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Enantioselective Component Selection in Multi-Component Supramolecular Gels

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ABSTRACT: We investigate a two-component acid-amine gelation system in which chirality plays a vital role. A carboxylic acid based on a second generation L-lysine dendron interacts with chiral amines and subsequently assembles into supramolecular gel fibers. The chirality of the amine controls the assembly of the resulting diastereomeric complexes, even if this chirality is relatively 'poor quality'. Importantly, the selective incorporation of one enantiomer of an amine over the other into the gel network has been demonstrated, with the *R* amine that forms complexes which assemble into the most stable gel being primarily selected for incorporation. Thermodynamic control has been proven by forming a gel exclusively with an *S* amine, then allowing the *R* enantiomer to diffuse through the gel network, displacing it from the solid-like fibers, demonstrating these gels adapt and evolve in response to chemical stimuli to which they are exposed. Excess amine, which remains unincorporated within the 'solid-like' gel fiber network, can diffuse out and be reacted with an isocyanate, allowing us to quantify the enantioselectivity of component selection, but also demonstrating how gels can act as selective reservoirs of potential reagents, releasing them on demand to undergo further reactions – hence component-selective gel assembly can be coupled with controlled reactivity.

Introduction

Supramolecular gels are soft materials with self-assembled nanoscale fibrillar architectures which are being explored for a wide range of different applications - from formulation science through to high-tech multi-functional materials.¹ Recently there has been particular focus on multi-component gels, in which several different molecular-scale building blocks participate in gel formation.² These gels often rely on a complex forming between different components before gelation can take place.³ In multi-component gels, complex/gelator formation and/or fibre self-assembly can sometimes drive a 'component selection' event. When this occurs, certain molecules are selected from a mixture because their favoured selfassembly thermodynamically drives the evolution of the mixed molecular library.⁴ Furthermore, gelators programmed with different molecular-scale information, may be able to independently self-sort into their own nanoscale networks.⁴ Understanding how self-assembly operates within complex systems such as these is of fundamental importance in understanding how non-covalent chemistry can effectively program the emergence of order from chaos.⁶ Furthermore, these complex, yet well-organised, multi-component gels are particularly interesting, because the presence of multiple molecular-scale species offers the possibility of introducing multi-functionality to these materials.

Chiral gels have been of particular interest, as the chiral information programmed in at the molecular-scale can be translated through to the nanoscale assembly of chiral architectures, and ultimately have an impact on the macroscopic performance of the gel.⁷ There have been a number of studies in which mixtures of enantiomers have been investigated, and in general terms, mixing enantiomeric gelators suppresses gela-

tion.⁸ In some cases, if homochiral recognition is preferred the enantiomers can self-sort to form their own chirally sorted nanostructures.^{5,9} In rare cases, the two enantiomers interact preferentially with each other to form a complex which is an even better gelator than either individual enantiomer.¹⁰ In general terms, however, rules about chiral selectivity in gelation are still emerging. It has been demonstrated, for example that enantiopure gelators can express their chiral assembly preferences on an achiral analogue in a 'sergeants and soldiers' type manner.¹¹ An achiral gelator has even been shown to undergo a mirror symmetry breaking event on gelation leading to spontaneous amplification of chirality.¹² Furthermore, there have been several reports in which chiral gels respond in an enantioselective manner to chiral analytes (and solvents) with changes in properties signalling the recognition event.¹² There have also been several examples in which a chiral compound can be induced to form a gel if it complexes with one guest enantiomer but a precipitate when bound to the other.¹⁴



Fig. 1. Chiral gelation system of G2-Lys and C6R/S.

We have recently been working on a simple, and highly effective organogelation system comprised of two components (e.g. Fig. 1) in a 1:1 ratio; (i) a chiral second generation dendron based on L-lysine (**G2-Lys**) with a carboxylic acid at the focal point, and (ii) a primary amine.¹⁵ These two soluble components can form instant gels on mixing, and we recently demonstrated how **G2-Lys**, if challenged with mixture of different amines, would select those which had the highest pKa values and/or which formed the complexes best able to self-assemble into nanoscale fibers. These systems were shown to be adaptive and responsive to chemical stimuli. We therefore reasoned that if chiral **G2-Lys** was challenged with enantiomeric amines, we may observe interesting enantioselective uptake effects. This would potentially leave one enantiomer unincorporated within the gel and available for further reaction, enabling gel-mediated enantioselective derivatization.

Results and Discussion Gelation with Different Enantiomers

In our previous work,¹⁵ hexylamine was one of the most effective amines for inducing gelation, and we therefore chose chiral amines **C6R/S** to study enantioselection. Compound **G2-Lys** was tested with each enantiomer individually in a 1:1 mixture (both 10 mM) in toluene (0.5 mL). For reproducibility of mixing kinetics, all gels formed in this paper were made using a heat-cool cycle. Both enantiomers were able to induce gelation when mixed with **G2-Lys**. Interestingly, however, the enantiomers produced gels with markedly different T_{gel} values.¹⁶ The gel formed with **C6R** was more thermally stable (80°C) than that formed with **C6S** (67°C), an intriguing result, given that in this complex (mass of over 900 Da), the orientation of just one methyl group has such a pronounced effect on gel stability – a significant impact of relatively low quality chiral information upon the assembly of these complexes.



Fig, 2. CD spectra of **G2-Lys** in the presence of **C6R** and **C6S**, in methylcyclohexane/dioxane (95:5).

Mixtures of **G2-Lys** with each enantiomer were investigated using circular dichroism (CD) spectroscopy in a 95:5 mixture of methylcyclohexane/dioxane (Fig. 2) – the thermal characteristics of the gels in this solvent mix were analogous to those in toluene. The CD spectrum recorded for **G2-Lys** with **C6R** shows a negative Cotton effect band, while that made with **C6S** produces a different spectrum, with a simpler, less intense, broad negative CD band. The peak maximum at 220 nm corresponds to absorbance of the peptides within **G2-Lys** – hydrogen bond interactions between these groups are primarily responsible for gel fibre assembly here.¹⁷ It should be noted that although C6R and C6S are enantiomeric, they produce gels with a diastereomeric relationship, because G2-Lys has the same chirality in each complex. CD clearly demonstrates different chiral organization which – as expected for diastereomeric samples – are not mirror images. Interestingly, the CD signature with C6R is similar to that previously observed with hexylamine (which formed good gels),¹⁵ whereas the CD signature with C6S is similar to that previously observed with octylamine (which formed less effective gels).¹⁵ This therefore suggests that the nanoscale chiral organisation of G2-Lys can be optimised with C6R (and hexylamine) but not with C6S (and octylamine).

