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Multiscale mapping of plant functional groups and plant traits in the High Arctic using field spectroscopy, UAV imagery and Sentinel-2A data

Eleanor R Thomson^{1,*}, Marcus P Spiegel¹, Inge H J Althuizen², Polly Bass³, Shuli Chen⁴, Adam Chmurzynski⁴, Aud H Halbritter², Jonathan J Henn⁵, Ingibjörg S Jónsdóttir^{6,11}, Kari Klanderud⁷, Yaoqi Li^{4,8}, Brian S Maitner⁴, Sean T Michaletz⁹, Pekka Niittynen¹⁰, Ruben E Roos⁷, Richard J Telford², Brian J Enquist⁴, Vigdis Vandvik², Marc Macias-Fauria¹

and Yadvinder Malhi¹ 💿

- School of Geography and the Environment, University of Oxford, Oxford, United Kingdom
- ² Department of Biological Sciences and Bjerknes Centre for Climate Research, University of Bergen, Bergen, Norway
- ³ Matanuska-Susitna College, University of Alaska, Anchorage, AK, United States of America
- ⁴ Department of Ecology and Evolutionary Biology, University of Arizona, Tucson, AZ, United States of America
- ⁵ Department of Integrative Biology, University of Wisconsin-Madison, Madison, WI, United States of America
- ⁶ Faculty of Life and Environmental Sciences, University of Iceland, Reykjavik, Iceland
- ⁷ Faculty of Environmental Sciences and Natural Resource Management, Norwegian University of Life Sciences, Aas Norway
 ⁸ Institute of Ecology and Key Laboratory for Earth Surface Processes of the Ministry of Education, College of Urban and
- Environmental Sciences, Peking University, Beijing, People's Republic of China
- ⁹ Department of Botany and Biodiversity Research Centre, University of British Columbia, Vancouver, Canada
- ¹⁰ Department of Geosciences and Geography, University of Helsinki, Helsinki, Finland
- ¹¹ Arctic Biology, The University Centre in Svalbard, Longyearbyen, Norway
- * Author to whom any correspondence should be addressed.

E-mail: eleanor.thomson@oriel.ox.ac.uk

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Abstract

The Arctic is warming twice as fast as the rest of the planet, leading to rapid changes in species composition and plant functional trait variation. Landscape-level maps of vegetation composition and trait distributions are required to expand spatially-limited plot studies, overcome sampling biases associated with the most accessible research areas, and create baselines from which to monitor environmental change. Unmanned aerial vehicles (UAVs) have emerged as a low-cost method to generate high-resolution imagery and bridge the gap between fine-scale field studies and lower resolution satellite analyses. Here we used field spectroscopy data (400-2500 nm) and UAV multispectral imagery to test spectral methods of species identification and plant water and chemistry retrieval near Longyearbyen, Svalbard. Using the field spectroscopy data and Random Forest analysis, we were able to distinguish eight common High Arctic plant tundra species with 74% accuracy. Using partial least squares regression (PLSR), we were able to predict corresponding water, nitrogen, phosphorus and C:N values ($r^2 = 0.61-0.88$, RMSEmean = 12%-64%). We developed analogous models using UAV imagery (five bands: Blue, Green, Red, Red Edge and Near-Infrared) and scaled up the results across a 450 m long nutrient gradient located underneath a seabird colony. At the UAV level, we were able to map three plant functional groups (mosses, graminoids and dwarf shrubs) at 72% accuracy and generate maps of plant chemistry. Our maps show a clear marine-derived fertility gradient, mediated by geomorphology. We used the UAV results to explore two methods of upscaling plant water content to the wider landscape using Sentinel-2A imagery. Our results are pertinent for high resolution, low-cost mapping of the Arctic.

1. Introduction

The Arctic is the fastest warming region on earth [1]. Air temperatures have risen at twice the global rate [2], driving changes to the structure and functioning of tundra ecosystems [3–7]. Increased temperatures lead to shifts in vegetation cover and composition and accelerate belowground nutrient cycling and mineralization rates, with potentially important effects on global climate [8–14].

Plant traits are a primary control on the distribution and functioning of plants and underpin vegetation-climate responses (e.g. [15-17]). Traits related to the uptake and allocation of resources, such as leaf mass per area, leaf water content, leaf nitrogen content (N) and leaf phosphorus content (P) can affect growth rates, plant longevity, primary productivity, decomposition rates and biogeochemical cycling [18-22]. Simultaneously, morphologyrelated traits, such as leaf area and plant height, can influence aboveground biomass, surface albedo and snow dynamics [14, 23, 24]. Establishing spatial environmental-trait relationships could provide a quantitative basis to forecast the effects of climate change in the Arctic [25, 26]. Yet observational plot-studies often fail to isolate environmental drivers or take place at a smaller scale than the community/landscape they represent. Hence, the biotic consequences of warming in the coldest environments remain poorly predicted [27].

When variations in vegetation characteristics affect spectral reflectance, these changes can be measured using optical remote sensing techniques. In comparison to traditional field-based methods, using remote sensing to generate vegetation data is less laborious, less invasive, more cost-effective and spatially continuous. Using multispectral satellite-based sensors, the distribution of Arctic vegetation has been well-established at broad scales through datasets such as the Circumpolar Arctic Vegetation Map (CAVM [28]), which classifies 16 Arctic vegetation types at 1 km resolution. Using coarse-resolution satellite imagery, widespread vegetation change at high latitudes has also been well-documented, including pan-Arctic 'greening' and 'browning' trends [29-32], with important implications for albedo, active layer depth, permafrost dynamics, carbon cycling and wildlife [10, 33–41]. However, vegetation cover in the Arctic is extremely heterogeneous, varying at very fine spatial scales ([42–44]), and observations do not always agree at the satellite and plot level [32, 45, 46]. Thus, while remote sensing offers enormous potential to generate large-scale vegetation data across the Arctic, such coarse spatial and spectral resolution is limited in its ability to capture the fine-scale dynamics of tundra plants or resolve the key drivers of observed vegetation trends [32, 47, 48].

In lower-latitude environments, highresolution field spectroscopy (visible-near infrared

(VNIR: 400-1100)) or visible-short-wave infrared (VSWIR: 400-2500 nm) is a well-established technique for gathering plant taxonomic and trait data with a high degree of accuracy [49-52]. Specific spectral reflectance and absorption features have been directly linked to concentrations of cellulose, lignin, chlorophyll, nutrients and water (see [53] for a comprehensive synthesis). These plant traits can be combined to create unique chemical 'fingerprints' to distinguish individual genera or species [54-57]. Compared to multispectral remote sensing, field spectroscopy provides numerous spectral bands that can capture fine-scale spectral differences and improve discrimination of vegetation features. Spectroscopy studies are rarely extended to the tundra however, likely due to the challenging logistics and the fact that field spectrometers are not designed to accommodate the small size of tundra plant leaves. Spectroscopy studies carried out in the Arctic have mainly focused on classifying tundra communities [58–62], vegetation cover fraction [63], biophysical plant traits, such as vegetation height and biomass [61, 64], and leaf chlorophyll content or 'greenness' [61, 65], but not individual species or plant chemical traits. Remote sensing of high-latitude vegetation would benefit from more spectroscopy studies, which can link plant characteristics to spectral features at specific wavelengths, as well as establish remote sensing baselines (i.e. if a vegetation characteristic cannot be determined using field spectroscopy, it is unlikely to be distinguished using coarser spatial and spectral resolution imagery).

By itself however, spectroscopy cannot generate the spatial and temporal data required to map and monitor biotic change in the Arctic. In order to bridge this 'scale-gap', unmanned aerial vehicles (UAVs) have emerged as platforms to upscale field data and provide nuance to satellite observations [47, 48, 66]. Due to the miniaturization of technology and decreasing costs, UAV-mounted sensors can facilitate the mapping and monitoring of vegetation at previously unachievable spatial, spectral and temporal resolutions. Using hyper and multispectral imagery, UAVs have been used to map plant communities [67, 68], individual species [69-71], plant traits [72-74] and plant health [69, 75-77]. While some UAV studies have applied structurefrom-motion methods to analyse biophysical plant characteristics in the Arctic (e.g. [78, 79]), or applied spectral techniques to classify high latitude plant communities (e.g. [59, 67, 68, 80]), to the best of our knowledge, this is the first study to use field spectroscopy and UAVs to identify high Arctic plant species and map vegetation chemistry across the tundra.

In this study, we investigate the use of field spectroscopy for predicting species identity and retrieving plant water and chemistry concentrations across a range of common species at three sites found near Longyearbyen, Svalbard. Based on the demonstrated feasibility of generating plant taxonomic and trait data using field spectra, we develop analogous models using UAV five-band multispectral imagery. Additionally, we explore methods of upscaling our UAV results to the wider landscape using Sentinel-2A imagery. Specifically, we aim to:

- (a) develop predictive models of species identity and biochemical plant traits (water, nitrogen, phosphorus and C:N concentrations) using field spectroscopy,
- (b) develop predictive models of species identity and the same plant traits using UAV spectra,
- (c) apply the UAV models to UAV imagery to derive spatially continuous maps of plant species and their corresponding trait values,
- (d) explore two methods of upscaling the UAV models to the wider landscape using Sentinel-2A imagery.

2. Methods

2.1. Study sites

The data were collected at three High Arctic study sites around Longyearbyen on the Svalbard Archipelago, Norway (figure 1). The sites comprised two contrasting environmental gradients and an experimental warming site. The sites are characterized by a dry Arctic climate with a mean average temperature of -2.6 °C and an average maximum and minimum monthly temperature of 7.6 °C and -11.4 °C (2005–2018). Precipitation is around 190 mm per year (all data obtained from the National Center of Environmental Information [81]). In all cases, underlying soils were typical cryosols with a thin organic layer on top of inorganic sediments [82].

2.1.1. The Birdcliffs (78.14° N, 15.2° E)

Located near Bjørndalen, up the slope of Platåfjellet, the Birdcliffs is a 175 m high, 450 m long nutrient and elevation gradient, stretching from the base of a Little Auk (Alle alle) and Black-legged Kittiwake (Rissa tridactyla) colony to sea level at Isfjorden. The top of the gradient receives high nutrient input in the form of bird guano and other associated biological material [83]. The majority of the site is characterised by steep slopes and large alluvial fans made up of a mix unconsolidated material of various sizes. Vegetation is dominated by dry dwarf shrub tundra in topographically elevated areas, particularly Salix polaris and Dryas octopetala. Mosses and graminoids dominate in moist areas, especially around the edge of the alluvial fans and towards the flatter end of the gradient near the shore.

2.1.2. The control gradient (78.12° N, 15.4° E)

The Control Gradient is an 850 m long colluvial slope located up the north face of Lindholmhøgda, a spur at the mouth of the Adventdalen valley. The gradient runs from 200 m.a.s.l to a reservoir (Isdammen) at sea level. The area is characterised by dry dwarf shrub tundra that alternates with ridge communities of scarce vegetation. Habitats with thin winter snow cover are dominated by *Dryas octopetala* and *Salix polaris* (*Dryas* heath), while habitats of intermediate snow depth are dominated by *Cassiope tetragona* (*Cassiope* heath).

