



University of HUDDERSFIELD

University of Huddersfield Repository

Ding, W., Stewart, D.I., Humphreys, P.N., Rout, S.P. and Burke, I.T.

Role of an organic carbon-rich soil and Fe(III) reduction in reducing the toxicity and environmental mobility of chromium(VI) at a COPR disposal site

Original Citation

Ding, W., Stewart, D.I., Humphreys, P.N., Rout, S.P. and Burke, I.T. (2016) Role of an organic carbon-rich soil and Fe(III) reduction in reducing the toxicity and environmental mobility of chromium(VI) at a COPR disposal site. *Science of The Total Environment*, 541. pp. 1191-1199. ISSN 0048-9697

This version is available at <http://eprints.hud.ac.uk/28667/>

The University Repository is a digital collection of the research output of the University, available on Open Access. Copyright and Moral Rights for the items on this site are retained by the individual author and/or other copyright owners. Users may access full items free of charge; copies of full text items generally can be reproduced, displayed or performed and given to third parties in any format or medium for personal research or study, educational or not-for-profit purposes without prior permission or charge, provided:

- The authors, title and full bibliographic details is credited in any copy;
- A hyperlink and/or URL is included for the original metadata page; and
- The content is not changed in any way.

For more information, including our policy and submission procedure, please contact the Repository Team at: E.mailbox@hud.ac.uk.

<http://eprints.hud.ac.uk/>



Role of an organic carbon-rich soil and Fe(III) reduction in reducing the toxicity and environmental mobility of chromium(VI) at a COPR disposal site



Weixuan Ding^a, Douglas I. Stewart^{b,*}, Paul N. Humphreys^c, Simon P. Rout^c, Ian T. Burke^{a,*}

^a School of Earth and Environment, University of Leeds, Leeds, LS2 9JT, UK

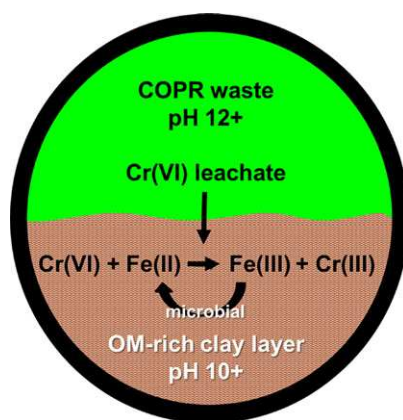
^b School of Civil Engineering, University of Leeds, Leeds LS2 9JT, UK

^c Department of Biological Sciences, School of Applied Sciences, University of Huddersfield, Huddersfield HD1 3DH, UK

HIGHLIGHTS

- A Fe(II)-containing organic rich soil is found beneath a COPR waste tip.
- Cr(VI) is removed from COPR leachate by reaction with Fe(II), producing Cr(OH)₃.
- The soil contains alkaliphilic fermentative and Fe(III)-reducing bacteria.
- The bacterial community maintains a redox barrier reducing the spread of Cr(VI).
- Organic matter addition to OM poor soils is a low cost, long-term treatment.

GRAPHICAL ABSTRACT



ARTICLE INFO

Article history:

Received 13 August 2015

Received in revised form 29 September 2015

Accepted 29 September 2015

Available online 11 November 2015

Editor: D. Barcelo

Keywords:

COPR
Alkaline
Chromium
Iron
Bioremediation

ABSTRACT

Cr(VI) is an important contaminant found at sites where chromium ore processing residue (COPR) is deposited. No low cost treatment exists for Cr(VI) leaching from such sites. This study investigated the mechanism of interaction of alkaline Cr(VI)-containing leachate with an Fe(II)-containing organic matter rich soil beneath the waste. The soil currently contains 0.8% Cr, shown to be present as Cr(III)(OH)₃ in EXAFS analysis. Lab tests confirmed that the reaction of Cr(VI) in site leachate with Fe(II) present in the soil was stoichiometrically correct for a reductive mechanism of Cr accumulation. However, the amount of Fe(II) present in the soil was insufficient to maintain long term Cr(VI) reduction at historic infiltration rates. The soil contains a population of bacteria dominated by a *Mangroviflexus*-like species, that is closely related to known fermentative bacteria, and a community capable of sustaining Fe(III) reduction in alkaline culture. It is therefore likely that in situ fermentative metabolism supported by organic matter in the soil produces more labile organic substrates (lactate was detected) that support microbial Fe(III) reduction. It is therefore suggested that addition of solid phase organic matter to soils adjacent to COPR may reduce the long term spread of Cr(VI) in the environment.

© 2015 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

* Corresponding authors.

E-mail addresses: D.I.Stewart@leeds.ac.uk (D.I. Stewart), I.T.Burke@leeds.ac.uk (I.T. Burke).

1. Introduction

Chromium chemicals are produced for a range of industries including; leather tanning, chromium plating, pigments, refractories and magnetic media (Darrie, 2001). Chromium has two environmentally stable oxidation states; Cr(III) and Cr(VI). Cr(III) is an essential trace element for mammals (Vincent, 2000; Cefalu and Hu, 2004), which has low solubility at pH 4–12 (Fendorf, 1995), whereas Cr(VI) compounds are toxic and carcinogenic and easily mobilised in water as the chromate (CrO_4^{2-}) anion (James, 1994; Farmer et al., 2006; Tinjum et al., 2008). Hexavalent chromium is, therefore, an important environmental contaminant associated with a wide range of industrial processes and wastes, including the production of chromate chemicals from chromite ore (FeCr_2O_4) by the high-lime process. In this process, chromate is extracted from chromite by heating to 1150 °C with an alkali-carbonate to oxidise Cr(III) to Cr(VI), which is extracted with water upon cooling (Darrie, 2001; Burke, 1991) (lime (CaO) or limestone is added to the kiln to increase air penetration (Farmer et al., 2002)). The high-lime process is notoriously inefficient and produces large volumes (up to 4 tonnes per tonne of product) of caustic waste with pH > 12 (Deakin et al., 2001a; Higgins et al., 1998; Walawska and Kowalski, 2000). This waste, known as Chromium Ore Processing Residue (COPR), typically contains 2–7% chromium as a mixture of Cr(III) and Cr(VI) compounds (Geelhoed et al., 2002). COPR mineral phases typically include portlandite, calcite, ettringite, hydrogarnet and brownmillerite (Chrysochoou et al., 2010). When water comes into contact with high-lime COPR, portlandite and other soluble phases dissolve. As a result COPR pore water has a characteristically high pH of ~12, and contains up to 1.6 mmol·L⁻¹ Cr(VI) as chromate (Farmer et al., 2002; Geelhoed et al., 2002; Fuller et al., 2013).

The high lime method was superseded by lime-free processing in more developed countries in the 1960s, but continued in China, Russia, India and Pakistan (40% of world production in 2001 (Darrie, 2001)). Prior to last few decades the hazards arising from COPR were not fully recognised, and as a result vast quantities of COPR have been used as a fill material for roads and other construction projects (Burke, 1991; Graham et al., 2006; Watts et al., 2015), or simply dumped in unlined tips (Stewart et al., 2010; Stewart et al., 2007; Breeze, 1973; Deakin et al., 2001b; Jeyasingh and Philip, 2005), in and around urban areas at, or near, the production facility (e.g. Glasgow and Bolton, UK; Baltimore, Maryland and Hudson County, NJ, USA; Cracow, Poland; Hubei and Hunan Provinces, China, and; several sites in India (Burke, 1991; Higgins et al., 1998; Walawska and Kowalski, 2000; Geelhoed et al., 2002; Breeze, 1973; Jeyasingh and Philip, 2005; Wang et al., 2013; Brose and James, 2013; Sreeram et al., 2003)). Unless the waste is properly capped, rainwater will readily infiltrate COPR, therefore, there are numerous sites around the world where COPR leachate is contaminating the surrounding area with Cr(VI).

Recently, at a legacy site in the North of England where COPR had been dumped, Cr(VI)-containing water was found to be leaching from the waste into adjacent soils (Whittleston et al., 2011a). However, chromium was found to be accumulating in the original topsoil layer directly beneath the waste exclusively in its reduced Cr(III) form associated with Fe(III) oxy-hydroxides in the soil. It was found that Cr immobilisation by reduction from Cr(VI) to Cr(III) was intrinsically linked to the microbially mediated generation of Fe(II), suggesting that the soil organic matter (SOM) was providing an electron donor for microbial metabolism including iron reduction.

