- 1 Title: The broad-scale analysis of metals, trace elements, organochlorine pesticides and
- 2 polycyclic aromatic hydrocarbons) in wetlands along an urban gradient, and the use of a high
- 3 trophic snake as a bioindicator.

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# 21 Abstract:

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Wetlands and their biodiversity are constantly threatened by contaminant pollution from urbanisation. Despite evidence suggesting that snakes are good bioindicators of environmental health, the bioaccumulation of contaminants in reptiles is poorly researched in Australia. We conducted the first broad-scale analysis of 17 metals and trace elements, 21 organochlorine pesticides and 14 polycyclic aromatic hydrocarbons in the sediments (four samples per site, December 2018) from four wetlands along an urban gradient in Perth, Western Australia and from the livers (five livers per site, February - April 2019) of Western tiger snakes Notechis scutatus occidentalis at those sites. All 17 metals and trace elements were detected in the sediments of every wetland, and 15 in the livers of tiger snakes. Arsenic, Cu, Hg, Pb, Se, Zn were at concentrations exceeding government trigger values in at least one sediment sample. Two organochlorine pesticides and six of seven polycyclic aromatic hydrocarbons were detected in the sediments of a single wetland, all exceeding government trigger values, but not in tiger snakes. Metals and trace elements were generally in higher concentration in sediments and snake livers from more heavily urbanised wetlands. The least urbanised site had some higher concentrations of metals and trace elements, probably due to agriculture contaminated groundwater. Concentrations of nine metals and trace elements in snake livers were statistically different between sites. Arsenic, Cd, Co, Hg, Mo, Sb and Se near paralleled the pattern of contamination measured in the wetland sediments; this supports the use of high trophic wetland snakes, such as tiger snakes, as bioindicators of wetland contamination. Contamination sources and impacts on these wetland ecosystems and tiger snakes are discussed herein.

## 1. Introduction

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Wetlands are biodiversity hotspots threatened by urbanisation, worldwide. Wetlands are often the only remaining fragments of habitat within an urban landscape (Ehrenfeld 2004; Garden et al. 2006), and provide islands of water storage, ground water replenishment, supply and transformation of nutrients, high biodiversity, and recreational enjoyment for humans (Lee et al. 2006; Novitski et al. 1996; Zedler and Kercher 2005). As land development increases wetland ecosystems degrade through changes in hydrology (Chadwick et al. 2006), structure (Faulkner 2004; Lee et al. 2006), floral and faunal biodiversity (Gibbs 2000; McKinney 2008), and water and sediment chemistry (Brown et al. 2010; Fitzpatrick et al. 2007; Panno et al. 1999). Environmental contamination, as a consequence of rapid urbanisation, industrialisation and poor waste management practices (Nriagu 1990; Rodriguez Martin et al. 2015), is particularly severe for urban wetlands and threatens the health of biological communities (Spurgeon and Hopkin 1999; Zhang et al. 2017). Due to their topography, urban wetlands are susceptible to contamination from several primary source points: urban runoff (Zhang et al. 2012), stormwater drains feeding into wetlands (Clarke et al. 1990), groundwater (Roy and Bickerton 2011), and pest and weed treatment (Gentilli and Bekle 1993). Perth is the largest city of the west coast of Australia, and is built almost entirely on the Swan Coastal Plain bioregion (Davis and Froend 1999). The Swan Coastal Plain is characterised by sandy soil dunes systems interlaced with a chain of inter-connected ephemeral wetlands and lakes (Simpson and Newsome 2017). For the past near 200 years urbanisation and agriculture has drained or filled in an estimated 70% of the original wetland area of the Swan Coastal Plain, and the remaining wetlands have been subject to structural and hydrological modifications, isolation, biodiversity loss and pollution (Davis and Froend 1999; Gentilli and Bekle 1993). The remnant wetlands of urban Perth have a history of contamination from primary sources such as inflowing stormwater drains, as well as historical industrial dumping and pest management through pesticides (Clarke et al. 1990; Department of Water 2009; ESRI 1983; Gentilli and Bekle 1993). Some of the larger wetlands in Perth still support populations of Western tiger snakes (Notechis scutatus occidentalis), a top predator of these ecosystems. Snakes are an important organism in the ecosystem, providing predator and prev

functions that shift as they climb the trophic tiers as they age. Wetland snakes in particular represent an interface between aquatic and terrestrial habitats; thus they are susceptible to the bioaccumulation of contaminants and can be used as bioindicators for environmental pollution (Campbell and Campbell 2001; Drewett et al. 2013; Lemaire et al. 2018).

Despite the historical contamination and close proximity of infrastructure to urban wetlands in Perth, there are little monitoring data available on the contamination of water, sediment or fauna (see table 1 for summary). Wetlands can be polluted with a plethora of contaminants, the more frequently assessed contaminants are metals and trace elements, organochloride pesticides (OCPs) and

polycyclic aromatic hydrocarbons (PAHs) (Cooper 1993; Gambrell 1994; Haarstad et al. 2012). As wetland structure plays an important role in contaminant retention, permanently enclosed wetlands such as lakes are more susceptible to the accumulation of contaminants compared to flowing water

systems (Burger et al. 2007; Schulz and Peall 2001). As a result, fauna communities restricted to

enclosed and isolated urban wetlands might be vulnerable to bioaccumulation from continuous

exposure to contaminants. Three permanent and enclosed wetlands that differ in degree of

urbanisation persist in Perth and contain abundant populations of tiger snakes. By comparing these

wetlands and tiger snake populations with a similar wetland within a national park we are presented

with a rare and ideal system to study the contamination of wetlands and a top predator snake along an

urban gradient.

This study presents and examines a snapshot of concentrations of 52 contaminants in both sediment and tiger snake livers from four wetlands around Perth, Western Australia. The objective of this study was threefold: (1) quantify contaminants present in the wetlands, (2) measure contaminants that are bioaccumulating in the top predator: tiger snakes, and (3) investigate if contaminant concentrations parallel the degree of urbanisation of wetlands? Research on contaminants in snakes is an emerging field (Burger et al. 2017; Drewett et al. 2013; Gavric et al. 2015; Quintela et al. 2019; Schwabenlander et al. 2019) yet as far as we are aware this study analyses the largest range of contaminants in any species of snake, and is only the third study to present data on the contaminants of both the snake tissue and ecosystem they were collected from (Ford and Hill 1991; Soliman et al.

2019). We also present the first published contaminants in Australian snakes in over 40 years (Beck

96 1956; Best 1973).

## 2. Material and methods:

2.1 Study sites

We examined a suite of contaminants in four wetlands in the Swan Coastal Plain of Western Australia (Fig. 1.): Herdsman Lake, Bibra Lake, Lake Joondalup and Loch McNess. Loch McNess is located in Yanchep National Park. We selected these wetlands as study sites based upon presence and abundance of tiger snakes. Historically, these wetland lakes were partially linked and ephemeral (Gentilli and Bekle 1993) yet the development of Perth city lead to the draining of some wetlands while others were dredged to become permanent (Halse 1989). These wetlands share similar climatic factors and modification from naturally ephemeral to permanently-filled, yet differ in degrees of urbanisation (Davis and Froend 1999). Table 1 describes these wetlands and summarises the few studies on contaminants reported at these wetlands. As there is no recent and detailed research available on the contaminants of these wetlands, we initially tested the sediments for a suite of contaminants listed in Table A. 1.

# 2.2 Study species

The tiger snake (*Notechis scutatus*) is a polymorphic Australian elapid occurring in disjunct populations that vary in diet, habitat and ecology (Aubret et al. 2004; Aubret et al. 2006). However, across most of its range, including the mainland Western Australian subspecies *N. scutatus occidentalis*, tiger snakes are an abundant higher trophic reptile predator in wetlands that have a preference for frogs (Aubret et al. 2006; Shine 1987). There is no long-term monitoring data available for tiger snakes to determine life expectancy, yet the oldest captive record was up to 24 years (Fearn and Norton 2011). Tiger snakes and dugites (*Pseudonaja affinis*) are the only two large snake species to persist within urban Perth and dugites prefer woodland and heath over wetlands, thus we selected tiger snakes as our bioindicator model species based on their abundance, habitat preference, high trophic position and prey preference. Frogs are known to bioaccumulate contaminants indirectly

through their food or directly through their water permeable skin when in contact with contaminated waters or sediments (Bruhl et al. 2011; Hopkins 2007; Ohlendorf et al. 1988), and therefore provide a crucial link for contaminant transfer throughout the food web.

## 2.3 Sediment collection and analysis

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Sediment samples were collected 18 December 2018 during the dry season when Perth receives little rainfall. Each wetland was sampled in two locations and sediments were collected in duplicates, each duplicate was collected three metres away from each other. Sampling locations are shown in Fig. 1 and corresponded to areas of the wetland with the highest capture rate of tiger snakes. The first 10 cm of sediment was collected using a metal scoop and glass jar for organic contaminant samples and a plastic scoop and jar for metal and trace element samples. Sediment samples were kept cool until submitted for chemical analysis at the end of the day of collection. All samples were analysed for 17 metals and trace elements (from hereon in collectively referred to as metals), 21 organochlorine pesticides and 14 PAHs by ChemCentre (Perth, Western Australia). These contaminants were chosen based on the limited historical data and regular screening of this suite by ChemCentre for environmental monitoring (Leif Cooper, pers. comm.). The specific contaminants, method of analysis, detection limits and quality assurance are reported in Appendix 1. Metals were determined using methods iMET2SAMS and iMET2SAMS based on US EPA method 3051A (US EPA 2007). Sediment samples were extracted with concentrated nitric and hydrochloric acid, then analysed for metals using a combination of inductively coupled plasma-atomic emission spectroscopy (ICP-AES) and inductively coupled plasma-mass spectrometry (ICP-MS). Organochlorine pesticides were determined using method ORG141S and PAHs were determined using method ORG100S, both based on US EPA method 8270D (US EPA 1998). Contaminant concentrations are reported as mg/kg dry weight, and are compared to the Australian and New Zealand guidelines (ANZECC & ARMCANZ 2000) and the revised ANZECC/ARMCANZ Sediment Quality Guidelines (Simpson et al. 2013). The lower trigger value (sediment quality guideline value (SQGV)) represents the threshold for biological effects, and the higher sediment quality guideline trigger value (SQGV-High) represents the high probability of biological effects (Simpson et al. 2013). If there was no guidelines for a particular

contaminant they are compared to alternative guidelines (Lemly 1996; Ontario Ministry of Environment and Energy 1993).

