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***Achatina (Lissachatina) fulica* BOWDICH: ITS
MOLECULAR PHYLOGENY, GENETIC VARIATION IN
GLOBAL POPULATIONS, AND ITS POSSIBLE ROLE IN
THE SPREAD OF THE RAT LUNGWORM
Angiostrongylus cantonensis (CHEN)**

IAN KENDRICH C. FONTANILLA, M.Sc.

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ABSTRACT

The Giant African Snail, *Achatina (Lissachatina) fulica* Bowdich, 1822, is a tropical crop pest species with a widespread distribution across East Africa, the Indian subcontinent, Southeast Asia, the Pacific and the Caribbean. It is also a known intermediate host of the rat lungworm, *Angiostrongylus cantonensis*, which can infect humans and cause eosinophilic meningitis. The phylogenetic position of *A. fulica* within the Achatinoidea and the Achatinidae was investigated using segments of the nuclear ribosomal (r) RNA cluster, actin and histone 3 genes and the mitochondrial CO1 and 16S rRNA genes. Results from molecular data support the monophyly of the Achatinidae based on the taxa surveyed as well as the morphological distinction of the Eastern *Achatina (Lissachatina)* from the Western and Central *Achatina (Achatina)*; *Lissachatina* should therefore be elevated to genus status. The results also show non-monophyly of the Coeliacidae, Ferussaciidae and Subulinidae; the taxonomy of these families must therefore be reassessed. The extent of genetic diversity in global *A. fulica* populations was also determined using an SSCP molecular marker developed from the 16S rRNA gene. Results reveal only one haplotype (C) emerged from East Africa and spread globally. The rat lungworm (*Angiostrongylus cantonensis*) has a parallel distribution with *A. fulica*, and the possible role of the snail in the spread of the parasite is investigated using a molecular marker derived from the small subunit (SSU) rRNA gene. A survey of the parasite within the route of dispersal of *A. fulica* detected *A. cantonensis* only in the Philippines and the French Polynesian territory of Tahiti, the latter of which being the first reported case of *A. cantonensis* infection for *Achatina fulica* in that territory. Due to the limited sampling of the snail and the patchy distribution of the parasite, there are insufficient data at this time to assess the role of *Achatina fulica* in the spread of *Angiostrongylus cantonensis*.

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TABLE OF CONTENTS

CHAPTER 1 – Introduction	1
1.1. Taxonomy of <i>Achatina fulica</i> : Mollusca, Gastropoda, Stylommatophora, Achatinoidea, Achatinidae	2
1.2.1. Classification of the Stylommatophora based on morphological data	3
1.2.2. Relationships within the Stylommatophora based on molecular data	6
1.2.3. The ‘achatinoid clade’ and the Achatinoidea	10
1.2.4. The Achatinidae	12
1.3. The biology of <i>Achatina fulica</i>	13
1.4. The biology of introduced species	13
1.5. <i>Achatina fulica</i> as an introduced species and its dispersal from East Africa	15
1.6. Parasites of molluscs	18
1.7. Nematode parasites of <i>Achatina fulica</i>	19
1.8. <i>Angiostrongylus cantonensis</i> : the nematode that hitched along	20
1.8.1. The taxonomy of <i>Angiostrongylus cantonensis</i> : Nematoda, Rhabditea, Strongylida, Metastrongyloidea, Angiostrongylidae	21
1.8.2. Biology of <i>Angiostrongylus cantonensis</i>	23
1.8.3. The life cycle of <i>Angiostrongylus cantonensis</i>	24
1.8.4. Natural intermediate hosts of <i>Angiostrongylus cantonensis</i>	25
1.8.5. Medical importance of <i>Angiostrongylus cantonensis</i>	26
1.8.6. The dispersal of <i>Angiostrongylus cantonensis</i>	28
1.8.7. Molecular identification of <i>A. cantonensis</i> and other nematodes	28
1.9. Major objectives	29
1.10. Literature cited	31
CHAPTER 2 – General Protocols	37
2.1. DNA extraction methods	37
2.1.1. DNA extraction of snail tissues using the CTAB protocol	37
2.1.2. DNA extraction of snail tissues using the Nucleon® PhytopureDNA™ Purification Kit	39
2.1.3. DNA extraction of snail tissues using the NaOH-lysis method	40
2.1.4. DNA extraction of single nematodes using NaOH digestion	41
2.2. Polymerase chain reaction (PCR)	42
2.2.1. PCR primers	43
2.2.1.1. Snail PCR primers	45
2.2.1.1.1. Nuclear genes	45
2.2.1.1.1.1. Ribosomal RNA (rRNA) gene cluster	45
2.2.1.1.1.2. Actin gene	48
2.2.1.1.1.3. Histone 3 gene	50
2.2.1.1.2. Mitochondrial primers	51
2.2.1.1.2.1. Cytochrome oxidase subunit 1 gene	51
2.2.1.1.2.2. 16S ribosomal (r) RNA gene	52
2.2.1.2. Nematode PCR primers	53
2.2.2. PCR components	54
2.2.3. PCR running conditions	55
2.2.3.1. For rRNA 1 ⁰ PCR	56
2.2.3.2. For rRNA 2 ⁰ PCR and all other genes	56
2.3. Gel electrophoresis	56
2.4. Gel extraction	57
2.5. DNA quantification	59
2.6. Cycle sequencing	59
2.6.1. Cycle sequencing components	60
2.6.2. Cycle sequencing running conditions	60
2.6.3. Precipitation of cycle sequencing samples	60
2.7. Cloning	61
2.7.1. Agar plate preparation	63
2.7.2. Setting up the TOPO™ Cloning Reaction	64
2.7.3. Plasmid isolation	64

2.8.	Sequencing of the insert	66
2.8.	Single strand conformational polymorphism analysis	66
2.8.1.	Native polyacrylamide gel preparation	67
2.8.2.	Preparation of PCR products for SSCP analysis	68
2.8.3.	Loading of samples in the polyacrylamide gel	68
2.8.4.	Silver staining of SSCP gels	69
2.9.	DNA sequence analyses	70
2.9.1.	Aligning sequences	70
2.9.2.	Checking for contamination	71
2.9.3.	Assigning ambiguous sites	72
2.9.4.	Estimating the average base frequencies and the number of variable and parsimony-informative sites	73
2.9.5.	Identifying the optimal model for DNA sequence evolution	73
2.9.6.	Evaluating the sequences for evidence of saturation	77
2.9.7.	Testing for phylogenetic signal	81
2.9.8.	Tree construction	82
2.9.9.	Estimating reliabilities of clades in phylogenetic trees	85
2.9.10.	Combining datasets as a single concatenated sequence	86
2.9.11.	Hypothesis testing	89
2.9.12.	Inferring the presence of paralogous sequences in the dataset	89
2.9.13.	Assessing the utility of the actin gene for phylogenetic analyses	90
2.10.	Literature cited	91
CHAPTER 3 – Phylogeny of the Achatinoidea		97
3.1.	Introduction	97
3.1.1.	The Achatinoidea	97
3.1.2.	Some questions about the Achatinoidea	102
3.1.3.	Use of molecular markers to determine phylogeny	104
3.1.4.	Objectives of this study	108
3.2.	Materials and methods	109
3.2.1.	Taxa used	109
3.2.2.	DNA extraction, PCR amplification and sequencing	110
3.2.3.	Sequence analysis	111
3.3.	Results	114
3.3.1.	Molecular data	114
3.3.2.	Sequence analyses	118
3.3.2.1.	Evaluating for saturation and phylogenetic signal	118
3.3.2.2.	Inferring the presence of paralogous sequences in the actin dataset and assessing its utility for phylogenetic analysis	155
3.3.2.3.	Partition homogeneity test for combining datasets	159
3.3.3.	Molecular phylogeny	160
3.3.3.1.	Individual gene phylogenies	169
3.3.3.2.	Combined gene phylogenies	172
3.3.3.3.	Hypothesis testing of groups expected to be monophyletic based on taxonomy	174
3.4.	Discussion	177
3.4.1.	Single versus combined gene phylogenies	177
3.4.2.	Phylogenetic relationships within the Achatinoidea	179
3.4.2.1.	The monophyly and phylogenetic position of the Achatinidae	179
3.4.2.2.	The monophyly and phylogenetic position of the Ferussaciidae	180
3.4.2.3.	Evaluation of the taxonomic designation of the Subulinidae, Coeliacidae, Glessulidae and Thyrophorellidae	181
3.4.2.4.	Evaluation of the monophyly of the different subulinid subfamilies	184
3.4.3.	The next step	185
3.5.	Summary	186
3.6.	Literature cited	187

CHAPTER 4 – Phylogeny of the Achatinidae	192
4.1. Introduction	192
4.1.1. The Achatinidae	192
4.1.2. Distribution and habitat of the Achatinidae	193
4.1.3. The search for the phylogeny of the Achatinidae using morphological characters	193
4.1.4. Taxonomic revision of the genus <i>Achatina</i> based on morphological characteristics	198
4.1.5. Objective of the study	199
4.2. Materials and methods	200
4.2.1. Taxa used	200
4.2.2. DNA extraction, PCR amplification and sequencing	202
4.2.3. Sequence analysis	202
4.3. Results	204
4.3.1. Molecular data	204
4.3.2. Sequence analyses	208
4.3.2.1. Evaluating for saturation and phylogenetic signal	208
4.3.2.2. Inferring the presence of paralogous sequences in the actin dataset and assessing its utility for phylogenetic analyses	247
4.3.2.3. Partition homogeneity test for combining datasets	250
4.3.3. Molecular phylogeny	250
4.3.3.1. Single gene phylogenies	259
4.3.3.2. Combined gene phylogenies	260
4.3.3.3. Hypothesis testing of <i>Achatina</i> (<i>Achatina</i>) based on taxonomy	263
4.4. Discussion	264
4.4.1. Single versus combined gene phylogenies	264
4.4.1. Evolutionary history of the Achatinidae based on molecular data	265
4.4.2. Limitations of the study and the next step	270
4.5. Summary	270
4.6. Literature cited	270
CHAPTER 5 - Genetic Variation in Global Populations of <i>Achatina fulica</i>	273
5.1. Introduction	273
5.1.1. The dispersal of <i>Achatina fulica</i> from East Africa	273
5.1.2. Genetic variation in introduced species	275
5.1.3. Mitochondrial molecular markers for evaluating genetic variation in snail populations and elucidating their population history	277
5.1.4. Objective of the study	278
5.2. Materials and methods	278
5.2.1. Sample collection	278
5.2.2. PCR and SSCP analysis using the 16S rRNA gene	280
5.2.3. Phylogenetic and network analyses of the 16S rRNA haplotypes	281
5.3. Results	281
5.3.1. PCR and SSCP analysis using the 16S rRNA gene	281
5.3.2. Phylogenetic and network analyses of the 16S rRNA haplotypes	285
5.4. Discussion	287
5.5. Summary	293
5.6. Literature cited	293
CHAPTER 6 - The SSU rRNA gene as a genetic marker for identifying infective 3 rd juvenile stage <i>Angiostrongylus cantonensis</i> from known gastropod intermediate hosts	297
6.1. Introduction	297
6.1.1. Problems with identifying nematodes: the angiostrongylids as examples	298
6.1.2. Identifying nematodes using molecular markers	299
6.1.3. Objectives of the study	302

6.2.	Materials and methods	304
6.2.1.	Sequence variation in angiostrongylid worms	304
6.2.2.	Using the SSU rRNA gene to identify <i>Angiostrongylus cantonensis</i> in a trial population	305
6.2.3.	Phylogeny of the Angiostrongylidae	307
6.3.	Results	310
6.3.1.	Sequence variation in the angiostrongylid worms	310
6.3.2.	Using the SSU rRNA gene to identify <i>Angiostrongylus cantonensis</i> in a trial population	311
6.3.3.	Phylogeny of the Angiostrongylidae	315
6.4.	Discussion	318
6.4.1.	Suitability of the 5' end of the SSU rRNA gene as a molecular marker for <i>Angiostrongylus cantonensis</i>	318
6.4.2.	Molecular identification of nematodes from a trial population of <i>Achatina fulica</i> and <i>Laevicaulis alte</i> in the Philippines	319
6.4.3.	Phylogenetic position of the Angiostrongylidae and the <i>Angiostrongylus</i> species within the Metastrongylina	320
6.5.	Conclusion	321
6.6.	Literature cited	321
CHAPTER 7 - Survey of <i>Angiostrongylus cantonensis</i> (and other nematodes) from global populations of the Giant African Snail, <i>Achatina fulica</i> (and other snails)		325
7.1.	Introduction	325
7.1.1.	Dispersal of <i>Angiostrongylus cantonensis</i> from East Africa: Alicata's hypothesis	326
7.1.2.	Perceived role of <i>Achatina fulica</i> in the spread of <i>A. cantonensis</i>	328
7.1.3.	Dispersal of <i>Angiostrongylus cantonensis</i> from South and Southeast Asia: Drozd <i>et al.</i> 's alternative hypothesis	329
7.1.4.	Other nematodes in <i>Achatina fulica</i>	330
7.1.5.	Objectives of the study	331
7.2.	Materials and methods	332
7.2.1.	Samples and localities	332
7.2.2.	DNA extraction, PCR amplification and sequencing	333
7.2.3.	Sequence analyses	333
7.3.	Results	335
7.4.	Discussion	345
7.4.1.	<i>Angiostrongylus cantonensis</i> infection in snail and slug populations	345
7.4.2.	The presence of other worms in snail and slug populations	349
7.4.3.	Limitations and the next step	351
7.5.	Summary	352
7.6.	Literature cited	353
CHAPTER 8 – Summary		357
APPENDIX		360
Appendix 2.1.	Command lines for generating likelihood scores for the 12 models in PAUP*	360
Appendix 2.2.	Command lines in PAUP* for estimating the uncorrected and corrected distances as well as the transition and transversion values	367
Appendix 2.3.	Command lines for the different tree-construction methods	368
Appendix 2.3.1.	Maximum likelihood (ML) method	368
Appendix 2.3.2.	Bayesian inference	369
Appendix 2.3.3.	Neighbor-joining (NJ) method	371
Appendix 2.3.4.	Maximum parsimony (MP) method	371
Appendix 2.4.	Command lines for the partition homogeneity test in PAUP*	373
Appendix 2.5.	Command lines to run the Shimodaira-Hasegawa test in PAUP*	374
Appendix 3.1.	Alignment of the LSU rRNA gene for the Achatinoidea and three streptaxid outgroup taxa	375

Appendix 3.2.	Alignment of the actin gene for the Achatinoidea and three streptaxid outgroup taxa	411
Appendix 3.3.	Alignment of the histone 3 gene for the Achatinoidea and three streptaxid outgroup taxa	419
Appendix 3.4.	Alignment of the CO1 gene for the Achatinoidea and three streptaxid outgroup taxa	422
Appendix 3.5.	Alignment of the 16S rRNA gene for the Achatinoidea and three streptaxid outgroup taxa	428
Appendix 3.6.	Log likelihood scores of the different models used for the Achatinoidea	433
Appendix 3.7.	Summary of ambiguous sites for the actin gene for the Achatinoidea and the three streptaxid outgroup taxa	434
Appendix 3.8A.	Summary of cloned actin sequences from three achatinoid species	435
Appendix 3.8B.	Variable sites across the actin sequence for the different clones of (A) <i>Coeliaxis blandii</i> , (B) <i>Cochlitoma granulata</i> and (C) <i>Leptinaria lamellata</i>	436
Appendix 3.9.	Neighbor-joining phylogenetic tree of the Achatinoidea (including taxa from the Achatinidae) based on the actin and showing the clones of the three representative taxa	438
Appendix 3.10.	Phylogenetic trees of the Achatinoidea based on the large subunit rRNA gene	439
Appendix 3.11.	Phylogenetic trees of the Achatinoidea based on the actin gene	441
Appendix 3.12.	Phylogenetic trees of the Achatinoidea based on the histone 3 gene	443
Appendix 3.13.	Phylogenetic trees of the Achatinoidea based on the 1 st and 2 nd codon positions of the CO1 gene	445
Appendix 3.14.	Phylogenetic trees of the Achatinoidea (taxa with complete datasets only) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1 st and 2 nd codon positions of the CO1 gene	447
Appendix 3.15.	Phylogenetic trees of the Achatinoidea (all taxa) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1 st and 2 nd codon positions of the CO1 gene	449
Appendix 4.1.	Alignment of the LSU rRNA gene for the Achatinidae and the subulinid outgroup taxon <i>Rumina decollata</i>	451
Appendix 4.2.	Alignment of the actin gene for the Achatinidae and the subulinid outgroup <i>Rumina decollata</i>	486
Appendix 4.3.	Alignment of the histone 3 gene for the Achatinidae and the subulinid outgroup <i>Rumina decollata</i>	494
Appendix 4.4.	Alignment of the CO1 gene for the Achatinidae and the subulinid outgroup <i>Rumina decollata</i>	497
Appendix 4.5.	Alignment of the 16S rRNA gene for the Achatinidae and the subulinid outgroup <i>Rumina decollata</i>	503
Appendix 4.6.	Log likelihood scores of the different models used for the Achatinidae	508
Appendix 4.7.	Summary of ambiguous sites for the actin gene for the Achatinidae	509
Appendix 4.8.	Maximum likelihood phylogenetic tree of the Achatinidae based on unambiguously aligned nucleotide sites of the actin gene	510
Appendix 4.9.	Phylogenetic trees of the Achatinidae based on the large subunit rRNA gene	511
Appendix 4.10.	Phylogenetic trees of the Achatinoidea based on the histone 3 gene	513
Appendix 4.11.	Phylogenetic trees of the Achatinidae based on the 1 st and 2 nd codon positions of the CO1 gene	515
Appendix 4.12.	Phylogenetic trees of the Achatinidae based on the 16S rRNA gene	517
Appendix 4.13.	Phylogenetic trees of the Achatinidae (taxa with complete datasets only) based on the combined dataset of the LSU rRNA, histone 3, the 1 st and 2 nd codon positions of the CO1 gene and the 16S rRNA	519
Appendix 4.14.	Phylogenetic trees of the Achatinidae (all taxa) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1 st and 2 nd codon positions of the CO1 gene	521

Appendix 5.1.	Pair-wise uncorrected distances of the East African <i>Achatina</i> species using the SSCP fragment of the 16S rRNA gene (293 nucleotides for <i>A. fulica</i>)	523
Appendix 5.2.	Alignment of the 15 haplotypes for the global <i>Achatina fulica</i> populations based on 293 nucleotides of the 16S rRNA gene	523
Appendix 5.3.	Log likelihood scores of the different models used for comparison for the 16S haplotypes of the global <i>A. fulica</i> populations using 293 nucleotides of the 16S rRNA gene	525
Appendix 6.1.	Nematode species used for the <i>Angiostrongylus cantonensis</i> survey as found in GenBank	526
Appendix 6.2.	Alignment of the 5' end of the SSU rRNA gene for the Nematoda	537
Appendix 6.3.	Log likelihood scores of the different models used for comparison for the (A) the Nematoda and (B) the Rhabditida within Nematoda using the SSU rRNA gene	604
Appendix 6.4.	Alignment of the full-length SSU rRNA gene for the Metastrongylina and two Trichostrongylina outgroups	605
Appendix 6.5.	Log likelihood scores of the different models used for comparison for the Suborder Metastrongylina using 1628 unambiguously aligned nucleotide sites from the near full-length SSU rRNA gene	616
Appendix 7.1.	Log likelihood scores of the different models used for comparison for the (A) the Nematoda and (B) the Rhabditida within Nematoda using SSU rRNA gene	617

LIST OF FIGURES

Figure 1.1.	The Pilsbry-Baker system of classification of the Stylommatophora based on the structures of the excretory organ	5
Figure 1.2.	Neighbor-joining phylogenetic tree of the Stylommatophora based on the large subunit rRNA gene (823 unambiguously aligned nucleotide sites)	9
Figure 1.3.	Adult male bursa of (A) <i>Angiostrongylus</i> , i.e. <i>A. vasorum</i> , and (B) <i>Angiostrongylus</i> (= <i>Parastrongylus</i>), i.e. <i>A. cantonensis</i>	23
Figure 1.4.	Life cycle of <i>Angiostrongylus cantonensis</i>	25
Figure 2.1.	Schematic diagram of the rRNA gene cluster based on the complete sequence of <i>Rattus norvegicus</i> (GenBank X00133) and showing the position of the primers used in this study	48
Figure 2.2.	Schematic diagram of the SSU rRNA gene based on the complete sequence of <i>Caenorhabditis elegans</i> (GenBank X00133) and showing the position of the primers used in this study	54
Figure 3.1.	Phylogeny of the Achatinoidea based on 823 unambiguously aligned nucleotide sites of the rRNA gene cluster	104
Figure 3.2.	Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the rRNA cluster in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.	119
Figure 3.3.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the rRNA cluster in (A) the Achatinoidea and outgroup taxa and (B) the Achatinoidea only	121
Figure 3.4.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the rRNA cluster in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only	122
Figure 3.5.	Plots of pairwise uncorrected distance against corrected distance for the actin gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only	123
Figure 3.6.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the actin in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only	125
Figure 3.7.	Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	126
Figure 3.8.	Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	127
Figure 3.9.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	129
Figure 3.10.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	130
Figure 3.11.	Plots of pairwise uncorrected distance versus corrected (TN93) distance for the 1 st and 2 nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	131
Figure 3.12.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1 st and 2 nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	132
Figure 3.13.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1 st and 2 nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	133

Figure 3.14.	Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 3 rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only.	134
Figure 3.15.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3 rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	136
Figure 3.16.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3 rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	137
Figure 3.17.	Plots of pairwise uncorrected distance against corrected distance for the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	138
Figure 3.18.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	140
Figure 3.19.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	141
Figure 3.20.	Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 1 st and 2 nd codon positions of the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	143
Figure 3.21.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1 st and 2 nd codon positions of the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	145
Figure 3.22.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1 st and 2 nd codon positions of the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	146
Figure 3.23.	Plots of pairwise uncorrected distance against corrected distance for the 3 rd codon position of the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	147
Figure 3.24.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3 rd codon position of the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	149
Figure 3.25.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3 rd codon position of the CO1 gene in (A) the Achatinoidea and the streptaxid outgroup taxon <i>Gibbulinella dewinteri</i> and (B) the Achatinoidea only	150
Figure 3.26.	Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the 16S rRNA gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only	151
Figure 3.27.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 16S rRNA gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only	153
Figure 3.28.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only	154
Figure 3.29.	Maximum likelihood phylogenetic trees of the Achatinoidea	161

Figure 3.30.	Maximum likelihood phylogenetic trees of the Achatinoidea based on a concatenated sequence of 5028 nucleotides derived from the combined dataset of the rRNA cluster, actin and H3 genes as well as the 1 st and 2 nd codon positions of the CO1 gene	166
Figure 4.1.	Generalized reproductive structure of achatinids	195
Figure 4.2.	Schematic diagrams of how the penis sheath envelops certain parts of achatinid genitalia	197
Figure 4.3.	Schematic illustrations of the reproductive organs of the West and Central African <i>Achatina</i> (<i>Achatina</i>) (A) and the East African <i>Achatina</i> (<i>Lissachatina</i>) (B)	199
Figure 4.4.	Plots of pairwise uncorrected distance against corrected (TN93) distance for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	209
Figure 4.5.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	211
Figure 4.6.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	212
Figure 4.7.	Plots of pairwise uncorrected distance against corrected distance for the actin gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	214
Figure 4.8.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the actin in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	216
Figure 4.9.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the actin gene in (A) the the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	217
Figure 4.10.	Plots of pairwise uncorrected distance versus corrected (GTR+I) distance for the histone 3 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	218
Figure 4.11.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the histone 3 gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	220
Figure 4.12.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the histone 3 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae	221
Figure 4.13.	Plots of pairwise uncorrected distance versus corrected (HKY85) distance for the 1 st and 2 nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	222
Figure 4.14.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1 st and 2 nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	224
Figure 4.15	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1 st and 2 nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only.	225

Figure 4.17.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3 rd codon position of the histone 3 gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> ; (B) the Achatinidae only; and (C) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> but excluding <i>Archachatina marginata</i>	229
Figure 4.18.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3 rd codon position of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> ; (B) the Achatinidae only; and (C) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> but excluding <i>Archachatina marginata</i>	230
Figure 4.19.	Plots of pairwise uncorrected distance against corrected distance for the CO1 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	231
Figure 4.20.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the CO1 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	232
Figure 4.21.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the CO1 gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	233
Figure 4.22.	Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 1 st and 2 nd codon positions of the CO1 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	234
Figure 4.23.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1 st and 2 nd codon positions of the CO1 gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	236
Figure 4.24.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1 st and 2 nd codon positions of the CO1 gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	237
Figure 4.25.	Plots of pairwise uncorrected distance against corrected distance for the 3 rd codon position of the CO1 gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	238
Figure 4.26.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3 rd codon position of the CO1 gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	240
Figure 4.27.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3 rd codon position of the CO1 gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	241
Figure 4.28.	Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the 16S rRNA gene in (A) the Achatinidae and the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	243
Figure 4.29.	Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 16S rRNA gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	245
Figure 4.30.	Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 16S rRNA gene in (A) the Achatinidae and the the subulinid outgroup <i>Rumina decollata</i> and (B) the Achatinidae only	246
Figure 4.31.	Maximum likelihood phylogenetic trees of the Achatinidae	251
Figure 4.32.	Maximum likelihood phylogenetic trees of the Achatinidae based on a concatenated sequence of 4403 nucleotides derived from the combined dataset of the rRNA cluster, H3 genes and the 1 st and 2 nd codon positions of the CO1 gene	256

