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4	NICHE SEGREGATION BETWEEN TWO CLOSELY RELATED
5	GAMMARIDS (CRUSTACEA, AMPHIPODA) – NATIVE VS. NATURALISED
6	NON-NATIVE SPECIES
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8	BY
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17	
18	ABSTRACT
19	Closely related species may occupy similar niches, but are often found to diverge
20	by one or more traits when they inhabit the same habitat. In this study, we examined
21	how two co-occurring gammarids - the native Gammarus fossarum and the
22	naturalised G. roeselii – are distributed among microhabitats, depending on their
23	sympatric or allopatric distribution. We hypothesized that the larger body sized
24	species (G. roeselii), exploiting their advantages in competition, restrict smaller
25	species to microhabitats with smaller particle sizes. Four headwaters were sampled in

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Mecsek Mountains (SW Hungary) in May, July and October 2009, and 37 local scale 26 environmental variables at each site were measured. Although G. fossarum is smaller 27 in size, significantly more individuals were collected from the more favourable lithal 28 and biotic microhabitats, whereas a strong negative association was observed between 29 the two species. Gammarus roeselii occurred at sites characterised by degraded 30 31 riparian vegetation, which indicates stronger anthropogenic impacts, but still has a disadvantage in competition in mountainous streams under anthropogenic influence. 32 33 Keywords: Gammarus fossarum, Gammarus roeselii, habitat segregation, co-34 existence, environmental variables;

INTRODUCTION

37 In the last one hundred years, through the accelerated industrialization, the state and conditions of watercourses have worsened a lot, that opened the gate for the 38 39 spread of invasive species (Van der Velde et al 2000), which may have been contributed the extinction of native species from the middle section of river Danube 40 (Bódis et al. 2012). This process could be responsible for the increase of the numbers 41 of invasive species in large European rivers like, Oder, Rhine or Vistula (Jazdzewski 42 43 1980, Jazdzewski & Konopacka 2000, Bij de Vaate et al. 2002, Konopacka and Jazdzewski 2002, Borza 2009). Native gammarid communities are exposed to the 44 45 impacts of invasive species (MacNeil & Platvoet 2005). That process is remarkable, 46 because gammarids are common and play important functional role in fresh and brackish running water systems across Europe (Jazdzewski 1980). They could be 47 48 considered as key species in aquatic assemblages, especially in food web interactions (Piscart et al. 2011). These species owe their success to their relatively short 49 50 generation time, fast sexual maturation and high reproductive ability (Bij de Vaate et al. 2002, Grabowski et al. 2007). Invasive gammarids have restricted native species 51 from numerous large rivers, to smaller mountainous streams, where the absence of 52 53 invasive species presumably related to the special physico-chemical condition of 54 habitats (e.g. high water velocity, low level of salinity) (Wijnhoven et al. 2003, Piscart et al. 2009). Changes in amphipod assemblages after the colonization by invasive 55 species have been well known (e.g. Dick 1996, Jazdzewski et al. 2005, Josens et al. 56 57 2005, Grabowski et al. 2006, Piscart et al 2011, Mayer et al. 2012), but interactions among native species assemblages are poorly known. 58 59 Among gammarids, several closely related species share highly similar ecological

60 niches (van Riel et al. 2009). Thus, strong interactions could occur between them (van

61	Riel et al. 2007). On the other hand, it is well known that changes in resource
62	allocation could help to avoid interference or competitive exclusion (Schoener 1983).
63	Among gammarids, shifts in life-cycles and microhabitat preference could limit the
64	niche overlap and competition between species (Korpinen & Westerbom 2009). Such
65	shifts could create horizontal (Czarnecka et al. 2010) and, if the depth permits,
66	vertical separations (Kley & Maier 2005). An interesting phenomenon of horizontal
67	separations is that the native gammarid species are usually restricted to biotic habitats
68	(roots and leaf litter) after the colonization of an invasive species (Dick 1996; Piscart
69	et al. 2007; van Riel et al. 2007). Nevertheless, precisely describing the structure of
70	native communities is important for estimating the effects of further invasions. To
71	survey changes in habitat segregation among gammarids, we chose two sympatric
72	species, which are showing similarity in several ecological traits; the native G.
73	fossarum Koch, in Panzer 1836 and the non-indigenous, but naturalised G. roeselii
74	Gervais, 1835. Gammarus fossarum is generally considered as a widely distributed
75	freshwater species in Europe (e.g. Karaman & Pinkster 1977) and also the most
76	abundant native amphipod in freshwater streams of mountainous areas in Central
77	Europe (Pöckl et al. 2003). Gammarus roeselii was introduced from the Balkans
78	(Karaman & Pinkster 1977, Jazdzewski & Roux, 1988) as an early invader, and today
79	is considered as a naturalised species (Piscart et al. 2009) in the Eastern- and Central-
80	European rivers, where it usually occupies the lower parts of watercourses
81	(Jazdzewski & Roux 1988; Janetczky 1994). The appearance of this species in
82	mountain streams is relatively unusual (Nesemann et al. 1995).
83	For this reason the aims of this study were to examine (1) how two closely related
84	gammarid species - \underline{G} . fossarum and \underline{G} . roeselii – are distributed among
85	microhabitats, depending on their sympatric or allopatric distribution, and (2) the

