



UNIVERSITY OF LEEDS

This is a repository copy of *Expression of phosphorylated eIF4E-binding protein 1, but not of eIF4E itself, predicts survival in male breast cancer.*

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/100407/>

Version: Accepted Version

Article:

Millican-Slater, RA, Sayers, CD, Hanby, AM et al. (1 more author) (2016) Expression of phosphorylated eIF4E-binding protein 1, but not of eIF4E itself, predicts survival in male breast cancer. *British Journal of Cancer*, 115 (3). pp. 339-345. ISSN 0007-0920

<https://doi.org/10.1038/bjc.2016.178>

© 2016 Cancer Research UK. All rights reserved. This is an author produced version of a paper published in *British Journal of Cancer*. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Unless indicated otherwise, fulltext items are protected by copyright with all rights reserved. The copyright exception in section 29 of the Copyright, Designs and Patents Act 1988 allows the making of a single copy solely for the purpose of non-commercial research or private study within the limits of fair dealing. The publisher or other rights-holder may allow further reproduction and re-use of this version - refer to the White Rose Research Online record for this item. Where records identify the publisher as the copyright holder, users can verify any specific terms of use on the publisher's website.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Expression of phosphorylated eIF4E-binding protein 1, but not of eIF4E itself, predicts survival in male breast cancer

Rebecca A Millican-Slater^{1,3}, Craig D Sayers², Andrew M Hanby^{1,3}, Thomas A Hughes^{3*}

¹Department of Cellular Pathology, St. James's University Hospital, Leeds, LS9 7TF, UK

²Histopathology Department, Mid Yorkshire Hospitals NHS Trust, Dewsbury, WF13 4HS, UK

³School of Medicine, University of Leeds, Leeds, LS9 7TF, UK

*Corresponding author: Thomas A Hughes (+44 113 3431984; t.hughes@leeds.ac.uk);

Wellcome Trust Brenner Building, St. James's University Hospital, Leeds, LS9 7TF, UK

Running title: eIF4E and 4E-BPs in male breast cancer

ABSTRACT

Background: Male breast cancer is rare and treatment is based on data from females. High expression/activity of eukaryotic initiation factor 4E (eIF4E) denotes a poor prognosis in female breast cancer, and the eIF4E pathway has been targeted therapeutically. eIF4E activity in female breast cancer is deregulated by eIF4E over-expression and by phosphorylation of its binding protein, 4E-BP1, which relieves inhibitory association between eIF4E and 4E-BP1. The relevance of the eIF4E pathway in male breast cancer is unknown.

Methods: We have assessed expression levels of eIF4E, 4E-BP1, 4E-BP2 and phosphorylated 4E-BP1 (p4E-BP1) using immunohistochemistry in a large cohort of male breast cancers (n=337) and have examined correlations with prognostic factors and survival.

Results: Neither eIF4E expression or estimated eIF4E activity were associated with prognosis. However, a highly significant correlation was found between p4E-BP1 expression and disease-free survival, linking any detectable p4E-BP1 with poor survival (univariate log rank $p=0.001$; multivariate HR 8.8, $p=0.0001$).

Conclusions: Our data provide no support for direct therapeutic targeting of eIF4E in male breast cancer, unlike in females. However, as p4E-BP1 gives powerful prognostic insights that are unrelated to eIF4E function, p4E-BP1 may identify male breast cancers potentially suitable for therapies directed at the upstream kinase, mTOR.

Keywords: male breast cancer, translational regulation, predictive markers

Introduction

Male breast cancer (MBC) is a rare disease, accounting for less than 1% of all breast cancers and less than 1% of all male cancers diagnosed in the UK in 2009 (Cancer Research UK, 2010). There is relatively little research into MBC, presumably as a result of its rarity, and much of the published work has focused on comparisons with female breast cancer (FBC). Comparisons demonstrate that MBC is more likely to be estrogen receptor positive (92% positivity vs 78% for FBC (Ruddy & Winer, 2013)), and has some differences in genetic (Johansson *et al*, 2011; Kornegoor *et al*, 2012; Piscuoglio *et al*, 2016), transcriptomic (Callari *et al*, 2011; Johansson *et al*, 2012), and protein expression profiles (Shaaban *et al*, 2012) (reviewed in (Deb *et al*, 2016)). Incidence trends in terms of geographical location and impact of patient age for both diseases are broadly similar (Kreiter *et al*, 2014). These studies have not given insights that suggest different treatment approaches are appropriate, either in terms of which prognostic or predictive markers might be useful, or which therapies should be used. Also, there are no prospective randomised controlled trials for MBC that could inform treatment decisions (Bratman *et al*, 2012). Consequently, MBC management is based on data from FBC. One key difference, however, is that the vast majority of MBC patients undergo mastectomies (Korde *et al*, 2010), whereas breast conserving surgery is prevalent for FBC. It is worth noting that this difference is not based on evidence concerning treatment outcomes, rather on practical issues relating to the size of breast tissue. Adjuvant therapies, including radiotherapy (Ruddy & Winer, 2013), tamoxifen (Fogh *et al*, 2011; Ribeiro & Swindell, 1992) and chemotherapy (Korde *et al*, 2010), are essentially the same.

The eukaryotic translation initiation factor 4E (eIF4E) is a key component of the translational machinery and has two specific functions. Firstly, it recognizes and binds to mRNA caps within the cytoplasm allowing initiation of cap-dependent translation (Sonenberg, 2008), the