2.4 Snake liver collection and analysis

Five tiger snakes were collected from each site by hand between February and April 2019, and euthanised humanely by blunt force trauma to the head. Sex, snout-vent length (SVL), total mass and liver mass were recorded for all snakes (Table 2). We selected livers for analysis due to their potential to retain contaminants taken up by feeding. Whole livers were extracted using ceramic blades and frozen, and were submitted to ChemCentre for chemical analysis. All liver samples were analysed for the same contaminants as were the sediments; the specific contaminants, method of analysis, detection limits and quality assurance are listed in Appendix 2. Whole livers were homogenised and extracted with concentrated nitric and hydrochloric acid, then analysed for metals were determined using methods iMETBTMS and iMETBTICP based on APHA methods 3120 and 3125 (APHA 1998) using a combination of inductively coupled plasma-atomic emission spectroscopy (ICP-AES) and inductively coupled plasma-mass spectrometry (ICP-MS). Organochlorine pesticides and PAHs were determined using method ORG100B based on US EPA methods 8270D (US EPA 1998). Contaminant concentrations are reported as mg/kg wet weight.

# 164 2.5 Data analysis

Concentration means and standard deviation were calculated for each contaminant for each site. For statistical analysis samples that were recorded below detectable limits (BDL) were entered as half the detection limit (ANZECC & ARMCANZ 2000; Zeghnoun et al. 2007). Due to the limited number of animals allowed to be sacrificed at each site, no statistical differences could be established between male and female contaminant burdens. Consequently contaminant data were pooled for male and female snakes. The Shapiro-Wilk test was used to determine all data (sediment and livers) had a non-parametric distribution. Hence a Kruskal-Wallis test was used for both sediments and tiger snake livers to determine if there were significant differences (p <0.05) in contaminant concentrations between sites. Following, a Dunn post-hoc test was performed to identify the pairs of sites that

showed significant differences. P-values were adjusted using the Benjamini-Hochberg method. We used Spearman rank correlations to explore the relationship between SVL (age) of snakes and each metal and trace element within each site. We present and consider rho values over 0.5 as moderate positive correlations and allow the reader to assess the significance themselves. All statistical analysis were conducted in R Studio (R Core Team 2018).

## 3. Results and Discussion

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#### 3.1 Wetland sediment contaminants

All 17 metals tested for were detected in Herdsman Lake, 16 were detected in Bibra Lake and Loch McNess, and 15 were detected in Lake Joondalup (discussed data presented in Table 3). Four metals (As, Cu, Pb and Zn) were detected exceeding trigger values in at least one sample, and Zn was detected exceeding the high trigger value in one sample from Herdsman Lake. Selenium was detected exceeding the trigger value in one sample at Bibra Lake and two samples at Loch McNess. Mercury was detected exceeding the trigger value in two samples in Lake Joondalup and one sample in Loch McNess. Generally, mean concentrations of contaminants decreased with less urbanised sites except for nine metals that were high at Loch McNess. Herdsman Lake generally had the highest mean concentration of metals including significantly higher concentrations of Pb, As, Co, Cu, Mo, Ni, Sn and Zn compared to at least one other site, as well as the only detection of Ag. Herdsman Lake is particularly susceptible to contamination from several point sources. Since urbanisation began in Perth the wetland has suffered from considerable changes in land use including: stock grazing (1850s), market gardening (1910s), drainage for irrigation and land reclamation (1920s), sanitary landfill (1930s), compensation basin for urban drainage (1930s+), intense pesticide treatment (1950 – 1980s), reserved for public recreational space (1970s+), dredging (1980s) and periodic illegal rubbish disposal (Clarke et al. 1990; Davis and Garland 1986; Department of Water 2009; ESRI 1983; Gentilli and Bekle 1993; Kobryn 2001). Currently, the wetland is divided into three main interconnected water bodies which receive drainage from five major and an unknown number of minor drainage systems, including the bordering industrial area. Urban and industrial

development began to encroach the lake in the 1950s and currently virtually no original fringing vegetation remains. The high concentrations of As, Cu, Pb and Zn are likely to originate from industrial stormwater, urban runoff, and leaching from historical dumping. Although our sampling did not detect any pesticides they have been detected in more comprehensive studies in the past (Clarke et al. 1990; Davis and Garland 1986).

Bibra Lake generally had the second highest mean concentrations of metals, and the only detections of OCPs and PAHs (discussed later). There is limited information available on the history of Bibra Lake but it has not suffered from the same degree of urbanisation as Herdsman Lake. Although it has a much larger buffer of remnant vegetation than Herdsman Lake (Fig. 1) it is still in close proximity

of Water 2009). The northern and southern edge of the lake are bordered closely to main roads, and

the southern edge of the lake has buried sanitary landfill (Burkett 2005). Bibra Lake receives water

to an industrial area which began development in conjunction with suburbia in the 1970s (Department

via direct rainfall and surface runoff from the urban catchments (Sinang et al. 2015), which is might

be the source of its moderate concentration of metals.

Joondalup Lake had the lowest concentration of most of the metals detected, including significantly lower concentrations of Pb, As, Co, Cu, Mo, Ni and Zn than Herdsman Lake, the latter being the most contaminated and urbanised site. Urbanisation of Joondalup only began to rapidly increase around the lake in the 1980s, which is later than the other study sites (Kinnear et al. 1997); and the wetland has a relatively large buffer of remnant vegetation surrounding the lake compared to the other urban sites, protecting the lake from urban runoff as most of its water is recharged from rainfall (Newport and Lund 2014). Interestingly, Joondalup had the highest mean and maximum concentrations of total Hg, both exceeding the guideline's low trigger value. Monitoring of Joondalup's surface water in recent years has detected Hg exceeding guideline concentrations in winter months; while the point source is still unknown the annual spike suggests that the source is runoff from winter rains (Gonzalez-Pinto et al. 2017; Newport and Lund 2014).

Despite being outside the urban matrix, Yanchep National Park's wetland Loch McNess had elevated concentrations of As, Ba, Cd, Cr, Co, Mo and Zn. In addition, Hg and Se were detected at trigger

value concentrations higher (Table 3). Loch McNess is defined as a 'flow-through lake', receiving water from the groundwater system to which it was connected, potentially receiving contaminants from bordering agricultural land. Yet over the last two decades it has been suffering from a severe surface water and groundwater decline resulting in it's almost permeant disconnection from the groundwater (Department of Water 2011). As a result, large areas of lakebed sediment are now exposed and suffer from drying and erosion, then re-flooding with rain events. This process can release the sediments accumulated contaminants back into the wetland ecosystem (Al-Maarofi et al. 2013). Arsenic compounds are common and naturally occurring in many soils and wetlands of the Swan Coastal Plain (Appleyard et al. 2006), yet elevated concentrations in the sediment of Loch McNess could potentially be enhanced by groundwater contaminated from the historic use of pesticides on sheep (Arnold and Oldham 1997; Davis et al. 1993). Although Se is a necessary element for the normal development of organisms (Kapustka et al. 2004) it was detected at levels of concern in the sediments of Loch McNess (mean 4.04, max 7.9 mg/kg dry weight), and significantly higher concentrations than Bibra and Joondalup Lakes (Table 3). A point source cannot be determined but contamination might be from groundwater passing beneath agricultural irrigation and fertiliser use (Gardiner and Gorman 1963). Only one sediment sample at one site (Bibra Lake) contained OCPs or PAHs, and the two detected OCPs and six out of seven PAHs exceeded sediment guidelines. The OCPs aldrin and its metabolite dieldrin have been banned from manufacture, importation, and use in Australia since the internationally legally binding agreement for OCPs (The Stockholm Convention on Persistent Organic Pollutants) was enacted in 2004 (DEH 2004; UNEP 2011). However, OCPs and their metabolites are highly persistent compounds that can be still present in the environment (Bai et al. 2015; Wu et al. 1999), and have been historically detected in the sediment of Perth's estuaries (Nice 2009) and wetlands (Davis et al. 1993). The presence of dieldrin and other OCPs detected in Perth's urban wetlands can be attributed to the state government program to control Argentine ants (from 1950s to 1980s) and periodic termite control (Davis and Froend 1999; Davis and Garland 1986). Bibra Lake and other urban wetlands of Perth have been subject to heavy pesticide treatment in the past in an

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attempt to control non-biting midge and mosquito levels (City of Cockburn 2015; Davis and Froend 1999). PAHs have historically been found in the sediments of Bibra Lake (Department of Water 2009); and six out of seven detected PAHs exceeded the SQG trigger values. The hypothesized source is used vehicle oil and machinery fluids. The sampling site of our sediment containing both OCPs and PAHs is within 50 meters down slope of a main road and may be frequently exposed to road pollution during rain events. Although this research only detected OCPs and PAHs in one sediment sample in one lake, the small sample sizes and large intra-site variation of contaminant concentrations between sediment samples should not rule out the possibility of OCPs and PAHs persisting in sediments at concentrations of concern in our other study sites or other wetlands of Perth.

Despite a high degree of intra-site variation in contaminant concentrations, generating knowledge on the sediment contaminant burden is important for highlighting the extent and history of wetland pollution (Förstner 2004). Generally, contaminants were in higher concentrations in the sediments of heavily urbanised compared to less-urbanised wetlands except for some metals in Loch McNess. The concentrations of Loch McNess sediments suggest that despite being outside of the urban matrix and surrounded by a protected National Park, contamination is still possible. High concentrations may originate from agriculture-contaminated groundwater that potentially contributes to wetland sediment contamination at levels comparable to highly urbanised wetlands. We recognise our number of sediment samples is both small and limited to the areas where snakes were captured, thus might not be an accurate representation of contamination of the entire wetlands. Nevertheless the samples do present a snapshot of contamination present in areas abundant with tiger snakes.

