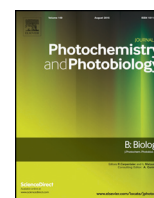




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journal homepage: [www.elsevier.com/locate/jphotobiol](http://www.elsevier.com/locate/jphotobiol)Alkyl and aryl sulfonyl *p*-pyridine ethanone oximes are efficient DNA photo-cleavage agentsNicolaos-Panagiotis Andreou<sup>a</sup>, Konstantinos Dafnopoulos<sup>a</sup>, Christos Tortopidis<sup>a</sup>, Alexandros E. Koumbis<sup>b</sup>, Maria Koffa<sup>c</sup>, George Psomas<sup>d</sup>, Konstantina C. Fylaktakidou<sup>a,\*</sup><sup>a</sup> Laboratory of Organic, Bioorganic and Natural Product Chemistry, Molecular Biology and Genetics Department, Democritus University of Thrace, University Campus, Dragana, 68100 Alexandroupolis, Greece<sup>b</sup> Laboratory of Organic Chemistry, Chemistry Department, Aristotle University of Thessaloniki, 54124 Thessaloniki, Greece<sup>c</sup> Laboratory of Cellular Biology and Cell Cycle, Molecular Biology and Genetics Department, Democritus University of Thrace, University Campus, Dragana, 68100 Alexandroupolis, Greece<sup>d</sup> Laboratory of Inorganic Chemistry, Chemistry Department, Aristotle University of Thessaloniki, 54124 Thessaloniki, Greece

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## ABSTRACT

Sulfonyloxyl radicals, readily generated upon UV irradiation of *p*-pyridine sulfonyl ethanone oxime derivatives, effectively cleave DNA, in a pH independent manner, and under either aerobic or anaerobic conditions. *p*-Pyridine sulfonyl ethanone oxime derivatives were synthesized from the reaction of *p*-pyridine ethanone oxime with the corresponding sulfonyl chlorides in good to excellent yields. All compounds, at a concentration of 100 μM, were irradiated at 312 nm for 15 min, after incubation with supercoiled circular pBluescript KS II DNA and resulted in extended single- and double- strand cleavages. The cleavage ability was found to be concentration dependent, with some derivatives exhibiting activity even at nanomolar levels. Besides that, *p*-pyridine sulfonyl ethanone oxime derivatives showed good affinity to DNA, as it was observed with UV interaction and viscosity experiments with CT DNA and competitive studies with ethidium bromide. The compounds interact to CT DNA probably by non-classical intercalation (i.e. groove-binding) and at a second step they may intercalate within the DNA base pairs. The fluorescence emission spectra of pre-treated EB-DNA exhibited a significant or moderate quenching. Comparing with the known aryl carbonyloxyl radicals the sulfonyloxyl ones are more powerful, with both aryl and alkyl sulfonyl substituted derivatives to exhibit DNA photo-cleaving ability, in significantly lower concentrations. These properties may serve in the discovery of new leads for “on demand” biotechnological and medical applications.

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## 1. Introduction

Light as an “on-off trigger” has long been used as a tool to promote and control reactions. It offers distinct advantages in several fields of chemistry, including biomedicine, where it has been employed in “on demand” therapeutic applications with profound benefits. Among others, cancer treatment has been realized using photodynamic therapy [1], combined in several cases with nanotechnological approaches [2].

Small synthetic organic molecules have been recognized as tools for targeting a variety of DNA structural features, aiming to cause cancer cell death [3]. Among DNA cleaving agents, those causing oxidative cleavage upon UV light activation are called “photo-cleavers” [4]. *O*-Acyl oximes represent one such class of compounds [5–7], which yield photo-generated carbonyloxyl radicals (CRs) capable of causing oxidative DNA damage. However, their use is so far mainly limited to aryl

conjugates, since the alkyl ones suffer rapid decarboxylation and produce less drastic radicals [8].

Sulfonyl ketoximes (SKs) are useful organic precursors, which, upon the Neber [9] or Beckmann [10] rearrangement reactions, provide access to medicinally interesting scaffolds [11]. However, very limited studies have been performed for the evaluation of the biological activity of SKs themselves [12]. In the field of photosensitive polymeric systems, SKs have found applications in microlithography, polymer, material and computer sciences [13], and are considered photo-acid generators. The production of the corresponding UV photo-generated sulfonyloxyl radicals (SRs), which abstract a hydrogen atom to form sulfonic acid derivatives, has been well documented [13].

In continuation of our interest in the chemistry and biology of oximes [7,14–17], we have decided to investigate the photo-activating behavior of SKs towards plasmid DNA. *p*-Pyridine ethanone oxime was selected as the model carrier scaffold of the sulfonyl moiety in order to facilitate comparison of the obtained results with the ones previously reported for the *p*-pyridine oxime and amidoxime ester conjugates [7]. Additionally, after evaluating the derivative which exhibits the best

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cleaving activity, the *p*-nitro-phenyl ester conjugate of *p*-pyridine ethanone oxime was also synthesized, allowing, thus, the direct reactivity comparison between sulfonyloxy and carbonyloxy radicals. A variety of both alkyl and aryl sulfonyl conjugates were prepared to further perform structure activity relationship (SAR) studies. Nevertheless, since SKs have the  $S_0 \rightarrow S_1$  and  $S_0 \rightarrow T_1$  transition states localized on the oxime moiety [18], similar activity is expected to be observed with other SKs as well, as long as they absorb at least partially at the wavelength of irradiation, and have some kind of affinity to DNA [3,4].

## 2. Experimental

### 2.1. Materials, Physical Measurements and Instrumentation

All commercially available reagent-grade chemicals and solvents were used without further purification. Dry solvents were prepared by literature methods and stored over molecular sieves. Calf thymus (CT) DNA, ethidium bromide (EB), NaCl and trisodium citrate were purchased from Sigma-Aldrich Co. DNA stock solution was prepared by dilution of CT DNA to buffer (containing 15 mM trisodium citrate and 150 mM NaCl at pH 7.0) followed by 3 day stirring and kept at 4 °C for no longer than two weeks. The stock solution of CT DNA gave a ratio of UV absorbance at 260 and 280 nm ( $A_{260}/A_{280}$ ) of 1.87, indicating that the DNA was sufficiently free of protein contamination [19]. The DNA concentration was determined by the UV absorbance at 260 nm after 1:20 dilution using  $\epsilon = 6600 \text{ M}^{-1} \text{ cm}^{-1}$  [20].

Melting points (mps) were measured on a Kofler hot-stage apparatus or a melting point meter M5000 Krüss, and are uncorrected. FT-IR spectra were obtained in a Perkin-Elmer 1310 spectrometer using potassium bromide pellets. NMR spectra were recorded on an Agilent 500/54 (500 MHz and 125 MHz for  $^1\text{H}$  and  $^{13}\text{C}$  respectively) spectrometer using  $\text{CDCl}_3$ , and/or  $\text{DMSO}-d_6$  as solvent. *J* values are reported in Hz. High resolution mass spectra (HRMS) were recorded on micrOTOF GC-MS QP 5050 Shimadzu single-quadropole mass spectrometer. Mass spectra were determined on a Shimadzu LCMS-2010 EV system under Electrospray Ionization (ESI) conditions. UV-vis spectra were recorded on a Hitachi U-2001 dual beam spectrophotometer. Fluorescence emission spectra were recorded in solution on a Hitachi F-7000 fluorescence spectrophotometer. Viscosity experiments were carried out using an ALPHA L Fungilab rotational viscometer equipped with an 18 mL LCP spindle. Samples containing plasmid DNA were irradiated in a Macrovue 2011 transilluminator LKB BROMMA at 312 nm, T-15.M 90 W, 0.225 W/cm<sup>2</sup>, and 10 cm distance.

