

Manuscript version: Author's Accepted Manuscript

The version presented in WRAP is the author's accepted manuscript and may differ from the published version or Version of Record.

Persistent WRAP URL:

<http://wrap.warwick.ac.uk/152080>

How to cite:

Please refer to published version for the most recent bibliographic citation information.

Copyright and reuse:

The Warwick Research Archive Portal (WRAP) makes this work by researchers of the University of Warwick available open access under the following conditions.

Copyright © and all moral rights to the version of the paper presented here belong to the individual author(s) and/or other copyright owners. To the extent reasonable and practicable the material made available in WRAP has been checked for eligibility before being made available.

Copies of full items can be used for personal research or study, educational, or not-for-profit purposes without prior permission or charge. Provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way.

Publisher's statement:

Please refer to the repository item page, publisher's statement section, for further information.

For more information, please contact the WRAP Team at: wrap@warwick.ac.uk.

1
2
3 **Biochemical abnormalities in coronavirus disease 2019 (COVID-19): a comparison of White vs.**
4
5 **ethnic minority populations in the U.K.**
6
7

8 David R. Taylor¹, Devon Buchanan¹, Wiaam Al-Hasani¹, Jessica Kearney², Tina Mazaheri¹, Ruvini N.K

9
10 Ranasinghe¹, Georgios K. Dimitriadis² and Royce P. Vincent¹
11

12
13 Departments of ¹Clinical Biochemistry (Viapath) and ²Endocrinology, King's College Hospital NHS

14
15 Foundation Trust, London, SE5 9RS
16
17

18
19
20
21 **Short Title:** Biochemistry at presentation in ethnic minority and White COVID-19 patients at hospital
22
23 presentation
24
25

26
27
28 **Address for correspondence:**

29 David Taylor

30
31 Department of Clinical Biochemistry (Viapath),

32
33 First Floor, Bessemer Wing,

34
35 King's College Hospital

36
37 Denmark Hill

38
39 London SE5 9RS

40
41 Tel: 0203 299 3009

42
43 Email: davidtaylor8@nhs.net
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 **Acknowledgements:** None
4

5
6 **Declaration of conflicting interests:** The authors declared no potential conflicts of interest with
7
8 respect to the research, authorship and/or publication of this article.
9

10
11 **Funding:** The author(s) received no financial support for the research, authorship and/or publication
12
13 of this article
14

15
16 **Ethical approval:** Approved by the King's College Hospital Clinical Audit Committee. Ref ENDOC01
17

18
19 **Guarantor:** DRT
20

21
22 **Contributorship:** All authors contributed to design and execution of the study. All authors reviewed
23
24 and edited the article and approved the final version of the manuscript.
25

26
27 **ORCID iD**
28

29 David R Taylor <https://orcid.org/0000-0003-1385-3961>
30

31 Devon Buchanan <https://orcid.org/0000-0003-1175-7212>
32

33 Tina Mazaheri <https://orcid.org/0000-0001-8655-2372>
34

35 Jessica Kearney <https://orcid.org/0000-0001-7216-4408>
36

37 Ruvini N K Ranasinghe <https://orcid.org/0000-0002-6603-8322>
38

39 Georgios K Dimitriadis <https://orcid.org/0000-0002-6662-804X>
40

41 Royce P Vincent <https://orcid.org/0000-0003-0743-4189>
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Abstract

Aims: Public Health England have identified that in coronavirus disease 2019 (COVID-19), death rates amongst ethnic minorities far exceeds that of the White population. Whilst the increase in ethnic minorities is likely to be multi-factorial; to date, no studies have looked to see whether values for routine clinical biochemistry parameters differ between ethnic minority and White individuals.

Methods: Baseline biochemical data for 22 common tests from 311 SARs-CoV-2 positive patients presenting to hospital in April 2020 in whom ethnicity data was available was retrospectively collected and evaluated. Data comparisons between ethnic minority and White groups were made for all patient data and for the subset of patients subsequently admitted to intensive care.

Results: When all patient data were considered, the ethnic minority population had statistically significant higher concentrations of CRP, AST and GGT, whilst troponin T was higher in the White group. A greater proportion of ethnic minority patients were subsequently admitted to intensive care, but when the presenting biochemistry of this subset of patients was compared, no significant differences were observed between ethnic minority and White groups.

