Four distinct trajectories of tau deposition identified in Alzheimer's disease

Jacob W. Vogel[1,], **Alexandra L. Young**[2], **Neil P. Oxtoby**[3,4], **Ruben Smith**[5,6], **Rik Ossenkoppele**[5,7], **Olof T. Strandberg**[5], **Renaud La Joie**[8], **Leon M. Aksman**[3,9], **Michel J Grothe**[10,11], **Yasser Iturria-Medina**[1], **the Alzheimer's Disease Neuroimaging Initiative**[*], **Michael J. Pontecorvo**[12], **Michael D. Devous**[12], **Gil D. Rabinovici**[8,13], **Daniel C. Alexander**[3,4], **Chul Hyoung Lyoo**[14], **Alan C. Evans**[1], **and**

Oskar Hansson[5,15]

1 Montreal Neurological Institute, McGill University, Montréal, QC, Canada

2 Department of Neuroimaging, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, UK.

3 Centre for Medical Image Computing, University College London, London, WC1E 6BT, UK

4 Department of Computer Science, University College London, London, WC1E 6BT, UK

5 Clinical Memory Research Unit, Lund University, Lund, Sweden

6 Department of Neurology, Skåne University Hospital, Lund, Sweden

7 Alzheimer Center Amsterdam, Department of Neurology, Amsterdam Neuroscience, Vrije Universiteit Amsterdam, Amsterdam UMC, Amsterdam, The Netherlands

8 Memory and Aging Center, Department of Neurology, Weill Institute for Neurosciences, University of California, San Francisco, San Francisco, CA

9 Department of Medical Physics and Biomedical Engineering, University College London, London, WC1E 6BT, UK

10 Wallenberg Centre for Molecular and Translational Medicine, University of Gothenburg, Gothenburg, Sweden

11 Instituto de Biomedicina de Sevilla, Hospital Universitario Virgen del Rocío/CSIC/Universidad de Sevilla, Seville, Spain

12 Avid Radiopharmaceuticals, Philadelphia, PA, USA

13 Department of Radiology and Biomedical Imaging; University of California, San Francisco, San Francisco, CA

14 Departments of Neurology, Gangnam Severance Hospital, Yonsei University College of Medicine, Seoul, Republic of Korea

15 Memory Clinic, Skåne University Hospital, Malmö, Sweden

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Abstract: Alzheimer's disease (AD) is characterized by spread of tau pathology throughout the cerebral cortex. The spreading pattern was thought to be fairly consistent across individuals, though recent work has demonstrated substantial variability in the AD population. Using tau-PET scans from 1612 individuals, we identified four distinct spatiotemporal trajectories of tau pathology, ranging in prevalence from 18 to 33%. We replicated previously described limbic-predominant and medial temporal lobe-sparing patterns, while also discovering posterior and lateral temporal patterns resembling atypical clinical variants of AD. These "subtypes" were stable during longitudinal follow-up, and were replicated in a separate sample using a different radiotracer. The subtypes presented with distinct demographic and cognitive profiles, and differing longitudinal outcomes. Additionally, network diffusion models implicated that pathology originates and spreads through distinct corticolimbic networks in the different subtypes. Together, our results suggest variation in tau pathology is common and systematic, perhaps warranting a re-examination of the notion of "typical AD", and a revisiting of tau pathological staging.

Correspondence: *jacob.vogel@mail.mcgill.ca; oskar.hansson@med.lu.se*

Introduction

Alzheimer's disease (AD) is the leading cause of dementia worldwide and prevalence is expected to double in the next twenty years[1]. At autopsy, AD presents with diffuse extracellular and neuritic amyloid-β (Aβ) plaques, and intracellular neurofibrillary tangles and neuropil threads of hyperphosphorylated tau, along with extensive neurodegeneration[2,3]. Leading hypotheses have postulated these two hallmark proteins, Aβ and tau, either alone or in combination, are causative agents in disease etiology and progression[4,5]. Cortical tau colocalizes with cortical atrophy and predicts future neurodegeneration[6], while the appearance of tau in specific cognitive networks leads to domain-specific cognitive impairments[7]. Recently, the focus of treatment discovery has shifted to tau, and numerous therapeutic interventions are currently undergoing research and development. A better understanding of tau pathophysiology is therefore of imminent need in order to aid development of these interventions.

51   Tau tangles are thought to exhibit a stereotypical pattern of cortical spread, which has been formal-
52   ized into the Braak staging system[8,9]. The six Braak stages describe the first appearance of cortical tau
53   tangles in the transentorhinal cortex, subsequent spread throughout the medial and basal temporal lobes,
54   then into neocortical associative regions, and finally into the unimodal sensory and motor cortex[9]. While
55   this stereotyped progression was derived from histopathological staining at autopsy, tau can now be
56   measured *in vivo* in the human brain using positron emission tomography (PET). Early tau-PET imaging
57   studies described average spatial patterns that have mostly converged with the Braak staging system[10,11].
58   However, many examples have emerged of individual tau patterns that do not fit neatly into the
59   Braak staging system. A medial temporal lobe (MTL)-sparing phenotype with extensive cortical tau burden
60   but limited MTL burden has been described, as well as a limbic-predominant phenotype with most promi-
61   nent tau pathology in limbic and medial temporal cortex, which were found to be associated with specific
62   patient profiles[12–14]. In addition, clinical variants of AD have been described that exhibit specific patterns
63   of pathology that deviate from the Braak staging scheme[15], e.g., posterior cortical atrophy (PCA) [16], logo-
64   penic primary progressive aphasia (lvPPA)[17], and others[18]. These latter clinical variants of AD are relatively
65   uncommon and most frequently associated with early-onset AD, but represent another example of atypi-
66   cal tau patterning.
67   Taken together, the examples above suggest that, while the Braak staging system appears to be a
68   good description of tau spreading at the population level, it does not account for systematic variability at
69   the individual level. Variation in tau patterning may be indicative of distinct underlying neurobiology [19,20],
70   which may affect treatment response. Different subtypes may also have distinct rates and profiles of cog-
71   nitive decline [21,22], potentially affecting clinical trials. For these reasons, a systematic description of varia-
72   tion in AD pathological spread is needed. Previous studies have provided invaluable information toward
73   this effort [12,13,23–26], but carry certain limitations. Pathology studies, for example, are limited by spatial
74   sampling and semi-quantitation. Neuroimaging studies overcome some of those limitations, but often use
75   non-specific measurements, and rely on methods designed to parse spatial rather than spatiotemporal
76   variation.
77   Here we present a systematic characterization of heterogeneity in tau patterning in AD. We
78   amassed the largest and most diverse sample of tau-PET data to date (n=2324), covering the full clinical
79   spectrum from asymptomatic through mild cognitive impairment (MCI) to AD dementia, allowing unprec-
80   edented power to detect and characterize AD subtypes. We fit this data using the Subtype and Stage In-
81   ference (SuStaIn) model, a paradigm-shifting algorithm that combines disease progression modeling with
82   traditional clustering to achieve probabilistic spatiotemporal partitioning and classification[23]. SuStaIn re-
83   quires only cross-sectional datasets to automatically detect multiple spatiotemporal trajectories, and it
84   provides probabilistic and quantitative information for individualized inference. We apply SuStaIn to our
85   multi-cohort sample of tau-PET data to discover systematic spatiotemporal variation in tau spreading. We
86   validate the subtypes across different PET radiotracers, and we validate the progression patterns using
87   serial longitudinal tau-PET data.

88
89   **Results**
90   We compiled an initial sample of 1667 individuals with flortaucipir-PET tau images, spanning five separate
91   cohorts. 1143 individuals were identified as either cognitively normal (n=707), or showed biomarker evi-
92   dence for Aβ pathology (Aβ+ MCI, n=223; Aβ+ AD dementia, n=213), and were used as a discovery sample
93   for subsequent analysis. Demographic information and cross-cohort comparisons can be found in Table
94   S1.

95
96   **Spatiotemporal subtypes of Alzheimer's disease.** We applied the SuStaIn algorithm (Extended Data Fig.
97   S1a) to the 1143 flortaucipir-PET images in order to extract distinct spatiotemporal trajectories of tau
98   spreading. As expected, many individuals (n=700; 61%) did not demonstrate any abnormal tau-PET signal,
99   and were therefore automatically assigned to a tau-negative group (S0) (see Supplementary Note 1). Us-
100   ing cross-validation, we determined a four-subtype solution to best represent the remaining data (n=443;

See Methods, Extended Data Fig. S1c-f). The four-subtype model was applied to probabilistically assign individuals to one of 30 progressive stages along one of the four subtype trajectories (Fig 1).

The distribution of clinical diagnoses across stages and subtypes can be found in Extended Data Fig. S2f,g,i. 145 (32.7%) individuals exhibited a limbic-predominant phenotype, with a Braak-like spatial progression across SuStaIn stages (S1: Limbic). An additional 79 individuals (17.8%) expressed a parietal-dominant and MTL-sparing phenotype, where early precuneus binding accumulated across temporoparietal and frontal cortex, but with relative sparing of the MTL (S2: MTL-Sparing). The third subtype composed 135 (30.5%) individuals with a predominant posterior occipitotemporal phenotype, involving early occipital lobe binding and gradual anterior progression across SuStaIn stage (S3: Posterior). The remaining 84 (19.0%) individuals showed a temporoparietal phenotype with distinct left-sided lateralization, characterized by early left-temporal tau eventually spreading to parietal and frontal cortex across disease stage (S4: Lateral [L] Temporal). The differences highlight inconsistencies between tau-PET binding and pathological sequencing of specific brain regions found in previous studies, such as the hippocampus, lingual gyrus and insula[10,11,27], which exhibited different binding patterns across subtypes (Extended Data Fig. S3).

Stability of AD subtypes. While variation in subtype proportion was observed (and expected) across cohorts, all subtypes were represented across all cohorts (Extended Data Fig. S4). Most individuals fell neatly into the stereotypical progression of each subtype (Fig 1b), allowing a clean stepwise progression across tau abnormality events to be observed across each subtype population (Extended Data Fig. S5). 12% of individuals did not fall cleanly into any subtype due to having either too little or too much pathology, both of which are uninformative for subtype (Fig 1b, Extended Data Fig. S2d,h). In general, early stage and cognitively normal individuals were assigned to subtypes with less confidence, though median subtype probability neared 100% by SuStaIn stage 7 (Extended Data Fig. S2e), and by the MCI clinical stage (Extended Data Fig. S2h). This provides evidence that the earliest phases of each subtype may overlap, or that they are difficult to distinguish above measurement error. We further confirmed that the subtypes produced by SuStaIn were not driven by, or specific to, arbitrary user inputs relating to the anchoring of regional pseudotimes (Methods, Extended Data Fig. S6).

We next assessed whether the same subtypes could be derived within a separate replication sample of 469 individuals scanned with the RO948 tau-PET tracer. The replication cohort, BioFINDER II[28], is described in Supplementary Table S1. SuStaIn was run separately on these individuals, constraining the analysis to produce four subtypes. Three of the four replication subtypes greatly resembled those derived in the discovery sample (Fig 2). The only exception involved the S4: L Temporal subtype, which had a similar overall tau-PET pattern but involved right-sided rather than left-sided lateralization. Further analysis determined that this related to the smaller sample size rather than the differing radiotracer, and further suggested that the S4: L Temporal subtype has a consistent overall pattern but a high propensity for marked lateralization (see Supplemental Note 2, Extended Data Fig. S7).

Subtypes characterized by distinct clinical profiles. Next, we compared demographic, cognitive and genetic (i.e. APOE4 status) variables between the subtypes and the tau-negative S0 group (Table 1). Individuals across all four subtypes expressed worse MMSE and worse memory scores compared to S0 individuals. In addition, all subtypes except S1 (Limbic) had worse global cognitive composite scores, individuals across all subtypes except S2 (MTL-Sparing) were more likely to be *APOE4* carriers, and all subtypes except S4 (L Temporal) were more likely to be female, compared to S0 individuals. Compared to tau-negative individuals in S0, S1 and S3 were older, S2 exhibited poorer executive function, S2 and S3 exhibited poorer visuospatial function, and S4 had worse language scores.