To gain further insight into the structure of the gels formed with the different amine enantiomers, dried xerogels were formed under ambient conditions and analysed by FEG-SEM and TEM. The images produced (see supp. info.) indicated that the different amine chiralities gave rise to quite different nanoscale morphologies – with **C6R** giving rise to very small diameter poorly defined fibres, and **C6S** yielding significantly larger and better defined nanofibers. The smaller fibres associated with **C6R** will constitute a network with a larger number of contact points and greater degree of entanglement, supportive of the higher T_{gel} value.¹⁸ Crucially, the different morphologies, must result from differing amine chirality leading to diastereomeric complexes with differing assembly modes.

Gelation using Mixtures of Enantiomeric Amines in Overall 1:1 Stoichiometry with G2-Lys

We then went on to explore how gels made with mixtures of enantiomeric amines would behave, and how the ratio of **C6R** to **C6S** would control this. In all of these initial experiments, the concentration of **G2-Lys** was held at 10 mM and the total amine concentration was also 10 mM (i.e. one stoichiometric equivalent). This ensures that all of the amine should be bound by **G2-Lys** in these experiments.

A series of gels with a 1:1 mix of G2-Lys (10 mM) and varying ratios of C6R/S (10 mM in total) were formed and their $T_{\rm gel}$ values measured. Overall the $T_{\rm gel}$ values show that the thermal stability decreases as an increasing amount of C6S is present and incorporated into the network (Fig. 3A). It takes ca. 20% of C6S before the gel is significantly disrupted and stability starts decreasing. Similarly it takes ca. 20% of C6R being incorporated into the gel network before the thermal stability of the gel increases. It therefore appears that the complex present in the majority can direct the thermal stability of the gel. To investigate whether this change in thermal stability was linked to a change in chiral organization, the same mixtures of G2-Lys with varying ratios of C6R and C6S were analysed by CD spectroscopy in 95:5 methylcyclohexane:dioxane. Surprisingly, the spectra from samples with from 0% to 90% C6S are very similar. Only when the sample is made with entirely C6S did we see a significant change in the CD spectrum (Fig. 3B).

We propose that the reason for the different responses of thermal stability (macroscopic) and CD intensity (nanoscale chirality) lies in the fact that the thermal stability depends on the packing of the whole acid-amine complex, whereas the CD signal corresponds specifically only to the **G2-Lys** component. The presence of the 'wrong' amine therefore impacts on the thermal stability, because this depends on the overall packing of the complex. However, the CD only directly reports on the chiral nano-environment experienced by **G2-Lys** and would suggest that **C6S** can only change the chiral environment ex-

perienced by **G2-Lys** when it is present in very large amounts. This indicates that **G2-Lys** is better suited to achieve its optimal nanoscale chiral assembly mode with **C6R** rather than with **C6S**. As such, the assembly of the two-component complex into the gel (T_{gel}) and the chiral nano-environment experienced by **G2-Lys** (CD) and are not directly correlated. Similar non-correlations have been observed before by Maitra and co-workers¹⁹ highlighting the complex and hierarchical nature of gel formation.



Fig. 3. A: Effect of mixing enantiomers on macroscopic thermal stability of the gel. [G2Lys] = 10 mM, [C6R] + [C6S] = 10 mM. B: Effect of mixing enantiomers on the nanoscale chiral organisation of the gel as recorded in the CD spectrum [G2Lys] = 0.625 mM, [C6R] + [C6S] = 0.625 mM.

We wanted to confirm whether these mixtures produced a single network (a co-assembly) made with **G2-Lys** and both **C6R** and **C6S**, rather than two separate networks, each made from a different diastereomeric complex. This was first probed using differential scanning calorimetry (DSC). In self-sorting gels, where two separate networks are formed, two separate thermal transitions can sometimes be observed.^{5f} Toluene gels (10 mM) were placed in a DSC pan and analysed, but the gels gave either a very small signal or no signal at all, as only a relatively small part of the sample is the gelator. 1,2,3,4-Tetrahydronaphthalene was thus used as a solvent – it is chemically very similar to toluene but with a much higher boiling point and 50 mM gels could now be analysed. These more concentrated samples produced more easily detectable heat changes.

Three samples were measured, using G2-Lys and either C6R, C6S or a 50/50 mixture of both (Table 1). The calorimetry

traces recorded were still of low quality but did show reproducible endo/exotherm peaks for each gel, representative of a phase transition (the exotherms for gel formation were more reproducible than the endotherms for gel breakdown). As expected the gel formed with C6R had thermal transitions at higher temperatures than that formed with C6S, and well separated from it. When a gel with a 50:50 mixture of C6R and C6S was analysed, it showed a single transition, in agreement with a mixed co-assembled network being present rather than two separate, independently melting networks. This transition occurs at temperatures intermediate between those for the gels formed with either C6R or C6S individually. Neither the endotherm nor exotherm of this mixed sample were any broader than those of the other samples, further supporting the conclusion that a co-assembled network is formed. Co-assembly of the different systems into a single network was broadly supported by the FEG-SEM images of the xerogel formed from the mixed gel which shows a single network similar to both the xerogels made with C6R or C6S (see supp. info.)

Table 1. DSC data for G2-Lys (50 mM) with C6R (50 mM) or C6S (50 mM) or both C6R and CSR (25 mM of each).

