#### **RESEARCH ARTICLE**



# Association of microbiota-derived propionic acid and Alzheimer's disease; bioinformatics analysis

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#### **Abstract**

**Purpose** Microbiota-derived metabolites could alter the brain tissue toward the neurodegeneration disease. This study aims to select the genes associated with Propionic acid (PPA) and compromise Alzheimer's disease (AD) to find the possible roles of PPA in AD pathogenesis.

**Methods** Microbiota-derived metabolites could alter the brain tissue toward the neurodegeneration disease. This study aims to select the genes associated with Propionic acid (PPA) and compromise Alzheimer's disease (AD) to find the possible roles of PPA in AD pathogenesis.

**Results** Amongst all genes associated with PPA and AD, 284 genes to be shared by searching databases and were subjected to further analysis. AD-PPA genes mainly involved in cancer, bacterial and virus infection, and neurological and non-neurological diseases. Gene Ontology and pathway analysis covered the most AD hallmark, such as amyloid formation, apoptosis, proliferation, inflammation, and immune system. Network analysis revealed hub and bottleneck genes. MCODE analysis also indicated the seed genes represented in the significant subnetworks. *ICAM1* and *CCND1* were the hub, bottleneck, and seed genes.

**Conclusions** PPA interacted genes implicated in AD act through pathways initiate neuronal cell death. In sum up, AD-PPA shared genes exhibited evidence that supports the idea PPA secreted from bacteria could alter brain physiology toward the emerging AD signs. This idea needs to confirm by more future investigation in animal models.

**Keywords** Alzheimer's disease · Propionic acid · Microbiota · Bioinformatics · System biology

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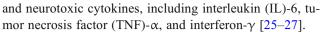
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### Introduction

Alzheimer's disease (AD) is known as the most common neurodegenerative disorder that its primitive hallmarks are the accumulation of beta-amyloid peptide (AB) and neurofibrillary tangles [1]. In the recent decade, another hypothesis explains AD pathogenesis that named the infection hypothesis. This hypothesis explains that microorganisms might have an essential role in AD progression [2]. This hypothesis reconstructed by pioneering work of Itzhaki's group, who showed that plaques contain residues of HSV-1 viral DNA and some experimental data intimate that other viruses, such as CMV, may also be involved in the pathogenesis of AD [3, 4]. The epidemiological studies show that the presence of antiviral antibodies has been correlated with the long-term development of AD [5]. Other groups of investigators suggested a role for spirochetes in the pathogenesis of AD or the presence of Borrelia burgdorferi in the post-mortem brains of many AD patients [6, 7]. Cutibacterium acnes(propionibacterium acnes) was detected in the cortex of patients with AD [8]. Scientists at Bristol University also used 16S ribosomal NGS to assess the bacterial component of the microbiome in post-mortem tissue from the temporal cortex of AD and control. This study suggests that AD brains tend to have large bacterial loads compared to controls. They reported phyla such as Firmicutes and most consistently Actinobacteria, especially Cutibacterium acnes [9]. Cutibacterium acnes is a Gram-positive and anaerobic human skin commensal that involved in the pathogenesis of acne. It was formerly named *Propionibacterium acnes* for its ability to generate propionic acid (PPA) [10, 11]. In addition to microbiota found in post-mortem brain tissue of AD and related them to neurological pathologies [9], we, therefore, hypothesized the microbiota-derived metabolites such as PPA might be a significant risk factor for AD.

PPA is found in the gut, along with other short-chain fatty acids, such as acetate and butyrate, which are major metabolic products of enteric bacteria, following fermentation of dietary carbohydrates and some amino acids [12, 13]. Many bacteria existing in the oral mucosa also produce PPA [14–16]. PPA is taken up by neuroglia and neurons and enters the CNS, where it is thought to comprise a significant energy source in cellular metabolism, particularly during early brain development [17, 18]. Besides, PPA changes several physiological processes such as neurotransmitter synthesis and release, cell signaling, free radical production, mitochondrial function, lipid metabolism, immune function, gap junction gating, intracellular pH maintenance, and modulation of gene expression through phosphorylation and histone acetylation [19, 18, 20–22]. PPA inoculation induced abnormal neural cell organization, which may have led to autism-like neurobehaviors [23, 24]. Studies showed that following PPA administration in rats elevates levels of microglia (CD68 positive)



Tanzi and Moir have produced a compelling piece of evidence demonstrating amyloid β that accumulated in the AD brain is an antimicrobial peptide [28]. These observations were supported by the in vivo testing of the antimicrobial activity of Aß [29]. Antimicrobial activity of Aß in the presence of pathogens or pathogen-derived metabolites like PPA could be a crucial trigger of starting AD pathogenesis. Multiple studies focus on the positive effect of PPA on treated cells in animal models [25–27], or determined its association as an environmental contributor in autism spectrum disorders [23, 24, 30]. Since there is no investigation directly shows the AD pathology and neuronal toxicity of PPA in experimental models, this study used bioinformatic analysis to identify PPA complications link to emerge AD pathophysiology and signs. Herein, we aimed to combine the multi-source PPA-related data in a meaningful manner as retrieved from multiple databases to uncover the mechanism associated with these critical genes interacted with PPA and involved in AD. This study will improve our ability to understand and diagnose the possible microbiota-derived PPA and consequences of AD pathology and it will help to prevent long-term complications.

# Methodology

### Study design and prioritization of AD and PPA genes

To find the possible relationship of PPA secreted by microbiota from variable resources and AD pathogenesis, we selected the genes shared in PPA and AD for further functional and structural analysis. There are different public resources that contain the interactions of genes with chemicals, drugs, and other different agents. These kinds of data emerged from experimental [31, 32] and computational knowledge [32, 33]. Besides, database repositories of high throughput gene expression, including Gene Expression Omnibus (GEO) [34] and ArrayExpress [35] contain data, have been produced from the direct effect of drugs and chemicals on cell lines, animal models, or the result of human clinical trials. In this study, collected genes associated with PPA and AD separately from several of the following resources are explained in data gathering part and selected the shared genes (named AD-PPA shared genes or PPA-AD shared genes) for the next analysis. Used Gene Ontology (GO) and Pathway enrichment tools to find the mechanism related to these genes as the functional analysis of data and applied network analysis as the structural analysis of data to find the crucial genes. An outline of the workflow of this study was summarized in Fig. 1. To prioritize the AD-PPA genes, first focused on the data gathering from gene expression databases and compared significant data with data obtained from public databases [36]. Public databases generally collected data from different molecular databases and,



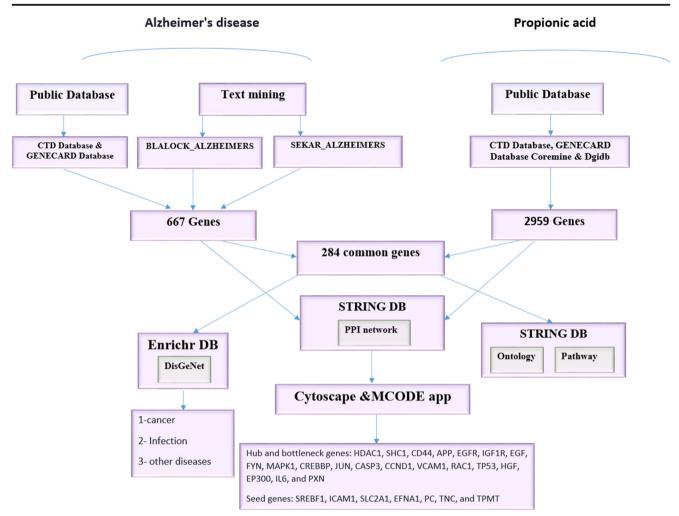


Fig. 1 The workflow of data gathering related to AD and PPA and their functional and structural analysis

based on unique algorithms designed for their analysis; report the output with significant value. In this study, it was almost selected output from different databases with significant value for both AD and PPA keywords. The other prioritization of AD-PPA genes, analysis of AD, and PPA network separately and compare the output with the network of AD-PPA genes. For analysis gene-diseases association, gene ontology, and KEGG pathway, used genes that are shared in AD and PPA.

# Data gathering related to Alzheimer disease and propionic acid

The parent terms "Alzheimer disease" was used to find the genes and proteins. The AD data was collected from public databases (CTD, and GeneCards) and gene expression data from text mining. Comparative Toxicogenomics Database (CTD, http://ctdbase.org) is a public resource that carries information about interactions between gene products and environmental chemicals. Over 15 million toxicogenomic relationships in CTD provide a user-friendly database that helps to find the effect of these interactions on human diseases [36]. In CTD, the input is

a gene list or gene name, drug, chemical agent, and disease term. The output will be chemicals interact with genes, genes interact with the drug and chemical agent, and genes involved in the disease. We searched the keyword "Alzheimer's disease" in the search box related to keyword queries and selected Alzheimer's disease between 2 queries obtained from the input. Then we opened the gene box in Alzheimer's disease page to download the genes associated with AD. We used all data retrieved from this database without considering the inference score. For prioritizing inferences of the different database used in CTD, CTD provides a statistic named "Inference Score" that it reflects the degree of similarity between CTD chemical—gene-disease networks and a similar scale-free random network [36].

