

This is a repository copy of *Has the sensitivity of soybean cultivars to ozone pollution increased with time? : An analysis of published dose–response data.*

White Rose Research Online URL for this paper:
<https://eprints.whiterose.ac.uk/102871/>

Version: Published Version

Article:

Osborne, Stephanie, Mills, Gina, Hayes, Felicity et al. (3 more authors) (2016) Has the sensitivity of soybean cultivars to ozone pollution increased with time? : An analysis of published dose–response data. *Global Change Biology*. pp. 3097-3111. ISSN 1354-1013

<https://doi.org/10.1111/gcb.13318>

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here:
<https://creativecommons.org/licenses/>

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

Has the sensitivity of soybean cultivars to ozone pollution increased with time? An analysis of published dose–response data

STEPHANIE A. OSBORNE^{1,2}, GINA MILLS¹, FELICITY HAYES¹, ELIZABETH A. AINSWORTH³, PATRICK BÜKER² and LISA EMBERSON²

¹Centre for Ecology and Hydrology, Environment Centre Wales, Bangor LL57 2UW, UK, ²Stockholm Environment Institute, University of York, York YO10 5NG, UK, ³USDA ARS Global Change and Photosynthesis Research Unit, Urbana, IL 61801, USA

Abstract

The rising trend in concentrations of ground-level ozone (O₃) – a common air pollutant and phytotoxin – currently being experienced in some world regions represents a threat to agricultural yield. Soybean (*Glycine max* (L.) Merr.) is an O₃-sensitive crop species and is experiencing increasing global demand as a dietary protein source and constituent of livestock feed. In this study, we collate O₃ exposure–yield data for 49 soybean cultivars, from 28 experimental studies published between 1982 and 2014, to produce an updated dose–response function for soybean. Different cultivars were seen to vary considerably in their sensitivity to O₃, with estimated yield loss due to O₃ ranging from 13.3% for the least sensitive cultivar to 37.9% for the most sensitive, at a 7-h mean O₃ concentration (M7) of 55 ppb – a level frequently observed in regions of the USA, India and China in recent years. The year of cultivar release, country of data collection and type of O₃ exposure used were all important explanatory variables in a multivariate regression model describing soybean yield response to O₃. The data show that the O₃ sensitivity of soybean cultivars increased by an average of 32.5% between 1960 and 2000, suggesting that selective breeding strategies targeting high yield and high stomatal conductance may have inadvertently selected for greater O₃ sensitivity over time. Higher sensitivity was observed in data from India and China compared to the USA, although it is difficult to determine whether this effect is the result of differential cultivar physiology, or related to local environmental factors such as co-occurring pollutants. Gaining further understanding of the underlying mechanisms that govern the sensitivity of soybean cultivars to O₃ will be important in shaping future strategies for breeding O₃-tolerant cultivars.

Keywords: China, crops, cultivar, food security, *Glycine max*, India, response functions, yield

Received 24 September 2015; revised version received 24 March 2016 and accepted 30 March 2016

Introduction

Ensuring that the rising global population has access to a sufficient and stable food supply is a key international priority for the 21st century. At a time when an estimated 795 million people worldwide are undernourished (FAO, 2015), agricultural productivity is being limited by several factors, including *inter alia*, rising water scarcity (Falkenmark, 2013), the limited land available for cultivation (Zabel *et al.*, 2014), widespread soil erosion and degradation (FAO, 2011), and the impacts of climate change (Parry *et al.*, 2004). A further threat to agricultural yield comes from rising concentrations of ground-level ozone (O₃) (Fuhrer, 2009) – a common air pollutant and phytotoxin (Krupa *et al.*, 2001). Ozone is a secondary pollutant formed in photochemical reactions from precursor compounds, the most important of which are nitrogen oxides (NO_x), methane

(CH₄) and carbon monoxide (CO) (Royal Society, 2008). The global surface background concentration of O₃ more than doubled between the early 1900s and the end of the 20th century (Hough & Derwent, 1990; Parrish *et al.*, 2014), most likely as a result of rising anthropogenic emissions of O₃ precursor compounds from fossil fuel combustion, biomass burning and paddy field cultivation (Brasseur *et al.*, 2001). Projected changes in global surface O₃ for the period 2000–2050 range from a decrease in the 24-h mean of 2.5–7.2 ppb under the optimistic emission scenarios (RCP2.6, RCP4.5, RCP6.0, B1) to an increase in 1.5–6.2 ppb under the more pessimistic RCP8.5 and A2 emission scenarios (IPCC, 2013). Trends in surface O₃ are, however, highly variable geographically, and the most rapid increase is currently occurring in South Asia where surface O₃ concentrations are expected to continue to rise until 2050 under all but one of the emission scenarios (Beig & Singh, 2007; IPCC, 2013). Establishing a thorough understanding of crop and cultivar responses to O₃, and the incorporation of

Correspondence: Stephanie A. Osborne, tel. +44 07906877798, fax +44 01904322898, e-mail: stepo@ceh.ac.uk

these responses into crop production models, is therefore needed to quantify the potential impact of O₃ on food supply in different world regions.

Soybean (*Glycine max.* (L.) Merr.) ranks among the most O₃-sensitive agricultural crops (Mills *et al.*, 2007). It is the fifth most significant crop in terms of global production (FAO, 2012), is a key source of vegetable protein for humans (Mateos-Aparicio *et al.*, 2008), provides approximately 30% of the world's processed vegetable oil (Graham & Vance, 2003), accounts for 77% of global nitrogen fixation by crop legumes (Herridge *et al.*, 2008) and is an important feed constituent for the livestock and aquaculture industries (Hartman *et al.*, 2011). The crop holds significant economic importance for a number of world economies including the USA, Brazil, Argentina, China and India (FAO, 2014), and world soybean demand is increasing by an average of 2.2% annually (Masuda & Goldsmith, 2009). Ozone exposure reduces the photosynthetic rate, stomatal conductance (g_s), leaf chlorophyll content and leaf starch concentration of soybean (Morgan *et al.*, 2003). Ground-level O₃ pollution over agricultural land has been estimated to cause an annual reduction in soybean yield ranging between 6 and 16%, and financial losses of \$2.0–5.8 billion annually, based on analysis of year 2000 data conducted in two separate global crop loss assessments (Van Dingenen *et al.*, 2009; Avnery *et al.*, 2011a). Soybean crop yield reduction for the year 2030 as a result of O₃ is estimated to be 9.5–15% under the optimistic (B1) scenario or 15–19% under the pessimistic (A2) emission scenario (Avnery *et al.*, 2011b).

The magnitude of O₃ damage to soybean is dependent on the timing of exposure, with greater reductions in photosynthesis and yield being observed when exposure occurs during the reproductive stages of growth (Morgan *et al.*, 2003). Co-occurrence of seasonal peaks in O₃ surface concentrations and the flowering and pod-filling stages could therefore be particularly damaging for yield. Ozone damage occurs when the gaseous pollutant enters the leaf *via* the stomatal pores, and interacts with cell membranes and walls in the apoplast to yield reactive oxygen species (ROS) (Wilkinson *et al.*, 2012); these directly damage plant tissue through protein oxidation, leading to accelerated senescence and cell death (Fiscus *et al.*, 2005). The widely observed reduction in photosynthetic rate in response to O₃ is not fully understood, but is in part the result of a reduction in the leaf concentrations of chlorophyll and Rubisco (Glick *et al.*, 1995; Fiscus *et al.*, 2005). Ozone has also been observed to reduce nodulation in a range of legume species including soybean (Tingey & Blum, 1973; Reinert & Weber, 1980; Zhao *et al.*, 2012), although this effect is largely thought to be a secondary response as a result of reduced total carbon assimilation

and the diversion of assimilates away from the roots (Hewitt *et al.*, 2016).

Dose–response studies for a range of crops have revealed that O₃ sensitivity is a heritable trait (Reinert & Eason, 2000) and is highly variable among species and among cultivars (Ariyaphanphitak *et al.*, 2005; Mills *et al.*, 2007; Mills & Harmens, 2011). The maximum stomatal conductance which a species or cultivar can reach (g_{max}) is thought to play a role in determining O₃ sensitivity, because greater conductance results in greater O₃ uptake. This view is supported by the observation that wheat cultivar sensitivity to O₃ is positively correlated with g_{max} (Biswas *et al.*, 2008). Furthermore, modern wheat varieties are more sensitive to O₃ than older varieties; this may be a result of selective breeding programmes targeting varieties with a higher g_s , as these have a higher rate of CO₂ fixation leading to higher yields (Biswas *et al.*, 2008; Roche, 2015). The detoxification and repair capacity of a plant species or variety is also thought to be important in determining sensitivity (Fiscus *et al.*, 2005): for example, O₃ tolerance of a number of plant species has been seen to positively correlate with greater apoplastic concentrations of ascorbic acid, an antioxidant (Frei *et al.*, 2008, 2010; Frei, 2015). A thorough understanding of how O₃ sensitivity varies among cultivars of the same species – and the factors which drive these differences – is key in improving assessments of current and future O₃-induced crop losses. Previous studies in soybean investigating inter-cultivar variation in O₃ response have typically compared a relatively small number of cultivars from the same geographical region: examples include studies of USA cultivars by Betzelberger *et al.* (2010, 2012) and an investigation of Chinese cultivars by Zhang *et al.* (2014). Knowledge of which cultivars are most resistant to the effects of O₃ could potentially help plant breeders to develop O₃-tolerant soybean varieties, which, if adopted by farmers, could mitigate O₃-induced crop losses.

Much of the research relating to soybean-O₃ responses conducted to date has taken place in the USA, as part of the US National Crop Loss Assessment Network (NCLAN) programme in the 1970s and 1980s (Heagle, 1989) and more recently at the University of Illinois, Urbana-Champaign and USDA Agricultural Research Service SoyFACE facility in Illinois (Long *et al.*, 2005; Betzelberger *et al.*, 2010, 2012; Gillespie *et al.*, 2012). Groups in India and China have also studied O₃ responses to soybean in recent years, but these data have, to date, not been pooled to produce dose–response relationships. Response functions for soybean used in global crop loss assessments have therefore been based on experimental data collected only in the USA. Two dose–response functions for soybean have been published: one by Lesser *et al.* (1990), synthesized from the

NCLAN data set; and one by Mills *et al.* (2007), who combined some of the NCLAN data with more recent dose–response data collected in the USA to update the function. These functions have been applied in a number of different studies in order to estimate O₃-induced soybean yield reduction globally and the associated financial loss to farmers. Producing these estimates involves combining a dose–response function for soybean with crop distribution and yield maps, growing season dates and modelled O₃ concentrations. The Mills *et al.* (2007) function was used by Avnery *et al.* (2011a) in their global assessment of O₃-induced soybean crop losses. The Lesser *et al.* (1990) function was used by Wang & Mauzerall (2004) in their soybean yield loss assessment for East Asia and by Van Dingenen *et al.* (2009) in their global assessment. Both functions were used by Tai *et al.* (2014) in their analysis of combined O₃ and climate change effects on future soybean production. All of these assessments applied a soybean dose–response function based on data from North America to model yield impacts in Asia. However, a comparison by Emberson *et al.* (2009) of wheat and rice dose–response data from North America and Asia has shown that Asian wheat and rice cultivars appear to be more sensitive to O₃ than their North American counterparts, possibly due to locally occurring physiological traits associated with sensitivity, such as high *g_s* and low antioxidative capacity (Emberson *et al.*, 2009). The application of North American dose–response functions in global yield loss assessments for wheat and rice may have therefore underestimated O₃-induced yield losses in Asia.

