1	PERSPECTIVES
2	The value of artificial stimuli in behavioral research:
3	Making the case for egg rejection studies in avian brood parasitism
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24 Abstract

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Experimentation is at the heart of classical and modern behavioral ecology research. The 26 manipulation of natural cues allows us to establish causation between aspects of the 27 environment, both internal and external to organisms, and their effects on animals' 28 behaviors. In recognition systems research, including the quest to understand the 29 30 coevolution of sensory cues and decision rules underlying the rejection of foreign eggs by 31 hosts of avian brood parasites, artificial stimuli have been used extensively, but not without 32 controversy. In response to repeated criticism about the value of artificial stimuli, we 33 describe four potential benefits of using them in egg recognition research, two each at the proximate and ultimate levels of analysis: (1) the standardization of stimuli for 34 35 developmental studies and (2) the disassociation of correlated traits of egg phenotypes used for sensory discrimination, as well as (3) the estimation of the strength of selection on 36 37 parasitic egg mimicry and (4) the establishment of the evolved limits of sensory and cognitive plasticity. We also highlight constraints of the artificial stimulus approach, and 38 39 provide a specific test of whether responses to artificial cues can accurately predict 40 responses to natural cues. Artificial stimuli have a general value in ethological research beyond research in brood parasitism, and may be especially critical in field studies involving 41 42 the manipulation of a single parameter, where other, confounding variables are difficult or 43 impossible to control experimentally or statistically. Keywords: artificial stimuli, brood 44 parasitism, egg rejection, recognition systems, research methods, unnatural

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#### 48 Background

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50 Over 50 years ago, Niko Tinbergen performed classic experiments to determine whether egg size and coloration affected avian parents' choices to incubate eggs or remove broken 51 eggshells and thereby reduce predation on nests (Tinbergen 1951, Tinbergen et al. 1962). By 52 53 testing competing predictions, drawn from alternative hypotheses at the same level of 54 analysis, Tinbergen endorsed and illustrated the value of the alternative hypothesis-testing 55 framework for evolutionary and mechanistic studies of animal behavior in the wild. A critical component of these experimental approaches was the use of artificial stimuli that were 56 inspired by natural forms, but they either mimicked or exaggerated aspects of those 57 58 through the use of artificial materials (e.g., oversized model eggs, and brighter painted 59 colors, and artificially larger spots, than seen in natural eggs). In this way, the experiments 60 limited and defined both the modality and the degree of variation within and among egg traits in order to best isolate those features that predictably elicited natural behaviors in 61 62 wild animals.

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Researchers have frequently and productively used painted model eggs, as well as dyed natural eggs in the search for the recognition cues used by hosts of brood parasitic birds to reject foreign eggs: well over 10,000 such egg rejection experiments have been completed (reviewed in Grim 2007). In most of these studies, a model or painted-over natural 'parasitic egg' is placed into an active nest and monitored for several days in order to determine 69 whether the egg is accepted, pecked, or ejected, or the nest is abandoned (e.g., Davies and
70 Brooke 1989, Antonov et al. 2009, Moskát et al. 2014).

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72 Yet, in recent years, both the value, and the general applicability of evolutionary conclusions 73 drawn about natural behaviors, through the use of artificial stimuli in egg rejection research, 74 have been repeatedly and openly questioned. Here, we define an artificial egg stimulus, as 75 any material and pigment that is not taken directly from nature; for example, according to 76 this definition, a natural or model egg dyed blue with a human-manufactured paint, to 77 resemble the avian-perception of the immaculate egg of an American robin Turdus 78 migratorius, is still an artificial stimulus (Croston and Hauber 2014). Accordingly, Honza and colleagues (2007) used artificial dyes, to test the chromatic basis of foreign egg rejection by 79 80 song thrush T. philomelos. Avian visual modelling (Avilés 2008) was then applied to the 81 reflectance spectra of the artifial colors used, and combined with experimental rates of egg 82 rejection to characterize, for the first time, the sensory-perceptual basis of egg recognition in birds (Cassey et al. 2008). Several studies followed these early works, including those 83 84 using conspecific eggs to characterize the fine scale perceptual cues causing egg rejection behavior in other host species (e.g., Avilés et al. 2010, Spottiswoode and Stevens 2010, 85 86 Stevens et al. 2013a), but some of these also included pointed criticism that experiments 87 with artificial egg colors, and the resulting perceptual modelling, were not relevant to 88 evolutionary and ecological studies of brood parasitism in natural contexts. Recently, we 89 prepared a new manuscript inspired by Honza et al. (2007), and eventually published it (Bán 90 et al. 2013) but during peer-review, we repeatedly encountered several incarnations of a knee-jerk reaction to our use of artificial stimuli to infer not only mechanistic but also 91

evolutionary conclusions from the study. David Lahti's commentary (2014) in this journal
unpacks some of these concerns to complement our commentary below.

