

# Synthesis and structure of novel ferrocene-containing $\beta$ -carboline derivatives with the elements of planar-, central- and conformational chirality

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**Abstract:** – Employing tryptamine or tryptophane methylester hydrochloride and (*S<sub>p</sub>*)-2-formylferrocene-1-carboxylate as precursors by means of Pictet-Spengler reactions and subsequent intramolecular acylation effected by cyanuric fluoride or carbonyldiimidazole (CDI), the first representatives of ferrocene-containing  $\beta$ -carboline derivatives including polycyclic ferroceno/fused lactames, were prepared. In the course of CDI-mediated tandem cyclization of a tryptophane-derived carboxyferrocenyl-substituted  $\beta$ -carboline, a ring enlargement simultaneously taking place with the loss of the  $\eta^5$ -C<sub>5</sub>H<sub>5</sub>Fe<sup>+</sup> fragment of the fused ferrocene moiety, effected by the coordinating imidazole released from the reagent, led to a 4*H*-cyclopenta[7,8]azonino[5,4-*b*]indole as a minor product. The constitution and relative configuration of the new compounds with the elements of planar-, central- and conformational chirality were established by NMR methods including HMQC, HMBC and NOESY measurements supported by DFT modeling studies.

**Keywords:** Condensed ferrocene;  $\beta$ -carboline; Tandem diastereoselective cyclization; Loss of iron from ferrocene; Planar chirality; Conformational chirality; DFT modeling; NMR spectroscopy

## 1. Introduction

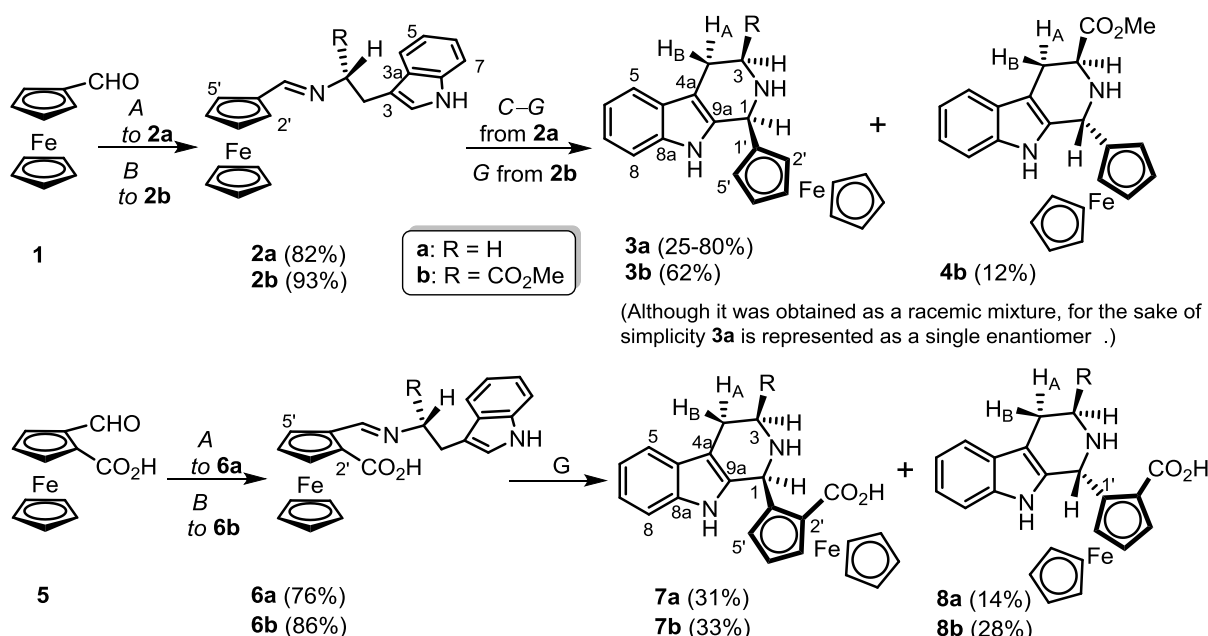
There is a continuous interest in the  $\beta$ -carboline alkaloids representing an important class of synthetic and naturally occurring heterocycles with a wide spectrum of valuable pharmacological properties [1–6]. In the recent decades it was also demonstrated that  $\beta$ -carboline alkaloids may have a pronounced potential as anti-cancer agents [7–16], which can act through different mechanisms including eg. DNA-intercalation [17], inhibition of Topoisomerase I and II [18] and CDK (cyclin-dependent kinases) [19, 20]. On the other hand, it is well documented that the replacement of an aromatic nucleus in certain organic compounds for a ferrocene unit can lead to such products which possess enhanced biological activity relative to that displayed by the parent molecule [21], and a variety of functionalized ferrocenes with relatively simple structures has been shown to exhibit valuable biological effects [22–32]. In this regard our group has also synthesized and characterized a wide variety of ferrocene-containing heterocycles of potential biological importance [33–46] and – in four cases [47–50] – with proved *in vitro* anticancer activity against human malignant cell lines. Since to our best knowledge metallocene-based  $\beta$ -carbolines are not known in the literature, in the frame of our ongoing research we targeted the preparation and structural analysis of ferrocene-containing heterocycles with 2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole residue including diastereomeric 6,7,12,12*b*-tetrahydro-4*H*-ferroceno[1,2]indolizino[8,7-*b*]indol-4-ones, the first representatives of novel classes of alkaloid-like fused metallocenes with the elements of planar-, central- and conformational chirality which will be subjected to *in vitro* anti-cancer assays on selected human malignant cell lines.

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## 2. Results and Discussion

### 2.1. Optimization of Pictet-Spengler conditions

At the beginning of the synthetic pathway we searched for optimal conditions for the crucial Pictet-Spengler cyclizations of imines **2a,b** and **6a,b** resulted from facile condensations of tryptamine-based amine components with formylferrocene (**1**) and (*S<sub>p</sub>*)-2-formylferrocene-1-carboxylic acid (**5**) [46], respectively, (cf. Methods A and B, Scheme 1). The cyclization of **2a**, selected as model precursor with the simplest structure in hand, was first attempted by four methods (C–G) employing different acidic conditions to construct 1-ferrocenyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole **3a**. Since the prolonged treatment (24 h) of **2a** with acetic acid at reflux temperature proved to be the most efficient protocol (Method G) to promote the formation of **3a** (isolated yield 80%), the same conditions were applied for the cyclization of iminoester **2b** affording an approximately 5:1 mixture of diastereomers **3b** and **4b**. The cyclizations of tryptophane-based imines **6a,b**, were also performed under the conditions of Method G resulting in zwitterionic β-carboline carboxylates as mixtures of diastereomeric pairs **7a/8a** and **7b/8b**, respectively. Although **8a** could be separated as a minor product, the ester derivatives **7b** and **8b** were isolated in comparable yields (33% and 28%, respectively).



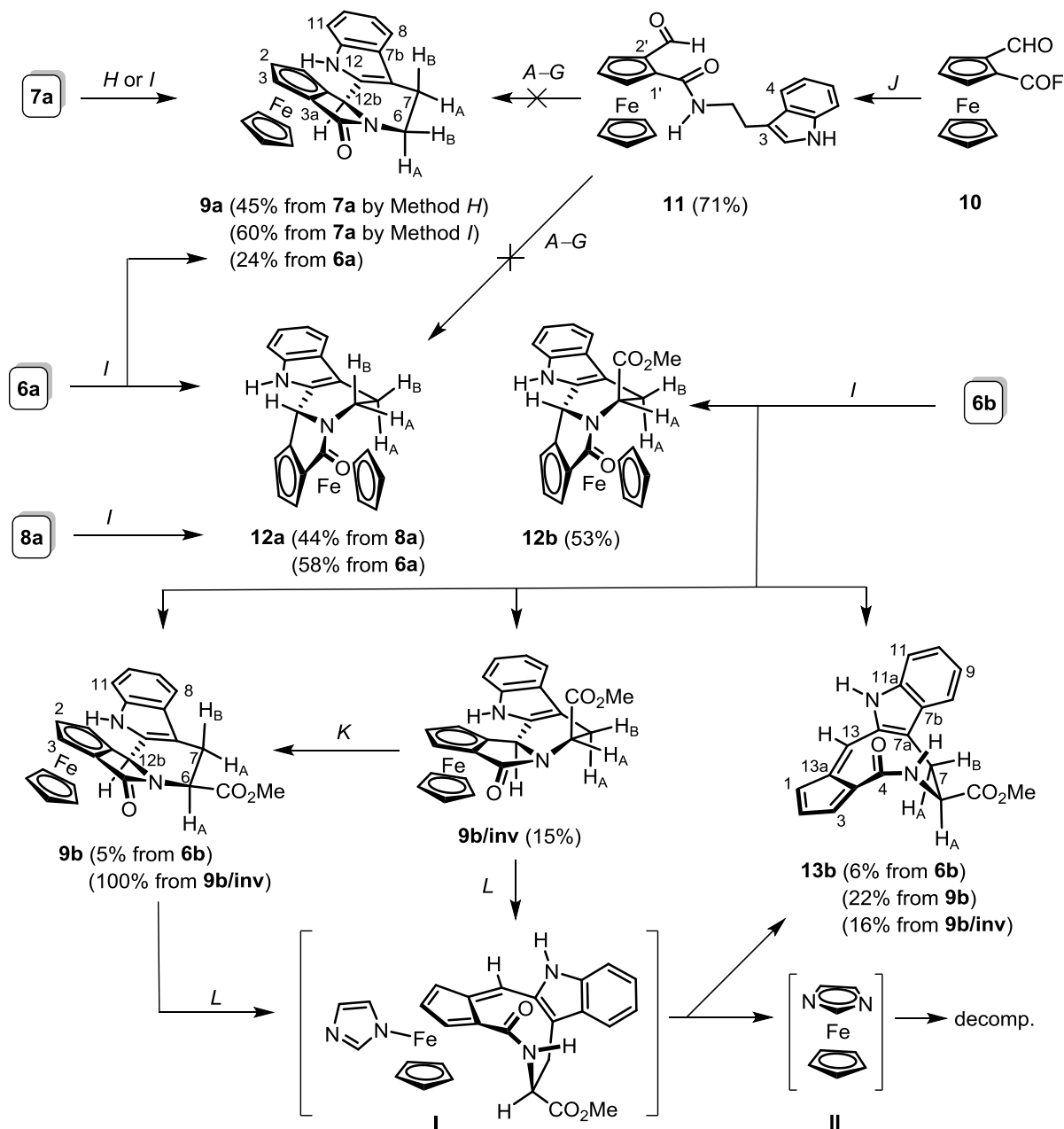
Reaction conditions: A) Tryptamine, EtOH, reflux 4 h, Ar; B) L-Tryptophan methylester hydrochloride, DCM, Na<sub>2</sub>CO<sub>3</sub>, reflux 30 min. Ar; C) HClO<sub>4</sub>/EtOH, reflux, 24h, Ar; D) HClO<sub>4</sub>/AcOH, reflux, 4h, Ar; E) TFA/AcOH, reflux, 4h, Ar; F) H<sub>3</sub>BO<sub>3</sub>/AcOH, reflux, 4h, Ar; G) AcOH, reflux, 24 h, Ar.

### Scheme 1

With the intention of constructing novel ferroceno-fused ring systems with alkaloid-like structures of potential biological interest first we attempted the cyclization of **7a** by two versions of carboxyl-activation effected by cyanuric fluoride and carbonyldiimidazole (CDI) in the presence of catalytic amount of triethylamine hydrochloride representing Methods H and I, respectively (Scheme 2). The reactions gave the fused lactame **9a**, with “*S*”-configuration at the C12b stereogenic center and “*M*” helical chirality associated with the conformation adopted by the tetrahydropyridine ring (referred as ring C in the subsequent discussion), in mediocre yields (45% and 60%). The analogous cyclization

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of **8a** was carried out by Method *I* resulting in diastereomeric lactame **12a** with “*R*”-configuration at the C12b stereogenic center and “*P*” helical chirality of ring *C* in acceptable yield (44%). (The structure determination of the fused lactames will be discussed in detail in the next session.) We also checked the accessibility of **9a** and **12a** by a reverse sequence involving primary acylation followed by acid-catalyzed tandem cyclization. However, amide **11** obtained by a facile reaction of tryptamine with (*S<sub>p</sub>*)-2-formyl-1-fluorocarbonylferrocene (**10**) [46] in THF (Method *J*) resisted to any further conversions under each condition proved to be efficient in promoting Pictet-Spengler cyclization of imine-type substrates (Scheme 2).