Figure 4.33.	Geographic distribution of the achatinid taxa used in this study	263
Figure 5.1.	Dispersal route of <i>Achatina fulica</i> from East Africa	275
Figure 5.2.	Gel profiles of the 15 haplotypes based on a 293 bp fragment found in the domain IV of the secondary structure of the 16S rRNA gene	281
Figure 5.3.	Seventeen variable sites across the 15 haplotypes	282
Figure 5.4.	Distribution map of the 15 <i>Achatina (Lissachatina) fulica</i> populations and their 16S rRNA haplotypes	284
Figure 5.5.	Neighbor-joining tree of the 15 <i>Achatina fulica</i> 16S rRNA haplotypes based on unambiguously aligned nucleotide sites and using the optimal HKY model	286
Figure 5.6.	Median-joining network of the 15 <i>Achatina fulica</i> 16S haplotypes	287
Figure 6.1.	Posterior portion of the infective third juvenile stage of (A) <i>Angiostrongylus cantonensis</i> and (B) <i>Angiostrongylus vasorum</i>	299
Figure 6.2.	Variable sites across the SSU rDNA region for the previously sequenced <i>Angiostrongylus cantonensis</i> (Carreno & Nadler, 2003) and the 4 other angiostrongylid worms sequenced in this study	311
Figure 6.3.	Neighbor-joining (NJ) phylogenetic trees based on 376 unambiguously aligned nucleotide sites of the SSU rRNA gene. (A) NJ tree of the Nematoda (349 taxa excluding the new nematode sequences from the Philippines) rooted on 4 non-nematode taxa	314
Figure 6.4.	Phylogenetic trees of the angiostrongylids and worms in other families of the suborder Metastrongylina based on the SSU rRNA gene	315
Figure 7.1.	Probable dispersal route of <i>Angiostrongylus cantonensis</i>	327
Figure 7.2.	Neighbor-joining (NJ) phylogenetic trees based on 376 unambiguously aligned nucleotide sites of the SSU rRNA gene for the Nematoda and the Rhabditida	344

LIST OF TABLES

Table 1.1.	The Achatinoidea according to various authors in comparison with the Achatinoidea by Wade <i>et al.</i> (2006) based on molecular data (rRNA cluster)	11
Table 2.1.	Summary of the rRNA primers	47
Table 2.2.	Summary of the actin primers	50
Table 2.3.	Summary of the histone 3 primers	51
Table 2.4.	Summary of the cytochrome oxidase subunit 1 primers	52
Table 2.5.	Summary of the 16S rRNA primers	53
Table 2.6.	The four overlapping primer pairs used to amplify nearly the entire region (approximately 1670 nucleotides) of the SSU rDNA gene	54
Table 2.7.	PCR components used and their concentrations for the different genes	55
Table 2.8.	Codes for ambiguous positions in DNA sequences	73
Table 2.9.	Summary of the number of parameters of the different models of DNA substitution	76
Table 2.10.	Critical values for g1 measure of skewness at $P=0.05$	82
Table 2.11.	Comparison of methods	85
Table 3.1.	Comparison of the composition and distribution of the Rumininae according to Zilch (1959) and Schileyko (1999).	102
Table 3.2.	Taxa used for the phylogenetic study of the Achatinoidea	110
Table 3.3.	Summary of molecular data across all genes used for the Achatinoidea and three streptaxid outgroup taxa (rRNA gene cluster, actin and 16S) and one streptaxid outgroup taxon (histone 3 and CO1)	117
Table 3.4.	Comparison of the actin ambiguous sites and nucleotide differences for each achatinoid and its closest relative	159
Table 3.5.	Hypothesis testing for the monophyly of selected taxa of the Achatinoidea using the Shimodaira-Hasegawa test	176
Table 4.1.	Taxa used for the phylogenetic study of the Achatinidae	201
Table 4.2.	Summary of molecular data across all genes used (rRNA gene cluster, actin, histone 3, CO1 and 16S) for the Achatinidae and the subulinid outgroup <i>Rumina decollata</i>	208
Table 4.3.	Comparison of the actin ambiguous sites and nucleotide differences for each achatinid and its closest relative.	249
Table 4.4.	Hypothesis testing for the monophyly of <i>Achatina</i> (<i>Achatina</i>) using the Shimodaira-Hasegawa test	264
Table 5.1.	Locality, collector and sample size of global populations of <i>Achatina fulica</i> used in the study	279
Table 5.2.	Summary of the geographic distribution and frequency of the 16S rRNA haplotypes from <i>Achatina fulica</i> populations as evaluated by SSCP analysis	283
Table 6.1.	Detailed information of the angiostrongylid worms used in this study	304
Table 6.2.	Taxa used for the sequence analysis of the angiostrongylid worms	309
Table 6.3.	Matrix showing the number of differences between any two <i>Angiostrongylus</i> species	311
Table 6.4.	Summary of the distribution of the 2 Philippine nematode sequences from the University of the Philippines, Diliman campus in Quezon City	312
Table 7.1.	Snail samples, collectors and localities	332
Table 7.2.	Population sizes and percentage of nematode infection	335
Table 7.3.	Distribution and frequencies of the nematode sequences across global populations of <i>Achatina fulica</i> , West African populations of <i>Achatina</i> sp. and Philippine populations of <i>Laevicaulis alte</i>	337
Table 7.4.	Snails infected with more than one type of worm based on the sequence of the 5' end of the SSU rRNA gene	338
Table 7.5.	GenBank BLAST results for the nematode type sequences	340

CHAPTER 1 – Introduction

The tropical Giant African Land Snail *Achatina (Lissachatina) fulica* Bowdich, 1822 is one of the most extensively studied snails because of its economic, ecological and medical importance (Mead, 1979). It belongs to a family of African snails, the Achatinidae (Mollusca, Gastropoda, Stylommatophora, Achatinoidea), that includes more than 200 species in 13 genera (Schileyko, 1999). *Achatina fulica* is a major crop pest species that originated in East Africa but has been spreading across the globe since before the 1800's primarily through human activities (Mead, 1961, 1979; Raut & Barker, 2002). The World Conservation Union (IUCN) has listed *Achatina fulica* as one of the world's 100 most invasive species (Lowe *et al.*, 2000). The snail also serves as an intermediate host of the rat lungworm *Angiostrongylus cantonensis* (Chen) (see Alicata, 1966). Humans get accidentally infected by ingesting the 3rd juvenile stage of this parasite, derived from the snail intermediate host, which leads to eosinophilic meningoencephalitis (EME) or angiostrongyliasis, a disease of the central nervous system (Marquardt *et al.*, 2000). Although much is already known about *Achatina fulica*, many issues regarding the snail remain unaddressed. These include its phylogenetic relationship with other members of the Achatinoidea and the Achatinidae, the validity of the subgenus *Lissachatina* based on molecular data, the genetic variation present in introduced populations of *A. fulica* across the globe, and its potential role in the spread of the parasite *Angiostrongylus cantonensis*.

1.1. Taxonomy of *Achatina fulica*: Mollusca, Gastropoda, Stylommatophora, Achatinoidea, Achatinidae

Achatina fulica belongs to the phylum Mollusca, the second largest animal phylum in the world, with an estimated total species diversity ranging from less than 50,000 to as much as 200,000 and inhabiting freshwater, marine and terrestrial habitats (van Bruggen, 1995). Although members of this phylum exhibit diversity in form as exemplified by snails, clams, octopods, squids, chitons and the tusk shells, this group possesses three unique hallmarks that distinguish its members from other animal phyla. These hallmarks are: (1) a muscular foot for locomotion; (2) a fleshy skin fold called a mantle that secretes a calcareous shell; and (3) a feeding organ called a radula. Some molluscan groups may have lost one or more of these hallmarks during the course of evolution, but their ancestors clearly had all three as shown by fossil records (Ruppert *et al.*, 2004). Other characteristics of molluscs are the lack of true segmentation, the reduction of the body cavity, and the presence of spiral cleavage during early development (van Bruggen, 1995).

Terrestrial slugs and snails like *Achatina fulica* belong to the Gastropoda, the largest taxonomic class in the Mollusca, in which members have a characteristic head-foot region and a visceral mass (Ruppert *et al.*, 2004). Gastropods undergo torsion during larval development that leads to the 180⁰ rotation of the visceral mass and brings the ctenidia or gills and the excretory organs to the anterior region above and behind the head (Barker, 2001). There are an estimated 30,000-35,000 species of terrestrial snails and slugs (Solem, 1984), and many of these have become very important to Man either as a food source, parasite vectors, ornaments or even tools (Barker, 2001). The majority of terrestrial snails and slugs, including *A. fulica*, do not have gills but instead have a vascularised chamber or a 'lung' within the mantle cavity, a characteristic of the

subclass Pulmonata. The groupings within the pulmonates are still in a state of flux as disagreements still persist regarding interpretations of evolutionary pathways based on conchological and anatomical characters, but most workers agree on the monophyly of the (sub)order Stylommatophora. Stylommatophorans like *A. fulica* characteristically have two pairs of caudal tentacles, with the upper pair bearing eyes at the tip (Ruppert *et al.*, 2004); a contractile pneumostome that minimises contact between the environment and the pallial cavity that houses the lung, kidney and ureter; and the lack of an operculum that covers the aperture (Barker, 2001). Around 71-92 families constitute the Stylommatophora (Emberton *et al.*, 1990). Recent molecular evidence from (1) sequences from the rRNA cluster (Wade & Mordan, 2000); (2) primary sequence data of mitochondrial genes, particularly the absence of entire stem/loop structures in some domains of the mitochondrial 16S rRNA gene (Lydeard *et al.*, 2000); (3) sequence data provided by the cytochrome c oxidase subunit I (COI) gene (Remigio & Hebert, 2003); and (4) rare genomic changes in the trnP and COI genes (Grande *et al.*, 2004) all support the monophyly of the Stylommatophora.

1.2.1. Classification of the Stylommatophora based on morphological data

There is considerable debate as to how the Stylommatophora should be subdivided into groups that reflect true evolutionary relationships. The widely used but criticised Pilsbry-Baker System divides the Stylommatophora into four infraorders based on the excretory system (Pilsbry, 1900; Baker, 1955). The main excretory organ is made up of three parts: (1) a nephridium or nephridial sac (=kidney); (2) a distal ureteric pouch or orthureter; and (3) a separate ureter represented either as an open ciliated groove or a closed tube. Based on variations of these structures, the four infraorders are as follows: (1) Orthurethra (those with nephridium, orthureter with an

anterior nephropore, and a ureter developed as a groove, the proximal part of which lies along the rectal face of the nephridium, Figure 1.1A); (2) Mesurethra (those with neither an orthureter on the nephridium nor a closed ureter, Fig. 1.1B); (3) Sigmurethra (those without an orthureter but with a ureter that runs along the anterior portion of the nephridium and connects to the pallial cavity before terminating at the pneumostome, Fig. 1.1C); and (4) Heterurethra (those without an orthureter, with the nephridium extending transversely, and a ureter that runs along the face of the kidney before following the rectum and terminating at the pneumostomal opening Fig. 1.1D). It has been hypothesised that the orthurethran type of excretory system is the most 'primitive' or ancient while the other types are derived from it (Pilsbry, 1900; Baker, 1955; Barker, 2001). Based on this classification system, *A. fulica* is included in the Sigmurethra.

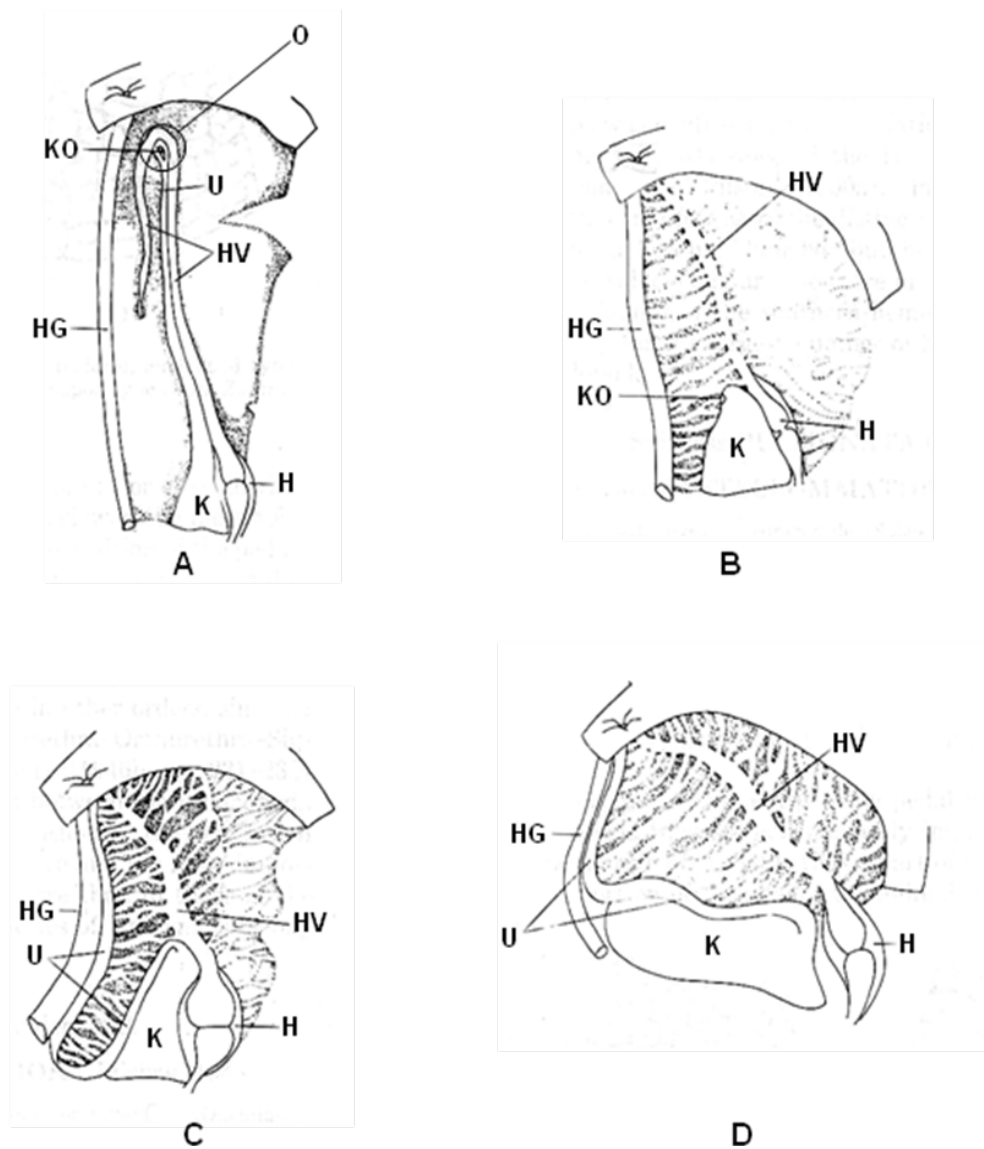


Figure 1.1: The Pilsbry-Baker system of classification of the Stylommatophora based on the structures of the excretory organ. K – nephridium (=kidney); U – ureter; KO – renal orifice or nephropore; O – orthureter; HG – hindgut; HV – principal pulmonary vein; H – heart. The four infraorders of the Stylommatophora are (A) – Orthurethra; (B) – Mesurethra; (C) – Sigmurethra; (D) – Heterurethra. (From Solem, 1959)

Pilsbry's classification system is not without its critics (Simroth & Hoffmann, 1908-1928; Thiele, 1929-1935). Tillier (1989) proposed an alternative classification system based on the differentiation within the renal organ. He retained the Orthurethra as one suborder but divided the other stylommatophorans into two different suborders: the Brachynephra in which the shortening of the kidney took place followed by closure

of the ureter; and the Dolichonephra, in which the ureter closed first before the kidney shortened. He also based his classification scheme on the various groups' current distributions that corresponded to regions that were once part of either of the two hypothetical supercontinents that resulted from the breakup of the Pangaeon landmass 200 million years ago. The Orthurethra have a mixed Laurasian (Europe, North America and Asia) and Gondwanian (South America, Africa, India, Australia) distribution, the Brachynephra includes representatives that are wholly or partly Gondwanian, and the Dolichonephra have members with Laurasian links. In this classification scheme, *A. fulica* is placed in the Dolichonephra. However, Nordsieck (1992) criticised Tillier's system and believed that overemphasis was given to the differentiation within the renal organ. Instead, he divided the Stylommatophora into two suborders: the Orthurethra that bear an orthurethran excretory system and have a primarily Laurasian origin, and the Sigmurethra with a non-orthurethran system and mixed Laurasian and Gondwanian origins. He agreed with Pilsbry's assumption of the basal position of the Orthurethra. Nordsieck's Sigmurethra also includes *Achatina fulica*.

1.2.2. Relationships within the Stylommatophora based on molecular data

Molecular markers are increasingly being used to infer phylogenetic relationships among groups of taxa because of the advantages they provide. Molecular data are derived from discrete heritable characters, are unambiguous, can easily be subjected to quantitative analyses and homology assessment, may provide characters that can be used to compare distantly related taxa, and are abundant (Graur & Li, 2000).

Although stylommatophoran taxa have been used in a variety of molecular studies examining evolutionary relationships within the Mollusca, relatively few studies

have focused on the relationships within the Stylommatophora themselves. Studies that include a handful of representative stylommatophorans, with emphasis on relationships at deeper levels within the Mollusca and the position of the Stylommatophora therein, include those of Winnepenninckx *et al.* (1998) on the small subunit (SSU) ribosomal (r) RNA gene; Thollessen (1999) on the 16S rRNA gene; Lydeard *et al.* (2000) on the secondary structures of the 16S rRNA gene; Wade & Mordan (2000) on the rRNA cluster; Remigio & Hebert (2003) on the COI gene; Colgan *et al.* (2003) on the SSU rRNA, COI and histone 3 genes; Grande *et al.* (2004) on the rare genomic changes in the trnP and COI genes; and Passamanek *et al.* (2004) on the LSU and SSU rRNA genes.

Studies focusing on the evolutionary relationships within the Stylommatophora include those of Armbruster *et al.* (2005) and Wade *et al.* (2001; 2006). Armbruster *et al.* (2005) surveyed 18 species from eight families of the Stylommatophora using the combined coding regions of the histone 3 and histone 4 genes. Their study demonstrated the basal position of the Helicidae relative to Punctidae, Clausiliidae, Pupillidae, Enidae, Vertiginidae, Cochlicopidae, and Valloniidae. *Achatina fulica* was not represented in their study. Wade *et al.* (2001, 2006) conducted a more comprehensive phylogenetic study of the Stylommatophora with initially 104 species from 50 families including *A. fulica* (2001), which they expanded to 160 species encompassing 61 families (2006). They utilised DNA sequence data from the nuclear rRNA gene cluster spanning a 1460 bp region of the 5.8S rRNA gene, the internal transcribed spacer (ITS) 2 region and the large subunit (LSU) ribosomal (r) RNA gene and comprising 823 unambiguously aligned nucleotides used for phylogenetic analysis (Figure 1.2). Although the results they obtained were in general agreement with currently accepted taxonomic families, a clear and unexpected dichotomy of the

Stylommatophora into two clades was also revealed: the strongly supported ‘achatinoids’ (99% NJ bootstraps for both the 2001 and 2006 studies) and the weakly supported ‘non-achatinoids’ (65% NJ bootstraps for the 2001 study and 63% NJ bootstraps for the 2006 study). Both clades included members with Laurasian and Gondwanian distributions, implying that the radiation of the Stylommatophora predated the breakup of Pangaea into Laurasia and Gondwana some 200 million years ago. This also implies that the sigmurethran type of excretory system is ancestral with all the other types being derived from it, contradicting Pilsbry’s assumption of a basal Orthurethra and suggesting that the Orthurethra is a derived group.

Additionally, several studies have focused on specific taxonomic groups within the Stylommatophora, as exemplified by Dutra-Clarke *et al.* (2001) on the Succineidae using the SSU rRNA gene; Holland & Hadfield (2004) on endemic Hawaiian Achatinellinae using the COI gene; Steinke *et al.* (2004) on the western Palearctic Helicidae also using the COI gene; and Tongkerd *et al.* (2004) on the Thai Pupillidae using the LSU and the 16S rRNA genes.

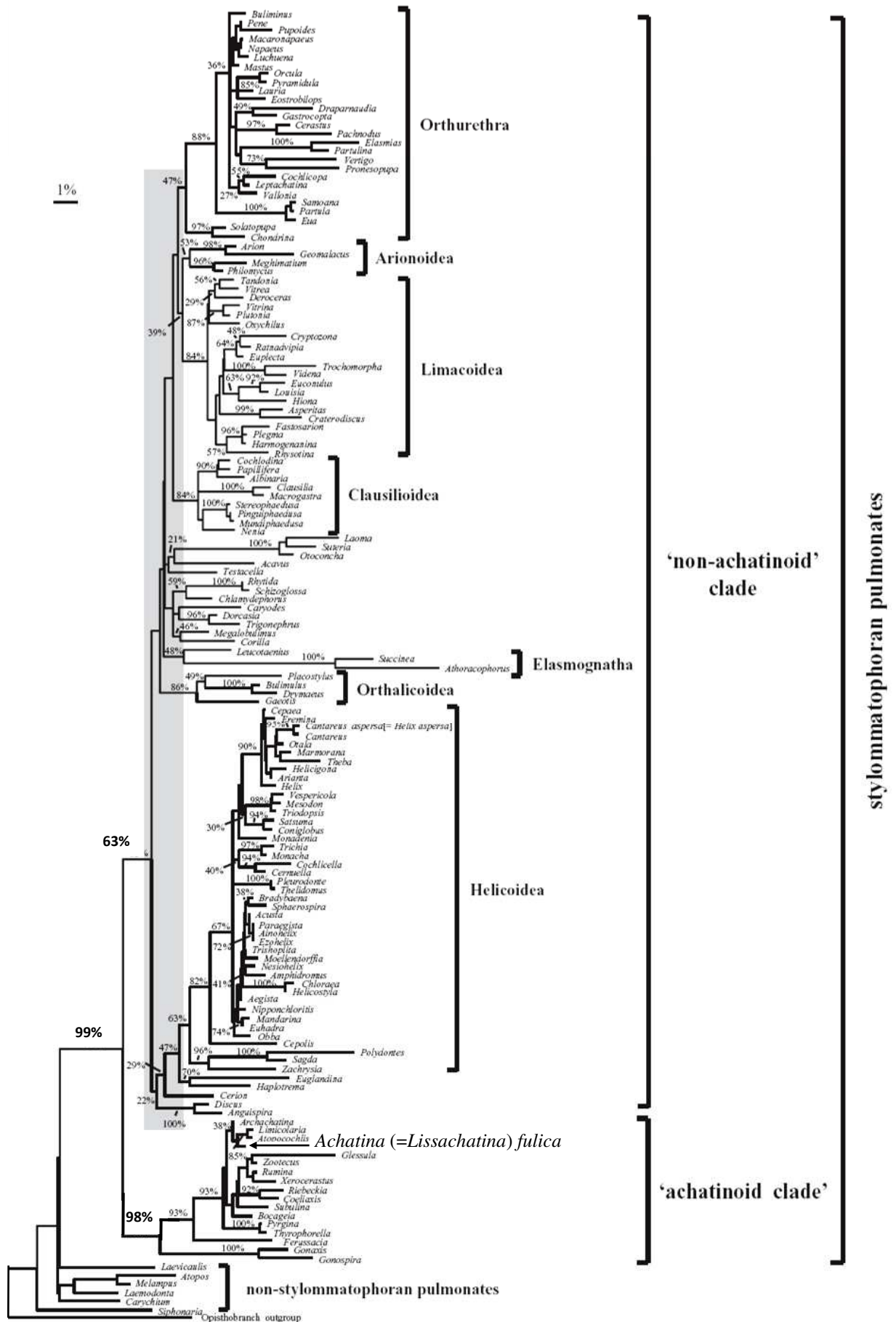


Figure 1.2: Neighbor-joining phylogenetic tree of the Stylommatophora based on the large subunit rRNA gene (823 unambiguously aligned nucleotide sites). The phylogeny shows dichotomy of the group into the 'achatinoïd' (bootstrap support=99%) and the 'non-achatinoïd' (bootstrap support=63%) clades. (From Wade et al., 2006.)

1.2.3. The ‘achatinoid clade’ and the Achatinoidea

The ‘achatinoid clade’ of Wade *et al.* (2001, 2006) based on the rRNA cluster consisted of two superfamilies: the Streptaxoidea and the Achatinoidea. Traditionally, the Streptaxoidea comprises only a single family, the Streptaxidae. They are the only carnivorous group within the ‘achatinoid clade,’ and have a long buccal mass and short oesophagus reflecting their carnivorous adaptation (Tillier, 1989). The other families in Wade *et al.*’s ‘achatinoid clade’ belong to the superfamily Achatinoidea, which included representatives of the Achatinidae, Coeliacidae, Ferussaciidae, Subulinidae, Glessulidae and Thyrophorellidae. Various authors have different views as to which families constitute the Achatinoidea. Solem’s (1978) Achatinacea (=Achatinoidea) comprised the Achatinidae, Ferussaciidae, Megaspiridae, Spiraxidae and Subulinidae (including the coeliacids and glessulids as the subfamilies Coeliacinae and Glessulinae, respectively) while Abbott (1989) regarded Coeliacidae as a family in its own right while additionally including the monotypic Thyrophorellidae. Tillier’s (1989) Achatinoidea comprised the Achatinidae, Ferussaciidae, Subulinidae (including the subfamilies Coeliacinae and Glessulinae), Thyrophorellidae as well as the Streptaxidae, Succineidae and the Oleacinidae (Spiraxidae and Testacellidae). Tillier treated the Achatinoidea as those with a closed ureter, symmetrical cerebro-pedal connectives, short cerebral commissure, and contiguous left parietal and visceral ganglia; however, he also included the Succineidae because of superficial features it shares with the Ferussaciidae and the presence of a heterurethran type of excretory organ. Vaught (1989) incorporated the Achatinidae, Coeliacidae, Ferussaciidae, Subulinidae, (including the Glessulidae which Vaught regarded as subfamily Glessulinae) and Thyrophorellidae in her Achatinoidea. Schileyko (1999) placed only the Achatinidae in the Achatinoidea, whereas Bouchet & Rocroi (2005) included the Achatinidae,

Ferussaciidae, Subulinidae (which also incorporated the coeliacids and glessulids as the subfamilies Coeliacinae and Glessulinae) and Micractaeonidae. In Wade *et al.*'s (2006) study, the Spiraxidae (represented by *Euglandina*) and Succineidae (represented by *Succinea*) fell within the 'non-achatinooid' clade and not within the Achatinoidea, thus contradicting Solem (1978) and Abbott (1989) for their inclusion of the Spiraxidae in the Achatinoidea and Tillier (1989) for his inclusion of the Oleacinidae (Spiraxidae and Testacellidae) and the Succineidae. The Micractaeonidae, which are presumably distantly related to the Ferussaciidae (Schileyko, 1999), were not represented in the Wade *et al.* study. Table 1.1 summarises the various authors' groupings of the Achatinoidea.

Table 1.1: The Achatinoidea according to various authors in comparison with the Achatinoidea by Wade *et al.* (2006) based on molecular data (rRNA cluster).

