

## ***In silico* analysis of hypermethylation in CpGislands of *UCHL1* gene's promoter in nasopharyngeal carcinoma**

Lao Duc Thuan<sup>1</sup>, Thieu Hong Hue<sup>1</sup>, Hoang Van Nam<sup>1</sup>, Nguyen Hoang Danh<sup>1</sup>, Le Huyen Ai Thuy<sup>1\*</sup>, Ngo Dong Kha<sup>2</sup>, Nguyen Hoang Anh Tuan<sup>2</sup>

<sup>1</sup>Ho Chi Minh City Open University, Vietnam

<sup>2</sup>University of Science, Vietnam National University Ho Chi Minh City, Vietnam

\*Corresponding author: [thuy.lha@ou.edu.vn](mailto:thuy.lha@ou.edu.vn)

---

### ARTICLE INFO

### ABSTRACT

DOI:10.46223/HCMCOUJS.tech.en.8.1.333.2018

Received: February 11<sup>th</sup>, 2018

Revised: March 20<sup>th</sup>, 2018

Accepted: April 16<sup>th</sup>, 2018

**Keywords:**

methylation, MSP, NPC, UCHL-1 gene

**Background and Objective:** The methylation of the Ubiquitin C-Terminal Hydrolase L1 (*UCHL1*) gene has been reported in many human cancers including nasopharyngeal carcinoma (NPC). In Vietnam, the methylation of the *UCHL1* gene's promoter in NPC has not been demonstrated yet. In this study, a systematic literature revision was carried out to summarize the current evidence about the frequencies of the *UCHL-1* gene's promoter methylation in NPC for further application in the Vietnamese population.

**Methods:** A systematic literature analysis was conducted based on comprehensive studies. Moreover, many bioinformatic tools such as Methprimer, TFsearch, IDT OligoAnalyzer 3.1 were used to predict the CpG islands, transcriptional factors, and to pick up the MSP (Methylation-Specific PCR) primers.

**Results:** A total of three previous studies were summarized and accessed for eligibility from literature research. As the results, the average weight methylated frequencies were 72.4% and 13.0% for NPC and non-cancerous samples, respectively. The significant association between *UCHL-1* promoter methylation and NPC with the OR of 10.459 (95% CI = 4.915 - 22.254,  $p < 0.001$ ) and RR of 4.117 (95% CI = 1.958 - 6.645,  $p < 0.0001$ ) based on the random-effects model, was observed. Moreover, we were successful in predicting the CpG islands as well as identifying transcriptional factor binding sites that served as "hot spots" for ideal primer pick up and located in the gene promoter.

**Conclusion:** The methylation of the *UCHL-1* gene promoter was significantly associated and contributed to NPC development in which could be further applied in the evaluation of *UCHL-1* gene promoter status in the Vietnamese population.

---

## 1. Introduction

The DNA methylation that refers to a covalent modification of Cytosine ring at the 5' position of CpG dinucleotide by adding a methyl group in the 5<sup>th</sup> Carbon of the ring by S-adenosyl methionine has been identified as the common mechanism of tumor suppressor genes (TSGs) silenced, involves in many human diseases (Le, Lao, & Truong, 2017; Yuanyuan, Haiyang, Haiyan, Xiaokun, & Shulin, 2014). It is noted that the CpG dinucleotides are concentrated in the upstream promoter regions of many genes, therefore, the promoter methylation of genes involved in many cells progresses such as DNA repair, cell-cycle regulation, signal transduction has already confirmed in various tumor types, including nasopharyngeal carcinoma (NPC) (Le et al., 2017; McCabe & Caudill, 2005; Tsao et al., 2014). A series of TSGs are silenced by promoter CpG methylation identified in NPC, including *UCHL1*, which play a key role in NPC pathogenesis (Ayadi et al., 2008; Chan et al., 2010; Kwong et al., 2007; Li et al., 2010; Tian et al., 2013).