mechanism responsible for most protein synthesis (Gray & Wickens, 1998). Secondly, eIF4E binds to some mRNAs within the nucleus and regulates their nuclear export (Culjkovic *et al*, 2007; Culjkovic *et al*, 2005). Activity of eIF4E is controlled largely by the eIF4E binding proteins (4E-BPs), of which there are three, although only 4E-BP1 and 2 have been studied in any detail. eIF4E function is inhibited when bound by 4E-BPs (Matsuo *et al*, 1997), but this interaction is itself regulated by a series of sequential phosphorylations to the 4E-BPs mediated via the mTORC1 complex (Gibbons *et al*, 2009). Phosphorylated 4E-BPs are unable to bind to eIF4E. Thus, eIF4E activity is defined by a subtle balance of expression levels of eIF4E and the 4E-BPs, and the phosphorylation status of the 4E-BPs (Coleman *et al*, 2009). Activity of eIF4E is frequently increased in a wide range of cancers (De Benedetti & Graff, 2004), resulting in enhanced translation (and potentially nuclear export) of a subset of mRNAs that contains many cancer-related transcripts. In FBC, eIF4E is frequently expressed at higher levels in breast cancers compared to normal or benign breast tissue (Kerekatte *et al*, 1995; Norton *et al*, 2004) and higher levels of eIF4E are associated with poorer prognoses (Byrnes *et al*, 2006; Li *et al*, 2002). In addition, higher levels of the phosphorylated form of 4E-BP1 (p4E-BP1) are also seen in FBC compared to normal and benign tissue (Zhou *et al*, 2004), and these levels are positively associated with grade, lymph node metastasis and disease recurrence (Rojo *et al*, 2007). Our own work has demonstrated that combined analysis of expressions of eIF4E, 4E-BP1, 4E-BP2 and p4E-BP1 predicts breast cancer survival in females and represents an estimate of eIF4E activity (Coleman *et al*, 2009). The influential role that eIF4E plays in neoplasia has made it an attractive anti-cancer drug target. Therapeutic approaches that have been explored include knock-down of eIF4E expression (Graff *et al*, 2007; Hong *et al*, 2011), blocking of eIF4E-cap binding (Assouline *et al*, 2009; Pettersson *et al*, 2011), inhibition of eIF4E phosphorylation in an effort to reduce its activity (Wheater *et al*, 2010), and - most commonly - inhibition of mTORC1 activity leading to 4E-BP hypophosphorylation and inhibitory binding to eIF4E (Chan

et al, 2005; Wazir *et al*, 2014). It should be noted that inhibition of mTORC1, or more generally the mTOR kinase component of this complex, clearly has anti-cancer influence that are independent of 4E-BP1 through other targets of the complex (Laplane & Sabatini, 2009), and therefore this approach is in no way equivalent to direct targeting of eIF4E. Currently nothing is known about the prognostic relevance of eIF4E and the 4E-BPs in MBC, and there is no evidence base from which novel eIF4E-directed therapies might be considered in this disease; our aim was to perform the first investigation of the importance of these molecules in this cancer type.

Materials and Methods

Patients and tissue microarrays

Ethical approval was obtained from Leeds (West) (ref 06/Q1205/156) and Leeds (East) Research Ethics Committees (ref 05/Q1206/136). Archival resection samples of invasive breast cancers from male breast cancer patients (n=337) and associated clinical and pathological data were collected from the United Kingdom (157; 46.6%), Italy (50; 14.8%), Hungary (41; 12.2%), Poland (30; 9.5%), Canada (50; 14.8%) and Nigeria (9; 2.7%). Clinico-pathological characteristics are shown in Table 1. Survival data were available for 187 cases. Tissue microarrays (TMAs) were constructed from tissues; this process has been described in detail previously (Shaaban *et al*, 2012). In summary, H+E stained tumour sections were reviewed by specialist breast consultant histopathologists (RAM-S, AMH, Dr Abeer Shaaban [Queen Elizabeth Hospital, Birmingham, UK]) in order to confirm diagnoses and select representative areas of invasive carcinoma from which TMA cores would be taken. TMAs were constructed of duplicate or triplicate 0.6mm tumour cores from each individual case. 7 TMA blocks were used for the cohort, each including a perimeter wall of non-breast tissue (liver, sheep lung, placenta and brain) to minimise edge effects and to provide internal controls.

Immunohistochemistry

Immunohistochemistry was carried out as previously described (Coleman *et al*, 2009). In summary, 5µm sections were taken from blocks, and were deparaffinised and re-hydrated. Appropriate antigen retrieval (see below) was performed and sections were treated with 1% hydrogen peroxide-methanol to inhibit endogenous peroxidase activity. Sections were stained overnight with primary antibodies (see below) diluted in antibody diluent solution (Invitrogen; Carlsbad, USA). Signals were visualised using the DAB based Envision System (Dako;

Glostrup Denmark). All case TMAs, and a control TMA of female breast cancers, were stained for each antibody as a single batch. Female cores served as positive and negative controls. Antibodies, dilutions and antigen retrieval: eIF4E (mouse monoclonal sc9976; Santa Cruz Biotechnology; Santa Cruz, USA; 1:100; 2min pressure cooker in antigen un-masking solution, Vector, USA); 4E-BP1 (rabbit polyclonal 9452; Cell Signalling Technology [Danvers, USA]; 1:100; no antigen retrieval); 4E-BP2 (rabbit polyclonal 2845; Cell Signalling Technology [Danvers, USA]; 1:100; 12min full power microwave in pH6 citrate buffer); p4E-BP1 Thr37/46 (rabbit polyclonal 2855; Cell Signalling Technology [Danvers, USA]; 1:25; 12min full power microwave in pH6 citrate buffer). The specificities of these antibodies have been validated previously and they have all been used successfully for immunohistochemistry in breast tissue previously (Coleman *et al*, 2009; Satheesha *et al*, 2011; Zhou *et al*, 2006).

Scoring and statistics

Stained TMAs were digitally scanned (Aperio; Oxford, UK), and cores were scored independently by two consultant histopathologists (RAM-S, CDS) from the same digital images. Cytoplasmic and nuclear immunoreactivity were separated and given individual scores. The scoring system incorporated scores for staining intensity in tumour cells (0 no staining, 1 weak staining, 2 moderate staining and 3 strong staining) added to scores for proportions of tumour cells staining positively (1 <5%, 2 6-25%, 3 26-75% and 4 >75%), giving totals of either 0 or from 2 to 7, as has been used previously for these antigens (Coleman *et al*, 2009; Zhou *et al*, 2006). Analyses were performed in SPSS (SPSS; Chicago, USA) unless stated otherwise. Correlations between antigen expression scores and clinical factors were examined by calculating Spearman rho correlation coefficients. Associations with disease recurrence and survival were analysed by Kaplan-Meier survival curves and log rank tests following ROC curve analysis to dichotomise the expression scores into low and high expression appropriately. Kappa

calculations were performed using Analyse-it for Excel (Microsoft; Redmond, USA). All tests were two-sided.