Solem (1978)	Abbott (1989)	Tillier (1989)	Vaught (1989)	Schileyko (1999)	Bouchet & Rocroi (2005)	Wade <i>et al.</i> (2006)
Achatinidae Ferussaciidae Megaspiridae Spiraxidae Subulinidae (including Coeliacinae & Glessulinae)	Achatinidae Coeliacidae Ferussaciidae Spiraxidae Subulinidae Thyrophorellidae	Achatinidae Ferussaciidae Oleacinidae* Streptaxidae Subulinidae (including Coeliacinae & Glessulinae) Succineidae Thyrophorellidae	Achatinidae Coeliacidae Ferussaciidae Subulinidae (including Glessulinae) Thyrophorellidae	Achatinidae	Achatinidae Ferussaciidae Micractaeonidae Subulinidae (including Coeliacinae & Glessulinae)	Achatinidae Coeliacidae Ferussaciidae Glessulidae Subulinidae Thyrophorellidae

* Oleacinidae – Spiraxidae + Testacellidae

The Achatinoidea in Wade *et al.*'s (2006) study were represented by 14 taxa, and their relationship was evaluated using only a short fragment (823 unambiguously aligned nucleotides) of the rRNA cluster. Increasing the number of taxa and using more genes could shed more light on the relationships of the various achatinooid groups and the validity of the taxonomic designation of the families under the Achatinoidea.

1.2.4. The Achatinidae

The family Achatinidae is a group of land snails confined to Sub-Saharan Africa, with the exception of *Achatina fulica*, which has been introduced by Man to areas outside Africa (Mead, 1979; Raut & Barker, 2002). Their shells are mostly dextral, higher than wide and are fusiform, ovoid or pillar-shaped (Schileyko, 1999). Classification within the family is based on conchological features (Bequaert, 1950) and the highly variable reproductive tract (Mead, 1991). There are three subfamilies within the Achatinidae: (1) the Callistopeplinae in which members have medium sized shells with a truncated columellar margin and a vas deferens that is not attached to the penis sheath (Mead, 1994); (2) the Limicolarinae in which members also have medium sized shells but with a non-truncated columellar margin and a vas deferens that is attached to the penis sheath (Schileyko, 1999); and (3) the Achatininae with medium to large shells with truncated columellar margin and a vas deferens that is also attached to the penis sheath (Mead, 1994). Variation also exists within the subfamilies. In the Achatininae, for instance, the East African *Achatina (Lissachatina)*, to which *A. fulica* belongs, has a smooth nepionic whorl (Bequaert, 1950) and a half-dome apex (Mead, 1995) as well as a muscular bulboid enlargement of the basal vagina (Mead, 1991; 1995) while the West and Central African *Achatina (Achatina)* has a sculptured nepionic whorl (Bequaert, 1950), a plateaued apex (Mead, 1995) and no muscular bulboid enlargement in the basal vagina (Mead, 1991; 1995).

Despite the huge amount of morphological data available on the Achatinidae, no systematic molecular approach has yet been attempted to correlate molecular data with the morphological data, in particular to assess the designation of *Achatina fulica* under the subgenus *Lissachatina* based on conchological and anatomical features.

1.3. The biology of *Achatina fulica*

Achatina fulica is a large snail with a shell length ranging from 5 to 10 cm, with some specimens even reaching 20 cm. The conical shell is light brown in colour, though the colour pattern may vary (Schotman, 1989). The presence of streaks is associated with a dominant allele such that homozygous recessive individuals have unstreaked shells (Allen, 1983). However, variation in shell morphology in terms of size, shape and colour exists and has been largely attributed to environmental conditions (Mead, 1961).

A typical *A. fulica* has a life span of 5-6 years, becoming sexually mature as early as five months. Although hermaphroditic, *A. fulica* cross-fertilises and lays eggs 8-20 days after mating. The number of eggs laid can vary depending on the age of the snail but can reach up to 1800 in a year in a tropical setting. If conditions become unfavourable, the snail can aestivate by burrowing underground and covering its shell opening with a calcareous membrane, called an epiphragm, until such time as the environment improves (Mead, 1979, Raut & Barker, 2002).

1.4. The biology of introduced species

Achatina fulica is a classic example of an introduced species. Introduced species, also known as exotic species, are those found outside their natural range due to human activity (Primack, 2006). Species may be introduced deliberately to benefit Man, with examples including agricultural plants and animals for human consumption, decorative plants for gardening, and animals for hunting or fishing. Other species may be introduced unintentionally such as parasites or pests found in deliberately introduced species and those that “hitchhike” with transported goods (Freeland, 2005).

Species introduced by Man to new areas can have a profound effect on the ecosystem of that area as well as on the introduced species itself. They can prey upon, infect, outcompete or hybridise with native species or alter the habitat at the expense of the native species (Simberloff *et al.*, 2005), thus providing additional selective pressure on the native species (Suarez & Tsutsui, 2008). A balancing act must therefore come into play between the native species and the introduced species, and sometimes even between the introduced species and the local environment of the invaded habitat; otherwise, either the native or introduced species will face displacement or extinction (Suarez & Tsutsui, 2008). The disturbance caused by introduced species has become problematic in many areas such that they have been regarded as the second greatest threat to biodiversity, next only to habitat loss (Freeland, 2005). In some cases, however, introduced species simply spread into new areas that are human-disturbed from which most native species have already disappeared due to habitat loss (Cowie & Robinson, 2003).

Not all species become easily established once translocated into a new area, but characteristics such as a rapid reproduction rate, high fecundity and generalist food and habitat requirements can increase the success of an introduced species (Cowie, 2000). Organisms that become invasive are also most likely to possess traits that facilitate their transport by humans, the ability to withstand the rigours of transport, the capacity to tolerate varying environmental conditions, and the predilection to thrive in human disturbed areas (Suarez & Tsutsui, 2008). As mentioned previously, species may be introduced deliberately or inadvertently, although those in the former category may have a greater chance of being established, particularly if the introduction is perceived to have some economic benefits. Deliberate introductions involve individuals that are

cared for and are brought to new areas in large numbers, thus improving their chance of survival once they are released in the wild (Cowie & Robinson, 2003).

The success of an introduced species in a new area can also be influenced by the genetic composition of its population. In many cases, introduced species are represented by a few individuals with a reduced amount of genetic variation when compared to their source population, a phenomenon called a founder effect. After many generations, a population bottleneck ensues where genetic variation is considerably reduced and allele frequencies undergo massive shifts (Dlugosch & Parker, 2008). As a consequence, some beneficial adaptive traits that could otherwise improve the survival and fitness of the species in the new habitat may be lost (Kolbe *et al.*, 2007). However, this low genetic variability as a result of founder events and bottlenecks could be counteracted by multiple introductions from different source populations (Dlugosch & Parker, 2008), as was shown in the cheatgrass *Bromus tectorum* (Novack & Mack, 1993) and the *Anolis* lizards (Kolbe *et al.*, 2007).

1.5. *Achatina fulica* as an introduced species and its dispersal from East Africa

Man has always been drawn to the Giant African Land Snail for reasons including its large size, supposed medicinal properties and its potential as a human or animal food source (Mead, 1979; Kliks & Palumbo, 1992; Raut & Barker, 2002). It is for these reasons that *Achatina fulica* has been spreading globally primarily through human factors, and its success as an introduced species can be attributed to several factors.

First, the biology of *Achatina fulica* makes it eminently suitable as an introduced species. The snail has a high reproductive capacity, producing between 10 and 400 eggs per clutch and as many as 1800 eggs per year; they also become sexually mature

between 5 and 8 months (Raut & Barker, 2002). *Achatina fulica*'s high reproductive capacity, in addition to the tendency of people to release the snails into the wild, would help to explain the rapid spread of *Achatina fulica* into new areas, as in Brazil where it was introduced as recently as 1988 but has since spread (Thiengo *et al.*, 2007). The Giant African Land Snail is a voracious herbivore that feeds on a wide range of cultivated plants and even weeds and indigenous plants, thus making it a serious crop pest (see Raut & Barker, 2002 for a comprehensive list of economically important food crops and ornamental and medicinal plants that are susceptible to *Achatina fulica*). It also scavenges its food from detritus and decaying plant material, which may comprise up to 75% of its diet (Raut & Barker, 2002). Its non-specific food requirements further leads to its success as an introduced species.

Second, *Achatina fulica* possesses traits that facilitate its transport by humans. For instance, the snails can easily be transported in consigned cargoes, whether accidentally or on purpose, and survive the journey of several days with little adverse effect on the "hitchhikers." This was demonstrated by a tourist who came from Hawaii and inadvertently brought a live snail to the mainland USA over a period of ten days (Mead, 1979). During these periods of long distance travel, the snails can undergo aestivation to avoid desiccation (Mead, 1961). Furthermore, *A. fulica* has a wide tolerance for different environmental conditions despite being a tropical snail (Mead, 1979; Raut & Barker, 2002). They have been found to survive temperatures as low as 2⁰ C in India and altitudes as high as 1500 meters in Malaysia (Raut & Barker, 2002). In addition, they can thrive in a range of soil pH, moisture and plant cover conditions as well as withstand a diverse group of predators (Mead, 1979). Lastly, *A. fulica* has frequently been associated with human disturbed areas such as agricultural lands and

gardens, though they have also been found in primary and secondary forests in Hawaii, the Bonin Islands, India, Southeast Asia and New Caledonia (Raut & Barker, 2002).

Third, *Achatina fulica* is commonly introduced deliberately and is therefore transported in large numbers and properly cared for, which then increases its chance of survival. In Brazil, *Achatina fulica* was introduced in 1988, probably from Indonesia, when it was heralded as an alternative source of meat. These snails were then distributed for commercial purposes but were subsequently released when people eventually lost interest. As a consequence, Brazil is currently experiencing an explosive stage of the invasion that is characterised by large individuals that are prevalent in urban areas, particularly in gardens (Thiengo *et al.*, 2007). Other deliberate introductions include those in Borneo where duck farmers used the snails as feed (Jarrett, 1931), those in Nepal where the snails were introduced in local gardens and venerated for their religious significance (Budha & Naggs, 2008) and those in the Indo-Pacific where Japanese soldiers and merchants before and during the Second World War used the snails as food and sometimes as pets (Kliks & Palumbo, 1992; Civeyrel & Simberloff, 1996).

Although currently distributed in many areas around the globe, it is possible to elucidate the route of dispersal of *Achatina fulica* since before the 1800's. The snail is thought to be indigenous to East Africa, specifically Kenya and Tanzania. It is believed to have been introduced into Madagascar and Mauritius in the early 19th century (Bequaert, 1950) and subsequently to have spread to the Indian subcontinent, Southeast Asia, most islands of the Pacific (Mead, 1961; Kliks & Palumbo, 1992), the Caribbean (Schotman, 1989) and South America (Paiva, 1999; Thiengo *et al.*, 2007; Borrero *et al.*, 2009). It has also been recently detected in West Africa (Raut & Barker, 2002). There are several pathways or activities that are available to the snail that enables it to spread

quickly to new areas. These are the accidental transfer by humans, usually by hitchhiking onto agricultural machinery and even in cars and trucks (Cowie & Robinson, 2003) and the intentional transfer by humans as pets, for food consumption or for other uses (Mead, 1979; Kliks & Palumbo, 1992; Cowie & Robinson, 2003).

To date, no systematic evaluation of the genetic variation of *Achatina fulica* across global populations has been undertaken. Such data could indicate the number of genetic types that have been introduced in the snail's new range. Furthermore, results from such a global survey would reveal if introduced populations of *A. fulica* are undergoing genetic bottlenecks as a consequence of a lack of genetic variation, or if multiple introductions from various source populations took place that could potentially counteract the effects of bottlenecking.

1.6. Parasites of molluscs

Parasitism is a relationship between two organisms where the parasite thrives on or within the host, which is harmed in some way (Roberts & Janovy, 2005). Many organisms are known to parasitise molluscs. These parasites use molluscs either as intermediate hosts, where the parasite develops but does not reach sexual maturity, as definitive or final hosts where the parasite reaches the adult stage, or as permanent hosts where the entire development of the parasite takes place in the host (Malek & Cheng, 1974; Roberts & Janovy, 2005). Examples of known parasites of molluscs include bacteria such as *Mycobacterium* in the gastropod *Helisoma anceps* (Malek & Cheng, 1974) and *Aeromonas hydrophila* in *Achatina fulica* (Dean *et al.*, 1970), protozoa such as *Hartmanella* sp. in the gastropod *Biomphalaria* (Malek & Cheng, 1974), trematodes such as *Schistosoma japonicum* in the gastropod *Oncomelania quadrasi* (Malek & Cheng, 1974) and *Echinostoma* in the apple snail *Pomacea* (Hollingsworth & Cowie,

2006), and copepod arthropods such as *Mytilocola intestinalis* in the blue mussel *Mytilus edulis* (Malek & Cheng, 1974) and nematodes such as *Oslerus ostratus* in the pulmonate slug *Laevicaulis alte* and *Nemhelix bakeri* in the pulmonate snail *Helix aspersa* (Grewal *et al.*, 2003). Molluscs act as intermediate hosts for several medically important parasites, as exemplified by trematodes such as *Schistosoma* that leads to schistosomiasis (a liver disease characterised by an enlarged liver and spleen, diarrhea, and bloody urine) (Roberts & Janovy, 2005), *Fasciola* that induces fasciolariasis (necrosis of the liver) (Roberts & Janovy, 2005), *Echinostoma* that causes echinostomiasis (an intestinal disease that leads to headache, dizziness, gastric pain, anemia and diarrhea) (Hollingsworth & Cowie, 2006) as well as nematodes such as the rat lungworm *Angiostrongylus cantonensis* that leads to eosinophilic meningioencephalitis (see below).

1.7. Nematode parasites of *Achatina fulica*

Various nematodes are associated with molluscs (Grewal *et al.*, 2003; Morand *et al.*, 2004), including *Achatina fulica*. Members of the phylum Nematoda are typically bilaterally elongated and tapering at both ends. They also have a characteristic body cavity called a pseudocoelom that is derived embryologically from the blastocoel, which normally disappears during gastrulation in eucoleomate animals but not in nematodes. These animals possess a non-cellular body covering called the cuticle that is shed four times during their lifetime through moulting; these worms therefore have four juvenile stages (sometimes incorrectly referred to as larval stages) that resemble the final adult stage in form. Nematodes include sexually dimorphic species (e.g. the rat lungworm *Angiostrongylus cantonensis*) as well as hermaphroditic species (e.g. the soil nematode *Caenorhabditis elegans*). Other characteristics include a complete digestive system,

absence of circular muscles in the body cavity, and in the case of sexually dimorphic species, the presence of females that are generally larger and males that have a more curled tail (Roberts & Janovy, 2005).

Molluscs are infected by nematodes either as intermediate hosts of the juvenile worms or as definitive or final hosts of the adult worms (Grewal *et al.*, 2003; Morand *et al.*, 2004). In total, 108 species have been found to infect gastropods, with 61 using these gastropods as intermediate hosts and 47 using gastropods as final hosts (Grewal *et al.*, 2003). Evolutionarily speaking, it has been postulated that parasitic nematodes started out as facultative parasites in which free-living forms accidentally infected animals (Adamson, 1986). Morand *et al.* (2004) used Blaxter *et al.*'s (1998) phylogeny of the Nematoda to map the occurrence of parasitism in terrestrial molluscs among the different nematode groups. They hypothesised that nematode parasitism in terrestrial molluscs occurred independently at least five times.

Very limited data are available on nematode species associated with *Achatina fulica*. Nematodes shown to be associated with *A. fulica* include *Oslerus ostratus*, which uses *A. fulica* as an intermediate host (Grewal *et al.*, 2003), *Rhabditis* sp., most likely a facultative parasite of the snail (Viyada, 2005), and the medically important *Angiostrongylus cantonensis*, the rat lungworm, which uses the snail as an intermediate host (see Mead, 1979 for a detailed list of *A. fulica* survey for *A. cantonensis*).

1.8. *Angiostrongylus cantonensis*: the nematode that hitched along

Angiostrongylus cantonensis is a parasite of rodents that requires a gastropod intermediate host such as *Achatina fulica* to complete its life cycle. The nematode's definitive or final hosts are murid rodents such as the black rat (*Rattus rattus*), the brown rat (*Rattus norvegicus*), the Pacific rat (*Rattus exulans*), the oriental house rat

(*Rattus tanezuni*), the Philippine forest rat (*Rattus everetti*) and the malabaric bandicoot rat (*Bandicota malabarica*) (Alicata, 1966; Westerlund & Chamberlain, 1969; Marquardt *et al.*, 2000). Humans can also get infected by acquiring the 3rd juvenile stage of the worm from several of its snail or slug intermediate hosts, including *Achatina fulica* (Alicata, 1966; Marquardt *et al.*, 2000), which leads to a disease of the nervous system called eosinophilic meningoencephalitis (EME) or angiostrongyliasis (Marquardt *et al.*, 2000).

1.8.1. The taxonomy of *Angiostrongylus cantonensis*: Nematoda, Rhabditea, Strongylida, Metastrongyloidea, Angiostrongylidae

Angiostrongylus cantonensis belongs to the phylum Nematoda, a group including some of the most abundant animals in the world. There are more than 25,000 described species of nematodes, with 10,000 known to be free-living and more than 15,000 known to be parasitic on animals (Poulin & Morand, 2000; Hugot *et al.*, 2001). Roberts & Janovy (2005) predicted that current figures pertaining to the diversity of the Nematoda are underestimates and that there may be more species of nematodes than there are of insects, which Ruppert *et al.* (2004) estimate to be around 30 million. Nematodes can be divided into two main classes, the Enoplea and the Rhabditea, based on the presence of certain sensilla or small sense organs. Members of the Enoplea possess pouch-like anterior sensilla called amphids whereas members of the Rhabditea, including *Angiostrongylus*, have ventrally coiled amphids; many rhabditeans also possess sensilla called phasmids near the posterior end. Within the Rhabditea is the order Strongylida that includes long, slender worms with males having the characteristic copulatory bursa supported by sensory rays (Roberts & Janovy, 2005). Within the Strongylida is the superfamily Metastrongyloidea whose members utilise mammals as

definitive hosts, with many occupying the host lungs during the adult stage (Anderson, 2000).

The genus *Angiostrongylus* belongs to the family Angiostrongylidae in the Metastrongyloidea. Members of this family have an adult stage that possesses a posterior vulva (Anderson, 2000) but not a buccal cavity or lips at the mouth. They reside in the lungs of the mammalian final hosts and require an invertebrate intermediate host (Roberts & Janovy, 2005). *Angiostrongylus* itself was regarded by Ubelaker (1986) as a heterogeneous group, and he suggested that, on the basis of the morphological differences in the copulatory bursa in adult males as well as differences in their mammalian final host specificity, the genus should be split into five distinct genera including *Parastrongylus* (which Ubelaker was resurrecting and was first described by Baylis in 1928 using *Parastrongylus tateronae* as the type species) as well as *Angiostrongylus* (first described by Baillet in 1866 using *Angiostrongylus vasorum* as type species). In *Parastrongylus*, the lateral rays of the copulatory bursa arise from a single common trunk in adult males whereas those in *Angiostrongylus* arise separately (Fig. 1.3). In terms of their final hosts, *Parastrongylus* utilises murid rodents whereas *Angiostrongylus* infects carnivores like dogs, foxes and cats (Ubelaker, 1986). Based therefore on these characteristics, Ubelaker reclassified *Angiostrongylus cantonensis* as *Parastrongylus cantonensis*. Other members of *Parastrongylus*, according to Ubelaker, include *Parastrongylus costaricensis* (Central and South America, Cuba and southern North America), *P. dujardini* (Europe) and *P. malaysiensis* (Malaysia). Species retained by Ubelaker in the genus *Angiostrongylus* are *A. vasorum* in foxes and dogs (Africa, Europe, North America and South America) (Anderson, 2000) and *A. chabaudi* in wild cats (central Italy) (Ubelaker, 1986).

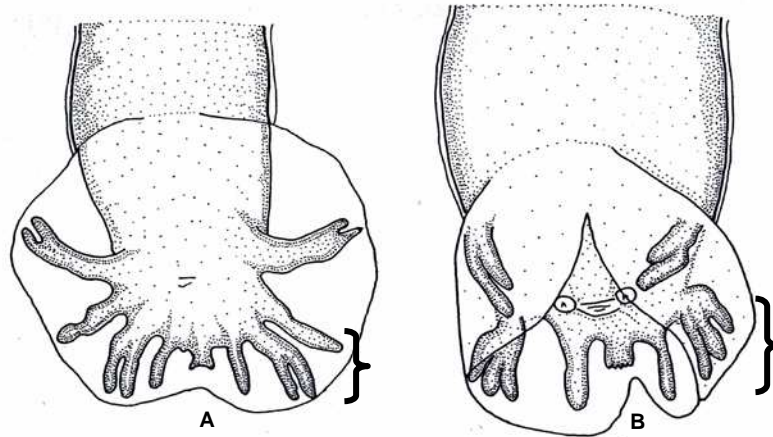


Figure 1.3: Adult male bursa of (A) *Angiostrongylus*, i.e. *A. vasorum*, and (B) *Angiostrongylus* (=Parastrongylus), i.e. *A. cantonensis*. Brackets show lateral rays. Note that the lateral rays arise from a common trunk in (B) but not in (A). From Ubelaker (1986).

The recognition of *Parastrongylus* as a separate genus, however, is not widely accepted and the name *Angiostrongylus* is still used to refer to the genus of *A. cantonensis* and the other species mentioned in the previous paragraph (Kliks & Palumbo, 1992). Molecular data to confirm or refute Ubelaker's designation are also wanting.

1.8.2. Morphology of *Angiostrongylus cantonensis*

Adult female *A. cantonensis* range from 17 to 33 mm long and 0.28 to 0.5 mm wide, whereas the males tend to be smaller, being only 15 to 22 mm long and 0.25-0.35 mm wide. The worms feed on blood, thus giving a red appearance to their intestines. In females, the intestines intertwine with the uterine tubules, which lend a characteristic barber pole appearance (Marquardt *et al.*, 2000; Roberts & Janovy, 2005). The infective 3rd juvenile stage of the parasite is considerably smaller, around 425-524 µm long and 23-34 µm wide. Although the juvenile is similar morphologically to other

species of lungworms like *Aulurostrongylus abstrutus* and *Anafilaroides rostratus*, it can be subtly distinguished by the fine point termination of the tail (Ash, 1970).

1.8.3. The life cycle of *Angiostrongylus cantonensis* (Figure 1.4)

The mature adult worms reside for approximately two weeks in the subarachnoid space separating the meninges of the rat brain. After that period, they move into the circulatory system through the venous system until they reach the heart where the females begin laying their eggs. The arterial blood then brings these eggs to the lungs where they get lodged in the alveoli. As the 1st juvenile stage emerges from the eggs, they are passed from the lungs into the trachea until they reach the gut and stay there for 42-45 days before being eliminated through the faeces (Marquardt *et al.*, 2000; Kliks & Palumbo, 1992). A snail or slug intermediate host then gets infected by these juveniles either by ingesting contaminated rodent faeces or by burrowing of the nematodes through the gastropod body wall or respiratory pores (Hollingsworth & Cowie, 2006). These nematodes then progress into the 2nd and 3rd juvenile stages after about 18 days (Marquardt *et al.*, 2000) where they reside mostly in the head-foot region of the gastropod intermediate host, although they can also be found in the lungs, liver and kidneys (Hollingsworth & Cowie, 2006). The 3rd stage juveniles are passed on to definitive hosts such as rats when they ingest infected gastropod intermediate hosts. In the absence of a definitive host ingesting the intermediate host, the 3rd stage juveniles can become quiescent and remain in the intermediate host tissue for months (Hollingsworth & Cowie, 2006). Occasionally, crabs, prawns and planarians act as paratenic hosts in which they passively carry 1st to 3rd juvenile stage worms; consumption of such paratenic hosts can pass on the 3rd stage juvenile worms (Kliks & Palumbo, 1992; Hollingsworth & Cowie, 2006). Once inside the definitive host, the

nematodes normally take only one to two days before they reach the central nervous system through the blood where they mature into adult worms after about 13 days (Kliks & Palumbo, 1992; Marquardt *et al.*, 2000).

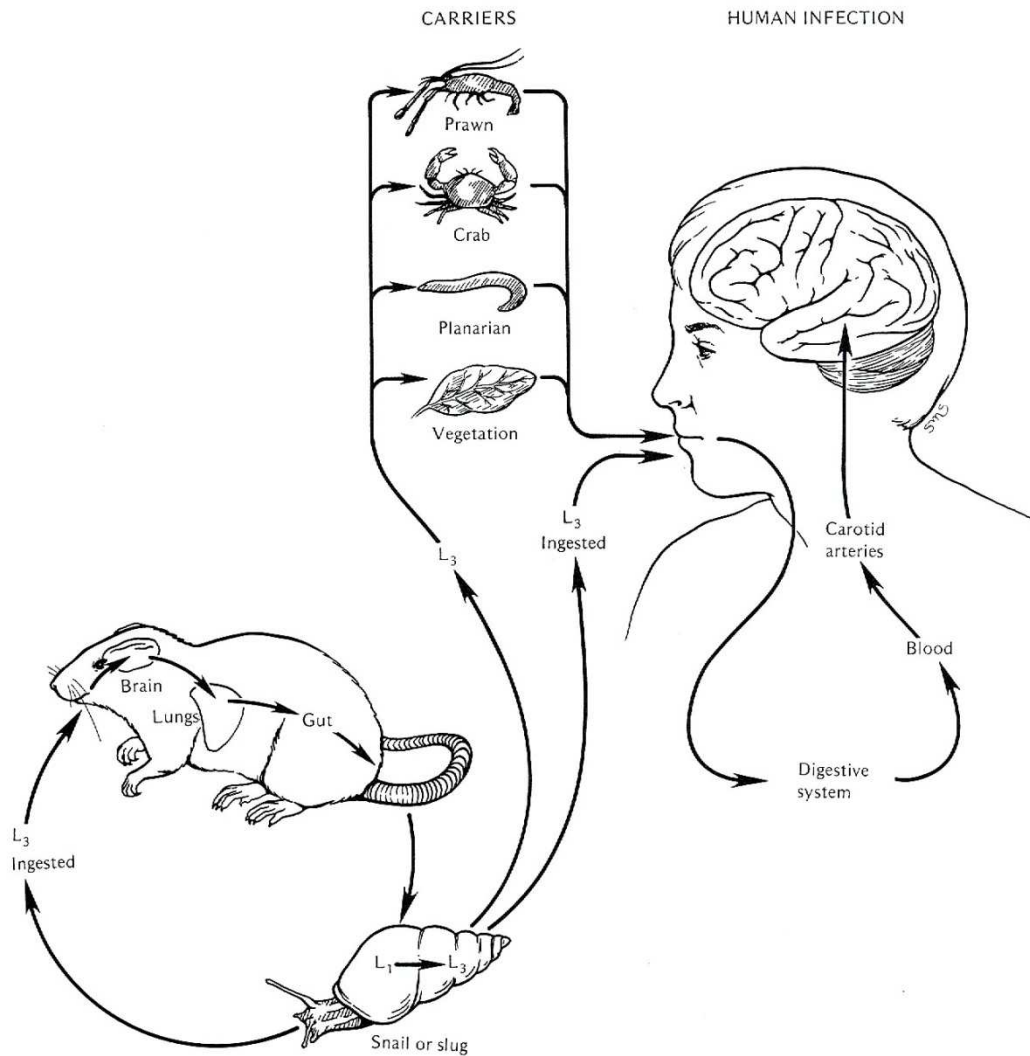


Figure 1.4: Life cycle of *Angiostrongylus cantonensis*. (From Marquardt *et al.*, 2000).