**Reaction conditions:**

A–G: cf. Scheme 1; *H*: cyanuric fluoride, DCM, rt, 24 h, Ar; *I*: i.) CDI (1.2 eq.) / Et<sub>3</sub>N.HCl (0.1 eq.), DMF 50 °C, 30 min., Ar; ii.) MeOH, rt, Ar; *J*: tryptamine, THF, rt, 24 h, Ar; *K*: DMSO-*d*<sub>6</sub>, 90 °C, 30 min.; *L*: imidazole/DMF 110 °C, 20 min. Ar.

**Scheme 2**

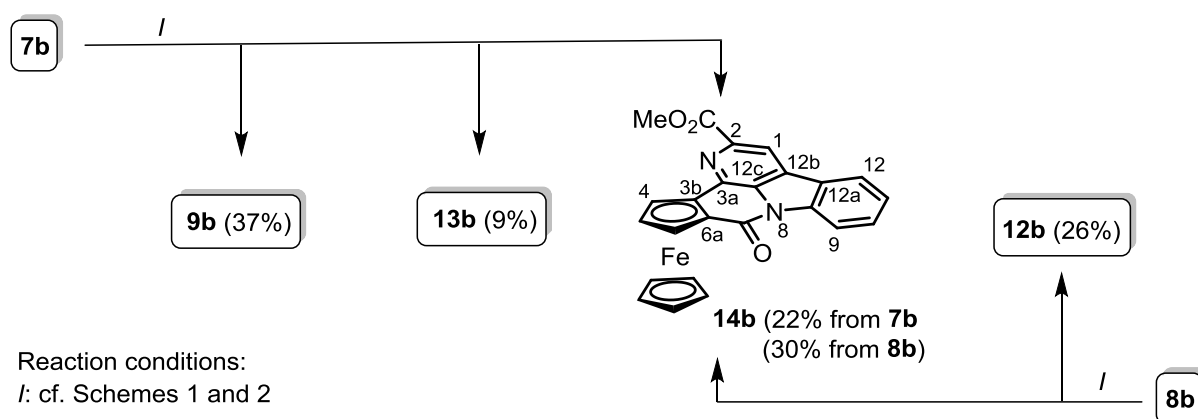
Finally, a third version of sequential cyclization procedure employing Method *I* based on primary carboxyl-activation was attempted to convert imine **6a** into fused lactames (**9a** and **12a**) in a

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single operation. To our surprise, the reaction resulted in diastereomer **12a** as the major product (58%), while **9a** was formed as the minor component isolated in low yield (24%). This one-pot protocol also proved to be successful in promoting a tandem cyclization of tryptophane-based imine **6b** to construct diastereomeric esters **12b** (53%), **9b** (5%) and **9b/inv** (15%). Pointing to the relative stability of the separable conformers, on heating in DMSO-*d*<sub>6</sub> solution in an NMR tube at 90 °C under an argon atmosphere for ca. 30 min (Method *K*), **9b/inv** with “*P*” helical chirality of ring *C* was completely transformed into **9b** with “*M*” helical chirality of ring *C*.

Besides the ferroceno-fused  $\gamma$ -lactams a fulvene-fused azoninone (**13b**) with *E*-lactame unit could also be isolated in low yield (6%) from the reaction mixture. The formation of this purely organic product takes place by *trans*-annular opening of the fused  $\gamma$ -lactame fragment along the C12b-N5 bond associated with decomposition of the ferrocene residue, probably induced by imidazole of CDI-origin. This view gains support from the imidazole-mediated transformations of **9b** and **9b/inv** carried out in DMF at 110 °C affording **13b** in low yields (22% and 16%, respectively). The modified conditions employing elevated temperatures or prolonged reaction times did not led to higher yields of **13b**, but resulted in the formation of substantial amounts of tarry substances. This transformation can be interpreted by the coordination of imidazole to the  $\eta^5$ -C<sub>6</sub>H<sub>5</sub>Fe<sup>+</sup> residue to give intermediate **I** which undergoes fragmentation affording **13b** and 1,3-diazaferrocene **II**. Probably due to its uncontrolled decomposition processes **II** could not be isolated from the reaction mixture.

The cyclization reactions of esters **7b** and **8b** were also accomplished under the conditions of Method *I* employing CDI as coupling reagent. Accordingly, the reaction of **7b** afforded a mixture of **9b** (37%), azoninone **13b** (9%) and  $\delta$ -lactame **14b** (22%) as a result of the acylation of N2- and N9 atoms, respectively (Scheme 3). The decreased propensity of N2 to undergo intramolecular acylation is probably due to the proximity of the bulky methoxycarbonyl group, thus increasing the chance of indole N9 atom to be involved in the cyclization step accompanied by spontaneous dehydrogenation of the tetrahydropyridine ring finally affording **14b**. Under the same conditions  $\beta$ -carboline **8b** got converted again into  $\delta$ -lactame **14b** and  $\gamma$ -lactame **12b** in comparable yields (30% and 26%, respectively). Since in **12b** the  $\eta^5$ -C<sub>6</sub>H<sub>5</sub>Fe<sup>+</sup> fragment is situated in the *endo* position of decreased accessibility, this  $\gamma$ -lactame seems to resist the imidazole-mediated *trans*-annular ring opening finally leading to **13b**. It must be pointed out here that – contrary to **9b** and **9b/inv** – **9a** did not undergo an analogous transformation with the cleavage of the C12b-N5 bond, which is probably facilitated by the electron withdrawing effect of the adjacent methoxycarbonyl group.



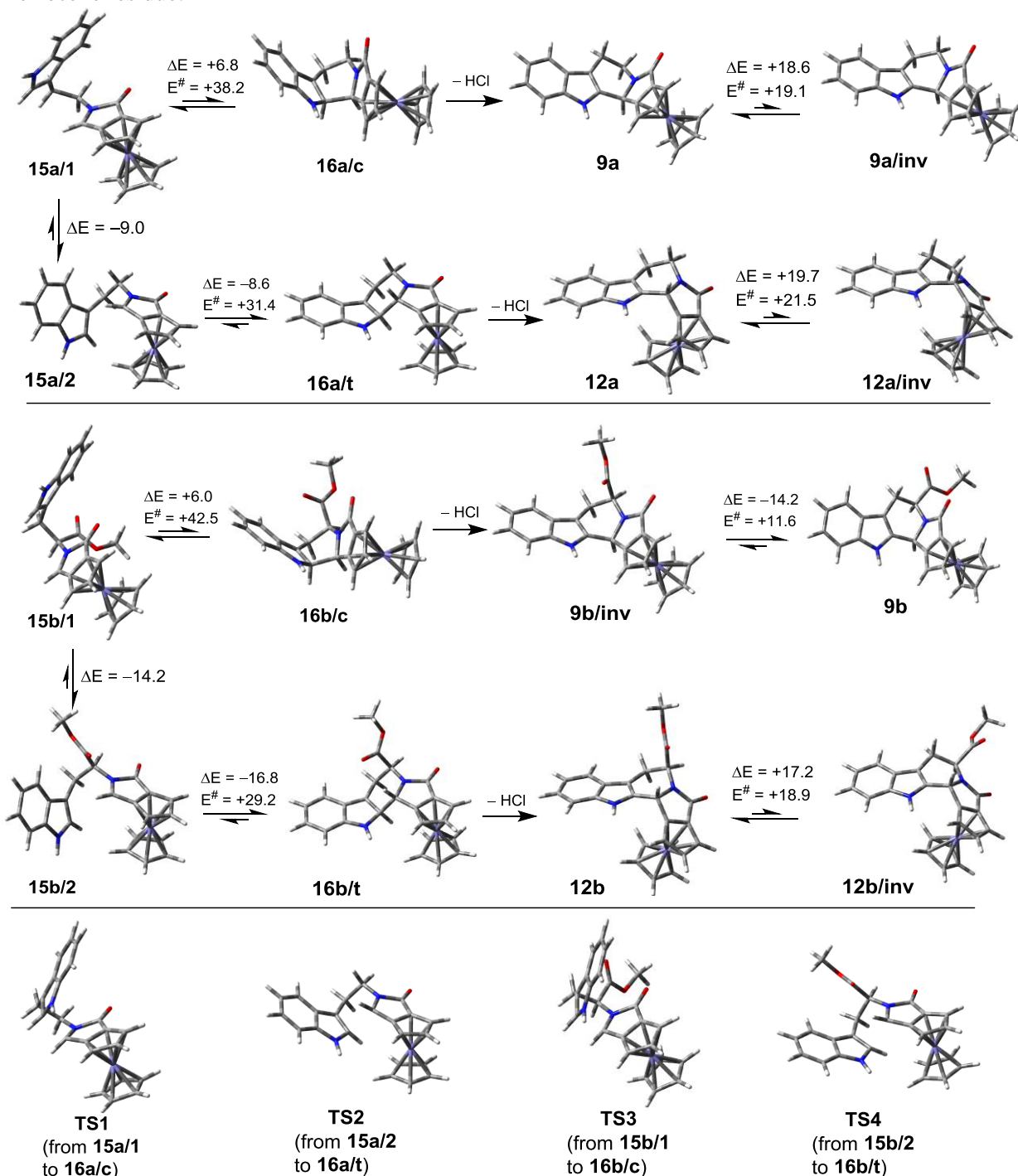
### Scheme 3.

From the aspect of the stereochemical outcome of the tandem cyclizations of imines **6a,b**, it must be pointed out that under the conditions promoting the primary activation of the carboxyl group, the formation of the isomers with “*R*”-configuration at the C12b stereogenic center is preferred over that of the diastereomers with “*S*”-configuration at the same stereogenic center irrespective of the helical chirality of ring *C* (isolable yields: 58%/24% for **12a/9a** and 53%/(15%+5%) for **12b/(9b/inv+9b)**, respectively). On the basis of the results of the experiment conducted under forced conditions (cf. Method *K*: Scheme 2) and supported by the modeling studies discussed below, it seems

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that it is **9b/inv** which is formed as the primary product in the tandem cyclization reaction of **6b**. In order to disclose the pathways of CDI-promoted tandem cyclization reactions accounting for this diastereoselectivity, we undertook comparative DFT modeling studies [51] carried out at B3LYP level of theory [52] with 6-31 G(d,p) basis set [53] on the cyclic *N*-acyliminium intermediates **15a** and **15b** supposed to be resulted from the cyclization of **6a,b**, and stabilized by the highly electron-donor ferrocene residue.



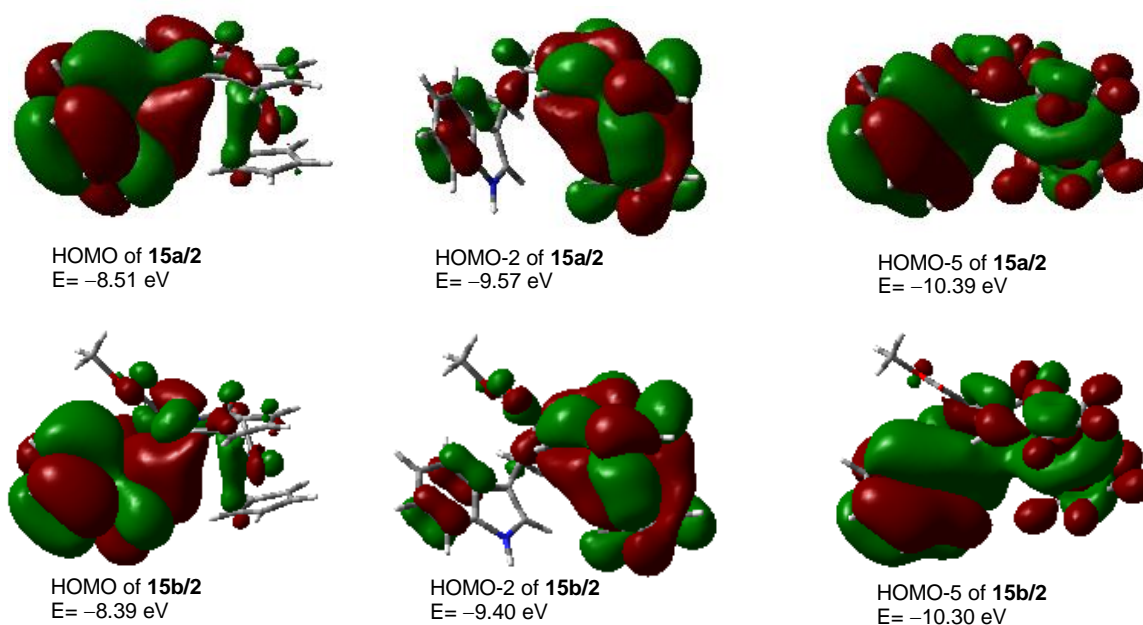
Intermediates types **15** and **16** as well as TS structures **TS1-4** are cations with chloride as counterion (not presented for clarity).