%C6R	%C6S	Endotherm peak max / °C	Exotherm peak max / °C
100	0	104	93
50	50	95	81
0	100	85	65

Further examination of these gels was then conducted using VT-NMR experiments. Three samples were measured, using G2-Lys (10 mM) and either C6R (10 mM), C6S (10 mM) or a mixture of both (5 mM of each). All of these samples also contained diphenylmethane (10 mM) as a mobile internal standard. If molecules are immobile in the 'solid-like' fibre network, they will not be observed by NMR, whereas if they are in the mobile 'liquid-like' phase they will have quantifiable resonances.^{15,20} The temperature of the sample was increased and ¹H NMR spectra recorded at 5°C intervals. The concentration of mobile G2-Lys at each temperature was plotted as a way of following dissolution of the gel network (see supp info). This allowed us to quantify $T_{100\%}$ (the temperature at which all the gelator is mobile and "visible" in the ¹H NMR). The $T_{100\%}$ and T_{gel} values are similar for each sample, with the $T_{100\%}$ values being slightly higher in each case as this represents the temperature at which the gelator network is completely disbanded on the molecular scale, whereas T_{gel} is the point at which the macroscopic gel network can no longer support itself against the force of gravity. Importantly, the molecular scale data from this NMR experiment is in full agreement with the macroscopic observations, indicating that the thermal stability of the 50/50 gel was intermediate between that containing 100% C6R and that with 100% C6S.

The thermodynamic parameters associated with the gel-sol transition, ΔH_{diss} and ΔS_{diss} values could also be found using the van't Hoff method plotting ln[Sol] against 1/T.^{15,22c} Both values for the sample made with **C6R** are larger than those for the gel made with **C6S**. This would suggest that the **C6R** gel, with a larger entropic gain upon dissolution is likely a highly organised, more rigid and closely packed structure. The larger endothermic change upon dissolution of this **C6R** network

indicates that this network is better stabilised by hydrogen bonding interactions, again indicative of a more closely packed structure. The C6S sample is less well organised and less able to take advantage of hydrogen bonding. Interestingly, the gel produced with a mixture of C6R and C6S has much lower $\Delta H_{\rm diss}$ and $\Delta S_{\rm diss}$ values than for either of the gels with individual enantiomers. We suggest that this is due to the network having to accommodate both C6R and C6S into the fibres and the diastereomeric complexes formed being unable to pack as efficiently into overall supramolecular aggregates. At first, it might therefore seem surprising that the gel based on the C6R/S mixture is not also thermally less stable than either of the gels made with individual enantiomers given it has a smaller $\Delta H_{\rm diss}$. However, the fact that it has a $T_{\rm gel}$ value between those of the gels formed with either C6R or C6S alone is a result of the balance between ΔH_{diss} and ΔS_{diss} . The entropic cost of gelating 50/50 C6R and C6S drops very significantly, and therefore even though the enthalpy of gelation is lower, the relative lack of order within the mixed co-assembled gel more than compensates for it in thermodyamic terms.

Table 2. Comparison of molecular $(T_{100\%})$ and materials (T_{gel}) properties for gels formed with enantiomeric amines C6R/S and the thermodynamic parameters associated with the gel-sol transition.

C6R	C6S	r _{gel} / ⁰C	<i>T</i> _{100%} / ⁰C	$\Delta H_{\rm diss}$ / kJmol ⁻¹	$\Delta S_{\rm diss}$ / J mol ⁻¹ K ⁻¹
100%	0%	80	83	78.3	181
50%	50%	74	77	45.4	91
0%	100%	67	69	66.9	157

In summary, the chirality of the amine mixed with **G2-Lys** has a significant effect on the assembly of the resulting complexes and a pronounced effect on the gel that is produced. The chirality of the amine profoundly affects the molecular-scale assembly of complexes in solution (CD, NMR), the nanoscale morphology of fibrous network formed (FEG-SEM and TEM) and ultimately the macroscopic stability of the material produced (T_{gel}). Furthermore, when a gel is formed from a mixture of **G2-Lys** and **C6R/S** with varying ratios of enantiomers, a co-assembled network appears to be formed rather than individual self-sorted assemblies.

Component Selection Experiments

In all of the systems described above, there was stoichiometric equivalence between acid (**G2-Lys**) and total amine (**C6R/S**), meaning all of the amine should be bound in each case. An alternative experimental approach would provide **G2-Lys** with a choice between different amines – a 'component selection experiment'.⁴ To the best of our knowledge, this has not previously been done with regard to chiral selection within gels.

We therefore made gels made with a 1:1:1 mixture of G2-Lys, C6R and C6S at concentrations ranging from 2-10 mM and the T_{gel} values were measured. In each experiment, G2-Lys effectively has a choice between the two amines – it could bind all of one enantiomer, all of the other, or any ratio in between. The T_{gel} values were compared to those of gels formed with G2-Lys and C6R or C6S only (Fig. 4). The gels made from a mixture of both enantiomers had T_{gel} values almost identical to the more stable gels formed with only the C6R enantiomer. This gives a strong indication that G2-Lys has selected to as-

semble its gel network primarily with **C6R** rather than **C6S**, which we would propose remains unselected, and mobile in solution (see below). Interestingly, we know from the analysis of the 1:0.5:0.5 mixture described above, in which **G2-Lys** is effectively forced by stoichiometry to interact with 50% of each of the amines, that the T_{gel} value was only 74°C (10 mM). As such, we can be confident that in the 1:1:1 component selection system, we are indeed seeing significant enantioselectivity, with T_{gel} being 79°C – much closer to the value for 100% **C6R** (80°C) than for the gel in which 50% each of **C6R** and **C6S** have been taken up (74°C).



Fig, 4. T_{gel} values measured for **G2-Lys** (1 eq.) with **C6R** (1 eq.), **C6S** (1 eq.) or **C6R** and **C6S** (1 eq. of each).

