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A novel *RLIM/RNF12* variant disrupts protein stability and function to cause severe Tonne–Kalscheuer syndrome

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Tonne–Kalscheuer syndrome (TOKAS) is an X-linked intellectual disability syndrome associated with variable clinical features including craniofacial abnormalities, hypogenitalism and diaphragmatic hernia. TOKAS is caused exclusively by variants in the gene encoding the E3 ubiquitin ligase gene *RLIM*, also known as *RNF12*. Here we report identification of a novel *RLIM* missense variant, c.1262A>G p.(Tyr421Cys) adjacent to the regulatory basic region, which causes a severe form of TOKAS resulting in perinatal lethality by diaphragmatic hernia. Inheritance and X-chromosome inactivation patterns implicate *RLIM* p.(Tyr421Cys) as the likely pathogenic variant in the affected individual and within the kindred. We show that the *RLIM* p.(Tyr421Cys) variant disrupts both expression and function of the protein in an embryonic stem cell model. *RLIM* p.(Tyr421Cys) is correctly localised to the nucleus, but is readily degraded by the proteasome. The *RLIM* p.(Tyr421Cys) variant also displays significantly impaired E3 ubiquitin ligase activity, which interferes with *RLIM* function in *Xist* long-non-coding RNA induction that initiates imprinted X-chromosome inactivation. Our data uncover a highly disruptive missense variant in *RLIM* that causes a severe form of TOKAS, thereby expanding our understanding of the molecular and phenotypic spectrum of disease severity.

Tonne–Kalscheuer syndrome (TOKAS; MIM #300978) is a recently described X-linked recessive multiple congenital anomaly disorder^{1–3}. Male patients display global developmental delay apparent from early infancy, impaired intellectual development, speech delay, behavioural abnormalities, and abnormal gait. Affected individuals also display dysmorphic facial features, anomalies of the hands, feet and nails, abnormal pulmonary development, and urogenital abnormalities with hypogenitalism. In a subset of severely affected patients, development of congenital diaphragmatic hernia may result in perinatal or premature death. Carrier females may display minor clinical manifestations including very mild skeletal or hormonal abnormalities¹.

In all reported cases, TOKAS is caused by variants in the RING finger type E3 ubiquitin ligase *RLIM/RNF12*, which ubiquitylates transcription factor substrates to control key developmental processes including imprinted X-chromosome inactivation⁴, stem cell maintenance and differentiation^{5,6}. In most cases, patient-derived *RLIM* TOKAS variants lead to impaired E3 ubiquitin ligase activity^{1,6} without major impact on other aspects of *RLIM* expression and function, including protein stability, phosphorylation, subcellular localisation and protein:protein interactions⁶. Specific disruption of *RLIM* activity by TOKAS variants result in deregulated stem cell differentiation to neurons⁶, providing insight into the cellular processes that may underpin TOKAS etiology.

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Thus far, 8 variants that cause TOKAS in distinct unrelated families have been reported^{1–3}. *RLIM* variants have been reported in 4 cases out of a cohort of 405 cases of unresolved syndromic X-linked intellectual disability with no known genetic or environmental basis². This data suggests that a significant proportion of unresolved X-linked intellectual disability cases may be caused by *RLIM* variants, and that other *RLIM* TOKAS variants await identification.

Here, we report identification of a novel *RLIM* missense variant, p.(Tyr421Cys), adjacent to the regulatory basic region, which causes a severe form of TOKAS leading to perinatal lethality by diaphragmatic hernia. Inheritance and X-chromosome inactivation patterns clearly implicate *RLIM* p.(Tyr421Cys) as the causative variant in the affected individual and kindred. We show that the *RLIM* p.(Tyr421Cys) variant severely disrupts protein expression and function, and is readily degraded by the proteasome. *RLIM* p.(Tyr421Cys) also displays significantly impaired E3 ubiquitin ligase activity. Together, defects in *RLIM* p.(Tyr421Cys) protein expression and activity profoundly interfere with *RLIM* function in *Xist* long-non-coding RNA induction, a key step in initiating imprinted X-chromosome inactivation. Our data uncover a highly disruptive missense variant in *RLIM* that causes a severe form TOKAS, thereby expanding our understanding of the molecular and phenotypic spectrum of disease severity.

Results

Clinical information for a patient with an undiagnosed developmental disorder. The male proband was the first child of unrelated parents. The first trimester nuchal translucency/morphology scan and second trimester fetal morphology scan were reported as normal. Polyhydramnios was detected at 36 weeks' gestation and intrauterine growth restriction (IUGR) was reported at 37 weeks. Labour was induced at 39 weeks and delivery was via emergency Caesarean section for subsequent fetal distress. The baby became cyanosed immediately after birth, was unable to be resuscitated and died at 30 min of age.

On examination the baby was symmetrically small for gestational age with a birth weight of 2285 g (<3rd percentile; Z score – 2.57), length 48.5 cm (~5th percentile; Z score – 1.67) and head circumference 32.5 cm (<3rd percentile; Z score – 2.16). Craniofacial anomalies were identified, including hypertelorism, broad nasal bridge with flat nasal tip, and a high arched palate. He had distal limb hypoplasia, with brachytelephalangy, soft tissue syndactyly between the 2nd and 3rd toes bilaterally and absent or hypoplastic nails on 2nd–3rd fingers and 2nd–4th toes. There were no ocular abnormalities.