Conclusion: Our data show for the first time that routine biochemistry at hospital presentation in COVID-19 differs between ethnic minority and White groups. Amongst the markers identified, CRP was significantly higher in the ethnic minority group pointing towards an increased tendency for severe inflammation in this group.

Keywords: SARs-CoV-2, COVID-19, ethnic minority, ethnicity, routine biochemistry

Abbreviations: ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; BMI, body mass index; CK, creatine kinase; CKD, chronic kidney disease; COPD, chronic obstructive pulmonary disease; COVID-19, coronavirus disease 2019; CRP, C-reactive protein;

1
2
3 CVD, cardiovascular disease; eGFR, estimated glomerular filtration rate; GGT, gamma-
4
5 glutamyltransferase; IL-6, interleukin-6; ITU, intensive care unit; LDH, lactate dehydrogenase; NT-
6
7 proBNP, N-terminal prohormone of brain natriuretic peptide; SARS-CoV-2, severe acute respiratory
8
9 syndrome coronavirus 2.
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Confidential: For Review Only

Introduction

In December 2019 a new highly infectious disease, coronavirus disease 2019 (COVID-19), was first reported in Wuhan, Hubei Province, China.^{1,2} The causative agent of COVID-19, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has since spread worldwide resulting in 88 387 352 cases and 1 919 204 deaths as of January 10 2021, according to a weekly epidemiological update from the World Health Organization.³

Severe or fatal COVID-19 infection has been associated with gross changes in Clinical Biochemistry parameters. To date, common findings include increases of markers of tissue damage (creatinine kinase (CK), lactate dehydrogenase (LDH), myoglobin, troponin), inflammation (C-reactive protein (CRP), ferritin, procalcitonin), renal impairment (increased creatinine and urea) and liver dysfunction (increases of amino transferases and bilirubin, decreased albumin).^{4,5} Severe COVID-19 infection has also been associated with low serum sodium, potassium and calcium.⁶ Biochemical data can be predictive in COVID-19; parameters which are predictive of death include increased CRP, LDH, aspartate aminotransferase (AST), troponin I and creatinine and low albumin.⁷⁻¹⁰

In a review by Public Health England, death rates amongst Black, Asian and other minority ethnicities COVID-19 positive people were shown to be significantly higher than in White British people.¹¹ Death rates in those of Bangladeshi background were twice as high, and for other ethnic minority groups between 10-50% higher, after taking into account age, sex, deprivation, and region. The cause of these discrepancies is unclear, but likely to be multifactorial. We have previously assessed differences in cardiac markers at hospital presentation in ethnic minority and White groups in COVID-19¹², but to date, no study has investigated whether more routine biochemistry broadly differ in this setting, nor whether any differences provide prognostic value. To address these questions, we retrospectively reviewed admission biochemistry for a cohort of COVID-19 positive patients in whom ethnicity data

1
2
3 was available. We compared all data between ethnic minority and White groups and in a subset of
4
5 patients who were subsequently admitted to an intensive care unit (ITU).
6
7
8
9

10 **Methods**

11
12
13 This retrospective observational study was conducted at King's College Hospital NHS Foundation Trust,
14
15 a busy teaching hospital located in South London. Patients with baseline biochemistry data were
16
17 included in the analysis if they were admitted between the 1st and 28th of April 2020 (the period of
18
19 peak admission rates in London during the first wave of the pandemic), had positive (RT-PCR) SARS-
20
21 CoV-2 serology and if ethnicity data was available. The first available result after hospital admission
22
23 for the following 22 biochemical tests were obtained for each patient: albumin, alkaline phosphatase
24
25 (ALP), alanine aminotransferase (ALT), aspartate aminotransferase (AST), total bilirubin, adjusted
26
27 calcium, CK, creatinine, CRP, estimated glomerular filtration rate (eGFR), ferritin, gamma-glutamyl
28
29 transferase (GGT), LDH, magnesium, sodium, N-terminal prohormone of brain natriuretic peptide (NT-
30
31 proBNP), procalcitonin, phosphate, potassium, troponin T, total protein and urea. Inclusion of
32
33 admission biochemical data in the ITU analyses was made if the individual was subsequently
34
35 transferred to ITU within 28 days of admission. Electronic patient records were accessed for body
36
37 mass index (BMI) data and to assess for the presence of pre-existing common comorbidities in each
38
39 patient at baseline (defined as histories of diabetes mellitus, cardiovascular disease (CVD), chronic
40
41 kidney disease (CKD), hypertension and chronic obstructive pulmonary disease ((COPD) or asthma).
42
43 Patients were classified as "White" or "ethnic minority" using the Office for National Statistics list of
44
45 ethnic groups. All biochemical data were generated using Roche c-702 and e-801 analytical platforms
46
47 (Roche, Burgess Hill, U.K.), using blood samples collected into serum separator tubes (Greiner Bio-One
48
49 Ltd, Stonehouse, U.K.). All tests are accredited by the United Kingdom Accreditation Service to
50
51 iso15189. The methods used for ALT and AST included pyridoxal phosphate, with the LDH assay being
52
53 measured in the L-lactate to pyruvate direction. Test requests on samples for which haemolysis, icteric
54
55
56
57
58
59
60