Results

eIF4E, 4E-BP1, 4E-BP2 and p4E-BP1 expression varies widely in male breast cancer

Tissue microarrays (TMAs) containing duplicate or triplicate samples from 337 male breast tumours were stained using immunohistochemistry to analyse expression of eIF4E, 4E-BP1, 4E-BP2 and p4E-BP1. Cores were scored by two independent histopathologists in terms of expression intensity and proportions of cells staining positively. To take into account the potentially different roles of these protein species in different cellular compartments (Culjkovic *et al*, 2006; Sonenberg, 2008), cytoplasmic and nuclear immunoreactivity were separated and given individual scores. Scores from the two histopathologists were highly concordant, demonstrating robust and reproducible scoring; quadratic weighted kappa statistics were 0.85-0.96 for cytoplasmic scores and 0.74-0.95 for nuclear (depending on antigen; see Table S1). Core loss, an expected and documented occurrence in TMA-based research (Parsons & Grabsch, 2009), or lack of tumour cells meant that staining was not assessable in some cases; however a mean of 2.3 cores were successfully scored for each case for each antibody. We analysed variability in scores between multiple cores representing individual tumours in order to assess potential heterogeneity within individual tumours and therefore the representative nature of TMA cores. Spearman's rho correlation coefficients for duplicate scores for each tumour and antigen were all 0.79 ($p < 0.001$) or over, demonstrating that there was relatively little heterogeneity in marker expression within individual tumours and that TMA based analyses were appropriate. Having determined that inter-scorer and core-to-core variability were low, we took mean values of all the scores available for each case/antigen/sub-cellular location to create single scores for further analysis. Representative staining and the frequency distributions of these scores (rounded to the nearest whole number) are shown in Fig 1. The full range of expression patterns were seen for each antigen, ranging from no detectable expression through

to strongly expressed in more than 75% of tumour cells. The distributions of cytoplasmic and nuclear scores were broadly similar for each antigen, and expressions in the two compartments were strongly associated (Spearman's rho correlation coefficients 0.85 to 0.95, $p < 0.0008$), suggesting that separating the two scores gave relatively little additional information.

Expression of eIF4E correlates weakly with ER status in male breast cancer

Associations between marker expression and established prognostic factors were examined. The factors tested were: 1) histological tumour grade (1, 2 or 3); 2) tumour size (categorised as 2cm or less, >2cm but less than or equal to 5cm, or >5cm); 3) lymph node status (negative or positive); and, 4) oestrogen receptor alpha (ER α) status (negative [Allred 0 or 2] or positive [Allred >2]). Spearman's rho correlation coefficients (r) were calculated for each potential association. The only associations with Spearman's coefficients >0.2, which is weak at best, were both cytoplasmic and nuclear eIF4E expression being positively associated with ER α status ($r=0.231$ and $r=0.202$ respectively; $p < 0.002$).

Expression of p4E-BP1, but not eIF4E, is strongly associated with male breast cancer survival

Kaplan-Meier survival analyses were performed to determine whether expression of the markers was significantly related to disease free survival (DFS). Cut offs were applied to dichotomise patients into two groups based on low or high expression of each marker. These cut offs were defined objectively using receiver operator curve analyses (Zlobec *et al*, 2007) to give the best balance between sensitivity and specificity for prediction of the relevant clinical outcome (i.e. breast cancer recurrence). The cut off values are shown in Table S2. Kaplan-Meier survival analyses were performed and log rank tests were used to assess the significance of relationships. A stringent value of $p < 0.003$ was defined as indicating significance, after Bonferroni correction

for multiple tests from an initial value of $p < 0.05$. Only cytoplasmic expression of p4E-BP1 demonstrated a significant relationship with survival (Fig 2), with patients with low p4E-BP1 expression having a longer DFS than those with high expression (215 vs. 95 months, $p = 0.001$). It is important to note that the cut off to dichotomise p4E-BP1 expression was 0.83, meaning that tumours in the two groups were those without detectable p4E-BP1 (negative) or those with any detectable expression (positive). Cytoplasmic and nuclear expression of 4E-BP1, and - surprisingly - nuclear expression of 4E-BP2, also showed trends towards significant relationships with survival, although these fell short of our stringent significance test. Expression of eIF4E itself showed no such trend (Table S3).

Estimated eIF4E activity is not associated with male breast cancer survival

We have previously demonstrated in FBC that assessments of expression of these markers could be combined to estimate eIF4E activity, an estimated value that was significantly associated with survival (Coleman *et al*, 2009). Activity (referred to as “z”) was estimated as $X - BP1/4 + pBP1/2 - BP2/4$, where X represents the eIF4E score, BP1 the 4E-BP1 score, BP2 the 4E-BP2 score, and pBP1 the p4E-BP1 score. This estimate was determined for these MBC cases using the cytoplasmic scores, and receiver operator curve analysis was performed to determine a suitable cut off to split the cohort in groups with high and low z scores. Kaplan Meier survival analyses were performed. Estimated eIF4E activity was not significantly associated with DFS (Table S3).

Cytoplasmic p4E-BP1 is significantly associated with survival in multivariate analysis

Multivariate regression was performed to assess whether cytoplasmic expression of p4E-BP1 was an independent prognostic factor with regards to DFS. The other variables put into the model were the currently used prognostic factors of grade, tumour size, lymph node status and

ER α status. Both cytoplasmic p4E-BP1 expression and tumour size were significantly associated with DFS on univariate and multivariate analyses, although cytoplasmic p4E-BP1 expression consistently showed the greater significance and the more informative hazard ratio (Table 2).