The xerogel produced from the 1:1:1 mixture of **G2-Lys**, **C6R** and **C6S** was imaged by FEG-SEM (see supp info). The images show a sample with a very ill-defined morphology. No distinct fibres are easily visible – similar to the SEM images seen with **C6R** alone. However, given the limitations of FEG-SEM for these very narrow nanofibers, we also made use of small angle x-ray scattering experiments (SAXS) to probe the morphology further. SAXS data for the solvated gels showed the gels made with **C6R** and **C6S** had different cylinder form factors of 4 and 3 nm respectively. The component selecting 1:1:1 gel had a cylinder form factor of 4 nm – the same as the gel made with only **C6R**. In addition, the 1:1:1 xerogel had Bragg peaks which were more similar to the gel formed with **C6R** alone than with **C6S** (see supp info).

Further analysis of the gel with a 1:1:1 mixture of **G2-Lys**, **C6R** and **C6S** was performed using VT-NMR in toluene- d_8 (all components 10 mM). The concentration of **G2-Lys** visible in solution at each point was used to determine ΔH_{diss} and ΔS_{diss} . The gel has $\Delta H_{\text{diss}} = 56.0 \text{ kJ mol}^{-1}$ and $\Delta S_{\text{diss}} = 122 \text{ J}$ mol⁻¹K⁻¹ These values are lower than for **G2-Lys** made with either single enantiomer, but higher than for the 1:0.5:0.5 mixture. This suggests that when **G2-Lys** has a choice between **C6R** and **C6S**, it is not identical to the gel formed with **C6R**, but neither is it anywhere near a 50:50 mix of enantiomers. It should also be noted that the additional equivalent of amine present in the gel will be in dynamic exchange with the amine bound to the solid-like fibres, which might be expected to decrease the thermodynamic stability and order of the gel.

We wanted to use NMR methods to directly quantify the amount of each amine free in solution (and by inference that immobilised in the solid-like fibres), as we hoped to determine the enantioselectivity of this self-assembling system. We attempted to do this using a chiral shift reagent approach, but were unable to get sufficient peak separation between the diastereomers formed from **C6R** and **C6S** in the liquid-like phase. This led us to consider alternative approaches.

We decided to use a chiral derivatization reagent to probe these component-selecting chiral gels further. Gels are fascinating media for organic reactions – they are solvated and porous, hence reagents and catalysts can be diffused in and out of them very simply.²¹ In this case, we hoped to use the gelation event to facilitate enantiomer separation by preferentially immobilising one enantiomer into the gel nanofibers, allowing the mobile enantiomer to diffuse out of the gel and react with a chiral substrate. We reasoned this would allow us to infer how much of each enantiomer was immobilised within the gel.



Fig, 5, Reaction between **C6R/S** and (*S*)-methylbenzyl isocyanate, giving rise to diastereoisomeric products.

We formed the 1:1:1 gel with **G2-Lys**, **C6R** and **C6S** in toluene (0.5 mL). After gelation, a further amount of toluene (0.5 mL) was gently pipetted on top of the gel and the sample was left for 24 hours to allow any amine not included in the solidlike gel network to diffuse throughout the entire volume of toluene. The supernatant solvent was removed using a pipette and placed in a round bottom flask and an excess of (*S*)methylbenzyl isocyanate was added to derivatize all of the chiral amine that had diffused into the toluene. This converts the amine enantiomers into two diastereoisomeric ureas (Fig. 5), which, it was hoped, could be distinguished and quantified by ¹H NMR.

After reaction, the solvent was evaporated and reaction success determined by NMR and MS analysis. The solid was redissolved in CDCl₃, analysed by ¹H NMR and compared to samples prepared using the same method but with either **C6R** or **C6S** alone. The difference in chemical shift between the peaks of the CH₃CH protons (originally on the amine) of each diastereomeric urea was 0.085 ppm. There was also a measurable difference in the chemical shift of the peak of the terminal CH₃CH₂ group of each diastereomer ($\Delta \delta = 0.072$ ppm). Therefore, when analysing the mixed gel, the resonances asso-

ciated with both diastereomers were easily resolved and the relative amounts of each could be simply quantified (Fig. 6). Of all the urea, 20% was derived from **C6R** and 80% from **C6S**. This would indicate that the solid-like gelator fibre network is formed from the inverse composition (80% **C6R** and 20% **C6S**). This result demonstrates unambiguously that there is indeed selective uptake of the **C6R** enantiomer that forms the most stable gel network into the gel fibres by **G2-Lys**. We suggest that gels of this type maybe of interest for applications in chiral resolution and enantioselective reaction pathways – especially given that they can select between relatively low quality chiral information.



Fig. 6. NMR spectra of diasteromeric mixture arising from reaction of **C6R/S** mixture diffused out of gel made with **G2-Lys** after reaction with (*S*)-methylbenzyl isocyanate

Self-assembly of these multi-component gel nanofibers occurs in several hierarchical steps, with the initial key steps being: (i) formation of acid-base complexes, (ii) uni-directional self-assembly of these complexes (Fig. 7).¹⁵ We wanted to determine whether the apparent selectivity of **G2-Lys** for the *R* amine was associated with the initial formation of the acid-base complex (step (i)) or self-assembly of the diastereomeric complexes into gel fibres (step (ii)).

To examine the acid-base formation step, NMR titration experiments were carried out in which the concentration of either **C6R** or **C6S** remained constant while the concentration of **G2-Lys** was increased. This titration was carried out in a solvent ($CDCl_3$) which did not support self-assembly of the complexes and therefore effectively isolated the initial complexation event (step (i).



Fig. 7. Schematic of self-assembly showing step (i) acid-base formation and step (ii) self-assembly of the complexes formed.

The change in chemical shift of the CH peak of **C6R** or **C6S** as the concentration of **G2-Lys** increases was almost identical (Fig. 8). To quantify binding, stability constants were fitted using WinEQNMR2²² and a 1:1 binding model. With **C6R** $\log K = 4.30$, with **C6S** $\log K = 4.37 (\pm 15\%)$, clearly showing that, within error, the stability constant of the complex is the same in each case. Acid-base complex formation (step(i)) is therefore not responsible for the selective uptake of one amine enantiomer, and step (ii) must be more important.



Fig. 8. NMR titration of **G2-Lys** with **C6R** or **C6S** (2.0 mM) in $CDCl_3$ indicating how the CH proton at the chiral centre of the amine is perturbed on addition of the carboxylic acid.