**Scheme 4** DFT representation of the interconversions of possible diastereomeric intermediates and lactame products involved in the CDI-mediated tandem cyclizations. The relative total electronic energy values and activation barriers [kJ/mol] were calculated at B3LYP/6-31 G(d,p) level of theory using IEFPCM to imitate the experimental conditions ( $\epsilon_{\text{DMF}}=36.7$ ).

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The geometry optimization found conformers **15a,b/1** and **15a,b/2** (Scheme 4) as local minima on the potential energy surfaces (PES) preformed for the crucial bond formation between the C12a and C12b atoms to yield intermediates **16a,b/c** and **16a,b/t** of which deprotonation finally leads to two types of diastereomeric products **9a,b** and **12a,b**, respectively. Using QST2 method [54] the transition states of the crucial cyclization steps **15/1**→**16/c** and **15/2**→**16/t** were identified on the PES as saddle points and confirmed by IRC analysis [55]. The total electron energy values for all optimized structures were recalculated with IEFPCM solvent model [56] employing the dielectric constant of DMF ( $\epsilon=36.7$ ) to provide a more realistic representation of the experimental conditions. Finally, considering the  $\Delta E$  and  $\Delta E^\ddagger$  values presented on Scheme 4 it can be stated that the results of modeling studies provide a satisfactory support for the proposed reaction pathways outlined above, as both the relative energetics of the relevant conformers of the primarily formed *N*-acyliminium intermediates (**15a,b/1** and **15a,b/2**) and the activation barriers of their ring closures, ultimately determining the relative configuration of the products, are in good agreement with the diastereoselectivity experienced in the CDI-mediated tandem cyclization reactions of imines **6a,b**. On the other hand, the higher activation barriers of the ring closures of **15a,b/1** (+38.2 kJ/mol and +42.5 kJ/mol) relative to those of **15a,b/2** (+31.4 kJ/mol and +29.2 kJ/mol) seem to correlate with the opposite signs of the energetics calculated for these elementary steps [ $\Delta E(16a/c-15a/1)/(16b/c-15b/1)=+6.8$  kJ/mol/+6.0 kJ/mol, vs.  $\Delta E(16a/t-15a/2)/(16b/t-15b/2)=-8.6$  kJ/mol/-16.8 kJ/mol]. The total electron energy values of the possible conformer pairs of the lactame products and the activation barriers of their interconversions were also calculated by the aforementioned DFT method and solvent model (Scheme 4). Although the results indicate that - except for **9b/inv**→**9b** - the ring inversions of the isolated lactames **9a** and **12a,b** are not favored processes in terms of relative energetics, due to their low activation barriers the presence of minimal amounts of conformers **9a/inv** and **12a,b/inv** (<1%) cannot be ruled out in the appropriate reaction mixtures or in the dissolved samples subjected to NMR measurements. However, since the transition state was located with a fixed position of the methoxycarbonyl group, the calculated activation barrier for ring inversion **9b/inv**→**9b** (+11.6 kJ/mol) is certainly lower than the real one of which height must substantially be increased by the rotation of the bulky substituent which, sweeping through the plane of the lactame carbonyl group, strongly interferes with the skeletal atoms.



**Figure 1** Selected bonding orbitals of conformers **15a,b/2**, the precursors of ring closures finally leading to lactames **12a,b**, determined by B3LYP/6-31 G(d,p) analysis performed on optimized structures.

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The relative energetics of the conformer pairs **15a/1–15a/2** and **15b/1–15b/2**, respectively, performed for the crucial cyclization steps responsible for the stereochemical outcome of the overall reactions, are also worth for discussion based on the analysis of relevant bonding orbitals (Figure 1). Accordingly, it might be concluded that the extra stabilization of conformers type **15/2** relative to that of alternative conformers type **15/1** can at least partially be attributed to the delocalization of HOMO-5 presenting a weak bonding between the iron center and the proximal indole ring. Iminium ions **15a,b/2** are also stabilized by interactions of the cationic center with the donor indole and ferrocene moieties, as demonstrated by HOMO and HOMO-2, respectively.

### 2.3. Structure determination of the novel compounds

The spectroscopic data listed in the Experimental part are consistent with the structures of novel compounds **2a,b**, **3a,b**, **4b**, **6a,b**, **7a,b**, **8a,b**, **9a,b**, **9b/inv**, **11**, **12a,b**, **13b** and **14b**, however the following points are necessary to be highlighted.

Due to the presence of incorporated ferrocene- and/or tryptophane fragments with fixed elements of chirality, the absolute configurations of **3b**, **4b**, **6b**, **7a,b**, **8a,b**, **9a,b**, **9b/inv**, **12a,b** and **13b** follow from their relative configurations.

The characteristic cross peaks resulted from the  $^2J$ - and  $^3J$  couplings between the signals of nucleus pairs H1/C4a, H1/C9a, H9/C4a and H9/C9a detected by  $^1\text{H}$ - $^{13}\text{C}$ -HMBC method for **3a,b**, **4b**, **7a,b** and **8a,b** indicate the presence of the  $\beta$ -carboline skeleton, i.e. the condensation of the tetrahydropyridine ring to the indole residue. In a similar way, the polycyclic constitution of **9a,b**, **9b/inv** and **12a,b** is unequivocally proved by the cross peak correlations between the signals originated from atom pairs H12b/C7a, H12b/C12a, H12b/C4 and H6/C4 discernible in their  $^1\text{H}$ - $^{13}\text{C}$ -HMBC spectra. In accord with their relative configuration, the NOESY spectra of **9a**, **9b** and **9b/inv** disclosed that H12b is situated in the proximity of the protons of the freely rotating  $\eta^5$ -C<sub>5</sub>H<sub>5</sub> residue. In **9b** and **9b/inv** the “S” configuration of the C6 stereogenic center was evidenced by NOE’s detected between protons H12b and H6<sub>A</sub> being in relative 1,3-*cis* position on the ring system. On the other hand, in **12a** and **12b** the “R” configuration of the C12b stereogenic center associated with the *endo* orientation of the  $\eta^5$ -C<sub>5</sub>H<sub>5</sub> ring is evidenced by the NOE’s generated by its interactions with H7<sub>A</sub> and the indole NH proton, respectively. In keeping with the results of NOESY experiments, in the optimized structures of these compounds H7<sub>A</sub>, attached to ring C with half chair conformation of “P”-helicity, is situated in *endo-axial* position in the proximity of the  $\eta^5$ -C<sub>5</sub>H<sub>5</sub> ring. Being in the shielding region of the indole ring,  $^1\text{H}$  resonances of this residue measured for **12a** and **12b** (3.68 and 3.70 ppm, resp.) are significantly upfield-shifted relative to those measured for diastereomers **9a,b** and **9b/inv** carrying the  $\eta^5$ -C<sub>5</sub>H<sub>5</sub> ring in *exo* site of the polycyclic skeleton (4.06–4.31 ppm). The NOESY experiments carried out for **9a,b** and **9b/inv** detected an additional interaction between H12b and H6<sub>A</sub> atoms. Supporting the results of DFT modeling studies on **9a,b**, the coupling patterns of H6- and H7 multiplets discernible in their  $^1\text{H}$ -NMR spectra refer to “M” helical chirality of ring C with a half chair conformation carrying H6<sub>A</sub> and H7<sub>B</sub> in relative 1,2-*di-axial* position [ $J(\text{H6}_A/\text{H7}_B)$ =11.6 Hz and 11.8 Hz for **9a** and **9b**, resp.]. On the other hand, the opposite helical chirality (“P”) of ring C in **9b/inv** and **12b**, carrying the methoxycarbonyl group in *axial* position, is reflected from the simple broadened doublet split of the signal of the H6<sub>A</sub> proton (7.1 Hz and 7.5 Hz, resp.) due to an interaction with the H7<sub>A</sub> proton and a negligible coupling with the H7<sub>B</sub> proton. In **12a** the same conformation of ring C associated with helical chirality “P” is also evidenced by the coupling pattern of the signals originated from the skeletal protons, which indicates only a weak interaction characterized by 2.0 Hz between H6<sub>A</sub> and H7<sub>B</sub>. In the  $^1\text{H}$ -NMR spectrum of **12a** the assignments of H6<sub>A</sub>- and H6<sub>B</sub> signals were supported by a NOESY experiment disclosing a significant interaction between protons H6<sub>B</sub> and H12b. Accordingly, the signal of H6<sub>B</sub> is significantly upfield-shifted relative to that of the *equatorial* H6<sub>A</sub> positioned in the proximity of the lactame carbonyl group exerting a significant anisotropic deshielding effect on the latter resonance (3.54 ppm and 4.98 ppm, resp.). It must be pointed out that in the  $^1\text{H}$ -NMR spectrum of **9a** the resonance due to *equatorial* proton H6<sub>B</sub>, (5.18 ppm) situated nearly in the plane of the lactame carbonyl group, is significantly downfield shifted relatively to that measured for H6<sub>A</sub> (3.79 ppm) in *axial* position.

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In order to detect the particular conformers of the fused  $\gamma$ -lactames we attempted to register their  $^1\text{H-NMR}$  spectra at low temperature using  $\text{CD}_2\text{Cl}_2$  as solvent, however these efforts failed due to severe solubility problems. Instead, the almost exclusive presence of the preferred conformers of **9a**, and **12a,b** was supported by comparative NMR-GIAO calculations [57] carried out for the optimized structures of the corresponding conformer pairs (Table 1) by B3LYP functional using the extended 6-311 ++G(2d,p) basis set [58]. Although no exact data can be given for the ratio of the particular components involved in the possible equilibrium ring inversion processes, in each studied case the difference between the calculated- and experimental chemical shifts and separation of the skeletal H6 signals (cf. **9a-9a/inv** and **12a-12a/inv**) unambiguously refer to a strong dominance of the diastereomer supposed to be detected by NMR experiments over its counterparts with reversed helicity of ring C. Supporting this view, the highly diagnostic vicinal coupling constants  $J(\text{H6}_A/\text{H7}_B)$  and  $J(\text{H6}_A/\text{H7}_A)$  are in good accord with the values calculated by a particular version of Karplus-equation using a parameter set composed for a general N-CH<sub>2</sub>-CH<sub>2</sub>-C sequence [59] and the dihedral angles found in the modeled structures of the isolated  $\gamma$ -lactames (Table 1). It is also important to note here that the substantial mismatch between the calculated and measured values of  $J(\text{H6}_A/\text{H7}_B)$  rules out the presence of diastereomers **9a/inv**, **12a/inv** and **12b/inv** in significant amounts in the solutions subjected to NMR studies. Finally, the alternative conformations of ring C with opposite helicity in diastereomers **9b** and **9b/inv** are conclusively evidenced by the highly similar calculated- and experimental NMR parameter pairs listed in Table 1. This stereochemical relation was also confirmed by ring inversion **9b/inv**→**9b** carried out in a separate experiment under the conditions of Method K, as discussed above.

The relative configurations of carboxyferrocenyl-substituted  $\beta$ -carboline **7a,b** and **8a,b**, the precursors of CDI-mediated cyclization reactions assumed to take place with the preservation of the configuration of the particular stereogenic centers, were deduced from those determined for the corresponding lactame products types **9** and **12**.

In compound **13b** the constitution and the bent conformation of the nine-membered azonine ring with “E” lactame residue were established on the basis of NOESY cross-peak correlations between proton pairs H3/H6<sub>A</sub>, H5/H7<sub>B</sub>, H7<sub>B</sub>/H8, H12/H13 and H1/H13.

The significant downfield shift of the H9 signal in the  $^1\text{H-NMR}$  spectrum of **14b** (8.62 ppm) refers to the proximity of the oxygen atom of the carbonyl group incorporated in the  $\delta$ -lactame ring fused to the ferrocene moiety. The broadened doublet signals due to H9 and H12 were unambiguously assigned on the basis of a NOESY interaction detected between proton pair H1/H12 being in spatial proximity on the rigid ring system.