SOM is a complex mixture of organic compounds (polysaccharides, lignin, protein, humics) that derive from plant and microbial biomass (Fierer et al., 2009). The in-situ biodegradation of SOM is controlled by the prevailing physical, chemical and biological conditions (Dungait et al., 2012), and SOM generally becomes more stable lower in the soil profile (Dungait et al., 2012; VandenBygaart et al., 2015). Consequently, the buried soil horizons at the COPR sites could be expected to be relatively stable. However the presence of an alkaline leachate promotes the process of alkaline hydrolysis, where cellulosic materials degrade under

anoxic conditions to generation of a range of soluble low molecular weight products dominated by saccharinic acids (Humphreys et al., 2010; Knill and Kennedy, 2002). These degradation products are then potentially available to support a wide range of microbial processes, including iron reduction (Bassil et al., 2015; Rout et al., 2014; Rout et al., 2015).

There are currently no effective low cost treatments for COPR tips that offer long term environment protection. This paper reports a field investigation of COPR tip where initial investigations suggest a naturally occurring organic-rich soil beneath the waste has accumulated high concentrations of chromium. The aim of the work was to use chemical analysis, characterise of the soluble organic fraction, microbial community analysis and X-ray absorption spectroscopy, to better understand the mechanism controlling Cr accumulation in alkaline affected soils. This new understanding is then used to provide an assessment of the potential for organic matter addition to enhance natural Cr attenuation processes occurring at COPR sites.

2. Material and methods

2.1. Field sampling

COPR waste, which was dumped into a gravel extraction site next to the former production site in the north of England (Fig. 1), now forms a domed hill covered with gravel and topsoil (the site was redeveloped as a secondary school and recreational area). Springs containing alkaline leachate occur at the edge of the waste which flow in all directions away from the site (only the spring that was sampled is shown in Fig. 1).

Three boreholes (BH1, BH2 and BH3; Fig. 1.) were advanced at the site in March 2013 using a Van Walt Cobra percussion drill fitted with a 75 mm diameter core sampler. Solid material recovered from the sampler was stored in 150 mL plastic containers. Selected containers (from below the level of water strike) were stored in 2 L glass jars containing anaerogen sachets to prevent sample oxidation. Approximately 1 g sub-samples of each horizon was transferred to 1.5 mL tubes and stored at -80 °C or -20 °C for synchrotron and microbial community analysis, respectively. Sub-samples from each depth were suspended (1:1 ratio w/w) in deionised water for pH measurement in the field (ASTM, D4972-01, 2006). Leachate was collected from a spring located at the edge of the COPR and stored in 1 L polypropylene bottles. The pH of soil suspensions and the leachate were determined using a Myron Ultimeter2 portable multimeter.

2.2. Sample characterisation

X-ray powder diffraction (XRD) analysis of selected solid samples (dried at 65 °C; ground to <75 µm) was performed on a Bruker D8 XRD using Cu K-alpha radiation. X-ray fluorescence (XRF) analysis was performed on powdered samples on an Olympus Innovex X-5000 XRF. Total organic carbon content was determined using a LECO SC-144DR elemental analyser after pre-treated with 20% HCl to remove carbonates (Schumacher, 2002). Extractable Fe(II) and Fe(total) in solids were determined using field moist samples after extraction by 0.5 N HCl and reaction with Ferrozine (Lovley and Phillips, 1986). Sulphate concentrations were determined by ion chromatography using a Dionex DX500 and AS9 column.

2.3. Characterisation of soluble organic fraction

Acid soluble organic substances were determined in the BH2-180 cm sample after extraction with 2 M HCl (1 g soil in 5 mL of 2 M HCl for 3 days at 4 °C). The extractant was then separated by centrifugation at 8000 g for 10 min, pH neutralised by drop-wise addition of 2 M NaOH, evaporated to dryness; and finally the resulting solid dissolved in ultra-pure water at 1 g·L⁻¹. Total organic carbon in the extractant was determined using a Shimadzu total organic carbon analyser

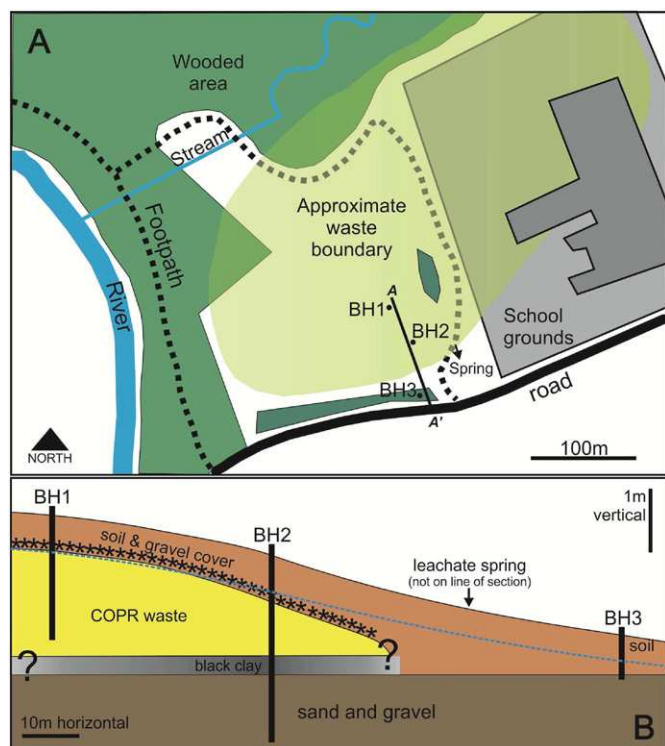


Fig. 1. A. Map of the sampling site showing the approximate boundary of COPR waste, and the location of boreholes and the sampled leachate spring. B. Approximate ground model along the line of section A–A' showing the relationship of soil and waste horizons found in this part of the site. The blue dashed line joins the observed height of water strike in the boreholes. Question marks indicate areas of uncertainty over the position and structure of inferred boundaries.

5050A. Alkaline cellulose degradation products; α , β and xylo-isosaccharinic acids, α and β metasaccharinic acids, were determined by high performance anion exchange chromatography (Rout et al., 2014) that was also capable of detecting a wide range of sugars ranging from glycerol (C-3) to maltose (C-12). Glucose/lactate, ethanol and formic acid were measured by enzymatic methods using a Picotracer glucose/lactate sensor and QuantiChrom™ ethanol assay kit a commercial formic assay kit. Remaining volatile fatty acids were detected using gas chromatography (Rout et al., 2015) capable of determining C2–C8 volatile fatty acids.

2.4. Microcosm experiments

To investigate the interaction of COPR leachate with sediments found beneath the waste, microcosm experiments were established where 10 g sediment was incubated with 100 mL site leachate in 120 mL glass serum bottles. Three sediments recovered from 180 ± 10 , 235 ± 20 and 295 ± 30 cm below ground level (bgl) from BH2 were used, providing a range of initial pH values between 8 and 10. Triplicate microcosms were purged with N_2 gas, sealed with butyl rubber stoppers and aluminium crimps, and stored at 21 °C in the dark. Samples (3 mL) of sediment/water suspension were periodically removed over a 30 day period using sterile syringes and needles. After sampling the solid and aqueous phase were separated by centrifugation (5mins; 16,000 g), and pH was measured using an Orion bench top metre and calibrated electrodes (pH calibrated at 7 and 10). Aqueous chromium (VI) concentrations were determined using a standard UV/VIS spectroscopy method based on the reaction with diphenylcarbazide using a Jenway 6715 UV/Visible spectrophotometer (USEPA, 1992). Solid Fe phases were analysed using the Ferrozine method previously described (Lovley and Phillips, 1986) (the solids content of the suspension was

$100 \pm 1 \text{ g} \cdot \text{L}^{-1}$ and 1.5 mL suspension was used in each measurement, therefore Fe(II) concentrations ($\mu\text{mol} \cdot \text{g}^{-1}$) were calculated on the basis that 0.15 g was used). To test for the presence of a viable Fe(III)-reducing bacteria, 0.1 g samples of the 180 ± 10 cm soil was added to triplicate 100 mL serum bottle containing a defined Fe(III)-citrate growth media at pH 9.2 (see (Fuller et al., 2014) for details of the media composition). After 14 days the media was scored for growth with formation of a black Fe(II) phosphate precipitate indicating a positive result.

2.5. X-ray absorption spectroscopy (XAS)

Cr K-edge XANES spectra were collected on beamline I20, Diamond Light Source, UK, from 13 soil and COPR samples. EXAFS spectra were collected from soil sample BH2-180 cm. Spectra were compared to standard laboratory chemicals, natural chromite ore, and precipitated Cr-hydroxide. All spectra were summed and normalised using Athena v0.8 (Ravel & Newville, 2005). Background subtracted EXAFS spectra were prepared using PySpline v1.1 (Tenderholt et al., 2007), and were modelled using DLexcur v1.0 (Tomic et al., 2005) (see Supplementary material for details).