1.8.4. Natural intermediate hosts of *Angiostrongylus cantonensis*

Apart from *Achatina fulica*, there are several other gastropods that serve as natural intermediate hosts of *A. cantonensis* (Anderson, 2000). Gastropods so far identified as hosts include the following: *Hemiplecta sagittifera*, *Helicostyla*

macrostoma, *Cyclophorus* sp., *Chlorea fibula* (Westerlund & Chamberlain, 1969), *Imerinia plebia* (Salazar & Cabrera, 1969), *Bellamya ingallsiana*, *Bradybaena similaris*, *Cipangopaludina chinensis*, *Deroceras laeve*, *Euglandina rosea*, *Girasia peguensis*, *Indoplanorbis exustus*, *Laevicaulis alte* (also referred to by the name of *Veronicella altae* in the literature), *Macrochlamys resplendens*, *Microparmarion malayanus*, *Opeas japonicum*, *Pupina complanata*, *Pila ampullacea*, *P. scutata*, *Quantula striata*, *Sarasinula plebeia* (= *Vaginalus plebeius* in the literature) *Subulina octona* (Anderson, 2000) and *Pomacea canaliculata* (= *Ampullarium canaliculatus* in the literature) (Tsai *et al.*, 2001b). Other gastropods that were found experimentally to be susceptible to *A. cantonensis* infection could potentially become intermediate hosts. They are: *Biomphalaria* spp. *Bithynia* sp., *Bradybaena oceania*, *Bulinis* spp., *Deroceras reticulatum*, *Drepanotrema simmonsii*, *Euglandina rosea*, *Euhadra hickonsis*, *Ferrissia tenuis*, *Fossaria ollula*, *Fruticola despecta*, *Helicina orbiculata*, *Helisoma* sp., *Indoplanorbis exustus*, *Lanistes carinatus*, *Limax arborum*, *L. flavus*, *L. maximus*, *L. marginalis*, *Lymnaea* spp., *Marisa cornuarietis*, *Mesodon thyroidus*, *Onchidium* sp., *Physa acuta*, *Planorbis planorbis*, *Plesiophysa hubendicki*, *Segmentina hemisphaerula*, *Semisalcospira libertina*, *Stagnicola elodes* and *Succinea lauta* (Anderson, 2000).

1.8.5. Medical importance of *Angiostrongylus cantonensis*

Angiostrongylus cantonensis is the causative agent of eosinophilic meningoencephalitis (EME) or angiostrongyliasis in humans. In recent years, the geographic range of the nematode and the number of types of animal it infects have been growing rapidly, making EME an 'emerging' disease (Prociv *et al.*, 2000). People accidentally acquire the 3rd juvenile stage through any of the following routes of infection: (1) eating raw or undercooked snails or slugs (Marquardt *et al.*, 2000); (2)

drinking water or food such as raw vegetables or salads contaminated with the parasite such as raw vegetables for salads (Wallace & Rosen, 1969; Marquardt *et al.*, 2000); (3) eating raw or undercooked paratenic hosts like crabs, prawns and (by accident) planarians such as *Platydemus* (Kliks & Palumbo, 1992; Marquardt *et al.*, 2000); or (4) handling infected snails followed by failure to wash hands (Wan & Weng, 2004). Since humans are not the natural definitive hosts of *A. cantonensis*, these worms do not mature into adults inside the human body; however, their presence elicits a whole range of clinical manifestations typical of eosinophilic meningoencephalitis. Symptoms of this disease include headaches, stiffness of the neck, vomiting, paresthesia or abnormal sensation, fever, paralysis and tremors. In response to the presence of the worm, there is also a marked increase in the cerebrospinal fluid and peripheral blood of eosinophils, white blood cells responsible for combating infection and parasites (Kliks & Palumbo, 1992; Marquardt *et al.* 2000; Lee, 2002). Cases of EME are rarely fatal, with patients recovering after the symptoms subside within ten weeks (Hollingsworth & Cowie, 2006). It is not known how many infective 3rd stage juveniles are necessary to elicit EME in humans (Prociv *et al.*, 2000), although fatal cases often involve infection of hundreds or thousands of nematodes, as in the case of one Korean fisherman who died after consuming infected *Achatina fulica* in Pago Pago, American Samoa (Kliks *et al.*, 1982). To date, there is no known antihelminthic drug against *A. cantonensis*, although it is possible to treat infection at an early stage using thiabendazole (Roberts & Janovy, 2005). Most doctors are wary of recommending this drug, however, since worms may be more dangerous dead than alive as they tend to elicit a strong inflammatory reaction that could lead to the formation of a large granuloma and cause further damage (Marquardt *et al.*, 2000; Roberts & Janovy, 2005). On the other hand, mebendazole combined with corticosteroids have been successfully used to treat the symptoms of *A.*

cantonensis-induced meningoencephalitis and shorten the course of infection (Tsai *et al.*, 2001a; Wan & Weng, 2004). Aside from humans, other mammals such as dogs (Mason, 1987), horses (Costa *et al.*, 2000), tamarins (Carlisle *et al.*, 1998) and even a captive white-handed gibbon (Duffy *et al.*, 2004) have also been reported to be susceptible to *A. cantonensis* infection.

1.8.6. The dispersal of *Angiostrongylus cantonensis*

The expanding range of *Angiostrongylus cantonensis* has been attributed largely to the global distribution of rats, although the role of the intermediate hosts such as the Giant African Land Snail should not be ignored. The nematode is believed to have originated either in East Africa (Alicata, 1966) or South or Southeast Asia (Drozd *et al.*, 1975). Alicata (1966) noted the nearly parallel distribution of *A. cantonensis* and *A. fulica* and postulated that the rapid dispersal of the snail brought about the current geographical distribution of the parasite. Drozd *et al.* (1975), on the other hand, argued that murid rats were responsible, owing to the long association of the worm with its definitive hosts and the dispersal of the murid rats from tropical Asia. It would be valuable to determine which hypothesis is correct as this would identify which host is primarily responsible for the expanding range of the nematode; hence, appropriate measures could be set in place to control the host. Testing the hypothesis could be addressed by conducting a comprehensive survey to identify *A. cantonensis* among global populations of the nematode's definitive and intermediate hosts.

1.8.7. Molecular identification of *A. cantonensis* and other nematodes

It is, however, difficult to identify nematodes such as *A. cantonensis* to named species. Few taxonomic experts are available (Floyd *et al.*, 2002), and many worms

lack suitable morphological characters to facilitate identification, particularly in the infective juvenile stages (Newton *et al.*, 1998). An alternative to morphological identification is the use of DNA ‘barcodes’ that are unique to individual species. DNA barcoding makes use of a small section of a DNA sequence from a standardised region of the genome to identify species (Dasmahapatra & Mallet, 2006). A 650-bp fragment of the 5’ end of the mitochondrial cytochrome *c* oxidase subunit I gene (COI) is most commonly used for barcoding in animals (Hajibabaei *et al.*, 2007), although other genes have also been used. For instance, the small subunit (SSU) rRNA gene was employed to rapidly identify individual free-living marine nematodes in Southwest England (Bhadury *et al.*, 2006); the gene was likewise used on the parasitic nematodes of the sardine *Sardinella pilchardus* (Santos *et al.*, 2006). Through DNA barcoding, unidentified individuals of nematodes such as *Angiostrongylus cantonensis* could be assigned to species, irrespective of life cycle stage. This tool is applied here to rapidly identify 3rd stage juvenile *A. cantonensis* from their gastropod intermediate hosts.

1.9. Major objectives

This thesis will address five major issues concerning *Achatina fulica* over five chapters. A separate chapter, Chapter 2, will summarise all the general protocols to be used for the five chapters.

In Chapter 3, the phylogenetic relationships of the six achatinoid families (Achatinidae, Coeliacidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophorellidae) will be evaluated using an expanded coverage of the rRNA gene cluster as well as the nuclear actin and histone 3 genes and the mitochondrial cytochrome *c* oxidase subunit I and the 16S rRNA genes. The monophyly and validity of these taxonomic families will likewise be assessed.

In Chapter 4, comprehensive molecular-based phylogenetic analyses will be provided for the first time on the Achatinidae using the same molecular markers to be used in Chapter 3 in order to correlate molecular data with the available morphological data and to provide evidence of relationships among the members of this family. The designation of *Lissachatina* as a subgenus of *Achatina* will likewise be evaluated based on molecular data.

In Chapter 5, genetic variation among global populations of *Achatina fulica* will be examined using the 16S rRNA gene. This study will investigate the movement of *A. fulica* out of Africa and its subsequent spread throughout the tropics. It will establish if more than one genetic type has been introduced in the snail's new range and whether genetic bottlenecks are taking place in these populations. The lack of genetic variation, if any, could have a profound impact on the success of *A. fulica* as an invasive species.

In Chapters 6 and 7, focus will be on the nematode parasite of *A. fulica*, *Angiostrongylus cantonensis*. A survey of global populations of *A. fulica* for the parasite will be conducted to determine the possible role of the snail in the spread of the parasite. In Chapter 6, a rapid molecular method of identification for *A. cantonensis* from Philippine samples of *A. fulica* as well as the black slug *Laevicaulis alte* will be developed using a segment of the small subunit (SSU) rRNA gene. Using the same marker, a phylogenetic analysis will also be conducted on representative taxa of the genus *Angiostrongylus* to assess if Ubelaker's designation of *Parastrongylus* has merit based on molecular data. In Chapter 7, global populations of *A. fulica* will be screened for the nematode parasite using the molecular marker developed in Chapter 6. The role of the snail in spreading the parasite will be evaluated in this chapter. In addition, the presence of other types of nematodes in global populations of *A. fulica* will also be reported.

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CHAPTER 2 - General Protocols

Summarized in this chapter are general protocols that are referred to in the succeeding chapters. A brief description, along with some basic concepts, accompanies each method.

2.1. DNA extraction methods

DNA from fresh or preserved samples of snails and nematodes was extracted using several extraction protocols. Each method is described below, along with details of the rationale for their use. As the DNA extracts were used for subsequent PCR amplification, the methods emphasized the removal of polysaccharides and other PCR inhibitors (Demeke & Adams, 1992) as well as nuclease enzymes that could cleave DNA (Rolfs *et al.*, 1992).

2.1.1. DNA extraction of snail tissues using the CTAB protocol (modified from Hillis *et al.* (1996))

This standard and cost-effective protocol was used to extract DNA from snail tissues used for phylogenetic analyses in Chapters 3 and 4. The protocol was applied to fresh, fresh frozen and ethanol-preserved specimens. Traditionally, this technique also includes phenol (Rolfs *et al.*, 1992) but this was excluded here as phenol is a very toxic substance.

1. For ethanol preserved tissues, the tissue slices were soaked in 1 ml TE buffer (10 mM Tris-HCl, 1mM EDTA) for approximately 1 hour in order to remove excess ethanol to soften the tissue prior to DNA extraction.
2. Tissue was cut into small pieces, placed into 500 µl of CTAB solution [100mM Tris-HCl pH 8, 20mM EDTA pH 8, 1.4 mM NaCl, CTAB 2% (w/v)] and ground

using sterile glass beads and a plastic pestle. CTAB (cetyltrimethylammonium bromide) is a non-ionic detergent that precipitates polysaccharides and lyses cells (Richards *et al.*, 1995).

3. 20 μ l of Proteinase K (10mg/ml), an enzyme that digests proteins such as nucleases that cleave naked DNA, was added to each tube. This was followed by 10 μ l of β -mercaptoethanol, which precipitates polyphenolics (Rolfs *et al.*, 1992). The tubes were then vortexed then incubated at 55⁰ C for at least one hour until the tissue slices were completely digested.
4. 500 μ l of ice-cold chloroform-isoamyl alcohol (24:1) was added, after which the tubes were inverted several times for 5 minutes. This separated the DNA from proteins (Rolfs *et al.*, 1992). The tubes were then centrifuged for 10 minutes at 13,000 rpm, after which the aqueous phase was transferred to a new tube (~400 μ l). When necessary (i.e. there were a lot of proteins), step 4 was repeated.
5. 2.5 volumes (~1 ml) of ice-cold 95% ethanol and 1/10 volume (~40 μ l) of 3M sodium acetate (NaOAc) were added to the mix followed by overnight incubation at -80⁰ C to precipitate the DNA (Rolfs *et al.*, 1992). The tubes were centrifuged for 15 minutes at 13,000 rpm, then the supernatant was carefully removed.
6. The remaining pellets were washed with 500 μ l of ice-cold 70% ethanol and centrifuged for five minutes at 13,000 rpm to remove salts and small organic molecules. The ethanol was carefully removed afterwards.
7. The remaining pellets were air-dried on a heat block at 45°C for a maximum of 15 minutes. The pellets were then resuspended in 150 μ l TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.5).

8. Each suspension was then boiled for 15 min at 100°C to denature any remaining enzymes in order to prolong the life of the DNA extract.
9. The DNA extracts were stored at -80⁰ C until use.

2.1.2. DNA extraction of snail tissues using the Nucleon® PhytopureDNA™ Purification Kit

Because of its relatively fast and simple use, the Nucleon® PhytopureDNA™ kit was used as an alternative to the CTAB protocol (Section 2.1.1) in extracting DNA from snail tissues used for phylogenetic analyses (Chapters 3 and 4) as well as for the global genetic survey of *A. fulica* populations (Chapter 5). It produced no discernible difference in DNA quality in comparison to the CTAB method. Like CTAB, this kit allowed for the precipitation of polysaccharides. The protocol below generally followed the manufacturers' instructions except that volumes indicated were half of what was recommended while the times prescribed were lengthened.

1. 300 µl of reagent 1 was added to a tube containing approximately 0.1 g snail tissue from the foot muscle that was previously ground using sterile glass beads and plastic pestle. After mixing thoroughly, 100 µl of reagent 2 was added. The tubes were inverted several times until a homogenous mixture was obtained.
2. The tubes were placed in a 65⁰ C water bath for at least 30 minutes with regular manual agitation until the tissues were fully digested.
3. The tubes were then placed in ice for at least 30 minutes.
4. 250 µl of ice-cold chloroform was added followed by 50 µl Phytopure DNA extraction resin suspension that was thoroughly shaken prior to use. The tubes were inverted for 10 minutes followed by centrifugation at 13,000 rpm for 10

minutes. The DNA-containing phase above the resin layer was then transferred into a fresh tube.

5. An equal volume (~400 μ l) of cold isopropanol was added, after which the tubes were gently inverted several times to precipitate the DNA. The tubes were centrifuged at 13,000 rpm for 5 minutes to pellet the DNA. The isopropanol was pipetted out and discarded.
6. The DNA pellet was washed with 1.0 ml 70% ethanol, then centrifuged at 13,000 rpm for 5 minutes. The ethanol was pipetted out and discarded.
7. The remaining DNA pellets in the tubes were air-dried for a maximum of 15 minutes at 45-55⁰ C on a heat block.
8. The DNA pellets were resuspended in 150 μ l TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.5) and boiled for 15 minutes at 100°C.
9. The DNA extracts were stored at -80⁰ C until further use.

2.1.3. DNA extraction of snail tissues using the NaOH-lysis method

This method was used on the *Achatina fulica* population samples (Chapter 5) and was chosen over the two previous methods because it was considerably cheaper to use even if the quantity of DNA extracted was slightly less than that extracted in the previous methods. This disadvantage was offset by using more tissue, which could be readily obtained when using *A. fulica* population samples.

1. Tissue slices were ground in microfuge tubes containing 200 μ l of 0.1 N NaOH using sterile glass beads and a plastic pestle. The samples were boiled at 95-100⁰ C for 20 minutes to lyse the cells.
2. 100 μ l of sterile distilled water and 300 μ l of chloroform-isoamyl alcohol (24:1) were added. The tubes were vortexed, then centrifuged at 13,000 rpm for 10

minutes. The upper phase (~300µl) was collected and transferred into new tubes.

3. An equal volume of isopropanol (~300µl) was added to precipitate the DNA. The tubes were inverted several times, then stored at -80⁰ C for at least one hour.
4. The tubes were centrifuged at 13,000 rpm for 15 minutes, after which the isopropanol was carefully removed.
5. The pellets were washed with 500 µl of 70% ice cold ethanol then centrifuged for 5 minutes at 13,000 rpm to remove salts and small organic molecules. The ethanol was carefully removed.
6. The DNA pellets were air dried on a heat block at 45°C for a maximum of 15 minutes, after which they were resuspended in 150 µl TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.5). Each suspension was then boiled for 15 minutes at 100°C.
7. The DNA extracts were stored at -80⁰ C until further use.

2.1.4. DNA extraction of single nematodes using NaOH digestion

This method is a modification of the NaOH lysis method used to extract DNA from individual nematodes (Floyd *et al.*, 2002). The method was used on nematodes extracted from the gastropod intermediate hosts (Chapters 6 and 7) as well as on the adult angiostrongylid nematodes used for phylogenetic analyses and as references for molecular barcoding (Chapter 6).

1. Each nematode was placed in a separate tube containing 20 µl 0.25 M NaOH to digest the cuticle and cells. The tubes were centrifuged briefly to ensure that the nematodes were fully immersed in NaOH. This was followed by overnight

- incubation at 25⁰ C (or 3-16 hours; beyond that, over-digestion could lead to less intense or infrequent bands during PCR amplification).
2. The tubes were heated for three minutes at 95⁰ C, then cooled to room temperature. The tubes were centrifuged briefly to collect any liquid along the sides and the lid.
 3. The following were added in succession: 4 µl 1.0 M HCl to neutralize the NaOH; 10 µl 0.5 M Tris-HCl, and 5 µl 2% Triton X-100, a strong detergent that disrupts the cell membranes and effectively lyses the cells.
 4. The tubes were mixed and centrifuged briefly, then heated for 3 minutes at 95⁰C. The samples were cooled to room temperature (pH of digest should be between 8 and 9).
 5. The DNA extracts were stored at -80⁰ C until further use.

2.2. Polymerase chain reaction (PCR)

Polymerase chain reaction (PCR) entails the rapid amplification *in vitro* of specific DNA segments using the same principles involved in DNA replication. This technique was used to amplify fragments of the nuclear ribosomal (r)RNA cluster, actin and histone 3 genes of snails and the small subunit (SSU) rRNA of nematodes as well as the mitochondrial cytochrome c oxidase subunit I (COI) and 16S rRNA genes of snails. Crucial in any successful PCR is the pair of primers that bind to specific regions of the complementary strands and the DNA polymerase that facilitates the amplification. Other components of PCR are the deoxynucleotide triphosphates (dNTPs) that are incorporated in the growing chain, and MgCl₂ that acts as the enzyme co-factor of the DNA polymerase. There are three stages in PCR that are repeated over a number of cycles; these are: (1) denaturation of the double stranded DNA at 94⁰ C; (2)

annealing of the PCR primers to target sites in the DNA at specific temperatures; and (3) extension of the growing strand of synthesized DNA at 72⁰ C through the activity of the DNA polymerase (Reece, 2004).

2.2.1. PCR primers

Primers are short oligonucleotides (17-30 nucleotides) that bind to specific targets in the DNA (Reece, 2004). Several factors need to be considered for PCR primers to successfully amplify their intended targets. First, the melting temperature (T_m) of each primer in the primer pair, calculated as $2(A+T)+4(C+G)$, where A, C, G and T refer to the nitrogenous bases of the oligonucleotide, should be approximately equal so that they anneal to their target sites at almost the same time (Reece, 2004). Second, primers should not have strings of repeated nucleotides within their sequences in order to prevent annealing to non-specific targets that exhibit strings of complementary repeated nucleotides (Reece, 2004). Third, primers should not contain complementary sequences with each other or within themselves as these would result in primer dimers and secondary structures, respectively, which would lead to non-amplification (Reece, 2004). Fourth, the 3' end of the primers should match perfectly the target region for the polymerase enzyme to properly extend the primer beginning at the 3' end (Reece, 2004). Fifth, provided that the 3' ends of the primers match with their targets, some mismatched pairing could form partial bonds between the primer and the target. For example, G-T bonds also occur with some stability. Such a phenomenon could be taken advantage of when designing primers that need to be applied across many taxa in which ambiguous positions could potentially be present in the primer-binding sites. If a position in the target region could either be a C or a T, then the primer would be assigned a G for that position in order to bind to either C or T of the

target. Likewise, if the target contains a position that could either be an A or a G, then the primer that binds to it should preferentially contain a T for that corresponding position (Palumbi, 1996). Sixth, the length of the product being amplified by the primer pair would determine the duration of the extension step of the PCR. For instance, 30 seconds are generally needed to amplify products less than 500 bp, 60 seconds for products between 500 and 1500 bp, and 90 seconds for products longer than 1500 bp (Palumbi, 1996). Lastly, more than one round of PCR could be undertaken to ensure the success of amplification of the target segment. For instance, the product of the first round of PCR using one set of primers could be used as template for the second round using an internal set of primers in a two-step process called nested PCR (Aurelius *et al.*, 1991). This was applied to the rRNA cluster and the actin gene in which the first pair of primers amplified a larger fragment (approximately 4000 bp for the rRNA cluster and around 900 bp for the actin) while the second round made use of primer pairs that bound to targets within the larger fragments. This process facilitated the annealing of the primers in the second round where the target regions were more readily available.

2.2.1.1. Snail PCR primers

2.2.1.1.1. Nuclear genes

2.2.1.1.1.1. Ribosomal RNA (rRNA) gene cluster

The nuclear ribosomal RNA gene family encodes the small and large subunit rRNA genes that are incorporated into the small subunit and large subunit, respectively, of the ribosome that facilitates gene translation into amino acid sequences (Lewin, 2008). The gene family consists of five parts that are transcribed as a single unit: (1) the small subunit rRNA gene (SSU or 16-18S); (2) the first internal transcribed spacer (ITS 1); (3) the 5.8S gene; (4) the second internal transcribed spacer (ITS-2); and (5) the LSU rRNA gene (26-28S). A 5S gene found downstream of the LSU gene is separated from this array by an intron or a non-transcribed spacer and is independently transcribed in eukaryotes (Graur & Li, 2000; Lafontaine & Tollervey, 2001; Klug *et al.*, 2007). The ribosomal RNA gene cluster is repeated in a tandem array over the genome, with the actual number of copies varying across different taxa; for instance, nematodes have 50 to 100 copies, some mammals up to several hundred copies, while plants have up to thousands (Long & Dawid, 1980). The function of these genes in protein translation means that the genes are required in many copies and that these copies should be homogenous. The identity of these copies within a species is maintained by concerted evolution through purifying selection in which new variants are eliminated, thus leading to sequence homogeneity within a species (Gasser & Newton, 2000; Graur & Li, 2000). Variation between copies of the rRNA array in a species is rare; in land snails, variation has only ever been found in the ITS as detected by ambiguous positions in direct sequences of the ITS (Wade, pers comm.). The choice of this gene cluster for

phylogenetic study is ideal for several reasons: (1) it is easy to PCR amplify because more copies are available for the PCR primers to anneal to; (2) the genes are present in all animal taxa because of their conserved function in RNA translation into proteins; and (3) regions within the genes exhibit variable evolutionary rates, with some regions that are very conserved and are thus ideal for primer binding sites, while other regions are variable enough to be phylogenetically informative (Hillis & Dixon, 1991).

The primers listed in Table 2.1 were used to amplify an approximately 4000 nucleotide fragment of the rRNA gene cluster that was employed as a marker for the snail phylogenetic analyses (Chapters 3 and 4). The amplified region included nearly the entire large subunit (LSU) rRNA gene as well as the internal transcribed spacer 2 (ITS 2) region and about 80 nucleotides of the 5.8S rRNA gene (Figure 2.1). Actual differences in length of the amplified products among taxa were predominantly due to variations in length in the ITS 2 region and to a lesser extent the minor variations in the variable regions in the LSU. The rRNA cluster was amplified using nested PCR. The first round made use of the LSU-1 and LSU-12 or the LSU-2 and LSU-12 primer pairs. This served as the 1⁰ PCR for the rRNA gene fragment. Using the primary PCR products as template, the succeeding rounds of PCR (2⁰ PCR) amplified the internal rRNA fragments (fragment A using primers LSU-1/1iii and LSU-3/3iii; fragment B using LSU-2 and LSU-5; fragment C using primers LSU-4ii and LSU-7/7i; fragment D using LSU-6/6ii and LSU-9/9ii; fragment E using LSU-8/8ii and LSU-11/11ii; and fragment F using LSU-10/10i and LSU 12/12i; see also Table 2.1).