Postmortem examination revealed congenital diaphragmatic hernia with aplasia of the posterolateral left hemi-diaphragm, displacement of the mediastinum to the right, herniation of the small and large intestine, stomach, spleen, left lobe of liver, and pancreas into the left hemithorax, absent middle lobe right lung and severe bilateral pulmonary hypoplasia. There was an accessory spleen. The cardiovascular system showed aberrant aortic arch branching with the left subclavian, left common carotid and right common carotid arteries arising from a single brachiocephalic trunk. Urogenital abnormalities included cryptorchidism with pelvic testes, but kidneys were normal. A cavum septum pellucidum was present. The clinical features were considered to be most consistent with Fryns syndrome (MIM #229850). Pallister-Killian syndrome (tetrasomy 12p; MIM #601803) was considered, but fluorescence in-situ hybridisation for chromosome 12p on fetal lung imprints, standard karyotype on cultured fibroblasts and chromosome microarray on DNA extracted from stored fetal tissue (Agilent Sureprint G3 ISCA Targeted Microarray 8 × 60 K) were normal.

Identification of a novel missense *RLIM* variant and diagnosis of Tonne–Kalscheuer syndrome (TOKAS). In order to investigate the genetic basis of this case, we performed whole exome sequencing on genomic DNA from the proband and his parents, and identified a maternally inherited hemizygous variant in the X-linked *RLIM* gene (NM_016120.3) c.1262A>G, p.(Tyr421Cys) (Fig. 1a). *RLIM* encodes the *RLIM*/*RNF12* E3 ubiquitin ligase that is mutated in the recessive X-linked disorder Tonne–Kalscheuer syndrome (TOKAS)^{1–3}, which is characterised by clinical features that significantly overlap with Fryns syndrome. There were no variants in *PIGN*, in which biallelic loss of function mutations have been reported to cause a Fryns-like syndrome. We hypothesised that the individual was affected by a severe form of TOKAS that is caused by this novel *RLIM* variant. Consistent with this notion, the p.(Tyr421Cys) variant is located adjacent to the basic region of the *RLIM* protein, a key regulatory domain that is mutated in several TOKAS kindreds^{1–3}. *RLIM* p.(Tyr421Cys) has not been observed in affected individuals in the literature and was not reported in the ClinVar archive of genomic variation in human health (<https://www.ncbi.nlm.nih.gov/clinvar>) or the gnomAD population database (<https://gnomad.broadinstitute.org/>). As a result, the variant was submitted to ClinVar under the accession number SCV001435291.

Sanger sequencing confirmed the presence of the *RLIM* c.1262A>G p.(Tyr421Cys) variant in the proband, whilst the mother and maternal grandmother were confirmed as carriers (Fig. 1b). The variant was absent in the mother's unaffected brother (Fig. 1b), consistent with an X-linked inheritance pattern. Furthermore, X-chromosome inactivation analysis showed the mother had a highly skewed X-chromosome inactivation pattern (95%; Supplementary Figure 1), which is characteristic of female carriers of *RLIM* TOKAS variants^{1,3}. The clinical features and identification of this novel *RLIM* variant inherited by the affected individual led to a revised diagnosis of a severe form of TOKAS.

***RLIM* p.(Tyr421Cys) is poorly expressed and readily degraded by the proteasome.** The diagnosis of severe TOKAS prompts the hypothesis that the p.(Tyr421Cys) variant impacts on *RLIM* protein expression and/or function. This variant is predicted to be damaging to the protein by multiple in silico programs (SIFT (v6.20)⁷: deleterious (score: 0); PolyPhen2⁸: probably damaging (score: 0.994); MutationTaster (v2013)⁹: disease causing (p-value: 1)), and is classified as a variant of uncertain significance (VUS) by ACMG guidelines¹⁰.

