




Review

# Applied Nanotechnologies in Anticoagulant Therapy: From Anticoagulants to Coagulation Test Performance of Drug Delivery Systems

Yuri B. G. Patriota<sup>1</sup>, Luíse L. Chaves<sup>1</sup>, Evren H. Gocke<sup>2</sup>, Patricia Severino<sup>3,4,5</sup> , Mônica F. R. Soares<sup>1</sup>, José L. Soares-Sobrinho<sup>1,\*</sup> and Eliana B. Souto<sup>6,7,\*</sup>

- <sup>1</sup> Quality Control Core of Medicines and Correlates, Department of Pharmaceutical Sciences, Federal University of Pernambuco, Professor Arthur de Sá Avenue, Recife, PE 50740-521, Brazil; yuribgp@hotmail.com (Y.B.G.P.); luise@fps.edu.br (L.L.C.); monica.soares@ufpe.br (M.F.R.S.)
- <sup>2</sup> Department of Pharmaceutical Technology, Faculty of Pharmacy, Ege University, Bornova-Izmir 35100, Turkey; evrenhomangokce@gmail.com
- <sup>3</sup> Laboratory of Nanotechnology and Nanomedicine (LNMed), Institute of Technology and Research (ITP), Av. Murilo Dantas, 300, Aracaju, SE 49010-390, Brazil; pattypharma@gmail.com
- <sup>4</sup> Institute of Technology and Research (ITP), University of Tiradentes (Unit), Av. Murilo Dantas, 300, Aracaju, SE 49010-390, Brazil
- <sup>5</sup> Tiradentes Institute, 150 Mt Vernon St, Dorchester, MA 02125, USA
- <sup>6</sup> Faculty of Pharmacy, University of Coimbra, Pólo das Ciências da Saúde, Azinhaga de Santa Comba, 3000-548 Coimbra, Portugal
- <sup>7</sup> CEB-Centre of Biological Engineering, Campus de Gualtar, University of Minho, 4710-057 Braga, Portugal
- \* Correspondence: jose.ssobrinho@ufpe.br (J.L.S.-S.); ebsouto@ff.uc.pt (E.B.S.)



**Citation:** Patriota, Y.B.G.; Chaves, L.L.; Gocke, E.H.; Severino, P.; Soares, M.F.R.; Soares-Sobrinho, J.L.; Souto, E.B. Applied Nanotechnologies in Anticoagulant Therapy: From Anticoagulants to Coagulation Test Performance of Drug Delivery Systems. *Appl. Nano* **2021**, *2*, 98–117. <https://doi.org/10.3390/applnano2020009>

Academic Editor: Angelo Maria Taglietti

Received: 9 March 2021  
Accepted: 22 April 2021  
Published: 1 May 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

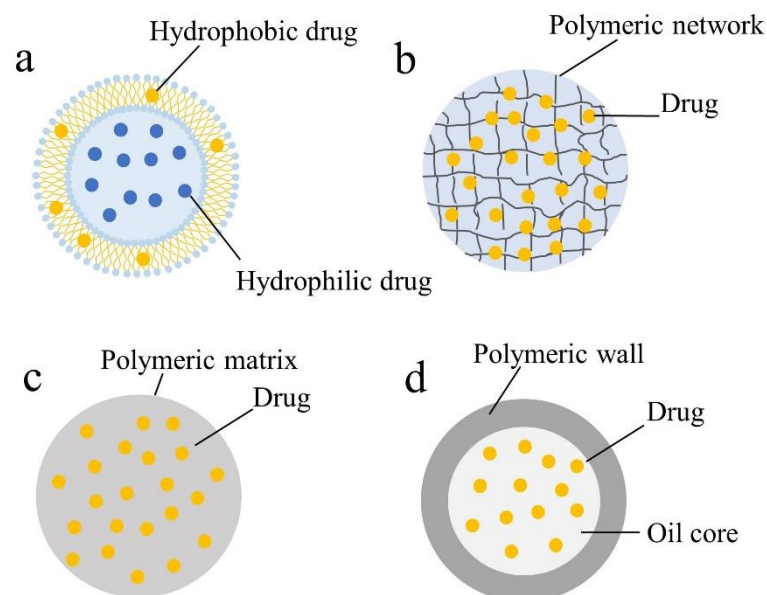
**Abstract:** Heparin-based delivery systems have been explored to improve their therapeutic efficacy and to reduce toxicity for different administration routes. Regardless of the applied drug delivery system (DDS), the evaluation of anticoagulant performance is instrumental for the development of a suitable DDS. The understanding of the range of anticoagulant assays, together with their key applications and limitations, is essential both within the context of scientific research and for clinical usage. This review provides an overview of the current anticoagulant therapy and discusses the advantages and limitations of currently available anticoagulant assays. We also discuss studies involving low-molecular-weight heparin (LMWH)-based nanocarriers with emphasis on their anticoagulation performance. Conventional anticoagulants have been used for decades for the treatment of many diseases. Direct oral anticoagulants have overcome some limitations of heparins and vitamin K antagonists. However, the lack of an accurate laboratory assessment, as well as the lack of a factor “xaban” (Xa) inhibitor reversal agent, remains a major problem associated with these anticoagulants. LMWHs represent anticoagulant agents with noteworthy efficacy and safety, and they have been explored to improve their outcomes with various nanocarriers through several administration routes. The main problems related to LMWHs have been surmounted, and improved efficiency may be achieved through the use of DDSs.

**Keywords:** heparin; anticoagulant drugs; drug delivery systems; anticoagulant assays

## 1. Introduction

Investments in terms of money and time in the development of new chemical compounds have led pharmaceutical companies to explore several delivery approaches in order to improve the efficacy and minimize the side effects of drugs that are already on the market [1]. One example is heparin. This natural compound has been clinically used for the treatment and prophylaxis of coagulation disorders (e.g., venous thromboembolism (VTE) [2]) for more than 60 years [3]. Though heparins, including unfractionated heparin (UFH) and low-molecular-weight heparins (LMWHs), have also been clinically used for a

long time, they show limitations because they are available only for parenteral administration, a costly and invasive method that leads to low patient compliance [4]. Non-invasive drug delivery systems are being explored to overcome these limitations. Nanotechnology-based drug carriers (nanocarriers), such as liposomes, nanogels, nanospheres, and nanocapsules, have gained increasing attention in the last few decades as a promising strategy for improving drug delivery [5] (Figure 1). Compared to conventional systems, nanocarriers have a number of advantages: (i) They are able to pass through the capillary vessels due to their submicron size and avoid clearance by phagocytes, prolonging the bloodstream circulation time; (ii) they can permeate cells and tissues to reach target organs; (iii) they can deliver the drugs in a controlled manner, reducing the frequency of administration and side effects, and they improve patient compliance [6]. Several heparin-based drug delivery systems have been developed to enhance the therapeutic efficacy of these drugs [7,8]. Although the oral route is the most desirable, it is noteworthy that there have been efforts to investigate heparin delivery through other pathways, such as nasal [9], pulmonary [10], and transdermal [11] routes.



**Figure 1.** Schematic representation of different types of nanocarriers: (a) liposomes, (b) nanogels, (c) nanospheres, and (d) nanocapsules.

The evaluation of nanocarrier efficacy comprises a critical step in the development of DDSs. In particular, heparin-based nanocarriers' performance is evaluated through several anticoagulant assays based on the type of drug used. The suitability and relevance of a specific anticoagulant assay for a certain objective are also important questions [12], thus stressing the need to know their advantages and limitations [13]. Anticoagulant assays, however, have not been comprehensively described from the perspective of DDS performance. The aim of our work was to provide a comprehensive overview of anticoagulant drugs, together with an outline of the techniques used for monitoring the anticoagulant plasma level. The most important heparin-based DDSs are also addressed, highlighting the most relevant techniques used to evaluate their anticoagulant performance.

## 2. Anticoagulant Therapy

The main anticoagulant drugs are categorized into four classes: heparins, vitamin K antagonists (VKAs), direct thrombin inhibitors (DTIs), and direct factor "xaban" (Xa) inhibitors. For clarity, we subdivided this section into conventional anticoagulant agents (heparins and VKAs) and novel anticoagulant drugs (DTIs and direct factor Xa inhibitors).

## 2.1. Conventional Anticoagulant Agents

### 2.1.1. Unfractionated Heparin (UFH), Low-Molecular-Weight Heparin (LMWH), and Ultra-Low-Molecular-Weight Heparin (ULMWH)

Unfractionated heparin (UFH) is one of the oldest biopolymeric drugs that is still used in therapeutics. It is a highly sulfated glycosaminoglycan (GAG), presenting the highest negative charge density of any known biological macromolecule [14]. The anticoagulant activity of UFH is related to its affinity with a naturally occurring serine protease inhibitor, antithrombin III (ATIII), resulting in an increase in the ATIII thrombin inhibition rate, as well as an inhibition of other serine proteases involved in the coagulation process [15]. UFH is indicated for treatment and prophylaxis of several conditions, including VTE, a dangerous disorder that includes deep vein thrombosis (DVT) and pulmonary embolism (PE) [2]. UFH is administered parenterally through either intravenous or subcutaneous injection; the latter is associated with a poor bioavailability. The non-specific binding of UFH to plasma proteins explains the unpredictable anticoagulant activity among individuals, which requires continuous monitorization. UFH is rapidly cleared from the body through endothelial and macrophage cells' depolymerization mechanisms, while the kidneys are responsible for a slower UFH clearance mechanism [16]. UFH is an effective, inexpensive, and relatively safe anticoagulant agent; however, it has numerous limitations. Hemorrhage is its main life-threatening adverse effect [17]. Although bleeding can be fatal, protamine sulfate administration could reverse this adverse event. Moreover, the binding of UFH to osteoblasts and platelet factor 4 leads to the tendency to induce osteoporosis and thrombocytopenia, respectively [18,19].

The discovery of the pentasaccharide sequence [20,21], the most active core, which is involved in ATIII binding and activation and associated with the need to overcome some heparin limitations, resulted in the development of LMWH in the late 1970s to early 1980s [22,23]. LMWHs are oligosaccharide fragments that are highly sulfated, water-soluble, and negatively charged compounds, and like UFH, they are indicated for the treatment and prophylaxis of VTE [24]. LMWHs show a mean molecular weight ( $M_w$ ) between 3500 and 6000 Da, with at least 60% of the material with  $M_w$  below 8000 Da. Hence, as the minimum  $M_w$  for anti-IIa activity is approximately 5000 Da, the LMWH anti-IIa activity is dramatically reduced [17]. Currently available LMWHs are produced by several controlled chemical or enzymatic depolymerization reactions of UFH, as seen in Table 1.

These controlled reactions afford drugs with more predictable pharmacokinetic and pharmacodynamic profiles, making them more dose-dependent than UFH. As a consequence, LMWHs require less monitorization [25]. Nevertheless, LMWHs are heterogeneous compounds and have different pharmacological and biochemical features; thus, they are not clinically interchangeable [26]. Additionally, LMWHs present less adverse effects and higher bioavailability when administered subcutaneously in comparison with UFH. However, protamine sulfate administration partially reverts the anticoagulant effect, thus increasing the risk of bleeding due to overdose. LMWHs are eliminated by the kidneys; consequently, the administration in patients with renal failure prolongs its half-life [22,27].