GeneCards (https://www.genecards.org/) includes comprehensive useful and annotative information of known and predicted human genes that categorized in different terms including, genetic loci, gene clusters, RNA genes, pseudogenes, and protein coding. These data integrate automatically from over 125 data sources [37]. In this database, also search for genes, biological process, or human disease and output will be all genes linked to the searching



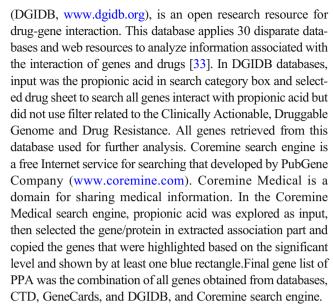
term. To find the gene associated with Alzheimer's disease, search this keyword as input in the search box of the GeneCards database and exported all data related to Alzheimer's disease. For selecting the genes in this database without considering scoring, applied all data as output. GeneCards used GIFtS score linked to the GIFtS algorithm that reflected the degree of a gene's functionality. [37].

Gene expression data were obtained from text mining. We found papers that analyzed the microarray and RNAseq data [38, 39]. Brandon L. Pearson et al. used previously published neuro-degenerative disease data, including these two data named BLALOCK\_ALZHEIMERS and SEKAR\_ALZHEIMERS. An Excel list of brain disorders gene sets and genes was accessible in Supplementary Data 2 [40]. We picked up BLALOCK\_ALZHEIMERS and SEKAR\_ALZHEIMERS data (were statistically significant (FDR < 0.1)) and combined these two AD datasets and introduced them as "text mining data."

Blalock EM et al. have been generated microarray data that used snap-frozen hippocampal specimens of 31 subjects at different levels of ADs that assessed with Braak stages, adjusted Mini-mental Status Exam (MMSE) test and neurofibrillary tangle (NFT) count. The control tissues were matched for age, and neurodegenerative disorders (MMSE > 25, < Braak stage II). The samples were provided by Brain Bank of the Alzheimer's Disease Research Center at the University of Kentucky. To generate gene expression data used, human GeneChips (HG-U133A) and MICROARRAY SUITE 5 (MAS5; 50) [38].

Sekar Sh. et al. in a study with dbGaP accession NO. phs000745.v1.p1 also has been used ten post-mortem brain samples from late-onset AD subjects(Braak stages ranging from IV to VI) and ten healthy elderly control subjects(Braak stages ranging from I to IV). They were collected at the Banner Sun Health Research Institute's Brain and Body Donation Program. Library pools were sequenced by synthesis on the Illumina HiSeq2000 using Illumina's Truseq PE Cluster Kit v3 and Illumina's TruSeq SBS Kits v3 for paired 83 bp read lengths [39].

The final gene list related to AD was obtained from the genes that were shared between CTD and GeneCards databases and text mining. The parent term "propionic acid" was used to find the genes that interact with propionic acid. In GEO or ArrayExpress databases, there was not found high-throughput data that have been directly produced from the effect of PPA on cell line or animal models, so to collect PPA data used the public databases, CTD and GeneCards, DGIDB, and Coremine search engine. In CTD, searched propionic acid in keyword queries and selected propionic acid between 316 results to find the genes associated with propionic acid. Then opened the gene box in the propionic acid page and downloaded all the genes as output. GeneCards also searched the propionic acid in the keyword search box and exported all genes associated with propionic acid as output. Here for CTD and GeneCards, selected all genes without considering a specific threshold for scoring. The Drug Gene Interaction Database



AD genes and genes interacted PPA were downloaded in December 2019.

### **AD-PPA shared genes**

To determine the shared AD-related genes from databases and text mining applied Venn diagram software (http://bioinformatics.psb.ugent.be/webtools/Venn/). To find the shared genes between AD and PPA (shared genes) also used Venn diagram software. All of these shared genes of AD and PPA were selected for further analysis.

# **Gene-disease association analysis**

In order to find the comorbidity disease-related to a gene set, the Enrichr database use several disease databases, OMIM, DSigDB, dbGaP, MSigDB, GeneSigDB, to analyze genes for association with diseases. The input of this database is a gene set, and output will be a table of significant diseases and associated genes [41]. To identify the disease possibly related to the AD-PPA genes used Enrichr database (http://amp. pharm.mssm.edu/Enrichr). That is an open-source database that used different libraries to enrich a gene list [41]. For gene-disease association analysis, selected the DisGeNet database in Enrichr database. We insert the AD-PPA genes as input to the search box, then used the DisGeNet database between disease databases in the disease page of Enrichr and exported the table contain different statistical analyses for the relationship between genes and diseases. We selected the diseases that their P-values < 0.05.

### Gene ontology and KEGG pathway analysis

Functional analysis was performed using the String database (version 11.0) (https://string-db.org). This database



determined the protein-protein interaction and held the enrichment tools to analyze the ontologies and pathways. The Gene Ontology (GO) categories are GO Biological Process, GO Molecular Function, and GO Cellular Component. String database connected to the KEGG database to analyze genes for the enriching pathway. We inserted the AD-PPA genes as input to search box related to multiple proteins and selected homo sapience for the organism. The output contains network and several characteristics information linked to genes in the network. Then analysis box opened and find the different enrichment analyses that we selected the Gene Ontology and KEGG pathway analysis and downloaded. Enrichment analyses in the String database contain only significant terms with P-value < 0.05, and it is not necessary to select significant output manually.

# Protein-protein interaction network construction and topological analysis

Network topology indicates the topological structure of a network [42]. In protein-protein interaction network is a layout of the physical connections between proteins [43]. In the analysis of biological networks used topological structure analysis to identify some aspects such as hubs and bottleneck nodes or modules as the groups of nodes with high topological overlap. A biological network is a scale-free network contains a few highly connected nodes (hubs) which link the rest of the less connected nodes and are sensitive to deletion hubs. Also, hub nodes in the protein-protein interaction network are more likely to be essential for a vital cellular process [43]. All genes related to PPA, AD, and AD-PPA shared genes were fed into the String database to construct the protein-protein interaction networks for each gene set. Then, the protein-protein interaction network (tsv file) was imported to Cytoscape software (version 3.5.1) (http://www.cytoscape.org/) to decipher the crucial protein from the analysis of the network. The Cytoscape is open-source software that constructs and analysis network [44]. In order to introduce the hub and bottleneck genes, it was employed network analysis that assesses the top 10% of degree and betweenness-centrality of every node, respectively. Hub genes are defined as highly connected nodes in the protein-protein interaction network. The network is sensitive to delete the hub genes [45]. Betweenness-centrality is a measure of centrality in a network based on shortest paths. In network analysis, the genes with higher betweennesscentrality are crucial nodes contain valuable information that passes through these nodes, and whole network connectivity depends on these nodes [46]. Hub and bottleneck genes play essential roles in the characteristics and the development of the disease [45]. To find the seed gene used the MCODE app in Cytoscape. MCODE used the topological feature to find highly interconnected regions in a network and introduce numerous clusters and seed genes. The seed genes are interest

genes in identified subnetworks and defined as the highestscoring gene in a gene cluster [47]. The input for analysis in MCODE was selecting every network and then separately run the MCODE. The output was that the sub-networks of each network were significant and contained a score. MCODE parameter was selected to analyze data included, Node Score Cutoff: 0.2, Haircut: true, Fluff: false, K-Core: 2, Max. Depth from Seed: 100. The clusters with the highest score selected for functional analysis (GO and KEGG pathway analysis). Consequently, a protein-protein interaction network is composed of many significant genes (hub and bottleneck genes) and the seed genes allocated to the subnetwork analysis. In this study, constructed networks from AD, PPA, and AD-PPA shared genes, determined the hub and bottleneck nodes in each network, then by MCODE determined the subnetworks and compared the seed genes between all networks. The data selected based on hub-bottleneck-seed genes were listed for functional analysis by GO and pathway analysis.

### Statistical analysis

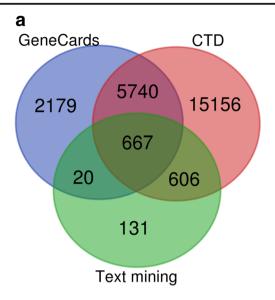
All data with P-values of < 0.05 were considered to be statistically significant.

