Screening of bat faeces for arthropod-borne apicomplexan protozoa: *Babesia canis* and *Besnoitia besnoiti*-like sequences from Chiroptera

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Abstract

Background:

Microbats (Chiroptera: Microchiroptera) are among the most eco-epidemiologically important mammals, owing to their presence in human settlements and animal keeping facilities. Roosting of bats in buildings may bring pathogens of veterinary-medical importance into the environment of domestic animals and humans. In this context bats have long been studied as carriers of various pathogen groups. However, despite their close association with arthropods (both in their food and as their ectoparasites), only a few molecular surveys have been published on their role as carriers of vector-borne protozoa. The aim of the present study was to compensate for this scarcity of information.

Findings:

Altogether 221 (mostly individual) bat faecal samples were collected in Hungary and the Netherlands. The DNA was extracted, and analysed with PCR and sequencing for the presence of arthropod-borne apicomplexan protozoa. *Babesia canis canis* (with 99-100% homology) was identified in five samples, all from Hungary. Because it was excluded with an Ixodidae-specific PCR that the relevant bats consumed ticks, these sequences derive either from insect carriers of *Ba. canis*, or from the infection of bats. In one bat faecal sample from the Netherlands a sequence having the highest (99%) homology to *Besnoitia besnoiti* was amplified.

Conclusions:

These findings suggest that some aspects of the epidemiology of canine babesiosis are underestimated or unknown, i.e. the potential role of insect-borne mechanical transmission and/or the susceptibility of bats to *Ba. canis*. In addition, bats need to be added to future studies in the quest for the final host of *Be. besnoiti*.

Keywords: vector-borne, Chiroptera, faecal DNA, Apicomplexa, *Dermacentor*, *Stomoxys*

Background

Microbats, known for their nocturnal activity and echolocation, belong to the second largest order (Chiroptera) of mammals and have a world-wide geographical distribution except arctic areas and deserts [1]. The great majority of their species are insectivorous, and therefore ecologically and economically important regulators of natural insect populations. Microbats also have a high epidemiological significance, due to their ability of "true flying" (frequently connected to migratory habit) and their presence in human settlements. In particular, roosting of bats in buildings (attics, cellars, stables) may bring pathogens of veterinary-medical importance into the environment of domestic animals and humans, thus increasing the chance of acquiring related infections. In this scenario bats have features that may further enhance their eco-epidemiological role, as exemplified by ubiquitous occurrence, long life-span, social behaviour (close contacts and allogrooming in colonies) and tendency for persistent infections [2].

Accordingly, bats are increasingly recognized as reservoirs or carriers (vectors) of various pathogen groups [3]. However, while numerous studies focused on emerging viruses (e.g. [2]) and bacteria (e.g. [4]) associated with bats, only a few recent, molecular surveys have been reported on their role as carriers of vector-borne protozoa [5] – despite the close association of bats with arthropods (both in their food and as their ectoparasites). Therefore, the present study was initiated to screen bat samples for arthropod-borne protozoa (Apicomplexa: Piroplasmida and related groups).

For this molecular survey bat faeces was chosen as the sample source, in part because of its non-invasive availability (that is a primary concern when handling small bodied, highly protected animal species). In addition, molecular investigation of bat faeces proved to be useful in taxonomical identification of macroscopic prey insects [6]. On the other hand, to the best of our knowledge, this method was hitherto not used to reveal the presence of arthropod-

borne protozoa bats may have contact with. Demonstration of microbial/protozoan DNA from bat faeces is not only informative on prey insect (or bat intestinal) pathogens. It may also have relevance to the role bats may play as potential reservoirs of extraintestinal apicomplexans, because invasive stages or intracellular forms of these may cross the gut barrier. In this way the DNA of haemotropic protozoa may pass in detectable amounts with the faeces, as exemplified by *Plasmodium* spp. in primates [7].