Concerning to NPC, NPC has a striking and geographic distribution, which gravitates toward China and Vietnam (Da Costa, Marques-Silva, & Moreli, 2015; Dai, Zheng, Cheung, & Lung, 2016; Pathmanathan, Prasad, Sadler, Flynn, & Raab-Traub, 1995). Notably, Vietnam is considered as one of most panic areas which are contributed to these indices, due to the total number were 4,931 cases (ASR = 5.4/100,000) and deaths were 2,885 cases (ASR = 3.3/100,000) (Globocan, 2012). The major obstacle to early diagnosis and screening of NPC are the different access due to the deeply seated location of the nasopharynx, as well as the unclear presenting symptoms, such as hearing loss, nosebleeds, headache, trouble opening the mouth, etc.,(ref). Consequently, to achieve favorable treatment and increase patient survival, early diagnosis and screening are necessary to be established. The major etiological factors proposed for nasopharyngeal cancer include viral infection, genetic susceptibility, the aberrant methylation of CpG islands belonged to the TSGs' promoter, and environmental factors (Tsao et al., 2014). Herein, we summarize some recent studies on the CpG aberrant methylation, as well as the prediction of CpG islands in the *UCHL1* gene for further using methylated genes as a molecular marker. This may provide a useful biomarker for future diagnosis and prognosis in Vietnamese cancer patients.

## 2. Materials and methods

### 2.1. Search strategy, selection criteria, eligibility and data extraction

Firstly, a systematic literature analysis was conducted in the Medline database, using PubMed, was carried out for the *UCHL1* promoter methylation in NPC, published before October 2017. The following keywords: Nasopharyngeal carcinoma, *UCHL1*, methylation, epigenetics, and nasopharyngeal cancer were used for literature research. For these articles, the type of study separating prospective studies from others, mainly based on case-control studies were extracted. When data were available, the range and average methylation frequencies of the candidate gene in both case-control were reported. Additionally, the detection of methylated genes in NPC is technically feasible by numerous techniques invented for the mapping of Cytosine methylation. Thus, kinds of the method were systematically enrolled into our current *in vitro* studies to have a general vision in techniques permitting the highly specific and sensitive identification of CpG methylation.

## 2.2. CpG island prediction

For picking primer, CpG islands were necessary to be predicted with the promoter sequence of the candidate gene. The promoter sequence and information of the *UCHL1* gene were downloaded from the Ensemble database (<https://asia.ensembl.org/index.html>). Several bioinformatics programs such as Methprimer ([www.urogen.org](http://www.urogen.org)), TFsearch ([http://cbrc3.cbrc.jp/papia/howtouse/howtouse\\_tfsearch.html](http://cbrc3.cbrc.jp/papia/howtouse/howtouse_tfsearch.html)) were applied for predicting CpG islands and transcriptional factor binding sites located in CpG islands in promoter regions of *UCHL1* gene. For evaluation of MSP primers, prime physical characteristics were computed by IDT OligoAnalyzer 3.1 (<http://sg.idtdna.com/calc/analyzer>).

## 3. Results and discussion

### 3.1. Systematic literature analysis

After exclusion of studies that did not meet the inclusion criteria, up to now, only three studies were carried out for the analysis of the *UCHL1* promoter aberrant methylation in NPC. The frequencies of *UCHL1* promoter were observed in different range of methylation. Overall, the mean weighted average of methylated *UCHL1* were 72.40% and 13.00% in NPC samples and non-cancerous samples, respectively (Table 1). These results indicated that the frequency of *UCHL1* promoter in NPC samples was higher than normal samples. Concerning to methods, the MSP assay was the majority technique for mapping methylated candidate gene status in NPC. Additionally, *UCHL1* promoter methylation was associated with NPC risk with OR of 10.459 (95% CI = 4.915 - 22.254,  $p < 0.001$ ) and RR of 4.117 (95% CI = 1.958 - 6.645,  $p < 0.0001$ ) based on the random effects model (Table 2).