2.6. DNA extraction and sequencing of the 16S rRNA gene

Microbial DNA was extracted from sample BH2-180 cm, and a 16S rRNA gene fragment (~1.5 kb) was amplified by polymerase chain reaction (PCR) using broad specificity primers. The PCR product was ligated into a standard cloning vector, and transformed into *Escherichia coli* competent cells to isolate plasmids containing the insert, which were sent for sequencing (see SI for details). The quality of each gene sequence was evaluated using UCHIME implemented within USEARCH 6.0.310 (Edgar et al., 2011), and non-chimeric sequences were grouped into operational taxonomic units (OTUs) using MOTHUR 1.30.2 (Schloss et al., 2009) and classified using the Ribosomal Database Project (RDP) naïve Bayesian Classifier (Wang et al., 2007). Sequences were submitted to the GenBank database (GenBank Numbers: accession request #Hx2000047540 submitted 07-May-2015). Phylogenetic trees were constructed using the MEGA5.2.2 (Tamura et al., 2011).

3. Results

3.1. Sample recovery and characterisation

The upper parts of BH1 consisted of a soil layer over a gravel capillary barrier. Waste material was encountered at 75 cm bgl. From 75 to 135 cm the waste primarily consisted of furnace clinker mixed with gravel and some pale white/green COPR. COPR was then recovered from 135 to 200 cm, where the borehole terminated. BH2 consisted of a similar soil and gravel cover to 90 cm bgl. COPR was recovered from 90 to 170 cm. immediately below the COPR was a black clay layer (170–190 cm), then interlayered brown sand and gravel to 320 cm, where the borehole terminated. BH3 consisted of a dark brown clay soil layer to 40 cm bgl, which then graded into brown sands and gravels to 75 cm, where the borehole terminated (see Fig. 1B).

All soil samples had essentially the same bulk mineralogy dominated by quartz, feldspars (albite, orthoclase) and clays (muscovite, kaolinite, chlorite). The COPR was dominated by portlandite and calcite with ettringite and quartz components. The leachate had a pH of 11.2 and contained $384 \pm 17 \mu\text{mol} \cdot \text{L}^{-1}$ Cr(VI) and $135 \pm 4 \mu\text{mol} \cdot \text{L}^{-1}$ sulphate.

A detailed geochemical characterisation was produced using solid samples recovered from BH2 (Fig. 2). Surface soil pH was 9.0, but increased to 9.7 with depth. The COPR horizon pH was 12.5 ± 1 . The black clay horizon immediately below the COPR had a pH value of 10.4, below this layer the pH decreased to pH 7.9 in the deepest sample. In addition to high pH, the COPR horizon was characterised by high Ca (>20%) and Cr (>2%) content, low TOC (<0.5%). The black clay horizon also contained a high Cr content (~0.8%), was rich in organic matter

(TOC = 4.8%), and had an acid extractable Fe(II) content of $75 \pm 14 \mu\text{mol}\cdot\text{g}^{-1}$ (equivalent to 66% of total extractable Fe). Acid extractable organic carbon in the black clay was $2740 \text{ mg}\cdot\text{kg}^{-1}$ and lactate was detected at $0.48 \text{ mg}\cdot\text{kg}^{-1}$. None of the other organic substrates considered were detectable in the extract. Soils found below the black clay horizon contained <1% total organic carbon, $<7 \mu\text{mol}\cdot\text{g}^{-1}$ extractable Fe(II) (<45% of total extractable Fe) and Cr concentrations where <0.1% in all samples.

XANES analysis (Fig. 3) showed that between 50 and 70% of the Cr present in the COPR waste horizons was present in the Cr(VI) oxidation state (based on analysis of the pre-edge peak present at 5995 eV (Parsons et al., 2007); shown in Fig. 2). Most of the soil samples analysed, however, contained predominately Cr(III), although the sand from below the black clay occasionally contains up to 20% Cr(VI) as part of a moderate overall Cr concentrations. Similarly, in the soil cover up to 20% of the Cr was Cr(VI).

The EXAFS spectra collected from the black clay sample BH2-180 cm (Fig. 4) could be well-fitted with a model using three shells of 6 O backscatters @ 1.98 Å and two additional shells of Cr backscatters at 3.00 and 3.94 Å (Table 1), consistent with Cr K-edge EXAFS analysis of Cr(III)(OH)₃ presented by Fendorf et al. (1997).

3.2. Microcosm tests

The experimental pH in the BH2-180 cm black clay, –235 cm sand and –295 cm sand microcosms equilibrated to $\text{pH } 10.2 \pm 0.2$, 9.2 ± 0.2 and 7.9 ± 0.4 respectively within 1 day then remained constant throughout the tests. The concentration of extractable Fe(II) in the black clay was initially $75 \pm 14 \mu\text{mol}\cdot\text{g}^{-1}$ and reduced to around $57 \pm 12 \mu\text{mol}\cdot\text{g}^{-1}$ at the first sampling point, but recovered to $70 \pm 2 \mu\text{mol}\cdot\text{g}^{-1}$ by 30 days. The two deeper aquifer sands initially contained $<7 \mu\text{mol}\cdot\text{g}^{-1}$ extractable Fe(II), which was consumed by 3 days

(Fig. 5). Aqueous Cr(VI) (from the site leachate) was removed from solution rapidly in the microcosms containing the black clay, with ~60% removed after 1 day, but very little Cr(VI) removal was observed during 30 days incubation in tests using either of the aquifer sand samples. Sulphate removal did not occur in any microcosm system, indicating that sulphate reduction did not occur in these tests.

3.3. Fe(III) citrate media tests

When the BH2-180 cm black clay samples were incubated in alkaline Fe(III)-citrate media at pH 9.2, the orange Fe(III) colour of the media was removed within two weeks and black precipitate formed (identified as the Fe(II)-phosphate mineral, vivianite, in similar previous tests (Fuller et al., 2014)) indicating a positive score for both microbial growth and Fe(III)-reduction.

3.4. DNA characterisation of microbial communities

59 rRNA gene sequences were obtained from BH2-180 cm. These were assigned to just 3 different bacterial phyla with a confidence threshold of 100% (see Fig. 6 and SI Table S1). The population was dominated Bacteroidetes sequences (70%), with Proteobacteria and Firmicutes sequences representing 21% and 9% of the population, respectively. MOTHUR analysis indicated that the sequences could be grouped into 12 OTUs based on >98% nearest neighbour similarity (97% identity is commonly used to group at “species-level” (Zaura et al., 2009; Fierer et al., 2008)). The most abundant OTU contained 68% of the sequences, each of which was classified as belonging to the genus *Mangroviflexus*, within the class *Bacteroidia*, with a >95% confidence value (an 80% confidence value results in 98.7% of sequences being correctly classified to genus level (Claesson et al., 2009)). A phylogenetic tree (Fig. 7) compares the characteristic member of the