The theory that originated the therapeutic interest in ULMWH was based on the idea that compounds with a high anti-activated factor X (FXa) to anti-activated factor II (FIIa) activity ratio would achieve similar or better efficacy than LMWH products, but with a lower risk of bleeding and thrombocytopenia [28]. ULMWHs are obtained through a more extensive controlled depolymerization reaction, which preserves the pentasaccharide active site. They contain higher percentages of short chains ( $M_w < 3000$  Da), showing better efficacy and safety profiles. Fondaparinux was the first synthetic ULMWH, which is an analogue of the pentasaccharide sequence marketed by Sanofi in 2002 and now sold by GlaxoSmithKline under the trade name Arixtra© [29]. It has an  $M_w$  of 1728 Da, with a higher specific anti-FXa activity and longer half-life after subcutaneous injection than LMWHs. It is rapidly absorbed and 100% bioavailable after subcutaneous administration and has a half-life of approximately 14 to 18 h. It has a minimal non-specific binding

and does not require monitorization. It is cleared only through the renal route; therefore, clearance may be diminished in elderly patients [30]. An advantage of fondaparinux is that it presents a low risk of virus or prion contamination because it is synthetic. On the other hand, it requires an expensive synthetic process (approximately 50 steps) with a low yield [15]. This fact has stimulated research on less expensive synthetic routes.

Idraparinux is a long-acting hypermethylated fondaparinux analogue developed by Sanofi-Aventis that binds to AT with high affinity. After subcutaneous injection, idraparinux is almost completely absorbed and shows a plasma half-life of 80 h, allowing once-weekly injection without monitoring. Idraparinux has no antidote and is excreted unaltered through the renal route. The development of idraparinux was discontinued in phase III trials due to excessive intracranial bleeding [22,31]. The absence of an antidote for the clinical use of a long-acting anticoagulant is too dangerous. To solve this problem, a biotinylated idraparinux version was synthesized and clinically investigated. Idrabiota-parinux is a long-acting synthetic pentasaccharide analogue developed by Sanofi-Aventis. It displays the same anticoagulant properties as idraparinux. The biotin moiety allows rapid anticoagulant neutralization following the avidin infusion [31–33]. There is only one commercially available ULMWH (bemiparin). It is obtained through alkaline depolymerization from UFH, and it has been available since 1998. Bemiparin is approved for once-daily subcutaneous use in the prophylaxis of VTE. Pharmacokinetics studies reported that bemiparin showed a dose-dependent effect in healthy volunteers [34,35]. Semuloparin is a hemisynthetic ULMWH obtained through selective and controlled depolymerization. It is designed for once-daily subcutaneous administration in VTE prophylaxis in patients with cancer and to produce a higher anti-factor Xa and minimal anti-factor IIa activity compared with LWMHs. However, in 2012, the manufacturer suspended the development of semuloparin [28].

**Table 1.** Comparative features of LMWHs, ULMWHs, and their synthetic analogues.

Agents	M <sub>w</sub> <sup>a</sup> (Da <sup>b</sup> )	Production Process	Trade Name (Pharma Company)	Anti-Factor Xa:IIa Ratio	References
<b>LMWH</b>					
Ardeparin	6000	Peroxidative depolymerization	Normiflo® (Pfizer)	1.9	[15,22,36]
Certoparin	6000	Deamine cleavage with isoamyl nitrate degradation	Sandoparin®, Mono-Embolex®, Sandoz® (Novartis)	2.4	[15,22,36]
Dalteparin	6000	Nitrous acid depolymerization	Fragmin® (Pfizer)	2.7	[15,22,36]
Enoxaparin	4500	Benzylation and alkaline depolymerization	Clexane®/Lovenox® (Sanofi-Aventis)	3.8	[15,22,37]
Nadroparin	5000	Nitrous acid depolymerization	Fraxiparin® (Choay/Aspen)	3.6	[15,22]
Parnaparin	4500	Hydrogen peroxide and cupric salt depolymerization	Fluxum® (Wasserman)	3.0	[22]
Reviparin	4000	Nitrous acid depolymerization followed by chromatographic purification	Clivarine® (Abbott)	3.5	[15,22]
Tinzaparin	6000	Heparinase digestion	Innohep® (Novo/Leo)	1.5	[15,22,24]

Table 1. Cont.

Agents	M <sub>w</sub> <sup>a</sup> (Da <sup>b</sup> )	Production Process	Trade Name (Pharma Company)	Anti-Factor Xa:IIa Ratio	References
<b>ULMWH</b>					
Bemiparin	3600	β-eliminative cleavage through alkaline depolymerization	Hibor®, Ivor®, Zivor® (Sigma Tau)	8.1	[15,22,24]
Semuloparin	2400	β-eliminative cleavage through selective and controlled depolymerization using a phosphazene base	(Sanofi-Aventis)	80	[22,28]
<b>Synthetic analogues</b>					
Fondaparinux	1728	Chemical synthesis	Arixtra® (GlaxoSmithKline)	~850 UI anti-Xa/mg	[22,28]
Idraparinux	1728	Chemical synthesis	(Sanofi-Aventis)	~1600 UI anti-Xa/mg	[22]
Idrabiotaparinux	2052	Chemical synthesis	(Sanofi-Aventis)	~1600 UI anti-Xa/mg	[22]

<sup>a</sup> Mean molecular weight (M<sub>w</sub>); <sup>b</sup> Daltons (Da).

### 2.1.2. Vitamin K Antagonists (VKAs)

VKAs, such as warfarin, have been the standard anticoagulant therapy for more than 60 years, and they remain the most commonly prescribed oral anticoagulants worldwide [38]. VKAs are indicated for the long-term prevention of arterial and venous thromboembolic disorders [39]. Antagonizing the vitamin K, VKAs inhibit the formation of clotting factors dependent on vitamin K, such as factor II (FII), factor VII (FVII), factor IX (FIX), factor X (FX), and proteins C and S [40]. After oral administration, VKAs are rapidly absorbed from the gastrointestinal tract; they show high bioavailability and reach the plasma concentration peak within a few hours. Furthermore, VKAs have a mean plasma half-life of 40 h, and are highly bound to plasma proteins during circulation. Biotransformation and inactivation occur through different enzymes of the cytochrome P450 system, including CYP2C9, CYP1A2, and CYP3A4 [41–43]. Excretion of unchanged VKAs is negligible; thus, its elimination relies on hepatic metabolism [44]. Moreover, VKA antidotes are available, and a reversal effect could be achieved through vitamin K administration or clotting factor infusion [45]. Although the VKAs' advantages are well established in several thromboembolic disorders, they have several remarkable drawbacks. Bleeding is the most common and severe complication of VKAs [46]. In addition, the delay of onset and offset of action narrows the therapeutic index, and high inter- and intra-variability in the dose–response relationship are other issues presented by VKAs, leading to the need for frequent monitoring and dose adjustment [27,47].

### 2.2. Novel Anticoagulant Drugs

Over the last two decades, several direct oral anticoagulants (DOACs) have been developed. The research has moved directly toward the search for small molecules that bind reversibly with thrombin or FXa and are suitable for oral administration [48]. Megalotran (the prodrug was ximelagatran), developed by Hassle, followed by Astra and, subsequently, Astra-Zeneca, became the first available oral thrombin inhibitor. It shows an efficacy and bleeding rate comparable to those of LMWH and had the advantage of not needing parenteral administration or laboratory monitoring. However, ximelagatran was removed from the market due to liver toxicity [49]. Currently, there are two types of DOACs licensed for use in thromboembolic disorders: FXa inhibitors (apixaban, betrixaban, edoxaban, and rivaroxaban) and a thrombin inhibitor: dabigatran. The comparable characteristics of these

agents are shown in Table 2. Dabigatran etexilate (Pradaxa<sup>®</sup>), the only direct thrombin inhibitor (DTI) licensed [47], is a selective and reversible oral nonpeptide prodrug developed by Boehringer Ingelheim Pharmaceuticals [42]. After being taken orally, dabigatran etexilate is converted into its active form, dabigatran, through esterase-mediated hydrolytic cleavage [50]. It has a rapid-action onset and low bioavailability, and it is predominantly excreted through the renal route [51]. Dabigatran etexilate has been approved for the prevention of venous thromboembolism (VTE), stroke, or systemic embolism [19]. Rivaroxaban (Xarelto<sup>®</sup>), the first direct oral FXa inhibitor clinically approved by the Food and Drug Administration (FDA) in 2008 [52], is a selective, dose-dependent, and competitive inhibitor of free and clot-based factor Xa, which was developed by Janssen Pharmaceuticals and Bayer HealthCare [53]. It is rapidly absorbed, and the blood concentration peak is achieved after 2–4 h. It has a half-life of 5–13 h, and it is excreted through the renal route (66%) and through the fecal/biliary route (28%) [54]. Rivaroxaban has been approved for the prevention of VTE [19]. Apixaban (Eliquis<sup>®</sup>) was the second FXa inhibitor to be clinically approved by the European Medicines Agency (EMA) and by the FDA for thromboembolic prophylaxis, as well as for preventing blood clots in DVT and PE [52]. Apixaban is a direct, reversible, competitive, and selective FXa inhibitor designed by Bristol Myers Squibb and Pfizer (US). It shows good oral absorption, reaching its peak plasma concentration in 1–4 h. A total of 25% of the apixaban is excreted through the kidneys, and the remnant by the liver [55]. Edoxaban (Savaysa<sup>®</sup> in USA and Lixiana<sup>®</sup> in Canada) is another direct FXa inhibitor developed by Daiichi-Sankyo, approved in Japan and in the US for stroke and VTE prophylaxis [56,57]. It is well absorbed, reaching its plasma concentration peak after 1–2 h. Elimination of edoxaban takes place via the renal and hepatobiliary routes [58]. Betrixaban (Bevixxa<sup>®</sup>) is a more recent direct FXa inhibitor developed by Portola Pharmaceuticals Inc. (San Francisco, CA, USA). It was approved by the FDA in 2017 for the prevention of thromboembolism in high-risk hospitalized individuals [59]. Betrixaban inhibits FXa in a competitive, reversible, selective, and dose-dependent way. It is rapidly absorbed and shows its peak plasma concentration within 3–4 h. Its bioavailability is 34%, and it shows the longest half-life among the FXa inhibitors. Betrixaban is mainly excreted through the hepatobiliary system and via a P-glycoprotein efflux pump [60,61].

**Table 2.** Pharmacokinetic and pharmacodynamic properties of DOACs.

Features	Factor IIa Inhibitor		Factor Xa Inhibitor			References
	Dabigatran	Apixaban	Rivaroxaban	Edoxaban	Betrixaban	
M <sub>w</sub> <sup>a</sup> (Da <sup>b</sup> )	628	460	436	548	452	[62]
Bioavailability (%)	6	50	80–100	62	34	[62,63]
Protein binding (%)	35	87	92–95	55	60	[61,63]
T <sub>max</sub> <sup>c</sup> (h)	1–3	1–4	2–4	1–2	3–4	[61]
Half-life (h)	12–17	12	5–13	9–11	19–27	[53,60]
Reversal agents	Yes	No	No	No	No	[62]
Renal clearance (%)	>80	25	66	35–50	<7	[61]

<sup>a</sup> Mean molecular weight (M<sub>w</sub>); <sup>b</sup> Daltons (Da); <sup>c</sup> Time to reach the maximum plasma concentration (T<sub>max</sub>).