#### Results

# Genes associated with AD; genes associated with PPA and shared genes between AD and PPA

There is no data that explain the role of microbiota-derived PPA in AD pathophysiology. Here, by using available data related to the AD and PPA, try to find how genes interacted with PPA could affect and change the brain cells to promote AD. In order to meet this aim applied several filters to prioritize genes and mechanisms. First, it was in data gathering that integrated both gene expression data and data obtained from public databases. In order to find the accurate genes related to AD, we selected the expressed genes available in text mining data produced from BLALOCK ALZHEIMERS (1218 genes) and SEKAR ALZHEIMERS (226 genes), and genes available in public databases, CTD (22668 genes), and GeneCards (8605 genes) databases. All shared genes were obtained by Venn Diagram software (Supplementary Data S1 and Fig. 2a). It was determined 667-shared genes in AD. To find the interaction of the genes with PPA, we integrated data available in CTD (17 genes), DGIDB (3 genes), and GeneCards (2719 genes) databases and 513 genes found significantly in the Coremine search engine. It was collected 2959 genes interact with PPA in different databases in which found 284 shared genes with AD (Fig. 2b). These genes named AD-PPA shared genes that contain genes associated with both AD and PPA. All detected genes related to AD and





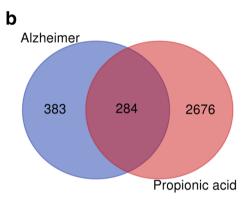
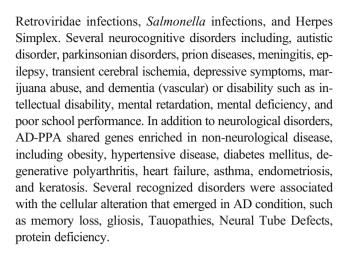


Fig. 2 Venn Diagram of genes associated with AD (a) and the common genes detected in AD and PPA (b)

PPA and AD-PPA shared genes were listed in Supplementary Data S1. The second step in prioritization was the analysis of AD, PPA, and AD-PPA genes separately, particularly network analysis. AD network constructed from 667 genes whereas the PPA network constructed from 2959 genes.

#### **Gene-disease association**

Enrichr database was used for gene-disease association analysis and selected DisGeNet database to determine the possible disease enriched for 284 AD-PPA shared genes. The results of Enrichr analysis within the DisGeNet database indicated that AD-PPA shared genes are related to some diseases that organized in three categories, infection, cancer, and other diseases (represented in Table 1 and Supplementary data S2). The most significant disease enriched was allocated to cancer progression and metastatic process and represented these genes are dysregulated in several cancer types. The next significant category related to infection by several bacteria and viruses such as HIV, Cytomegalovirus, Respiratory syncytial virus, *Helicobacter pylori*, Epstein-Barr virus, Enterovirus infections,



# **Enriched Gene Ontology**

Gene ontology enrichment for AD-PPA shared genes that participated in the biological processes revealed in 1997 terms that linked to the different hallmarks of AD. The biological processes related to AD represented briefly in Table 2 and completed shown in Supplementary Data S2. Hallmark of AD was selected based on Kelly N. H. Nudelman et al. study [48]. They have included AD neuropathology, cell death, proliferative signaling, growth suppressors, angiogenesis, cell adhesion, genomic instability, inflammation, immune function, and cellular energetics. GO molecular function was enriched 219 terms that represented the top 20 in Table 3 and shown Supplementary Data S2. Most significant molecular functions were protein binding (P-value = 2.27E-48), signaling receptor binding (P-value = 2.28E-26) and kinase activity (P-value = 6.97E-15). Out of them, 5 genes (CLU, INSR, LDLR, SCARB1, TGFB2) associated with amyloid-beta (P-value = 0.0114). The significant cellular components terms enriched from AD-PPA shared genes were 177 that listed the top 20 in Table 4. The main cellular components are the cytoplasmic part, extracellular region, cell surface, vesicle, Golgi, ER, mitochondrion, cytoskeleton, or various parts of the neuron (cell body, dendrite, synapse, myelin sheath). Supplementary Data S2 contains all cellular components enrichment related to AD-PPA shared genes.

# **Enriched KEGG Pathway**

The KEGG pathway analysis demonstrated that 175 pathways are linked to AD-PPA genes. Table 5 contains the top 20 pathways and complete result represented in Supplementary Data S2. The most important pathways are included, pathways in cancer, microRNAs in cancer, proteoglycans in cancer, MAPK signaling pathway, PI3K-Akt signaling pathway,



 Table 1
 Top significant of DisGeNet disease related to PPA-AD genes that organized in 3 categories, infections, cancer and other diseases

Term	Adjusted P-value	Genes
Infections		
HIV Infections	1.26E-25	APP;CDKN1A;MCM7;CD81;HFE;TAT;TNC;SLC2A1;GPT;ICAM1;EDNRA;
Infection	9.58E-23	APP;SCARB1;KHDRBS1;CD81;HFE;PDGFB;SLC2A1;LPL;EGFR;ICAM1;PTGS1;
Infection caused by Helicobacter pylori	3.90E-16	HFE;ODC1;TGFA;EGFR;ICAM1;GJA1;KHSRP;CASP3;PIM1;HMOX1;RAC1;JUN;
Herpes Simplex Infections	7.27E-15	HDAC4;APP;ITIH4;CDKN1A;SPARC;TAT;ODC1;LEF1;SLC2A1;NOLC1;COMT;EGFR;
Respiratory syncytial virus (RSV) infection in conditions classified elsewhere and of unspecified site		ABCA1;TGFB1;VCAM1;IL15;IL18;CLU;CXCL2;EGFR;NFKB1;ICAM1;NFKBIA;IL1A;IL6;
Cytomegalovirus Infections	3.10E-12	LRP1;ODC1;EGFR;ICAM1;EDNRA;GJA1;MAPK1;NCAM1;CD34;MICA;SREBF1;JUN;
Bacterial Infections	3.45E-11	ITIH4; CD163; TGFB1; HFE; HGF; IL18; TGFA; PLA2G4A; CFLAR; CXCL2; EGFR; NFKB1; NFKB1A
Epstein-Barr Virus Infections	6.07E-11	TGFB1;VCAM1;IL18;EGFR;NFKB1;DCN;ICAM1;IL1A;IL6;CCND1;BCL6;IFNG;CASP3;
Cancer		
Neoplasm Metastasis	3.89E-77	APP;SPARC;MYLK;ICAM1;AQP1;EDNRA;GJA1;PSMD4;RPS6KA1;DPYSL3;TNFSF10;
Tumor Progression	4.82E-53	SPARC;CSF1;TNC;IRS2;FGF1;ELK1;CLU;ICAM1;IGF1R;EDNRA;GJA1;ZFP36;CCND3;
Other Diseases		
Rheumatoid Arthritis	1.52E-51	APP;TPMT;CSF1;HFE;TNC;FGF1;CLU;ICAM1;IGF1R;C4B;EDNRA;ZFP36;CCND3
Atherosclerosis	3.47E-50	APP;SCARB1;SPARC;CSF1;HFE;TNC;IRS2;FGF1;ELK1;CLU;MYLK;ICAM1;
Obesity	2.11E-48	APP;SCARB1;FAAH;SPARC;CSF1;CD81;HFE;INPPL1;TNC;IRS2;FGF1;
Hypertensive disease	2.16E-46	APP;SPARC;HFE;INPPL1;TNC;IRS2;FGF1;COMT;CLU;MYLK;ICAM1;AQP1;
Diabetes Mellitus	7.58E-41	APP;SCARB1;SPARC;HFE;INPPL1;IRS2;COMT;CLU;ICAM1;AQP1;IGF1R;EDNRA;
Degenerative polyarthritis	1.34E-38	DDR1;APP;CDKN1A;CSF1;THRA;HFE;TNC;SLC2A1;PTPRK;FGF1;CLU;CXCL2;
Heart failure	1.01E-36	DDR1;GSK3B;CDKN1A;CSF1;HFE;TNC;SLC2A1;GPT;ECE1;BRCA1;PRL;CXCL2;
Asthma	5.29E-35	CDKN1A;TNC;CTSS;MYLK;ICAM1;CASP9;AKAP13;EDNRA;ZFP36;TNFSF10;
Endometriosis	1.86E-34	CSF1;THRA;TGFB1I1;TNC;SLC2A1;IRS2;BRCA1;PRL;FGF1;COMT;ICAM1;
Autistic Disorder	4.84E-11	APP;FAAH;TNC;PRL;COMT;AKAP1;C4B;GJA1;ERBB4;EP300;MAPK1;DMD;
Parkinsonian Disorders	5.94E-10	PDGFRB;APP;NQO1;TBP;PDGFB;PRL;PLA2G6;COMT;SOD2;CP;GFAP;TPO;
Neurocognitive Disorders	1.63E-06	ITIH4;ZFP36;IFNG;TAT;F2R;HMOX1;TP53;MAP3K11
Intellectual Disability	1.87E-06	APP;THRA;TAT;ARHGAP1;INPPL1;SLC2A1;ELK1;IGF1R;EDNRA;GJA1;SPTLC2;
Mental Retardation	1.95E-06	HDAC4;APP;THRA;TAT;ARHGAP1;SLC2A1;PLA2G6;ASCL1;IGF1R;PURA;GJA1;TPO;
Depressive Symptoms	1.95E-06	TGFB1;TAT;BRCA1;COMT;CYP3A4;EGFR;IL6;FOLH1;DAO;MAPK1;NCAM1;DRD2;CD34
Prion Diseases	1.97E-06	ABCA1;APP;IL1A;TGFB2;IL6;GSN;CYBB;SOD2;CLU;NFKB1
Marijuana Abuse	2.14E-06	ABCA1;SCARB1;FAAH;EDNRA;NCAM1;COMT;DRD2;TP53;EGFR
Memory Loss	2.52E-06	PDGFRB;ABCA1;APP;IL6;PAH;HSF1;PDGFB;BCL2;COMT;DRD2;EGFR;CTSB
Meningitis	6.05E-06	C4B;IL6;TGFB1;IFNG;TNFSF10;CYBB;FLNA;RAC1;JAK3;ICAM1
Dementia, Vascular	1.65E-05	APP;IL1A;IL6;TGFB1;EGF;MMP2;TP53;SREBF2;FGFR1
Epilepsy, Temporal Lobe	1.73E-05	SLC2A1;NFKB1;ICAM1;PLD2;TNFRSF1A;IL1A;GJA1;CASP3;TNFSF10;BCL2;MBP;FYN;TP53
Transient Cerebral Ischemia	1.75E-05	CASP9;MMP12;GSK3B;BAD;GPX3;SOD2;TP53
Autism Spectrum Disorders	6.17E-05	DDR1;FAAH;EGF;HFE;HGF;IL18;CASK;IRS2;PRL;SOD2;GFAP;IL1A;EDNRA;MMP16;IFNG;
Keratosis	9.53E-05	CAST;MMP12;CCND1;BCL2;FGFR3;TP53
Gliosis		TBP;HGF;ITGA1;MAPK1;PLA2G6;EGFR;LMNB1;GFAP
Poor school performance		HDAC4;APP;TAT;SLC2A1;PLA2G6;ASCL1;IGF1R;PURA;GJA1;TPO;FLNA;EP300;MAPK1; DMD;GGT1;
Tauopathies	5.76E-04	APP;KHDRBS1;GSK3B;IL1A;TPO;IL6;HMOX1;MAPK1;TP53
Neural Tube Defects		C5;FOLH1;TCN2;IFNG;SNAI2;NCAM1;COMT;TP53;LMNB1;LRP6
Basal cell carcinoma		LATS1;IL6;TGFB1;CFLAR;TP53;EGFR;IGF1R;MCL1
Mental deficiency		HDAC4;APP;TAT;SLC2A1;PLA2G6;ASCL1;IGF1R;PURA;GJA1;TPO;FLNA;EP300;MAPK1;
Protein Deficiency		C4B;APP;SCARB1;IFNG;SLC2A1;TP53;LDLR



