1	Cerebrospinal fluid parameters of horses with West Nile virus neuroinvasive disease

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21	The authors declare no conflict of interest.

22 Abstract

23 Objective: To compare biochemical and cytological findings of cerebrospinal fluid (CSF)

samples in horses with acute neuroinvasive West Nile virus (WNV) infections with those of

25 control healthy horses.

26 Design: Retrospective case-control study.

Samples: Fifteen CSF samples from horses with acute WNV neuroinvaisve disease (WNVND)and twenty from healthy horses.

29 Procedures: WNVND was diagnosed based on acute neurologic symptoms and positive IgM

30 ELISA results. CSF samples were collected either from the atlanto-occipital or the lumbosacral
31 sites.

32 Results: CSF results of the WNV affected group did not follow normal distribution. Protein,

33 creatine-kinase, aspartate-aminotransferase, lactate-dehydrogenase, alkaline-phosphatase,

34 magnesium, glucose, and lactate concentrations showed abnormal levels in a number of WNV

cases. None of the 6 horses with elevated glucose concentrations survived (<=0.36, modified

36 Wald method). Opposite to previous equine studies we have found neutrophilic pleocytosis in

37 54% of cases. Measured data also indicates that CSF neutrophilia is more likely to be found

parallel with high protein content (Fisher exact test, p = 0.1026).

Conclusions and clinical relevance: The CSF findings with WNVND are nonspecific and
variable. Neutrophils likely play a role in the development of inflammatory response and brain
damage. Increased enzyme activities and changes in the electrolyte concentrations reflect CNS
cellular injury rather than blood-brain barrier leakage. Although elevated glucose levels reliably
predicted outcome, these results might be the consequences of increased plasma levels and reflect

- 44 general stress rather than any CNS pathophysiology. Examination of CSF is most useful when the
- 45 results are correlated with history, clinical findings and ancillary laboratory studies.

46 Abbreviations

- 47 CSF cerebrospinal fluid
- 48 WNV West Nile virus
- 49 WNVND West Nile Virus Neuroinvasive Disease
- 50 CNS central nervous system
- 51 AST aspartate-aminotransferase
- 52 ALP alkaline-phosphatase
- 53 CK creatine-kinase
- 54 LDH lactate-dehydrogenase
- 55 RT-PCR reverse transcriptase polymerase chain reaction
- 56 CI confidence interval

57 Introduction

58 West Nile virus (WNV) is a mosquito-borne zoonotic arbovirus belonging to the genus Flavivirus

in the family Flaviviridae¹, and transmitted in natural cycles between mosquitoes, (mainly the

60 genus Culex), and wild birds^{2,3}. Horses and human beings are incidental and dead-end hosts, but

- 61 severe neurological disorders can develop in horses^{4,5}. Phylogenetic studies have identified 2
- 62 main lineages of WNV strains. The Hungarian equine WNV outbreak reported in 2008 was the
- first to be caused by a lineage 2 sub-Saharan strain in Europe. The pathogenicity of this lineage 2

strain resembled that of lineage 1 strains, and its sudden spread was unpredictable^{2,4}. During the 64 last decade, the epidemiology of WNV in human beings has changed in the southern regions of 65 Europe, with high incidence of West Nile fever cases, but also of WNVND⁶. 66 Depending on the level of viremia WNV can cross the blood-brain barrier into the brain and 67 cause meningo-encephalitis⁵. Due to its close contact with the extracellular fluid of the brain, 68 analysis of cerebrospinal fluid (CSF) composition can reflect biological central nervous system 69 70 (CNS) impairments enabling the diagnosis and understanding of various neurodegenerative CNS disorders⁷. To obtain accurate results when evaluating equine CSF samples, apart from the 71 precise sampling technique, the application of the correct laboratory analytical methods is also 72 73 important. However, due to the varied methods that different laboratories use today, it has become imperative for each laboratory to establish its own reference ranges based on the 74 calibrated measurement techniques⁸. Until now, few studies investigated the association between 75 the imbalance of CSF elements and the severity of WNV infection. The aim of the present study 76 was to compare biochemical and cytological findings of CSF in horses with acute neuroinvasive 77 WNV infections with those of control healthy horses. 78

79 Materials and methods

The data were obtained performing an observational retrospective case-control study between
2008 and 2014. The study was permitted by the Animal Health and Welfare Directorate of
National Food Chain Safety Office (22.1./1606/003/2009).

