



Genetics polymorphism of poplars from Moscow region based on high-throughput sequencing of ITS

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Poplars are widely used in landscaping of Moscow due to the ability to effectively purify the air from harmful impurities and to release a large amount of oxygen. The genus *Populus* is characterized by a high level of intraspecies polymorphism, as well as the presence of natural interspecies hybrids. The aim of our work was to evaluate the genetic diversity of poplars, which are growing on the territory of Moscow city by high-throughput sequencing of internal transcribed spacers of 45S rRNA genes (ITS sequences). Sequencing of ITS of 40 poplar plants was performed on Illumina platform (MiSeq) and about 3 000 reads were obtained for each sample in average. Bioinformatics analysis was performed using CLC Genomics Workbench tool. The involved set of poplars had a high level of genetic diversity – the number of single nucleotide polymorphisms (SNPs) detected in each genotype relative to the reference ITS1 and ITS2 sequences of *P. trichocarpa* varying from 4 to 44. We showed that even trees which had been planted on the same territory and, probably, at the same time had significant genetic differences. It can be speculated that highly polymorphic plant material was used for planting poplars in Moscow. For some sites with SNPs, several variants of nucleotides were found in the same individual and the ratio of SNPs was different. We assume that close to 50/50 ratio is observed in interspecific hybrids due to genetic differences in the ITS sequences between maternal and paternal genotypes. For SNPs with a predominance of one of the variants, the presence of paralogues among numerous genomic copies of ITS sequences is more likely. The results of our work can provide a framework for molecular genetic markers application with the purpose of *Populus* species and interspecific hybrids identification, determination the origin of a number of natural hybrids, and monitoring the diversity of genus *Populus* in the Moscow city.

Key words: *Populus*; poplar; Moscow; high-throughput sequencing; ITS; polymorphism; genetic diversity.

Генетический полиморфизм тополей Московского региона на основе высокопроизводительного секвенирования ITS-последовательностей

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Тополь широко используется в озеленении Москвы благодаря способности эффективно очищать воздух от вредных примесей и выделять большое количество кислорода. Роду Тополь (*Populus*) свойствен высокий уровень внутривидового полиморфизма, а также наличие естественных межвидовых гибридов. Целью настоящей работы была оценка генетического разнообразия тополей, растущих на территории города Москвы, с использованием высокопроизводительного секвенирования внутренних транскрибируемых спейсеров генов 45S рРНК (ITS-последовательностей). На платформе Illumina (MiSeq) проведено секвенирование ITS-последовательностей 40 растений тополя и в среднем получено около 3000 прочтений для каждого образца. Биоинформатическая обработка данных проведена с использованием программы CLC Genomics Workbench. Исследованная выборка тополей имела высокий уровень генетического разнообразия: число выявленных в каждом генотипе однонуклеотидных полиморфизмов (SNP) относительно референсных последовательностей ITS1 и ITS2 *P. trichocarpa* варьировало от 4 до 44. Показано, что даже деревья, посаженные на одной территории и, вероятно, в одно время, значительно различаются генетически. Можно предположить, что при посадке тополей в Москве использовался крайне полиморфный растительный материал. Для некоторых сайтов с SNP у одного и того же индивидуума выявлено несколько вариантов нуклеотидов, соотношение которых было различным. Мы предполагаем, что соотношение, близкое к 50/50, наблюдается в межвидовых гибридах и является следствием генетических различий в ITS-последовательностях между материнским и отцовским

генотипами. Для SNP с преобладанием одного из вариантов вероятнее наличие паралогов среди многочисленных геномных копий ITS-последовательностей. Результаты работы закладывают основу для применения молекулярно-генетических маркеров с целью идентификации видов и межвидовых гибридов тополя, определения происхождения ряда естественных гибридов, а также мониторинга разнообразия представителей рода *Populus*, растущих на территории города Москвы.

Ключевые слова: *Populus*; тополь; Москва; высокопроизводительное секвенирование; ITS; полиморфизм; генетическое разнообразие.

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Moscow is one of the largest megalopolises of the world with a developed infrastructure, in which there are more than 12 million inhabitants, that is associated with an unfavorable ecology in the city. To improve the situation, effective landscaping of the city is necessary. Poplar is actively used in the landscaping of Moscow due to the ability to purify the air from pollutants, and release a large amount of oxygen.