All reactions were monitored on commercial available pre-coated TLC plates (layer thickness 0.25 mm) of Kieselgel 60 F<sub>254</sub>. Silica gel Merck 60 (40–60 μm) has been used for column chromatography. Yields were calculated after recrystallization.

### 2.2. General Procedure for the Synthesis of Ethanone Sulfonyl Oximes

A solution of (*E*)-1-(pyridin-4-yl)ethanone oxime 1 [21] (274 mg, 2 mmol) in dry chloroform (15 mL) was cooled to 0 °C under an argon atmosphere. Triethylamine (0.28 mL, 2 mmol) was added, followed by the required sulfonyl chloride (2 mmol) and the mixture was stirred for 1–3 h allowing the temperature to slowly rise to 25 °C. The reaction was monitored by TLC and, upon completion, water (70 mL) was added and the mixture was extracted with dichloromethane (2 × 70 mL). The combined organic extracts were dried ( $\text{Na}_2\text{SO}_4$ ) and concentrated under vacuum (rotary evaporator) to give a residue which was purified using column chromatography and a mixture of hexanes/ethyl acetate as eluent.

The same procedure was applied for the synthesis of the *p*-nitrobenzoyl oxime 15, but, instead of sulfonyl chloride, PNP-benzoyl chloride was used. Reaction time: 1 h.

### 2.3. Data of Sulfonyl Ketoximes 3–14 and Carbonyl Ketoxime 15

#### 2.3.1. (*E*)-1-(pyridin-4-yl)ethanone *O*-propylsulfonyl oxime (3)

White crystals, yield 90%, mp 135–136 °C (ethyl acetate/methanol); IR (KBr): 3045, 2973, 1621, 1372 and 1172 ( $\text{SO}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 μHz,  $\text{CDCl}_3$ )  $\delta$  1.10 (t, *J* = 7.5 Hz, 3H), 1.95 (sext, *J* = 7.6 Hz, 2H), 2.42 (s, 3H), 3.38 (t, *J* = 7.7 Hz, 2H), 7.57 (d, *J* = 6.2 Hz, 2H), 8.72 (d, *J* = 6.1 Hz, 2H);  $^{13}\text{C}$  NMR (125 μHz,  $\text{CDCl}_3$ )  $\delta$  12.9, 13.9, 17.1, 51.1, 120.8, 141.0, 150.5, 162.1; HRMS (ESI) Calc  $\text{C}_{10}\text{H}_{14}\text{N}_2\text{O}_3\text{S}$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 243.0798; found 243.0799.

#### 2.3.2. (*E*)-1-(pyridin-4-yl)ethanone *O*-decylsulfonyl oxime (4)

Beige crystals, yield 58%, mp 60 °C (ethyl acetate/methanol); IR (KBr): 2957, 2923, 2855, 1596, 1364 and 1169 ( $\text{SO}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 μHz,  $\text{CDCl}_3$ )  $\delta$  0.86 (t, *J* = 6.7 Hz, 3H), 1.22–1.35 (m, 12H), 1.45 (quin, *J* = 7.4 Hz, 2H), 1.90 (quin, *J* = 7.7 Hz, 2H), 2.41 (s, 3H), 3.38 (t, *J* = 7.8 Hz, 2H), 7.58 (d, *J* = 6.1 Hz, 2H), 8.71 (d, *J* = 5.9 Hz, 2H);  $^{13}\text{C}$  NMR (125 μHz,  $\text{CDCl}_3$ )  $\delta$  13.9, 14.0, 22.6, 23.2, 28.1, 28.9, 29.1, 29.4, 31.8, 49.5, 120.8, 141.2, 150.4, 162.1; HRMS (ESI) Calc  $\text{C}_{17}\text{H}_{28}\text{N}_2\text{O}_3\text{S}$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 341.1893; found 341.1894.

#### 2.3.3. (*E*)-1-(pyridin-4-yl)ethanone *O*-hexadecylsulfonyl oxime (5)

White crystals, yield 53%, mp 66–68 °C (ethyl acetate/methanol); IR (KBr): 2957, 2921, 2851, 1366 and 1168 ( $\text{S} = \text{O}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 μHz,  $\text{CDCl}_3 + \text{DMSO}-d_6$ )  $\delta$  0.79 (t, *J* = 6.7 Hz, 3H), 1.10–1.30 (m, 24H), 1.38 (quin, *J* = 7.3 Hz, 2H), 1.79 (quin, *J* = 7.6 Hz, 2H), 2.38 (s, 3H), 3.34 (t, *J* = 7.7 Hz, 2H), 7.71 (d, *J* = 5.5 Hz, 2H), 8.70 (d, *J* = 5.7 Hz, 2H);  $^{13}\text{C}$  NMR (125 μHz,  $\text{CDCl}_3 + \text{DMSO}-d_6$ )  $\delta$  13.5, 13.7, 22.8, 27.7, 28.5, 28.8, 28.9, 29.0, 29.13, 29.17, 29.19, 29.20, 29.21, 29.22, 31.5, 49.0, 120.5, 140.7, 150.0, 161.8; HRMS (ESI) Calc  $\text{C}_{23}\text{H}_{40}\text{N}_2\text{O}_3\text{S}$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 425.2832; found 425.2833.

#### 2.3.4. (*E*)-1-(pyridin-4-yl)ethanone *O*-phenylsulfonyl oxime (6)

Beige crystals, yield 93%, mp 156.3 °C (ethyl acetate/ethanol); IR (KBr): 3048, 1592, 1377 and 1195 ( $\text{SO}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 μHz,  $\text{CDCl}_3$ )  $\delta$  2.33 (s, 3H), 7.42 (bd, *J* = 4.6 Hz, 2H), 7.56 (t, *J* = 7.9 Hz, 2H), 7.66 (t, *J* = 8.1 Hz, 1H), 8.02 (d, *J* = 8.1 Hz, 2H), 8.63 (d, *J* = 4.6 Hz, 2H);  $^{13}\text{C}$  NMR (125 μHz,  $\text{CDCl}_3$ )  $\delta$  13.6, 120.7, 128.9, 129.0, 134.2, 135.2, 141.1, 150.3, 161.5; HRMS (ESI) Calc  $\text{C}_{13}\text{H}_{12}\text{N}_2\text{O}_3\text{S}$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 277.0641; found 277.0641.

#### 2.3.5. (*E*)-1-(pyridin-4-yl)ethanone *O*-tosyl oxime (7) [22]

Brown crystals, yield 89%, mp 162.3 °C (ethyl acetate/ethanol); IR (KBr): 1594, 1379 and 1195 ( $\text{SO}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 μHz,  $\text{CDCl}_3$ )  $\delta$  2.35 (s, 3H), 2.44 (s, 3H), 7.37 (d, *J* = 8.2 Hz, 2H), 7.54 (d, *J* = 6.2 Hz, 2H), 7.91 (d, *J* = 8.3 Hz, 2H), 8.67 (d, *J* = 5.9 Hz, 2H);  $^{13}\text{C}$  NMR (125 μHz,  $\text{CDCl}_3$ )  $\delta$  13.6, 21.7, 121.2, 128.9, 129.7, 132.0, 142.6, 145.6, 149.0, 160.8; HRMS (ESI) Calc  $\text{C}_{14}\text{H}_{14}\text{N}_2\text{O}_3\text{S}$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 291.0798; found 291.0798.