Compared to other subtypes (i.e., other tau-positive individuals), individuals within the S1 (Limbic) subtype were more likely to be *APOE4* carriers, had less overall tau with a more right-sided pattern, and had better overall cognition, but worse memory relative to their overall cognition. S2 (MTL Sparing) individuals were younger, less likely to carry an *APOE4* allele, had more overall tau burden, had a more right-sided tau pattern and had worse relative executive function, compared to other subtypes. S4 (L Temporal) individuals had more overall tau with a more left-lateralized pattern. These individuals also trended at

152 having worse overall cognition, but had better relative memory and worse relative language scores com-
153 pared to other subtypes. Finally, individuals with the S3 (Posterior) subtype did not exhibit any significant
154 cognitive, demographic or *APOE4* differences compared to the other subtypes. These relationships (after
155 adjustment for demographics, diagnosis, cohort and SuStaIn stage) are described in Table 1 and visualized
156 in Extended Data Fig. S8.
157   Each individual was assigned a stage along their respective subtype trajectory. As expected, increas-
158 ing SuStaIn stage was associated with worse global cognition as measured with MMSE (r=0.54, p<0.0001;
159 Fig 3a). This relationship was consistent across all subtypes (S1: r = -0.51, S2: r = -0.53, S3: r = -0.64, S4: r =
160 -0.40, all p<0.001). A strong negative relationship between SuStaIn stage and age was also observed, such
161 that individuals at later SuStaIn stages tended to be younger (r = -0.59, p<0.0001). This relationship was
162 again consistent across all subtypes, though less prominent for S1 (S1: r = 0.20, S2: r = -0.68, S3: r = -0.64,
163 S4: r = -0.73, all p<0.05; Fig 3b). This inverse relationship was also present among individuals both 65 and
164 younger (n = 100, r = -0.43, p <0.0001) and individuals older than 65 (n = 342, r = -0.28, p < 0.0001), sug-
165 gesting the effect is not driven purely by early onset cases. Lateralization also increased with increasing
166 SuStaIn stage (Extended Data Fig. S9). However, despite trends in lateralization at higher SuStaIn stage,
167 many individuals were observed with a "reversed" lateralization compared to the group average tau lat-
168 eralization patterns for their subtype (Extended Data Fig. S9), suggesting lateralization to be at least par-
169 tially orthogonal with subtype.
170
171 **Cognitive prognosis of AD subtypes.** Longitudinal MMSE data was available for a subset of 697 individuals
172 (mean follow-up = 1.74 years from PET scan, sd = 0.64). Individuals with the S3 (Posterior) subtype had
173 significantly slower decline compared to all other subtypes independently (S1: t=2.03, p=0.043; S2: t=2.88,
174 p=0.004; S4: t=4.83, p<0.0001), as well as in a one vs all analysis (t=3.64,p=0.0003; Fig 3c). This finding
175 persisted across different clinical diagnoses (Fig 3d, Supplemental Table S2), and was confirmed through a
176 meta-analysis across the five cohorts, which also showed a significantly slower decline for the S3 (Posteri-
177 or) group (t=1.67,p=0.047; Fig 3e). Individuals with the S4 (L Temporal) subtype additionally showed
178 steeper cognitive decline compared to S1 (Limbic) subtype individuals (t=3.40, p=0.0008), and generally
179 showed faster decline compared to other subtypes in a one vs all analysis (t=-4.49,p<0.0001) and across
180 clinical diagnoses (Fig 3d, Supplementary Table S2). A meta-analysis once again confirmed a significant
181 overall effect (t=1.88,p=0.031; Fig 3e).
182
183 **Stability and progression of AD subtypes over time.** SuStaIn uses cross-sectional data to infer longitudinal
184 trajectories for the tau data, so evaluating how well longitudinal data fits the model is a key aspect of val-
185 idation. 519 individuals from the discovery sample also had follow-up flortaucipir-PET scans (mean follow-
186 up time = 1.42, sd = 0.58, years). Overall, 88.5% of individuals exhibited the same subtype at both baseline
187 and follow-up, or progressed from S0 into a subtype (Fig 3f). Stability when excluding individuals classified
188 as S0 at baseline (tau-positive stability) and follow-up was 83.9%. Stable individuals were classified with a
189 higher degree of confidence at baseline compared to individuals whose subtype changed at follow-up
190 (stable mean = 0.91, sd = 0.17; change mean = 0.74, sd = 0.27; t = 5.26, p < 0.0001; Fig 3g). Supplementary
191 Table S3 shows longitudinal tau-positive stability (i.e. excluding S0) when excluding individuals using vari-
192 ous subtype probability thresholds.
193   We next examined how SuStaIn stage changed over time for each subtype. Across the whole sample,
194 we observed significant yearly increase in SuStaIn stage (mean Δ/year = 0.8, t[148]=6.54, p<0.0001) (Fig
195 3h, Table S4), and a significant difference in mean annual rate of SuStaIn stage change was seen across
196 subtypes (details in Supplementary Note 3). The annual SuStaIn stage increased faster in S4 (L Temporal)
197 compared to S2 (MTL-Sparing) and S3 (Posterior) subtypes (Fig 3h, Supplementary Note 3). Younger age
198 (r=-0.22, p=0.006), but not higher baseline SuStaIn stage (r = 0.12, p=0.15), was associated with faster
199 annual change in stage. As a final validation, we used SuStaIn to forecast longitudinal rate of regional tau-
200 PET change at the individual level. On average, predictions were significantly better than chance for all
201 subtypes (S1 (Limbic): t[78]=5.00, p<0.0001; S2 (MTL-Sparing): t[52]=2.16, p=0.035; S3 (Posterior):
202 t[45]=3.05, p=0.0039; S4 (L Temporal): t[29]=4.93, p<0.0001; Fig 3i).

203
Subtype patterns resemble distinct cortico-limbic networks. Based on our previous work[29], we used net-
work diffusion models to examine the possibility that the observed subtype-specific tau spreading pat-
terns may be driven by spread through distinct networks. We found that an entorhinal cortex epicenter
was optimal for the S1 (Limbic) subtype tau pattern and strongly replicated the pattern of tau spreading
(r^2=0.70), but did not reproduce other subtype patterns nearly as well (S2: r^2=0.04; S3: r^2=0.41; S4:
r^2=0.37). Models using different epicenters substantially improved fit for these others subtypes (Fig
4a,b,e): best fitting models used the middle temporal gyrus (r^2=0.27) for S2 (MTL-Sparing), the fusiform
gyrus (r^2=0.59) for S3 (Posterior) and the inferior temporal gyrus (r^2=0.50) for S4 (L Temporal) (Fig 4c),
suggesting a possible predominance of these regions in secondary tau seeding for different subtypes.
Highly similar results were found using a different brain atlas and different connectivity data (Extended
Data Fig. S10). We further tracked how the best-fitting epicenter changed at higher disease stages, per-
haps reflecting participation of different regions as secondary seeding points with advancing disease pro-
gression (Fig 4d). All but the S2 (MTL-Sparing)_subtype exhibited MTL spreading in earlier stages, whereas
early stages of S2 involved parietal spread. Later stages involved secondary seeding in the temporal lobes,
as well as subtype-specific regions. Together, these results suggest that distinct tau patterns across differ-
ent subtypes may be driven in part by vulnerability of, or selective spread through, distinct temporal lobe
networks.

Discussion

For the last thirty years, the progression of tau pathology in AD has principally been described by a single
model of spatiotemporal evolution[8,9], despite frequent examples of nonconforming cases[12]. We show that
the cortical cascade of tau pathology is better described by a data-driven model including multiple spatio-
temporal patterns (Fig. 5). Importantly, our findings may reconcile atypical AD variants with common vari-
ations of typical AD into a single unified model of pathological progression. First, the model reaffirms the
existence of observed cortical-predominant and limbic-predominant pathological patterns as distinct sub-
types of tau progression, rather than phases along a continuum. In addition, the model also accounts for
the most frequently occurring atypical clinical variants of AD, PCA and lvPPA, as the extremes of regularly
occurring posterior and lateral-temporal AD subtypes. Together, our data align with a recent model[14] to
suggest variation in the pathological expression of AD along two orthogonal axes: subtype and severity,
the latter of which is strongly and inversely correlated with age (Fig 5). Given that no dominant pattern
emerged, our data suggest the existence of multiple common AD subtypes, challenging the notion that
there is such a pathological entity that can be described as "typical" AD. Rather, the spatial pattern of tau
spreading appears to vary along at least four archetypes, depending on factors such as age and genotype.
Therefore, we propose heterogeneity in AD is best represented as a quadrilateral axis (Fig 5).
        Our results are robust across datasets and radiotracers. We found individuals representing each
of four subtype patterns in each of the five contributing cohorts, and we reproduced a very similar set of
subtypes in a totally separate sample using a different radiotracer. Further, most individuals were confi-
dently assigned into one subtype pattern, which was consistent over time. The limbic subtype was the
most frequent, and presented with many characteristics typically associated with AD, including a greater
proportion of *APOE4* carriers, a strongly amnestic phenotype, and medial temporal pathology with a
Braak-like progression of tau spread. However, this subtype represented only a third of all tau-positive
cases in our dataset (though the earliest stages of three of the four subtypes featured prominent MTL
binding, Fig 4d). Our data suggest instead that, at older onset ages or earlier disease stages, the subtypes
may present with subtle differences that may be difficult to detect in the clinic, while at younger onset
ages or later stages, the more aggressive phenotype can amplify the distinct subtype expressions. The
existence of these phenotypes, if further validated, may necessitate a reform in pathological tau staging,
where key regions are surveyed to increase sensitivity to detect subtype-specific patterns.
        Many pioneering studies have noted variation in AD pathology. For example, limbic-predominant
and MTL-sparing phenotypes are contrasted against "typical" phenotypes that express tau pathology in

253 both the MTL and neocortex[12,13]. In contrast to this notion, we found a subtype of individuals expressing
254 both cortical and MTL tau exhibiting a more aggressive phenotype with marked lateralization, the latter
255 being a feature that has not been well characterized in histopathological studies of AD, which typically
256 assess only one hemisphere. In addition, our model allows the concurrence of MTL and cortical pathology
257 at later stages of several distinct progressions, perhaps suggesting that solely contrasting cortical and MTL
258 tau (e.g. [21,22]) may not be sufficient to describe AD heterogeneity. Indeed, while some spatial convergence
259 could be observed in our AD subtypes, particularly at early or late stages, subtle regional variation con-
260 sistently distinguishes individuals of one subtype from another.
261 We reproduce previous reports describing a strong negative correlation between age and tau pro-
262 gression[30–34], as well as previous reports that a younger age of onset of AD is associated with a more rapid
263 progression of tau pathology[35,36]. Interestingly, in our study, this phenomenon was observable across all
264 subtypes (Fig 3b). Previous work has noted that early-onset AD (EOAD) is more likely to present with an
265 atypical (i.e. nonamnestic) phenotype[37]. This may be a specific characteristic of EOAD. However, ours and
266 others studies[26,38,39] suggest that posterior or left-lateralized temporal binding are not uncommon across
267 the age spectrum, but our data suggest that the phenotype is more pronounced at earlier ages. There-
268 fore, atypical variants of AD may represent an accelerated and intensified manifestation of common AD
269 subtypes, though this will require further validation.
270 Our findings complement other supervised and unsupervised AD subtyping studies from the imaging
271 and pathology literature[12–14,21,22,26,38], though our analysis also produced some novel findings worth fur-
272 ther investigation. Despite the extreme of the posterior subtype being represented by PCA, an aggressive
273 disease variant, the posterior subtype overall demonstrated slower cognitive decline compared to all oth-
274 er subtypes. These individuals exhibited considerable tau pathology in posterior (including occipital) brain
275 regions, but also relatively less MTL and frontal binding. These findings, however, are in agreement with
276 pathology literature describing common variation in occipital tau pathology in both preclinical and symp-
277 tomatic AD [2,40–42]. These studies, variously surveying Brodmann areas 17, 18 and 19, find evidence for oc-
278 cipital lobe tau in 24-52% of sampled brains, including in cognitively normal individuals. Our study sug-
279 gests this population variation may indeed be systematic, and could be associated with a specific progres-
280 sion pattern. However, tau in the occipital lobe remains understudied, and future studies will be neces-
281 sary to validate the precise characteristics of this posterior subtype. It is still unclear if the posterior sub-
282 type is related to PCA beyond a shared predominance of posterior tau, though it may at least signify the
283 existence of a posterior cortical network selectively vulnerable to tau pathology.
284 Different manifestations of AD may represent subtle variations in the spread of pathology, or could
285 signal the influence of highly distinct processes relevant to treatment intervention. For example, a recent
286 pathology study found increased NFT pathology and neuronal loss in the cholinergic basal forebrain spe-
287 cifically in patients with a MTL-sparing phenotype, and that earlier disease onset was associated with
288 more NFT pathology in these subjects[43]. Furthermore, another recent study indicated that a targeted ba-
289 sal forebrain treatment could be most effective for patients with a MTL-sparing phenotype[44]. This re-
290 search may suggest a unique role of the basal forebrain in certain subtypes of AD. Meanwhile, APOE has
291 been consistently associated with limbic manifestations of AD[12,34], including the present study, and APOE
292 or hippocampus-focused therapies could prove more effective for these individuals. Together, these re-
293 sults point to the possibility that clinical trials may benefit from stratification or enrichment based on AD
294 subtype, or as a first step, post-hoc identification of within-subtype effects.
295 There are currently very few explanations as to why subtypes of AD manifest. Fascinating work has
296 found PCA and lvPPA patients are more likely to exhibit learning disabilities in childhood[45,46], perhaps me-
297 diated by abnormalities during brain development[47]. While lvPPA and PCA may represent extremes along
298 the AD continua (as indicated by the present results), this points to the possibility that distinct subtypes
299 may be influenced by variation in cognitive development or other premorbid factors. Another possible
300 explanation for subtypes is interactions between post-translational tau modification and synaptic tau
301 spreading. Several studies have shown that the regional pattern of pathological tau expression in mice is
302 dependent on conformation and injection site of tau seeds[35,48,49]. It is therefore possible that subtypes of
303 tau spread may simply be dictated by distinct tau conformations and/or systematic variation in the human

