

1 **Morphological and molecular characterisation of *Aporcelaimellus***
2 ***nigeriensis* sp. n. (Dorylaimida, Aporcelaimidae), a remarkable**
3 **dorylaim from Nigeria**

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16 **Summary** – A new species of the genus *Aporcelaimellus*, collected in a watermelon field
17 [in](#) Nigeria is described, including its morphological and molecular (D2-D3 28S-rDNA,
18 18r-DNA) characterisation. *Aporcelaimellus nigeriensis* sp. n. is distinguishable by its
19 2.76-3.55 [mm length](#), very coarse ventral body pores, lip region offset by deep
20 constriction and 24-27 μm broad, odontostyle 30-36 μm long at its dorsal side, neck 648-
21 779 μm long, pharyngeal expansion occupying 54-60% of total neck length, uterus 300-
22 473 μm or 2.1-3.2 body diameters long and tripartite, $V = 49-54$, tail short and convex
23 conoid (27-41 μm , $c = 72-115$, $c' = 0.5-0.7$), spicules 108-137 μm long, and 9-10 [\$\mu\text{m}\$](#)
24 spaced ventromedian supplements with hiatus. LSU analysis revealed a close
25 [similarity between](#) *A. nigeriensis* sp. n. [and](#) other *Aporcelaimellus* species,
26 [which questions](#), once more, the monophyly of Aporcelaimidae. SSU
27 phylogenetic tree [analysis was not able to](#) resolve the [relationship](#)
28 [between the new species and](#) other [closely related](#) species.

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Keywords – Description, LSU, morphology, morphometry, phylogeny, SSU.

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35 With more than 3300 described species, dorylaims are probably the most diverse
36 nematode Order, despite their distribution being restricted to continental (soil and
37 freshwater) habitats. With the exception of species identified from
38 South African, which have been extensively studied for over
39 half a century, the dorylaimid fauna of Africa remains poorly explored. However, plant
40 parasitic and virus vector species of the family
41 Longidoridae have received more
42 attention due to their applied aspects. Nigerian dorylaimid fauna are no exception
43 of this general panorama as no monographic contribution was devoted to characterise it.
44 Nevertheless, two dozen species from 20 genera are currently recorded from
45 Nigeria by means of 15 reports (Table 1), proving its
46 tentatively high nematode diversity.

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47 During a nematological survey conducted to characterise the nematode community
48 associated with watermelon fields in Nigeria, interesting specimens belonging
49 to the genus *Aporcelaimellus* Heyns, 1965 were collected. A detailed study
50 revealed that such specimens represent a non-described species that is herein
51 presented.

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54 **Material and methods**

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56 NEMATODE EXTRACTION AND PROCESSING

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58 Rhizosphere soil samples were collected from watermelon fields during a survey
59 conducted in the south-western agricultural areas of Nigeria during 2016. Nematodes
60 were extracted from soil using a modified pie-pan method (Coyne *et al.*, 2007), fixed in
61 a hot 4% formaldehyde solution (Nico *et al.*, 2002), and subsequently mounted in
62 anhydrous glycerine as permanent slides (De Grisse, 1963). Specimens for molecular
63 analysis were stored in DESS solution.

64 Nematodes were observed, measured and photographed using a Nikon Eclipse 80i
65 microscope equipped with DIC optics, a drawing tube (*camera lucida*) and a Nikon DS
66 digital camera. Morphometrics include Demanian indices and other usual measurements

67 and ratios. Position of pharyngeal gland nuclei presented according with Loof and
68 Coomans (1970). Spicules were described following Peña-Santiago *et al.* (2014).
69 Microphotographs were edited using Adobe® Photoshop® CS.