**Table 1**

Studies included in the systematic review

Studies	Year	Country	Method	SCS	SC	MT	MTF (%)	MC	MCF (%)
Li et al.	2010	China	MSP	Tissue	Tissue	34/41	82.93	3/9	33.33
Loyo et al.	2011	HongKong	qMSP	Biopsy	Biopsy	32/50	64.00	2/28	7.14
Tian et al.	2013	China	MSP	Serum	Serum	24/37	64.86	8/41	19,52
Average weight frequency (%)							72.40		13.00

Note: SCS: Source of cancer sample; SC: Source of control; MT: Methylation Tumor; MC: Methylation control; MSP: Methylation specific PCR; MTF: Methylation frequency in NPC samples; MCF: Methylation frequency in non-cancerous samples; qMSP: quantitative Methylation specific PCR

Source: The researcher's data analysis

**Table 2**

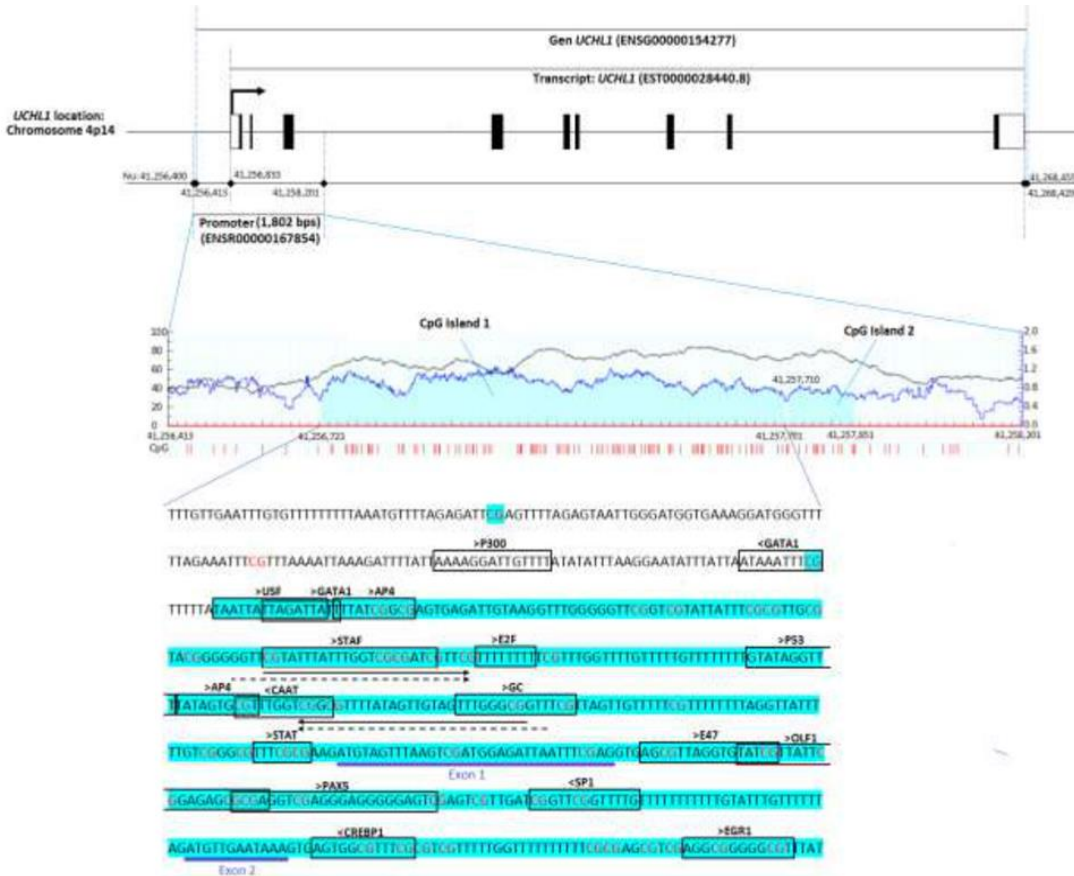
Odds ratio and Relative risk of *UCHL1* gene promoter using Random-effects model based on previous studies