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95 We disagree with the premise and conclusion about whether artificial stimuli can be used to explore evolutionary questions of brood parasites. Specifically, Tinbergen and his followers, 96 including ourselves, clearly recognize(d) that the mechanisms themselves are an evolved 97 98 phenotype that in turn influences the expression and outcome of selective pressures. Thus, 99 mechanistic and evolutionary questions are never uninformative about each other 100 (Taborsky 2014). Furthermore, for evolutionary studies aimed at understanding the causes 101 and consequences of natural variation of cues and responses, it is assumed and understood that extant variation is the result of evolutionary forces that have constrained it (Samaš et 102 103 al. 2014). In turn, extending or exploring the phenotype's variable space beyond the natural 104 range is precisely what we need to do to probe how selection might be acting on novel 105 traits.

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107 Additionally, from an evolutionary perspective, whether a parasitic egg is rejected because it is recognized as an egg or a non-egg (e.g., detritus, flower petal) in the nest cup, is 108 109 equivalent at the level of the fitness outcomes of responding to brood parasitism (i.e., egg 110 rejection: beneficial; egg acceptance: costly). In other words, no matter how and what hosts 111 perceive/interpret about the different objects (including eggs) that they see in the nest, the 112 only thing that matters from an ultimate/evolutionary perspective is the resulting fitness of 113 the host and the parasite. Conceptually, the same criticism can also apply for the use of a natural, non-mimetic cuckoo egg: it, too, may be rejected because the host considers it a 114

piece of flower petal or other detritus fallen in the nest, or it may accepted it because its 115 appearance is so different from the hosts' own eggs so as to not be considered an egg, but 116 117 instead an integral nest construction material. What and whether artificial (and natural) eggs placed into the nest are considered as "eggs" is an empirical question that requires 118 detailed and careful experimental analyses (reviewed by Guigueno and Sealy 2012) but 119 these questions should not be answered based on human (peer-reviewer's) a priori 120 121 interpretation of what a naturalistic stimulus should look like and what constitutes a 122 *caricature of nature'*. If anything, recent brood parasitism research has taught us that over 123 the course a handful of decades, hosts can evolve brand new egg coloration to evade the 124 costs of accepting mimetic parasite eggs (Spottiswoode and Stevens 2012), thus what may 125 be a caricature today, might be reality tomorrow.

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To illustrate our argument in the context of the aims of ethological research, we highlight four potential benefits of the use of artificial colors in the study of avian egg rejection behaviors; critically, again, these benefits span both the ultimate and proximate levels of analysis (Tinbergen 1963). We also use published data to illustrate to fellow researchers, and to respond to critics, how to assess whether experiments with artificial stimuli may be used to interpret natural variation in host responses to natural stimuli.

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134 Four potential benefits of artificial stimuli in egg rejection research:

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**136 1. The standardization of stimuli for developmental studies, with a focus on repeatability** 

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Individual hosts of brood parasites may consistently reject or accept naturally laid parasitic 138 eggs across repeated parasitism events, may switch from being acceptors to being rejecters, 139 140 (or vice versa), or may vary their responses based on other ecological cues. For example, 141 older oriental reed warblers Acrocephalus orientalis are more likely to reject common cuckoo Cuculus canorus eggs than are younger warblers (Lotem et al. 1992). To understand 142 the ontogenetic basis of egg recognition and rejection, including its experience dependence, 143 144 and the roles of learning and maternal effects, requires experimentation with a standardized set of stimuli across different time points of the host's lifespan (Samaš et al. 145 146 2011, Grim et al. 2014, Moskát et al. 2014). Because natural egg coloration changes within 147 days of laying in the nest (Moreno et al. 2011), as well as in storage under controlled 148 conditions (Cassey et al. 2010), and natural nests may be difficult to find in a timely manner 149 and the donor-species may be a protected or otherwise vulnerable taxon, it is not always 150 possible, and/or ethically justifiable, to use natural eggs as consistent stimuli for 151 developmental studies, including the study of repeatability. For example, repeatability estimation requires the use of identical stimulus across repeated experiments with the same 152 153 individual; as any two natural eggs are never identical, the only way to test repeatability robustly is through the use of artificial models (for details see Grim et al. 2014). 154