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71 MOLECULAR IDENTIFICATION

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73 Fixed specimens in DESS solution were rinsed using double distilled water
74 (ddH₂O), [after which](#) one specimen was then transferred into 1.5 ml Eppendorf tube
75 containing 20 µl ddH₂O for molecular characterisation. [The deoxyribonucleic acid](#)
76 [\(DNA\) of the specimen](#) was extracted using the chelex-100 [protocol](#) as described by
77 Rashidifard *et al.* (2019). [And](#) amplified using a Vacutec
78 thermocycler (www.vacutec.co.za). The amplification reaction was made up by adding
79 12.5 µl ready to use master mix (Promega Corporation), 1 µl forward primer (10 µM), 1
80 µl reverse primer (10 µM), 5 µl DNA and 5.5 µl ddH₂O. The following primers were
81 used for amplification of partial [large subunits \(LSU\)](#) (D2-D3) rDNA: D2A (5'-
82 ACAAGTACCGTGAGGGAAAGTTG-3'), D3B (5'-
83 TCGGAAGGAACCAGCTACTA-3') (Subbotin *et al.*, 2006) and partial SSU: SSU F04
84 (GCTTGTCTCAAAGATTAAGCC), [and](#) [small subunits \(SSU\)](#) R26
85 (CATTCTTGGCAAATGCTTTTCG) (Blaxter *et al.*, 1998). [Polymerase chain reaction](#)
86 [\(PCR\)](#) amplification was carried out using the following steps: 3 min initial denaturation
87 at 94 °C, 35 cycles of denaturation for 45 s at 94 °C, annealing temperature (54 °C and
88 56 °C for SSU and LSU, respectively) for 45 s and finally a 6 min extension cycle at 72
89 °C followed by a holding temperature of 4 °C.

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91 PHYLOGENETIC ANALYSES

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93 The newly obtained sequences for LSU and SSU were compared using the
94 BLASTN search with those of other species available in GenBank. Taxa selection for
95 reconstruction of the LSU tree, was [conducted](#) based on Álvarez-Ortega *et al.*
96 (2013a) and for SSU the currently available SSU rDNA sequences of the genera within
97 the family Aporcelaimidae were obtained from GenBank. The sequences of selected taxa
98 as well as outgroups were aligned using the MUSCLE alignment tool (Edgar, 2004)

99 implemented in Geneious version 7.1 (Kearse *et al.*, 2012). The jModelTest 2.1.10
100 (Darriba *et al.*, 2012) program^{me} was used to identify the most appropriate nucleotide
101 substitution model. The identified model was General Time Reversible with proportion
102 of invariable sites and a Gamma distribution (GTR+I+G) for LSU and SSU genes.
103 Bayesian inference (BI) was performed using MrBayes 3.2.2 (Huelsenbeck **and** Ronquist,
104 2001) implemented in Geneious 7.1, running the chain for 3×10^6 generations. Markov
105 Chains Monte Carlo (MCMC) algorithm was used to estimate the posterior probabilities
106 of the Bayesian phylogenetic trees (Larget **&** Simon, 1999) using the 50% majority rule.
107 The Markov chain was sampled every 100 generations and a 25% burn-in samples was
108 implemented.

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111 Results

112 *Aporcelainellus nigeriensis*¹ sp. n.

113 (Figs 1-3)

114 MATERIAL EXAMINED

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116 Five females and four males from two locations.

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118 MEASUREMENTS

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120 See Table 2.

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122 DESCRIPTION

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124 *Adult*

125 Moderately slender ($a = 20-28$) nematodes of medium to large size, 2.76-3.55 mm
126 long. Body cylindrical, tapering towards both ends but substantially towards the anterior
127 region as the tail is short. Upon fixation, habitus regularly curved **ventrad**, C-shaped in
128 females, J-shaped in males. Cuticle three-layered, very thick throughout the entire body,
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¹ The specific epithet refers to the geographical origin of the new species in Nigeria.

