

1 **SYNTHESIS OF NEW *N*-PHENYL-*N*-(1-PHENYLHEX-5-EN-1-YL)ACETAMIDES**
2 **AND THEIR ¹H-NMR CONFORMATIONAL STUDY**

3
4 **SÍNTESIS DE NOVAS *N*-FENIL-*N*-(1-FENILHEX-5-EN-1-IL)ACETAMIDAS E O**
5 **SEU ESTUDO CONFORMACIONAL POR ¹H-RMN**

6
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15
16 **Abstract**

17 The synthesis and characterization of different *N*-phenyl-*N*-(1-phenylhex-5-en-1-yl)acetamides is presented. Two conformational isomers were observed for one of the
18 compounds in their ¹H/¹³C-NMR spectra. Computational calculations and dihedral angle
19 comparison using the allylic system coupling constants (*J*) were carried out to determine
20 the isomeric structures responsible for signals duplicity and chemical shifting.
21

22
23 **Key words:** acetamides, conformational isomers, computational calculation, NMR,
24 Garbisch equation.

25

Resumo

26 A síntese e caracterização de diferentes *N*-fenil-*N*-(1-fenilhex-5-en-1-il) acetamidas é
27 apresentada. Dois isómeros conformacionais foram observados para um dos compostos no
28 seu espectro de $^1\text{H}/^{13}\text{C}$ -RMN. Cálculos computacionais e comparação de ângulos diedros
29 usando as constantes de acoplamento (*J*) para o sistema alílico foram realizadas para
30 determinar as estruturas isoméricas responsáveis pela duplicidade de sinais e o
31 deslocamento químico.

32

33 **Palavras-chave:** acetamidas, isómeros conformacionais, cálculo computacional, RMN,
34 equação de Garbisch.

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Introduction

38

39 Target oriented synthesis (TOS) is one of the most important methodologies in organic
40 chemistry to access biologically active compounds (1). Molecules including quinoline and
41 tetrahydroquinoline derivatives are widely known for their biological and pharmacological
42 activity as well as for their uses in organic electronics (2). Previous reports from our
43 research group have shown both antifungal and antiparasitic activities of *N*-phenyl- α -2-
44 propen-1-yl benzenepropanamines **1a-e** (3-4). The use of **1a** as synthon for different *N*-
45 heterocycles containing the tetrahydrolepidine and quinolinemoiety is well known. These
46 substances are similar to isolated compounds from *Galipealongiflora* and
47 *Galipeaofficinalis*, which have been studied and used against fever, dysentery, malaria and
48 leishmaniasis treatment, among others (5-12).

49 Literature reports have shown that acetylation of *N*-(prop)butenylamines turns these
50 compounds into biologically active *N*-(prop)butenylacetamides (3,13-14). Thus, new *N*-
51 phenyl-*N*-(1-phenylhex-5-en-1-yl)acetamides **2a-f** were prepared by *N*-acetylation of
52 compounds **1a-f**. Their synthesis and ¹H/¹³C-NMR data is discussed and presented.
53 Biological activity of compounds **2a-f** is currently under study.

54

55

56 Experimental section (Materials and methods)

57

58 IR spectra were obtained on a FT-IR Bruker Tensor 27, using KBr windows. IR main
59 signals are condensed in **Table 1**. GC/MS data were acquired on a HP5890A Series II gas
60 chromatography equipped with a HP-5MS column (5% methyl phenyl siloxane, 30m x
61 0.25mm x 0.25µm) and a selective mass detector HP5972 (EI, 70 eV). NMR spectra were
62 recorded on a Bruker Avance-400. Coupling constants *J* are reported in Hertz. See **Scheme**
63 **1** for ¹H, ¹³C assignment.

64

65 *General procedure for the synthesis of N-phenyl-N-(1-phenylhex-5-en-1-yl) acetamides 2a-f*

66

67 A round bottom flask with a reflux condenser, a thermometer and a magnetic stirrer was
68 filled with 0.35 g (1.24 mmol) of *N*-phenyl- α -2-propen-1-ylbenzenpropanamine (previously
69 prepared) (**3**) and 3.80 g (37.02 mmol) of acetic anhydride. The reaction mixture was
70 refluxed for 4-6 h and neutralized using NaHCO₃. NaOH 0.1 M was used to adjust the
71 mixture to pH 12. Ethyl acetate was used for extraction (20 mL x 3). The organic layer was
72 dried over Na₂SO₄, the solvent removed and the crude product purified by column

73 chromatography on SiO₂ using *n*-heptane and ethyl acetate gradient mixtures. Compounds
74 characterization was carried out using IR, ¹H, ¹³C NMR and GC-MS.