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## **2. Disassociation of correlated phenotypic traits of eggs used for sensory discrimination**

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Once it has been established from observational and experimental studies whether and to what extent hosts reject natural parasitic eggs, further use of natural eggs to understand the sensory basis of egg recognition is a heuristically limited approach (de la Colina et al. 2012).

Natural stimuli often show limited variability overall in multidimensional trait space, but 161 exhibit extensive covariation between specific traits (e.g., avian feather colors: Stoddard 162 163 and Prum 2011); for example, eggs of brown-headed cowbirds Molothrus ater, that are 164 always rejected by American robins, are always smaller in size, beige in background, and heavily maculated, compared to the larger and blue immaculate eggs of this host species 165 (Friedmann 1929). Any of these differing egg traits, or their combinations, may be the 166 167 possible recognition cue(s) for egg rejection, but these traits might simply be physiologically or structurally constrained to co-vary. Thus, using natural cowbird eggs exclusively as egg 168 169 rejection stimuli prevents testing the relative contribution of size, color, and maculation in 170 American robin's egg recognition process (Rothstein 1982, Croston and Hauber 2014). Instead, using unnatural combination of natural variation (e.g, small blue model eggs), 171 172 generates novel (artificial) models which can critically aid the characterization of the 173 proximate basis of the egg rejection cues used by hosts to eliminate parasitic eggs in the 174 nest.

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## 176 **3.** The estimation of the strength of selection on parasitic egg mimicry

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The rejection of parasitic eggs by hosts represents a critical selective pressure in the coevolutionary arms race that drives parasites to evolve increasingly mimetic eggs, which required increasingly fine-tuned sensory systems to be detected by hosts (Davies 2000). This is because female parasites have nil fitness when their eggs are rejected and, thus, represent an evolutionary dead end. Yet, some parasites lay highly mimetic eggs, many of which are still rejected, whereas other parasites lay inaccurately or poorly mimetic eggs,

most of which are accepted (Stoddard and Stevens 2011). To characterize comparative 184 patterns of egg rejection behaviors, and to reconstruct the evolutionary trajectories of how 185 186 rejection behaviors have changed with exposure to brood parasitism, requires a 187 standardized metric of egg rejection responses (Grim et al. 2011). These, by definition, cannot be based on responses to natural parasitic eggs, because the coevolutionary 188 hypothesis assumes a reciprocal and dynamic process between hosts and parasites, which 189 190 will result in varying degrees of host-brood parasite egg mimicry across different systems 191 (Igic et al. 2012). Instead, using a specific, variably rejected model egg color, can provide a 192 metric of egg rejection directly comparable across host populations and species. 193 Accordingly, analyzing the responses of different species of common cuckoo hosts in Europe against the same artificial egg color, revealed that more discriminating and rejecting hosts 194 195 are parasitized by perceptually more mimetic parasite eggs (Stoddard and Stevens 2011).

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### **4. Establishment of the evolved limits of sensory and cognitive plasticity**

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199 Presenting hosts exclusively with foreign eggs that are within the natural range of variation 200 can also lead to incorrect conclusions about whether hosts recognize and reject foreign 201 eggs. For example, some hosts, including the common redstart *Phoenicurus phoenicurus* 202 accept virtually all naturally laid parasitic common cuckoo eggs (Rutila et al. 2006). When 203 experimentally testing such a host's egg discrimination ability by introducing natural 204 parasite eggs laid elsewhere, model eggs painted to resemble them, or host eggs only 205 partially dyed, this host accepts most of these foreign egg types, too (Rutila et al. 2002, Hauber et al. 2014). The results would then lead to the conclusion that egg rejection as a 206