130 5.5-7.5 μm in anterior region, 8.5-14.5 μm at mid-body and 11-15 μm on tail, consisting
131 of a very thin outer layer with nearly smooth surface, and two much thicker intermediate
132 and inner layers; both intermediate and inner layers are equally thick at caudal and mid-
133 body regions but the former is distinctly thicker than the latter in the anterior region;
134 the intermediate layer bears very conspicuous radial striation that it is not perceptible in
135 the inner layer. Lateral chord very narrow, 5.5-11.5 μm wide or 4-8% of mid-body diam.
136 Ventral body pores coarse, very conspicuous, 56-64 μm in total: 19-20 μm at neck region,
137 13-20 μm from neck base to vulva, and 22-25 μm from vulva to anus (n = 5 females);
138 dorsal pores reduced to cervical region, not so coarse as the ventral ones; lateral pores
139 small. Lip region offset by a distinct constriction, 2.7-3.0 times wider than high, and 17-
140 24% of body diam. at neck base; lips mostly separate, with moderately protruding
141 papillae. Amphid fovea stirrup-shaped, its opening 11-12.5 μm or nearly one-half (42-
142 52%) of lip region diam. Odontostyle strong, 3.9-4.6 times longer than wide, hardly
143 longer (1.1-1.3 times) than lip region diam., and 0.87-1.18% of total body length, with
144 aperture 20-22 μm long or two-thirds (64-71%) of its total length. Guiding ring simple
145 but distinct, plicate. Odontophore linear, rod-like, 1.8-2.2 times longer than odontostyle.
146 Pharynx entirely muscular, enlarging very gradually, with its basal expansion 5.4-6.9
147 times as long as wide, 2.7-3.7 times the body diam. at neck base, and occupying 54-60%
148 of total neck length; pharyngeal gland nuclei located as follows (n = 3): DO = 48-52, DN
149 = 51-57, S₁N₁ = 65-70, S₁N₂ = 72-75, S₂N = 84-85. Cardia conical, 20-47 x 14-23 μm ,
150 surrounded by intestinal tissue. Tail short, convex conoid, its ventral side visibly
151 straighter than the dorsal side; caudal pores two pairs, subdorsal, at the middle of tail.

152

153 *Female*

154 Genital system didelphic-amphidelphic, with well-developed genital branches, the
155 anterior 440-678 μm and the posterior 498-726 μm long. Ovaries comparatively large,
156 reaching and surpassing the oviduct-uterus junction, the anterior 162-310 μm long, the
157 posterior 185-335 μm long. Oviduct 112-203 μm or 0.8-1.4 body diameters long,
158 consisting of a slender distal portion made of prismatic cells and a distinct *pars dilatata*
159 with perceptible lumen. Oviduct and uterus separate by a distinct sphincter. Uterus 300-
160 473 μm or 2.1-3.2 body diameters long, and tripartite as it consists of a narrower
161 intermediate section between the proximal and distal parts, these both more dilate. Uterine

162 egg ovoid, 119-135 x 64-72 μm (n = 4). Vagina extending inwards 56-60 μm , occupying
163 two-fifths (38-43%) of body diam.: *pars proximalis* 38-44 x 20-25 μm , with somewhat
164 sigmoid walls and surrounded by a relatively weak musculature, *pars refringens*
165 consisting of two drop-shaped sclerotized pieces measuring 9-11 x 6.5-7.5 μm and a
166 combined width of 13.5-16 μm , and *pars distalis* very short, 5-6 μm long. Vulva a
167 transverse slit. Prerectum 1.7-2.4, rectum 1.1-1.4 times the anal body diam. long.

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168

169 *Male*

170 Genital system diorchic, with opposed testes. Prerectum 2.4-3.6, cloaca 1.4-1.5
171 times the body diam. at level of cloacal aperture. In addition to the ad-cloacal pair,
172 situated at 16-19 μm from the cloacal aperture, there is a series of 9-10 widely but
173 irregularly spaced, 23-51 μm apart, ventromedian supplements, the most posterior of
174 which is located at 70-85 μm from the ad-cloacal pair, a short distance behind the level
175 of anterior end of spicules. Spicule dorylaimid, 5.9-7.3 times as long as wide and 1.9-2.4
176 times longer than body diam. at level of cloacal aperture: head 11-13 x 8-9.5 μm ,
177 occupying 9-10% of total spicule length, with its dorsal side strongly curved at its anterior
178 tip, whereas the ventral side is very short and straight; median piece occupying 31-39%
179 of spicule maximum width; posterior tip 7.5-9.5 μm broad; ventral hump and hollow
180 weak but perceptible, the former situated at 31-41% of total spicule length; curvature 110-
181 126°. Lateral guiding piece 25-30 μm long, 7.1-9.3 times longer than wide, with furcate
182 tip.

183

184 MOLECULAR CHARACTERISATION

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186 One 769 bp long D2-D3 28S rDNA [sequence](#) and one 776 bp long partial 18S rDNA
187 sequence (GenBank access codes MN685820 and MN505320, respectively) were
188 obtained. Their analysis allowed the elucidation of evolutionary relationships of the new
189 species. The results are presented in Figs. 4 and 5, and discussed below.