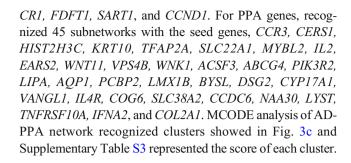
Ras signaling pathway, HIF-1 signaling pathway, focal adhesion, EGFR tyrosine kinase inhibitor resistance, Jak-STAT signaling pathway.

# Protein-protein interaction networks construction and analysis

The Cytoscape software was used for network analysis. We constructed three protein-protein interaction networks, AD, PPA, and AD-PPA, then analyzed every network separately and finally compared the crucial genes related to each network. The primary protein-protein interaction network obtained from the String database was imported into Cytoscape. Constructed AD network contains 591 nodes and 5076 edges, while the PPA network contains 2768 nodes and 104202 edges. The network analysis determined hub and bottleneck genes for AD and PPA network that represented in Supplementary Table S1 and S2 respectively. We used the lists obtained from the hub, and bottleneck nodes of AD and PPA network to prioritization the genes belong to AD and PPA and then compare the list will obtain from the network of AD-PPA shared genes. The constructed network from AD-PPA shared genes (284 genes) contains 280 nodes, which are connected to 2859 edges. The overview figure of integrating 3 networks was represented in Supplementary Figure S1. All three networks were a scale-free network that included a few numbers of hub genes with the most number of interactions and many numbers of nodes with a few interactions. Figure 3a represented the protein-protein interaction network of AD-PPA genes in which hub and bottleneck nodes have a more prominent size with different colors and located in the center, and Fig. 3b represented the node degree distribution. Besides, the analysis of the AD-PPA network revealed the several parameters that are containing; the clustering coefficient is 0.402, network density is 0.073, the network diameter is 5, and the connected component is 1, the shortest paths are 78120. The network hub and bottleneck nodes were also listed in Table 6. Out of them, HDAC1, SHC1, CD44, APP, EGFR, IGF1R, EGF, FYN, MAPK1, CREBBP, JUN, CASP3, CCND1, VCAM1, RAC1, TP53, HGF, EP300, IL6, and PXN are the genes shared between hub and bottleneck and introduce as relevant genes in the protein-protein interaction network.

### Construction and analysis of subnetworks

The subnetworks were studied for protein-protein interaction networks, AD, PPA, and AD-PPA through MCODE app in Cytoscape to identify the seed genes. AD genes could be clustered in 14 subnetworks with seed genes, including ARL17B, ITGA6, HDAC4, CYP3A4, RUNX2, TAT, TCF3,



# Identification and analysis of hub, bottleneck and seed nodes in AD-PPA shared genes

As depicted in Supplementary Figure S2, 80 genes of 284 AD-PPA shared genes at least had overlap with hub or bottleneck genes in one of two AD and the PPA networks. Between these overlap genes, 51 genes were shared between 3 networks. When comparing the hub or bottleneck genes in 3 networks (represented in Supplementary Figure S3), 32 genes were shared between them.

Amongst 280 AD-PPA shared genes, only seven genes, i.e., SREBF1, ICAM1, SLC2A1, EFNA1, PC, TNC, and TPMT, were identified as seed genes. When compared, the seed genes of the three networks, found no shared genes between them. As represented in the Venn diagram of Supplementary Figure S4, when compared the seed genes of AD and PPA with the all 284 shared genes of AD-PPA, found two shared genes with PPA and seven shared genes with AD. This data represented two genes; TFAP2A and AQP1 in the PPA network are crucial genes that could not distinguish as the seed genes in topological analysis of the AD-PPA subnetworks. TFAP2A was not the hub and bottleneck in the PPA network. AQP1 also was not hub and bottleneck in the PPA network but seen in the AD network. Seven genes, HDAC4, TAT, ITGA6, CCND1, CYP3A4, CR1, and RUNX2, are seed genes of AD network that could not be detected as seed genes in AD-PPA subnetwork. CCND1 seen in all three networks AD, PPA, and AD-PPA, belongs to the hub and bottleneck nodes. CCND1 was also observed in most of GO categories or KEGG pathways. CYP3A4, also seen in three networks, had high betweenness centrality. CYP3A4 was also seen in several GO categories and diseases. RUNX2 detected as hub and bottleneck nodes in the PPA network and recognized as a hub in the AD network. This gene was not a hub and bottleneck nodes in the AD-PPA network. HDAC4, ITGA6, CR1 seen in three networks but not the hub and bottleneck nodes. TAT was seen in AD but was not a hub and bottleneck nodes.

*ICAM1* is a seed gene of AD-PPA subnetwork that belongs to the hub and bottleneck genes. This gene also allocated to several biological processes and diseases. *SLC2A1* is a seed gene of AD-PPA subnetwork that also detected as a bottleneck gene. *SLC2A1* is a part of genes attributed to biological processes, including cellular response to glucose starvation



**Table 2** Biological process related to common genes in PPA-AD potentially involved by Ontology with default parameters. Observed gene count indicate the number of genes from our list and background gene

count indicate the number of genes allocated to the specific GO term in its library. The head title obtained from the important hallmarks of cancer that observed in AD

Gene Onthology ID	Description	Observed gene count	Background gene count	False discovery rate
AD Neuropathology		1	·	
GO:1904645	response to amyloid-beta	5	29	0.00063
GO:1900221	regulation of amyloid-beta clearance	4	10	0.00023
GO:1902003	regulation of amyloid-beta formation	4	18	0.0012
GO:1902947	regulation of tau-protein kinase activity	3	10	0.0033
Cell Death				
GO:0043523	regulation of neuron apoptotic process	23	195	2.08E-12
GO:0043067	regulation of programmed cell death	100	1516	4.65E-36
GO:1903201	regulation of oxidative stress-induced cell death	7	65	0.00044
GO:1901030	positive regulation of mitochondrial outer membrane permeabilization involved in apoptotic signaling pathway	4	34	0.0078
GO:2001238	positive regulation of extrinsic apoptotic signaling pathway	4	51	0.0238
Proliferative Signaling				
GO:0048666	neuron development	38	758	1.17E-09
GO:0031175	neuron projection development	31	616	5.69E-08
GO:0048699	generation of neurons	64	1422	2.33E-14
GO:0030182	neuron differentiation	45	940	8.53E-11
GO:0042127	regulation of cell population proliferation	115	1594	5.60E-46
GO:2000177	regulation of neural precursor cell proliferation	6	79	0.0056
GO:0007265	Ras protein signal transduction	12	155	4.03E-05
GO:0043406	positive regulation of MAP kinase activity	27	264	3.48E-13
Growth Suppressors				
GO:0045787	positive regulation of cell cycle	31	376	7.98E-13
GO:0007346	regulation of mitotic cell cycle	31	608	4.28E-08
GO:0045786	negative regulation of cell cycle	19	517	0.0014
GO:0007050	cell cycle arrest	9	149	0.0022
GO:0090400	stress-induced premature senescence	2	8	0.0253
GO:0090399	replicative senescence	2	13	0.0482
Angiogenesis				
GO:0001568	blood vessel development	29	464	2.07E-09
GO:0008015	blood circulation	22	373	7.13E-07
GO:0003073	regulation of systemic arterial blood pressure	7	92	0.0025
GO:0007596	blood coagulation	11	288	0.0136
GO:1903589	positive regulation of blood vessel endothelial cell proliferation involved in sprouting angiogenesis	2	9	0.0296
GO:0090049	regulation of cell migration involved in sprouting angiogenesis	4	37	0.01
GO:0001525	angiogenesis	17	297	2.46E-05
GO:0001666	response to hypoxia	26	288	1.36E-11
Cell adhesion				
GO:0022409	positive regulation of cell-cell adhesion	22	238	4.63E-10
GO:0007155	cell adhesion	39	843	5.40E-09
GO:0001952	regulation of cell-matrix adhesion	9	105	0.00024
GO:0048041	focal adhesion assembly	3	24	0.0206
GO:0001764	neuron migration	7	118	0.0081
Genomic Instability				
GO:0006974	cellular response to DNA damage stimulus	22	749	0.0067