Discussion

This study is the first in which expressions and prognostic relevance of eIF4E and the 4E-BPs have been examined in MBC. Our analysis involved one of the largest MBC cohorts assembled (n=337) and thorough immunohistochemical analyses with multiple tissue samples per case, very robust histopathological scoring, and well-validated antibodies. It is also worth noting that our work is the first in any cancer to separately investigate the prognostic worth of eIF4E and its regulatory molecules in cytoplasmic and nuclear compartments, in accordance with their different reported roles in these locations (De Benedetti & Graff, 2004; Siddiqui & Borden, 2012). We found expression in these compartments to be tightly correlated, and separate prognostic insights were not gained from the compartment analysis. Interestingly, some individual cases with prominent nuclear only, or cytoplasmic only expression were noted, suggesting that sub-cellular regulation may take place in some circumstances; however cases were infrequent and analysis of their common clinicopathological features was flawed on this basis. A rare precedent for separating different subcellular localisation of these molecules in cancer is, remarkably, also in the context of MBC. Nuclear and cytoplasmic distributions of p4E-BP1 have been reported previously in 56 familial MBCs, showing expression in the two compartments to be highly associated and positive in 52 and 55% of cases respectively (slightly more than we find) (Deb *et al*, 2013).

Surprisingly, and in marked contrast to FBC (Coleman *et al*, 2009), no association was found between eIF4E expression and survival. Expression of eIF4E has been associated with prognosis in a wide range of cancers (De Benedetti & Graff, 2004), but there are specific cancers where this is not the case, for example, in acute myeloid leukaemia (Green *et al*, 2012) or osteosarcoma (Osborne *et al*, 2011). In addition, there is likely to be a publication bias against

such findings, so it may be that this lack of association is more common than currently appreciated. Strikingly, however, we identified a strong prognostic association for p4E-BP1, with any detectable p4E-BP1 expression correlated with poor survival in both univariate and multivariate analyses (Fig 2; Table 2). This association was far stronger than previously found in FBC (Coleman *et al*, 2009). 4E-BP1 phosphorylation breaks 4E-BP1's inhibitory interaction with eIF4E resulting in increased eIF4E activity (De Benedetti & Graff, 2004), therefore one might expect that p4E-BP1 could only be functionally associated with prognosis through the eIF4E pathway. Yet, here we show that the wide variations in expression of eIF4E itself, or in estimated eIF4E activity, do not impact on prognosis (Tables S2 and S3) rendering this expectation incompatible with our data. We interpret this to suggest that p4E-BP1 is acting as a biomarker for functionally-relevant activity of the upstream kinase, the mTORC1 complex, rather than having a direct functional impact on prognosis itself. In support of this, it is well established that levels of p4E-BP1 correlate with mTORC1 activity in various contexts, and accordingly p4E-BP1 has frequently been used as a pharmacodynamic marker for mTORC1 activity in trials of mTORC1-targeting therapeutics (Spunt *et al*, 2011; Taberero *et al*, 2008).

The kinase within the mTORC1 complex is mTOR, up-regulation of which is associated with development of many cancers (Shaw & Cantley, 2006). The mTORC1 complex acts on a large number of different molecular substrates (Hsu *et al*, 2011; Laplante & Sabatini, 2009), although the functional importance of two have been studied in considerably more detail than the others with regard to cancer: 4E-BP1 and S6 kinase 1 (S6K1). In MBC we believe that 4E-BP1 may not be a functionally-relevant substrate, therefore it seems likely that deregulated mTOR acts at least in part through S6K1 and its downstream effectors. Phosphorylated (activated) S6K1 can induce oncogenic increases in overall protein translation, and changes in sterol, lipid and mitochondrial metabolism via a variety of complex signalling pathways (Alayev & Holz, 2013).

Expression levels of both eIF4E and mTOR have been noted in a previous analysis of gene expression profiles in MBC (n=37) as compared to FBC (Callari *et al*, 2011). Both proteins were found to be more highly expressed in MBC than in FBC, and the authors commented that the eIF4E pathway may therefore present an attractive therapeutic target in MBC. Our findings impact on this suggestion, in that we find eIF4E itself to be unrelated to prognosis, while we infer that mTOR activity within the mTORC1 complex may well relate to prognosis. Thus, our data do not support use of therapies directed at eIF4E itself, such as knock-down of eIF4E expression (Graff *et al*, 2007; Hong *et al*, 2011), or function (Assouline *et al*, 2009; Pettersson *et al*, 2011; Wheater *et al*, 2010), but do support potential use of therapies directed at the upstream kinase, mTOR.

These findings may delineate potential differences in appropriate treatments between female and male breast cancers. For example, the eIF4E-directed therapies LY2275796 (anti-sense oligonucleotides directed against eIF4E) and ribavirin (which reduces eIF4E-dependent translation) have shown some promise in pre-clinical or clinical trials (Hong *et al*, 2011; Pettersson *et al*, 2015), and are under-going evaluation for FBC. Our data suggest that these may have limited efficacy in MBC since eIF4E activity appears relatively unimportant in determining prognosis in this disease. However, by contrast, the growing list of therapies targeting mTOR (Sun, 2013), such as everolimus or temsirolimus that have already shown promise in FBC trials (Baselga *et al*, 2012; Wolff *et al*, 2013), may well be suitable therapies in both female and male cancers. Interestingly, there is a single case report describing a favourable response of a MBC patient to temsirolimus (Katayama *et al*, 2013), but unfortunately it seems unlikely that a MBC trial will take place due to the rarity of the disease overall. A further issue would be that fewer than 50% of MBC cases expressed detectable p4E-BP1 in our data (Fig 1),

and therefore only a minority may potentially be suitable for this approach. Nevertheless, we conclude that mTOR-targeted therapies may be worth considering in p4E-BP1 positive MBC.

Acknowledgements

We thank the national and international investigators who contributed samples and data to the MBC cohort, and Dr Abeer Shaaban (Queen Elizabeth Hospital, Birmingham, UK) and Prof Valerie Speirs (University of Leeds), who assembled the cohort. Thanks also to Prof Speirs, for providing the tissue and data through a material transfer agreement.