190

191 DIAGNOSIS AND RELATIONSHIPS

192

193 The new species is characterised by its 2.76-3.55 [mm](#) long body, very coarse ventral
194 body pores, lip region offset by deep constriction and 24-27 μm broad, odontostyle 30-
195 36 μm long at its dorsal side and 28-32 μm at its ventral side, neck 648-779 μm long,
196 pharyngeal expansion 347-424 μm long or 54-60% of total neck length, female genital
197 system didelphic-amphidelphic, uterus 300-473 μm or 2.1-3.2 body diameters long and
198 tripartite, $V = 49-54$, tail short and convex conoid (27-41 μm , $c = 72-115$, $c' = 0.5-0.7$),
199 spicules 108-137 μm long, and [9-10](#) spaced ventromedian supplements with hiatus.

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200 In its general morphology and particularly in having coarse ventral body pores, the
201 new species resembles *A. porosus* Álvarez-Ortega, Ahmad & Peña-Santiago, 2011,
202 known to occur in West Africa too, but it significantly differs from this in its larger
203 general size (body 2.76-3.55 vs 2.51-2.81 mm long, neck 648-779 vs 589-626 μm long),
204 thicker cuticle (for instance, 11-15 vs 6.5-10.5 μm on caudal region) with different
205 layering (intermediate and inner layers with equal thickness vs inner layer much thicker
206 than the others), wider lip region (24-27 vs 19.5-21.5 μm), longer odontostyle (30-36 vs
207 26-30 μm), and much longer spicules (108-137 vs 81-87 μm). In having [a](#) comparatively
208 strong odontostyle, broad lip region and tripartite uterus, it resembles *A. castaneanus*
209 Álvarez-Ortega, Abolafia, Liébanas & Peña-Santiago, 2012, from Spain but it can be
210 easily distinguished from this in its larger general size (body 2.76-3.55 vs 2.18-2.83 mm
211 long), presence (vs absence) of coarse ventral body pores, much thicker cuticle (for
212 instance, 11-15 vs 5-7 μm on caudal region) with different layering (intermediate and
213 inner layers with equal thickness vs inner layer much thicker than the others), broader lip
214 region (24-27 vs 21-22 μm), much longer odontostyle (30-36 vs 22-24 μm), shorter caudal
215 region (27-41 vs 44-52 μm , $c = 72-115$ vs 48-60, $c' = 0.5-0.7$ vs 0.8-1.1), longer spicules
216 (108-137 vs 94-103 μm), and less ventromedian supplements (9-10 vs 14-15).

217 Evolutionary relationships of *A. nigeriensis* sp. n. as obtained using Bayesian
218 Inference based on partial LSU and SSU sequences are shown in two trees (Figs. 4 & 5).
219 The most relevant results of LSU analysis (Fig. 4) is the inclusion of the new species
220 sequence in a very highly supported clade with other *Aporcelaimellus* sequences; *A.*
221 *obtusicaudatus* (Bastian, 1865) Altherr, 1968 being its closest relative.
222 [Results from the current study](#) agree with [those of](#) previous studies (Álvarez-Ortega *et*
223 *al.*, 2013a,b) concerning the non-monophyly of the family Aporcelaimidae Heyns, 1965
224 as sequences of the genera *Aporcella* Andrassy, 2002, *Metaporcelaimus* Lordello, 1965

225 and *Sectonema* Thorne, 1930 form part of respective separate highly supported clades.
226 Unfortunately, SSU analysis (Fig. 5) did not provide any satisfactory new insight into the
227 phylogeny of the new species or the aporcelaims since the corresponding branching did
228 not reach good support.

229

230 TYPE LOCALITY AND HABITAT

231

232 Southwest Nigeria, Ibarapa, a peri-urban locality (coordinates: 7°30'55.87" N
233 3°27'51.27" E) with a history of maize and vegetable production, where the species was
234 collected from a watermelon field on sandy-loam soil (sand = 68%, silt = 12%, clay =
235 20%, organic matter = 14.17 %, pH = 6.05).

236

237 OTHER LOCALITY AND HABITAT

238

239 Southwest Nigeria, Modakeke, an agrarian locality (coordinates: 7°24'1.99" N
240 4°15'41.55" E) with a history of leafy vegetable production, where the species was
241 collected from a watermelon field on sandy-loam soil (sand = 76%, silt = 10%, clay =
242 14%, OM = 4.73%).