304 connectome, perhaps at key synaptic junctures. Supporting the latter hypothesis, we found the tau-PET
305 pattern of AD subtypes resembled macroscale neuronal networks seeded from different brain regions.
306 These findings do not presuppose tau pathology necessarily starts in different regions, but instead that
307 different regions may play a more prominent role in tau propagation across subtypes as "amplifying
308 nodes". This could be mediated by involvement of distinct neuronal cell subtypes[50], which may incur dis-
309 rupted development due to environmental or genetic factors, leading to network abnormality during life
310 and network vulnerability in late life.
311   This study has a number of limitations. The SuStaIn method fits data based on the assumption that
312 several discrete sequences are represented within the data, and it uses cross-sectional information to
313 create pseudo-longitudinal sequences. This framework is based off of the same logic as most pathological
314 staging schema (e.g. [8]) and hypotheses of biomarker trajectories (e.g. [5]), but does so in an automated
315 fashion. It is therefore possible that a SuStaIn subtype trajectory could be created by "appending" or
316 "stitching" unrelated disease states together. However, we did find most individuals to remain the same
317 subtype at longitudinal follow up, and we could predict regional individual tau accumulation greater than
318 chance using just the SuStaIn model. While the use of tau-PET imaging is a great improvement over using
319 MRI to measure AD pathology, there is still some discrepancy between tau-PET signal and true tau pa-
320 thology[51]. While flortaucipir binds to paired-helical filament tau, off-target binding is an issue with flor-
321 taucipir, particularly in the striatum, white matter and choroid plexus[52]. We mitigated this issue by regres-
322 sion of choroid plexus signal, exclusion of subcortical ROIs and non-AD dementia patients, and region-
323 specific normalization against non-specific binding, as well as replication with RO948 which exhibits less
324 off-target binding[53]. Similarly, recent reports question whether elevated flortaucipir binding is detectable
325 before advanced stages of tau accumulation[54–56]. However, SuStaIn's modeling is based on relative re-
326 gional differences in pathology, and regional variation in tau-PET and tau pathology are correlated[55–57].
327 Still, while the unbiased spatial sampling of tau-PET data across the brain aided our discovery of these
328 subtype patterns, they must still be validated using histopathology studies. Sample size was an obvious
329 strength of our study, but it comes with the caveat of mixing data from multiple cohorts, scanners, and
330 cognitive batteries. We addressed this issue somewhat by examining subtypes in each cohort separately,
331 replicating our results in a separate sample and adjusting for cohort in our comparisons. In addition, de-
332 spite our study boasting the largest tau-PET sample to date, even larger samples would be preferable in
333 order to elucidate the spatiotemporal progression of each subtype in more detail. We arrived at a four-
334 subtype solution to describe our data using established statistical methodology to identify a solution the
335 data supports with confidence. However, this does not preclude the possibility that other, more subtly
336 disctinct subtypes exist (Extended Data Fig. S1f).
337   In conclusion, we describe four distinct but stable spatiotemporal phenotypes of tau accumulation
338 in AD. These subtypes exhibit differing clinical profiles and longitudinal outcomes, and their tau patterns
339 resemble distinct temporal lobe networks. Our data-driven results call into question whether "typical AD"
340 is a quantifiable entity, rather suggesting that several AD subtypes exist, and that their individual differ-
341 ences are exacerbated by more aggressive phenotypes with younger onset ages. Future studies should
342 seek to validate the existence and temporal evolution of these subtypes, as well as identify genetic, cellu-
343 lar and developmental factors that may influence their expression. This may include identifying differ-
344 ences in brain activity and connectivity between individuals, as well as differences in regional vulnerabil-
345 ity. This framework may also be useful for enrichment of clinical trials, for providing more individualized
346 clinical care, and eventually for more individualized treatment.
347
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Author Contributions

JWV, OH & ACE conceptualized the work. JWV, ALY, NPO, LMA, MJP & OH contributed to analytic design. MJP, MDD, GDR, CHL & OH acquired and provided the data. RS, RO, OTS & RLJ contributed to data curation and processing. JWV analyzed the data. ALY & DCA originally created the SuStaIn algorithm, and NPO & LMA contributed to its execution. YIM created the ESM algorithm. JWV & OH drafted the manuscript. All authors interpreted the data and substantively contributed to revising the manuscript.

Competing interests

MJP and MDD are employees of Avid Radiopharmaceuticals, a wholly owned subsidiary of Eli Lilly and Company and are minor stockholders in Eli Lilly. OH has acquired research support (for the institution)

from AVID Radiopharmaceuticals, Biogen, Eli Lilly, Eisai, GE Healthcare, Pfizer, and Roche. In the past 2 years, he has received consultancy/speaker fees from AC Immune, Alzpath, Biogen, Cerveau and Roche. The remaining authors declare no competing interests.

Data Availability statement

Tau-PET data contributing to this study was sourced from six different cohorts. One of them, ADNI, is a public access dataset and can be obtained through an application at http://adni.loni.usc.edu/. Data from the other datasets are not publicly available for download, but access requests can be made to the respective study Investigators: BioFINDER 1,2 – Oskar Hansson; UCSF Memory and Aging Center – Gil D Rabinovici; Gangnam Severence Hospital, Seoul – Chul Hyoung Lyoo; AVID Radiopharmaceuticals – Michael J Pontecorvo, Michael D Devous. Additionally, CMU60 DTI data used to create template DTI connectomes are publicly available, and can be accessed at https://www.cmu.edu/dietrich/psychology/cognitiveaxon/data.html.

Code Availability statement

Python and MatLab implementations of the SuStaIn algorithm are available on the UCL-POND github page: https://github.com/ucl-pond. The ESM algorithm is available for academics as part of an open-access, user-friendly software (for further details, visit https://www.neuropm-lab.com/).

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546 **Figure Legends**
547
548 **Figure 1**
549 Spatiotemporal subtypes of tau progression. A) Tau-PET pattern of tau-positive (subtyped) individuals. B) Quarter-
550 nary plot showing probability each individual is classified as each subtype. Dots are labeled by final subtype classi-
551 fication: S1 (blue), S2 (green), S3 (orange) or S4 (pink). Inset box shows individuals that had a probability < 0.5 to
552 be classified as any of the four subtypes (i.e. showing poor fit). C) Average tau-PET pattern for each subtype. The
553 colorbar is the same as Panel A. D) Regions showing significant difference between one subtype and all other sub-
554 types using OLS linear models adjusting for SuStaIn stage, after FDR correction. E) Progression of each subtype
555 through SuStaIn stages. Each image is a mean of individuals classified at the listed stage and up to four stages low-
556 er. Only the left hemisphere is shown.
557
558 **Figure 2**
559 Subtype stability: AD spatiotemporal subtypes replicate in another cohort using a different PET tracer. A) For both
560 the discovery (Orig) and replication (Repl) cohorts, maps showing regions significantly different between one sub-
561 type and all others (excluding S0) within the cohort (after FDR correction). Similar spatial patterns were observed,
562 except for a reversed pattern in S4. B) Confusion matrix comparing subtypes identified in the original (discovery)
563 sample (y-axis), and subtypes separately identified in the replication sample (x-axis). Values represent spatial cor-
564 relation between average regional tau for each subtype. Values along the diagonal indicates similarity between the
565 same subtype across both cohorts.
566
567 **Figure 3**
568 Progression of AD subtypes. Increasing SuStaIn stage is associated with lower age a) and worse cognition b) across
569 all subtypes. c) Rate of longitudinal decline in MMSE for each subtype. The x-axis was jittered for visualization pur-
570 poses only. The y-axis shows MMSE across all observations as predicted by linear mixed models adjusted for co-
571 variates. d) Boxplots showing the distribution of predicted MMSE slopes for each subtype, stratified by clinical di-
572 agnosis (stats in Supplementary Table S2). e) Cross-cohort meta-analysis for the effects of S4: L Temporal declining
573 faster (left) and S3: Posterior declining slower (right) than other subtypes, respectively. Diamonds represent effect
574 sizes, while diamond size reflects relative sample size. Red diamonds indicate significant effects. Error bars = SEM.
575 f) Confusion matrix showing longitudinal stability of subtypes. Each row shows the number of subjects from a given
576 subtype at Visit 1 that were classified as each subtype at Visit 2. The diagonal represents the number of subjects
577 that were classified as the same subtype at Visit 1 and Visit 2. g) Individuals with a higher probability of being clas-
578 sified into their subtype at baseline were more likely to show a stable subtype over time (two-sided
579 t[156,53]=5.26, p=3.6e-07). h) Annual change in SuStaIn stage for each subtype, in individuals with stable subtypes
580 over time (stats in Supplementary Note 3). i) SuStaIn was used to predict longitudinal change in regional tau accu-
581 mulation. Each dot represents a subject, and the y-axis represents the spatial correlation between the true region-
582 al tau change and the predicted regional tau change. Average predictions were significantly greater than chance
583 based on a two-tided, one-sample t-test against 0 (S1: t[78]=5.00,p=3.5e-06; S2: t[52]=2.16,p=0.035; S3:
584 t[45]=3.05,p=0.0039; S4: t[29]=4.93,p=3.1e-05) . *p(unc.)<0.05, *** p(unc.)<0.001. Error bars in a-c represent 95%
585 CI of model fit across 1000 bootstrap samples. For boxplots in d, g-i, center line=median, box=inner quartiles,
586 whiskers=extent of data distribution except *=outliers
587
588 **Figure 4**
589 Application of epidemic spreading model to determine subtype-specific corticolimbic circuit vulnerability. An epi-
590 demic spreading model was fit separately for each subtype; once using an entorhinal cortex epicenter (a, blue),
591 and once with a subtype-specific best-fitting epicenter (b, red). For each plot, each dot represents a region. The x-
592 axis represents the mean simulated tau-positive probabilities across the population, while the y-axis represents the
593 mean observed tau-positive probability. Each row represents a subtype. Error bars in a-c represent 95% CI of mod-
594 el fit across 1000 bootstrap samples. c) For each subtype, the probability that each region is the best fitting epicen-

595 ter for that subtype, based on bootstrap resampling. d) For each subtype, the proportion of individuals at various
596 stages that had best-fitting epicenters within each of five major brain divisions: medial temporal lobe (MTL, blue),
597 temporal lobe (yellow), parietal lobe (purple), occipital lobe (gray) and frontal lobe (turquoise). e) For each subype,
598 spatial representation of ESM results from panel B using best-fitting epicenter. From left to right, observed region-
599 al tau-PET probabilities (tau-P), regional connectivity to best-fitting epicenter (Cx), tau-PET probabilities predicted
600 by the ESM. These images show the degree to which constrained diffusion of signal through a connectome (Pred.),
601 starting in a given epicenter and its associated fiber network (Cx.), recapitulates the tau patterns of each subtype
602 (Obs.).
603
604 **Figure 5**
605 A theoretical model summarizing variation in the spread of tau pathology in AD. Tau pathology varies along an axis
606 of severity (vertical in the diagram), which is inversely associated with onset age. In addition, tau varies along a
607 spatiotempral dimension (horizontal plane in the diagram), such that an individual can be described by their fit
608 along one of at least four trajectories. Text indicates clinical characteristics of each subtypes. Emboldened text re-
609 flects robust differences between subtypes, while normal text reflects less-robust characteristics that differentiate
610 subtypes from tau-negative individuals
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623 **Table 1**
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	S0: No Tau	S1: Limbic	S2: MTL-Sparing	S3: Posterior	S4: L Temporal
n	687	137	73	131	80
Age	71.52 (8.1)	75.28 (7.7)#	71.34 (8.3)*	75.06 (7.3)#	73.41 (6.9)
Prop. Female	0.49	0.70#	0.60‡	0.64#	0.56
Education	15.17 (2.9)	14.42 (3.9)	14.29 (4.0)	14.6 (3.0)	14.82 (2.9)
Prop. APOE4	0.26	0.75#*	0.47*	0.63#	0.59#