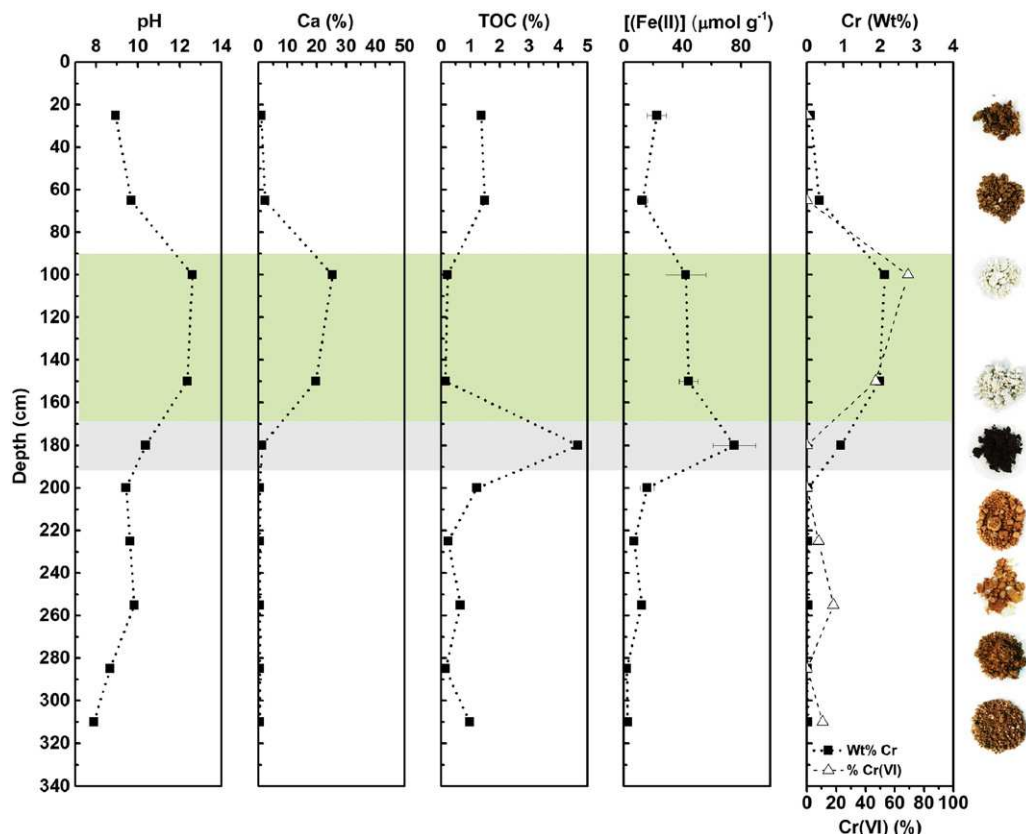
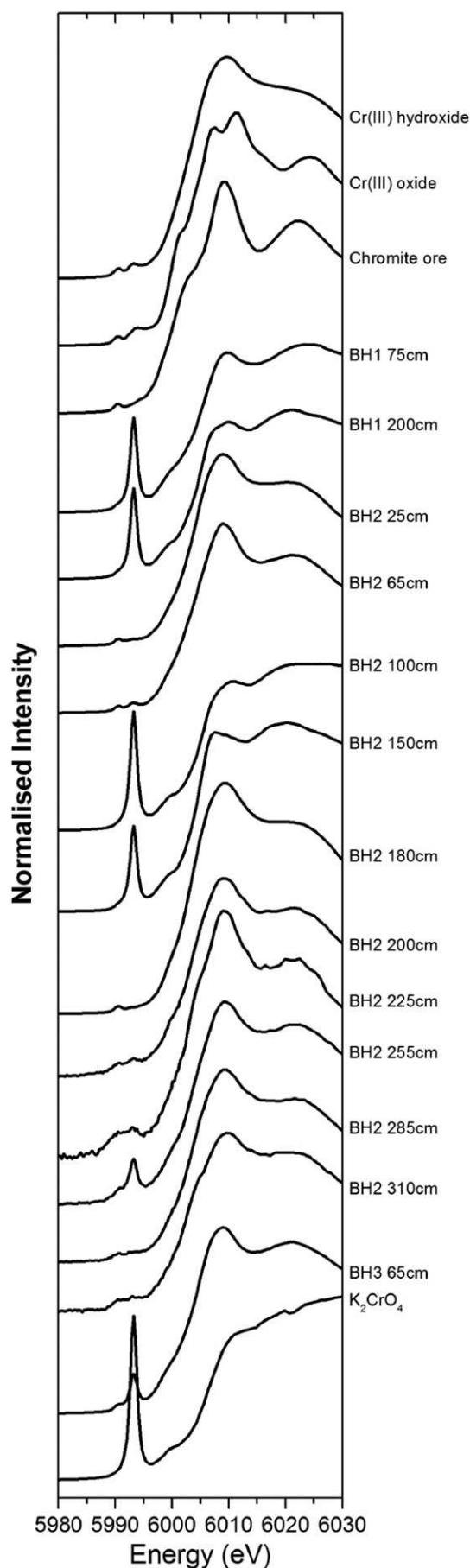


Fig. 2. Geochemical (and colour) variation in solid samples recovered from BH2. COPR waste and black organic-rich clay horizons marked by green and grey bands respectively.



Mangroviflexus-like OTU with other members of the order *Bacteroidales*. The next most abundant OTU contained 10% of the sequences were classified as belonging to the genus *Pseudomonas* (confidence 100%), within the class γ -proteobacteria.

4. Discussion

4.1. The black clay horizon acts as an in situ reactive zone

The COPR present on this site contained around 2% (w/w) Cr, with 50–70% present in the toxic Cr(VI) oxidation state. Water entering the COPR (primarily as rainwater infiltration) buffers with portlandite to produce a high pH (pH 12+) leachate containing high concentrations of Cr(VI) due to slow dissolution of a range of sparingly soluble Cr(VI)-containing minerals (Chrysochoou et al., 2010; Deakin et al., 2001b; Hillier et al., 2003). The black clay horizon found directly beneath the COPR is acting (to a certain extent) as an aquitard (resulting in springs at the base of the waste), but it also has elevated pH and Cr concentrations due to long-term interaction with COPR leachate.

The accumulated Cr was present as Cr(III)-hydroxide. Although, dissimilatory Cr(VI)-reduction using organic matter as an electron donor is possible, it is quite a fragile process under alkaline conditions (Fuller et al., 2015). The black clay was also relatively rich in solid phase Fe(II), which is known to readily react with aqueous Cr(VI) (Stewart et al., 2007; Whittleston et al., 2011b). Therefore, the redox reaction of Fe(II) and Cr(VI) is proposed as the likely mechanism for Cr(VI) reduction (Eq. (1)).



To test if this reaction mechanism was feasible, Cr(VI)-containing site water was reacted with sediments recovered from below the COPR. Cr(VI) was only removed in experiments using the black clay. Stoichiometrically, in order to remove all the Cr(VI) present in these experiments by reaction with soil associated Fe(II), $\sim 12 \mu\text{mol} \cdot \text{g}^{-1}$ Fe(II) are required. Dissolved oxygen present in the leachate (calculated at $10 \text{ mg} \cdot \text{L}^{-1}$ at 15°C) could have consumed an additional $\sim 6 \mu\text{mol} \cdot \text{g}^{-1}$ Fe(II). The observed initial removal of Fe(II) in the black clay microcosms ($\sim 18 \mu\text{mol} \cdot \text{g}^{-1}$), was therefore, broadly consistent with the total amount of oxidants present (notwithstanding the relatively large analytical errors associated with the Fe(II) measurements). Microcosms tests using the aquifer sand materials contained $< 7 \mu\text{mol} \cdot \text{g}^{-1}$ Fe(II), therefore this Fe(II) could be oxidised by the dissolved oxygen present alone. The aquifer sands also contained low organic carbon content, therefore, no Fe(III)-reduction was supported and consequentially no appreciable Cr(VI) removal occurred.

4.2. The source of Fe(II) and energy for microbial processes within the black clay

The total Cr concentration of black clay layer was $\sim 0.8\%$ ($\sim 150 \mu\text{mol} \cdot \text{g}^{-1}$). Uncontaminated soil Cr concentrations are $< 0.01\%$ (Banks et al., 2006; Madrid et al., 2006; Lado et al., 2008), therefore, this excess Cr can be attributed almost completely to infiltration of Cr(VI) leached from the COPR and in situ reduction and storage in this soil. The waste was deposited in its present location in the 1970s (~ 40 years prior to sampling), therefore, $\sim 4 \mu\text{mol} \cdot \text{g}^{-1} \cdot \text{yr}^{-1}$ of Cr have accumulated in the black clay, implying an Fe(II) utilisation rate of $\sim 12 \mu\text{mol} \cdot \text{g}^{-1} \cdot \text{yr}^{-1}$. An implication of this calculation is that the (0.5 N HCl extractable) Fe(III) currently present in the black clay ($\sim 40 \mu\text{mol} \cdot \text{g}^{-1}$), even if originally present as Fe(II), can only account for a few years of Cr(III) accumulation. Indeed, the Fe(II) currently remaining in the black clay will be

Fig. 3. Cr K-edge XANES spectra collected from samples recovered from BH1–3 and several Cr(III) and Cr(VI) containing standards.