Unlike VKAs, the DOACs have a rapid onset and offset of action and a low potential for food interaction. In addition, DOACs have a wide therapeutic window and predictable anticoagulant response, and there is no need for laboratory monitoring or dose adjustments according to body weight [48]. Despite the enthusiasm for DOACs, a number of situations are noteworthy and should be considered. Pregnant patients were excluded from the randomized trials and, consequently, the efficacy and safety concerns (e.g., teratogenic risk) in this group are unknown [39]. Cost is another issue. Drugs such as UH and VKAs are cheaper compared to DOACs [64]. Moreover, the lack of available reversal agents is another important problem. Currently, dabigatran is the only DOAC with a licensed antidote [48].

Although the DOACs have been licensed without the need for laboratory monitoring, some circumstances may require that the drugs be monitored: (i) unconscious patients, where the presence of anticoagulants may influence the management decision; (ii) in the case of suicide attempts, it may be important to establish whether there was an anticoagulant drug overdose; (iii) bridging from one anticoagulant to another; (iv) establishing whether co-medication effects are altering the anticoagulant efficacy [64]. Therefore, the lack of a reliable method for monitoring the drug level is potentially problematic [65].

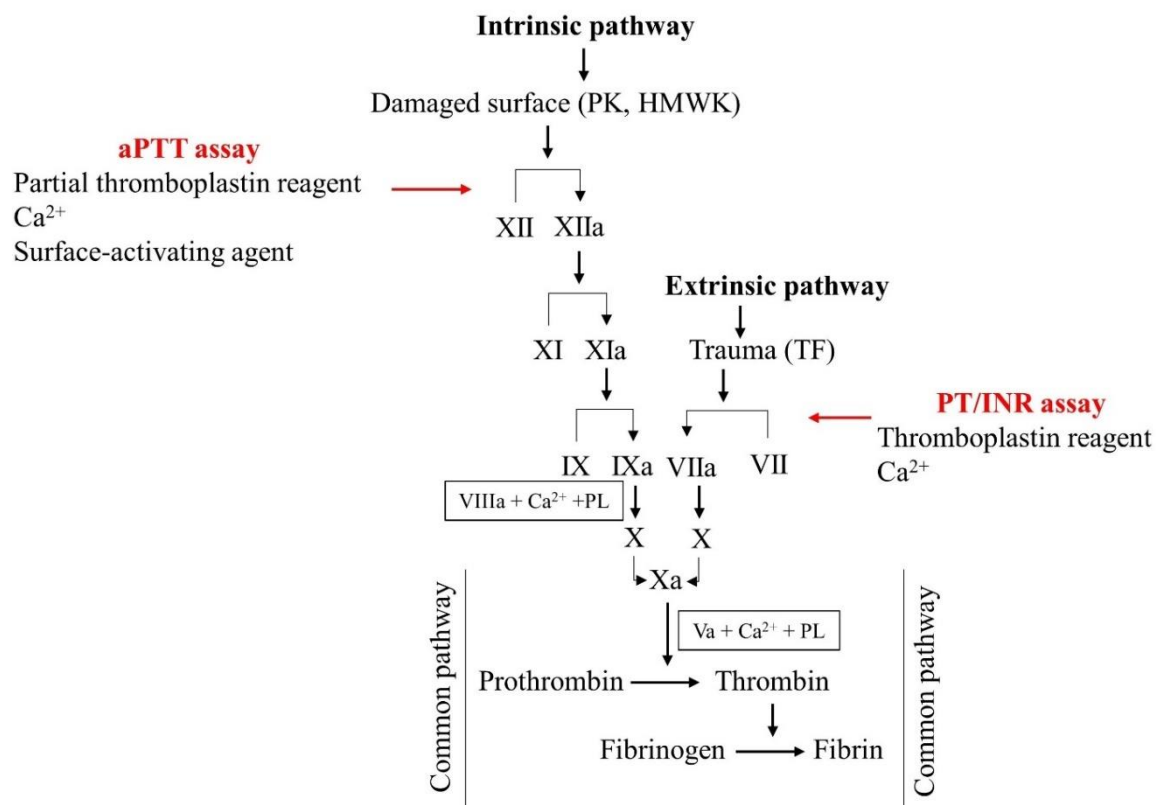
### 3. Laboratory Assessment of Anticoagulant Therapy

There are several anticoagulant drugs available, as discussed in the previous section. Each anticoagulant agent differs in its effects on coagulation tests, and each drug may require different laboratory assays to measure/monitor drug concentration.

#### 3.1. Monitoring VKAs and UFH

Routine coagulation tests are represented according to their prothrombin time/international normalized ratio (PT/INR) and their activated partial thromboplastin time (aPTT). The PT/INR and aPTT are sometimes obtained to investigate dysfunctions in secondary hemostasis; however, they are more frequently used for monitoring anticoagulant therapy [64]. The PT assay is a straightforward test that evaluates the extrinsic and common pathway factors of coagulation. It is defined as the time in seconds for plasma to clot after the addition of calcium and the thromboplastin reagent, which uses an extrinsic activator pathway (Figure 2) [66]. Commercial thromboplastin reagents have different sources, methods of preparation by their different manufacturers, and, consequently, varying sensitivities to coagulation factors, resulting in uncertainty in PT results [13]. In addition, there are various coagulation analyzers available from several manufacturers. The influence of these two factors is responsible for the variability in PT results among laboratories [66]. To bypass the variability and standardize the PT results, in 1983, the World Health Organization (WHO) introduced the INR concept. The INR is calculated as follows:  $INR = (\text{patient PT}/\text{MNPT})^{ISI}$  [64], where MNPT is the mean normal PT, which is obtained from at least 20 healthy volunteers, and ISI is the international sensitivity index of the thromboplastin. The ISI reflects the ability of thromboplastin to reduce the vitamin-K-dependent factor in comparison with a standard thromboplastin defined by the WHO [13]. While the INR approach has reduced the variability of patients' interlaboratory results, on the other hand, the determination of ISI has generated a source of variability depending on the reagent/instrument utilized in the laboratory and the source of plasma used to generate MNPT [66].

Despite the fact that INR has brought an important means of standardization for interlaboratory results, the need for improvements in the technique to ensure more reliable results still exists. The PT/INR is sensitive to the presence or activity of FII, factor V (FV), FVII, FX, and fibrinogen. In that way, anticoagulant agents that alter the levels of these factors can be monitored with the PT/INR if a dose–response relationship could be defined [64]. The PT/INR responds to a lowering of the level of FII, VII, and FX caused by VKAs, leading to a prolongation of PT and, thus, a higher INR [13]. The target INR therapeutic limits are 2.0 to 3.0 for most indications, and it is well defined that subtherapeutic INR levels are related with an increased thrombotic risk. On the other hand, supratherapeutic levels correlate with an increased bleeding risk [48]. Monitoring VKA therapy is crucial due to the highly variable anticoagulant response and the narrow therapeutic window. VKAs interfere with three of the five factors of the extrinsic pathway. Thus, the PT/INR is the most sensitive technique for monitoring the VKA levels, as well as, possible fluctuations in VKA levels due to several interactions presented by these drugs [66].



**Figure 2.** Intrinsic and extrinsic activator pathways.

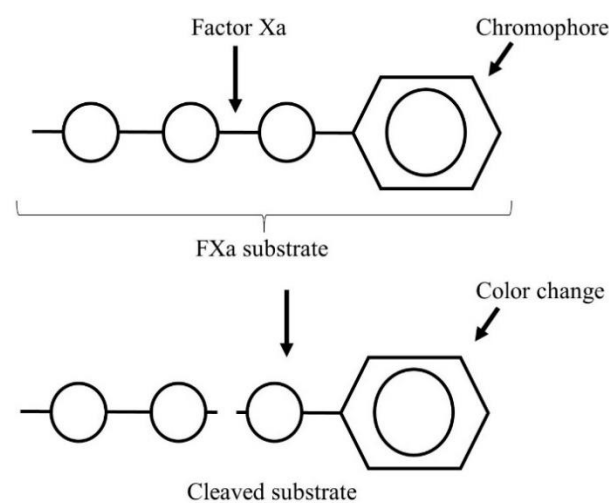
The aPTT is a clotting assay that evaluates the intrinsic and common pathway factors of coagulation. This test was first developed by Langdell, Wagner, and Brinkhous in 1953 [67] through a modification of the whole blood clotting time test (WBCT) developed by Lee and White to test patients for hemophilia [68]. The aPTT is defined as the time in seconds for the recalcified plasma to form a clot after adding a platelet substitute (partial thromboplastin, named this way because the absence of TF) and a surface-activating agent (such as celite, ellagic acid, kaolin, or micronized silica) (Figure 2) [69,70]. The aPTT is sensitive to the presence or activity of FII, FV, factor VIII (FVIII), FIX, FX, factor XI (FXI), factor XII (FXII), and fibrinogen. Therefore, anticoagulant agents that affect the levels of these factors can be monitored with the aPTT if a dose–response relationship can be determined [64]. Most aPTT reagents shows linear sensitivity to UFH over the therapeutic range, so the aPTT is considered the main assay for monitoring UFH therapy [71]. Monitoring UFH therapy is important for avoiding plasma variation, preventing both toxic levels, which are related with an increased risk of bleeding, thrombocytopenia, and osteoporosis, or subtherapeutic levels, leading to the inefficacy of thrombosis treatment [47]. The aPTT test offers advantages, such as its low cost, ease of performing, wide availability, and automation. These explain why it is widely accepted by clinicians [72]. However, there are some limitations that might influence the technique and are noteworthy. Regarding pre-analytical conditions, several variables might affect the test results, namely, the duration of sample collection, citrate concentration, sample transport temperature, and the time between collection and plasma separation [73]. Concerning analytical conditions, the reagents, instruments, and methods of end-point detection are a source of test variability. The same reagents produced in different lots can yield variability in the test’s sensibility. The partial thromboplastin reagent is available through many suppliers with either vegetable or animal phospholipid sources [74]. Thus, the high phospholipid heterogeneity, either in composition or in concentration, is another source of variation in the sensitivity of aPTT assays. Moreover, the choice of the surface-activating agent modifies the aPTT time in normal plasma [75]. The biological conditions are another factor influencing the



aPTT results. Intravascular volume, protein level, coagulation factor concentration, and AT deficiency contribute to the variability in the aPTT [72]. From all of the limitations discussed above, it is remarkable that the major problem of aPTT assays is the absence of standard methods. Consequently, the aPTT shows a high interlaboratory variation, and even in-house analysis should be carefully interpreted [76]. UFH and VKAs require rigorous laboratory control because they have a very narrow therapeutic window; therefore, the correct dose is strictly related to an accurate aPTT or PT/INR assay control, respectively.