75

76

77 *N*-phenyl-*N*-(1-phenylhex-5-en-1-yl)acetamide**2a**

78

79 From *N*-phenyl- α -2-propen-1-ylbenzenepropanamine**1a** (0.33 g, 1.31 mmol) and acetic
80 anhydride (4.32 g, 42.30 mmol). Pure compound **2a** was obtained after column
81 chromatography as a brownish oil. R_f= 0.50 (*n*-heptane:AcOEt, 5:2). MS [EI, 70 eV] (*m/z*,
82 %): 293 (M⁺, 2), 252 (38), 210 (100), 117 (13), 91 (47). δ_{H} ppm (CDCl₃, 400 MHz): 1.62
83 (td, ³*J*= 8.0, 7.3, 2H₅), 1.72 (s, 3H_{4c}), 2.03-2.19 (m, 2H₃), 2.63 (ddd, ²*J*= 14.4; ³*J*= 8.3, 7.6,
84 1H₆), 2.71 (ddd, ²*J*=14.4; ³*J*= 8.3, 7.6, 1H₆), 4.90-5.05 (m, 3H_{1,4}), 5.76 (dddd, ³*J*= 17.7, 9.5,
85 6.8, 6.4, 1H₂), 6.94-7.39 (m, 10H_{Arom}). δ_{C} ppm (CDCl₃, 100 MHz): 23.63_(4c), 33.13₍₆₎,
86 34.57₍₅₎, 37.75₍₃₎, 54.07₍₄₎, 117.11₍₁₎, 125.86_(p), 128.25_(2xm), 128.36_(2xo), 128.74_(p'),
87 129.36_(2xm'), 129.86_(2xo'), 135.66₍₂₎, 139.37_(i'), 141.81_(i), 171.03_(4b).

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90 *N*-Phenyl-*N*-(4-methylphenylhex-5-en-1-yl)acetamide**2b**

91

92 From of *N*-(4-methylphenyl)- α -2-propen-1-ylbenzenepropanamine **1b** (1.07 g, 4.03 mmol)
93 and acetic anhydride (10.79 g, 105.75 mmol). Pure compound was obtained after
94 chromatography column as a brownish oil. R_f= 0.50 (*n*-heptane:AcOEt, 5:2). MS [EI, 70
95 eV] (*m/z*, %): 307 (M⁺, 2), 266 (37), 224 (100), 150 (11), 91 (40). δ_{H} ppm (CDCl₃, 400
96 MHz): 1.68 (td, ³*J*= 8.1, 7.3, 2H₅), 1.79 (s, 3H_{4c}), 2.08-2.29 (m, 2H₃), 2.39 (s, 3H_{4d}), 2.70

97 (ddd, $^2J= 14.5$; $^3J= 8.1, 7.5, 1H_6$), 2.77 (ddd, $^2J= 14.5$; $^3J= 8.1, 7.5, 1H_6$), 4.93-5.18 (m,
98 $3H_{1,4}$), 5.83 (dddd, $^3J= 17.5, 9.3, 6.9, 6.5, 1H_2$), 7.06 (d, $^3J= 8.2, 2H_{6'}$), 7.14-7.33 (m,
99 $7H_{Arom}$). δ_C ppm (CDCl₃, 100 MHz): 20.98_(4d), 23.47_(4c), 33.04₍₆₎, 34.46₍₅₎, 37.69₍₃₎, 53.80₍₄₎,
100 116.95₍₁₎, 125.75_(p), 128.17_(2xm), 128.27_(2xo), 129.51_(2xo'), 129.90_(2xm'), 135.64₍₂₎, 136.49_(p'),
101 138.18_(i'), 141.79_(i), 171.16_(4b).

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103

104 *N*-Phenyl-*N*-(4-methoxyphenylhex-5-en-1-yl)acetamide**2c**

105

106 From *N*-(4-methoxyphenyl)- α -2-propen-1-ylbenzenepropanamine **1c** (0.35 g, 1.24 mmol)
107 and acetic anhydride (3.67 g, 35.96 mmol). Pure product was obtained after
108 chromatography column as a brownish oil. $R_f= 0.40$ (*n*-heptane:AcOEt, 5:2). MS [EI, 70
109 eV] (*m/z*, %): 323 (M^+ , 2), 282 (28), 240 (100), 91 (51). δ_H ppm (CDCl₃, 400 MHz): 1.67
110 (td, $^3J= 8.1, 7.3, 2H_5$), 1.80 (s, $3H_{4c}$), 2.06-2.30 (m, $2H_3$), 2.70 (ddd, $^2J= 14.4$; $^3J= 8.3, 7.6$,
111 $1H_6$), 2.77 (ddd, $^2J=14.4$; $^3J= 8.3, 7.6, 1H_6$), 3.83 (s, $3H_{4d}$), 4.98-5.14 (m, $3H_{1,4}$), 5.83
112 (dddd, $^3J=17.6, 9.5, 6.8, 6.5, 1H_2$), 6.92 (d, $^3J= 9.2, 2H_{6'}$), 7.04-7.35 (m, $7H_{Arom}$). δ_C ppm
113 (CDCl₃, 100 MHz): 23.55_(4c), 33.12₍₆₎, 34.54₍₅₎, 37.72₍₃₎, 53.78₍₄₎, 55.43_(4d), 114.43_(2xm'),
114 117.06₍₁₎, 125.87_(p), 128.27_(2xm), 128.39_(2xo), 130.87_(2xo'), 131.75_(i'), 135.76₍₂₎, 141.88_(i),
115 159.20_(p'), 171.57_(4b).