This study, to our knowledge, synthesizes all existing data in the scientific literature describing soybean yield response to O₃, in order to produce a comprehensive and up-to-date dose–response function. We also analyse intercultural differences in O₃ sensitivity, allowing the most O₃-sensitive and O₃-tolerant soybean cultivars to be identified. Additional analysis is also conducted on the dose–response data set, to investigate potential correlations between the degree of O₃ sensitivity observed and (i) the year in which the soybean cultivar was released, to identify temporal trends in sensitivity; (ii) the geographical location of the dose–response experiment, to determine whether sensitivity varies geographically; and (iii) the method of O₃ fumigation used in experimentation, to assess whether experimental design influences the sensitivity observed.

Materials and methods

Literature search

A search of the published scientific literature was performed between October 2013 and September 2014 to find all O₃

exposure studies conducted on soybean. The search was conducted using the Science Citation Index Expanded® (Thomson-ISI, Philadelphia, PA, USA). The criteria for inclusion were as follows:

1. Ozone exposure concentrations must have been presented as either the seasonal 7-h (M7), 8-h (M8), 12-h (M12) or 24-h (M24) means or as the 3-month AOT40.
2. The exposure experiments must have taken place in the open air, either within open-top chambers (OTC) or using free air concentration enrichment (FACE). For experiments which included one or more additional experimental variables alongside O₃ concentration (e.g. watering regime, nitrogen concentration), only the yield data from the control treatment were used.
3. The duration of O₃ exposure must have spanned at least 60% of the total growing season. Soybean takes approximately 3 months (90 days) from sowing to maturity (Pedersen & Lauer, 2004). About 60% of this period is equal to 7.7 weeks, which was rounded to a minimum exposure duration of 8 weeks for the purpose of this study.
4. Yield must have been measured directly, as the pod or seed weight. Response parameters such as total aboveground biomass, photosynthetic rate, percentage leaf damage or the 100-seed weight were not considered to represent the yield response.

The literature search found 28 studies that met the inclusion criteria and were included in the analysis. These studies included experiments investigating 48 cultivars, and when combined, produced a data set comprising 379 data points. A list of all the experimental studies included in our analysis can be found in Table 1, alongside information relating to study sites, cultivars tested and experimental design. Experiments which had used pot-grown soybean were included in our analysis; this was justified given that we found no significant difference in the dose–response relationships exhibited by pot-grown and field-grown soybean (See Appendix S1).

Standardization of O₃ and yield parameters

Dose–response data in the literature were presented using a number of different concentration metrics and yield parameters, as listed above. All O₃ concentration data had to be converted into a standard metric to enable the data to be combined for analysis. The M7 was selected to act as the common O₃ metric in our analysis, because this was most frequently reported in the literature. O₃ values presented in the form of the AOT40, M12 and M24 were converted to the M7 metric using conversion functions calculated using The ICP Vegetation database (described in Fig. 1), which contains O₃ observations measured at the same time and location but using a range of different O₃ metrics. The three different conversion functions which we used to standardize our data to the M7 metric, calculated from the ICP Vegetation database, are shown in Fig. 1. For each of the separate years and measurement stations for which there were seasonal O₃ data, the 3-month M7, M12, M24 and AOT40 were calculated for the summer season (1 May – 31 August). Concentration values

Table 1 List of experimental studies included in the analysis, with information regarding the study site, experimental design and cultivars used

References	Study site	Exposure type used	Method of soybean cultivation	Cultivars tested	O ₃ range (M7, ppb)	Calculated theoretical yield at zero O ₃	Parameter used for reporting yield
Betzlberger <i>et al.</i> (2010)	Champaign, USA	FACE	Field	A3127; Clark; Dwight; Holt; HS93-4118; IA-3010; LN97-15076; Loda; NE3399; Pana	37.9–82.5	5048.2–2785.7	Seed yield, kg ha ⁻¹
Betzlberger <i>et al.</i> (2012)	Champaign, USA	FACE	Field	93B15; Dwight; HS93-4118; IA-3010; LN97-15076; Loda; Pana	38.1–120.6	5005.3–3206.3	Seed yield, kg ha ⁻¹
Booker <i>et al.</i> (2005)	Raleigh, USA	OTC	Pot	Essex	26.0–76.0	72.7–33.3	Seed yield, g plant ⁻¹
Bou Jaoudé <i>et al.</i> (2008)	Bari, Italy	OTC	Field	Casa	31.2–44.7	0.58	Seed yield, kg m ⁻²
Chernikova <i>et al.</i> (2000)	Beltsville, USA	OTC	Field	Forest; Essex	24.2–62.9	491.8–414.3	Seed yield, g m ⁻²
Fiscus <i>et al.</i> (1997)	Raleigh, USA	OTC	Pot	Essex	23.7–94.7	212.2–167.4	Seed yield, g plant ⁻¹
Heagle & Letchworth (1982)	Raleigh, USA	OTC	Pot	Forest; Davis; Ransom; Bragg	26.0–100.0	123.6–67.3	Seed yield, g plant ⁻¹
Heagle <i>et al.</i> (1983a)	Raleigh, USA	OTC	Field	Davis	24.5–124.7	468.0	Seed yield, g m ⁻¹ of row
(Heagle <i>et al.</i> , 1983b)	Raleigh, USA	OTC	Field	Davis	25.0–98.0	89.8–67.3	Seed yield, g plant ⁻¹
Heagle <i>et al.</i> (1986)	Raleigh, USA	OTC	Field	Davis	19.0–92.0	560.9	Seed yield, g m ⁻¹ of row
Heagle <i>et al.</i> (1987)	Raleigh, USA	OTC	Field	Davis	30.0–107.0	529.4–465.0	Seed yield, g m ⁻¹ of row
Heagle <i>et al.</i> (1991)	Raleigh, USA	OTC	Pot	Forrest; Davis; Bragg; Ransom	25.0–96.8	287.2–158.9	Seed weight, g pot ⁻¹
Heagle <i>et al.</i> (1998)	Raleigh, USA	OTC	Pot	Essex; Holladay; NK 6955	21.4–78.4	166.4–123.2	Seed yield, g plant ⁻¹
Heggestad <i>et al.</i> (1985)	Beltsville, USA	OTC	Field	Williams-79; Forrest; Corsoy-79	16.0–51.0	8140.3–3765.5	Seed yield, kg ha ⁻¹
Heggestad <i>et al.</i> (1988)	Beltsville, USA	OTC	Field	Williams-79; Corsoy-79	19.0–32.0	38.4–30.1	Seed yield, g plant ⁻¹
Heggestad & Lesser (1990)	Beltsville, USA	OTC	Field	Williams-79; Essex; Forrest; Corsoy-79	15.0–99.0	5867.7–4441.6	Seed yield, kg ha ⁻¹
Kohut <i>et al.</i> (1986)	Ithaca, USA	OTC	Field	Hodgson	17.0–122.0	12.8	Seed yield, g plant ⁻¹
Kress & Miller (1983)	Chicago, USA	OTC	Field	Corsoy	22.0–115.0	3097.4	Seed yield, kg ha ⁻¹
Kress <i>et al.</i> (1986)	Raleigh, USA	OTC	Field	Amsoy-71; Corsoy-79	23.0–92.0	6.0–4.8	Seed yield, g plant ⁻¹
Miller <i>et al.</i> (1989)	Raleigh, USA	OTC	Field	Young	15.5–94.7	596.2	Seed yield, g m ⁻¹
Miller <i>et al.</i> (1994)	Raleigh, USA	OTC	Pot	Essex; NK 6955; S 53–34	14.4–94.7	200.3–136.0	Seed yield, g pot ⁻¹
Morgan <i>et al.</i> (2006)	Champaign, USA	FACE	Field	93B15	50.0–75.0	800.9–563.9	Seed yield, g m ⁻²
Mulchi <i>et al.</i> (1988)	Beltsville, USA	OTC	Field	Calland; Cumberland; Pella; Williams; Miles; Sparks; Union; Ware; Bay; Essex; Forrest; York	22.7–67.3	564.7–246.0	Seed yield, g cm ⁻²
Mulchi <i>et al.</i> (1995)	Beltsville, USA	OTC	Field	Clark	26.0–72.7	253.7	Seed yield, g m ⁻²
Robinson & Britz (2000)	Beltsville, USA	OTC	Field	Essex; Forrest	24.0–58.0	28.6–21.4	Seed yield, g plant ⁻¹
Singh <i>et al.</i> (2010)	Varanasi, India	OTC	Field	PK-472; Bragg	10.0–61.7	6.6–5.3	Pod yield, g plant ⁻¹
Singh & Agrawal (2011)	Varanasi, India	OTC	Field	Pusa 9712; Pusa 9814	4.0–74.7	8.3–5.2	Seed yield, g plant ⁻¹
Troiano <i>et al.</i> (1983)	Ithaca, USA	OTC	Field	Beeson	8.0–27.0	14.9	Seed yield, g plant ⁻¹
Zhang <i>et al.</i> (2014)	Harbin, China	OTC	Pot	Hefeng25; Hefeng35; Hefeng55; Heinong35; Heinong37; Heinong65; Suinong22; Suinong26; Suinong31	19.1–58.6	24.3–15.7	Seed yield, g plant ⁻¹

OTC, Open-top chamber; FACE, Free air concentration enrichment.