#### 2.3.6. (*E*)-1-(pyridin-4-yl)ethanone *O*-[(4-nitrophenyl)sulfonyl] oxime (8)

Beige crystals, yield 90%, mp 117–119 °C (ethyl acetate/ethanol); IR (KBr): 3105, 1531, 1381 and 1193 ( $\text{SO}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 μHz,  $\text{CDCl}_3$ )  $\delta$  2.39 (s, 3H), 7.42 (d, *J* = 5.4 Hz, 2H), 8.24 (d, *J* = 8.5 Hz, 2H), 8.42 (d, *J* = 8.4 Hz, 2H), 8.68 (d, *J* = 5.4 Hz, 2H);  $^{13}\text{C}$  NMR (125 μHz,  $\text{CDCl}_3$ )  $\delta$  14.0, 120.6, 124.2, 130.4, 140.6, 140.9, 150.6, 151.0, 162.9; HRMS (ESI) Calc  $\text{C}_{13}\text{H}_{11}\text{N}_3\text{O}_5\text{S}$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 322.0492; found 322.0493.

#### 2.3.7. (*E*)-1-(pyridin-4-yl)ethanone *O*-[(4-methoxyphenyl)sulfonyl] oxime (9)

White crystals, yield 74%, mp 139–140 °C (ethyl acetate); IR (KBr): 1596, 1376 and 1174 ( $\text{SO}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (500 μHz,  $\text{CDCl}_3$ )  $\delta$  2.34 (s, 3H), 3.88 (s, 3H), 7.02 (d, *J* = 9.0 Hz, 2H), 7.52 (d, *J* = 6.2 Hz, 2H), 7.96 (d, *J* = 9.0 Hz, 2H), 8.68 (d, *J* = 5.8 Hz, 2H);  $^{13}\text{C}$  NMR (125 μHz,  $\text{CDCl}_3$ )  $\delta$  13.6, 55.7, 114.3, 121.1, 126.3, 131.2, 142.4, 149.2, 160.7,

164.4; HRMS (ESI) Calc  $C_{14}H_{14}N_2O_4S$   $[M + H]^+$  307.0747; found 307.0747.

2.3.8. (*E*)-1-(pyridin-4-yl)ethanone *O*-[(4-(trifluoromethyl)phenyl)sulfonyl] oxime (**10**)

Off white crystals, yield 79%, mp 115–116 °C (ethyl acetate/hexanes); IR (KBr): 3103, 3054, 1597, 1318, 1383 and 1196 (SO)  $cm^{-1}$ ;  $^1H$  NMR (500  $\mu$ Hz,  $CDCl_3$ )  $\delta$  2.39 (s, 3H), 7.53 (d,  $J = 3.8$  Hz, 2H), 7.85 (d,  $J = 7.8$  Hz, 2H), 8.17 (d,  $J = 7.8$  Hz, 2H), 8.71 (d,  $J = 3.8$  Hz, 2H);  $^{13}C$  NMR (125  $\mu$ Hz,  $CDCl_3$ )  $\delta$  13.9, 120.7, 122.6 (q,  $J = 272$  Hz), 126.2 (q,  $J = 3.7$  Hz), 129.5, 135.8 (q,  $J = 34$  Hz), 138.8, 140.9, 150.4, 162.4; HRMS (ESI) Calc  $C_{14}H_{11}F_3N_2O_3S$   $[M + H]^+$  345.0515; found 345.0518.

2.3.9. (*E*)-1-(pyridin-4-yl)ethanone *O*-[(4-bromophenyl)sulfonyl] oxime (**11**)

Off white crystals, yield 79%, mp 118–120 °C (ethyl acetate/hexanes); IR (KBr): 3086, 3015, 1569, 1176, 1379 and 1194 (SO)  $cm^{-1}$ ;  $^1H$  NMR (500  $\mu$ Hz,  $CDCl_3$ )  $\delta$  2.37 (s, 3H), 7.52 (d,  $J = 6.0$  Hz, 2H), 7.72 (d,  $J = 8.5$  Hz, 2H), 7.89 (d,  $J = 8.6$  Hz, 2H), 8.69 (d,  $J = 5.3$  Hz, 2H);  $^{13}C$  NMR (125  $\mu$ Hz,  $CDCl_3$ )  $\delta$  13.8, 121.1, 129.8, 130.4, 132.5, 134.1, 142.1, 149.3, 161.6; HRMS (ESI) Calc  $C_{13}H_{11}BrN_2O_3S$   $[M + H]^+$  354.9747; found 354.9747, 356.9726.

2.3.10. (*E*)-1-(pyridin-4-yl)ethanone *O*-[(4-chlorophenyl)sulfonyl] oxime (**12**)

Beige crystals, yield 72%, mp 108.2 °C (ethyl acetate); IR (KBr): 3092, 3013, 1592, 1175, 1382 and 1194 (SO)  $cm^{-1}$ ;  $^1H$  NMR (500  $\mu$ Hz,  $CDCl_3$ )  $\delta$  2.37 (s, 3H), 7.51 (d,  $J = 5.5$  Hz, 2H), 7.55 (d,  $J = 8.5$  Hz, 2H), 7.97 (d,  $J = 8.6$  Hz, 2H), 8.69 (d,  $J = 5.8$  Hz, 2H);  $^{13}C$  NMR (125  $\mu$ Hz,  $CDCl_3$ )  $\delta$  13.8, 121.1, 129.5, 130.4, 133.5, 141.2, 142.1, 149.4, 161.6; HRMS (ESI) Calc  $C_{13}H_{11}ClN_2O_3S$   $[M + H]^+$  311.0252; found 311.0258, 313.0227.

2.3.11. (*E*)-1-(pyridin-4-yl)ethanone *O*-[(2,5-dichlorophenyl)sulfonyl] oxime (**13**)

Beige crystals, yield 41%, mp 106–108 °C (ethyl acetate/ethanol); IR (KBr): 3083, 3060, 1593, 1389 and 1192 (SO)  $cm^{-1}$ ;  $^1H$  NMR (500  $\mu$ Hz,  $DMSO-d_6$ )  $\delta$  2.47 (s, 3H), 7.50 (d,  $J = 6.1$  Hz, 2H), 7.82 (d,  $J = 8.6$  Hz, 1H), 7.89 (dd,  $J = 8.6, 2.6$  Hz, 1H), 8.12 (d,  $J = 2.6$  Hz, 1H), 8.66 (d,  $J = 6.1$  Hz, 2H);  $^{13}C$  NMR (125  $\mu$ Hz,  $DMSO-d_6$ )  $\delta$  14.5, 121.4, 130.8, 132.1, 133.0, 134.4, 134.5, 136.5, 140.6, 150.8, 165.0; HRMS (ESI) Calc  $C_{13}H_{10}Cl_2N_2O_3S$   $[M + H]^+$  344.9862; found 344.9864, 346.9832, 348.9801.