86	variability in their biotic interactions if they are sympatric. Our aim was to assess the
87	primary influencing factors (at temporal and different spatial scales) structuring the
88	distribution patterns of <u>G. fossarum</u> and <u>G. roeselii</u> . We hypothesized that, as the
89	result of interspecific competition, the two species show spatial segregation at the
90	level of microhabitat, if they co-occur. We also sought the abiotic environmental
91	variables that could determine the occurrence of <u>G. fossarum</u> and <u>G. roeselii</u> at the
92	reach scale. We examined the biotic interactions between the two species within
93	different microhabitats and seasons.
94	
95	MATERIAL AND METHODS
96	Sampling sites and data collection
97	The study area is located in Mecsek Mountains in south-western Hungary. The
98	350 km2 area is considerably isolated from other mountainous regions and bordered
99	by low plain and hilly territories. Macroinvertebrate samples were taken at four sites
100	(fig. 1) from second-order headwaters running in deep, cool and shadowed valleys. In
101	two of the four streams, the only occurring species is G. fossarum. Investigating the
102	biotic interactions and the changes in habitat preference of the two gammarid species
103	were possible in the two other streams (fig. 1). Samples were taken from a 100 m long
104	section of each site between 6-20 of May, 21-25 of July and 19-23 of October in
105	2009. The macroinvertebrates were quantitatively collected according to Integrated
106	Assessment System for the Ecological Quality of Streams and Rivers throughout
107	Europe using Benthic Macroinvertebrates (AQEM) protocol (AQEM Consortium,
108	2002), which focuses on a multihabitat scheme designed for sampling major habitats
109	in proportion to their presence within 100 m long sampling reaches. A sample
110	consisted of 20 'sampling units' taken from all microhabitat types at each sampling

111	site with a share of at least 5% coverage. The 20 'sampling units' were distributed
112	according to the proportion of microhabitats. A 'sampling unit' was taken from a total
113	of 0,25 * 0,25 m2 area by 'kick and sweep' sampling method using a handnet (1 mm
114	mesh size). In case of macrolithal type samples were collected from the given surface
115	of the rocks and among the fine sediment between them. Thus, a total of 1.25 m^2 area
116	was sampled for benthic macroinvertebrates at each site. In this study, seven different
117	types of habitat were determined according to the original AQEM microhabitats (table
118	1). The 'sampling units', which contained the complete assortment of
119	macroinvertebrates, were fully and separately sorted in the field. The collected
120	gammarids were identified in laboratory from the sorted samples, based on the keys
121	and descriptions of Cărăușu et al. (1955) and Kontschán et al. (2002).
122	
400	
123	Environmental variables
123 124	At each site, 37 local scale environmental variables were measured in all seasons
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136	vegetation (Equisetum telmateia, Petasites hybridus, Urtica dioica), secondary bushes
137	(e.g. of Cornus sanguinea, Corylus avellana), and secondary riparian forests (Salix
138	alba, Salix fragilis).

140

Statistical analyses

141 Prior to all analysis, the abundance data of the two species were log ₁₀	(x+1)
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142 transformed to reduce heteroskedasticity.

143 To identify the seasonal shifts in microhabitat preferences, we calculated

additional Kruskal-Wallis tests within each season. To compare the abundances of <u>G.</u>

145 <u>fossarum</u> directly from sites with and without the other species were carried out with

146 Mann-Whitney U tests. For these analyses, we reduced the seven available

147 microhabitat types into three wider habitat groups (table 1). Thus, we cumulated the

abundance of each species and referred them to $1m^2$ because different numbers of

sampling units of single microhabitat types were available at sampling sites.

150 Furthermore, to explore the differences between the abiotic factors of the two types of

151 site, independent samples t-tests were used. These analyses were implemented with

software R ver. 2.14.0 (R Development Core team 2011).

Coexistence analyses were made in Microsoft Excel Macro, based on the work of 153 Schmera et al. (2007). Co-existence indices (CI_{ii}) between the species were calculated 154 using the formula $CI_{ij} = \sum_{a=1}^{N} x_{ai} * x_{aj}$, where CI_{ij} is the co-existence index (i.e. degree 155 of association) between i and j species, x_{ai} is the relative abundance of species 'i' in 156 sample 'a', x_{ai} is the relative abundance of species 'j' in sample 'a', N is the total 157 number of samples. Altogether 1000 random pseudo-assemblages were generated, 158 species' abundances were kept constant in a sample. If the observed value falls in the 159 upper marginal tail of the random distribution, it presumes a positive association (e.g. 160

aggregation). If the observed value falls in the lower marginal tail of the distribution,
then the species pair shows negative association (e.g. competition) (Schmera et al.
2007).

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- 165

166

RESULTS

Temporal and spatial distribution patterns

During the survey, a total of 27,445 specimens were identified. There was no

168 significant difference in abundances of the species among sites (Kruskal-Wallis test:

169 $\chi^2 = 6.385$, df = 2 p = 0.094).

170 The temporal shift in the microhabitat preference, based on the Kruskal-Wallis171 tests did not show significant differences in quantitative distribution between

microhabitat types at sites where only <u>G. fossarum</u> was present (spring: $\chi^2 = 0.432$, df

173 = 2, p = 0.806; summer: χ^2 = 3,545, df = 2, p = 0.170; autumn: χ^2 = 1.054, df = 2, p =

174 0.590) (fig. 2). However, when both species were present (fig. 3) a significant

175 difference occurred in the microhabitat preference of <u>G. fossarum</u> in spring and

summer (spring: $\chi^2 = 10.744 \text{ df} = 2$, p = 0.005; summer: $\chi^2 = 14.617$, df = 2, p =

177 0.001). In autumn, no significant difference was found in habitat preference ($\chi^2 =$

178
$$1.618, df = 2, p = 0.445$$
 (fig. 3)

The abundance of <u>G. roeselii</u> was low in spring, and no difference was found in the abundance between microhabitats ($\chi^2 = 0.614$, df = 2, p = 0.736) (fig. 4), whereas significant differences were found in the abundance between the microhabitat types in summer and autumn (summer: $\chi^2 = 11.349$, df = 2, p = 0.003; autumn: $\chi^2 = 14.080$, df = 2, p = 0.001) (fig. 4).

