

This is a repository copy of Use of habitat odour by host-seeking insects.

White Rose Research Online URL for this paper: http://eprints.whiterose.ac.uk/103032/

Version: Accepted Version

Article:

Webster, B. and Cardé, R.T. (2017) Use of habitat odour by host-seeking insects. Biological Reviews, 92 (2). pp. 1241-1249. ISSN 1464-7931

https://doi.org/10.1111/brv.12281

This is the peer reviewed version of the following article: Webster, B. and Cardé, R. T. (2016), Use of habitat odour by host-seeking insects. Biological Reviews, which has been published in final form at 10.1111/brv.12281. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving (http://olabout.wiley.com/WileyCDA/Section/id-828039.html)

Reuse

Unless indicated otherwise, fulltext items are protected by copyright with all rights reserved. The copyright exception in section 29 of the Copyright, Designs and Patents Act 1988 allows the making of a single copy solely for the purpose of non-commercial research or private study within the limits of fair dealing. The publisher or other rights-holder may allow further reproduction and re-use of this version - refer to the White Rose Research Online record for this item. Where records identify the publisher as the copyright holder, users can verify any specific terms of use on the publisher's website.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



Biological Reviews



Use of habitat odour by host-seeking insects

Journal:	Biological Reviews	
Manuscript ID	BRV-10-2015-0202.R1	
Manuscript Type:	Original Article	
Date Submitted by the Author:	n/a	
Complete List of Authors:	Webster, Ben; University of Sheffield, Animal and Plant Sciences Cardé, Ring T.; University of California, Department of Entomology	
Keywords:	habitat cues, host location, olfaction, semiochemicals, insect-host interactions, insect behaviour	

SCHOLARONE™ Manuscripts

I	USE OF HABITAT ODOUR BY HOST-SEEKING INSECTS
2	Ben Webster* ¹ and Ring T. Cardé ²
3	¹ Department of Animal and Plant Sciences, University of Sheffield, Western Bank, Sheffield,
4	S3 7AD, UK
5	² Department of Entomology, University of California Riverside, Riverside, CA 92521, USA
6	*Corresponding author: Webster, B. (b.webster@sheffield.ac.uk)
7	
8	

9 ABSTRACT

Locating suitable feeding or oviposition sites is essential for insect survival. Understanding how insects achieve this is crucial, not only for understanding ecology and evolution of insect-host interactions, but also for the development of sustainable pest control strategies that exploit insects' host-seeking behaviours. Volatile chemical cues are used by foraging insects to locate and recognise potential hosts but in nature these resources usually are patchily distributed, making chance encounters with host odour plumes rare over distances greater than 10s of meters. The majority of studies on insect host-seeking have focussed on short-range orientation to easily-detectable cues and it is only recently we have begun to understand how insects overcome this challenge. Recent advances show that insects from a wide range of feeding guilds make use of 'habitat cues', volatile chemical cues released over a relatively large area that indicate a locale where more specific host cues are most likely to be found. Habitat cues differ from host cues in that they tend to be released in larger quantities, are more easily detectable over longer distances, and may lack specificity, yet provide an effective way for insects to maximise their chances of subsequently encountering specific host cues. This review brings together recent advances in this area, discussing key examples and similarities in strategies used by haematophagous insects, soil-dwelling insects and insects that forage around plants. We also propose and provide evidence for a new theory that general and non-host plant volatiles can be used by foraging herbivores to locate patches of vegetation at a distance in the absence of more specific host-cues, explaining some of the many discrepancies between lab and field trials that attempt to make use of plant-derived repellents for controlling insect pests.

- 31 Key words: habitat cues, host location, olfaction, semiochemicals, insect-host interactions,
- 32 insect behaviour.

CONTENTS

34	I. Introduction	3
35	II. Use of habitat cues by foraging insects	6
36	(1) Soil-dwelling insects	6
37	(2) Haematophagous insects	8
38	(3) Predatory and parasitic insects	0
39	(4) Pollinators and above-ground foraging herbivores	11
10	(a) Green leaf volatiles	12
1 1	(b) Non-host volatiles	13
12	III. Implications and future work	15
13	IV. Conclusions	17
14	V. Acknowledgements	18
15	VI. References	19

I. INTRODUCTION

Over the years considerable knowledge has accumulated on how insects use volatile chemical cues to locate and recognise their respective host species. These cues usually consist of specific-characteristic blends of volatile compounds or, in some cases, individual volatiles that are restricted to a narrow range of related host species (Bruce, Wadhams & Woodcock, 2005; Bruce & Pickett, 2011). These cues offer an effective means of locating a host at short range but, due to the physical properties of odour plumes, chance encounters with host odour at longer distances are rare. Volatiles emanating from an odour source in wind form an odour plume that meanders downwind. Molecular diffusion occurs at too small a scale to contribute significantly to plume structure and the distribution of odours within the plume's overall boundaries is mainly dictated by the forces of turbulence, which creates discrete filaments of

relatively undiluted odour interspersed with clean air (Murlis, Elkinton & Cardé, 1992;

Voskamp, Den Otter & Noorman, 1998; Koehl, 2006). Many insects are adept at following these plumes (Murlis et al., 1992; Cardé & Willis, 2008; Bau & Cardé, 2015). Voskamp et al. (1998) showed that tsetse flies detected odour plumes up to 10-20 m downwind of an odour source in an open field and up to 60 m in woodland. At longer distances, however, odourant concentration can fall below insect detection thresholds (Murlis, Willis & Cardé, 2000; Koehl, 2006) and, together with increased intermittency of plume encounter (Koehl, 2006; Beyaert & Hilker, 2014), this means that insects' abilities to use host-originating odour to locate a feeding or oviposition site becomes increasingly difficult. This presents an enormous challenge to host-seeking insects. In areas with high plant species diversity, suitable hosts for phytophagous insects may be patchily distributed (Randlkofer et al., 2010) and insects may not come close enough to detect odour plumes using random foraging movements alone, particularly if hosts are hidden within dense patches of non-host vegetation that may obstruct or adsorb odour (Beyaert & Hilker, 2014). Haematophagous insects face a similar problem since their animal hosts may occupy large home ranges with distances of up to many kilometres between individuals or groups of individuals (Potts & Lewis, 2014). Where host odour cues are difficult to locate, the use of 'habitat odour cues' provides insects with a means of increasing their foraging success. As opposed to 'host cues' in the traditional sense, which are used to locate specific feeding or oviposition sites, habitat cues indicate a general area where such sites or associated cues are most likely to be found (Bell, 1990; Meiners, 2015). From a behavioural ecology perspective, habitat odour may be many things. Habitat odour may comprise the collective volatile emissions of all organisms inhabiting a

potential foraging patch. Many of the volatile compounds that insects can detect are found

ubiquitously across the plant or animal kingdoms, produced by host and non-host alike. For

example, green leaf volatiles and other ubiquitous plant volatiles can indicate the presence of

a patch of vegetation in a heterogeneous landscape. Respiratory CO₂ emissions produced by plant roots or animals can provide information on the presence of a patch of vegetation to below-ground feeding herbivores or a group of animals to blood-feeding insects. Alternatively, habitat odour may be associated with the collective secretions or excretions of host organisms, for example odours associated with dwellings or nests of animals that are hosts to blood-feeding insects. Since they tend to be produced over larger areas or by many different organisms within a habitat, habitat odour is generally emitted in greater quantities and detectable at greater distances than host odour, providing insects with information on a location to search for more host-specific volatile cues. The main ways in which habitat and host odour cues differ are summarised in Table 1.