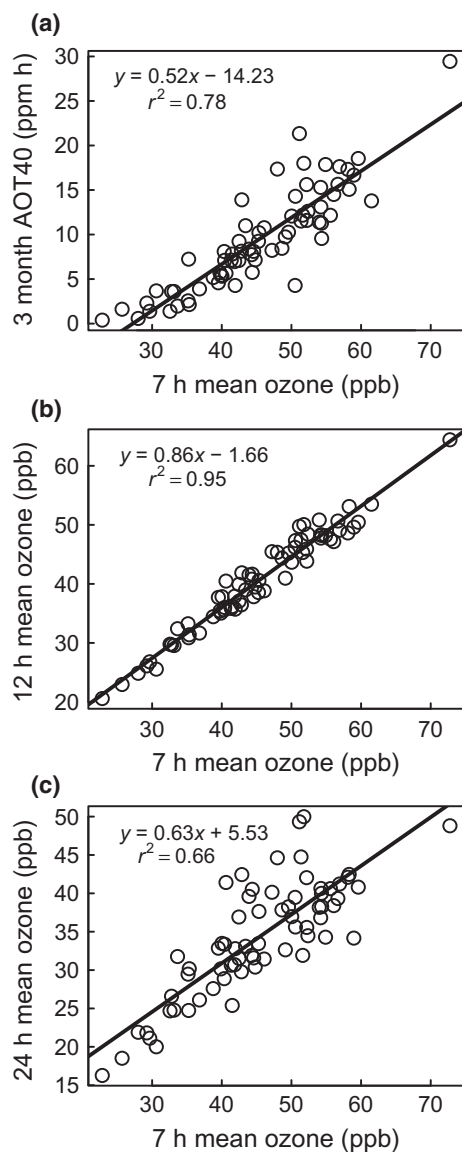


Fig. 1 Conversion functions used to convert between (a) 3-month AOT40 and 7 h mean, (b) 12 h mean and 7 h mean, (c) 24 h mean and 7 h mean O_3 concentrations. Data points represent summer season measurements of O_3 concentration at 35 stations between 2001 and 2013, recorded in the ICP Vegetation database. Measurement stations were located in Austria, Belgium, China, France, Germany, Greece, Hungary, Italy, Japan, Latvia, Poland, Portugal, Slovenia, Spain, Switzerland and the United Kingdom.

represented using the different O_3 metrics were then plotted against each other, and conversion functions were derived using linear regression.

During standardization of the reported O_3 concentrations to the M7, concentrations presented as the M8 were considered to be equivalent to the M7, as the small difference between the two was considered unlikely to add significant uncertainty to our analysis. A total of 205 O_3 concentration values were presented in the soybean data set using the M7 or M8 metrics and

did not need to be converted. A total of 125 and 49 data points were presented using the AOT40 and M12 metrics, respectively, and were converted to M7. Not all of the studies included in our analysis used a full 3-month O_3 exposure; for studies which had shorter exposure durations, it was assumed that the 3-month mean would not radically differ from the mean covering a shorter duration, as O_3 exposure in all studies was artificial and therefore would not follow natural seasonal patterns in O_3 concentration. No study which had used an exposure duration of less than 8 weeks (60% of the soybean growing season) was included in our analysis. Only one study – Betzelberger *et al.* (2012) – required conversion of the AOT40 to the M7, and this study used an exposure duration of 3 months. The process of conversion to the M7 metric had the potential to introduce some error into our data set, which we tested for in our statistical analysis.

As with O_3 concentration, yield was reported in the literature using a range of different metrics, and the control O_3 concentration varied considerably between the different experiments. Yield data were therefore standardized following the method described by Fuhrer *et al.* (1997). For each separate O_3 exposure experiment, linear regression was used to determine the theoretical yield at 0 ppb O_3 , expressed as the M7 metric. In a second step, the theoretical yield at 0 ppb O_3 was used as the reference (relative yield of 1) for calculating relative yields. The range of theoretical yields at 0 ppb O_3 for each study is included in Table 1.

Derivation of species and cultivar dose–response functions

All statistical analyses were carried out using R software (R Development Core Team, 2015). To calculate the overall dose–response function for soybean, relative yield data from all studies which met the inclusion criteria were pooled and plotted against the seasonal M7. The shape of the distribution was determined by fitting linear, quadratic and Weibull functions to the combined dose–response data set. Goodness of fit of the model best-fit lines was compared by eye and using the Akaike information criterion (AIC). The linear model was found to be the best fit to the data (AIC values for linear, quadratic and Weibull models are reported in the results section). Linear modelling was therefore chosen as the method to be used in the derivation of independent dose–response functions for individual soybean cultivars which had three or more supporting data points. A mixed model was used when deriving the overall dose–response function for soybean, and in the derivation of individual cultivar dose–response functions, with experimental study included as a random effect to account for the nonindependence of data points originating from the same study. During model fitting, the intercept was allowed to vary and was not forced through a relative yield value of 1. This decision was made to better allow for comparisons of the O_3 sensitivity of the different soybean cultivars based on their dose–response functions. Allowing the intercept to vary around 1 did not result in any systematic bias in the calculated slopes of the dose–response functions (See Appendix S2).

Analysis of the effect of cultivar release date, country of study and fumigation method on O₃ sensitivity

Stepwise model selection was used to determine whether the cultivar release date, country of data collection and method of O₃ fumigation were important explanatory variables in the model describing the response of soybean to O₃. A fourth explanatory variable describing whether the O₃ concentration values had been reported as the M7 or had been converted was also included, to test for bias in the data introduced through standardization to the M7 metric. A mixed-effect model structure was used to allow experimental study to act as a random effect. Model fit was assessed using Akaike's information criterion (AIC), a goodness-of-fit parameter calculated from the number of fitted parameters in a model and the maximum likelihood estimate (Symonds & Moussalli, 2011). Cultivar release dates were taken from Specht & Williams (1984), the USDA Germplasm database (USDA, 2015) and the Indian Council of Agricultural Research Oilseed report (ICAR, 2012). Data transformation of the response variable (relative yield) was carried out before analysis by taking the base-10 logarithm, to correct for non-normality observed in model residuals.

Before beginning the analysis, a diagnostic test was carried out on the data set to test the degree of collinearity between the explanatory variables. The presence of collinearity can be a concern in multiple regression due to difficulties differentiating the separate influence of variables that are partially correlated with each other (Belsley *et al.*, 1980). The variance inflation factor (VIF), a widely used measure of the degree of collinearity of independent variables in a regression model (O'Brien, 2007), was calculated for each explanatory variable (See Appendix S3 and Table S1). Calculated VIF values ranged from 1.1 to 6.1, falling well below the value of 10 considered to be a threshold above which it is recommended that measures are taken to counter the effects of collinearity (Mason & Perreault, 1991; Smith *et al.*, 2009). The diagnostic test, however, reveals the presence of a certain degree of collinearity in the data, meaning that we cannot with complete certainty rank the explanatory variables in order of their relative importance. Nevertheless, we are able to identify which of the candidate explanatory variables are likely to be important in describing the dose–response of soybean to O₃.

Multivariate regression analysis was stepwise and began with the simplest model (yield ~ O₃), with variables sequentially added to create a more complex model, and goodness-of-fit assessment at each step to determine whether variables should be kept or removed. The order of variable addition was determined by adding each explanatory variable individually to the simplest model, to identify the single variable which gave the greatest improvement to model fit; this model was then carried forward and the process was repeated until the best model was found. A complete list of all the model configurations tested during stepwise selection is given in Appendix S4 and Table S2 of the supporting information.

Candidate explanatory variables which were present in the 'best' model describing the response of soybean yield to O₃ were investigated further by subsequent graph plotting and separate individual regression analyses, which also used a mixed model structure.

Linear regression to determine how soybean cultivar sensitivity has changed with year of cultivar release

Soybean cultivars represented in the data set by three or more data points (25 cultivars in total, 22 tested in USA and 3 tested in India – listed in Table 2) were included in a separate linear regression analysis to determine whether cultivar sensitivity (represented by the slope of the dose–response function) was related to the year of cultivar release. The regression analysis was carried out twice, once on all cultivars and once excluding the cultivars from India, to ensure that any geographical differences in sensitivity were not biasing the observed relationship between sensitivity and year of release.

Reporting yield reductions predicted by dose–response functions

The standardization of reported yield data from the literature was achieved by scaling all data to yield at 0 ppb O₃. However, when reporting the yield reductions predicted by our dose–response functions in the results and discussion sections of this manuscript, we reasoned that it would be more useful to express yield reductions relative to the naturally occurring background O₃ concentration. Yield reduction estimates presented in the results and discussion of this manuscript have therefore been calculated relative to pre-industrial O₃ levels in Europe, which are thought to have averaged around 20 ppb M24, or 23 ppb M7 (Vingarzan, 2004). The O₃ concentration used to represent present-day background levels was 55 ppb M7 – a background concentration which has been commonly exceeded in the last 20 years across different world regions (Jaffe & Ray, 2007; Wang *et al.*, 2007; Chakraborty *et al.*, 2015). Relative yield reduction at the present-day O₃ concentration relative to the pre-industrial concentration will hereafter be referred to as RYL_{c,p} in this paper. A graphical representation of the method used to calculate RYL_{c,p} is shown in Fig. 2.

Results

The overall yield response of soybean to O₃, combined across all cultivars, regions and exposure types, is shown in Fig. 3a. Fitting quadratic and Weibull functions to the data set did not improve the model goodness of fit, suggesting that the soybean response to O₃ was linear across the range of M7 index values examined here (linear AIC = –458, quadratic AIC = –456, Weibull AIC = –453). The combined soybean dose–response function in Fig. 3, calculated using a mixed-effect model, estimates a RYL_{c,p} of 17.3%. For comparison with earlier studies, the response function for the same data set but using AOT40 as the O₃ metric is provided in Fig. 3b.