2.1.3. The international tundra experiment (78.18° N, 15.77° E)

The International Tundra Experiment (ITEX [84]) is an experimental warming site located in Endalen. Data were collected outside the warming chambers from three predominant habitats: an exposed and relatively dry *Dryas* heath with shallow snow cover in winter, a mesic *Cassiope* heath with intermediate winter snow depth and a moist snowbed community dominated by bryophytes, *Salix polaris, Bistortavivipara* and graminoids.

2.2. UAV imagery acquisition

All fieldwork was conducted as part of the Plant Functional Trait Course 4 (https:// plantfunctionaltraitscourses.w.uib.no/), from 16 July 2018 to 27 July 2018. See figure 2 for a diagrammatic representation of the data collection and analysis.

UAV imaging data were acquired from all three sites. The data were acquired using a 3DR Solo drone equipped with a MicaSense RedEdge-MX multispectral camera and MicaSense RedEdge Downwelling Light Sensor (DLS). The MicaSense RedEdge-MX camera captures surface reflectance at five narrow spectral bands: blue (475 nm), green (560 nm), red (668 nm), Red Edge (717 nm) and NIR (840 nm). The DLS points upwards and captures illumination conditions, which are embedded in the metadata of each image. The drone was flown at an altitude of 40-60 m, resulting in <10 cm resolution imagery. To map the entirety of the study sites, multiple overlapping flights were required, which were taken over the course of one day at each site. Radiometric calibration images were recorded using a MicaSense reflectance panel as the calibration target. Ground control points for georeferencing were taken using the Emlid Reach+ differential GNSS system (Emlid, Hong Kong).

The imagery was processed in Pix4Dmapper (v.4.3.31, Pix4D, Lausanne, Switzerland) using a standard structure-from-motion (SfM) workflow. For each site, images from all flights were processed in the same project to form a single orthomosaic. The GCPs were manually identified in the images and georeferenced using their field-collected RTK–GNSS coordinates. Across the three sites, this resulted in RMS errors of 0.10–0.14 m. Radiometric calibration was performed in Pix4D using the MicaSense reflectance target images and the metadata from the DLS.

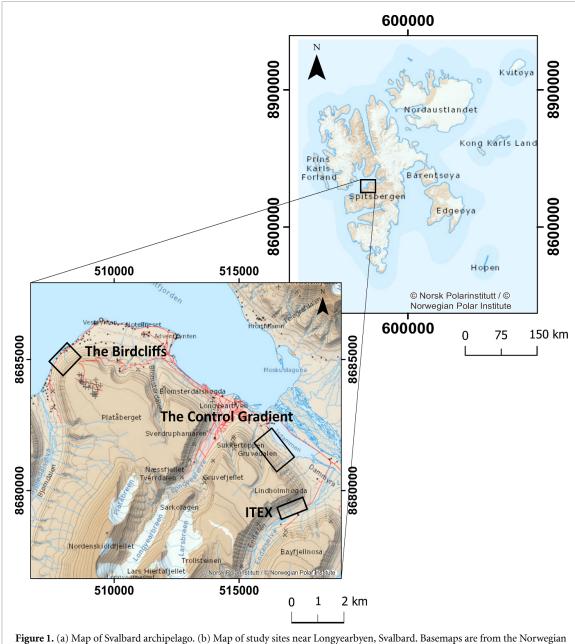


Figure 1. (a) Map of Svalbard archipelago. (b) Map of study sites near Longyearbyen, Svalbard. Basemaps are from the Norwegian Polar Institute. Coordinates in UTM Zone 33 N, WGS84 ellipsoid.

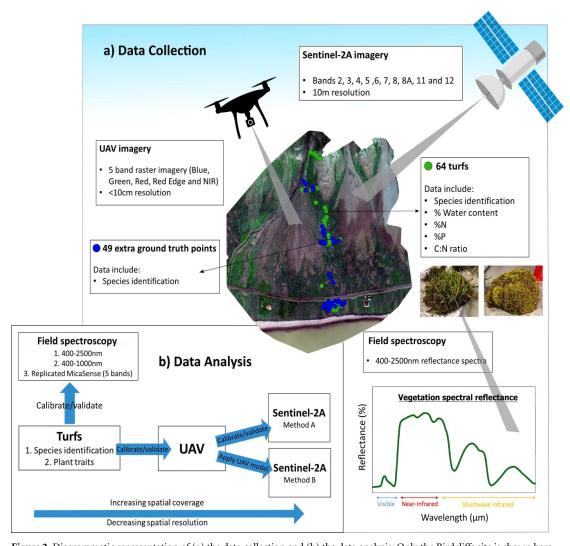
2.3. Vegetation sampling

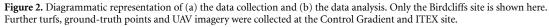
After the UAV imagery acquisition, $68\ 20 \times 20\ cm$ single-species turfs were collected from across the Birdcliffs and ITEX sites. The turfs were selected to represent the most common plant functional types identified across all sites: mosses, graminoids and dwarf shrubs (table 1, figure 2). High-accuracy GNSS coordinates were taken from the locations of the extracted turfs with an Emlid Reach+differential GNSS system. Additional vegetation ground-truthing points were taken from all three sites (table 2, figure 2). The turfs were cut to a substrate depth of approximately 5 cm, sealed inside plastic bags and transported back to the University Centre in Svalbard (UNIS) for species identification and analysis.

Some shrub samples collected from the dry and mesic heaths at ITEX showed signs of tissue degradation, probably due to drought or frost damage [85]. These samples were included in the plant trait analyses, as part of a continuum of trait values, but excluded from the species classification analyses. Mapping plant health falls outside the scope of this study and only healthy shrub communities were observed over the UAV upscaling areas.

2.3.1. Field spectroscopy measurements

Field spectroscopy measurements of the turf samples (350–2500 nm) were taken at UNIS using an ASD Fieldspec Pro with fibre optic cable and contact probe (Analytical Spectral Devices, Boulder, CO, USA). Turfs were stored outside and reflectance





measurements were taken within 24 h of turf cutting. Spectroscopy measurements were not taken 'in situ' at study sites, due to the non-portable set-up of the spectrometer. Turfs were selected to be as homogenous as possible. If multiple plant species were present across the turf, measurements were only taken from areas where the main species dominated. The contact probe was pushed firmly down onto the turf so all extraneous light was excluded from the measurement. Five measurements were taken at different locations across each turf. Each measurement consisted of 40 internally averaged reflectance readings to increase the signal-to-noise ratio. The spectrometer was optimised and calibrated for dark current and white light after every turf. For all statistical analyses, the spectral data were trimmed to the 400-2500 nm range. For some analyses, the five measurements were averaged to form one spectrum per turf.

2.3.2. Plant trait sampling

After the field spectroscopy measurements had been taken, three 5 \times 5 cm vegetation samples were

cut from each turf. For each sample, all vegetation above the substrate was harvested. The samples were weighed for fresh mass, dried at 60 °C for 48 h and re-weighed for dry mass. Vegetation water content was calculated as ((fresh mass – dry mass)/fresh mass) \times 100.

Nutrient analysis of the dried samples was carried out at the University of Arizona. The protocol to obtain total phosphorus concentration involved using persulfate oxidation followed by the acid molybdate method (APHA, 1992), after which P concentration was determined via colorimetric analysis with a spectrophotometer (ThermoScientific Genesys20, United States). Carbon, nitrogen, and their stable isotope ratios were measured at the Department of Geosciences Environmental Isotope Laboratory on a continuous-flow gas-ratio mass spectrometer.

The three samples were averaged to form one trait value per turf. See table S1 (available online at stacks.iop.org/ERL/16/055006/mmedia) for average trait values.

Turf samples					
Site	Plant functional type	Genus	Species	N. of turf samples	N. of Spectra
Birdcliffs	Moss	Aulacomnium	Palustre	3	15
		Aulacomnium	Turgidum	4	20
		Polytrichastrum	Alpinum	2	10
		Polytrichum	Hyperboreum	1	5
		Polytrichum	Strictum	1	5
		Racomitrium	Canescens	2	10
		Racomitrium	Lanuginosum	2	10
Birdcliffs	Graminoid	Alopecurus	Ovatus	7	35
		Luzula	Confusa	6	30
		Luzula	Nivalis	1	5
Birdcliffs	Dwarf shrub	Cassiope	Tetragona	2	10
		Dryas	Octopetala	5	25
		Salix	Polaris	9	45
ITEX	Moss	Identity not recorded		5	25
ITEX	Graminoid	Identity not recorded		1	5
ITEX	Dwarf shrub	Cassiope	Tetragona	2	10
	Dwarf Shrub	Dryas	Octopetala	3	15
	Dwarf Shrub	Salix	Polaris	3	15
	Senesced Shrub	Cassiope	Tetragona	6	30
	Senesced Shrub	Dryas	Octopetala	3	15
				Total: 68	Total: 340

Table 1. Information on turf samples, including site location, plant functional type, genus and species. Turfs measured 20×20 cm. Five independent spectrum readings were taken per turf.

 Table 2. Information on ground-truthing vegetation survey,

 including site, plant functional type and number of data points.

 GNSS locations were taken with an Emlid Reach+ differential

 GNSS.

	Plant functional	N. data points	
Site	type		
Birdcliffs	Moss	15	
	Graminoid	23	
	Dwarf Shrub	24	
Control Gradient	Moss	9	
	Graminoid	0	
	Dwarf shrub	9	
ITEX	Moss	6	
	Graminoid	1	
	Dwarf shrub	30	
		Total: 117	

2.4. Field spectroscopy analyses

2.4.1. Field spectroscopy-species analysis

The turf spectra were divided into eight vegetation classes (below) representing a mixture of families, genera and species. Some of the moss and graminoid species were combined into their families or genera to increase the class sample size and because they were assumed to be spectrally similar. Mosses were classified into Aulacomnium spp. (containing Aulacomnium palustre and Aulacomnium turgidum); Polytrichaceae spp. (containing Polytrichum hyperboreum, Polytrichastrum alpinum and Polytrichastrum strictum); and Racomitrium spp. (containing *Racomitrium canescens* and *Racomitrium lanuginosum*). The graminoids were classified into *Alopecurus ovatus* (a grass) and *Luzula spp.* (a rush, containing *Luzula confusa* and *Luzula nivalis*). The dwarf shrubs were separated into *Cassiope tetragona*, *Dryas octopetala* and *Salix polaris*.

To quantify similarity or differences between the spectra, agglomerative hierarchical cluster analysis was performed using the eight classes described above and one averaged spectrum per turf. Cluster analysis works by calculating the distance (difference) between each possible pair of spectra. The two spectra closest to each other are merged and the process repeats, with the number of clusters reduced by one each cycle. Spectral distances were calculated using Euclidean Distance in MATLAB (MathWorks, Natick, MA, USA).