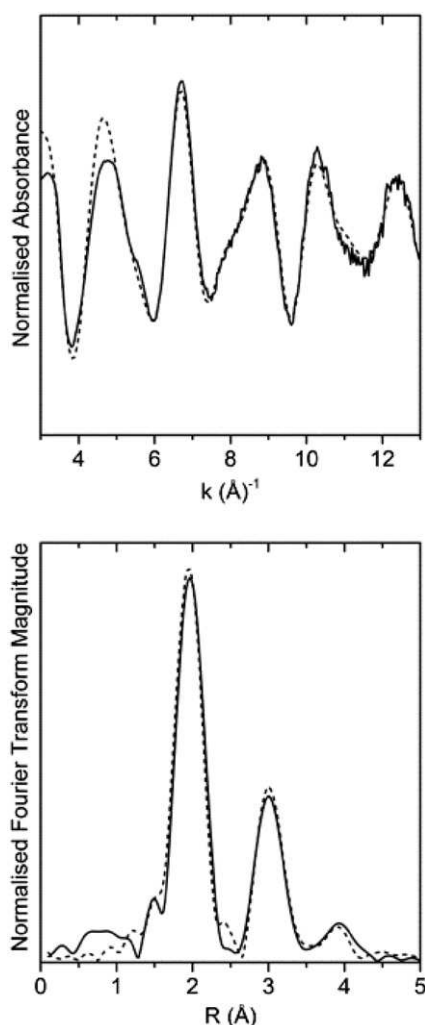


Fig. 4. Cr K-edge EXAFS spectra (top) and corresponding Fourier transformation (bottom) collected from the BH2-180 cm sample. Dotted line represents a Cr(OH)₃ model fit using the parameters listed in Table 1.

completely depleted in substantially less than 10 years, if future Cr(VI) infiltration continues at historical rates.

Therefore, it is very likely that the Fe(II) present in the black clay is replenished over time by on-going microbial Fe(III) reduction occurring in this soil despite the high pH. At pH 10.4, microbial Fe(III) reduction is energetically marginal, and predicted respiration and growth rates are slow (Williamson et al., 2013; Burke et al., 2012; Rizoulis et al., 2012). However, the rate of Fe(II) reduction required to maintain current concentrations is also relatively slow, and the black clay contains a viable population of alkaliphilic anaerobes capable of sustaining Fe(III) reduction at pH 9.2 in Fe(III)-citrate containing media. Therefore, it is conceivable that in situ Fe(III) reduction will maintain Fe(II) at favourable concentrations as long as sufficient labile organic matter is present to support dissimilatory Fe(III) reduction.

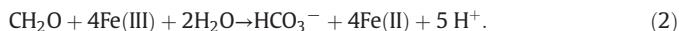
Table 1

Black Clay Cr K-edge EXAFS Fits, where *N* is the Occupancy, *r* is interatomic distance, $2\sigma^2$ is the Debye–Waller Factor and *R* is the Least Squares Residual goodness of fit parameter. Uncertainties in the last digit shown in parentheses.

Shell	<i>N</i>	<i>r</i> (Å)	$2\sigma^2$ (Å ²)	<i>R</i> (%)
O	6 ^a	1.98(2)	0.007(2)	21.3
Cr	3 ^a	3.00(5)	0.013(3)	
Cr	1 ^a	3.94(5)	0.014(4)	

^a Values fixed during fitting.

The black clay contained ~4.8% (4,000 μmol·g⁻¹) total organic carbon. According to equation 2 below (a generalised equation for dissimilatory Fe(III)-reduction using carbohydrate as an electron donor; (Lovley, 1997)), this amount of organic carbon might support the production of up to 16,000 μmol·g⁻¹ Fe(II) over time.



That amount of Fe(II) would treat Cr(VI) at historic infiltration rates for over 1000 years. However, this assumes that all of the organic matter present both is available for dissimilatory metabolism, and, that dissimilatory Fe(III) reduction is the dominant metabolism occurring in the soil – neither of which are realistic assumptions. Indeed, the lack of detectable soluble organic matter fractions (except lactate) suggests that most of the organics present are cellulosic and therefore sparingly soluble. Therefore, either the cellulosic materials are being degraded directly by microbes or that alkaline degradation products (expected to form at the pH present; (Kniill & Kennedy, 2002)) are being rapidly utilised by microbes to ensure there is no residual to detect.

The microbial community in the black clay was dominated by a *Mangroviflexus*-like specie from the family Marinilabiliaceae, in the order Bacteroidales. It is interesting to note that type species from the family Marinilabiliaceae are obligately anaerobic, fermentative bacteria (Zhao & Chen, 2012; Zhao et al., 2012; Zhilina et al., 2004; Denger et al., 2002; Gao et al., 2013). Also, the most closely related type species, *Mangroviflexus xiamenensis*, *Alkalitalea saponilacus*, and *Alkaliflexus imshenetskii*, were all isolated from anaerobic cellulose-degrading environments (Zhao & Chen, 2012; Zhao et al., 2012; Zhilina et al., 2004). *A. saponilacus*, and *A. imshenetskii* are alkaliphiles that can degrade xylan (Zhao & Chen, 2012; Zhao et al., 2012), which is a group hemicelluloses that common in plant cell walls (Erlandson et al., 2001), whereas *M. xiamenensis*, is a halo-tolerant bacterium that can degrade mono and disaccharides derived from the breakdown of cellulose and hemicelluloses (Zhao et al., 2012). Therefore, the black clay layer contains a population of bacteria that could produce labile organic matter as a by-product of fermentation that could potentially support other metabolic processes such as dissimilatory iron reduction. Extractable lactate was present in the black clay at low concentrations. Lactate is a potent electron donor source for dissimilatory metal reduction (Lovley, 1995), so its presence in the soil was probably a result of on-going fermentative respiration, possibly coupled to on-going dissimilatory Fe(III)-reduction.

4.3. Implications for management of COPR contaminated sites

It is clear that the organic rich clay horizon, that fortuitously is present at this site, is acting as an in situ redox barrier, effectively preventing the downwards spread of Cr(VI) that has leached from the COPR. The continued existence of the barrier depends on the continual production of Fe(II) over time to maintain a supply of reductant in the soil to react with Cr(VI) in the infiltrating COPR leachate. The supply of Fe(II), in turn, depends on electron donor availability, both in terms of total organic carbon present, and the availability of the organic carbon source for bioreduction. In the black clay layer alkaline hydrolysis reactions are predicted at high pH, which, coupled to the actions of alkaliphilic fermentative anaerobes, could maintain a long term supply of labile organic matter to support Fe(III)-reduction. As the soil naturally contains relatively high concentrations of organic matter, it is expected that these beneficial circumstances will be maintained for many decades to come, possibly even until all the available Cr(VI) has been leached from the COPR.

However, the sandy soils found beneath, and adjacent to, the COPR contains relatively low amounts of organic carbon (< 1%), and also low concentrations of protective Fe(II). Although total Cr concentrations in these soils are relatively low, some of that Cr occurs as Cr(VI). Therefore, the natural barrier provided by the black clay horizon is probably not

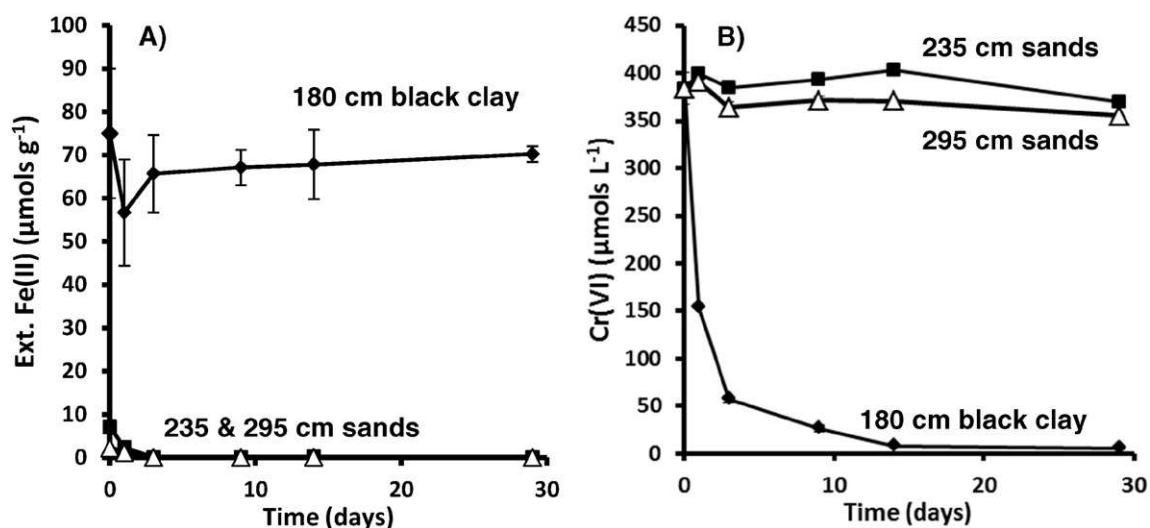


Fig. 5. Results from microcosm tests using sediments recovered from below the COPR waste in BH2 mixed with Cr(VI)-containing site leachate. A) Variation in 0.5 N HCl extractable Fe(II) and, B) variation in aqueous Cr(VI) concentrations. Error bars are 1 σ of triplicate microcosms and were not shown are less than the size of the symbol used.

continuous across the whole site and other soils present do not contain sufficient organic matter to support Fe(III) bioreduction processes at sufficient rates to counteract the effects of Cr(VI) infiltration.

