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6 **The relative importance of spatial and environmental processes in distribution of**

7 **benthic chironomid larvae within a large and shallow lake**

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18

19 **Abstract** Although chironomids are popular model organisms in ecological research and
20 indicators of bioassessment, the relative role of dispersal and environmental filtering in their
21 community assembly is still poorly known, especially at fine spatial scales. In this study we
22 applied a metacommunity framework and used various statistical tools to examine the relative
23 role of spatial and local environmental factors in distribution of benthic chironomid taxa and
24 their assemblages in large and shallow Lake Balaton, Hungary. Contrary to present
25 predictions on the metacommunity organization of aquatic insects with winged terrestrial
26 adults, we found that dispersal limitation can considerably affect distribution of chironomids
27 even at lake scale. However, we also revealed the predominant influence of environmental
28 filtering, and strong taxa-environment relationships were observed especially along sediment
29 type, sediment organic matter content and macrophyte coverage gradients. We account that
30 identified reference conditions and assemblages along with specified optima and tolerances of
31 the abundant taxa can contribute to our understanding of chironomid ecology and be utilized
32 in shallow lake bioassessment. Further, we propose that predictive models of species-
33 environment relationships should better take into account pure spatial structuring of local
34 communities and species-specific variability of spatial processes and environmental control
35 even at small spatial scales.

36

37 **Keywords** Chironomidae assemblages, Dispersal limitation, Environmental gradients,
38 Indicator species, Optima and tolerances, Within lake spatial pattern.

39

40 **Introduction**

41

42 Most organisms are distributed non-randomly in space and the analysis of these spatial
43 patterns is one of the most beneficial tools of ecological research and bioassessment based
44 environmental management. For studying the spatial distribution of species and their
45 assemblages, the metacommunity concept (Leibold et al., 2004; Logue et al., 2011) has been
46 increasingly applied because of its comprehensive approach. This concept acknowledges that
47 the distribution of organisms is influenced by a series of processes related to species identity,
48 regional (dispersal and demographic stochasticity among sites) and/or local (species sorting
49 along biotic and abiotic environmental filters) factors. In their conceptual work, Leibold et al.
50 (2004) introduced four metacommunity perspectives accounting for local community
51 structures depending on the relative role of stochastic, regional (i.e. dispersal) and local (i.e.
52 environmental filtering) processes. However, development of the concept has resulted in a
53 much more continuous approach of metacommunity organization characteristics (Gravel et
54 al., 2006). The recent framework emphasizes the fundamental and practical importance of
55 testing the relative role of spatial and local factors implying on the relevance of dispersal and
56 environmental filtering processes, respectively (Logue et al., 2011), which can be dissected by
57 recently developed statistical tools (Borcard et al., 2004; Dray et al., 2006).

58 Thanks to their wide distribution, huge species richness, various environmental optima and
59 tolerances and important role in food webs, chironomids have been in the focus of aquatic
60 ecological research for a long time (Porinchu & MacDonald, 2003; Milošević et al., 2013). In
61 addition, chironomids have rapid generation cycle (generally from 3 weeks to few months;
62 Porinchu & MacDonald, 2003) and relatively good dispersal and colonization capacity by
63 their winged terrestrial adults (Armitage, 1995; Čerba et al., 2010). The above mentioned
64 features make them one of the most popular indicators of bioassessment studies at various
65 spatial and temporal scales (Brundin, 1958; Seather, 1979; Brodersen & Anderson, 2002;
66 Gajewski et al., 2005; Milošević et al., 2013) and ideal model organisms for studying
67 colonization patterns and metacommunity organization in dynamic and heterogeneous
68 environment. However, there is a clear gap in our knowledge about the relative importance of
69 dispersal and local environmental filtering processes in the spatial distribution of
70 chironomids, which information is substantial for understanding their ecology and reinforce
71 fundamentals of their bioassessment application.

72 Environmental factors affecting distributions of chironomids at various spatial scales have
73 been extendedly studied in both lotic and lentic habitats (Silver et al., 2000; Mousavi, 2002;

74 Bitušik & Svitok, 2006; Ferrington Jr., 2008; Puntí et al., 2009; Tóth et al., 2012; Jyväsjärvi et
75 al., 2013; Tóth et al., 2013). Nevertheless, the quantitative role of spatial (dispersal) processes
76 in the distribution of chironomids over various spatial scales is hardly known. According to
77 recent theories, the relative role of dispersal processes decreases from broad geographical
78 towards fine microhabitat scale and with the increasing dispersal capacity of organisms
79 (Cottenie, 2005; Beisner et al., 2006; Van de Meutter et al., 2007; Capers et al., 2009).
80 Applying the metacommunity framework, Grönroos et al. (2013) and Heino (2013a,b) argued
81 that the relative role of spatial and local environmental factors in assembly of local
82 communities of aquatic invertebrates depend not only on the spatial scale scanned but also on
83 species traits, especially on dispersal mode. In line with the conclusions of the above studies,
84 it could be supposed that fine-scale (e.g. within lake) spatial distributions of chironomids are
85 decisively determined by local environmental factors and hardly affected by pure spatial
86 processes (i.e. dispersal limitation). However, the metacommunity framework in aquatic
87 insects with winged adults have been applied only at medium to broad spatial scales (i.e.
88 across drainage areas and geographical regions; Puntí et al., 2009; Grönroos et al., 2013;
89 Heino, 2013a,b), whereas it was not tested how spatial and local environmental processes
90 actually contribute to distribution of these organisms at small spatial scale such for instance
91 within a lake.

92 In this study, we examine the relative role of spatial and various local environmental
93 factors in the spatial distribution of benthic chironomid larvae in large and shallow Lake
94 Balaton, Hungary. Previous investigations focused on the largely homogenous open water
95 area of the lake and emphasised the spatio-temporal relationship between the phytoplankton
96 production and population dynamics of the dominant chironomid species (Specziár & Bíró,
97 1998; Specziár & Vörös, 2001). However, considering the extended area of the lake and the
98 high heterogeneity of its littoral and surrounding habitats, we assumed that organization of
99 chironomid assemblages at the lake scale is influenced by various other environmental factors
100 and complex processes likely including dispersal as well. Presumably, dispersal of adults that
101 is largely depend on atmospheric motions (Armitage, 1995; Heino, 2013a,b) is not
102 homogeneous over the lake area, and also dispersal capacity (i.e. colonization distance and
103 density) may vary among species (Armitage, 1995).

104 Accordingly, we hypothesised that both spatial and local environmental factors have
105 significant effect on the distribution of chironomids within Lake Balaton and the relative role
106 of these two processes is species specific. Moreover, if spatial and local environmental
107 processes present simultaneously, it is also important to apply the metacommunity framework

108 when deriving species indicator values based on species-environment data for bioassessment
109 applications. To assess these issues, we performed an extensive sampling covering entire
110 ranges of presumed habitat types and environmental gradients over the lake within a short
111 period of time (i.e. three weeks of sampling) and used a series of uni- and multivariate
112 statistical tools to identify spatial and local biotic and abiotic components of assemblage and
113 taxon level variability in chironomid data. Specific aims of this study are to analyse: (i) what
114 relative importance spatial and local environmental factors have in structuring local
115 chironomid communities; (ii) what environmental gradients are influential in structuring
116 chironomid metacommunities; (iii) what groups of microhabitats (i.e. functional habitats) and
117 related indicator taxa may be identified based on taxon-environmental relationships if spatial
118 processes are also accounted for; and (iv) what optima and tolerances characterize the
119 abundant taxa regarding the most influential environmental factors in Lake Balaton.