	Carrier				
Cortical Tau SUVR	1.04 (0.1)	1.41 (0.1)#*	1.44 (0.1)#	1.44 (0.1)#	1.47 (0.1)#*
Laterality	0.0 (0.2)	-0.28 (1.3)*#R	-0.13 (1.6)‡*R	0.04 (1.5)	1.95 (1.2)#*L
MMSE	28.9 (1.5)	24.33 (3.0)#	24.32 (4.2)#	24.19 (3.0)#	23.33 (5.0)#
Global Cognition	0.36 (0.5)	-0.03 (0.8)*	-0.29 (0.8)#	-0.23 (0.8)#	-0.39 (0.9)#†
Abs. Memory	0.48 (0.7)	-0.62 (0.7)#†	-0.36 (0.7)#	-0.55 (0.7)#	-0.3 (0.8)#†
Abs. Language	0.22 (0.7)	-0.11 (0.8)	0.01 (0.9)	-0.18 (0.8)	-0.64 (1.1)#*
Abs. Executive	0.19 (0.6)	0.02 (0.9)	-0.33 (0.9)#	0.03 (0.8)	-0.17 (1.0)‡
Abs. Visuospatial	0.19 (0.6)	0.08 (1.0)	-0.25 (1.2)#	-0.23 (1.2)#	-0.09 (1.0)
Rel. Memory	0.26 (0.8)	-0.61 (1.0)#*	-0.14 (1.0)	-0.37 (1.0)#	-0.06 (1.1)*
Rel. Language	-0.02 (0.8)	0.05 (1.0)	0.31 (1.2)#	0.06 (1.0)	-0.51 (1.3)#*
Rel. Executive	-0.14 (0.8)	0.25 (1.0)‡	-0.22 (1.0)‡*	0.38 (1.1)#	0.22 (1.2)
Rel. Visuospatial	-0.1 (0.7)	0.31 (1.1)‡	0.03 (1.3)	0.0 (1.3)	0.27 (1.3)

Table 1 Comparison of means of different variables between subtypes in the discovery sample, after correction for age (except in the case of age), sex (except in the case of sex), education (except in the case of education), cohort, clinical diagnosis (i.e. CN, MCI, AD), and SuStaIn stage

629 (except comparisons with S0). Standard deviations are given in parentheses where relevant. All p-values were corrected for multiple compari-
630 sons.
631 MMSE = Mini-Mental State Examination; Abs. = Absolute; Rel. = Relative; Prop. = Proportion
632 * = Adj. p$<$0.05 (vs all other subtypes, not including S0)
633 † = Adj. p$<$0.1 (vs all other subtypes, not including S0)
634 # = Adj. p$<$0.05 (vs S0)
635 ‡ = Adj. p$<$0.1 (vs S0).
636 R = Significant right-sided laterality in this subtype compared to others
637 L = Significant left-sides laterality in this subtype compared to other subtypes.
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644 **Online Methods**
645 Unless otherwise noted, all data analysis was conducted, and all figures were created, using Python
646 v.3.7.3, mostly using the numpy, scipy, pandas, scikit-learn, nilearn, matplotlib, seaborn and statsmodels
647 libraries.
648
649 **Sample Characteristics.** The total sample for the following analyses comprised of flortaucipir tau-PET scans
650 from 1667 individuals from five different cohorts (BioFINDER I, Seoul, AVID, UCSF, ADNI), and RO948 PET
651 scans from 657 individuals from a sixth cohort (BioFINDER II). Information pertaining to recruitment, diag-
652 nostic criteria and Aβ positivity assessment for the BioFINDER I (BioF)[46], ADNI[27], AVID[32], Seoul[59], UCSF[6] and
653 BioFINDER II (BF2)[28] cohorts have been previously reported. Informed written consent was provided for all
654 participants or their designated caregiver, and all protocols were approved by each cohort's respective
655 institutional ethical review board. Specifically: All BioFINDER subjects provided written informed consent
656 to participate in the study according to the Declaration of Helsinki; ethical approval was given by the Eth-
657 ics Committee of Lund University, Lund, Sweden, and all methods were carried out in accordance with the
658 approved guidelines. Approval for PET imaging was obtained from the Swedish Medicines and Products
659 Agency and the local Radiation Safety Committee at Skåne University Hospital, Sweden. For UCSF, the
660 study was approved by the University of California (San Francisco and Berkeley) and Lawrence Berkeley
661 National Laboratory institutional review boards for human research. Data from the AVID sample were
662 collected in compliance with the Declaration of Helsinki and the International Conference on Harmoniza-
663 tion guideline on good clinical practice. Data collection for the Gangnam Severance hospital sample was
664 approved by the institutional review board of Gangnam Severance Hospital. Information related to partic-
665 ipant consent in ADNI can be found at (ADNI; http://adni.loni.usc.edu). Some of the data used in the
666 preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI)
667 database (adni.loni.usc.edu). The ADNI was launched in 2003 as a public-private partnership, led by Prin-
668 cipal Investigator Michael W. Weiner, MD. The primary goal of ADNI has been to test whether serial mag-
669 netic resonance imaging (MRI), positron emission tomography (PET), other biological markers, and clinical
670 and neuropsychological assessment can be combined to measure the progression of mild cognitive im-
671 pairment (MCI) and early Alzheimer's disease (AD). For up-to-date information, see www.adni-info.org.
672   From this total sample of 1667 subjects with flortaucipir scans, a subsample was derived including i)
673 all cognitively unimpaired individuals older than 40 years; and ii) individuals who had both a diagnosis of
674 MCI or AD, *and* imaging or fluid evidence of brain Aβ pathology. All subjects with a primary diagnosis oth-
675 er than cognitively unimpaired (which included subjective cognitive decline), MCI or AD were excluded.
676 This subsample, used for all subsequent analysis, comprised 1143 individuals. The same screening proce-
677 dures were used to filter individuals from BioFINDER II, reducing the samples size from 657 to 469. Char-
678 acteristics of all samples, including inter-cohort differences, are detailed in Table S1.
679

680 **Image Acquisition and Preprocessing.** Tau-PET data acquisition procedures for each cohort have been pre-
681 viously described[6,27,28,32,46,59]. All tau-PET data were processed centrally in Lund by analysts blinded to de-
682 mographic and clinical data, in a manner previously described[46]. Briefly, resampling procedures were used
683 to harmonize image size and voxel dimension across sites. Each image underwent motion correction using
684 AFNI's 3dvolreg (https: //afni.nimh.nih.gov/), and individual PET volumes were averaged within-subject.

685 Each subject's mean PET image next underwent rigid coregistration to it's respective skull-stripped native
686 T1 image, and images were intensity normalized using an inferior cerebellar gray reference region, result-
687 ing in standardized update value ratio (SUVR) images. T1 images were processed using Freesurfer v6.0
688 (https://surfer.nmr.mgh.harvard.edu/), resulting in native space parcellations of each subject's brain using
689 the Desikan-Killiany (freesurfer) atlas. These parcellations were used to extract mean SUVR values within
690 different regions of interest (ROIs) for each subject in native space.
691
692 **Subtype and Stage Inference.** Typical efforts to perform data-driven subtyping of neuroimages in AD are
693 limited by the confound of disease stage. In a sample spanning the AD spectrum from healthy to dement-
694 ed such as ours, disease progression represents the main source of variation in MR and PET images.
695 Therefore, unless disease stage is somehow accounted for, most clustering algorithms will partition indi-
696 viduals based on their disease stage. This is not useful for parsing heterogeneous patterns related to pro-
697 gression subtypes, which are theoretically orthogonal to disease progression itself. The Subtype and Stage
698 Inference (SuStaIn)[23] algorithm surmounts this limitation by combining clustering with disease progression
699 modeling. Detailed formalization of SuStaIn has been published previously[23].
700   SuStaIn models linear transition across discrete points along a progression of indices of severity (typ-
701 ically z-scores), separately across different ROIs (Fig. S1a). Input requires a subject x feature matrix where,
702 in this case, features represent mean tau-PET signal within different ROIs. In addition, "severity scores",
703 indicating different waypoints along the natural progression of ROI severity, must be provided. Whereas
704 the choice of ROI constrains the spatial dimensions along which individuals may vary, the severity scores
705 instead constrain the temporal dimension of variation. The total number of features is therefore repre-
706 sented by the product of N ROIs by N ROI-specific severity scores. A balance must thus be struck between
707 resolution in the spatial and temporal dimensions, with respect to overall sample size.
708   Our discovery sample boasts scans from 1143 individuals, but even given our inclusion criteria, we
709 expected from previous work[29] that the majority of individuals (50-60%) will have minimal tau binding
710 (note that SuStaIn will automatically detect these individuals and exclude them from progression model-
711 ing). We therefore expect the modeling to be performed on a sample of closer to N~450-550. We there-
712 fore decided on ten different ROIs (spatial features), each with three severity scores (temporal dimen-
713 sion), totalling 30 features. Given an arbitrary rule of 10-20 observations per feature, 300-600 observa-
714 tions should provide sufficient power, and our sample size should therefore be sufficient.
715   For the ten spatial features, we opted for left and right lobar regions of interest: parietal, frontal,
716 occipital, temporal and medial temporal lobe (MTL). This choice is justified as follows: i) previous imaging
717 and pathology subtyping studies have revealed variation in AD pathology to often occur within specific
718 lobes, e.g. limbic-predominant (MTL), MTL-sparing (parietal), posterior cortical atrophy (occipital), logo-
719 penic aphasia (temporal) and behavioral variant AD (perhaps frontal)[18]; ii) hemispheric laterality in AD is
720 understudied, perhaps due to pathological staining often occurring on single hemispheres. However,
721 some laterality has been observed in AD clinical variants (i.e. lvPPA[15]) and may point to differing pheno-
722 types in typical AD; iii) These lobar regions maintain some orthogonality to disease progression, as multi-
723 ple lobes are involved in Braak stages IV - VI[8].
724   To define severity score cutoffs, we first sought to normalize SUVR values to account for regional
725 differences in PET signal (due to nonuniformity of off-target binding, perfusion, etc. across the brain)[29].
726 Two-component Gaussian mixture models were used to define, for each ROI, a normal (Gaussian-shaped
727 noise) and abnormal distribution. We then created tau Z-scores by normalizing all values using the mean
728 of the normal distribution (Extended Data Fig. S1b). This procedure centered the Z-score values on the

729 normal distribution to allow for more interpretable values (i.e. 2=2 SDs from normal), and also accounted
730 for region-specific differences in normal and abnormal SUVR distributions. Uniform values of Z = 2, 5, 10
731 were arbitrarily chosen as severity score control points for all ROIs (Extended Data Fig. S1)B. However,
732 analyses were also run with alternative z-score values, see the Replication Analysis section below.
733   The number of subtypes (i.e. distinct spatiotemporal progressions) was determined through cross-
734 validation. Separately for each k=1-7 subtypes, 10-fold cross-validation was performed where, for each
735 fold, SuStaIn was fit to 90% of the data, and this model was used to evaluate sample likelihood for the
736 10% left-out subjects. For each left-out set, model fit was evaluated using the cross-validation information
737 criterion (CVIC; as described in [23]), as well as out-of-sample log-likelihood. In addition, we used the inner-
738 fold SuStaIn model to assign all outer-fold individuals to a subtype, and we evaluated the probability of
739 the maximum-likelihood subtype. In theory, a better fit model should produce more high probability as-
740 signments of left-out data, though more subtypes will also make assignment more challenging. k was cho-
741 sen by evaluating these three metrics in concert (Extended Data Fig. S1c-e). CVIC increased significantly
742 with increasing k, indicating better fit to the data as the number of subtypes increased, though the curve
743 flattened somewhat after k=4 (Extended Data Fig. S1c). Similarly, log-likelihood increased indicating better
744 model fit, up until k=4, after which no improvement was seen (Extended Data Fig. S1d). In contrast to the-
745 se fit statistics, cross-validated maximum-likelihood subtype probability decreased with increasing k, indi-
746 cating less-confident assignment of left-out data with more subtypes. This decline was steady, though the
747 median probability dropped below 0.5 after k=4. Taken together, k=4 appeared to be the best solution to
748 maximize model fit but minimize detriment to subtype confidence. We also noted that no subtypes after
749 k=4 had more than one "parent" subtype. In other words, solutions 3 and 4 feature subtypes that were
750 composed of multiple parent subtypes, whereas all solutions thereafter featured only subtypes that split
751 off from a single parent subtype. This could be indicative of a certain level of hierarchical convergence at
752 k=4 (Extended Data Fig. S1f).
753   Finally, SuStaIn was run on the whole sample with the selected k=4. Note that for model fitting, SuS-
754 taIn uses a uniform prior on disease subtype and stage (i.e. assumes all subtype and stage combinations
755 equally likely). Note also that the model is initialized with an expectation-maximization algorithm, and
756 therefore does not require a burn-in period.[23] The model was fit using 10000 Markov chain Monte Carlo
757 (MCMC) iterations. SuStaIn calculates the probability that each individual falls into each stage of each sub-
758 type, and individuals are assigned to their maximum likelihood subtype and stage. Note that individuals
759 that do not express abnormal tau in any region are classified by SuStaIn as "Stage 0", and are not assigned
760 to a subtype. The proportion of individuals classified into each subtype was quantified. We also stratified
761 this quantification by clinical diagnosis, and to cohort to assess the frequency of subtypes in each contrib-
762 uting dataset. Finally, we quantified the proportion of subjects that did not fall well into any subtype (no
763 subtype probability >50%).
764
765 **Post-hoc subtype correction.** Manual inspection of subtype progressions suggested that the early stages of
766 one subtype (S2: MTL-Sparing; see Results) were composed mostly of cognitively normal individuals with
767 abnormally high tau-PET binding throughout the cortex, but little-to-no tau in typical early-mid AD re-
768 gions, i.e. false (tau) positives. Specifically, these individuals showed elevated binding throughout the cor-
769 tex, including sensorimotor and frontal regions (regions where tau typically accumulates only in the latest
770 stages of AD[8]), but had low tau levels in the temporal lobes (Extended Data Fig. S1a). On an individual
771 basis, such individuals showed tau-PET signal that was slightly but globally elevated, with several small
772 "hotspots" distributed diffusely throughout frontal, parietal and occipital cortex. While it is unclear
773 whether this elevated binding represents off-target binding, diffuse low-level target binding, or other
774 methodological issues, consensus among co-authors was that these individuals were not consistent with
775 an AD phenotype. We used Gaussian mixture modeling across all individuals as described in [29] to define
776 the probability of abnormal tau-positivity in each of the left and right entorhinal cortex and precuneus,
777 respectively. We then marked individuals who had <90% probability of tau in all four regions as low-
778 probability tau individuals (T-). These individuals also underwent manual inspection. Next, we identified T-
779 individuals in the MTL-Sparing subtype, finding 40.6% of this subtype was composed of this group, and all