1
2
3 or lipaemic indices exceeded the manufacturer's limits for that particular test were cancelled and not
4 included in analyses. Biochemical test data were tested for normality using the Kolmogorov-Smirnov
5 test, and with the exception of albumin, adjusted calcium, globulin, LDH, magnesium, potassium and
6 total protein, were found to be not normally distributed so group comparisons of biochemical data
7 were made using a two tailed Mann-Whitney U test. Data are reported as median (interquartile
8 range). Values of $P < 0.05$ were taken as statistically significant. The proportion of abnormal results
9 for each test studies was compared using a Chi-square test. Data comparisons were made using R
10 version 3.6.2 (R Foundation for Statistical Computing, Vienna, Austria).¹³

23 24 **Results**

25
26
27 Ethnic minority patients presenting in April 2020 with COVID-19, in comparison to those from White
28 ethnic groups, were younger (median age 65 vs. 75 years) and predominantly male (64% vs. 52%, Table
29 1). Ethnic minority individuals were more likely to subsequently require ITU admission (19% patients
30 vs. 13% White patients, Table 1). A history of diabetes mellitus was more common in the ethnic
31 minority group, whilst CVD history was more common in the White group. Frequencies of
32 hypertension, CKD and respiratory disorders (COPD or asthma) were similar in ethnic minority and
33 White groups (Table 1). BMI data was incomplete across the two groups. Of available data (151/211
34 ethnic minority patients and 70/100 White patients), BMI was similar in the two groups. Median BMI
35 (interquartile range) in the ethnic minority group was 28.8 kg/m² (24.7 – 33.5) and in the White group
36 28.7 kg/m² (23.3 – 32.0).

37
38
39 Review of baseline biochemistry data for all patients revealed statistically significant differences
40 between the two groups for a number of common analytes (Table 2). Of these results, 86% were
41 obtained on hospital admission day, with 97% of test results being acquired within the first 3 days of
42 the patients hospital stay. The majority of patients in the study had abnormal CRP results, however,
43 CRP concentrations were higher in the ethnic minority group (median value 111.2 mg/L, interquartile
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 range 66.5 – 181.1) than in the White group (48.1 mg/L, 22.1 – 112.9). Increase of this inflammatory
4
5 marker was not reflected by ethnicity-related differences in procalcitonin, nor ferritin concentration;
6
7 almost all patients had elevated ferritin. The median cardiac troponin T was higher, and a greater
8
9 proportion of patients had abnormal results, in the White group than in the ethnic minority group.
10
11 Median NT-proBNP concentration was also higher in the White group, although this association did
12
13 not reach statistical significance. Of markers of liver function, AST and GGT were higher in the ethnic
14
15 minority group, but ALP lower. A greater proportion of the ethnic minority population also had AST
16
17 concentrations falling outside the reference interval. ALT, albumin and total bilirubin were not
18
19 different between the two groups. No differences between ethnic minority and White groups were
20
21 noted for sodium, potassium, adjusted calcium or phosphate, nor for markers of renal function
22
23 (creatinine, eGFR and urea). The tissue damage markers, LDH and CK, also showed no ethnicity-specific
24
25 differences at presentation, with the majority of patients having abnormal values.
26
27
28