Addition of labile organic electron donors (e.g. acetate, lactate etc.) to organic matter poor soils and sediments has been proposed as a way to promote in situ dissimilatory Fe(III)-reduction and Cr(VI) removal (Whittleston et al., 2011b; Lovley, 1995; Marsh & McInerney, 2001). Indeed, several recent trials using alkaline effected soils have shown this to be possible in laboratory tests (Stewart et al., 2007; Whittleston et al., 2011b), especially if implemented with soils at pH < 10 (or in conjunction with bicarbonate or acid addition designed to control soil pH (Whittleston et al., 2013)). However, addition of labile organic matter would be unlikely to produce a long-lived effect. Infiltration of oxygenated Cr(VI)-containing waters would over time consume any Fe(II) produced, and the subsurface geochemistry would return to the initial pre-treatment conditions. This would result in the need for extended periods of planned groundwater monitoring, and potentially additional 'top-up' treatment phases, applied as required.

Alternatively, an organic matter source containing mixture of labile and refractory organic carbon could be used, preferably as an immobile solid phase (e.g. sewage sludge, injected as a water suspension using in

situ jet grouting methods). In this situation, much like the natural black clay, conditions might be established where in situ populations of alkali-tolerant fermentative bacteria develop and use the refractory part of the added organic matter slowly over time. This would in turn produce a long lived source of labile metabolic by-products to support Fe(III) reduction for decades (or even centuries) to come. Emplacing and evenly distributing solid phase organic matter additions in the correct locations, would likely add significant costs and technical challenge in comparison to addition of a dissolved electron donor addition dispersed by groundwater. However, if achieved, the benefits of reduced future monitoring and no requirement for future interventions may be cost effective when calculated over the long term.

5. Conclusions

Cr(VI) leaching from COPR waste enters an organic-carbon rich and Fe(II)-containing soil horizon beneath the waste. Cr(VI) in leachate is reduced to Cr(III) by reaction with Fe(II) in the soil and accumulates as Cr(OH)₃. Despite a pH of 10.4 the soil contains a viable community of alkaliphilic microbes dominated by a *Mangroviflexus*-like specie of fermentative bacteria. Members of this microbial community are also

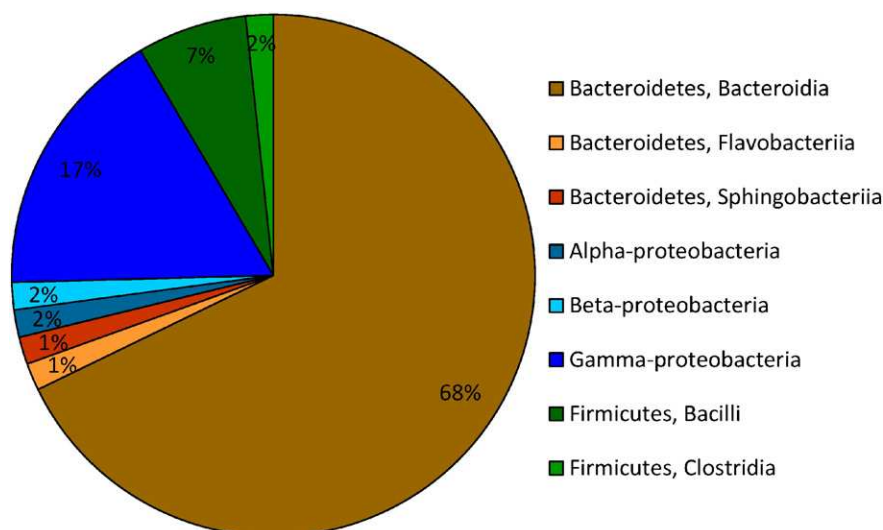


Fig. 6. Phylogenetic affiliation of the 16S rRNA gene sequences obtained from sample BH2-180 cm (assignments to both phylum and class were made with 100% confidence).

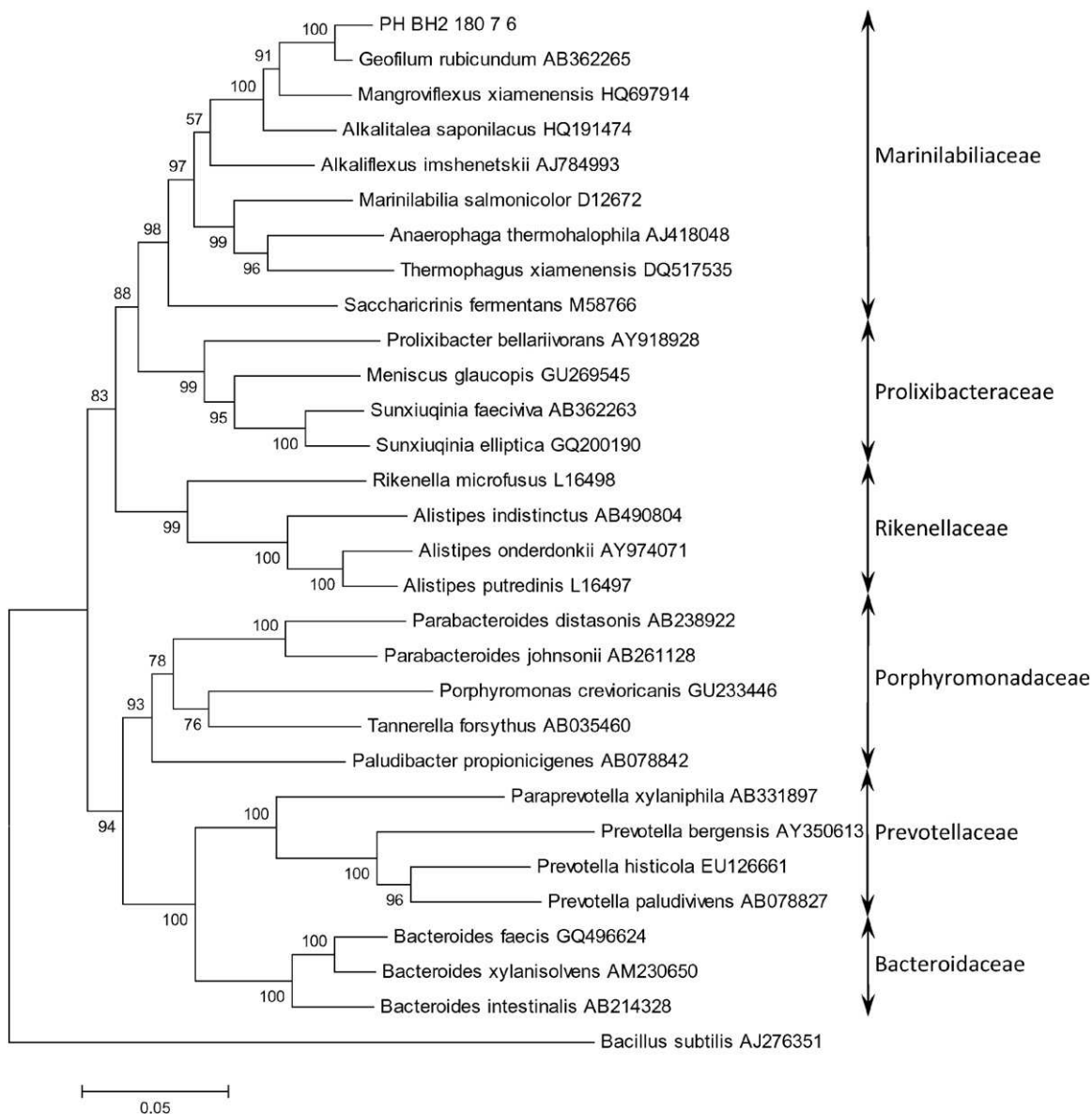


Fig. 7. Phylogenetic tree showing the relationship between characteristic sequence from the *Mangroviflexus*-like OTU and other members of the order *Bacteroidales*. *Bacillus subtilis* (*Firmicutes*) is included as an out-group. The scale bar corresponds to 0.05 nucleotide substitutions per site. Bootstrap values (%) based on 2000 replications are shown at branch points.

capable of Fe(III) reduction at pH > 9. The organic carbon and Fe(II)-rich soil, therefore, act as a potent in-situ reactive zone, effectively trapping and preventing the spread of Cr(VI) at the site. It is predicted that addition of organic matter to soils below or adjacent to COPR waste could be a viable strategy to prevent the spread of Cr(VI) at COPR disposal sites.