2.3.12. (*E*)-4-[1-(pyridin-4-yl)ethylideneaminoxysulfonyl]benzoxime (**14**)

Beige crystals, yield 54%, mp 108.2 °C (ethyl acetate); IR (KBr): 3097, 3050, 2236, 1596, 1168, 1384 and 1195 (SO)  $cm^{-1}$ ;  $^1H$  NMR (500  $\mu$ Hz,  $CDCl_3 + DMSO-d_6$ )  $\delta$  2.36 (s, 3H), 7.52 (d,  $J = 5.4$  Hz, 2H), 8.05 (d,  $J = 8.3$  Hz, 2H), 8.15 (d,  $J = 8.4$  Hz, 2H), 8.63 (d,  $J = 5.5$  Hz, 2H);  $^{13}C$  NMR (125  $\mu$ Hz,  $CDCl_3 + DMSO-d_6$ )  $\delta$  12.1, 115.4, 115.8, 119.3, 127.7, 131.7, 137.0, 139.0, 148.2, 161.6; HRMS (ESI) Calc  $C_{14}H_{11}N_3O_3S$   $[M + H]^+$  302.0594; found 302.0599.

2.3.13. (*E*)-1-(pyridine-4-yl)ethan-1-one *O*-[(4-nitrobenzoyl)oxime] (**15**)

Off white crystals, yield 89%, mp 205.1 °C (ethyl acetate); IR (KBr): 3110, 3084, 1744, 1595  $cm^{-1}$ ;  $^1H$  NMR (500  $\mu$ Hz,  $CDCl_3$ )  $\delta$  2.55 (s, 3H), 7.70 (d,  $J = 5.8$  Hz, 2H), 8.30 (d,  $J = 8.7$  Hz, 2H), 8.37 (d,  $J = 8.7$  Hz, 2H), 8.75 (d,  $J = 5.8$  Hz, 2H);  $^{13}C$  NMR (125  $\mu$ Hz,  $CDCl_3$ )  $\delta$  14.3, 121.1, 123.9, 130.8, 134.1, 141.8, 150.5, 150.9, 161.6, 162.5; HRMS (ESI) Calc  $C_{14}H_{11}N_3O_4$   $[M + H]^+$  286.0822; found 286.0820.

## 2.4. DNA Cleavage Experiments

### 2.4.1. Cleavage of Supercoiled Circular pBluescript KS II DNA by Sulfonyl Ketoximes

The reaction mixtures (20  $\mu$ L) containing supercoiled circular pBluescript KS II DNA stock solution (Form I, 50  $\mu$ M/base pair, ~500 ng), compounds, and Tris buffer (25  $\mu$ M, pH 6.8) in Pyrex vials were incubated for 30 min at 37 °C, centrifuged, and then irradiated with UV light (312 nm, 90 W) under aerobic conditions at room temperature for 15 min. After addition of the gel-loading buffer (6X Orange DNA Loading Dye 10 mM Tris-HCl (pH 7.6), 0.15% orange G, 0.03% xylene cyanol FF, 60% glycerol, and 60 mM EDTA, by Fermentas), the reaction mixtures were loaded on a 1% agarose gel with ethidium bromide staining. The electrophoresis tank was attached to a power supply at a constant current (65 V for 1 h). The gel was visualized by 312 nm UV transilluminator and photographed by an FB-PBC-34 camera vilber lourmat. Quantitation of DNA-cleaving activities was performed by integration of the optical density as a function of the band area using the program "Image J" available at the site <http://rsb.info.nih.gov/ij/download.html>.

### 2.4.2. Calculation of Single-strand Damage (ss)% and Double-strand Damage (ds)%

ss% and ds% damage was calculated according to the following Eqs. (1) and (2).

$$ss\% = \frac{\text{Form II}}{(\text{Form I} + \text{Form II} + \text{Form III})} \times 100 \quad (1)$$

$$ds\% = \frac{\text{Form III}}{(\text{Form I} + \text{Form II} + \text{Form III})} \times 100 \quad (2)$$

As Form II we consider Form II of each series minus Form II of the irradiated control DNA. As Form I we consider Form I of each series.

## 2.5. Interaction With CT DNA

### 2.5.1. Study With UV Spectroscopy

The interaction of compounds **3** and **8** with CT DNA was studied by UV spectroscopy in order to investigate the possible binding modes to CT DNA and to calculate the binding constants to CT DNA ( $K_b$ ). The UV spectra of the compounds ( $1 \times 10^{-4}$  M) were recorded in the presence of CT DNA at diverse [compound]/[DNA] mixing ratios ( $= r$ ). Control experiments with DMSO were performed and no changes in the spectra of CT DNA were observed.

The binding constant,  $K_b$ , can be obtained by monitoring the changes in the absorbance at the corresponding  $\lambda_{max}$  with increasing concentrations of CT DNA and it is given by the ratio of slope to the y intercept in plots  $[DNA] / (\epsilon_A - \epsilon_f)$  versus  $[DNA]$ , according to the Wolfe–Shimer equation [23]:

$$\frac{[DNA]}{(\epsilon_A - \epsilon_f)} = \frac{[DNA]}{\epsilon_b - \epsilon_f} + \frac{1}{K_b(\epsilon_b - \epsilon_f)} \quad (3)$$

where  $[DNA]$  is the concentration of DNA in base pairs,  $\epsilon_A = A_{obsd} / [compound]$ ,  $\epsilon_f$  = the extinction coefficient for the free compound and  $\epsilon_b$  = the extinction coefficient for the compound in the fully bound form.

### 2.5.2. Viscometry

Viscosity experiments were carried out using an ALPHA L Fungilab rotational viscometer equipped with an 18 mL LCP spindle and the measurements were performed at 100 rpm. The viscosity of DNA ( $[DNA] = 0.1$  mM) in buffer solution (150 mM NaCl and 15 mM trisodium citrate at pH 7.0) was measured in the presence of increasing amounts of the compounds (up to the value of  $r = 0.35$ ). All measurements were

performed at room temperature. The obtained data are presented as  $(\eta / \eta_0)^{1/3}$  versus  $r$ , where  $\eta$  is the viscosity of DNA in the presence of the compound, and  $\eta_0$  is the viscosity of DNA alone in buffer solution.

### 2.5.3. Competitive Studies With EB

The competitive studies of each compound with EB were investigated with fluorescence emission spectroscopy in order to examine whether the compound can displace EB from its DNA–EB complex. The DNA–EB complex was prepared by adding 20  $\mu\text{M}$  EB and 26  $\mu\text{M}$  CT DNA in buffer (150 mM NaCl and 15 mM trisodium citrate at pH 7.0). The possible intercalating effect of the compounds was studied by adding a certain amount of a solution of the compound step by step into a solution of the DNA–EB complex. The influence of the addition of each compound to the DNA–EB complex solution was monitored by recording the variation of fluorescence emission spectra with excitation wavelength at 540 nm.

The Stern–Volmer constant ( $K_{SV}$ ) is used to evaluate the quenching efficiency for each compound according to the Stern–Volmer equation [24]:

$$\frac{I_0}{I} = 1 + K_{SV}[Q] \quad (4)$$

where  $I_0$  and  $I$  are the emission intensities in the absence and the presence of the quencher, respectively,  $[Q]$  is the concentration of the quencher (compounds **3** and **8**);  $K_{SV}$  is obtained from the Stern–Volmer plots by the slope of the diagram  $I_0/I$  versus  $[Q]$ . Subsequently, the quenching constant  $k_q$  may be calculated by the equation:

$$K_{SV} = k_q \tau_0 \quad (5)$$

where  $\tau_0$  = the average fluorescence lifetime of the EB–DNA system.