120

121 **Material and methods**

122

123 Study area and sampling sites

124 Balaton is the largest shallow lake (surface area: 596 km²; mean depth: 3.2 m) in Central
125 Europe, situated at 46° 42' - 47° 04' N, 17° 15' - 18° 10' E and 104.8 m above sea level. This
126 slightly alkaline (400 mg l⁻¹ of Ca²⁺ and Mg²⁺(HCO₃⁻)₂) lake can be divided into four basins
127 (Keszthely, Szigliget, Szemes, Siófok) with slightly increasing mean depth (from 2.3 to 3.7
128 m) and decreasing mean planktonic chlorophyll-*a* concentration (from 26.6 to 9.7 µg l⁻¹, mean
129 data of 2008-2012; Ministry of Environmental Protection and Water Management of
130 Hungary, http://www.ktm.hu/balaton/lang_en/index.htm) from the major inflow, River Zala
131 (Keszthely basin) toward the only outflow, Sió Canal (Siófok basin; Istvánovics et al., 2007).
132 A majority of the lake area (>85%) is largely homogeneous open water with soft, silt
133 sediment, little variable physico-chemical features and with no macrovegetation. The littoral
134 zone is more heterogeneous (for ranges of some environmental variables see Appendix A). At
135 present, only 47% of the lake shore is in a natural or semi-natural state and these sections are
136 covered by reed grass *Phragmites australis*, dispersedly supplemented with small patches of
137 lesser bulrush *Typha angustifolia* and common club-rush *Schoenoplectus lacustris*.
138 Submerged (most abundant species: perfoliate pondweed *Potamogeton perfoliatus*, sago
139 pondweed *Potamogeton pectinatus*, Eurasian watermilfoil *Myriophyllum spicatum* and rigid
140 hornwort *Ceratophyllum demersum*) and floating leaved (e.g. yellow water-lily *Nuphar lutea*)
141 macrophytes occur sparsely in the littoral zone. Significant part of the lake shore has been

142 stabilized with rocks, and these riprap habitats are covered by filamentous algae (mainly
143 *Cladophora* sp.) up to 0.5 m water depth. There are also several large, commercial boat
144 harbours sheltered by ripraps from swash and many small harbours established within the
145 reed grass stand for anglers. Since northern winds dominate in the area, northern and southern
146 littorals are markedly different; the former has dominantly soft, silt and the latter hard, sandy
147 sediment.

148 In order to capture the longest possible length of the major environmental gradients, we
149 sampled 128 sites distributed across the four basins and characteristic habitat types of the lake
150 (Fig. 1), between 26. June and 13. July 2012. Examined habitats included offshore area (along
151 the longitudinal axis of the lake), natural-like littoral habitat transects from the riparian edge
152 of the reed grass stand towards the macrophyte-free inshore area (typically one to three sites
153 within the reed grass stand, one in the edge of the reed grass stand and the open water, and at
154 50 and 200 m distances from the reed grass stand; Fig. 1), small boat harbours situated within
155 the reed grass stand, stands of the most characteristic submerged and floating leaved
156 macrophytes, and modified littoral areas, large sailing-vessel and ship harbours, ripraps and
157 nearby littoral areas. Samplings covered both the northern and southern littorals.

158

159 Chironomid survey

160 At each sampling site, three sediment samples were taken using an Ekman grab and merged
161 for analyses (total sampled area per site: 0.036 m²). Sediment samples were washed through a
162 0.25 mm mesh sieve and transported to the laboratory in a cooling box. Riprap habitats were
163 sampled by cleaning and washing algal coating and sediment from a measured rock surface
164 being equivalent to Ekman grab samples into plastic containers. In the laboratory,
165 chironomids were sorted from sediment samples alive by sugar flotation method (Anderson,
166 1959), euthanized and preserved in 70% ethanol for later identification. Chironomids were
167 cleared by digestion in 10% KOH (potassium hydroxide) and slide-mounted in Euparal[®].
168 Identification was performed to species or the lowest possible taxonomic level according the
169 keys of Bíró (1981), Cranston (1982), Wiederholm (1983), Janecek (1998), Vallenduuk
170 (1999), Sæther et al. (2000) and Vallenduuk & Moller Pillot (2002).

171

172 Local environmental variables

173 We measured a number of local physical-, chemical- and biotic variables (Appendix A) that
174 have been found influencing abundance and assemblage structure of chironomids (e.g. Real et
175 al., 2000; Rae, 2004; Free et al., 2009; Puntí et al., 2009; Tóth et al., 2012). At each sampling

176 site, we recorded water depth, temperature and redox potential of the uppermost sediment
177 layer, and dissolved oxygen, pH and conductivity of the water close to the bottom. Visual
178 estimates of emergent (dominantly reed grass), submerged and floating leaved macrophytes
179 and filamentous algae (*Cladophora* sp.) coverage (%) was made within a circle of 3 m
180 diameter around the origin of chironomid samples and the area of the submerged and floating
181 leaved macrophyte stand was recorded by GPS and calculated by MapSource version 6.16.3.
182 software (Garmin Ltd., www.garmin.com). The substratum of the lake bed was inspected for
183 percent occurrence of clay, silt, sand, rock, mollusc shells and pure reed grass root
184 (characteristic in some degrading reed grass stands), and the sediment was examined for the
185 occurrence of fine and coarse particle decomposing plant material and leaves of reed grass
186 and rated on a six category scale (0-5). Additional sediment samples were taken for laboratory
187 analyses. Chlorophyll-*a* was extracted from the upper 2 cm sediment layer by hot methanol
188 method (Iwamura et al., 1970) and measured spectrophotometrically (Shimadzu UV-1601
189 spectrophotometer). Percentage organic matter content was assessed from dry (at 50 °C for 72
190 hours) samples of the upper 2 cm sediment layer according to the loss-on-ignition method at
191 550 °C for 1 hour (LOI550; Heiri et al., 2001).

192

193 Spatial variables

194 We established two sets of spatial variables (Appendix A). First, to model directional
195 environmental gradients of the lake, position of each sampling site was characterized with
196 lake-scale geographical (LSG) variables including lake basin (i.e. Keszthely-, Szigliget-,
197 Szemes- and Siófok-basins), location along the north-to-south transect of the lake (i.e.
198 northern littoral, offshore and southern littoral), and distances from the closest shore, reed
199 grass stand, floating leaved or submerged macrophyte meadow and open water. Second, to
200 capture remaining spatial pattern in the chironomid data not described by the local
201 environmental and LSG variables, we composed a set of theoretical variables modelling broad
202 to fine scale spatial patterns among sampling sites by performing principal coordinates of
203 neighbour matrices (PCNM) analysis based on GPS coordinates of sampling sites (Borcard et
204 al., 2004; Dray et al., 2006) using Past version 2.17 software (Hammer et al., 2001). This
205 method practically models position of each sampling site relative to all the other sites
206 similarly as they distribute on the map by means of artificial PCNM variables. During this
207 procedure a matrix of Euclidean distances between all pairs of sampling sites was constructed
208 from the GPS coordinates and truncated at a threshold value of 6 km corresponding to the
209 largest minimum distance between neighbouring sites (i.e. offshore to littoral distance).

210 Distances >6 km were replaced with an arbitrary large value of 24 km equal to four times the
211 threshold distance (Borcard & Legendre, 2002). Then, a principal coordinate analysis of the
212 truncated distance matrix was performed and the derived principal coordinate functions with
213 positive eigenvalues (in total 76 functions were obtained) were retained as PCNM variables
214 (Borcard et al., 2004).

215

216 Statistical analyses

217 Aims (i) and (ii): we performed partial direct gradient and partial multiple second degree
218 polynomial regression analyses (MPRA) followed by a variance partitioning approach
219 (Cushman & McGarigal, 2002; Peres-Neto et al., 2006) to evaluate the role of spatial (i.e.
220 LSG and PCNM variables) and local environmental factors in within lake distribution of
221 benthic chironomids at the assemblage and individual taxon levels, respectively.

222 Extremely rare taxa with <0.1% representation in the total abundance or occurring in <5%
223 of the samples were excluded to reduce their disproportionate effect in the multivariate
224 analyses (Legendre & Legendre, 2012). For analyses, chironomid abundance data were
225 $\ln(x+1)$ transformed to improve their normality and reduce heteroscedasticity. Of LSG
226 variables, lake basin and location along the north-to-south transect were treated as categorical
227 factors and re-coded into binary dummy variables (Lepš & Šmilauer, 2003), whereas others
228 were $\ln(x+1)$ transformed. Local environmental variables measured on continuous scales
229 (including variables representing percentage distribution) were transformed depending on
230 their scale of measurement. However, PCNM variables and local environmental variables
231 which scaled categorically were not transformed (see Appendix A).