The eight vegetation classes were predicted using five spectra per turf and Random Forest classification [86]. The five spectra per turf were used as independent samples, as the spectral curves were highly diverse due to the heterogeneous nature of the turfs. Random Forest is a machine learning classifier that grows as an ensemble of decision trees [87, 88]. It is useful for classifying hyperspectral data as it is robust against overfitting and can be used when the number of predictor variables (e.g. 2150 wavelength bands) is greater than the number of samples (e.g. 470 spectra). All the Random Forest analyses were carried out in MATLAB using the Treebagger function.

2.4.2. Field spectroscopy-trait analysis

Plant water, nitrogen, phosphorous and C:N values were predicted using one averaged spectrum per turf and partial least squares regression (PLSR [89]). The PLSR method is effective, as it uses the continuous spectrum as a single measurement, rather than carrying out a band-by-band analysis and reduces a large predictor matrix (2150 spectral bands) down to a few relatively uncorrelated factors (known as latent variables). For the trait predictions, we chose to use PLSR instead of Random Forest Regression because, unlike Random Forest Regression, PLSR is able to extrapolate beyond its calibration data. While Random Forest is a strong tool for classification, it cannot predict values outside of its training range, limiting its accuracy when scaling up continuous data over wider areas that may contain trait values outside the range of the sample data.

PLSR analyses were carried out using the PLSregress function in MATLAB. The PLSR models were validated using an unseen 30% of the dataset. Due to the random nature of the 70:30 split, 1000 PLSR iterations were made for each spectra-trait analysis. The PLSR equations resulting in robust models (r^2 > the mean value of the 1000 iterations) were evaluated using r^2 for the independent testing (val) dataset and RMSE as a percentage of the sample mean (as in [90, 91]).

All Random Forest and PLSR analyses were carried out using (a) the full hyperspectral range from 400 to 2500 nm, (b) just the VIS–NIR region from 400 to 1000 nm and (c) downscaled spectra to match the five bands represented by the MicaSense camera (blue, green, red, red edge and near-infrared). The three analyses were carried out in order to act as a baseline from which the effects of decreased spectral information and the effects of using an airborne camera could be accurately separated.

2.5. UAV spectral analysis

2.5.1. UAV spectra-species analysis

UAV spectra were extracted from the orthomosaic at all the points where the turfs and ground-truthed species coordinates were taken. The spectra were extracted using the 'extract' function in the Raster package [92] in R (v.3.6.0, R Core Team, 2019). First, an NDVI mask was applied to the imagery to mask out any pixels with an NDVI of <0.5. The threshold of 0.5 was chosen as it represents the lower limit of NDVI values measured for vegetation in the Arctic [93]. The UAV spectra were extracted using three methods: simple (only the pixel value the coordinate falls in is returned), bilinear (the returned value is interpolated from the values of the four nearest pixels) and buffer (the returned value is averaged from all the pixels that fall within a specified 3×3 pixel buffer area). Across all analyses, the buffer method demonstrated the most robust results, hence only the results from these analyses are shown.

Using the UAV spectra, Random Forest classification was used to predict three broad plant functional groups: mosses, graminoids and dwarf shrubs. These plant functional groups represented the finest classification of vegetation that could be achieved using the UAV data and the Random Forest method of prediction. The Random Forest model that demonstrated the best result on the training/validation data was scaled up and applied to each pixel of the Birdcliff imagery to generate a continuous ecosystem map of plant functional type. The models were only applied to the Birdcliff site, as this was where the majority of the turfs were collected. As such, we only had confidence in the model validation for the species and environmental conditions found at this site.

2.5.2. UAV spectra-trait analysis

Trait values were predicted using the same UAV spectra and PLSR method described above. The PLSR model that maximised the r^2 and minimised the %RMSE was chosen and applied to each pixel of the UAV imagery to generate continuous maps of trait information. Trait values were also compared to an NDVI map generated from the UAV imagery to quantify the benefits of using a multispectral machine learning approach to trait prediction, compared to an NDVI-based prediction (figure S3).

2.6. Sentinel-2A scaling analysis

Sentinel-2A imagery was used to explore methods of upscaling the UAV maps to the wider region. Plant water content was chosen as an example trait, as water has a strong direct expression in spectra and the Birdcliff site was assumed to be a nutrient anomaly in the landscape.

The Sentinel image used in the analysis was a Level-2A cloud-free, atmospherically-corrected image taken on 30th July 2018, one week after the field-collection. Only vegetation pixels were included in the analysis, identified using the Sentinel-2 scene classification layer (SCL). Analyses were carried out in R (v.3.6.0, R Core Team, 2019) and Google Earth Engine (https://earthengine.google.org/).

Two methods of upscaling from UAV to satellite data were tested:

(a) the UAV-generated map of plant water content (figure 8) was resampled to match Sentinel-2A resolution (10 m) and gridlines using bilinear interpolation. Lower-resolution Sentinel bands were also resampled to 10 m using bilinear interpolation. The resampled UAV water values were used as a calibration and validation dataset for the Sentinel imagery. Thus, each Sentinel pixel (n = 329) had a single water value as well as a reflectance value for each relevant Sentinel band (Band 2 (492 nm), 3 (560 nm), 4 (665 nm), 5 (704 nm), 6 (741 nm), 7 (783 nm), 8 (833 nm), 8A (865 nm), 11 (1614 nm) and 12 (2202 nm)).

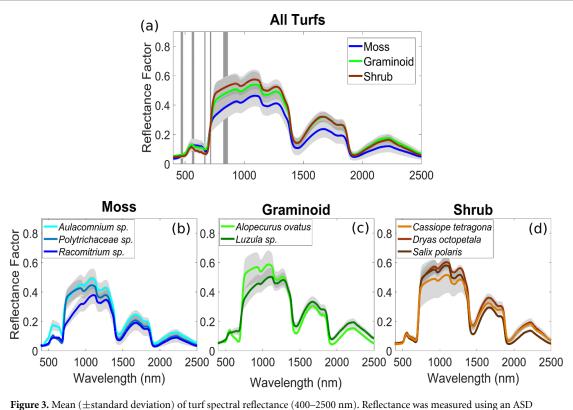


Figure 3. Mean (±standard deviation) of turf spectral reflectance (400–2500 nm). Reflectance was measured using an ASD Fieldspec Pro with fibre optic cable and contact probe. (a) Spectra were grouped into three plant functional types; mosses, graminoids and dwarf shrubs. The MicaSense RedEdge-MX five spectral bands (blue, green, red, red edge and near–infrared) are shown in grey for comparison purposes. (b) Mosses were classified into *Aulacomnium spp. (A. palustre* and *A turgidum), Polytrichaceae spp. (P. hyperboreum, P. alpinum* and *P. strictum)*, and *Racomitrium spp. (R. canescens* and *R. lanuginosum).* (c) Graminoids were classified into *Aluzula spp. (L. confusa* and *L. nivalis*). (d) Shrubs were separated into *C. tetragona, D. octopetala* and *S. polaris.*

Using this dataset and the same PLSR method described above, model coefficients were generated for each Sentinel band and applied to each pixel of the Sentinel image.

(b) the coefficients from the PLSR model used to create the UAV map of water content were applied to the Sentinel bands that best matched the MicaSense bands (Sentinel bands 2,3,4,5 and 8) for each pixel of the Sentinel image.

3. Results

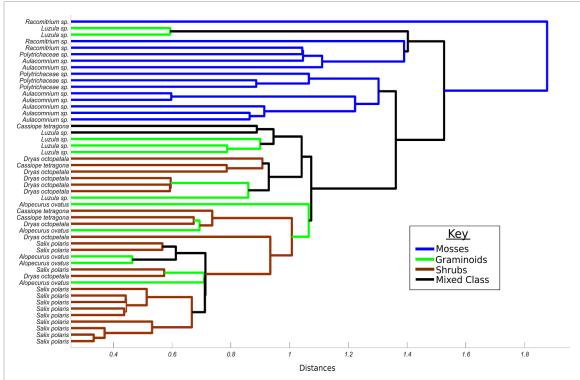
3.1. Field spectroscopy identification of plant functional types and species

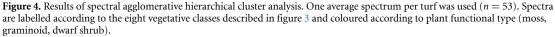
Average reflectance spectra for three plant functional types (moss, graminoids and dwarf shrubs) showed distinct spectral separability, especially in the NIR (750–1300 nm) and SWIR regions (1500–2500 nm, figure 3(a)). Moss displayed the lowest reflectance across the NIR–SWIR region, especially in the water absorption bands near 1450, 1940 and 2500 nm [53]. Within each plant functional type, unique spectral signatures were displayed by genera and species (figures 3(b)-(d)). Shrubs were more spectrally similar to each other, but showed some divergence in the NIR–SWIR. Intraspecific spectral variability was low across most of the mosses, while *Alopecurus ovatus*

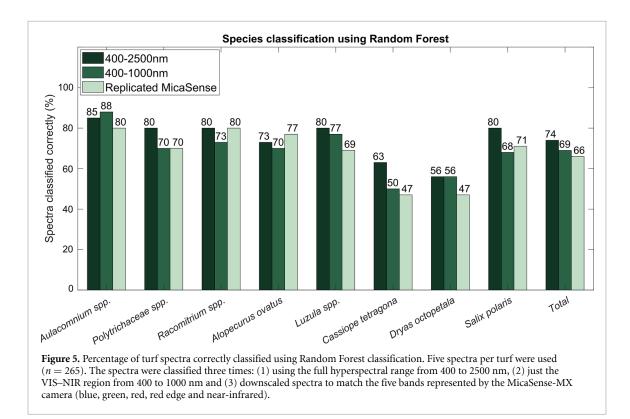
and *Cassiope tetragona* displayed the highest spectral variability, especially across the NIR.

Using cluster analysis to further quantify spectral separability, mosses emerged as a distinct spectral family (figure 4). All moss spectra clustered together, with some stratification of genera, especially *Aulacomnium* and *Racomitrium*. There was less guild clustering of graminoids and shrubs, however *Salix polaris* emerged as a tight cluster and most *Dryas octopetala* spectra clustered together. Despite some grouping of *Luzula*, the graminoids did not cluster together as a functional group. The majority of the spectral confusion was created by *Alopecurus ovatus* and *Cassiope tetragona*, whose high spectral variability meant they were commonly paired with other shrub and graminoid species.