Acknowledgements

The authors acknowledge funding for an EPSRC Doctoral Training Award to WD, NERC grant NE/L01405X/1, and beamtime proposal SP2471/6006 at the Diamond Light Source, UK. We thank Lesley Neve and Andrew Connelly (University of Leeds) for help with XRD, XRF and TOC analysis.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <http://dx.doi.org/10.1016/j.scitotenv.2015.09.150>.

References

- ASTM, D4972-01, 2006. Standard test method for pH of soils. Annual Book of ASTM standards. American Society for Testing and Materials, pp. 963–965.
- Banks, M.K., Schwab, A.P., Henderson, C., 2006. Leaching and reduction of chromium in soil as affected by soil organic content and plants. *Chemosphere* 62, 255–264.
- Bassil, N.M., Bryan, N., Lloyd, J.R., 2015. Microbial degradation of isosaccharinic acid at high pH. *ISME J.* 9, 310–320.
- Breeze, V.G., 1973. Land reclamation and river pollution problems in the Croal Valley caused by waste from chromate manufacture. *J. Appl. Ecol.* 10, 513–525.
- Brose, D.A., James, B.R., 2013. Hexavalent chromium reduction in solution and in chromite ore processing residue-enriched soil by tartaric acid with isopropyl alcohol and divalent manganese as co-reductants. *J. Environ. Qual.* 42, 766–773.
- Burke, T., 1991. Chromite Ore Processing Residue in Hudson County, New Jersey. *Environ. Health Perspect.* 92, 131–137.
- Burke, I.T., Mortimer, R.J.G., Palaniyandi, S., Whittleston, R.A., Lockwood, C.L., Ashley, D.J., Stewart, D.I., 2012. Biogeochemical reduction processes in a hyper-alkaline leachate affected soil profile. *Geomicrobiol J.* 29, 769–779.
- Cefalu, W.T., Hu, F.B., 2004. Role of chromium in human health and in diabetes. *Diabetes Care* 27, 2741–2751.
- Chrysochoou, M., Dermatas, D., Grubb, D.G., Moon, D.H., Christodoulatos, C., 2010. Importance of mineralogy in the geoenvironmental characterization and treatment of chromite ore processing residue. *J. Geotech. Geoenviron.* 136, 510–521.
- Claesson, M.J., O'Sullivan, O., Wang, Q., Nikkilä, J., Marchesi, J.R., Smidt, H., de Vos, W.M., Ross, R.P., O'Toole, P.W., 2009. Comparative analysis of pyrosequencing and a