**Table 1:**  $^1\text{H-NMR}$  shifts and separation of H6<sub>A</sub> and H6<sub>B</sub> signals calculated for diastereomer pairs **9a-9a/inv**, **9b-9b/inv**, **12a-12a/inv** and **12b-12b/inv** (rows 1-6, reference: TMS)<sup>a</sup>, dihedral angles between skeletal protons (9: rows 7 and 10)<sup>b</sup> and vicinal coupling constants  $J(\text{H6}_A/\text{H7}_B)$  and  $J(\text{H6}_A/\text{H7}_A)$  (calculated values: rows 8 and 11, measured values: rows 9 and 12).

	<b>9a</b>	<b>9a/inv</b>	<b>9b</b>	<b>9b/inv</b>	<b>12a</b>	<b>12a/inv</b>	<b>12b</b>	<b>12b/inv</b>
1. $\delta\text{H6}_A$ [ppm] (calcd.)	<b>3.84</b> <sup>c</sup>	3.47	4.18	5.28	<b>5.00</b> <sup>c</sup>	3.68	<b>5.85</b> <sup>c</sup>	4.84
2. $\delta\text{H6}_A$ [ppm] (meas.)	<b>3.79</b> <sup>c</sup>		4.26	5.49	<b>4.98</b> <sup>c</sup>		<b>5.66</b> <sup>c</sup>	
3. $\delta\text{H6}_B$ [ppm] (calcd.)	<b>4.88</b> <sup>c</sup>	4.01	-	-	<b>3.24</b> <sup>c</sup>	4.02	-	-
4. $\delta\text{H6}_B$ [ppm] (meas.)	<b>5.18</b> <sup>c</sup>		-	-	<b>3.54</b> <sup>c</sup>		-	-
5. $\Delta(\delta\text{H6}_A-\delta\text{H6}_B)$ [ppm] (calcd.)	<b>-1.04</b> <sup>c</sup>	-0.54	-	-	<b>1.96</b> <sup>c</sup>	-0.34	-	-
6. $\Delta(\delta\text{H6}_A-\delta\text{H6}_B)$ [ppm] (meas.)	<b>-1.39</b> <sup>c</sup>		-	-	<b>1.76</b> <sup>c</sup>		-	-
7. $\vartheta(\text{H6}_A/\text{H7}_B)$ [°] (calcd.)	162.8	79.1	167.7	73.6	74.9	157.2	77.7	150.5
8. $J(\text{H6}_A/\text{H7}_B)$ [Hz] (calcd.) <sup>d</sup>	<b>10.9</b> <sup>c</sup>	1.5	11.4	1.9	<b>1.8</b> <sup>c</sup>	10.1	<b>1.6</b> <sup>c</sup>	9.1
9. $J(\text{H6}_A/\text{H7}_B)$ [Hz] (meas.)	<b>11.6</b> <sup>c</sup>		11.8	~0	<b>2.0</b> <sup>c</sup>		<b>~0</b> <sup>c</sup>	
10. $\vartheta(\text{H6}_A/\text{H7}_A)$ [°] (calcd.)	44.8	35.8	51.9	39.8	42.0	41.2	35.4	38.1
11. $J(\text{H6}_A/\text{H7}_A)$ [Hz] (calcd.) <sup>d</sup>	<b>5.9</b> <sup>c</sup>	7.3	4.8	6.8	<b>6.4</b> <sup>c</sup>	6.5	<b>7.4</b> <sup>c</sup>	7.0
12. $J(\text{H6}_A/\text{H7}_A)$ [Hz] (meas.)	<b>5.2</b> <sup>c</sup>		4.3	7.1	<b>5.4</b> <sup>c</sup>		<b>7.5</b> <sup>c</sup>	

<sup>a</sup>. The calculations were performed by B3LYP/NMR-GIAO method using the extended 6-311 ++G(2d,p) basis set.

<sup>b</sup>. The values are extracted from the structures optimized by B3LYP functional employing 6-31 G(d,p) basis set.

<sup>c</sup>. The better match between the calculated and measured values is highlighted by **bold-italic** fonts.

<sup>d</sup>. Obtained by Karplus equation using  $\vartheta$  values and a parameter set composed for a general N-CH<sub>2</sub>-CH<sub>2</sub>-C sequence.

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### 3. Conclusion

By means of sequential use of Pictet-Spengler and *N*-acylation protocols 4-oxo-6,7,12,12b-tetrahydro-4*H*-ferroceno[1,2]indolizino[8,7-*b*]indoles, the first representatives of a novel  $\beta$ -carboline-based alkaloid-like ring system incorporating fused metallocene and  $\gamma$ -lactame moieties were prepared as single enantiomers with the elements of planar- and central chirality. The conformation and the relative configuration of these products were determined by combined use of one- and two-dimensional versions of  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR methods supported by theoretical NMR calculations. Besides the targeted  $\gamma$ -lactame products the ring closures of carboxyferrocenyl-substituted  $\beta$ -carbolines carrying methoxycarbonyl group in position 3 also afforded the planar chiral  $\delta$ -lactame ( $S_p$ )-methyl 7-oxo-7*H*-ferroceno[*c*]indolo[3,2,1-*ij*][1,5]naphthyridine-2-carboxylate of which formation was prevented by implementing an alternative method for the construction of the desired  $\gamma$ -lactames based on CDI-mediated tandem cyclization of simple carboxyferrocenyl-substituted tryptamine/tryptophane-derived imines. Possible reaction pathways accounting for the diastereoselectivity of the overall processes were proposed on the basis of the results obtained by comparative DFT modeling studies. Employing further imine precursors suitable to form a five- or six-membered cyclic *N*-acyliminium intermediate capable of acting as efficient electrophilic component in intramolecular Pictet-Spengler reaction, this one-pot procedure may allow the facile synthesis of a range of related polyheterocyclic alkaloid analogues of potential biological interest. The evidenced propensity of metallocene-fused lactame products to undergo *trans-anullar* ring opening, taking place with simultaneous decomposition of the organometallic fragment, can be utilized in design of improved protocols to provide a relatively easy access to novel classes of medium-size heterocyclic scaffolds.

### 4. Experimental

All chemicals were obtained from commercially available sources (Aldrich, Fluka) and used without further purification. Melting points (uncorrected) were determined with a Boethius microstage. Merck Kieselgel (230–400 mesh, 60 Å) was used for flash column chromatography. The  $R_f$  values given for the separated products were determined using DCM-MeOH (20:1) as eluent on silica plate. The IR spectra were run by ATR (Attenuated Total Reflectance) method [60] on a Bruker IFS-55 FT-spectrometer controlled by Opus 3.0 software. Optical rotations were measured with a Zeiss Polamat A polarimeter. The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra were recorded in DMSO- $d_6$  or  $\text{CDCl}_3$  solution in 5 mm tubes at RT, on a Bruker DRX-500 spectrometer at 500 ( $^1\text{H}$ ) and 125 ( $^{13}\text{C}$ ) MHz, with the deuterium signal of the solvent as the lock and TMS as internal standard ( $^1\text{H}$ ,  $^{13}\text{C}$ ) The 2D-COSY, HSQC, HMBC and NOESY spectra were obtained by using the standard Bruker pulse programs. All calculations were carried out with the Gaussian 09 suite of programs [61]. Optimized structures are available from the authors.

#### 4.1. Preparation of imines **2a** and **6a** (Method A)

Tryptamine (3.204 g, 20 mmol) and formylferrocene (4.281 g, 20 mmol) or ( $S_p$ )-2-formyl-ferrocene carboxylic acid (5.161 g, 20 mmol) were heated in MeOH (20 mL) at reflux temperature for 1 h. The reaction mixture was evaporated to half of its volume and diluted with water (30 mL). The precipitated solid was filtered off, washed with a cold mixture of MeOH-water (2:3) and dried to obtain the appropriate imine **2a** or **6a**. Analytical samples of the products were recrystallized from mixtures of DCM-hexane.

##### 4.1.1. (*E*)-*N*-(2-(1*H*-Indol-3-yl)ethyl)-1-ferrocenylmethanimine (**2a**)

Dark yellow solid. Yield: 5.840 g (82%); mp.: 67–70 °C; IR ( $\text{cm}^{-1}$ ): ~3300–2900, 1643, 1446, 1245, 1105, 1043, 1021, 810, 737.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ): 8.30 (s, 1H, NH); 8.03 (s, 1H, CH=N); 7.72 (d,  $J=7.4$  Hz, 1H, H4); 7.36 (d,  $J=7.4$  Hz, 1H, H7); 7.21 (t,  $J=7.4$  Hz, 1H, H5); 7.17 (t,  $J=7.4$  Hz, 1H, H6); 7.05 (br s, 1H, H2); 4.64 (t,  $J=1.8$  Hz, 2H, H2',5'); 4.37 (t,  $J=1.8$  Hz, 2H, H3',4'); 4.06 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 3.84 (t,  $J=6.8$  Hz, 2H, N-CH<sub>2</sub>); 3.17 (t,  $J=6.8$  Hz, 2H, C-CH<sub>2</sub>);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ): 162.0 (CH=N); 136.8 (C7a); 128.0 (C3a); 122.6 (C2); 122.3 (C6); 119.6 (C5); 119.4 (C4); 114.4 (C3); 111.7 (C7); 80.5 (C1'); 71.0 (C3',4'); 69.5 ( $\eta^5\text{-C}_5\text{H}_5$ ); 69.0 (C2',5'); 62.3 (N-CH<sub>2</sub>); 27.3 (C-CH<sub>2</sub>); Anal. Calcd. for  $\text{C}_{21}\text{H}_{20}\text{FeN}_2$  (356.25): C, 70.80; H, 5.66; N, 7.86. Found: C, 70.70; H, 5.74; N, 7.91%.

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#### 4.1.2. (*S<sub>p</sub>*,*E*)-2-(((2-(1*H*-Indol-3-yl)ethyl)imino)methyl)ferrocene carboxylic acid (**6a**)

Dark red solid. Yield: 6.084 g (76%); mp.: 85-87 °C;  $[\alpha]_{\text{D}}^{25}$ : +42.2° (EtOH  $c$  = 0.084 g/100 mL); IR (cm<sup>-1</sup>): ~3600-2700, 1710, 1645, 1585, 1437, 1340, 1230, 1106, 1003, 821, 741. <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>): 10.91 (s, 1H, NH); 8.49 (s, 1H, CH=N); 7.63 (d,  $J$ =7.3 Hz, 1H, H4); 7.33 (d,  $J$ =7.3 Hz, 1H, H7); 7.07 (t,  $J$ =7.3 Hz, 1H, H6); 7.01 (t,  $J$ =7.3 Hz, 1H, H5); 7.23 (br s, 1H, H2); 5.11 (br s, 1H, H5'); 4.82 (br s, 1H, H3'); 4.70 (t,  $J$ =1.8 Hz, 1H, H3'); 4.05 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 3.90 (t,  $J$ =6.4 Hz, 2H, N-CH<sub>2</sub>); 3.10 (t,  $J$ =6.4 Hz, 2H, C-CH<sub>2</sub>); due to exchange processes the signal of CO<sub>2</sub>H proton could not be detected in the spectrum; <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>): 171.5 (CO<sub>2</sub>H); 167.5 (CH=N); 137.2 (C7a); 127.9 (C3a); 124.0 (C2); 121.9 (C6); 119.3 (two coalesced lines, C4 and C5); 112.3 (C7); 112.0 (C3); 76.4 (C5'); 75.9 (C1'); 75.5 (C4'); 73.6 (C3'); 71.9 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 70.2 (C2'); 57.8 (N-CH<sub>2</sub>); 26.5 (C-CH<sub>2</sub>); Anal. Calcd. for C<sub>22</sub>H<sub>20</sub>FeN<sub>2</sub>O<sub>2</sub> (400.26): C, 66.02; H, 5.04; N, 7.00. Found: C, 65.94; H, 5.12; N, 7.08%.

#### 4.2. Preparation of imines **2b** and **6b** (Method B)

A mixture of (*L*)-tryptophan methylester hydrochloride (5.094 g, 20 mmol), Na<sub>2</sub>CO<sub>3</sub> (2.650 g, 25 mmol), water (40 mL) and DCM (80 mL) was intensively stirred for ca. 5 min., the phases were separated. The aqueous solution was extracted with DCM (3x30 mL). To the combined organic phase formylferrocene (4.281 g, 20 mmol) or (*S<sub>p</sub>*)-2-formyl-ferrocene carboxylic acid (5.161 g, 20 mmol) was added. The solution was dried on Na<sub>2</sub>SO<sub>4</sub> then evaporated to dryness to obtain a solid residue which was triturated with cold MeOH, filtered off washed thoroughly with water and dried. Analytical samples of the products were recrystallized from mixtures of DCM-hexane.