Table 2.1: Summary of the rRNA primers

LSU Fragment	Primers	Reference	Fragment Size (bp)
A	LSU-1 (sense): 5'-CTAGCTGCGAGAATTAATGTGA-3' LSU-3 (anti-sense): 5'-ACTTTCCTCACGGTACTTG-3'	Wade & Mordan (2000); Wade <i>et al.</i> (2001); Wade <i>et al.</i> (2006)	~900-1200 (difference due mostly to variable size of the ITS2 region; see also Fig. 2.1)
	LSU-1iii (sense): 5'-TGCGAGAATTAATGTGAATTGC-3' LSU-3iii (anti-sense): 5'-ACGGTACTTGTCCGCTATCG-3'	designed by C. Wade	
B	LSU-2 (sense): 5'-GGGTTGTTTGGGAATGCAGC-3' LSU-5 (anti-sense): 5'-GTTAGACTCCTTGGTCCGTG-3'	Wade & Mordan (2000); Wade <i>et al.</i> (2001); Wade <i>et al.</i> (2006)	~580
C	LSU-4ii (sense): 5'-GTCGGCATTCCACCCGACC-3' LSU-7 (anti-sense): 5'-GCAGGTGAGTTGTTACACACTC-3' LSU-7i (anti-sense): 5'-GTTGTTACACACTCCTTAGCGG-3'	designed by C. Wade	~700
D	LSU-6 (sense): 5'-AAGGTGCCAAACGCTGACGC-3' LSU-6ii (sense): 5'-GTGCCAAACGCTGACGCTCA-3' LSU-9 (anti-sense): 5'-CAGTCCTCAGAGCCAATCCTT-3' LSU-9ii (anti-sense): 5'-ACCCAGTCCTCAGAGCCAATC-3'	designed by C. Hudelot	~850
E	LSU-8 (sense): 5'-CCATATCCGCAGCAGGTCTC-3' LSU-8ii (sense): 5'-GTGCACAGCCTCTAGTCGATA-3' LSU-11 (anti-sense): 5'-CTGAGCTCGCCTTAGGACAC-3' LSU-11ii (anti-sense): 5'-TCCTCCTGAGCTCGCCTTAG-3'	designed by C. Hudelot	~850
F	LSU-10 (sense): 5'-ATCCGCTCTGAAGACAGTGTC-3' LSU-10i (sense): 5'-GGCCGCGATCCGTCTGAAGA-3' LSU-12 (anti-sense): 5'-TTCTGACTTAGAGGCGTTTCAG-3' LSU-12i (anti-sense): 5'-GGCTTCTGACTTAGAGGCGTT-3'	designed by C. Hudelot	~500

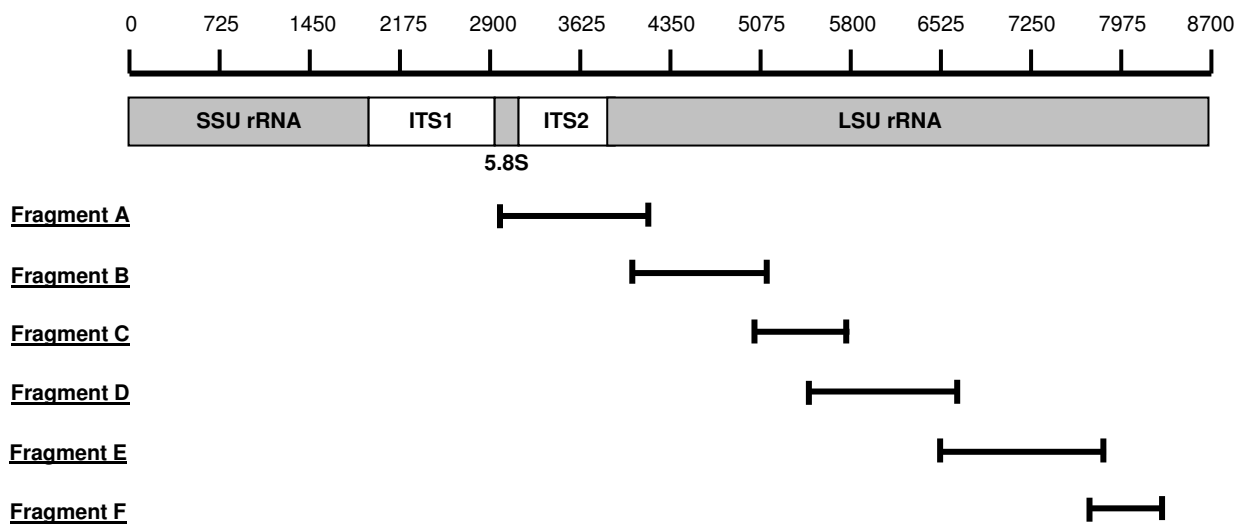


Figure 2.1: Schematic diagram of the rRNA gene cluster based on the complete sequence of *Rattus norvegicus* (GenBank X00133) and showing the position of the primers used in this study.

2.2.1.1.1.2. Actin gene

Actin is a protein involved in various functions such as muscle contraction, cell division and differentiation (Hightower & Meagher, 1986; Hernan, 1993) and is highly conserved across animal groups (Hightower & Meagher, 1986; Palumbi, 1996). Actin is coded by several genes that arose from gene duplication and divergence, resulting in the presence of different forms (isoforms) of the actin protein within an organism (Hightower & Meagher, 1986). These protein isoforms can be classified into two broad categories: the muscle (α) type and the cytoplasmic (β) type (Vandekerckhove & Weber, 1978; Adema, 2002), with the number of isoforms for each category varying between animal groups. In mammals, there are two cytoplasmic actins and four muscle-type actins (Vandekerckhove & Weber, 1978); the same numbers were also found in *Drosophila* (Fyrberg *et al.*, 1981). In molluscs, the number and type of actin isoforms appear to vary among taxa based on the few that have been evaluated to date. DesGroseillers *et al.* (1994) found between three and five genes in the sea hare *Aplysia*

californica that probably coded for the muscle type actin, while Patwary (1996) found 12-15 genes in the bivalve *Placopecten magellanicus* that were also of the muscle type. Carlini *et al.* (2000) surveyed 44 taxa among the coleoid cephalopods and found at least one gene each of the muscle type and cytoplasmic type actins and a third that was most probably a pseudogene. Adema (2002) sequenced one gene of a cytoplasmic actin isoform in each of six planorbid basommatophoran species using cytoplasmic actin-specific primers. However, Southern blotting and hybridization experiments using probes specific for actin yielded up to five genes in *Biomphalaria glabrata* and *Helisoma trivolis*, prompting Adema not to discount the possibility that some of these genes could code for muscle type actin isoforms. Morgan *et al.* (2002) extended the actin survey of the planorbids to 38 taxa using the cytoplasmic actin to correlate the relationship of these snails to the evolution of their *Schistosoma* trematode parasite. Using actin for phylogenetic analyses could pose a problem if the sequences being compared are paralogous, which resulted from gene duplication, rather than orthologous, as in the case of muscle and cytoplasmic actins that arose from gene duplication in the distant past or even within each type due to more recent duplications. Using paralogous sequences could lead to inferring incorrect relationships; awareness of their presence in the sequence data is therefore crucial in any phylogenetic analysis (Bailey *et al.*, 2003). Donald *et al.* (2005) acknowledged the problem of paralogs in actin, but they justified the use of actin to elucidate the phylogeny of marine top shells by employing primers specific for the gene that codes for a single isoform of the cytoplasmic actin and by showing agreement of the actin phylogeny with the 16S rRNA and COI phylogenies. How to address the issues of paralogous sequences in the actin gene is discussed further in Sections 2.9.12 and 2.9.13, pp. 89-91.

The primers listed in Table 2.2 amplified an approximately 900 bp fragment of the actin gene; this fragment was used for the snail phylogenetic analyses in Chapters 3 and 4. Nested PCR was carried out in which a primary PCR product was amplified using the primers ActF2 and ActR1; this fragment was then used as template for the secondary PCR using ActF1 and ActR. If amplification failed, several combinations of primers from the primary and secondary PCR were tried until a product was amplified.

Table 2.2: Summary of the actin primers.

Primers	Reference	Fragment Size (bp)
ActF1 (sense): 5' -TATGTTGGTGATGAGGCTCAG-3'	Morgan <i>et al.</i> (2002)	~900
ActF2 (sense): 5' -GGTATGGGTCAGAAGGACAGCTATG-3' ActR1 (anti-sense): 5' -GAAGCATTTCCTGTGGTCAATG-3' ActR (anti-sense): 5' -GATCCACATCTGTTGGAAGGT-3'	Designed by C. Hudelot	

2.2.1.1.1.3. Histone 3 gene

Histone 3 (H3) forms part of the histone core protein octamer that packs the eukaryotic chromatin into bead-like structures (Lewin, 2008). Histone core proteins are some of the most conserved proteins known, suggesting a function that is identical across eukaryotes (Lewin, 2008). Histone genes occur as a tandemly repeating unit comprising the different genes that code for the subunits of the core protein octamer. In *Drosophila*, the repeating unit consists of five genes in the following order, H1, H2A, H2B, H3 and H4; this repeating unit occurs 110 times (Lifton *et al.*, 1977). In bivalve molluscs, a different order was found in the genome of the bivalve mussel *Mytilus galloprovinialis*: H4, H2B, H2A, H3 and H1, with the copy number averaging more

than 200 per haploid genome (Eirin-Lopez *et al.*, 2004). Transcription of the H3 and H4 genes in the stylummatophorans occurs in opposite directions as in other protostomate animals (Armbruster *et al.*, 2005) but not in the bivalve *Mytilus* (Eirin-Lopez *et al.*, 2004). Among the gastropods, Colgan *et al.* (2000) noted a high codon usage bias for the H3 gene. As with the rRNA genes, H3 and H4 genes are subject to concerted evolution, thus leading to sequence homogeneity among copies of the genes within the species (Liao, 1999). The primers listed in Table 2.3 amplified a 331 bp fragment that was also used for the snail phylogenetic analyses in Chapters 3 and 4. Different primer combinations amplified roughly the same fragment.

Table 2.3: Summary of the histone 3 primers

Primers	Reference	Fragment Size (bp)
H3aF (sense): 5'-ATGGCTCGTACCAAGCAGACVGC-3' H3aR (anti-sense): 5'-ATATCCTTRGGCATRATRAGTGAC-3'	Colgan <i>et al.</i> (1998)	328
H3Fm (sense): 5'-ATGGCTCGTACCAAGCAGAC-3' H3Fm1 (sense): 5'-ATGGCTAGAACGAAGCAGAC-3' H3Rm (anti-sense): 5'-TCCTTGGGCATGATGGTGAC-3' H3Rm1 (anti-sense): 5'-CCAACCTGAATATCTTTGGGCAT-3'	designed by C. Hudelot	331-340

2.2.1.1.2. Mitochondrial primers

2.2.1.1.2.1. Cytochrome c oxidase subunit I gene

Cytochrome c oxidase is an enzyme that transfers electrons from cytochrome c to O₂ during the electron transport chain in the mitochondrion (Zubay *et al.*, 1995). A typical enzyme has three functional subunits in which subunits I and II contain the

electron carriers (Alberts *et al.*, 2008). Although the amino acid sequence of the subunit I is conserved across the different animal phyla, the nucleotide sequence is subject to silent mutations (Palumbi, 1996), particularly in the 3rd codon positions. In fact, its variability that yields phylogenetic signal and its robust universal primers have made the COI a marker of choice for DNA barcoding in animals (Hebert *et al.*, 2003). Several primer pairs listed in Table 2.4 amplified a fragment that was used in Chapters 3 and 4. Different primer combinations amplified roughly the same fragment.

Table 2.4: Summary of the cytochrome c oxidase subunit I primers

Primers	Reference	Fragment Size (bp)
LCO 1490 (sense): 5' -GGTCAACAAAATCATAAAGATATTGG-3' HCO 2198 (anti-sense): 5' -TAAACTTCAGGGTGACCAAAAAATCA-3'	Folmer <i>et al.</i> (1994)	655
STY_LCOi (sense): 5' -TCAACGAATCATAAGGATATTGG-3' STY_LCOii (sense): 5' -ACGAATCATAAGGATATTGGTAC-3' STY_LCOiii (sense): STY_HCO: (anti-sense) 5' -GAATTAAAAATATATACTTCTGGGTG-3'	designed by I. Fontanilla	628-667

2.2.1.1.2.2. 16S ribosomal (r) RNA gene

The 16S rRNA gene transcribes a ribosomal RNA that folds into a secondary structure following base pairing of the nucleotides within it, after which it is incorporated in the mitochondrial ribosome and is used for translation of proteins (Lewin, 2008). Among the Mollusca, 16S exhibits extreme variation in length, with those of the Stylommatophora being the shortest (Lydeard *et al.*, 2000). Two primer pairs are listed in Table 2.5. The first pair (STY_16Sarm and STY_16Sbrm) amplified a 420-450 bp fragment that was used for the snail phylogenies in Chapters 3 and 4. The second pair (16S1i and 16S_SSCP2i) amplified a 293 bp fragment that was used for the

global population survey of *Achatina fulica* in Chapter 5.

Table 2.5: Summary of the 16S rRNA primers

Primers	Reference	Fragment Size (bp)
(for the phylogeny studies-chapters 3 & 4) STY_16Sarm (sense): 5'-CTTCTCGACTGTTTATCAAAAACA-3' STY_16Sbrm (anti-sense): 5'-GCCGGTCTGAACTCAGATCAT-3'	Bonnaud <i>et al.</i> (1994)	420 – 500
(for the SSCP study-chapter 5) 16S1i: 5'-TGACTGTGCAAAGGTAGCATAA-3' 16S_SSCP2i: 5'-CCTAGTCCAACATCGAGGTC-3'	designed by I. Fontanilla	293

2.2.1.2. Nematode PCR primers

The small subunit (SSU) rRNA, which is a part of the ribosomal gene family, has been used to elucidate the phylogeny of the Nematoda (Blaxter *et al.*, 1998; Meldal *et al.*, 2006). The 5' end of the SSU has also been employed as a marker to identify marine nematodes (Bhadury *et al.*, 2006). The primers listed in Table 2.6 amplified four overlapping fragments that included nearly the entire region of the small subunit (SSU) rRNA gene in nematodes (Blaxter *et al.*, 1998); this fragment was used to infer the phylogeny of the Angiostrongylidae relative to other metastrongylids as well as to determine the most suitable marker to identify *Angiostrongylus cantonensis* in Chapter 6. The relative locations of the binding sites for these primers are shown in Figure 2.2.

Table 2.6: The four overlapping primer pairs used to amplify nearly the entire region (approximately 1670 nucleotides) of the SSU rDNA gene.

	Primers	Reference	Fragment size (bp)
Set A	SSU_F_07 (sense): 5' -AAAGATTAAGCCATGCATG-3' SSU_R_09 (anti-sense): 5' -AGCTGGAATTACCGCGGCTG-3'	Blaxter <i>et al.</i> (1998)	480
Set B	SSU_F_11 (sense): 5' -AAGTCTGGTGCCAGCAGCCGC-3' SSU_R_26 (anti-sense): 5' -CATTCTTGGCAAATGCTTTTCG-3'	Blaxter <i>et al.</i> (1998)	360
Set C	SSU_F_24 (sense): 5' -AGRGGTGAAATYCGTGGACC-3' SSU_R_23 (anti-sense): 5' -TCTCGCTCGTTATCGGAAT-3'	Blaxter <i>et al.</i> (1998)	390
Set D	SSU_F_02 (sense): 5' -GGAAGGGCACCACCAGGAGTGG-3' SSU_R_81 (anti-sense): 5' -TGATCCWKC YGCAGGTTTCAC-3'	Blaxter <i>et al.</i> (1998)	600

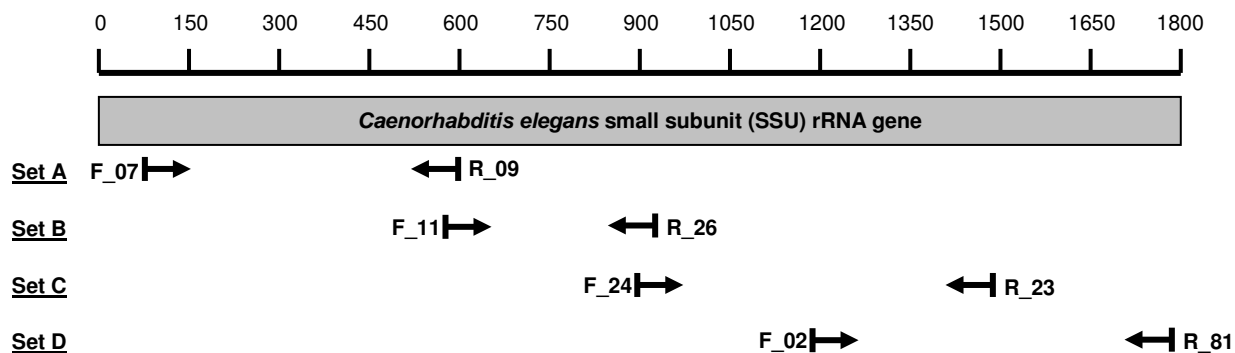


Figure 2.2: Schematic diagram of the SSU rRNA gene based on the complete sequence of *Caenorhabditis elegans* (GenBank X00133) and showing the position of the primers used in this study.

2.2.2. PCR components

Listed in Table 2.7 are the PCR components used and their concentrations. The optimal concentrations varied depending on the size of the gene and the number of copies available per cell. In particular, the concentration of MgCl₂ differed between the nuclear (1.5 mM) and the mitochondrial (2.5 mM) genes.

Table 2.7: PCR components used and their concentrations for the different genes

Components	Initial concentration	Final concentration		Volume (μ l)	
Buffer	10X	1X		5.0	
dNTPs	1.25 mM	1 ⁰ PCR for LSU: 300 μ M	2 ⁰ PCR for LSU/1 ⁰ & 2 ⁰ PCR for actin/ other genes: 200 μ M	1 ⁰ PCR for LSU: 12.0	2 ⁰ PCR for LSU/1 ⁰ & 2 ⁰ PCR for actin/ other genes: 8.0
^a MgCl ₂	50 mM	LSU rRNA/ Actin/ Histone3: 1.5 mM	16S/ Cytochrome c oxidase I: 2.5 mM	LSU rRNA/ Actin/ Histone3: 1.5	16S/ Cytochrome c oxidase I: 2.5
Primer 1	10 mM	200 μ M		1.0	
Primer 2	10 mM	200 μ M		1.0	
^b Q-solution	5X	1X		10	
Taq	1 unit/ μ l	0.01 unit/ μ l		0.5	
Sterile distilled H ₂ O	--	--		To make final volume of 50 μ l	

^aSome manufacturers provided PCR buffers that already contained MgCl₂ (e.g. QIAGEN™ with 25 mM concentration), while other manufacturers separated the buffer and the MgCl₂ (e.g. Bioline™, which separately provided 50 mM MgCl₂). Care was taken in order to guarantee that the recommended final MgCl₂ concentration was followed.

^bProvided by QIAGEN™

2.2.3. PCR running conditions

Different reaction cycles were employed depending on the size of the expected PCR product. For the rRNA gene cluster, a very long extension step (5 minutes) was used in the 1⁰ PCR reaction in order to ensure that the approximately 4000 bp fragment was amplified. For the rRNA 2⁰ PCR reactions, which amplified the shorter internal rRNA fragments, a shorter extension step was used. For all other genes, reaction cycles followed those used for the rRNA 2⁰ PCR reactions.

2.2.3.1. For rRNA 1⁰ PCR

hot start at 96⁰ C for 2 minutes

94 ⁰ C for 30 sec	}	35 cycles
50 ⁰ C for 30 sec		
72 ⁰ C for 5 minutes		

2.2.3.2. For rRNA 2⁰ PCR and all other genes:

hot start at 94⁰ C for 2 minutes

94 ⁰ C for 30 sec	}	38 cycles
^a 45 ⁰ C for 30 sec		
65 ⁰ C for 60 sec		

^a45⁰ C was the optimal annealing temperature for the mitochondrial primers; this temperature also worked well for the nuclear primers

2.3. Gel electrophoresis

Once the PCR products were amplified, they were then separated and visualized through gel electrophoresis. PCR products were loaded in wells in an agarose gel, after which an electric current was applied to move the DNA towards the positive electrode. Different-sized DNA fragments migrate at different rates with smaller fragments moving rapidly through the gel matrix and larger fragments moving more slowly. Different sized PCR products would therefore be found at different positions of the gel at the end of electrophoresis. The DNA molecules were visualized when the ethidium bromide in the agarose gel intercalated between the nitrogen bases of the DNA and fluoresced under UV light (Reece, 2004). Below is the general protocol for agarose gel electrophoresis.

1. For a 1.25% agarose gel, 1.25 g agarose was dissolved in 100 ml TBE buffer [54 g Tris, 27.5 g boric acid and 20 ml 0.5 EDTA dissolved in 1000 ml distilled water to make 5X TBE buffer, then diluted to 1X before use]. The agarose was melted in a microwave oven and the molten gel cast in a container. The combs were added to make the wells.
2. Once the gel had set, the combs were removed and the PCR products loaded on the wells, after which the gel was submerged in a tank containing 1X TBE.
3. A current (100-120 V) was applied through the gel using a portable power supply.
4. Once the PCR products had run their course through the gel, the gel was removed from the tank and placed on a UV transilluminator to visualize the bands.

2.4. Gel extraction

Since most of the PCR amplifications yielded more than one band, it was necessary to separate these bands by electrophoresis prior to purifying the band of the correct size from the agarose gel in preparation for sequencing. Each band in the gel represented a PCR product of a particular size, thus the nucleic acid purified from a particular band is assured of having no other PCR product or primer of a different size. For bands that were very close to each other, care was taken when cutting these bands.

The protocol follows the manufacturers' instructions for the Eppendorf Perfectprep™ Gel Extraction Kit.

1. The PCR product was excised from the agarose gel with a sterile, sharp scalpel and placed in a 1.5ml microfuge tube.

2. The gel slice was weighed. Three volumes of Binding Buffer were added to every volume of the excised gel (100 mg ~ 100 μ l).
3. The gel slices were incubated at 50 $^{\circ}$ C for 10 minutes (or until the gel slice had completely dissolved). To help dissolve the gel, the tubes were inverted every 2-3 minutes during the incubation.
4. One gel volume of isopropanol was added and mixed to precipitate the PCR product.
5. A spin column with a membrane filter was placed in a 2 ml collection tube. To bind the PCR product, the sample was applied to the spin column provided and centrifuged at 13,000 rpm for 1 minute. The flow-through was discarded and the column was placed back into the same collection tube. (Maximum volume capacity of spin column was 800 μ l. For sample volumes of more than 800 μ l, the remaining samples were also loaded and the collection tube centrifuged again until all the samples were used up.)
6. 750 μ l of the wash buffer were added to the column and centrifuged at 13,000 rpm for 1 minute. After discarding the flow-through the column was replaced in the collection tube and centrifuged at 13,000 rpm for another minute to completely remove traces of the Wash Buffer.
7. The spin column was placed into a clean 2 ml collection tube. 30 μ l of the Elution Buffer (10 mM Tris-Cl, pH 8.5) was added to the center of the membrane filter and the collection tube centrifuged at 13,000 rpm for 1 minute to elute the PCR product.
8. The spin column was discarded, and the eluted product was stored at -20 $^{\circ}$ C until further use.

2.5. DNA quantification

Eluted PCR products were quantified using a NanoDrop® ND-1000 spectrophotometer prior to DNA sequencing. For cycle sequencing using the ABI PRISM™ Big Dye Version 3.1 Kit (see next section), approximately 1-3 ng concentration was recommended for every 100-200 bp of the PCR product to be sequenced.

2.6. Cycle sequencing

Cycle sequencing was employed to derive the sequence of the purified PCR products. This method uses the principle of dideoxy chain termination (also known as the Sanger method) in determining DNA sequences. In this procedure, a template DNA such as the double stranded PCR product was subjected to amplification of each strand in separate tubes using one primer each. A small amount of fluorescently labeled dideoxynucleotide triphosphates (ddNTPs) were included in the sequencing mix. These ddNTPs differed from the normal dNTPs in that they had an H group rather than an OH group on the 3rd carbon of the ribose sugar. When a ddNTP was incorporated instead of a dNTP in the growing chain, no other dNTP would bind to the 3rd carbon of the ribose sugar in the absence of the OH group, thereby prematurely terminating the sequence. This resulted in a mixture of different sized fragments all terminating in a fluorescently labeled ddNTP. After the sequencing reaction, these fragments were run on a gel in an automated DNA sequencer where the fragments separated according to size. As the samples passed through a certain point along the course of the gel, an argon ion laser fluoresced the dye attached to the samples. Each ddNTP (A, C, G and T) carried a dye that fluoresced at a particular wavelength, which were picked up and registered by a computer as a unique color. A sequence complementary to the template DNA was then

generated as a series of differently colored peaks (Reece, 2004). The protocol below follows the manufacturers' instructions for the ABI PRISM™ Big Dye Version 3.1 Kit except that volumes were reduced by half for most components while less than 20% of the Big Dye was used.

2.6.1. Cycle sequencing components

Listed below are the components used for each sample sequenced. Sequencing of the forward and reverse strands was carried out in separate tubes.

1. 1.5 µl Big Dye Mix (for 500-1000 bp PCR product)
2. 1.6 µl primer (1 µM)
3. purified PCR product (approximately 1-3 ng/ 100-200 bp)
4. sterile distilled water to make a final volume of 10 µl.

2.6.2. Cycle sequencing running conditions

96 ⁰ C for 5 minutes	} 25 cycles
96 ⁰ C for 10 sec	
50 ⁰ C for 5 sec	
60 ⁰ C for 4 minutes	

2.6.3. Precipitation of cycle sequencing samples

Once the cycle sequencing was complete, the samples were precipitated prior to being run in an automated sequencer such as the Applied Biosystems 3730 DNA sequencer. The instructions below follow those recommended by the manufacturers of the ABI PRISM™ Big Dye Version 3.1 Kit but with some modifications.

1. For each 1.5 ml microfuge tube, 2.0 µl 3M NaOAc (pH 4.6) and 50 µl 95% ethanol was added. The entire cycle sequencing mix was then added into the

NaOAc-ethanol mixture. The tubes were vortexed and set aside for at least 30 minutes. This step precipitated the different-sized PCR products in the tube.

2. The tubes were centrifuged for 20 minutes at 13,000 rpm. The supernatant was then carefully removed.
3. The pellet was rinsed with 190 μ l 70% ethanol, then vortexed briefly.
4. The tubes were centrifuged for 5 minutes at 13,000 rpm, after which the supernatant was carefully removed.
5. The tubes were placed with the lid open in a heat block or thermal cycler at 90^o C for one minute. The samples were then sent to the Department of Biochemistry of Oxford University or the DNA Sequencing Facility of the Natural History Museum for sequencing.