Genus *Populus*, according to the Eckenwalder classification (Eckenwalder, 1996), includes 29 species predominantly distributed in the Northern hemisphere. Poplars are dioecious wind-pollinated plants that leads to high intraspecies diversity (Rae et al., 2007). It is known that various species of poplar are easily crossed forming natural interspecific hybrids (Roe et al., 2014; Jiang et al., 2016) that poses difficulties in identifying their taxonomic status. Genome of *P. trichocarpa* was sequenced in 2006 being the first genome of a tree (Tuskan et al., 2006). It is shown that the use of nucleotide sequences of internal transcribed spacers (ITS) of 45S ribosomal RNA (rRNA) genes (Hamzeh, Dayanandan, 2004) is efficient for genetic polymorphism evaluation, taxonomic classification, and determination of phylogenetic relationships in poplars. The ITS region includes highly variable ITS1 and ITS2 sequences located on both sides of highly conserved sequence encoding 5.8S rRNA. ITS sequences, unlike chloroplast and mitochondrial markers, are inherited from both parents and have high variability, while the procedure for their amplification is standardized (Poczai, Hyvonen, 2010). All of the above promotes the active use of ITS sequences for plant barcoding (Li et al., 2011).

ITS sequences are represented by many copies in a genome and different ITS paralogs may be present in one individual that requires special attention in data analysis and may even hinder obtaining of reliable data by Sanger sequencing (Hollingsworth et al., 2011). High-throughput sequencing can overcome the mentioned above difficulties because hundreds of ITS are sequenced for one individual and sample preparation does not require cloning. In the present work, high-throughput sequencing of ITS was

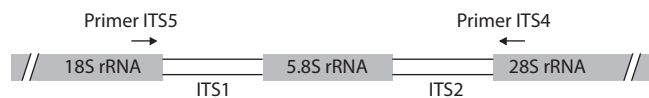
performed and genetic polymorphism of poplars growing on the territory of Moscow city was evaluated.

Materials and methods

Plant material was collected during the poplar flowering in the south and north of Moscow city. Young leaves were frozen in liquid nitrogen and stored at -70°C . DNA isolation was performed as described previously (Melnikova et al., 2014). The DNA quality was evaluated by electrophoresis on 1 % agarose gel. DNA concentration was measured on Qubit 2.0 fluorometer (Life Technologies, USA). For further work, a test set of DNA from 40 poplar plants was used.

Two-stage polymerase chain reaction (PCR) was used to prepare DNA libraries for high-throughput sequencing: the first stage included amplification of selected regions of the genome and the addition of universal sequences to the amplicons; at the second stage, the addition of sequences necessary for high-throughput sequencing and dual indexes for sample identification was performed. To amplify the ITS region, we used the primers proposed by Hsiao and White (White et al., 1990; Hsiao et al., 1995) (see Figure) with the universal adapters added. For the second PCR, Nextera XT v2 primers were used (Table 1). Primer design was proceeded according to the recommendations of the Illumina protocol (https://support.illumina.com/content/dam/illumina-support/documents/documentation/chemistry_documentation/16s/16s-metagenomic-library-prep-guide-15044223-b.pdf).

PCR for DNA library preparation was performed on Geneamp 9700 (Applied Biosystems, USA). The reaction volume was 25 μl and included 1x Tersus polymerase (Evrogen, Russia), 1x buffer for Tersus polymerase, 0.5 μM of each primer (Evrogen), 200 μM dNTP (Thermo Fisher Scientific, USA), and 40 ng of total DNA. The following reaction conditions were used: 3 min – 95°C ; 10 cycles for the first PCR and 35 cycles for the second PCR: 15 s – 95°C , 30 s – 58°C , and 1 min – 72°C ; 3 min – 72°C . The quality and concentration of 40 DNA libraries were evaluated on Agilent 2100 bioanalyzer (Agilent Technologies) and



Primer localization for amplification of ITS sequences of 45S rRNA gene.

Qubit 2.0 fluorometer (Life Technologies). DNA libraries were sequenced on a MiSeq sequencer (Illumina, USA) using MiSeq Reagent Kit v2 (500 cycles). The reading length was 250 nucleotides from each side for the target sequence; double indexes (Multiplexed Paired-End Run) were also sequenced.