207 defence against parasites has not evolved in the redstart. However, there is a biologically critical, alternative functional explanation: that even if such hosts have evolved sensory 208 209 mechanisms to recognize increasingly similar foreign eggs, their recognition mechanisms 210 may be circumvented by the high accuracy of the coevolved mimicry of the parasitic egg's appearance; in other words, the cuckoo eggs are such a good match of the redstart eggs 211 that they cannot be discriminated and, thus, rejected by this host. This alternative 212 213 hypothesis can be directly tested solely through the use of artificial eggs that deviate in a 214 known direction from the phenotypic range of natural host and parasite eggs; using natural 215 eggs of other species, or even conspecifics, would introduce both tractable (measured) and 216 intractable (unmeasured) sources of variation. Once the host's ability to reject such non-217 matching eggs has been established, experimenters can move onto the use of better 218 matching (more mimetic) eggs in order to meaningfully isolate more proximate drivers of 219 egg rejection. Similarly, most of the grassland passerines that lay beige and spotted eggs, 220 accept all or nearly all beige and spotted cowbird-like eggs, but reject blue model eggs (Klippenstine and Sealy 2008). Importantly, in hosts that do reject non-mimetic eggs, the use 221 222 of increasingly mimetic models is needed to establish the sensory thresholds of these 223 discrimination abilities, and then to test whether these perceptual acceptance thresholds 224 function adaptively, i.e. allow the rejection foreign eggs to reduce the fitness costs of brood 225 parasitism (e.g. Croston and Hauber 2014).

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Responses to artificial stimuli can predict behaviors in response to natural stimuli: the case of
egg rejection by a brood parasite host

We recognize here that the use of artificial colors and/or materials can also be a severe 230 constraint on the utility of these experiments in evolutionary interpretations of egg 231 232 rejection data, for example when using treatments which only change the color of the egg in 233 spectral ranges not perceived by the subject (Avilés et al. 2006), or when model eggs are made from materials that cannot be pierced or grasped for successful ejection, despite 234 repeated rejection attempts by hosts (Antonov et al. 2009). Nonetheless, to evaluate our 235 236 specific claim that, contrary to our critics, experimentally induced behaviors in response to 237 artificial stimuli can help to explain both causation and pattern in fitness-relevant responses 238 to natural cues, we focused on our own published data (Bán et al. 2013, Moskát et al. 2014). 239 Specifically, we tested for a predictive relationship between the evolved behavior (egg ejection) and the artificial stimuli (dyed egg colors) at nests of the free-living great reed 240 241 warblers Acrocephalus arundinaceus, an intermediate rejecter host species of the common 242 cuckoo Cuculus canorus in central Hungary (Bán et al. 2013). From that study, we obtained 243 host responses to experimental parasitism with a single foreign egg (host egg dyed with a highlighter pen of one of five colors, n = 12-16 nests), and contrasted them with egg 244 245 rejection rates of a natural conspecific egg (moved a different host's nest, n = 16; Bártol et al. 2002), and a natural parasite egg (a cuckoo egg moved from a parasitized to a non-246 247 parasitized nest, n = 13 nests; C. Moskát, unpublished data). We then calculated a stimulus 248 metric that can be applied to both artificial and natural color stimuli: we measured avian-249 visible spectral reflectance (300-700 nm), and used perceptual modelling to estimate chromatic contrast distances between natural host eggs' background coloration and 250 251 stimulus egg coloration (Moskát et al. 2014).

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Our data points did not include the limits of rejection probabilities (0%, 100%), and so we 253 254 used a linear regression analysis between egg rejection rates and pairwise just noticeable 255 differences (chromatic JNDs, n = 8 randomized egg-pairs per color type; Fig. 1); the result 256 showed a significantly positive relationship between perceivable chromatic contrasts and egg rejection rates ( $R^2$  = 0.29,  $F_{5, 38}$  = 15.3, P = 0.0004). When we also plotted the mean 257 values of JNDs and experimentally induced rejection rates of single, natural conspecific eggs 258 259 or single, natural parasitic eggs amongst the data points from these artificial colors, the 260 natural eggs fell within the 95% confidence interval of the predicted means (Fig. 1); the combined model, including both artificial and natural eggs, was also significant ( $R^2 = 0.32$ , 261  $F_{6,45}$  = 20.7, P < 0.0001). The implication is that behavioral responses to natural stimuli are 262 within the range predicted by variation in behavioral responses elicited by diverse artificial 263 264 stimuli.