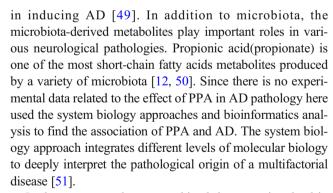
Table 2 (continued)

Gene Onthology ID	Description	Observed gene count	Background gene count	False discovery rate
GO:0006978	DNA damage response, signal transduction by p53 class mediator resulting in transcription of p21 class mediator	3	17	0.0101
GO:0006268	DNA unwinding involved in DNA replication	2	8	0.0253
GO:0006975	DNA damage induced protein phosphorylation	2	8	0.0253
GO:0032392	DNA geometric change	4	63	0.0408
GO:0000077	DNA damage checkpoint	6	138	0.0451
GO:2001252	positive regulation of chromosome organization	9	160	0.0033
Inflammation				
GO:0006954	inflammatory response	41	482	2.20E-17
GO:0002526	acute inflammatory response	8	73	0.00014
GO:0002248	connective tissue replacement involved in inflammatory response wound healing	3	5	0.00082
GO:0006925	inflammatory cell apoptotic process	2	5	0.0138
GO:0002544	chronic inflammatory response	2	13	0.0482
GO:0001774	microglial cell activation	3	20	0.0138
GO:0034097	response to cytokine	77	1035	3.86E-30
GO:0019221	cytokine-mediated signaling pathway	55	655	2.51E-23
Immune Function				
GO:0045824	negative regulation of innate immune response	5	49	0.0044
GO:0002376	immune system process	105	2370	1.19E-24
GO:0002250	adaptive immune response	24	280	2.53E-10
GO:0006959	humoral immune response	14	252	0.0002
Cellular Energetics				
GO:0043467	regulation of generation of precursor metabolites and energy	7	96	0.0031
GO:0042593	glucose homeostasis	7	169	0.0367
GO:0005979	regulation of glycogen biosynthetic process	5	29	0.00063

and response to peptide. Moreover, *SLC2A1* was recognized in most of the diseases enriched by DisGeNet analysis but not the virus and bacterial infection. When assessed functionally 40 hubs, bottlenecks, and seed genes of AD-PPA network in the String database, these genes were found that enriched all biological process, molecular function, cellular component, and pathways obtained from all genes. The most significant GO and Pathways listed in Table 7. For two top ranked AD-PPA subnetworks obtained from MCODE (score 16.146 and score 13.125 contain 59 genes) included 29 genes had overlap with 40 hub-bottleneck-seed genes. In addition, the functional analysis in Enrichr for GO and KEGG pathway enrichments found a similar result to the analysis of 40 hub-bottlenecks-seed genes.

# **Discussion**

The infection hypothesis is an interesting theory in Alzheimer's disease onset and explains the role of microbiota



In the present study, we combined the genes involved in PPA available in CTD, DISEASES, and Gene Cards databases and the Coremine search engine by Venn diagram software. As a result, according to the Fig. 1, 284 PPA genes were found to be shared with AD genes (extracted from text mining, CTD, and GeneCards databases). We used network and enrichment analysis to uncover critical molecular mechanisms and relationships between PPA and AD. DisGeNet enriched diseases that arranged in categories related to infection, cancer, and neurological and non-neurological diseases. As



Table 3 Top 20 of Molecular function obtain from enrichment of PPA- AD common genes. Observed gene count indicate the number of genes from our list and background gene count indicate the number of genes allocated to the specific GO term in its library

Gene Onthology ID	Description	Observed gene count	Background gene count	False discovery rate
GO:0005515	protein binding	221	6605	2.27E-48
GO:0005488	binding	266	11878	1.97E-34
GO:0005102	signaling receptor binding	87	1513	2.28E-26
GO:0019899	enzyme binding	98	2197	2.95E-22
GO:0004672	protein kinase activity	46	635	2.48E-16
GO:0016773	phosphotransferase activity, alcohol group as acceptor	50	767	3.44E-16
GO:0044877	protein-containing complex binding	54	968	6.79E-15
GO:0016301	kinase activity	50	835	6.97E-15
GO:0019900	kinase binding	44	678	3.82E-14
GO:0042802	identical protein binding	72	1754	8.55E-14
GO:0019901	protein kinase binding	40	599	3.29E-13
GO:0098772	molecular function regulator	69	1793	6.63E-12
GO:0008134	transcription factor binding	38	610	1.05E-11
GO:0004713	protein tyrosine kinase activity	22	180	1.08E-11
GO:0003824	catalytic activity	140	5592	1.43E-11
GO:0043167	ion binding	145	6066	1.44E-10
GO:0043168	anion binding	85	2696	1.44E-10
GO:0140096	catalytic activity, acting on a protein	74	2176	1.77E-10
GO:0008144	drug binding	63	1710	4.28E-10
GO:0097367	carbohydrate derivative binding	72	2163	9.22E-10

Table 4 Top 20 of cellular component enrichment detected from common AD-PPA genes. Observed gene count indicate the number of genes from our list and background gene count indicate the number of genes allocated to the specific GO term in its library

Gene Onthology ID	Description	Observed gene count	Background gene count	False discovery rate
GO:0044444	cytoplasmic part	214	9377	4.25E-18
GO:0005576	extracellular region	97	2505	1.46E-17
GO:0005737	cytoplasm	235	11238	2.71E-17
GO:0009986	cell surface	49	690	2.71E-17
GO:0005615	extracellular space	62	1134	4.48E-17
GO:0044421	extracellular region part	68	1375	7.88E-17
GO:0031982	vesicle	90	2318	1.44E-16
GO:0044459	plasma membrane part	95	2651	1.71E-15
GO:0005829	cytosol	138	4958	3.91E-15
GO:0031410	cytoplasmic vesicle	83	2226	3.69E-14
GO:0071944	cell periphery	139	5254	1.63E-13
GO:0005886	plasma membrane	137	5159	2.17E-13
GO:0044464	cell part	277	16244	3.10E-13
GO:0012505	endomembrane system	120	4347	2.48E-12
GO:0098805	whole membrane	63	1554	5.50E-12
GO:0045121	membrane raft	27	300	8.19E-12
GO:0044422	organelle part	191	9111	6.90E-11
GO:0043227	membrane-bounded organelle	219	11244	1.20E-10
GO:0044446	intracellular organelle part	187	8882	1.20E-10
GO:0005622	intracellular	254	14286	2.61E-10



Table 5 Top 20 of KEGG pathways of common AD-PPA genes. Observed gene count indicate the number of genes from our list and background gene count indicate the number of genes allocated to the specific KEGG pathway term in its library