##### 4.2.1. Methyl (*S,E*)-2-(ferrocenylmethyleneamino)-3-(1*H*-indol-3-yl)propanoate (**2b**)

Dark yellow solid. Yield: 7.883 g (93%); mp.: 78-81 °C;  $[\alpha]_{\text{D}}^{25}$ : -57.5° (EtOH  $c$  = 0.053 g/100 mL); IR (cm<sup>-1</sup>): ~3600-2700, 1742, 1600, 1580, 1437, 1340, 1244, 1100, 1012, 833, 746. <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>): 10.82 (s, 1H, NH, indole); 8.36 (s, 1H, CH=N); 7.61 (d,  $J$ =7.6 Hz, 1H, H4); 7.30 (d,  $J$ =7.6 Hz, 1H, H7); 7.08 (t,  $J$ =7.6 Hz, 1H, H6); 6.96 (t,  $J$ =7.6 Hz, 1H, H5); 7.09 (br s, 1H, H2); 4.74 (t,  $J$ =18.0 Hz, 2H, H2',5'); 4.40 (t,  $J$ =1.8 Hz, 2H, H3',4'); 4.51 (dd,  $J$ =9.4 and 4.4 Hz, H, N-CH); 4.10 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 3.76 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.45 (dd,  $J$ =14.7 and 4.4 Hz, 1H, C-CH<sub>A</sub>H<sub>B</sub>); 3.22 (dd,  $J$ =14.7 and 9.7 Hz, 1H, C-CH<sub>A</sub>H<sub>B</sub>); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>): 171.8 (CO<sub>2</sub>CH<sub>3</sub>); 161.2 (CH=N); 136.4 (C7a); 127.4 (C3a); 124.1 (C2); 121.5 (C6); 119.0 (C5); 118.6 (C4); 112.5 (C7); 110.0 (C3); 80.1 (C1'); 71.0 (C3',4'); 69.2 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 68.3 (C2',5'); 71.2 (N-CH); 53.6 (CO<sub>2</sub>CH<sub>3</sub>); 29.1 (C-CH<sub>2</sub>); Anal. Calcd. for C<sub>23</sub>H<sub>22</sub>FeN<sub>2</sub>O<sub>2</sub> (414.29): C, 66.68; H, 5.35; N, 6.76. Found: C, 66.85; H, 5.41; N, 6.65%.

##### 4.2.2. (*S<sub>p</sub>*,*S,E*)-2-(((1-Methoxycarbonyl-2-(1*H*-indol-3-yl)ethyl)imino)methyl)ferrocene carboxylic acid (**6b**)

Dark red solid. Yield: 7.883 g (86%); mp.: 114-117 °C;  $[\alpha]_{\text{D}}^{25}$ : -23.9° (CHCl<sub>3</sub>  $c$  = 0.071 g/100 mL); IR (cm<sup>-1</sup>): ~3600-2800, 1735, 1690, 1570, 1458, 1443, 1369, 1254, 1106, 1005, 823, 742. <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>): 10.89 (s, 1H, NH); 8.43 (s, 1H, CH=N); 7.56 (d,  $J$ =7.3 Hz, 1H, H4); 7.32 (d,  $J$ =7.3 Hz, 1H, H7); 7.06 (t,  $J$ =7.3 Hz, 1H, H6); 6.99 (t,  $J$ =7.3 Hz, 1H, H5); 7.15 (d,  $J$ =1.8 Hz, 1H, H2); 5.07 (br s, 1H, H5'); 4.83 (br s, 1H, H3'); 4.70 (t,  $J$ =2.0 Hz, 1H, H4'); 4.49 (dd,  $J$ =8.9 and 4.6 Hz, H, N-CH); 4.13 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 3.72 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.38 (dd,  $J$ =14.5 and 4.6 Hz, 1H, C-CH<sub>A</sub>H<sub>B</sub>), partly overlapped by the HDO signal of the solvent); 3.19 (dd,  $J$ =14.5 and 8.9 Hz, 1H, C-CH<sub>A</sub>H<sub>B</sub>) due to exchange processes the signal of CO<sub>2</sub>H proton could not be detected in the spectrum; <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>): 171.4 (CO<sub>2</sub>H); 171.0 (CO<sub>2</sub>CH<sub>3</sub>); 167.5 (CH=N); 136.6 (C7a); 127.3 (C3a); 124.5 (C2); 121.5 (C6); 119.1 (C5); 118.7 (C4); 112.1 (C7); 109.5 (C3); 76.5 (C5'); 75.5 (C3'); 73.5 (C4'); 73.2 (C1'); 71.9 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 70.2 (C2'); 70.7 (N-CH); 53.2 (CO<sub>2</sub>CH<sub>3</sub>); 28.6 (C-CH<sub>2</sub>); Anal. Calcd. for C<sub>24</sub>H<sub>22</sub>FeN<sub>2</sub>O<sub>4</sub> (458.30): C, 62.90; H, 4.84; N, 6.11. Found: C, 63.04; H, 4.77; N, 6.20%.

#### 4.3. Pictet-Spengler cyclizations of imine **2a** by Methods C-G: synthesis of 1-ferrocenyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole (**3a**)

Imine **2a** (3.563 g, 10 mmol) was added to the corresponding acidic mixture (Method C: 70% HClO<sub>4</sub>/EtOH (1 mL/15 mL); Method D: 70% HClO<sub>4</sub>/AcOH (1 mL/15 mL); Method E: TFA/AcOH (1 mL/15 mL); Method F: H<sub>3</sub>BO<sub>3</sub>/AcOH (0.350 g/15 mL); Method G: AcOH (15 mL)) The resulted solution was stirred and heated at reflux (for 24 h by Methods C and G and for 4h by Methods D-F) under argon and evaporated to dryness. The residue was triturated with water and the pH of the resulted suspension was adjusted to 8-9 by addition of solid Na<sub>2</sub>CO<sub>3</sub> in small portions. The solid precipitate was collected by filtration, washed with cold EtOH and dried to obtain **3a**. An analytical sample was recrystallized from EtOH. Yield: 1.283 g (36% by Method C); 0.891 g (25% by Method D); 1.603 g (45% by Method E); 1.960 g (55% by Method F); 2.850 g (80% by Method G); mp.: 98-100 °C;  $R_f$ : 0.30; IR (cm<sup>-1</sup>): 3423, 3232, 1642, 1620, 1584, 1455, 1415, 1294,

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1103, 999, 820, 743.  $^1\text{H}$  NMR (DMSO- $d_6$ ): 10.17 (s, 1H, NH); 7.34 (dd,  $J=7.4$  and 1.8 Hz, 1H, H5); 7.28 (br d,  $J=7.4$  Hz, 1H, H8); 6.98 (td,  $J=7.4$  and 1.8 Hz, 1H, H7); 6.92 (td,  $J=7.4$  and 1.8 Hz, 1H, H6); 4.84 (s, 1H, H1); 4.48, 4.31, 4.17, 4.09 (4xbr s, 4x1H, H2'-5'); 4.19 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 3.30 (dt,  $J=12.1$  and 3.1 Hz, 1H, H3<sub>A</sub>, partly overlapped by the HDO signal of the solvent); 2.99 (ddd,  $J=12.1$ , 7.6 and 4.8 Hz, 1H, H3<sub>B</sub>); 2.69-2.57 (overlapping m's, 2H, H4<sub>A</sub> and H4<sub>B</sub>);  $^{13}\text{C}$  NMR (DMSO- $d_6$ ): 137.4 (C9a); 136.6 (C8a); 127.6 (C4b); 121.2 (C7); 119.1 (C6); 118.5 (C5); 112.2 (C8); 107.4 (C4a); 92.2 (C1'); 68.6, 67.9, 67.8, 66.7 (C2'-5'); 69.3 ( $\eta^5\text{-C}_5\text{H}_5$ ); 52.6 (C1); 43.5 (C3); 23.3 (C4); Anal. Calcd. for C<sub>21</sub>H<sub>20</sub>FeN<sub>2</sub> (356.25): C, 70.80; H, 5.66; N, 7.86. Found: C: 70.68; H: 5.60; N, 7.77%.

#### 4.4. Pictet-Spengler cyclizations of imines **2b** and **6a,b** by Method G:

The conversion of the appropriate imine (10 mmol) was performed in AcOH (15 mL). The workup procedure described in the previous section led to the isolation of diastereomeric mixtures of the appropriate  $\beta$ -carboline products (**3b/4b**, **7a/8a** and **7b/8b**). (After the reactions of **6a** and **6b** the pH of the suspensions obtained by trituration of the evaporated reaction mixture with water was adjusted to 7-8 by addition of solid Na<sub>2</sub>CO<sub>3</sub> in small portions.) The diastereomeric pairs were subjected to flash column chromatography on silica using DCM-MeOH (20:1 for **3b/4b**, 10:1 for **7a/8a**, and 15:1 for **7b/8b**) as eluents. The solid residues obtained by evaporation of separated bands were triturated by cold MeOH, filtered off and dried to give  $\beta$ -carbolines as single diastereomers. Analytical samples were recrystallized from MeOH.

##### 4.4.1. Methyl (1*S*,3*S*)-1-ferrocenyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylate (**3b**)

Dark yellow solid. Yield: 2.569 g (62%); mp.: 97-100 °C;  $R_f$ : 0.41;  $[\alpha]_D^{25}$ : -62.0° (CHCl<sub>3</sub>  $c$  = 0.064 g/100 mL); IR (cm<sup>-1</sup>): 3354, 3204, 1622, 1573, 1423, 1380, 1287, 1085, 975, 810, 748.  $^1\text{H}$  NMR (DMSO- $d_6$ ): 10.24 (s, 1H, NH); 7.46 (br d,  $J=7.6$  Hz, 1H, H5); 7.19 (br d,  $J=7.6$  Hz, 1H, H8); 7.02 (td,  $J=7.6$  and 1.8 Hz, 1H, H7); 6.95 (td,  $J=7.6$  and 1.8 Hz, 1H, H6); 4.84 (s, 1H, H1); 4.68, 4.31, 4.17, 4.10 (4xbr s, 4x1H, H2'-5'); 4.32 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 4.07 (dd,  $J=11.9$  and 4.4 Hz, 1H, H3); 3.90 (s, 3H, OCH<sub>3</sub>); 3.05 (dd,  $J=14.5$  and 4.8 Hz, 1H, H4<sub>B</sub>); 2.83 (dd,  $J=14.5$  and 11.9 Hz, 1H, H4<sub>A</sub>);  $^{13}\text{C}$  NMR (DMSO- $d_6$ ): 138.3 (C8a); 135.8 (C9a); 127.0 (C4b); 122.0 (C7); 120.3 (C6); 118.6 (C5); 112.9 (C8); 106.5 (C4a); 91.8 (C1'); 72.0 ( $\eta^5\text{-C}_5\text{H}_5$ ); 69.3, 68.4, 68.1, 66.9 (C2'-5'); 50.0 (C1); 54.7 (C3); 25.2 (C4); Anal. Calcd. for C<sub>23</sub>H<sub>22</sub>FeN<sub>2</sub>O<sub>2</sub> (414.29): C, 66.68; H, 5.35; N, 6.76%. Found: C: 66.80; H: 5.31; N, 6.70%.