Studies	OR	95% CI	P	W (%)	RR	95% CI	P	W (%)
Li et al.	9.714	1.948 - 48.455		22.08	2.49	0.977 - 6.333		28.99
Loyo et al.	23.11	4.906 - 108.871		23.74	8.96	2.319 - 34.618		13.85

Studies	OR	95% CI	P	W (%)	RR	95% CI	P	W (%)
Tian et al.	7.62	2.730 - 21.241		54.19	3.32	1.709 - 6.467		57.16
Total	10.459	4.915 - 22.254	< 0.001	100.00	4.117	1.958 - 6.645	< 0.001	100.00

Source: The researcher's data analysis

### 3.2. Prediction of CpG islands



**Figure 1.** (A): Schematic structure of UCHL1 gene (locus: 4p13) and its promoter and exons (9 exons); (B): CpG Islands located in the promoter regions; (C) The predictive transcription factors and MPS primers identified in the CpG1. Line arrow: methylated primer; Dash arrow: an unmethylated primer

The *UCHL1* gene's sequence, promoter and transcript were collected from the Ensembl database via accession number: ENSG00000154277, ENSR00000167854 and EST0000028440.8, respectively. *UCHL1* gene locates in 4p11-14, Nu. 41,256,833 to Nu. 41,268,455. The promoter of the *UCHL1* gene locates in Nu. 41,256,413 to Nu. 41,258,201 (Figure 1A). The CpG sites located in the *UCHL1* gene promoter were identified by using MethPrimer. As the result, two CpG sites in the promoter of the candidate gene were identified to be located in Nu. 41,256,723 to Nu. 41,257,701 (yielded 979 bps) and Nu. 41,257,710 to Nu. 41,257,851 (yielded 142 bps) (Figure 1B). In the current study, CpG Island 1 (CpG1) was chosen to be searched for transcriptional factors and picked up the MSP primers. The results of

transcription factor prediction showed many transcription factor binding sites were identified, such as STAF, CAAT, E2F, GC (Figure 1C). Notably, the CpG1 covered exon 1 (Nu. 41,256,977 to Nu. 41,257,009), exon 2 (Nu. 41,257,115 to Nu. 41,257,126) and a partial exo (Nu. 41,257,609 to Nu. 41,257,737). For MSP primer, the forward methylated/unmethylated primer and reverse methylated/unmethylated primer were picked up as covered five CpG sites, and three CpG sites, respectively (Figure 1C). The primer sequences were shown in Table 3.

**Table 3**

The sequences of MSP primers

Primer	Sequence (5' - 3')
M_F	CGTATTTATTTGGTC <b>CGCG</b> ATCGTTC
M_R	CGCCCAA <b>ACTACA</b> ACTATA <b>AAACG</b> CCG
U_F	GGTTTGTATTTATTTGGTT <b>GTG</b> ATTGTTT
U_R	AACC <b>ACCCAA</b> ACTACA <b>ACTATA</b> AAACACCA