The CLC Genomics Workbench software package (Qiagen, USA) was used for bioinformatics analysis of the data. The reads were mapped to ITS sequence of *P. trichocarpa* (GenBank: AJ006440.1), the genome of which is the reference one for *Populus*. The parameters were as follows: window length – 11, maximum number of gaps and mismatches – 2, minimum average quality of surrounding bases – 15, minimum quality of central base – 20, minimum coverage – 500, minimum paired-end coverage – 0, maximum coverage – 20 000, minimum variant frequency – 20 % or 50 reads.

Results

We performed high-throughput sequencing of ITS of 40 poplar plants growing on the territory of Moscow city. Sequence length was 250 nucleotides (paired-end reads), and, on average, about 3 000 reads were obtained for each sample. A bioinformatics analysis of the ITS sequences was carried out. The results are presented in Table 2 and Supplementary Materials¹.

The investigated set of trees was characterized by a high level of genetic diversity, the number of detected single nucleotide polymorphisms (SNPs) varied from 4 to 44 relative to ITS sequences of *P. trichocarpa* (GenBank: AJ006440.1). One of the subgroups of trees (numbers 17–28) had been planted in one territory and, probably, at the same time on both sides of the pedestrian road. Table 2 shows even this group of plants to be extremely heterogeneous – the number of detected SNPs varied from 6 to 44.

For some sites with SNPs, more than one nucleotide variant was detected. For these SNPs, in some cases, the ratio of allelic variants was close to 50/50, while in other cases, the distribution was unequal (Supplementary materials). It might be assumed that the 50/50 ratio is observed in hybrids and is a result of genetic differences in ITS sequences between paternal and maternal plants. For SNPs with a significant prevalence of one nucleotide variant over another, polymorphism within numerous copies of ITS in the genome is more likely.

Table 1. Primer sequences for DNA library preparation

Primer	Nucleotide sequence
ITS5_IIIu_F	TCGTCGGCAGCGTCAGATGTGTATAAGAGA CAGTCGTAACAAGGTTTCCGTAGGTG
ITS4_IIIu_R	GTCTCGTGGGCTCGGAGATGTGTATAAGAG ACAGTCCTCCGCTTATTGATATGC
Nextera XT v2 (i7)	CAAGCAGAAGACGGCATACGAGAT[i7] GTCTCGTGGGCTCGG
Nextera XT v2 (i5)	AATGATACGGCGACCACCGAGATCTACAC [i5]TCGTCGGCAGCGTC

Note: Primer sequences, which are necessary for amplification of ITS, are italicized. i7 and i5 are the sequences for dual indexing of samples.

Discussion

Poplar is a model object for biological research in trees (Jansson, Douglas, 2007). Over the last decades, numerous approaches have been developed and applied for the analysis of poplar genome, the study of interaction between genotype and environment, and the identification of inter- and intraspecific polymorphism in *Populus* (Jansson et al., 2010; Melnikova et al., 2017). Morphology analysis is actively used in studies of poplars growing on the territory of Moscow city and the Moscow region. High heterogeneity of poplar populations in Moscow and widespread distribution of interspecific hybrids were shown (Kostina, Nasimovich, 2014; Kostina et al., 2017).

In addition to morphological features, the use of molecular markers is effective for plant diversity evaluation (Melnikova et al., 2009–2011; Khadeeva et al., 2011; Bolsheva et al., 2015). In our work, we first applied high-throughput sequencing of ITS to assess the genetic diversity of poplars in Moscow. ITS sequences were already used to study the polymorphism and barcoding of poplar species growing in western China and the number of detected SNPs (38) was high (Feng et al., 2013); that is comparable to the obtained by us data. It should be noted that high-throughput sequencing of ITS performed in the present work allowed us to obtain a much more complete picture of the genetic polymorphism of poplars growing in Moscow by contrast to Sanger sequencing. Thus a high level of genetic diversity of the studied plants was revealed. It can be assumed that such heterogeneous populations of poplars are highly adaptive and have the advantage of surviving in ecologically unfavorable urban conditions. We also showed that while planting poplars in Moscow, an extremely polymorphic plant material was probably used, and even trees planted at the same time in one limited territory were considerably genetically different.