29
30 When baseline biochemistry data for ethnic minority and White patients that were subsequently
31
32 admitted to ITU were compared, no statistically significant differences between these groups were
33
34 observed for any of the analytes studied (Table 3).
35
36
37
38
39
40

41 **Discussion**

42
43 For the first time, we have shown that there are significant differences at hospital presentation
44
45 between ethnic minority and White populations in results for a number of routine biochemistry tests.
46
47 The results of this study are therefore important because they may contribute towards greater
48
49 understanding of why ethnic minority individuals are at increased risk of death due to COVID-19. The
50
51 most striking finding from our study is the increase at presentation of CRP in ethnic minority
52
53 individuals versus White individuals. CRP measurement is now well established as a marker of disease
54
55 severity in COVID-19.¹⁴ Zhang *et al.*¹⁵ showed that in 140 hospitalized patients with confirmed SARS-
56
57 CoV-2 infection, in non-severe disease CRP concentrations ranged from 9.5–52.1 mg/L, whilst in
58
59
60

1
2
3 severe disease values ranged from 20.6–87.1 mg/L. In another study, 56.4% of patients with non-
4
5 severe COVID-19 had CRP above the reference interval, which rose to 81.5% in those with severe
6
7 disease.¹⁶ Around 20% of patients infected with SARS-CoV-2 progress to having associated life-
8
9 threatening complications involving acute inflammation associated with a cytokine storm,
10
11 coagulopathy, septic shock and multiple organ failure.¹⁷ Increased concentrations of interleukin-6 (IL-
12
13 6) are associated with severe COVID-19 and positively correlate with adverse outcomes.^{18,19} The
14
15 increased concentrations of IL-6 directly result in the liver increasing synthesis of CRP.¹⁸ The results
16
17 from this study raise the interesting possibility that the higher concentrations of CRP in ethnic minority
18
19 patients at hospital presentation may mark an increased susceptibility to severe inflammation during
20
21 their COVID-19 disease course. These differences appear not to be accounted for by a genetic
22
23 predisposition of ethnic minority individuals to higher CRP levels; median value of CRP in Blacks was
24
25 3.0 mg/L vs. 2.3 mg/L in Whites in one study of healthy individuals.²⁰ The absence of differences
26
27 between the ethnic minority and White groups for the two other markers of inflammation studied
28
29 (procalcitonin and ferritin) supports the hypothesis of a specific increased susceptibility to cytokine-
30
31 mediated adverse incidents in the ethnic minority population.²¹ Pre-existing comorbidities including
32
33 CVD, COPD, diabetes and hypertension are established risk factors for severe disease in the
34
35 pandemic²². Many of these comorbidities are known to promote a pro-inflammatory state, but of
36
37 these, only diabetes mellitus prevalence was higher in the ethnic minority group than in the White
38
39 group in our study. It is hypothesised that existing diabetes mellitus may accentuate the inflammation
40
41 associated with viral infection²³, therefore the increased prevalence of diabetes in the ethnic minority
42
43 group in our study may be a contributory factor to the increased CRP concentrations at presentation
44
45 in this group. Like CRP, diabetes has also been shown to be significantly associated with in-hospital
46
47 mortality²⁴.
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 Liver injury in COVID-19 is also associated with disease severity²⁵. It is characterised by increased
4 transminases, with increase of AST dominating over ALT⁵. In our study, AST tended to be higher in the
5 ethnic minority population than in the White population whilst ALT did not show any difference
6 between the two groups. However, the increases of transminases seen in both groups were generally
7 mild and unlikely to represent significant liver injury. Although GGT tended to be higher in the ethnic
8 minority population, this difference is likely related to known ethnic differences in this marker.²⁶ We
9 have previously reported higher, troponin T and NT-ProBNP in the White population than in ethnic
10 minorities¹², although the latter association did not reach statistical significance. Likely contributory
11 factors to these findings are that the White population were older with a higher prevalence of CVD.
12 Elevated troponin has been associated with worse outcomes in COVID-19²⁷.