##### 4.4.2. Methyl (1*R*,3*S*)-1-ferrocenyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylate (**4b**)

Dark yellow solid. Yield: 0.497 g (12%); mp.: 113-115 °C;  $R_f$ : 0.53;  $[\alpha]_D^{25}$ : -71.7° (EtOH  $c$  = 0.062 g/100 mL); IR (cm<sup>-1</sup>): 3408, 3222, 1615, 1573, 1445, 1364, 1186, 1103, 988, 814, 740.  $^1\text{H}$  NMR (DMSO- $d_6$ ): 10.52 (s, 1H, NH); 7.42 (br d,  $J=7.6$  Hz, 1H, H5); 7.36 (br d,  $J=7.6$  Hz, 1H, H8); 7.00 (br t,  $J=7.6$  Hz, 1H, H7); 6.85 (br t,  $J=7.6$  Hz, 1H, H6); 5.24 (s, 1H, H1); 4.53, 4.29, 4.22, 4.07 (4xbr s, 4x1H, H2'-5'); 4.13 (dd,  $J=5.4$  and 4.2 Hz, 1H, H3); 3.96 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 3.73 (s, 3H, OCH<sub>3</sub>); 3.19 (dd,  $J=15.6$  and 4.2 Hz, 1H, H4<sub>B</sub>); 2.83 (dd,  $J=15.6$  and 5.4 Hz, 1H, H4<sub>A</sub>);  $^{13}\text{C}$  NMR (DMSO- $d_6$ ): 136.1 (C8a); 132.3 (C9a); 127.4 (C4b); 122.4 (C7); 120.2 (C6); 119.4 (C5); 111.6 (C8); 106.2 (C4a); 90.3 (C1'); 70.7 ( $\eta^5\text{-C}_5\text{H}_5$ ); 68.1, 67.5, 67.0, 66.5 (C2'-5'); 50.0 (C1); 51.2 (C3); 24.0 (C4); Anal. Calcd. for C<sub>23</sub>H<sub>22</sub>FeN<sub>2</sub>O<sub>2</sub> (414.29): C, 66.68; H, 5.35; N, 6.76%. Found: C: 66.77; H: 5.44; N, 6.82%.

##### 4.4.3. (*S*<sub>p</sub>)-((*S*)-5-(2,3,4,9-Tetrahydro-1*H*-pyrido[3,4-*b*]indol-1-yl)ferrocene-1-carboxylic acid (**7a**)

Orange solid. Yield: 1.241 g (31%); mp.: 193-195 °C;  $R_f$ : 0.36;  $[\alpha]_D^{25}$ : +20.9° (CHCl<sub>3</sub>  $c$  = 0.033 g/100 mL); IR (cm<sup>-1</sup>): ~3600-2700, 1612, 1563, 1440, 1365, 1265, 1106, 1005, 815, 735.  $^1\text{H}$  NMR (DMSO- $d_6$ ): 10.20 (s, 1H, NH); 7.45 (br d,  $J=7.5$  Hz, 1H, H5); 7.35 (br d,  $J=7.5$  Hz, 1H, H8); 7.09 (td,  $J=7.5$  and 1.8 Hz, 1H, H7); 7.04 (td,  $J=7.5$  and 1.8 Hz, 1H, H6); 5.88 (s, 1H, H1); 4.80 (br s, 1H, H3'); 4.57 (br s, 1H, H4'); 4.51 (br s, 1H, H5'); 4.37 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 3.51 (ddd,  $J=12.6$ , 6.9 and 5.0 Hz, 1H, H3<sub>A</sub>); 3.35 (ddd,  $J=12.6$ , 6.2 and 4.5 Hz, 1H, H3<sub>B</sub>); 3.01-2.79 (overlapping m's, 2H, H4<sub>A</sub> and H4<sub>B</sub>);  $^{13}\text{C}$  NMR (DMSO- $d_6$ ): 171.0 (CO<sub>2</sub>H); 137.8 (C8a); 133.3 (C9a); 127.5 (C4b); 122.2 (C7); 119.7 (C6); 119.2 (C5); 112.9 (C8); 104.7 (C4a); 90.8 (C1'); 73.8 (C5'); 71.5 (C2'); 71.0 ( $\eta^5\text{-C}_5\text{H}_5$ ); 70.6 (C4'); 70.2 (C3'); 53.1 (C1); 43.5 (C3); 20.2 (C4); Anal. Calcd. for C<sub>22</sub>H<sub>20</sub>FeN<sub>2</sub>O<sub>2</sub> (400.26): C, 66.02; H, 5.04; N, 7.00; Found: C: 65.97; H: 5.13; N, 6.94%.

##### 4.4.4. 2-(*S*<sub>p</sub>)-((1*S*,3*S*)-3-(Methoxycarbonyl)-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indol-1-yl)ferrocene-1-carboxylic acid (**7b**)

Red solid. Yield: 1.512 g (33%); mp.: 181-184 °C;  $R_f$ : 0.55;  $[\alpha]_D^{25}$ : -27.5° (CHCl<sub>3</sub>  $c$  = 0.060 g/100 mL); IR (cm<sup>-1</sup>): ~3500-2900, 1746, 1603, 1540, 1438, 1364, 1267, 1219, 1106, 1002, 915, 819, 737.  $^1\text{H}$  NMR (DMSO- $d_6$ ): 10.32 (s, 1H, NH, indole); 7.44 (br d,  $J=7.7$  Hz, 1H, H5); 7.25 (br d,  $J=7.7$  Hz, 1H, H8); 7.03 (br t,

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$J=7.7$  Hz, 1H, H7); 6.97 (td,  $J=7.7$  and 1.7 Hz, 1H, H6); 5.17 (s, 1H, H1); 4.78 (br s, 1H, H5'); 4.72 (br s, 1H, H3'); 4.61 (t,  $J=2.1$  Hz, 1H, H4'); 4.46 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 4.10 (dd,  $J=11.7$  and 4.3 Hz, 1H, H3); 3.87 (s, 3H,  $\text{CO}_2\text{CH}_3$ ); 3.18 (dd,  $J=14.9$  and 4.3 Hz, 1H, H4<sub>A</sub>); 2.88 (ddd,  $J=14.9$ , 11.7 and 1.9 Hz, 1H, H4<sub>B</sub>);  $^{13}\text{C}$  NMR (DMSO- $d_6$ ): 172.4 ( $\text{CO}_2\text{CH}_3$ ); 171.2 ( $\text{CO}_2^-$ ); 137.0 (C8<sub>A</sub>); 133.1 (C9<sub>A</sub>); 126.5 (C4<sub>B</sub>); 121.7 (C7); 119.4 (C6); 118.5 (C5); 111.9 (C8); 105.5 (C4<sub>A</sub>); 89.2 (C1'); 74.5 (C5'); 73.5 (C4'); 72.3 (C3'); 70.9 (C2'); 70.6 ( $\eta^5\text{-C}_5\text{H}_5$ ); 55.9 (C3); 52.8 ( $\text{CO}_2\text{CH}_3$ ); 51.4 (C1); 24.3 (C4); Anal. Calcd. for  $\text{C}_{24}\text{H}_{22}\text{FeN}_2\text{O}_4$  (458.30): C, 62.90; H, 4.84; N, 6.11; Found: C, 62.84; H, 4.95; N, 6.14%.

#### 4.4.5. (*S<sub>p</sub>*)-((*R*)-5-(2,3,4,9-Tetrahydro-1*H*-pyridol[3,4-*b*]indol-1-yl)ferrocene-1-carboxylic acid (**8a**))

Orange solid. Yield: 0.112 g (14%); mp.: 162-164 °C;  $R_f$ : 0.49;  $[\alpha]_D^{25}$ :  $-20.1^\circ$  ( $\text{CHCl}_3$   $c = 0.028$  g/100 mL); IR ( $\text{cm}^{-1}$ ):  $\sim 3650\text{-}2750$ , 1621, 1560, 1451, 1357, 1270, 1103, 995, 808, 740;  $^1\text{H}$  NMR (DMSO- $d_6$ ): 10.68 (s, 1H, NH, indole); 7.46 (br d,  $J=7.5$  Hz, 1H, H5); 7.25 (br d,  $J=7.5$  Hz, 1H, H8); 7.05 (td,  $J=7.5$  and 1.8 Hz, 1H, H7); 7.01 (td,  $J=7.5$  and 1.8 Hz, 1H, H6); 5.45 (s, 1H, H1); 4.85 (br s, 1H, H3'); 4.61 (br s, 1H, H5'); 4.48 (br s, 1H, H4'); 4.02 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 3.73 (ddd,  $J=12.5$ , 6.2 and 4.0 Hz, 1H, H3<sub>A</sub>); 3.23 (ddd,  $J=12.5$ , 5.2 and 4.1 Hz, 1H, H3<sub>B</sub>); 2.94-2.74 (overlapping m's, 2H, H4<sub>A</sub> and H4<sub>B</sub>);  $^{13}\text{C}$  NMR (DMSO- $d_6$ ): 170.8 ( $\text{CO}_2\text{H}$ ); 137.0 (C8<sub>A</sub>); 132.9 (C9<sub>A</sub>); 127.1 (C4<sub>B</sub>); 121.7 (C7); 120.0 (C6); 118.6 (C5); 112.5 (C8); 106.9 (C4<sub>A</sub>); 89.1 (C1'); 74.6 (C5'); 72.0 (C2'); 70.8 ( $\eta^5\text{-C}_5\text{H}_5$ ); 70.6 (C4'); 70.0 (C3'); 52.2 (C1); 41.3 (C3); 20.6 (C4);

#### 4.4.6. 2-(*S<sub>p</sub>*)-((1*R*,3*S*)-3-(Methoxycarbonyl)-2,3,4,9-tetrahydro-1*H*-pyridol[3,4-*b*]indol-1-yl)ferrocene-1-carboxylic acid (**8b**))

Red solid. Yield: 1.283 g (28%); mp.: 174-178 °C;  $R_f$ : 0.48;  $[\alpha]_D^{25}$ :  $-63.4^\circ$  ( $\text{CHCl}_3$   $c = 0.040$  g/100 mL); IR ( $\text{cm}^{-1}$ ):  $\sim 3450\text{-}2900$ , 1742, 1607, 1553, 1441, 1362, 1259, 1224, 1102, 998, 905, 822, 741.  $^1\text{H}$  NMR (DMSO- $d_6$ ): 10.34 (s, 1H, NH, indole); 7.44 (br d,  $J=7.7$  Hz, 1H, H5); 7.30 (br d,  $J=7.7$  Hz, 1H, H8); 7.06 (br t,  $J=7.7$  Hz, 1H, H7); 6.98 (td,  $J=7.7$  and 1.7 Hz, 1H, H6); 5.56 (s, 1H, H1); 4.74 (br s, 1H, H3'); 4.57 (br s, 1H, H5'); 4.55 (br s, 1H, H4'); 4.27 (dd,  $J=5.7$  and 4.2 Hz, 1H, H3<sub>A</sub>); 3.85 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 3.68 (s, 3H,  $\text{CO}_2\text{CH}_3$ ); 3.15-3.11 (overlapping m's, 2H, H4<sub>A</sub> and H4<sub>B</sub>);  $^{13}\text{C}$  NMR (DMSO- $d_6$ ): 173.4 ( $\text{CO}_2\text{CH}_3$ ); 170.3 ( $\text{CO}_2\text{H}$ ); 137.3 (C8<sub>A</sub>); 133.4 (C9<sub>A</sub>); 127.0 (C4<sub>B</sub>); 122.2 (C7); 119.7 (C6); 118.9 (C5); 112.4 (C8); 105.2 (C4<sub>A</sub>); 89.7 (C1'); 74.4 (C5'); 71.2 ( $\eta^5\text{-C}_5\text{H}_5$ ); 70.9 (C2'); 70.5 (C4'); 70.2 (C3'); 53.1 (C3); 52.3 ( $\text{CO}_2\text{CH}_3$ ); 49.7 (C1); 23.6 (C4); Anal. Calcd. for  $\text{C}_{24}\text{H}_{22}\text{FeN}_2\text{O}_4$  (458.30): C, 62.90; H, 4.84; N, 6.11; Found: C, 62.98; H, 4.90; N, 6.05%.