Note: F: Forward primer; R: Reverse primer; CpG sites were in bold characters

Source: The researcher's data analysis

#### 4. Discussion

To our knowledge, no research related to characterizing the methylation status of the *UCHL1* gene promoter in NPC has been carried out in the Vietnamese population. In our study, we systematically evaluated the promoter methylation profile of the *UCHL1* gene, which was considered as an important mechanism of its inactivation in NPC, based on previous studies, in order to have further studied on methylation of *UCHL1* gene in Vietnamese NPC samples. Even though, limited researches were carried out on the evaluation of methylation of *UCHL1* gene's promoter, a moderate-sized case-control group that had indicated the significant higher methylation in NPC samples (counting for 72.40%, based on the calculation of average weight frequency) than in non-cancerous samples (counting for 13.00%). It could be explained that previous studies have shown that mechanism by which *UCHL1* functions as a tumor suppressor gene is through ubiquitinating oncoproteins and deubiquitinating tumor suppressor genes (Li et al., 2010). Additionally, *UCHL1* has been shown to promote the degradation of cell cycle inhibitor p27<sup>kip1</sup> and stabilize NF- $\kappa$ B and p53 (Caballero et al., 2002; Li et al., 2010; Shen, Sikorska, Leblanc, Walker, & Liu, 2006; Takami et al., 2007; Tokumaru et al., 2008). By collecting the methylation/unmethylation of *UCHL1* frequencies in both NPC samples/non-cancerous, the OR and RR value were calculated, indicating that the *UCHL1* promoter methylation was significantly associated with an increased NPC risk with a pooled OR of 10.459 ( $p < 0.0001$ ) and RR of 4.117 ( $p < 0.0001$ ). The OR represents the odds (10.459 times) for cancer risk in case of *UCHL1* promoter methylation occurred, compared to the absence of *UCHL1* promoter methylation. Thus, it suggested that methylation of the *UCHL1* promoter would appear the promising methylated gene to obtain an appropriate performance for NPC screening, and further applied in clinical analysis.

Several bioinformatics tools were applied to identify CpG islands located in the promoter region, as well as many key transcription factors located in the CpG island. In the

current study, the CpG1 was chosen for methylation analysis, the CpG1 covers many CpG sites than CpG2 (Data not shown). Additionally, exon 1, exon 2 and partial exon were covered by CpG1. We confirmed that CpG sites located in several transcription factors, such as STAF (contains four CpG sites), GC (contains two CpG sites), thus, it was chosen for picking MSP primers, including forwarding primer covers five CpG sites, reverse primer covers three CpG sites, for evaluating the methylation of *UCHL1* gene's promoter. In the current study, the MSP primers were similar to Tian et al. (2013), however, in our methylated primer (forward) and unmethylated primer (both forward and reverse primer) were covered more than one CpG site, which located in the transcription binding factor: CAAT, GC, than the research Tian et al. (2013), which may be increased the specificity of evaluation of the methylated status of *UCHL1* gene.

Therefore, based on the analysis of previous studies, we confirmed that the methylation of the *UCHL1* gene's promoter was significantly associated and contributed to NPC tumorigenesis. Those data will be useful for further experiments related to analyzing the *UCHL1* gene's promoter methylation status in the Vietnamese population, that whether or not could be served as a promising biomarker for prognosis and early diagnosis NPC.

## 5. Conclusion

A significant association between *UCHL1* promoter methylation and NPC was shown and confirmed by a systematic analysis of previous studies. Additionally, CpG island and MSP were successfully identified that covering many transcriptional factors, in which methylation may cause nasopharyngeal tumorigenesis. Further studies will be looked at an experiment carried on clinical NPC samples/non-cancerous samples in the Vietnamese population.

---

## References

- Ayadi, W., Karray-Hakim, H., Khabir, A., Feki, L., Charfi, S., Boudawara, T., ... Hammami, A. (2008). Aberrant methylation of p16, DLEC1, BLU and E-cadherin gene promoters in nasopharyngeal carcinoma biopsies from Tunisian patients. *Anticancer Research*, 28(4B), 2161-2167.
- Caballero, O. L., Resto, V., Patturajan, M., Meerzaman, D., Guo, M. Z., Engles, J., ... Jen, J. (2002). Interaction and colocalization of PGP9.5 with JAB1 and p27(Kip1). *Oncogene*, 21(19), 3003-3010.
- Chan, W. H., Chang, K. P., Yang, S. W., Yao, T. C., Ko, T. Y., Lee, Y. S., ... Tsai, C. N. (2010). Transcriptional repression of DLEC1 associates with the depth of tumor invasion in oral squamous cell carcinoma. *Oral Oncology*, 46(12), 874-879.
- Da Costa, V. G., Marques-Silva, A. C., & Moreli, M. L. (2015). The Epstein-Barr virus latent membrane protein-1 (LMP1) 30-bp deletion and XhoI-polymorphism in nasopharyngeal carcinoma: a meta-analysis of observational studies. *Systematic Reviews*, 2015, 13-46.
- Dai, W., Zheng, H., Cheung, A. K., & Lung, M. L. (2016). Genetic and epigenetic landscape of nasopharyngeal carcinoma. *Chinese Clinical Oncology*, 5(2), 1-13.