### 3.2. Monitoring LMWH Therapy

Despite the advantageous pharmacokinetic and pharmacodynamic properties of LMWHs, which grant a lesser need for laboratory monitoring in some circumstances, drug monitoring can bring some benefits for specific populations. The most commonly applied test for LMWH monitoring is the chromogenic anti-factor Xa or anti-Xa assay, mainly due to its low cost and broad availability [77]. The chromogenic anti-factor Xa is considered the gold standard for assessing the plasma LMWH level. In general, the anti-Xa assay measures the enzymatic activity of a patient's plasma in cleaving an exogenous FXa [77]. A synthetic FXa substrate linked with a chromophore is added to the plasma sample. Endogenous FXa cleaves the chromogenic substrate to release the chromophore unit, leading to a color change that can be detected spectrophotometrically (Figure 3) [78]. In the case of a plasma sample containing LMWH, it will inhibit FXa, reducing the FXa level in order to cleave the synthetic chromophore. By comparing the results with the corresponding standard curve (samples containing known amounts of LMWH), the anticoagulant concentration can be calculated. The extent of colorimetric change is directly proportional to the enzymatic activity and inversely proportional to the anticoagulant concentration [79].



**Figure 3.** Schematic representation of the endogenous FXa cleavage of the chromogenic substrate, thus releasing the chromophore unit and leading to a color change.

Drawbacks in LMWH monitoring with the anti-factor Xa assay include the lack of standardization in the (1) anti-Xa reagent, (2) standard curve construction, and (3) instrument for analysis [80]. There are numerous commercially available anti-Xa assay kits that use diverse synthetic substrates to measure FXa activity. Thus, analyses of the same sample yield discrepant results [77]. Two different anti-Xa test designs with distinct aims are available. One applies exogenous AT to correct a possibly low *in vivo* AT concentration. However, this approach may lead to overestimation of the *in vivo* anticoagulation activity because unbound LMWH, which does not produce anticoagulant activity, could bind to exogenous AT, causing a false positive result [12]. On the other hand, excess dextran sulfate may be added to mitigate *in vivo* protein binding to the drugs. This strategy may result in overestimated anticoagulant activity, although this is a minor concern for LMWH due to their reduced binding to plasma proteins [77]. The construction of standard curves to calcu-

late the drug's concentration in the plasma is essential. The same brand and lot number of LMWH administered to patients should be utilized in order to prepare the standard curve. Moreover, different reagents and instrument systems produce variable results [12,80]. The anti-FXa/anti-FIIa ratios among LMWH formulations and the time needed to obtain a blood sample are another pitfall related to the anti-factor Xa assay. In fact, because the peak plasma concentration of LMWH is at about 4 h, blood samples should be collected as close as possible to 4 h after LMWH administration [78,79]. The target therapeutic range for anti-Xa activity depends on the dosage regime and the LMWH agent utilized. With once-daily dosing, the therapeutic range of LMWH is 1.0 to 2.0 U/mL, while for a twice-daily regime, the therapeutic range of LMWH is between 0.5 and 1.0 U/mL [81]. Although anti-Xa is considered the main choice for plasma LMWH level measurement, the anti-Xa level may not correlate with the clinical results, so the results should be evaluated with caution.

### 3.3. Monitoring DOAC Therapy

Although routine laboratory monitoring of DOACs is not necessary, in some special situations, laboratory measurements may be helpful [82], such as for emergency surgeries, overdoses, extreme body weights, pregnancies, drug interactions, renal impairments, and treatment failures [83]. An ideal assay that measures DOACs accurately should show a great level of linearity within a wide range of drug concentrations. Moreover, the assay should be sensitive to low therapeutic concentrations and should be specific for the drug of interest so that no interferences affect the drug's determination. Finally, the assay should be widely available and able to give results in a short time. Currently, there is no test with these ideal characteristics [82].

Liquid chromatography/mass spectrometry (LC/MS), calibrated for each drug to be measured, is considered the standard method for DOAC measurement [84]. This technique has a superior accuracy and precision over a broad concentration range compared to coagulation-activity-based assays [85,86]. However, this technique has limited availability, is time consuming and expensive, and requires high technical expertise. Therefore, although LC/MS is a gold-standard method, it is unsuitable for routine DOAC measurement. Conventional anticoagulant assays, such as PT and aPTT, are inappropriate for accurate DOAC measurement. However, recent guidelines and recommendations for DOAC assessment state that aPTT and PT may be utilized as a screening test for DTI and anti-Xa inhibitor (only rivaroxaban), respectively, particularly when rapid and reliable information about a possible overcoagulation is necessary [87].

Conventional monitoring tests are not suitable for apixaban, edoxaban, and betrixaban. Thrombin time (TT) is a rapid, simple, and inexpensive test that assesses the thrombin activity through conversion of fibrinogen into fibrin [88,89]. TT is highly sensitive to dabigatran, and it is not used for accurate measurement of it. Depending on the reagent and the dabigatran concentration, TT may be unmeasurable. Nevertheless, TT has an important use: Normal values are suggestive of little or no clinically relevant dabigatran levels [82]. The plasma dabigatran level can be measured with different assays, such as diluted thrombin time (dTT) and ecarin-based assays: ecarin clotting time (ECT) and ecarin chromogenic assay (ECA) [71,87]. The dTT, or its commercial version, Hemoclot<sup>®</sup> Thrombin Inhibitor (Hyphen Biomed, Neuville-sur-Oise, France), is a TT modification assay for overcoming the exquisite sensitivity to dabigatran. Diluting the test plasma with normal plasma and purified human  $\alpha$ -thrombin (to initiate clotting) ensures a rapid, simple, and commercially available quantitative method for determining plasma DTI concentration [88]. When dTT is calibrated for dabigatran measurement, the assay shows a high linearity with clinical dabigatran concentration with a good reproducibility [90]. The dTT assay has been considered the gold standard for measurement of plasma dabigatran concentration [91]. ECT (Ecarin Reagent, Diagnostica Stago, Asnieres, France) is a meizothrombin generation assay that enables accurate DTI measurement [92]. Ecarin is a metalloproteinase purified from snake venom, *Echis carinatus*, which, when added to citrated plasma, specifically

cleaves prothrombin into an active intermediate, meizothrombin, and the time until clot formation is measured [82]. With DTI therapy, the meizothrombin generated is inactivated as long as DTI is present in the plasma. Thus, coagulation will not occur until all DTI is consumed, thus prolonging the time until clot formation [88,92]. In the ECA (ECA-T Kit, Diagnostica Stago, Asnieres, France), the meizothrombin cleaves a chromogenic substrate, which releases the chromophore unit (paranitroalanine), resulting in absorbance changes. With DTI therapy, the cleavage reaction is inhibited, and by comparing the results with the corresponding standard curve, the anticoagulant concentration can be measured [93]. Direct FXa inhibitors can be accurately measured through a modified chromogenic anti-Xa assay. It is advocated that the anti-Xa assay calibrated with the same drug being taken by the patient is the most reliable technique for accurately measuring the anticoagulation effect of direct oral FXa inhibitors [82,87].

Although the development and introduction of DOACs in the market have revolutionized anticoagulant therapy due to their several advantages over VKAs and heparins, they have brought some challenges in laboratory monitoring [85]. Due to the differences in the sensitivity of routine coagulation tests, none of them are able to accurately measure DOACs. Nevertheless, qualitative information about the presence of DOACs or lack thereof could be obtained [88]. The absence of FDA-approved calibrators and reliable methods for quantifying DOACs has generated a lack of interest in the development of a quantitative assay. Alternative methods, such as ECT and ECA, may be useful, but have not been widely implemented. The use of commercial calibrators, as well as the use of specialty coagulation tests, to measure DOACs should be interpreted with attention [85].

#### 4. Coagulation Test Performance of Drug Delivery Systems

##### 4.1. Liposomes

Liposomes are defined as phospholipid vesicles formed by concentric lipid bilayers surrounding an aqueous inner phase [94]. Their special structure, hydrophobic and hydrophilic characteristics, and biocompatibility make liposomes a promising system for drug delivery [95].

LMWH-loaded flexible liposomes (flexosomes) (size~83 nm) and LMWH-loaded ethosomes (size~85 nm) were prepared for transdermal delivery [96]. Their physicochemical features and pharmacokinetic parameters were compared. The plasma anti-Xa activity of flexosomes, ethosomes, and LMWH solution was measured through a chromogenic anti-Xa assay (Coatest<sup>®</sup>, Chromogenix, Instrumentation Laboratory, Milano, Italy). After transdermal application, the [anti-Xa]<sub>max</sub> of the LMWH-loaded flexosomes was 1.11 IU/mL, while the [anti-Xa]<sub>max</sub> of the LMWH-loaded ethosomes was 0.32 UI/mL. The authors stated that the improved skin permeability of the flexosomes was due to their reduced size and the higher deformability than that of ethosomes (76.7% versus 46.8%). Moreover, they explained that flexosomes are able to change their shape to pass through skin barriers. In another study, Bai and Ahsan [97] prepared ardeparin-loaded conventional liposomes and PEGylated liposomal formulations with the hydration method for the respiratory route. Pulmonary absorption of formulations was monitored through plasma anti-Xa activity and compared with LMWH saline formulations and subcutaneous administration. The antifactor Xa levels (>0.2 UI/mL) for all liposomal formulations were at therapeutic levels for more than 6 h. Moreover, the prolongation of the half-life of the drug was significantly higher when compared to that for the plain LMWH or subcutaneous route.

##### 4.2. Hydrogels

Hydrogels are three-dimensional cross-linked hydrophilic polymer networks that are capable of retaining large amounts of water or biological fluids while remaining insoluble in these media [98]. Their hydrophilicity is responsible for some of the desirable features of hydrogels, such as biocompatibility and the capability of encapsulating hydrophilic molecules [99]. Because the hydrogels were mostly prepared in aqueous solutions, the drug denaturation and aggregation possibilities were mitigated. Moreover, the hydrogel

network was able to protect the entrapped drug against the harsh environment [100]. Loira-Pastoriza et al. prepared nanogels to incorporate four commercial LMWHs for topical delivery [11]. For bemparin measurement in plasma after topical application, they used a chromogenic anti-Xa assay (Stachrom<sup>®</sup> heparin, Diagnostica Stago, Asnières-sur-Seine, France). The authors could not detect bemparin in plasma after 24 h of topical application. However, they stated that, for the route of administration and for the treatment of superficial thrombosis and hematomas, the drug should not pass into the systemic blood circulation in order to avoid a systemic effect. Matanovic et al. developed a thermoresponsive poloxamer-based platform that formed in situ combined with pH-responsive polyelectrolyte heparin/chitosan nanocomplexes for prolonged subcutaneous release of heparin. The plasma heparin concentration was measured after subcutaneous injection of selected individual formulations (thermoreponsive hydrogel and heparin/chitosan nanocomplexes) and a dual system (nanocomplexes incorporated into thermoresponsive hydrogel) through a chromogenic FXa assay and Berichrom<sup>®</sup> Heparin kit (Siemens Healthcare Diagnostics Inc., Erlanger, Germany). Plasma heparin concentration–time profiles showed a double-peak phenomenon, and this dual behavior was probably related to diffusion of heparin inside the polymer network and gel dissolution. It was demonstrated that thermoresponsive hydrogel with heparin/chitosan nanocomplexes showed sustained release of heparin for 5 days [101]. The aPTT is the most preferred assay for monitoring the therapeutic dose of UFH in many hospitals and laboratories, mainly due to its low cost, wide availability, and ease of performing, rather than the specific advantages of this technique. However, the anti-Xa assay may be necessary either in some patients (e.g., elevated baseline aPTT or FXII deficiency) or because it offers another option with fewer interferences [71,87].