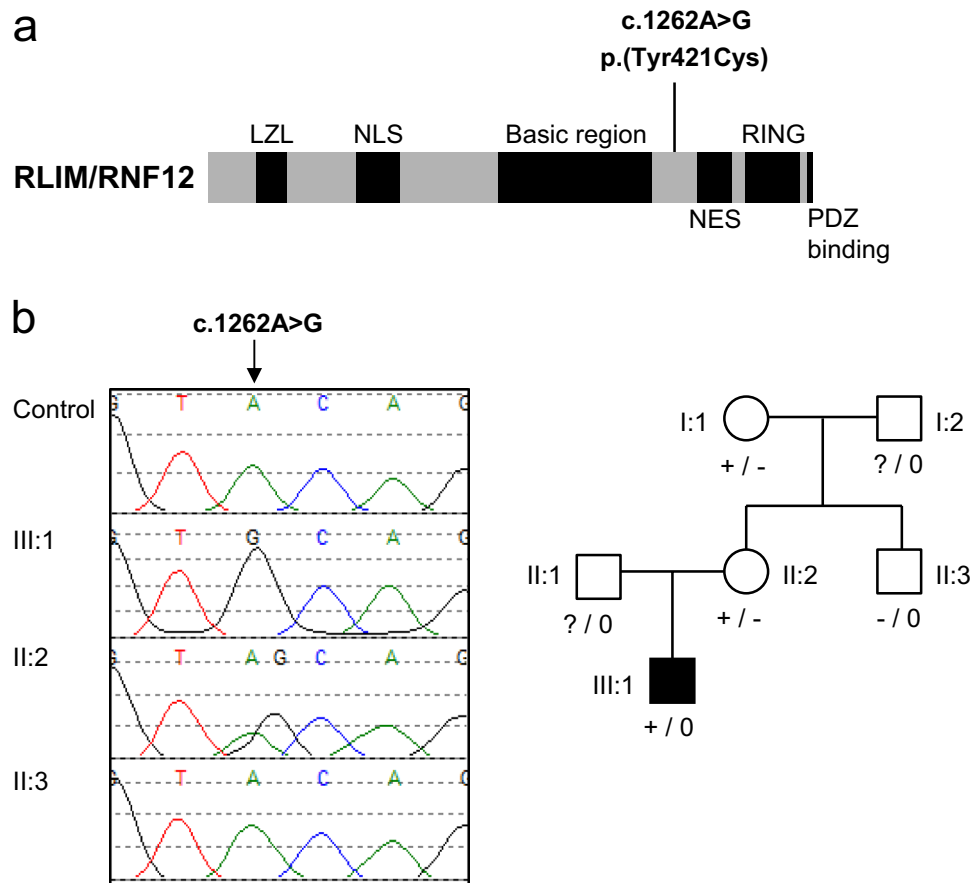


Figure 1. Identification of *RLIM* p.(Tyr421Cys), a novel variant in a severe form of Tonne–Kalscheuer Syndrome. (a) Schematic diagram of the *RLIM* protein domain structure, with the position of the c.1262A>G p.(Tyr421Cys) variant identified by exome sequencing indicated. (b) (Left panel) Genomic DNA sequencing electropherogram identifying the *RLIM* c.1262A>G p.(Tyr421Cys) variant within the kindred. Figure was generated using SoftGenetics Mutation Surveyor v5.01 (<https://softgenetics.com/mutationSurveyor.php>). (Right panel) Family pedigree showing inheritance of the *RLIM* c.1262A>G p.(Tyr421Cys) variant.

Therefore, we exploited *RLIM*-deficient male mouse Embryonic Stem Cells (mESCs) to investigate expression and function of *RLIM* p.(Tyr421Cys) in a cellular model. In comparison to wild-type human *RLIM*, the p.(Tyr421Cys) variant is poorly expressed (Fig. 2a). Quantitative analysis indicates that *RLIM* p.(Tyr421Cys) expression is reduced to $28.7 \pm 11.5\%$ of wild-type (Fig. 2b). However, the mRNA is expressed at similar levels, (Fig. 2c) and the protein is correctly localised to the nucleus (Fig. 2d), suggesting that the *RLIM* p.(Tyr421Cys) variant specifically interferes with translation and/or stability. We explored whether *RLIM* p.(Tyr421Cys) is more readily turned over by the proteasome. Indeed, treatment of *RLIM* p.(Tyr421Cys) expressing mESCs with the proteasome inhibitor MG132 expression rescues *RLIM* p.(Tyr421Cys) expression to levels approaching that of wild-type *RLIM* (Fig. 2e), indicating that *RLIM* p.(Tyr421Cys) more readily undergoes proteasomal degradation. Taken together, these data indicate that the *RLIM* p.(Tyr421Cys) variant destabilises the protein.

***RLIM* p.(Tyr421Cys) variant interferes with E3 ubiquitin ligase activity.** As other *RLIM* TOKAS variants located within the proximal basic region (Fig. 1a) have been shown to disrupt E3 ubiquitin ligase activity^{1,6}, we next sought to determine the impact of p.(Tyr421Cys) on *RLIM* catalytic activity. We expressed and purified *RLIM* wild-type and p.(Tyr421Cys) from *E. coli*, and examined the ability of these variants to transfer ubiquitin from a cognate E2 (UBE2D1) to the prototypic substrate REX1, which is a transcription factor that is ubiquitylated by *RLIM* to initiate X-chromosome inactivation¹¹. Wild-type *RLIM* efficiently promotes REX1 ubiquitylation, as evidenced by appearance of ubiquitylated species of increasing molecular weight upon addition of substrate (Fig. 3a). However, *RLIM* p.(Tyr421Cys) significantly impairs REX1 ubiquitylation activity (Fig. 3a), suggesting that the p.(Tyr421Cys) variant also has a negative impact on *RLIM* catalytic activity. Quantitative analysis indicates that *RLIM* p.(Tyr421Cys) activity is reduced to $32 \pm 3.7\%$ of wild-type (Fig. 3b). Therefore, similar to other previously reported *RLIM* TOKAS variants, p.(Tyr421Cys) displays impaired E3 ubiquitin ligase activity.