2.7. Cloning

There are instances in which PCR amplifies more than one copy of a particular gene fragment. This happens if an individual is either a heterozygote for a particular gene or actually possesses several genes. Once the PCR products are sequenced, variable sites among the different copies are manifested as ambiguous sites as PCR does not discriminate between copies as long as they have the same primer-binding sites. Such is the case for actin where several genes coding for actin are sometimes found within the same individual (Hernan, 1993). Where possible, all the different genes should be amplified, cloned and then sequenced in order to identify the presence of paralogous sequences. However, cloning was not possible for the snail phylogenetic analyses in Chapters 3 and 4 due to time and cost constraints, so direct-sequencing from PCR products was employed instead, with ambiguity codes assigned to positions of ambiguity in the direct sequences (see Section 2.9.3, pp. 72-73). These ambiguous sites

were then utilized as ambiguities in the tree-building process. Sequences obtained via cloning from the different copies from a few selected taxa served as a reference in assigning codes for the ambiguous sites. One taxon that did not exhibit any ambiguous sites (i.e. *Leptinaria lamellata*, see Appendix 3.8C, p. 436) was also used for cloning in order to check if any ambiguity was missed in the direct sequence. It should be emphasized that, in the absence of sequencing all the actin clones for each taxon, using direct sequence with ambiguous sites implies using an “average” or consensus sequence for all the possible copies of the actin gene for that species. Caution must therefore be taken in interpreting trees involving taxa with potentially more than one actin gene.

Sequencing different copies of the actin gene separately was achieved by first cloning the PCR products in bacteria using a plasmid vector. A special plasmid used in cloning called the *pUC* plasmid included an ampicillin-resistant gene and a *lacZ* gene that coded for a subunit of the β -galactosidase enzyme that breaks down galactose. Foreign DNA can be inserted into this plasmid within the *lacZ* gene. Once the insert is incorporated in the plasmid, the plasmid was allowed to be taken in through chemical transformation by a *lacZ* mutant *E. coli* that could not produce a functional β -galactosidase enzyme. These bacteria were then allowed to grow in an agar plate containing ampicillin and galactose. Bacteria that took in the plasmid were able to grow on the medium because of the presence of the ampicillin-resistant gene in the plasmid. Bacteria that harbored plasmids with inserts would not exhibit β -galactosidase activity in the presence of galactose because the inserts were placed within the *lacZ* gene, thereby disrupting its transcription. Colonies of these bacteria appeared white when compared to bacteria with functional *lacZ* in the plasmid, which appeared as blue colonies. These white colonies were then picked from the agar and were subcultured in a liquid medium to allow them to grow, thereby producing a vast number of copies of

the insert. The plasmids were then isolated from the bacteria and the inserts direct-sequenced using plasmid specific primers that bound to the flanking regions of the annealing site of the insert. As only one molecule of foreign DNA could be inserted into a plasmid, the inserts derived from a single colony represented a single copy of that insert (Reece, 2004). In this way individual copies of the actin gene were obtained for sequencing.

The protocol below follows the manufacturers' instructions for the Invitrogen™ TOPO Cloning Kit except that the volume of competent cells used was reduced by half.

2.7.1. Agar plate preparation

1. Around 15-20 ml LB agar was poured into a Petri dish and set aside at room temperature to solidify the agar. The agar plate was left slightly open in a drying cabinet for 30 minutes to remove condensation from the lid.
2. 17 μ l of 50 mg/ml ampicillin was spread on top of the agar plate before replacing the lid.
3. The agar plate was then warmed at 37⁰ C for 30 minutes.
4. 80 μ l of 20 mg/ml X-gal was spread on the agar plate, after which it was incubated at 37⁰ C until use.

2.7.2. Setting up the TOPO™ Cloning Reaction

1. The following were mixed gently and incubated for five minutes at room temperature:

PCR product	1.0 µl	}	provided in the kit
Salt solution	0.5 µl		
water	1.0 µl		
TOPO	0.5 µl		

2. 1 µl of the TOPO cloning reaction from (1) was added to 25 µl of One-Shot™ chemically competent cells and incubated on ice for 5 minutes.
3. The cells were heat-shocked for 30 seconds at 42⁰ C without shaking. This allowed the cells to take in the plasmids. The tubes were immediately transferred in ice.
4. 125 µl of previously defrosted SOC medium from Box 2 was added, and the tube was then capped tightly and shaken horizontally (200 rpm) at 37⁰C for one hour.
5. 50 µl from each transformation was spread on a pre-warmed (37⁰ C) selective agar plate containing X-gal and incubated overnight at 37⁰ C.
6. White colonies were picked and cultured overnight in 5-ml LB medium containing 50 ug/ml ampicillin.

2.7.3. Plasmid isolation

1. The culture was transferred into a microfuge, after which it was centrifuged for five minutes at 13,000 rpm to discard the supernatant.
2. The process was repeated until most of the culture was used up. [A small amount (~500 µl) was set aside in a separate tube where ~125 µl of 80% sterile

glycerol was added. The tubes were then stored at -80°C so that the culture could be re-grown in the near future should the need arise.]

3. 1.0 ml of cold TE buffer was added, after which the tube was vortexed. The tube was then centrifuged for 5 minutes at 13,000 rpm to discard the supernatant.
4. 100 μl of cold TE buffer was added and the tube was then vortexed. Incubation on ice for a minimum of 15 minutes followed.
5. 200 μl of 0.2 N freshly prepared NaOH / 1% SDS solution was added. This mixed the bacterial cells.
6. 150 μl of 3M KOAc, pH 4.8 was added followed by gentle mixing. The tube was mixed gently and left in ice for 5 minutes. This stage precipitated the proteins and neutralized the alkaline pH.
7. The tube was centrifuged for 5 minutes at 13,000 rpm. The supernatant was moved to a separate tube while the pellet was discarded.
8. 10 μl of 2 mg/ml RNase was added, and the tube was then incubated at 37°C for 30 minutes. This degraded RNA.
9. An equal volume ($\sim 450\ \mu\text{l}$) of phenol-chloroform was added, after which the tube was inverted several times. It was then centrifuged for 5 minutes at 13,000 rpm. The aqueous phase was transferred to a new tube.
10. An equal volume ($\sim 400\ \mu\text{l}$) of chloroform was added, after which the tube was inverted several times. It was then centrifuged for five minutes at 13,000 rpm. The aqueous phase was transferred to a new tube.
11. 2.5 volumes ($\sim 1\ \text{ml}$) of ice-cold 95% ethanol were added. The tube was placed in -80°C for at least 20 minutes or in -20°C for at least 2 hours and 30 minutes.

It was then centrifuged for 10 minutes at 13,000 rpm. The aqueous phase was discarded.

12. 1.0 ml of 70% ice-cold ethanol was added, and the tube was then centrifuged for 5 minutes at 13,000 rpm before the aqueous phase was discarded.

13. The pellets were dried at 45⁰ C for 15 minutes. These were then eluted in 20 µl sterile distilled water and stored at -20⁰ C until use.

2.7.4. Sequencing of the insert

The protocols for sequencing of the insert were the same as those in Section 2.6 (see pp. 59-61) except that the M13 forward primer (5' – GTAAAACGAC GGCCAGTGAA – 3') and the M13 reverse primer (5' – CAGGAAACAG CTATGACCATGA – 3') were used. These primers bound to sites that flank the annealing site of the insert in the plasmid. This ensured that the sequence, which included some segments of the plasmid DNA, was derived from the insert.

2.8. Single strand conformational polymorphism analysis

Determining variation at the DNA level among populations need not require sequencing every sample. One method of detecting variation is single strand conformational polymorphism analysis (SSCP), which was used to evaluate the variation in global populations of *Achatina fulica* (Chapter 5). Amplified PCR products from a marker gene such as the 16S rRNA was compared for nucleotide variation by denaturing them into single strands and then allowing them to migrate slowly over a long period in a polyacrylamide gel. Single stranded DNA underwent conformational changes that depended on their nucleotide composition. Therefore, similar sized fragments that varied even in just a single nucleotide assumed different conformations

and hence exhibited different migration rates. The DNA was visualized by silver staining in which silver nitrate bound to the DNA. The pattern generated by the two single stranded DNA's on a polyacrylamide gel represented the haplotype of the individual for the amplified genetic marker (Orita *et al.*, 1989). The SSCP technique, however, only works for fragments that are 400 bp or shorter (Sunnucks *et al.*, 2000); in this study, the amplified PCR products derived from the 16S primers were 293 bp long. The subsequent protocol follows that of Orita *et al.* (1989) with some modifications.

2.8.1. Native polyacrylamide gel preparation:

1. The following were mixed in a beaker:

2X MDE™ gel solution (Cambrex Bio Science®)	7.5 ml
1X TBE buffer	18 ml
sterile distilled water	4.0 ml
TEMED	40 µl
ammonium persulfate	400 µl

Note: The ammonium persulfate was added last as this catalyzed the polymerization of the gel.

2. Once the reagents were mixed, the solution was immediately poured into two glass plates stacked on top of each other and separated by 0.4 mm spacers. The gel set-up was then tapped at the bottom to ensure that no bubbles formed. The combs were subsequently added on top of the gel set-up.
3. The gel was allowed to set for at least 40 minutes, after which the combs were gently removed and the wells washed with water.
4. The gel plates were secured to the rig, which was filled with 2.0 L of 0.6X TBE.

2.8.2. Preparation of PCR products for SSCP analysis

1. An equal volume of isopropanol was added to each PCR product. The tubes were then inverted several times, then placed in ice for at least 30 minutes to precipitate the DNA.
2. The tubes were centrifuged for 15 minutes at 13,000 rpm, after which the supernatant was discarded. The pellets were dried at 45-55⁰ C for 15 minutes.
3. The pellets were resuspended in formamide-heavy loading buffer. [Loading buffer: 0.2 g bromophenol blue, 0.2 g CFF, 20 ml 0.5 M EDTA; one part loading buffer diluted in nine parts formamide]
4. The tubes were incubated at 95⁰ C for 10 minutes to denature the DNA into single strands, then the tubes were placed in ice to prevent gradual reannealing.

2.8.3. Loading of samples in the polyacrylamide gel

1. 15 µl of formamide-heavy loading buffer was loaded in each well. The gel was allowed to run at 180 volts for at least one hour to ensure that the gel was both consistent and equilibrated.
2. Once the gel was equilibrated, the samples were then loaded directly from ice into the wells.
3. The samples were allowed to migrate along the gel for 24 hours at 180 V.

2.8.4. Silver staining of SSCP gels

1. The following solutions were prepared:

Fix-stop solution

10% acetic acid	200 ml
Distilled water	1800 ml

Silver nitrate solution

Silver nitrate	2.0 g
Distilled water	1997 ml
Formaldehyde	3.0 ml

Developer

Sodium carbonate	60 ml
Distilled water	1997 ml
Formaldehyde	3.0 ml

2. After the run was complete, the gel was transferred into a staining tray containing 200 ml of fix-stop solution. Under the fume hood, the gel was agitated for 45 minutes to ensure complete fixation of the bands, after which the fix-stop solution was removed.
3. The gel was washed with distilled water three times for 20 minutes or until it attained a smooth appearance, indicating that it was already free of fix-stop solution. The excess water was removed.
4. 200 ml of silver nitrate solution was carefully added on the side of the tray and not directly onto the gel itself to prevent it from blackening. The gel was agitated for 40 minutes to give adequate time for the silver nitrate to bind with the DNA. The silver nitrate solution was removed afterwards into a suitable waste jar containing NaOH pellets to neutralize the solution.

5. The gel was washed with distilled water to remove excess silver nitrate, then the water was discarded into the silver nitrate waste jar.
6. 200 ml of developer was added and the gel agitated until bands were clearly visible.
7. The reaction was stopped by pouring 200 ml of fix-stop solution onto the gel, which was then agitated for an additional 5 minutes to release all the CO₂.
8. The gel was repeatedly washed with 200 ml distilled water until the smell of the acetic acid was no longer detected, after which the excess water was removed.
9. The gel was transferred onto a clean filter paper and covered with cling film. The gel was allowed to dry in a gel dryer for approximately 3 hours at 70 °C.

2.9. DNA sequence analyses

DNA sequences were assembled using the STADEN package version 1.5.3 (Staden, 2000) and aligned manually within the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994). The sequences were then checked for ambiguous nucleotide sites, saturation, and phylogenetic signal before being subjected to phylogenetic estimation. To this end, the optimal model that best fits the dataset was first identified, then a range of tree construction methods were employed.

2.9.1. Aligning sequences

Sequences were brought into the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994) where they were aligned manually. Positions were identified as unambiguously aligned when there was absolutely no question over their positional homology. All other positions for which homology could not be ascertained were excluded from subsequent analyses. For the nuclear rRNA cluster and the

mitochondrial 16S rRNA, the sequences were aligned according to their secondary structure. Initial secondary structure-based alignments for the nuclear rRNA and mitochondrial 16S rRNA for the Stylommatophora were obtained by C. Hudelot with all other sequences brought and aligned into these initial alignments. The initial alignment of the SSU for the Nematoda was provided by M. Blaxter. This alignment was updated to include all nematode SSU sequences available in GenBank (GB Release Numbers 157-158) as well as the new sequences obtained in this study.

2.9.2. Checking for contamination

Every precaution was taken to avoid contamination, which could have occurred either during DNA extraction, PCR amplification of the gene fragment of interest, or through cycle sequencing. These precautions included maintaining a clean work bench, regularly cleaning pipettes with 0.1 N HCl to destroy any contaminant DNA present, and using sterile pipette tips, scalpel blades and reagents. However, sequences obtained were still checked for contamination. A cursory inspection of the alignment was conducted to see if a sequence varied greatly or was more difficult to align than the rest. A BLAST check through NCBI was also carried out to verify if indeed the sequence originated from a contaminant (i.e. the sequence had a closest identity to a non-mollusc such as a human gene or a cross-contaminant with another mollusc). Sequences obtained in this study were also crosschecked against each other to determine whether sequences from different taxa (or even segments of sequences as in the case of the rRNA cluster) exhibited identical sequences; this might indicate cross contamination (though it should be noted that among closely related taxa, relatively conserved regions might reasonably be expected to be identical). In cases of contamination, the entire process from PCR amplification to cycle sequencing was repeated. Where the same

result was obtained, the process was again repeated, this time starting from the DNA extraction.

2.9.3. Assigning ambiguous sites

Sequence positions were scored as ambiguous either because the sequence could not be read reliably (due to poor quality sequence reads), or because there were several copies of the gene (as in the case of actin for which several copies were identified in some achatinoid species; see Chapters 3 and 4). For the actin gene, the identification of ambiguous sites (positions where different copies of the actin gene differed in sequence) was facilitated by using as guides the sequences obtained through cloning from representative taxa. For perfectly clear forward and reverse sequences, an ambiguous site was assigned to a position where two overlapping peaks were seen for that position in both strands. In cases where one strand was clear and the other strand had background noise, then the clear strand was used as the basis for deciding whether the site was ambiguous or not. If both strands exhibited background noise and yet both had the same clear overlapping peaks for a site, then that site was also assigned as ambiguous. Table 2.8 lists the codes for ambiguous nucleotides.

Table 2.8: Codes for ambiguous positions in DNA sequences (Cornish-Bowden, 1985)

Base ambiguity	Code
A or C	M
A or G	R
A or T	W
C or G	S
C or T	Y
G or T	K
A or C or G (not T)	V
A or C or T (not G)	H
A or G or T (not C)	D
C or G or T (not A)	B
A or C or G or T	N

2.9.4. Estimating the average base frequencies and the number of variable and parsimony-informative sites

Sequences for DNA very rarely exhibit identical frequencies for each base due to nucleotide bias. An extreme example of this is exhibited at the 3rd codon position of insects and crustaceans where 95% of the bases are either A or T (Palumbi, 1996). For all sequences under consideration, base frequencies were estimated using the basefreq command in the PAUP* (version 4.0b10) package (Swofford, 2002). The numbers of variable and parsimony-informative sites were calculated using the randtrees command in PAUP*.

2.9.5. Identifying the optimal model for DNA sequence evolution

Multiple ‘hits’, where pre-existing mutations are masked by more recent mutations that occur at the same site, will lead to an underestimate of the actual number of changes that have taken place at a particular site, thus obscuring the phylogenetic relationship of the taxa being compared. It is therefore necessary to apply a model of sequence evolution in order to ‘correct’ for such multiple ‘hits’ (Graur & Li, 2000). Models require certain assumptions as to how variations in DNA sequences evolve. All

possible assumptions for a given situation that are taken into account form a ‘conceptual model’ in which phylogenetic estimation is made. As more assumptions or parameters are incorporated in the model, the more complex it becomes. Several models can be used to account for DNA sequence evolution; these include the JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P (Kimura, 1980), HKY85 (Hasegawa, *et al.* 1985), TN93 (Tamura & Nei, 1993) and general time-reversible (GTR) models (Rodriguez *et al.*, 1990). The JC69 is the simplest model and assumes that all types of change (all substitutions) are equally likely, base frequencies are equal, all sites are equally likely to change and change independently of each other, and base composition is at equilibrium among all the sequences under consideration (Jukes & Cantor, 1969). The K2P is an extension of the JC69 model but allows transitions and transversions to have different substitution rates (Kimura, 1980). Likewise, the F81 model is an extension of the JC69 but allows for unequal base frequencies (Felsenstein, 1981). The HKY85 model allows for different rates of substitution for transitions and transversions as well as allowing for unequal base frequencies (Hasegawa *et al.*, 1985). The TN93 model is an extension of the HKY model but distinguishes between transition rates of purines and pyrimidines (Tamura & Nei, 1993). Finally, the GTR model allows all six pairs of substitution to have different substitution rates as well as allowing for unequal base frequencies (Rodriguez *et al.*, 1990). Rate heterogeneity between sites can also be accounted for by incorporating gamma distributed rates (Γ) into the models (Yang, 1993). Gu *et al.* (1995) proposed to take into account the proportion of invariant sites (I) in the gamma distributed rates, hence the ‘ Γ +I’ model. Yang (2006) describes this model as “pathological” as gamma distribution with an α shape parameter less than 1 already accounts for the invariant sites. Depending on the model under consideration, the base frequencies, rate matrix and shape parameter (α) of the gamma distribution using 16 rate

categories were estimated using likelihood by iteration from an initial neighbor-joining (NJ) tree. The parameters derived from the initial tree were then used to build a new neighbor-joining tree and the parameters re-estimated, repeating the process until no noticeable improvement is seen in the likelihood.

Models are generally selected based on their fit to the sequence data as measured by likelihood values (Kelchner & Thomas, 2007). Normally, addition of parameters in a model increases the likelihood score; this, however, increases complexity and thus the data are spread more thinly, so if there is no significant improvement in likelihood score then there is no justification for using the more complex model. One way to identify which model to use is through a likelihood ratio test (LRT). The LRT is a statistical test that determines the goodness of fit of any two models being compared with a particular dataset. This can be applied to models that are nested since twice the difference in the likelihood scores between two nested models is approximately Chi squared distributed. The formula for this test is given as: $LR = 2 * (\ln L1 - \ln L2)$; where $\ln L1 - \ln L2$ is the difference in the log likelihood scores between any two nested models being compared. The LRT can then be used to determine if there is a significant difference between the log likelihood scores of the two models by identifying the degrees of freedom and checking for the *P* value in a Chi square table. The number of degrees of freedom is the difference between the number of parameters used by the two models being compared. For example, the GTR and the GTR+ Γ models differ by one parameter (addition of the gamma distribution in the latter); therefore, the number of degrees of freedom for comparing these two models is 1 (Huelsenbeck & Crandall, 1997). Table 2.9 summarizes the number of parameters for a given model of DNA substitution. The model with the best likelihood score was selected but only if it was significantly better than a less complex model; otherwise, the simpler model was used.

Table 2.9: Summary of the number of parameters of the different models of DNA substitution (taken from Morrison, 2006)

Model	Number of Parameters
JC69	0
JC69+ Γ	1
K2P	1
K2P+ Γ	2
F81	3
F81+ Γ	4
HKY85	4
HKY85+ Γ	5
TN93	5
TN93+ Γ	6
GTR	8
GTR+ Γ	9

Twelve different models were evaluated; these were: (1) JC69, (2) JC69+ Γ , (3) K2P, (4) K2P+ Γ , (5) F81, (6) F81+ Γ , (7) HKY85, (8) HKY85+ Γ , (9) TN93, (10) TN93+ Γ , (11) GTR, (12), and GTR+ Γ . Since the F81 and K2P models are not nested, they could not be compared with each other. F81 and K2P could, however, be compared with any other model. Although it was tempting to restrict the model search to the parameter-rich HKY85, TN93 and GTR models as they were determined to be the optimal models by the ModelTest program in 80% of 208 published datasets in 2004 alone (Kelchner & Thomas, 2007), it was more prudent to check the less parameter-rich models to confidently rule them out if the more complex models had significantly higher likelihood scores. The likelihood scores for these models were computed in PAUP*, with the command lines summarized in Appendix 2.1, pp. 360-366.

The application of the LRT described here is similar to that which is applied in the Modeltest program by Posada and Crandall (1998) except that the LRT used in this study allows for a comprehensive comparison of all models under consideration (apart

from non-nested models) whereas Modeltest ‘traverses’ a model space through a series of pairwise comparisons of the different models. For instance, if Modeltest compares the likelihood scores of JC69 and F81 and found the latter to be significantly better, then F81 is selected and compared with HKY85. If HKY85 is better than F81, then HKY85 is selected and compared with GTR. If GTR is better than HKY, then GTR is compared with GTR+ Γ . Otherwise, HKY and TN93 will instead be compared. The problem with this approach is that it does not allow for a comprehensive comparison of all the different models being considered. In the above example, the GTR and TN93 models were not compared, and there is the possibility that TN93 is not significantly better than GTR.

2.9.6. Evaluating the sequences for evidence of saturation

Sequence data could suffer from extreme substitution saturation to such a degree that an optimal model of DNA sequence evolution could no longer correct for multiple hits (Xia *et al.*, 2003). The effect of this phenomenon of substitution saturation in phylogenetic analyses is twofold. First, it underestimates the amount of evolutionary change that took place. Second, homoplasy could occur where descendant sequences appear similar even if the similarity is not brought about by descent from a common ancestor (Graur & Li, 2000). For protein-coding genes, the third codon position has been found to be the most variable, with the 1st and 2nd codon positions more conserved. This is because substitution in either the 1st or 2nd codon position is more likely to lead to amino acid replacement that could affect the function of the protein being coded by the gene; hence, substitution tends to occur at a higher frequency at the 3rd codon position where substitution is mostly synonymous (Graur & Li, 2000). Including the 3rd codon position in phylogenetic analyses is beneficial when close relationships are

examined as it is the most variable position and therefore contains much of the phylogenetic information; removing it, on the other hand, would leave only a few variable sites in the 1st and 2nd codon positions to work with. In the presence of substitution saturation, however, using the 3rd codon position could be more disadvantageous since 3rd codon positions would saturate first (Xia *et al.*, 2003).

Constructing different types of plots based on substitutions have been the standard procedure to check for evidence of saturation in sequence datasets (Morisson, 2006; Xia *et al.*, 2009), and their use abounds in the literature. Three types of plots were constructed. Plotting pairwise uncorrected distances against pairwise corrected distances based on an optimal model (Plot 1), for instance, would normally result in a curve. When distances are small there would be little difference between the uncorrected and corrected distances. As more divergent sequences are compared, however, multiple hits would lead to an underestimate of the actual amount of change in the uncorrected distances relative to the corrected distances where a correction is made to account for these unseen changes. At some point, the sequences being compared would become too divergent to allow correction for multiple hits to be made reliably leading to substitution saturation; when this happens, a plateau is observed, and the actual amount of change taking place would be underestimated (Strimmer & von Haeseler, 2003). Morisson (2006) used this type of plot on the actin and HSP70 genes of *Cryptosporidium*. If both transitions and transversions are plotted against total distance (Plot 2), on the other hand, both would be expected to increase linearly. However, as more divergent sequences are compared, transitions (which generally occur more frequently than transversions, Brown *et al.*, 1982) would be expected to saturate prior to transversions, leading to the transition line curving and eventually reaching a plateau. Transversions (which generally occur less frequently than transitions, Brown *et*

al., 1982) would continue to increase linearly for longer but eventually as sequences become even more divergent even the transversion substitutions would saturate, eventually leading to a curved transversion line and ultimately a plateau. This type of plot was used by Jorgensen *et al.* (2008) on the 16S rRNA and COI genes of apple snails. Plotting transitions against transversions (Plot 3) would result in a linear relationship when sequences are unsaturated. As divergence increases and the transitions saturate, the relationship between transitions and transversions would be non-linear, resulting in a curve in the plot. Tsigenopolous *et al.* (2002) employed this type of plot on the cytochrome b of southern African barb. It was noted that interpretation of the plots could be somewhat subjective, particularly in determining when a plot has plateaued; it was imperative that all three types of plots were considered alongside one another in evaluating the level of saturation of each dataset.

The command lines in PAUP* for computing the uncorrected and corrected distances as well as the transition and transversion values are listed in Appendix 2.2, p. 367.

In addition to the plots mentioned above, there are other ways to detect the presence of full substitution saturation in a given dataset. One method is the randomization or permutation test in which a set of tree lengths are generated from randomized set of characters derived from the same dataset. If the dataset is saturated, then the tree lengths will not be significantly different from one another (Archie, 1989). Another method is the tree-independent relative apparent synapomorphy analysis (RASA) where synapomorphies or shared characteristics between any two given taxa are statistically evaluated (as expressed by the sum total of all shared character states between two taxa at the exclusion of third taxon) to determine if they result from shared evolutionary history or from convergence (Lyons-Weiler *et al.*, 1996). Both methods,

however, can incorrectly conclude that there is no saturation taking place if there are two very closely related taxa in the dataset even if the other taxa have reached full substitution saturation (Xia, 2009). A third method is the frequency-dependent significance test based on parsimony (see discussion on parsimony in Section 2.9.8, p. 84) in which the distribution of phylogenetically informative sites is statistically tested against the hypothesis that the sequence variation is random, as in the case of full substitution variation (Steel *et al.*, 1993, 1995). This method, however, is affected by long branch attraction, a typical problem of parsimony, where highly divergent taxa tend to group together by chance not because of shared homology but precisely because of homoplasy (Xia, 2009). A fourth method makes use of the index of saturation based on the concept of entropy in information theory where entropy is defined as the measure of uncertainty pertaining to a random variable, in this case the frequency of a nucleotide for a nucleotide site (Xia & Xie, 2001, Xia *et al.*, 2003). The index of substitution saturation (Iss) is the ratio of the observed mean entropy of all the sites in a gene of specific length over the expected entropy of the sequence at full saturation. Once the Iss has reached a critical value (or is not statistically significantly different from this critical value) in which the sequences could no longer be expected to recover the true tree, then the dataset is said to be saturated. However, applying this method becomes problematic if more than 32 taxa are used (Xia, 2009). It is also limited to determining if saturation has taken place for a perfectly symmetrical and completely asymmetrical tree but not anything in between. For all the reasons mentioned, these four methods were therefore not used for testing for evidence of saturation.