KEGG- ID	Description	Observed gene count	Background gene count	False discovery rate
hsa05200	Pathways in cancer	63	515	5.19E-35
hsa04010	MAPK signaling pathway	43	293	3.02E-26
hsa05206	MicroRNAs in cancer	32	149	8.06E-24
hsa04151	PI3K-Akt signaling pathway	41	348	6.08E-22
hsa04014	Ras signaling pathway	32	228	4.91E-19
hsa05167	Kaposi's sarcoma-associated herpesvirus infection	28	183	1.58E-17
hsa05215	Prostate cancer	22	97	6.66E-17
hsa04066	HIF-1 signaling pathway	21	98	9.72E-16
hsa05169	Epstein-Barr virus infection	26	194	3.77E-15
hsa05205	Proteoglycans in cancer	26	195	3.80E-15
hsa04380	Osteoclast differentiation	22	124	4.05E-15
hsa04510	Focal adhesion	26	197	4.05E-15
hsa05210	Colorectal cancer	19	85	1.03E-14
hsa05166	HTLV-I infection	28	250	1.04E-14
hsa01521	EGFR tyrosine kinase inhibitor resistance	18	78	3.30E-14
hsa04630	Jak-STAT signaling pathway	23	160	3.34E-14
hsa05211	Renal cell carcinoma	17	68	5.75E-14
hsa04933	AGE-RAGE signaling pathway in diabetic complications	19	98	7.24E-14
hsa04722	Neurotrophin signaling pathway	20	116	9.34E-14
hsa04015	Rap1 signaling pathway	24	203	3.29E-13

depicted in Table 1, several bacteria and virus infections were disclosed. Out of them, Salmonella typhimurium able to generate PPA during fermentation [52]. Prior microbiome researches have also shown oral and genital herpes, Epstein Barr virus, cytomegalovirus, HIV, gut bacteria, liver bacteria, Helicobacter pylori, periodontal pathogens, and Chlamydophila pneumonia present in AD pathogenesis [49, 53, 54]. These pathogens may cross the blood-brain barrier or brain-CSF barrier and attack the CNS [55, 56]. Also, it was determined that microbiotas influence CNS by microbiotaderived metabolites and inflammation [49, 55, 57-60]. Since we used the AD-PPA genes to detect the diseases, this data suggests that PPA can induce the same effect provided by microbiota. The most significant diseases were related to cancer progression and metastatic processes that also validated by the KEGG pathway analysis and previously were mentioned as familiar hallmarks of cancer and AD [48]. In the analysis of biological process also enriched cancer as the first gene ontology. Our results are supporting this evidence that PPA could able to promote brain tissue toward AD pathology like seen in cancer. AD-PPA genes also identified disease-related Neurocognitive Disorders and several disabilities. It has been reported excessive PPA has implied disadvantageous effects such as propionic acidemia, a neurodevelopmental metabolic disorder, that identified by elevation of PPA levels in the blood, cerebrospinal fluid, and neurons [61, 62]. For instance,

several studies showed that intraventricular inoculation of PPA created behavioral and brain abnormalities in rats similar to autism spectrum disorder [53–56].

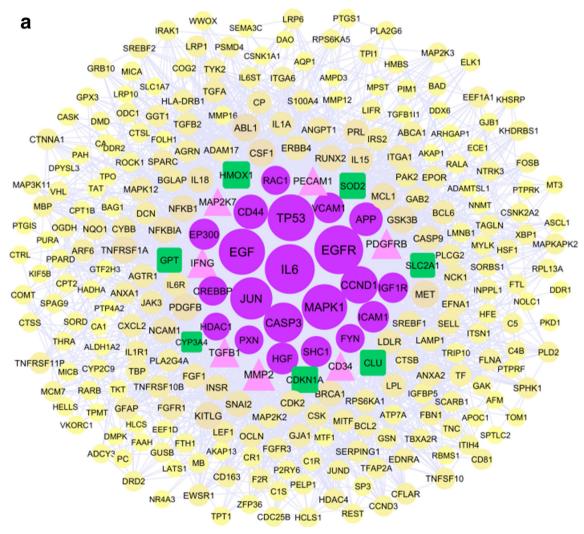
Dysregulation in the cell cycle was another significant KEGG pathway that previously has been contributed to surviving neuron cells and accumulating the amyloid fibril, which eventually undergoes apoptosis [63]. Dysregulation in proliferative signaling and evade growth suppressor have been identified as an important hallmark of AD [64]. The shared genes of AD-PPA enriched the biological processes related to neuron generation, development, projection, and differentiation. Besides, they contain genes that describe the positive and negative regulation of proliferation. MAP kinase pathway that enriched in our data is the primary signaling activity in neuron and glial cells that promote phosphorylation of tau deposit [65]. Also, another proliferative signaling is RAS that the pathway-related this GTPase also enriched and previously suggested in tau phosphorylation [66]. It has been determined that neurodegeneration is associated with improper cell cycle progression that increases neuropathological processes and finally leads to apoptosis [67]. Out of genes, 19 genes found the negative cell cycle regulators while there existed 31 genes with function in positive regulation of cell cycle that most of them were shared in apoptosis and proliferative process. In AD patients, Aβ42 causes hyperactivation of MEK-ERK signaling lead to cell death by possibly mediating Tau



hyperphosphorylation. In addition, the A\u00e342-mediated aberrant MEK-ERK signaling pathway may promote S-phase cell cycle reentry through inducing expression of cyclin D1(CCND1) and neuronal cell death [68, 69]. CCND1 as a crucial gene in cell cycle reentry of postmitotic neurons was an important hub and bottleneck gene in three networks AD, PPA, and AD-PPA. Other impressive gene ontology enrichment results had a direct link to the amyloid formation and tau pathology. This finding was also confirmed by gene-disease association analysis. Tauopathies, Neural Tube Defects, Protein Deficiency were recognized in DisGeNet analysis. These results confirm the role of PPA in AD as a risk factor could able to enhance the process, leading to cell cycle deregulation and contribute to neuronal loss and neurodegeneration observed in the AD brains. This finding was also supported by the enrichment output of AD-PPA genes that associated with

apoptosis activated by the p53 signaling pathway or cell death through both intrinsic and extrinsic apoptosis signaling pathways. Apoptosis activated by the p53 signaling pathway has implicated by the treatment of neuronal cells by PPA *in vitro* [70]. Lobzhanidze G et al. reported that a low level of PPA could change amygdala cells toward apoptosis [30]. Previously indicated enhanced Caspase-3 mRNA expression and inhibited Bcl-2 mRNA expression in the brain of rat pups that had been exposed to PPA [71].

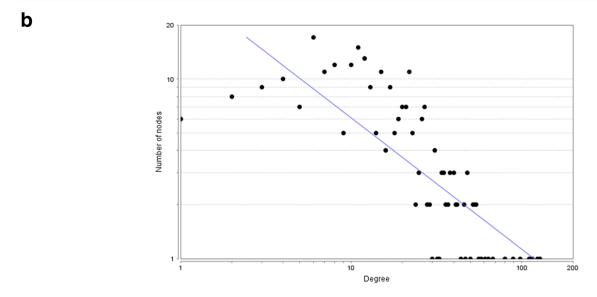
Another mediator of brain injury in AD is Oxidative stress [72] that regulation of oxidative stress-induced cell death was the significant biological processes detected by AD-PPA genes. Increased reactive oxygen species (ROS) production may lead to apoptosis [73]. Alteration of PPA treatment in the enzymatic antioxidant capacity in rat brains has been represented a significant decrease in superoxide dismutase (SOD)



**Fig. 3** The protein-protein interaction networks of 284 genes are common in AD and PPA (a) that constructed by Cytoscape software. The size of nodes was determined by their degree. Hub and bottleneck genes are purple ellipse nodes. The genes that are only hub are pink triangle nodes

while the genes that are only bottleneck represented as green rectangle nodes. Degree distribution related nodes in network (b) revealed the AD-PPA network is a scale free network. MCODE analysis detected 12 clusters that seed detected as diamond nodes with yellow color(c)





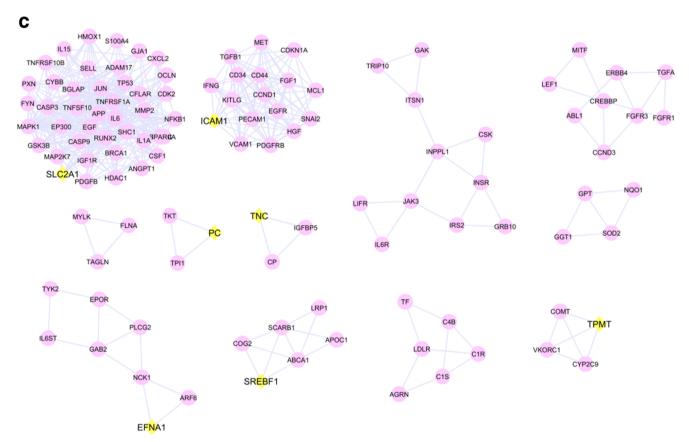


Fig. 3 (continued)

and catalase(CAT) activities [71, 74]. According to Table 6, *SOD2* is the bottleneck genes distinguished in network analysis, play role as a critical antioxidant. Accumulating evidence disclosed that ROS produced by various enzymatic reactions and chemical processes had implicated the pathogenesis of neurodegenerative disorders such as AD [75]. ROS involved

in the expression of well-defined inflammatory mediators, including MMPs, cPLA2, COX-2, and adhesion molecules. We recognized several pro-inflammatory genes and the protein response to this event, including *APP*, *CASP3*, *EGF*, *EGFR*, *EP300*, *HMOX1*, *ICAM1*, *IL6*, *JUN*, *MMP2*, *PDGFRB*, and *VCAM1* that were belonged to the hub- bottleneck- seed



Table 6 Hub genes and bottleneck related the common genes of AD and PPA network obtained from Cytoscape software