Methods

Between May and September, 2014, 196 individual and 25 pooled bat faecal samples were collected (192 on 38 locations in Hungary, and 29 on 10 locations in the Netherlands: Figure 1). The study involved the following 19 bat species (sample number): *Nyctalus noctula* (21), *N. leisleri* (9), *Myotis alcathoe* (23), *M. daubentonii* (49), *M. bechsteini* (21), *M. emarginatus* (6), *M. myotis* (8), *M. dasycneme* (4), *M. brandtii* (6), *M. nattereri* (13), *M. blythii* (5), *Rhinolophus ferrumequinum* (3), *R. hipposideros* (2), *Pipistrellus nathusii* (3), *P. pipistrellus* (14), *P. pygmaeus* (1), *Barbastella barbastellus* (6), *Miniopterus schreibersii* (1), *Plecotus auritus* (1). These bats were caught (as part of a monitoring program) at the entrance of caves between sunset and dawn, using standard Ecotone mist-nets (Gdynia, Poland) with 12 m length, 2.5 m height and 14 × 14 mm mesh. After ringing the bats were individually held in sterile paper bags (i.e. one bat per one bag) until sufficient defecation. The standard sample size was three to five faecal pellets for each individual bat. The individual faecal pellets were transferred into numbered, screw cap plastic tubes and stored frozen at -20 °C until evaluation.

DNA was extracted with the QIAamp Fast DNA Stool Mini Kit (QIAGEN, Hilden, Germany) according to the manufacturer's instructions and including extraction controls.

All samples were molecularly screened with a conventional PCR that amplifies an approx. 500 bp long part of the 18S rDNA gene of piroplasms [8]. This method also detects other apicomplexan genera, including vector-borne haemogregarines and certain cystogenic coccidia [9]. The primers BJ1 (forward: 5'-GTC TTG TAA TTG GAA TGA TGG-3') and BN2 (reverse: 5'-TAG TTT ATG GTT AGG ACT ACG-3') were used. The reaction volume was 25 µl, i.e. 5 µl of extracted DNA was added to 20 µl of reaction mixture containing 0.5 unit HotStarTaq Plus DNA polymerase (5U/ µl), 200 µM PCR nucleotid mix, 1 µM of each primer and 2.5 µl of 10× Coral Load PCR buffer (15 mM $MgCl_2$ included). For amplification an initial denaturation step at 95 °C for 10 min was followed by 40 cycles of denaturation at 95 °C for 30 s, annealing at 54 °C for 30 s and extension at 72 °C for 40 s. Final extension was performed at 72 °C for 5 min.

Electrophoresis and visualization of the PCR product was done in a 1.5% agarose gel, followed by sequencing (Biomi Inc., Gödöllő, Hungary). Representative sequences were deposited in the GenBank (accession numbers are shown in Table 1). Phylogenetic analyses were conducted according to the Tamura-Nei model [10] and Maximum Composite Likelihood method by using MEGA version 5.2 [11].

In addition, the presence of hard tick (Acari: Ixodidae) DNA in the bat faeces was evaluated by a conventional PCR that amplifies a 460 bp portion of the mitochondrial 16S rDNA gene of Ixodidae, with the forward primer 16S+1 (5'-CTG CTC AAT GAT TTT TTA AAT TGC TGT GG-3') and reverse primer 16S-1 (5'-CCG GTC TGA ACT CAG ATC AAG T-3'). The original method [12] was slightly modified by using 1.0 unit of HotStartTaq Plus DNA polymerase in a reaction mixture as above, and a thermal profile of initial denaturation step at 95 °C for 5 min, followed by 40 cycles of denaturation at 94 °C for 40 s, annealing at 51 °C for 1 min, extension at 72 °C for 1 min, and final extension at 72 °C for 10 min.

Exact confidence interval (CI) for the prevalence rate was calculated at the 95% level.

147

148 **Ethical approval**

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150 Authorization for bat capture was provided by the National Inspectorate for Environment,
151 Nature and Water (No. 14/2138-7/2011). Bat banding licence number is TMF-14/
152 32/2010 (DK) and 59/2003 (PE).