# 3.2 Occurrence of contaminants in snakes

Sixteen metals were detected in the livers of tiger snakes collected across Perth's wetlands in 2019 (discussed data and significant differences presented in Table 4). Beryllium wasn't analysed above detection limits and Sn was only above detection limits in a single liver from a snake captured at Lake Joondalup. Antimony, As, Ba, Cd, Co, Hg, Mo and Se concentrations were significantly different between sites, mostly higher at Herdsman Lake compared to Joondalup Lake. Lead was detected in only four snakes, three from Loch McNess and one from Herdsman Lake, and Sn was detected in only

one snake from Lake Joondalup. Molybdenum concentrations in snakes from Herdsman Lake were significantly higher than in snakes at all other sites, and are as far as we can tell, the highest reported liver concentration in a reptile. These contaminants have all been documented in snakes before; however, the published literature has only one other report of a comprehensive suite of metals analysed in snakes (Wylie et al. 2009). Our study is the most comprehensive study to date on bioaccumulation in any terrestrial reptile in Australia. There were no significant differences between Cr, Mn, Ni, Ag and Zn liver concentrations between sites. No OCPs or PAHs were quantified above detection limits; however, both were detected from only one sediment sample at a single wetland, and PAHs are known to be metabolised quickly in vertebrates (Hylland 2006). In addition, due to small liver tissue mass (<10g) and broad-scale analysis detection limits had to be up to 2 mg/kg, thus we cannot conclude that these were not present under those concentrations. There is no quantitative toxicity thresholds available for OCPs and PAHs in reptiles; however, relative to other vertebrates toxicity results, <2 mg/kg is unlikely to induce adverse biological impact on snakes (Ball and Truskewycz 2013; Weir et al. 2013). Currently, the information available on the bioaccumulation of contaminants and their effects on reptiles is a limited but is a growing research field. Although there are many studies reporting various contaminants in snakes (Albrecht et al. 2007; Burger et al. 2007; Burger et al. 2017; Campbell et al. 2005; Drewett et al. 2013; Heydari Sereshk and Riyahi Bakhtiari 2015; Quintela et al. 2019), we could only directly compare our data to seven metals (As, Cd, Cr, Hg, Mn, Pb, Se) commonly analysed as wet weight in the livers of three other wetland snakes Nerodia fasciata, Thamnophis gigas, Agkistrodon piscivorus conanti (see summary table in Wylie et. al. 2009 and (Hopkins et al. 1999; Rainwater et al. 2005; Wixon 2013)). Mean and maximum concentrations of these metals in Perth's tiger snakes were similar to concentrations reported in other wetland snakes, besides As and Se that were much lower in Perth's tiger snakes compared to other snakes from contaminated sites, and Perth's tiger snakes had less than half the concentration of Mn and Hg reported in other wetland snakes regardless of site contamination. For the less frequently analysed metals we could only compare the liver concentrations of Perth's tiger snakes to livers from A. piscivorus and T. gigas

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(Wixon 2013; Wylie et al. 2009). Mean liver concentrations of Sb, Co, Ag and Sn were similar in tiger snakes compared to those reported in *A. piscivorus* and *T. gigas*, while Ba, Cu, Ni and Zn concentrations varied between the three species. Molybdenum was similar for all wetland snakes except for tiger snakes from Herdsman Lake, where it were much higher (mean 8.32, max 13.0 mg/kg wet weight).

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Very few metals were positively correlated with body size except for Sb (rho 0.6, p 0.35), Hg (rho 0.87, p 0.05) and Ni (rho 0.8, p 0.13) in Herdsman Lake snakes; Sb (rho 0.6, p 0.35), Cr (rho 0.7, p 0.23) and Ag (rho 1, p 0.02) in Bibra Lake snakes; As (rho 0.8, p 0.13) in Lake Joondalup snakes; and Sb (rho 0.82, p 0.09) in Loch McNess snakes. We consider these results to be exploratory at best, and low significances is reflective of small sample sizes due to financial and ethical limitations. These results, however, suggest the uptake of most metals does not appear to be related to body size, a common observation in bioaccumulation research (Albrecht et al. 2007; Fontenot et al. 2000; Quintela et al. 2019). Although our study didn't detect any metals at alarmingly high concentrations, we identified nine metals of interest based on their significant difference between sites in the sediment or liver samples, and we consider these to be the contaminants of most concern. We use Fig. 2 to compare the inter-site differences in metal levels between sediment and liver concentrations. Antimony, Cd, Co, Mo and Se had almost identical inter-site patterns between the mean contaminant concentrations of the sediment and livers. Copper, As and Ba had mostly similar inter-site differences although there was high variation in the snake's livers. Mercury concentrations in snakes from Loch McNess reflected the concentrations in the sediment, although both had high inter-sample variation; concentrations were lower in snakes from Joondalup than the sediment and much higher in Herdsman Lake snakes than the sediment. Lead and Sn showed generally inverse inter-site relationships between sediment and snake liver concentrations. Despite Zn exceeding the SQG higher trigger value in sediment from Herdsman Lake, it was found at consistent concentrations in Perth's tiger snake livers as it is metabolically regulated in vertebrates (Sandstead 2014).

The intra-site variation in contaminant concentration was generally higher in sediment than it was in snake livers (Tables 3 & 4). Variation is naturally high in sediment due to different point sources of

contaminants, seasonal changes of water chemistry and soil grain-sizes. Biological tissues such a livers are more homogenous in composition than sediments and consequently show a lesser interindividual and inter-site variability in contaminant burdens than sediments do, thus the use of fauna as bioindicators of environmental health. The use of wetland snakes as bioindicators has been proposed for decades (Campbell and Campbell 2001; Haskins et al. 2019; Stafford et al. 1977) based on their trophic status and site fidelity (Campbell et al. 2005). Despite limited positive correlations between body size and metals Fig. 2 suggests there is a strong relationship between sediment and snake liver concentrations within sites. These results are consistent with the general pattern of fauna contaminated by metals increases if the population is exposed to a source of contamination (Nasri et al. 2017), in this case the sediment. Therefore, our results support the use of tiger snakes as a bioindicators of wetland ecosystem health based on (1) the evidence that they accumulate a range of metals, (2) their long life expectancy of up to 24 years (Fearn and Norton 2011), (3) their small home ranges (~0.05 km<sup>2</sup>) (Butler et al. 2005) especially in wetlands isolated by urbanisation, and (4) their high trophic position in the food chain as adults (thus being exposed to all of their prey's contaminants). Tiger snakes are directly exposed to the contaminants in their wetland through drinking the water, foraging in sediment (Orange 2007) when wetland waters annually recede, and through the consumption of their prey. Frogs, being tiger snakes' preferred prey (Aubret et al. 2006; Lettoof et al. 2020), are particularly sensitive to contaminant accumulation as they feed in sediment as tadpoles, have the ability to absorb chemicals through their skin (Bruhl et al. 2011; Smalling et al. 2015), and stay in close proximity to waterbodies (eg. ~20 m for *Litoria raniformis*), especially in urban isolated wetlands (Hamer and Organ 2008; Hitchings and Beebee 1997). Lemaire et al. (2018) compared the Hg concentrations in scales of various populations of viperine snakes (Natrix maura) that predate on frogs or on aquaculture-raised fish, and found lower mean contaminant concentrations and accumulation rates in frog-eating snake populations. This difference may be due to frogs generally being at a lower trophic tier than fish and potential Hg contamination from the aquaculture farms from which snakes were sampled (Lemaire et al. 2018). We believe frogs are an important vector for

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contaminants in the food web, and recommend testing them to better understand the food chain transfer of contaminants in Perth's urban wetlands, a research area lacking in Australia (Mann et al. 2009; Sievers et al. 2019). Due to financial limitations and the wetlands being inhabited by several species of frogs we chose to investigate bioaccumulation in tiger snakes as they should reflect the contamination of that particular food web.

In addition, tiger snakes may have a shorter lifespan in urban wetlands as a result of exposure to multiple stressors and therefore may not have the opportunity to accumulate large concentrations of contaminants. Such stressors include harassment from the public, pet dogs, vegetation management and parasitism, as tiger snakes often have large burdens of gastric nematodes (Lettoof et al. 2020). Although the high frequency of parasitism suggests tolerance, the influence of contaminants and parasites on tiger snake populations may be more complex. Chronic symptoms from synergistic contaminants may result in immunosuppression and increasing nematode parasitism (Rohr et al. 2008), or a reduction in feeding behaviour (Alonso et al. 2009; Forrow and Maltby 2000) which may imperil individual snakes who cannot consume enough food to feed both themselves and the nematodes. From another point of view, a large parasite burden may have beneficial impacts on tiger snakes, as parasites are able to uptake contaminants directly and reduce the concentrations in their host (Evans et al. 2001).

# 3.3 Potential impacts to snakes

As with other high trophic predators, the presence and persistence of higher trophic snakes represents a healthy ecosystem, and yet ecotoxicological research on snakes is still relatively rare compared to research on other predators (Campbell and Campbell 2001). Snakes can provide crucial insight into the distribution and movement of contaminants through wetland food webs in comparison to most other top predators (usually mammals and birds) based on several characteristics: entirely carnivorous, comparatively small home ranges, longevity and progression up trophic tiers throughout their life history. Six metals, two OCPs and seven PAHs of the 52 screened contaminants were detected at concentrations of concern in the sediment of wetlands across Perth; and of the 16 metals detected in tiger snakes from these wetlands, nine were of significantly different concentrations

between sites. Although we did not detect very high concentrations of metals that may cause acute toxicity in tiger snakes, the generally higher concentrations of most tested contaminants in snakes from the most urbanised wetland, Herdsman Lake, may have synergistic chronic effects on snake health. Only a handful of studies have been conducted on the acute or chronic impacts metal exposure and accumulation has on reptiles. For example, the presence of blood Hg concentrations have been found to negatively correlate with lymphocyte and B-cell proliferation in Loggerhead sea turtles (Caretta caretta), suggesting suppression of the immune system (Day et al. 2007). Lead and Cd, both detected in snake livers at higher concentrations in Loch McNess and Herdsman Lake than other sites, have reported lethal doses. The approximate concentration of Pb and Cd required to kill western fence lizards (Sceloporus occidentalis) is 2000 mg/kg and 1480 mg/kg of body weight respectively (Brasfield et al. 2004; Salice et al. 2009), which is considered mid-range sensitivity when compared to other taxa. The concentrations we detected in Perth's tiger snakes were much lower than these and are unlikely to have acute clinical impacts on the populations. Experimental exposure of several metals to reptiles has received more interest. Dietary exposure to Se, for example, can lower reproduction likelihood and potentially reduce egg and total mass of clutches in brown house snakes (Boaedon fuliginosus) (Hopkins et al. 2004); reduce average food intake and growth in body mass and length in leopard geckos (Eublepharus macularius) (Rich and Talent 2009). Banded water snakes (N. fasciata) from polluted wetlands with high liver concentrations of As and Se (mean As: 134 ppm; Se 140 ppm dry weight) exhibited mean standard metabolic rates 32% higher than snakes from unpolluted sites (Hopkins et al. 1999). Corn snakes (Elaphe guttata) feed methylmercury experienced a reduced growth rate (Bazar et al. 2002), while new-born northern water snakes (N. sipedon) contaminated with maternally-transferred Hg had less motivation to feed and had reduced strike efficiency (Chin et al. 2013). Developing embryos can also be exposed to metals through the egg shell (Marco et al. 2004). Consequently, absorption of Cd can cause malformation of eye development in the Italian wall lizard (Podarcis sicula) (Simoniello et al. 2014) and the running speed of hatchling Iberian rock lizard (*Iberolacerta monticola*) was negatively

correlated with embryonic As absorption (Marco et al. 2004). Metal body burdens or exposure levels