## 3. Results and Discussion

### 3.1. Chemistry

Photolysis causes homolytic cleavage of the N–O bond of SKs and production of iminyl and sulfonyloxy radicals (Scheme 1). As it has been demonstrated the iminyl radical may abstract a hydrogen atom to give an imine, which is further hydrolyzed to give the corresponding ketone. The same ketone is obtained when the iminyl radical reacts with molecular oxygen. Finally, a third option is its dimerization to a bisalkylidene hydrazine derivative. On the other hand, the SR abstracts a hydrogen atom and gives the corresponding acid [13].

The mixture of oxime sulfonyl esters photo-cleaved material is expected to contain iminyl and sulfonyloxy radicals, which, potentially, are both capable of attacking DNA, under various mechanisms. Since there is no precedent of SRs acting as DNA photo-cleaving agents, we wish to provide data which correspond to the reactivity of SRs themselves. For this reason we have synthesized derivatives **3–14** in order to establish a structure–activity relationship study within the SRs series. Additionally, based on the fact that derivative **8** exhibited the best

activity, we have synthesized its carbonyl analogue (**15**), in order to compare the reactivity of the sulfonyl and the carbonyl radicals of similar origin.

Compounds **3–14** were synthesized in good yields, as a single product, upon the reaction of *p*-pyridine ethanone oxime **1** [21] with the corresponding alkyl or aryl sulfonyl chlorides **2a–l** in dry  $\text{CHCl}_3$ , in the presence of  $\text{Et}_3\text{N}$  and under an argon atmosphere (Scheme 2).

Compound **15** was similarly obtained, using the corresponding benzoyl chloride, Scheme 3.

All these compounds are new, with the exception of **7**, which has been previously reported [22].

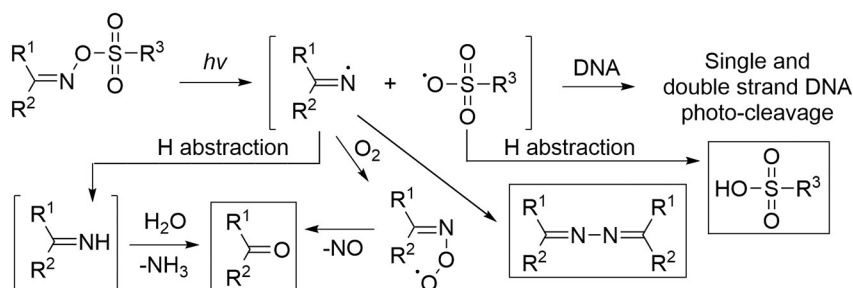
### 3.2. DNA Cleavage Studies

DMF solutions of SKs **3–14** (100  $\mu\text{M}$ ) were mixed with a Tris buffer solution (25  $\mu\text{M}$ , pH = 6.8) containing the supercoiled circular pBluescript KS II DNA (Form I) and this mixture was irradiated with UV light (312 nm) at room temperature for 15 min, under aerobic conditions. Plasmid DNA was analyzed by gel electrophoresis on 1% agarose stained with ethidium bromide, Fig. 1. All experiments were performed at least three times. None of the tested compounds showed any activity towards DNA in the absence of UV irradiation.

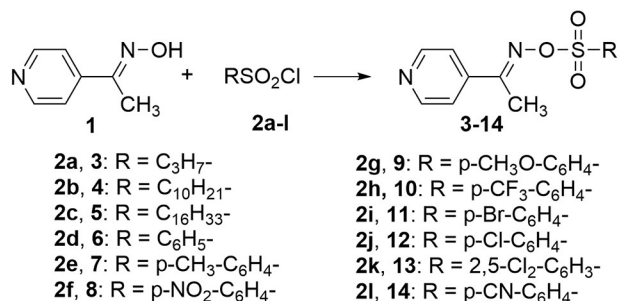
In the presence of compounds **3–14** the supercoiled plasmid DNA (Form I) sustained single-stranded (ss) nicks of the double helix, generating the relaxed circular DNA (Form II) found in all experiments, whereas the linear DNA (Form III) was formed as well, generated by double-stranded (ds) nicks, Fig. 1.

The results presented in Fig. 1 show that most of the aryl SKs are in general considerably reactive. The most important observation is that alkyl SRs are also effective, and, comparing to the aryl ones, they are similarly efficient at the given concentration, contrary to what was observed for the carbonyl analogues [7,8]. As for the aromatic ones, a SAR study revealed that lack of substitution on the aromatic ring (**6**) results in reduced activity. The activity also decreases when the  $\text{CH}_3$  group is replaced with the electron withdrawing  $\text{CF}_3$  group (**7** and **10**, respectively). Additionally, regarding the *p*-halogenated compounds, the bromo-derivative **11** is not as efficient comparing to the chloro-one **12** [4], whereas the dichloro-derivative **13** was the less reactive. Calculation of the energies of the N–O bonds of these SKs indicates that their variation lies within  $\sim 2$  Kcal/mol (45.4–47.7 Kcal/mol).

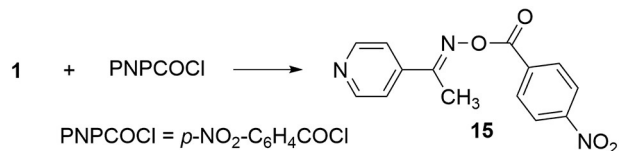
The observations that a) the energy provided by UV irradiation is more than sufficient for the cleavage of all N–O bonds; b) the length of the aliphatic chain is almost irrelevant to the activity; c) the substituents of the aromatic ring with electron donating effect (**7** and **9**) and electron withdrawing effect (**8**) have approximately the same activity (Form II + Form III); and d) all aromatic substituents with electron withdrawing effects have not the same activity (**8** and **10–14**) shows that the variations of activity might also be dependable on affinity DNA interactions of the sulfonyl conjugates. This is in accordance to previously reported conclusions regarding the dependence of DNA cleavage mechanism on the structure of the photo-cleavage agent, primarily because of the geometrical requirements for H-abstraction reactions [4a].



Scheme 1. The fate of radicals generated by the photo-cleavage of the N–O bond of oxime sulfonyl esters.



**Scheme 2.** Synthesis of *p*-pyridine-ethanone oxime sulfonyl esters **3–14**. Reagents and conditions: RSO<sub>2</sub>Cl (1.1 eq.), dry Et<sub>3</sub>N (1.1 eq.), dry CHCl<sub>3</sub>, 0 °C to r.t., 1–3 h, 41–93%.



**Scheme 3.** Synthesis of the *p*-pyridine-ethanone PNP oxime **15**. Reagents and conditions: PNP-COCl (1.1 eq.), dry Et<sub>3</sub>N (1.1 eq.), dry CHCl<sub>3</sub>, 0 °C to r.t., 1 h, 89% (PNP = *p*-NO<sub>2</sub>-C<sub>6</sub>H<sub>4</sub>-).