232 Because a detrended correspondence analysis (DCA) indicated relatively long gradient
233 length (4.11 in standard deviation units) in our data, we chose canonical correspondence
234 analysis (CCA) for further analyses (Lepš & Šmilauer, 2003). Potential explanatory variables
235 (i.e. LSG, PCNM and local environmental variables) were filtered for collinearity at $r>0.7$
236 and subjected to a forward stepwise selection procedure (at $P<0.05$) based on Monte Carlo
237 randomization test with 9,999 unrestricted permutations. This selection resulted in two LSG,
238 13 PCNM and 16 local environmental variables for the final overall CCA model (Appendix
239 A). Finally, a series of CCA and partial CCAs were conducted to partition the effects of
240 significant variable groups on chironomid assemblages (Cushman & McGarigal, 2002). DCA
241 and CCA analyses were performed using CANOCO version 4.5 software (ter Braak &
242 Šmilauer, 2002).

243 At the individual taxon level, we followed the same approach (i.e. variable selection
244 procedure followed by variation partitioning based on the final model) and used the same
245 explanatory variables as described above, but by applying MPRA including pure and
246 quadratic form of each explanatory variable. This type of regression enables modelling of
247 both linear and unimodal responses of organisms along different gradients (Legendre &
248 Legendre, 2012). During the forward stepwise variable selection in MPRA, pure and
249 quadratic forms of each potential explanatory variable were considered as independent
250 variables. We performed regression analyses for the most abundant chironomid taxa occurring
251 in ≥ 25 samples by using STATISTICA 8.0 software (www.statsoft.com).

252 Aim (iii): a k-means cluster analysis was performed to obtain ecologically relevant groups
253 of sites based on their scores along the first three CCA axes by using STATISTICA 8.0
254 software (www.statsoft.com). Clustering sampling sites into four groups gave the best
255 interpretable results. Differences in the environmental conditions between the four sample-
256 groups were analysed by Kruskal-Wallis ANOVA followed by Mann-Whitney pairwise post
257 hoc test. The most characteristic taxa for each sample group were identified with the indicator
258 value method using IndVal 2.0 software (Duf rene & Legendre, 1997;
259 <http://old.biodiversite.wallonie.be/outils/indval/>). This method calculates an indicator value
260 (IV) that may range between 0 and 100 for each taxon by defined groups of samples and tests
261 its significance by Monte Carlo permutations (9,999 permutations in this case).

262 Aim (iv): optima and tolerances of the abundant chironomid taxa regarding the influential
263 environmental factors were assessed by weighted averaging regression method applied widely
264 in ecological (Punt  et al., 2009) and especially in palaeoecological studies (Brodersen &
265 Anderson, 2002) using C2 version 1.7.4 software (Juggins, 2007;
266 <http://www.staff.ncl.ac.uk/staff/stephen.juggins/software/C2Home.htm>). Optima and
267 tolerances were calculated for taxa occurring in ≥ 10 samples and only for environmental
268 factors which measured on continuous scales.

269

270 **Results**

271

272 Chironomidae assemblages

273 Altogether 13,804 specimens of 40 taxa were identified from 3 subfamilies: Tanypodinae (7),
274 Orthocladiinae (4) and Chironominae (29) (Appendix B). The average taxon richness was 8
275 with a range of 2 to 22 taxa per sample. Most abundant taxa were *Cladotanytarsus mancus*

276 gr., *Polypedilum nubeculosum*, *Chironomus balatonicus*, *Chironomus dorsalis*, *Cricotopus*
277 *sylvestris* gr. and *Procladius choreus*.

278

279 Components of assemblage level variance

280 The overall CCA model based on all significant and non-collinear LSG, PCNM and
281 environmental variables explained 61.9% (eigenvalue: 1.546) of the total variance
282 (eigenvalue: 2.496) in the chironomid relative abundance data. First CCA axis explained
283 15.9% of taxon variation and represented gradients of substrate type from hard (i.e. rock and
284 sand) to soft (i.e. silt) and oxygen concentration, as well as captured some broad spatial
285 structure described by PCNM1 and PCNM3 and distance from the shore (Fig. 2). Second
286 CCA axis explained 12.1% of variance in chironomid data and represented a gradient of
287 organic matter; it correlated positively with LOI550, submerged macrophyte coverage and
288 amount of reed leaves on the sediment, and negatively with the oxygen concentration, pH and
289 of LSG variables with distances from the reed-grass stand and shore. Third CCA axis
290 explained 8.3% of variance in chironomid data and emphasized the difference between rock
291 (i.e. riprap habitats) and sandy substrates (i.e. southern littoral zone).

292 Local environmental variables explained 45.4%, LSG variables 12.7% and PCNM
293 variables 29.1% of total variance in chironomid assemblage data (Fig. 3). However, the
294 shared effect of local environmental and spatial (i.e. both LSG and PCNM) variables was
295 relatively high (21.8%) suggesting a marked spatiality in the local environmental conditions
296 as well. Retained LSG variables (i.e. distances from reed-grass stand and shore) modelled
297 variance related to inshore towards offshore patterns, but had little individual effect (2.2%)
298 compared to PCNM (13.5%) and local environmental (23.6%) variables. Local environmental
299 effects could be divided into three components: 1) substrate type (25.0%, including shared
300 effect), 2) plant material (26.7%) and 3) chemical properties (14.3%). The influence of water
301 chemistry on chironomid assemblages was highly overlapped by the effects of substrate type
302 and plant material. Live (i.e. macrophytes) and decomposing plant material had
303 approximately equal influence on the variability of chironomid assemblages.

304

305 Components of individual taxon level variance

306 At the individual taxon level, local environmental condition had generally higher explanatory
307 power in abundance patterns of benthic chironomids than spatial variables (i.e. LSG and
308 PCNM variables; Table 1, Fig. 4). *C. sylvestris* gr., *C. balatonicus*, *C. virescens* and
309 *Microtendipes chloris* agg. proved to be especially sensitive to local environmental factors

310 and were hardly influenced by spatiality. Local environmental factors also captured a large
311 proportion of variance in the abundance of *P. choreus* and *C. mancus* gr., but in high overlap
312 with spatial factors. Spatial processes clearly predominated in the distribution of *C. defectus*,
313 *P. nubeculosum* and *Stictochironomus* sp. and the considerable proportion of purely PCNM
314 related variance indicated the presence of dispersal limitation in these taxa.

315

316 Functional habitats and their indicator taxa

317 Based mainly on the type (i.e. silt, sand and rock) and organic matter content of the substrate,
318 the CCA indicated the separation of sites into four (functional) habitat types and associated
319 chironomid assemblages. This grouping of sites was supported by k-means clustering (Fig. 5)
320 as follows: 1) northern littoral sites and boat harbours with macrophytes; 2) ripraps; 3)
321 offshore and northern littoral sites without macrophytes; and 4) southern littoral sites (for
322 distribution of site groups in the lake see Fig. 1). Environmental conditions and indicator taxa
323 of these habitat types are presented in Tables 2 and 3, respectively. Briefly, group 1 sites were
324 mainly covered by macrophytes, had silt sediment with high organic matter content, low to
325 medium oxygen concentration and redox potential. These sites were indicated by ten taxa
326 including *Cladopelma virescens*, *C. balatonicus*, *Tanypus kraatzi*, *C. dorsalis* and
327 *Paratanytarsus* sp. Group 2 sites represented shallow riprap habitats (rock: 100%) covered by
328 *Cladophora* sp. algae, had high oxygen concentration and redox potential. Indicator taxa for
329 these sites were *Cricotopus reversus*, *C. sylvestris* gr. and *Orthocladius oblidens*. Group 3
330 sites were in the offshore area and in the macrophyte free parts of the northern littoral,
331 generally had silt sediment, moderate organic matter content, medium to high oxygen
332 concentration and low to medium redox potential. Indicator taxa of this site group were *P.*
333 *choreus*, *Microchironomus tener* and *Tanypus punctipennis*. Group 4 sites had sandy
334 sediment, low amount of decomposing organic matter, medium to high oxygen concentration
335 and medium redox potential. Indicator taxa of this site group were *C. mancus* gr.,
336 *Cryptochironomus defectus* and *Stictochironomus* sp.