Gene name	Degree (Hub)	Gene name	Betweenness centrality (Bottleneck)
IL6	128	EGFR	0.097698
EGFR	124	IL6	0.093625
TP53	116	TP53	0.075256
EGF	114	MAPK1	0.069349
MAPK1	111	EGF	0.063399
JUN	98	APP	0.038658
CASP3	89	JUN	0.038026
CCND1	80	CASP3	0.036808
CD44	68	CCND1	0.029613
APP	64	RAC1	0.02678
MMP2	61	EP300	0.022104
HGF	58	GPT	0.022064
ICAM1	56	SOD2	0.021618
EP300	54	SLC2A1	0.019361
SHC1	54	PXN	0.018438
VCAM1	53	HGF	0.017373
PDGFRB	53	CYP3A4	0.016507
IGF1R	52	CD44	0.015733
IFNG	52	CREBBP	0.015148
TGFB1	50	SHC1	0.014951
CREBBP	48	HDAC1	0.014795
CD34	48	IGF1R	0.012673
PECAM1	48	HMOX1	0.01255
RAC1	47	ICAM1	0.012427
PXN	46	CDKN1A	0.012121
FYN	46	CLU	0.010988
MAP2K7	44	FYN	0.010582
HDAC1	42	VCAM1	0.010404

genes. The previous research on natural products such as resveratrol, curcumin, berberine has been shown that they could able to elicit anti-cancer and antiaging by intracellular signaling mechanisms. Many of the beneficial effects have been attributed to their anti-inflammatory properties [76]. Between the genes underlying these effects, we detected APP, CASP3, GPT, HMOX1, ICAM1, IL6, JUN, MAPK1, TGFB1, TP53, and VCAM1 that are significant genes of our gene list. As mentioned, they belong to inflammatory genes that also targeted in antiaging investigations. Within this list, ICAM1 was the only gene distinguished as a hub-bottleneck-seed node. Several studies have been emphasized the importance of ICAM1 gene polymorphism and its expression levels in AD pathogenesis [77–79].

Animal studies showing that oxidative damage to proteins may be involved in the pathophysiology of PPA [80]. The alterations of protein structure by oxidants may affect the function of receptors, enzymes, and transport proteins, resulting in a partial or complete loss of protein functionality [81]. The most crucial molecular function enrichments link to

protein binding and kinase activity that this oxidation could change the protein interaction, and unfavorable results would contain alteration seen in the protein-protein interaction network. Previously determined, the increase of phosphorylation promotes pathogenesis-related TAU protein [1]. In AD-PPA genes enriched protein binding as the most significant molecular function. It has been previously indicated the aberrant protein binding in AD [63]. In addition to protein oxidation, PPA animates lipid peroxidation in the rat brain and the plasma of patients with propionic academia that are finally resulting in cell damage [82, 83]. PPA inhibits the antioxidant enzyme activities and induces Malondialdehyde, which may have happened in mitochondrial dysfunction [84, 85]. Mitochondrial hypometabolism characterizes brain aging and AD. Redox dysregulation and chronic neuroinflammation are observed in brain aging and AD that linked to energy metabolism and inflammatory responses. The metabolicinflammatory axis describes the dynamic interaction of these systems in the brain [86]. Changing in brain metabolism and cellular energy are the hallmark of AD that increased levels of



**Table 7** Functional analysis of 40 genes including hub, bottlenecked and seed genes in String database. Observed gene count indicate the number of genes from our list and background gene count indicate the number of genes allocated to the specific GO and KEGG pathway term in its library

		Permit and the month			
#term ID	Description	Observed gene count	Background gene count	False discovery rate	Proteins
Biological process GO:0007166 c	ess cell surface receptor signaling pathway	29	2198	6.92E-16	APP,CASP3,CCND1,CD44,CDKN1A,CREBBP,EFNA1,EGF,EGFR,EP300, FYN,HGF,HMOX1,ICAM1,IFNG,IGF1R,IL6,JUN,MAPK1,MMP2, PDGFRB,PXN,RAC1,SHC1,SOD2,SREBF1,TGFB1,TP53,VCAM1
GO:0001934	positive regulation of protein phosphorylation	21	941	2.17E-14	APP,CCND1,CD44,CDKN1A,CLU,EFNA1,EGF,EGFR,FYN,HGF,ICAM1, IFNG,IL6,IUN,MAP2K7,MAPK1,PDGFRB,RAC1,SHC1,TGFB1,TP53
GO:2000377	regulation of reactive oxygen species metabolic process	13	169	2.46E-14	APP,CD34,CDKN1A,CLU,EGFR,FYN,ICAM1,IFNG,PDGFRB,RAC1,SHC1, TGFB1,TP53
GO:0006950	response to stress	31	3267	2.63E-14	APP,CASP3,CCND1,CD34,CD44,CDKN1A,CLU,CREBBP,EGFR,EP300, FYN,HDAC1,HMOX1,ICAM1,IFNG,IL6,JUN,MAP2K7,MAPK1,MMP2, PDGFRB,PXN,RAC1,SHC1,SLC2A1,SOD2,SREBF1,TGFB1,TNC,TP53, VCAM1
GO:0031401	positive regulation of protein modification process	22	1149	2.63E-14	APP,CCND1,CD44,CDKN1A,CLU,EFNA1,EGF,EGFR,FYN,HGF,ICAM1, IFNG,IL6,JUN,MAP2K7,MAPK1,PDGFRB,RAC1,SHC1,SREBF1,TGFB1, TP53
GO:0070887	cellular response to chemical stimulus	29	2672	2.63E-14	APP,CASP3,CCND1,CD44,CDKN1A,CREBBP,CYP3A4,EGFR,EP300,FYN, HGF,HMOX1,ICAM1,IFNG,IGF1R,IL6,JUN,MAPK1,MMP2,PDGFRB, PXN,RAC1,SHC1,SOD2,SREBF1,TGFB1,TNC,TP53,VCAM1
GO:0010033	response to organic substance	29	2815	4.54E-14	APP,CASP3,CCND1,CD44,CDKN1A,CLU,EGFR,EP300,FYN,HGF,HMOX1, ICAM1,IFNG,IGF1R,IL6,JUN,MAP2K7,MAPK1,MMP2,PDGFRB,PXN, SHC1,SLC2A1,SOD2,SREBF1,TGFB1,TNC,TP53,VCAM1
GO:0042221	response to chemical	33	4153	5.15E-14	APP,CASP3,CCND1,CD44,CDKN1A,CLU,CREBBP,CYP3A4,EFNA1, EGFR,EP300,FYN,HGF,HMOX1,ICAM1,IFNG,IGF1R,IL6,JUN,MAP2K7, MAPK1,MMP2,PDGFRB,PXN,RAC1,SHC1,SLC2A1,SOD2,SREBF1, TGFB1,TNC,TP53,VCAM1
GO:0010941	regulation of cell death	24	1638	5.19E-14	APP,CASP3,CD34,CD44,CDKN1A,CLU,CREBBP,EFNA1,EGFR,FYN, HDAC1,HGF,HMOX1,ICAM1,IFNG,IGF1R,IL6,JUN,MAP2K7, PDGFRB,SHC1,SOD2,TGFB1,TP53
Molecular function	ion				
GO:0005515	protein binding	36	9099	8.21E-11	APP, CASP3, CCND1, CD34, CDKN1A, CLU, CREBBP, CYP3A4, EFNA1, EGF, EGR, EP300, FYN, HDAC1, HGF, HMOX1, ICAM1, IFNG, IGF1R, IL6, JUN, MAP2K7, MAPK1, PC, PDGFRB, PECAM1, PXN, RAC1, SHC1, SLC2A1, SOD2, SREBF1, TGFB1, TNC, TP53, VCAM1
GO:0019899	enzyme binding	23	2197	4.01E-10	APP,CASP3,CCND1,CDKN1A,CLU,CYP3A4,EGF,EGFR,FYN,HDAC1,HGF, HMOX1,JUN.MAP2K7,MAPK1,PDGFRB,PXN,RAC1,SHC1,SLC2A1, SREBF1,TGFB1,TP53
GO:0005102	signaling receptor binding	19	1513	2.91E-09	APP,CASP3,CLU,EFNA1,EGF,EGFR,EP300,FYN,HGF,ICAM1,IFNG,IGF1R, IL6,PDGFRB,PXN,SHC1,TGFB1,TP53,VCAM1

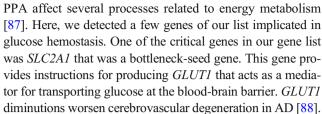


Table 7 (continued)