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266 Conclusions

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268 Conceptually, our arguments go far beyond studies on egg rejection by hosts of avian brood parasites, as similar dyeing treatments are also used for experimental studies on nest 269 predation (Weidinger 2001), nest mate recognition (Tibbetts 2002), and in many other 270 271 experimental fields of animal ecology, evolution, and behavior (Ferrari et al. 2008). For 272 example, artificial stimuli that fall far outside of range of natural stimuli proved to be useful in non-brood parasitism studies, e.g., camouflage (Stevens et al. 2013b) and sexual selection 273 274 (Safran et al. 2010). Here, we argue that experimental studies with wild animals should not be classified a priori as strictly mechanistic, and discarded as irrelevant to fitness, on the 275

basis that manipulations involve artificial stimuli in quantity, in quality, or in both. Instead, 276 artificial stimuli should be appreciated and utilized when these allow for the careful design, 277 alteration, and delivery of exact cues and triggers that elicit fitness-relevant responses in 278 279 freely behaving animals. This is especially relevant for studies in the wild, where other social 280 and ecological cues and contexts are typically uncontrolled, and most also remain unmeasured. In turn, the possibility to design specific stimulus types that vary (only) along 281 282 known trait dimension(s), remains the core strength of behavioral experimentation. 283 Implementing diverse, and yet standardized stimuli can be informative for both proximate, 284 mechanistic questions about developmental and cognitive processes, and for ultimate, 285 comparative analyses of predicted behavioral responses induced by these stimuli, and their consequences on fitness. However, we also recognize that there are limits to the use and 286 utility of artificial stimuli in the study of evolutionary processes (see Lahti 2014 287 288 commentary). To address these concerns empirically, we recommend (and illustrate above) 289 the use of statistical checks to assess whether chosen stimuli, and/or the behavioral 290 responses elicited by these, fit or predict the known range of responses elicited by natural 291 stimuli.

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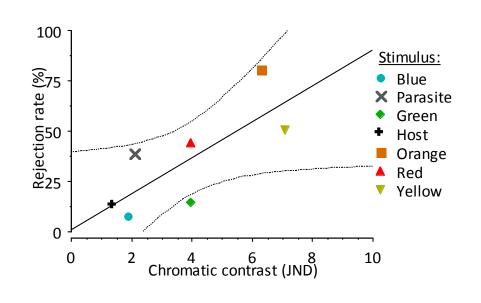
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*Figure 1.* The relationship between egg rejection rates of great reed warblers in response to experimentally introduced eggs, and avian perceivable distances (chromatic JNDs) between natural coloration of the host's own eggs and the artificial coloration of artificially dyed natural eggs, as well as of natural conspecific and natural parasitic, common cuckoo eggs. The graph depicts the mean JND and the percent of rejection per egg type, the regression line (solid), and its 95 % confidence intervals (dotted lines).

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317 *References:* 

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319	Antonov A, Stokke BG, Mosknes A, Røskaft E 2009. Evidence for egg discrimination
320	preceding failed rejection attempts in a small cuckoo host. Biology Letters 5: 169-
321	171.
322	Avilés JM 2008. Egg color mimicry in the common cuckoo Cuculus canorus as revealed by
323	modelling host retinal function. Proceedings of the Royal Society of London B 275:
324	2345-2352.
325	Avilés JM, Soler JJ, Pérez-Contreras T, Soler M, Møller AP 2006. Ultraviolet reflectance of
326	great spotted cuckoo eggs and egg discrimination by magpies. Behavioral Ecology 17:
327	310-314.
328	Avilés JM, Vikan JR, Fossøy F, Antonov A, Moksnes A, Røskaft E, Stokke BG. 2010. Avian
329	colour perception predicts behavioural responses to experimental brood parasitism
330	in chaffinches. Journal of Evolutionary Biology 23: 293-301.
331	Bán M, Moskát C, Barta Z, Hauber ME 2013. Simultaneous viewing of own and parasitic eggs
332	is not required for egg rejection by a cuckoo host. Behavioral Ecology 24: 1014-1021.
333	Bártol I, Karcza Z, Moskát C, Røskaft E, Kisbenedek T 2002. Responses of great reed
334	warblers Acrocephalus arundinaceus to experimental brood parasitism: the effects of
335	a cuckoo Cuculus canorus dummy and egg mimicry. Journal of Avian Biology 33: 420-
336	425.