780   were classified as stage 5 (of 31) or below. Furthermore these individuals showed many other indications
781   of being false (tau) positives: they had normal MMSE scores, were older, were less likely to be Aβ+ and
782   less likely to be MCI or AD (Extended Data Fig. S2b,c). We assume SuStaIn appended this specific group of
783   T- individuals to the MTL-Sparing subtype because the individuals i) had abnormally high tau in at least
784   one ROI as per our calculations (even if that abnormal signal was not driven by pathology); ii) the abnor-
785   mal tau was located mainly in the isocortex inclusive of the parietal lobe; iii) these individuals did not have
786   elevated MTL binding. As SuStaIn is an unsupervised algorithm, the pathological MTL-sparing phenotype
787   became conflated with this specific profile of T- individuals. To correct this issue, we converted all T- indi-
788   viduals classified as MTL-sparing to Subtype 0 for all further analysis.

790   **Visualization of subtype-specific tau-PET patterns.** To visualize tau-PET patterns for each subtype, we calcu-
791   lated the mean tau Z-score for each Desikan-Killiany (freesurfer) atlas ROI. To visualize the progression of
792   the subtype pattern across SuStaIn stages, for each subtype, we created mean images for all individuals
793   falling into the following SuStaIn stage bins: 2-6, 7-11, 12-16, 17-21, 22-26. To deduce regions with rela-
794   tively greater or less tau signal for each subtype, we created region-wise one-vs-all ordinary least squares
795   (OLS) linear models comparing regional tau in one subtype to all others. This analysis was performed to
796   visualize subtype models inferred by SuStaIn using individual data, and to explore differences between
797   subtypes. Each model included ROI tau Z-scores as the dependent variable, a one-hot dummy variable
798   representing membership in the reference subtype, and SuStaIn stage as a covariate. These models were
799   FDR-corrected for the number of comparisons (i.e. number of ROIs).

801   **Subtype Characterization.** Several demographic, cognitive and genetic variables were available for nearly
802   all individuals across the five cohorts in our main (discovery) cohort. These variables included clinical di-
803   agnosis (100%), age (99.8%), sex (100%), years of education (97.1%), mini mental state examination
804   (MMSE) score (97.7%) and *APOE4* allele carriage (89.5%). Only the UCSF sample provided diagnoses of
805   clinical AD variants such as PCA[16] and lvPPA[17]. In addition, most individuals underwent extensive cohort-
806   specific cognitive batteries assessing multiple domains of cognition. In order to utilize this rich cognitive
807   data, we created cognitive domain scores separately within each cohort by taking the mean of several z-
808   scored tests within the following cognitive domains: memory, executive function, language and visuospa-
809   tial function. Supplemental Table S5 indicates which cognitive tests were used in each cognitive domain
810   score across each cohort. We calculated global cognition as the mean between the four domain scores.
811   Finally, we additionally regressed global cognition out of each domain score to create "relative" cognitive
812   domain scores. These scores are useful for assessing the degree of domain-specific impairment above and
813   beyond global impairment. The various absolute and relative domain scores were then aggregated across
814   all cohorts to maximize the sample size available for cognitive tests: memory (86.6%), language (81.3%),
815   executive function (85.5%), visuospatial function (82.0%). While aggregating scores of different composi-
816   tions across cohorts of different compositions presents a suboptimal solution, we rest on sample sizes and
817   statistical correction helping to overcome these limitations.
818   Subtypes were statistically compared to one another, and to tau-negative (i.e. Stage 0) individuals,
819   in order to determine subtype-specific characteristics. These analyses compared age, sex, education, AP-
820   OE4 carriage, MMSE, global cognition, total tau, and total tau asymmetry. Comparisons between subtypes
821   and Stage 0 individuals additionally included the four cognitive domain scores, while comparison between
822   subtypes instead included the four "relative" cognitive domain scores. This statistical comparison involved
823   three steps: 1) Comparison to tau-negative individuals: Tau-negative individuals were those characterized
824   as "Subtype 0" by SuStaIn, i.e. those individuals that did not demonstrate any abnormal tau events. An
825   OLS linear model was fit with each variable described above as the dependent variable, and with dummy-
826   coded subtype entered as the independent variable (with S0 as the reference subtype). The model also
827   included age, sex, education, clinical status (CN, MCI, AD) and cohort as covariates (except when that co-
828   variate was the dependent variable). Model t- and p-values were stored for each model and the latter
829   were FDR-corrected. 2) Comparison between subtypes. A one-vs-all approach was applied to subtyped
830   individuals only to assess how different tau-progression subtypes differed from one another. Separately

831 for each subtype, models were fit for each variable with a single dummy variable entered indicating
832 membership to that subtype. Models once again covaried for age, sex, education, clinical status (CN, MCI,
833 AD), cohort, and, this time, SuStaIn stage. T and p values were stored, and the latter was FDR-corrected
834 for the number of variables assessed. 3) Finally, each subtype was compared directly to each other sub-
835 type (i.e. one-vs-one comparison). OLS models were fit with dummy coded subtype variables as the de-
836 pendent variable, cycling each subtype as the reference subtype. T and p values for each of these models
837 were stored, and the latter was FDR-corrected for number of comparisons (i.e. number of dependent var-
838 iables). These models were also adjusted for age, sex, education, clinical status (CN, MCI, AD), cohort and
839 SuStaIn stage. For space reasons, the results of 3) are only visible in Extended Data Fig. S8.
840   All models were subjected to diagnostics to ensure our data fulfilled assumptions of OLS regression
841 models. We found the residuals of all models to be normally distributed (Anderson-Darling tests p>0.05).
842 Further, we found no strong evidence for autocorrelation (Durbin Watson test, 1.5 < all models < 2.5),
843 outliers (Cooks distance of all subjects < 0.5), multicollinearity (variance inflation factor (VIF) for all covari-
844 ates < 100; besides age [23-27], sex [8-12] and education [13-17], all VIF < 10) or heteroscedasticity (visual
845 assessment of distribution around mean of residuals) in any of our models.
846   We also assessed the relationship between SuStaIn stage and two variables: age and MMSE. For
847 these analyses, stage was correlated with age and MMSE, and the results were visualized across the
848 whole sample and also stratified by subtype. As a post-hoc analysis, we separated individuals into differ-
849 ent age groups: 65 or younger, and older than 65. We then reassessed age by SuStaIn stage correlations
850 within each of these age groups.
851   Longitudinal MMSE data was also available for individuals from all cohorts, totalling 735 individuals
852 with at least two timepoints. 195 individuals had an additional third timepoint, 29 had a fourth, and 3 had
853 a fifth. Mean latest follow-up was 1.72 years from PET scan (sd = 0.64). Linear mixed effect models were
854 used to assess difference in longitudinal MMSE change between subtypes. All models were fit using the
855 lme4 library in R, using type-III sum of squares, unstructured covariance matrices, and Satterthwaite's
856 approximation to calculate the denominator degrees of freedom for p-values. Models featured MMSE
857 measurements as the dependent variable, interactions between time from baseline and dummy coded
858 subtype variables as the independent variables of interest (cycling the reference subtype), subject ID as a
859 random effect (allowing for random intercepts and slopes), and age, sex, education, cohort, dummy cod-
860 ed variables for MCI and AD, and SuStaIn stage as covariates of no interest. One vs all models were also fit
861 for each subject using dummy coded subtype variables, and significant effects were reported. We addi-
862 tionally repeated the one vs. all subtype models within each cohort separately, and used this to calculate
863 a meta-analysis by finding a weighted mean of the t-values and standard errors. Since this analysis was
864 confirmatory, we used a one-tailed significance test to calculate the p-values.
865
866 **Replication Analysis.** We performed two types of replication analysis. To ensure that our results were not
867 driven by arbitrary z-score cutoffs, we reran models with completely different cutoffs. To ensure the re-
868 sults were not driven by our sample or unique to the flortaucipir radiotracer, we repeated the analysis *de*
869 *novo* in a separate cohort using a different tau-PET radiotracer.
870   SuStaIn require z-score values to anchor the psuedotime for each ROI (see section Subtype and
871 Stage Inference above), and we chose values of 2,5,10 for all ROIs so as to not let any region bias or influ-
872 ence the model unduly, and to aid comparability across different regions. To ensure our results were not
873 driven by this choice, we reran the model with a different set of z-score values chosen in a data-driven
874 manner. The object was to allow the distribution of tau-PET data in each region to define natural way-
875 points in the data. For each input region, we fit Gaussian mixture models to the data, varying the number
876 of components between 1 and 5. We used the model fit (AIC) to decide the optimal number of compo-
877 nents for each region. Finally, we used five-fold cross-validation to determine the boundaries of these
878 Gaussians to define anchors for each regions. We did this separately for each ROI, and as a result, differ-
879 ent ROIs had different waypoints, and even different numbers of waypoints (Table S6). We then refit the