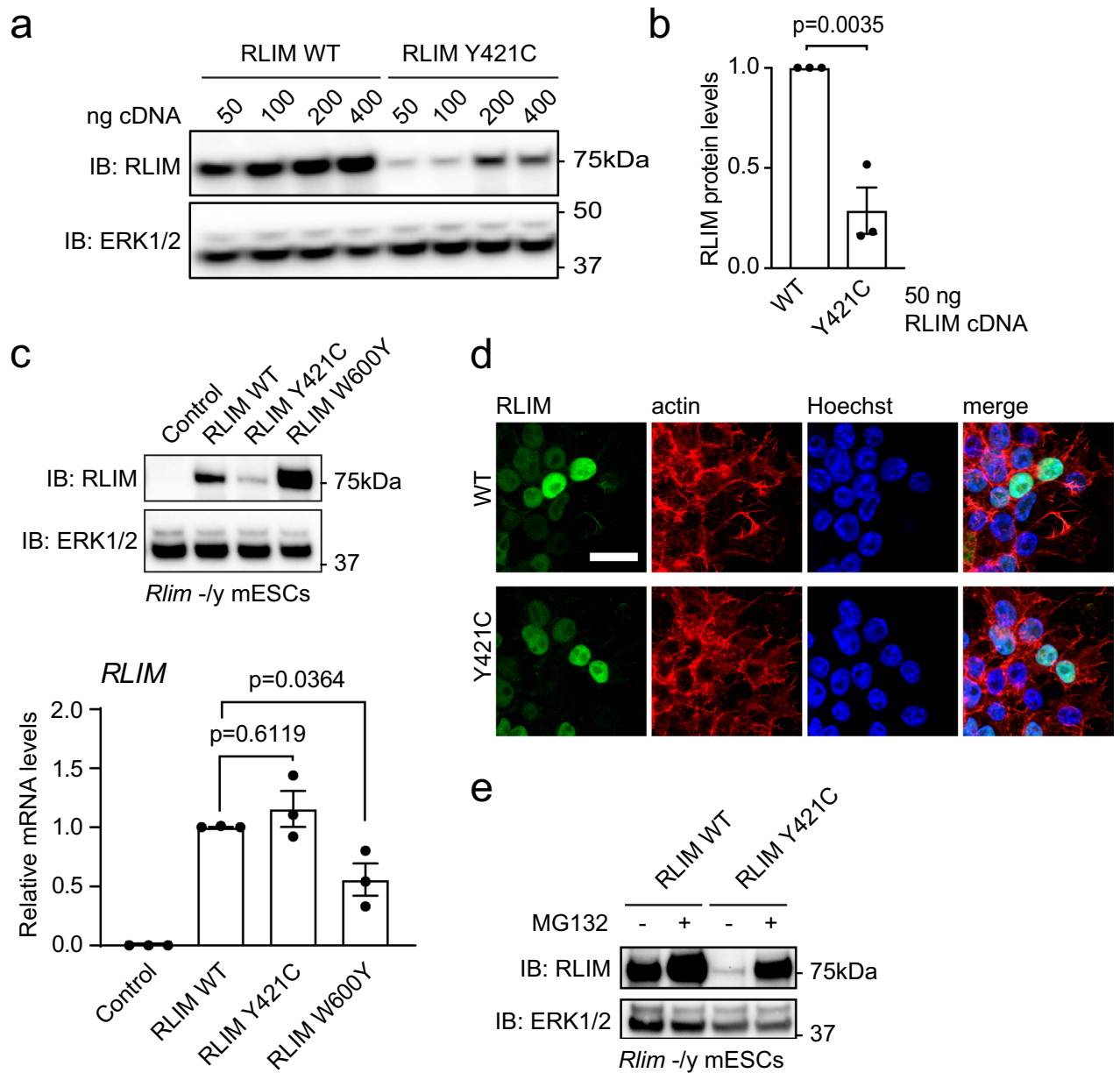


Figure 2. RLIM p.(Tyr421Cys) is poorly expressed and readily degraded by the proteasome. **(a)** Human RLIM wild-type (WT) or Y421C TOKAS variant were expressed at increasing amounts (50–400 ng plasmid DNA) in *Rlim*^{-/-} mESCs. RLIM expression was determined by immunoblotting and ERK1/2 expression analysed as a loading control. **(b)** Quantification of human RLIM wild-type (WT) or Y421C TOKAS variant protein expression from 50 ng plasmid cDNA in *Rlim*^{-/-} mESCs. RLIM expression was determined by immunoblotting. Normalised data are represented as mean \pm standard error of the mean (n = 3). Statistical significance was determined by student's t-test. Figure was generated using BioRad ImageLab v6.0.1 (<https://www.bio-rad.com/en-uk/product/image-lab-software?ID=KRE6P5E8Z>) and Graphpad Prism v8.0 (<https://www.graphpad.com/scientific-software/prism/>). **(c)** Human RLIM wild-type (WT), Y421C TOKAS variant or W600Y catalytically-inactive variant were expressed in *Rlim*^{-/-} mESCs. (Top panel) Human RLIM protein expression was determined by immunoblotting and ERK1/2 expression analysed as a loading control. (Bottom panel) Human *RLIM* mRNA expression was determined by qRT-PCR and normalised to *Gapdh* mRNA expression. Data are represented as mean \pm standard error of the mean (n = 3). Statistical significance was determined by one-way ANOVA. Figure was generated using BioRad ImageLab v6.0.1 (<https://www.bio-rad.com/en-uk/product/image-lab-software?ID=KRE6P5E8Z>) and Graphpad Prism v8.0 (<https://www.graphpad.com/scientific-software/prism/>). **(d)** Human RLIM wild-type (WT) and Y421C TOKAS variant were expressed in *Rlim*^{-/-} mESCs. RLIM localisation was determined by immunofluorescence. Scale Bar = 20 μ m. Microscopy images were acquired using Zeiss ZEN 2012 SP5 (black edition) software (<https://www.zeiss.com/microscopy/us/products/microscope-software/zen.html>) and processed using NIH Image J v1.48 (<https://imagej.net/>) and Adobe Photoshop CC v19.1.8 (<https://www.adobe.com/uk/products/photoshop.html>). **(e)** Human RLIM wild-type (WT) or Y421C TOKAS variant were expressed in *Rlim*^{-/-} mESCs treated with vehicle or 10 μ M MG132 for 4 h. RLIM expression was determined by immunoblotting and ERK1/2 expression analysed as a loading control.