#term ID	Description	Observed gene count	Background gene count	False discovery rate	Proteins
GO:0019901	protein kinase binding	11	599	1.39E-06	CCND1,CDKN1A,EGFR,MAP2K7,MAPK1,PDGFRB,PXN,RAC1,SHC1, SREBF1,TP53
GO:0004672	protein kinase activity	11	635	2.19E-06	CCND1,CDKN1A,EGF,EGFR,FYN,HGF,IGF1R,MAP2K7,MAPK1, PDGFRB,RAC1
GO:0070851	growth factor receptor binding	9	131	1.19E-05	APP, EGF, FYN, IL6, PDGFRB, SHC1
GO:0005178	integrin binding	5	122	0.00011	APP,EGFR,ICAM1,PXN,VCAM1
GO:0005088	Ras guanyl-nucleotide exchange factor activity	9	243	0.00019	EGF, EGFR, FYN, HGF, PDGFRB, SHC1
GO:0033613	activating transcription factor binding	4	69	0.00025	CREBBP, EP300, HDAC1, JUN
GO:0002039	p53 binding	4	73	0.0003	CREBBP, EP300, HDAC1, TP53
Cellular component	nent				
GO:0045121	membrane raft	6	300	2.98E-06	APP, CASP3, EGFR, FYN, HMOX1, ICAM1, MAPK1, PECAM1, SLC2A1
GO:0044421	extracellular region part	15	1375	3.09E-06	APP,CD34,CLU,EGF,EGFR,HGF,HMOX1,ICAM1,IFNG,IL6,MMP2, PECAM1,TGFB1,TNC,VCAM1
GO:0044444	cytoplasmic part	35	9377	1.02E-05	APP,CASP3,CCND1,CD34,CD44,CDKN1A,CLU,CYP3A4,EGF,EGFR,EP300, FYN,GPT,HDAC1,HGF,HMOX1,IL6,JUN,MAP2K7,MAPK1,MMP2, PC,PDGFRB,PECAM1,PXN,RAC1,SHC1,SLC2A1,SOD2,SREBF1,TGFB1, TNC,TP53,TPMT,VCAM1
GO:0031982	vesicle	16	2318	0.00018	APP,CD44,CLU,EGF,EGFR,FYN,HGF,ICAM1,MAPK1,PDGFRB,PECAM1,RAC1,SLC2A1,SREBF1,TGFB1,VCAM1
GO:0009986	cell surface	6	069	0.00019	APP, CD34, CD44, CLU, EGFR, ICAM1, PDGFRB, TGFB1, VCAM1
GO:0097386	glial cell projection	2	18	0.0047	APP,FYN
GO:0030425	dendrite	5	531	0.0212	APP,CLU,FYN,MAPK1,RAC1
GO:0000123	histone acetyltransferase complex	2	92	0.0412	CREBBP, EP300
GO:0005739	mitochondrion	~	1531	0.0417	CLU,FYN,MAPK1,MMP2,PC,SHC1,SOD2,TP53
GO:0070062	extracellular exosome	2	80	0.0435	EGF,ICAM1
KEGG pathways	S				
hsa05200	Pathways in cancer	21	515	1.54E-20	CASP3,CCND1,CDKN1A,CREBBP,EGF,EGFR,EP300,HDAC1,HGF,HMOX1, IFNG,IGF1R,IL6,JUN,MAPK1,MMP2,PDGFRB,RAC1,SLC2A1,TGFB1,TP53
hsa05206	MicroRNAs in cancer	14	149	7.73E-18	CASP3,CCND1,CD44,CDKN1A,CREBBP,EGFR,EP300,HDAC1,HMOX1, MAPK1,PDGFRB,SHC1,TNC,TP53
hsa04066	HIF-1 signaling pathway	11	86	8.67E-15	CDKN1A,CREBBP,EGF,EGFR,EP300,HMOX1,IFNG,IGF1R,IL6,MAPK1,SLC2A1
hsa04510	Focal adhesion	13	197	8.67E-15	CCND1,EGF,EGFR,FYN,HGF,IGF1R,JUN,MAPK1,PDGFRB,PXN,RAC1,SHC1, TNC
hsa05205	Proteoglycans in cancer	13	195	8.67E-15	CASP3,CCND1,CD44,CDKN1A,EGFR,HGF,IGF1R,MAPK1,MMP2,PXN,RAC1, TGFB1,TP53
hsa04068	FoxO signaling pathway	11	130	9.14E-14	



#term ID	Description	Observed gene count	Observed Background False gene count discor	False discovery rate	Proteins
					CCND1,CDKN1A,CREBBP,EGF,EGFR,EP300,IGF1R,IL6,MAPK1,SOD2, TGFB1
hsa04010	MAPK signaling pathway	13	293	3.56E-13	CASP3,EFNA1,EGF,EGFR,HGF,IGF1R,JUN,MAP2K7,MAPK1,PDGFRB, RAC1,TGFB1,TP53
hsa04151	PI3K-Akt signaling pathway	13	348	1.81E-12	CCND1,CDKN1A,EFNA1,EGF,EGFR,HGF,IGF1R,IL6,MAPK1,PDGFRB, RAC1,TNC,TP53
hsa04360	Axon guidance	4	173	0.00086	EFNA1,FYN,MAPK1,RAC1
hsa05100	Bacterial invasion of epithelial cells	3	72	0.00088	PXN,RAC1,SHC1



PPA induced not only lipid damage but also DNA damage [71]. In this study, AD- PPA genes enrich the process response to DNA damage, DNA geometric change, DNA damage checkpoint, or activated apoptotic pathway in response to DNA damage and positive regulation of chromosome organization. Previously determine the association of genomic instability observed in AD that one of evidence is emerging early-onset AD in a patient with down syndrome [89].

Inflammation process and response to acute and chronic inflammation or cytokine production by the immune system are processes that enriched AD-PPA genes. Inflammation in glia cells "Gliosis" is the hallmark of AD that enhances the amyloidogenic process [90] and detected by analysis of our genes in DisGeNet. Innate and adaptive immune response or humoral immune system enriched by 284 genes. The immune system plays an essential role in the progression, or maybe it is a risk factor for AD [91]. Lobzhanidze G et al. have been recognized the significant structural alteration in the amygdala beyond the administration of PPA in adolescent rats. They reported the Glial alterations, the activation of astrocytes and microglia, and axons demyelination [30]. Microglia plays an essential role in the clearance of tau oligomers and the actin cytoskeleton for phagocytosis in AD [92]. In addition, the pathway-related proliferation immune system response and inflammation induced by PPA noticed in autism spectrum disorder [25].

There was the amount of biological process linked to angiogenesis blood circulating and allocated the 26 genes to the hypoxia-induced response. Previous studies have indicated the vascular dysfunctions in the early year increase regionally blood flow as a compensatory mechanism while observed decrease eventually in the later stage of AD. The capillary amyloid angiopathy induced by hypoxia through activating  $\beta$ - and  $\gamma$ -secretases can be attributed to AD pathology [93]. Hypoxia condition modulates hypometabolism in several regions of the brain by overexpression of amyloid precursor protein and decreases the clearance of A $\beta$ . This event promotes inflammation and, ultimately, neuronal cell death [94]. HIF-1 signaling pathway supports cellular adaptation in hypoxic conditions that were seen in the KEGG pathway of AD-PPA genes.

Cell adhesion that detected in AD-PPA genes enriched biological processes related to cell-cell adhesion and cell-matrix adhesion. It has been determined that the genes with pleiotropic roles in cell adhesion highlighted in AD pathogenesis [95] and PPA also had been prepared induced extensive



Table 7 (continued)

alterations in gene expression, including neuronal cell adhesion molecules [90].

Cellular component analysis enriched the critical part of a cell that promotes AD pathogenesis. Cytoskeleton change in neuron cell that enriched by AD-PPA genes effect on the synapse. Rush T et al. reported that Aβ oligomers induce aberrant actin stabilization and synaptic loss and impairment [96]. Dysfunctionality was observed in several parts of the neuron, i.e., synapse or cellular secretion [90, 97]. Elevated PPA could also produce sensitivity to oxidative stress and, in turn, increase the damage caused by other toxic environmental factors such as metals or infectious agents [87]. Mitochondria also implicated alteration specific in function, size, and form by PPA treatment *in vivo* [30, 90, 98].

### **Conclusions**

AD-PPA genes analysis unveiled the comorbidity with diseases that surprisingly were related to the effect of bacteria and virus infection or enriched the neurological disease that previously reported in PPA intervention, such as meningitis or autism disorder. It could be pinpointed that AD-PPA genes carry biological processes that cover almost all of AD pathogenesis hallmarks. Functional analysis of hub-bottleneck-seed genes represented the role of these crucial genes in redox signaling, neuroinflammation, and cell cycle, and cell death. Since it has not been produced high throughput data directly obtained from the effect of PPA on cell line or animal models, our analysis opens the view of the possible effect of PPA on AD pathogenesis. Therefore, it is necessary to design further empirical investigations to attain deep insights into the PPA metabolite secreted by microbiota and implicated in AD pathogenesis.

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Author contributions Morteza Aliashrafi design the study carried out data collection and statistical analysis drafted the manuscript and interpreted the data; Mohammad Nasehi designed and supervised the study; Mohammad-Reza Zarrindast designed and supervised the study; Mohammad Taghi Joghataei design the study; Hakimeh Zali carried out data collection and statistical analysis; Seyed Davar Siadat designed and supervised the study. All authors read and approved the final manuscript.

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# **Compliance with ethical standards**

**Conflict of interest** The authors declare that they have no conflict of interests.

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