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117

118 *N*-Phenyl-*N*-(4-bromophenylhex-5-en-1-yl) acetamide**2d**

119

120 From *N*-(4-bromophenyl)- α -2-propen-1-ylbenzenpropanamine **1d** (0.68 g, 2.06 mmol) and
121 acetic anhydride (7.34 g, 71.91 mmol). Pure product was obtained after column
122 chromatography as a brownish oil. R_f = 0.47 (*n*-heptane:AcOEt, 5:2). MS [EI, 70 eV] (*m/z*,
123 %): 373 (M^+ , 2), 330 (33), 290 (95), 288 (100), 91 (82). δ_H ppm ($CDCl_3$, 400 MHz): 1.61-
124 1.73 (m, 2H₅), 1.79 (s, 3H_{4c}), 2.17 (ta, $^3J = 7.0$, 2H₃), 2.65-2.79 (m, 2H₆), 4.94-5.13 (m,
125 3H_{4,1}), 5.81 (dddd, $^3J = 17.2$, 10.1, 6.6, 6.3, 1H₂), 7.06 (d, $^3J = 8.8$, 2H_{0'}), 7.14-7.31 (m,
126 5H_{Arom}), 7.55 (d, $^3J = 8.8$, 2H_m). δ_C ppm ($CDCl_3$, 100 MHz): 23.63_(4c), 33.07₍₆₎, 34.59₍₅₎,
127 37.55₍₃₎, 54.07₍₄₎, 117.31₍₁₎, 122.39_(p'), 125.95_(p), 128.21_(2xm), 128.41_(2xo), 131.55_(2xo'),
128 132.62_(2xm'), 135.44₍₂₎, 138.42_(i'), 141.53_(i), 170.67_(4b).

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130

131 *N*-Phenyl-*N*-(4-fluorophenyl)hex-5-en-1-ylacetamide **2e**

132

133 From *N*-(4-fluorophenyl)- α -2-propen-1-ylbenzenpropanamine **1e** (0.63 g, 2.34 mmol) and
134 acetic anhydride (6.80 g, 66.62 mmol). Pure product was obtained after column
135 chromatography as a brownish oil. R_f = 0.43 (*n*-heptane:AcOEt, 5:2). MS [EI, 70 eV] (*m/z*,
136 %): 311 (M^+ , 2), 270 (36), 228 (100), 117 (15), 91 (51). δ_H ppm ($CDCl_3$, 400 MHz): 1.71-
137 1.82 (m, 2H₅), 1.87 (s, 3H_{4c}), 2.22-2.28 (m, 2H₃), 2.79 (ddd, $^2J = 13.8$; $^3J = 9.6$, 6.9, 1H₆),
138 2.84 (ddd, $^2J = 13.8$; $^3J = 9.6$, 6.9, 1H₆), 5.06-5.22 (m, 3H_{1,4}), 5.91 (dddd, $^3J = 17.0$, 10.6, 6.6,
139 6.3, 1H₂), 7.16-7.40 (m, 9H_{Arom}). δ_C ppm ($CDCl_3$, 100 MHz): 23.55_(4c), 33.03₍₆₎, 34.53₍₅₎,
140 37.52₍₃₎, 53.89₍₄₎, 116.28_(2xm') (d, $^2J = 22.5$), 117.21₍₁₎, 125.91_(p), 128.18_(2xm), 128.38_(2xo),
141 131.55_(2xo'), 135.19_(i') (d, $^4J = 3.4$), 135.49₍₂₎, 141.57_(i) 162.07_(p') (d, $^1J = 249.1$), 171.02_(4b).

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143

144 *N*-Phenyl-*N*-(2-methylphenyl)hex-5-en-1-yl)acetamide **2f**

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146 From *N*-(2-methylphenyl)- α -2-propen-1-ylbenzen propanamine **1f** (0.67 g, 2.52 mmol) and
147 acetic anhydride (7.23 g, 70.81 mmol). Pure product was obtained after column

148 chromatography as a brownish oil. R_f = 0.53 (*n*-heptane:AcOEt, 5:2). GC showed a single

149 signal. MS [EI, 70 eV] (*m/z*, %): 307 (M^+ , 2), 266 (39), 224 (100), 118 (13), 91 (51). NMR

150 data for α conformer: δ_H ppm ($CDCl_3$, 400 MHz): 1.73 (s, 3 H_{4c}), 1.74-1.82 (m, 2 H_5), 2.26

151 (s, 3 H_{4d}), 2.45-2.51; 2.52-2.59 (m, 2 H_3), 2.59-2.67 (m, 2 H_6), 4.75-4.86 (m, 1 H_4), 5.08-5.20

152 (m, 2 H_1), 5.89 (dddd, 3J = 17.3, 9.9, 6.7, 6.5, 1 H_2), 7.07-7.34 (m, 9 H_{Arom}). δ_C ppm ($CDCl_3$,

153 100 MHz): 18.44_(4d), 23.15_(4c), 33.45₍₆₎, 33.57₍₅₎, 38.23₍₃₎, 55.87₍₄₎, 117.08₍₁₎, 125.81_(p),

154 126.87_(m'), 128.21_(p'), 128.25_(2xo), 128.32_(2xm), 129.64_(o'), 131.54_(m''), 135.93₍₂₎, 136.87_(i'),

155 139.35_(o''), 141.67_(i), 171.06_(4b). NMR data for β conformer: δ_H ppm ($CDCl_3$, 400 MHz):