Based on the Mann-Whitney U tests, the comparison of the abundances of <u>G.</u>
fossarum with and without G. roeselii showed no differences in most cases. We found

186	exceptions in spring in the lithal and gravel microhabitats and in the biotic one in
187	summer (table 3.)
188	
189	Coexistence
190	Throughout the year, negative associations were found between the species in
191	almost every habitat type, which indicates interference. The only exception was the
192	gravel microhabitat, where positive associations were observed in autumn (table 4).
193	
194	Comparing abiotic conditions of the two groups of sites with different species
195	composition
196	Comparing the two different groups of sites we found difference in some factors
197	of bed morphology and degradation state together with an ion concentration and the
198	proportion of the xylal microhabitat type (t = -2.684, p = 0.028). The biggest
199	difference among factor of bed morphology was found in water depth (t = -3.556, p =
200	0.007). Furthermore, significant difference was found in the degradation state of
201	riparian vegetation. The secondary (degraded) site number was higher at sites where
202	<u>G. roeselii</u> was present at the scale of valley floor (t = -3.437, p = 0.009) and slope (t
203	= -5.817, p = 0.001). Additionally concentration (mg/l) of Ca^{2+} ion was higher (t = -
204	4.530, $p = 0.002$) at sites where both species were present. Also, significant difference
205	was found in coverage of riparian shrub (t = -2.623, p = 0.031) and forest (t = 3.217, p
206	= 0.012) between the two sites.
207	

DISCUSSION

209	Our study provides information about the spatial niche segregation of two
210	gammarid species (G. fossarum, G. roeselii) in small headwaters. We also made an
211	attempt to identify factors affecting the distribution patterns.
212	We revealed different microhabitat preference of the gammarid species at sites
213	where they co-occurred (fig. 3, 4). We also showed that differences in microhabitat
214	preference did not exist during the whole year and its rate changed among seasons
215	(fig. 3, 4). Gammarus fossarum showed microhabitat preference only, if it co-
216	occurred with G. roeselii. In those cases, G. fossarum was mostly abundant in the
217	optimal lithal and biotic microhabitats, whereas it was less dominant in gravel ones.
218	On the contrary, G. roeselii was almost completely absent from the lithal
219	microhabitats, but it was usually frequent in the biotic ones. Our results partly
220	confirmed some previous studies in which competition could be observed between \underline{G} .
221	fossarum and G. roeselii at stream reaches if they co-occurred (e.g. Pöckl &
222	Humpesch 1990). Besides, the reproduction is not restricted to a short period and may
223	occur throughout the year (Beracko et al. 2012), thus we consider that the seasonal
224	investigation might not be sufficient to show the shift of their life-cycles.
225	Nevertheless, former studies (e.g. van Overdijk et al. 2003, McGrath et al. 2007,
226	Korpinen & Westerbom 2009) also focused mostly on the spatial resource partition,
227	especially on habitat separation. In a laboratory experiment, G. pulex and G. roeselii
228	did not show any change in habitat preference when co-occurring (van Riel et al.
229	2007). On the contrary, a high degree of divergence in substrate choice was found
230	between the aggressive invader <u>Dikerogammarus villosus</u> and the non-indigenous <u>G.</u>
231	roeselii (Kley et al. 2009). The weak interactions between G. roeselii and G. pulex
232	were supported by a field survey, which provided for <u>G. roeselii</u> to colonize the more
233	favourable habitats that are less acceptable to <u>G. pulex</u> (Kaldonski et al. 2008).

Thereby, G. roeselii could permanently co-exist with other native species. On the 234 235 other hand, Túri et al. (2003) observed a competitive exclusion and checkerboard pattern between two native species (G. fossarum, G. balcanicus) in mountainous 236 237 small streams in NE Hungary. Regarding the habitat segregation, several previous studies revealed that larger species, exploiting their advantages in competition, 238 restrict smaller species to microhabitats with smaller particle sizes (e.g. Hacker & 239 Steneck 1990, Olyslager & Williams 1993). Despite that, G. fossarum is smaller in 240 body size (Pöckl 1992), even though we collected significantly more individuals from 241 larger grained lithal microhabitats; moreover G. fossarum was relatively frequent in 242 the biotic microhabitats. According to several investigations, G. roeselii prefers 243 slower and warmer stream sections (e. g. Meijering 1972; Dahl & Greenberg 1996; 244 Toman & Dall 1998), and it is not able to colonize springs and spring outlets (e.g. 245 246 Wijnhoven et al. 2003, Piscart et al. 2009). Furthermore, our results show that G. roeselii occurred at sites characterised by abiotic habitat features resembling 247 248 downstream sections of streams with degraded riparian vegetation, which indicates stronger anthropogenic impacts (table 2, fig. 5). It is well known that environmental 249 factors have non-negligible effects on the distribution pattern (Früh et al. 2012), 250 besides that G. roeselii shows some kind of expansion as it appears in sufficiently 251 252 degraded second order streams. Nevertheless, we supposed that small, mountainous headwaters were less optimal habitats for G. roeselii; thereby the typical mountain 253 species G. fossarum could be a stronger competitor. This presumption was also 254 confirmed, according to the permanence microhabitat preference of G. fossarum 255 (table 3). Several authors (Dick 1996; Piscart et al. 2007; van Riel et al. 2007) found 256 257 similar distribution patterns of native and presumably weaker competitor species being restricted to biotic microhabitats and excluded from the, lithal ones following 258

invasions. Based on the previous statement, we can assume that, being the stronger
competitor, <u>G. fossarum</u> restricts <u>G. roeselii</u> from optimal lithal microhabitats, while
still dominant in the biotic ones, characterised by higher amounts of detritus
deposition.