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that cause physiological or behavioural alterations in the aforementioned studies are well above what we observed in the livers of our adult tiger snakes from the Swan Coastal Plain; however, chronically contaminated reptiles are susceptible to a variety of pernicious developmental and behavioural disorders. Snakes from the urban sites Herdsman and Bibra Lake seem to be in poor health conditions (observation by the authors), which parallels sediment and liver contaminant levels. We have not as part of the present study measured behavioural parameters or biochemical markers of health however complimentary studies are initiated to investigate potential health impacts of contaminants on tiger snakes (Lettoof et al, in prep).

Interestingly, tiger snakes from Herdsman Lake had the highest mean liver concentration of Mo reported in a reptile  $(8.32 \pm 3.458 \text{ mg/kg})$ . Molybdenum is an essential element for vertebrate nutrition, however, excessive exposure to laboratory rats, guinea pigs and rabbits has been associated with anaemia, diarrhoea, deformities of joints and long bones, mandibular exostoses and morphological changes to the liver, kidneys and spleen (Tallkvist and Oskarsson 2015). Molybdenum exists naturally in various oxidation states but is mined as a principle ore for use as an alloy in steel and cast iron production, and also used in lubricants, chemical reagents and dyes. Stormwater drainage and urban runoff from the bordering industrial area are likely point-sources for Mo contamination in Herdsman Lake.

## 3.4 Future research and recommendations

Laboratory testing for ecotoxicology is expensive and usually charged per contaminant (for example, the cost of this study was over \$11 000 AUD). This warrants a trade-off between broad-scale screening with small sample sizes, or targeted screening focussing on contaminants of concern and much larger samples. Most other studies choosing the latter had contamination spill events or access to published recent monitoring data allowing a focus on contaminants of concern. Despite the study sites being recognised as critical wetlands for the environment, major tourist and local attractions, and situated throughout a developed country's major city, contaminant monitoring of these wetlands has largely been disregarded; and as a result sample sizes were sacrificed for broad-scale screening. Periodic monitoring of contaminants should be conducted in urban wetlands to prevent researchers

from investing limited resources on broad-scale screening and rather focus on contaminants of concern.

Ecotoxicological research on reptiles (especially snakes) is minimal at present, and fundamentally non-existent in Australia, despite consistent evidence of wetland snakes in the United States and other countries being good bioindicators of environmental contamination (Burger et al. 2007; Drewett et al. 2013; Lemaire et al. 2018; Quintela et al. 2019; Wixon 2013). Australia has several major cities with snakes persisting in urban wetlands or estuaries which should be susceptible to bioaccumulation of contaminants, and would be suitable systems to model the ecotoxicological impacts on snakes. Other model species include: the red-bellied blacksnake (*Pseudechis porphyriacus*), lowlands copperhead (*Austrelaps superbus*), common tree snakes (*Dendrelaphis punctulatus*), slaty-grey snake (*Stegonotus australis*), keelback (*Tropidonophis mairii*), Australian bockadam (*Cerberus australis*), and Richardson's mangrove snake (*Myron richardsonii*).

## 4. Conclusions

This study conducted a broad-scale analysis for 52 contaminants in sediment samples of four wetlands spanning across an urban gradient of Perth, Australia. We detected six metals, two OCPs and six PAHs exceeding SQG trigger values in sediments, with generally higher mean concentrations of contaminants found in more heavily urbanised wetlands. The natural site, Loch McNess, despite being the least modified and furthest from urbanisation was an exception, as the sediments contained second highest concentrations for 8 of the 16 of the metals analysed. We hypothesize the source of contamination in Loch McNess is from inflowing groundwater contaminated by radial agricultural land, specifically the historic use of pesticides and fertilisers.

Arsenic, Cd, Co, Hg, Mo, Sb and Se measured in the livers of tiger snakes paralleled the pattern of contamination measured in the wetland sediments where snakes were collected, and should be the focus of future research. Hence we propose the use of high trophic wetland snakes, such as tiger snakes, as bioindicators of wetland contamination. Although these contaminants may not be the only ones accumulating in tiger snakes (e.g. other environments might have other persistent contaminants),

we present the first data on these particular ecosystems. The potential impacts of these metal mixtures is unknown; however, research is currently being conducted on the physical and biochemical markers of health in tiger snakes at these wetlands (Lettoof et al., in prep.). Despite wetland degradation related to urbanisation, urban wetlands often provide the last refuge for a wide diversity of wildlife species including higher trophic snakes.

Although our data are limited we chose to sacrifice sample size to allow screening of more contaminants, and quantify both environment and predator contaminants. Therefore this study presents a snapshot of the first broad-scale contaminant screening of an Australian snake, and one of few studies, to our knowledge, that complements contaminants in snakes with contaminants in wetland sediment. It was also the first detailed investigation of contaminants in four iconic wetlands of a capital city in a developed country, reinforcing urgent government monitoring of urban wetland pollution to inform environmental management on actions required to preserve these highly productive and biodiverse habitats.

## References

Al-Maarofi SS, Alhello AZAR, Fawzi NA-M, Douabul AAZ, Al-Saad HT (2013) Desiccation versus Re-Flooding: Heavy Metals Mobilization—Part 1 Journal of Environmental Protection 4:27
Albrecht J, Abalos M, Rice TM (2007) Heavy metal levels in ribbon snakes (Thamnophis sauritus) and anuran larvae from the Mobile-Tensaw River Delta, Alabama, USA Arch Environ Contam Toxicol 53:647-654 doi:10.1007/s00244-006-0175-3
Alonso A, De Lange HJ, Peeters ET (2009) Development of a feeding behavioural bioassay using the freshwater amphipod Gammarus pulex and the Multispecies Freshwater Biomonitor Chemosphere 75:341-346 doi:10.1016/j.chemosphere.2008.12.031
ANZECC & ARMCANZ (2000) Australian and New Zealand guidelines for fresh and marine water quality. Australia and New Zealand Environment and Conservation Council & Agriculture and Resource Management, Council of Australia and New Zealand,

495	APHA (1998) Standard methods for the examination of water and wastewater. American Public
496	Health Association, American Water Works Association and Water Environment Federation
497	Washington DC
498	Appleyard SJ, Angeloni J, Watkins R (2006) Arsenic-rich groundwater in an urban area experiencing
499	drought and increasing population density, Perth, Australia Appl Geochem 21:83-97
500	doi:10.1016/j.apgeochem.2005.09.008
501	Arnold TN, Oldham CE (1997) Trace-element contamination of a shallow wetland in Western
502	Australia Mar Freshw Res 48:531-539 doi:Doi 10.1071/Mf96088
503	Aubret F, Bonnet X, Maumelat S, Bradshaw D, Schwaner T (2004) Diet divergence, jaw size and
504	scale counts in two neighbouring populations of tiger snakes (Notechis scutatus) Amphib-
505	Reptilia 25:9-17
506	Aubret F, Burghardt GM, Maumelat S, Bonnet X, Bradshaw D (2006) Feeding preferences in 2
507	disjunct populations of tiger snakes, Notechis scutatus (Elapidae) Behav Ecol 17:716-725
508	doi:10.1093/beheco/arl004
509	Bai J, Lu Q, Zhao Q, Wang J, Gao Z, Zhang G (2015) Organochlorine pesticides (OCPs) in wetland
510	soils under different land uses along a 100-year chronosequence of reclamation in a Chinese
511	estuary Sci Rep 5:17624 doi:10.1038/srep17624
512	Ball A, Truskewycz A (2013) Polyaromatic hydrocarbon exposure: an ecological impact ambiguity
513	Environ Sci Pollut Res Int 20:4311-4326 doi:10.1007/s11356-013-1620-2
514	Bazar M, Holtzman D, Adair B, Gresens S Effects of dietary methylmercury in juvenile corn snakes
515	(Elaphe guttata). In: Abstracts, SETAC 23rd Annual Meeting, Salt Lake City, Utah,
516	November, 2002. pp 16-20
517	Beck A (1956) The copper content of the liver and blood of some vertebrates Aust J Zool 4:1-18
518	Best SM (1973) Some organochlorine pesticide residues in wildlife of the Northern Territory,
519	Australia, 1970-71 Aust J Biol Sci 26:1161-1170
520	Brasfield SM, Bradham K, Wells JB, Talent LG, Lanno RP, Janz DM (2004) Development of a
521	terrestrial vertebrate model for assessing bioavailability of cadmium in the fence lizard