Habitat cues may serve to increase foraging efficiency in a number of ways (Fig. 1). Insects may fly upwind in response to a habitat-odour plume before engaging in hierarchical plume switching (Beyaert& Hilker, 2014), abandoning the former long-range cue in favour of following the host plume to its source. Alternatively, habitat cues present as background odour may induce non-directional localised searching behaviours, for example through increases in rates of turning or changes in speed of movement in order to increase the probability of intercepting a host plume, after which movement upwind toward the host can occur. In these first two scenarios, habitat and host odour are encountered sequentially and encounter with host odour must override any behavioural response to the habitat odour cue. Habitat cues may also act in conjunction with host cues, reinforcing behavioural responses when detected by insects at the same time by providing important contextual information. Here, detection of habitat odour may sensitise insects to host volatiles, enhancing their responsiveness to these cues (Dekker, Geier & Cardé, 2005; Schröder & Hilker, 2008). This sensitization may work in conjunction with upwind flight and localised search behaviours in

response to habitat cues (Fig 1a & b), ensuring that insects respond strongly to host cues once they are encountered.

Since habitat odour is generally released over a relatively large area and in large quantities, foraging insects will find themselves exposed to habitat odour cues for lengthy periods of time. Constant exposure to these background odourants can eventually result in olfactory adaptation and habituation (Schröder & Hilker, 2008), meaning insects may become less responsive to habitat cues over time. This may serve to allow insects to 'give up' on a resource patch after failing to locate any host cues within it. Alternatively, constant exposure to habitat odour could lead to sensitisation, reinforcing the behavioural response over time.

II. USE OF HABITAT CUES BY FORAGING INSECTS

The majority of studies on insect host location have focussed on easily detectable, short-range cues originating from the host and it is only recently that evidence of habitat cue use has emerged. Their use now appears to be exceptionally widespread, employed by insects from a diverse range of feeding guilds including soil-dwelling insects, haematophagous insects, predatory and parasitic insects, above-ground herbivores, and pollinators. The seemingly widespread use of these cues suggests they are a fundamental component of insect host location. Key examples from each of these insect feeding guilds are described below.

(1) Soil-dwelling insects

Herbivorous insects that dwell within the soil make use of exudates from roots to locate suitable feeding sites (Johnson & Gregory, 2006a; Johnson & Nielsen, 2012) and these often confer species-specific information on host identity (Soni & Finch, 1979; Rogers & Evans, 2013a). Respiratory CO₂ emissions have also been shown to elicit behavioural responses from a range of root-feeding insects, suggesting a role in host location (Johnson *et al.*, 2012).

Carbon dioxide is generally produced in much higher quantities than other root exudates and diffuses relatively rapidly through the soil (Payne & Gregory, 1988), making it detectable at greater distances (Johnson & Nielsen., 2012). Carbon dioxide emission from roots is ubiquitous, produced by all respiring tissue, and also exhibits strong vertical gradients between the upper soil and air and, in areas of high root density, horizontal concentration gradients may not always be perceptible to soil-dwelling insects (Johnson et al., 2006b). This led Johnson and Gregory (2006a) to question the role of CO₂ in host location, particularly for specialist root herbivores for which CO₂ is unlikely to confer sufficient information. Instead, they proposed that CO₂ serves to inform as to the presence of a nearby patch of plants where more specific root exudates may subsequently be searched for, thus functioning as a 'search trigger' rather than a host cue. This hypothesis was supported by behavioural studies on the larvae of the root-feeding clover weevil, Sitona lepidus (Johnson et al., 2006b). In behavioural experiments, no evidence of oriented movement towards point emissions of CO₂ was observed, regardless of emission rates. In the presence of constant CO₂ emissions, however, larvae made more tortuous and intensive searching movements compared to CO₂free experiments, allowing insects to increase their chances of intercepting other root-derived chemical cues. Similar effects were observed for larvae of the wheat bulb fly, Delia coarctata, which displayed increased rates of turning and track length in elevated CO₂ but did not orientate toward point emissions in arena-based behavioural experiments (Rogers et al., 2013b).

Reinecke *et al.* (2008) showed that exudates from undamaged roots of dandelion inhibited behavioural responses of the European cockchafer (*Melolontha melolontha*) to CO₂. This was interpreted as a possible plant defence strategy, with exudates serving to mask the attractiveness of the long range cue. An alternative hypothesis is that these exudates may be used by host-seeking larvae to switch off responses to CO₂ when close enough to a plant

patch to be able to make use of more specific host cues or localised searching behaviours. Once host root exudates reach sufficient concentrations for inhibition of CO₂ detection, they are presumably also in sufficient concentrations to be used for host orientation, making CO₂ a redundant and, due to large horizontal gradients in CO₂ concentration close to dense plant patches, potentially disruptive signal at short range (Johnson & Gregory, 2006a). These two hypotheses are not mutually exclusive but further work is needed to validate either, preferably involving realistic plant densities that would be encountered by root-feeding larvae in field conditions. The use of host-originating CO₂ inhibitors may be a widespread phenomenon and future studies may uncover their use by other soil-dwelling insects.

(2) Haematophagous insects

Like root-feeding herbivores, haematophagous insects make use of a combination of CO₂ and more specific host volatiles, as well as heat, to locate a feeding site. The malaria mosquito, *Anopheles gambiae*, prefers to feed on human hosts and readily responds to human body odour in a wind tunnel by flying upwind but tends not to land on the source of emission unless it is heated (Spitzen *et al.*, 2013). We recently found that, when placed in a screen cage containing a source of human odour female *An. gambiae* did not land on the human odour source unless CO₂ was delivered through the side of the cage, instead preferring to rest on the walls and ceiling in the absence of CO₂ (Webster, Lacey & Cardé, 2015). Similar observations were made for the yellow fever mosquito, *Aedes aegypti*, which was found not to feed through a membrane when presented with human odour alone but nevertheless flew upwind upon detection of human odourants (Lacey, Ray & Cardé, 2014; McMeniman *et al.*, 2014). Anthropophilic mosquitoes such as *An. gambiae* search for blood meals in and around human dwellings. In our study we suggested that human odour, in the absence of carbon dioxide or heat, serves as a means for mosquitoes to locate a human dwelling since these continuously emit human odour even when its occupants are absent (Webster *et al.*, 2015).