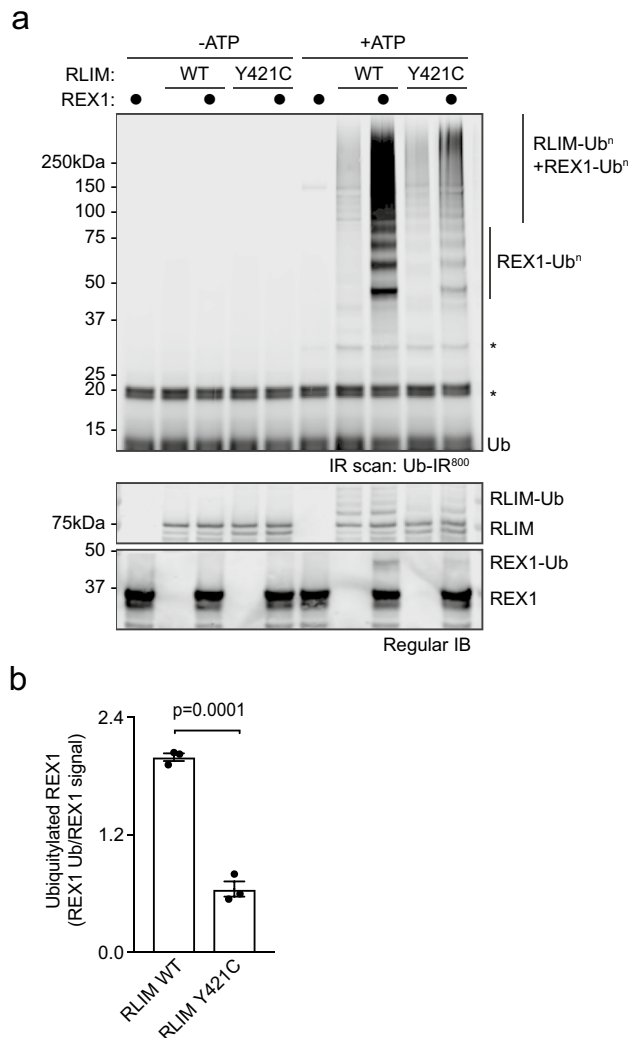


Figure 3. The p.(Tyr421Cys) variant impairs RLIM catalytic E3 ubiquitin ligase activity and substrate ubiquitylation. **(a)** Recombinant human RLIM wild-type (WT) or Y421C TOKAS variant were assayed for E3 ubiquitin ligase activity by incubating with UBE1 E1, UBE2D1 E2 conjugating enzyme in the presence or absence of ATP and recombinant REX1 substrate. REX1-specific substrate ubiquitylation (REX1-Ubⁿ) and RLIM, REX1 and/or free ubiquitin chains (RLIM-Ubⁿ+REX1-Ubⁿ) are indicated. RLIM and REX1 protein levels were determined by immunoblotting, and ubiquitylated RLIM and REX1 indicated. Infrared images were generated by LI-COR Biosciences Image Studio v5.2 software (<https://www.licor.com/bio/image-studio/>). **(b)** Quantification of E3 ubiquitin ligase activity of RLIM wild-type (WT) or Y421C TOKAS variant. Data are represented as mean \pm standard error of the mean (n = 3). Statistical significance was determined by student's t-test. Figure was generated using Graphpad Prism v8.0 (<https://www.graphpad.com/scientific-software/prism/>).

Functional disruption of the RLIM signalling pathway by the p.(Tyr421Cys) variant. Finally, we explored the functional impact of the RLIM p.(Tyr421Cys) variant in a cellular model of RLIM signalling. RLIM plays a key role in imprinted X-chromosome inactivation¹², which is initiated by RLIM-dependent transcriptional induction of the *Xist* long non-coding RNA (lncRNA)¹³. Thus, we used *Xist* expression as a readout for RLIM function. As expected, *Xist* expression in RLIM-expressing *Rlim*^{+/-} mESCs is very low (Fig. 4a). However, *Xist* expression is induced by expression of wild-type human RLIM, but not by a catalytically inactive variant (W600Y; Fig. 4a). When compared to wild-type RLIM, *Xist* induction is severely impaired upon expression of RLIM p.(Tyr421Cys) (Fig. 4a), demonstrating that the p.(Tyr421Cys) variant profoundly interferes with RLIM function. In order to control for expression, we analysed RLIM levels in *Rlim*^{+/-} mESCs, which do not express endogenous RLIM (Fig. 4b). We also show using *Rlim*^{-/-} mESCs that loss of RLIM expression and function does not significantly alter cell number accumulation under standard mESC culture conditions (Supplementary Figure 2), suggesting that reduced expression/function variants such as RLIM p.(Tyr421Cys) are unlikely to significantly impact on cell viability, proliferation and survival. Taken together, our data indicate that the RLIM p.(Tyr421Cys) variant disrupts RLIM stability and E3 ubiquitin ligase catalytic activity, which leads to significantly impaired RLIM function in vivo.