Coexistence analyses confirmed the influence of biotic interactions on the 263 distribution patterns. Presumably, G. fossarum could be the stronger competitor, 264 whereas a strong negative association was found between the two species, which 265 confirms the habitat segregation between the two species. Positive association could 266 be found only between the species in autumn in the gravel microhabitats. Throughout 267 the year, the fine particulate substrate was characterised by small numbers of 268 individuals. Therefore, when large quantities of allochthonous organic matter (e.g. 269 leaf litter) appeared in autumn, habitat structure changes for gammarids, providing 270 271 shelter and nutritive as well.

To sum up, our aim was to describe the effect of an expanding species (G. 272 273 roeselii) on a closely related species (G. fossarum) belonging to the same functional guild and utilizing similar niches (Nesemann et al. 2002). Thus strong competition 274 could be assumed between them. The most notable is that a simple practical model 275 based on a field study supported the previous theoretical models that described the 276 277 main opportunity to avoid competitive exclusion based on niche segregation (Tilman 1987). In this particular case, this segregation appeared as spatial resource allocation. 278 Our study revealed whether the native or the 'expanding' species has the advantage 279 280 in the competition in a certain environmental condition (small mountainous streams). Since we worked in mostly natural habitats, G. roeselii which arrived from the lower 281 sections of streams still had a disadvantage in competition. In fact, we did not expect 282 the appearance of G. roeselii in these natural stream sections; however, it spread 283

284	already at the moderately disturbed reaches. Lastly, we conclude, that the
285	degradation of near-pristine headwater sites could thus enable the naturalized non-
286	native species to get into action for further expansion of its area.
287	
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294	
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461 TABLES

462	Table 1.		
	Groups	Туре	Definition
	Gravel	Psammal	Sand; diameter 6µm–2 mm
		Akal	Fine to medium-sized gravel; diameter 2mm-2 cm
	Lithal	Microlithal	Coarse gravel with medium to fine gravel; diameter 2-6 cm
		Mesolithal	Cobbles with a variable percentage of gravel and sand; diameter 6-20 cm
		Macrolithal	Coarse cobbles, gravel and sand; diameter 20-40 cm
	Biotic	Xylal	Tree trunks, dead wood, branches, roots
		CPOM	Deposits of coarse particulate organic matter

Table 2.