#### 4.3. Polymeric Nanoparticles (PNPs)

PNPs are colloidal dispersions that have been used for drug and biomolecule delivery [102]. PNPs have several advantages: (i) They have improved stability; (ii) they can avoid the phagocyte system, thus prolonging their blood stream duration; (iii) because of their sub-cellular size, they can pass through capillary vessels; (iv) they can improve the drug properties and reduce side effects; (v) they can release the drug or biomolecule in a controlled/sustained fashion [103–105]. Dong et al. prepared chitosan-based polymer-lipid hybrid nanoparticles (PLNs) with a self-assembly method to incorporate and improve the oral absorption of enoxaparin (Enox). The PLNs were composed of chitosan as the polymer and glyceryl monooleate (GMO) as the lipid, with an optimized lipid/polymer mass ratio of 0.2 and pluronic<sup>®</sup> F127 as a stabilizer. Nanoparticles with the optimal composition were submitted to an in vivo study. The Enox anticoagulant activity was measured through the aPTT assay. The optimized Enox/PLNs showed a 4.5-fold increase in oral Enox bioavailability in comparison with an Enox solution. The authors claimed that the stability of nanoparticles is a crucial parameter for improved oral drug delivery efficiency [7]. The authors utilized the aPTT assay to measure plasma LMWH concentration; however, the aPTT was not helpful for this purpose because the anticoagulant effect of LMWH is produced mainly through FXa inhibition, and the prolongation of the aPTT is highly dependent on low thrombin activity [77]. In another work, Ramadan et al. produced and characterized cationic lipid nanocapsules (cLNCs) with a phase-inversion method intended for fondaparinux (Fp) oral absorption. In vivo studies were performed after a single oral dose in rats, and the plasma Fp measurements were carried out with the anti-Xa activity assay (HYPHEN, Biomed). The authors reported that the cLNCs significantly increased oral Fp bioavailability and improved pharmacokinetic profile in a dose-dependent manner [106].

#### 4.4. Other Drug Delivery Systems

Solid lipid nanoparticles (SLNs) are lipid-based colloidal nanocarriers consisting of a hydrophobic core composed of lipids that are solid at both room and body temperatures

and stabilized by a surfactant layer. SLNs are capable of affording a controlled drug release because the release of the active compound from the solid lipid matrix is much slower when compared to liquid oils [107,108]. Paliwal et al. synthesized biomimetic LMWH–lipid conjugate-loaded SLNs for oral bioavailability studies. An in vivo study was performed in albino rats, and the plasma LMWH concentration was measured with a colorimetric anti-Xa assay using a Chromogenix Coatest LMW heparin kit (Diapharma). The authors demonstrated that SLNs were within the nanometric range and were safe for oral administration. Furthermore, improved bioavailability of LMWH–lipid conjugates was observed in comparison to LMWH-loaded SLNs and LMWH solution. They attributed this result to the high loading capacity and better lymphatic drainage of the LMWH–lipid conjugate SLNs, thus achieving an improved systemic LMWH concentration [109]. Self-nano emulsifying drug delivery systems (SNEDDSs) have been explored as drug delivery systems because they are able to spontaneously form oil-in-water (o/w) nanoemulsions after administration in an aqueous environment, followed by mild agitation (e.g., the gastric motility) [110,111]. Zupancic et al. prepared and characterized various enoxaparin-loaded self-emulsifying drug delivery systems (SEDDSs) with lipids of different chain lengths for oral administration [112]. An in vivo study was carried out in rats, and the plasma enoxaparin concentration was measured with a Biophen<sup>®</sup> Heparin anti-Xa assay kit. Medium-chain lipids (MC-SEDDS) and no lipids (NL-SEDDS) showed greater mucus diffusion. MC-SEDDS and NL-SEDDS showed sustained enoxaparin release in vitro and an oral absolute enoxaparin bioavailability of 2.02% and 2.25%, respectively. A literature review focusing on anticoagulant therapies using different drug delivery systems is given in Table 3.

**Table 3.** Anticoagulant drug delivery systems for different routes of administration.

Drug Delivery System	Active (Class)	Composition	Size Range	Major Outcomes	Ref
Liposomes	(LMWH)	Egg PC Tween <sup>®</sup> 20 Ethanol	80–90 nm	LMWH-loaded flexosomes showed higher antifactor Xa (Anti-Xa) max than LMWH-loaded ethosome.	[96]
	Ardeparin (LMWH)	PC Chol DSPE DSPE-PEG-2000 and DSPE-PEG-5000	100–150 nm	Liposomal formulations showed sustained release and longer half-life compared to the plain solution or subcutaneous route.	[97]
	Enoxaparin (LMWH)	Soybean PC Chol SA Eudragit <sup>®</sup> S 100	100–200 nm	Eudragit-coated liposomes showed higher permeation and oral bioavailability when compared to uncoated liposomes.	[113]
Hydrogels	Bemiparin (LMWH) Nadroparin (LMWH) Tinzaparin (LMWH)	Eudragit <sup>®</sup> RS 30D	130 nm	Gel formulations were able to deliver LMWHs across the skin barrier, and after 24 h, the drug was not detected in plasma.	[11]
	Heparin (UFH)	CS hydrochloride Lutrol <sup>®</sup> F127 Lutrol <sup>®</sup> F68 HPMC	150–400 nm	The dual system enabled the lowest absorption rate of heparin into systemic circulation and provided heparin concentrations above the prophylaxis threshold for 5 days.	[101]

Table 3. Cont.

Drug Delivery System	Active (Class)	Composition	Size Range	Major Outcomes	Ref
	Enoxaparin (LMWH)	CS Pluronic® F127 PEI Plys	100–1000 nm	Thermo-sensitive hydrogels were able to prolong the enoxaparin release.	[114]
	Heparin (UFH)	Gelatin MBA TEA	NA	Heparin-loaded hydrogels showed sustained release for 60 h and platelet adhesion was significantly reduced.	[115]
	Enoxaparin (LMWH)	CS GMO Pluronic® F127	290–320 nm	The optimized formulation showed higher oral bioavailability compared with the drug solution.	[7]
	Fondaparinux (Synthetic analogues)	Labrafac® WL 1349 Lipoïd® S75-3 Lipoïd® GMBH Solutol® HS 15 CTAB SA	40–65 nm	Cationic lipid nanocapsules showed increased oral bioavailability and longer half-life when compared to fondaparinux control solutions (oral and intravenous).	[106]
PNP	Enoxaparin (LMWH)	Pluronic® F-68 CTAB Dextran sulfate PLGA Precirol ATO 5 E80 Tween 80 Poloxamer 407	180–195 nm	Enoxaparin/CTAB nanoparticles showed three-fold improved gastrointestinal permeation when compared with the drug solution.	[116]
	Enoxaparin (LMWH)		145–160 nm	LPHNs improved the drug's intestinal permeation, enhanced the oral bioavailability, and showed therapeutic efficacy.	[117]
SLN	(LMWH)	Compritol 888 ATO Stearic, palmitic and myristic acid PC	280–380 nm	The SLNs were able to improve the LMWH bioavailability in comparison to the free drug solution.	[109]
SNEDDDS	Enoxaparin (LMWH)	Capmul MCM EP, Capmul PG-8 EP/NF Captex 8000, Peceol Labrafil M 1944 CS, Labrasol Maisine 35-1, Transcutol HP Myglyol 840, Cremophor EL PEG, triacetin, olive and sesame oil	30–245 nm	SEDDS formulations showed sustained enoxaparin release and two-fold bioavailability.	[118]

Table 3. Cont.

Drug Delivery System	Active (Class)	Composition	Size Range	Major Outcomes	Ref
	Rivaroxaban (Factor Xa inhibitor)	IPM Ethyl oleate, Tween20, and Tween80 Cremophor, Cremophor HEL, and Transutol	50–105 nm	SNEDDS showed higher dissolution than the commercial formulation. The SNEDDS technology used in rivaroxaban successfully enhanced drug bioavailability in fasting conditions, and no food effects were observed in the rivaroxaban–SNEDDS formulation.	[119]
	Rivaroxaban (Factor Xa inhibitor)	Transcutol HP Capryol TM 90 Maisine TM 35-1 Castor oil, oleic acid, triacetin, IPM Cremophore EL PEG 300 and PEG 400 Tween 20, Tween 80, and Span 80	10–115 nm	Safe SNEDDS formulations enhanced oral and intravenous bioavailability in comparison to the drug suspension. Moreover, SNEDDS exhibited anticoagulant efficacy in a rat thrombosis model.	[120]

## 5. Conclusions

Conventional anticoagulant agents are well-known drugs with reliable anticoagulant assays that have been used for decades. On the other hand, after a long time without advances in anticoagulation management, DOACs have emerged in the last decade. Despite the remarkable advances made in anticoagulant therapy with the introduction of DOACs, there are still some important issues that should be addressed, such as the lack of specific antidotes and the absence of an accurate and standardized coagulation test. The anticoagulant tests were found to be suitable for measuring the anticoagulant performance of heparin-based nanocarriers through several administration routes.