Of the 49 cultivars reported in the literature, 25 had three or more data points supporting their dose–response relationship and therefore were analysed independently using linear regression. The dose–

Table 2 Dose–response functions for individual soybean cultivars which were represented in the data set by three or more data points

Cultivar	Dose–response function	<i>P</i> -value	Number of refs	Number of data points	O ₃ exposure type	Country of study	Year of cultivar release	Cultivar growth habit	References
93B15	$Y = -0.0053x + 0.91$	<0.001	2	22	FACE	USA	2000	I	Morgan <i>et al.</i> (2006), Betzelberger <i>et al.</i> (2012)
Amsoy-71	$Y = -0.0046x + 0.99$	<0.05	1	4	OTC	USA	1972	I	Kress <i>et al.</i> (1986)
Bragg	$Y = -0.0022x + 0.97$	0.21	3	7	OTC	USA and India	1964	D	Singh <i>et al.</i> (2010), Heagle <i>et al.</i> (1991), Heagle & Letchworth (1982)
Clark	$Y = -0.0052x + 1.02$	<0.01	2	5	OTC and FACE	USA	1952	I	Betzelberger <i>et al.</i> (2010), Mulchi <i>et al.</i> (1995)
NK 9655	$Y = -0.0038x + 1.03$	0.17	2	9	OTC	USA	1989	D	Miller <i>et al.</i> (1994), Heagle <i>et al.</i> (1998)
Corsoy	$Y = -0.0049x + 1.00$	<0.001	1	5	OTC	USA	1970	I	Kress & Miller (1983)
Corsoy-79	$Y = -0.0052x + 1.04$	<0.001	3	11			1975	I	Kress <i>et al.</i> (1986), Heggstad <i>et al.</i> (1985, 1988)
Davis	$Y = -0.0045x + 0.99$	<0.001	6	36	OTC	USA	1966	D	Heagle <i>et al.</i> (1986, 1991, 1983a), Heagle & Letchworth (1982), Heagle <i>et al.</i> (1983b, 1987)
Dwight	$Y = -0.0049x + 1.00$	<0.001	2	22	FACE	USA	1997	I	Betzelberger <i>et al.</i> (2010, 2012)
Essex	$Y = -0.0043x + 1.05$	<0.001	8	36	OTC	USA	1972	D	Mulchi <i>et al.</i> (1988), Fiscus <i>et al.</i> (1997), Booker <i>et al.</i> (2005), Chernikova <i>et al.</i> (2000), Robinson & Britz (2000), Miller <i>et al.</i> (1994), Heagle <i>et al.</i> (1998), Heggstad & Lesser (1990)
Forrest	$Y = -0.0046x + 1.02$	<0.01	7	17	OTC	USA	1972	D	Mulchi <i>et al.</i> (1988), Chernikova <i>et al.</i> (2000), Heggstad <i>et al.</i> (1985), Robinson & Britz (2000), Heagle <i>et al.</i> (1991), Heggstad & Lesser (1990), Heagle & Letchworth (1982)
Hodgson	$Y = -0.0038x + 1.00$	<0.001	1	5	OTC	USA	1973	I	Kohut <i>et al.</i> (1986)
Holladay	$Y = -0.0054x + 0.99$	0.11	1	3	OTC	USA	1993	D	Heagle <i>et al.</i> (1998)
HS93-4118	$Y = -0.0057x + 1.05$	<0.001	2	22	FACE	USA	2000	I	Betzelberger <i>et al.</i> (2010, 2012)
IA-3010	$Y = -0.0045x + 0.97$	<0.001	2	21	FACE	USA	1998	U	Betzelberger <i>et al.</i> (2010, 2012)
LN97-15076	$Y = -0.0054x + 1.04$	<0.001	2	22	FACE	USA	2003	I	Betzelberger <i>et al.</i> (2010, 2012)
Loda	$Y = -0.0059x + 1.04$	<0.001	2	23	FACE	USA	2000	I	Betzelberger <i>et al.</i> (2010, 2012)
Pana	$Y = -0.0059x + 1.03$	<0.001	2	21	FACE	USA	1997	I	Betzelberger <i>et al.</i> (2010, 2012)
PK472	$Y = -0.0065x + 1.00$	0.068	1	3	OTC	India	1986	D	Singh <i>et al.</i> (2010)
Pusa 9712	$Y = -0.0083x + 1.00$	0.051	1	3	OTC	India	2005	D	Singh & Agrawal (2011)
Pusa 9814	$Y = -0.0093x + 1.00$	<0.05	1	3	OTC	India	2006	D	Singh & Agrawal (2011)
Ransom	$Y = -0.0036x + 1.00$	0.19	2	4	OTC	USA	1973	D	Heagle <i>et al.</i> (1991), Heagle & Letchworth (1982)
S53-34	$Y = -0.0054x + 0.96$	<0.001	1	6	OTC	USA	1980	D	Miller <i>et al.</i> (1994)
Williams-79	$Y = -0.0047x + 1.01$	<0.001	3	18	OTC	USA	1978	I	Heggstad <i>et al.</i> (1985, 1988), Heggstad & Lesser (1990)
Young	$Y = -0.0044x + 1.00$	<0.05	1	5	OTC	USA	1987	D	Miller <i>et al.</i> (1989)

Significant *P*-values are highlighted in bold. Study reference, type of O₃ fumigation used, country in which data were collected, release year and growth habit (D = determinate, I = indeterminate, U = unknown) of each cultivar are also shown. Growth habit information for the soybean cultivars was derived either from the respective dose–response papers, or from the USDA Germplasm database (USDA, 2015).

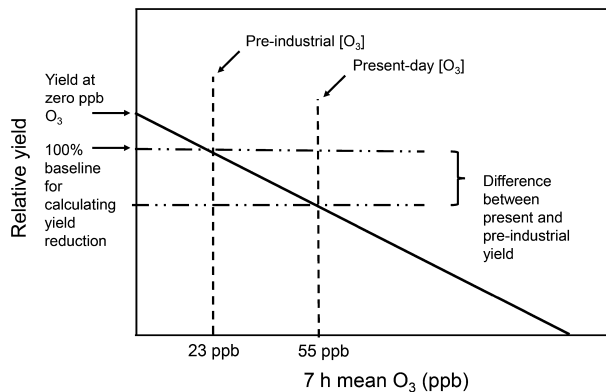


Fig. 2 Diagram illustrating how % relative yield reduction estimates reported in the results and discussion of this manuscript were calculated. Pre-industrial yield, predicted by the dose–response function, was treated as the 100% baseline yield (relative yield = 1), relative to which yields at present-day O_3 concentrations were expressed.

response functions for these 25 cultivars are shown in Table 2. Nineteen cultivars exhibited a statistically significant decline in yield with increasing O_3 concentration. Within those 19 cultivars, sensitivity to O_3 varied widely, with $RYL_{c,p}$ ranging from 13.3% for the least sensitive cultivar, ‘Hodgson’, to 37.9% for the most sensitive cultivar ‘Pusa 9814’. The three most sensitive cultivars in the data set – ‘PK472’, ‘Pusa 9712’ and ‘Pusa 9814’ – were from India. The most recently released USA cultivar in the data set, ‘LN97-15076’ released in 2003, exhibited a $RYL_{c,p}$ of 18.8%.

The AIC values for all of the different model configurations tested in the stepwise multiple regression analysis are reported in Appendix S4 and Table S2 of the supporting information. The model that performed best in describing the response of soybean to O_3 included the year of cultivar release, country of study and type of O_3 exposure as interacting variables. The AIC value for the best model shows a far greater model fit when compared to the simple model of relative yield versus O_3 concentration (delta-AIC = 42.1). It is therefore likely that the year of cultivar release, country of study and type of exposure all have some separate influence on the sensitivity of the response of soybean to O_3 . The presence of some collinearity between the candidate explanatory variables, and the observation that many of the AIC values representing different model configurations are very similar, means that we should be cautious when trying to rank the variables in order of influence. The metric conversion variable was not present in the ‘best’ model, and it is therefore likely that only minimal error was introduced to the data set through O_3 concentration metric conversions.

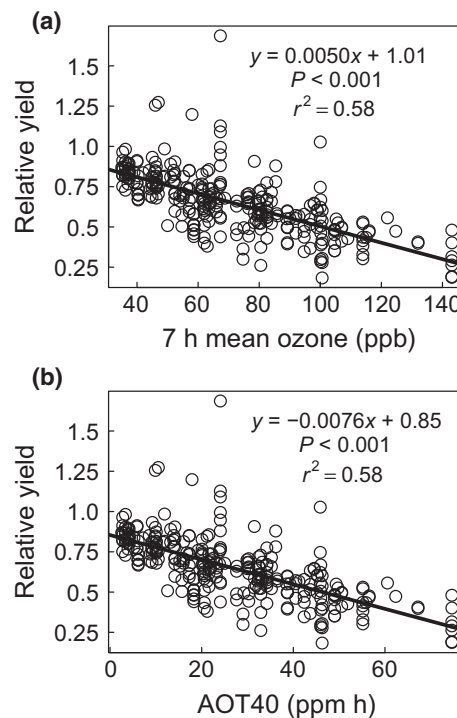


Fig. 3 Dose–response functions for soybean and O_3 , expressed using (a) 7 h mean O_3 (ppb) and (b) seasonal AOT40 (ppm h). Data comprise 379 data points from 28 studies. The regression equations and P -values describing the mixed-effect models are displayed on the two plots. The r^2 values displayed on the plots are derived from simple linear regressions fitted to the same data sets; these are included here to aid in visual interpretation of model fit.

The effect of country of study on soybean sensitivity to O_3 was investigated further by fitting separate regression lines to the combined dose–response data set according to country. Dose–response data from Indian and Chinese studies were seen to exhibit a steeper decline in yield with increasing O_3 concentration than the data from the USA (Fig. 4a). The response function based on USA data alone predicts a $RYL_{c,p}$ of 16.5%, relative to pre-industrial levels. The Indian and Chinese functions predict a $RYL_{c,p}$ of 30.3% and 33.3%, respectively. The interaction between O_3 concentration and country was highly statistically significant in a separate regression analysis carried out to investigate the individual country effect ($P = 0.0015$, $F = 6.625$, $df = 348$). There was no significant difference between the dose–response functions for India and China ($P = 0.79$ when the India- O_3 and China- O_3 interactions are compared). Their data was therefore combined to produce a more robust ‘Asia’ function based on more data points (Fig. 4b).

The individual effect of exposure method on the observed sensitivity of soybean to O_3 was also

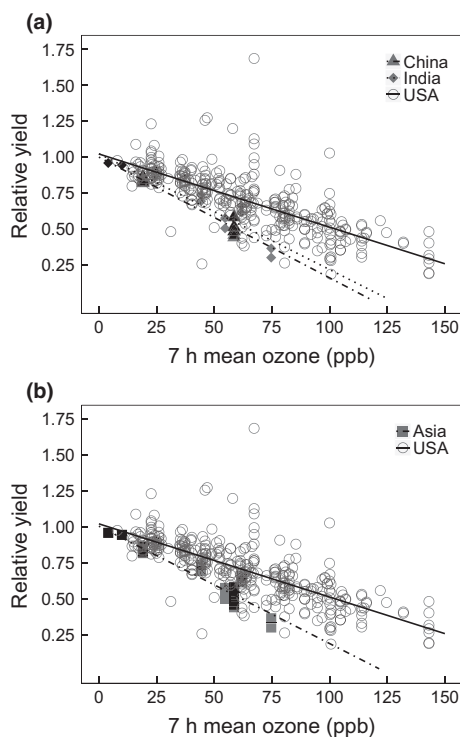


Fig. 4 (a) Subdivision of soybean dose–response data by the country in which data collection took place and (b) with the data for China and India combined into one dose–response function ('Asia'). Dose–response functions are as follows: USA, $y = -0.0047x + 1.020$ ($df = 323$, $P < 0.001$). India, $y = -0.0079x + 1.015$ ($df = 9$, $P < 0.001$). China, $y = -0.0084x + 1.00$ ($df = 16$, $P < 0.001$). Asia, $y = -0.0081x + 1.01$ ($df = 26$, $P < 0.001$)

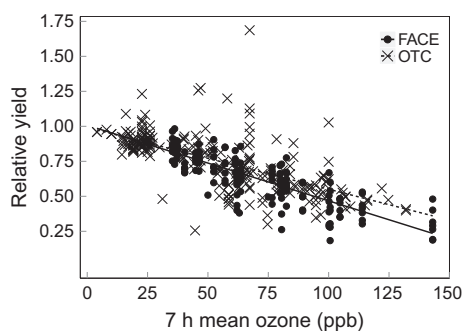


Fig. 5 Plot showing regression lines when data are subdivided by the exposure method. Dose–response functions are as follows: OTC, $y = -0.0045x + 1.00$. FACE, $y = -0.0053x + 0.97$.

investigated. Data from FACE experiments were seen to exhibit a steeper dose–response relationship than data collected in OTC's (Fig. 5). A linear regression analysis to investigate the individual effect of exposure type found the interaction of exposure type and O_3 concentration to be of borderline statistical significance ($P = 0.048$, $F = 3.93$, $df = 364.17$).