Using Random Forest Classification and the full hyperspectral range (400–2500 nm), our results show that eight common Arctic tundra species can be classified with 74% accuracy (much above the expected statistical random accuracy value of 12.5%, given eight classes, figure 5). Reflecting the results of the cluster analysis, moss species displayed the highest classification accuracy values (85, 80 and 80%), along with *Salix polaris* (80%) and *Luzula sp.* (80%). Reducing the amount of spectral information led to a small decrease in accuracy to 69 and 66% for the







400–1000 nm spectral range and downscaled Mica-Sense bands, respectively. Across the eight classes, the greatest source of uncertainty came from the classification of *Cassiope tetragona* and *Dryas octopetala*, which were commonly confused with each other (see *figure S1* for confusion matrices). **3.2. Field spectroscopy identification of plant traits** Using the full reflectance spectra (400–2500 nm), PLSR results show that plant water content was estimated with the highest accuracy ($r^2 = 0.88$, RMSE = 12%), followed by N ($r^2 = 0.77$, RMSE = 28%), P ($r^2 = 0.67$, RMSE = 64%) and C:N

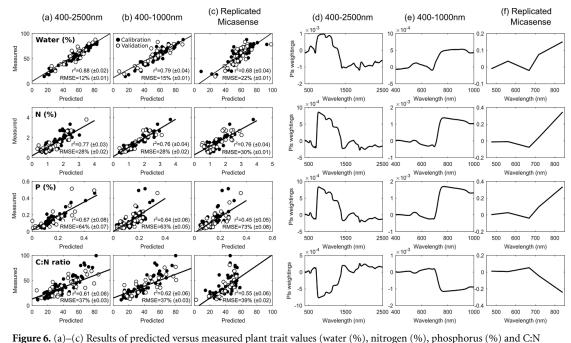


Figure 6. (a)–(c) Results of predicted versus measured plant trait values (water (%), nitrogen (%), phosphorus (%) and C:N ratios) using PLSR analysis. (d)–(f) PLSR spectral weightings for each plant trait. Deviation from 0 indicates the parts of the spectrum that contribute most strongly to prediction. Spectra were analysed three times, using: (1) the full hyperspectral range from 400 to 2500 nm, (2) the VIS–NIR region from 400 to 1000 nm and (3) downscaled spectra to simulate the five bands represented by the MicaSense-MX camera (blue, green, red, red edge and near-infrared).

ratios ($r^2 = 0.61$, RMSE = 37%, figure 6(a)). Removing spectral information from the SWIR range had the greatest impact on water prediction. Decreased SWIR spectral information had almost no impact on N prediction, and little impact on C:N ratio prediction. P estimation showed little change with reduced SWIR information but was strongly influenced by fewer VIS–NIR spectral bands (figures 6(b) and (c)). The spectral weightings showed that all regions of the spectrum are important to trait prediction when the full spectrum is used, but the red-edge and NIR region are particularly important when SWIR information is lacking (figures 6(d)–(f)).

3.3. UAV spectral mapping of plant functional types

When using the UAV MicaSense camera, the eight vegetation classes could not be classified with the same accuracy as when using the downscaled hyperspectral data. Instead, vegetation cover was classified into three plant functional types; mosses, graminoids and dwarf shrubs (figure 7(b)). Overall classification accuracies were 72% across the validation dataset, much above the statistical random accuracy of 33% expected for three classes. Dwarf shrubs were classified with the highest accuracy (83%), followed by graminoids (61%) and mosses (57%, figure 7(c)). Vegetation cover across the Birdcliff site was dominated by dwarf shrubs. Graminoids were shown as present at the confluence of runoff channels and on some of the flatter, wetter areas before the fjord. Mosses showed speckling between the shrubs and

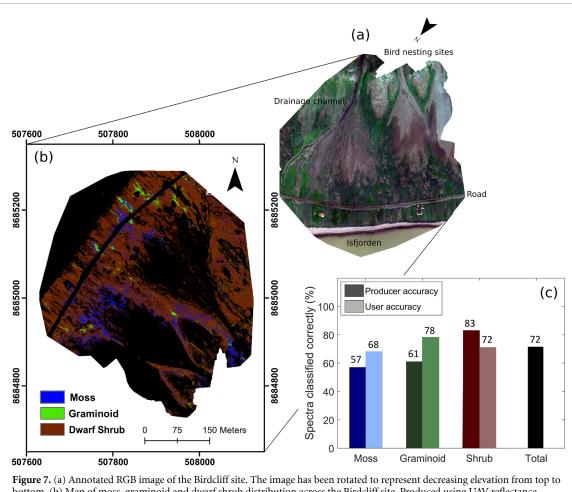
were found in dense communities around the bottom of the colluvial fans.

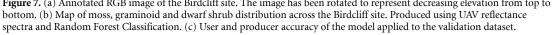
3.4. UAV spectral mapping of plant traits

Using the UAV five-band data, average correlations with water and N were significantly lower (p < 0.001) than when using the downscaled hyperspectral data (figure 8). Plant water trait correlations decreased from $r^2 = 0.68$ to $r^2 = 0.50$ and N accuracies decreased from $r^2 = 0.76$ to $r^2 = 0.50$. The exception was P, where the average correlations between the UAV spectroscopy and field measurements increased from $r^2 = 0.45$ to $r^2 = 0.54$ compared to the downscaled hyperspectral data. Scaling up the most robust models, there was a clear plant water and nutrient gradient across the site. Plant water and nutrient values were generally highest at the top of the nutrient and elevation gradient and declined downslope. Nutrient concentrations also appeared to follow drainage channels across the site and were tightly correlated to each other.

3.5. Sentinel-2A spectral mapping of plant water content

Scaling from the UAV to the satellite level, calibrating a Sentinel-specific PLSR model on the UAV plant water map values (figure 9(a)) and applying the UAV plant water model directly to the Sentinel imagery (figure 9(b)) produced similar patterns but a very different magnitude of result. The maps were correlated with each other at r = 0.86, with a *y*-intercept of 41%. For method A, plant water content values





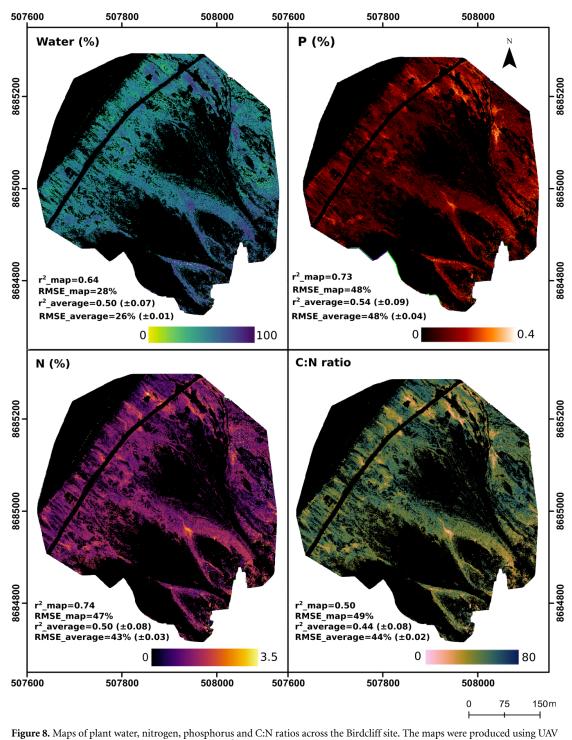
ranged between 30% and 80%, while method B overestimated water content, with values ranging from 80% upwards.

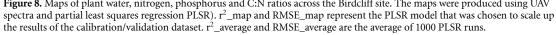
4. Discussion

Our results show that UAV multispectral data can be used to map fine-scale vegetation cover (moss, graminoids and dwarf shrubs) across the High Arctic and monitor changes in plant water and nutrient traits. Using field spectroscopy data, we were able to distinguish eight common tundra species at 74% accuracy, suggesting good prospects for near-future vegetation mapping at the species level with the increasing commercialisation of UAV-hyperspectral systems and next generation of hyperspectral satellites (CHIME, EnMAP, HISUI, HypXIM, HyspIRI, PRISMA etc [94-99]). Although only CHIME, EnMAP and PRISMA cover the high latitudes (extending to 84° N, 80° N and 70° N respectively), hyperspectral technology in this area is expanding.

Mosses in particular displayed a unique spectral signature and distinct spectral separability within tundra vegetation. Average moss reflectance across the NIR and SWIR was lower than for graminoids and shrubs, reflecting their lack of vascular system and high water content. All three families/genera of moss, Aulacomnium, Polytrichaceae and Racomitrium, displayed high prediction accuracies in the Random Forest analysis (85, 80% and 80% respectively), which is promising for the automatic detection of moss taxa in future hyperspectral studies. In northern environments, mosses can dominate aboveground primary productivity and have been shown to display a wide range of traits and life history strategies [100–103]. Yet, due to the difficulty of identifying species, mosses are often ignored by ecologists in favour of vascular plants, or conglomerated into groups that may or may not be meaningful [104]. Thus, the ability to distinguish moss taxa at broad scales would facilitate research focusing on this significant but understudied plant group.

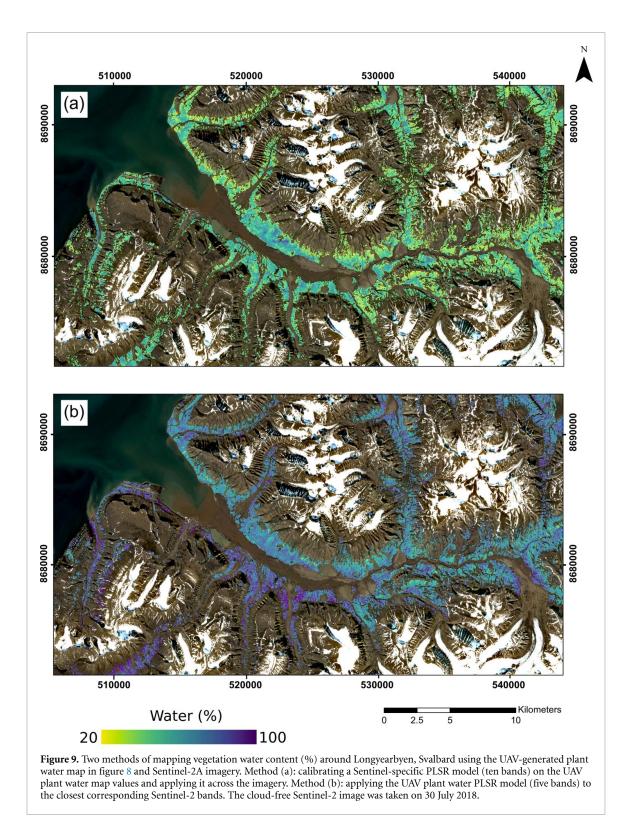
Graminoids showed similar spectral separability with a 73% and 80% prediction accuracy for *Alopecurus* ovatus and *Luzula*, respectively. *Salix polaris* also demonstrated a high prediction accuracy of 80%, whereas some spectral confusion was created between the two evergreen dwarf shrubs, *Cassiope* and *Dryas*, likely due to *Cassiope*'s woody vertical structure that created high variation in its spectral signature (figure 3). Developing spectroscopic techniques





to improve the identification of dwarf shrub species would be useful at broad scales as, along with snow depth, the litter quality of *Cassiope, Dryas* and *Salix* has been linked to differences in soil organic matter and respiration rates [105]. *Cassiope, Dryas* and *Salix* also display contrasting evergreen and deciduous strategies, which govern different relationships between soil nutrient availability, photosynthesis and growth rates [106]. Overall, these results are highly encouraging with respect to the spectral separability of tundra vegetation and application of fine-scale remote sensing in the Arctic.