On its own, human odour therefore likely serves as a habitat cue for these species, inducing location of and arrestment within a human dwelling, an ideal habitat within which to subsequently search for a blood meal. Once CO₂ or heat indicates a human is present, human odour acts together with these other cues to function as a host cue. This is an example of how the same volatile compounds can function as both habitat and host cue depending upon the context in which they are detected. Similar habitat cues are also used by mosquitoes feeding on non-human animals. The southern house mosquito, *Culex quinquefasciatus*, preferentially obtains blood meals from birds and is attracted by the odour of fresh chicken faeces, an effective cue indicating a physical location recently occupied by potential hosts (Cooperband *et al.*, 2008). Behavioural responses of haematophagous insects to urine and faecal odour are widespread and these offer effective and sometimes host-characteristic cues indicating a nest or general area regularly inhabited by potential hosts (Becker *et al.*, 1995; Baldacchino *et al.*, 2013; Nordéus *et al.*, 2014).

Exhaled CO₂ is generally considered a host cue for haematophagous insects (Cardé & Gibson, 2010) but recent studies have led us to question this hypothesis. Exhaled CO₂ offers little information regarding host species but is detectable over large distances (Zöllner *et al.*, 2004; Lorenz *et al.*, 2013) and, since many haematophagous insects tend to prefer social animals as hosts (Lehane, 2005), combined exhalations of a group of animals offers an effective long-range cue indicating a general area inhabited by potential hosts. Whilst CO₂ elicits upwind flight and plume following, haematophagous insects tend not to orient toward or land on the source at close range (Spitzen, Smallegange & Takken, 2008), instead initiating rapid 'zigzag' flight behaviour in the general vicinity of the CO₂ source (Spitzen *et al.*, 2008; Lacey *et al.*, 2014). This suggests a switch to localised searching behaviour in order to make contact with more specific host cues indicating potential feeding sites. Carbon dioxide therefore seems to function more as a 'habitat' cue by indicating a general area

occupied by potential hosts where more specific host cues may be subsequently located, similar to the model proposed by Johnson and Gregory (Johnson *et al.*, 2006a) used by root-feeding herbivores. Support for this hypothesis comes from that fact that at close range CO₂ is almost completely ignored in favour of skin odour by most haematophagous insects. *Aedes aegypti* readily flies upwind along a plume of CO₂ but, when presented with a human foot-odour plume in parallel, the CO₂ plume was completely ignored (Lacey *et al.*, 2014). Similar observations were made for *Cx. quinquefasciatus*, which also seemed to orient at long range to CO₂ but only used human odour at close range for landing (Lacey & Cardé, 2011).

As with root-feeding herbivores, volatiles that inhibit detection of CO₂ by haematophagous insects have recently been uncovered (Tauxe *et al.*, 2013). These compounds were identified using *in-silico* screening to predict chemical structures likely to interact with the CO₂ receptor (Boyle, McInally & Ray, 2013), providing a large range of compounds only a few of which have been tested and so the possible ecological function of these CO₂ inhibitors remains unclear for now. As with root-feeding herbivores, inhibition of long-range habitat cue detection may represent a defensive strategy by the host or, alternatively, a mechanism employed by the insect to facilitate switching from habitat cues to host cues at shorter ranges. Further work is needed to test either hypothesis and could lead to novel strategies for controlling these important public health pests.

(3) Predatory and parasitic insects

Among the most widely-recognised examples of use of habitat cues comes from predatory and parasitic insects. This topic has already been reviewed extensively (Vet & Dicke, 1992; Hare, 2011; Heil, 2014; Pierik, BallarÉ & Dicke, 2014; Hilker & Fatouros, 2015; Meiners, 2015) and so is only discussed briefly here. When searching for prey or insect hosts for oviposition, predatory and parasitic insects can make use of volatiles directly emanating from

their host's body or emitted as pheromones (Afsheen et al., 2008). Due to the small size of such odour sources these are often emitted in minute quantities, however, and so use of such cues in long-range host location is difficult. The plant on which the host is feeding represents a larger and far more easily detectable target at long range. Predatory and parasitic insects use plant odours to locate their hosts' habitat at a distance and subsequently engage in more localised foraging behaviour once on the plant (Bukovinszky et al., 2012; de Rijk, Dicke & Poelman, 2013). For example, the rove beetle *Aleochara bipustulata*, which feeds on and whose larvae parasitise the pupae of cabbage root flies, *Delia radicum*, uses volatiles emitted from fly-infested roots to locate a suitable area for foraging at a distance. Once in the vicinity of the root beetles can then make use of volatile cues from larval tracks and pupae (Goubert et al., 2013). Plants can benefit from recruitment of natural enemies of their herbivores and consequently tend to produce elevated quantities of volatiles upon herbivory (Vet & Dicke, 1992; Heil, 2014) or in response to herbivore egg deposition (Hilker & Fatouros, 2015). Herbivore-induced volatile blends may also provide specific information reflecting infestation by specific herbivore species (De Moraes et al., 1998; McCormick, Unsicker & Gershenzon, 2012), greatly facilitating eventual host-location by specialist parasitic insects. While herbivore-induced volatiles can substantially increase the detectability of prey at shortmid range, at distances of more than a few 10s of meters predatory and parasitic insects face the same challenge as other insects in that plume encounters may be too rare to provide an effective means of locating an infested plant. Larger-scale habitat cues that indicate an area of vegetation provide a solution to this problem, and evidence for use of such cues comes from pollinators and above-ground foraging herbivores.

(4) Pollinators and above-ground foraging herbivores

The use of habitat cues by insects foraging above-ground around plants remains largely unexplored but tantalizing indirect evidence exists for their use. Foliage offers a large source

of odour but at greater distances downwind falls in concentration within odour plumes (Murlis *et al.*, 2000; Koehl, 2006) and increasing intermittency (Koehl, 2006; Beyaert & Hilker, 2014) means they may be difficult to detect. Host plants located within the middle of a patch may be even harder to detect due to obstruction or adsorbtion of volatiles onto surfaces of downwind vegetation (Beyaert & Hilker, 2014). In the absence of specific host plumes, orienting first toward a broad patch of vegetation could substantially increase the foraging insect's chances of subsequently encountering host cues. This is particularly true in areas where vegetation coverage is not complete and broken up by bodies of water, rocky areas, urban constructions etc. Even where vegetation coverage is fairly extensive, localised regions containing higher abundances/diversity of plants will generally offer the most promising locations to search for a suitable host. Many plant volatiles are ubiquitous, produced by a wide range of different plant species in large quantities and can indicate such areas of vegetation. Even non-host volatiles, normally avoided at short-range (Bruce *et al.*, 2005), may be used to indicate such areas at long range and may facilitate eventual host encounter.