83 West Nile virus neuroinvasive cases were defined based on seasonality (August-November),

84 acute neurologic clinical signs (less than 5 days), positive serum IgM ELISA test^a and the

85 absence of any WNV vaccination in their history. Only clinically healthy horses without any

86	neurologic signs and with hematological and biochemistry parameters within reference interval
87	were included in the control group. Age, breed, gender characteristics and sampling sites are
88	described in table 1.

All WNV cases were sampled within 36 h of clinical admission, sampling site was determined 89 based on the clinical signs. Horses with characteristics of more pronounced brainstem and 90 cerebral involvement were sampled by atlanto-occipital puncture during general anesthesia (1 91 92 mg/kg [2.2 mg/lb] xylazine iv., 0.02 mg/kg [0.045 mg/lb] butophanol iv. and 1 mg/kg [2.2 mg/lb] 93 ketamine iv.) and horses with spinal cord involvement and 12 control horses were sampled in sedation (0.3-0.4 mg/kg xylazine [0.66-0.88 mg/lb] iv. and 0.02 mg/kg [0.045 mg/lb] butorphanol 94 iv.) with local anesthesia (lidocaine) on the lumbosacral site as previously described⁹. In case of 95 diffuse CNS involvement both locations were sampled under general anesthesia. Altogether 96 fifteen samples were collected and 2 horses were sampled both by lumbosacral and atlanto-97 occipital punctures. 98

99 CSF was first analyzed macroscopically for color and turbidity in front of a white paper.

100 Cytological analysis was performed within 6 h of sampling after cytocentrifugation and Wright-

101 Giemsa staining. Protein content was measured with spectrophotometry^b and other biochemical

102 parameters like aspartate-aminotransferase (AST), alkaline-phosphatase (ALP), glucose, lactate,

103 urea, creatine-kinase (CK), lactate-dehydrogenase (LDH), sodium, potassium, calcium, chloride,

anorganic phosphate, and magnesium were determined spectrophotometrically using

105 commercialy available test kits on an a chemistry analyzer^c.

106 In each case we attempted to characterize the virus from peripheral blood leukocytes or brain and

107 spinal cord samples by virus isolation, nested reverse transcriptase polymerase chain reaction

108 (RT-PCR), real-time RT-PCR, and sequencing techniques as described previously⁴.

When analyzing our results, first we evaluated normality of our data, whether parametric or nonparametric statistics should be used. We used Wald method and Fisher exact test to evaluate which measured variables are predictors of the outcome. Finally Fisher exact test was applied to establish relationships between the measured parameters^d.

113 **Results**

All CSF sampling procedures were performed without any complications. Horses of the control group recovered quickly from the anesthesia without having any sequela. Three horses from the WNV group were euthanized right after the sampling procedure and three other were euthanized on human grounds within the next 5 days. Seven horses survived the neuroinvasive disease, five without any residual symptoms. In all 6 cases where it was identified with PCR or virus isolation, lineage 2 strain was responsible for the infection.

On macroscopical examination the CSF was transparent and non-turbid in all control animals and in 9 WNV cases and slightly hazy in 6 WNV cases. Cytological analysis revealed normal cell counts within reference intervals with exclusively small and large mononuclear cells on all control samples and on three WNV samples. There were 4/15 cases with mononuclear and 8/15 cases with neutrophilic pleocytosis in the diseased group.

Most of the data obtained from the WNV neuroinvasive cases did not seem to follow a normal distribution, their mean, mode and median were not close to being equal, hence t-test based comparison of the means with the control group was not feasible. Instead we opted to establish reference ranges (95% prediction intervals) based on the control groups and count the number of cases for each variable that fall outside of this range. We have found that protein, CK, AST,