13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29 In our cohort, although non-White ethnicity and male gender were predictive of ITU admission, no
30 statistically significant differences in biochemistry at presentation were noted for those who were
31 subsequently admitted to ITU compared to the White population, including for CRP. This may suggest
32 that there is no difference in the pathological mechanisms underlying severe COVID-19 which are
33 reflected by routine biochemistry, but that ethnic minority subjects are at an increased risk of
34 developing them.¹⁹ Of course, the association between ethnic minority and severe COVID-19 disease
35 is likely to be complex and incorporate multiple demographic and socioeconomic factors not already
36 captured in this study. In one study, Raisi-Estabragh *et al.*²⁸ suggested that both the sex and ethnic
37 patterns of COVID-19 are not adequately explained by variation in cardio-metabolic factors, 25(OH)-
38 vitamin D concentrations or socio-economic factors; clearly there is a need for more research required
39 to define the mechanism of increased ethnic minority risk.

40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57 This study has some limitations. Although the facts that 86% of total test results were obtained on
58 admission day and 97% within three days of admission would argue against the possibility of
59
60

1
2
3 differences in care between ethnic groups whilst in hospital, we cannot exclude the possibility that
4 biochemical differences are not contributed to by variance in the disease stage at which different
5 ethnic groups accessed hospital care. In the patients who were admitted to ITU during their hospital
6 stay, the number of results for some tests may be too low to identify true differences between the
7 two groups. The outcomes of ITU patients (requirement for mechanical ventilation/continuous
8 positive airway pressure support and morbidity/mortality) were also not available to allow
9 interrogation of the biochemistry with respect to endpoint. In addition, the majority of the ethnic
10 minority group were of Black ethnicity, (149/211 of all ethnic minority cases, all ethnic minority
11 individuals in ITU), so caution is advised in making generalisations across all non-White ethnic groups.
12 Finally, for some tests in the study, such as procalcitonin, only a small number of tests were performed
13 overall.
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30

31 In conclusion, the major finding of this study is that ethnic minority patients have higher CRP
32 concentrations at presentation, indicating a more severe acute inflammation. This may augment
33 existing co-morbidities characterised by chronic inflammation which may be more prevalent in the
34 ethnic minority population such as diabetes, pointing towards an increased tendency for severe
35 inflammation in this group.
36
37
38
39
40
41
42
43
44
45

46 Key messages:

- 47
- 48
- 49 • In the U.K, it has been shown that ethnic minorities have poorer outcomes in COVID-19
50 relative to those of the White population, including an increased risk of death
- 51
- 52
- 53 • In this study, we show that there are significant differences between ethnic minority and
54 White populations in routine clinical biochemistry parameters at presentation to hospital
55 with COVID-19
- 56
- 57
- 58 • Amongst the markers identified, C-reactive protein was significantly higher in the ethnic
59 minority group, pointing towards an increased tendency for severe inflammation in this
60 group, which may contribute towards the poorer outcomes in this group reported previously

References

1. Zhu N, Zhang D, Wang W, et al; China Novel Coronavirus Investigating and Research Team. A novel coronavirus from patients with pneumonia in China, 2019. *N Engl J Med* 2020; 382: 727-733.
2. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020; 395: 497-506.
3. World Health Organization. Weekly epidemiological update - 12 January 2021. Accessed January 17, 2021. <https://www.who.int/publications/m/item/weekly-epidemiological-update---12-january-2021>
4. Henry BM, de Oliveira MHS, Benoit S, et al. Hematologic, biochemical and immune biomarker abnormalities associated with severe illness and mortality in coronavirus disease 2019 (COVID-19): a meta-analysis. *Clin Chem Lab Med* 2020; 58: 1021-1028.
5. Bloom PP, Meyerowitz EA, Reinus Z, et al. Liver Biochemistries in Hospitalized Patients With COVID-19. *Hepatology* 2020; May 16. doi: 10.1002/hep.31326. Online ahead of print.
6. Lippi G, South AM and Henry BM. Electrolyte imbalances in patients with severe coronavirus disease 2019 (COVID-19). *Ann Clin Biochem* 2020; 57: 262–265.
7. Bonetti G, Manelli F, Patroni A, et al. Laboratory predictors of death from coronavirus disease 2019 (COVID-19) in the area of Valcamonica, Italy. *Clin Chem Lab Med* 2020; 58: 1100-1105.
8. Galloway JB, Norton S, Barker RD, et al. A clinical risk score to identify patients with COVID-19 at high risk of critical care admission or death: An observational cohort study. *J Infect* 2020; 81: 282-288.
9. Lei F, Ye-Mao Liu Y-M, Feng Zhou F et al. Longitudinal Association Between Markers of Liver Injury and Mortality in COVID-19 in China. *Hepatology* 2020; doi: 10.1002/hep.31301. Online ahead of print.