337

338 Optima and tolerances of chironomid taxa

339 The optima and tolerances of taxa occurred in >10 samples are shown for some important
340 factors in Fig. 6. Indicator taxa of site group 1, *T. kraatzi*, *Paratanytarsus* sp. and *C. dorsalis*
341 had high optima for silt, LOI550 and emerged and submerged macrophyte coverage, and
342 relatively low for dissolved oxygen and redox potential. It is not surprising that indicator taxa
343 of ripraps (site group 2: *C. reversus*, *C. sylvestris* gr. and *O. oblidens*) and offshore habitats

344 (site group 3: *P. choreus*, *M. tener* and *T. punctipennis*) were found at the opposite ends of
345 some optima diagrams. *C. reversus*, *C. sylvestris* gr. and *O. oblidens* were restricted to
346 shallow water, with high oxygen concentration, redox potential, algae coverage and low
347 percentage of silt, while *P. choreus*, *M. tener* and *T. punctipennis* preferred deep water with
348 silt sediment, moderate oxygen concentration and redox potential and low algae coverage.
349 Indicator taxa of site group 4, *C. mancus* gr., *C. defectus* and *Stictochironomus* sp., preferred
350 shallow water and sandy sediment, had medium optima for redox potential and emerged and
351 submerged macrophyte coverage, and low optima for LOI550 and algae coverage. Regarding
352 pH, *T. kraatzi* had the lowest, whereas *C. mancus* gr. the highest optima. Tolerance values
353 varied considerably across taxa and environmental factors. For instance, regarding water
354 depth, LOI550, algae, emergent and submerged macrophyte coverage highest tolerances were
355 observed in taxa with high optima, whereas regarding silt, taxa with medium optima were the
356 most tolerant.

357

358 **Discussion**

359

360 Relative role of spatial processes and environmental filtering

361 Metacommunity theory assumes that local assemblages are structured by a series of spatial
362 processes related to the migration of species among neighbouring sites and by local
363 environmental control, and the relative role of these two types of processes changes with the
364 spatial scale and varies among taxonomic groups (Leibold et al., 2004; Cottenie, 2005). At
365 small-scale (i.e. within a lake) and in organisms with relatively good dispersal capacity, such
366 as chironomids with winged adults, recent theorems predict overwhelming role of local
367 environmental filtering and consider that spatial processes may have only marginal
368 importance (Leibold et al., 2004; Cottenie, 2005; Beisner et al., 2006; Heino, 2013a,b). Our
369 results show some divergence from these theories. We could prove the predominant role of
370 local environmental filtering at assemblage level and in several individual taxa. However, we
371 also revealed that a substantial part of the non-random spatial patterning in the within lake
372 distribution of chironomids was unrelated to environmental conditions and 13.5% of the total
373 and 21.8% of the explained variance of relative abundance data purely related to PCNM
374 variables. Moreover, in line with the conclusion of other studies (Pandit et al., 2009; Verberk
375 et al., 2010), our results reinforce the inter-specific variability of the relative role of these two
376 type of processes (i.e. spatial and environmental).

377 We consider that LSG variables in a metacommunity context captured both some coarse
378 scale within lake environmental patterns and spatiality related to dispersal. Distances from the
379 shore and emergent macrophyte stand represent coarse patterns of the obvious littoral to
380 offshore habitat gradient with increasing water depth, decreasing macrophyte abundance and
381 sediment organic matter content, etc. On the other hand, distance from the shore correlates
382 with the distance of nearby aquatic habitats serving potential species pools for colonization.
383 Distance from these sites may play a crucial role because it filters the species that are able to
384 reach and colonize particular lake sites (Patrick & Swan, 2011). However, LSG variables
385 purely explained only minor proportion (2.2%) of the total variance in the relative abundance
386 data and most of the variance they captured was also explained by local environmental and
387 PCNM variables.

388 Pure spatial (PCNM related) pattern in metacommunities is a strong indicator of uneven
389 dispersal of organisms among sites (Borcard et al., 2004; Cottenie, 2005). Chironomids with
390 winged adults are considered as medium to good dispersers, although their dispersal over
391 longer distances depends on atmospherical motions and dispersal potential of particular
392 species can also be highly variable due to behavioural and habitat factors (Armitage, 1995;
393 Heino, 2013a,b). Specziár and Vörös (2001) and Specziár (2008) showed that *C. balatonicus*,
394 which is an abundant large-bodied species in Lake Balaton, has high dispersal capacity and
395 can very densely colonize extended lake areas within one generation time when food
396 condition (i.e. production of planktonic algae) becomes favourable. However, distributions of
397 other smaller-bodied open water species (*P. choreus*, *T. punctipennis* and *M. tener*) had
398 substantial purely PCNM related variance indicating dispersal limitation even at lake scale.
399 Similarly, littoral taxa proved to be highly variable in their relative sensitivity to spatial and
400 environmental filtering processes. Although inter-specific variability of the influence of
401 spatial processes seems to be evident in chironomids the underlying mechanisms are not
402 ascertained yet. Dispersal capacity of an organism depends on its mobility and abundance and
403 distribution of species pools (Palmer et al., 1996; Shurin et al., 2000; Heino, 2013b). Since
404 adults of open water species may hatch over extended lake area and thus may have larger and
405 more evenly dispersed stock of potential colonizers than species restricted to relatively small
406 and separated microhabitat patches in the littoral of Lake Balaton. Consequently, lesser spatial
407 effect could be supposed in the former group. However, our results did not unequivocally
408 support this hypothesis. Based on observations on variety of aquatic organisms, Pandit et al.
409 (2009) and Verberk et al. (2010) argued that habitat generalists are governed mainly by
410 spatial (dispersal) processes and habitat specialist are governed mainly by environmental

411 control. However, we could not unequivocally classify individual chironomid taxa into such
412 categories and thus could not test this hypothesis.

413 The high ratio of shared effect between environmental and PCNM and LSG variables
414 indicates that environmental conditions have a considerable spatial structure in the lake as
415 well. This environmental patterning includes a littoral to open water gradient and the marked
416 difference between northern and southern littorals. Previous studies also reported a consistent
417 longitudinal gradient in the open water chironomid assemblages related to the abundance
418 trend of planktonic algae (Specziár & Bíró, 1998; Specziár & Vörös, 2001). However, in the
419 present study, the more pronounced effects of the littoral and the littoral to open water
420 gradients likely outcompeted the weak longitudinal trend in the statistical analyses, and
421 neither chlorophyll-*a* concentration, nor lake basin (LSG variable group) were retained in the
422 final model.

423 As within lake distribution of chironomid assemblages is governed by both environmental
424 control and dispersal limitation (e.g. distance among sites, weather conditions, species pool
425 and behaviour of adults; Armitage, 1995), a comprehensive metacommunity framework
426 seems to have better conceptual responsibility than pure niche based models even at this
427 spatial scale (Leibold et al., 2004). However, Balaton is a relatively large lake, and
428 consequently the question arises whether spatial processes still have relevance in smaller
429 lakes (i.e. <10 km² lake area) or ponds. Further challenge could be specifying underlying
430 spatial (dispersal) processes and their relative importance in structuring chironomid
431 metacommunities over various spatial scales. For instance, there is a clear gap in our
432 knowledge to what extent species-specific mobility and drifting by wind contributes to
433 dispersion of chironomid species across different spatial scales and various landscapes, and
434 whether this inter-specific differences could be related to specific morphological, life-history
435 or functional (i.e. food and habitat preference) traits.