**Author Contributions:** All authors substantially contributed to the conceptualization, methodology, formal analysis, investigation, resources, and data curation. Y.B.G.P., L.L.C., M.F.R.S., and J.L.S.-S. contributed to the writing—original draft, data search, and literature research. M.F.R.S., J.L.S.-S., E.H.G., P.S., and E.B.S. contributed to the writing—review, validation, and editing. E.B.S., P.S., M.F.R.S., and J.L.S.-S. contributed to the project administration, supervision, and funding acquisition. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was supported by the Banco do Nordeste (grant FUNDECI/2016.0015), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Fundação de Apoio à Pesquisa e à Inovação Tecnológica do Estado de Sergipe (Fapitec), and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES). Eliana B. Souto would like to acknowledge the Portuguese Science and Technology Foundation (FCT/MCT) and the European Funds (PRODER/COMPETE) for the project UIDB/04469/2020 (strategic fund), co-financed by FEDER, under the Partnership Agreement PT2020.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

CS	chitosan
Chol	cholesterol
CTAB	hexadecyltrimethyl ammonium bromide
DSPE	1,2-distearoyl-sn-glycero-3-phosphoethanolamine
DSPE-PEG	1,2-distearoyl-sn-glycero-3-phosphoethanolamine polyethyleneglycol
E80	egg yolk lecithin
Egg PC	phosphatidylcholine from egg yolk
GMO	glyceryl monooleate
HPMC	hydroxypropylmethylcellulose
IPM	isopropyl myristate
MBA	N,N-methylenebis (acrylamide)
NA	not available
PEI	polyethylenimine
PC	phosphatidylcholine
Plys	$\epsilon$ -polylysine
PNP	polymeric nanoparticle
SA	stearyl amine
SEDDS	self-emulsifying drug delivery system
SLN	solid lipid nanoparticles
SNEDDS	self-nanoemulsifying drug delivery system
TEA	triethylamine

## References

- Patel, V.F.; Liu, F.; Brown, M.B. Advances in oral transmucosal drug delivery. *J. Control. Release* **2011**, *153*, 106–116. [[CrossRef](#)] [[PubMed](#)]
- Aláez-Versón, C.R.; Lantero, E.; Fernández-Busquets, X. Heparin: New life for an old drug. *Nanomedicine* **2017**, *12*, 1727–1744. [[CrossRef](#)] [[PubMed](#)]
- Motlekar, N.A.; Youan, B.-B.C. The quest for non-invasive delivery of bioactive macromolecules: A focus on heparins. *J. Control. Release* **2006**, *113*, 91–101. [[CrossRef](#)] [[PubMed](#)]
- Das Kurmi, B.; Tekchandani, P.; Paliwal, R.; Rai Paliwal, S. Nanocarriers in Improved Heparin Delivery: Recent Updates. *Curr. Pharm. Des.* **2015**, *21*, 4509–4518. [[CrossRef](#)] [[PubMed](#)]
- Lee, J.H.; Yeo, Y. Controlled drug release from pharmaceutical nanocarriers. *Chem. Eng. Sci.* **2015**, *125*, 75–84. [[CrossRef](#)] [[PubMed](#)]
- Shi, J.; Votruba, A.R.; Farokhzad, O.C.; Langer, R. Nanotechnology in drug delivery and tissue engineering: From discovery to applications. *Nano Lett.* **2010**, *10*, 3223–3230. [[CrossRef](#)]
- Dong, W.; Wang, X.; Liu, C.; Zhang, X.; Zhang, X.; Chen, X.; Kou, Y.; Mao, S. Chitosan based polymer-lipid hybrid nanoparticles for oral delivery of enoxaparin. *Int. J. Pharm.* **2018**, *547*, 499–505. [[CrossRef](#)]
- Charoongchit, P.; Suksiriworapong, J.; Mao, S.; Sapin-Minet, A.; Maincent, P.; Junyaprasert, V.B. Investigation of cationized triblock and diblock poly( $\epsilon$ -caprolactone)-co-poly(ethylene glycol) copolymers for oral delivery of enoxaparin: In vitro approach. *Acta Biomater.* **2017**, *61*, 180–192. [[CrossRef](#)]
- Rawat, A.; Majumder, Q.H.; Ahsan, F. Inhalable large porous microspheres of low molecular weight heparin: In vitro and in vivo evaluation. *J. Control. Release* **2008**, *128*, 224–232. [[CrossRef](#)]
- Patel, B.; Gupta, V.; Ahsan, F. PEG-PLGA based large porous particles for pulmonary delivery of a highly soluble drug, low molecular weight heparin. *J. Control. Release* **2012**, *162*, 310–320. [[CrossRef](#)]
- Loira-Pastoriza, C.; Sapin-Minet, A.; Diab, R.; Grossiord, J.L.; Maincent, P. Low molecular weight heparin gels, based on nanoparticles, for topical delivery. *Int. J. Pharm.* **2012**, *426*, 256–262. [[CrossRef](#)] [[PubMed](#)]
- Walenga, J.; Hoppensteadt, D. Monitoring the new antithrombotic drugs. *Semin. Thromb. Hemost.* **2005**, *30*, 683–695. [[CrossRef](#)] [[PubMed](#)]
- Samama Meyer, M.; Guinet, C. Laboratory assessment of new anticoagulants. *Clin. Chem. Lab. Med.* **2011**, *49*, 761–772. [[CrossRef](#)]
- Mulloy, B.; Hogwood, J.; Gray, E.; Lever, R.; Page, C.P. Pharmacology of heparin and related drugs. *Pharmacol. Rev.* **2016**, *68*, 76–141. [[CrossRef](#)] [[PubMed](#)]
- Torri, G.; Naggi, A. Heparin centenary—An ever-young life-saving drug. *Int. J. Cardiol.* **2016**, *212*, S1–S4. [[CrossRef](#)]
- Onishi, A.; St Ange, K.; Dordick, J.S.; Linhardt, R.J. Heparin and anticoagulation. *Front. Biosci. (Landmark Ed.)* **2016**, *21*, 1372–1392. [[CrossRef](#)] [[PubMed](#)]
- Gray, E.; Mulloy, B.; Barrowcliffe, T.W. Heparin and low-molecular-weight heparin. *Thromb. Haemost.* **2008**, *99*, 807–818. [[CrossRef](#)] [[PubMed](#)]



18. Harter, K.; Levine, M.; Henderson, S.O. Anticoagulation drug therapy: A review. *West. J. Emerg. Med.* **2015**, *16*, 11–17. [[CrossRef](#)] [[PubMed](#)]
19. Vene, N.; Mavri, A. An overview of the anticoagulant drugs used in routine clinical practice. In *Anticoagulant Drugs*; Božič-Mijovski, M., Ed.; IntechOpen: London, UK, 2018; p. 10. [[CrossRef](#)]
20. Casu, B.; Oreste, P.; Torri, G.; Zoppetti, G.; Choay, J.; Lormeau, J.C.; Petitou, M.; Sinaç, P. The structure of heparin oligosaccharide fragments with high anti-(factor Xa) activity containing the minimal antithrombin III-binding sequence. *Biochem. J.* **1981**, *197*, 599–609. [[CrossRef](#)] [[PubMed](#)]
21. Lindahl, U.; Bäckström, G.; Höök, M.; Thunberg, L.; Fransson, L.-A.; Linker, A. Structure of the antithrombin-binding site in heparin. *Proc. Natl. Acad. Sci. USA* **1979**, *76*, 3198–3202. [[CrossRef](#)]
22. Cosmi, B.; Palareti, G. Old and new heparins. *Thromb. Res.* **2012**, *129*, 388–391. [[CrossRef](#)] [[PubMed](#)]
23. Kakkar, A.K. Low- and ultra-low-molecular-weight heparins. *Best Pract. Res. Clin. Haematol.* **2004**, *17*, 77–87. [[CrossRef](#)] [[PubMed](#)]
24. Ibrahim, S.S.; Osman, R.; Awad, G.A.S.; Mortada, N.D.; Geneidy, A.-S. Low molecular weight heparins for current and future uses: Approaches for micro- and nano-particulate delivery. *Drug. Deliv.* **2016**, *23*, 2661–2667. [[CrossRef](#)] [[PubMed](#)]
25. Park, J.; Byun, Y. Recent advances in anticoagulant drug delivery. *Expert Opin. Drug Deliv.* **2016**, *13*, 421–434. [[CrossRef](#)]
26. Merli, G.J.; Vanscoy, G.J.; Rihn, T.L.; Groce Iii, J.B.; McCormick, W. Applying scientific criteria to therapeutic interchange: A balanced analysis of low-molecular-weight heparins. *J. Thromb. Thrombolysis* **2001**, *11*, 247–259. [[CrossRef](#)]
27. Hirsh, J.; O'Donnell, M.; Eikelboom, J.W. Beyond unfractionated heparin and warfarin. *Circulation* **2007**, *116*, 552–560. [[CrossRef](#)]
28. Walenga, J.M.; Lyman, G.H. Evolution of heparin anticoagulants to ultra-low-molecular-weight heparins: A review of pharmacologic and clinical differences and applications in patients with cancer. *Crit. Rev. Oncol. Hematol.* **2013**, *88*, 1–18. [[CrossRef](#)]
29. Masuko, S.; Linhardt, R.J. Chemoenzymatic synthesis of the next generation of ultralow MW heparin therapeutics. *Future Med. Chem.* **2012**, *4*, 289–296. [[CrossRef](#)]
30. Bauer, K.A. Fondaparinux sodium: A selective inhibitor of factor Xa. *Am. J. Health Syst. Pharm.* **2001**, *58*, S14–S17. [[CrossRef](#)]
31. Gómez-Outes, A.; Suárez-Gea, M.L.; Lecumberri, R.; Rocha, E.; Pozo-Hernández, C.; Vargas-Castrillón, E. New parenteral anticoagulants in development. *Ther. Adv. Cardiovasc. Dis.* **2011**, *5*, 33–59. [[CrossRef](#)]
32. Savi, P.; Herault, J.P.; Duchaussoy, P.; Millet, L.; Schaeffer, P.; Petitou, M.; Bono, F.; Herbert, J.M. Reversible biotinylated oligosaccharides: A new approach for a better management of anticoagulant therapy. *J. Thromb. Haemost.* **2008**, *6*, 1697–1706. [[CrossRef](#)]
33. Paty, I.; Trellu, M.; Destors, J.-M.; Cortez, P.; Boëlle, E.; Sanderink, G. Reversibility of the anti-FXa activity of idrabiotaparinux (biotinylated idraparinux) by intravenous avidin infusion. *J. Thromb. Haemost.* **2010**, *8*, 722–729. [[CrossRef](#)]
34. Falkon, L.; Garí, M.; Barbanj, M.; Amiral, J.; Fontcuberta, J. Tissue factor pathway inhibitor and anti-FXa kinetic profiles of a new low-molecular-mass heparin, bemiparin, at therapeutic subcutaneous doses. *Blood Coagul. Fibrinolysis* **1998**, *9*, 137–141. [[CrossRef](#)]
35. Ciccone, M.M.; Cortese, F.; Corbo, F.; Corrales, N.E.; Al-Momen, A.K.; Silva, A.; Zito, A.; Pinto, M.; Gesualdo, M.; Scicchitano, P. Bemiparin, an effective and safe low molecular weight heparin: A review. *Vasc. Pharmacol.* **2014**, *62*, 32–37. [[CrossRef](#)]
36. Akhtar, F.; Wan, X.; Wu, G.; Kesse, S.; Wang, S.; He, S. Low-Molecular-Weight Heparins: Reduced Size Particulate Systems for Improved Therapeutic Outcomes. *Molecules* **2018**, *23*, 1757. [[CrossRef](#)]
37. Schluter, A.; Lamprecht, A. Current Developments for the Oral Delivery of Heparin. *Curr. Pharm. Biotechnol.* **2014**, *15*, 640–649. [[CrossRef](#)]
38. Pirmohamed, M.; Kamali, F.; Daly, A.K.; Wadelius, M. Oral anticoagulation: A critique of recent advances and controversies. *Trends Pharm. Sci.* **2015**, *36*, 153–163. [[CrossRef](#)]
39. Ferreira, J.L.; Wipf, J.E. Pharmacologic therapies in anticoagulation. *Med. Clin. N. Am.* **2016**, *100*, 695–718. [[CrossRef](#)]
40. Ibrahim, T.F.; Maxwell, S.; Iqbal, O. Current anticoagulation drugs and mechanisms of action. In *Anticoagulation and Hemostasis in Neurosurgery*; Loftus, C.M., Ed.; Springer: Cham, Switzerland, 2016; pp. 33–46. [[CrossRef](#)]
41. Hirsh, J. Oral anticoagulant drugs. *N. Engl. J. Med.* **1991**, *324*, 1865–1875. [[CrossRef](#)]
42. Ageno, W.; Gallus, A.S.; Wittkowsky, A.; Crowther, M.; Hylek, E.M.; Palareti, G. Oral anticoagulant therapy: Antithrombotic therapy and prevention of thrombosis, 9th ed: American college of chest physicians evidence-based clinical practice guidelines. *Chest* **2012**, *141*, e44S–e88S. [[CrossRef](#)]
43. Wadelius, M.; Pirmohamed, M. Pharmacogenetics of warfarin: Current status and future challenges. *Pharmacogenomics J.* **2006**, *7*, 99–111. [[CrossRef](#)]
44. Ufer, M. Comparative pharmacokinetics of vitamin K antagonists. *Clin. Pharmacokinet.* **2005**, *44*, 1227–1246. [[CrossRef](#)] [[PubMed](#)]
45. Mega, J.L.; Simon, T. Pharmacology of antithrombotic drugs: An assessment of oral antiplatelet and anticoagulant treatments. *Lancet* **2015**, *386*, 281–291. [[CrossRef](#)]
46. Wysowski, D.K.; Nourjah, P.; Swartz, L. Bleeding complications with warfarin use: A prevalent adverse effect resulting in regulatory action. *Arch. Intern. Med.* **2007**, *167*, 1414–1419. [[CrossRef](#)]
47. Franchini, M.; Liumbruno, G.M.; Bonfanti, C.; Lippi, G. The evolution of anticoagulant therapy. *Blood. Transfus.* **2016**, *14*, 175–184. [[CrossRef](#)]
48. Riva, N.; Ageno, W. Pros and cons of vitamin K antagonists and non-vitamin K antagonist oral anticoagulants. *Semin. Thromb. Hemost.* **2015**, *41*, 178–187. [[CrossRef](#)]
49. Schulman, S. Advantages and limitations of the new anticoagulants. *J. Intern. Med.* **2014**, *275*, 1–11. [[CrossRef](#)]
50. Bauer, K.A. Pros and cons of new oral anticoagulants. *Hematol. Am. Soc. Hematol. Educ. Program.* **2013**, *2013*, 464–470. [[CrossRef](#)]