We used infrared (IR) spectroscopy to probe this further. We measured IR spectra of gels formed in toluene from G2-Lys (10 mM) with C6R (10 mM) or C6S (10 mM). The IR spectra were almost identical, reflecting the fact that both diastereomeric complexes give rise to gelation. However, there was a reproducible difference in the IR absorbance associated with the N-H (amide) stretch (ca. 3300 cm⁻¹), with the N-H peak being clearly split in two in the presence of C6R (see supp info for data). This reflects that the chirality of the amine has a direct effect on the self-assembly of G2-Lys which is underpinned by intramolecular amide-amide hydrogen bond interactions. Furthermore, when we measured the IR spectrum of the gel formed from G2-Lys (10 mM) with both C6R and C6S (both 10 mM), the N-H stretch was identical to that observed for C6R alone (see supp info). We can therefore conclude the selective uptake of one amine enantiomer is driven by preferential self-assembly of the complex formed with C6R rather than that with C6S (step (ii)). We suggest that the steric influence of the methyl group attached to the chiral centre on C6R/C6S has a significant influence on the way these complexes can pack, with C6R enabling better interaction between G2-Lys peptides, while C6S compromises G2-Lys assembly and enforces a chiral adjustment.

We then wanted to prove that this component selection was a true thermodynamic preference, not simply the result of a kinetically trapped gel forming on cooling the sample. Kinetic trapping could occur if, as the sample was cooled, the network with **G2-Lys** and **C6R** preferentially formed simply because it has the higher T_{gel} value: i.e., on reaching the temperature at which the network with **C6S** could actually start to form, all of the **G2-Lys** would already have been "used" in forming a network with **C6R**. To test the reversibility of component selection and gel-assembly, a gel with **G2-Lys** and **C6S** was preformed and a solution of **C6R** pipetted onto the gel and allowed to diffuse into the sample for 5 days (Fig. 9). If the

system is kinetically trapped, it should not change. The proportion of each amine in the network was calculated by derivatizing the excess solution-phase amine with (S)methylbenzyl isocyanate as already described. After equilibration, an excess of C6S was found in solution - clearly it has been displaced from the gel network by C6R. In the solution phase only 33% of the amine is C6R and 67% is C6S. Therefore the gelator network is 67% C6R and 33% C6S, demonstrating that the preference for C6R is primarily thermodynamic and that these gels are responsive - adapting and evolving their compositions in response to chemical stimulus. The selective uptake of C6R is slightly lower than observed for the gel formed directly from the 1:1:1 mixture with a heatcool cycle (80% C6R, 20% C6S). We suggest that for the gel formed by displacement the network needs to re-organise to accommodate the new amine, slightly lowering selectivity.



Fig. 9. Schematic of thermodynamically controlled gel evolution on addition of **C6R** to a gel made from **G2-Lys** and **C6S**.

In summary, when a 1:1:1 mixture of **G2-Lys**, **C6R** and **C6S** is used to form a gel, the resulting gelator network is mainly composed of **G2-Lys** and **C6R**, whilst most of the **C6S** is left in the liquid-like phase – enantioselective component selection. Most importantly, this combination of experimental approaches allows us to directly connect macroscopic performance (T_{gel}) with the molecular level behaviour (NMR). We have also clearly demonstrated that these gels are adaptive and responsive to changes in their external environment.

Fascinatingly, the ability of chiral gels to induce differential uptake and reactivity in a mixture of enantiomeric amines demonstrates how chirality can be simply passed on from one source to another, with the gel matrix acting to preferentially remove one enantiomer from the system. Porous gels are thus fascinating media in which chiral information may be transferred and/or amplified.²³ Such a mechanism may have been relevant in prebiotic evolution of homochiral systems – it has often been noted that the interior of a cell is a gel-like matrix, and it has been suggested that simple gels may have played a pre-biotic role before the evolution of membranes.²⁴

Probing Component Selection with a Range of Amines

We then applied these techniques to mixtures of other chiral amines (Fig. 10). In each case, we used the chirality of **G2-Lys** to select between enantiomeric amines. This was rapidly tested using T_{gel} evaluation and the reaction of excess amine with (*S*)-methyl isocyanate combined with NMR characterisation. We took care to choose amines which gave rise to diastereomeric products with (*S*)-methyl isocyanate having good solubilities and distinguishable NMR peaks – for examples of amines where this was not possible, see the supporting information. We hoped to determine if:

a) molecular-level chiral selectivity is a general rule in these systems,

- b) the sense of chiral preference for *R* enantiomeric amines is retained,
- c) macroscopic thermal performance can be rationalised in terms of molecular level chiral selectivity.



Fig. 10. Selection of amines used successfully in component selection experiments.

Initially, we tested chiral aliphatic amines, which have a methyl group adjacent to the primary amine as the source of chirality. The NMR derivatization experiment indicated that C4iR, C8R and C9R are selected by G2-Lys in preference to C4iS, C8S and C9S respectively (Table 3). As such, we note that the chiral selectivity in all of these systems matches that for C6R/S in which the *R* enantiomer is preferred because the resulting complex better assembles into gel fibres.

The chiral preference observed by the derivatization approach was also reflected in the thermal stability of the mixed gels. Firstly, it should be noted that for all of these amines, the Renantiomer forms a more thermally stable gel than the S (as for C6R/S, Table 3). Furthermore, the 1:0.5:0.5 gels, in which G2-Lys is forced to interact with both enantiomers equally (i.e., 50% R, 50% S) had T_{gel} values somewhere in between the R and S extremes. Considering the T_{gel} values for the component selection 1:1:1 experiment ($T_{gel obs}$), in which G2-Lys has a choice of which amine to interact with, it is evident that, in all cases, these $T_{gel obs}$ values lie between those for the 50/50 mixture and those for 100% R (Table 3). As such, these macroscopic observations are in agreement with the molecular scale information which indicates preferential incorporation of R amines into the gel. As such, we propose the molecularscale chiral information, enantioselected by G2-Lys, is being read through into the macroscopic performance of the gel.