Figures / Tables and legends

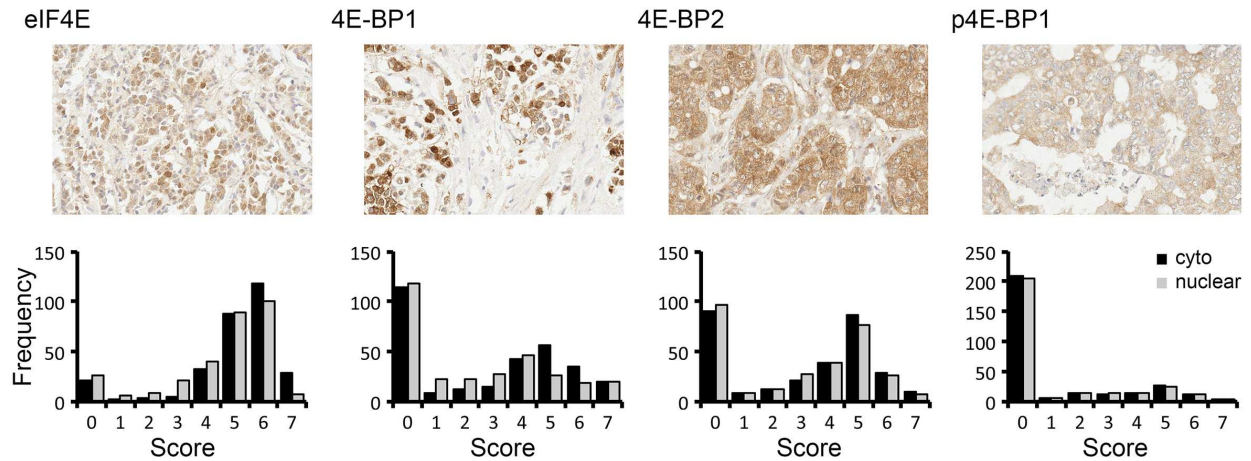


Figure 1. Male breast cancer (MBC) has a full range of expression patterns for eIF4E, 4E-BP1, 4E-BP2 and p4E-BP1. Tissue microarrays containing multiple tumour cores from 337 MBCs were stained as indicated using immunohistochemistry. Cytoplasmic and nuclear expressions in tumour cells were assessed as 0 (negative) or 2-7 (positive, increasing intensity/proportion of positive cells). Representative positive staining is shown at the top of the panel for each antigen. Images shown were scored for cytoplasmic, c, and nuclear, n, expression as follows: eIF4E - c 7, n 6; 4E-BP1 - c 4, n 5; 4E-BP2 - c 5, n 0; p4E-BP1 - c 6, n 3. Frequency distributions of cytoplasmic (black) or nuclear (grey) expression across the cohort are shown below. Mean scores for each case were determined and are represented rounded to the nearest whole number.

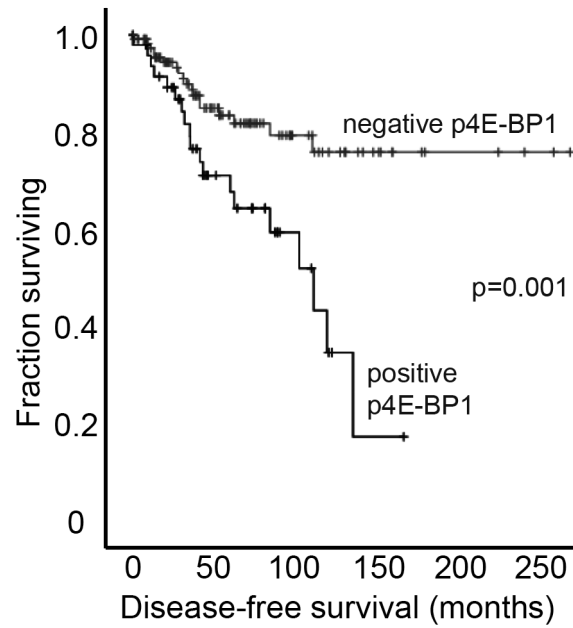


Figure 2. Expression of p4E-BP1 is significantly associated with disease-free survival in male breast cancer ($p=0.001$). Kaplan–Meier survival analyses for patient groups with tumours with either no detectable (negative; grey line) or any detectable (positive; black line) expression of p4E-BP1.

Characteristic	Number (%) n=337
Histological type:	
Ductal no-special type	275 (81.6)
Papillary/encysted papillary	17 (5.1)
Mucinous	11 (3.3)
Lobular	3 (0.9)
Other special type	5 (1.5)
Mixed	8 (2.4)
Unknown	8 (2.4)
Tumour grade:	
1	44 (13.1)
2	158 (46.9)
3	121 (35.9)
Ungraded	14 (4.2)
Tumour size:	
1 (<2cm)	70 (20.8)
2 (2-5cm)	65 (19.3)
3 (>5cm)	14 (4.2)
Unknown	188 (55.8)
LN status:	
At least 1 positive node	112 (33.2)
No positive nodes	101 (30.0)
Unknown	124 (36.8)
ER status:	
Positive (Allred score >2)	238 (70.6)
Negative	52 (15.4)
Unknown	47 (13.9)
PR status:	
Positive (Allred score >2)	238 (70.6)
Negative	48 (14.2)
Unknown	51 (15.1)

Table 1. Clinical and pathological features of the cohort. LN – lymph node; ER – oestrogen receptor alpha; PR – progesterone receptor

	Univariate analysis		Multivariate analysis	
	Hazard ratio	p value	Hazard ratio	p value
Cytoplasmic p4E-BP1	3.073	0.001	8.755	<0.0005
Tumour size	1.963	0.048	2.923	0.016
Tumour grade	0.704	0.165	0.432	0.129
LN status	1.494	0.326	4.976	0.018
ER status	1.792	0.277	1.176	0.788

Table 2. Cytoplasmic p4E-BP1 is significantly associated with survival in univariate and multivariate regression analysis. LN – lymph node; ER – oestrogen receptor alpha

	Quadratic Kw (95% CI)
Cytoplasmic eIF4E	0.85 (0.83 to 0.88)
Nuclear eIF4E	0.74 (0.71 to 0.78)
Cytoplasmic 4E-BP1	0.95 (0.94 to 0.96)
Nuclear 4E-BP1	0.93 (0.92 to 0.95)
Cytoplasmic 4E-BP2	0.95 (0.94 to 0.96)
Nuclear 4E-BP2	0.91 (0.89 to 0.92)
Cytoplasmic p4E-BP1	0.96 (0.95 to 0.97)
Nuclear p4E-BP1	0.95 (0.94 to 0.96)

Table S1. Concordance values for the two independent scorers for each antigen and sub-cellular location. Quadratic weighted kappa statistics (Kw) are shown with 95% confidence intervals (CI).