436

437 Environmental filters and bioindication value of chironomid assemblages

438 Environmental control is one of the strongest ecological regulator theorems in community
439 ecology (Chase & Leibold, 2003) and explored species-environment relationships also
440 underlie bioassessment applications (Brundin, 1958; Seather, 1979; Brodersen & Anderson,
441 2002). In Lake Balaton, we identified number of relatively long environmental gradients and
442 revealed that the optima and tolerances of individual chironomid taxa distributed quite evenly
443 along them supporting the significance of environmental filtering. Of environmental factors,
444 physical attributes of the sediment and the occurrence of different plant materials (i.e. algae,

445 macrophytes, and their decomposing particles) received high explanatory power, whereas
446 water chemistry had lower individual influence on chironomid assemblages. The effect of
447 chemical factors (i.e. dissolved oxygen concentration, pH and conductivity of the water and
448 redox potential of the sediment) greatly overlapped with the effect of sediment and plants. A
449 substrate gradient from silt to sand and the presence of rock affect the distribution of
450 burrowing and tube building larvae and larvae living on the sediment surface (Wolfram, 1996;
451 Rae, 2004). Moreover, physical characteristic of the sediment usually correlates with other
452 environmental factors affecting chironomid assemblage structure (Ruse, 1994; Rae, 2013). In
453 Lake Balaton, sand and especially rock substrates coincide with swash, low amount of
454 decomposing organic matter and high oxygen concentration. The ratio of silt in the sediment
455 provided one of the best separations of taxa optima. For instance, *T. kraatzi*, *P. varus*, *T.*
456 *punctipennis*, *C. balatonicus* and *P. choreus* showed the highest preference whereas *O.*
457 *oblidens*, *C. reversus*, *C. mancus* gr. and *Stictochironomus* sp. the highest avoidance of silt.
458 Likewise larvae of *T. punctipennis* and *Chironomus* gr. *plumosus*, which group *C. balatonicus*
459 belongs to, were found to associate with muddy substrate in Neusiedler See (Wolfram, 1996).
460 On the other hand, *P. choreus* did not show a clear sediment preference there. The preference
461 of *C. mancus* gr. for sandy sediment is widely approved as well as the two other indicator
462 species of the southern littoral in Lake Balaton, the *C. defectus* and *Stictochironomus* sp. are
463 usually associated with sand or other compact sediments (Wolfram, 1996; Na & Bae, 2010).
464 The particular importance of algae, macrophytes and decomposing plant material in
465 structuring chironomid assemblages is also well known. Plants provide chironomids with
466 substrate and specific environmental conditions, while their decomposing parts offer directly
467 (live tissues or detritus) and indirectly (via bacteria and fungi growing on detritus) food
468 sources (Tokeshi & Pinder, 1985; Papas, 2007; Čerba et al., 2010). *Cladophora* sp. algae
469 coating on the substrate was tolerated only by few of the abundant taxa and among them *C.*
470 *reversus*, *O. oblidens* and *M. chloris* agg. had the highest optima and thus probably represent
471 the highest indicative value for bioassessments. Most abundant taxa in Lake Balaton showed
472 low optima but relatively high tolerance for macrophytes. Dense emergent vegetation was
473 most preferred and tolerated by *T. kratzii* and *C. viridulum*. Whereas *P. varus* and *C. mancus*
474 gr. were two good benthic indicator taxa of dense submerged macrophyte stands in the silty
475 northern and sandy southern littorals, respectively. Present results are in good agreement with
476 other information on the habitat preference of the above taxa as their occurrences are usually
477 associated with algal coating on the substrate surface or with macrophytes (Čerba et al., 2010;
478 Tóth et al., 2012, 2013). Moreover, *Cricotopus* species require plant surfaces free of sediment

479 as increasing amounts of sedimented particles reduces the availability of the periphyton
480 (Čerba et al., 2010; Tarkowska-Kukuryk, 2010) which criterion is best met in swashed riprap
481 habitats in Lake Balaton. The amount of particulate organic matter was found to affect the
482 distribution of chironomids even at very fine microhabitat scale (Ruse, 1994; Ali et al., 2002;
483 Syrovátka et al., 2009). Although they are important food resources, decomposing organic
484 matters usually produce unfavourable chemical environment for most macroinvertebrates in
485 standing waters (Carpenter & Lodge, 1986; Porinchu & MacDonald, 2003; Papas, 2007). The
486 highest LOI550 values were measured at the riparian edge of dense reed-grass stands along
487 the northern littoral, where the dissolved oxygen concentration and the redox potential were
488 the lowest. These specific circumstances were best tolerated by *T. kraatzi* and *Tanytarsus* and
489 *Paratanytarsus* species.

490 Based mainly on physical characteristics of the sediment, and density of macrophytes and
491 decomposing plant materials four site groups and associated chironomid assemblages could
492 be separated. These habitat types and related chironomid assemblages provided ecologically
493 meaningful primary reference conditions for this lake. The four habitat types are (1) northern
494 macrophyted littoral and sheltered boat harbours with silt sediment and high LOI550, (2)
495 ripraps (rocks) with algal coating, (3) open water with silt sediment and low LOI550, and (4)
496 southern littoral with sand sediment and low LOI550. Indicator species analysis also
497 supported this habitat classification. Our results prove the presence of a consequent north to
498 south natural habitat and associated chironomid assemblage gradients in the lake, which is
499 mainly related to the effect of dominant northern winds in the region. However, human
500 induced changes were also obviously recognizable. The building of ripraps to stabilize large
501 parts of the lake shore and to provide sheltered areas for boats, resulted in the establishment
502 of a new habitat type (site group 2) and specific chironomid assemblages (indicator taxa: *C.*
503 *reversus*, *C. sylvestris* gr. and *O. oblidens*). While, benthic chironomid assemblages of the
504 sheltered boat harbours were similar to natural assemblages of the macrophyted northern
505 littoral. The establishment of these two new habitats (i.e. ripraps and boat harbours) also
506 affects consistent patterns of the northern and southern littorals. All ripraps, either locate in
507 the northern or southern littorals, clustered to the habitat group 2. Whereas boat harbours
508 enable for assemblages characterized originally only the northern littoral (group 1) to locate
509 also in the southern littoral (originally belonging exclusively to group 4).

510 Water depth is an important predictor of chironomid assemblage attributes in many studies
511 (e.g. Verneaux & Aleya, 1998; Ali et al., 2002; Luoto, 2012) and depth optima of several
512 species prove to be consequent across broad spatial scales (Luoto, 2012). However, in this

513 study, the water depth tightly correlated with several potential LSG and local environmental
514 factors and thus was excluded from CCA and MPRA analyses. Optima and tolerances of taxa
515 are clearly separated along this gradient in Lake Balaton as well. Most obviously *C.*
516 *balatonicus*, *T. punctipennis*, *M. tener* and *P. choreus* as the most abundant open water
517 species separate from the other taxa along this gradient. However, since most environmental
518 factors scale with water depth (e.g. sediment characteristics, oxygen concentration,
519 macrovegetation, composition and production of benthic algae, temperature and fish
520 assemblages) it is very difficult to disentangle the individual effect of water depth on
521 chironomid and other macroinvertebrate communities (Real et al., 2000; Wilson & Gajewski,
522 2004). Therefore, water depth itself has probably little indicative value in chironomid based
523 bioassessment in shallow lake habitats (Brodersen & Quinlan, 2006; but see Luoto, 2012).

524

525 **Conclusions**

526

527 We found a clear support that within lake local chironomid assemblages are structured
528 according to metacommunity principles and both spatial processes and environmental filtering
529 have strong relevance at fine spatial scale as well. Moreover, assembly of within lake
530 chironomid metacommunities is further complicated by a marked inter-specific variability in
531 the relative role of spatial and environmental processes. Accordingly, we propose that
532 predictive models of species-environment relationships should better take into account pure
533 spatial structuring of local assemblages even at lake scale. Disentangling the underlying
534 reasons of dispersal limitation and its inter-specific variability is clearly one of the most
535 critical problems in this respect. In addition, since the relative role of spatial and
536 environmental processes depends on the spatial scale and taxa examined, we argue that the
537 reliability of predictive bioassessment models can also vary among regions and habitats, if
538 they do not consider spatial processes as well. Future challenges thus also include to explore
539 to what extent disregarded spatial processes may bias conclusions of monitoring studies
540 depending on the assemblage metric (i.e. relative abundance, presence-absence) and
541 taxonomic and/or functional resolution used.

542

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544

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550

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748 Lakes. *Arctic, Antarctic, and Alpine Research* 34: 446–455.
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750 shallow, alkaline lake (Neusiedler See, Austria). *Hydrobiologia* 318: 103–115.

751 **Table 1** Summary results of multiple second degree polynomial regression models for abundant chironomid taxa in Lake Balaton, Hungary. The
 752 relative contribution of spatial and local environmental variable groups to the total variance is shown on Figure 4. Note that pure and quadratic
 753 forms of each explanatory variable entered the forward stepwise variable selection procedure as independent variables. Abbreviations for
 754 explanatory variables are explained in Appendix A.