Shearmate composition % Macrolithal $0.0042213.22$ (6.28) $0.000.953.17$ (1.45) $arcsin(x/100)^{9.5}$ % Mesolithal $0.7910.3918.73$ $4.7615.1828.44$ $arcsin(x/100)^{9.5}$ % Microlithal $0.7910.3918.73$ $4.7615.1828.44$ $arcsin(x/100)^{9.5}$ % Akal $0.798.8117.62$ $0.0007824.60$ $arcsin(x/100)^{9.5}$ % Akal $0.5212.1426.70$ $6.8715.4939.68$ $arcsin(x/100)^{9.5}$ % Xylal $0.5212.1426.70$ $6.8715.4939.68$ $arcsin(x/100)^{9.5}$ % CPOM $0.0065.320.08$ $(0.4.72)$ $arcsin(x/100)^{9.5}$ % CPOM $0.0065.320.08$ $(0.0055.0700$ $bn(x+1)$ Number of pols in 100 3.00433800 40055.0700 $bn(x+1)$ Number of pols in 100 3.00433800 40055.0700 $bn(x+1)$ Number of bonds in 100 2.00333600 $3.0055.0700$ $bn(x+1)$ Number of bonds in 100 2.00333600 $5.0065.3700$ $bn(x+1)$ Number of bonds in 100	Variables	Sites with <u>G. fossarum</u> only	Sites with <u>G. fossarum</u> and <u>G. roeselii</u>	Transform.
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Number of bends in 100 m2.003.836.00 (1.72)3.004.838.00 (2.13) $\ln(x+1)$ Number of woods in 100 m3.005.169.00 (2.99)6.0010.3.00 (2.80) $\ln(x+1)$ % Detritus $6.7024.4754.00$ (18.39) $19.0535.1252.38$ (13.99) $\arctan(x+1)$ % Washaway $4.5520.0442.86$ (17.34) $4.7619.2135.71$ (15.54) $\arcsin(x/100)^{0.5}$ Physicochemical attribute (17.34) (15.54) $\arctan(x+1)$ pH $7.978.399.00$ (0.38) (2.67) (0.50) $\ln(x+1)$ Water temperature (°C) $6.4013.0119.40$ (5.15) (2.67) (2.67) $\ln(x+1)$ Conductivity (μ S/cm) $572.00722.23886.00$ (1.58.43) $649.70795.93899.30$ (89.60) $\exp(x/100)$ Concentration of dissolved oxygen (mg L ⁻¹) $0.020.491.00$ (0.43) $0.040.411.20$ (0.01) $\ln(x+1)$ NO ₂ ⁻¹ (mg L ⁻¹) $0.020.932.10$ (0.78) $0.040.552.70$ (0.28) $\ln(x+1)$ NO ₃ ²⁻¹ (mg L ⁻¹) $0.180.460.84$ (0.28) $0.140.400.76$ (0.29) $\ln(x+1)$ NO ₄ ²⁻¹ (mg L ⁻¹) $0.3041.84120.00$ (0.78) $10.557.37140.00$ (0.20) $\ln(x+1)$ $O_4^{2^2+1}$ (mg L ⁻¹) $0.3041.84120.00$ (0.28) $10.557.37140.00$ (52.15) $\ln(x+1)$ $O_4^{2^2+1}$ (mg L ⁻¹) $0.3041.84120.00$ (0.28) $10.557.37140.00$ (52.15) $\ln(x+1)$	Water depth (m)			ln(x+1)
Number of woods in 100 m $3.005.169.00$ (2.99) $6.0010.3313.00$ (2.80) $\ln(x+1)$ % Detritus $6.7024.4754.00$ (18.39) $19.0535.1252.38$ (13.99) $\arcsin(x/100)^{0.5}$ arcsin(x/100)^{0.5}% Washaway $4.5520.442.86$ (17.34) $4.7619.2135.71$ (15.54) $\arcsin(x/100)^{0.5}$ Physicochemical attribute $17.748399.00$ (0.38) $7.127.918.50$ (0.50) $\exp(x/100)$ Water temperature (°C) $6.4013.0119.40$ (5.15) $10.1012.6517.83$ (2.67) $\ln(x+1)$ Conductivity (μ S/cm) $572.00722.23886.00$ (15.843) $649.70795.93899.30$ (89.60) $\exp(x/100)$ Concentration of dissolved oxygen $5.106.918.10$ (1.17) $5.206.508.20$ (0.33) $\ln(x+1)$ NH4^+ (mg L^{-1}) $0.020.491.00$ (0.01) $0.040411.20$ (0.53) $\ln(x+1)$ NO2^{-2} (mg L^{-1}) $0.180.460.84$ (0.28) $0.140400.76$ (0.29) $\ln(x+1)$ NO3^{2-} (mg L^{-1}) $0.3041.84120.00$ (47.09) $10.557.37140.00$ (52.15) $\ln(x+1)$ SO4^{2-} (mg L^{-1}) $0.3041.84120.00$ (47.09) $10.557.16134.59165.18$ ($n(x+1)$		2.003.836.00	3.004.838.00	ln(x+1)
$\begin{tabular}{ c c c c c c c c c c c c c c c c c c c$		3.005.169.00	6.0010.3313.00	ln(x+1)
76° washaway (17.34) (15.54) atesh((x)100) Physicochemical attribute (0.38) (15.54) atesh((x)100) PH $7.978.399.00$ $7.127.918.50$ $exp(x/100)$ Water temperature (°C) $6.4013.0119.40$ $10.1012.6517.83$ $ln(x+1)$ Conductivity (μ S/cm) $572.00722.23886.00$ $649.70795.93899.30$ $exp(x/100)$ Concentration of dissolved oxygen (158.43) $5106.918.10$ $5.206.508.20$ $ln(x+1)$ NH ₄ ⁺ (mg L ⁻¹) $0.020.491.00$ $0.040.411.20$ $ln(x+1)$ NO ₂ ⁻ (mg L ⁻¹) $0.010.020.03$ $0.010.050.32$ $ln(x+1)$ NO ₂ ⁻ (mg L ⁻¹) $0.180.460.84$ $0.140400.76$ $ln(x+1)$ SO ₄ ²⁻ (mg L ⁻¹) $0.3041.84120.00$ $1.0557.37140.00$ $ln(x+1)$ SO ₄ ²⁻ (mg L ⁻¹) $0.3041.84120.00$ $1.0557.37140.00$ $ln(x+1)$ Co ²⁺ (mg L ⁻¹) $0.3041.84120.00$ $1.0557.37140.00$ $ln(x+1)$	% Detritus			$\arcsin(x/100)^{0.5}$
attributepH $7.978.399.00$ (0.38) $7.127.918.50$ (0.50) $exp(x/100)$ Water temperature (°C) $6.4013.0119.40$ (5.15) $10.1012.6517.83$ (2.67) $ln(x+1)$ Conductivity (μ S/cm) $572.00722.23886.00$ (158.43) $649.70795.93899.30$ (89.60) $exp(x/100)$ Concentration of dissolved oxygen ($mg L^{-1}$) $5.106.918.10$ (1.17) $5.206.508.20$ (1.34) $ln(x+1)$ NH4+ (mg L^1) $0.020.491.00$ (0.43) $0.040.411.20$ (0.53) $ln(x+1)$ NO2^{-} (mg L^{-1}) $0.010.020.03$ (0.01) $0.010.050.32$ (0.01) $ln(x+1)$ NO3^{2^{-}} (mg L^{-1}) $0.180460.84$ (0.28) $0.140.400.76$ (0.20) $ln(x+1)$ SO4^{2^{-}} (mg L^{-1}) $0.3041.84120.00$ (47.09) $1.0557.37140.00$ (52.15) $ln(x+1)$ $Ca^{2^{+}}$ (mg L^{-1}) $5.00483.57110.43$ $115.46134.59165.18$ $ln(x+1)$	% Washaway			$\arcsin(x/100)^{0.5}$
pH (0.38) (0.50) $exp(x/100)$ Water temperature (°C) $6.4013.0119.40$ (5.15) $10.1012.6517.83$ (2.67) $ln(x+1)$ Conductivity (µS/cm) $572.00722.23886.00$ (158.43) $649.70795.93899.30$ (89.60) $exp(x/100)$ Concentration of dissolved oxygen (mg L ⁻¹) $5.106.918.10$ (1.17) $5.206.508.20$ (1.34) $ln(x+1)$ NH4+ (mg L ⁻¹) $0.020.491.00$ (0.43) $0.040.411.20$ (0.53) $ln(x+1)$ NO2 ⁻ (mg L ⁻¹) $0.010020.03$ (0.01) $0.010050.32$ (0.03) $ln(x+1)$ NO3 ²⁻ (mg L ⁻¹) $0.200932.10$ (0.78) $0.401.552.70$ (0.28) $ln(x+1)$ SO4 ²⁻ (mg L ⁻¹) $0.3041.84120.00$ (47.09) $1.0557.37140.00$ (52.15) $ln(x+1)$	•			
Water temperature (°C) $6.4013.0119.40$ (5.15) $10.1012.6517.83$ (2.67) $ln(x+1)$ Conductivity (µS/cm) $572.00722.23886.00$ (158.43) $649.70795.93899.30$ (89.60) $exp(x/100)$ Concentration of dissolved oxygen (mg L ⁻¹) $5.106.918.10$ (1.17) $5.206.508.20$ (1.34) $ln(x+1)$ NH4+ (mg L ⁻¹) $0.020.491.00$ (0.43) $0.040.411.20$ (0.53) $ln(x+1)$ NO2^{-} (mg L^{-1}) $0.010.020.03$ (0.01) $0.010.050.32$ (0.05) $ln(x+1)$ NO3^{2-} (mg L^{-1}) $0.200.932.10$ (0.78) $0.401.552.70$ (0.93) $ln(x+1)$ PO4^{3-} (mg L^{-1}) $0.180.460.84$ (0.28) $0.140.400.76$ (0.20) $ln(x+1)$ SO4^{2-} (mg L^{-1}) $0.3041.84120.00$ (47.09) $1.0557.37140.00$ (52.15) $ln(x+1)$	pН			exp(x/100)
Conductivity (µS/cm) (158.43) (89.60) $exp(x/100)$ Concentration of dissolved oxygen (mg L ⁻¹) $5.106.918.10$ (1.17) $5.206.508.20$ (1.34) $ln(x+1)$ NH4+ (mg L ⁻¹) $0.020.491.00$ (0.43) $0.040.411.20$ (0.53) $ln(x+1)$ NO2- (mg L ⁻¹) $0.010.020.03$ (0.01) $0.010.050.32$ (0.05) $ln(x+1)$ NO3- (mg L ⁻¹) $0.200.932.10$ (0.78) $0.401552.70$ (0.93) $ln(x+1)$ PO4-3- (mg L ⁻¹) $0.180.460.84$ (0.28) $0.140.400.76$ (0.20) $ln(x+1)$ SO4-2- (mg L ⁻¹) $0.3041.84120.00$ (47.09) $1.0557.37140.00$ (52.15) $ln(x+1)$ Ca ²⁺ (mg L ⁻¹) $50.0483.57110.43$ $115.46134.59165.18$ $ln(x+1)$	Water temperature (°C)	6.4013.0119.40	10.1012.6517.83	ln(x+1)
dissolved oxygen (mg L ⁻¹) $5.106.918.10$ (1.17) $5.206.508.20$ (1.34) $ln(x+1)$ NH4+ (mg L ⁻¹) $0.020.491.00$ (0.43) $0.040.411.20$ (0.53) $ln(x+1)$ NO2- (mg L ⁻¹) $0.010.20.03$ (0.01) $0.010.050.32$ (0.05) $ln(x+1)$ NO3- (mg L ⁻¹) $0.200.932.10$ (0.78) $0.401.552.70$ (0.93) $ln(x+1)$ PO4-3- (mg L ⁻¹) $0.180.460.84$ (0.28) $0.140.400.76$ (0.20) $ln(x+1)$ SO4-2- (mg L ⁻¹) $0.3041.84120.00$ (47.09) $1.0557.37140.00$ (52.15) $ln(x+1)$ Ca ²⁺ (mg L ⁻¹) $50.0483.57110.43$ $115.46134.59165.18$ $ln(x+1)$	Conductivity (µS/cm)			exp(x/100)
NH4 (mg L ') (0.43) (0.53) $ln(x+1)$ NO2 (mg L') $0.010.20.3$ $0.010.50.32$ $ln(x+1)$ NO32 (mg L') $0.200.932.10$ $0.401.552.70$ $ln(x+1)$ NO32 (mg L') $0.200.932.10$ $0.401.552.70$ $ln(x+1)$ PO43 (mg L') $0.180.460.84$ $0.140.400.76$ $ln(x+1)$ SO42 (mg L') $0.3041.84120.00$ $1.0557.37140.00$ $ln(x+1)$ SO42 (mg L') $50.0483.57110.43$ $115.46134.59165.18$ $ln(x+1)$	dissolved oxygen			ln(x+1)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$NH_4^+ (mg L^{-1})$			ln(x+1)
NO3 (mg L)(0.78)(0.93) $\ln(x+1)$ PO43 (mg L-1)0.180.460.840.140.400.76 $\ln(x+1)$ SO42 (mg L-1)0.3041.84120.001.0557.37140.00 $\ln(x+1)$ Co22+ (mg L-1)50.0483.57110.43115.46134.59165.18 $\ln(x+1)$	NO_2^- (mg L ⁻¹)	0.010.020.03	0.010.050.32	ln(x+1)
PO4 (mg L) (0.28) (0.20) In(x+1) SO_4^{2-} (mg L ⁻¹) 0.3041.84120.00 1.0557.37140.00 In(x+1) (47.09) (52.15) In(x+1) Ca^{2+} (mg L ⁻¹) 50.0483.57110.43 115.46134.59165.18 In(x+1)	$NO_3^{2-}(mg L^{-1})$	0.200.932.10	0.401.552.70	ln(x+1)
$SO_4^{2-} (mg L^{-1}) \qquad \begin{array}{c} 0.3041.84120.00 & 1.0557.37140.00 \\ (47.09) & (52.15) \\ 50.0483.57110.43 & 115.46134.59165.18 \\ 1n(x+1) \\ 1n(x+$	$PO_4^{3-}(mg L^{-1})$			ln(x+1)
$Ce^{2^+}(mg I^{-1})$ 50.0483.57110.43 115.46134.59165.18 $ln(x+1)$	$SO_4^{2-}(mg L^{-1})$	0.3041.84120.00	1.0557.37140.00	ln(x+1)
	$Ca^{2+}(mg L^{-1})$		115.46134.59165.18	ln(x+1)