A high ratio of ds to ss cleavages is a very important characteristic of DNA cleaving ability [25], since ds cleavages are more difficult to be repaired and may trigger self-programmed cell death. The UV generated SRs cause ss and ds cleavages, with the one derived from *p*-pyridine *p*-nitrophenyl sulfonyl ethanone oxime **8** yielding the best ratio of double- to single-strand damages [Fig. 1(A), Lane 8 and Fig. 1(B)]. Moreover, compared to the *p*-pyridine *p*-nitrobenzoyl oxime and amidoxime derivatives, the sulfonyl ones were found to exhibit a superior effect on cleavage: approximately 10-fold lower concentrations were sufficient to lead to the same activity, along with similar ds/ss ratio cleavage (the effective concentration was 100 μM and 1000 μM, respectively) [7]. This is in accordance with previous literature kinetic results where SRs are reported to be more reactive compared to CRs [26]. The role of the *p*-nitro-phenyl conjugate was also found to be important in both types of radicals [4,7,27]. DNA photo cleavage of carbonyl compound

**15** was 86(±5.8)%, including a ~3% of double-strand cleavage, at a concentration of 100 μM [Fig. 2(A), Lane 9]. Although both **8** and **15** completely consume supercoiled plasmid DNA at the given concentration, sulfonyl ester **8** causes many more nicks, which eventually leads to the formation of linear DNA and a high ds/ss ratio.

Furthermore, we have also examined the lowest concentration which is sufficient for the 50% cleavage of the supercoiled plasmid DNA caused by each one of compounds **3–8**. The results at concentrations of 10 and 1 μM are given in Fig. 2(A) and (B), respectively. The diagram with the average cleavage for all experiments is depicted in Fig. 2(C). All three aromatic sulfonyl conjugates exhibit good cleavage ability (~70–80%) at a concentration of 10 μM, whereas the aliphatic ones were found less reactive at both concentrations.

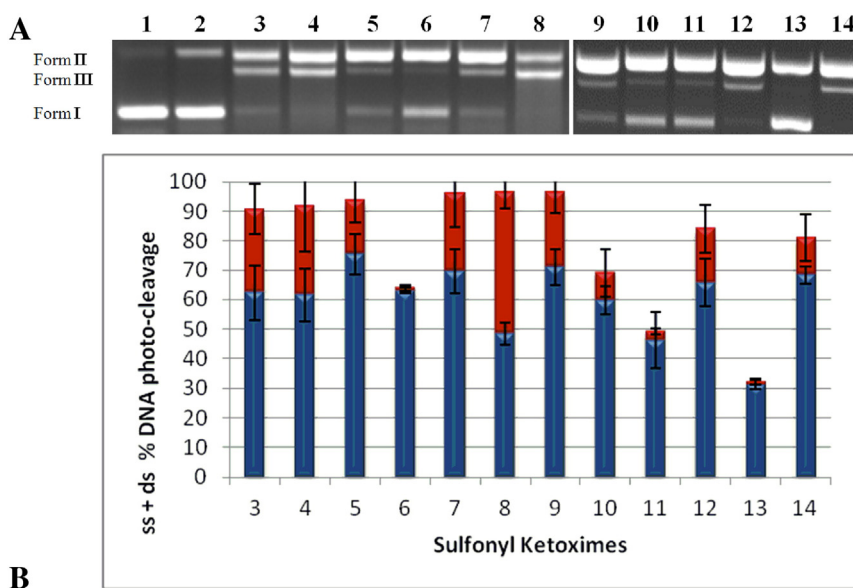
In brief, the cleavage caused by SKs is concentration dependent. Comparing the activity of carbonyl analogue **15** at a concentration of 100 μM with the sulfonyl analogue **8** at a concentration of 10 μM we conclude that the latter has about the same activity in approximately 10-fold lower concentration [86(±5.8)% and 77.5(±1.1)%, respectively], Fig. 2(A) lanes 8 and 9. Compound **8** is effective for 50% DNA photo-cleavage in a concentration of approximately 1 μM. Most importantly, according to these results all compounds are expected to have some activity even in the nanomolar range.

Mechanistic studies performed with compound **8** (Fig. 3, left part) showed that SKs can react under anaerobic conditions (Lane 4) and that the mode of action under aerobic conditions most probably involves hydroxyl radicals and singlet oxygen (reaction with DMSO—Lane 5 and NaN<sub>3</sub>—Lane 6, respectively). Nevertheless, the cleavage of plasmid DNA is not enhanced when D<sub>2</sub>O was used as solvent (Lane 7). These results indicate that not a single mechanistic pathway is implicated in the cleavage. Additional experiments to clarify the behavior of the system are in due course. Finally, a variation of pH (range 5–8) does not affect the photo-cleavage activity (Fig. 2, right part, Lanes 8–11, respectively).

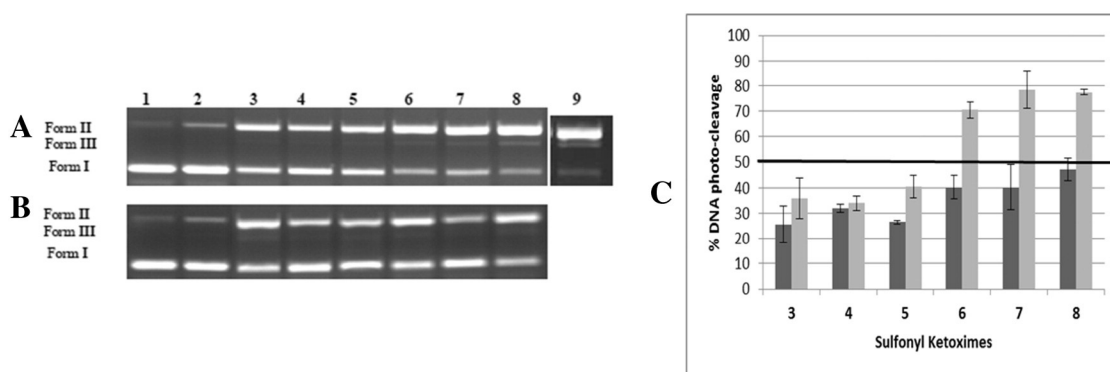
The ability of SKs to act under anaerobic conditions and acidic pH may be very useful in the treatment of solid tumors where hypoxic conditions as well as low acidity dominate [28].

### 3.3. DNA Binding Studies

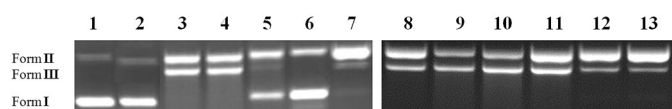
The interaction of SKs with CT DNA, which proves the affinity of SKs with DNA, was examined by UV spectroscopic titrations and DNA



**Fig. 1.** (A) Comparative dose measurement results from DNA cleavage by the SKs derivatives **3–14** at a concentration of 100 μM. Gel electrophoresis data: Lane 1: DNA without UV irradiation; Lane 2: DNA with UV irradiation; Lanes 3–14: DNA + **3–14**, respectively, and with UV irradiation. (B) (ss + ds) % DNA photo-cleavage caused by SKs **3–14** (100 μM). Blue column ss%, red column ds%.



**Fig. 2.** (A) Comparative dose measurement results from DNA cleavage by the SKs derivatives **3–8** at concentration of 10  $\mu\text{M}$ . Gel electrophoresis data: Lane 1: DNA without UV irradiation; Lane 2: DNA with UV irradiation; Lanes 3–14: DNA + **3–8**, respectively, and with UV irradiation; Lane 9: DNA + **15** (100  $\mu\text{M}$ ). (B) Comparative dose measurement results from DNA cleavage by the SKs derivatives **3–8** at concentration of 1  $\mu\text{M}$ . Gel electrophoresis data: Lane 1: DNA without UV irradiation; Lane 2: DNA with UV irradiation; Lanes 3–14: DNA + **3–8**, respectively, and with UV irradiation. (C) % DNA photo-cleavage caused by SKs **3–8** (1  $\mu\text{M}$  and 10  $\mu\text{M}$ , dark gray and light gray, respectively).