The results of our work lay the foundation for the development of molecular markers for poplars species and interspecific hybrids of poplars growing on the territory of Moscow identification, as well as for determination of the origin of a number of natural hybrids. In addition, recent studies showed that poplar sex is genetically determined

¹ Supplementary Materials are available in the online version of the paper: <http://www.bionet.nsc.ru/vogis/download/pict-2018-22/appx8.pdf>

Table 2. Single nucleotide polymorphisms of poplars growing in Moscow based on ITS sequence

No.	Number			No.	Number		
	paired-end reads	SNPs*	SNPs with several nucleotide variants		paired-end reads	SNPs*	SNPs with several nucleotide variants
1	4886	11	0	21	2530	7	2
2	4447	16	11	22	2169	10	8
3	3274	8	3	23	1606	6	2
4	4465	4	0	24	2740	16	12
5	7812	30	13	25	4463	44	37
6	5920	29	13	26	2586	9	7
7	1329	6	2	27	2619	10	8
8	1685	7	5	28	3086	39	37
9	4863	16	2	29	2727	10	8
10	321	10	10	30	2443	10	8
11	3171	13	9	31	1482	8	6
12	2597	18	6	32	4008	39	31
13	2829	10	8	33	2722	25	21
14	2740	9	6	34	3017	9	5
15	2567	9	7	35	1784	9	6
16	3908	25	21	36	2916	18	15
17	6592	30	14	37	2944	12	10
18	2150	8	6	38	1117	8	6
19	2373	9	7	39	1186	8	6
20	2163	10	8	40	1217	22	20

* ITS sequences of *P. trichocarpa* (GenBank: AJ006440.1) were used as references.

and only a small percentage of trees with recombination in the sex-associated genome region could change the sex (Geraldès et al., 2015; Borkhert et al., 2017; McKown et al., 2017). These data open up new opportunities for molecular marker development so as to use in the landscaping only male poplars, which do not produce fluff, while barcoding using ITS will allow evaluation of polymorphism and maintenance the diversity of populations adaptive to unfavorable urban conditions.

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Conflict of interest

The authors declare no conflict of interest.

References

Bolsheva N.L., Zelenin A.V., Nosova I.V., Amosova A.V., Samatadze T.E., Yurkevich O.Y., Melnikova N.V., Zelenina D.A., Volkov A.A., Muravenko O.V. The diversity of karyotypes and genomes within section *Syllinum* of the genus *Linum* (Linaceae) revealed by

molecular cytogenetic markers and RAPD analysis. *PLoS One*. 2015;10(4):e0122015. DOI 10.1371/journal.pone.0122015.

Borkhert E.V., Dmitriev A.A., Krasnov G.S., Bolsheva N.L., Kudryavtseva A.V., Melnikova N.V. Sex associated single nucleotide polymorphisms in the poplar genome. *Acta Naturae*. 2017;Spec. Iss.1(9):61. (in Russian)

Eckenwalder J.E. Systematics and evolution of *Populus*. In: Stettler R.F., Bradshaw H.D., Heilman P.E., Hinckler T.M. (Eds.). *Biology of Populus and its Implications for Management and Conservation*. Ottawa: Can. Government Publ., 1996;7-32.

Feng J., Jiang D., Shang H., Dong M., Wang G., He X., Zhao C., Mao K. Barcoding poplars (*Populus* L.) from western China. *PLoS One*. 2013;8(8):e71710. DOI 10.1371/journal.pone.0071710.

Geraldès A., Hefer C.A., Capron A., Kolosova N., Martínez-Núñez F., Soolanayakanahally R.Y., Stanton B., Guy R.D., Mansfield S.D., Douglas C.J., Cronk Q.C. Recent Y chromosome divergence despite ancient origin of dioecy in poplars (*Populus*). *Mol. Ecol*. 2015; 24(13):3243-3256. DOI 10.1111/mec.13126.

Hamzeh M., Dayanandan S. Phylogeny of *Populus* (Salicaceae) based on nucleotide sequences of chloroplast *trnT-trnF* region and nuclear rDNA. *Am. J. Bot.* 2004;91(9):1398-1408. DOI 10.3732/ajb.91.9.1398.

Hollingsworth P.M., Graham S.W., Little D.P. Choosing and using a plant DNA barcode. *PLoS One*. 2011;6(5):e19254. DOI 10.1371/journal.pone.0019254.