(a) Green leaf volatiles

To be of any use, habitat cues should be detectable at relatively large distances and thus emitted from a larger area and/or in higher concentrations. Green leaf volatiles (GLVs) are C-6 fatty acid derivatives, produced ubiquitously throughout the plant kingdom, and may offer such a cue. Whilst GLVs are generally only produced in large quantities following herbivory or physical stress (Mwenda & Matsui, 2014), in nature such stresses are common and so most plant patches produce large quantities of GLVs. Strong behavioural responses of herbivorous insects to GLVs have been shown by a number of insect species, even for those which are generally thought to recognise their hosts using highly species-specific blends. The black bean aphid, *Aphis fabae*, is attracted to specific blends of volatiles emitted by its host *Vicia*

faba (Webster et al., 2008a; Webster et al., 2008b) and the requirement for a blend is so pronounced that, when presented with individual host volatiles outside the context of the complete blend, aphids were repelled (Webster et al., 2010a). Notable exceptions to this trend, however, were the GLVs (Z)-3-hexenol and 1-hexenol, both of which were attractive on their own at levels similar to those emitted by plants (Webster et al., 2010b). Given the strict preference for host-specific blends by this aphid, and the total lack of host-specific information provided by GLVs, these responses are at first glance surprising. Such responses to GLVs are fairly widespread, however, with many insects responding positively to them despite normally showing preferences for highly host-specific volatile blends (Birkett et al., 2004; Ruther & Mayer, 2005; Alagarmalai et al., 2009; Li et al., 2014). Their possible role as habitat cues, indicating general areas of vegetation worthy of closer inspection by hostseeking insects, may explain this pattern. GLVs may also be used to inform of habitat suitability once the insect is already within a habitat. (Z)-3-hexenol is not attractive to the leaf beetle Cassida denticollis but the presence of this volatile as background odour dramatically increased the speed at which beetles discriminate host tansy (Tanacetum vulgare) stems from non-odourous dummy stems (Muller & Hilker, 2000), possibly by informing on overall habitat quality.

(b) Non-host volatiles

Aside from GLVs, general vegetative odours from plants other than hosts may serve to inform as to the location of a plant patch. Unmated female cotton leafworm (*Spodoptera littoralis*) feed from nectar-rich lilac flowers and respond to their odour in a wind tunnel (Saveer *et al.*, 2012). Cotton plants, which are used for oviposition and are only attractive to mated females, were not landed on by unmated females but their odour elicited increased take-off flights compared to dummy plants (Saveer *et al.*, 2012). This suggests that 'non-host' odour may play a role in long-range orientation even if they do not induce orientation at short

range. In fact, non-host volatiles that are repellent at short range may be attractive over longer distances when used as habitat cues. The idea that non-host volatiles may be used in host location is contrary to many long-held assumptions in insect behavioural ecology. A huge number of studies have demonstrated that insects will move away from non-host odours but the overwhelming majority of these studies used short-range olfactometers. Olfactometers are simple walking assays that are extremely efficient at screening short-range behavioural responses but do not account for long-range behavioural responses to volatile cues. Few studies have compared long- and short-range responses to the same odours but those that did showed surprising contradictions. Calatavud et al. (2014) showed that female cereal stem borers (Busseola fusca) avoided non-host Napier grass in preference of host maize in a Ytube olfactometer (short-range orientation) but showed no such preference in a wind tunnel (longer-range orientation). Even more striking discrepancies come from *Drosophila* for which, in an olfactometer designed to assess walking behaviour, addition of CO₂ to vinegar odour decreased its attractiveness whereas in a free-flying cage assay the addition of CO₂ raised the attractiveness of vinegar (Faucher, Hilker & de Bruyne, 2013). It is impossible to draw broad conclusions from the few studies that use both short- and long-range behaviour assays but in these examples at least, odours which are avoided at short range at the host location stage may elicit different, or opposite, responses at long range during the habitat location stage.

Use of non-host odours in the field to deter insect pests have met with mixed results. Although there are numerous examples of non-host plants being extraordinarily effective at reducing pest numbers when planted alongside hosts (Pickett *et al.*, 2014), there are many more that fail to have any effect in the field or that deliver opposite than expected results. In an attempt to use a range of non-host plant odours to protect roses against Japanese beetle (*Popillia japonica*) it was found that addition of supposedly repellent non-host species

actually increased numbers of invading beetles (Held, Gonsiska & Potter, 2003). Similar effects were observed when plants deterrent to the Colorado potato beetle (*Leptinotursa decemlineata*) were planted amidst potato plants, resulting in larger number of beetles than in untreated plots (Moreau, Warman & Hoyle, 2006). If non-host volatiles are avoided at short range but used to indicate the presence of a plant patch at longer ranges this may explain why attempts to incorporate non-host volatiles into integrated pest management strategies have often had opposite than expected outcomes in these, and other (Legaspi, Simmons & Legaspi, 2011; Moreno & Racelis, 2015) field experiments.

III. IMPLICATIONS

The use of habitat cues by foraging insects has now been demonstrated in a number of systems spanning several different feeding guilds. Despite this, use of habitat cues by insects is still widely overlooked. This is probably in part due to the difficulty of identifying such cues. Most studies use olfactometers that only record simple attractive/repellent behaviours used in short-range orientation. Some habitat cues may only operate at long range (thus walking assays may not always be appropriate) and may also elicit more complex nondirectional searching behaviours that are difficult to detect without using advanced tracking techniques in suitably large arenas. Video-tracking technology has advanced considerably in recent years allowing detailed three-dimensional flight paths to be constructed for small insects both inside the lab and outdoors (Spitzen et al., 2013; Manoukis et al., 2014). Insect movement can also be tracked effectively using radar or by fitting insects with active transmitters and tracking using radio telemetry (Chapman, Drake & Reynolds, 2011; Kissling, Pattemore & Hagen, 2014). These techniques have been successfully employed to track insect movement in the field, often over large distances, and can provide important insights into how insects move within and between different habitats (Negro et al., 2008; Hagen, Wikelski & Kissling, 2011; Lihoreau et al., 2012). As use of these advanced tracking

techniques becomes more widespread, we predict that identification of new habitat cues will accelerate.