522	(Sceloporus undulatus) and in ovo effects on hatchling size and thyroid function
523	Chemosphere 54:1643-1651
524	Brown JS, Sutula M, Stransky C, Rudolph J, Byron E (2010) Sediment contaminant chemistry and
525	toxicity of freshwater urban wetlands in southern California 1 JAWRA Journal of the
526	American Water Resources Association 46:367-385
527	Bruhl CA, Pieper S, Weber B (2011) Amphibians at risk? Susceptibility of terrestrial amphibian life
528	stages to pesticides Environ Toxicol Chem 30:2465-2472 doi:10.1002/etc.650
529	Burger J et al. (2007) Metal levels in blood, muscle and liver of water snakes (Nerodia spp.) from
530	New Jersey, Tennessee and South Carolina Sci Total Environ 373:556-563
531	doi:10.1016/j.scitotenv.2006.06.018
532	Burger J, Gochfeld M, Jeitner C, Zappalorti R, Pittfield T, DeVito E (2017) Arsenic, Cadmium,
533	Chromium, Lead, Mercury and Selenium Concentrations in Pine Snakes (Pituophis
534	melanoleucus) from the New Jersey Pine Barrens Arch Environ Contam Toxicol 72:586-595
535	doi:10.1007/s00244-017-0398-5
536	Burkett D (2005) Nutrient contribution to hyper-eutrophic wetlands in Perth, Western Australia.
537	Deakin University
538	Butler H, Malone B, Clemann N (2005) The effects of translocation on the spatial ecology of tiger
539	snakes (Notechis scutatus) in a suburban landscape Wildl Res 32:165-171
540	doi:10.1071/Wr04020
541	Campbell KR, Campbell TS (2001) The accumulation and effects of environmental contaminants on
542	snakes: a review Environ Monit Assess 70:253-301 doi:10.1023/a:1010731409732
543	Campbell KR, Campbell TS, Burger J (2005) Heavy metal concentrations in northern water snakes
544	(Nerodia sipedon) from East Fork Poplar Creek and the Little River, East Tennessee, USA
545	Arch Environ Contam Toxicol 49:239-248 doi:10.1007/s00244-004-0200-3
546	Chadwick MA, Dobberfuhl DR, Benke AC, Huryn AD, Suberkropp K, Thiele JE (2006) Urbanization
547	affects stream ecosystem function by altering hydrology, chemistry, and biotic richness Ecol
548	Appl 16:1796-1807

549	Chin SY, Willson JD, Cristol DA, Drewett DV, Hopkins WA (2013) Altered behavior of neonatal
550	northern watersnakes (Nerodia sipedon) exposed to maternally transferred mercury Environ
551	Pollut 176:144-150 doi:10.1016/j.envpol.2013.01.030
552	City of Cockburn (2015) Integrated midge control strategy: Version 5. City of Cockburn, Perth,
553	Western Australia
554	Clarke K, Davis J, Murray F (1990) Herdsman Lake water quality study. Murdoch University, Perth,
555	Australia
556	Congdon R (1986) Nutrient loading and phytoplankton blooms in Lake Joondalup, Wanneroo,
557	Western Australia. Department of Conservation and Environment,
558	Cooper CM (1993) Biological Effects of Agriculturally Derived Surface Water Pollutants on Aquatic
559	Systems—A Review Journal of Environment Quality 22:402-408
560	doi:10.2134/jeq1993.00472425002200030003x
561	Davis JA, Froend R (1999) Loss and degradation of wetlands in southwestern Australia: underlying
562	causes, consequences and solutions Wetlands Ecol Manage 7:13-23
563	Davis JA, Garland M (1986) Herdsman Lake Pesticide Study. Department of Conservation and Land
564	Management, Perth, Western Australia
565	Davis JA, Rosich RS, Bradley JS, Growns JE, Schmidt LG, Cheal F (1993) Wetland classification on
566	the basis of water quality and invertebrate community data. Wetlands of the Swan Coastal
567	Plain vol 6. Water Authority of Western Australia and Enviornmental Protection Authority,
568	Perth, Western Australia
569	Day RD, Segars AL, Arendt MD, Lee AM, Peden-Adams MM (2007) Relationship of blood mercury
570	levels to health parameters in the loggerhead sea turtle (Caretta caretta) Environ Health
571	Perspect 115:1421-1428 doi:10.1289/ehp.9918
572	DEH (2004) National chemical reference guide – standards in the Australian environment.
573	Department of Environment and Heritage, Australian Government, Canberra
574	Department of Water GoWA (2009) A snapshot of contaminants in drains of Perth's industrial areas:
575	Industrial contaminants in stormwater of Herdsman Lake, Bayswater Drain, Bickley Brook
576	and Bibra Lake between October 2007 and January 2008.

577	Department of Water GoWA (2011) Perth Shallow Groundwater Systems Investigation: Loch
578	McNess. Perth, Western Australia
579	Drewett DV, Willson JD, Cristol DA, Chin SY, Hopkins WA (2013) Inter- and intraspecific variation
580	in mercury bioaccumulation by snakes inhabiting a contaminated river floodplain Environ
581	Toxicol Chem 32:1178-1186 doi:10.1002/etc.2157
582	Ehrenfeld JG (2004) The expression of multiple functions in urban forested wetlands Wetlands
583	24:719-733 doi:Doi 10.1672/0277-5212(2004)024[0719:Teomfi]2.0.Co;2
584	ESRI (1983) Herdsman Industrial Estate Western Australia. Phase 1. Environmental Monitoring
585	Report September 1982 to June 1983. ESRI Australia Pty. Ltd., Perth, Western Australia
586	Evans DW, Irwin SWB, Fitzpatrick S (2001) The effect of digenean (Platyhelminthes) infections on
587	heavy metal concentrations in Littorina littorea J Mar Biol Assoc U K 81:349-350 doi:Doi
588	10.1017/S0025315401003873
589	Faulkner S (2004) Urbanization impacts on the structure and function of forested wetlands Urban
590	Ecosyst 7:89-106
591	Fearn S, Norton I (2011) The oldest captive Australian snake? A longevity record for a chappell
592	island tiger snake (Notechis scutatus) in Tasmania Herpetofauna 41:7-8
593	Fitzpatrick M, Long D, Pijanowski B (2007) Exploring the effects of urban and agricultural land use
594	on surface water chemistry, across a regional watershed, using multivariate statistics Appl
595	Geochem 22:1825-1840
596	Fontenot LW, Noble GP, Akins JM, Stephens MD, Cobb GP (2000) Bioaccumulation of
597	polychlorinated biphenyls in ranid frogs and northern water snakes from a hazardous waste
598	site and a contaminated watershed Chemosphere 40:803-809
599	Ford WM, Hill EP (1991) Organochlorine Pesticides in Soil Sediments and Aquatic Animals in the
600	Upper Steele Bayou Watershed of Mississippi Arch Environ Contam Toxicol 20:161-167
601	doi:Doi 10.1007/Bf01055900
602	Forrow DM, Maltby L (2000) Toward a mechanistic understanding of contaminant-induced changes
603	in detritus processing in streams: Direct and indirect effects on detritivore feeding Environ
604	Toxicol Chem 19:2100-2106 doi:Doi 10.1897/1551-5028(2000)019<2100:Tamuoc>2.3.Co;2

605	Förstner U (2004) Sediments — resource or waste J Soils Sediments 4:3-3 doi:10.1007/bf02990821
606	Gambrell R (1994) Trace and toxic metals in wetlands—a review J Environ Qual 23:883-891
607	Garden J, McAlpine C, Peterson A, Jones D, Possingham HP (2006) Review of the ecology of
608	Australian urban fauna: A focus on spatially explicit processes Austral Ecol 31:126-148
609	doi:10.1111/j.1442-9993.2006.01578.x
610	Gardiner M, Gorman R (1963) Further observations on plant selenium levels in Western Australia
611	Australian Journal of Experimental Agriculture 3:284-289
612	Gavric JP et al. (2015) Effects of Metals on Blood Oxidative Stress Biomarkers and
613	Acetylcholinesterase Activity in Dice Snakes (Natrix Tessellata) from Serbia Archives of
614	Biological Sciences 67:303-315 doi:10.2298/Abs141203047g
615	Gentilli J, Bekle H (1993) History of the Perth lakes Early Days: Journal of the Royal Western
616	Australian Historical Society 10:442
617	Gibbs JP (2000) Wetland loss and biodiversity conservation Conserv Biol 14:314-317 doi:DOI
618	10.1046/j.1523-1739.2000.98608.x
619	Gonzalez-Pinto J, Lund M, Quintero Vasquez M (2017) Yellagonga Regional Park wetlands water
620	quality monitoring 2016/17 report. Mine Water and Environment Research Centre, Edith
621	Cowan University, Perth, Western Australia
622	Haarstad K, Bavor HJ, Maehlum T (2012) Organic and metallic pollutants in water treatment and
623	natural wetlands: a review Water Sci Technol 65:76-99 doi:10.2166/wst.2011.831
624	Halse S Wetlands of the Swan Coastal Plain past and present. In: Proceedings of the Swan Coastal
625	Plain Groundwater Management Conference'.(Ed. G. Lowe.) pp, 1989. pp 105-112
626	Hamer A, Organ A (2008) Aspects of the ecology and conservation of the Growling Grass Frog
627	Litoria raniformis in an urban-fringe environment, southern Victoria Aust Zool 34:393-407
628	Haskins DL, Gogal RM, Tuberville TD (2019) Snakes as Novel Biomarkers of Mercury
629	Contamination: A Review
630	Heydari Sereshk Z, Riyahi Bakhtiari A (2015) Concentrations of trace elements in the kidney, liver,
631	muscle, and skin of short sea snake (Lapemis curtus) from the Strait of Hormuz Persian Gulf
632	Environ Sci Pollut Res Int 22:15781-15787 doi:10.1007/s11356-015-4631-3

633	Hitchings SP, Beebee TJ (1997) Genetic substructuring as a result of barriers to gene flow in urban
634	Rana temporaria (common frog) populations: implications for biodiversity conservation
635	Heredity (Edinb) 79 ( Pt 2):117-127 doi:10.1038/hdy.1997.134
636	Hopkins WA (2007) Amphibians as models for studying environmental change ILAR J 48:270-277
637	doi:DOI 10.1093/ilar.48.3.270
638	Hopkins WA, Rowe CL, Congdon JD (1999) Elevated trace element concentrations and standard
639	metabolic rate in banded water snakes (Nerodia fasciata) exposed to coal combustion wastes
640	Environ Toxicol Chem 18:1258-1263 doi:Doi 10.1897/1551-
641	5028(1999)018<1258:Etecas>2.3.Co;2
642	Hopkins WA, Staub BP, Baionno JA, Jackson BP, Roe JH, Ford NB (2004) Trophic and maternal
643	transfer of selenium in brown house snakes (Lamprophis fuliginosus) Ecotoxicol Environ Saf
644	58:285-293 doi:10.1016/S0147-6513(03)00076-9
645	Hylland K (2006) Polycyclic aromatic hydrocarbon (PAH) ecotoxicology in marine ecosystems J
646	Toxicol Environ Health A 69:109-123 doi:10.1080/15287390500259327
647	Kapustka LA, Clements WH, Ziccardi L, Paquin PR, Sprenger M, Wall D Issue paper on the
648	ecological effects of metals. In: US Environmental Protection Agency, Risk Assessment
649	Forum, 2004.
650	Kinnear A, Garnett P, Bekle H, Upton K (1997) Yellagonga Wetlands: a study of the water chemistry
651	and aquatic fauna
652	Kobryn H (2001) Land use changes and the properties of stormwater entering a wetland on a sandy
653	coastal plain in Western Australia (PhD Thesis). Murdoch University
654	Lee S et al. (2006) Impact of urbanization on coastal wetland structure and function Austral Ecol
655	31:149-163
656	Lemaire J et al. (2018) Determinants of mercury contamination in viperine snakes, Natrix maura, in
657	Western Europe Sci Total Environ 635:20-25 doi:10.1016/j.scitotenv.2018.04.029
658	Lemly A (1996) Assessing the toxic threat of selenium to fish and aquatic birds Environ Monit Assess
659	43