At the trait level, we found that four plant traits (water, N, P and C:N values) could be estimated with varying accuracy using field spectroscopy. Plant water and N concentrations were strongly correlated with the full range spectra and showed good agreement with previous PLSR studies at the leaf and canopy scale (e.g. [91, 107, 108]). The estimation of P was slightly lower than previous studies



(e.g. [91, 106, 109, 110]), although the PLSR models still captured a substantial amount of variation across the data. Unlike water and N, P has no direct expression across the 400–2500 nm spectrum. Predictions result from correlations with other traits, especially nitrogen, attributed to the stoichiometric link between N and P—a link which has been used to map leaf P across large areas of the tropics and identify crop-level P deficiencies ([111–113]). In contrast to forest or agricultural studies however, N:P ratios may

differ more widely among our samples, through different plant groups having fundamentally different N:P ratios ([114]) and high nutrient deposition from bird colonies leading to uneven N:P soil ratios.

When downscaling the hyperspectral data, estimations of plant water content showed a corresponding decrease in accuracy with the number of reduced spectral bands. The nutrient traits were less sensitive to reduced SWIR information, reflecting that water is expressed in a wide range of spectral bands across the VNIR-SWIR spectrum [53], whereas N is a major component of chlorophyll with strong expression in blue and green VIS bandwidths (430-660 nm [53, 115]). P displayed similar spectral weightings to N although, when downscaled to the five Mica-Sense bands, P values were highly sensitive to a reduction in VNIR information (as similarly found by [110, 116]). In contrast, N showed no significant reduction in accuracy across the downscaled analyses, further suggesting that, in Arctic environments, estimations of P arise from collinearities with traits other than N. We have confidence that the models are detecting trait variation, rather than species-level differences, given the observed spectral weightings and high intraspecific trait variation within Arctic plant functional groups (see table S1 and [117]).

Using the multispectral UAV data, we were able to map moss, graminoids and dwarf shrubs at 72% accuracy. Forbs were not included in the model, as the vegetation cover of any individual forb species was not extensive enough to sample sufficiently. In future studies, it would be useful to test whether forbs can be spectrally distinguished from graminoids (as found by [80] using multispectral satellite imagery, LiDAR and phenology), or whether their similar functionality render forbs and graminoids too related to be differentiated by spectra alone.

Across the Birdcliff site, modelled vegetation distribution was dominated by shrub cover. High nutrient input from nesting colonies generally leads to the formation of moss tundra below bird cliffs on Svalbard [118, 119], but along warmer coastlines, such as Bjørndalen, vascular plants have been found to proliferate [120]. Although moss only constituted 10% of the vegetation cover across the site, the ability to separate moss-dominated pixels from shrubs and graminoids is a key strength of high-resolution mapping over satellite-scale analyses. Regular monitoring of moss communities can act as an early warning system for climatic shifts and associated biotic and abiotic effects in polar environments [121–123]. However, it is important to remember that this study represented a snapshot in time, where moisture conditions, illumination conditions and phenological phase were unique. Time-series data is required to investigate whether moss and other vegetation types can be distinguished with the same accuracy at different phenological stages and under varying abiotic conditions [124, 125].

Using the UAV imagery, we were unable to distinguish vegetation at the species level. This is despite the downscaled hyperspectral data displaying species level accuracy of 64%. We attribute this loss in accuracy to two main reasons. Firstly, UAV imagery suffers from quality issues to a greater extent than field-measured spectra. These include bidirectional reflectance distribution function effects, variability in illumination conditions during a flight, poor radiometric calibration and spectral mixing, even at high resolutions [126, 127]. These effects are even more pronounced in the Arctic due to low sun angle, variable cloud cover and heterogeneous tundra. While the UAV imagery shows a correlation with the field-collected spectra (figure S2), there is substantial variation in the agreement, with the UAV spectra likely exhibiting more statistical error. This may explain why the highest accuracies in our analyses were achieved by averaging spectra from a square of nine pixels (18×18 cm) rather than extracting single pixels, due to the increased signal-to-noise ratio of the averaged spectra.

Secondly, accurate geolocation of the groundtruthed vegetation points can be challenging. Across the Arctic, species distribution is extremely heterogeneous and ground-truthing points may only define species presence within a radius of <20 cm. Although RMS errors for the orthomosaics were low (<0.14 m), this could still have resulted in an offset between the ground-truthed vegetation points and the imagery, leading to some inaccuracies in the calibration and validation points. Thus, in environments with high spatial heterogeneity, extremely precise georeferencing equipment is required to accurately calibrate and validate the information collected from UAVs. We recommend covering vegetation around groundtruthing points to create an exclusion zone, or similar methods to reduce the spectral effect of nearby species. In addition, when dealing with mixed plant communities, novel spectral methods are required to capture the co-occurrence of plant types. Rather than discrete vegetation categories, a more flexible model may be required, that removes individual classifications and instead places a vegetation pixel on a continuum of values between moss, graminoid or shrub.

At the landscape level, plant trait distributions displayed a steep water and fertility gradient from the bird cliff colony to the sea, mediated by geomorphology. Plant water content was highest directly underneath the bird colonies and at the confluence of drainage channels. N, P and C:N values indicated well-fertilised vegetation under nesting sites and reflected the transport of nutrients downslope through a network of drainage channels. For example, plant water and nutrient levels were extremely high in the depositional area below the snowmelt channel (figure 7), demonstrating the role that run-off processes play in distributing marine-derived organic matter across the otherwise nutrient-limited tundra. While graminoids constituted just 4% of the vegetation cover across the mapped area, they were mostly found in moist, fertile areas, reflecting their fast growth rates and ability to take swift advantage of available nutrients [128]. This suggests that increased mineralization rates under a warming climate may preferentially benefit graminoid species (as found by [129–131]).

Adding to our confidence that the models were predicting traits explicitly, rather than through correlations with plant type, vegetation cover or productivity, multispectral methods of plant trait retrieval showed a clear independence from NDVIbased predictions (figure S3). With the exception of C:N ratios, neither measured nor predicted trait values were significantly correlated with NDVI, demonstrating the value of a multispectral mapping approach and the lack of physical basis for using NDVI to predict plant traits in the Arctic.

Scaling from the UAV to satellite level, we found different methods of upscaling had a large effect on the magnitude of results, but little influence on the spatial pattern of trait values. Although we do not present either map in figure 9 as accurate, or have the data to validate either map, we found that method A (calibrating Sentinel imagery on UAV-derived plant water values) produced more varying and realistic trait values than method B (applying a UAV plant water model to Sentinel imagery). The higher variation in trait values using method A is probably due to the greater number of spectral bands used in the analysis. In method A, all the relevant Sentinel bands were included as predictors (ten in total), whereas in method B, only the five Sentinel bands that best matched the spectral bands of the MicaSense camera could be used by the UAV plant water model. The overestimation of trait values in method B demonstrates the difference in reflectance captured by UAVs and satellites (see figure S4). Cross-sensor differences arise from atmospheric effects (e.g. haze), differences in viewing and illumination angles, sensor specifications (e.g. bandwidths), and non-linear spectral mixing at different spatial resolutions [32, 127, 132, 133], meaning predictive models generated from one type of imagery cannot be directly applied to another with the same accuracy.

To upscale from UAV to satellite imagery therefore, we recommend creating satellite-specific models calibrated on UAV maps (e.g. method A). Similar methods have been used for a variety of ecologicalmapping tasks, such as tree species classification, percentage vegetation cover, agricultural assessments, habitat mapping and fire severity quantification [66, 90, 134–139]. To the best of our knowledge however, this is the first time that upscaling from UAV to satellite imagery has been used to quantify plant traits. With greater UAV coverage across the landscape and more accurate UAV-models ($r^2 > 0.9$), using UAV information to calibrate satellite-based models may provide a valuable method for expanding spatially limited plot studies across the Arctic.

5. Conclusion

Using field spectroscopy, we showed that eight common Arctic plants can be distinguished at the species level at 74% accuracy. These results are relevant for UAV-hyperspectral systems, the first Arctic passenger aircraft equipped with hyperspectral instruments [140] and upcoming hyperspectral satellites, which should be able to use similar methods explored here to advance these results and expand them over a wider geographical area.

Using UAV multispectral information, we have shown that High Arctic moss-, graminoid- and dwarf shrub-dominated vegetation can be mapped at high resolution (<10 cm) with 72% accuracy, alongside corresponding water, nitrogen, phosphorus and C:N values. We present two methods of upscaling these results using Sentinel-2A imagery. The majority of Arctic tundra regions remain under-investigated and difficult to access for scientific research. We must continue to develop methods to expand spatiallylimited plot studies, which inherently cannot capture landscape-level vegetation patterns or functional trait variation, in order to monitor environmental change across the Arctic and understand the mechanisms driving large-scale climate-vegetation feedbacks.

Data availability statement

The data that support the findings of this study are openly available at the following URL/DOI: https://osf.io/smbqh/.

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ORCID iDs

Eleanor R Thomson () https://orcid.org/0000-0003-1670-8970

Marcus P Spiegel b https://orcid.org/0000-0001-5879-5465 Inge H J Althuizen () https://orcid.org/0000-0003-3485-9609

Aud H Halbritter lo https://orcid.org/0000-0003-2597-6328

Jonathan J Henn in https://orcid.org/0000-0003-1551-9238

Ingibjörg S Jónsdóttir b https://orcid.org/0000-0003-3804-7077

Kari Klanderud © https://orcid.org/0000-0003-1049-7025

Yaoqi Li [©] https://orcid.org/0000-0001-6540-395X Brian S Maitner [©] https://orcid.org/0000-0002-2118-9880