130	LDH, ALP, magnesium, glucose, and lactate concentrations showed abnormal levels in a number
131	of cases. The following table (2. table) describes our results.
132	We also studied if any of the measured variables are a good predictor of the outcome
133	(death/survival) of the disease. Most noteworthy was that none of the 6 horses with elevated
134	glucose levels survived the disease $(0/6, \le 0.36, \text{modified Wald method with } 90\% \text{ CI})$ and all of
135	the 6 horses with normal glucose levels have survived $(6/6, \geq 0.64, \text{ modified Wald method with})$
136	90% CI). The dependence of the outcome on the glucose level was also verified with a Fisher
137	exact test (two-tailed, p=0.0022).
138	Measured data also indicated that neutrophilic pleocytosis in CSF was more likely in the cases
139	with high total protein content (Fisher exact test, two tailed, $p = 0.1026$).
140	In the two WNV cases were both samples were collected, results differed based on the location.
141	In one case atlanto-occipital sample cytology was negative, while lymphocytic pleocytosis was
142	found on lumbosacral puncture also showing higher protein, glucose and lactate levels. In the
143	other case lymphocytic pleocytosis was found in the lumbosacral sample, and more neutrophils
144	with higher protein content, CK, LDH, AST and lower urea were identified in the atlantoocipital
145	one, while glucose levels were similar. None of these horses survived.

146

147 **Discussion**

148 Limitations of the study were the relative low sample number according to sampling sites and

that blood biochemistry and hematology parameters were not measured and evaluated parallel.

150 The reference ranges set up by our control group were in concordance with previous

151 findings^{10,11,12}, except that lactate concentration being slightly and LDH value being moderately

higher in our reference group. This could be attributed to different methodology used by ourlaboratory.

According to previous studies in human beings, the CSF findings in patients with WNVND are 154 nonspecific and include pleocytosis (neutrophil or lymphocyte predominance) with elevated 155 protein and normal glucose levels¹³. Our findings are very similar except we have found high 156 glucose levels in nonsurvival patients. On the other hand previous reports in horses most 157 commonly described mononuclear pleocytosis with lymphocytic predominance¹⁴. Although 158 WNV disease was caused by lineage 1 strains in those cases and lineage 2 strains were 159 160 responsible in the present report, lineage differences are not likely to be the reasons for these 161 discrepancies. Other studies on human CSF also resulted variable data, where patients did not present with the typical lymphocytic pleocytosis often quoted when discussing a viral 162 meningitis/encephalitis; rather most presented with a cerebrospinal fluid neutrophilia^{13,15,16}. 163 Previous results in mice suggest that neutrophils are the predominant immune cells that are 164 initially and rapidly recruited to sites of infection with WNV⁵. According to a study in human 165 patients¹⁵ mean total leukocyte counts and mean neutrophil fractions were greater in individuals 166 sampled within the first 3 days of symptoms than in those sampled beyond day 3. Sampling time 167 168 might also be responsible for the different findings. Most of our horses were sampled relative 169 early (all horses within 5 days and 8 within 3 days of onset of clinical signs) in the disease process. In another study it was found that older WNV patients were more likely to have 170 neutrophils in their CSF¹⁷. The average age of our patients with high neutrophil numbers was 8 171 172 years (4-13 years) and with mononuclear pleocytosis it was 6.6 years (4-9 years). Furthermore, 173 neutrophil-related proteins were found at higher levels in CSF of WNVND patients, underlining the likely key role played by neutrophils in the development of the inflammatory response and 174

brain damage⁶. We have also found that CSF neutrophilia is more likely to be found parallel with 175 high protein content. Albumin was lower in most cases and total protein was increased suggesting 176 the presence of increased inflammatory proteins like globulins in the CSF of diseased animals. 177 Opposite to some previous data, we have detected normal or high CSF glucose concentration 178 similarly to a study of seasonal human epidemic West Nile Virus meningo-encephalitis¹⁵. 179 Cerebrospinal glucose concentrations might reflect changes of blood glucose which could be 180 increased because of critical illness causing a stress response^{11,18}. Human and animal studies 181 suggest that this is not benign, and that stress-induced hyperglycemia is associated with a high 182 risk of mortality^{19,20}. Increased lactate levels were found in most of the cases as well increased 183 184 LDH levels in half of the samples in the WNV affected group. L-lactate is formed during normal anaerobic glycolysis by interconversion from pyruvate via the actions of LDH²¹. Lactate 185 concentrations in the CSF largely represent its production by the brain but it is also increased in 186 case of low glucose concentrations to meet the energy requirements by the anaerobic pathway²¹. 187 Hypoglycemia was not present in our cases. Increases in CSF lactate concentration reportedly 188 occur with bacterial infections but not with nonseptic meningitis⁹. On the other hand CSF lactate 189 increases also occur with any condition that results in reduced brain oxygenation and/or increased 190 191 intracranial pressure. CNS tissue hypoxia could be the result of inflammatory processes 192 secondary to the WNV infection.