156 1.49-1.61 (m, 2 H_5), 1.77 (s, 3 H_{4c}), 1.86-2.07; 2.36-2.48 (m, 2 H_3), 2.27 (s, 3 H_{4d}), 2.77 (ta,

157 3J = 8.4, 2 H_6), 4.75-4.86 (m, 1 H_4), 4.95-5.01 (m, 2 H_1), 5.69 (dddd, 3J = 16.6, 10.8, 7.9, 6.0,

158 1 H_2), 7.07-7.34 (m, 9 H_{Arom}). δ_C ppm ($CDCl_3$, 100 MHz): 18.49_(4d), 23.06_(4c), 33.14₍₆₎,

159 33.31₍₅₎, 36.61₍₃₎, 55.83₍₄₎, 117.18₍₁₎, 125.81_(p), 126.95_(m'), 128.17_(p'), 128.24_(2xo), 128.34_(2xm),

160 129.32_(o'), 131.60_(m''), 135.42₍₂₎, 137.02_(i'), 139.22_(o''), 141.97_(i), 171.04_(4b).

161

162

163 *Conformational analysis*

164

165 Ten chemically reasonable structures for compound **2f** were used as the starting point for

166 energy minimization using the Parameterized Model 3 (PM3) semiempirical method (15).

167 Geometry optimization and vibrational frequency calculations (for verifying that the

168 structures correspond to minima on the potential energy surface) were then carried out
169 using the B3LYP functional with the 6-31+G(2d,p) basis set as implemented in Gaussian
170 09 (16-24). Seven different possible conformer structures **2f₁-2f₇** were found. Energy
171 differences [kcal/mol] for all minima were calculated with and without vibrational zero
172 point energies (VZPE).

173

174

175

Results and discussion

176

177 **Scheme 1** shows the synthesis of new 1-phenylhex-5-en-1-ylacetamides **2a-f** from *N*-
178 butenylamines **1a-f** using acetic anhydride as both reagent and solvent, under reflux (140
179 °C). Acetamides **2a-f** were easily obtained with yields between 66-92%. Optimal reaction
180 time was established by both TLC and GC in about 4-6 h. Main IR vibration bands of allyl
181 C=C-H and acetamide C=O are listed in **Table 1**. IR N-H tension and flexion bands in
182 compounds **1a-f** clearly disappeared after acetyl protection.

183

184

185 **Scheme 1**. Synthesis of acetamides **2a-f** (see labels for ¹H and ¹³C-NMR assignments in
186 experimental section).

187

188

189 **Table 1.** IR [ν , cm^{-1}] main signals for compounds **2a-f**.

	$\nu(\text{C}=\text{O})$	$\nu(\text{C}=\text{CH})$	$\nu(\text{C}=\text{H})$ Aromatic <i>p</i> -disubstituted	$\nu(\text{C}=\text{H})$ Aromatic monosubstituted
2a	1655	915	----	745-701
2b	1655	915	824	749-700
2c	1653	916	837	750-700
2d	1658	917	833	749-700
2e	1658	917	844	750-700
2f	1658	917	763*	749-700

190 * *o*- disubstitued

191

192 GC-MS (EI, 70eV) data are condensed in **Table 2**. Molecular ions corresponding to
 193 molecular mass of **2a-f** appeared in all cases with low intensity; base peak Φ_2 results after
 194 consecutive allyl [Φ_1] and acetyl [$(\Phi_1\text{-C}_2\text{H}_3\text{O})^+$] fragmentation. Benzyl fragment Φ_3 (m/z 91)
 195 is characteristic in all compounds.

196

197 **Table 2.** MS (EI, 70eV) [m/z (int. %)] main fragments for compounds **2a-f**.