$Mg^{2+}(mg L^{-1})$	19.5633.9645.24 (10.47)	14.9743.91102.05 (31.54)	ln(x+1)
Cl ⁻	1.014.9710.19 (3.99)	7.769.9712.12 (1.69)	ln(x+1)
HCO ₃	238.90365.93415.80 (66.30)	360.90442.92525.60 (69.63)	ln(x+1)
Chemical oxygen demand	3.043.704.75 (0.59)	0.771.963.12 (1.03)	ln(x+1)
m alkalinity	5.506.256.80 (0.52)	5.897.338.60 (1.08)	ln(x+1)
Riparian vegetation			
% Tree (0-1m)	8.7519.6029.76 (7.97)	14.6720.0126.12 (3.71)	$\arcsin(x/100)^{0.5}$
% shrubs (0-1m)	1.193.866.81 (2.03)	6.3317.0424.93 (7.29)	$\arcsin(x/100)^{0.5}$
% herbaceous (0-1m)	46.4354.9965.09 (8.62)	48.4760.8969.43 (7.70)	$\arcsin(x/100)^{0.5}$
% Forest coverage at the valley floor	90.0095.00100.00 (5.44)	100.00	$\arcsin(x/100)^{0.5}$
% of natural habitat of vegetation at valley floor	100.00	50.00	$\arcsin(x/100)^{0.5}$
% of degraded habitat of vegetation at valley floor	0.00	50.00	$\arcsin(x/100)^{0.5}$
% Forest coverage of at the nearest slopes	90.0095.00100.00 (5.44)	60.0075.0090.00 (16.43)	$\arcsin(x/100)^{0.5}$
% of natural habitat of vegetation at slope	50.0075.00100.00 (27.38)	33.0054.160.75 (22.82)	$\arcsin(x/100)^{0.5}$
% of degraded habitat of vegetation at slope	0.0025.0050.00 (27.38)	25.0045.8366.00 (22.82)	$\arcsin(x/100)^{0.5}$
Hydrology			
Distance from source (km)	1.472.323.17 (1.20)	0.202.083.96 (2.65)	ln(x+1)
Altitude (m)	218.00268.50319.00 (71.41)	187.00203.00219.00 (22.62)	ln(x+1)
Aspect	96.12138.75181.37 (60.27)	108.60124.77140.94 (22.86)	ln(x+1)
Slope	4.006.509.00 (3.53)	7.008.009.00 (1.41)	ln(x+1)