Sean T Michaletz in https://orcid.org/0000-0003-2158-6525

Pekka Niittynen () https://orcid.org/0000-0002-7290-029X

Ruben E Roos (a) https://orcid.org/0000-0002-1580-6424

Brian J Enquist lhttps://orcid.org/0000-0002-6124-7096

Vigdis Vandvik lo https://orcid.org/0000-0003-4651-4798

Marc Macias-Fauria https://orcid.org/0000-0002-8438-2223

Yadvinder Malhi liktps://orcid.org/0000-0002-3503-4783

References

- Box J E, Colgan W T, Christensen T R, Schmidt N M, Lund M, Parmentier F-J W, Brown R, Bhatt U S, Euskirchen E S and Romanovsky V E 2019 Key indicators of arctic climate change: 1971–2017 *Environ. Res. Lett.* 14 045010
- [2] Stocker T F, Qin D, Plattner G-K, Tignor M, Allen S K, Boschung J, Nauels A, Xia Y, Bex V and Midgley P M 2013 Climate change 2013: the physical science basis Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change (Cambridge: Cambridge University Press) p 1535
- [3] Joshua L A, Klein E S, Oberbauer S F and Welker J M 2016 Coupled long-term summer warming and deeper snow alters species composition and stimulates gross primary productivity in tussock tundra *Oecologia* 181 287–97
- [4] Park T, Ganguly S, Tømmervik H, Euskirchen E S, Høgda K-A, Karlsen S R, Brovkin V, Nemani R R and Myneni R B 2016 Changes in growing season duration and productivity of northern vegetation inferred from long-term remote sensing data *Environ. Res. Lett.* 11 084001
- [5] Khorsand Rosa R, Oberbauer S F, Starr G, Parker La Puma I, Pop E, Ahlquist L and Baldwin T 2015 Plant phenological responses to a long-term experimental extension of growing season and soil warming in the tussock tundra of Alaska *Glob. Change Biol.* 21 4520–32
- [6] Oberbauer S F, Elmendorf S C, Troxler T G, Hollister R D, Rocha A V, Bret-Harte M S, Dawes M A, Fosaa A M, Henry G H R and Høye T T 2013 Phenological response of tundra plants to background climate variation tested using the International Tundra Experiment *Phil. Trans. R. Soc.* B 368 20120481
- [7] Wahren C H A, Walker M D and Bret-Harte M S 2005 Vegetation responses in Alaskan arctic tundra after 8 years of a summer warming and winter snow manipulation experiment *Glob. Change Biol.* 11 537–52

- [8] Davidson S J, Sloan V L, Phoenix G K, Wagner R, Fisher J P, Oechel W C and Zona D 2016a Vegetation type dominates the spatial variability in CH₄ emissions across multiple arctic tundra landscapes *Ecosystems* 19 1116–32
- [9] Higuera P E, Brubaker L B, Anderson P M, Brown T A, Kennedy A T and Hu F S 2008 Frequent fires in ancient shrub tundra: implications of paleorecords for arctic environmental change PLoS One 3 e0001744
- [10] Loranty M M, Goetz S J and Beck P S A 2011 Tundra vegetation effects on pan-Arctic albedo *Environ. Res. Lett.* 6 024014
- [11] Nadelhoffer K J, Giblin A E, Shaver G R and Laundre J A 1991 Effects of temperature and substrate quality on element mineralization in six Arctic soils *Ecology* 72 242–53
- [12] Nauta A L, Heijmans M M P D, Blok D, Limpens J, Elberling B, Gallagher A, Li B, Petrov R E, Maximov T C and Van Huissteden J 2015 Permafrost collapse after shrub removal shifts tundra ecosystem to a methane source *Nat. Clim. Change* 5 67–70
- [13] Pearson R G, Phillips S J, Loranty M M, Beck P S A, Damoulas T, Knight S J and Goetz S J 2013 Shifts in Arctic vegetation and associated feedbacks under climate change *Nat. Clim. Change* 3 673–7
- [14] Sturm M and Douglas T 2005 Changing snow and shrub conditions affect albedo with global implications *J. Geophys. Res.* 110 1–13
- [15] Bjorkman A D, Myers-Smith I H, Elmendorf S C, Normand S, Rüger N, Beck P S A, Blach-Overgaard A, Blok D, Cornelissen J H C and Forbes B C 2018 Plant functional trait change across a warming tundra biome *Nature* 562 57–62
- [16] Mekonnen Z A, Riley W J and Grant R F 2018 Accelerated nutrient cycling and increased light competition will lead to 21st century shrub expansion in North American Arctic Tundra J. Geophys. Res. Biogeosci. 123 1683–701
- [17] Soudzilovskaia N A, Elumeeva T G, Onipchenko V G, Shidakov I I, Salpagarova F S, Khubiev A B, Tekeev D K and Cornelissen J H C 2013a Functional traits predict relationship between plant abundance dynamic and long-term climate warming *Proc. Natl Acad. Sci.* 110 18180–4
- [18] Cornwell W K, Cornelissen J H C, Amatangelo K, Dorrepaal E, Eviner V T, Godoy O, Hobbie S E, Hoorens B, Kurokawa H and Pérez-Harguindeguy N 2008 Plant species traits are the predominant control on litter decomposition rates within biomes worldwide *Ecol. Lett.* 11 1065–71
- [19] Díaz S, Kattge J, Cornelissen J H C, Wright I J, Lavorel S, Dray S, Reu B, Kleyer M, Wirth C and Prentice I C 2016 The global spectrum of plant form and function *Nature* 529 167–71
- [20] Reich P B, Walters M B and Ellsworth D S 1997 From tropics to tundra: global convergence in plant functioning *Proc. Natl Acad. Sci.* 94 13730–4
- [21] Shipley B, Lechowicz M J, Wright I and Reich P B 2006 Fundamental trade-offs generating the worldwide leaf economics spectrum *Ecology* 87 535–41
- [22] Wright I J, Reich P B, Westoby M, Ackerly D D, Baruch Z, Bongers F, Cavender-Bares J, Chapin T, Cornelissen J H C and Diemer M 2004 The worldwide leaf economics spectrum *Nature* 428 821–7
- [23] Hudson J M G and Henry G H R 2009 Increased plant biomass in a high Arctic heath community from 1981 to 2008 Ecology 90 2657–63
- [24] Juszak I, Erb A M, Maximov T C and Schaepman-Strub G 2014 Arctic shrub effects on NDVI, summer albedo and soil shading *Remote Sens. Environ.* 153 79–89
- [25] Myers-Smith I H, Thomas H J D and Bjorkman A D 2019 Plant traits inform predictions of tundra responses to global change *New Phytol.* 221 1742–8
- [26] Thomas H J et al 2020 Global plant trait relationships extend to the climatic extremes of the tundra biome Nat. Commun. 11 1–12

- [27] Metcalfe D B, Hermans T D G, Ahlstrand J, Becker M, Berggren M, Björk R G, Björkman M P, Blok D, Chaudhary N and Chisholm C 2018 Patchy field sampling biases understanding of climate change impacts across the Arctic Nat. Ecol. Evol. 2 1443–8
- [28] Raynolds M K, Walker D A, Balser A, Bay C, Campbell M, Cherosov M M, Daniëls F J A, Eidesen P B, Ermokhina K A and Frost G V 2019 A raster version of the circumpolar Arctic vegetation map (CAVM) *Remote Sens. Environ.* 232 111297
- [29] Myneni R B, Keeling C D, Tucker C J, Asrar G and Nemani R R 1997 Increased plant growth in the northern high latitudes from 1981 to 1991 *Nature* 386 698–702
- [30] Bhatt U S, Walker D A, Raynolds M K, Bieniek P A, Epstein H E, Comiso J C, Pinzon J E, Tucker C J and Polyakov I V 2013 Recent declines in warming and vegetation greening trends over pan-Arctic tundra *Remote Sens.* 5 4229–54
- [31] Zhang Y, Song C, Band L E, Sun G and Junxiang L 2017 Reanalysis of global terrestrial vegetation trends from MODIS products: browning or greening? *Remote Sens. Environ.* 191 145–55
- [32] Myers-Smith I H, Kerby J T, Phoenix G K, Bjerke J W, Epstein H E, Assmann J J, John C, Andreu-Hayles L, Angers-Blondin S and Beck P S A 2020 Complexity revealed in the greening of the Arctic *Nat. Clim. Change* 10 106–17
- [33] Blok D, Schaepman-Strub G, Bartholomeus H, Heijmans M M P D, Maximov T C and Berendse F 2011 The response of arctic vegetation to the summer climate: relation between shrub cover, NDVI, surface albedo and temperature *Environ. Res. Lett.* 6 035502
- [34] Frost G V, Epstein H E, Walker D A, Matyshak G and Ermokhina K 2018 Seasonal and long-term changes to active-layer temperatures after tall shrubland expansion and succession in Arctic tundra *Ecosystems* 21 507–20
- [35] Loranty M M and Goetz S J 2012 Shrub expansion and climate feedbacks in Arctic tundra *Environ. Res. Lett.* 7 011005
- [36] Bonfils C J W, Phillips T J, Lawrence D M, Cameron-Smith P, Riley W J and Subin Z M 2012 On the influence of shrub height and expansion on northern high latitude climate *Environ. Res. Lett.* 7 015503
- [37] Blok D, Heijmans M M P D, Schaepman-strub G, Kononov A V, Maximov T C and Berendse F 2010 Shrub expansion may reduce summer permafrost thaw in siberian tundra *Glob. Change Biol.* 16 1296–305
- [38] Arndt K A, Santos M J, Ustin S, Davidson S J, Stow D, Oechel W C, Tran T T P, Graybill B and Zona D 2019 Arctic greening associated with lengthening growing seasons in Northern Alaska *Environ. Res. Lett.* 14 125018
- [39] Fauchald P, Park T, Tømmervik H, Myneni R and Hausner V H 2017 Arctic greening from warming promotes declines in Caribou populations *Sci. Adv.* 3 e1601365
- [40] Ims R A, Henden J-A, Strømeng M A, Thingnes A V, Garmo M J and Jepsen J U 2019 Arctic greening and bird nest predation risk across tundra ecotones *Nat. Clim. Change* 9 607–10
- [41] Tape K D, Christie K, Carroll G and O'Donnell J A 2016 Novel wildlife in the a rctic: the influence of changing Riparian ecosystems and shrub habitat expansion on snowshoe hares *Glob. Change Biol.* 22 208–19
- [42] Opedal Ø H, Armbruster W S and Graae B J 2015 Linking small-scale topography with microclimate, plant species diversity and intra-specific trait variation in an alpine landscape *Plant Ecol. Divers.* 8 305–15
- [43] Suvanto S, Le Roux P C and Luoto M 2014 Arctic-alpine vegetation biomass is driven by fine-scale abiotic heterogeneity *Geogr. Ann.* A 96 549–60
- [44] Räsänen A and Virtanen T 2019 Data and resolution requirements in mapping vegetation in spatially heterogeneous landscapes *Remote Sens. Environ.* 230 111207