337	Cassey P, Honza M, Grim T, Hauber ME 2008. The modelling of avian visual perception
338	predicts behavioural rejection responses to foreign egg colours. Biology Letters 4:
339	515-517.

- Cassey P, Maurer G, Duval C, Ewen JG, Hauber ME (2010) Differential impacts of time since
- 341 collection on eggshell colour components relevant to perceptual modeling in
- museum collections of song thrush (*Turdus philomelos*) eggs. Behavioral Ecology and
   Sociobiology 64: 1711–1720.
- 344 Croston R, Hauber ME 2014. Spectral tuning and perceptual differences do not explain the
- rejection of brood parasitic eggs by American robins (*Turdus migratorius*). Behavioral
- 346 Ecology and Sociobiology 68: 351-362.

347 Davies NB 2000. Cuckoos, cowbirds and other cheats. Poyser, London.

348 Davies NB, Brooke MDL 1989. An experimental study of co-evolution between the cuckoo,

- Cuculus canorus, and its hosts. I. Host egg discrimination. Journal of Animal Ecology
  58: 207-224.
- de la Colina MA, Pompillo L, Hauber ME, Reboreda JC, Mahler B 2012. Different recognition
- 352 cues reveal the decision rules used for egg rejection by hosts of a variably mimetic
   353 avian brood parasite. Animal Cognition 15: 881-889.

354 Ferrari MCO, Messier F, Chivers DP 2008. Can prey exhibit threat-sensitive generalization of

- 355 predator recognition? Extending the predator recognition continuum hypothesis.
- 356 Proceedings of the Royal Society of London B 275: 1811-1816.
- 357 Friedmann H 1929. The cowbirds: a study in the biology of social parasitism. Charles C.

358 Thomas, Springfield, IL.

359 Grim T 2007. Equal rights for chick brood parasites. Annales Zoologici Fennici 44: 1-7.

360	Grim T, Samaš P, Moskát C, Kleven O, Honza M, Moksnes A, Røskaft E, Stokke BG 2011.
361	Constraints on host choice: why do parasitic birds rarely exploit some common
362	potential hosts? Journal of Animal Ecology 80: 508-518.
363	Grim T, Samaš P, Hauber ME (2014) The repeatability of avian egg ejection behaviors across
364	different temporal scales, breeding stages, female ages and experiences. Behavioral
365	Ecology and Sociobiology 68: 749-759.
366	Guigueno MF, Sealy SG (2012) Nest sanitation in passerine birds: implications for egg
367	rejeciton in hosts of brood parasites. Journal of Ornithology 153: 35-52.
368	Hauber ME, Moskát, C, Bán M 2006. Experimental shift in hosts' acceptance threshold of
369	inaccurate-mimic brood parasite eggs. Biology Letters 2: 177-180.
370	Hauber ME, Samaš P, Anderson MG, Rutila J, Low J, Cassey P, Grim T (2014) Life-history
371	theory predicts host behavioural responses to experimental brood parasitism.
372	Ethology Ecology & Evolution (online).
373	Honza M, Polaciková L, Procházka P 2007. Ultraviolet and green parts of the colour
374	spectrum affect egg rejection in the song thrush (Turdus philomelos). Biological
375	Journal of the Linnean Society 92: 269-276.
376	lgic B, Cassey P, Grim T, Greenwood DR, Moskát C, Rutila J, Hauber ME 2012. A shared
377	chemical basis of avian host-parasite egg colour mimicry. Proceedings of the Royal
378	Society of London B 279: 1068-1076.
379	Klippenstine DR, Sealy SG 2008. Differential ejection of Cowbird eggs and non-mimetic eggs

- by grassland passerines. Wilson Journal of Ornithology 120: 667-673.
- Lahti D (2014) The limits of artificial stimuli in behavioral research: the umwelt gamble.