As most enzymes are relatively large molecules, there is very little diffusion across the intact and normal blood-CSF barrier and increased activities of the enzymes in the CSF are assumed arise from cells within the CNS²². Potential sources of the increased enzyme activity in the WNV group horses are the release of these enzymes from the inflammatory cells or directly from the damaged nerve cells and myelin. Recently, it has been shown that WNV induced the expression of interleukin-1 β , -6, -8, and tumor necrosis factor- α , where neurons were one of the potential sources of pro-inflammatory cytokines, and these pro-inflammatory mediators were one of the main factors driving WNV-induced neurotoxicity, cell death and CNS tissue damage⁵. Based on previous histologic findings of WNV encephalitis including perivascular inflammation, microgliosis, variable degree of necrosis, and loss of neurons it is less likely that the source of increased enzymes is secondary to a damaged blood-brain barrier or blood-CSF barrier and increased leakage from the plasma.

The most reliable increase was demonstrated by the alkaline phosphatase enzyme. The CSF of 205 patients without neurological disorders contains little or no activity as it was also shown in our 206 study based on our control group²³. According to a previous human study the CSF alkaline 207 phosphatase activities of patients with meningitis and other neurological disorders varied directly 208 with the number of polymorphonuclear leukocytes present and with the protein concentration 23 . 209 We could not demonstrate a clear relationship between the number of neutrophils, the level of 210 protein and the alkaline phosphatase activity. The reason for this might have been the low sample 211 number. 212

There is surprisingly little information about urea levels in the normal CSF, although an increase
would have significance in differentiating uremic encephalopathy. Since urea is readily
diffusible, therefore urea levels should be parallel that found in the serum²¹. Decreased urea
levels in some of our patients' sample could be secondary to reduced hepatic synthesis of urea
from ammonia in case of severe systemic disease. None of our patients had increased urea
concentration in their CSF.

Electrolyte composition of the CSF has been only sparsely reported, but in general the CSF
sodium, chloride are similar or slightly higher, potassium concentrations are similar or slightly

lower and magnesium concentrations are slightly higher than those in the serum^{10,12}. Our 221 reference ranges based on the control group were concordant with these results previously 222 published. In some of the WNV affected horses low sodium and increased magnesium 223 concentration could be detected. These electrolyte abnormalities can originate from the cellular 224 225 damage in the CNS, where intracellular solutes may leak out of the cell because of an increased membrane permeability and may lead to redistribution of sodium and an increased magnesium. 226 Based on studies in humans, phosphate is found in normal CSF at levels of 50-60% of expected 227 228 serum concentration and it has also previously been observed that CSF anorganic phosphate concentrations increase in direct proportion to total CSF protein levels²¹. Although we have 229 measured increased anorganic phosphate concentration in three cases, similar relationship could 230 not be demonstrated. 231

When we collected both lumbosacral and atlanto-occipital samples from the same patient, we got 232 different results. This can be attributed to the different locations of the more severe CNS damage 233 causing more significant alterations in the sample collected from the closer site. Although there is 234 a synchronous appearance of WNV at many sites in the brain and spinal cord and pathological 235 alterations can be detected in many parts of the CNS, but the severity of these damages can differ 236 237 which is also reflected in the detectable clinical signs and disease progression. On the other hand 238 values of certain parameters differ depending on sampling sites even in the healthy horses because of different blood-CSF permeability and flow rates between the atlanto-occipital and the 239 lumbar regions⁸. 240

CSF analysis is a general index of neurological health and often provides evidence of the
presence of a disease. Similar to a complete blood count, CSF analysis has reasonable sensitivity
but low specificity. The CSF findings with WNVND are nonspecific and variable and possibly

depend on the age of the patient, the sampling time, the site of sampling in relation to the location 244 245 of the most severe lesions and also on previous treatments. Neutrophils likely play a role in the 246 development of inflammatory response and brain damage but further examinations would be required to fully elucidate their role in the pathogenesis of WNVND. Increased enzyme activities 247 248 and changes in the electrolyte concentrations reflect CNS cellular injury rather than blood-brain barrier damage. Higher sample number would be required to demonstrate relationships between 249 250 inflammation, CNS damage and changes of the CSF parameters. Although elevated glucose 251 levels reliable predicted the outcome, these results might be secondary to increased plasma levels 252 and reflect general stress to serious illness rather than any CNS pathophysiology. Based on all these findings, examination of CSF is most useful when the results are correlated with history, 253 clinical findings and ancillary laboratory studies. 254