	Cut off values
Cytoplasmic eIF4E	4.92
Nuclear eIF4E	4.21
Cytoplasmic 4E-BP1	0.75
Nuclear 4E-BP1	0.50
Cytoplasmic 4E-BP2	0.17
Nuclear 4E-BP2	0.17
Cytoplasmic p4E-BP1	0.83
Nuclear p4E-BP1	1.63
Cytoplasmic z score	5.17

Table S2. Cut off values determined using receiver operator curve analyses for each antigen and sub-cellular localisation for the end point of breast cancer recurrence.

		Mean DFS in months (95% CI)	p value
Cytoplasmic eIF4E	Low	145 (116-175)	0.120
	High	199 (168-231)	
Nuclear eIF4E	Low	161 (121-200)	0.048
	High	195 (165-224)	
Cytoplasmic 4E-BP1	Low	225 (198-251)	0.007
	High	119 (101-136)	
Nuclear 4E-BP1	Low	228 (199-256)	0.004
	High	122 (105-139)	
Cytoplasmic 4E-BP2	Low	152 (117-187)	0.040
	High	203 (172-234)	
Nuclear 4E-BP2	Low	147 (112-181)	0.006
	High	206 (175-237)	
Nuclear p4E-BP1	Low	212 (188-235)	0.008
	High	99 (79-118)	
Cytoplasmic z score	Low	196 (166-225)	0.064
	High	141 (91-192)	

Table S3. Non-significant relationships with disease free survival (DFS) for markers. Mean DFS for low and high expression groups are shown (with 95% confidence intervals). Note after Bonferroni correction for multiple testing, a stringent threshold of $p < 0.003$ was defined as indicating significance.

References

Alayev A, Holz MK (2013) mTOR signaling for biological control and cancer. *Journal of cellular physiology* **228**(8): 1658-64

Assouline S, Culjkovic B, Cocolakis E, Rousseau C, Beslu N, Amri A, Caplan S, Leber B, Roy DC, Miller WH, Jr., Borden KL (2009) Molecular targeting of the oncogene eIF4E in acute myeloid leukemia (AML): a proof-of-principle clinical trial with ribavirin. *Blood* **114**(2): 257-260

Baselga J, Campone M, Piccart M, Burris HA, 3rd, Rugo HS, Sahmoud T, Noguchi S, Gnant M, Pritchard KI, Lebrun F, Beck JT, Ito Y, Yardley D, Deleu I, Perez A, Bachelot T, Vittori L, Xu Z, Mukhopadhyay P, Lebwohl D, Hortobagyi GN (2012) Everolimus in postmenopausal hormone-receptor-positive advanced breast cancer. *N Engl J Med* **366**(6): 520-9

Bratman SV, Kapp DS, Horst KC (2012) Evolving trends in the initial locoregional management of male breast cancer. *Breast* **21**(3): 296-302

Byrnes K, White S, Chu Q, Meschonat C, Yu H, Johnson LW, Debenedetti A, Abreo F, Turnage RH, McDonald JC, Li BD (2006) High eIF4E, VEGF, and microvessel density in stage I to III breast cancer. *Ann Surg* **243**(5): 684-690; discussion 691-692

Callari M, Cappelletti V, De Cecco L, Musella V, Miodini P, Veneroni S, Gariboldi M, Pierotti MA, Daidone MG (2011) Gene expression analysis reveals a different transcriptomic landscape in female and male breast cancer. *Breast Cancer Res Treat* **127**(3): 601-610

Chan S, Scheulen ME, Johnston S, Mross K, Cardoso F, Dittrich C, Eiermann W, Hess D, Morant R, Semiglazov V, Borner M, Salzberg M, Ostapenko V, Illiger HJ, Behringer D, Bardy-Bouxin N, Boni J, Kong S, Cincotta M, Moore L (2005) Phase II study of temsirolimus (CCI-779), a novel inhibitor of mTOR, in heavily pretreated patients with locally advanced or metastatic breast cancer. *J Clin Oncol* **23**(23): 5314-5322

Coleman LJ, Peter MB, Teall TJ, Brannan RA, Hanby AM, Honarpisheh H, Shaaban AM, Smith L, Speirs V, Verghese ET, McElwaine JN, Hughes TA (2009) Combined analysis of eIF4E and 4E-binding protein expression predicts breast cancer survival and estimates eIF4E activity. *Br J Cancer* **100**(9): 1393-9

CRUK (2010) Breast Cancer Incidence Statistics

Culjkovic B, Topisirovic I, Borden KL (2007) Controlling gene expression through RNA regulons: the role of the eukaryotic translation initiation factor eIF4E. *Cell Cycle* **6**(1): 65-9

Culjkovic B, Topisirovic I, Skrabanek L, Ruiz-Gutierrez M, Borden KL (2005) eIF4E promotes nuclear export of cyclin D1 mRNAs via an element in the 3'UTR. *J Cell Biol* **169**(2): 245-56

Culjkovic B, Topisirovic I, Skrabanek L, Ruiz-Gutierrez M, Borden KL (2006) eIF4E is a central node of an RNA regulon that governs cellular proliferation. *The Journal of cell biology* **175**(3): 415-426

De Benedetti A, Graff JR (2004) eIF-4E expression and its role in malignancies and metastases. *Oncogene* **23**(18): 3189-99

Deb S, Do H, Byrne D, Jene N, kConFab I, Dobrovic A, Fox SB (2013) PIK3CA mutations are frequently observed in BRCA1 but not BRCA2-associated male breast cancer. *Breast Cancer Res* **15**(4): R69

Deb S, Lakhani SR, Ottini L, Fox SB (2016) The cancer genetics and pathology of male breast cancer. *Histopathology* **68**(1): 110-8