Evidence for the use of habitat cues by above-ground-foraging herbivorous insects remains tentative with much work still to do. The possibility that non-host odours can function as habitat cues at long range deserves particular attention since this has obvious implications for the use of non-host volatiles as deterrents in integrated pest management strategies. The occurrence of habitat cue inhibitors in both soil-dwelling and haematophagous insects (Reinecke et al., 2008; Turner & Ray, 2009; Tauxe et al., 2013) suggests a widespread phenomenon and could fuel new pest management strategies. Attempts could be made to screen host volatiles for their ability to inhibit behavioural responses to known habitat cues when presented to insects simultaneously. Any identified habitat cue inhibitors could then potentially be used to disrupt habitat location at a distance if placed around the perimeter of an area to be protected, reducing the influx of pests. This would probably be more effective in situations where points of entry are limited, such as vents in a glasshouse or other man-made structure, where inhibitors can be most easily applied. A better understanding of habitat cues may also help with monitoring or mass-trapping strategies. For example, some haematophagous insects use CO₂ as a long-range habitat cue but may not orient towards it at close range, yet many modern CO₂ commercial traps still aim to trap insects at or very near to the point of CO₂ release (Vaidyanathan & Feldlaufer, 2013). More careful arrangement of habitat and host cues in such traps may lead to far more effective trapping rates (Spitzen et al., 2008; Cooperband & Cardé, 2006).

Future studies on host-location behaviour should seek to better distinguish between habitat and host cues. This could easily be achieved by using a combination of long- (e.g. wind tunnel) and short- (e.g. olfactometer) range behavioural assays to observe differences in responses to volatiles at different spatial scales and also by precisely tracking movement

paths of insects rather than simply recording their final destinations. Cues that elicit upwind orientation at long range but are ignored or avoided at short range are more likely to be habitat than host cues. Volatiles that elicit non-directional searching behaviour rather than directed movement to the point of emission are also more likely to be habitat cues, as are those whose presence as background odour enhances insects' responses to other host cues.

Whilst this review has focussed on examples from insects, use of habitat cues has also been recorded in vertebrates (e.g. use of dimethyl sulphide to indicate regions of biodiversity by procellariform seabirds (Nevitt, 2008)), suggesting they are a near-universal component of host location. An improved understanding of habitat cue use would greatly improve our understanding of insect foraging behaviour and ecology and may also lead to improved

development of pest control strategies that aim to exploit or disrupt insect host-seeking

IV. CONCLUSIONS

behaviours.

- 1. Habitat cues are used to improve chances of ultimately locating a host cue by inducing orientation toward, or triggering foraging behaviour within, a physical area that is likely to harbour hosts. Habitat cues differ from host cues in that they tend to be produced in larger quantities and detectable at greater distances than host cues and tend to provide less host-specific information.
- 2. Habitat odour comprises the collective volatile emissions of a habitat or physical region.

 These volatiles may originate collectively from the different organisms that inhabit the area.

 Many habitat cues are ubiquitous volatiles that, while offering little information on species identity, tend to be produced in large quantities from a wide range of plants/animals including host and non-host alike (e.g. GLVs from foliage, respiratory CO₂ emissions from roots or animals) and thus may be used to indicate patches of vegetation or groups of animals.

- Alternatively, habitat cues may originate from excretions or secretions of host organisms, for example indicating a dwelling or nesting area of a group of animals.
- 3. In recent years evidence of habitat cue use by insects has accelerated. There are now numerous clear examples of their use by soil-dwelling herbivores, haematophagous insects, predatory and parasitic insects, above-ground foraging herbivorous insects, and even birds.
- Use of habitat cues is thus rapidly emerging as an essential component of host location for many host-seeking organisms.
- 4. Numerous field and laboratory experiments suggest that non-host plant volatiles may
 function as habitat cues for host-seeking herbivorous insects, used to indicate patches of
 vegetation where potential hosts may be searched for if host-specific cues are unavailable.
- Non-host volatiles that are repellent at short range can be attractive over larger distances, further evidencing their role as habitat cues with important implications for their use in integrated pest management.
- 5. Research into host-location behaviour has largely involved use of short-range olfactometer behavioural assays that, while possessing many advantages for rapid screening of behavioural responses and assessment of short-range orientation, are unsuitable for identification of habitat cues. This is likely a reason for the slow progress made in identifying new habitat cues and future research making use of state of the art tracking technologies will undoubtedly result in discoveries of new habitat cues.

V. ACKNOWLEDGEMENTS

This work was funded in part by an R56AI099778 (NIAID) grant to Anandasankar Ray and Ring Cardé. The granting agencies had no role in the preparation of this work.

VI. REFERENCES

- 427 AFSHEEN S., WANG, X., LI, R., ZHU, C.-S. & LOU, Y.-G. (2008). Differential attraction of
- parasitoids in relation to specificity of kairomones from herbivores and their by-
- 429 products. *Insect Science* **15,** 381-397.
- 430 Alagarmalai J., Nestel, D., Dragushich, D., Nemny-Lavy, E., Anshelevich, L., Zada,
- A. & SOROKER, V. (2009). Identification of host attractants for the Ethiopian fruit fly,
- 432 Dacus ciliatus Loew. Journal of Chemical Ecology **35**, 542-551.
- 433 BALDACCHINO F., CADIER, J., PORCIANI, A., BUATOIS, B., DORMONT, L. & JAY-ROBERT, P.
- 434 (2013). Behavioural and electrophysiological responses of females of two species of
- 435 tabanid to volatiles in urine of different mammals. Medical and Veterinary
- 436 Entomology **27**, 77-85.
- 437 BAU J. & CARDÉ, R. T. (2015). Modeling optimal strategies for finding a resource-linked,
- windborne odor plume: theories, robotics, and biomimetic lessons from flying insects.
- *Integrative and Comparative Biology* **55,** 461-477.
- 440 BECKER N., ZGOMBA, M., PETRIC, D. & LUDWIG, M. (1995). Comparison of carbon dioxide.
- octenol and a host-odor as mosquito attractants in the upper Rhine valley, Germany.
- *Medical and Veterinary Entomology* **9,** 377-380.
- 443 BELL W. J. (1990). Locating patches and distant resources. In Searching behaviour: the
- behavioural ecology of finding resources, pp. 69-82. Chapman and Hall.
- BEYAERT I. & HILKER, M. (2014). Plant odour plumes as mediators of plant-insect
- interactions. *Biological Reviews* **89**, 68-81.
- High Birkett M. A., Bruce, T. J. A., Martin, J. L., Smart, L. E., Oakley, J. & Wadhams, L. J.
- 448 (2004). Responses of female orange wheat blossom midge, Sitodiplosis mosellana, to
- wheat panicle volatiles. *Journal of Chemical Ecology* **30,** 1319-1328.