**Fig. 3.** Gel electrophoresis data. Left part: Mechanistic studies involved at the DNA cleavage by derivative **8** (100  $\mu\text{M}$ ). Lane 1: DNA without UV irradiation; Lanes 2–13: DNA with UV irradiation; Lane 3: DNA + **8**; Lane 4: DNA + **8** + argon; Lane 5: DNA + **8** + DMSO (20%); Lane 6: DNA + **8** + NaN<sub>3</sub>; Lane 7: DNA + **8** + D<sub>2</sub>O. Right part: Effect of pH. Lanes 8–13: DNA + **8** at pH 5, 6, 7, 8, 9, and 10, respectively.

viscosity measurements and was carried out representatively for the alkyl derivative **3** and the aryl one **8**, which exhibited the best activity. The slight hypochromism (2% for **3** and 3% for **8**) observed in the UV spectra of the compounds in the presence of increasing amounts of CT DNA (Fig. 4) is indicative of the existing interaction.

The binding constant of the compounds with CT DNA ( $K_b$ ) was calculated by the Wolfe–Shimer equation (Eq. (3)) [23] and plots  $\{[DNA] / (\epsilon_A - \epsilon_f)\}$  versus  $[DNA]$  (Insets in Fig. 4). The  $K_b$  value of **3** [ $K_b = 4.82(\pm 0.25) \times 10^5 \text{ M}^{-1}$ ] was much higher than that of **8** [ $K_b = 2.44(\pm 0.10) \times 10^5 \text{ M}^{-1}$ ], suggesting that **3** exhibits higher affinity than **8** for CT DNA. These  $K_b$  values are of the same order to that of classic DNA-intercalator ethidium bromide (EB) ( $K_b = 1.23 \times 10^5 \text{ M}^{-1}$ ) as previously reported by our group [29].

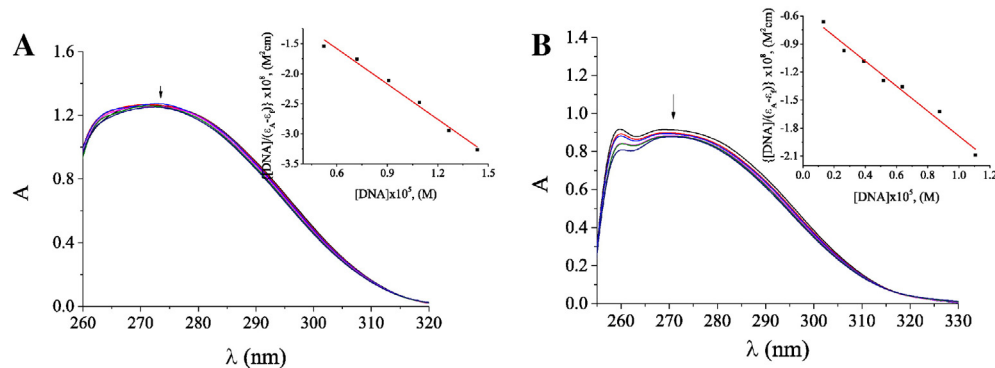
However, it is generally believed that it is not quite safe to propose the exact mode of binding to CT DNA merely from UV spectroscopic titration studies [30]. Thus, the viscosity of a CT DNA solution (0.1 mM) was monitored upon addition of increasing amounts of the compounds **3** and **8** (up to the value of  $r = 0.35$ ) (Fig. 5). Initially and up to  $r$  value of

0.12, the viscosity did not exhibit any appreciable change in the presence of the compounds; for higher concentrations, however, the viscosity presented a significant increase.

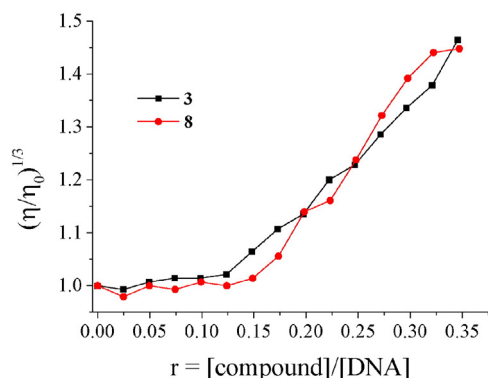
Considering the overall changes of the DNA viscosity in the presence of compounds **3** and **8**, we may suggest that, initially, the compounds interact to CT DNA probably by non-classical intercalation (i.e. groove-binding) and at a second step they may intercalate within the DNA base pairs [31]. This behavior may elucidate the findings from the UV spectroscopic studies.

The ability of the compounds to displace EB from the EB–DNA complex may shed light in their intercalating ability. The competition between compounds **3** and **8** with EB for the intercalation sites of DNA was monitored by fluorescence emission spectroscopy with  $\lambda_{\text{excitation}} = 540 \text{ nm}$ . The fluorescence emission spectra of pre-treated EB–DNA ( $[\text{EB}] = 20 \mu\text{M}$ ,  $[\text{DNA}] = 26 \mu\text{M}$ ) as recorded in the presence of increasing amounts of compounds **3** or **8** [Fig. 6(A) and (B)] exhibit a significant or moderate quenching [60.2% for **3** and 48% for **8**, Fig. 6(C)], respectively. This quenching is due to the ability of the compounds to displace EB from the DNA–EB complex and indirectly reveals that their interaction with CT DNA via an intercalative mode may exist [31].

Furthermore, the observed quenching is in good agreement ( $R = 0.99$ ) with the linear Stern–Volmer equation (Eq. (4)) [24]. The values of the Stern–Volmer constant ( $K_{SV}$ ) as calculated from the Stern–Volmer plots (Insets in Fig. 6) are  $6.07(\pm 0.18) \times 10^4 \text{ M}^{-1}$  for **3** and  $3.72(\pm 0.10) \times 10^4 \text{ M}^{-1}$  for **8**; they may be considered rather high and verify the tight binding of compounds **3** and **8** to DNA [29,32]. Taking  $\tau_0 = 23 \text{ ns}$  as the fluorescence lifetime of EB–DNA system [33], the quenching constants of the compounds as calculated



**Fig. 4.** UV spectra of a DMSO solution ( $1 \times 10^{-4} \text{ M}$ ) of compounds (A) **3** and (B) **8** in the presence of increasing amounts of CT DNA. The arrows show the changes upon increasing amounts of CT DNA. Insets: Plot of  $\frac{[DNA]}{(\epsilon_A - \epsilon_f)}$  versus  $[DNA]$ .



**Fig. 5.** Relative viscosity  $(\eta/\eta_0)^{1/3}$  of CT DNA (0.1 mM) in buffer solution (150 mM NaCl and 15 mM trisodium citrate at pH 7.0) in the presence of compounds **3** and **8** at increasing amounts ( $r = [\text{compound}]/[\text{DNA}]$ ).

with Eq. (5) are  $k_{q(3)} = 2.64(\pm 0.08) \times 10^{12} \text{ M}^{-1} \text{ s}^{-1}$  and  $k_{q(8)} = 1.62(\pm 0.04) \times 10^{12} \text{ M}^{-1} \text{ s}^{-1}$ ; the values of the  $k_q$  constants are significantly higher than  $10^{10} \text{ M}^{-1} \text{ s}^{-1}$  suggesting, thus, the quenching via a static mechanism [34].