243

244 TYPE MATERIAL

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246 Female holotype, two female and one male paratypes deposited with nematode
247 collection of the University of Jaén, Spain.

248

249 REMARKS

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251 One of the most remarkable traits of the new species herein
252 described is the presence of a row of coarse ventral body pores throughout the entire body,
253 a very atypical and infrequent feature within the genus *Aporcelaimellus* and
254 representatives of the family Aporcelaimidae. Nevertheless, this rare trait also occurs in
255 *A. porosus*, which, interestingly, was also described from Western Africa.
256 Unfortunately, molecular data of this second species are not available, therefore it is not

257 possible to confirm [the](#) evolutionary relationship between them [but which is proposed to](#)
258 [be conducted in the](#) future.

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265

266 **References**

267 Álvarez-Ortega, S., Abolafia, J., Liébanas, G. & Peña-Santiago, R. (2012). Studies on the
268 genus *Aporcelaimellus* Heyns, 1965 (Nematoda, Dorylaimida, Aporcelaimidae).

269 Four new species with complex uterus from Southeastern Iberian Peninsula. *Zootaxa*
270 3551, 1-24.

271 Álvarez-Ortega, S., Ahmad, W. & Peña-Santiago, R. (2011). Studies on the genus
272 *Aporcelaimellus* Heyns, 1965 (Dorylaimida: Aporcelaimidae). An interesting new
273 species from Western Africa. *Nematology* 13, 911-917.

274 Álvarez-Ortega, S., Subbotin, S.A. & Peña-Santiago, R. (2013a). Morphological and
275 molecular study of Californian species of the genus *Aporcelaimellus* Heyns, 1965
276 (Dorylaimida: Aporcelaimidae). *Nematology* 15, 431-449. DOI: 0.1163/15685411-
277 00002691

278 Álvarez-Ortega, S., Subbotin, S.A. & Peña-Santiago, R. (2013b). Morphological and
279 molecular study of Californian species of the genus *Metaporcelaimus* Lordello, 1965
280 (Dorylaimida: Aporcelaimidae). *Nematology* 15, 431-449. DOI: 0.1163/15685411-
281 00002691

282 Andrassy, I. (2002). New genera and species of nematodes from southern Chile. *Opuscula*
283 *Zoologica Budapestinensis* 34, 5-22.

284 Andrassy, I. (2009). *Free-living nematodes of Hungary. III*. Pedozoologica Hungarica n
285 5. Hungarian Natural History Museum. Budapest, Hungary. 608 pp.

286 Blaxter, M.L., De Ley, P., Garey, J.R., Liu, L.X., Scheldeman, P., Vierstraete, A.,
287 Vanfleteren, J.R., Mackey, L.Y., Dorris, M., Frisse, L.M., Vida, J.T. & Thomas,

288 W.K. (1998). A molecular evolutionary framework for the phylum Nematoda.
289 *Nature* 392, 71-75.

290 Carbonell, E. & Coomans, A. (1986). Observations on *Opisthodorylaimus*, with
291 descriptions of three new species (Nematoda: Dorylaimoidea). *Nematologica*
292 31(1985), 379-409.

293 Carbonell, E. & Coomans, A. (1987). The genus *Thornenema* Andr ssy, 1959
294 (Nematoda: Thornenematidae). *Nematologica* 32(1986), 125-179.

295 Coyne, D.L., Nicol, J.M. & Claudius-Cole, B. (2007). *Practical plant nematology: a field*
296 *and laboratory guide*. SP-IPM Secretariat, Cotonou, Benin. International Institute of
297 Tropical Agriculture (IITA).

298 Darriba, D., Taboada, G.L., Doallo, R. & Posada, D. (2012). jModelTest 2: more models,
299 new heuristics and parallel computing. *Nature Methods* 9, 772.

300 Das, V.M., Khan, E. & Loof, P.A.A. (1969). Revision of the genus *Discolaimoides*
301 Heyns, 1963 with description of two new species reminiscent of this genus.
302 *Nematologica* 15, 473-491.

303 De Grisse, A. (1963). A counting dish for nematodes excluding border effect.
304 *Nematologica* 9, 162. DOI: 10.1163/187529263X00313.