Table 3. Quantification of % amine incorporation in fibres & enantiomeric excess (ee) of uptake into fibers using molecular-scale derivatisation (NMR), and macroscopic-scale analysis of $T_{\rm gel}$ data.

	Molecular Scale			Macroscopic Behaviour			
amine	% <i>R</i> in fibres	%S in fibres	ee	$T_{\rm gel}$ (R)	T _{gel} (R/S)	$T_{\rm gel}$ (S)	$T_{\rm gel}$ obs
C4i	68%	32%	36%	54	48	37	50
C6	80%	20%	60%	80	74	67	79
C8	83%	17%	66%	70	60	54	64
C9	79%	21%	58%	61	52	46	54
Tol	68%	32%	36%	60	54	53	59
1-Nap	62%	38%	24%	79	71	54	74
2-Nap	54%	46%	8%	57	53	53	58

We then tested some chiral primary amines with pendant aromatic groups, **TolR/S**, **1-NapR/S** and **2-NapR/S**. Similarly to the aliphatic amines, the *R* enantiomer was preferentially taken into the gel over the *S* version and the thermal stability reflected this chiral preference (Table 3). However, in all cases, the degree of chiral selectivity was somewhat lower than observed for aliphatic amines and had all but disappeared for **2-NapR/S**. We propose that the greater steric hindrance of the aromatic groups may hinder the chiral directing preference of **G2-Lys** during self-assembly.

Conclusions

In conclusion, we have demonstrated that the chirality of the amine used to form a gel with G2-Lys has a large bearing on the assembly of the resulting diastereomeric complexes into self-assembled gel networks. This has been investigated most thoroughly using C6R/S but has also been observed using a range of other amines, all of which have what would otherwise be regarded as poor quality chiral centres. This demonstrates the remarkable and powerful effect of chirality on gelation of these systems. Importantly, the selective incorporation of one enantiomer of an amine over the other into the gel network has been demonstrated, and in all cases, the R amine that forms the most stable gel network is primarily selected for incorporation into the gel. The thermodynamic control over this process has been proven by forming a gel exclusively with C6S and then allowing C6R to diffuse through the sample and displace C6S from the solid-like nanofibers. This forms a new nanoscale network and shows that these gels can adapt and evolve in response to chemical stimuli to which they are exposed. Finally, it has been demonstrated that excess amine - which remains unincorporated within the gel network - can diffuse out and selectively react with a chiral isocyanate. This allowed us to quantify the enantioselectivity of component selection within these gels, but also illustrates how gels can act as selective reservoirs of potential reagents, releasing them on demand to yield (in this case) one enantiomer (of amine) in preference to another. We suggest that the lessons learned in this research may go on to be applied in enantioseparation, asymmetric synthesis, or the development of hydrogels which can play active roles in pre-biotic reaction pathways.

SUPPORTING INFORMATION AVAILABLE

Full details of amine gelation studies including: T_{gel} data for all amines at different enantiomeric ratios, FEG-SEM imaging, VT-NMR data, IR data, full data from selectivity studies including NMR spectra of diastereomeric products. This information is available free of charge via the Internet at http://pubs.acs.org/.

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References

1. (a) Molecular Gels: Materials with Self-Assembled Fibrillar Networks, ed. Terech, P.; Weiss, R. G. Springer, Dordrecht, Netherlands, 2006. (b) van Esch, J. H. Langmuir **2009**, 25, 8392-8394. (c)

Steed, J. W. Chem. Commun. 2011, 47, 1379-1383. (d) Banerjee, S.; Das, R. K.; Maitra, U. J. Mater. Chem. 2009, 19, 6649-6687. (e) Hirst, A. R.; Escuder, B.; Miravet, J. F.; Smith, D. K. Angew. Chem. Int. Ed. 2008, 47, 8002-8018. (f) Dawn, A.; Shiraki, T.; Haraguchi, S.; Tamaru, S.-i.; Shinkai, S.; Chem. Asian J. 2011, 6, 266-282.

2. (a) Hirst, A. R.; Smith, D. K. *Chem. Eur. J.* **2005**, *11*, 5496-5504. (b) Buerkle, L. E.; Rowan, S. J. *Chem. Soc. Rev.* **2012**, *41*, 6089-6102.

3. For early examples see: (a) Hanabusa, K.; Miki, T.; Taguchi, Y.; Koyama, T.; Shirai, H. *J. Chem. Soc., Chem. Commun.* **1993**, 1382-1384. (b) Inoue, K.; Ono, Y.; Kanekiyo, Y.; Ishi-I, T.; Yoshihara, K.; Shinkai, S. *J. Org. Chem.* 1999, **64**, 2933-2937. (c) Partridge, K. S.; Smith, D. K.; Dykes, G. M.; McGrail, P. T. *Chem. Commun.*, **2001**, 319-320.

4. (a) Sreenivasachary, N.; Lehn, J.-M. Proc. Natl. Acad. Sci. USA 2005, 102, 5938-5943. (b) Buhler, E.; Sreenivasachary, N.; Candau, D.-J.; Lehn, J.-M. J. Am. Chem. Soc. 2007, 129, 10058-10059. (c) Hirst, A. R.; Miravet, J. F.; Escuder, B.; Noirez, L.; Castelletto, V.; Hamley, I. W.; Smith, D. K. Chem. Eur. J. 2009, 15, 372-379. (d) Wang, G.-T.; Lin, J.-B.; Jiang, X.-K.; Li, Z.-T. Langmuir 2009, 25, 8414-8418. (e) R. J. Williams, A. M. Smith, R. Collins, N. Hodson, A. K. Das and R. V. Ulijn, Nat. Nanotechnol., 2009, 4, 19-24. (f) A. K. Das, A. R. Hirst and R. V. Ulijn, Faraday Discuss., 2009, 143, 293-303.(g) Buchs, B.; Fieber, W.; Vigoroux-Elie, F.; Sreenivasachary, N.; Lehn, J.-M.; Herrmann, A. Org. Biomol. Chem. 2011, 9, 2906-2919. (h) Smith, M. M.; Edwards, W.; Smith, D. K. Chem. Sci. 2013, 4, 671-676.