	No. of PCNM variables, including pure and quadratic forms	Lake-scale geographical variables	Environmental variables in the model	$R^2_{adj.}$	df	F	P
<i>Procladius choreus</i>	2	distr.	(pH) ² , (cond.) ² , silt, (silt) ² , EMC	0.590	8, 119	23.9	<0.001
<i>Tanypus kraatzi</i>	4	-	DO, (DO) ² , (SMC) ² , FMC	0.587	8, 119	23.6	<0.001
<i>Tanypus punctipennis</i>	8	dists.	silt, (LOI550) ² , (SMC) ²	0.542	12, 115	13.5	<0.001
<i>Cricotopus sylvestris</i> gr.	1	-	(RP) ² , cond., sand, (sand) ² , rock, (LOI550) ² , SMC, FMC, (FMC) ²	0.647	10, 117	24.3	<0.001
<i>Chironomus balatonicus</i>	4	-	(pH) ² , (RP) ² , cond., sand, rock, root, LOI550, (LOI550) ² , (FMC) ²	0.712	13, 114	25.2	<0.001
<i>Chironomus dorsalis</i>	3	distr., (distr.) ²	(DO) ² , RP, (RP) ² , silt	0.476	9, 118	13.8	<0.001
<i>Cladopelma virescens</i>	-	-	DO, (DO) ² , FMC, macarea, (macarea) ² , (FOM) ² , COM, (COM) ² , leaves	0.532	9, 118	17.0	<0.001
<i>Cladotanytarsus mancus</i> gr.	1	dists.	sand, (sand) ²	0.561	4, 123	41.6	<0.001
<i>Cryptochironomus defectus</i>	4	-	RP, sand	0.498	6, 121	22.0	<0.001
<i>Dicrotendipes nervosus</i>	4	dists.	cond., (SMC) ²	0.349	7, 120	10.7	<0.001
<i>Microchironomus tener</i>	3	dists.	(silt) ² , algae, LOI550	0.451	7, 120	15.9	<0.001
<i>Microtendipes chloris</i> agg.	-	-	(rock) ² , (root) ² , (algae) ² , COM	0.265	4, 123	12.5	<0.001
<i>Polypedilum nubeculosum</i>	4	(distr.) ² , dists., (dists.) ²	(RP) ² , (LOI550) ²	0.464	9, 118	13.2	<0.001
<i>Stictochironomus</i> sp.	4	dists.	sand, algae	0.567	7, 120	24.8	<0.001

755 PCNM = spatial variables derived from principal coordinate analysis of neighbouring matrix.

756 **Table 2** Median, minimum and maximum values of the main explanatory variables for the four site groups established by k-means clustering
757 from constrained canonical correspondence analysis site scores based on spatial and environmental variables versus relative chironomid
758 abundance data in Lake Balaton, Hungary. Differences in the environmental conditions between the four sample groups were tested by Kruskal-
759 Wallis ANOVA (K-W). Median values without any common letter in their markings are statistically different according to Mann-Whitney
760 pairwise post hoc test ($P < 0.05$). Abbreviations for explanatory variables are explained in Appendix A.

	Group 1 (38 sites)			Group 2 (8 sites)			Group 3 (34 sites)			Group 4 (48 sites)			K-W	
	median	min.	max.	median	min.	max.	median	min.	max.	median	min.	max.	$H_{df=3}$	P
Distr. (m)	1 ^a	0	1000	100 ^b	3	400	245 ^b	1	5500	2 ^a	0	500	49.2	<0.001
Distm. (m)	5 ^a	0	200	5 ^a	1	50	270 ^b	0	5500	15 ^a	0	450	31.2	<0.001
Disto. (m)	20 ^b	0	129	0 ^{ab}	0	50	0 ^a	0	50	0 ^a	0	100	27.6	<0.001
Dists. (m)	38 ^b	0	190	1 ^a	0	1	303 ^d	5	5500	54 ^c	0	380	50.1	<0.001
Depth (m)	0.7 ^c	0.1	2.3	0.1 ^a	0.0	0.2	2.3 ^d	0.3	4.5	0.6 ^b	0.1	2.2	64.1	<0.001
T (°C)	25.4 ^b	18.8	27.4	28.0 ^{bc}	19.7	29.8	22.9 ^a	21.6	27.1	26.5 ^c	19.4	33.3	20.5	<0.001
pH	7.2 ^a	5.6	8.2	7.4 ^{ab}	6.7	7.9	7.8 ^b	6.7	8.4	7.7 ^b	6.5	9.0	25.2	<0.001
DO (mg l ⁻¹)	8.9 ^a	3.6	14.1	15.5 ^b	9.6	17.0	12.9 ^b	11.1	16.1	14.4 ^b	6.7	20.6	67.7	<0.001
RP (mV)	36 ^a	-67	149	319 ^c	289	365	137 ^b	-48	184	90 ^b	-15	279	42.9	<0.001
Cond. (μS cm ⁻¹)	847 ^c	760	914	832 ^{abc}	784	858	841 ^b	806	868	814 ^a	634	864	25.2	<0.001
Clay (%)	0	0	0	0	0	0	0	0	50	0	0	0	2.8	0.429
Silt (%)	100 ^d	20	100	0 ^a	0	0	100 ^c	0	100	10 ^b	0	70	96.9	<0.001
Sand (%)	0 ^a	0	60	0 ^a	0	0	0 ^a	0	0	90 ^b	30	100	114	<0.001
Moll. (%)	0 ^a	0	80	0 ^a	0	0	0 ^b	0	60	0 ^a	0	40	12.4	0.006
Rock (%)	0 ^a	0	0	100 ^b	100	100	0 ^a	0	10	0 ^a	0	20	103.4	<0.001
Root (classes 0-5)	0 ^a	0	3	0 ^{ab}	0	0	0 ^d	0	5	0 ^b	0	5	12.6	0.006
Algae (%)	0 ^a	0	80	50 ^b	30	100	0 ^a	0	0	0 ^a	0	100	45.8	<0.001
EMC (%)	8 ^c	0	100	0 ^{ab}	0	0	0 ^a	0	30	0 ^{bc}	0	80	21.9	<0.001
SMC (%)	0 ^a	0	100	0 ^a	0	0	0 ^a	0	30	0 ^a	0	90	8	0.046
FMC (%)	0	0	80	0	0	0	0	0	0	0	0	30	5	0.175