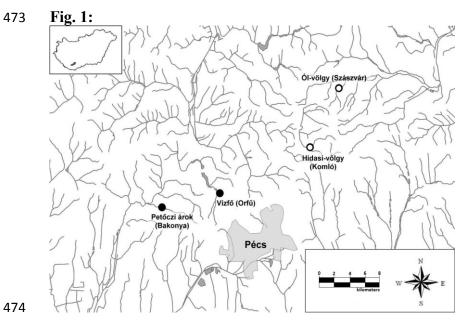
Table 3.

Season / m.habitat	lithal	gravel	biotic
Spring	Z = -2.559, p = 0.009	Z = -3.249, p = 0.001	Z = -0.387, p = 0.755
Summer	Z = -1.846, p = 0.065	Z = -1.736, p = 0.088	Z = -2.492, <i>p</i> = 0.011
Autumn	Z = -0.66, p = 0.948	Z = -1.827, p = 0.067	Z = -0.31, p = 0.976

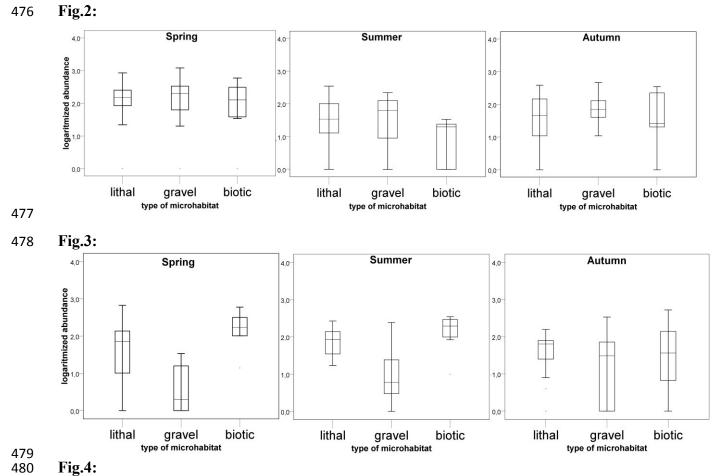
Table 4.