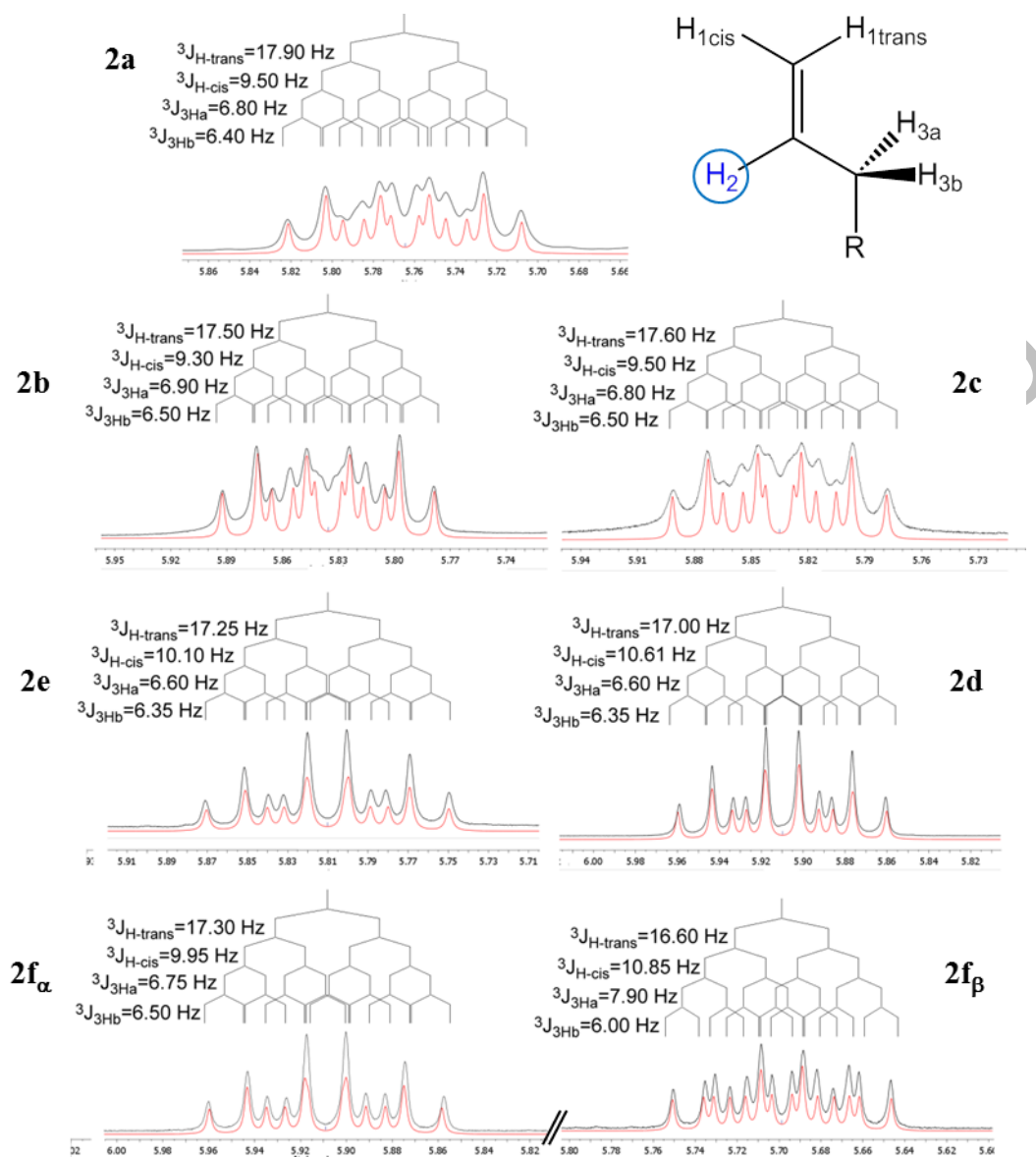
	M^+	$\Phi_1, [\text{M}-\text{C}_3\text{H}_5]^+$	$\Phi_2, [\Phi_1\text{-C}_2\text{H}_3\text{O}]^+$	$\Phi_3, [\text{C}_7\text{H}_7]^+$
2a	293 (2)	252 (38)	210 (100)	91 (47)
2b	307 (2)	266 (37)	224 (100)	91 (40)
2c	323 (2)	282 (28)	240 (100)	91 (51)
2d	373 (2)	330 (33)	288(100)	91 (82)
2e	311 (2)	270 (33)	228 (100)	91 (51)
2f	307 (2)	266 (39)	224 (100)	91 (51)

198

199 ^1H , ^{13}C and 2D NMR experiments confirmed the structures of compounds **2a-f**. Due to a
 200 short relaxation time in ^{13}C -APT NMR spectra for *Co'*, a broad signal appeared; thus, in **2e**

201 it was not possible to assign the correct $C o'-F$ coupling constant. Diastereotopic protons
202 H_{3a} and H_{3b} appeared in 1H -NMR as a multiplet from 1.86 ppm to 2.59 ppm in all cases.
203 Correct assignment of allyl- H_2 coupling constants and multiplicity for compounds **2a-f** was
204 completed by comparative simulation considering a first order spectra ($\Delta\nu/J > 8$) of an
205 AA'MXX' spin system (25); spin simulation was carried out using Mestrenova v7.1.1 (test
206 version) (26). Determined experimental values of chemical shifts and coupling constants
207 range for the allyl system were used to simulate the H_2 signal until an identical
208 multiplet was obtained by direct comparison, as shown in **Figure 1**.
209

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210

211 **Figure 1.** Obtained (black) and simulated (red) spectra for allyl- H_2 proton considering an
 212 AA'MXX' spin system.

213

214 H_2 proton chemical shifts and coupling constants for **2a-f** allyl system are shown in **Table**
 215 **3.** Compound **2f** showed a single GC signal but a double group of signals in 1H and ^{13}C -
 216 NMR; characteristic coupling constants for the allyl H_2 proton were observed due to the
 217 existence of two different conformational isomers (α/β).