- BOYLE S. M., McInally, S. & Ray, A. (2013). Expanding the olfactory code by in silico decoding of odor-receptor chemical space. *eLife* 2, e01120.
 BRUCE T. J. A. & PICKETT, J. A. (2011). Perception of plant volatile blends by herbivorous insects Finding the right mix. *Phytochemistry* 72, 1605-1611.
- BRUCE T. J. A., WADHAMS, L. J. & WOODCOCK, C. M. (2005). Insect host location: a volatile situation. *Trends in Plant Science* **10**, 269-274.
- BUKOVINSZKY T., POELMAN, E. H., KAMP, A., HEMERIK, L., PREKATSAKIS, G. & DICKE, M. (2012). Plants under multiple herbivory: consequences for parasitoid search behaviour and foraging efficiency. *Animal Behaviour* **83,** 501-509.
- CALATAYUD P. A., AHUYA, P. & LE RU, B. (2014). Importance of the experimental setup in research on attractiveness of odours in moths: an example with *Busseola fusca*.

 Entomologia Experimentalis Et Applicata 152, 72-76.
- CARDÉ R. T. & GIBSON, G. (2010). Host finding by female mosquitoes: mechanisms of
 orientation to host odours and other cues. In *Olfaction in vector-host interactions* (ed.
 W. Takken and B. G. J. Knols), pp. 115-142. Wageningen Academic Publishers,
- Wageningen.
- CARDÉ R. T. & WILLIS, M. A. (2008). Navigational strategies used by insects to find distant,
 wind-borne sources of odor. *Journal of Chemical Ecology* 34, 854-866.
- 468 CHAPMAN, J. W., DRAKE, V. A. & REYNOLDS. R. (2011). Recent insights from radar studies of insect flight. *Annual Review of Entomology* **56**, 337–356.
- COOPERBAND M. F. & CARDÉ, R. T. (2006). Orientation of Culex mosquitoes to carbon dioxide-baited traps: flight manoeuvres and trapping efficiency. *Medical and Veterinary Entomology* **20**, 11-26.

473	COOPERBAND M. F., MCELFRESH, J. S., MILLAR, J. G. & CARDÉ, R. T. (2008). Attraction of
474	female Culex quinquefasciatus Say (Diptera: Culicidae) to odors from chicken feces.
475	Journal of Insect Physiology 54, 1184-1192.
476	DEKKER, T., GEIER, M., & CARDÉ, R. T. (2005). Carbon dioxide instantly sensitizes female
477	yellow fever mosquitoes to human skin odours. Journal of Experimental Biology 208,
478	2963-2972.
479	DE MORAES C. M., LEWIS, W. J., PARE, P. W., ALBORN, H. T. & TUMLINSON, J. H. (1998).
480	Herbivore-infested plants selectively attract parasitoids. <i>Nature</i> 393 , 570-573.
481	DE RIJK M., DICKE, M. & POELMAN, E. H. (2013). Foraging behaviour by parasitoids in
482	multiherbivore communities. Animal Behaviour 85, 1517-1528.
483	FAUCHER C. P., HILKER, M. & DE BRUYNE, M. (2013). Interactions of carbon dioxide and
484	food odours in Drosophila: Olfactory hedonics and sensory neuron properties. Plos
485	One 8 , e56361.
486	GOUBERT C., JOSSO, C., LOUÂPRE, P., CORTESERO, A. M. & POINSOT, D. (2013). Short- and
487	long-range cues used by ground-dwelling parasitoids to find their host.
488	Naturwissenschaften 100, 177-184.
489	HAGEN, M., WIKELSKI, M. & KISSLING, W. D. (2011). Space use of bumblebees (Bombus
490	spp.) revealed by radio-tracking. <i>PLoS One</i> 6 , e19997.
491	HARE J. D. (2011). Ecological role of volatiles produced by plants in response to damage by
492	herbivorous insects. Annual Review of Entomology 56, 161-180.
493	HEIL M. (2014). Herbivore-induced plant volatiles: targets, perception and unanswered
494	questions. New Phytologist 204, 297-306.
495	HELD D. W., GONSISKA, P. & POTTER, D. A. (2003). Evaluating companion planting and non-
496	host masking odors for protecting roses from the Japanese beetle (Coleoptera:
497	Scarabaeidae). Journal of Economic Entomology 96, 81-87.

Science **63**, 36-44.

HILKER M. & FATOUROS, N. E. (2015). Plant responses to insect egg deposition. Annual Review of Entomology 60, 493-515. JOHNSON S. & NIELSEN, U. (2012). Foraging in the dark – chemically mediated host plant location by lelowground insect herbivores. *Journal of Chemical Ecology* **38**, 604-614. JOHNSON S. N. & GREGORY, P. J. (2006a). Chemically mediated host plant location and selection by root feeding insects. *Physiological Entomology* **31,** 1-13. JOHNSON S. N., ZHANG, X. X., CRAWFORD, J. W., GREGORY, P. J., HIX, N. J., JARVIS, S. C., MURRAY, P. J. & YOUNG, I. M. (2006b). Effects of carbon dioxide on the searching behaviour of the root-feeding clover weevil Sitona lepidus (Coleoptera: Curculionidae). Bulletin of Entomological Research 96, 361-366. KISSLING W. D., PATTEMORE, D. E. & HAGEN, M. (2014). Challenges and prospects in the telemetry of insects. *Biological Reviews* **89**, 511-530. KOEHL M. A. R. (2006). The fluid mechanics of arthropod sniffing in turbulent odor plumes. *Chemical Senses* **31,** 93-105. LACEY E. S. & CARDÉ, R. T. (2011). Activation, orientation and landing of female Culex quinquefasciatus in response to carbon dioxide and odour from human feet: 3D flight analysis in a wind tunnel. Medical and Veterinary Entomology 25, 94-103. LACEY E. S., RAY, A. & CARDÉ, R. T. (2014). Close encounters: contributions of carbon dioxide and human skin odour to finding and landing on a host in Aedes aegypti. Physiological Entomology **39**, 60-68. LEGASPI J. C., SIMMONS, A. M. & LEGASPI, B. (2011). Evaluating mustard as a potential companion crop for collards to control the silverleaf whitefly, Bemisia argentifolii (Hemiptera: Aleyrodidae): Olfactometer and outdoor experiments. Subtropical Plant