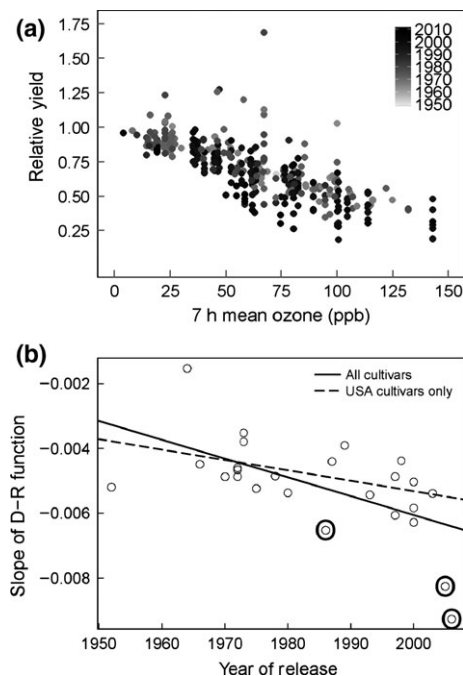


Fig. 6 (a) Greyscale gradient plot showing the time of release of cultivars in the combined data set. (b) Dose–response slope of 25 soybean cultivars expressed using the M7 metric, plotted against the year in which they were released to market. Two regression lines are shown: one which has been fitted to all cultivars ($df = 23$, $P = 0.0019$, $r^2 = 0.32$), and one which has been fitted to cultivars tested in the USA only ($df = 20$, $P = 0.0271$, $r^2 = 0.18$), excluding the data for Indian cultivars which are circled. Linear equation for all cultivars: $y = -0.000058x + 0.11$. USA-only linear equation: $y = -0.000032x + 0.06$.

Figure 6a distinguishes the data points in the combined soybean dose–response data set by the decade of cultivar release. Modern cultivars, represented on the plot by darker colours, tend to represent the steeper side of the dose–response distribution. A separate linear regression analysis on the 25 soybean cultivars with three or more supporting data points showed that cultivar sensitivity to O_3 has increased over time (Fig. 6b). The regression analysis was carried out twice, once with and once without the Indian cultivars. The sensitivity–time function comprising data exclusively from the USA is the one that avoids the possibility of bias due to geographical differences in sensitivity. This function estimates that the average slope of the soybean dose–response relationship would have been -0.0040 in 1960 and -0.0053 in 2000, representing an increase in the dose–response slope of 32.5%, over a period of 40 years.

Discussion

The combined dose–response function for soybean in Fig. 3 predicts similar yield reductions at current O_3

levels as previously published functions. $RYL_{c,p}$ is estimated to be 17.2% using our function, compared to 16.2% and 18.9% predicted by the functions of Mills *et al.* (2007) and Lesser *et al.* (1990), respectively. However, the dose–response relationship presented in this manuscript is linear, with 100% relative yield occurring at a theoretical background O_3 M7 value of zero. This is in contrast to the Mills *et al.* (2007) function which is based on the AOT40 metric and therefore assumes that O_3 concentrations below 40 ppb are not contributing to effects. The dose–response function for soybean published by Lesser *et al.* (1990) is in Weibull form and is therefore nonlinear, although the curve is very slight and much closer to a linear model when compared to other crop dose–response functions calculated from the NCLAN experiments (Wang & Mauzerall, 2004). Both of the previously published soybean dose–response functions are based only on data from the USA and do not include any data published after 1998. Our dose–response function is therefore the most comprehensive published to date and predicts that some soybean yield reduction will occur even at low concentrations of ambient O_3 , consistent with the previously published Weibull function for soybean (Lesser *et al.*, 1990).

The critical level for soybean – defined as the O_3 concentration threshold at which statistically significant yield reduction (5%) can be observed (Mills *et al.*, 2007) – is predicted using our dose–response function to be 32.3 ppb M7, when calculated relative to pre-industrial O_3 levels (M7 of 23 ppb). This is in line with the 32.4 ppb M7 critical level estimated by the function of Lesser *et al.* (1990) but a lower estimate than the 40.3 ppb M7 level predicted by the function in Mills *et al.* (2007), when both are converted to the M7 metric using the conversion functions presented in Fig. 1. The dose–response functions presented in this manuscript for India and China predict slightly lower critical levels of 28.3 ppb and 27.8 ppb M7, respectively.

Further analysis of cultivar sensitivity within the dose–response data set has revealed several important trends. The first is the significant positive correlation observed between soybean cultivar sensitivity and the year of release. Based on the sensitivity–time relationship calculated from the USA cultivars only, O_3 -induced $RYL_{c,p}$ is estimated to be on average 14.1% for cultivars released in 1960, compared to 19.3% for cultivars released in 2000. This change in cultivar sensitivity is considered to be a conservative estimate. The sensitivity–time relationship which includes the Indian cultivars estimates a greater change in cultivar sensitivity over time, with $RYL_{c,p}$ increasing from 13.1% in 1960 to 22.6% in 2000. However, this steeper sensitivity–time function incorporating the Asian cultivars could be artificially steep if differences in sensitivity due to

geographical location are also influencing the values. The trend we have identified in cultivar sensitivity to O_3 over time is in line with the results of a number of studies conducted for wheat, which found modern wheat cultivars to have greater O_3 -sensitivity than older ones (Barnes *et al.*, 1990; Velissariou *et al.*, 1992; Pleijel *et al.*, 2006; Biswas *et al.*, 2008), although this study is to our knowledge the first evidence of this phenomenon in soybean.

The mechanism underlying this temporal trend in sensitivity is unclear, although it may be linked to varietal improvement strategies. Selective breeding across different world regions has transformed the agronomic characteristics of soybean cultivars over the last half-century (Morrison *et al.*, 2000; Jin *et al.*, 2010; Agarwal *et al.*, 2013; Koester *et al.*, 2014; Rincker *et al.*, 2014). As well as having dramatically higher seed yield, modern varieties also have higher net photosynthetic rate, chlorophyll content and transpiration rate and have lower leaf area index and shorter maturation periods compared to older varieties (Liu *et al.*, 2012; Miladinović *et al.*, 2015). It is possible that agronomic traits which have been targeted by crop breeders are mechanistically linked to physiological traits associated with O_3 sensitivity, such as a low antioxidative capacity and high g_{max} (Fiscus *et al.*, 2005; Biswas *et al.*, 2008). For example, selection for high yield could have simultaneously targeted a high g_{max} to facilitate greater CO_2 fixation (Roche, 2015). This hypothesis is supported by results from a study on 24 soybean cultivars with release dates spanning 1923 to 2007, which observed an increase in g_s with year of release in cultivars which also exhibited increasing instantaneous rates of carbon uptake with year of release (Koester *et al.*, 2014). The g_s of wheat cultivars has also been reported to progressively increase with their year of release and correlates positively with O_3 sensitivity (Biswas *et al.*, 2008). Breeding for a high harvest index and rapid maturation over recent decades (Morrison *et al.*, 2000; Jin *et al.*, 2010) may have also played a role in the greater O_3 sensitivity of modern cultivars of soybean, by selecting for a trade-off which prioritizes vegetative and reproductive growth over antioxidant synthesis, which could be associated with a metabolic cost under O_3 enriched conditions (Huot *et al.*, 2014; Frei, 2015).

A net increase in the yield of soybean cultivars has taken place over recent decades despite their increasing sensitivity to O_3 . The heterogeneity of O_3 concentrations temporally and geographically may explain the lack of sufficient natural selection pressure for O_3 tolerance at cultivar breeding sites (Ainsworth *et al.*, 2008). Cultivar breeding programmes focussing on enhancing the ability of varieties to detoxify O_3 would increase tolerance and improve yield further (Frei, 2015). Another

approach for breeding O_3 tolerance would be to select for reduced g_{max} to reduce the rate of O_3 flux into the plant, and faster stomatal dynamics to allow leaves to close their stomata more rapidly in response to O_3 stress (Morgan *et al.*, 2003). While the reduction in photosynthetic gas exchange associated with excluding O_3 could result in a small yield penalty during less polluted years, cultivars with reduced g_{max} would likely perform better during years with high levels of air pollution, perhaps resulting in an average yield gain over time. A similar strategy in soybean with drought tolerant traits has shown early signs of success, with a 50-year simulation based on US weather data showing a significant improvement in average yields, despite some of the traits being detrimental in wet years (Sinclair *et al.*, 2010).

A second important pattern identified in the data analysis relates to the observed geographical variation in O_3 sensitivity. A steeper decline in soybean yield with increasing O_3 was observed in experimental data collected in India and China, compared to data from the USA. Unfortunately, a limited amount of dose–response data was available for the Asian region: two studies from India and one from China met the inclusion criteria for analysis, with Asian cultivars comprising 12 of the 49 cultivars and 30 of the 379 data points included in the complete data set. Despite the small number of data points representing the Asian region in our analysis, the interaction between O_3 concentration and country of data collection exhibited a high level of statistical significance in the individual regression describing the variation in soybean yield response to O_3 ($P = 0.0015$), and country of study emerged as an important variable in the stepwise multiple regression.