218

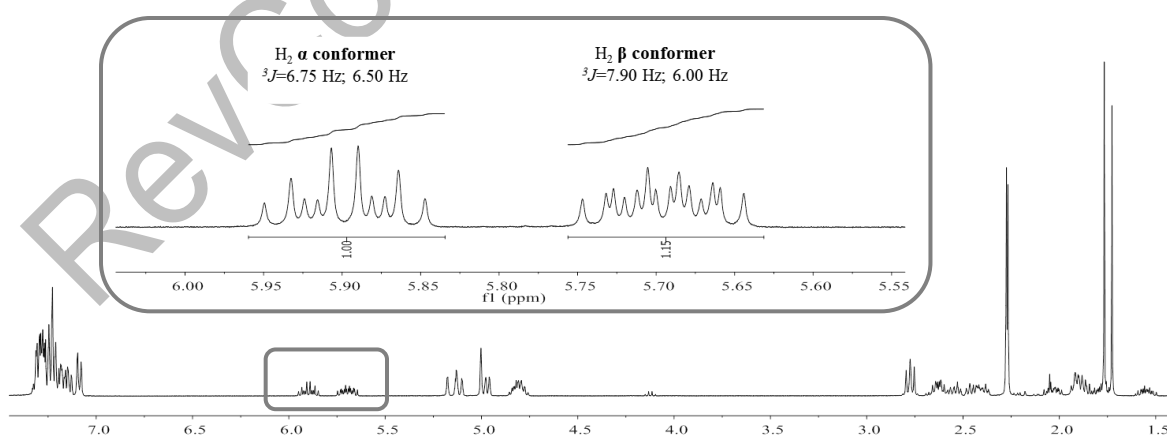
219 **Table 3.** $^1\text{H-NMR}$ chemical shifts [ppm] and coupling constants 3J [Hz] for the allyl
 220 H_2 nuclei of **2a-f**.

Compound	δ [ppm]	3J (H_{trans})	3J (H_{cis})	$^3J(\text{H}_{3a})$	$^3J(\text{H}_{3b})$
2a	5.765	17.90	9.50	6.80	6.40
2b	5.834	17.50	9.30	6.90	6.50
2c	5.834	17.60	9.50	6.80	6.50
2d	5.811	17.25	10.10	6.60	6.35
2e	5.907	17.00	10.61	6.60	6.35
2fα	5.898	17.30	9.95	6.75	6.50
2fβ	5.696	16.60	10.85	7.90	6.00

221

222 $^1\text{H-NMR}$ integral relation of conformers **2f α** and **2f β** was 1:1.15 (**Figure 2**). $^1\text{H-NMR}$
 223 experiments showed that α and β conformational isomers ratio does not change in time at
 224 room temperature. Conformers **2f α** and **2f β** also showed diastereotopic protons H_{3a} and H_{3b} as
 225 separated multiplets.

226



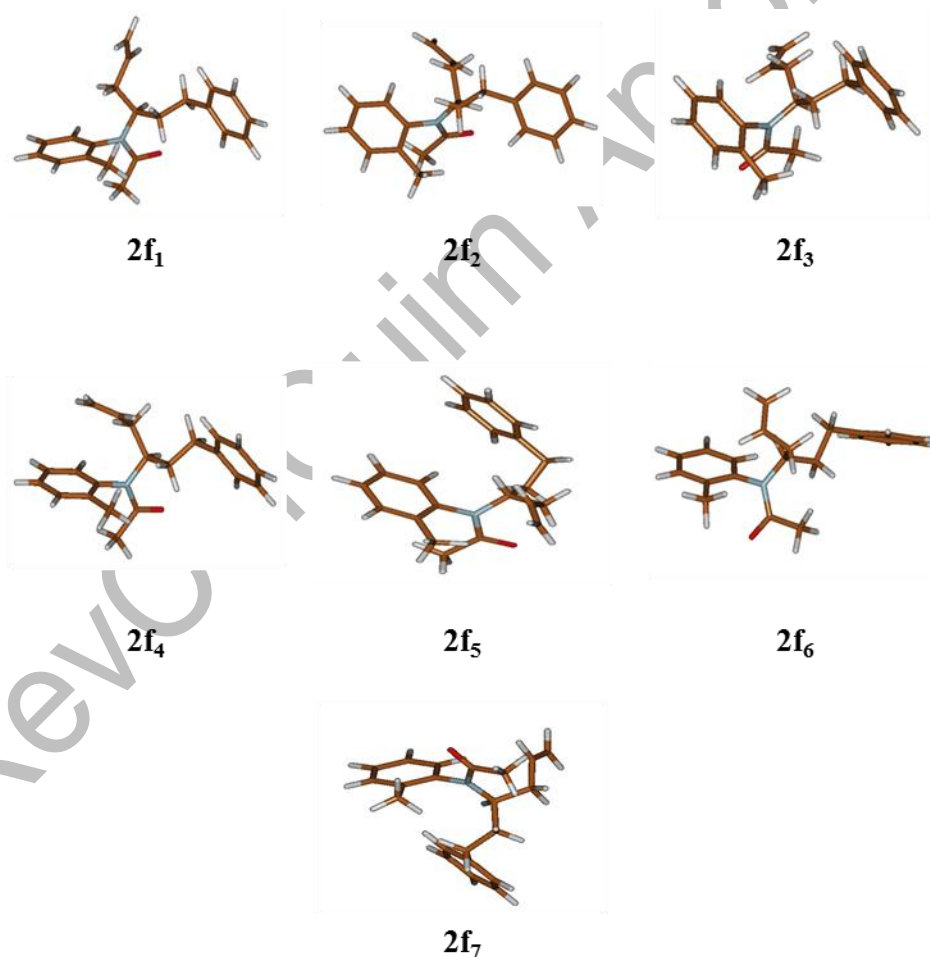
227

228 **Figure 2.** $^1\text{H-NMR}$ (CDCl_3 , 400MHz) spectra of **2f**; the two different allyl- H_2 signals (in
 229 expansion) determine the α/β conformers.