- Globocan. (2012). *Estimated cancer incidence, mortality and prevalence worldwide in 2012*. Retrieved January 10, 2018, from [http://globocan.iarc.fr/Pages/fact\\_sheets\\_population.aspx](http://globocan.iarc.fr/Pages/fact_sheets_population.aspx)
- Kwong, J., Chow, L. S., Wong, A. Y., Hung, W. K., Chung, G. T., To, K. F., ... Lo, K. W. (2007). Epigenetic inactivation of the deleted in lung and esophageal cancer 1 gene in nasopharyngeal carcinoma. *Genes Chromosomes Cancer*, 46(2), 171-180.
- Le, H. A. T., Lao, D. T., & Truong, K. P. (2017). DNA hypermethylation in breast cancer. In P. Pham (Ed.), *Breast cancer - From biology to medicine*. London, UK: IntechOpen.
- Li, L., Tao, Q., Jin, H., van Hasselt, A., Poon, F. F., Wang, X., ... Cao, Y. (2010). The tumor suppressor UCHL1 forms a complex with p53/MDM2/ARF to promote p53 signaling and is frequently silenced in nasopharyngeal carcinoma. *Clinical Cancer Research*, 16(11), 2949-2958.
- Loyo, M., Brait, M., Kim, M. S., Ostrow, K. L., Jie, C. C., Chuang, A. Y., ... Sidransky, D. (2011). A survey of methylated candidate tumor suppressor genes in nasopharyngeal carcinoma. *International Journal of Cancer*, 128(6), 1393-1403.
- McCabe, D. C., & Caudill, M. A. (2005). DNA methylation, genomic silencing, and links to nutrition and cancer. *Nutrition Reviews*, 63(6), 183-195.
- Pathmanathan, R., Prasad, U., Sadler, R., Flynn, K., & Raab-Traub, N. (1995). Clonal proliferations of cells infected with Epstein-Barr virus in pre-invasive lesions related to nasopharyngeal carcinoma. *The New England Journal of Medicine*, 333, 693-698.
- Shen, H., Sikorska, M., Leblanc, J., Walker, P. R., & Liu, Q. Y. (2006). Oxidative stress regulated expression of ubiquitin Carboxyl-terminal Hydrolase-L1: Role in cell survival. *Apoptosis*, 11(6), 1049-1059.
- Takami, Y., Nakagami, H., Morishita, R., Katsuya, T., Cui, T. X., Ichikawa, T., ... Kaneda, Y. (2007). Ubiquitin carboxyl-terminal hydrolase L1, a novel deubiquitinating enzyme in the vasculature, attenuates NF-kappaB activation. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 27(10), 2184-2190.
- Tian, F., Yip, S. P., Kwong, D. L., Lin, Z., Yang, Z., & Wu, V. W. (2013). Promoter hypermethylation of tumor suppressor genes in serum as potential biomarker for the diagnosis of nasopharyngeal carcinoma. *Cancer Epidemiol*, 37(5), 708-713.
- Tokumaru, Y., Yamashita, K., Kim, M. S., Park, H. L., Osada, M., Mori, M., & Sidransky, D. (2008). The role of PGP9.5 as a tumor suppressor gene in human cancer. *International Journal of Cancer*, 123(4), 753-759.
- Tsao, S. W., Yip, Y. L., Tsang, C. M., Pang, P. S., Lau, V. M., Zhang, G., & Lo, K. W. (2014). Etiological factors of nasopharyngeal carcinoma. *Oral Oncology*, 50(5), 330-338.
- Yuanyuan, D., Haiyang, Z., Haiyan, L., Xiaokun, L., & Shulin, Y. (2014). DNA methylation as an early diagnostic marker of cancer (Review). *Biomedical Reports*, 2(3), 326-330. doi:10.3892/br.2014.237.