#### 4.5. Cyclization of **7a** with cyanuric fluoride: synthesis of (1*2bS*,*S<sub>p</sub>*,*P*)-4-Oxo-6,7,12,12*b*-tetrahydro-4*H*-ferroceno[1,2]indolizino[8,7-*b*]indole-5-one (**9a**) by Method H

$\beta$ -Carboline **7a** (4.003 g, 10 mmol) was dissolved in dry DCM (50 mL). To this solution pyridine (1.650 mL) and cyanuric fluoride (3.605 mL) were added at 0 °C. After a few minutes the cooling bath was removed and the mixture was stirred at rt for 90 minutes under Ar. After addition of crushed ice the resulted suspension was filtered, the organic phase was separated, washed with cold water (3x50 mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered and evaporated. The crude lactame was purified by flash column chromatography on silica using DCM-MeOH (30:1) as eluent. The orange band was collected, evaporated and triturated with cold EtOH to obtain **9a** in pure form as red needles. Yield: 1.477 g (45%); mp.: 103-105 °C;  $R_f$ : 0.74;  $[\alpha]_D^{25}$ :  $+55.4^\circ$  ( $\text{CHCl}_3$   $c = 0.048$  g/100 mL); IR ( $\text{cm}^{-1}$ ): 3222, 1621, 1521, 1447, 1368, 1303, 1247, 1158, 1106, 1003, 829, 742.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ): 9.11 (s, 1H, NH); 7.59 (br d,  $J=7.6$  Hz, 1H, H8); 7.48 (br d,  $J=7.6$  Hz, 1H, H11); 7.27 (td,  $J=7.6$  and 1.9 Hz, 1H, H10); 7.18 (td,  $J=7.6$  and 1.9 Hz, 1H, H9); 6.95 (s, 1H, H12<sub>b</sub>); 5.18 (br dd,  $J=13.1$  and 5.2 Hz, 1H, H6<sub>B</sub>); 4.96 (br s, 1H, H1); 4.59 (t,  $J=2.0$  Hz, 1H, H2); 4.55 (br s, 1H, H3); 4.06 (s, 5H,  $\eta^5\text{-C}_5\text{H}_5$ ); 3.79 (ddd,  $J=13.1$ , 11.6 and 5.2 Hz, 1H, H6<sub>A</sub>); 3.13-3.12 (overlapping m's, 2H, H7<sub>A</sub> and H7<sub>B</sub>);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ): 168.3 (C4); 135.7 (C11<sub>a</sub>); 133.1 (C12<sub>a</sub>); 125.8 (C7<sub>b</sub>); 123.2 (C10); 120.2 (C9); 119.1 (C8); 111.4 (C11); 107.9 (C7<sub>a</sub>); 92.3 (C12<sub>c</sub>); 74.2 (C3); 72.5 (C1); 71.6 (C2); 71.5 ( $\eta^5\text{-C}_5\text{H}_5$ ); 60.0 (C3<sub>a</sub>); 50.0 (C12<sub>b</sub>); 41.3 (C6); 21.3 (C7); Anal. Calcd. for  $\text{C}_{22}\text{H}_{18}\text{FeN}_2\text{O}$  (382.24): C, 69.13; H, 4.75; N, 7.33; Found: C, 69.23; H, 4.80; N, 7.24%.

#### 4.6. Cyclization of **6a,b**, **7a,b** and **8a,b** with carbonyldiimidazole by Method I

The mixture of 5 mmol of the corresponding imine (**6a,b**) or  $\beta$ -carboline (**7a,b** and **8a,b**), carbonyldiimidazole (CDI, 0.474 g, 6 mmol) and  $\text{NEt}_3\cdot\text{HCl}$  (0.069 g, 1 mmol) dissolved in dry DMF (10 mL) was stirred under an Ar atmosphere at 50 °C for 30 min. The reaction mixture was poured on ice-water and the resulted precipitate was filtered off, washed with water, dried and subjected to flash column chromatography on silica using DCM-MeOH (30:1) as eluent. Lactames **9a,b**, **9b/inv 12a,b**, **14b** and cyclopenta[7,8]azonino[5,4-*b*]indole **13b** obtained by the evaporation of the separated bands were crystallized by cold MeOH. The analytical and spectral data of lactame **9a** (1.147 g, 60% from **7a** and 0.456 g, 24% from **6a**) were practically identical to those measured for the sample prepared by Method H.

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#### 4.6.1. (12bR,S<sub>p</sub>,M)-4-Oxo-6,7,12,12b-tetrahydro-4H-ferroceno[1,2]indolizino[8,7-b]indole-5-one (12a)

Red solid. Yield: 2.219 g (58% by from **6a**); 1.682 g (44% by from **8a**); mp.: 126-128 °C; *R*<sub>f</sub>: 0.82; [α]<sub>D</sub><sup>25</sup>: +56.3° (CHCl<sub>3</sub> *c* = 0.024 g/100 mL); IR (cm<sup>-1</sup>): ~ 3400-2950, 1685, 1611, 1559, 1451, 1375, 1270, 1233, 1106, 901, 830, 749. <sup>1</sup>H NMR (CDCl<sub>3</sub>): 9.24 (s, 1H, NH); 7.54 (br d, *J*=7.8 Hz, 1H, H8); 7.36 (br d, *J*=7.8 Hz, 1H, H11); 7.21 (br t, *J*=7.8, 1H, H10); 7.14 (td, *J*=7.6 and 1.9 Hz, 1H, H9); 6.33 (s, 1H, H12b); 4.98 (ddd, *J*=14.6, 5.4 and 2.0 Hz, 1H, H6<sub>A</sub>); 4.78 (br s, 1H, H1); 4.52 (t, *J*=2.2 Hz, 1H, H2); 4.41 (br s, 1H, H3); 3.68 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 3.54 (ddd, *J*=14.6, 11.2 and 5.4 Hz, 1H, H6<sub>B</sub>); 3.25-3.18 (overlapping m's, 2H, H7<sub>A</sub> and H7<sub>B</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>): 167.1 (C4); 131.6 (C11a); 129.8 (C12a); 124.6 (C7b); 123.0 (C10); 121.0 (C9); 119.4 (C8); 112.1 (C11); 106.7 (C7a); 90.9 (C12c); 74.0 (C3); 71.1 (C1); 70.6 (C2); 70.2 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 63.1 (C3a); 48.2 (C12b); 41.0 (C6); 21.6 (C7); Anal. Calcd. for C<sub>22</sub>H<sub>18</sub>FeN<sub>2</sub>O (382.24): C, 69.13; H, 4.75; N, 7.33; Found: C: 69.01; H: 4.82; N, 7.19%.

#### 4.6.2. Methyl (6S,12bS,S<sub>p</sub>,P)-4-oxo-6,7,12,12b-tetrahydro-4H-ferroceno[1,2]indolizino[8,7-b]indole-6-carboxylate (9b)

Red solid. Yield: 0.220 g (5% from **6b**); 1.631 g (37% from **7b**); mp.: 101-104 °C; *R*<sub>f</sub>: 0.57; [α]<sub>D</sub><sup>25</sup>: -32.9° (CHCl<sub>3</sub> *c* = 0.039 g/100 mL); IR (cm<sup>-1</sup>): ~ 3400-2850, 1740, 1628, 1543, 1451, 1369, 1262, 1230, 1105, 992, 900, 811, 751. <sup>1</sup>H NMR (CDCl<sub>3</sub>): 8.81 (s, 1H, NH); 7.58 (dd, *J*=7.6 and 1.8 Hz, 1H, H8); 7.47 (br d, *J*=7.6 Hz, 1H, H11); 7.06 (td, *J*=7.6 and 1.8 Hz, 1H, H10); 6.98 (td, *J*=7.6 and 1.9 Hz, 1H, H9); 5.89 (s, 1H, H12b); 4.91 (br s, 1H, H1); 4.55 (br s, 1H, H3); 4.27 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 4.22 (t, *J*=2.0 Hz, 1H, H2); 4.18 (dd, *J*=11.8 and 4.3 Hz, 1H, H6<sub>A</sub>); 3.97 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.29 (dd, *J*=15.9 and 11.8 Hz, 1H, H7<sub>B</sub>); 3.09 (dd, *J*=15.9 and 4.3 Hz, 1H, H7<sub>A</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>): 173.2 (C4); 170.2 (CO<sub>2</sub>CH<sub>3</sub>); 136.3 (C11a); 130.7 (C12a); 126.4 (C7b); 122.4 (C10); 119.9 (C9); 118.6 (C8); 111.4 (C11); 105.7 (C7a); 94.0 (C12c); 75.5 (C3a); 72.6 (C2); 70.2 (C3); 68.1 (C1); 71.0 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 55.6 (C6); 52.5 (CO<sub>2</sub>CH<sub>3</sub>); 51.1 (C12b); 23.4 (C7); Anal. Calcd. for C<sub>24</sub>H<sub>20</sub>FeN<sub>2</sub>O<sub>3</sub> (440.28): C, 65.47; H, 4.58; N, 6.36; Found: C: 65.42; H: 4.68; N, 6.50%.

#### 4.6.3. Methyl (6S,12bS,S<sub>p</sub>,M)-4-oxo-6,7,12,12b-tetrahydro-4H-ferroceno[1,2]indolizino[8,7-b]indole-6-carboxylate (9b/inv)

Red solid. Yield: 0.661 g (15% from **6b**); mp.: 92-95 °C; *R*<sub>f</sub>: 0.65; [α]<sub>D</sub><sup>25</sup>: -27.6° (CHCl<sub>3</sub> *c* = 0.041 g/100 mL); IR (cm<sup>-1</sup>): ~ 3450-2850, 1742, 1634, 1543, 1472, 1381, 1242, 1222, 1121, 1008, 919, 835, 772. <sup>1</sup>H NMR (CDCl<sub>3</sub>): 8.29 (s, 1H, NH); 7.50 (br d, *J*=7.7 Hz, 1H, H8); 7.35 (br d, *J*=7.7 Hz, 1H, H11); 7.19 (td, *J*=7.7 and 1.8 Hz, 1H, H10); 7.13 (td, *J*=7.7 and 1.8 Hz, 1H, H9); 6.30 (s, 1H, H12b); 5.64 (d, *J*=7.1 Hz, 1H, H6<sub>A</sub>); 4.94 (br s, 1H, H1); 4.70 (br s, 1H, H3); 4.31 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 4.29 (t, *J*=2.0 Hz, 1H, H2); 3.78 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.41 (d, *J*=15.9 Hz, 1H, H7<sub>B</sub>); 3.22 (dd, *J*=15.9 and 7.1 Hz, 1H, H7<sub>A</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>): 172.3 (C4); 172.0 (CO<sub>2</sub>CH<sub>3</sub>); 136.1 (C11a); 130.7 (C12a); 126.6 (C7b); 122.6 (C10); 120.1 (C9); 118.7 (C8); 111.1 (C11); 106.7 (C7a); 95.1 (C12c); 76.5 (C3a); 72.6 (C2); 71.0 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 64.0 (C1); 62.4 (C3); 53.5 (C12b); 52.6 (CO<sub>2</sub>CH<sub>3</sub>); 50.1 (C6); 24.1 (C7); Anal. Calcd. for C<sub>24</sub>H<sub>20</sub>FeN<sub>2</sub>O<sub>3</sub> (440.28): C, 65.47; H, 4.58; N, 6.36; Found: C: 65.30; H: 4.70; N, 6.29%.

#### 4.6.4. Methyl (6S,12bR,S<sub>p</sub>,M)-4-oxo-6,7,12,12b-tetrahydro-4H-ferroceno[1,2]indolizino[8,7-b]indole-6-carboxylate (12b)

Red solid. Yield: 1.167 g (53% from **6b**); 0.572 g (26% from **8b**); mp.: 116-118 °C; *R*<sub>f</sub>: 0.68; [α]<sub>D</sub><sup>25</sup>: +75.0° (CHCl<sub>3</sub> *c* = 0.046 g/100 mL); IR (cm<sup>-1</sup>): ~ 3500-2850, 1748, 1630, 1549, 1442, 1370, 1284, 1220, 1113, 1002, 905, 820, 764; <sup>1</sup>H NMR (CDCl<sub>3</sub>): 9.17 (s, 1H, NH); 7.59 (br d, *J*=7.8 Hz, 1H, H8); 7.47 (br d, *J*=7.8 Hz, 1H, H11); 7.06 (br t, *J*=7.8 Hz, 1H, H10); 6.98 (br t, *J*=7.8 Hz, 1H, H9); 5.82 (s, 1H, H12b); 5.66 (d, *J*=7.5 Hz, 1H, H6<sub>A</sub>); 4.73 (br s, 1H, H1); 4.71 (br s, 1H, H3); 4.33 (t, *J*=2.2 Hz, 1H, H2); 3.70 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 3.67 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.56 (d, *J*=15.9 Hz, 1H, H7<sub>B</sub>); 3.24 (dd, *J*=15.9 and 7.5 Hz, 1H, H7<sub>A</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>): 171.6 (CO<sub>2</sub>CH<sub>3</sub>); 171.3 (C4); 136.1 (C11a); 132.4 (C12a); 126.56 (C7b); 122.4 (C10); 119.9 (C9); 118.6 (C8); 111.3 (C11); 105.4 (C7a); 94.8 (C12c); 78.3 (C3a); 72.4 (C2); 69.8 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 62.9 (C1); 61.8 (C3); 52.4 (CO<sub>2</sub>CH<sub>3</sub>); 51.9 (C12b); 50.2 (C6); 24.3 (C7); Anal. Calcd. for C<sub>24</sub>H<sub>20</sub>FeN<sub>2</sub>O<sub>3</sub> (440.28): C, 65.47; H, 4.58; N, 6.36; Found: C: 65.62; H: 4.51; N, 6.45%.