10. Du RH, Liang LR, Yang CQ, et al. Predictors of mortality for patients with COVID-19 pneumonia caused by SARS-CoV-2: a prospective cohort study. *Eur Respir J* 2020; 55: 2000524.
11. Public Health England. Disparities in the risk and outcomes of COVID-19. PHE, 2020.
12. Ranasinghe RNK, Taylor DR, Mazaheri T, et al. Cardiac markers in Black, Asian and minority ethnic (BAME) patients with COVID-19. *J Clin Pathol* 2020; Dec 10 jclinpath-2020-207188. doi: 10.1136/jclinpath-2020-207188. Online ahead of print.
13. R Core Team. *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing 2019.
14. Potempa LA, Rajab IM, Hart PC, et al. Insights Into the Use of C-Reactive Protein as a Diagnostic Index of Disease Severity in COVID-19 Infections. *Am J Trop Med Hyg* 2020; doi: 10.4269/ajtmh.20-0473. Online ahead of print.
15. Zhang JJ, Dong X, Cao YY, et al. Clinical characteristics of 140 patients infected with SARS-CoV-2 in Wuhan, China. *Allergy* 2020; 75: 1730-1741.
16. Guan WJ, Ni ZY, Hu Y, et al. Clinical characteristics of coronavirus disease 2019 in China. *N Engl J Med* 2020; 382: 1708–1720.
17. He F, Deng Y and Li W. Coronavirus Disease 2019: What We Know? *J Med Virol* 2020; 92: 719-725.
18. Moore JB and June CH. Cytokine Release Syndrome in Severe COVID-19. *Science* 2020; 368: 473-474.
19. Aziz M, Fatima R and Assaly R. Elevated interleukin-6 and Severe COVID-19: A Meta-Analysis. *J Med Virol* 2020; 10.1002/jmv.25948. doi: 10.1002/jmv.25948. Online ahead of print.
20. Khera A, McGuire DK, Murphy SA, et al. Race and Gender Differences in C-reactive Protein Levels. *J Am Coll Cardiol* 2005;46(3):464-9. doi: 10.1016/j.jacc.2005.04.051.
21. Vepa A, Bae JP, Ahmed F, et al. COVID-19 and ethnicity: A novel pathophysiological role for inflammation. *Diabetes Metab Syndr* 2020; 14: 1043–1051.

- 1
2
3 22. Del Sole F, Farcomeni A, Loffredo L, et al. Features of severe COVID-19: A systematic review
4 and meta-analysis. *Eur J Clin Invest* 2020; 50: e13378.
5
6
7
8 23. Lim S, Bae JH, Kwon H-S, et al. COVID-19 and diabetes mellitus: from pathophysiology to
9 clinical management. *Nat Rev Endocrinol* 2021; 17: 11-30.
10
11
12 24. Silverio A, Di Maio M, Citro R, et al. Cardiovascular risk factors and mortality in hospitalized
13 patients with COVID-19: systematic review and meta-analysis of 45 studies and 18,300
14 patients. *BMC Cardiovasc Disord* 2021; 21(1):23.
15
16
17
18 25. Anirvan P, Bharali P, Gogoi M, et al. Liver injury in COVID-19: The hepatic aspect of the
19 respiratory syndrome - what we know so far. *World J Hepatol* 2020; 12(12):1182-1197.
20
21
22
23 26. Stranges S, Freudenheim JL, Muti P, et al. Greater hepatic vulnerability after alcohol intake in
24 African Americans compared with Caucasians: a population-based study. *J Natl Med Assoc*
25 2004; 96: 1185-92.
26
27
28
29 27. Tersalvi G, Vicenzi M, Calabretta D, et al. Elevated troponin in patients with Coronavirus
30 Disease 2019 (COVID-19): possible mechanisms. *J Card Fail* 2020; 26: 470-5.
31
32
33
34 28. Raisi-Estabragh Z, McCracken C, Bethell MS, et al. Greater risk of severe COVID-19 in Black,
35 Asian and Minority Ethnic populations is not explained by cardiometabolic, socioeconomic or
36 behavioural factors, or by 25(OH)-vitamin D status: study of 1326 cases from the UK
37 Biobank. *J Public Health (Oxf)* 2020; fdaa095. doi: 10.1093/pubmed/fdaa095. Online ahead
38 of print. PMID: 32556213
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 **Tables**
4
5
6