880 SuStaIn model to the data and compared the results to the original model using spatial correlation (see
881 below).
882   While the five cohorts from the main discovery sample all use flortaucipir as the tau-PET tracer, a
883 sixth cohort (BioFINDER II; BF2) was available that instead used the RO948 radiotracer. While the two
884 tracers have similar binding patterns, RO948 tends to have less off-target binding in the basal ganglia and
885 better MTL signal, but frequently boasts high meningeal signal that can affect cortical SUVR measure-
886 ment[53]. Because of these differences, we opted to leave BF2 out of the discovery sample, and instead use
887 it as a replication cohort. This strategy allowed us to not only evaluate the stability of the subtypes in a
888 new cohort, but also allowed us to evaluate whether the subtypes are robust to tau-PET radiotracer.
889   We reran SuStaIn *de novo* in the BF2 sample, using identical procedures to those described above
890 (Methods: Subtype and Stage Inference), although using the discovery sample to inform the number of
891 subtypes. The resulting subtypes were visualized and quantitatively assessed using spatial correlations.
892 Specifically, mean within-subtype SUVRs were computed for each (freesurfer) ROI, and each discovery
893 subtype ROI-vector was correlated to each replication (BF2) subtype ROI-vector. To account for whether
894 different sample sizes contribute to differing results between the discovery and replication datasets, we
895 performed a split-half analysis with the discovery sample. Specifically, we split the discovery sample in
896 half and ran SuStaIn separately on each half, once again using the original discovery sample to inform the
897 number of subtypes. We then compared each half, which had a sample size comparable to that of BF2, to
898 the BF2 samples using spatial correlations.
899
900 **Assessment of Longitudinal Stability.** Longitudinal PET data was available for individuals across all cohorts
901 except for UCSF, totaling 519 individuals with at least two time points (mean follow-up time = 1.42, sd =
902 0.58, years). These longitudinal scans were used to validate the stability of subtypes over time, under the
903 hypothesis that individuals should remain the same subtype, but should advance (or remain stable) in
904 SuStaIn stage over time. ROIs for the longitudinal datasets were Z-scored as described above, but using
905 the cross-sectional cohort as the cohort for normalization. The SuStaIn model fitted to the cross-sectional
906 dataset was used to infer subtype and stage of longitudinal data (all timepoints). Confusion matrices were
907 built to assess subtype stability between baseline and first follow-up. Stability was calculated as propor-
908 tion of individuals classified as the same subtype at follow-up, or who advanced from Stage 0 into a sub-
909 type, compared to the total number of individuals. Stability was also calculated excluding individuals who
910 were classified as Stage 0 at baseline or follow-up. We also assessed the influence of subtype probability
911 (i.e. the probability a subject falls into their given subtype) on individual subtype stability. Specifically, we
912 compared the subtype probability of stable individuals to unstable individuals with a t-test. We additional-
913 ly calculated overall model stability after excluding individuals using various subtype probability thresh-
914 olds.
915   Subtype progression was assessed by observing change in SuStaIn stage over time in stable subtype
916 individuals. We calculated the proportion of individuals who advanced, were stable, or regressed in dis-
917 ease stage over time, before and after accounting for model uncertainty. Specifically, while stages are
918 generally characterized by advancing abnormality in a given region, uncertainty leads to some stages be-
919 ing characterized by probabilities of progressing abnormalities in more than one region. Therefore, indi-
920 viduals who advanced or regressed to a stage with event probabilities overlapping with their previous
921 stage were considered to be stable. We also calculated annual change in SuStaIn stage by dividing total
922 change in SuStaIn stage by number of years between baseline and final available timepoint. We used one-
923 sample t-test against zero to assess whether significant change over time was observed across the whole
924 sample, and within each subtype. We used ANOVAs and Tukey's post-hoc tests to assess differences in
925 annual change in stage across the different subtypes. We also correlated annual change in stage with
926 baseline stage, and with age.
927
928 **Individual forecasting of longitudinal tau progression.** SuStaIn models spatiotemporal subtype progressions,
929 but does so using only cross-sectional data. Therefore, longitudinal data can be used as "unseen" or "left-

930   out" data, which can be used to test whether and to what extent individuals follow the trajectories pre-
931   dicted by SuStaIn. We accomplish this by using an individual's subtype and stage probability to generate a
932   predicted second time point, and comparing the change between baseline and predicted follow-up to
933   change between baseline and actual follow-up.
934   To do this, we first sought to predict the rate of change of stage for each individual. We trained a
935   Lasso model to predict individual annualized change in SuStaIn stage (Δstage) using available data, and
936   cross-validation to get out-of-sample predictions for each individual. Features included age, sex, educa-
937   tion, amyloid status, APOE4 status, baseline stage MMSE and dummy coded variables for MCI, AD, and
938   each subtype. For each fold, the model was trained on 90% of the data, and this model was used to pre-
939   dict Δstage in the 10% left out subjects. This process was repeated until predictions were made for each
940   subject. The mean absolute error between the predicted and true Δstage was 0.91 stages/year. The pre-
941   dicted Δstage was used for subsequent aspects of the tau prediction. This is important, as we are there-
942   fore minimizing the amount of longitudinal information leaking into the forecast.
943   Using this predicted Δstage, we were then able to predict an individual's stage at follow-up $k_{i,new}$ giv-
944   en any stage at baseline k, as $k_{i,new} = k + \Delta_{stage} t_i$, where t_i is the time between follow-up visits in years.
945   We can then evaluate the SuStaIn-predicted pattern of regional tau deposition at baseline $Y_{i,j}$ as

$$Y_{ij} = \sum_{c=1}^{C} \sum_{k=0}^{K} A_{j,c,k} P_{i,c,k}$$

946   or at follow-up $Z_{i,j}$ as

$$Z_{ij} = \sum_{c=1}^{C} \sum_{k=0}^{K} A_{j,c,k_{i,new}} P_{i,c,k}$$

947   where $A_{j,c,k}$ is an 'archetype' indicating the expected amount of tau deposition for biomarker j at stage k of
948   subtype c and $P_{i,c,k}$ is the probability subject i is at stage k of subtype c. The archetype $A_{j,c,k}$ is estimated
949   probabilistically from the Markov chain Monte Carlo (MCMC) samples of uncertainty provided by the SuS-
950   taIn algorithm, giving an average archetypal pattern accounting for the uncertainty in the progression
951   pattern of each subtype. This means that each SuStaIn-predicted pattern $Y_{i,j}$ accounts for both uncertainty
952   in the progression pattern of each subtype as well as uncertainty in the subtype and stage of each individ-
953   ual.
954   We can therefore represent the predicted change in tau as $Z_{i,j} - Y_{i,j}$. This vector represents the pre-
955   dicted change in tau Z-score in each of the ten spatial input features to SuStain (i.e. left and right tem-
956   poral, parietal, occipital, frontal and medial temporal lobes). We evaluate the prediction by computing,
957   for each individual, the correlation between the predicted and true regional tau change vectors. We eval-
958   uate the overall prediction across the whole sample, and within-subtypes, by comparing the average pre-
959   diction against chance using one-sample t-tests against a correlation of zero.
960
961   **Epidemic spreading model.** Perhaps the most prominent hypothesis of tau spread suggests tau oligomers
962   spread directly from neuron to neuron through axonal connections. Under this hypothesis, diverse but
963   systematic variations in tau spreading may be driven by variability in macroscale connectivity, network
964   organization or vulnerable circuits. We test this idea by investigating whether a network diffusion model
965   simulating tau spread through the human connectome can recapitulate the various subtype patterns dis-
966   covered by SuStaIn. We have previously applied the epidemic spreading model (ESM)[58] to tau-PET data,
967   showing diffusion of an agent through human connectivity data (measured with diffusion imaging-based
968   tractography) can explain a majority of the variance of spatial tau patterns across a population of individ-
969   uals along the AD spectrum[29]. We here conduct the exact same analysis separately for each subtype iden-
970   tified through SuStaIn. We further allow the ESM to identify regional epicenters separately for each sub-
971   type, under the hypothesis that different subtype patterns may be driven by prominence of different cor-
972   ticolimbic networks.

973   As described in [29], each tau-PET ROI was converted to tau-positive probabilities using mixture mod-
974 eling. This process is similar to the Z-scoring procedure (Extended Data Fig. S1), though in this case, the
975 probability that values fall onto the abnormal distribution is ascertained using five-fold cross-validation.
976 These measures represent the probability that a given ROI exhibits tau in the abnormal range. Connectivi-
977 ty was measured from a dataset of 60 young healthy subjects from the CMU-60 DSI Template
978 (http://www.psy.cmu. edu/coaxlab/data.html). Deterministic tractography was calculated for each indi-
979 vidual by finding connections between ROIs using orientation distribution functions, and connectivity was
980 measured using the anatomical connection density (ACD) metric[58]. Images were assessed for quality and
981 connectomes were averaged across all 60 individuals. For each subtype separately, the ESM was fitted
982 across all individuals, cycling through the average of each left-right pair of cortical ROIs (including hippo-
983 campus and amygdala, 33 pairs in total) as the model epicenter. The best fitting epicenter was selected by
984 finding the model with the minimum mean euclidian distance between model predicted and observed tau
985 spatial pattern across subjects. Model accuracy was represented as the r^2 between the mean observed
986 ROI-level tau-PET probabilities and mean predicted probabilities across subjects. For each subtype, we
987 compared the r^2 of the model using the best-fitting epicenter to the r^2 of models using an entorhinal epi-
988 center. To gain confidence in the subject-specific epicenter, we bootstrapped the sample 1000 times and
989 recomputed the best-fitting epicenter for each subtype. Epicenter probability was calculated as the fre-
990 quency that an epicenter was selected as best epicenter across bootstrap samples.
991   We were additionally interested in how secondary seeding evolved over the course of each subtype
992 progression. While the ESM is designed to ascertain the true pathological epicenter, the selected epicen-
993 ter reflects the seeding point that best matches the spatial pattern of the dependent variable. As such, it
994 is likely that "secondary epicenters" become important for disease spread at later disease stages. We
995 binned individuals for each subtype into disease stage bins, as with Fig 1e. Individual epicenters were as-
996 certained for each subject, and were aggregated based on lobe (MTL, temporal, frontal, parietal, occipi-
997 tal). We then calculated epicenter frequency among individuals in each stage bin for each subtype. This
998 allowed us to track how the secondary epicenter evolve throughout the disease course for each subtype
999 trajectory.
1000   We repeated this same analysis with a different connectome based on rsfMRI connectivity from an
1001 elderly population, and using a higher-resolution atlas. The sample consisted of rsfMRI scans from 422
1002 healthy elderly controls (166 Aβ-positive), 138 individuals with subjective cognitive decline but without
1003 objective impairment (48 Aβ-positive), and 83 Aβ-positive MCI patients. 57 individuals overlapped be-
1004 tween this sample and the tau-PET discovery sample used for analysis. Functional data was processed
1005 using modified CPAC pipeline[60] involving slice time correction, bandpass filtering at 0.01-0.1 Hz, regression
1006 of motion, white matter and CSF signal, compcor physiological noise, and the 24 Friston parameters. The
1007 timeseries also underwent adaptive censoring of volumes for which DVARS jumps above median+1.5*IQR
1008 were observed. Timeseries were averaged within ROIs of the 246-ROI Brainnetome cortical/subcortical
1009 atlas (https://atlas.brainnetome.org/), nodewise connectivity was calculated using either Fisher's Z trans-
1010 formed correlations or partial-correlation (see below). The ESM was fit using the bilateral A35/36r ROI as
1011 model epicenter, and the following combinations of parameters were varied: regions (cortical only or all
1012 regions), subject-base (Aβ-negative only vs. all subjects), density (edgewise thresholding at 0.02, 0.5, 0.1,
1013 0.25, 1, or partial correlation with no thresholding, and normalization (whether connectivity matrices
1014 were normalized after density thresholding). The only parameter strongly affecting model performance
1015 was density threshold – partial correlation far outperformed all other conditions. Using all regions over
1016 only cortical regions bore slight advantages, as did using all subjects over only Aβ-negative. Normalization
1017 had no effect on outcomes. The best-fitting model was used for further analysis. The ESM was fit to each
1018 subject separately, and epicenter bootstrapping was performed, both as described above.
1019
1020
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1022

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1031
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1033 **Alzheimer's Disease Neuroimaging Initiative**

1034 Michael Weiner[17], Paul Aisen[18], Ronald Petersen[19], Clifford R. Jack Jr.[19], William Jagust[20], John

1035 Q. Trojanowki[21], Arthur W. Toga[22], Laurel Beckett[23], Robert C. Green[24], Andrew J. Saykin[25],

1036 John Morris[26], Leslie M. Shaw[27], Enchi Liu[28], Tom Montine[29], Ronald G. Thomas[18], Michael

1037 Donohue[18], Sarah Walter[18], Devon Gessert[18], Tamie Sather[18], Gus Jiminez[18], Danielle Harvey[23],

1038 Matthew Bernstein[19], Nick Fox[30], Paul Thompson[31], Norbert Schuff[32], Charles DeCArli[23], Bret

1039 Borowski[19], Jeff Gunter[19], Matt Senjem[19], Prashanthi Vemuri[19], David Jones[19], Kejal Kantarci[19],

1040 Chad Ward[19], Robert A. Koeppe[33], Norm Foster[34], Eric M. Reiman[35], Kewei Chen[35], Chet

1041 Mathis[36], Susan Landau[20], Nigel J. Cairns[26], Erin Householder[26], Lisa Taylor Reinwald[26],

1042 Virginia Lee[37], Magdalena Korecka[37], Michal Figurski[37], Karen Crawford[22], Scott Neu[22], Tatiana

1043 M. Foroud[25], Steven Potkin[38], Li Shen[25], Faber Kelley[25], Sungeun Kim[25], Kwangsik Nho[25],

1044 Zaven Kachaturian[39], Richard Frank[40], Peter J. Snyder[41], Susan Molchan[42], Jeffrey Kaye[43],

1045 Joseph Quinn[43], Betty Lind[43], Raina Carter[43], Sara Dolen[43], Lon S. Schneider[44], Sonia

1046 Pawluczyk[44], Mauricio Beccera[44], Liberty Teodoro[44], Bryan M. Spann[44], James Brewer[45], Helen

1047 Vanderswag[45], Adam Fleisher[35], Judith L. Heidebrink[33], Joanne L. Lord[33], Sara S. Mason[19],

1048 Colleen S. Albers[19], David Knopman[19], Kris Johnson[19], Rachelle S. Doody[46], Javier Villanueva

1049 Meyer[46], Munir Chowdhury[46], Susan Rountree[46], Mimi Dang[46], Yaakov Stern[47], Lawrence S.

1050 Honig[47], Karen L. Bell[47], Beau Ances[48], John C. Morris[48], Maria Carroll[48], Sue Leon[48], Mark A.

1051 Mintun[48], Stacy Schneider[48], Angela OliverNG[49], Randall Griffith[49], David Clark[49], David

1052 Geldmacher[49], John Brockington[49], Erik Roberson[49], Hillel Grossman[50], Effie Mitsis[50], Leyla

1053 deToledo-Morrell[51], Raj C. Shah[51], Ranjan Duara[52], Daniel Varon[52], Maria T. Greig[52], Peggy