660	Lettoof D, von Takach B, Bateman PW, Gagnon MM, Aubret F (2020) Investigating the role of
661	urbanisation, wetlands and climatic conditions in nematode parasitism in a large Australian
662	elapid snake International Journal for Parasitology: Parasites and Wildlife 11:32-39
663	doi:10.1016/j.ijppaw.2019.11.006
664	Mann RM, Hyne RV, Choung CB, Wilson SP (2009) Amphibians and agricultural chemicals: review
665	of the risks in a complex environment Environ Pollut 157:2903-2927
666	doi:10.1016/j.envpol.2009.05.015
667	Marco A, Lopez-Vicente M, Perez-Mellado V (2004) Arsenic uptake by reptile flexible-shelled eggs
668	from contaminated nest substrates and toxic effect on embryos Bull Environ Contam Toxicol
669	72:983-990
670	McKinney ML (2008) Effects of urbanization on species richness: A review of plants and animals
671	Urban Ecosyst 11:161-176 doi:10.1007/s11252-007-0045-4
672	Nasri I, Hammouda A, Hamza F, Zrig A, Selmi S (2017) Heavy metal accumulation in lizards living
673	near a phosphate treatment plant: possible transfer of contaminants from aquatic to terrestrial
674	food webs Environ Sci Pollut Res Int 24:12009-12014 doi:10.1007/s11356-015-5390-x
675	Newport M, Lund M (2014) Yellagonga Regional Park wetlandswater quality monitoring 2013-2014.
676	Mine Water and Environment Research Centre, Edith Cowan University, Perth, Western
677	Australia
678	Nice HE (2009) A baseline study of contaminants in the sediments of the Swan and Canning
679	estuaries: Water Science Technical Series Report No. 6. Department of Water, Government of
680	Western Australia,
681	Novitski R, Smith RD, Fretwell JD (1996) Wetland functions, values, and assessment National
682	Summary on Wetland Resources USGS Water Supply Paper 2425:79-86
683	Nriagu JO (1990) Global metal pollution: poisoning the biosphere? Environment: Science and Policy
684	for Sustainable Development 32:7-33
685	Ohlendorf HM, Hothem RL, Aldrich TW (1988) Bioaccumulation of Selenium by Snakes and Frogs
686	in the San-Joaquin Valley, California Copeia:704-710 doi:Doi 10.2307/1445391

687	Ontario Ministry of Environment and Energy (1993) Guidelines for the protection and management of
688	aquatic sediment quality in Ontario. <a href="https://www.ontario.ca/document/guidelines-identifying-">https://www.ontario.ca/document/guidelines-identifying-</a>
689	$\underline{assessing\text{-}and\text{-}managing\text{-}contaminated\text{-}sediments\text{-}ontario/identification\text{-}and\text{-}assessment}.$
690	Accessed 24 February 2019
691	Orange P (2007) Fossorial frog foraging by the western tiger snake, Notechis scutatus (Elapidae)
692	Herpetofauna 37:16-21
693	Panno SV, Nuzzo VA, Cartwright K, Hensel BR, Krapac IG (1999) Impact of urban development on
694	the chemical composition of ground water in a fen-wetland complex Wetlands 19:236-245
695	doi:Doi 10.1007/Bf03161753
696	Quintela FM, Lima GP, Silveira ML, Costa PG, Bianchini A, Loebmann D, Martins SE (2019) High
697	arsenic and low lead concentrations in fish and reptiles from Taim wetlands, a Ramsar site in
698	southern Brazil Sci Total Environ 660:1004-1014 doi:10.1016/j.scitotenv.2019.01.031
699	R Core Team (2018) R: A language and environment for statistical computing. R Foundation for
700	Statistical Computing, Vienna, Austria
701	Rainwater TR, Reynolds KD, Canas JE, Cobb GP, Anderson TA, McMurry ST, Smith PN (2005)
702	Organochlorine pesticides and mercury in cottonmouths (Agkistrodon piscivorus) from
703	northeastern Texas, USA Environ Toxicol Chem 24:665-673
704	Rich CN, Talent LG (2009) Soil Ingestion May Be an Important Route for the Uptake of
705	Contaminants by Some Reptiles Environ Toxicol Chem 28:311-315 doi:Doi 10.1897/08-
706	035.1
707	Rodriguez Martin JA, De Arana C, Ramos-Miras JJ, Gil C, Boluda R (2015) Impact of 70 years urban
708	growth associated with heavy metal pollution Environ Pollut 196:156-163
709	doi:10.1016/j.envpol.2014.10.014
710	Rohr JR et al. (2008) Agrochemicals increase trematode infections in a declining amphibian species
711	Nature 455:1235-1239 doi:10.1038/nature07281
712	Roy JW, Bickerton G (2011) Toxic groundwater contaminants: an overlooked contributor to urban
713	stream syndrome? Environ Sci Technol 46:729-736

714	Salice CJ, Suski JG, Bazar MA, Talent LG (2009) Effects of inorganic lead on Western fence lizards
715	(Sceloporus occidentalis) Environ Pollut 157:3457-3464 doi:10.1016/j.envpol.2009.06.013
716	Sandstead H (2014) Zinc. In: Handbook on the Toxicology of Metals, vol 1. 4th edn. Elsevier,
717	London, UK,
718	Schulz R, Peall SK (2001) Effectiveness of a constructed wetland for retention of nonpoint-source
719	pesticide pollution in the Lourens River catchment, South Africa Environ Sci Technol
720	35:422-426
721	Schwabenlander M, Buchweitz JP, Smith CE, Wunschmann A (2019) Arsenic, Cadmium, Lead, and
722	Mercury Concentrations in the Livers of Free-Ranging Common Garter Snakes (Thamnophis
723	sirtalis) from Minnesota, USA J Wildl Dis
724	Shine R (1987) Ecological Comparisons of Island and Mainland Populations of Australian
725	Tigersnakes (Notechis, Elapidae) Herpetologica 43:233-240
726	Sievers M, Hale R, Swearer SE, Parris KM (2019) Frog occupancy of polluted wetlands in urban
727	landscapes Conserv Biol 33:389-402 doi:10.1111/cobi.13210
728	Simoniello P, Trinchella F, Filosa S, Scudiero R, Magnani D, Theil T, Motta CM (2014) Cadmium
729	contaminated soil affects retinogenesis in lizard embryos J Exp Zool A Ecol Genet Physiol
730	321:207-219 doi:10.1002/jez.1852
731	Simpson G, Newsome D (2017) Environmental history of an urban wetland: from degraded colonial
732	resource to nature conservation area Geo-Geography and Environment 4:e00030
733	doi:10.1002/geo2.30
734	Simpson S, Batley G, Chariton A (2013) Revision of the ANZECC/ARMCANZ Sediment Quality
735	Guidelines. CSIRO Land and Water Science Report 08/07. CSIRO Land and Water.,
736	Sinang SC, Reichwaldt ES, Ghadouani A (2015) Local nutrient regimes determine site-specific
737	environmental triggers of cyanobacterial and microcystin variability in urban lakes Hydrology
738	and Earth System Sciences 19:2179-2195 doi:10.5194/hess-19-2179-2015
739	Smalling KL, Reeves R, Muths E, Vandever M, Battaglin WA, Hladik ML, Pierce CL (2015)
740	Pesticide concentrations in frog tissue and wetland habitats in a landscape dominated by
741	agriculture Sci Total Environ 502:80-90 doi:10.1016/j.scitotenv.2014.08.114

742	Soliman M, El-Shazly M, Abd-El-Samie E, Fayed H (2019) Variations in heavy metal concentrations
743	among trophic levels of the food webs in two agroecosystems Afr Zool 54:21-30
744	Spurgeon DJ, Hopkin SP (1999) Seasonal variation in the abundance, biomass and biodiversity of
745	earthworms in soils contaminated with metal emissions from a primary smelting works J Appl
746	Ecol 36:173-183 doi:10.1046/j.1365-2664.1999.00389.x
747	Stafford D, Plapp F, Fleet R (1977) Snakes as indicators of environmental contamination: relation of
748	detoxifying enzymes and pesticide residues to species occurrence in three aquatic ecosystems
749	Arch Environ Contam Toxicol 5:15-27
750	Tallkvist J, Oskarsson A (2015) Molybdenum. In: Handbook on the Toxicology of Metals, vol 4.
751	Elsevier, pp 1077-1089
752	UNEP (2011) Draft revised guidance on the global monitoring plan for persistent organic pollutants,
753	UNEP/POPS/COP.5/INF/27. United Nations Environment Programme, UNEP Chemicals,
754	Geneva, Switzerland
755	US EPA (1998) Method 8270D: Semivolatile organic compounds by gas chromatography/mass
756	spectrometry (GC/MS). <a href="https://www.epa.gov/sites/production/files/2015-07/documents/epa-">https://www.epa.gov/sites/production/files/2015-07/documents/epa-</a>
757	<u>8270d.pdf</u> .
758	US EPA (2007) Method 3051A: Microwave assisted acid digestion of sediments, sludges, soils, and
759	oils. https://www.epa.gov/sites/production/files/2015-12/documents/3051a.pdf.
760	Weir SM et al. (2013) Organochlorine pesticides in squamate reptiles from southern Arizona, USA
761	Bull Environ Contam Toxicol 90:654-659 doi:10.1007/s00128-013-0990-y
762	Wixon JG (2013) Bioaccumulation of Metals in an Insular Population of Florida Cottonmouth Snakes
763	(Agkistrodon piscivorus conanti). UNIVERSITY OF FLORIDA
764	Wu Y, Zhang J, Zhou Q (1999) Persistent organochlorine residues in sediments from Chinese
765	river/estuary systems Environ Pollut 105:143-150 doi:Doi 10.1016/S0269-7491(98)00160-2
766	Wylie GD, Hothem RL, Bergen DR, Martin LL, Taylor RJ, Brussee BE (2009) Metals and trace
767	elements in giant garter snakes (Thamnophis gigas) from the Sacramento Valley, California,
768	USA Arch Environ Contam Toxicol 56:577-587 doi:10.1007/s00244-008-9265-8