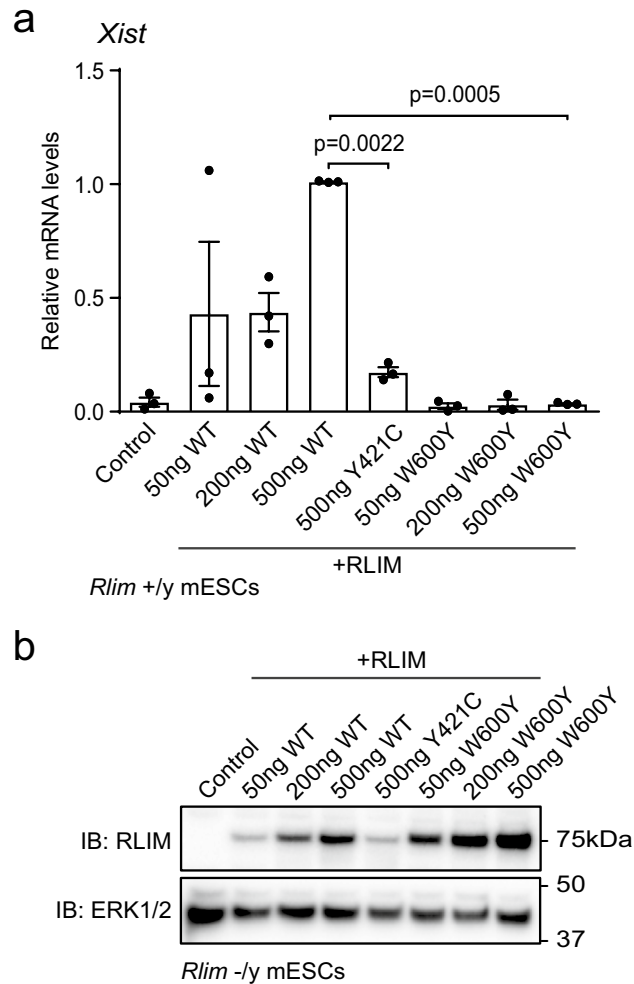


Figure 4. RLIM p.(Tyr421Cys) is functionally impaired in an assay for *Xist* lncRNA induction. **(a)** Human RLIM wild-type (WT), Y421C TOKAS variant or W600Y catalytically-inactive variant were expressed at increasing amounts (50–500 ng plasmid DNA) in *Rlim*^{+/y} mESCs. *Xist* lncRNA expression was determined by qRT-PCR and normalised to *Gapdh* mRNA expression. Data are represented as mean \pm standard error of the mean ($n = 3$). Statistical significance was determined by one-way ANOVA. Figure was generated using Graphpad Prism v8.0 (<https://www.graphpad.com/scientific-software/prism/>). **(b)** Human RLIM wild-type (WT) or W600Y catalytically-inactive variant were expressed at increasing amounts (50–500 ng plasmid DNA) in *Rlim*^{-/-} mESCs. The RLIM Y421C TOKAS variant was expressed only at the maximum amount of plasmid DNA (500 ng) to enable comparison with similar levels of RLIM WT or Y421C expressed at lower amounts of plasmid DNA. RLIM expression was determined by immunoblotting and ERK1/2 expression analysed as a loading control.

Discussion

Tonne–Kalscheuer syndrome (TOKAS) is a developmental disorder characterised by clinical features including intellectual disability, facial dysmorphism, velopharyngeal abnormalities and diaphragmatic hernia^{1–3}. In the most severe cases, diaphragmatic hernia causes death shortly after birth¹. TOKAS is caused by variants in the X-linked RLIM/RNF12 E3 ubiquitin ligase, which impair catalytic activity to varying extents^{1,6}. However, the extent to which phenotypic/disease severity correlates with genotypic severity (i.e. the extent of RLIM functional disruption) remains unclear.

Here, we provide a case report of a male patient who died shortly after birth with diaphragmatic hernia, facial dysmorphism and skeletal abnormalities. Cytogenetic studies showed no evidence of Pallister–Killian syndrome and an initial diagnosis of Fryns syndrome was thought most likely, although TOKAS was not considered as this baby was born several years before the association of diaphragmatic hernia with severe TOKAS was delineated¹. Research exome sequencing through the Care4Rare Canada Consortium uncovered a missense *RLIM* variant, p.(Tyr421Cys), which was heterozygous in the mother but not present in unaffected relatives, suggesting that the patient suffered a severe form of TOKAS. These findings raise the important question of whether other individuals diagnosed with Fryns syndrome and/or syndromic diaphragmatic hernia might actually represent incorrectly diagnosed TOKAS patients.

In this study, we explore the impact of the p.(Tyr421Cys) variant on RLIM protein expression and function. Strikingly, we find that RLIM p.(Tyr421Cys) is poorly expressed in an RLIM-deficient mouse embryonic stem cell model, and is prone to proteasomal degradation, although the protein is correctly localised in the nucleus. This is in contrast to other RLIM TOKAS variants, which show impaired catalytic activity but no major impact on stability^{1,6}. Interestingly, analysis of recombinant RLIM p.(Tyr421Cys) indicates that this variant also displays impaired E3 ubiquitin ligase catalytic activity. Although the RLIM p.(Tyr421Cys) variant lies outwith the core catalytic RING domain, this variant may disrupt a poorly understood catalytic function of the basic region^{1,6}.