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256 **Footnotes**

- ^a IDEXX IgM WNV Ab Test, Hoofddorp, The Netherlands
- ^b Olympus AU400, Beckman Coulter, Hamburg, Germany
- ^c Olympus AU640, Beckman Coulter, Hamburg, Germany
- 260 ^dIBM SPSS Statistics 20 Documentation, United States

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262 **References**

- 2631. Smithburn KC, Hughes TP, Burke AW et al. A neurotropic virus isolated from the blood
- of a native of Uganda. Am J Trop Med 1940;20:471–492.

265	2.	Bakonyi T, Ferenczi E, Erdélyi K, et al. Explosive spread of a neuroinvasive lineage 2
266		West Nile virus in Central Europe, 2008/2009. Vet Microbiol 2013;26(165):61-70.
267	3.	Szentpáli-Gavallér K, Antal L, Tóth M et al. Monitoring of West Nile virus in mosquitoes
268		between 2011-2012 in Hungary. Vector Borne Zoonotic Dis 2014;14:648-55.
269	4.	Kutasi O, Bakonyi T, Lecollinet S et al. Equine encephalomyelitis outbreak caused by a
270		genetic lineage 2 West Nile virus in Hungary. J Vet Intern Med 2011;25(3):586-91.
271	5.	Lim SM, Koraka P, Osterhaus AD et al. West Nile virus: immunity and pathogenesis.
272		Viruses 2011;3(6):811-28.
273	6.	Fraisier C, Papa A, Granjeaud S et al. Cerebrospinal Fluid Biomarker Candidates
274		Associated with Human WNV Neuroinvasive Disease. PLoS ONE 2014;9(4):e93637.
275	7.	Shahim P, Mansson JE, Darin N et al. Cerebrospinal fluid biomarkers in neurological
276		diseases in children. Eur J Paediatr Neurol 2013;17:7-13
277	8.	Vernau W, Vernau KA, Bailey CS. Cerebrospinal Fluid. In: Kaneko JJ, Harvey JW, Bruss
278		ML eds. Clinical biochemistry of domestic animals, 6th ed. USA: Elsevier Inc., 2008;769-
279		819.
280	9.	Schwarz B, PIERCY RJ. Cerebrospinal fluid collection and its analysis in equine
281		neurological disease. Equine Vet Educ 2006;18:243-248.
282	10	. Mayhew IG, Whitlock RH, Tasker JB. Equine Cerebrospinal Fluid: Reference Values of
283		Normal Horses. Am J Vet Res 1977;38:1271-1274.
284	11.	. Andrews FM, Matthews HK, Reed SM. The ancillary techniques and tests for diagnosing
285		equine neurologic disease. Vet Med 1990;85:1325-1330.
286	12	. Macwilliams PS. Cerebrospinal Fluid. In: Cowel RL, Tyler RD eds. Diagnostic cytology
287		and hematology of the horse. 2 nd ed. USA: Mosby, 2002;171-179.

288	13. Tyler KL, Pape J, Goody RJ et al. CSF findings in 250 patients with serologically
289	confirmed West Nile virus meningitis and encephalitis.Neurology 2006;66:361-365.
290	14. Wamsley HL, Alleman AR, Porter MB et al. Findings in cerebrospinal fluids of horses
291	infected with West Nile virus: 30 cases (2001). J Am Vet Med Assoc 2002;221(9):1303-
292	5.
293	15. Rawal A1, Gavin PJ, Sturgis CD. Cerebrospinal fluid cytology in seasonal epidemic Wes
294	Nile virus meningo-encephalitis. Diagn Cytopathol 2006;34(2):127-9.
295	16. Crichlow R, Bailey J, Gardner C. Cerebrospinal Fluid Neutrophilic Pleocytosis in
296	Hospitalized West Nile virus Patients. MD J Am Board Fam Pract 2004;17:470 –2.
297	17. Jordan M, Nagpal A, Newman W et al. Plasma cell cerebrospinal fluid pleocytosis does
298	not predict West Nile virus infection. J Biomed Biotechnol 2012;2012:697418.
299	18. Dean A, Seehusen MD, Mark M, et al. Cerebrospinal fluid analysis. Am Fam Physician
300	2003;68(6):1103-1109.
301	19. Capes SE, Hunt D, Malmberg K et al. Stress hyperglycemia and prognosis of stroke in
302	nondiabetic and diabetic patients: a systematic overview. Stroke 2001;32(10):2426–32.
303	20. Guo Y-J, Zhou Y, Zhang S-Y et al.: Optimal target range for blood glucose in
304	hyperglycaemic patients in a neurocritical care unit. Diabetes and Vascular Disease
305	Research 2014;11:352-358.
306	21. Irani D.N. Properties and composition of normal cerebrospinal fluid. In: Irani D.N. ed
307	Cerebrospinal Fluid in Clinical Practice. Elsevier Health Sciences, USA, 2009:69-93.
308	22. Furr M. Cerebrospinal fluid and the blood brain barrier, in Furr M, Reed S. eds. Equine
309	neurology, 2 nd ed, Blackwell Publishing, USA, 2008:21-36.