7 **Table 1.** Demographic data in BAME and White groups. Age data presented as median (interquartile range).
8

	Ethnic minority	White
All patient data		
Number	211	100
Age (years)	65 (55-78)	75 (66-84)
Male/female (%)	64/36	52/48
Pre-existing comorbidities:		
Diabetes (%)	54	31
CVD (%)	25	50
CKD (%)	31	28
Hypertension (%)	49	49
COPD/Asthma (%)	30	27
Patients subsequently admitted to intensive care		
Number (% total)	39 (18)	13 (13)
Age (years)	59 (52-65)	65 (57-71)
Male/female (%)	77/23	77/23

9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46

Table 2. Biochemistry at presentation (all patient data). P values <0.05 were taken as statistically significant. N/A = Chi-squared test was not applicable to data where only abnormal results were recorded.

Test	Number of results		Test result (Median (interquartile range))			Abnormal results (%)		
	Ethnic minority	White	Ethnic minority	White	P-value	Ethnic minority	White	P-value
Albumin (g/L)	211	100	37.0 (34.0 – 39.5)	37.5 (34.0 – 41.0)	0.247	30	33	0.668
ALP (IU/L)	209	100	74.0 (57.0 – 98.0)	84.0 (63.0 – 102.5)	0.039	12	17	0.302
ALT (IU/L)	104	33	41.0 (27.8 – 58.0)	32.0 (17.0 – 70.0)	0.078	29	27	1.000
AST (IU/L)	206	99	52.0 (35.2 – 76.0)	40.0 (28.0 – 71.5)	0.008	75	56	0.001
Total bilirubin (µmol/L)	211	100	9.0 (6.5 – 13.0)	9.0 (6.0 – 14.0)	0.617	6	6	1.000
Adjusted calcium (mmol/L)	183	81	2.3 (2.2 – 2.4)	2.3 (2.2 – 2.4)	0.537	19	22	0.604
CK (IU/L)	63	31	270.0 (111.5 – 1139.0)	163.0 (63.5 – 725.0)	0.447	67	52	0.236
Creatinine (µmol/L)	211	100	106.0 (80.0 – 159.5)	100.0 (76.8 – 138.5)	0.208	40	35	0.442
CRP (mg/L)	209	98	111.2 (66.5 – 181.1)	48.1 (22.1 – 112.9)	<0.001	97	93	0.153
eGFR (ml/min/1.73m ²)	210	99	58.0 (34.0 – 78.8)	55.0 (38.5 – 86.0)	0.447	56	65	0.222
Ferritin (µg/L)	91	35	916.0 (579.0 – 1815.5)	1,046.0 (386.5 – 1859.0)	0.208	90	86	0.699
GGT (IU/L)	209	100	57.0 (32.0 – 99.0)	41.0 (23.0 – 83.2)	0.030	51	40	1.000
LDH (IU/L)	48	12	467.0 (351.2 – 631.2)	503.5 (457.5 – 634.0)	0.432	92	92	1.000
Magnesium (mmol/L)	183	81	0.9 (0.8 – 1.0)	0.8 (0.8 – 1.0)	0.028	28	32	0.582
Sodium (mmol/L)	211	100	137.0 (134.0 – 140.0)	137.0 (134.0 – 140.30)	0.521	35	35	1.000
NT-proBNP (ng/L)	34	13	221.5 (90.8 – 1077.5)	427.0 (141.0 – 1678.0)	0.274	66	77	0.448
Procalcitonin (µg/L)	28	8	1.2 (0.4 – 11.4)	0.7 (0.3 – 2.5)	0.594	100	100	N/A
Phosphate (mmol/L)	181	81	1.0 (0.8 – 1.2)	1.0 (0.9 – 1.2)	0.145	36	30	0.395
Potassium (mmol/L)	208	98	4.2 (3.8 – 4.7)	4.3 (3.9 – 4.8)	0.215	24	19	0.502
Troponin T (ng/L)	122	45	19.0 (8.2 – 52.2)	35.0 (16.0 – 74.0)	0.023	62	82	0.024