305 Edgar, R.C. (2004). MUSCLE: multiple sequence alignment with high accuracy and high
306 throughput. *Nucleic Acids Research* 32, 1792-1797. DOI: 10.1093/nar/gkh340

307 Ferris, V.R., Goseco, C.G. & Ferris, J.M. (1980). Revisions in *Oxydirus* and *Tarjanius* n.
308 gen. in Oxydiridae, Belondiroidea (Nematoda: Dorylaimida); and *Oxydiroides* in
309 Prodorylaimidae, Dorylaimoidea. *Purdue University Agriculture Experimental*
310 *Station Research Bulletin* 965, 1-29.

311 Goseco, C.G. & Ferris, V.R. (1979). New data for *Leptonchus transvaalensis* from
312 Nigeria, and key to *Leptonchus*. *Journal of Nematology* 11, 108-110.

313 Heyns, J. (1965). On the morphology and taxonomy of the Aporcelaimidae, a new family
314 of dorylaimid nematodes. *Entomology Memoirs Department of Agriculture*
315 *Technical Services Republic of South Africa* 10, 1-51.

316 Heyns, J. (1968). A monographic study of the nematode families Nygolaimidae and
317 Nygolaimellidae. *Entomology Memoirs Plant Protection Research Institute*
318 *Pretoria, South Africa* 19, 1-144.

319 Huelsenbeck, J.P. & Ronquist, F. (2001). MRBAYES: Bayesian inference of
320 phylogenetic trees. *Bioinformatics* 17, 754-755.

321 Jairajpuri, M.S. (1968). Studies on the genus *Utahnema* Thorne, 1939 (Nematoda:
322 Dorylaimida). *Annales Épiphyties* 19, 727-735.

323 Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton,
324 S., Cooper, A., Markowitz, S., Duran, C., Thierer, T., Ashton, B., Meintjes, P. &
325 Drummond, A. (2012). Geneious Basic: an integrated and extendable desktop
326 software platform for the organization and analysis of sequence data. *Bioinformatics*
327 28, 1647-1649. DOI:10.1093/bioinformatics/bts199

328 Larget, B. & Simon, D.L. (1999). Markov chain Monte Carlo algorithms for the Bayesian
329 analysis of phylogenetic trees. *Molecular Biology and Evolution* 16, 750-759. DOI:
330 10.1093/

331 Lee, D.L. (1961). Two new species of cryptobiotic (anabiotic) freshwater nematodes,
332 *Actinolaimus hintoni* and *Dorylaimus keilini* sp. nov. (Dorylaimidae). *Parasitology*
333 51, 237-240.

334 Loof, P.A.A. & Coomans, A. (1970). On the development and location of the oesophageal
335 gland nuclei in Dorylaimina. *Proceedings of the IX International Nematology*
336 *Symposium (Warsaw, Poland, 1967)*, 79-161.

337 Loof, P.A.A. & Coomans, A. (1972). The oesophageal gland nuclei of Longidoridae
338 (Dorylaimida). *Nematologica* 18, 213-233.

339 Nico, A.I., Rapoport, H.F., Jiménez-Díaz, R.M. & Castillo, P. (2002). Incidence and
340 population density of plant-parasitic nematodes associated with olive planting
341 stocks at nurseries in southern Spain. *Plant Disease* 86, 1075-1079.

342 Peña-Santiago, R., Abolafia, J. & Álvarez-Ortega, S. (2014). New proposal for a detailed
343 description of the dorylaim spicule (Nematoda: Dorylaimida). *Nematology* 16, 1091-
344 1095.

345 Peña-Santiago, R., Peralta, M. & Siddiqi, M.R. (1993). Taxonomy of some new and
346 known species of the genus *Tylencholaimellus* with a proposal for *Margollus* gen. n.
347 (Nematoda: Dorylaimida). *Nematologica* 39, 218-233.

348 Rashidifard, M., Marais, M., Daneel, M.S., Mienie, C.M.S. & Fourie, H. (2019)
349 Molecular characterisation of *Meloidogyne enterolobii* and other *Meloidogyne* spp.

350 from South Africa. *Tropical Plant Pathology* 44, 213-224. DOI: 10.1007/s40858-
351 019-00281-4.

352 Siddiqi, M.R. (1971). *Oriverutus lobatus* gen. n., sp. n. and *Sicaguttur sartum* gen. n., sp.
353 n. (Nematoda: Dorylaimoidea) from cultivated soils in Africa. *Nematologica* 16
354 (1970), 483-491.