230

231 After conformational analysis of **2f**, despite of including both, the electronic energy and the
232 VZPE differences, the optimized structures **2f₁-2f₇** showed similar ΔE values, avoiding a
233 direct assignment to explain the duplicity of the signals in ¹H-NMR. Molden was used for
234 displaying the molecular structures (27). These structures are qualitatively similar, differing
235 mainly on the allyl group dihedral angles and slightly on rotation of other sigma bonds
236 (**Figure 3**).

237



238

239 **Figure 3.** Calculated structures for conformers **2f₁₋₇**

240

241 **Table 4** shows the energies and the allyl group dihedral angles for each **2f** conformer after
242 optimization, taking **2f₂** [lowest electronic energy at the B3LYP(6-31+G(2d,p)) level of
243 theory] as reference for energetic differences.

244

245 **Table 4.** Calculated energy values and allyl group dihedral angles (degrees) for seven
246 different conformers of **2f**.

	ΔE [Kcal/mol] (electronic energy)	Total ΔE [Kcal/mol] (including VZPE)	Dihedral angles [degrees °]	
			Φ_1	Φ_2
2f₁	0.47	0.46	63	179
2f₂	0	0	170	286
2f₃	1.79	2.02	64	179
2f₄	2.05	2.16	66	181
2f₅	1.08	1.11	173	295
2f₆	5.08	5.25	66	182
2f₇	4.85	5.46	62	178

247

248 To correlate the H_2 allyl 1H -NMR signals to a particular conformer, the proposed equation
249 by Garbisch for allyl compounds [**Eq. 1**] was used (28). Garbisch equation is a modified
250 Karplus equation involving also σ and π bonds contributions. Some authors suggest that
251 some modifications can be done to make this equation more precise (29-31). With the
252 Garbisch equation a relation between observed coupling constants (J) and the dihedral angle
253 (Φ) for the allylic proton H_2 and H_3 for each conformer was determined, as shown in **Tables**
254 **5** and **6**.

255

256 **Table 5.** Coupling constants 3J [Hz] and their corresponding dihedral angles (Φ_1 and Φ_2)
 257 from Garbisch equation for allyl protons H_2 and H_{3a}/H_{3b} in compounds **2a-e**.

Compound	${}^3J_{H_{3a}}$	${}^3J_{H_{3b}}$	$\Phi_1 [\pm 15^\circ]$	$\Phi_2 [\pm 15^\circ]$
2a	6.4	6.8	12.9	133
2b	6.5	6.9	9.1	134
2c	6.5	6.8	9.1	133
2d	6.3	6.6	0.4	130
2e	6.3	6.6	0.4	130

258

259 Dihedral angles for conformers **2f₁₋₇** found in our calculations and conformers **2f _{α}** and **2f _{β}**
 260 obtained from Garbisch equation are not equivalent, but approximated (**Table 6**)
 261 considering $\pm 15^\circ$ of uncertainty (27).

262

263 **Table 6.** Dihedral angles (Φ_1 y Φ_2) for allyl protons H_2 and H_{3a}/H_{3b} in seven different
 264 conformers for **2f**; calculated using the B3LYP(6-31+G(2d,p)) level of theory (left) and
 265 from the Garbisch equation (right).

$${}^3J \cong \begin{cases} 6.6 \cos^2 \Phi + 2.6 \sin^2 \Phi (0^\circ \leq \Phi \leq 90^\circ) \\ 11.6 \cos^2 \Phi + 2.6 \sin^2 \Phi (180^\circ \geq \Phi \geq 90^\circ) \end{cases} \text{ [Eq. 1]}$$

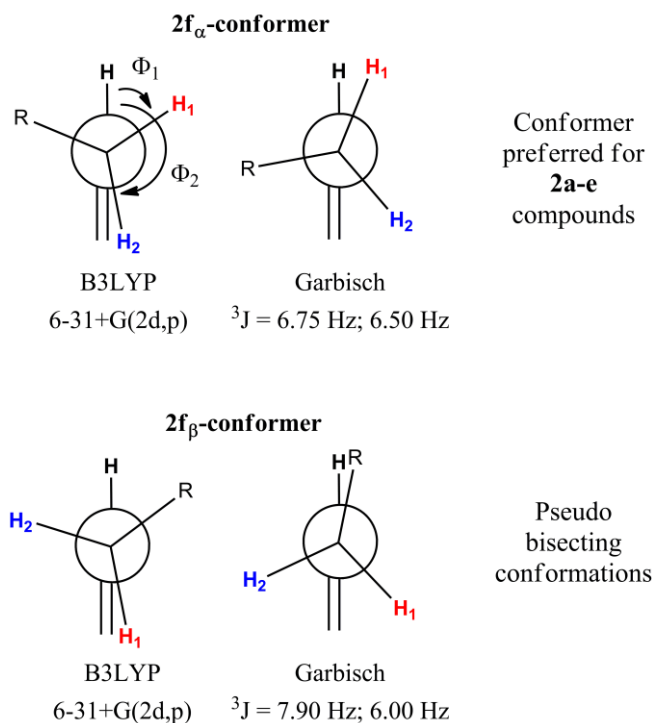
	Calculated results					Garbisch's results		α	β
	2f₁	2f₃	2f₄	2f₆	2f₇	2f₂	2f₅		
Φ_1	63	64	66	66	62	170	173	9.1	140
Φ_2	179	179	181	182	178	286	295	133	232

266

267 A comparison for both models (**Figure 4**) shows high similarity when a Newman
 268 representation of the allyl H_2 and vicinal methylenic protons H_{3a} and H_{3b} are depicted.