2.9.7. Testing for phylogenetic signal

If the tree lengths of all tree topologies generated through maximum parsimony (discussed in Section 2.9.8, p. 84-85) follow a normal distribution, then the differences in tree lengths are due to chance rather than homology; otherwise, the distribution is skewed enough that very few alternative solutions exist near the shortest tree, thus indicating the presence of a phylogenetic signal. A g_1 test can measure how skewed a dataset is when applied to a set of tree lengths based on parsimony. Critical values for the g_1 measure of skewness for such datasets (Table 2.10) were derived by Hillis and Huelsenbeck (1992). If g_1 test scores obtained are smaller (or more negative) than the critical values, then the tree length distributions are said to be skewed and therefore exhibit phylogenetic signal. It should be noted that the critical values change little beyond 15 taxa being investigated so the values derived for 25 taxa can be used to test datasets with more than 25 taxa (Hillis & Huelsenbeck, 1992). Where the number of parsimony-informative sites or the number of taxa being investigated is not available in the table, then the next lower number should be used as a basis for deriving the critical value. Increasing the number of parsimony-informative sites or the number of taxa will increase the critical value (make it less negative), so if a g_1 score for a particular number of parsimony-informative sites and taxa is less than the critical value for a lower number of parsimony-informative sites or lower number of taxa, then there is phylogenetic signal for the dataset.

Table 2.10: Critical values for g_1 measure of skewness at $P=0.05$.

No. of parsimony-informative sites	No. of Taxa							
	5	6	7	8	9	10	15	25
10	-0.95	-0.70	-0.59	-0.51	-0.44	-0.34	-0.23	-0.16
50	-0.78	-0.58	-0.45	-0.37	-0.25	-0.28	-0.19	-0.12
100	-0.66	-0.56	-0.40	-0.31	-0.25	-0.30	-0.20	-0.10
250	-0.81	-0.43	-0.39	-0.26	-0.22	-0.20	-0.16	-0.08

Caution was taken, however, when interpreting strong skewness as this could be misleading. The presence of duplicated taxa or even very closely related taxa (i.e. those with very similar sequences) in a group of random taxa could lead to a very strong left skew as the tree that would show monophyly of these taxa would be considerably shorter than the other trees generated (Swofford *et al.*, 1996).

The g1 test statistic was performed in PAUP* and applied to all the gene datasets using the randtrees command with 10,000 replicates in PAUP*.

2.9.8. Tree Construction

Phylogenetic trees are graphical representations of the evolutionary relationships among taxa derived from morphological or molecular datasets. For molecular datasets, tree-construction methods either convert aligned sequences into a matrix of pairwise distances among the taxa (distance methods), or they use the positions in the sequences directly (character-state methods). Tree construction methods can use an algorithm to search for the single best tree estimate (algorithmic methods) or they define a criterion by which several alternative trees are compared and scored in order to identify the best tree (optimality criterion methods) (Graur & Li, 2000).

The maximum likelihood (ML) method (Cavalli-Sforza & Edwards, 1967; Felsenstein, 1981) is a character-state method that finds the best tree with the highest probability of fit with the data (measured as the likelihood score) using a specific model of DNA sequence evolution (Graur & Li, 2000). The objective of maximum likelihood is to find the tree with the best or largest likelihood score. However, phylogenetic analyses involving many taxa would entail evaluating a large number of trees (e.g. there are more than 34 million possible rooted trees for only 10 taxa) so that conducting an exhaustive search is impractical. Instead, a heuristic search is done in which an initial

start tree (obtained by stepwise addition, with a random sequence input order and 10 replicates) is successively improved by swapping branches until the most likely tree with the best likelihood score is obtained. Unlike an exhaustive search, a heuristic search only evaluates a subset of all the possible trees and provides the best guess of the optimal tree (Hall, 2008). ML was undertaken in PHYML Version 2.4.4 (Guindon *et al.*, 2005). The command lines in PHYML are listed in Appendix 2.3.1, pp. 368-369.

Bayesian inference (BI) is similar to maximum likelihood in that it also utilises likelihood values. Unlike maximum likelihood, Bayesian inference determines the best hypothesis that maximises the posterior probability, which is proportional to the likelihood value multiplied by the prior probability of the hypothesis. A prior probability is based on what is initially known about the dataset (Holder & Lewis, 2004). Computer programs that conduct Bayesian inference such as MrBayes make use of the Markov Chain Monte Carlo algorithm to approximate posterior probability values of randomly generated trees through a number of generations until the chain attains a stable likelihood value (Ronquist & Huelsenbeck, 2003; Hall, 2008). Four chains of a Markov Chain Monte Carlo algorithm in BI were used to compute the posterior probability values with an optimized number of generations and heating temperature (Temp). The number of generations was optimized when the values of the posterior probabilities across the four chains converged or did not vary significantly from each other. The heating temperature was optimized when the acceptance rates of the Metropolis-Hastings mcmc sampler ranged between 0.1 and 0.7. (See also Appendix 2.3.2, pp. 369-370.) Bayesian inference was conducted in MrBayes Version 3.1.2 package (Ronquist & Huelsenbeck, 2003), and the command lines for this package are listed in Appendix 2.3.2., pp. 369-370.

The neighbor-joining (NJ) method (Saitou & Nei, 1987) is an algorithmic distance method that uses a modified distance matrix that adjusts the separation of each pair of sequences based on the pair's average divergence from all the other sequences. This method identifies the shortest tree by sequentially finding neighbors that minimize the total length of the tree (Graur & Li, 2000). NJ trees were built in PAUP*, with the command lines summarized in Appendix 2.3.3, p. 371.

The maximum parsimony (MP) method is a non-model based method that identifies a topology that requires the least number of possible changes (Eck & Dayhoff, 1966; Fitch, 1977). As with ML, a heuristic search for the most parsimonious tree or a set of equally most parsimonious trees is conducted by starting with an initial tree (obtained by stepwise addition, with a random sequence input order and 10 replicates) and swapping branches until the optimal tree is obtained. MP analysis was undertaken in PAUP* with a heuristic search using the tree bisection and reconnection (TBR) swapping routine and 10 random input orders. The command lines are listed in Appendix 2.3.4, pp. 371-372.

Advantages and disadvantages of the four tree methods are summarized in Table 2.11.

Table 2.11: Comparison of methods (table taken from Holder & Lewis, 2004)

Tree method	Advantages	Disadvantages
Neighbor-joining	Fast	Information is lost in compressing sequences into distances; reliable estimates of pairwise distances can be hard to obtain for divergent sequences
Maximum likelihood	The likelihood fully captures what the data tell us about the phylogeny under a given model	Can be prohibitively slow
Bayesian inference	Has a strong connection to the maximum likelihood method; might be a faster way to assess support for trees rather than the	The prior distributions must be specified; it can be difficult to determine whether the Markov chain Monte Carlo (MCMC)

	maximum likelihood bootstrapping	approximation has run for long enough
Maximum parsimony	Fast enough for the analysis of hundreds of sequences; robust if branches are short (closely related sequences or dense sampling)	Can perform poorly if there is substantial variation in branch length

2.9.9. Estimating reliabilities of clades in phylogenetic trees

A phylogenetic tree represents the best estimate of the true tree based on the sequences available and the method of tree construction. Reliability of the tree estimates, which can be measured by how many times the members of a particular clade are recovered as members of that clade, can be done through the bootstrap method in the maximum likelihood, neighbor-joining and maximum parsimony methods and through measuring posterior probabilities in the Bayesian inference method (Hall, 2008). The bootstrap method is a resampling technique used to estimate the confidence level of hypotheses in phylogenetic estimation. A measure of support for the branches in the tree is provided by bootstrap values, which are based on the percentage of the number of times a particular branch is generated in the bootstrap datasets (generated by random sampling from the dataset with replacement) (Graur & Li, 2000). For instance, a particular dataset is resampled 1000 times with replacement to produce 1000 resampled datasets, with each dataset used to generate a tree. Out of the 1000 trees, the percentage of the trees in which a particular branch appears becomes its measure of support. Bootstrap resampling with 1000 replicates was undertaken in PHYML for ML (see Appendix 2.3.1, pp. 368-369) and in PAUP* for NJ and MP (see Appendix 2.3.3, p. 371 for NJ and 2.3.4, pp. 371-372 for MP). In Bayesian inference analysis, the fraction of times a clade occurred among the last 1000 trees surveyed as part of the MCMC process was directly computed as posterior probabilities. Unlike the bootstrap method, the

underlying data are not resampled (Hall, 2008). The posterior probabilities were computed in MrBayes (see Appendix 2.3.2, pp. 369-370).

2.9.10. Combining datasets

As well as undertaking phylogenetic analyses for each gene independently, where possible the individual gene datasets were combined in order to get a better estimate of the phylogeny. It has been recognized that a phylogeny based only on a single gene, known as a gene tree, cannot truly represent a species tree (Cummings *et al.*, 1995; Graur & Li, 2000) as organisms that participate in the evolutionary process are a collection of genes, each with its own evolutionary history, that interact with each other and with the environment (Morrison, 2006). Combining data from different genes tends to increase phylogenetic signal and disperse noise while at the same time uncovering the underlying signal found in the different data partitions (Baldauf *et al.*, 2000). Using genes in concert that have different evolutionary rates and modes of inheritance (i.e. the relatively slower evolving nuclear genes and the faster evolving, maternally inherited mitochondrial genes) can also provide information at different levels of phylogeny (Graybeal, 1994). Combining datasets are also known to reduce the phenomenon of long-branch attraction in parsimony analysis (Gontcharov *et al.*, 2004) in which long branches in a tree tend to cluster together (Graur & Li, 2000).

Concatenation is the combination of all datasets into a single data matrix for the purpose of phylogenetic analysis (Morrison, 2006). However, simply concatenating the datasets under consideration and analyzing them using only one evolutionary model could produce a phylogeny with the wrong groupings if the different datasets represent very divergent evolutionary histories (Cunningham, 1997). The partition homogeneity test (also known as the incongruence length test) based on maximum parsimony in

PAUP* was used in this study to distinguish between separate datasets that could be combined to give an improved phylogenetic accuracy (with P normally greater than 0.01) and individual datasets that give poorer accuracy when combined (with $P < 0.001$) due to their very divergent evolutionary histories (Cunningham, 1997). The command lines for the partition homogeneity test in PAUP* are shown in Appendix 2.4, p. 373. As the test makes use of maximum parsimony to determine if datasets can be combined, its utility is hampered if at least one of the genes being considered exhibit polytomies as this considerably slows down PAUP*. For such a scenario, the decision to combine would rely on whether or not the single gene phylogenies exhibited concordance with each other.

Two sets of concatenated sequence alignments were prepared for the combined gene analyses. The first included only taxa with complete sequences across all genes evaluated. In order not to lose any taxon, a second set of concatenated sequences was also prepared in which taxa with missing datasets were also included. Though including taxa with incomplete datasets could be potentially problematic in phylogenetic analyses, Wiens (2006) found that these taxa could still be included and their phylogenetic position accurately determined so long as enough characters have been surveyed; their inclusion could also remove misleading long branches and potentially increase the accuracy of the combined phylogeny. The trees generated from the second set of concatenated sequences were compared for any similarity with the individual gene phylogenies as well as those derived from the first set of concatenated sequences.

An alternative to combining datasets by concatenating sequences is to construct from multiple trees a consensus tree derived from several genes with the same set of taxa or a supertree if the taxa sampled are not identical but there are taxa overlapping

between trees (Morrison, 2006). Two strategies employed to construct consensus trees and supertrees are strict consensus (tree generated contains all the groups that occur on all trees considered) and majority consensus (tree generated includes groups that occur in at least 50% of all the trees considered) (Felsenstein, 2004). However, the effectiveness of consensus methods is limited by the following: 1) their tendency to produce spurious groups that do not appear in any of the trees; 2) the non-avoidance of pseudoreplication as a result of using overlapping data in some of the trees that increases the weight of these data; 3) the inappropriateness of using branch support methods such as bootstrapping (Morrison, 2006); and 4) their treatment of each tree, no matter how bad, as equally good (Wade, pers. comm.). The consensus methods were therefore not used.

2.9.11. Hypothesis testing

An optimal tree may not necessarily produce the expected topologies, as in the case where a supposedly monophyletic group based on taxonomy turned out not to be monophyletic in the optimal tree. A tree can be constrained to reflect the expected topology, and this tree can be compared for significant difference with the optimal tree. If the optimal tree turns out to be not significantly better than the constrained tree, then the hypothesis being put forward by the optimal tree is not a strong hypothesis. The Shimodaira-Hasegawa test (Shimodaira & Hasegawa, 1999; Goldman *et al.*, 2000) was applied on the datasets found in Chapters 3 and 4 to compare for significant difference between constrained and optimal NJ trees using likelihood. The command lines for the Shimodaira-Hasegawa test, which was carried out in PAUP*, are listed in Appendix 2.5, p. 374.

2.9.12. Inferring the presence of paralogous sequences in the dataset

The presence of paralogous sequences due to gene duplication could yield misleading trees. All genes were therefore evaluated for the presence of paralogs. Of the genes used in this study, actin is most susceptible to the problem of paralogy since several copies of the actin gene, which arose from gene duplication events, abound. Although the primers used to amplify the actin gene were known to be specific for the cytoplasmic actin (see Section 2.2.1.1.1.2, pp. 48 – 50), this was no guarantee that only orthologous sequences were amplified as duplication events could also have taken place within the cytoplasmic actin. To infer the presence of paralogous sequences, the PCR-direct sequences were checked for the presence of ambiguous sites; the presence of ambiguous sites (where more than one peak is obtained at the same position in a DNA sequence) implies that more than one copy of the gene is present in that individual. All

sequences obtained in this study were evaluated for the presence of ambiguities. Where evidence of multiple copies is found, cloning the PCR products prior to sequencing enables each copy of the gene to be examined individually. Cloning of the actin gene (Chapters 3 and 4) was undertaken for a limited number of clones from three taxa (time and cost constraints precluded a more detailed analysis).

2.9.13. Assessing the utility of the actin gene for phylogenetic analyses

The inherent problem of actin in relation to the presence of paralogous sequences could limit its utility in phylogenetic analyses. However, the potential of actin to provide some phylogenetic signal should not be underestimated. Several steps were undertaken to evaluate the utility of the actin dataset. First, the clones derived from representative taxa were checked for monophyly; if all clones derived from a taxon fall as a monophyletic group within the actin tree, then they will not mislead organismal phylogeny. Second, the number of ambiguous sites within a taxon was compared with the total number of nucleotide differences between the taxon and its closest relative based on the closest sequence identity of the actin sequence (i.e. the least number of nucleotide differences between any two taxa). In principle, the time of divergence between two taxa should be earlier than the evolution of the different actin genes within these taxa, such that the number of ambiguous sites should not exceed the number of differences between the two taxa being compared. However, it is possible for two taxa to have a higher number of ambiguous sites than nucleotide differences if these ambiguous sites are due to a large number of multiple copies, in which each contains only a small number of variable sites (when this variation is added together it may exceed the number of nucleotide differences). These sequences will still cluster together in a tree and will not mislead phylogeny. Third, a partition homogeneity test (see

Section 2.9.10, pp. 86-88) was undertaken to determine whether or not the sequences from the actin gene exhibited a significant difference in terms of phylogenetic signal with the other genes being evaluated. If no significant difference was observed, this would suggest that the actin gene could be utilized alongside the other genes in phylogenetic analyses and that the presence of multiple copies of the actin gene was not having a significant effect on phylogenetic estimation. Lastly, the phylogenetic tree obtained from the actin dataset was checked for concordance with the other gene phylogenies. Concordance would suggest that the presence of multiple copies of the actin gene was not misleading phylogeny. Phylogenetic analyses involving the actin gene were therefore approached with caution, with reference to the limitations of using this gene.

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CHAPTER 3 – Phylogeny of the Achatinoidea

3.1. Introduction

The Giant African Land Snail, *Achatina fulica*, and other members of the superfamily Achatinoidea belong to the suborder Stylommatophora, whose members possess a vascularised mantle cavity that serves as a lung and two pairs of caudal tentacles, the upper pair of which bears the eyes at their tips (Ruppert *et al.*, 2004; Barker, 2001). Using the Pilsbry-Baker system of classification, the Stylommatophora can be divided into four infraorders based on the structure of the excretory system (Pilsbry, 1900; Baker, 1955). *Achatina fulica* and other members of the Achatinoidea belong to the infraorder Sigmurethra, in which the excretory system does not have a distal ureteric pouch or orthureter but possesses a ureter that runs along the anterior portion of the nephridium and connects to the pallial cavity before terminating at the pneumostome. Recent molecular phylogenetic studies (Wade *et al.*, 2001, 2006) based on the nuclear ribosomal (r) RNA gene cluster (including part of the 5.8S and LSU genes) revealed a principal division of the Stylommatophora into two major clades. The ‘achatinoid’ clade comprises the superfamilies Streptaxoidea, represented by the single family Streptaxidae, and the Achatinoidea, represented by the families Achatinidae, Coelioxidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophorellidae. All other stylommatophoran families fall within the ‘non-achatinoid’ clade.

3.1.1. The Achatinoidea

The composition of the Achatinoidea varies according to author (Solem, 1978; Abbott, 1989; Tillier, 1989; Vaught, 1989; Schileyko, 1999; Bouchet & Rocroi, 2005; Table 1.1). Solem (1978) included the Achatinidae, Ferussaciidae, Megaspiridae,

Spiraxidae and Subulinidae (including the coeliaxids and glessulids as the subfamilies Coeliaxinae and Glessulinae, respectively), while Abbott (1989) regarded the Coeliaxidae as a family in its own right and additionally included the monotypic Thyrophorellidae. Tillier (1989) classified land snails under the superfamily Achatinoidea based on certain morphological features such as a closed ureter in the excretory system and possession of symmetrical cerebro-pedal connectives, short commissure and contiguous left parietal and visceral ganglia in the nervous system. His Achatinoidea comprised the Achatinidae, Ferussaciidae, Subulinidae (including the subfamilies Coeliaxinae and Glessulinae), Thyrophorellidae as well as the Streptaxidae, Succineidae and the Oleacinidae (Spiraxidae and Testacellidae). Vaught (1989) did not include the Streptaxidae, Succineidae, Spiraxidae or Testacellidae in her Achatinoidea but instead included the Thyrophorellidae and the Coeliaxidae (elevated from a subfamily of subulinids) in addition to the Achatinidae, Ferussaciidae and Subulinidae. Schileyko (1999) placed only the Achatinidae in the Achatinoidea, whereas Bouchet & Rocroi (2005) included the Achatinidae, Ferussaciidae, Subulinidae (which also incorporated the coeliaxids and glessulids as the subfamilies Coeliaxinae and Glessulinae) and Micractaeonidae. Wade *et al.*'s (2001, 2006) molecular phylogenies based on a part of the rRNA gene cluster lend support to Vaught's Achatinoidea. In this study, I utilise an Achatinoidea that largely resembles Vaught's and comprising the Achatinidae, Coeliaxidae, Ferussaciidae, Glessulidae (a subfamily of the Subulinidae in Vaught's classification), Subulinidae and Thyrophorellidae.

The family Achatinidae, which includes the Giant African Land Snail, *Achatina fulica*, has shells that are mostly dextral, are generally higher than wide and range in size from medium to very large (Schileyko, 1999). The reproductive structures of the achatinids are highly variable (Mead, 1991), but all members possess a penis sheath

(Schileyko, 1999). With the exception of *Achatina fulica*, which has been introduced by Man into new areas (Mead, 1979; Raut & Barker, 2002), all other achatinids are restricted to Africa and nearby islands (Schileyko, 1999).

The monotypic family Thyrophorellidae is represented by *Thyrophorella thomensis* from São Thomé. The shell is sinistral and is made up of three nearly flat whorls so that it appears low conic on top and bulging at the bottom (Schileyko, 2001). *Thyrophorella thomensis* is one of the few stylommatophorans that have secondarily acquired a structure similar in function to the prosobranch gastropod operculum; in the case of *T. thomensis*, a lobe-shaped outgrowth of the parietal wall of the shell's aperture is hinged on a flexible periostracal layer to serve as a covering of the aperture (Barker, 2001). Bouchet and Rocroi (2005) included the Thyrophorellidae in the superfamily Punctoidea, although recent molecular data from the LSU rRNA gene (Wade *et al.*, 2006) clearly places this family within the Achatinoidea.

Members of the Ferussaciidae have very small dextral shells that are elongate-ovate to nearly cylindrical and appear translucent or transparent (Schileyko, 1999). Ferussaciids are unique among the 'achatinooids' because they superficially resemble the 'non-achatinooid' Succineidae in possessing a short but transversely elongate kidney (Tillier, 1989) with a complete ureter (Schileyko, 1999). The ferussaciids are largely found in the Mediterranean, with several in tropical Africa, the Indian subcontinent and Indian Ocean islands, the Philippines, Hawaii, Mexico, the Caribbean, Brazil (Schileyko, 1999), Hawaii (introduced, Cowie, pers. comm.), New Caledonia (introduced, Schileyko, 1999) and Northwest Europe (Kerney & Cameron, 1979).

The family Subulinidae is a large group with a pantropical distribution (Tillier, 1989). With very few exceptions, its members have characteristic slender and turreted shells and are generally dextral. The internal anatomy of members of the group is not

well studied, and most taxa within the family are grouped together based largely on the similarity of their shells and their geographic distribution. Such an approach is exemplified by Zilch (1959). In revising the Subulinidae, Schileyko (1999) attempted to restructure their classification by including characters of the reproductive tract, primarily those proximal to the genital orifice. As the only comprehensive review of the group since Pilsbry (1906-1907; 1908-1910), it seems logical to use Schileyko (1999) as the primary source when reviewing the group as was done by Bouchet & Rocroi (2005). According to Schileyko (1999), eight subfamily groups can be recognized. These are the Subulininae (dextral shell with the last whorl lacking internal sculpture and with a reproductive tract that has a reduced or absent epiphallus, a muscular organ responsible for the formation of the spermatophores, and a short or absent flagellum, an extension of the epiphallus; distributed in tropical and subtropical regions of the world), Petriolinae (dextral or sinistral shell similar to Subulininae but with a prominent epiphallus; found in Africa and St. Helena), Rishetiinae (dextral shell with a simple columella and a reproductive organ with a long flagellum; found in the Hindustan Peninsula, Ceylon, and Southeast Asia), Rumininae (dextral shell decollated or entire and with a reproductive organ lacking an epiphallus and a flagellum; found in South Africa and the Mediterranean), Opeatinae (with dextral shell and an enlarged penis; found in subtropical regions of the Old and New World), Obeliscinae (dextral shell slender turreted to subfusiform and with a reproductive organ lacking an epiphallus and a flagellum; found in South America, the Caribbean and Southeast Asia), Tristaniinae (sinistral shell; found in Tristan da Cunha Islands), and Perrieriinae (shell sinistral and subcylindrical or pillar-shaped; found in New Guinea) (Schileyko, 1999). Other subulinid groups have been accorded by some authors with full taxonomic family status such as the Coeliaxidae (Abbott, 1989; Vaught, 1989) and the Glessulidae (Schileyko,

1999). The coelioxids have slender dextral or sinistral shells and are distributed in South Africa (only one species), the Gulf Island of São Thomé in West Africa, the Caribbean, Colombia and Peru (Schileyko, 1999). The glessulids generally have glossy, oblong-conic dextral shells and are restricted to the Indian subcontinent and Sri Lanka (Schileyko, 1999).

However, closer examination of Schileyko's results raises doubts regarding his conclusions, and the taxonomy of the Subulinidae therefore warrants further scrutiny. For instance, rather than representing a subulinid subfamily, *Tristania* is a junior synonym of *Balea*, which is a member of the Clausiliidae (Preece & Gittenberger, 2003). *Rishetia* is anatomically close to *Glessula* (both have a shell with truncated columella and penis with epiphallus), and both are restricted to South Asia; therefore, there seems to be no basis for Schileyko's erection of the subfamily Rishetiinae. Shileyko's acceptance of Thiele's (1933) genus *Striosubulina* is supposedly justified by the reproductive anatomy described and figured by Shileyko. However, in describing the penis, Shileyko seems to have mistaken that part of the penis surrounded by the penial sheath for the complete penis, and most of what he describes as the penial retractor muscle is in fact the long thin penis almost identical to that of *Subulina octona*, the type species of *Subulina* (Naggs, pers. comm.). Thus, the arrangement of the Subulinidae into eight subfamilies by Shileyko (1999), subsequently accepted by Bouchet & Rocroi (2005), is unreliable and will need to be reassessed. Furthermore, some pieces of information on distributions of the subulinids based on published records were not included by Shileyko (1999). For example, in giving the range of *Glessula* as 'Hindustan peninsula and Ceylon (1999, page 541), Shileyko fails to mention that several species of *Glessula* occur in Myanmar, Yunnan (China), Thailand and east as far as Vietnam and southeast into the Indonesian peninsula (Pilsbry, 1908-

1910; Van Benthem Jutting, 1952, 1959; Hemmen & Hemmen, 2001; Vermeulen & Maassen, 2003). With the Rumininae, the geographical range for the group alters drastically if *Zootecus* is excluded (see Table 3.1) or included, when the range extends from circum-Mediterranean, Arabian and West African to encompass a large longitudinal range from the Cape Verde islands through Central and Northern India to Myanmar (Gude, 1914). It should be noted that Schileyko (1999) placed *Zootecus* under the Subulininae.

Table 3.1: Comparison of the composition and distribution of the Rumininae according to Zilch (1959) and Schileyko (1999).

	Rumininae sensu Zilch (1959)	Rumininae sensu Shileyko (1999)	Distribution according to Shileyko	Distribution according to Zilch
<i>Namibiella</i>	x	X	SW Africa	SW Africa
<i>Xerocerastus</i>	x	X	SW Africa	SW Africa
<i>Lubricetta</i>	x	X	SW Africa	SW Africa
<i>Krapfiella</i>		X	E. Africa	
<i>Riebeckia</i>		X	Sokotra	
<i>Balfouria</i>		X	Sokotra	
<i>Rumina</i>	x	X	Mediterranean	Europe and N. Africa
<i>Obeliscella</i>	x			Arabia
<i>Zootecus</i>	x			Cape Verde Islands, N Africa, N & C. India

3.1.2. Some questions about the Achatinoidea

Although the Achatinoidea is strongly supported in Wade *et al's* (2006) rRNA phylogeny with 93% NJ bootstraps (see Figure 3.1), their phylogeny was based on only 823 nucleotide sites from the rRNA gene cluster and incorporated only 15 achatinoid taxa. Clearly, the clade could be further refined by obtaining new sequence data (both

longer sequences of the rRNA as well as sequences from other new genes) as well as the inclusion of additional taxa.