5. (a) Hirst, A. R.; Huang, B.; Castelletto, V.; Hamley, I. W.; Smith, D. K. Chem. Eur. J. 2007, 13, 2180-2188 (b) Sugiyasu, K.; Kawano, S. I.; Fujita, N.; Shinkai, S. Chem. Mater. 2008, 20, 2863-2865. (c) Moffat, J. E.; Smith, D. K. Chem. Commun. 2009, 316-318. (d) Moffat, J. R.; Coates, I. A.; Leng, F. J.; Smith, D. K. Langmuir, 2009, 25, 8786-8793. (e) Cicchi, S.; Ghini, G.; Lascialfari, L.; Brandi, A.; Betti, F.; Berti, D.; Baglioni, P.; Di Bari, L.; Pescitelli, G.; Mannini, M.; Caneschi, A. Soft Matter 2010, 6, 1655-1661. (f) Smith, M. M.; Smith, D. K. Soft Matter 2011, 7, 4856-4860. (g) Das, A.; Ghosh, S. Chem. Commun. 2011, 47, 8922-8924. (h) Velazquez, D. G.; Luque, R. Chem. Eur. J. 2011, 17, 3847-3849. (i) Adhikari, B.; Nanda, J.; Banerjee, A. Soft Matter 2011, 7, 8913-8922. (j) Morris, K. L.; Chen, L.; Raeburn, J.; Sellick, O. R.; Cotanda, P.; Paul, A.; Griffiths, P. C.; King, S. M.; O'Reilly, R. K.; Serpell, L. C.; Adams, D. J. Nature Commun. 2013, 4, 1480. (k) Smith, M. M.; Edwards, W.; Smith, D. K. Chem. Sci. 2013, 4, 671-676. (1) Cornwell, D. J.; Okesola, B. O.; Smith, D. K. Soft Matter 2013, DOI: 10.1039/C3SM51967H.

6. (a) Lehn, J.-M. Angew. Chem. Int. Ed. **2013**, 52, 2836-2850. (b) Ludlow, R. F.; Otto, S. Chem. Soc. Rev. **2008**, 37, 101-108.

7. For reviews see: (a) Smith, D. K. Chem. Soc. Rev.2009, 38, 684-694. (b) Brizard, A.; Oda, R.; Huc, I. Top. Curr. Chem. 2005, 256, 167-218. (c) Wang, Y.; Xu, J.; Wang, Y.; Chen, H. Chem. Soc. Rev. 2013, 42, 2930-2962. For selected original research papers: (d) Ramanathan, N; Currie, A. L.; Ross Colvin, J. Nature, 1961, 190, 779-781. (e) Tachibana, T.; Kambara, H. J. Am. Chem. Soc. 1965, 87, 3015-3016. (f) Aggeli, A.; Nyrkova, I. A.; Bell, M.; Harding, R.; Carrick, L.; McLeish, T. C. B.; Semenov, A. N.; Boden, N. Proc. Natl. Acad. Sci. USA 2001, 98, 11857-11862. (g) Friggeri, A.; van der Pol, C.; van Bommel, K. J. C.; Heeres, A.; Stuart, M. C. A.; Feringa, B. L.; van Esch, J. Chem. Eur. J. 2005, 11, 5353-5361. (h) Iavicoli, P.; Xu, H.; Feldborg, L. N.; Linares, M.; Paradinas, M.; Stafstrom, S.; Ocal, C.; Nieto-Ortega, B. L.; Casado, J.; Navarette, J. T. L.; Lazzaroni, R.; De Feyter, S.; Amabilino, D. B. J. Am. Chem. Soc. 2010, 132, 9350-9362.

8. For typical examples see: (a) Koga, T.; Matsuoka, M.; Higashi, N. J. Am. Chem. Soc. 2005, 127, 17596-17597. (b) Hirst, A. R.; Smith, D. K.; Feiters, M. C.; Geurts, H. P. M. Chem. Eur. J. 2004, 10, 5901-5910. (c) Kim, J.-U.; Schollmeyer, D.; Brehmer, M.; Zentel, R. J. Coll. Interface Sci. 2011, 357, 428-433. (d) Xie, Z.; Stepanenko, V.; Radacki, K.; Würthner, F. Chem. Eur. J. 2012, 18, 7060-7070. (e) Bredikhin, A. A.; Bredikhina, Z. A.; Akhatova, F. S.; Gubaidullin, A. T. Chem. Commun. 2010, 46, 3523-3525.

9. (a) Messmore, B. W.; Sukerkar, P. A.; Stupp, S. I. J. Am. Chem. Soc. 2005, 127, 7992-7993. (b) Cicchi, S.; Pescitelli, G.; Lascialfari,

L.; Ghini, G.; Di Bari, L.; Brandi, A.; Busotti, L.; Aysbeha, T.; Marcelli, A.; Foggi, P.; Berti, D.; Mannini, M. *Chirality* **2011**, *23*, 833-840.

 (a) Frkanec, L.; Zinic, M. Chem. Commun. 2010, 46, 522-537.
(b) J. Makarević, M. Jokić, Z. Raza, Z. Štefanić, B. Kojić-Prodić and M. Žinić, Chem. Eur. J. 2003, 9, 5567-5580. (c) Kaplar, V.; Frkanec, L.; Vujicic, N. S.; Zinic, M. Chem. Eur. J. 2010, 16, 3066-3082. (d) Nagy, K. J.; Giano, M. C.; Jin, A.; Pochan, D. J.; Schneider, J. P. J. Am. Chem. Soc. 2011, 133, 14975-14977. (e) He, Y.; Bian, Z.; Kang, C.; Gao, L. Chem. Commun. 2011, 47, 1589-1591.