#### 4.6.5. Methyl (6S,Z)-4-oxo-5,6,7,12-tetrahydro-4H-cyclopenta[7,8]azonino[5,4-b]indole-6-carboxylate (13b)

Light yellow powder. Yield: 0.096 g (6% from **6b**); 0.144 g (9% from **7b**); 0.352 g (22% by Method *L* from **9b**); 0.256 g (16% by Method *L* from **9b/inv**); mp.: 198-201 °C; *R*<sub>f</sub>: 0.24; [α]<sub>D</sub><sup>25</sup>: +18.2° (CHCl<sub>3</sub> *c* = 0.026 g/100 mL); IR (cm<sup>-1</sup>): ~ 3300-2700, 1746, 1638, 1536, 1486, 1353, 1275, 1251, 1113, 1094, 958; <sup>1</sup>H NMR (CDCl<sub>3</sub>): 9.25 (s, 1H, indole NH); 7.42 (br d, *J*=7.7 Hz, 1H, H8); 7.37 (br d, *J*=7.7 Hz, 1H, H11); 7.06 (td, *J*=7.7

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and 1.9 Hz, 1H, H10); 7.00 (td,  $J=7.7$  and 1.9 Hz, 1H, H9); 6.70 (br s, 1H, H13); 6.76 (br d,  $J=4.6$  Hz, 1H, H3); 6.70 (br d,  $J=8.8$  Hz, 1H, H2); 6.64 (br d,  $J=8.8$  Hz, 1H, H1); 6.56 (d,  $J=7.4$  Hz, 1H, lactame NH); 5.09 (ddd,  $J=7.4$ , 5.5 and 2.5 Hz, 1H, H6); 3.68 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.45 (dd,  $J=14.7$  and 5.5 Hz, 1H, H7<sub>B</sub>); 3.39 (dd,  $J=14.7$  and 2.5 Hz, 1H, H7<sub>A</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>): 173.0 (CO<sub>2</sub>CH<sub>3</sub>); 166.7 ((C4); 135.8 (C13a); 132.3 (C11a); 130.4 (C12a); 127.6 (C7b); 123.5 (C13); 122.6 (C10); 122.1 (C1); 120.2 and 120.1 (C2 and C3); 119.4 (C9); 118.8 (C8); 114.5 (C3a); 111.8 (C11); 110.1 (C7a); 54.3 (CO<sub>2</sub>CH<sub>3</sub>); 52.7 (C6); 28.0 (C7); Anal. Calcd. for C<sub>19</sub>H<sub>16</sub>N<sub>2</sub>O<sub>3</sub> (320.35): C, 71.24; H, 5.03; N, 8.74; Found: C, 71.39; H, 4.97; 8.68%.

#### 4.6.6. Methyl 7-oxo-7H-ferroceno[c]indolo[3,2,1-ij][1,5]naphthyridine-2-carboxylate (**14b**)

Deep red solid. Yield: 0.480 g (22% from **7b**); 0.654 g (30% from **8b**); mp.: 202-203 °C;  $R_f$ : 0.82;  $[\alpha]_D^{25}$ : +32.0° (CHCl<sub>3</sub>  $c = 0.051$  g/100 mL); IR (cm<sup>-1</sup>): 2894, 1722, 1673, 1571, 1440, 1412, 1375, 1297, 1250, 1197, 1180, 1012, 806, 791; <sup>1</sup>H NMR (CDCl<sub>3</sub>): 8.90 (s, 1H, H1); 8.62 (br d,  $J=7.9$  Hz, 1H, H9); 8.51 (br d,  $J=7.9$  Hz, 1H, H12); 7.79 (td,  $J=7.9$  and 1.8 Hz, 1H, H10); 7.59 (td,  $J=7.9$  and 1.8 Hz, 1H, H11); 5.64 (dd,  $J=2.5$  and 1.6 Hz, 1H, H4); 5.44 (dd,  $J=2.5$  and 1.6 Hz, 1H, H6); 5.02 (t,  $J=2.5$  Hz, 1H, H5); 4.12 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 4.00 (s, 3H, OCH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>): 167.2 (C7); 166.0 (CO<sub>2</sub>CH<sub>3</sub>); 146.4 (C2); 142.7 (C3a); 139.0 (C8a); 133.5 (C12c); 131.1 (C10); 125.4 (C11); 125.0 (C12a); 124.0 (C12b); 123.9 (C12); 116.6 (C9); 116.5 (C1); 81.6 (C3b); 74.7 (C5); 73.6 (C6a); 71.7 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 70.0 (C6); 67.8 (C4); 53.0 (CO<sub>2</sub>CH<sub>3</sub>); Anal. Calcd. for C<sub>24</sub>H<sub>16</sub>FeN<sub>2</sub>O<sub>3</sub> (436.25): C, 66.08; H, 3.70; N, 6.42; Found: C, 65.97; H, 3.62; N, 6.48%.

#### 4.7. Acylation of tryptamine with (*S<sub>p</sub>*)-2-formylferrocenecarbonylfluoride (**10**): synthesis of (*S<sub>p</sub>*)-*N*-(2-(1*H*-indol-3-yl)ethyl)-2-formylferrocene-1-carboxamide (**11**) by Method J

The mixture of tryptamine (1.601 g, 10 mmol), **10** (2.601 g, 10 mmol) and dry THF (40 mL) was stirred under argon at 25 °C for 24 h then evaporated to dryness. The residue was triturated with 20 mL of EtOH-H<sub>2</sub>O (1:3), filtered off and dried to give amide **11** as a dark orange solid. Yield: 2.842 g (71%); mp.: 204-207 °C;  $[\alpha]_D^{25}$ : +34.2° (EtOH  $c = 0.037$  g/100 mL); IR (cm<sup>-1</sup>): ~ 3400-2700, 1695, 1641, 1453, 1337, 1249, 1226, 1125, 1104, 1020, 883, 827, 738; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>): 10.83 (br s, 1H, H1); 10.42 (s, 1H, CHO); 8.54 (t,  $J=5.6$  Hz, 1H, CONH); 7.62 (br d,  $J=7.8$  Hz, 1H, H4); 7.35 (br d,  $J=7.8$  Hz, 1H, H7); 7.23 (br d,  $J=2.0$  Hz, 1H, H2); 7.08 (br t,  $J=7.8$  Hz, 1H, H6); 7.01 (br t,  $J=7.8$  Hz, 1H, H5); 5.27 (dd,  $J=2.6$  and 1.5 Hz, 1H, H5'); 4.95 (dd,  $J=2.6$  and 1.5 Hz, 1H, H3'); 4.83 (t,  $J=2.6$  Hz, 1H, H4'); 4.23 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 3.59 and 3.53 (2xm, 2x1H, NCH<sub>A</sub>H<sub>B</sub> and NCH<sub>A</sub>H<sub>B</sub>); 2.99 (t,  $J=7.3$  Hz, 2H, CCH<sub>2</sub>); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>): 196.2 (CHO); 168.9 (CONH); 137.2 (C7a); 128.2 (C4a); 123.6 (C2); 121.8 (C6); 119.2 and 119.1 (C4 and C5); 112.7 (C3); 112.3 (C7); 80.1 (C1'); 79.2 (C2'); 75.5 (C5'); 74.1 (C4'); 72.8 (C3'); 72.0 (η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 40.6 (NCH<sub>A</sub>H<sub>B</sub>); 26.1 (CCH<sub>2</sub>); Anal. Calcd. for C<sub>22</sub>H<sub>20</sub>FeN<sub>2</sub>O<sub>2</sub> (400.26): C, 66.02; H, 5.04; N, 7.00; Found: C, 66.20; H, 5.12; N, 6.94%.

#### 4.8. Ring inversion **9b/inv** → **9b** monitored by <sup>1</sup>H-NMR spectroscopy

Lactame **9b/inv** (0.044g, 0.1 mmol) was dissolved in DMSO-*d*<sub>6</sub> (1 mL) and the resulted solution was subjected to <sup>1</sup>H-NMR studies. First the <sup>1</sup>H-NMR spectrum was registered at 300 K, then solution was incubated at 363 K for ca. 30 min then cooled down to 300 K, and the <sup>1</sup>H-NMR spectrum registered showed the presence of inverted lactame **9b** contaminated with a small amount (ca. 3-5%) of **9b/inv**.

**9b/inv**: <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>): 10.75 (br s, 1H, NH, indole), 7.45 (br d,  $J=7.6$  Hz, 1H, H8); 7.29 (br d,  $J=7.6$  Hz, 1H, H11); 7.20 (br t,  $J=7.6$  Hz, 1H, H10); 7.13 (td,  $J=7.7$  and 1.8 Hz, 1H, H9); 6.08 (s, 1H, H12b); 5.55 (d,  $J=7.3$  Hz, 1H, H6<sub>A</sub>); 5.02 (br s, 1H, H1); 4.65 (br s, 1H, H3); 4.22 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 4.30 (t,  $J=2.0$  Hz, 1H, H2); 3.76 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.51 (d,  $J=15.6$  Hz, 1H, H7<sub>B</sub>); 3.18 (dd,  $J=15.6$  and 7.3 Hz, 1H, H7<sub>A</sub>); 7.50 (br d,  $J=7.7$  Hz, 1H, H8); 7.35 (br d,  $J=7.7$  Hz, 1H, H11); 7.19 (td,  $J=7.7$  and 1.8 Hz, 1H, H10); 7.13 (td,  $J=7.7$  and 1.8 Hz, 1H, H9); 6.30 (s, 1H, H12b); 5.64 (d,  $J=7.1$  Hz, 1H, H6<sub>A</sub>); 4.94 (br s, 1H, H1); 4.70 (br s, 1H, H3); 4.31 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 4.29 (t,  $J=2.0$  Hz, 1H, H2); 3.78 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.41 (d,  $J=15.9$  Hz, 1H, H7<sub>B</sub>); 3.22 (dd,  $J=15.9$  and 7.1 Hz, 1H, H7<sub>A</sub>);

**9b**: <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>): 10.54 (s, 1H, NH, indole); 7.52 (br d,  $J=7.6$  Hz, 1H, H8); 7.40 (br d,  $J=7.6$  Hz, 1H, H11); 7.06 (br t,  $J=7.6$  Hz, 1H, H10); 6.98 (br t,  $J=7.6$  Hz, 1H, H9); 5.94 (s, 1H, H12b); 4.85 (br s, 1H, H1); 4.59 (br s, 1H, H3); 4.28 (s, 5H, η<sup>5</sup>-C<sub>5</sub>H<sub>5</sub>); 4.21 (t,  $J=2.0$  Hz, 1H, H2); 4.26 (dd,  $J=11.3$  and 4.6 Hz, 1H, H6<sub>A</sub>); 4.05 (s, 3H, CO<sub>2</sub>CH<sub>3</sub>); 3.31 (dd,  $J=15.7$  and 11.3 Hz, 1H, H7<sub>B</sub>); 3.11 (dd,  $J=15.7$  and 4.6 Hz, 1H, H7<sub>A</sub>). These signals were practically identical to those discernible in the <sup>1</sup>H-NMR spectrum of an original sample of **9b** registered in DMSO-*d*<sub>6</sub> solution in a separate experiment.

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#### 4.9. Imidazole-mediated ring enlargement of lactames **9b** and **9b/inv** associated with the decomposition of the ferrocene residue: synthesis of **13b** by Method L.

The corresponding lactame (0.440 g, 1 mmol) and imidazole (0.082 g, 1.2 mmol) were dissolved in DMF (4 mL). The solution was stirred at 110 °C for 20 min under argon. The dark reaction mixture was poured onto cold water (30 mL) and the resulted suspension was extracted with DCM (3x40 mL). The combined organic phase was washed with water (5x50 mL), dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated to dryness. The dark solid residue was subjected to flash column chromatography over silica using DCM-MeOH (10:1) as eluent. The evaporation of the first orange band recovered **9b** as evidenced by <sup>1</sup>H NMR data practically identical to those presented in section 4.5.2. Yield: 0.251 g (57% from **9b**) and 0.216 g (49% from **9b/inv**). (Under the employed conditions ring inversion **9b/inv**→**9b** obviously accompanies the conversion leading to **13b**.) Changing eluent to DCM-MeOH (5:1) allowed the isolation of **13b** as light yellow powder. Yield: 0.070 g (22% from **9b**) and 0.051 g (16% from **9b/inv**). The analytical and spectral data were practically identical to those measured for the sample prepared by Method I (cf. section 4.6.5).

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