51. Stangier, J. Clinical pharmacokinetics and pharmacodynamics of the oral direct thrombin inhibitor dabigatran etexilate. *Clin. Pharmacokinet.* **2008**, *47*, 285–295. [[CrossRef](#)] [[PubMed](#)]
52. Zacconi, F.C. FXa direct synthetic inhibitors. In *Anticoagulant Drugs*; Božič-Mijovski, M., Ed.; IntechOpen: London, UK, 2018; pp. 11–37. [[CrossRef](#)]
53. Hinojar, R.; Jiménez-Natcher, J.J.; Fernández-Golfín, C.; Zamorano, J.L. New oral anticoagulants: A practical guide for physicians. *Eur. Heart J. Cardiovasc. Pharmacother.* **2015**, *1*, 134–145. [[CrossRef](#)]
54. Laux, V.; Perzborn, E.; Kubitzka, D.; Misselwitz, F. Preclinical and clinical characteristics of rivaroxaban: A novel, oral, direct factor Xa inhibitor. *Semin. Thromb. Hemost.* **2007**, *33*, 515–523. [[CrossRef](#)] [[PubMed](#)]
55. He, K.; He, B.; Grace, J.E.; Xin, B.; Zhang, D.; Pinto, D.J.; Luettgen, J.M.; Knabb, R.M.; Lam, P.Y.S.; Wexler, R.R.; et al. Preclinical pharmacokinetic and metabolism of apixaban, a potent and selective factor Xa inhibitor. *Blood* **2006**, *108*, 910. [[CrossRef](#)]
56. Minguet, J.; Sims, H.M.; Smith, K.H.; Bramlage, P. The factor Xa inhibitor edoxaban for the prevention of stroke and systemic embolism in patients with atrial fibrillation. *Expert Rev. Clin. Pharmacol.* **2017**, *10*, 5–15. [[CrossRef](#)]
57. Poulakos, M.; Walker, J.N.; Baig, U.; David, T. Edoxaban: A direct oral anticoagulant. *Am. J. Health Syst. Pharm.* **2017**, *74*, 117–129. [[CrossRef](#)] [[PubMed](#)]
58. Parasrampur, D.A.; Truitt, K.E. Pharmacokinetics and pharmacodynamics of edoxaban, a non-vitamin K antagonist oral anticoagulant that inhibits clotting factor Xa. *Clin. Pharmacokinet.* **2016**, *55*, 641–655. [[CrossRef](#)] [[PubMed](#)]
59. Garland, S.G.; DeRemer, C.E.; Smith, S.M.; Gums, J.G. Betrixaban: A new oral factor Xa inhibitor for extended venous thromboembolism prophylaxis in high-risk hospitalized patients. *Ann. Pharmacother.* **2018**, *52*, 554–561. [[CrossRef](#)]
60. Huisman, M.V.; Klok, F.A. Pharmacological properties of betrixaban. *Eur. Heart J. Suppl.* **2018**, *20*, E12–E15. [[CrossRef](#)]
61. Thoenes, M.; Minguet, J.; Bramlage, K.; Bramlage, P.; Ferrero, C. Betrixaban—the next direct factor Xa inhibitor? *Expert Rev. Hematol.* **2016**, *9*, 1111–1117. [[CrossRef](#)]
62. Cosmi, B. An update on the pharmaceutical management of thrombosis. *Expert Opin. Pharmacother.* **2016**, *17*, 2149–2164. [[CrossRef](#)]
63. Gómez-Outes, A.; Suárez-Gea, M.L.; Lecumberri, R.; Terleira-Fernández, A.I.; Vargas-Castrillón, E. Direct-acting oral anticoagulants: Pharmacology, indications, management, and future perspectives. *Eur. J. Haematol.* **2015**, *95*, 389–404. [[CrossRef](#)]
64. Favalaro, E.J.; Lippi, G.; Koutts, J. Laboratory testing of anticoagulants: The present and the future. *Pathology* **2011**, *43*, 682–692. [[CrossRef](#)] [[PubMed](#)]
65. Czuprynska, J.; Patel, J.P.; Arya, R. Current challenges and future prospects in oral anticoagulant therapy. *Br. J. Haematol.* **2017**, *178*, 838–851. [[CrossRef](#)] [[PubMed](#)]
66. Moffat, K.; Lewis, C. Laboratory monitoring of oral vitamin K anticoagulation. *Semin. Thromb. Hemost.* **2017**, *43*. [[CrossRef](#)]
67. Langdell, R.D.; Wagner, R.H.; Brinkhous, K.M. Effect of antihemophilic factor on one-stage clotting tests. A presumptive test for hemophilia and a simple one-stage antihemophilic factor assay procedure. *J. Lab. Clin. Med.* **1953**, *41*, 637–647.
68. Kitchens, C.S. To bleed or not to bleed? Is that the question for the PTT? *J. Thromb. Haemost.* **2005**, *3*, 2607–2611. [[CrossRef](#)]
69. Chee, Y.L.; Greaves, M. Role of coagulation testing in predicting bleeding risk. *Hematol. J.* **2003**, *4*, 373–378. [[CrossRef](#)]
70. Bonhomme, F.; Fontana, P. Laboratory testing of hemostasis. In *Perioperative hemostasis*; Marcucci, C.E., Schoettker, P., Eds.; Springer: Berlin/Heidelberg, Germany, 2015; pp. 13–24. [[CrossRef](#)]
71. Avecilla, S.T.; Roshal, M.; Reyes Gil, M.; Erdman, P.A. Laboratory monitoring for heparins, fondaparinux, direct thrombin inhibitors, and oral anti-Xa medications. In *Transfusion Medicine and Hemostasis*, 3rd ed.; Shaz, B.H., Hillyer, C.D., Reyes Gil, M., Eds.; Elsevier: Amsterdam, The Netherlands, 2019; pp. 933–938. [[CrossRef](#)]
72. Cuker, A. Unfractionated heparin for the treatment of venous thromboembolism: Best practices and areas of uncertainty. *Semin. Thromb. Hemost.* **2012**, *38*, 593–599. [[CrossRef](#)]
73. Marlar, R.A.; Clement, B.; Gausman, J. Activated partial thromboplastin time monitoring of unfractionated heparin therapy: Issues and recommendations. *Semin. Thromb. Hemost.* **2017**, *43*, 253–260. [[CrossRef](#)]
74. Shapiro, G.A.; Huntzinger, S.W.; Wilson, J.E. Variation among commercial activated partial thromboplastin time reagents in response to heparin. *Am. J. Clin. Pathol.* **1977**, *67*, 477–480. [[CrossRef](#)]
75. Watson, H.G.; Greaves, M. Can we predict bleeding? *Semin. Thromb. Hemost.* **2008**, *34*, 97–103. [[CrossRef](#)]
76. Cuker, A.; Raby, A.; Moffat, K.; Flynn, G.; Crowther, M. Interlaboratory variation in heparin monitoring: Lessons from the quality management program of Ontario coagulation surveys. *Thromb. Haemost.* **2010**, *104*, 837–844. [[CrossRef](#)] [[PubMed](#)]
77. Babin, J.L.; Traylor, K.; Witt, D. Laboratory monitoring of low-molecular-weight heparin and fondaparinux. *Semin. Thromb. Hemost.* **2017**, *43*, 261–269. [[CrossRef](#)] [[PubMed](#)]
78. Gehrie, E.; Laposata, M. Test of the month: The chromogenic antifactor Xa assay. *Am. J. Hematol.* **2012**, *87*, 194–196. [[CrossRef](#)] [[PubMed](#)]
79. Bates, S.M.; Weitz, J.I. Coagulation assays. *Circulation* **2005**, *112*, 53–60. [[CrossRef](#)] [[PubMed](#)]
80. Newall, F. Anti-factor Xa (anti-Xa) assay. In *Haemostasis. Methods in Molecular Biology (Methods and Protocols)*; Monagle, P., Ed.; Humana Press: Totowa, NJ, USA, 2013; Volume 992, pp. 265–272.
81. Funk, D.M. Coagulation assays and anticoagulant monitoring. *Hematol. Am. Soc. Hematol. Educ. Program.* **2012**, *2012*, 460–465. [[CrossRef](#)]
82. Samuelson, B.T.; Cuker, A. Measurement and reversal of the direct oral anticoagulants. *Blood Rev.* **2017**, *31*, 77–84. [[CrossRef](#)]