- phylogenetic microarray for exploring microbial community structures in the human distal intestine. *PLoS ONE* 4.
- Darrie, G., 2001. Commercial extraction technology and process waste disposal in the manufacture of chromium chemicals from ore. *Environ. Geochem. Health* 23, 187–193.
- Deakin, D., West, L.J., Stewart, D.I., Yardley, B.W.D., 2001a. Leaching behaviour of a chromium smelter waste heap. *Waste Manag.* 21, 265–270.
- Deakin, D., West, L.J., Stewart, D.I., Yardley, B.W.D., 2001b. The leaching characteristics of chromite ore processing residue. *Environ. Geochem. Health* 23, 201–206.
- Denger, K., Warthmann, R., Ludwig, W., Schink, B., 2002. *Anaerophaga thermohalophila* gen. nov., sp. nov., a moderately thermohalophilic, strictly anaerobic fermentative bacterium. *Int. J. Syst. Evol. Microbiol.* 52, 173–178.
- Dungait, J.A.J., Hopkins, D.W., Gregory, A.S., Whitmore, A.P., 2012. Soil organic matter turnover is governed by accessibility not recalcitrance. *Glob. Chang. Biol.* 18, 1781–1796.
- Edgar, R.C., Haas, B.J., Clemente, J.C., Quince, C., Knight, R., 2011. UCHIME improves sensitivity and speed of chimera detection. *Bioinformatics* 27, 2194–2200.
- Erlanson, K.A., Delamarre, S.C., Batt, C.A., 2001. Genetic evidence for a defective xylan degradation pathway in *Lactococcus lactis*. *Appl. Environ. Microbiol.* 67, 1445–1452.
- Farmer, J.G., Thomas, R.P., Graham, M.C., Geelhoed, J.S., Lumsdon, D.G., Paterson, E., 2002. Chromium speciation and fractionation in ground and surface waters in the vicinity of chromite ore processing residue disposal sites. *J. Environ. Monit.* 4, 235–243.
- Farmer, J.G., Paterson, E., Bewley, R.J.F., Geelhoed, J.S., Hillier, S., Meeussen, J.C.L., Lumsdon, D.G., Thomas, R.P., Graham, M.C., 2006. The implications of integrated assessment and modelling studies for the future remediation of chromite ore processing residue disposal sites. *Sci. Total Environ.* 360, 90–97.
- Fendorf, S.E., 1995. Surface reactions of chromium in soils and waters. *Geoderma* 67, 55–71.
- Fendorf, S., Eick, M.J., Grossl, P., Sparks, D.L., 1997. Arsenate and chromate retention mechanisms on goethite. 1. Surface structure. *Environ. Sci. Technol.* 31, 315–320.
- Fierer, N., Hamady, M., Lauber, C.L., Knight, R., 2008. The influence of sex, handedness, and washing on the diversity of hand surface bacteria. *Proc. Natl. Acad. Sci. U. S. A.* 105, 17994–17999.
- Fierer, N., Grandy, A.S., Six, J., Paul, E.A., 2009. Searching for unifying principles in soil ecology. *Soil Biol. Biochem.* 41, 2249–2256.
- Fuller, S.J., Stewart, D.I., Burke, I.T., 2013. Chromate reduction in highly alkaline groundwater by zerovalent iron: implications for its use in a permeable reactive barrier. *Ind. Eng. Chem. Res.* 52, 4704–4714.
- Fuller, S.J., McMillan, D.G.G., Renz, M.B., Schmidt, M., Burke, I.T., Stewart, D.I., 2014. Extracellular electron transport-mediated Fe(III) reduction by a community of alkaliphilic bacteria that use flavins as electron shuttles. *Appl. Environ. Microbiol.* 80, 128–137.
- Fuller, S.J., Burke, I.T., McMillan, D.G.G., Ding, W., Stewart, D.I., 2015. Population changes in a community of alkaliphilic iron reducing bacteria due to changes in the electron acceptor: implications for bioremediation at alkaline Cr(VI) contaminated sites. *Water Air Soil Pollut.* 226, 180.
- Gao, Z.M., Liu, X., Zhang, X.Y., Ruan, L.W., 2013. *Thermophagus xiamenensis* gen. nov., sp. nov., a moderately thermophilic and strictly anaerobic bacterium isolated from hot spring sediment. *Int. J. Syst. Evol. Microbiol.* 63, 109–113.
- Geelhoed, J.S., Meeussen, J.C.L., Hillier, S., Lumsdon, D.G., Thomas, R.P., Farmer, J.G., Paterson, E., 2002. Identification and geochemical modeling of processes controlling leaching of Cr(VI) and other major elements from chromite ore processing residue. *Geochim. Cosmochim. Acta* 66, 3927–3942.
- Graham, M.C., Farmer, J.G., Anderson, P., Paterson, E., Hillier, S., Lumsdon, D.G., Bewley, R.J.F., 2006. Calcium polysulfide remediation of hexavalent chromium contamination from chromite ore processing residue. *Sci. Total Environ.* 364, 32–44.
- Higgins, T.E., Halloran, A.R., Dobbins, M.E., Pittignano, A.J., 1998. In situ reduction of hexavalent chromium in alkaline soils enriched with chromite ore processing residue. *J. Air Waste Manage. Assoc.* 48, 1100–1106.
- Hillier, S., Roe, M.J., Geelhoed, J.S., Fraser, A.R., Farmer, J.G., Paterson, E., 2003. Role of quantitative mineralogical analysis in the investigation of sites contaminated by chromite ore processing residue. *Sci. Total Environ.* 308, 195–210.
- Humphreys, P.N., Laws, A., Dawson, J., 2010. A review of cellulose degradation and the fate of degradation products under repository conditions. Serco Technical Report Prepared for the Nuclear Decommissioning Authority. Harwell, Didcot, Oxfordshire, UK.
- James, B.R., 1994. Hexavalent chromium solubility and reduction in alkaline soils enriched with chromite ore processing residue. *J. Environ. Qual.* 23, 227–233.
- Jeyasingh, J., Philip, L., 2005. Bioremediation of chromium contaminated soil: optimization of operating parameters under laboratory conditions. *J. Hazard. Mater.* 118, 113–120.
- Knill, C.J., Kennedy, J.F., 2002. Degradation of cellulose under alkaline conditions. *Carbohydr. Polym.* 51, 281–300.
- Lado, L.R., Hengl, T., Reuter, H.I., 2008. Heavy metals in European soils: a geostatistical analysis of the FOREGS geochemical database. *Geoderma* 148, 189–199.
- Lovley, D.R., 1995. Bioremediation of organic and metal contaminants with dissimilatory metal reduction. *J. Ind. Microbiol.* 14, 85–93.
- Lovley, D.R., 1997. Microbial Fe(III) reduction in subsurface environments. *FEMS Microbiol. Rev.* 20, 305–313.
- Lovley, D.R., Phillips, E.J.P., 1986. Availability of ferric iron for microbial reduction in bottom sediments of the freshwater tidal potomac river. *Appl. Environ. Microbiol.* 52, 751–757.
- Madrid, L., Diaz-Barrientos, E., Ruiz-Cortés, E., Reinoso, R., Biasioli, M., Davidson, C.M., Duarte, A., Grčman, H., Hossack, I., Hursthouse, A.S., 2006. Variability in concentrations of potentially toxic elements in urban parks from six European cities. *J. Environ. Monit.* 8, 1158–1165.
- Marsh, T.L., McNerney, M.J., 2001. Relationship of hydrogen bioavailability to chromate reduction in aquifer sediments. *Appl. Environ. Microbiol.* 67, 1517–1521.
- Parsons, J.G., Dokken, K., Peralta-Videa, J.R., Romero-Gonzalez, J., Gardea-Torresdey, J.L., 2007. X-ray absorption near edge structure and extended X-ray absorption fine structure analysis of standards and biological samples containing mixed oxidation states of chromium(III) and chromium(VI). *Appl. Spectrosc.* 61, 338–345.
- Ravel, B., Newville, M., 2005. Athena, artemis, Hephaestus: data analysis for X-ray absorption spectroscopy using IFEFFIT. *J. Synchrotron Radiat.* 12, 537–541.
- Rizoulis, A., Steele, H.M., Morris, K., Lloyd, J.R., 2012. The potential impact of anaerobic microbial metabolism during the geological disposal of intermediate-level waste. *Mineral. Mag.* 76, 3261–3270.
- Rout, S.P., Radford, J., Laws, A.P., Sweeney, F., Elmekawy, A., Gillie, L.J., Humphreys, P.N., 2014. Biodegradation of the alkaline cellulose degradation products generated during radioactive waste disposal. *PLoS ONE* 9.
- Rout, S.P., Charles, C.J., Garratt, E.J., Laws, A.P., Gunn, J., Humphreys, P.N., 2015. Evidence of the generation of isosaccharinic acids and their subsequent degradation by local microbial consortia within hyper-alkaline contaminated soils, with relevance to intermediate level radioactive waste disposal. *PLoS One* 10, e0119164.
- Schloss, P.D., Westcott, S.L., Ryabin, T., Hall, J.R., Hartmann, M., Hollister, E.B., Lesniewski, R.A., Oakley, B.B., Parks, D.H., Robinson, C.J., Sahl, J.W., Stres, B., Thallinger, G.G., Van Horn, D.J., Weber, C.F., 2009. Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Appl. Environ. Microbiol.* 75, 7537–7541.
- Schumacher, B.A., 2002. Methods for the determination of total organic carbon (TOC) in soils and sediments. United States Environmental Protection Agency, Las Vegas, USA.
- Sreeram, K.J., Tiwari, M.K., Ramasami, T., 2003. Some studies on recovery of chromium from chromite ore processing residues. *Indian J. Chem., Sect. A* 42, 2447–2454.
- Stewart, D.I., Burke, I.T., Mortimer, R.J.G., 2007. Stimulation of microbially mediated chromate reduction in alkaline soil-water systems. *Geomicrobiol. J.* 24, 655–669.
- Stewart, D.I., Burke, I.T., Hughes-Berry, D.V., Whittleston, R.A., 2010. Microbially mediated chromate reduction in soil contaminated by highly alkaline leachate from chromium containing waste. *Ecol. Eng.* 36, 211–221.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., Kumar, S., 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol. Biol. Evol.* 28, 2731–2739.
- Tenderholt, A., Hedman, B., Hodgson, K.O., 2007. PySpline: a modern, cross-platform program for the processing of raw averaged XAS edge and EXAFS data. AIP Conference Proceedings, pp. 105–107.
- Tinjum, J.M., Benson, C.H., Edil, T.B., 2008. Mobilization of Cr(VI) from chromite ore processing residue through acid treatment. *Sci. Total Environ.* 391, 13–25.
- Tomic, S., Searle, B.G., Wander, A., Harrison, N.M., Dent, A.J., Mosselmans, J.F.W., Inglesfield, J.E., 2005. New tools for the analysis of EXAFS: the DL_EXCURV package. CCLRC Technical Report DL-TR-2005-01, Daresbury, UK.
- USEPA, 1992. SW-846 Manual: Method 7196a. Chromium Hexavalent (Colorimetric).
- VandenBygaert, A.J., Gregorich, E.G., Helgason, B.L., 2015. Cropland C erosion and burial: is buried soil organic matter biodegradable? *Geoderma* 239–240, 240–249.
- Vincent, J.B., 2000. The biochemistry of chromium. *J. Nutr.* 130, 715–718.
- Walawska, B., Kowalski, Z., 2000. Model of technological alternatives of production of sodium chromate (VI) with the use of chromic wastes. *Waste Manag.* 20, 711–723.
- Wang, Q., Garrity, G.M., Tiedje, J.M., Cole, J.R., 2007. Naïve Bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy. *Appl. Environ. Microbiol.* 73, 5261–5267.
- Wang, Y., Yang, Z., Peng, B., Chai, L., Wu, B., Wu, R., 2013. Biotreatment of chromite ore processing residue by *Pannonibacter phragmitetus* BB. *Environ. Sci. Pollut. Res.* 20, 5593–5602.
- Watts, M.P., Coker, V.S., Parry, S.A., Patrick, R.A.D., Thomas, R.A.P., Kalin, R., Lloyd, J.R., 2015. Biogenic nano-magnetite and nano-zero valent iron treatment of alkaline Cr(VI) leachate and chromite ore processing residue. *Appl. Geochem.* 54, 27–42.
- Whittleston, R.A., Stewart, D.I., Mortimer, R.J.G., Tilt, Z.C., Brown, A.P., Geraki, K., Burke, I.T., 2011a. Chromate reduction in Fe(II)-containing soil affected by hyperalkaline leachate from chromite ore processing residue. *J. Hazard. Mater.* 194, 15–23.
- Whittleston, R., Stewart, D., Mortimer, R., Ashley, D., Burke, I., 2011b. Effect of microbially induced anoxia on Cr(VI) mobility at a site contaminated with hyperalkaline residue from chromite ore processing. *Geomicrobiol. J.* 28, 68–82.
- Whittleston, R.A., Stewart, D.I., Mortimer, R.J.G., Burke, I.T., 2013. Enhancing microbial iron reduction in hyperalkaline, chromium contaminated sediments by pH amendment. *Appl. Geochem.* 28, 135–144.
- Williamson, A.J., Morris, K., Shaw, S., Byrne, J.M., Boothman, C., Lloyd, J.R., 2013. Microbial reduction of Fe(III) under alkaline conditions relevant to geological disposal. *Appl. Environ. Microbiol.* 79, 3320–3326.
- Zaura, E., Keijsers, B.J., Huse, S.M., Crielaard, W., 2009. Defining the healthy “core microbiome” of oral microbial communities. *BMC Microbiol.* 9.
- Zhao, B., Chen, S., 2012. *Alkalitalea saponilacus* gen. nov., sp. nov., an obligately anaerobic, alkaliphilic, xylanolytic bacterium from a meromictic soda lake. *Int. J. Syst. Evol. Microbiol.* 62, 2618–2623.
- Zhao, C., Gao, Z., Qin, Q., Ruan, L., 2012. *Mangroviflexus xiamenensis* gen. nov., sp. nov., a member of the family Marinilibaliaceae isolated from mangrove sediment. *Int. J. Syst. Evol. Microbiol.* 62, 1819–1824.
- Zhilina, T.N., Appel, R., Probian, C., Brossa, E.L., Harder, J., Widdel, F., Zavarzin, G.A., 2004. *Alkaliflexus imshenetskii* gen. nov. sp. nov., a new alkaliphilic gliding carbohydrate-fermenting bacterium with propionate formation from a soda lake. *Arch. Microbiol.* 182, 244–253.