- [45] Pattison R R, Jorgenson J C, Raynolds M K and Welker J M 2015 Trends in NDVI and tundra community composition in the arctic of NE Alaska between 1984 and 2009 *Ecosystems* 18 707–19
- [46] Guay K C, Beck P S A, Berner L T, Goetz S J, Baccini A and Buermann W 2014 Vegetation productivity patterns at high northern latitudes: a multi-sensor satellite data assessment *Glob. Change Biol.* 20 3147–58
- [47] Assmann J J, Myers-Smith I H, Kerby J T, Cunliffe A M and Daskalova G N 2020 Drone data reveal heterogeneity in tundra greenness and phenology not captured by satellites *Environ. Res. Lett.* 15 125002
- [48] Siewert M B and Olofsson J 2020 Scale-dependency of Arctic ecosystem properties revealed by UAV Environ. Res. Lett. 15 094030
- [49] Asner G P, Martin R E and Suhaili A B 2012 Sources of canopy chemical and spectral diversity in lowland Bornean forest *Ecosystems* 15 504–17
- [50] Serbin S P, Singh A, McNeil B E, Kingdon C C and Townsend P A 2014 Spectroscopic determination of leaf morphological and biochemical traits for northern temperate and boreal tree species *Ecol. Appl.* 24 1651–69
- [51] Serbin S P, Jin W, Ely K S, Kruger E L, Townsend P A, Meng R, Wolfe B T, Chlus A, Wang Z and Rogers A 2019 From the Arctic to the tropics: multibiome prediction of leaf mass per area using leaf reflectance *New Phytol.* 224 1557–68
- [52] Roth K L, Roberts D A, Dennison P E, Alonzo M, Peterson S H and Beland M 2015 Differentiating plant species within and across diverse ecosystems with imaging spectroscopy *Remote Sens. Environ.* 167 135–51
- [53] Curran P J 1989 Remote sensing of foliar chemistry *Remote Sens. Environ.* 30 271–8
- [54] Asner G P, Knapp D E, Kennedy-Bowdoin T, Jones M O, Martin R E, Boardman J and Flint Hughes R 2008 Invasive species detection in hawaiian rainforests using airborne imaging spectroscopy and LiDAR *Remote Sens. Environ.* 112 1942–55
- [55] Durgante F M, Higuchi N, Almeida A and Vicentini A 2013 Species spectral signature: discriminating closely related plant species in the amazon with near-infrared leaf-spectroscopy For. Ecol. Manage. 291 240–8
- [56] Lopatin J, Fassnacht F E, Kattenborn T and Schmidtlein S 2017 Mapping plant species in mixed grassland communities using close range imaging spectroscopy *Remote Sens. Environ.* 201 12–23
- [57] Asner G P and Martin R E 2009 Airborne spectranomics: mapping canopy chemical and taxonomic diversity in tropical forests *Front. Ecol. Environ.* 7 269–76
- [58] Bratsch S N, Epstein H E, Buchhorn M and Walker D A 2016 Differentiating among four arctic tundra plant communities at Ivotuk, Alaska using field spectroscopy *Remote Sens.* 8 51
- [59] Davidson S J, Santos M J, Sloan V L, Watts J D, Phoenix G K, Oechel W C and Zona D 2016b Mapping Arctic tundra vegetation communities using field spectroscopy and multispectral satellite data in north Alaska, USA *Remote Sens.* 8 978
- [60] Beamish A L, Coops N, Chabrillat S and Heim B 2017 A phenological approach to spectral differentiation of low-Arctic tundra vegetation communities, north slope, Alaska *Remote Sens.* 9 1200
- [61] Buchhorn M, Walker D A, Heim B, Raynolds M K, Epstein H E and Schwieder M 2013 Ground-based hyperspectral characterization of Alaska tundra vegetation along environmental gradients *Remote Sens.* 5 3971–4005
- [62] Huemmrich K F, Gamon J A, Tweedie C E, Campbell P K E, Landis D R and Middleton E M 2013 Arctic tundra vegetation functional types based on photosynthetic physiology and optical properties *IEEE J. Sel. Top. Appl. Earth Obs. Remote Sens.* 6 265–75
- [63] Liu N, Budkewitsch P and Treitz P 2017 Examining spectral reflectance features related to arctic percent vegetation

cover: implications for hyperspectral remote sensing of Arctic tundra *Remote Sens. Environ.* **192** 58–72

- [64] Bratsch S, Epstein H, Buchhorn M, Walker D and Landes H 2017 Relationships between hyperspectral data and components of vegetation biomass in low Arctic tundra communities at Ivotuk, Alaska *Environ. Res. Lett.* **12** 025003
- [65] Kennedy B E, King D J and Duffe J 2020 Comparison of empirical and physical modelling for estimation of biochemical and biophysical vegetation properties: field scale analysis across an Arctic bioclimatic gradient *Remote Sens.* 12 3073
- [66] Riihimäki H, Luoto M and Heiskanen J 2019 Estimating fractional cover of tundra vegetation at multiple scales using unmanned aerial systems and optical satellite data *Remote Sens. Environ.* 224 119–32
- [67] Mora C, Vieira G, Pina P, Lousada M and Christiansen H H 2015 Land cover classification using high-resolution aerial photography in adventdalen, Svalbard *Geogr. Ann.* A 97 473–88
- [68] Tømmervik H, Karlsen S-R, Nilsen L, Johansen B, Storvold R, Zmarz A, Beck P S, Høgda K A, Goetz S and Park T 2014 Use of unmanned aircraft systems (UAS) in a multi-scale vegetation index study of Arctic plant communities in adventdalen on Svalbard *EARSeL eProc.* 13 47–52
- [69] Baena S, Moat J, Whaley O and Boyd D S 2017 Identifying species from the air: uAVs and the very high resolution challenge for plant conservation *PLoS One* 12 11
- [70] Lu B and Yuhong H 2017 Species classification using unmanned aerial vehicle (UAV)-acquired high spatial resolution imagery in a heterogeneous grassland *ISPRS J. Photogramm. Remote Sens.* **128** 73–85
- [71] Oldeland J, Große-Stoltenberg A, Naftal L and Strohbach B J 2017 The potential of UAV derived image features for discriminating savannah tree species *The Roles* of *Remote Sensing in Nature Conservation* (Berlin: Springer) pp 183–201
- [72] Constantin D, Rehak M, Akhtman Y and Liebisch F 2015 Detection of crop properties by means of hyperspectral remote sensing from a micro UAV Bornimer Agrartechnische Berichte 88 129–37
- [73] Capolupo A, Kooistra L, Berendonk C, Boccia L and Suomalainen J 2015 Estimating plant traits of grasslands from UAV-acquired hyperspectral images: a comparison of statistical approaches *ISPRS Int. J. Geo-Inf.* 4 2792–820
- [74] Thomson E R, Malhi Y, Bartholomeus H, Oliveras I, Gvozdevaite A, Peprah T, Suomalainen J, Quansah J, Seidu J and Adonteng C 2018 Mapping the leaf economic spectrum across West African tropical forests using UAV-acquired hyperspectral imagery *Remote Sens.* 10 1532
- [75] Dash J P, Pearse G D and Watt M S 2018 UAV multispectral imagery can complement satellite data for monitoring forest health *Remote Sens.* 10 1216
- [76] Lucieer A, Turner D, King D H and Robinson S A 2014 Using an unmanned aerial vehicle (UAV) to capture micro-topography of Antarctic moss beds Int. J. Appl. Earth Obs. Geoinf. 27 53–62
- [77] Turner D, Lucieer A, Malenovský Z, King D and Robinson S A 2018 Assessment of Antarctic moss health from multi-sensor uas imagery with random forest modelling *Int. J. Appl. Earth Obs. Geoinf.* 68 168–79
- [78] Fraser R H, Olthof I, Lantz T C and Schmitt C 2016 UAV photogrammetry for mapping vegetation in the low-Arctic *Arctic Sci.* 2 79–102
- [79] Palace M, Herrick C, DelGreco J, Finnell D, Garnello A J, McCalley C, McArthur K, Sullivan F and Varner R K 2018 Determining subarctic peatland vegetation using an unmanned aerial system (UAS) *Remote Sens.* 10 1498
- [80] Langford Z, Kumar J, Hoffman F M, Norby R J, Wullschleger S D, Sloan V L and Iversen C M 2016 Mapping Arctic plant functional type distributions in the barrow environmental observatory using WorldView-2 and LiDAR datasets *Remote Sens.* 8 733

- [81] Lawrimore J H, Ray R, Applequist S, Korzeniewski B and Menne M J 2016 Global Summary of the Month (GSOM), version 1.0. NOAA National Centers for Environmental Information (Accessed 26 October 2020)
- [82] Jones A, Stolbovay V, Tarnocai C, Broll G, Spaargaren O and Montanarella L 2010 Soil Atlas of the Northern Circumpolar Region (Luxembourg: European Commission)
- [83] Zwolicki A, Zmudczyńska-Skarbek K M, Iliszko L and Stempniewicz L 2013 Guano deposition and nutrient enrichment in the vicinity of planktivorous and piscivorous seabird colonies in spitsbergen *Polar Biol.* 36 363–72
- [84] Henry G H R and Molau U 1997 Tundra plants and climate change: the International Tundra Experiment (ITEX) *Glob. Change Biol.* 3 1–9
- [85] Bjerke J W, Treharne R, Vikhamar-Schuler D, Karlsen S R, Ravolainen V, Bokhorst S, Phoenix G K, Bochenek Z and Hans T 2017 Understanding the drivers of extensive plant damage in boreal and arctic ecosystems: insights from field surveys in the aftermath of damage *Sci. Total Environ.* 599 1965–76
- [86] Breiman L 2001 Random Forests Mach. Learn. 45 5-32
- [87] Ghimire B, Rogan J and Miller J 2010 Contextual land-cover classification: incorporating spatial dependence in land-cover classification models using random forests and the Getis statistic *Remote Sens. Lett.* 1 45–54
- [88] Kotsiantis S and Pintelas P 2004 Combining bagging and boosting Int. J. Comput. Intell. 1 324–33
- [89] Haaland D M and Thomas E V 1988 Partial least-squares methods for spectral analyses. 1. Relation to other quantitative calibration methods and the extraction of qualitative information *Anal. Chem.* **60** 1193–202
- [90] Asner G P, Martin R E, Anderson C B and Knapp D E 2015 Quantifying forest canopy traits: imaging spectroscopy versus field survey *Remote Sens. Environ.* 158 15–27
- [91] Asner G P and Martin R E 2015 Spectroscopic remote sensing of non-structural carbohydrates in forest canopies *Remote Sens.* 7 3526–47
- [92] Hijmans R J, Van Etten J, Cheng J, Mattiuzzi M, Sumner M, Greenberg J A, Lamigueiro O P, Bevan A, Racine E B and Shortridge A 2015 Package 'raster' *R Package*
- [93] Bokhorst S, Tømmervik H, Callaghan T V, Phoenix G K and Bjerke J W 2012 Vegetation recovery following extreme winter warming events in the sub-Arctic estimated using NDVI from remote sensing and handheld passive proximal sensors *Environ. Exp. Bot.* 81 18–25
- [94] Nieke J and Rast M 2018 Towards the copernicus hyperspectral imaging mission for the environment (CHIME) Proc. IEEE Int. Geosci. Remote Sens. Symp. (IGARSS) (July 2018) pp 157–9
- [95] Guanter L, Kaufmann H, Foerster S, Brosinsky A, Wulf H, Bochow M, Boesche N, Brell M, Buddenbaum H and Chabrillat S 2016 EnMAP Science Plan—Environmental Mapping and Analysis Program
- [96] Matsunaga T, Iwasaki A, Tsuchida S, Tanii J, Kashimura O, Nakamura R, Yamamoto H, Tachikawa T and Rokugawa S 2013 Current status of Hyperspectral Imager Suite (HISUI) Proc. IEEE Int. Geosci. Remote Sens. Symp. (IGARSS), 2013
- [97] Carrere V, Briottet X, Jacquemoud S, Marion R, Bourguignon A, Chami M, Dumont M, Minghelli-Roman A, Weber C and Marie-José L-F 2013 HYPXIM: a second generation high spatial resolution hyperspectral satellite for dual applications *Paper Presented at 5th Workshop on Hyperspectral Image and Signal Processing: Evolution in Remote Sensing (Gainsville, FL*, 25–28 June 2013)
- [98] Lee C M, Cable M L, Hook S J, Green R O, Ustin S L, Mandl D J and Middleton E M 2015 An introduction to the NASA Hyperspectral Infrared Imager (HyspIRI) mission and preparatory activities *Remote Sens. Environ.* 167 6–19
- [99] Pignatti S, Palombo A, Pascucci S, Romano F, Santini F, Simoniello T, Umberto A, Vincenzo C, Acito N and Diani M 2013 The PRISMA hyperspectral mission: science

activities and opportunities for agriculture and land monitoring *Proc. IEEE Int. Geosci. Remote Sens. Symp.* (IGARSS), 2013