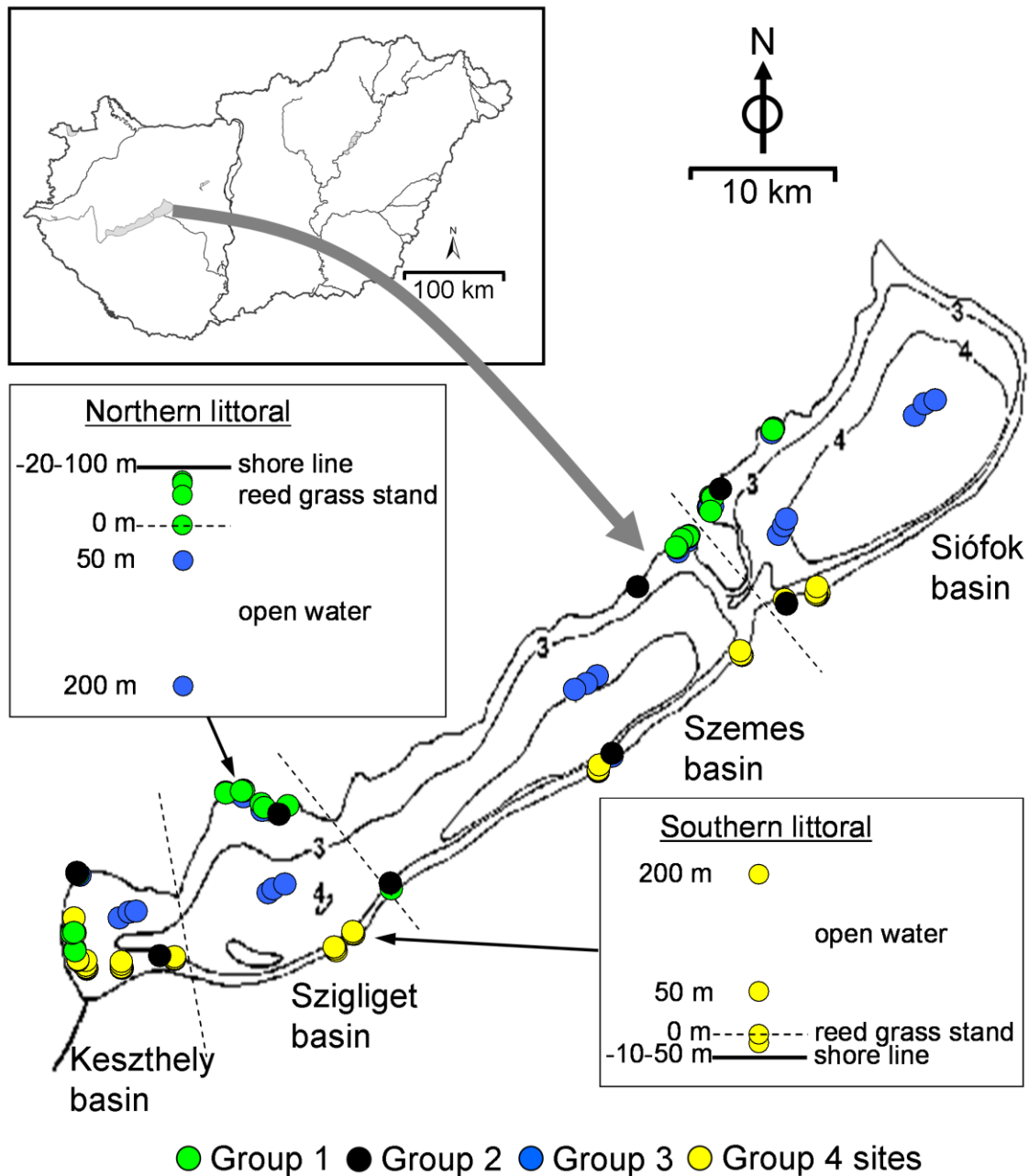
TMC (%)	55 ^c	0	100	0 ^{a0}	0	0	0 ^{a0}	0	60	30 ^b	0	95	46,4	<0.001
Macarea (m ²)	0	0	2000	0	0	0	0	0	500	0	0	5000	10.4	0.016
Chl-a (µg g sediment dwt ⁻¹)	51 ^b	9	1195	427 ^c	99	1078	17 ^a	1	744	17 ^a	2	104	47.9	<0.001
LOI550 (%)	16 ^c	4	81	19 ^c	8	45	8 ^b	5	27	1 ^a	1	26	88.1	<0.001
FOM (classes 0-5)	0 ^c	0	5	0 ^{ab}	0	0	0 ^b	0	1	0 ^a	0	1	24.9	<0.001
COM (classes 0-5)	0 ^b	0	5	0 ^a	0	0	0 ^a	0	2	0 ^a	0	2	23.1	<0.001
Leaves (classes 0-5)	2 ^b	0	5	0 ^a	0	0	0 ^a	0	2	0 ^a	0	5	32.9	<0.001

761

762 **Table 3** Indicator values (IV) of chironomid taxa and their statistical tests (based on 9999
 763 permutations of data) for the four site groups established by k-means clustering from
 764 constrained canonical correspondence analysis site scores based on spatial and environmental
 765 variables versus relative chironomid abundance data in Lake Balaton, Hungary. Group 1:
 766 northern littoral sites and boat harbours with soft sediment and macrophytes; group 2: ripraps;
 767 group 3: offshore and northern littoral sites with soft sediment and without macrophytes; and
 768 group 4: southern littoral sites with sandy sediment (for a more detailed description of
 769 environmental conditions see Table 2).

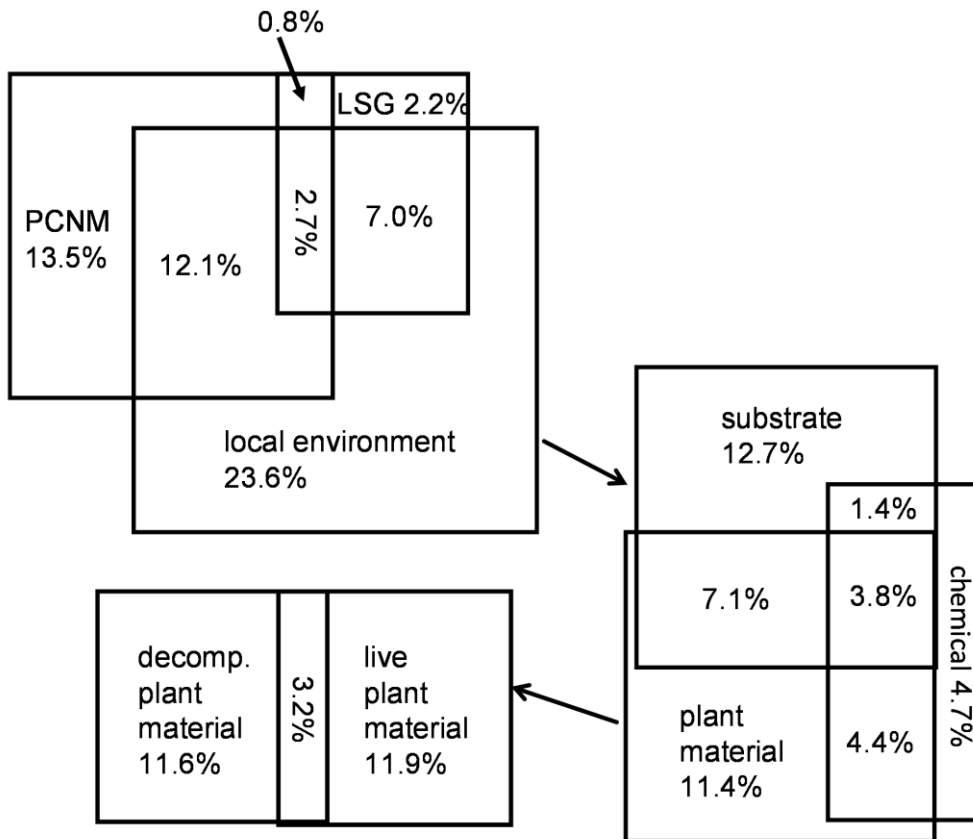
	Taxa	IV	t	Rank	P
Group 1	<i>Cladopelma virescens</i>	62.1	6.09	9	P<0.05
	<i>Chironomus balatonicus</i>	58.8	6.19	4	P<0.05
	<i>Tanypus kraatzi</i>	56.6	8.21	3	P<0.05
	<i>Chironomus dorsalis</i>	44.6	3.42	126	P<0.05
	<i>Paratanytarsus</i> sp.	33.4	3.83	103	P<0.05
	<i>Parachironomus varus</i>	31.4	4.85	40	P<0.05
	<i>Tanytarsus</i> sp.	30.5	3.29	152	P<0.05
	<i>Cladopelma viridulum</i>	21.1	2.27	395	P<0.05
	<i>Guttipelopia guttipennis</i>	21.1	3.58	127	P<0.05
	<i>Dicrotendipes lobiger</i>	18.4	3.06	177	P<0.05
Group 2	<i>Cricotopus reversus</i>	86.2	13.46	1	P<0.05
	<i>Cricotopus sylvestris</i> gr.	85.5	9.72	1	P<0.05
	<i>Orthocladus oblidens</i>	59.4	10.33	1	P<0.05
Group 3	<i>Procladius choreus</i>	68.8	8.71	1	P<0.05
	<i>Microchironomus tener</i>	62.7	7.13	4	P<0.05
	<i>Tanypus punctipennis</i>	38.4	4.00	82	P<0.05
Group 4	<i>Cladotanytarsus mancus</i> gr.	72.0	6.07	4	P<0.05
	<i>Cryptochironomus defectus</i>	42.8	3.84	66	P<0.05
	<i>Stictochironomus</i> sp.	38.1	4.85	41	P<0.05

770 t = the result of a t-test computing the weighted distance between randomized IV values and
 771 the observed value; Rank = the rank of the observed IV value among the decreasing ordered
 772 randomized value distribution (9999 permutations; as the observed value is always added to
 773 the randomized set, the minimum is 1).