11. (a) Nam, S. R.; Lee, H. Y.; Hong, J.-I. *Chem. Eur. J.* **2008**, *14*, 6040-6043. (b) Cai, W.; Wang, G.-T.; Du, P.; Wang, R.-X.; Jiang, X.-K.; Li, Z.-T. *J. Am. Chem. Soc.* **2008**, *130*, 13450-13459. (c) Rodriguez-Llansola, R.; Hermida-Merino, D.; Nieto-Ortega, B.; Ramirez, F. J.; Navarrete, J. T. L.; Casado, J.; Hamley, I. W.; Escuder, B.; Hayes, W.; Miravet, J. F. *Chem. Eur. J.* **2012**, *18*, 14725-14731.

12. (a) Zhang, S. Y.; Yang, S. Y.; Lan, J. B.; Yang, S. J.; You, J. S. *Chem. Commun.* **2008**, 6170-6172. (b) Azeroual, S.; Surprenant, J.; Lazzara, T. D.; Kocun, M.; Tao, Y.; Cuccia, L. A.; Lehn, J.-M. *Chem. Commun.* **2012**, *48*, 2292-2294.

13. (a) de Loos, M.; van Esch, J.; Kellogg, R. M.; Feringa, B. L. Angew. Chem. Int. Ed. **2001**, 40, 613-616. (a) Chen, X.; Huang, Z.; Chen, S.-Y.; Li, K.; Yu, X.-Q.; Pu, L. J. Am. Chem. Soc. **2010**, 132, 7297-7299. (b) Mukai, M.; Minamikawa, H.; Aoyagi, M.; Asakawa, M.; Shimizu, T.; Kogiso, M. Soft Matter **2012**, 8, 1197-11981. (c) Jintoku, H.; Takafuji, M.; Oda, R.; Ihara, H. Chem. Commun. **2012**, 48, 4881-4883. (d) Tu, T.; Fang, W.; Bao, X.; Li, X.; Dötz, K. H. Angew. Chem. Int. Ed. **2011**, 50, 6601-6605. (e) Cao, H.; Zhu, X.; Liu, M. Angew. Chem. Int. Ed. **2013**, 52, 4122-4126.

14. (a) Zheng, Y.-S.; Ran, S.-Y.; Hu, T.-J.; Liu, X.-X. *Chem. Commun.* **2009**, 1121-1123. (b) A. Tripathi, A. Kumar and P. S. Pandey, *Tetrahedron Lett.* **2012**, *53*, 5745-5748.

15. Edwards, W.; Smith, D. K. J. Am. Chem. Soc. 2013, 135, 5911-5920

16. Raghavan, S. R.; Cipriano, B. H. 'Gel Formation: Phase Diagrams using Tabletop Rheology and Calorimetry', Chapter 8 in *Molecular Gels, Materials with Self-Assembled Fibrillar Networks*, Ed. Weiss, R. G.; Terech, P. Springer, Dordrecht, Netherlands, 2006.

17. (a) Hirst, A. R.; Smith, D. K.; Feiters, M. C.; Geurts, H. P. M. J. Am. Chem. Soc. 2003, 125, 9010-9011. (b) Hirst, A. R.; Smith, D. K.; Feiters, M. C.; Geurts, H. P. M. Langmuir 2004, 20, 7070-7077. (c) Hirst, A. R.; Smith, D. K. Org. Biomol. Chem. 2004, 2, 2965-2971. (d) Huang, B.; Hirst, A. R.; Smith, D. K. Castelletto, V.; Hamley, I. W. J. Am. Chem. Soc. 2005, 127, 7130-7139. (e) Hirst, A. R.; Smith, D. K.; Smith, D. K.; Harrington, J. P. Chem. Eur. J. 2005, 11, 6552-6559. (f) Hardy, J. R.; Hirst, A. R.; Smith, D. K. Soft Matter 2012, 8, 3399-3406.

18. Hardy, J. G.; Hirst, A. R.; Ashworth, I. A.; Brennan, C.; Smith, D. K. *Tetrahedron* **2007**, *67*, 7397-7406.

19. Das, R. K.; Kandanelli, R.; Linnanto, J.; Bose, K.; Maitra, U. Langmuir **2010**, *26*, 16141-16149.

20. (a) Shapiro, Y. E.; *Progr. Polym. Sci.* 2011, **36**, 1184-1253. (b) Escuder, B.; LLusar, M.; Miravet, J. F. *J Org. Chem.* **2006**, 71, 7747-7752. (c) Hirst, A. R.; Coates, I. A.; Boucheteau, T. R.; Miravet, J. F.; Escuder, B.; Castelletto, V.; Hamley, I. W.; Smith, D. K. *J. Am. Chem. Soc.* **2008**, 130, 9113-9121. (d) Nebot, V. J.; Armengol, J.; Smets, J.; Prieto, S. F.; Escuder, B.; Miravet, J. F. *Chem. Eur. J.* **2012**, *18*, 4063-4072. (e) Makarević, J.; Jokić, M.; Perić, V.; Tomasić, V.; Kojić-Prodić, B.; Žinić, M. Chem. Eur. J. **2001**, *15*, 3328-3341.23.

21. (a) Diaz, D. D.; Kuhbeck, D.; Koopmans, R. J. *Chem. Soc. Rev.* **2011**, *40*, 427-448. (b) Escuder, B.; Rodriguez-Llansola, F.; Miravet, J. F. *New J. Chem.* **2010**, *34*, 1044-1054.

22. Hynes, M. J. J. Chem. Soc., Dalton Trans. 1993, 311-312.

23. (a) Dawn, A.; Fujita, N.; Haraguchi, S.; Sada, K.; Shinkai, S. *Chem. Commun.* **2009**, 2100-2102. (b) Rodriguez-Llansola, F.; Escuder, B.; Miravet, J. F. *Org. Biomol. Chem.* **2009**, *7*, 3091-3094. (c) Rodriguez-Llansola, F.; Miravet, J. F.; Escuder, B. *Chem. Eur. J.* **2010**, *16*, 8480-8486.

24. (a) Trevors, J. T.; Pollack, G. H. *Prog. Biophys. Mol. Biol.* **2005**, *89*, 1-8. (b) Trevors, J. T.; Pollack, G. H. *Biochimie*, **2012**, *94*, 258-262.

Graphical Abstract