1054 Roberts[52], Marilyn Albert[53], Chiadi Onyike[53], Daniel D'Agostino II[53], Stephanie Kielb[53], James

1055 E. Galvin[54], Dana M. Pogorelec[54], Brittany Cerbone[54], Christina A. Michel[54], Henry Rusinek[54],

1056 Mony J de Leon[54], Lidia Glodzik[54], Susan De Santi[54], P. Murali Doraiswamy[55], Jeffrey R.

1057 Petrella[55], Terence Z. Wong[55], Steven E. Arnold[27], Jason H. Karlawish[27], David Wolk[27], Charles

1058 D. Smith[56], Greg Jicha[56], Peter Hardy[56], Partha Sinha[56], Elizabeth Oates[56], Gary Conrad[56], Oscar

1059 L. Lopez[36], MaryAnn Oakley[36], Donna M. Simpson[36], Anton P. Porsteinsson[57], Bonnie S.

1060 Goldstein[57], Kim Martin[57], Kelly M. Makino[57], M. Saleem Ismail[57], Connie Brand[57], Ruth A.

1061 Mulnard[58], Gaby Thai[58], Catherine Mc Adams Ortiz[58], Kyle Womack[59], Dana Mathews[59], Mary

1062 Quiceno[59], Ramon Diaz Arrastia[59], Richard King[59], Myron Weiner[59], Kristen Martin Cook[59], Mi-

1063 chael DeVous[59], Allan I. Levey[60], James J. Lah[60], Janet S. Cellar[60], Jeffrey M. Burns[61], Heather

1064 S. Anderson[61], Russell H. Swerdlow[61], Liana Apostolova[62], Kathleen Tingus[62], Ellen Woo[62],

1065 Daniel H. S. Silverman[62], Po H. Lu[62], George Bartzokis[62], Neill R Graff Radford[63], Francine

1066 ParfittH[63], Tracy Kendall[63], Heather Johnson[63], Martin R. Farlow[25], Ann Marie Hake[25], Brandy

1067 R. Matthews[25], Scott Herring[25], Cynthia Hunt[25], Christopher H. van Dyck[64], Richard E. Carson[64],

1068 Martha G. MacAvoy[64], Howard Chertkow[65], Howard Bergman[65], Chris Hosein[65], Sandra

1069 Black[66], Bojana Stefanovic[66], Curtis Caldwell[66], Ging Yuek Robin Hsiung[67], Howard Feldman[67],

1070 Benita Mudge[67], Michele Assaly Past[67], Andrew Kertesz[68], John Rogers[68], Dick Trost[68], Charles

1071 Bernick[69], Donna Munic[69], Diana Kerwin[70], Marek Marsel Mesulam[70], Kristine Lipowski[70],

1072 Chuang Kuo Wu[70], Nancy Johnson[70], Carl Sadowsky[71], Walter Martinez[71], Teresa Villena[71],

1073 Raymond Scott Turner[72], Kathleen Johnson[72], Brigid Reynolds[72], Reisa A. Sperling[73], Keith A.

1074 Johnson[73], Gad Marshall[73], Meghan Frey[73], Jerome Yesavage[74], Joy L. Taylor[74], Barton Lane[74],

1075 Allyson Rosen[74], Jared Tinklenberg[74], Marwan N. Sabbagh[75], Christine M. Belden[75], Sandra A.

1076 Jacobson[75], Sherye A. Sirrel[75], Neil Kowall[76], Ronald Killiany[76], Andrew E. Budson[76],

1077 Alexander Norbash[76], Patricia Lynn Johnson[76], Thomas O. Obisesan[77], Saba Wolday[77], Joanne

1078 Allard[77], Alan Lerner[78], Paula Ogrocki[78], Leon Hudson[78], Evan Fletcher[79], Owen Carmichael[79],

1079 John Olichney[79], Charles DeCarli[79], Smita Kittur[80], Michael Borrie[81], T Y Lee[81], Rob Bartha[81],

1080 Sterling Johnson[82], Sanjay Asthana[82], Cynthia M. Carlsson[82], Steven G. Potkin[83], Adrian Preda[83],

1081 Dana Nguyen[83], Pierre Tariot[35], Stephanie Reeder[35], Vernice Bates[84], Horacio Capote[84], Michelle

1082 Rainka[84], Douglas W. Scharre[85], Maria Kataki[85], Anahita Adeli[85], Earl A. Zimmerman[86], Dzintra

1083 Celmins[86], Alice D. Brown[86], Godfrey D. Pearlson[87], Karen Blank[87], Karen Anderson[87], Robert

1084 B. Santulli[88], Tamar J. Kitzmiller[88], Eben S. Schwartz[88], Kaycee M. SinkS[89], Jeff D.

1085 Williamson[89], Pradeep Garg[89], Franklin Watkins[89], Brian R. Ott[90], Henry Querfurth[90], Geoffrey

1086 Tremont[90], Stephen Salloway[91], Paul Malloy[91], Stephen Correia[91], Howard J. Rosen[17], Bruce L.

1087 Miller[17], Jacobo Mintzer[92], Kenneth Spicer[92], David Bachman[92], Elizabether Finger[93], Stephen

1088 Pasternak[93], Irina Rachinsky[93], Dick Drost[93], Nunzio Pomara[94], Raymundo Hernando[94], Antero

Sarrael[94], Susan K. Schultz[95], Laura L. Boles Ponto[95], Hyungsub Shim[95], Karen Elizabeth

Smith[95], Norman Relkin[96], Gloria Chaing[96], Lisa Raudin[96], Amanda Smith[97], Kristin Fargher[97],

Balebail Ashok Raj[97]

[17] UC San Francisco, San Francisco, CA, USA

[18] UC San Diego, San Diego, CA, USA

[19] Mayo Clinic, Rochester, NY, USA

[20] UC Berkeley, Berkeley, CA, USA

[21] U Pennsylvania, Pennsylvania, CA, USA

[22] USC, Los Angeles, CA, USA

[23] UC Davis, Davis, CA, USA

[24] Brigham and Women's Hospital, Harvard Medical School, Boston, MA, USA

[25] Indiana University, Bloomington, IND, USA

[26] Washington University St. Louis, St. Louis, MO, USA

[27] University of Pennsylvania, Philadelphia, PA, USA

[28] Janssen Alzheimer Immunotherapy, South San Francisco, CA, USA

[29] University of Washington, Seattle, WA, USA

[30] University of London, London, UK

[31] USC School of Medicine, Los Angeles, CA, USA

[32] UCSF MRI, San Francisco, CA, USA

[33] University of Michigan, Ann Arbor, MI, USA

[34] University of Utah, Salt Lake City, UT, USA

[35] Banner Alzheimer's Institute, Phoenix, AZ, USA

[36] University of Pittsburgh, Pittsburgh, PA, USA

[37] UPenn School of Medicine, Philadelphia, PA, USA

[38] UC Irvine, Newport Beach, CA, USA

[39] Khachaturian, Radebaugh & Associates, Inc and Alzheimer's Association's Ronald and Nancy Reagan's Research Institute, Chicago, IL, USA

[40] General Electric, Boston, MA, USA

[41] Brown University, Providence, RI, USA

[42] National Institute on Aging/National Institutes of Health, Bethesda, MD, USA

[43] Oregon Health and Science University, Portland, OR, USA

[44] University of Southern California, Los Angeles, CA, USA

[45] University of California San Diego, San Diego, CA, USA

[46] Baylor College of Medicine, Houston, TX, USA

[47] Columbia University Medical Center, New York, NY, USA

1125   [48] Washington University, St. Louis, MO, USA

1126   [49] University of Alabama Birmingham, Birmingham, MO, USA

1127   [50] Mount Sinai School of Medicine, New York, NY, USA

1128   [51] Rush University Medical Center, Chicago, IL, USA

1129   [52] Wien Center, Vienna, Austria

1130   [53] Johns Hopkins University, Baltimore, MD, USA

1131   [54] New York University, New York, NY, USA

1132   [55] Duke University Medical Center, Durham, NC, USA

1133   [56] University of Kentucky, city of Lexington, NC, USA

1134   [57] University of Rochester Medical Center, Rochester, NY, USA

1135   [58] University of California, Irvine, CA, USA

1136   [59] University of Texas Southwestern Medical School, Dallas, TX, USA

1137   [60] Emory University, Atlanta, GA, USA

1138   [61] University of Kansas, Medical Center, Lawrence, KS, USA

1139   [62] University of California, Los Angeles, CA, USA

1140   [63] Mayo Clinic, Jacksonville, FL, USA

1141   [64] Yale University School of Medicine, New Haven, CT, USA

1142   [65] McGill Univ., Montreal Jewish General Hospital, Montreal, WI, USA

1143   [66] Sunnybrook Health Sciences, Toronto, ON, Canada

1144   [67] U.B.C. Clinic for AD & Related Disorders, British Columbia, BC, Canada

1145   [68] Cognitive Neurology St. Joseph's, Toronto, ON, Canada

1146   [69] Cleveland Clinic Lou Ruvo Center for Brain Health, Las Vegas, NV, USA

1147   [70] Northwestern University, Evanston, IL, USA

1148   [71] Premiere Research Inst Palm Beach Neurology, West Palm Beach, FL, USA

1149   [72] Georgetown University Medical Center, Washington, DC, USA

1150   [73] Brigham and Women's Hospital, Boston, MA, USA

1151   [74] Stanford University, Santa Clara County, CA, USA

1152   [75] Banner Sun Health Research Institute, Sun City, AZ, USA

1153   [76] Boston University, Boston, MA, USA

1154   [77] Howard University, Washington, DC, USA

1155   [78] Case Western Reserve University, Cleveland, OH, USA

1156   [79] University of California, Davis Sacramento, CA, USA

1157   [80] Neurological Care of CNY, New York, NY, USA

1158   [81] Parkwood Hospital, Parkwood, CA, USA

1159   [82] University of Wisconsin, Madison, WI, USA

1160   [83] University of California, Irvine BIC, Irvine, CA, USA

1161   [84] Dent Neurologic Institute, Amherst, MA, USA

1162   [85] Ohio State University, Columbus, OH, USA

1163   [86] Albany Medical College, Albany, NY, USA

1164   [87] Hartford Hosp, Olin Neuropsychiatry Research Center, Hartford, CT, USA

1165   [88] Dartmouth Hitchcock Medical Center, Albany, NY, USA

1166   [89] Wake Forest University Health Sciences, Winston-Salem, NC, USA

1167   [90] Rhode Island Hospital, Rhode Island, USA

1168   [91] Butler Hospital, Providence, RI, USA

1169   [92] Medical University South Carolina, Charleston, SC, USA

1170   [93] St. Joseph's Health Care, Toronto, Canada

1171   [94] Nathan Kline Institute, Orangeburg, SC, USA

1172   [95] University of Iowa College of Medicine, Iowa City, IA, USA

1173   [96] Cornell University, Ithaca, NY, USA

1174   [97] University of South Florida: USF Health Byrd Alzheimer's Institute, Tampa, FL 33613 USA

a Tau-Positive: 38.8%

b S1: Limbic | Inside: 12% | Outside: 88%
S2: MTL-Sparing | S4: L Temporal
S3: Posterior

c S1 (Limbic): 32.7% | S2 (MTL-Sparing): 17.8% | S3 (Posterior): 30.5% | S4 (L Temporal): 19.0%

d t: −8 −2 2 8

e SuStaIn Stage
6 11 16 21 26
S1
S2
S3
S4
Z: 2 7

a

S1: Limbic
Orig. Repl.

S2: MTL-Sparing
Orig. Repl.

S3: Posterior
Orig. Repl.