LEHANE M. (2005). The biology of blood-sucking in insects, 2nd edition. Cambridge University Press, New York. LI Y., ZHONG, S., QIN, Y., ZHANG, S., GAO, Z., DANG, Z. & PAN, W. (2014). Identification of plant chemicals attracting and repelling whiteflies. Arthropod-Plant Interactions 8, 183-190. LIHOREAU, M., RAINE, N. E., REYNOLDS, A. M., STELZER, R. J., LIM, K. S., SMITH, A. D., OSBORNE, J. L. & CHITTKA, L. (2012). Radar tracking and motion-sensitive cameras on flowers reveal the development of pollinator multi-destination routes over large spatial scales. *PLoS Biology* **10**, e1001392. LORENZ L. M., KEANE, A., MOORE, J. D., MUNK, C. J., SEEHOLZER, L., MSEKA, A., SIMFUKWE, E., LIGAMBA, J., TURNER, E. L., BISWARO, L. R., OKUMU, F. O., KILLEEN, G. F., MUKABANA, W. R. & MOORE, S. J. (2013). Taxis assays measure directional movement of mosquitoes to olfactory cues. Parasites & vectors 6, 131. MANOUKIS N. C., BUTAIL, S., DIALLO, M., RIBEIRO, J. M. C. & PALEY, D. A. (2014). Stereoscopic video analysis of *Anopheles gambiae* behavior in the field: Challenges and opportunities. Acta Tropica 132, S80-S85. MCCORMICK A. C., UNSICKER, S. B. & GERSHENZON, J. (2012). The specificity of herbivore-induced plant volatiles in attracting herbivore enemies. Trends in Plant Science 17, 303-310. MCMENIMAN C. J., CORFAS, R. A., MATTHEWS, B. J., RITCHIE, S. A. & VOSSHALL, L. B. (2014). Multimodal integration of carbon dioxide and other sensory cues drives mosquito attraction to humans. Cell 156, 1060-1071.

MEINERS T. (2015). Chemical ecology and evolution of plant-insect interactions: a

multitrophic perspective. Current Opinion in Insect Science 8, 22-28.

546	MOREAU T. L., WARMAN, P. R. & HOYLE, J. (2006). An evaluation of companion planting and
547	botanical extracts as alternative pest controls for the Colorado potato beetle.
548	Biological Agriculture & Horticulture 23, 351-370.
549	MORENO C. R. & RACELIS, A. E. (2015). Attraction, repellence, and predation: role of
550	companion plants in regulating Myzus persicae (Sulzer) (Hemiptera: Aphidae) in
551	organic kale systems of South Texas. Southwestern Entomologist 40, 1-14.
552	MULLER C. & HILKER, M. (2000). The effect of a green leaf volatile on host plant finding by
553	larvae of a herbivorous insect. Naturwissenschaften 87, 216-9.
554	MURLIS J., ELKINTON, J. S. & CARDÉ, R. T. (1992). Odor plumes and how insects use them.
555	Annual Review of Entomology 37, 505-532.
556	MURLIS J., WILLIS, M. A. & CARDÉ, R. T. (2000). Spatial and temporal structures of
557	pheromone plumes in fields and forests. Physiological Entomology 25, 211-222.
558	MWENDA C. M. & MATSUI, K. (2014). The importance of lipoxygenase control in the
559	production of green leaf volatiles by lipase-dependent and independent pathways.
560	Plant Biotechnology 31, 445-452.
561	NEVITT G. A. (2008). Sensory ecology on the high seas: the odor world of the procellariiform
562	seabirds. Journal of Experimental Biology 211, 1706-1713.
563	NEGRO, M., CASALE, A., MIGLIORE, L., PALESTRINI, C. & ROLANDO, A. (2008). Habitat use
564	and movement patterns in the endangered ground beetle species, Carabus olympiae
565	(Coleoptera: Carabidae). European Journal of Entomology 105, 105–112.
566	Nordéus K., Webster, B., Söderquist, L., Båge, R. & Glinwood, R. (2014). Cycle-
567	characteristic odour of cow urine can be detected by the female face fly (Musca
568	autumnalis). Reproduction in Domestic Animals 49, 903-908.
569	PAYNE D. & GREGORY, P. J. (1988). The soil atmosphere. In Russel's soil conditions and
570	plant growth (ed. A. Wild), pp. 298-314. Longman, Harlow, UK.

- PICKETT J. A., WOODCOCK, C. M., MIDEGA, C. A. O. & KHAN, Z. R. (2014). Push–pull farming systems. *Current Opinion in Biotechnology* **26**, 125-132.
- PIERIK R., BALLARÉ, C. L. & DICKE, M. (2014). Ecology of plant volatiles: taking a plant community perspective. *Plant, Cell & Environment* **37,** 1845-1853.
- POTTS J. R. & LEWIS, M. A. (2014). How do animal territories form and change? Lessons
- from 20 years of mechanistic modelling. Proceedings of the Royal Society B-
- 577 Biological Sciences **281**.
- 578 RANDLKOFER B., OBERMAIER, E., HILKER, M. & MEINERS, T. (2010). Vegetation
- complexity—The influence of plant species diversity and plant structures on plant
- chemical complexity and arthropods. *Basic and Applied Ecology* **11,** 383-395.
- REINECKE A., MÜLLER, F. & HILKER, M. (2008). Attractiveness of CO2 released by root
- respiration fades on the background of root exudates. Basic and Applied Ecology 9,
- 583 568-576.
- ROGERS C. D. & EVANS, K. A. (2013a). Wheat bulb fly (Delia coarctata, Fallen, Diptera:
- Anthomyiidae) larval response to hydroxamic acid constituents of host-plant root
- exudates. Bulletin of Entomological Research 103, 261-268.
- ROGERS C. D., EVANS, K. A., PARKER, J. & PAPPA, V. A. (2013b). Behavioural response of
- wheat bulb fly (*Delia coarctata*, Diptera: Anthomyiidae) larvae to the primary plant
- metabolite carbon dioxide. *Bulletin of Entomological Research* **103**, 675-82.
- 890 RUTHER J. & MAYER, C. J. (2005). Response of garden chafer, *Phyllopertha horticola*, to
- plant volatiles: from screening to application. Entomologia Experimentalis Et
- *Applicata* **115,** 51-59.
- 593 SAVEER A. M., KROMANN, S. H., BIRGERSSON, G., BENGTSSON, M., LINDBLOM, T.,
- BALKENIUS, A., HANSSON, B. S., WITZGALL, P., BECHER, P. G. & IGNELL, R. (2012).