769	Zedler JB, Kercher S (2005) Wetland resources: Status, trends, ecosystem services, and restorability
770	Annual Review of Environment and Resources 30:39-74
771	doi:10.1146/annurev.energy.30.050504.144248
772	Zeghnoun A, Pascal M, Fréry N, Sarter H, Falq G, Focant J-F, Eppe G (2007) Dealing with the non-
773	detected and non-quantified data. The example of the serum dioxin data in the French dioxin
774	and incinerators study Organohalog Compd 69:2288-2291
775	Zhang Q et al. (2017) Transboundary health impacts of transported global air pollution and
776	international trade Nature 543:705
777	Zhang Z, Cui B, Fan X (2012) Removal mechanisms of heavy metal pollution from urban runoff in
778	wetlands Frontiers of Earth Science 6:433-444 doi:10.1007/s11707-012-0301-7
779	

Table 1: Brief description and history of contaminants detected within the wetland sites used for study. Area was calculated using Google Earth Pro polygon tool based on the most recent aerial photographs. Sites are listed in order of most urbanised to least.

Site	Area	Brief description of	References	Contaminants detected	References
	(km <sup>2</sup> )	urbanisation		(S = sediment, W = water)	
Herdsman Lake	3.07	Heavily modified;	(Clarke et al. 1990;	(W) Cr, Cu, Pb, Ni, Zn, Fe, Mn	(ESRI 1983)
(HL)		• since 1850s has been	Gentilli and Bekle		
		subject to agriculture,	1993; Kobryn 2001)	(W) Cd, Cu, Pb, Zn;	(Clarke et al. 1990)
31° 55'12 S,		industrial dumping,		(W) Dieldrin, heptachlor	
115° 48'19 E		dredging and storm			
		water inflow;		(S) Cu, Zn, As;	(Davis et al. 1993)
		<ul> <li>located among heavy</li> </ul>		(S) Aldrin, Chlordane, dieldrin, DDT	
		urbanisation		(W) Cd, Cu, Pb, Zn	(Kobryn 2001)
				(W,S) Al, Cu, Pb, Zn, As;	(Department of Water
				(S) PCBs, PHCs	2009)
Bibra Lake	1.93	• Partially modified;	(Sinang et al. 2015)	(S) Dieldrin	(Davis et al. 1993)

(BI)		• some fringe is (W) Cu, Pb, Zn	(Burkett 2005)
		urbanised; (W,S) Al, Cr, Cu, Pb, Zn;	(Department of Water
32° 5'32 S,		• located among heavy (S) PHCs, PAHs	2009)
115° 49'27 E		urbanisation	
Lake Joondalup	7.33	• Partially modified; (Congdon 1986; (S) As	(Davis et al. 1993)
(JL)		• some fringe is Gonzalez-Pinto et al.  (W) Al, As, Hg, Zn  urbanised; 2017)	(Newport and Lund
31° 45'34 S,		• located on edge of	2014)
115° 47'33 E		recent urbanisation (W) Al, Cd, Hg, Zn	(Gonzalez-Pinto et al. 2017)
<b>Loch McNess</b>	0.36	• Minimally modified; (Department of (S) As	(Davis et al. 1993)
(LM)		• surrounded by Yanchep Water 2011)  (W) As, B, Cr, Mn, Ni  National Park;	(Department of Water 2011)
31° 32'44 S,		• located outside of	
115° 40'50 E		urbanisation	

Table 2. Morphological measurements (mean  $\pm$  one SE (range)) of tiger snakes (*Notechis scutatus occidentalis*) collected at each site.

Site	Sex (n)	SVL (mm)	Body mass (g)	Wet liver mass (g)
Herdsman	Male (4)	855 ± 12.6	250 ± 15.15	$5.2 \pm 0.6$
Lake		(825 - 877)	(216 - 282)	(4.0 - 6.3)
	Female (1)	776	277	3.9
Bibra Lake	Male (4)	$807 \pm 21.9$	$283 \pm 20.2$	$9.1 \pm 1.2$
		(746 - 847)	(233 – 329)	(6.6 – 11.7)
	Female (1)	704	212	4.8
Lake	Male (4)	$779 \pm 15.5$	$221 \pm 26.6$	$7.4 \pm 0.8$
Joondalup		(735 - 807)	(171 - 296)	(6.8 - 9.3)
	Female (1)	706	205.6	11.1
Loch McNess	Male (3)	$847 \pm 43.6$	$294 \pm 44.2$	$11.2 \pm 2.0$
(Yanchep)		(760 – 899)	(235 – 381)	(7.3 – 13.5)

Female (2)	$719 \pm 47.5$	$194 \pm 14.4$	$4.2 \pm 0.9$
	(671 – 766)	(180 -208)	(3.2 - 5.1)

85 SVL: snout-vent length

Figure 1. Wetlands in the Swan Coastal Plain where sediments and tiger snakes were collected for contaminant analyse. Red dots indicate collection points of sediment samples, yellow circles indicate tiger snakes collection areas. HL = Herdsman Lake, BI = Bibra Lake, JL = Lake Joondalup, LM = Loch McNess (located in Yanchep National Park). Satellite images were obtained from Google Earth Pro in 2019. Scale bar = 500m.

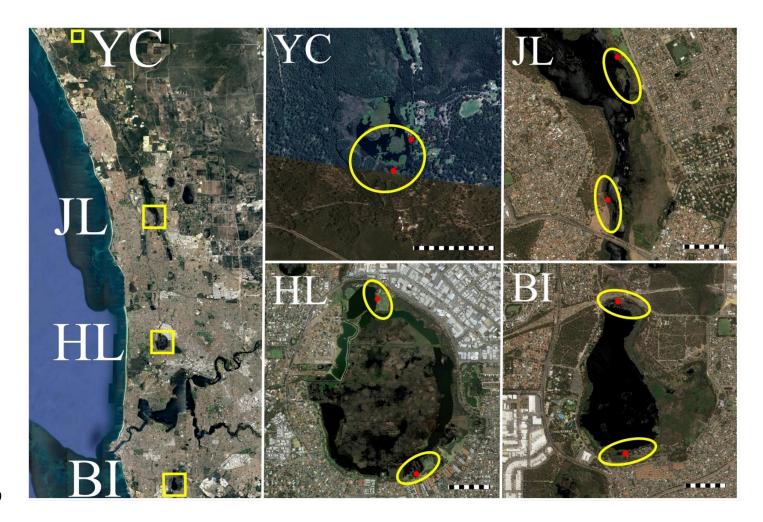


Table 3. Concentrations (mg/kg, dry weight) of contaminants detected in sediments. Given as: mean ± one SE (range). Concentrations in bold indicate values
 higher than the ANZECC sediment quality guideline value (SQGV).

Contaminant	Herdsman Lake	Bibra Lake	Lake Joondalup	Loch McNess	ANZECC SQGV
					(mg/kg, dry weight)
Aldrin	< 0.01	<b>0.14</b> ± 0.13	<0.01	< 0.01	SQGV: 0.002#
		(BDL-0.53)			SQG-High: NA
Dieldrin	<0.01	$0.65 \pm 0.65$	<0.01	<0.01	SQGV: 0.12
		(<0.01 – <b>2.60</b> )			SQG-High: 0.27
Phenanthrene	<0.5	<b>0.49</b> ± 0.24	<0.5	<0.5	SQGV: 0.24
		(<0.5 – <b>1.20</b> )			SQG-High: 1.5
Fluoranthene	<0.5	<b>0.86</b> ± 0.61	<0.5	<0.5	SQGV: 0.6
		(<0.5 – <b>2.70</b> )			SQG-High: 5.1
Pyrene	<0.5	<b>1.41</b> ± 1.17	<0.5	<0.5	SQGV: 0.665

		(<0.5 – <b>4.90</b> )			SQG-High: 2.6
Benz(a)anthracene	<0.5	$0.66 \pm 0.42$ (<0.5 – <b>1.90</b> )	<0.5	<0.5	SQGV: 0.261 SQG-High: 1.6
Chrysene	<0.5	$1.09 \pm 0.84$ $(<0.5 - 3.600)$	<0.5	<0.5	SQGV: 0.384 SQG-High: 2.8
Benzo(a)pyrene	<0.5	$0.69 \pm 0.44$ (<0.5 - <b>2.00</b> )	<0.5	<0.5	SQGV: 0.43 SQG-High: 1.6
Benzo(g,h,i)perylene	<0.5	$0.44 \pm 0.92$ (<0.5 – 1.00)	<0.5	<0.5	SQGV: NA SQG-High: NA
Antimony	$0.77 \pm 0.03^{a,b}$ $(0.68 - 0.83)$	$0.23 \pm 0.5$ $(0.10 - 0.32)$	$0.18 \pm 0.04^{a}$ (<0.05 – 0.15)	$0.20 \pm 0.05^{b}$ (<0.05 – 0.06)	SQGV: 2 SQG-High: 25