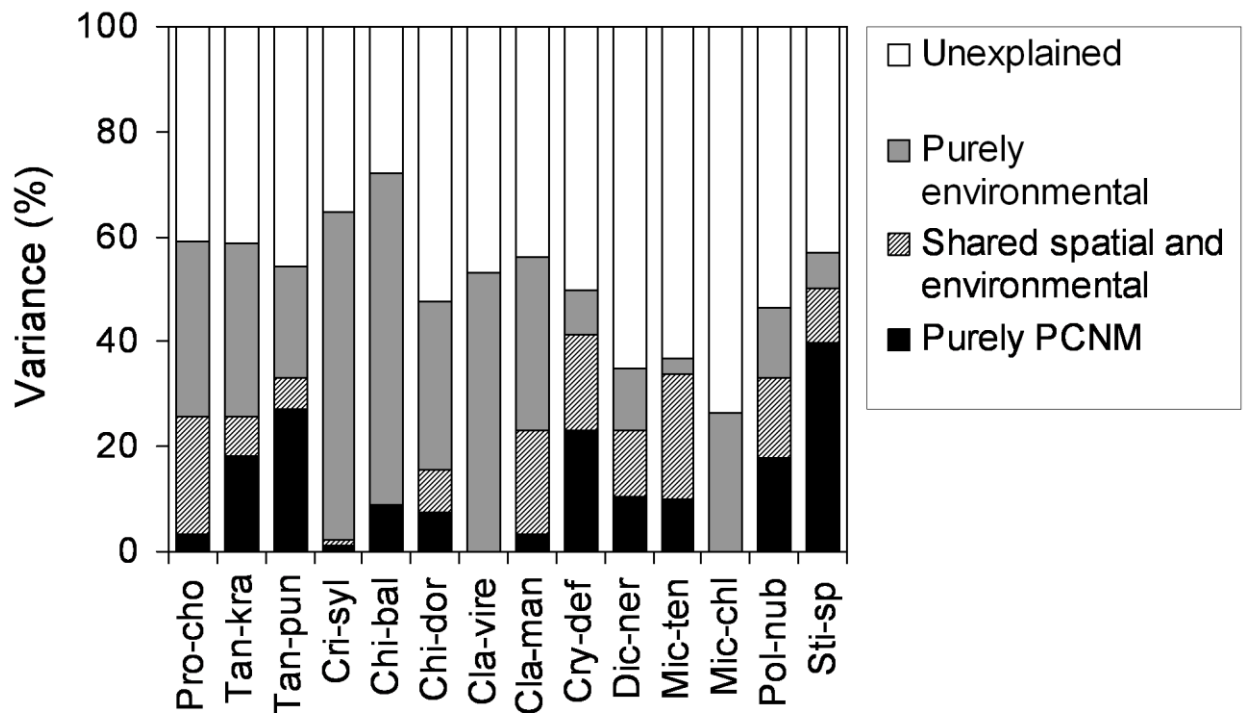


775
 776 **Fig. 1** Distribution of sampling sites in Lake Balaton, Hungary. Sampling sites clustered to
 777 four groups according to the *k*-means method based on their full model canonical
 778 correspondence analysis scores are differentiated by colouring: 1) northern littoral sites and
 779 boat harbours with soft sediment and macrophytes; 2) ripraps; 3) offshore and northern littoral
 780 sites with soft sediment and without macrophytes; and 4) southern littoral sites with sandy
 781 sediment. The small map of Hungary in the upper left corner indicates the location of Lake
 782 Balaton.
 783

791 brackets) by axes are indicated. Chironomid taxa and explanatory variables with scores close
 792 to the centre of the graph are clarified on the small graph in the upper left and right corners.
 793 Explanatory variable and taxa names abbreviations are explained in Appendixes A and B,
 794 respectively.
 795

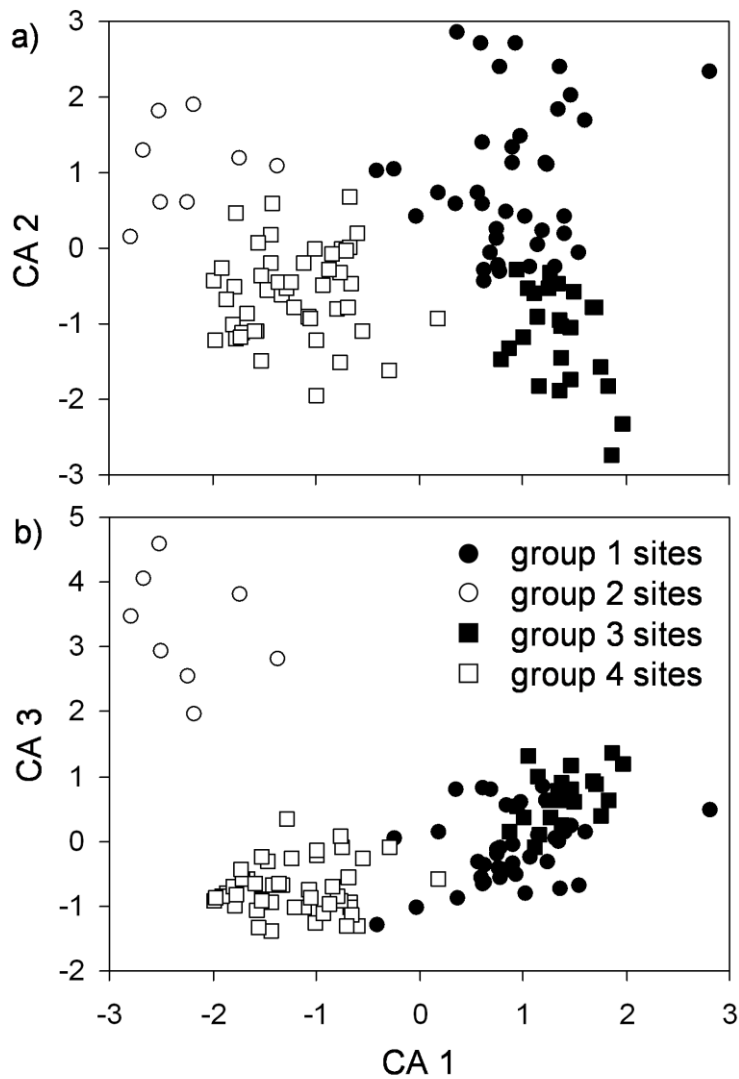


796
 797 **Fig. 3** Result of the variation partitioning of the influence of spatial and local environmental
 798 factors on the abundance of chironomid taxa in Lake Balaton, Hungary. The area of each
 799 rectangular cell is proportional to the variance accounted for by that component. The total
 800 explained variance proportion was 61.9%. The list of significant individual spatial and local
 801 environmental variables and their grouping is presented in Appendix A. PCNM = spatial
 802 variables derived from principal coordinate analysis of neighbouring matrix; LSG = lake-
 803 scale geographical variables; decomp. plant material = decomposing plant material.



804

805 **Fig. 4** Result of the variation partitioning showing the relative influence of spatial and local
 806 environmental factors on the abundance of individual chironomid taxa in Lake Balaton,
 807 Hungary. The effect of lake-scale geographical (LSG) variables is included in shared spatial
 808 and environmental variance proportion (see discussion). Taxa names abbreviations are
 809 explained in Appendix B. PCNM = spatial variables derived from principal coordinate
 810 analysis of neighbouring matrix.



811
 812 **Fig. 5** Canonical correspondence analysis plot of sampling sites along the first and second (a)
 813 and first and third (b) canonical axes (CA) describing the relationship between the abundance
 814 data of chironomid taxa and forward selected (at $P < 0.05$) spatial and local environmental
 815 variables in Lake Balaton, Hungary. Sampling sites were clustered to four groups by k -means
 816 method as follows: 1) northern littoral sites and boat harbours with soft sediment and
 817 macrophytes; 2) ripraps; 3) offshore and northern littoral sites with soft sediment and without
 818 macrophytes; and 4) southern littoral sites with sandy sediment. Environmental characteristics
 819 and indicator taxa of the site groups are presented in Table 2 and 3, respectively.

821 **Fig. 6** Optima and tolerances of individual chironomid taxa regarding some influential
822 environmental factors (a: water depth; b: pH; c: dissolved oxygen in the water; d: redox
823 potential of the sediment surface; e: percent silt in the sediment; f: loss-on-ignition of the
824 sediment – LOI550; g: algae coverage; h: percent emerged macrophyte coverage – EMC; i:
825 percent submerged macrophyte coverage - SMC) in Lake Balaton, Hungary. Taxa names
826 abbreviations are explained in Appendix B. Note that estimated tolerance ranges were cut at
827 the edge of the studied ranges of particular gradients.
828