Our data also support the notion that disrupted RLIM p.(Tyr421Cys) protein expression and catalytic activity contribute to impairment of RLIM function. In this regard, the RLIM p.(Tyr421Cys) variant significantly disrupts induction of the *Xist* lncRNA, which is a key initiating step of X-chromosome inactivation (XCI). However, XCI is unlikely to be a major feature of TOKAS pathogenesis, as this syndrome affects only male patients. Nevertheless, heterozygous female carriers of RLIM TOKAS variants display highly skewed XCI accompanied by relatively mild developmental anomalies¹, consistent with the presence of a deleterious X-linked variant. Indeed, we confirm that the mother, who is a carrier of the RLIM p.(Tyr421Cys) variant, displays a highly skewed XCI pattern.

A major question arising from this study concerns the developmental functions of RLIM that are disrupted to cause the major clinical features of TOKAS. Work from our group has previously identified a function for RLIM signalling in controlling expression of neuronal genes¹⁴. We hypothesise that RLIM TOKAS variants disrupt correct neuronal development leading to intellectual disability. However, the relevant gene expression programmes that are disrupted to cause the syndromic features of TOKAS, such as diaphragmatic hernia, are yet to be elucidated. In future, the mechanisms underpinning these syndromic anomalies will be uncovered by transcriptomic analyses of tissue-specific cellular and animal TOKAS models.

In summary, we introduce RLIM p.(Tyr421Cys) as the prototypic member of a new class of RLIM TOKAS variant that profoundly impacts on both protein stability and catalytic activity, which leads to severe TOKAS. This in turn expands our understanding of the molecular and phenotypic spectrum of TOKAS syndrome severity.

Methods

Genomic DNA sequencing and analysis. Trio exome sequencing of genomic DNA from proband and parents was performed through a collaboration with Care4Rare program (<http://care4rare.ca>). Target capture was performed with the Agilent CRE V1.0 and sequencing performed on the Illumina NextSeq 500 using 150 base-pair paired-end reads. Data analysis was done by standard methods and variants annotated using both Annovar and custom scripts to identify whether they affect protein coding sequence, and whether previously seen in dbSNP132, the 100 Genomes dataset (Nov 2011), the NHLBI GO exomes or in the approx. 1500 exomes previously sequenced at the center¹⁵. Variant reporting relevant to diagnosis was informed by reference to HGMD, dbSNP, online search engines e.g. PubMed and locus-specific databases. Variants seen in >20 of their controls, or with an allele frequency of >3% in 100 Genomes or NHBLI were removed. Confirmatory Sanger sequencing and familial testing was performed in the Sydney Genomic Diagnostic laboratory, Children's Hospital at Westmead.

Mouse embryonic stem cell (mESC) culture. *Rlim*^{+/-} and *Rlim*^{-/-} mESCs were described in⁶. Cells were cultured on 0.1% gelatin (w/v)-coated plates in DMEM containing 10% (v/v) fetal bovine serum (FBS), 5% (v/v) Knock-Out serum replacement, 20 ng/ml GST-tagged leukemia inhibitory factor (LIF), penicillin/streptomycin, 2 mM glutamine, 0.1 mM minimum essential media (MEM) Non-essential amino acids, 1 mM sodium pyruvate (all Thermo Fisher Scientific) and 0.1 mM β -mercaptoethanol (Sigma-Aldrich) in a controlled atmosphere at 5% CO₂ and 37 °C.

cDNA expression vectors and transfection. mESCs were transfected with Lipofectamine LTX (Thermo Fisher Scientific) according to manufacturer instructions. Plasmids used were pCAGGS human RLIM (DU53765), human RLIM Y421C (DU61099) and RLIM W600Y (DU53985). All cDNA clones were generated by MRC-PPU Reagents & Services; see <http://mrcppureagents.dundee.ac.uk> for detailed information and plasmid requests.

Immunoblotting. Cells were harvested in lysis buffer containing 20 mM Tris (pH 7.4), 150 mM NaCl, 1 mM EDTA, 1% Nonidet P-40 (NP-40) (v/v), 0.5% sodium deoxycholate (w/v), 10 mM β -glycerophosphate, 10 mM sodium pyrophosphate, 1 mM NaF, 2 mM Na₃VO₄, and Roche Complete Protease Inhibitor Cocktail Tablets. 10–30 μ g of cell lysate was loaded in SDS-PAGE gels and transferred to polyvinylidene fluoride (PVDF) membranes. Membranes were blocked with Tris buffered saline-tween 20 (TBS-T) 5% non-fat milk buffer (w/v).

Primary antibodies are anti-mouse RLIM amino acids 1–271 (S691D third bleed; see MRC-PPU Reagents & Services <http://mrcppureagents.dundee.ac.uk> for further information and requests), anti-ERK1/2 (Santa Cruz Biotechnology) and anti-REX1 (Abcam). Secondary antibodies are Sheep IgG-horseradish peroxidase (HRP), Mouse IgG-HRP (Cell Signaling Technology) and Rabbit IgG-HRP (Cell Signaling Technology). After secondary antibody incubation, membranes were subjected to chemiluminescence detection with Immobilon Western Chemiluminescent HRP substrate (Millipore) using a Gel-Doc XR + System (Bio-Rad) or to Infrared detection using a LI-COR Odyssey Clx system. Detected protein signals were quantified using Image J (NIH) or Image Studio (LI-COR Biosciences). All unprocessed immunoblots are provided in Supplementary Figure 3.