153

154 ## Results and Discussion

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156 ***Babesia canis* DNA in bat faeces**

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158 *Babesia canis canis* (referred to as *Ba. canis* onwards) DNA was shown to be present in five
159 individual samples (prevalence 2.7%, CI: 0.9-6.2%), all from Hungary (Table 1). Two
160 sequences were identified (accession numbers KP835549-50) with 2 nucleotide differences
161 (inversion of GA to AG at positions 151-152 in the 18S rDNA gene). These bat-derived
162 *Babesia* isolates showed 100% identity with two *Ba. canis* isolates from dogs in Croatia
163 (FJ209024 and FJ209025: [13]), and in phylogenetical comparison they clustered together
164 with other *Ba. canis* isolates (Figure 2). On the other hand, the relevant sequences exhibited
165 only 88% similarity to *Ba. vesperuginis* (AJ871610) known to infect bats (Figure 2). All five
166 bats with *Ba. canis* PCR positive faecal samples were caught within 50 km of the two regions
167 in Hungary (Figure 1), where the highest number of *Ba. canis* seropositive dogs were found in
168 a previous countrywide survey [14].

169 Taken together, this may be the first molecular evidence that both main European
170 genotypes of *Ba. canis* (group A, B: [15]) occur in Hungary.

There are three possible explanations for this unexpected finding. First, relevant bats may have eaten infected tick vectors of *Ba. canis*, i.e. *Dermacentor reticulatus*. To evaluate this possibility, the five *Babesia*-positive faecal DNA samples were molecularly analysed for the presence of tick DNA (mitochondrial 16S rDNA gene). All five samples were PCR negative. If relevant bats (with *Ba. canis* PCR positive faeces) have ingested infected tick vectors, the DNA of *D. reticulatus* should have been detected in their faeces, similarly to that of other prey arthropods [6]. This is supported by literature data: although bats also feed on arachnids, to the best of our knowledge ticks were never reported to be part of their diet (e.g. [6, 16]).

Alternatively, blood-sucking flies (e.g. *Stomoxys* spp.) are known to be incriminated as mechanical vectors in the transmission of *Babesia* spp. [17]. *Stomoxys calcitrans* (also called "dog fly") was reported to frequently bite dogs [18], and to be a predominant species in the diet of some bat species [19]. Therefore, *Ba. canis* DNA in bat faeces may have originated from haematophagous flies which had sucked blood on parasitaemic dogs (in an opportunity offered by the two regions highly endemic for *Ba. canis*), and were consequently eaten by the relevant bats. Unfortunately, two factors precluded to test this hypothesis in the present study, i.e. (1) the whole faecal sample of relevant bats was used for DNA extraction (thus morphological analysis of fly remnants was not possible), and (2) to the best of our knowledge PCR-based molecular methods specific for *S. calcitrans* are not available.

However, the presence of *B. canis* DNA in the faeces may also indicate the infection of relevant bats (i.e. parasitaemia), in which case *Babesia* DNA could get from the circulation into the gut contents (similarly to the DNA of other erythrocyte-infecting protozoa, e.g. *Plasmodium* spp. in primates: [7]). In support of this possibility, among the preferred rodent hosts of *D. reticulatus* larvae/nymphs [20] many *Apodemus* spp. are arboreal, i.e. known for their climbing habit on trees [21]. *Dermacentor* larvae and nymphs were reported to be