Arsenic	$20.78 \pm 6.54^{a}$	$1.80\pm0.35$	$0.88\pm0.17^{\rm a}$	$7.12 \pm 3.50$	SQGV: 20
	(9.10 – <b>34.00</b> )	(1.20 - 2.80)	(0.40 - 1.20)	(0.60 - 15.00)	SQG-High: 70
Barium	$91.75 \pm 30.10$	$85.50 \pm 9.32$	$31.00 \pm 9.19$	$86.75 \pm 49.74$	SQGV: NA
	(24.00 - 160.00)	(65.00 - 110.00)	(18.00 - 58.00)	(11.00 - 230.00)	SQG-High: NA
Beryllium	$0.18 \pm 0.12$	$0.22 \pm 0.07$	$0.04 \pm 0.01$	$0.08 \pm 0.04$	SQGV: NA
	(<0.05 – 0.28)	(0.09 - 0.41)	(<0.05-0.06)	(<0.05 – 0.16)	SQG-High: NA
Cadmium	$0.14 \pm 0.06$	$0.14 \pm 0.03$	$0.06\pm0.02$	$0.26 \pm 0.19$	SQGV: 1.5
	(<0.05 – 0.28)	(0.07 - 0.22)	(<0.05 – 0.10)	(<0.05-0.80)	SQG-High: 10
Chromium	$13.43 \pm 2.99$	$11.23 \pm 2.88$	$5.73 \pm 1.24$	$20.45 \pm 8.37$	SQGV: 60
	(8.70 - 22.00)	(6.10 - 19.00)	(3.60 - 8.60)	(2.80 - 38.00)	SQG-High: 370
Cobalt	$2.23\pm0.31^a$	$0.70 \pm 0.14$	$1.20\pm0.07^{\rm a}$	$1.40 \pm 0.64$	SQGV: NA
	(1.70 - 3.10)	(0.40 - 1.10)	(0.10 - 0.40)	(0.20 - 2.90)	SQG-High: 50#

Copper	$76.50 \pm 45.07^{a}$	$7.40\pm1.25$	$5.05\pm1.54^a$	$10.87 \pm 3.69$	SQGV: 65
	(20.00 - <b>210.00</b> )	(3.90 - 9.80)	(0.50 - 6.90)	(3.10 - 19.00)	SQG-High: 270
Lead	$43.75 \pm 8.84^{a,b}$	$20.50 \pm 1.66$	$14.08 \pm 3.57^{\rm a}$	$9.20\pm3.96^{b}$	SQGV: 50
	(31.00 – <b>69.00</b> )	(18.00 - 25.00)	(6.30 - 23.00)	(1.50 - 17.00)	SQG-High: 220
Manganese	$68.75 \pm 23.26$	$30.08 \pm 8.56$	$26.67 \pm 17.18$	$20.07 \pm 10.38$	SQGV: 460#
	(25.00 - 120.00)	(6.30 - 47.00)	(7.20 - 78.00)	(2.30 - 50.00)	SQG-High: 1100#
Mercury	$0.05\pm0.01$	$0.05 \pm 0.005$	$0.14 \pm 0.08$	$0.10 \pm 0.05$	SQGV: 0.15
	(0.01 - 0.07)	(0.04 - 0.06)	(0.01 - 0.35)	(0.01 - 0.21)	SQG-High: 1
Molybdenum	$1.19\pm0.27^{\rm a}$	$0.49 \pm 0.09$	$0.16\pm0.07^a$	$0.88 \pm 0.39$	SQGV: NA
	(0.65 - 1.7)	(0.27 - 0.71)	(0.07 - 0.37)	(0.19 - 1.7)	SQG-High: NA
Nickel	$7.48 \pm 1.18^a$	$3.80 \pm 0.59$	$0.55\pm0.16^a$	$2.55 \pm 0.99$	SQGV: 21

	(6.00 - 11.00)	(2.50 - 5.30)	(0.30 - 1.00)	0.50 - 4.60	SQG-High: 52
Selenium	$0.59 \pm 0.14$	$0.96\pm0.36^a$	$0.13 \pm 0.29^{a,b}$	$4.04 \pm 2.14^{b}$	SQGV: 2#
	(0.38 - 0.97)	(0.45 - 2.00)	(0.08 - 0.21)	(0.23 – <b>7.90</b> )	SQG-High: 4 <sup>#</sup>
Silver	$0.09 \pm 0.01$	< 0.05	< 0.05	< 0.05	SQGV: 1
	(0.07 - 0.11)				SQG-High: 3.7
Tin	$8.10 \pm 5.31^{a}$	$1.00 \pm 0.14$	< 0.5	$0.60 \pm 0.21^a$	SQGV: NA
	(2.00 - 24.00)	(0.60 - 1.30)		(<0.5 – 1.10)	SQG-High: NA
Zinc	$221.00 \pm 105.15^{a}$	$60.50 \pm 14.91$	$12.12 \pm 5.57^{a,b}$	$31.00 \pm 12.57^{b}$	SQGV: 200
	(41.00 – <b>510.00</b> )	(18.00 - 84.00)	(<5.00 – 28.00)	(11.00 - 67.00)	SQG-High: 410

Lower-case letter indicates Kruskal-Wallis significant difference (p = <0.05) between sites. # alternative guidelines (Lemly, 1996; Ontario Ministry of

<sup>793</sup> Environment and Energy, 1993).

Table 4. Concentrations (mg/kg, wet weight) of metals detected in tiger snake livers. N = 5 per site. Contaminants analysed at lower than detectable limits
 were not included. Given as: mean ± one SE (range); BDL = below detectable limits.

Contaminant	Herdsman Lake	Bibra Lake	Lake Joondalup	Loch McNess
Antimony (Sb)	$0.042 \pm 0.005^a$	$0.018 \pm 0.004$	$0.014 \pm 0.002$	$0.006 \pm 0.001^{a}$
	(0.025 - 0.052)	(0.007 - 0.03)	(0.010 - 0.019)	(0.003 - 0.008)
Arsenic (As)	$0.388 \pm 0.023^{\mathrm{a}}$	$0.098 \pm 0.012^{a}$	$0.188 \pm 0.082$	$0.276 \pm 0.049$
	(0.33 - 0.45)	(0.06 - 0.13)	(0.05 - 0.5)	(0.18 - 0.44)
Barium (Ba)	$0.122 \pm 0.015^{a}$	$0.039 \pm 0.009^a$	$0.32 \pm 0.270$	$0.089 \pm 0.023$
	(0.09 - 0.16)	(<0.05 – 0.06)	(<0.05 – 1.4)	(0.025 - 0.140)
Cadmium (Cd)	$0.023 \pm 0.004$	$0.015 \pm 0.004^{a}$	$0.02 \pm 0.006$	$0.057 \pm 0.013^{a}$
` ,	(0.014 - 0.035)	(<0.001 – 0.025)	(0.006 - 0.039)	(0.025 - 0.1)
Chromium (Cr)	$0.106 \pm 0.016$	$0.123 \pm 0.037$	$0.137 \pm 0.050$	$0.068 \pm 0.021$
	(0.06 - 0.15)	(<0.05-0.25)	(<0.05-0.28)	(<0.05-0.13)

Cobalt (Co)	$0.080 \pm 0.018^{a,b}$	$0.028 \pm 0.004^a$	$0.021 \pm 0.003^{b,c}$	$0.048 \pm 0.006^{c}$
	(0.054 - 0.15)	(0.019 - 0.041)	(0.013 - 0.031)	(0.032 - 0.064)
Copper (Cu)	$7.28 \pm 1.006^{\rm \ a,b}$	$3.9 \pm 0.376^{\rm  a,c}$	$3.68 \pm 0.676^{b,d}$	$9.84 \pm 1.518^{c,d}$
	(4.9 - 9.8)	(3.1 - 5.2)	(2.4 - 5.9)	(6.6 - 15.0)
Lead (Pb)	$0.007 \pm 0.004$	< 0.05	< 0.05	$0.047 \pm 0.028$
	(<0.05 – 0.024)			(<0.05 – 0.15)
Manganese (Mn)	$0.83 \pm 0.048$	$0.75 \pm 0.018$	$0.664 \pm 0.051$	$0.848 \pm 0.152$
	(0.7 - 0.94)	(0.70 - 0.79)	(0.49 - 0.76)	(0.54 - 1.4)
Mercury (Hg)	$0.164 \pm 0.009^{a,b}$	$0.061 \pm 0.02^{\mathrm{a,c}}$	$0.064 \pm 0.014^{b,d}$	$0.29 \pm 0.068^{c,d}$
	(0.14 - 0.19)	(BDL - 0.12)	(0.01 - 0.09)	(0.14 - 0.39)
Molybdenum (Mo)	$8.32 \pm 1.547^{\rm a,b,c}$	$1.36 \pm 0.214^{a}$	$1.98 \pm 0.666^{b}$	$1.36 \pm 0.144^{\circ}$

	(4.8 - 13.0)	(0.6 - 1.9)	(0.7 - 4.2)	(1.0 - 1.8)
Nickel (Ni)	$0.275 \pm 0.121$	$0.372 \pm 0.100$	$0.42 \pm 0.139$	$0.218 \pm 0.046$
	(<0.01 – 0.64)	(0.03 - 0.61)	(0.05 - 0.91)	(0.05 - 0.32)
Selenium (Se)	$0.988 \pm 0.061$	$1.03 \pm 0.139$	$0.49\pm0.058^a$	$1.53\pm0.208^a$
	(0.84 - 1.2)	(0.67 - 1.4)	(0.34 - 0.66)	(0.95 - 2.2)
Silver (Ag)	$0.014 \pm 0.002$	$0.007 \pm 0.002$	$0.006\pm0.002$	$0.008 \pm 0.001$
	(0.007 - 0.017)	(0.002 - 0.012)	(0.002 - 0.01)	(0.004 - 0.011)
Tin (Sn)	< 0.05	< 0.05	$0.058 \pm 0.033$	< 0.05
			(<0.05 – 0.19)	
Zinc (Zn)	$24 \pm 2.168$	$21.4 \pm 1.600$	$21.6 \pm 1.077$	$24.6 \pm 1.721$
	(20.0 - 32.0)	(17.0 - 27.0)	(19.0 - 24.0)	(21.0 - 31.0)

Lower-case letter indicates Kruskal-Wallis significant difference (p = <0.05) between sites.

Figure 2. The comparison between metals that were statistically significant in either the sediment or tiger snake livers between sites. Inter-site significant differences are listed in Tables 3 and 4. Left y axis represents sediment concentration and right y axis represents liver concentration, except for Hg and Se that share the same range. HL = Herdsman Lake, BI = Bibra Lake, JL = Lake Joondalup, LM = Loch McNess (located in Yanchep National Park). Patterned bars = mean sediment concentration; black bars = mean snake liver concentration. Sediment concentration corresponds to the left axis and liver to the right axis. Error bars = SE.