Fogh S, Hirsch AE, Langmead JP, Goldberg SI, Rosenberg CL, Taghian AG, Powell SN, Kachnic LA (2011) Use of tamoxifen with postsurgical irradiation may improve survival in estrogen and progesterone receptor-positive male breast cancer. *Clin Breast Cancer* **11**(1): 39-45

Gibbons JJ, Abraham RT, Yu K (2009) Mammalian target of rapamycin: discovery of rapamycin reveals a signaling pathway important for normal and cancer cell growth. *Semin Oncol* **36 Suppl 3**: S3-S17

Graff JR, Konicek BW, Vincent TM, Lynch RL, Monteith D, Weir SN, Schwier P, Capen A, Goode RL, Dowless MS, Chen Y, Zhang H, Sissons S, Cox K, McNulty AM, Parsons SH, Wang T, Sams L, Geeganage S, Douglass LE, Neubauer BL, Dean NM, Blanchard K, Shou J, Stancato LF, Carter JH, Marcusson EG (2007) Therapeutic suppression of translation initiation factor eIF4E expression reduces tumor growth without toxicity. *J Clin Invest* **117**(9): 2638-48

Gray NK, Wickens M (1998) Control of translation initiation in animals. *Annual review of cell and developmental biology* **14**: 399-458

Green AS, Grabar S, Tulliez M, Park S, Al-Nawakil C, Chapuis N, Jacque N, Willems L, Azar N, Ifrah N, Dreyfus F, Lacombe C, Mayeux P, Bouscary D, Tamburini J (2012) The eukaryotic initiating factor 4E protein is overexpressed, but its level has no prognostic impact in acute myeloid leukaemia. *British journal of haematology* **156**(4): 547-50

Hong DS, Kurzrock R, Oh Y, Wheler J, Naing A, Brail L, Callies S, Andre V, Kadam SK, Nasir A, Holzer TR, Meric-Bernstam F, Fishman M, Simon G (2011) A phase 1 dose escalation,

pharmacokinetic, and pharmacodynamic evaluation of eIF-4E antisense oligonucleotide LY2275796 in patients with advanced cancer. *Clin Cancer Res* **17**(20): 6582-91

Hsu PP, Kang SA, Rameseder J, Zhang Y, Ottina KA, Lim D, Peterson TR, Choi Y, Gray NS, Yaffe MB, Marto JA, Sabatini DM (2011) The mTOR-regulated phosphoproteome reveals a mechanism of mTORC1-mediated inhibition of growth factor signaling. *Science* **332**(6035): 1317-22

Johansson I, Nilsson C, Berglund P, Lauss M, Ringner M, Olsson H, Luts L, Sim E, Thorstensson S, Fjallskog ML, Hedenfalk I (2012) Gene expression profiling of primary male breast cancers reveals two unique subgroups and identifies N-acetyltransferase-1 (NAT1) as a novel prognostic biomarker. *Breast Cancer Res* **14**(1): R31

Johansson I, Nilsson C, Berglund P, Strand C, Jonsson G, Staaf J, Ringner M, Nevanlinna H, Barkardottir RB, Borg A, Olsson H, Luts L, Fjallskog ML, Hedenfalk I (2011) High-resolution genomic profiling of male breast cancer reveals differences hidden behind the similarities with female breast cancer. *Breast Cancer Res Treat* **129**(3): 747-760

Katayama K, Yamagishi J, Kashiwagi B (2013) [A case of male breast cancer for which mTOR inhibitor was effective with advanced renal cancer]. *Gan to kagaku ryoho Cancer & chemotherapy* **40**(3): 365-9

Kerekatte V, Smiley K, Hu B, Smith A, Gelder F, De Benedetti A (1995) The proto-oncogene/translation factor eIF4E: a survey of its expression in breast carcinomas. *Int J Cancer* **64**(1): 27-31

Korde LA, Zujewski JA, Kamin L, Giordano S, Domchek S, Anderson WF, Bartlett JM, Gelmon K, Nahleh Z, Bergh J, Cutuli B, Pruneri G, McCaskill-Stevens W, Gralow J, Hortobagyi G, Cardoso F (2010) Multidisciplinary meeting on male breast cancer: summary and research recommendations. *J Clin Oncol* **28**(12): 2114-2122

Kornegoor R, Moelans CB, Verschuur-Maes AH, Hogenes MC, de Bruin PC, Oudejans JJ, Marchionni L, van Diest PJ (2012) Oncogene amplification in male breast cancer: analysis by multiplex ligation-dependent probe amplification. *Breast Cancer Res Treat* **135**(1): 49-58

Kreiter E, Richardson A, Potter J, Yasui Y (2014) Breast cancer: trends in international incidence in men and women. *Br J Cancer*

Laplante M, Sabatini DM (2009) mTOR signaling at a glance. *J Cell Sci* **122**(Pt 20): 3589-94

Li BD, Gruner JS, Abreo F, Johnson LW, Yu H, Nawas S, McDonald JC, DeBenedetti A (2002) Prospective study of eukaryotic initiation factor 4E protein elevation and breast cancer outcome. *Ann Surg* **235**(5): 732-8; discussion 738-9

Matsuo H, Li H, McGuire AM, Fletcher CM, Gingras AC, Sonenberg N, Wagner G (1997) Structure of translation factor eIF4E bound to m7GDP and interaction with 4E-binding protein. *Nature structural biology* **4**(9): 717-24

Norton KS, McClusky D, Sen S, Yu H, Meschonat C, Debenedetti A, Li BD (2004) TLK1B is elevated with eIF4E overexpression in breast cancer. *The Journal of surgical research* **116**(1): 98-103

Osborne TS, Ren L, Healey JH, Shapiro LQ, Chou AJ, Gorlick RG, Hewitt SM, Khanna C (2011) Evaluation of eIF4E expression in an osteosarcoma-specific tissue microarray. *Journal of pediatric hematology/oncology* **33**(7): 524-8

Parsons M, Grabsch H (2009) How to make tissue microarrays. *Diagnostic Histopathology* **15**(3): 142-150