Total protein (g/L)	210	100	71.0 (67.0 – 74.0)	68.5 (64.0 – 71.2)	<0.001	12	13	0.929
Urea (mmol/L)	211	99	7.0 (4.6 – 14.3)	8.7 (5.9 – 13.9)	0.059	64	67	0.738

Confidential: For Review Only

Table 3. Biochemistry at hospital presentation in patients subsequently admitted to intensive care. P values <0.05 were taken as statistically significant. N/A = Chi-squared test was not applicable to data where only abnormal results were recorded.

Test	Number of results		Test result (Median (interquartile range))		P-value	Abnormal results (%)		
	Ethnic minority	White	Ethnic minority	White		Ethnic minority	White	P-value
Albumin (g/L)	39	13	38.0 (34.0 – 39.0)	35.0 (32.0 – 37.0)	0.167	28	38	0.729
ALP (IU/L)	39	13	81.0 (57.0 – 107.0)	83.0 (74.0 – 132.0)	0.459	21	31	0.704
ALT (IU/L)	36	10	44.5 (30.0 – 68.5)	76.0 (41.0 – 154.5)	0.432	36	50	0.667
AST (IU/L)	39	13	68.0 (48.5 – 136.0)	65.0 (44.0 – 183.0)	0.966	87	77	0.657
Total bilirubin (µmol/L)	39	13	8.0 (7.0 – 13.0)	11.0 (8.0 – 18.0)	0.203	5	23	0.175
Adjusted calcium (mmol/L)	39	13	2.3 (2.2 – 2.4)	2.3 (2.2 – 2.3)	0.958	23	31	0.853
CK (IU/L)	29	10	632.0 (161.0 – 1687.0)	142.5 (83.2 – 1174.2)	0.311	79	40	0.054
Creatinine (µmol/L)	39	13	110.0 (81.0 – 211.0)	130.0 (110.0 – 182.0)	0.310	41	62	0.335
CRP (mg/L)	39	13	157.0 (111.4 – 257.1)	168.4 (40.9 – 251.8)	0.616	100	100	N/A
eGFR (ml/min/1.73m ²)	39	13	59.0 (28.0 – 83.0)	39.0 (32.0 – 58.0)	0.295	51	85	0.073
Ferritin (µg/L)	34	10	1,054.5 (718.0 – 1981.0)	1,822.0 (1018.2 – 2507.5)	0.245	97	90	0.937
GGT (IU/L)	39	13	91.0 (46.0 – 136.0)	66.0 (48.0 – 100.0)	0.512	72	69	1.000
LDH (IU/L)	28	6	522.5 (399.8 – 687.0)	500.5 (402.2 – 672.2)	0.878	96	100	1.000

Magnesium (mmol/L)	39	13	0.9 (0.9 – 1.0)	1.0 (0.8 – 1.1)	0.657	33	46	0.618
Sodium (mmol/L)	39	13	136.0 (134.0 – 139.0)	137.0 (136.0 – 138.0)	0.367	38	23	0.501
NT-proBNP (ng/L)	23	5	400.0 (164.5 – 1867.0)	427.0 (126.0 – 3916.0)	0.676	74	80	1.000
Procalcitonin (µg/L)	22	8	1.6 (0.5 – 11.8)	0.7 (0.3 – 2.5)	0.241	100	100	N/A
Phosphate (mmol/L)	39	13	1.1 (0.8 – 1.5)	1.2 (1.0 – 1.7)	0.336	46	54	0.873
Potassium (mmol/L)	39	13	4.4 (4.0 – 4.9)	4.7 (3.8 – 5.4)	0.695	28	38	0.729
Troponin T (ng/L)	35	12	24.0 (14.5 – 54.0)	32.0 (16.0 – 90.2)	0.472	74	92	0.389
Total protein (g/L)	39	13	72.0 (67.0 – 73.0)	66.0 (60.0 – 71.0)	0.044	8	15	0.786
Urea (mmol/L)	39	13	8.6 (5.0 – 15.2)	17.6 (9.9 – 19.1)	0.076	59	85	0.178