83. Samuelson, B.T.; Cuker, A.; Siegal, D.M.; Crowther, M.; Garcia, D.A. Laboratory assessment of the anticoagulant activity of direct oral anticoagulants: A systematic review. *Chest* **2017**, *151*, 127–138. [[CrossRef](#)]
84. Eby, C. Novel anticoagulants and laboratory testing. *Int. J. Lab. Hematol.* **2013**, *35*, 262–268. [[CrossRef](#)]
85. Adcock, D.M.; Gosselin, R. Direct oral anticoagulants (DOACs) in the laboratory: 2015 review. *Thromb. Res.* **2015**, *136*, 7–12. [[CrossRef](#)]
86. Schmitz, E.M.H.; Boonen, K.; van den Heuvel, D.J.A.; van Dongen, J.L.J.; Schellings, M.W.M.; Emmen, J.M.A.; van der Graaf, F.; Brunsveld, L.; van de Kerkhof, D. Determination of dabigatran, rivaroxaban and apixaban by ultra-performance liquid chromatography—tandem mass spectrometry (UPLC-MS/MS) and coagulation assays for therapy monitoring of novel direct oral anticoagulants. *J. Thromb. Haemost.* **2014**, *12*, 1636–1646. [[CrossRef](#)]
87. Lippi, G.; Favaloro, E. Recent guidelines and recommendations for laboratory assessment of the direct oral anticoagulants (DOACs): Is there consensus? *Clin. Chem. Lab. Med.* **2015**, *53*, 185–197. [[CrossRef](#)]
88. Dale, B.J.; Chan, N.C.; Eikelboom, J.W. Laboratory measurement of the direct oral anticoagulants. *Br. J. Haematol.* **2016**, *172*, 315–336. [[CrossRef](#)]
89. van Ryn, J.; Stangier, J.; Härtter, S.; Liesenfeld, K.-H.; Wienen, W.; Feuring, M.; Clemens, A. Dabigatran etexilate—a novel, reversible, oral direct thrombin inhibitor: Interpretation of coagulation assays and reversal of anticoagulant activity. *Thromb. Haemost.* **2010**, *103*, 1116–1127. [[CrossRef](#)] [[PubMed](#)]
90. Pollack, C.V. Coagulation assessment with the new generation of oral anticoagulants. *Emerg. Med. J.* **2016**, *33*, 423–430. [[CrossRef](#)] [[PubMed](#)]
91. Douxflis, J.; Mullier, F.; Robert, S.; Chatelain, C.; Chatelain, B.; Dogne, J.M. Impact of dabigatran on a large panel of routine or specific coagulation assays. Laboratory recommendations for monitoring of dabigatran etexilate. *Thromb. Haemost.* **2012**, *107*, 985–997. [[CrossRef](#)]
92. Nowak, G. The ecarin clotting time, a universal method to quantify direct thrombin inhibitors. *Pathophysiol. Haemost. Thromb.* **2003**, *33*, 173–183. [[CrossRef](#)] [[PubMed](#)]
93. Gosselin, R.C.; Dwyre, D.M.; Dager, W.E. Measuring dabigatran concentrations using a chromogenic ecarin clotting time assay. *Ann. Pharmacother.* **2013**, *47*, 1635–1640. [[CrossRef](#)]
94. Ahmed, K.S.; Hussein, S.A.; Ali, A.H.; Korma, S.A.; Lipeng, Q.; Jinghua, C. Liposome: Composition, characterisation, preparation, and recent innovation in clinical applications. *J. Drug Target.* **2019**, *27*, 742–761. [[CrossRef](#)]
95. Liu, G.; Hou, S.; Tong, P.; Li, J. Liposomes: Preparation, Characteristics, and Application Strategies in Analytical Chemistry. *Crit. Rev. Anal. Chem.* **2020**, 1–21. [[CrossRef](#)]
96. Song, Y.-K.; Hyun, S.Y.; Kim, H.-T.; Kim, C.-K.; Oh, J.-M. Transdermal delivery of low molecular weight heparin loaded in flexible liposomes with bioavailability enhancement: Comparison with ethosomes. *J. Microencapsul.* **2011**, *28*, 151–158. [[CrossRef](#)]
97. Bai, S.; Ahsan, F. Inhalable liposomes of low molecular weight heparin for the treatment of venous thromboembolism. *J. Pharm. Sci.* **2010**, *99*, 4554–4564. [[CrossRef](#)] [[PubMed](#)]
98. Croisfelt, F.M.; Tundisi, L.L.; Ataide, J.A.; Silveira, E.; Tambourgi, E.B.; Jozala, A.F.; Souto, E.M.B.; Mazzola, P.G. Modified-release topical hydrogels: A ten-year review. *J. Mater. Sci.* **2019**, *54*, 10963–10983. [[CrossRef](#)]
99. Narayanaswamy, R.; Torchilin, V.P. Hydrogels and Their Applications in Targeted Drug Delivery. *Molecules (Basel, Switzerland)* **2019**, *24*, 603. [[CrossRef](#)] [[PubMed](#)]
100. Jacob, S.; Nair, A.B.; Shah, J.; Sreeharsha, N.; Gupta, S.; Shinu, P. Emerging Role of Hydrogels in Drug Delivery Systems, Tissue Engineering and Wound Management. *Pharmaceutics* **2021**, *13*, 357. [[CrossRef](#)] [[PubMed](#)]
101. Matanović, M.R.; Grabnar, P.A.; Voinovich, D.; Golob, S.; Mijovski, M.B.; Grabnar, I. Development and preclinical pharmacokinetics of a novel subcutaneous thermoresponsive system for prolonged delivery of heparin. *Int. J. Pharm.* **2015**, *496*, 583–592. [[CrossRef](#)] [[PubMed](#)]
102. Sur, S.; Rathore, A.; Dave, V.; Reddy, K.R.; Chouhan, R.S.; Sadhu, V. Recent developments in functionalized polymer nanoparticles for efficient drug delivery system. *Nano-Struct. Nano-Objects* **2019**, *20*, 100397. [[CrossRef](#)]
103. Singh, R.; Lillard, J.W. Nanoparticle-based targeted drug delivery. *Exp. Mol. Pathol.* **2009**, *86*, 215–223. [[CrossRef](#)]
104. Liu, Z.; Jiao, Y.; Wang, Y.; Zhou, C.; Zhang, Z. Polysaccharides-based nanoparticles as drug delivery systems. *Adv. Drug Deliv. Rev.* **2008**, *60*, 1650–1662. [[CrossRef](#)]
105. Rizvi, S.A.A.; Saleh, A.M. Applications of nanoparticle systems in drug delivery technology. *Saudi Pharma. J.* **2018**, *26*, 64–70. [[CrossRef](#)]
106. Ramadan, A.; Lagarce, F.; Tessier-Marteau, A.; Thomas, O.; Legras, P.; Macchi, L.; Saulnier, P.; Benoit, J.-P. Oral fondaparinux: Use of lipid nanocapsules as nanocarriers and in vivo pharmacokinetic study. *Int. J. Nanomed.* **2011**, *6*, 2941–2951. [[CrossRef](#)]
107. Souto, E.B.; Baldim, I.; Oliveira, W.P.; Rao, R.; Yadav, N.; Gama, F.M.; Mahant, S. SLN and NLC for topical, dermal and transdermal drug delivery. *Expert Opin. Drug Deliv.* **2020**, *17*, 357–377. [[CrossRef](#)] [[PubMed](#)]
108. Mahant, S.; Rao, R.; Souto, E.B.; Nanda, S. Analytical tools and evaluation strategies for nanostructured lipid carrier based topical delivery systems. *Expert Opin. Drug Deliv.* **2020**. [[CrossRef](#)]
109. Paliwal, R.; Paliwal, S.R.; Agrawal, G.P.; Vyas, S.P. Biomimetic solid lipid nanoparticles for oral bioavailability enhancement of low molecular weight heparin and its lipid conjugates: In vitro and in vivo evaluation. *Mol. Pharm.* **2011**, *8*, 1314–1321. [[CrossRef](#)] [[PubMed](#)]

110. Rehman, F.U.; Shah, K.U.; Shah, S.U.; Khan, I.U.; Khan, G.M.; Khan, A. From nanoemulsions to self-nanoemulsions, with recent advances in self-nanoemulsifying drug delivery systems (SNEDDS). *Expert Opin. Drug Deliv.* **2017**, *14*, 1325–1340. [[CrossRef](#)] [[PubMed](#)]
111. Cherniakov, I.; Domb, A.J.; Hoffman, A. Self-nano-emulsifying drug delivery systems: An update of the biopharmaceutical aspects. *Expert Opin. Drug Deliv.* **2015**, *12*, 1121–1133. [[CrossRef](#)]
112. Zupančič, O.; Grießinger, J.A.; Rohrer, J.; Pereira de Sousa, I.; Danninger, L.; Partenhauser, A.; Sündermann, N.E.; Laffleur, F.; Bernkop-Schnürch, A. Development, in vitro and in vivo evaluation of a self-emulsifying drug delivery system (SEDDS) for oral enoxaparin administration. *Eur. J. Pharm. Biopharm.* **2016**, *109*, 113–121. [[CrossRef](#)] [[PubMed](#)]
113. Lavanya, N.; Muzib, Y.I.; Aukunuru, J.; Balekari, U. Preparation and evaluation of a novel oral delivery system for low molecular weight heparin. *Int. J. Pharm. Investig.* **2016**, *6*, 148–157. [[CrossRef](#)] [[PubMed](#)]
114. Fang, G.; Zhou, J.; Qian, Y.; Gou, J.; Yang, X.; Tang, B. Development and evaluation of thermo-sensitive hydrogel system with nanocomplexes for prolonged subcutaneous delivery of enoxaparin. *J. Drug Deliv. Sci. Technol.* **2018**, *48*, 118–124. [[CrossRef](#)]
115. Gritsch, L.; Motta, F.L.; Contessi Negrini, N.; Yahia, L.H.; Farè, S. Crosslinked gelatin hydrogels as carriers for controlled heparin release. *Mater. Lett.* **2018**, *228*, 375–378. [[CrossRef](#)]
116. Eleraky, N.E.; Swarnakar, N.K.; Mohamed, D.F.; Attia, M.A.; Pauletti, G.M. Permeation-Enhancing Nanoparticle Formulation to Enable Oral Absorption of Enoxaparin. *AAPS PharmSciTech* **2020**, *21*, 88. [[CrossRef](#)]
117. Tang, B.; Qian, Y.; Fang, G. Development of Lipid-Polymer Hybrid Nanoparticles for Improving Oral Absorption of Enoxaparin. *Pharmaceutics* **2020**, *12*, 607. [[CrossRef](#)] [[PubMed](#)]
118. Soltani, Y.; Goodarzi, N.; Mahjub, R. Preparation and characterization of self nano-emulsifying drug delivery system (SNEDDS) for oral delivery of heparin using hydrophobic complexation by cationic polymer of  $\beta$ -cyclodextrin. *Drug Dev. Ind. Pharm.* **2017**, *43*, 1899–1907. [[CrossRef](#)] [[PubMed](#)]
119. Xue, X.; Cao, M.; Ren, L.; Qian, Y.; Chen, G. Preparation and Optimization of Rivaroxaban by Self-Nanoemulsifying Drug Delivery System (SNEDDS) for Enhanced Oral Bioavailability and No Food Effect. *AAPS PharmSciTech* **2018**, *19*, 1847–1859. [[CrossRef](#)] [[PubMed](#)]
120. Abouhoussein, D.M.N.; Bahaa El Din Mahmoud, D.; Mohammad, F.E. Design of a liquid nano-sized drug delivery system with enhanced solubility of rivaroxaban for venous thromboembolism management in paediatric patients and emergency cases. *J. Liposome Res.* **2019**, *29*, 399–412. [[CrossRef](#)]