S4: L Temporal
Orig. Repl.

t
-8 -2 2 8

b

	S0: Tau-Negative	S1: Limbic	S2: MTL-Sparing	S3: Posterior	S4: L Temporal
S0: Tau-Negative	0.91	-0.19	-0.5	-0.75	-0.86
S1: Limbic	-0.75	0.78	-0.34	0.046	0.72
S2: MTL-Sparing	-0.44	-0.38	0.8	0.67	0.45
S3: Posterior	-0.47	0.081	0.17	0.86	0.33
S4: L Temporal	-0.68	-0.032	0.7	0.4	0.7

Original Subtype (y-axis)

Validation Subtype (x-axis)

similarity (r)

a Entorhinal Epicenter

b Best Epicenter

c Epicenter Probability

d MTL · Parietal · Frontal · Occipital · Temporal

e Cx

S1: Limbic
- Entorhinal, $r^2=0.704$
- Best: Entorhinal, $r^2=0.704$
- Subtype 1
- Obs. · Cx. · Pred.

S2: MTL-Sparing
- $r^2=0.038$
- Best: Middle Temporal, $r^2=0.266$
- Subtype 2
- Obs. · Cx. · Pred.

S3: Posterior
- $r^2=0.413$
- Best: Fusiform, $r^2=0.592$
- Subtype 3
- Obs. · Cx. · Pred.

S4: L Temporal
- $r^2=0.369$
- Best: Inferior Temporal, $r^2=0.503$
- Subtype 4
- Obs. · Cx. · Pred.

SuStaIn Stage

Tau-P

Disease Severity

Onset Age

Posterior
Slow progressing
Visuospatial Impairment
Older onset

Limbic
APOE4+
Less tau
Amnestic
Older onset

MTL-Sparing
APOE4-
Younger onset
Dysexecutive

L Temporal
Rapidly progressive
Multi-domain impairment
More tau
Asymmetric presentation

S0: No Tau   S1: Limbic   S2: MTL-sparing   S3: Posterior   S4: L Temporal

ADNI   BioF   AVID   Seoul   UCSF

S1: Limbic

S2: L-sparing

S3: Posterior

S4: L Temporal

2   Z   7

a

Tau (z)

('R_MTL_2';), ('L_MTL_2';), ('R_Temporal_2';), ('R_Occipital_2';), ('L_Temporal_2';), ('L_Occipital_2';), ('L_Parietal_2';), ('L_Occipital_5';), ('L_Temporal_5';), ('R_Parietal_2';), ('R_Temporal_5';), ('R_Occipital_5';), ('R_Parietal_5';), ('R_Occipital_10';), ('L_Parietal_5';)

('L_Parietal_2';), ('R_Parietal_2';), ('L_Temporal_2';), ('R_Temporal_2';), ('R_Frontal_2';), ('L_Frontal_2';), ('R_MTL_2';), ('L_MTL_2';), ('L_Occipital_2';), ('L_Parietal_5';), ('R_Parietal_5';), ('L_Temporal_5';), ('R_Temporal_5';), ('R_Occipital_2';), ('L_Parietal_10';), ('R_Parietal_10';), ('R_Occipital_5';)

b

S1 (Limbic)

S2 (MTL-Sparing)

Subject

S3 (Posterior)

S4 (L Temporal)

Subject

SuStaIn Stage

SuStaIn Stage

a

S1: Limbic   S2: MTL-Sparing   S3: Posterior   S4: L Temporal

Orig.   DD   Orig.   DD   Orig.   DD   Orig.   DD

-8   -2 2   8

t

b

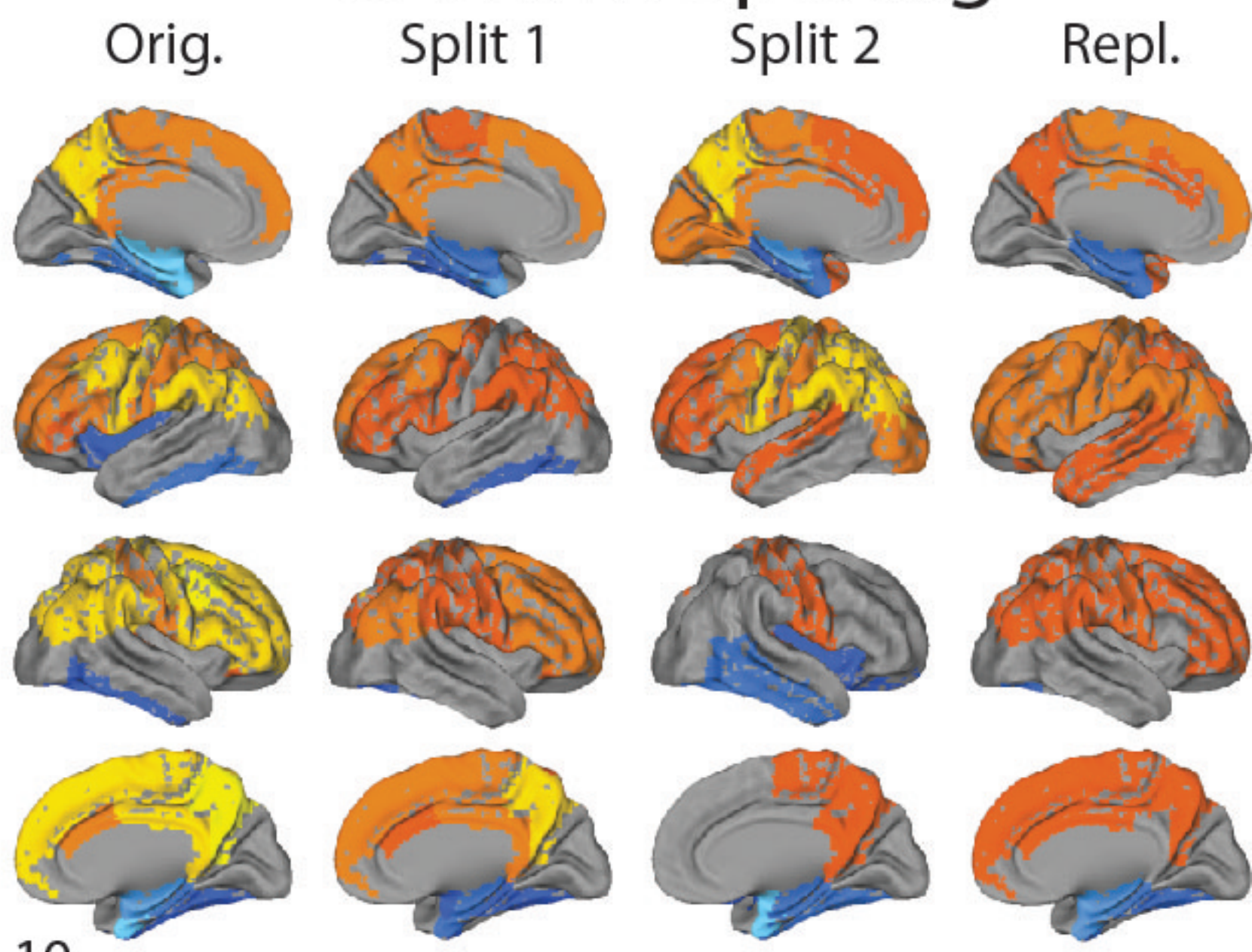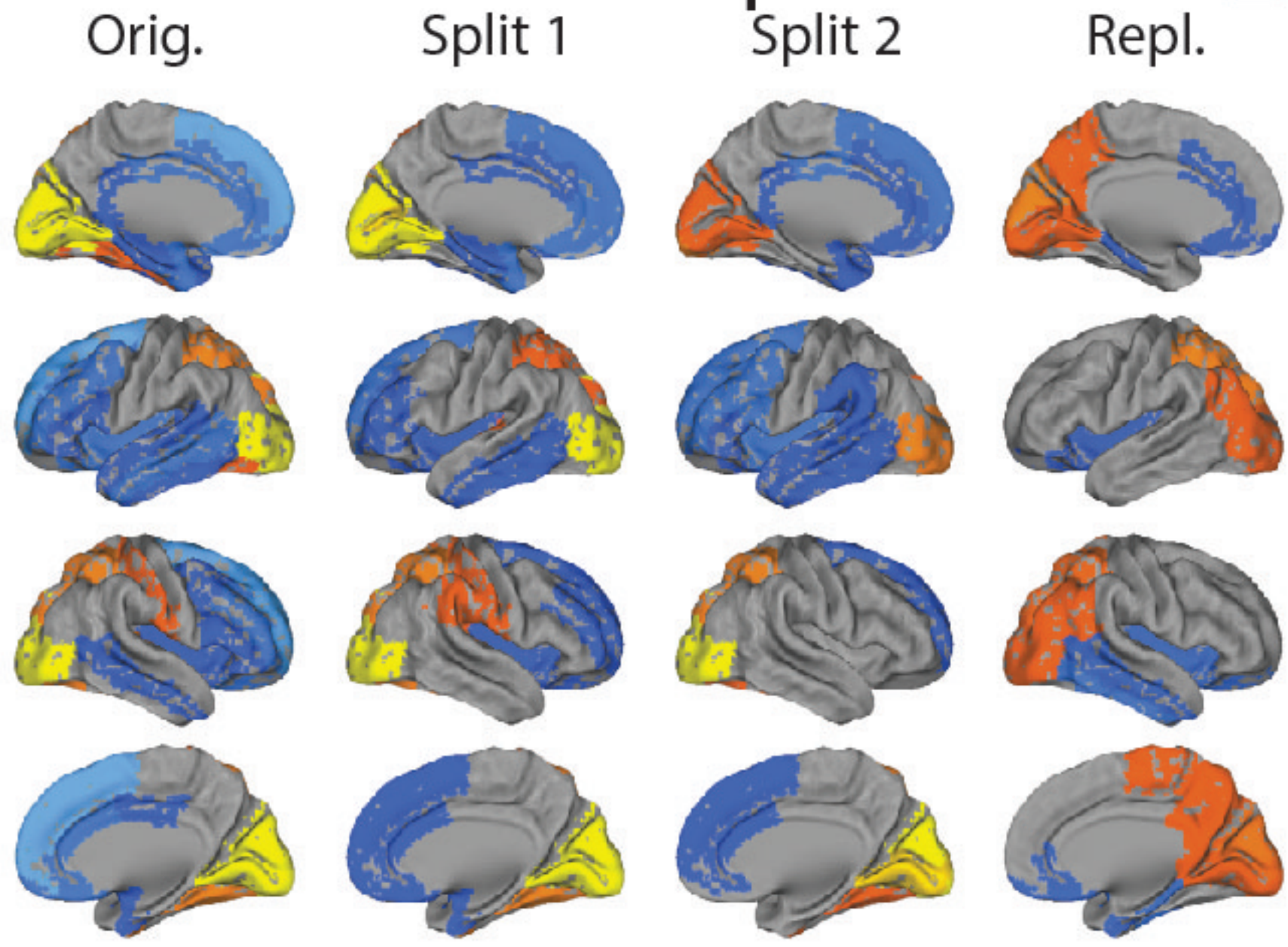|	S0: Tau-Negative	S1: Typical	S2: MTL-Sparing	S3: Occipital	S4: L Temporal
S0: Tau-Negative	1	-0.53	-0.72	-0.54	-0.77
S1: Typical	-0.51	0.98	-0.11	-0.16	0.17
S2: MTL-Sparing	-0.62	-0.22	0.98	0.46	0.69
S3: Occipital	-0.54	0.096	0.45	0.96	0.081
S4: L Temporal	-0.77	0.36	0.52	0.12	0.97

Subtype (Original Parameters)

Subtype (Data-Driven Parameters)

similarity (r)

0.9 0.6 0.3 0.0 -0.3 -0.6

S1: Typical

Orig.   Split 1   Split 2   Repl.

S2: MTL-Sparing

Orig.   Split 1   Split 2   Repl.

S3: Occipital

Orig.   Split 1   Split 2   Repl.

S4: L Temporal

Orig.   Split 1   Split 2   Repl.

t: -10   -2   2   10

similarity (r)

	S1: Limbic	S2: MTL-Sparing	S3: Posterior	S4: L Temporal
a Entorhinal	Entorhinal Epicenter $r^2=0.644$ mse=0.013	Entorhinal Epicenter $r^2=0.018$ mse=0.035	Entorhinal Epicenter $r^2=0.252$ mse=0.016	Entorhinal Epicenter $r^2=0.334$ mse=0.027
b Subtype-Specific	Best Epicenter (A20rv_R) $r^2=0.652$ mse=0.013	Best Epicenter (A37vl_L) $r^2=0.284$ mse=0.026	Best Epicenter (A20rv_R) $r^2=0.395$ mse=0.013	Best Epicenter (A20iv_L) $r^2=0.559$ mse=0.018

Axes: Observed Tau Probability (y), Predicted Tau Probability (x)

c Best Epicenter

d Epicenter Probability

Color scale: 0 0.17 0.35 0.52 0.7