196 present in such arboreal nests [22], and in this way may be shared between rodents and bats
197 [23]. All four bat species with *Ba. canis* PCR positive faeces (Table 1) are known for their
198 preference of tree holes as summer roosting places [1, 24], where they could thus have
199 become infested with *Dermacentor* larvae/nymphs (as reported for *Pipistrellus pipistrellus*
200 sampled in July: [25]). Therefore, it cannot be completely excluded that those bats, which
201 were PCR positive in their faeces, may have actually become infected with *Ba. canis* – a
202 protozoan hitherto reported from two mammalian orders (besides Carnivora also from
203 Perissodactyla: [26]), both taxonomically closely related to Chiroptera [27].
204
205 **_Besnoitia besnoiti_-like DNA in bat faeces**
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207 From one pooled faecal sample of a pond bat (*Myotis dasycneme*) colony roost in the
208 Netherlands another sequence was identified, having the highest (99%) homology with
209 *Besnoitia besnoiti* (Table 1). The sequence (accession number KP835555) had six nucleotide
210 difference from, but clustered together with *Be. besnoiti* and *Be. tarandi* (Figure 2). It showed
211 less (98%) homology with (i.e. nine nucleotide difference) and clustered separately (Figure 2)
212 from a cystogenic coccidium, *Nephroisospora eptesici* recently identified from New World
213 bats [28]. To the best of our knowledge, this is the first finding of a *Besnoitia*-like sequence
214 from a non-ungulate mammal in Europe, and from any bat species in a world-wide context.
215 The source of the *Be. besnoiti*-like sequence in the present study, the pond bat (*Myotis*
216 *dasycneme*) is known to be a long distant migratory species (up to 300 km seasonal migration:
217 [29]), and the closest endemic focus of bovine besnoitiosis in northern France is situated
218 within 300 km of the relevant sampling site [30]. In general, bats frequently use cattle stables
219 for roosting [31], where they may have access to the mechanical vectors of *Be. besnoiti*, i.e.
220 blood-sucking flies (*S. calcitrans*, *Tabanus* spp.) and mosquitoes [30]. In particular, *Tabanus*

spp. and mosquitoes develop in wet soil near water and in water, respectively, corresponding to the main habitat of the pond bat. Blood-sucking flies (especially *S. calcitrans*) were also reported to constitute a significant portion of bat prey insects [19]. Therefore, the *Be. besnoti*-like sequence in the present study may have originated from cattle via blood-sucking dipterans, or represents a novel *Besnoitia* genotype/species closely related to *Be. besnoiti*.

On the other hand, *Besnoitia* cystozoites (carried by flies) are able to penetrate mucosal surfaces [32]. Accordingly, the quest for the final host of *Be. besnoiti* should be extended to include chiropterans, particularly because experimental infection with another *Besnoitia* sp. was shown to establish in bats [33].

Conclusions

These findings suggest that some aspects of the epidemiology of canine babesiosis are underestimated or unknown, i.e. the potential role of insect-borne mechanical transmission and/or the susceptibility of bats to *Ba. canis*. In addition, bats need to be added to future studies in the quest for the final host of *Be. besnoiti*.

In the present study no mixed infections were detected. This can be explained by the relatively low prevalence of those apicomplexans, the DNA of which could be amplified with the applied method [8] from bat faeces.

Toxoplasma gondii was reported to infect at least some of the bat species evaluated in the present study [34]. This apicomplexan is able to invade most nucleated cells (including cells crossing the gut barrier), and it was shown to be present in bat liver as well [35], therefore its DNA is likely to be shed in bat faeces. However, *T. gondii* was not detected in the present study. This can be explained by the inability of the applied method [8] to amplify toxoplasma DNA, because the forward primer BJ1 cannot anneal to the 18S rDNA gene of *T. gondii* with its 3' end, unlike in the case of piroplasms, *Besnoitia* and *Sarcocystis* spp. [9].

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Authors' contributions

SH initiated and supervised the Hungarian part of the study, designed molecular analyses, wrote the manuscript. PE and DK collected the Hungarian individual bat faecal samples. BF participated in sample collection and extracted the DNA. NT performed the molecular and JK the phylogenetic analyses. KSZ participated in the sample collection. MG supervised additional sample analyses. AK and AJH were in charge of Dutch sample collection. AF and RF were consultants on besnoitiosis and babesiosis, respectively. HS supervised the Dutch part of the study.

Competing interests

No competing interests exist.

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Legends to figure:

Figure 1. Map of Hungary (A) and Neherlands (B) showing the sampling sites. Only places at least 10 km apart are shown. The red dots on the map of Hungary (A) indicate places, where *Babesia canis* PCR positive bat pellets were collected. The shaded red circles mark the highly endemic regions of *Babesia canis* according to [14]. The red dot on the map of Netherlands (B) indicates the location, where the *Besnoitia besnoiti*-like sequence originated.

Figure 2. Phylogenetic comparison of 18S rDNA sequences of arthropod-borne apicomplexan protozoa identified in the present study (inverse colour), with related sequences from the GenBank. Branch lengths correlate to the number of substitutions inferred according to the scale shown.