A current study from our group, concerning the photo-cleavage of sulfonyl amidoximes [35], show results which are in general agreement with the observations of the present. However, SKs presented herein

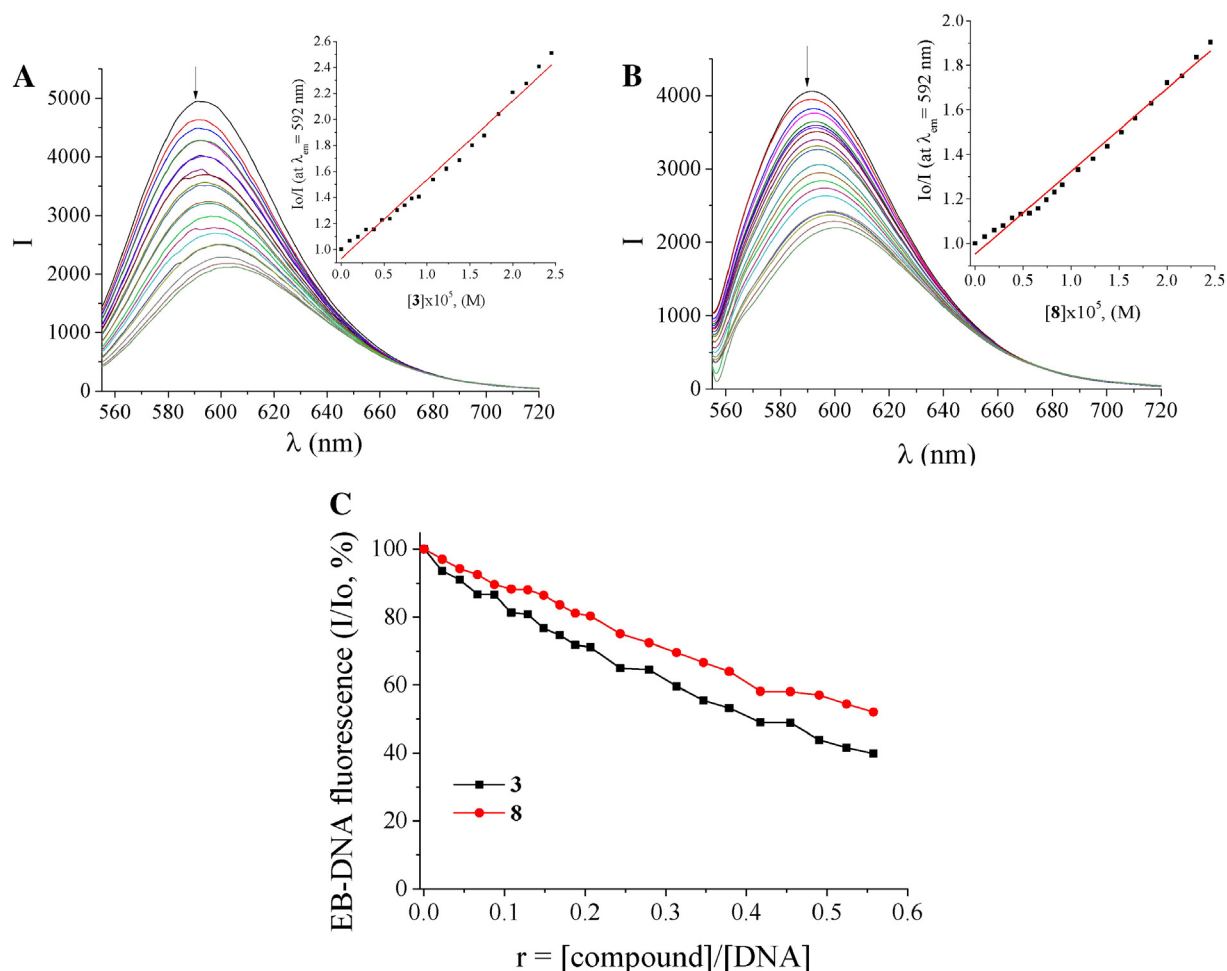
show better activity. The reasons may be that, at least in comparable derivatives, SKs show lower N–O bond energy ( $\sim 5 \text{ Kcal/mol}$ ), as well as better intercalation as it was measured in binding affinity studies.

#### 4. Conclusions

We have shown that SRs are effective DNA photo-cleaving agents. The herein provided experimental results evident a superiority of SRs compared to the known CRs as DNA photo-cleavers. SK **8** had similar ratio of ds to ss cleavage, in an approximately 10-fold lower concentration than a comparable oxime ester.

Additionally, another factor differentiating SRs from CRs, is that alkyl SRs are very potent DNA photo-cleavers. This property was not so far observed for the carbonyl derivatives. SAR and DNA binding affinity studies within the 4-pyridine SK series revealed that the aliphatic SK **3** exhibited better affinity to DNA than the aromatic SK **8**, but approximately the same total photo-cleavage ability, although with a lower ratio of ds to ss cleavage.

Sulfonyl ketoximes are organic compounds easily accessible in large scale from ketone derived oximes, and are stable in the absence of light for prolonged periods of time. The DNA cleavage is concentration dependent and occurs simply by irradiation without the need of external additives. 4-Pyridine ethanone oxime has been used as a model compound and it proved the efficacy of the SRs as novel DNA photo-cleavage agents, in concentrations close to the nanomolar scale.



**Fig. 6.** Emission spectra ( $\lambda_{\text{excit}} = 540 \text{ nm}$ ) for EB-DNA ( $[\text{EB}] = 20 \mu\text{M}$ ,  $[\text{DNA}] = 26 \mu\text{M}$ ) in buffer solution in the absence and presence of increasing amounts of (A) compound **3** and (B) compound **8** (up to the value of  $r = 0.06$ ). The arrows show the changes of intensity upon increasing amounts of the compounds. Insets: Stern–Volmer quenching plot of EB bound to CT DNA for (A) **3** (equation of linear fitting:  $y = 6.07(\pm 0.18) \times 10^4 \times x + 0.93$  ( $r = 0.993$ ,  $R^2 = 0.985$ )) and (B) **8** (equation of linear fitting:  $y = 3.72(\pm 0.10) \times 10^4 \times x + 0.95$  ( $r = 0.994$ ,  $R^2 = 0.987$ )). (C) Plot of EB relative fluorescence emission intensity at  $\lambda = 592 \text{ nm}$  ( $I/I_0$ , %) versus  $r$  ( $r = [\text{compound}]/[\text{DNA}]$ ) (150 mM NaCl and 15 mM trisodium citrate at pH = 7.0) in the presence of compound **3** and **8** (up to 39.8% of the initial EB-DNA fluorescence intensity for **3** and 52% for **8**).

The possibility of introducing a variety of properly designed SKs which, depending on their individual DNA interactions, could yield highly reactive SRs, may be fruitful in the fields of pharmacy, molecular biology and medicine. Thus, enriching the pool of possible applications with more powerful weapons, capable of acting at various pHs and under aerobic or anaerobic conditions, may provide novel biotechnological and therapeutic approaches for anticancer and antimicrobial therapies.

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