355 Siddiqi, M.R. (1982a). Six new genera of dorylaimid nematodes. *Nematologica* 27(1981),
356 397-421.

357 Siddiqi, M.R. (1982b). *Sicorinema* gen. n. and *Moshajia* gen. n. (Dorylaimida:
358 Crateronematidae) with descriptions of four new species. *Nematologia Mediterranea*
359 10, 157-166.

360 Siddiqi, M.R. (1983a). Taxonomy of the subfamily Dorylaimellinae (Nematoda:
361 Dorylaimida). *Pakistan Journal of Nematology* 1, 1-38.

362 Siddiqi, M.R. (1983b). Three new species of *Aculonchus* n. gen. and *Zetalaimus*
363 *bleferonchus* n. gen., n. sp. (Dorylaimida: Belonenchinae). *Indian Journal of*
364 *Nematology* 12(1982), 312-319.

365 Siddiqi, M.R. (1995). Nematodes of tropical rainforests. 5. Seven new genera and forty-
366 two new species of dorylaims. *Afro-Asian Journal of Nematology* 5, 72-109.

367 Subbotin, S.A., Sturhan, D., Chizhov, V.N., Vovlas, N. & Baldwin, J.G. (2006).
368 Phylogenetic analysis of Tylenchida Thorne, 1949 as inferred from D2 and D3
369 expansion fragments of the 28S rRNA gene sequences. *Nematology* 8, 455-474. DOI:
370 10.1163/156854106778493420

371 Thorne, G. (1930). Predaceous nemas of the genus *Nygolaimus* and a new genus
372 *Sectonema*. *Journal of Agricultural Research* 41, 445-466.

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Figure legends

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Figure 1. *Aporcelaimellus nigeriensis* sp. n. A: Entire body (male). B: Entire body (female). C, D: Anterior region in lateral median view. E: Lip region in lateral surface view. F: Vagina. G: Neck region. H: Female, posterior genital branch. I: Male, posterior body region. J: Spicule. K: Female, posterior body region. L: Lateral guiding piece. M: Female, caudal region. N: Male, caudal region. (Scale bars: A, B = 500 μm ; C, D, J = 20 μm ; E, F, M, N = 10 μm ; G, I = 100 μm ; H, K = 50 μm ; L = 5 μm .)

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Figure 2. *Aporcelaimellus nigeriensis* sp. n. (LM). A-C: Anterior region in lateral median view. D: Lip region in lateral surface view. E: Entire body (female). F-H: Coarse ventral pores at different regions of body. I: Pharyngeal enlargement showing DO and DN. J, M: Vagina. K: Female, posterior body region. L, N: Female, caudal region. (Scale bars: A-C = 20 μm ; D, F-H, J, L-N = 10 μm ; E = 500 μm , K = 50 μm .)

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Figure 3. *Aporcelaimellus nigeriensis* sp. n. (LM). Male. A: Entire. B: Caudal region. C: Lateral guiding piece. D: Posterior body region. E, F: Spicules. (Scale bars: A = 500 μm ; B = 10 μm ; C = 5 μm , D = 100 μm , E, F = 20 μm .)

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Figure 4. Bayesian inference (BI) with 50% majority rule of *Aporcelaimellus nigeriensis* sp. n. using LSU (D2-D3) ribosomal DNA under GTR+I+G model. Posterior probabilities more than 50% are given for the appropriate clade. Original sequence is indicated by bold font.

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Figure 5. Bayesian inference (BI) with 50% majority rule of *Aporcelaimellus nigeriensis* sp. n. using SSU ribosomal DNA under GTR+I+G model. Posterior probabilities more than 50% are given for the appropriate clade. Original sequence is indicated by bold font.

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Table 1. Dorylaimid species recorded in Nigeria.