The greater sensitivity observed in the Asian data suggests that the use of region-specific dose–response functions could potentially improve the accuracy of modelled crop loss estimates. It also highlights the urgent need for more O_3 exposure studies in India and China, which are currently significantly underrepresented in the dose–response literature compared to the USA. Historical and contemporary O_3 trends in India and China are not well documented (Cooper *et al.*, 2014), but both countries have seen a rapid increase in emissions of O_3 precursors as a result of rapid urbanization and industrialization (Granier *et al.*, 2011) and are likely to experience significant increases in surface O_3 concentrations by 2050 (Fiore *et al.*, 2012). Ozone modelling in South Asia by Engardt (2008) based on emissions for the year 2000 estimated surface O_3 concentrations during the soybean growing season (September–November) to be 40–45 ppb M7 over large areas of the state of Maharashtra, which produces over 30% of India's total soybean crop (DAC, 2014). The

dose–response function combined across all regions predicts relative yield reduction at this O_3 concentration to be 9.2–11.9% relative to pre-industrial levels, while the India-specific response function estimates yield reduction to be 16.2–20.9% – a large discrepancy of estimation. Over large areas of the agriculturally important Indo-Gangetic plain where soybean is also grown (Singh, 2006), modelled surface O_3 exceeds 49 ppb M7, with soybean yield reduction here estimated to be 24.8% using our Indian response function. Accurate estimates of potential O_3 effects on crop yield is arguably particularly important for the South Asian region, where 21% of the population are currently undernourished, an estimated 51% of soybean cropland is reported to be experiencing stagnating or declining yields (Ray *et al.*, 2012), and average soybean yield per hectare is less than half that in the USA (Panthee, 2010).

The mechanism underlying the differential sensitivity observed in North American and Asian dose–response data is unclear. Interestingly, greater O_3 sensitivity of Asian cultivars compared to North American ones has been previously observed in wheat and rice (Emberson *et al.*, 2009). Differences between the climate and environment of the different geographical regions could be one factor driving the observed difference in sensitivity. Large areas of China and India experience a humid subtropical climate (Rubel & Kottek, 2010), which facilitates high g_s and therefore high O_3 flux. Similarly, warm temperatures correlate with high g_s up to a species-specific optimum temperature, above which conductance falls (Emberson *et al.*, 2000). The co-occurrence of O_3 concentration peaks with periods of high humidity and optimum temperature – which follow seasonal and diurnal patterns specific to geographical regions – could therefore be a significant factor in determining the degree of crop loss. Unfortunately, the wide range of study locations, open-air experimental designs and seasonal duration of experiments included in this analysis meant that humidity and temperature could not be investigated when synthesizing the data.

Another important factor which must be considered when interpreting the Asia data is the possibility of interactions between O_3 and other ambient air pollutants. There is some evidence that the simultaneous or sequential occurrence of O_3 with SO_2 , NO_2 and NH_3 can have a greater than additive effect on the yield of crops (Fangmeier & Bender, 2002; Bender & Weigel, 2011). Two of the three experimental studies included in our analysis which took place in Asia added O_3 to nonfiltered air, and concentrations of other ambient air pollutants were not recorded during these experiments. The potential for O_3 interactions with other pollutants means we must interpret the higher sensitivity of soybean observed in the Asian studies with some caution.

However, all of the data points collected in Asia – including those from the experiment which added O₃ to carbon-filtered air (Singh *et al.*, 2010) and therefore removed other ambient pollutants – lie below the dose–response line fitted to USA-only data (Fig. S4), suggesting that multipollutant interactions are not the sole driver of the greater sensitivity of Asian dose–response data.

As discussed earlier in relation to temporal trends, crop breeding strategies may be partly driving the observed regional differences in O₃ sensitivity. Crop breeding strategies in the USA, China and India over the last half-century have shared the common aim of increasing yield and harvest index (Karmakar & Bhatnagar, 1996; Morrison *et al.*, 2000; Jin *et al.*, 2010), but other breeding targets are likely to have varied by region. For example, the high sensitivity of soybean to day-length means that maturation periods are highly tailored for different latitudes (Agarwal *et al.*, 2013). In addition, region-specific efforts to breed resistance to local diseases or pests could have increased the capacity of cultivars to upregulate antioxidants, potentially increasing their tolerance to O₃ (Bowler *et al.*, 1992).

The third key result from the data analysis is that the sensitivity of soybean cultivars to O₃ varies widely, and varieties introduced at a similar time and from the same geographical region also exhibit a certain degree of variation in sensitivity. For example, ‘Corsoy-79’ and ‘Hodgson’ – both released in the USA in the same decade (1970s) – are predicted using the functions calculated in this study to experience a RYL_{c,p} of 18.1% and 13.3%, respectively, relative to pre-industrial O₃. A wide range of within-species variation in O₃ sensitivity has been observed before in other crop species. Quarrie *et al.* (2007) studied 95 wheat cultivars and observed yield reduction ranging from 0% to 56% following season-long O₃ exposure at an M7 of 91 ppb. Further evidence of differential cultivar sensitivity in wheat has come from studies on Chinese (Biswas *et al.*, 2008) and Bangladeshi (Saitanis *et al.*, 2014) varieties. A similar range of sensitivity has also been observed in Thai rice cultivars (Ariyaphanphitak *et al.*, 2005). The variation in O₃ sensitivity among cultivars observed in this study suggests that there is substantial scope for breeding O₃-tolerant soybean varieties.

The difference in the yield response observed in FACE and OTC’s should be interpreted with caution, due to the marginal *P*-value in the individual regression (*P* = 0.048), and the presence of some collinearity. FACE data exhibited a marginally steeper dose–response slope compared to data collected in OTC’s. This result indicates that both methods of exposure produce dose–response data that is comparable and that the impact of the ‘chamber effect’ – the alteration

of the growth environment in OTC’s which can lead to heightened temperatures, altered air flow and greater vapour pressure deficit (Sanders *et al.*, 1991; Long *et al.*, 2005) – on the soybean yield response to O₃ is only small, if it exists. More work is needed to confirm or reject the possibility that exposure method impacts the yield response of crops in O₃ exposure studies.

In conclusion, this study has revealed a large degree of intercultivar variation in soybean O₃ sensitivity and has also identified temporal and geographical patterns in sensitivity. These patterns are relevant to efforts in breeding O₃-tolerant crop cultivars and also to those carrying out global modelling assessments of O₃ impacts on crop yield. This manuscript has discussed potential factors which might be playing a role in driving these patterns, but they are not yet fully understood. The derivation of flux-based dose–response relationships for soybean, which estimate O₃ exposure based on known relationships between climatic conditions and g_s (Emberson *et al.*, 2000), could shed light on the hypothesis that local climatic factors and particular physiological traits related to gas exchange are driving the observed regional and temporal patterns in sensitivity.

Acknowledgements

We would like to thank Dr. David Cooper for his valuable advice regarding statistical analyses, and The Natural Environment Research Council of the United Kingdom for funding this work.

References

- Agarwal DK, Billore S, Sharma A, Dupare B, Srivastava S (2013) Soybean: introduction, improvement, and utilization in India—problems and prospects. *Agricultural Research*, **2**, 293–300.
- Ainsworth EA, Rogers A, Leakey ADB (2008) Targets for crop biotechnology in a future high-CO₂ and high-O₃ world. *Plant Physiology*, **147**, 13–19.
- Ariyaphanphitak W, Chidthaisong A, Sarobol E, Bashkin VN, Towprayoon S (2005) Effects of elevated ozone concentrations on Thai Jasmine rice cultivars (*Oryza sativa* L.). *Water Air and Soil Pollution*, **167**, 179–200.
- Avnery S, Mauzerall DL, Liu JF, Horowitz LW (2011a) Global crop yield reductions due to surface ozone exposure: 1. Year 2000 crop production losses and economic damage. *Atmospheric Environment*, **45**, 2284–2296.
- Avnery S, Mauzerall DL, Liu JF, Horowitz LW (2011b) Global crop yield reductions due to surface ozone exposure: 2. Year 2030 potential crop production losses and economic damage under two scenarios of O₃ pollution. *Atmospheric Environment*, **45**, 2297–2309.
- Barnes J, Velissariou D, Davison A, Holevas C (1990) Comparative ozone sensitivity of old and modern Greek cultivars of spring wheat. *New Phytologist*, **116**, 707–714.
- Beig G, Singh V (2007) Trends in tropical tropospheric column ozone from satellite data and MOZART model. *Geophysical Research Letters*, **34**, L17801. doi: 10.1029/2007GL030460
- Belsley DA, Kuh E, Welsch RE (1980) *Regression Diagnostics: Identifying Influential Data and Sources of Collinearity*. Wiley, New York.
- Bender J, Weigel HJ (2011) Changes in atmospheric chemistry and crop health: a review. *Agronomy for Sustainable Development*, **31**, 81–89.
- Betzberger AM, Gillespie KM, McGrath JM, Koester RP, Nelson RL, Ainsworth EA (2010) Effects of chronic elevated ozone concentration on antioxidant capacity, photosynthesis and seed yield of 10 soybean cultivars. *Plant Cell and Environment*, **33**, 1569–1581.