269 Considering this fact, it was possible to associate these two different positions for H_{3a} and

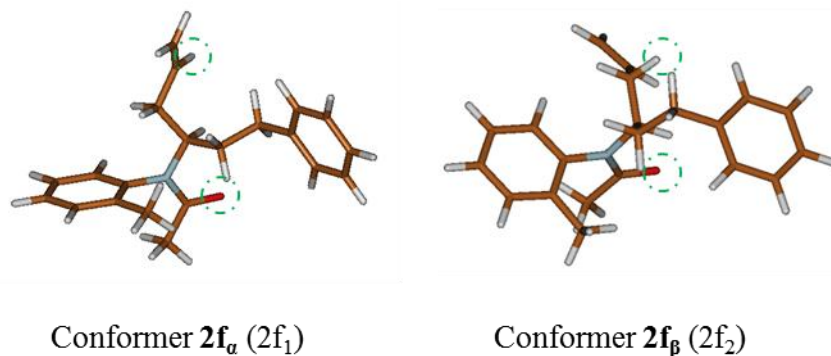
270 H_{3b} protons to conformers α and β according to the observed ^1H and ^{13}C -NMR spectra,
 271 concluding that bulky groups (R and C=C) are located in a pseudo bisecting conformation
 272 in Newman's projections (**Figure 4**).



273
 274 **Figure 4.** Newman projections for allyl group conformers $2f_\alpha$ and $2f_\beta$: left, calculated
 275 structures (B3LYP/6-31+G(2d,p)); right, according to [Eq. 1].

276
 277 Thus, possible structures of α/β conformers were established according to their calculated
 278 energy, in each case. Allyl group disposition according to Garbisch equation was within the
 279 approximation limits with calculated conformers $2f_1$ and $2f_2$ (**Figure 5**).

280

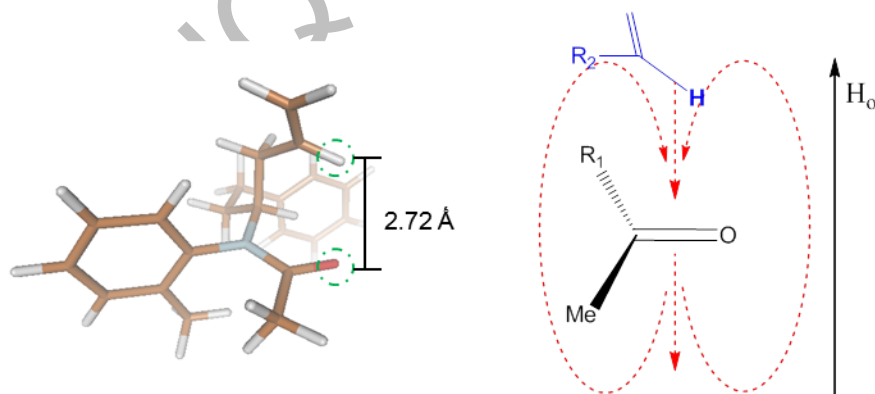


281

282 **Figure 5.** Calculated structures (B3LYP/6-31+G(2d,p) for conformers α and β of **2f**.

283

284 Analogous calculated structures for **2f** differ basically on the spatial distribution of the allyl
 285 group and its dihedral angle; a 2.72 Å dipolar interaction between the allyl proton and the
 286 acetyl oxygen atom was observed in conformer **2f β** (**Figure 6**), explaining the chemical
 287 shifting to high fields in the $^1\text{H-NMR}$ spectrum due to C=O anisotropic protection. Each
 288 conformer of **2f** (α and β) was totally elucidated using 2D-NMR experiments (COSY,
 289 HSQC and HMBC).



290

291 **Figure 6.** Left: Allyl- H_2 proton interaction at 2.72 Å distance with the carbonyl oxygen in
 292 conformer **2f β** . Right: the diamagnetic protection zone of double bond C=O.

293

294

295

Conclusions

296

297 An easy methodology to access new *N*-phenyl-*N*-(1-phenylhex-5-en-1-yl) acetamides **2a-f**
298 using acetic anhydride as reagent and solvent, including green chemistry principles was
299 used. Using quantum chemical calculations and Garbisch's approximation it was possible
300 to determine that compounds **2a-e** prefer a single conformation similar to conformer **2f_α**.
301 Existence of conformers **2f_α** and **2f_β** explain the double signals observed in both ¹H and ¹³C-
302 NMR spectra for compound **2f**. Coupling constants and chemical shifts values for *H*₂ allyl
303 proton signals of compounds **2a-f** were described for each conformer and can be used as a
304 comparative base in ¹H-NMR allyl-*H*₂ signal coupling constants assignment.

305

306

307

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308

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