595	Floral to green: mating switches moth olfactory coding and preference. <i>Proceedings</i>
596	of the Royal Society B-Biological Sciences 279, 2314-2322.
597	SONI S. & FINCH, S. (1979). Laboratory evaluation of sulphur-bearing chemicals as attractants
598	for larvae of the onion fly, Delia antiqua (Meigen)(Diptera: Anthomyiidae). Bulletin
599	of Entomological Research 69 , 291-298.
600	SPITZEN J., SMALLEGANGE, R. C. & TAKKEN, W. (2008). Effect of human odours and
601	positioning of CO(2) release point on trap catches of the malaria mosquito Anopheles
602	gambiae sensu stricto in an olfactometer. Physiological Entomology 33, 116-122.
603	SPITZEN J., SPOOR, C. W., GRIECO, F., TER BRAAK, C., BEEUWKES, J., VAN BRUGGE, S. P.,
604	Kranenbarg, S., Noldus, L. P. J. J., van Leeuwen, J. L. & Takken, W. (2013). A
605	3D analysis of flight behavior of Anopheles gambiae sensu stricto malaria mosquitoes
606	in response to human odor and heat. Plos One 8, e62995.
607	TAUXE G. M., MACWILLIAM, D., BOYLE, S. M., GUDA, T. & RAY, A. (2013). Targeting a dual
608	detector of skin and CO2 to modify mosquito host seeking. Cell 155, 1365-1379.
609	TURNER S. L. & RAY, A. (2009). Modification of CO ₂ avoidance behaviour in <i>Drosophila</i> by
610	inhibitory odorants. <i>Nature</i> 461 , 277-U159.
611	VAIDYANATHAN R. & FELDLAUFER, M. F. (2013). Bed bug detection: current technologies
612	and future directions. American Journal of Tropical Medicine and Hygiene 88, 619-
613	625.
614	VET L. E. M. & DICKE, M. (1992). Ecology of infochemical use by natural enemies in a
615	tritrophic context. Annual Review of Entomology 37, 141-172.
616	VOSKAMP K., DEN OTTER, C. & NOORMAN, N. (1998). Electroantennogram responses of
617	tsetse flies (Glossina pallidipes) to host odours in an open field and riverine
618	woodland. Physiological Entomology 23, 176-183.

619	WEBSTER B., BRUCE, T., PICKETT, J. & HARDIE, J. (2010a). Volatiles functioning as host cues
620	in a blend become nonhost cues when presented alone to the black bean aphid. Animal
621	Behaviour 79 , 451-457.
622	Webster B., Bruce, T. J. A., Dufour, S., Birkemeyer, C., Birkett, M. A., Hardie, J. &
623	PICKETT, J. A. (2008a). Identification of volatile compounds used in host location by
624	the black bean aphid, Aphis fabae. Journal of Chemical Ecology 34, 1153-1161.
625	WEBSTER B., BRUCE, T. J. A., PICKETT, J. A. & HARDIE, J. (2008b). Olfactory recognition of
626	host plants in the absence of host-specific volatile compounds. Communicative and
627	Integrative Biology 1, 167-169.
628	WEBSTER B., GEZAN, S., BRUCE, T., HARDIE, J. & PICKETT, J. (2010b). Between plant and
629	diurnal variation in quantities and ratios of volatile compounds emitted by Vicia faba
630	plants. Phytochemistry 71, 81-89.
631	WEBSTER B., LACEY, E. S. & CARDÉ, R. T. (2015). Waiting with bated breath: opportunistic
632	orientation to human odor in the malaria mosquito, Anopheles gambiae, is modulated
633	by minute changes in carbon dioxide concentration. Journal of Chemical Ecology 41,
634	59-66.
635	ZÖLLNER G. E., TORR, S. J., AMMANN, C. & MEIXNER, F. X. (2004). Dispersion of carbon
636	dioxide plumes in African woodland: implications for host-finding by tsetse flies.
637	Physiological Entomology 29 , 381-394.
638	
639	Figure 1. Use of habitat cues by foraging insects. Shown are three main ways in which habitat
640	odour can used for eventual location of a suitable host for feeding/oviposition (green circle):
641	a) The large red/green arrow represents odour from a potential resource patch. Insects follow
642	this toward the patch before eventually encountering host odour (small green arrow).
643	Encounter with host odour leads to a switch in behaviour, where insect ceases to move in

response to habitat odour and instead follows host odour to its source. b) Habitat odour (red/green area) present in the background induces non-directional localised searching behaviour in the form of increased rates of turning and changes in movement speed, increasing the probability of chance encounter with host odour. Once host odour is detected, the insect abandons this localised searching behaviour and instead follows the host odour toward its source. c) Habitat cues present as background odour gate behavioural responses to host cues. When relevant background odour is detected that indicates the insect is in a suitable habitat, insects become more responsive to host-odour cues. When this relevant background odour is lacking, host-odour cues that the insect detects are either responded to weakly or ignored.

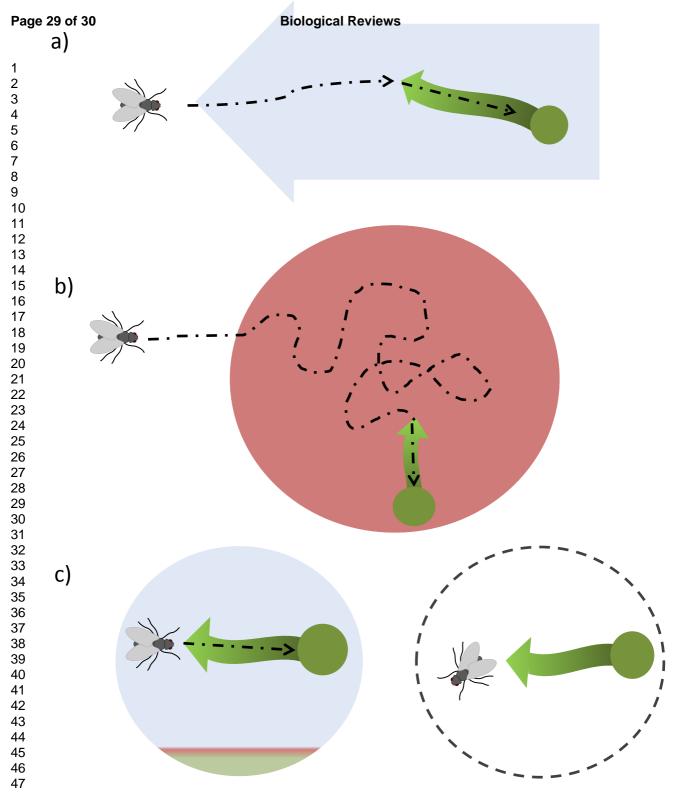


Table 1 Host cues vs. habitat cues. 'Host cue' has become a general term used to refer to any cue used at any stage in host location. There is an important distinction, however, between cues used to locate a feeding/oviposition site (host cues) and cues used to inform of a general area where host cues may subsequently be found (habitat cues). We propose the following broad criteria to help distinguish between the two. This list is not intended to be overly prescriptive since there are undoubtedly exceptions to each and should instead serve only as a general guide to distinguish between the two.

Feature	Host cues	Habitat cues
Function	Indicate location of a	Indicate a general area/location where
	feeding/oviposition site	host cues are most likely to be
		encountered
Source	Emitted from host	Emitted from host's habitat, which
		may include the host itself as well as
		non-hosts
Quantities	Lower quantities	Higher quantities
emitted in		
Detectability	Detectable at short distances	Detectable at longer distances
Specificity	Often host-specific	Not necessarily host-specific
Behaviours	Directed movement towards odour	General upwind movement, localised
elicited	source	searching behaviour, or enhanced
		responses to host cues