- Betzberger AM, Yendrek CR, Sun JD, Leisner CP, Nelson RL, Ort DR, Ainsworth EA (2012) Ozone exposure response for U.S. soybean cultivars: linear reductions in photosynthetic potential, biomass, and yield. *Plant Physiology*, **160**, 1827–1839.
- Biswas DK, Xu H, Li YG, Sun JZ, Wang XZ, Han XG, Jiang GM (2008) Genotypic differences in leaf biochemical, physiological and growth responses to ozone in 20 winter wheat cultivars released over the past 60 years. *Global Change Biology*, **14**, 46–59.
- Booker FL, Miller JE, Fiscus EL, Pursley WA, Stefanski LA (2005) Comparative responses of container- versus ground-grown soybean to elevated carbon dioxide and ozone. *Crop Science*, **45**, 883–895.
- Bou Jaoudé M, Katerji N, Mastrorilli M, Rana G (2008) Analysis of the ozone effect on soybean in the Mediterranean region: II. The consequences on growth, yield and water use efficiency. *European Journal of Agronomy*, **28**, 519–525.
- Bowler C, Vanmontagu M, Inze D (1992) Superoxide-dismutase and stress tolerance. *Annual Review of Plant Physiology and Plant Molecular Biology*, **43**, 83–116.
- Brasseur G, Müller J, Tie X, Horowitz L (2001) Tropospheric Ozone and Climate: Past, Present and Future. In: *Present and Future of Modeling Global Environmental Change: Toward Integrated Modeling*. (ed. Matsuno T, Kida H), pp. 63–76. Terrapub, Tokyo.
- Chakraborty T, Beig G, Dentener FJ, Wild O (2015) Atmospheric transport of ozone between Southern and Eastern Asia. *Science of the Total Environment*, **523**, 28–39.
- Chernikova T, Robinson JM, Lee EH, Mulchi CL (2000) Ozone tolerance and antioxidant enzyme activity in soybean cultivars. *Photosynthesis Research*, **64**, 15–26.
- Cooper OR, Parrish D, Ziemke J *et al.* (2014) Global distribution and trends of tropospheric ozone: an observation-based review. *Elementa: Science of the Anthropocene*, **2**, 000029.
- DAC (2014) *Agricultural Statistics at a Glance 2014*. Ministry of Agriculture, Department of Agriculture and Cooperation, Directorate of Economics and Statistics, New Delhi.
- Embersson LD, Ashmore MR, Cambridge HM, Simpson D, Tuovinen JP (2000) Modeling stomatal ozone flux across Europe. *Environmental Pollution*, **109**, 403–413.
- Embersson LD, Buker P, Ashmore MR *et al.* (2009) A comparison of North American and Asian exposure-response data for ozone effects on crop yields. *Atmospheric Environment*, **43**, 1945–1953.
- Engardt M (2008) Modelling of near-surface ozone over South Asia. *Journal of Atmospheric Chemistry*, **59**, 61–80.
- Falkenmark M (2013) Growing water scarcity in agriculture: future challenge to global water security. *Philosophical Transactions of the Royal Society A: Mathematical Physical and Engineering Sciences*, **371**, 20120410. doi: 10.1098/rsta.2012.0410
- Fangmeier A, Bender J (2002) Air pollutant combinations – Significance for future impact assessments on vegetation. *Phyton: Horn*, **42**, 65–72.
- FAO (2011) *The State of the World's Land and Water Resources for Food and Agriculture (SOLAW) – Managing Systems at Risk*. Rome and Earthscan, London.
- FAO (2012) Agricultural production and natural resource use. In: *World Agriculture Towards 2030/2050: The 2012 Revision*. ESA Working Paper No. 12-03. (ed. Alexandratos N, Bruinsma J), pp. 94–133. FAO, Rome.
- FAO (2014) FAOSTAT. FAO, Rome, Italy.
- FAO (2015) *The State of Food Insecurity in the World 2015*. Meeting the 2015 international hunger targets: taking stock of uneven progress. FAO, Rome, Italy.
- Fiore AM, Naik V, Spracklen DV *et al.* (2012) Global air quality and climate. *Chemical Society Reviews*, **41**, 6663–6683.
- Fiscus EL, Reid CD, Miller JE, Heagle AS (1997) Elevated CO₂ reduces O₃ flux and O₃-induced yield losses in soybeans: possible implications for elevated CO₂ studies. *Journal of Experimental Botany*, **48**, 307–313.
- Fiscus EL, Booker FL, Burkey KO (2005) Crop responses to ozone: uptake, modes of action, carbon assimilation and partitioning. *Plant Cell and Environment*, **28**, 997–1011.
- Frei M (2015) Breeding of ozone resistant rice: relevance, approaches and challenges. *Environmental Pollution*, **197**, 144–155.
- Frei M, Tanaka JP, Wissuwa M (2008) Genotypic variation in tolerance to elevated ozone in rice: dissection of distinct genetic factors linked to tolerance mechanisms. *Journal of Experimental Botany*, **59**, 3741–3752.
- Frei M, Tanaka JP, Chen CP, Wissuwa M (2010) Mechanisms of ozone tolerance in rice: characterization of two QTLs affecting leaf bronzing by gene expression profiling and biochemical analyses. *Journal of Experimental Botany*, **61**, 1405–1417.
- Fuhrer J (2009) Ozone risk for crops and pastures in present and future climates. *Naturwissenschaften*, **96**, 173–194.
- Fuhrer J, Skarby L, Ashmore MR (1997) Critical levels for ozone effects on vegetation in Europe. *Environmental Pollution*, **97**, 91–106.
- Gillespie KM, Xu FX, Richter KT *et al.* (2012) Greater antioxidant and respiratory metabolism in field-grown soybean exposed to elevated O₃ under both ambient and elevated CO₂. *Plant Cell and Environment*, **35**, 169–184.
- Glick RE, Schlagnhauser CD, Arteca RN, Pell EJ (1995) Ozone-induced ethylene emission accelerates the loss of ribulose-1,5-bisphosphate carboxylase oxygenase and nuclear-encoded messenger-rnas in senescing potato leaves. *Plant Physiology*, **109**, 891–898.
- Graham PH, Vance CP (2003) Legumes: importance and constraints to greater use. *Plant Physiology*, **131**, 872–877.
- Granier C, Bessagnet B, Bond T *et al.* (2011) Evolution of anthropogenic and biomass burning emissions of air pollutants at global and regional scales during the 1980–2010 period. *Climatic Change*, **109**, 163–190.
- Hartman GL, West ED, Herman TK (2011) Crops that feed the World 2. Soybean – worldwide production, use, and constraints caused by pathogens and pests. *Food Security*, **3**, 5–17.
- Heagle A (1989) Ozone and crop yield. *Annual Review of Phytopathology*, **27**, 397–423.
- Heagle A, Letchworth M (1982) Relationships among injury, growth, and yield responses of soybean cultivars exposed to ozone at different light intensities. *Journal of Environmental Quality*, **11**, 690–694.
- Heagle A, Heck WW, Rawlings JO, Philbeck RB (1983a) Effects of chronic doses of ozone and sulfur dioxide on injury and yield of soybeans in open-top field chambers. *Crop Science*, **23**, 1184–1191.
- Heagle A, Letchworth MB, Mitchell CA (1983b) Effects of growth medium and fertilizer rate on the yield response of soybeans exposed to chronic doses of ozone. *Phytopathology*, **73**, 134–139.
- Heagle A, Lesser VM, Rawlings JO, Heck WW, Philbeck RB (1986) Response of soybeans to chronic doses of ozone applied as constant or proportional additions to ambient air. *Phytopathology*, **76**, 6.
- Heagle A, Flagler RB, Patterson RP, Lesser VM, Shafer SR, Heck WW (1987) Injury and yield response of soybean to chronic doses of ozone and soil-moisture deficit. *Crop Science*, **27**, 1016–1024.
- Heagle A, Miller JE, Rawlings JO, Vozzo SF (1991) Effect of growth stage on soybean response to chronic ozone exposure. *Journal of Environmental Quality*, **20**, 562–570.
- Heagle A, Miller JE, Pursley WA (1998) Influence of ozone stress on soybean response to carbon dioxide enrichment: III. Yield and seed quality. *Crop Science*, **38**, 128–134.
- Heggstad H, Lesser VM (1990) Effects of ozone, sulfur dioxide, soil water deficit, and cultivar on yields of soybean. *Journal of Environmental Quality*, **19**, 488–495.
- Heggstad H, Gish T, Lee E, Bennett J, Douglass L (1985) Interaction of soil moisture stress and ambient ozone on growth and yields of soybeans. *Phytopathology*, **75**, 472–477.
- Heggstad H, Anderson EL, Gish TJ, Lee EH (1988) Effects of ozone and soil-water deficit on roots and shoots of field-grown soybeans. *Environmental Pollution*, **50**, 259–278.
- Herridge DF, Peoples MB, Boddey RM (2008) Global inputs of biological nitrogen fixation in agricultural systems. *Plant and Soil*, **311**, 1–18.
- Hewitt D, Mills G, Hayes F, Norris D, Coyle M, Wilkinson S, Davies W (2016) N-fixation in legumes—An assessment of the potential threat posed by ozone pollution. *Environmental Pollution*, **208**, 909–918.
- Hough AM, Derwent RG (1990) Changes in the global concentration of tropospheric ozone due to human activities. *Nature*, **344**, 645–648.
- Huot B, Yao J, Montgomery BL, He SY (2014) Growth-defense tradeoffs in plants: a balancing act to optimize fitness. *Molecular Plant*, **7**, 1267–1287.
- ICAR (2012) *Recent Varieties and Hybrids of Annual Oilseeds Recommended for Different States*. Directorate of Oilseeds Development, Himayat Nagar, Hyderabad.
- IPCC (2013) Annex II: Climate System Scenario Tables. In: *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change* [Stocker, T.F., D. Qin, G.-K. Plattner, M. Tignor, S.K. Allen, J. Boschung, A. Nauels, Y. Xia, V. Bex and P.M. Midgley (eds.)]. (eds Prather M, Flato G, Friedlingstein P, Jones C, Lamarque J-F, Liao H, Rasch P), Cambridge University Press, Cambridge, UK and New York, NY, USA.
- Jaffe D, Ray J (2007) Increase in surface ozone at rural sites in the western US. *Atmospheric Environment*, **41**, 5452–5463.
- Jin J, Liu X, Wang G, Mi L, Shen Z, Chen X, Herbert SJ (2010) Agronomic and physiological contributions to the yield improvement of soybean cultivars released from 1950 to 2006 in Northeast China. *Field Crops Research*, **115**, 116–123.
- Karmakar PG, Bhatnagar PS (1996) Genetic improvement of soybean varieties released in India from 1969 to 1993. *Euphytica*, **90**, 95–103.
- Koester RP, Skoneczka JA, Cary TR, Diers BW, Ainsworth EA (2014) Historical gains in soybean (*Glycine max* Merr.) seed yield are driven by linear increases in light interception, energy conversion, and partitioning efficiencies. *Journal of Experimental Botany*, **65**, 3311–3321.
- Kohut RJ, Amundson RG, Laurence JA (1986) Evaluation of growth and yield of soybean exposed to ozone in the field. *Environmental Pollution Series A*, **41**, 219–234.