Pettersson F, Del Rincon SV, Emond A, Huor B, Ngan E, Ng J, Dobocan MC, Siegel PM, Miller WH, Jr. (2015) Genetic and pharmacologic inhibition of eIF4E reduces breast cancer cell migration, invasion, and metastasis. *Cancer Res* **75**(6): 1102-12

Pettersson F, Yau C, Dobocan MC, Culjkovic-Kraljacic B, Retrouvey H, Puckett R, Flores LM, Krop IE, Rousseau C, Cocolakis E, Borden KL, Benz CC, Miller WH, Jr. (2011) Ribavirin treatment effects on breast cancers overexpressing eIF4E, a biomarker with prognostic specificity for luminal B-type breast cancer. *Clin Cancer Res* **17**(9): 2874-2884

Piscuoglio S, Ng CK, Murray MP, Guerini-Rocco E, Martelotto LG, Geyer FC, Bidard FC, Berman S, Fusco N, Sakr RA, Eberle C, De Mattos-Arruda L, Macedo GS, Akram M, Baslan T, Hicks J, King TA, Brogi E, Norton L, Weigelt B, Hudis CA, Reis-Filho JS (2016) The Genomic Landscape of Male Breast Cancers. *Clin Cancer Res*

Ribeiro G, Swindell R (1992) Adjuvant tamoxifen for male breast cancer (MBC). *Br J Cancer* **65**(2): 252-254

Rojo F, Najera L, Lirola J, Jimenez J, Guzman M, Sabadell MD, Baselga J, Ramon y Cajal S (2007) 4E-binding protein 1, a cell signaling hallmark in breast cancer that correlates with pathologic grade and prognosis. *Clin Cancer Res* **13**(1): 81-9

Ruddy KJ, Winer EP (2013) Male breast cancer: risk factors, biology, diagnosis, treatment, and survivorship. *Ann Oncol* **24**(6): 1434-1443

Satheesha S, Cookson VJ, Coleman LJ, Ingram N, Madhok B, Hanby AM, Suleman CA, Sabine VS, Macaskill EJ, Bartlett JM, Dixon JM, McElwaine JN, Hughes TA (2011) Response to

mTOR inhibition: activity of eIF4E predicts sensitivity in cell lines and acquired changes in eIF4E regulation in breast cancer. *Molecular cancer* **10**: 19

Shaaban AM, Ball GR, Brannan RA, Cserni G, Benedetto AD, Dent J, Fulford L, Honarpisheh H, Jordan L, Jones JL, Kanthan R, Maraqa L, Litwiniuk M, Mottolese M, Pollock S, Provenzano E, Quinlan PR, Reall G, Shousha S, Stephens M, Verghese ET, Walker RA, Hanby AM, Speirs V (2012) A comparative biomarker study of 514 matched cases of male and female breast cancer reveals gender-specific biological differences. *Breast Cancer Res Treat* **133**(3): 949-958

Shaw RJ, Cantley LC (2006) Ras, PI(3)K and mTOR signalling controls tumour cell growth. *Nature* **441**(7092): 424-30

Siddiqui N, Borden KL (2012) mRNA export and cancer. *Wiley interdisciplinary reviews RNA* **3**(1): 13-25

Sonenberg N (2008) eIF4E, the mRNA cap-binding protein: from basic discovery to translational research. *Biochemistry and cell biology = Biochimie et biologie cellulaire* **86**(2): 178-83

Spunt SL, Grupp SA, Vik TA, Santana VM, Greenblatt DJ, Clancy J, Berkenblit A, Krygowski M, Ananthkrishnan R, Boni JP, Gilbertson RJ (2011) Phase I study of temsirolimus in pediatric patients with recurrent/refractory solid tumors. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* **29**(21): 2933-40

Sun SY (2013) mTOR kinase inhibitors as potential cancer therapeutic drugs. *Cancer Lett* **340**(1): 1-8

Tabernero J, Rojo F, Calvo E, Burris H, Judson I, Hazell K, Martinelli E, Ramon y Cajal S, Jones S, Vidal L, Shand N, Macarulla T, Ramos FJ, Dimitrijevic S, Zoellner U, Tang P, Stumm M, Lane HA, Lebwohl D, Baselga J (2008) Dose- and schedule-dependent inhibition of the mammalian target of rapamycin pathway with everolimus: a phase I tumor pharmacodynamic study in patients with advanced solid tumors. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* **26**(10): 1603-10

Wazir U, Wazir A, Khanzada ZS, Jiang WG, Sharma AK, Mokbel K (2014) Current state of mTOR targeting in human breast cancer. *Cancer genomics & proteomics* **11**(4): 167-74

Wheater MJ, Johnson PW, Blaydes JP (2010) The role of MNK proteins and eIF4E phosphorylation in breast cancer cell proliferation and survival. *Cancer Biol Ther* **10**(7): 728-735

Wolff AC, Lazar AA, Bondarenko I, Garin AM, Brinca S, Chow L, Sun Y, Neskovic-Konstantinovic Z, Guimaraes RC, Fumoleau P, Chan A, Hachemi S, Strahs A, Cincotta M, Berkenblit A, Krygowski M, Kang LL, Moore L, Hayes DF (2013) Randomized phase III placebo-controlled trial of letrozole plus oral temsirolimus as first-line endocrine therapy in postmenopausal women with locally advanced or metastatic breast cancer. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* **31**(2): 195-202

Zhou S, Wang GP, Liu C, Zhou M (2006) Eukaryotic initiation factor 4E (eIF4E) and angiogenesis: prognostic markers for breast cancer. *BMC Cancer* **6**: 231

Zhou X, Tan M, Stone Hawthorne V, Klos KS, Lan KH, Yang Y, Yang W, Smith TL, Shi D, Yu D (2004) Activation of the Akt/mammalian target of rapamycin/4E-BP1 pathway by ErbB2 overexpression predicts tumor progression in breast cancers. *Clin Cancer Res* **10**(20): 6779-88

Zlobec I, Steele R, Terracciano L, Jass JR, Lugli A (2007) Selecting immunohistochemical cut-off scores for novel biomarkers of progression and survival in colorectal cancer. *J Clin Pathol* **60**(10): 1112-1116