Species	Habitat	Reference
<i>Afronygus longicaudatus</i> Heyns, 1968	Bush, maize	Heyns (1968)
<i>Discolaimoides bulbiferus</i> (Cobb, 1906) Heyns, 1963	?	Das <i>et al.</i> (1969)
<i>Dorylaimellus (Ibadanus) brevidens</i> Siddiqi, 1983	Maize	Siddiqi (1983a)
<i>Glochidorella brevicula</i> Siddiqi, 1982	Forest	Siddiqi (1982a)
<i>Helmabia campyla</i> Siddiqi, 1995	Rubber tree	Siddiqi (1995)
<i>Helmabia falcata</i> Siddiqi, 1995	Rainforest	Siddiqi (1995)
<i>Laimydorus keilini</i> (Lee, 1961) Andr�ssy, 1986	Freshwater-pool	Lee (1961)
<i>Leptonchus transvaalensis</i> Heyns, 1963	?	Goseco and Ferris (1979)
<i>Longidorus attenuatus</i> Hooper, 1961	?	Loof and Coomans (1970)
<i>Longidorus laevicapitatus</i> Williams, 1959	?	Loof and Coomans (1970)
<i>Longidorus macrosoma</i> Hooper, 1961	?	Loof and Coomans (1970)
<i>Longidorus pisi</i> Edward, Misra & Singh, 1964 ¹	?	Loof and Coomans (1972)
<i>Moshajia cultristyla</i> Siddiqi, 1982	Rice	Siddiqi (1982)
<i>Neoactinolaimus hintoni</i> (Lee, 1961) Thorne, 1967	Freshwater-pool	Lee (1961)
<i>Opisthodorylaimus cavalcantii</i> (Lordello, 1955) Carbonell & Coomans, 1986		Carbonell and Coomans (1986)
<i>Oriverutus lobatus</i> Siddiqi, 1971	Rice	Siddiqi (1971)
<i>Oxydirus gangeticus</i> Siddiqi, 1966	?	Andr�ssy (2009)
<i>Paraoxydirus cavenessi</i> (Ferris, Goseco & Ferris, 1980) Siddiqi, 1983	Banana	Ferris <i>et al.</i> (1980)
<i>Sicorinema sericatum</i> Siddiqi, 1982	Rubber tree	Siddiqi (1982b)
<i>Solididens bisexualis</i> (Thorne, 1930) Heyns, 1968	?	Heyns (1968)
<i>Thornedia nigerica</i> (Jairajpuri, 1968) Jairajpuri & Ahmad, 1992	<i>Mucuna utilis</i>	Jairajpuri (1968)
<i>Thornenema mauritianum</i> (Williams, 1959) Baqri & Jairajpuri, 1968	?	Carbonell and Coomans (1987)
<i>Tylencholaimellus cylindricus</i> Pe�a-Santiago, Peralta & Siddiqi, 1993	Sugarcane	Pe�a-Santiago <i>et al.</i> (1993)
<i>Zetalaimus blepheronchus</i> Siddiqi, 1983	Rainforest	Siddiqi (1983b)

¹Recorded as *Longidorus siddiqii* Aboul-Eid, 1970.

433 **Table 2.** Morphometrics of *Aporcelaimellus nigeriensis* sp. n.
 434 Measurements in μm except L in mm.

Character	Population	Type			Modakeke	
	n	Holotype	Paratypes		2♀♀	3♂♂
		♀	2♀♀	♂		
L		2.98	3.34, 3.41	3.17	2.76, 3.11	2.83-3.55
a		22	21, 24	26	20, 21	22-28
b		4.2	4.8, 4.8	4.2	4.0, 4.0	4.1-4.6
c		90	108, 97	77	89, 115	72-98
V		50	51, 50	-	54, 49	-
c'		0.6	0.5, 0.5	0.7	0.5, 0.5	0.5-0.7
Lip region diam.		26	24, 26	27	?, 27	26-27
Odontostyle length-dorsal side		34	33, 31	30	35, 36	35
ventral side		31	29, 28	28	30, 32	29-31
Odontophore length		63	62, 62	63	64, 60	58-64
Neck length		706	700, 708	758	691, 779	648-764
Pharyngeal expansion length		401	423, 399	418	392, 383	347-424
Body diam. at neck base		126	141, 123	113	129, 141	115-126
mid-body		135	158, 141	123	138, 148	127-130
anus/cloaca		59	60, 68	56	58, 54	56-59
Distance vulva – anterior end		1493	1706, 1718	-	1486, 1536	-
Prerectum length		128	143, 123	174	99, 126	143-210
Rectum/cloaca length		65	71, 79	83	78, 75	81-88
Tail length		33	31, 35	41	31, 27	30-41
Spicules length		-	-	137	-	108-132
Ventromedian supplements		-	-	10	-	9-10