- Kress LW, Miller JE (1983) Impact of ozone on soybean yield. *Journal of Environmental Quality*, **12**, 276–281.
- Kress LW, Miller JE, Smith HJ, Rawlings JO (1986) Impact of ozone and sulfur-dioxide on soybean yield. *Environmental Pollution Series A*, **41**, 105–123.
- Krupa S, McGrath MT, Andersen CP *et al.* (2001) Ambient ozone and plant health. *Plant Disease*, **85**, 4–12.
- Lesser VM, Rawlings JO, Spruill SE, Somerville MC (1990) Ozone effects on agricultural crops – statistical methodologies and estimated dose-response relationships. *Crop Science*, **30**, 148–155.
- Liu G, Chunwu Y, Kezhang X, Zhan Z, Dayong L, Zhihai W, Zhanyu C (2012) Development of yield and some photosynthetic characteristics during 82 years of genetic improvement of soybean genotypes in northeast China. *Australian Journal of Crop Science*, **6**, 1416–1422.
- Long SP, Ainsworth EA, Leakey AD, Morgan PB (2005) Global food insecurity. Treatment of major food crops with elevated carbon dioxide or ozone under large-scale fully open-air conditions suggests recent models may have overestimated future yields. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **360**, 2011–2020.
- Mason CH, Perreault WD Jr (1991) Collinearity, power, and interpretation of multiple regression analysis. *Journal of Marketing Research*, **28**, 268–280.
- Masuda T, Goldsmith PD (2009) World soybean production: area harvested, yield, and long-term projections. *International Food and Agribusiness Management Review*, **12**, 143–162.
- Mateos-Aparicio I, Redondo-Cuenca A, Villanueva-Suárez M, Zapata-Revilla M (2008) Soybean, a promising health source. *Nutrición Hospitalaria*, **23**, 305.
- Miladinović J, Vidić M, Đorđević V, Balešević-Tubić S (2015) New trends in plant breeding – example of soybean. *Genetika*, **47**, 131–142.
- Miller J, Heagle AS, Vozzo SF, Philbeck RB, Heck WW (1989) Effects of ozone and water stress, separately and in combination, on soybean yield. *Journal of Environmental Quality*, **18**, 330–336.
- Miller J, Booker F, Fiscus E, Heagle A, Pursley W, Vozzo S, Heck W (1994) Ultraviolet-B radiation and ozone effects on growth, yield, and photosynthesis of soybean. *Journal of Environmental Quality*, **23**, 83–91.
- Mills G, Harmens H (eds) (2011) Ozone pollution: a hidden threat to food security. In: *Convention on Long-Range Transboundary Air Pollution*, pp. 43–52. Centre for Ecology and Hydrology, Bangor, UK, ICP Vegetation.
- Mills G, Buse A, Gimeno B, Bermejo V, Holland M, Emberson L, Pleijel H (2007) A synthesis of AOT40-based response functions and critical levels of ozone for agricultural and horticultural crops. *Atmospheric Environment*, **41**, 2630–2643.
- Morgan PB, Ainsworth EA, Long SP (2003) How does elevated ozone impact soybean? A meta-analysis of photosynthesis, growth and yield. *Plant Cell and Environment*, **26**, 1317–1328.
- Morgan PB, Mies TA, Bollero GA, Nelson RL, Long SP (2006) Season-long elevation of ozone concentration to projected 2050 levels under fully open-air conditions substantially decreases the growth and production of soybean. *New Phytologist*, **170**, 333–343.
- Morrison MJ, Voldeng HD, Cober ER (2000) Agronomic changes from 58 years of genetic improvement of short-season soybean cultivars in Canada. *Agronomy Journal*, **92**, 780–784.
- Mulchi C, Lee E, Tuthill K, Olinick EV (1988) Influence of ozone stress on growth processes, yields and grain quality characteristics among soybean cultivars. *Environmental Pollution*, **53**, 151–169.
- Mulchi C, Rudorff B, Lee E, Rowland R, Pausch R (1995) Morphological responses among crop species to full-season exposures to enhanced concentrations of atmospheric CO₂ and O₃. *Water Air and Soil Pollution*, **85**, 1379–1386.
- O'Brien RM (2007) A caution regarding rules of thumb for variance inflation factors. *Quality & Quantity*, **41**, 673–690.
- Panthee D (2010) Varietal improvement in soybean. In: *The Soybean: botany, production and uses* (ed. Singh G), pp. 92–112. CABI, New Delhi.
- Parrish D, Lamarque JF, Naik V *et al.* (2014) Long-term changes in lower tropospheric baseline ozone concentrations: comparing chemistry-climate models and observations at northern midlatitudes. *Journal of Geophysical Research: Atmospheres*, **119**, 5719–5736.
- Parry ML, Rosenzweig C, Iglesias A, Livermore M, Fischer G (2004) Effects of climate change on global food production under SRES emissions and socio-economic scenarios. *Global Environmental Change-Human and Policy Dimensions*, **14**, 53–67.
- Pedersen P, Lauer JG (2004) Soybean growth and development in various management systems and planting dates. *Crop Science*, **44**, 508–515.
- Pleijel H, Eriksen AB, Danielsson H, Bondesson N, Sellden G (2006) Differential ozone sensitivity in an old and a modern Swedish wheat cultivar – grain yield and quality, leaf chlorophyll and stomatal conductance. *Environmental and Experimental Botany*, **56**, 63–71.
- Quarrie S, Kaminska A, Dodmani A, Gonzalez I, Gillespie C, Bilsborrow P, Barnes J (2007) QTLs governing ozone impacts on wheat yield. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, **146**, S261.
- R Development Core Team (2015) *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Ray DK, Ramankutty N, Mueller ND, West PC, Foley JA (2012) Recent patterns of crop yield growth and stagnation. *Nature Communications*, **3**, 1293.
- Reinert RA, Eason G (2000) Genetic control of O₃ sensitivity in a cross between two cultivars of snap bean. *Journal of the American Society for Horticultural Science*, **125**, 222–227.
- Reinert R, Weber D (1980) Ozone and sulfur dioxide-induced changes in soybean growth. *Phytopathology*, **70**, 914–916.
- Rincker K, Nelson R, Specht J *et al.* (2014) Genetic improvement of US soybean in maturity groups II, III, and IV. *Crop Science*, **54**, 1419–1432.
- Robinson JM, Britz SJ (2000) Tolerance of a field grown soybean cultivar to elevated ozone level is concurrent with higher leaflet ascorbic acid level, higher ascorbate-dehydroascorbate redox status, and long term photosynthetic productivity. *Photosynthesis Research*, **64**, 77–87.
- Roche D (2015) Stomatal Conductance Is Essential for Higher Yield Potential of C3 Crops. *Critical Reviews in Plant Sciences*, **34**, 429–453.
- Royal Society (2008) *Ground-Level Ozone in the 21st Century: Future Trends, Impacts and Policy Implications*. The Royal Society, London.
- Rubel F, Kottek M (2010) Observed and projected climate shifts 1901–2100 depicted by world maps of the Köppen-Geiger climate classification. *Meteorologische Zeitschrift*, **19**, 135–141.
- Saitanis C, Bari S, Burkey K, Stamatelopoulos D, Agathokleous E (2014) Screening of Bangladeshi winter wheat (*Triticum aestivum* L.) cultivars for sensitivity to ozone. *Environmental Science and Pollution Research*, **21**, 13560–13571.
- Sanders GE, Clark AG, Colls JJ (1991) The influence of open-top chambers on the growth and development of field bean. *New Phytologist*, **117**, 439–447.
- Sinclair TR, Messina CD, Beatty A, Samples M (2010) Assessment across the United States of the benefits of altered soybean drought traits. *Agronomy Journal*, **102**, 475–482.
- Singh BB (2006) Success of soybean in India: the early challenges and pioneer promoters. *Asian Agri-History*, **10**, 41–47.
- Singh S, Agrawal SB (2011) Cultivar-specific response of soybean (*Glycine max* L.) to ambient and elevated concentrations of ozone under open top chambers. *Water Air and Soil Pollution*, **217**, 283–302.
- Singh E, Tiwari S, Agrawal M (2010) Variability in antioxidant and metabolite levels, growth and yield of two soybean varieties: an assessment of anticipated yield losses under projected elevation of ozone. *Agriculture Ecosystems & Environment*, **135**, 168–177.
- Smith AC, Koper N, Francis CM, Fahrig L (2009) Confronting collinearity: comparing methods for disentangling the effects of habitat loss and fragmentation. *Landscape Ecology*, **24**, 1271–1285.
- Specht JE, Williams JH (1984) Contribution of genetic technology to soybean productivity - retrospect and prospect. In: *Genetic Contributions to Yield gains of five major crop plants* (ed. Fehr W.R.), pp. 49–74. Crop Science Society of America and American Society of Agronomy, Madison, USA.
- Symonds MR, Moussalli A (2011) A brief guide to model selection, multimodel inference and model averaging in behavioural ecology using Akaike's information criterion. *Behavioral Ecology and Sociobiology*, **65**, 13–21.
- Tai APK, Martin MV, Heald CL (2014) Threat to future global food security from climate change and ozone air pollution. *Nature Climate Change*, **4**, 817–821.
- Tingey DT, Blum U (1973) Effects of ozone on soybean nodules. *Journal of Environmental Quality*, **2**, 341–342.
- Troiano J, Colavito L, Heller L, HcCune DC, Jacobson JS (1983) Effects of acidity of simulated rain and its joint action with ambient ozone on measures of biomass and yield in soybean. *Environmental and Experimental Botany*, **23**, 113–119.
- USDA (2015) *Germplasm Resources Information Network*. National Germplasm Resources Laboratory, Beltsville, Maryland. Available at: <http://www.ars-grin.gov/>.
- Van Dingenen R, Dentener FJ, Raes F, Krol MC, Emberson L, Cofala J (2009) The global impact of ozone on agricultural crop yields under current and future air quality legislation. *Atmospheric Environment*, **43**, 604–618.
- Velissariou D, Barnes JD, Davison AW (1992) Has inadvertent selection by plant breeders affected the O₃ sensitivity of modern greek cultivars of spring wheat? *Agriculture Ecosystems and Environment*, **38**, 79–89.

- Vingarzan R (2004) A review of surface ozone background levels and trends. *Atmospheric Environment*, **38**, 3431–3442.
- Wang X, Mauzerall DL (2004) Characterizing distributions of surface ozone and its impact on grain production in China, Japan and South Korea: 1990 and 2020. *Atmospheric Environment*, **38**, 4383–4402.
- Wang X, Manning W, Feng Z, Zhu Y (2007) Ground-level ozone in China: distribution and effects on crop yields. *Environmental Pollution*, **147**, 394–400.
- Wilkinson S, Mills G, Illidge R, Davies WJ (2012) How is ozone pollution reducing our food supply? *Journal of Experimental Botany*, **63**, 527–536.
- Zabel F, Putzenlechner B, Mauser W (2014) Global agricultural land resources – a high resolution suitability evaluation and its perspectives until 2100 under climate change conditions. *PLoS ONE*, **9**, e107522.
- Zhang WW, Wang GG, Liu XB, Feng ZZ (2014) Effects of elevated O₃ exposure on seed yield, N concentration and photosynthesis of nine soybean cultivars (*Glycine max* (L.) Merr.) in Northeast China. *Plant Science*, **226**, 172–181.
- Zhao T, Cao Y, Wang Y, Dai Z, Liu Y, Liu B (2012) Effects of ozone stress on root morphology and reactive oxygen species metabolism in soybean roots. *Soybean Science*, **1**, 013.

Supporting Information

Additional Supporting Information may be found in the online version of this article:

Appendix S1. Methods and results from comparison of the dose-response observed in pot-grown and field-grown soybean.

Figure S1. Scatterplot of soybean dose-response data, subdivided by the method of soybean cultivation (pot versus field).

Appendix S2. Comparison of cultivar dose-response slopes when calculated using explicit or free intercept.

Figure S2. Plot of dose-response slope values calculated using an explicit and free intercept.

Appendix S3. Results of a diagnostic test to assess the degree of collinearity in the dataset, presented in Table S1.

Appendix S4. Results from stepwise multiple regression analysis, with model configurations and AIC values for all tested models reported in Table S2.

Figure S3. Plot of all data points included in our combined analysis, symbol-coded to show which values had to be converted to the M7 metric.

Figure S4. Plot showing measured soybean yield values in response to O₃ exposure from experiments carried out in Asia, relative to the USA-only regression line.