- 23. McComb RB, Bowers GN, Posen JR et al. Clinical Utilization of Alkaline Phosphatase
- 311 Measurements in McComb RB, Bowers GN, Posen JR et al. Alkaline phosphatase.
- 312 Plenum Press, New York and London. 1979:525-786.
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Table 1: Age, breed, gender characteristics and sampling sites

Group	Age	Breed	Gender	Sampling site	
WNV affected	M: 7.53 years,	12 warmbloods	9 mares	8 lumbosacral	
	SD: 2.84	1 pony	4 geldings	7 atlanto-occipital	
Control	M: 8.94 years,	16 warmbloods	10 mares	12 lumbosacral	
	SD: 3.57	3 draught horses	10 geldings	8 atlanto-occipital	
		1 thoroughbred			

321 M: mean, SD: standard deviation

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	inflammatory	inflammatory	albumin	total	AST	ALP	GGT	glucose	lactate	СК	LDH	urea	Na	К	Ca	Cl	Р	Mg
	proteins (total- albumin)	(mg/l)	protein (mg/l)	IU/I	IU/I	IU/I	mmol/l	mmol/l	ІИЛ	IU/I	mmol/l	mmol/l	mmol/l	mmol/l	mmol/l	mmol/l	mmol/l	
control reference	0-56	10-50	32,16- 75,55	6,0-14,0	0,1-3,5	0-4,0	2,54-3,81	1,89-3,07	0-4,6	14,7- 44,1	4,6-8,8	140-151,8	2,8-3,1	1,13-1,41	113- 128,2	0,02-0,38	0,53-1,34	
horses with norm values	4	3	6	6	1	12	6	3	5	6	9	8	11	6	7	9	7	
horses with abnorm values	8	9	7	6	11	0	6	7	7	6	3	4	1	0	5	3	5	
mean WNV group	59	11	98,36	12,85	8,16	1,10	3,89	3,81	20,07	46,57	4,80	136,96	2,92	1,24	120,53	0,43	1,15	
sd	0,32	0,10	54,76	5,45	4,80	0,93	1,54	1,88	24,32	31,20	1,61	40,25	0,85	0,51	36,22	0,24	0,55	
mean control group	27	26	50,32	9,9235	1,9	1,62	2,98	2,45	2,17	29,49	6,73	145,08	2,95	1,26	121,575	0,3095	0,89	
sd	15	12	11,85	2,22	1,01	1,65	0,32	0,30	1,46	13,02	1,29	3,10	0,08	0,07	3,49	0,24	0,19	
			5-100 ¹⁰	15-50 ¹⁰			1,67- 3,89 ^{10,12}											
other references			20-124 ¹¹ 20-80 ¹²	0-16 ¹¹ 7-24 ¹²			30-70% of blood glucose ¹¹	1,92-2,311	0-8 ^{10,11,12}	0-8 ^{10,11}	2/,	140- 150 ^{10,12}	2,5- 3,5 ^{10,12}		95- 123 ^{10,12}			

Table 2: Results of cerebrospinal fluid analysis compared to previously published data^{10,11,12}.

Note that inflammatory protein is a calculated value based on the total protein and albumin levels.

Na: sodium, K: potassium, Ca: calcium, Cl: chloride, P: phosphate, Mg: magnesium, sd: standard deviation