Microhabitat type	CI _{ij}	Upper 2.5%	Lower 2.5%	р	type of association
Spring					
Lithal	0.042	0.067	0.074	0.001	Negative
Gravel	0.074	0.156	0.173	0.001	Negative
Biotic	0.120	0.167	0.176	0.001	Negative
Summer					
Lithal	0.028	0.046	0.049	0.001	Negative
Gravel	0.109	0.211	0.223	0.001	Negative
Biotic	0.138	0.147	0.150	0.001	Negative
Autumn					
Lithal	0.035	0.056	0.060	0.001	Negative
Gravel	0.396	0.141	0.173	0.001	Positive
Biotic	0.228	0.288	0.308	0.001	Negative

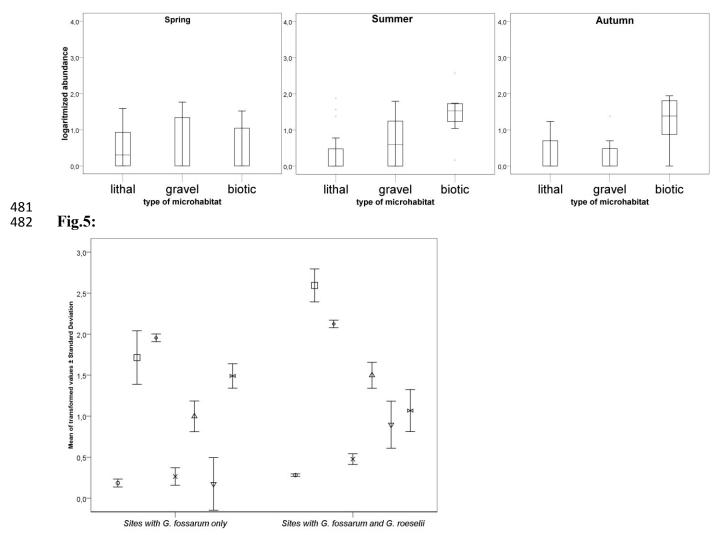
FIGURES 472











484 Table and figure captions

Table 1. List of microhabitats from AQEM protocol (following Hering et al. 2004.)

and the cumulative microhabitat groups used in this study.

487 **Table 2.** Minimum, mean and maximum (standard deviation) values of local

488 environmental variables used in analyses and grouped into four variable groups, and

489 their type of transformation.

490 **Table 3**. Comparison of the abundance data of <u>G. fossarum</u> with and without <u>G.</u>

491 <u>roeselii</u> in each season separated by microhabitats, the bold values are representing the

- 492 significant differences.
- 493 **Table 4**. Results of the co-existence analyses in seasonal partition, where CI_{ij} is the
- 494 co-existence index, upper 2.5% means the value of random distribution top 2.5%,
- lower 2.5% is the value of random distribution bottom 2.5%.

- 496 Fig. 1: Map of the study area. '•' marked sites where <u>G. fossarum</u> and <u>G. roeselii</u> co-
- 497 existed and ' \circ ' marked sites where *G. fossarum* is the only occurring gammarid.
- 498 Petőczi-árok: Petőczi stream (Bakonya; N 46°07'17" E 18°03'42"; 187m a.s.l.);
- 499 Vízfő: Vízfő spring (Orfű; N 46°08'21" E 18°09'37"; 219m a.s.l.); Hidasi-völgy:
- 500 Hidas stream (Komló; N 46°11'46" E 18°19'06"; 319m a.s.l.), Ól-völgy: Ól stream
- 501 (Szászvár, N 46°15'49" E 18°22'01"; 218m a.s.l.).
- 502 Fig. 2: Based on the mean of abundances, <u>Gammarus fossarum</u> showed no significant
- 503 microhabitat preference in the case of its single occurrence in each season (\Box :
- interquartile range T: standard error of mean SE, \circ outlier).
- **Fig. 3:** Based on the mean of abundances, it is clearly visible that <u>G. fossarum</u> showed
- a remarkable change in microhabitat preference in the case of co-existence with \underline{G} .
- 507 <u>roeselii</u> compared with the habitat choice of its single occurrences (\Box : interquartile
- 508 range T: standard error of mean SE , \circ outlier).
- 509 Fig. 4: Based on the mean of abundances, a definite preference of microhabitat of <u>G</u>.
- 510 <u>roeselii</u> was found in the case of co-existence with <u>G. fossarum</u> in each season (\Box :
- 511 interquartile range T: standard error of mean SE, \circ outlier).
- 512 Fig. 5: Comparison of the abiotic conditions of the two sites with different species
- 513 composition suggests the degraded state of the sites where both species co-occurred
- 514 (\circ : proportion of xylal microhabitat at the 100m section; \Box : water-depth; \diamond :
- 515 concentration of Ca^{2+} ion, \times : % shrubs (0-1m); \triangle : Number of degraded riparian
- habitats; ∇ : Number of degraded habitats at the nearest hill-side; $\triangleright \triangleleft$: the proportion
- 517 of the forest coverage at the nearest slopes).