{x} \pm S$. E.) from field samples.	96
19g	Mean surface area of leaves ($\bar{x} \pm S$. E.) from field samples.	98
20a	Above-ground and below ground biomass ($\bar{x} \pm S$. E.) of field samples	100
20b	Mean of ratio ($\bar{x} \pm S$. E.) of above-ground biomass to below ground biomass of field samples	100
21a	Theoretical diagram on the energy level flow during photosynthesis.	108
21b	Components of the antenna proteins involved in	108
22a	photosynthesis. Evidence of grazing on <i>H. ovalis</i> leaves by <i>Clithon</i> sp. (arrow).	129
22b	Clithon sp. that grazes on H. ovalis leaves.	129
23a	Sprorbid polychaete fouling on <i>H. ovalis</i> leaves	130



236	observed under a light microscope.	130
24	Tank placements layout of the culture shading study.	131
25	Node positions of the leaves taken for analysis.	134
26a	Photosynthetic rate ($\bar{x} \pm S$. E.) of field leaves by leaf fresh weight.	137
26b	Photosynthetic rate ($\bar{x} \pm S$. E.) of field leaves by leaf surface area.	139
26c	Photosynthetic rate ($\bar{x} \pm S$. E.) of field leaves by leaf chlorophyll content.	141
27a	Mean content of chlorophylls a and b ($\bar{x} \pm S$. E.) from culture samples by leaf fresh weight.	143
27b	Mean content of chlorophylls a and b ($\bar{x} \pm S$. E.) from culture samples by leaf surface area.	143
27c	Mean of ratio of chlorophyll a to b ($\bar{x} \pm S$. E.) of leaves from culture.	147
28	Mean content of reducing sugars ($\bar{x} \pm S$. E.) in above and below-ground plant portions per gram of dried leaf cultures.	149
29	Mean growth rate ($\bar{x} \pm S$. E.) of <i>Halophila ovalis</i> from cultures	154
30a	Mean length of leaves ($\bar{x} \pm S$. E.) from cultures.	156
30b	Mean width of leaves ($\bar{x} \pm S$. E.) from cultures.	156
30c	Mean ratio of leaf length to width ($\bar{x} \pm S$. E.) from cultures.	158
30d	Mean petiole length of leaves ($\bar{x} \pm S$. E.) from cultures.	158
30e	Mean number of cross-veins of leaves ($\bar{x} \pm S$. E.) from cultures.	161
30f	Mean fresh weight of leaves ($\bar{x} \pm S$. E.) from cultures.	161
30g	Mean surface area of leaves ($\bar{x} \pm S$. E.) from cultures.	164
31a	Mean values of above-ground and below-ground plant biomass ($\bar{x} \pm S$. E.) from cultures.	166
31b	Mean ratio ($\bar{x} \pm S$. E.) of above-ground and below-ground plant biomass from cultures.	166



32	investigations done in Chapters 4 and 5.	184
33	Sucrose and starch biosynthesis and catabolism in plant cells.	189
34	Illustration of the experimental setup used for the photosynthesis analysis by the oxygen electrode method.	238
35	An example of a single sprig of <i>Halophila ovalis</i> used for analyses.	242
36	Curve-fit regression analysis of values obtained in Chapter 4 from field experiments of photosynthesis by leaf fresh weight at low water level (Figure 36a); photosynthesis by leaf surface area at low water level (Figure 36b); and photosynthesis by leaf chlorophyll content at low water level (Figure 36c).	272
37	Curve-fit regression analysis of values obtained in Chapter 4 from field experiments of photosynthesis by leaf fresh weight at moderate water level (Figure 37a); photosynthesis by leaf surface area at moderate water level (Figure 36b); and photosynthesis by leaf chlorophyll content at moderate water level (Figure 37c).	273
38	Curve-fit regression analysis of values obtained in Chapter 4 from field experiments of chlorophyll a content by leaf fresh weight (Figure 38a-i); field experiments of chlorophyll b content by leaf fresh weight (Figure 38a-ii); chlorophyll a content by leaf surface area (Figure 38b-i); field experiments of chlorophyll b content by leaf surface area (Figure 38b-ii); and chlorophyll a to b ratio (Figure 38c).	274
39	Curve-fit regression analysis of values obtained in Chapter 4 from field experiments of glucose content (Figure 39a); sucrose content (Figure 39b); fructose content (Figure 39c) and maltose content (Figure 39d).	276
10	Curve-fit regression analysis of values obtained in Chapter 4 from field experiments of leaf length (Figure 40a); leaf width (Figure 40b); leaf length to width ratio (Figure 40c); leaf petiole length (Figure 40d); leaf cross-vein number (Figure 40e); leaf fresh weight (Figure 40f) and leaf surface area (Figure 40g).	278
1 1	Curve-fit regression analysis of values obtained in Chapter 4 from field experiments of above-ground biomass (Figure 41a); below-ground biomass (Figure 41b) and above-ground to below ground biomass ratio (Figure 41c).	281



LIST OF ABBREVIATIONS

α Photosynthetic efficiency

AG Above-ground
Area Leaf Surface Area
BG Below-ground

BOD Biological Oxygen Demand

Chl Chlorophyll
DW Leaf Dry Weight
FW Leaf Fresh Weight

HPLC High Performance Liquid Chromatography

 I_c Light compensation point I_k Light saturation point

IUCN The World Conservation Union

KEGG Kyoto Encyclopedia of Genes and Genomes

LHC Light-Harvesting Complex LHC II Light Harvesting Complex II

NCSS Number Cruncher Statistical System PAR Photosynthetically Active Radiation

P-I Photosynthesis-Irradiance

P_{max} Maximal photosynthetic capacity

PS I Photosystem complex I PS II Photosystem complex II

R_{dark} Dark respiration