Immunofluorescence. For localisation studies, mESCs were plated in 0.1% gelatin (v/v) coated coverslips and fixed in 4% PFA in PBS for 20 min at room temperature (RT). Cells were permeabilised with a 0.5% Triton X-100 in PBS solution for 5 min at RT and then blocked with 1% Fish gelatin (w/v) in PBS solution for 30 min at

RT. RLIM primary antibody (Novus Biologicals) was diluted 1:200 in blocking solution and added to cells for 2 h at RT. Anti-mouse Alexa-488 (Thermo Fisher Scientific) was used as a secondary antibody at 1:500 in blocking solution for 1 h at RT. Actin Red 555 reagent (Thermo Fisher Scientific, one drop per ml of blocking solution) was added together with secondary antibody for actin staining. Hoechst was added at 1:10,000 dilution in PBS for 5 min at RT as nuclear marker. Coverslips were mounted in glass slides using Fluorsave reagent (Millipore). Digital images were acquired in a Zeiss 710 confocal microscope and analysed and processed using ImageJ (NIH), Photoshop CC and Illustrator CC (Adobe).

In vitro ubiquitylation assay. All recombinant proteins were produced in *E. coli* by MRC-PPU Reagents and Services, purified via standard protocols and are available via <http://mrcppureagents.dundee.ac.uk/>. For in vitro ubiquitylation reactions, RLIM (140 nM) was incubated with a 20 μ l ubiquitylation mix containing 0.1 μ M UBE1, 0.05 μ M UBE2D1 (UbcH5a) and 1.5 μ g of REX1 (MRC-PPU Reagents and Services), 2 μ M DyLightTM 800 Maleimide fluorescently labelled ubiquitin (Ub-IR⁸⁰⁰), 0.5 mM TCEP (pH 7.5), 5 mM ATP (both from Sigma Aldrich), 50 mM Tris-HCl (pH 7.5), 5 mM MgCl₂ for 30 min at 30 °C. Reactions were stopped with SDS sample buffer and boiled for 5 min at 95 °C. Samples were loaded in 4–12% Bis-Tris gradient gels (Thermo Fisher Scientific). Gels were then scanned using an Odyssey CLx Infrared Imaging System (LICOR Biosciences). After scanning proteins were transferred to PVDF membranes and analysed by immunoblotting. All unprocessed gels and immunoblots are provided in Supplementary Figure 3.

RNA extraction and quantitative RT-PCR. RNA was extracted using Omega total RNA extraction kit (column-based system) and obtained RNA converted to cDNA using iScript cDNA synthesis Kit (Bio-Rad). qPCR was performed using SsoFast EvaGreen Supermix (Bio-Rad) in a CFX384 real time PCR system (Bio-Rad). Relative mRNA levels were expressed using the $\Delta\Delta$ CT method and normalized to *Gapdh* expression. Data was analysed in Excel software and plotted using GraphPad Prism v7.0c software (GraphPad Software Inc.) (3). Primers used were: human *RLIM*: Forward (5'–3'): ATCATCAGGCTCATCAGGTGC, Reverse (3'–5'): AAG GAAGGGCAAAGAGCCAC; mouse *Xist*: Forward (5'–3'): GGATCCTGCTTGAACACTACTGC, Reverse (3'–5'): CAGGCAATCCTTCTTCTTGAG; mouse *Gapdh*: Forward (5'–3'): CTCGTCCCGTAGACAAAA, Reverse (3'–5'): TGAATTTGCCGTGAGTGG. For *Xist* induction analysis, *Rlim*^{+/-} mESCs were cultured for 72 h in LIF-deficient mESC media prior to RNA extraction and analysis.

mESC proliferation assay. The CellTiter Glo kit (Promega) was used to measure mESC cell number accumulation (colorimetric MTS assay). *Rlim*^{+/-} and *Rlim*^{-/-} mESCs were plated into flat bottom 96 well plate (1000 cells/well) in 100 μ l medium. Cells were cultured for the indicated time, whereupon a titration of resuspended mESCs was used to calculate cell number. Each condition was measured in triplicate. CellTiter Glo was prepared as per manufacturer's instructions. 100 μ l CellTiter Glo was pipetted into each well and mixed for 2 min on an orbital shaker at room temperature to induce cell lysis. Plates were incubated for 10 min at room temperature to stabilise the luminescent signal. The plate was read on a Perkin Elmer Envision at a Peak emission wavelength of 560 nm, and data analysed in Graphpad Prism.

Approval for human experiments. This patient was enrolled under the research study “Enhanced Care for Rare Genetic Diseases in Canada”. Research ethics approval for the study was obtained from the Children's Hospital of Eastern Ontario Research Ethics Board (Ethics study number CTO 1577), which includes approval for sequencing and functional studies. All experiments were conducted in accordance within these ethical guidelines and regulations. Informed consent was obtained from all participants and/or their legal guardians.

Data availability

The datasets generated and/or analysed during the current study are available from the corresponding authors on request.

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Author contributions

A.J.E. and K.D.K. provided exome analysis, M.J.W. provided clinical history, diagnosis and management of the patient and L.G.R. helped coordinate functional studies and prepared some figures. G.M.F. coordinated and F.B., C.E.-S. and A.S.-F. performed biochemical and cell-based experiments, analysed data and prepared figures. R.T. performed molecular cloning. G.M.F. and L.G.R. wrote the paper with input from all authors.

Competing interests

The authors declare no competing interests.

Additional information

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