Hsiao C., Chatterton N.J., Asay K.H., Jensen K.B. Phylogenetic relationships of the monogenomic species of the wheat tribe, Triticeae (Poaceae), inferred from nuclear rDNA (internal transcribed spacer) sequences. *Genome*. 1995;38(2):211-223.

- Jansson S., Bhalerao R.P., Groover A.T. Genetics and Genomics of *Populus*. New York: Springer-Verlag, 2010. DOI 10.1007/978-1-4419-1541-2.
- Jansson S., Douglas C.J. *Populus*: a model system for plant biology. *Annu. Rev. Plant Biol.* 2007;58:435-458. DOI 10.1146/annurev.arplant.58.032806.103956.
- Jiang D., Feng J., Dong M., Wu G., Mao K., Liu J. Genetic origin and composition of a natural hybrid poplar *Populus × jrtyschensis* from two distantly related species. *BMC Plant Biol.* 2016;16:89. DOI 10.1186/s12870-016-0776-6.
- Khadeeva N.V., Goryunova S.V., Kochumova A.A., Yakovleva Y.Y., Mel'nikova N.V., Zholobova O.O., Korotkov O.I., Kudryavtsev A.M. Genetic monitoring of populations of *Matthiola fragrans* (Bunge) using RAPD and AFLP analysis. *Biol. Bull.* 2011;38(4):325-331. DOI 10.1134/S1062359011040078.
- Kostina M.V., Nasimovich J.A. On the taxonomy of *Populus* L. II. The importance of fruit characters for identification of cultivated and adventive species in the Moscow region. *Byulleten' Moskovskogo Obshchestva Ispytateley Prirody = Bulletin of the Moscow Society of Naturalists. Biol. Ser.* 2014; 119(5):74-79. (in Russian)
- Kostina M.V., Puzyryov A.N., Nasimovich J.A., Parshevnikova M.S. Representatives of the sections *Aigeiros* Duby and *Tacamahaca* Spach (genus *Populus* L., Salicaceae) and their hybrids in cities of central and eastern European Russia. *Skvortsovia.* 2017;3(3):97-119.
- Li D.Z., Gao L.M., Li H.T., Wang H., Ge X.J., Liu J.Q., Chen Z.D., Zhou S.L., Chen S.L., Yang J.B., Fu C.X., Zeng C.X., Yan H.F., Zhu Y.J., Sun Y.S., Chen S.Y., Zhao L., Wang K., Yang T., Duan G.W. Comparative analysis of a large dataset indicates that internal transcribed spacer (ITS) should be incorporated into the core barcode for seed plants. *Proc. Natl. Acad. Sci. USA.* 2011;108(49):19641-19646. DOI 10.1073/pnas.1104551108.
- McKown A.D., Klapste J., Guy R.D., Soolanayakanahally R.Y., La Mantia J., Porth I., Skyba O., Unda F., Douglas C.J., El-Kassaby Y.A., Hamelin R.C., Mansfield S.D., Cronk Q.C.B. Sexual homomorphism in dioecious trees: extensive tests fail to detect sexual dimorphism in *Populus*. *Sci. Rep.* 2017;7(1):1831. DOI 10.1038/s41598-017-01893-z.
- Melnikova N.V., Borhert E.V., Martynov S.P., Okuneva I.B., Molkano O.I., Upelnik V.P., Kudryavtsev A.M. Molecular genetic marker-based approaches to the verification of lilac *Syringa vulgaris* L. *in vitro* germplasm collections. *Russ. J. Genet.* 2009;45(1):85-90. DOI 10.1134/S1022795409010128.
- Melnikova N.V., Borkhert E.V., Snezhkina A.V., Kudryavtseva A.V., Dmitriev A.A. Sex-specific response to stress in *Populus*. *Front. Plant Sci.* 2017;8:1827. DOI 10.3389/fpls.2017.01827.
- Melnikova N.V., Kononov F.A., Kudryavtsev A.M. Long terminal repeat retrotransposon *Jeli* provides multiple genetic markers for common wheat (*Triticum aestivum*). *Plant Genet. Resour.* 2011;9(2):163-165. DOI 10.1017/S1479262111000487.