382 Ethology (this issue).

383	Lotem A, Nakamura H, Zahavi A 1992. Rejection of cuckoo egg in relation to host age: a
384	possible evolutionary equilibrium. Behavioral Ecology 3: 128-132.
385	Moreno J, Lobato E, Morales J 2011. Eggshell blue-green colouration fades immediately
386	after oviposition: a cautionary note about measuring natural egg colours. Ornis
387	Fennica 88: 51-56.
388	Moskát C, Takasu F, Muñoz RA, Nakamura H, Bán M, Barta Z 2012. Cuckoo parasitism on
389	two closely-related Acrocephalus warblers in distant areas: a case of parallel
390	coevolution? Chinese Birds 3: 320-329.
391	Moskát C, Zölei A, Bán M, Elek Z, Tong L, Geltsch N, Hauber ME 2014. How to spot a
392	stranger's egg? A mimicry-specific discordancy effect in the recognition of parasitic
393	eggs. Ethology 120: 616-626.
394	Moskát C, Bán M, Hauber ME 2014. Naïve hosts of avian brood parasites accept foreign
395	eggs, whereas older hosts fine-tune foreign egg discrimination during laying.
396	Frontiers in Zoology 11: 45.
397	Rothstein SI 1982. Mechanisms of avian egg recognition: which egg parameters elicit
398	responses by rejector species? Behavioral Ecology and Sociobiology 11: 229-239.
399	Rutila J, Latja R, Koskela K 2002. The Common Cuckoo Cuculus canorus and its cavity nesting
400	host, the Redstart Phoenicurus phoenicurus: a peculiar cuckoo-host system? Journal
401	of Avian Biology 33: 414-419.
402	Rutila J, Jokimäki J, Avilés JM, Kaisanlahti-Jokimäki ML 2006. Responses of currently
403	parasitized and unparasitized common redstart (Phoenicurus phoenicurus)
404	populations against artificial cuckoo parasitism. Auk 123: 259-265.

405	Safran RJ, Vitousek MN, Hauber ME, Ghalambor CK 2010. Sexual selection: a dynamic state
406	of affairs. Trends in Ecology and Evolution 25: 429-430.
407	Samaš P, Hauber ME, Cassey P, Grim T 2011. Repeatability of foreign egg rejection: Testing
408	the assumptions of co-evolutionary theory. Ethology 117: 606-619.
409	Samaš P, Hauber ME, Cassey P, Grim T 2014: The evolutionary causes of egg rejection in
410	European thrushes ( <i>Turdus</i> spp.): a reply to M. Soler. Frontiers in Zoology 11: 72.
411	Spottiswoode CN, Stevens M 2010. Visual modeling shows that avian host parents use
412	multiple visual cues in rejecting parasitic eggs. Proceedings of the National Academy
413	of Sciences USA 107: 8672-8676.
414	Spottiswoode CN, Stevens M 2012. Host-parasite arms races and rapid changes in bird egg
415	appearance. American Naturalist 179: 633-648.
416	Stevens M, Troscianko J, Spottiswoode CN 2013a. Repeated targeting of the same hosts by a
417	brood parasite compromises host egg rejection. Nature Communications 4: 2475.
418	Stevens M, Marshall KLA, Troscianko J, Finlay S, Burnand D, Chadwick SL 2013b. Revealed by
419	conspicuousness: distractive markings reduce camouflage. Behavioral Ecology. 24:
420	213-222.
421	Stoddard MC, Prum RO 2011. How colorful are birds? Evolution of the avian plumage color
422	gamut. Behavioral Ecology 22: 1042-1052.
423	Stoddard MC, Stevens M 2011. Avian vision and the evolution of egg color mimicry in the
424	common cuckoo. Evolution 65: 2004-2013.

- Taborsky M 2014. Tribute to Tinbergen: The four problems of biology. A critical appraisal.
- 426 Ethology 120: 224–227.
- 427 Tibbetts EA 2002. Visual signals of individual identity in the paper wasp *Polistes fuscatus*.

428 Proceedings of the Royal Society London B 269: 1423-1428.

- 429 Tinbergen N 1951. The study of instinct. Oxford University Press, New York.
- 430 Tinbergen N, Broekkhuysen GJ, Feekes F, Houghton JCW, Kruuk H, Szulc E 1962. Eggshell
- 431 removal by the Black-headed Gull, *Larus ridibundus* L.: a behaviour component of
- 432 camouflage. Behaviour 19: 74-117.
- 433 Tinbergen N 1963. On aims and methods of ethology. Zeitschrift für Tierpsychologie 20: 410-
- 434 433.
- 435 Weidinger K 2001. Does egg colour affect predation rate on open passerine nests?
- 436 Behavioral Ecology and Sociobiology 49: 456-464.