- [100] Turetsky M R, Mack M C, Hollingsworth T N and Harden J W 2010 The role of mosses in ecosystem succession and function in Alaska's boreal forest *Can. J. For. Res.* 40 1237–64
- [101] Longton R E 1988 Adaptations and strategies of polar bryophytes Bot. J. Linn. Soc. 98 253–68
- [102] Hedderson T A and Longton R E 1996 Life history variation in mosses: water relations, size and phylogeny Oikos 77 31–43
- [103] Elumeeva T G, Soudzilovskaia N A, During H J and Cornelissen J H C 2011 The importance of colony structure versus shoot morphology for the water balance of 22 subarctic bryophyte species J. Veg. Sci. 22 152–64
- [104] Turetsky M R, Bond-Lamberty B, Euskirchen E, Talbot J, Steve Frolking A, McGuire D and Tuittila E - S 2012 The resilience and functional role of moss in boreal and Arctic ecosystems New Phytol. 196 49–67
- [105] Elberling B 2007 Annual soil CO₂ effluxes in the high arctic: the role of snow thickness and vegetation type *Soil Biol. Biochem.* **39** 646–54
- [106] Baddeley J A, Woodin S J and Alexander I J 1994 Effects of increased nitrogen and phosphorus availability on the photosynthesis and nutrient relations of three Arctic dwarf shrubs from svalbard *Functional Ecology* 8 pp 676–85
- [107] Asner G P and Martin R E 2008 Spectral and chemical analysis of tropical forests: scaling from leaf to canopy levels *Remote Sens. Environ.* 112 3958–70
- [108] Ullah S, Skidmore A K, Ramoelo A, Groen T A, Naeem M and Ali A 2014 Retrieval of leaf water content spanning the visible to thermal infrared spectra *ISPRS J. Photogramm. Remote Sens.* 93 56–64
- [109] Asner G P, Martin R E, Carranza-Jiménez L, Sinca F, Tupayachi R, Anderson C B and Martinez P 2014 Functional and biological diversity of foliar spectra in tree canopies throughout the Andes to Amazon region *New Phytol.* 204 127–39
- [110] Mutanga O and Kumar L 2007 Estimating and mapping grass phosphorus concentration in an African savanna using hyperspectral image data *Int. J. Remote Sens.* 28 4897–911
- [111] Porder S, Asner G P and Vitousek P M 2005 Ground-based and remotely sensed nutrient availability across a tropical landscape *Proc. Natl Acad. Sci.* **102** 10909–12
- [112] Asner G P, Knapp D E, Anderson C B, Martin R E and Vaughn N 2016 Large-scale climatic and geophysical controls on the leaf economics spectrum *Proc. Natl Acad. Sci.* 113 E4043–51
- [113] Mahajan G R, Sahoo R N, Pandey R N, Gupta V K and Kumar D 2014 Using hyperspectral remote sensing techniques to monitor nitrogen, phosphorus, sulphur and potassium in wheat (Triticum Aestivum L.) Precis. Agric. 15 499–522
- [114] Mahajan G R, Pandey R N, Sahoo R N, Gupta V K, Datta S C and Kumar D 2017 Monitoring nitrogen, phosphorus and sulphur in hybrid rice (Oryza Sativa L.) using hyperspectral remote sensing *Precis. Agric.* 18 736–61
- [115] Kokaly R F 2001 Investigating a physical basis for spectroscopic estimates of leaf nitrogen concentration *Remote Sens. Environ.* **75** 153–61
- [116] Milton N M, Eiswerth B A and Ager C M 1991 Effect of phosphorus deficiency on spectral reflectance and morphology of soybean plants *Remote Sens. Environ.* 36 121–7
- [117] Thomas H J D, Myers-Smith I H, Bjorkman A D, Elmendorf S C, Blok D, Cornelissen J H C, Forbes B C, Hollister R D, Normand S and Prevéy J S 2019 Traditional plant functional groups explain variation in economic but not size-related traits across the tundra biome *Glob. Ecol. Biogeogr.* 28 78–95

- [118] Hofmann W 1968 Geobotanische Untersuchungen in Sudost-Spitzbergen 1960 Ergebn. Stauferland-Exped. 1959/60,8 pp 1–83
- [119] Vanderpuye A W, Elvebakk A and Nilsen L 2002 Plant communities along environmental gradients of high-arctic mires in Sassendalen, Svalbard J. Veg. Sci. 13 875–84
- [120] Elven R, Eriksen M B, Elvebakk A, Johansen B and Engelskjøn T 1990 Gipsdalen, central svalbard; flora, vegetation and botanical values *Environmental Atlas*, *Gipsdalen, Svalbard. II. Nor. Polarinst. Rapp. Ser.* vol 61 pp 27–66
- [121] Callaghan T V, Press M C, Lee J A, Robinson D L and Anderson C W 1999 Spatial and temporal variability in the responses of Arctic terrestrial ecosystems to environmental change *Polar Res.* 18 191–7
- [122] Robinson S A, Turnbull J D and Lovelock C E 2005 Impact of changes in natural ultraviolet radiation on pigment composition, physiological and morphological characteristics of the Antarctic Moss, Grimmia Antarctici *Glob. Change Biol.* 11 476–89
- [123] Royles J, Amesbury M J, Convey P, Griffiths H, Hodgson D A, Leng M J and Charman D J 2013 Plants and soil microbes respond to recent warming on the Antarctic Peninsula *Curr. Biol.* 23 1702–6
- [124] May J L, Parker T, Unger S and Oberbauer S F 2018 Short term changes in moisture content drive strong changes in normalized difference vegetation index and gross primary productivity in four arctic moss communities *Remote Sens*. *Environ.* 212 114–20
- [125] Van Leeuwen W J D and Huete A R 1996 Effects of standing litter on the biophysical interpretation of plant canopies with spectral indices *Remote Sens. Environ.* 55 123–38
- [126] Assmann J J, Kerby J T, Cunliffe A M and Myers-Smith I H 2018 Vegetation monitoring using multispectral sensors—best practices and lessons learned from high latitudes J. Unmanned Veh. Syst. 7 54–75
- [127] Stow D, Nichol C J, Wade T, Assmann J J, Simpson G and Helfter C 2019 Illumination geometry and flying height influence surface reflectance and NDVI derived from multispectral UAS imagery *Drones* 3 55
- [128] Wookey P A, Aerts R, Bardgett R D, Baptist F, Bråthen K A, Cornelissen J H C, Gough L, Hartley I P, Hopkins D W and Lavorel S 2009 Ecosystem feedbacks and cascade processes: understanding their role in the responses of arctic and alpine ecosystems to environmental change *Glob. Change Biol.* 15 1153–72
- [129] Van Der Wal R and Brooker R W 2004 Mosses mediate grazer impacts on grass abundance in Arctic ecosystems *Funct. Ecol.* 18 77–86
- [130] Theodose T A and Bowman W D 1997 The influence of interspecific competition on the distribution of an alpine graminoid: evidence for the importance of plant competition in an extreme environment Oikos 79 101–14
- [131] Tomassen H B M, Smolders A J P, Limpens J, Lamers L P M and Roelofs J G M 2004 Expansion of invasive species on ombrotrophic bogs: desiccation or high N deposition? *J. Appl. Ecol.* 41 139–50
- [132] Beamish A, Raynolds M K, Epstein H, Frost G V, Macander M J, Bergstedt H, Bartsch A, Kruse S, Miles V and Tanis C M 2020 Recent trends and remaining challenges for optical remote sensing of Arctic tundra vegetation: a review and outlook *Remote Sens. Environ.* 246 111872
- [133] Buchhorn M, Raynolds M K and Walker D A 2016
 Influence of BRDF on NDVI and biomass estimations of Alaska Arctic tundra *Environ. Res. Lett.* 11 125002
- [134] Pla M, Bota G, Duane A, Balagué J, Curcó A, Gutiérrez R and Brotons L 2019 Calibrating Sentinel-2 imagery with multispectral UAV derived information to quantify damages in mediterranean rice crops caused by western swamphen (Porphyrio Porphyrio) Drones 3 45

- [135] Kattenborn T, Lopatin J, Förster M, Braun A C and Fassnacht F E 2019 UAV data as alternative to field sampling to map woody invasive species based on combined Sentinel-1 and Sentinel-2 data *Remote Sens*. *Environ.* 227 61–73
- [136] Immitzer, Markus, Clement Atzberger, and Tatjana Koukal 2012 Tree species classification with random forest using very high spatial resolution 8-Band WorldView-2 satellite data *Remote Sens.* 4 2661–93
- [137] Fraser R H, Van Der Sluijs J and Hall R J 2017 Calibrating satellite-based indices of burn severity from UAV-derived metrics of a burned boreal forest in NWT, Canada *Remote Sens.* 9 279
- [138] Pla M, Duane A and Brotons L 2017 Potencial De Las Imágenes UAV Como Datos De Verdad Terreno

Para La Clasificación De La Severidad De Quema De Imágenes Landsat: aproximaciones a Un Producto Útil Para La Gestión Post Incendio *Rev. Teledetección* **49** 91–102

- [139] Leitão P J, Schwieder M, Pötzschner F, Pinto J R R, Teixeira A M C, Pedroni F, Sanchez M, Rogass C, Van Der Linden S and Bustamante M M C 2018 From sample to pixel: multi-scale remote sensing data for upscaling aboveground carbon data in heterogeneous landscapes *Ecosphere* 9 e02298
- [140] Sivertson A H, Stodle D, Solbo S and Storvold R A new airborne remote sensing platform at Svalbard Poster Session Presented at Svalbard Science Conf.: Svalbard in a pan–Arctic Perspective (Oslo, Norway, 5–6 November 2019)