- Melnikova N.V., Kudryavtseva A.V., Zelenin A.V., Lakunina V.A., Yurkevich O.Y., Speranskaya A.S., Dmitriev A.A., Krintsina A.A., Belenikin M.S., Uroshlev L.A., Snezhkina A.V., Sadritdinova A.F., Koroban N.V., Amosova A.V., Samatadze T.E., Guzenko E.V., Lemesh V.A., Savilova A.M., Rachinskaia O.A., Kishlyan N.V., Rozhmina T.A., Bolsheva N.L., Muravenko O.V. Retrotransposon-based molecular markers for analysis of genetic diversity within the genus *Linum*. *BioMed Res. Int.* 2014;2014:231589. DOI 10.1155/2014/231589.
- Melnikova N.V., Mitrofanova O.P., Liapounova O.A., Kudryavtsev A.M. Global diversity of durum wheat *Triticum durum* Desf. for alleles of gliadin-coding loci. *Russ. J. Genet.* 2010;46(1):43-49. DOI 10.1134/S1022795410010072.
- Poczai P., Hyvonen J. Nuclear ribosomal spacer regions in plant phylogenetics: problems and prospects. *Mol. Biol. Rep.* 2010;37(4):1897-1912. DOI 10.1007/s11033-009-9630-3.
- Roe A.M., Street N.R., Rodríguez-Acosta M. *Populus* Trees. In: Kole C. (Ed.). *Genome Mapping and Molecular Breeding in Plants. Vol. 7. Forest Trees.* Berlin; Heidelberg: Springer-Verlag, 2007;1-28. DOI 10.1007/978-3-540-34541-1_1.
- Roe A.D., MacQuarrie C.J., Gros-Louis M.C., Simpson J.D., Lamarche J., Beardmore T., Thompson S.L., Tanguay P., Isabel N. Fitness dynamics within a poplar hybrid zone. II. Impact of exotic sex on native poplars in an urban jungle. *Ecol. Evol.* 2014;4(10):1876-1889. DOI 10.1002/ece3.1028.
- Tuskan G.A., Difazio S., Jansson S., Bohlmann J., Grigoriev I., Hellsten U., Putnam N., Ralph S., Rombauts S., Salamov A., Schein J., Sterck L., Aerts A., Bhalerao R.R., Bhalerao R.P., Blaudez D., Boerjan W., Brun A., Brunner A., Busov V., Campbell M., Carlson J., Chalot M., Chapman J., Chen G.L., Cooper D., Coutinho P.M., Couturier J., Covert S., Cronk Q., Cunningham R., Davis J., Degroove S., DeJardin A., Depamphilis C., Detter J., Dirks B., Dubchak I., Duplessis S., Ehrling J., Ellis B., Gendler K., Goodstein D., Gribskov M., Grimwood J., Groover A., Gunter L., Hamberger B., Heinze B., Helariutta Y., Henrissat B., Holligan D., Holt R., Huang W., Islam-Faridi N., Jones S., Jones-Rhoades M., Jorgensen R., Joshi C., Kangasjarvi J., Karlsson J., Kelleher C., Kirkpatrick R., Kirst M., Kohler A., Kalluri U., Larimer F., Leebens-Mack J., Leple J.C., Locascio P., Lou Y., Lucas S., Martin F., Montanini B., Napoli C., Nelson D.R., Nelson C., Nieminen K., Nilsson O., Pereda V., Peter G., Philippe R., Pilate G., Poliakov A., Razumovskaya J., Richardson P., Rinaldi C., Ritland K., Rouze P., Ryabov D., Schmutz J., Schrader J., Segerman B., Shin H., Siddiqui A., Sterky F., Terry A., Tsai C.J., Uberbacher E., Unneberg P., Vahala J., Wall K., Wessler S., Yang G., Yin T., Douglas C., Marra M., Sandberg G., Van de Peer Y., Rokhsar D. The genome of black cottonwood, *Populus trichocarpa* (Torr. & Gray). *Science.* 2006;313(5793):1596-1604. DOI 10.1126/science.1128691.
- White T.J., Bruns T., Lee S., Taylor J.W. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis M.A., Gelfand D.H., Sninsky J.J., White T.J. (Eds.). *PCR Protocols: A Guide to Methods and Applications.* New York: Acad. Press, 1990;315-322. DOI 10.1016/B978-0-12-372180-8.50042-1.