For instance, the achatinids are a compact group based on the presence of a penis sheath (Schileyko, 1999) and their restricted geographic distribution to sub-Saharan Africa (Mead, 1979; Raut & Barker, 2002). Wade *et al.* (2006) used four taxa to represent the Achatinidae, but support for the monophyly of the group was equivocal (38% NJ bootstraps and $P=0.94$ BI) based on the small segment of the rRNA gene cluster used in their analyses.

The ferussaciid *Ferussacia folliculus* forms a distinct basal lineage in the Achatinoidea (Wade *et al.*, 2006). Whether this topology persists if other ferussaciids are surveyed remains to be seen.

The monophyly of the Subulinidae and its subfamilies also needs to be verified as, anatomically, this group is poorly studied (Schileyko, 1999), and both the coelioxids (Abbott, 1989; Vaught, 1989) and the glessulids (Abbott, 1989; Schileyko, 1999) have been regarded as separate families. In particular, the placement of *Zootecus* in the Subulininae according to Schileyko (1999) must be validated. The subulinids, coelioxids and glessulids were represented by nine taxa in the Wade *et al.* (2006) study, which formed several groups together with the thyrophorellid *Thyrophorella thomensis* but with no support apart from the *Riebeckia-Coeliaxis* group (92% NJ bootstraps and $P=1.0$ BI) and the *Pyrgina-Thyrophorella* group (100% NJ bootstraps and $P=1.0$ BI).

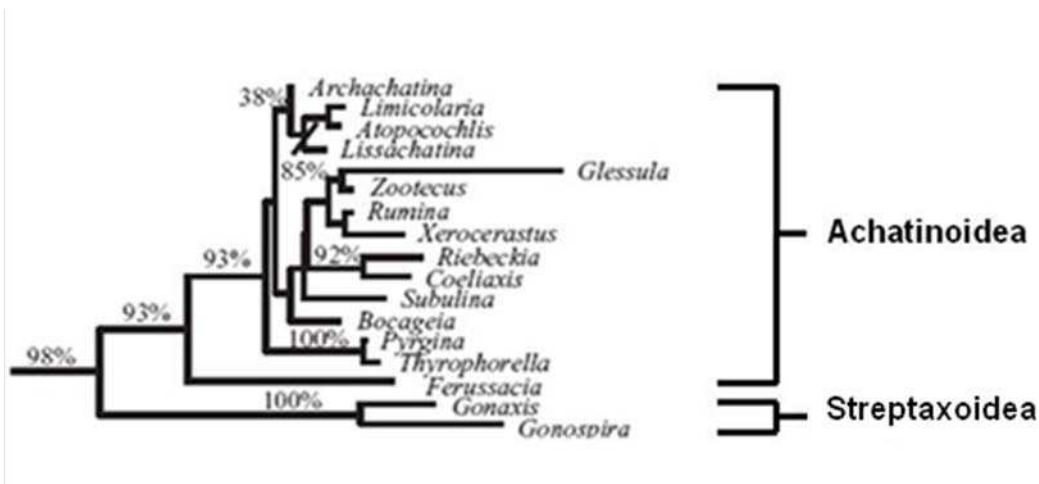


Figure 3.1: Phylogeny of the Achatinoidea based on 823 unambiguously aligned nucleotide sites of the rRNA gene cluster. (Taken from the stylommatophoran phylogeny of Wade *et al.*, 2006.)

3.1.3. Use of molecular markers to determine phylogeny

Various molecular markers are available for determining evolutionary relationships. These include the nuclear ribosomal RNA gene family (see Section 2.2.1.1.1.1 of Chapter 2, pp. 45-48). Already described is Wade *et al.*'s (2001, 2006) use of the nuclear ribosomal RNA gene cluster to infer phylogenetic relationships within the Stylommatophora. However, their results were derived from only a partial segment of the rRNA cluster. Wade *et al.* (2001, 2006) amplified an approximately 1460 nucleotide region of the rRNA (comprising approximately 80 nucleotides of the 5.8S gene, the complete internal transcribed spacer (ITS) 2 region and approximately 840 nucleotides of the LSU gene) of which 843 (2001 study) and 823 (2006 study) nucleotide sites could be aligned across all taxa and were therefore used in phylogeny reconstruction. Expanding the Wade *et al.* fragment to cover the entire LSU gene (approximately 4000 nucleotides) could provide better resolution of the Achatinoidea. The small subunit (SSU) rRNA gene has also been employed to elucidate evolutionary relationships among different gastropod groups. For example, Winnepeninckx *et al.*

(1998) applied the full-length SSU rRNA gene (approximately 1800 nucleotides), to reassess the groupings within the Gastropoda, demonstrating the basal position of the archaeogastropods. However, their results could not resolve whether the pulmonates or even the stylommatophorans were monophyletic.

In addition to the ribosomal genes, the nuclear actin and histone 3 genes have also been used to estimate phylogeny within the Mollusca. The actin gene encodes a protein that is involved in various functions such as muscle contraction, cell division and differentiation (Hernan, 1993, see also Section 2.2.1.1.1.2 of Chapter 2, pp. 48-50). However, several actin genes may exist in animals as a result of gene duplication and divergence (Hightower & Meagher, 1986); these genes code for protein isoforms that are broadly classified as either the muscle (α) type or the cytoplasmic (β) type (Vandekerckhove & Weber, 1984; Adema, 2002). Great care must therefore be taken to ensure that paralogous copies of the gene do not mislead the phylogenetic analysis. A 784 bp fragment of the actin gene has been successfully employed to show the monophyly of several groups within the coleoid cephalopods such as the Octopodiformes, the Decapodiformes, the Octopoda and the Incirrata (Carlini *et al.*, 2000). The actin gene was also used together with the mitochondrial 16S and cytochrome c oxidase subunit I (COI) genes to resolve the phylogeny within the ancestral archaeogastropod monodontine topshells (family Trochidae) from the southern hemisphere, with three species of *Austrocochlea* being transferred to the genus *Chlorodiloma* (Donald *et al.*, 2005). The actin fragment (764 bp) used in this study was amplified using primers designed for the cytoplasmic actin fragment of the planorbid snail *Biomphalaria glabrata* previously sequenced by Adema (2002). To date, the cytoplasmic type actin gene has not been used in the Achatinoidea, and the utility of this

gene for phylogenetic analysis of the group is worth investigating, though the potential presence of paralogous sequences could limit its utility.

The histone 3 (H3) gene codes for two subunits of the histone core protein octamer that packs the eukaryotic chromatin into bead-like structures (Lewin, 2008) (see Section 2.2.1.1.1.3 of Chapter 2, pp. 50-51). Histone 3 was used, in conjunction with the nuclear SSU and LSU rRNA genes and the mitochondrial COI gene, to resolve the incongruence between molecular and morphological data for the gastropod phylogeny (Colgan *et al.*, 2003). The results demonstrated the monophyly of the Euthyneura, which include the stylommatophoran pulmonates. The H3 fragment (331 bp) used in this study corresponds to the 3' end of the gene in the bivalve *Spisula solidissima*. Like the actin gene, the histone 3 gene has not been used in the Achatinoidea; thus, its usefulness is worth investigating.

Using mitochondrial genes in concert with nuclear genes is desirable for constructing phylogenetic trees as they tend to improve phylogenetic accuracy (Lake & Moore, 1998; Steinke *et al.*, 2004). Nuclear and mitochondrial genes evolve at different rates and are not inherited in the same way; as such, they provide information at different levels of phylogeny (Graybeal, 1994). Mitochondrial genes generally evolve faster than nuclear genes; they are also maternally inherited and are therefore not subject to recombination (Brown, 1985; Avise, 1994). Two mitochondrial genes commonly used for phylogenetic analyses are the cytochrome c oxidase subunit I (COI) gene, which codes for an enzyme that accepts electrons from cytochrome c during the electron transport chain in the mitochondrion (Zubay *et al.*, 1995; see also Section 2.2.1.1.2.1 of Chapter 2, pp. 51-52), and the 16S rRNA gene, which transcribes a ribosomal rRNA that is incorporated in the mitochondrial ribosome (Lewin, 2008; see also Section 2.2.1.1.2.2 of Chapter 2, pp. 52-53). Steinke *et al.* (2004) used a combined

data set of nuclear rRNA genes (18S and ITS-1) and mitochondrial genes (16S rRNA and COI) to separate the Western Palaearctic helicoid stylommatophorans into two families: Helicidae and Hygromiidae. They reasoned that the faster evolving mitochondrial genes were able to resolve the terminal taxa while the more conserved nuclear rRNA genes separated the basal groups. The 16S rRNA fragment (approximately 400 bp) was amplified using Palumbi *et al.*'s (1991) universal primers while the COI fragment (approximately 500 bp) was amplified using Folmer *et al.*'s (1994) universal primers. As mentioned previously, these same fragments were also used by Carlini *et al.* (2000) alongside the actin gene for the coleoid cephalopod phylogeny. Furthermore, both 16S (Thollesson, 1999) and COI (Remigio & Hebert, 2003) were employed with some degree of success to evaluate higher level phylogenies within the Gastropoda even though both genes are susceptible to significant levels of saturation. Neither Palumbi *et al.*'s (1991) universal 16S primers nor Folmer *et al.*'s (1994) universal COI primers have been applied previously to phylogenetic analyses of the Achatinoidea.

Aside from the COI and the 16S rRNA genes, several other mitochondrial genes have also been used for inferring deep level phylogenies within the gastropods. Grande *et al.* (2004) employed several mitochondrial genes, in addition to the 16S and COI, such as the tRNA-valine, tRNA-arginine, tRNA-proline and the NADH dehydrogenase subunits 5 and 6 genes in the study of the Euthyneura (opisthobranchs and pulmonates) in which their molecular data rejected the monophyly of the pulmonates.

At present, no study has attempted to use nuclear and mitochondrial genes in concert to address the issues regarding the relationships within the Achatinoidea. Using several genes together approximates the species tree that traces the evolutionary relationships of the species under consideration, which is more desirable than

representing their relationships based on the history of individual genes, as shown by a gene tree (Grauer & Li, 2000). Combining these genes could also resolve the phylogeny of the Achatinoidea at all levels, with the faster evolving mitochondrial genes (COI and 16S) resolving the shallower nodes and the slower evolving nuclear genes (rRNA, actin and possibly histone 3) resolving the deeper nodes (Graybeal, 1994; Johnson & Clayton, 2000). Combining genes could also increase phylogenetic signal and disperse noise (Baldauf *et al.* 2000; Gontcharov *et al.*, 2004).

3.1.4. Objectives of this study

This study had the following objectives: (1) to identify the primary divisions within the superfamily Achatinoidea and to elucidate the relationship of the different families; (2) to establish if the Achatinidae are monophyletic and to identify their position relative to the other families within the Achatinoidea; (3) to determine whether the Ferussaciidae is monophyletic and to establish its phylogenetic position; (4) to settle the taxonomic designation of the Subulinidae, Coelioxidae and Glessulidae; and (5) to validate the different subfamilies of the Subulinidae. To address these objectives, the phylogeny of the Achatinoidea was inferred by examining representative taxa from all major groupings across the superfamily using molecular data from three nuclear genes (rRNA cluster, actin, and histone 3) and two mitochondrial genes (16S rRNA and COI). The genes were analyzed separately and in combination.

3.2. Materials and methods

3.2.1. Taxa used

Twenty-four taxa from six achatinoid families (Achatinidae, Coeliacidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophoprellidae) and three streptaxid outgroup taxa, two of which were used by Wade *et al.* (2006) and were found to fall at the base of the ‘achatinoid clade,’ were used to evaluate the phylogeny of the Achatinoidea (see Table 3.2). Four subulinid subfamilies following Schileyko’s (1999) classification (Petriolinae, Rishetiinae, Rumininae and Subulininae) were also represented in this study. Thirteen taxa were entirely new to this study while the remaining 14 taxa were used by Wade *et al.* (2006) in their phylogeny of the land snails based on a partial fragment of the rRNA cluster.

Table 3.2: Taxa used for the phylogenetic study of the Achatinoidea. Taxa marked with (*) are new to this study and were solicited through email by myself, C. Hudelot, F. Naggs and C. Wade from the collectors shown in the table. The DNA for the remaining taxa was provided by C. Wade from the collections obtained for Wade *et al.* (2001, 2006).

Family	Species	Collection/Location	Collector/Provider
Achatinidae	* <i>Achatina achatina</i> (Linnaeus, 1758)	Unknown (Zool. Soc. Lond. Colln.)	NHM**
	<i>Achatina fulica</i> Bowdich, 1822	Captive bred, unknown origin	NHM**
	* <i>Achatina stuhlmanni</i> von Martens, 1892	Semuliki National Park, Uganda	B. Rowson
	* <i>Cochlitoma ustulata</i> (Lamarck, 1822)	Western Cape Prov., South Africa	A. Moussalli & D. Stuart-Fox
Coeliacidae	<i>Coeliaxis blandii</i> (Pfeiffer, 1852)	New Bradford, South Africa	N. Smith
	<i>Pyrgina umbilicata</i> Greeff, 1882	São Thomé	A. Gascoigne
Ferussaciidae	* <i>Cecilioides gokweanus</i> (Boettger, 1870)	Cape Vida, Bhangazi Hill, Zululand, South Africa	D. Herbert
	<i>Ferussacia folliculus</i> (Gmelin, 1791)	Los Alcornales, Prov Cadiz, Spain	M. Seddon
Glessulidae	<i>Glessula ceylanica</i> (Pfeiffer, 1845)	Colombo, Sri Lanka	P. Karunaratne
Subulinidae			
S.f. Petriolinae	<i>Bocageia</i> sp.	São Thomé	A. Gascoigne
	* <i>Subulona</i> sp.	Ossen Forest, Tupen Hills, Kenya	M. Pickford
S.f. Rishetiinae	* <i>Eutomopeas layardi</i> (Benson, 1863)	Koralegama, Sri Lanka	?
	* <i>Tortaxis erectus</i> (Pilsbry, 1906)	Guilin, Guangxi Prov., China	R. Anderson
S.f. Rumininae	<i>Riebeckia</i> sp.	Samha, Sokotra Archipelago	E. Neubert
	<i>Rumina decollata</i> (Linnaeus, 1758)	Sicily	A. Davison
	<i>Xerocerastus</i> sp.	Otjiwarongo, Namibia	W. Sirgel
S.f. Subulininae	* <i>Allopeas clavulinum</i> (Potiez & Michaud, 1838)	University of Sao Paulo, Brazil	F. Florens & C. Baider
	* <i>Leptinaria lamellata</i> (Potiez & Michaud, 1838)	Botanical Garden, Rio de Janeiro, Brazil	F. Florens & C. Baider
	* <i>Paropeas achatinaceum</i> (L. Pfeiffer, 1846)	Agra Gajaba's Garden Sri Lanka	D. Raheem
	* <i>Subulina octona</i> (Bruguere, 1789)	Island of Pulo Anna, Sonsorol, Southwest Islands, Republic of Palau	R. Rundell & A.M. Gawel
	<i>Subulina striatella</i> (Rang, 1831)	Kew Gardens (introduced)	F. Naggs
	* <i>Subulina vitrea</i> (Mousson, 1887)	Gauss, Namibia	?
	<i>Zootecus insularis</i> (Ehrenberg, 1831)	Dubai, United Arab Emirates	S. Green
	Thyrophorellidae	<i>Thyrophorella thomensis</i> Greeff, 1882	Zampala, São Thomé, West Africa
Streptaxidae (outgroups)	* <i>Gibbulinella dewinteri</i> Bank, Groh & Ripken, 2002	Puntas Coloradas, La Gomera Island	M. Ibañez
	<i>Gonaxis quadrilateralis</i> Preston, 1910	Reunion	O. Griffiths
	<i>Gonospira</i> sp.	Mauritius	O. Griffiths

** NHM-Natural History Museum

3.2.2. DNA extraction, PCR amplification and sequencing

For all new specimens, tissue slices (approximately eight mm³) from the foot muscle of the snail were obtained and the DNA was extracted using the standard CTAB method of DNA extraction (Section 2.1.1 of Chapter 2, pp. 37-39). The DNA was

provided by C. M. Wade for those specimens utilized in the Wade *et al.* (2001, 2006) studies.

Amplification by PCR, gel migration and purification of PCR products from agarose gels were carried out as described in Sections 2.2-2.4 of Chapter 2, pp. 42-59. Nearly the entire fragment of the nuclear LSU rRNA gene, the internal transcribed spacer 2 (ITS2) and 80 bp of the 5.8S rRNA gene (for a total of approximately 4000 bp) were amplified using six overlapping primer pairs listed in Table 2.1 of Chapter 2, p. 47. Sequences of the Morgan (2002) nuclear cytoplasmic actin gene fragment (approximately 900 bp), the Colgan *et al.* (2003) nuclear histone 3 fragment (328 bp), the Folmer *et al.* (1994) mitochondrial COI fragment (approximately 650 bp) and the Palumbi *et al.* (1991) mitochondrial 16S fragment (420-500 bp) were amplified using the primers listed in Tables 2.2-2.5 of Chapter 2, pp. 50-53. For all fragments, both sense and anti-sense strands were sequenced directly using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 termination cycle sequencing chemistry (see Section 2.6 of Chapter 2, pp. 59-61 for details).

3.2.3. Sequence analysis

Sequences were assembled using the STADEN package version 1.5.3 (Staden *et al.*, 2000) and aligned manually within the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994). The rRNA and 16S rRNA sequences were aligned following alignments generated by C. Hudelot for the Stylommatophora based on the secondary structure of these genes. Ambiguous sites due to sequencing errors (for all genes) or the presence of multiple copies (for the actin gene) were assigned as described in Section 2.9.3 of Chapter 2, pp. 72-73. Cloned sequences of actin gene copies from three

representative taxa of the Achatinoidea were used as a guide to assign ambiguous sites in the actin gene (see Section 2.7 of Chapter 2, pp. 61-66).

For each gene fragment, the average base frequencies as well as the numbers of variable and parsimony-informative sites were determined in PAUP* Version 4.0b10 package (Swofford, 2002) (Section 2.9.4 of Chapter 2, p. 73). Corrected distances were computed after determination of the optimal model for DNA sequence evolution. This was carried out using likelihood by estimating the log likelihood scores in the PAUP* for the JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P (Kimura, 1980), HKY85 (Hasegawa *et al.*, 1985), TN93 (Tamura & Nei, 1993) and the GTR (Rodriguez *et al.*, 1990) models as well as their variant that incorporated gamma distributed rates (Γ) (Yang, 1993), after which the scores were compared for significant differences using the Likelihood Ratio Test (LRT) (see Section 2.9.5 of Chapter 2, pp. 73-77 for details of the LRT).

In order to examine the sequences for evidence of saturation, the following plots were generated: pairwise corrected versus pairwise uncorrected distances (Plot 1); pairwise uncorrected transition and transversion distances versus pairwise uncorrected total distances (Plot 2); and pairwise uncorrected transition distances versus pairwise uncorrected transversion distances (Plot 3) (see Section 2.9.6 of Chapter 2, pp. 77-81). Gene datasets that demonstrated saturation were excluded from subsequent phylogenetic analyses. To test for the presence of phylogenetic signal, tree length distribution using parsimony was also determined for a $g1$ measure of skewness for 10,000 tree length replicates (see Section 2.9.7 of Chapter 2, pp. 81-82). Due to the potential problems posed by the presence of paralogous sequences the actin gene was assessed for its utility in phylogenetic analyses using the procedures outlined in Section 2.9.13, pp. 90-91.

Phylogenetic trees were generated from the aligned datasets using the model based maximum likelihood (ML), Bayesian inference (BI) and neighbor-joining (NJ) methods and the non-model based maximum parsimony (MP) method (Section 2.9.8 of Chapter 2, pp. 82-85). The trees for NJ and MP were generated in PAUP* while that of ML was generated using PHYML Version 2.4.4 (Guindon *et al.*, 2005). Bootstrap resampling (Felsenstein, 1985) with 1000 replicates for MP and NJ (using PAUP*) and 1000 replicates for ML (using PHYML) were also carried out. Bayesian inference (BI) was performed using the MrBayes (Version 3.1.2) package (Ronquist & Huelsenbeck, 2003) using four chains of a Markov Chain Monte Carlo algorithm. The number of generations to explore the tree space and the heating temperature used for each gene were optimised as described in the BI segment of Section 2.9.8 of Chapter 2, pp. 83-84 and on Appendix 2.3.2, pp. 369-370. A consensus tree for each gene (where unsaturated) was constructed using the last 1000 trees.

To determine if the sequences from the genes that did not exhibit saturation could be combined and analyzed as a single dataset of concatenated sequences, a partition-homogeneity test (Swofford, 2002) was carried out within PAUP* (Section 2.9.10 of Chapter 2, p. 86-88). Two sets of concatenated sequences were prepared. The first one included only those taxa with complete sequences for all the gene fragments under consideration while the second set included all taxa. Studies show that taxa with incomplete datasets could still be used and their phylogenetic positions accurately determined provided that a lot of characters were surveyed from the other sequences (Wiens, 2006). Inclusion of such taxa could also benefit the combined gene analyses by removing misleading long branches and potentially increase the accuracy of the phylogeny (Wiens, 2006).

Where taxonomic groups expected to be monophyletic based on taxonomy did not cluster together in the optimal trees, their monophyly was tested using the Shimodaira-Hasegawa (1999) test in PAUP* (Section 2.9.11 of Chapter 2, p. 89).

3.3. Results

3.3.1. Molecular data

Twenty four achatinoid and three streptaxid outgroup taxa (*Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp.) were examined. Sequences were obtained for the nuclear rRNA cluster, actin and histone 3 genes as well as the mitochondrial 16S and cytochrome c oxidase I genes. Sequence alignments are presented in Appendices 3.1-3.5 on pp. 375-432. All sequences were scanned for the presence of ambiguous positions in the direct sequences that might be indicative of the presence of multiple gene copies. Ambiguities indicative of multiple gene copies were detected only for the actin gene (see Section 3.3.2.2, p. 155-156 for details). A summary of the molecular data for all the sequences is shown in Table 3.2. For the protein-coding genes (actin, histone 3 and COI), summary information was obtained separately for the entire gene (all codon positions), the combined 1st and 2nd codon positions and the 3rd codon position only. Uncorrected and corrected pairwise distances and their optimal models were determined for all five genes and are summarized in Table 3.3. The GTR+ Γ model was found to be the optimal model by LRT for the nuclear rRNA cluster, the mitochondrial 16S RNA and for both the full (all codon positions) and 3rd codon position datasets for the actin, histone 3 and COI genes. For the combined 1st + 2nd codon position datasets, TNR93+ Γ was determined to be the

optimal model for the actin, TN93 for the histone 3, and GTR+ Γ for COI. (See Appendix 3.6, p. 433 for the summary of the LRT results).

Nearly the entire fragment of the LSU rRNA gene, the internal transcribed spacer 2 (ITS 2) and 80 nucleotides of the 5.8S rRNA gene was sequenced (approximately 4000 nucleotides) for the rRNA gene cluster, of which 3435 nucleotides could be aligned unambiguously. The ITS 2 region was too variable to align across all taxa and was therefore excluded from the analysis. A total of 260 (7.6%) variable sites were found, of which 151 were parsimony-informative. Pairwise distances across all taxa ranged from 0.001 to 0.034 (uncorrected) and 0.001 to 0.062 (corrected). The highest base frequency was obtained for G (0.318) followed by C (0.260), then by A (0.229) and finally by T (0.193). For the actin gene, 861 nucleotide positions were sequenced, of which all sites could be aligned unambiguously; of these, 288 (33.5%) were variable and 240 were parsimony-informative. Pairwise distances ranged from 0.009 to 0.173 (uncorrected) and 0.009 to 0.469 (corrected). Highest average base frequency was for A (0.274) followed by C (0.255) then T (0.250) and finally G (0.220). The majority of the variable sites (237) and parsimony-informative sites (210) were found at the 3rd codon position, while the combined 1st and 2nd codon positions only had 51 variable sites and 30 parsimony-informative sites. For the histone 3 gene, 328 nucleotides were sequenced and aligned unambiguously, with 103 (31.4%) variable positions and 71 parsimony-informative sites. Pairwise distances ranged from 0 to 0.183 (uncorrected) and 0 to 0.605 (corrected). Average base frequencies, in decreasing order, were as follows: C (0.321); G (0.271); A (0.242) and T (0.166). As with the actin gene, the 3rd codon position exhibited the most number of variable sites (93) as well as parsimony-informative sites (70) in comparison to the combined 1st and 2nd codon positions with only 10 and 1, respectively. Amplification of the histone 3 gene

fragment for *Cecilioides gokweanus* (Ferussaciidae), *Paropeas achatinaceum*, *Riebeckia* sp., and *Subulina vitrea* (Mousson) (Subulinidae) proved unsuccessful. Only a single streptaxid outgroup sequence (*Gibbulinella dewinteri*) was obtained for the histone 3 gene. For the COI gene, approximately 650 nucleotides were sequenced, with the length varying depending on the primers used. A total of 607 unambiguously aligned nucleotides were used; of which 279 (46.0%) were variable and 250 were parsimony-informative. Pairwise distances ranged from 0.160 to 0.265 (uncorrected) and 7.215 to 57.369 (corrected). Average base frequencies, in decreasing order, were as follows: T (0.395); A (0.249); G (0.194) and C (0.163), indicating a strong bias for T; this was heightened in the 3rd codon position where average frequency of T rose to 0.462. The 3rd codon position had 201 variable sites and 188 parsimony-informative sites, whereas the combined 1st and 2nd codon positions only had 78 and 62, respectively. Amplification of the COI gene for *Cecilioides gokweanus* was also unsuccessful. As with the histone 3 gene, only a single streptaxid outgroup sequence (*Gibbulinella dewinteri*) was obtained for the COI gene. The 16S rRNA gene product ranged from 420-500 nucleotides, with 294 that were unambiguously aligned, 139 (47.3%) that were variable, and 120 that were parsimony-informative. Pairwise distances ranged from 0.078 to 0.279 (uncorrected) and 0.107 and 1.082 (corrected). Base frequencies were as follows: T (0.306); A (0.296); G (0.217) and C (0.181), showing bias for A and T. Sequencing of the 16S rRNA gene fragment for *Eutomopeas layardi* and *Subulona* sp. (Subulinidae) was unsuccessful.

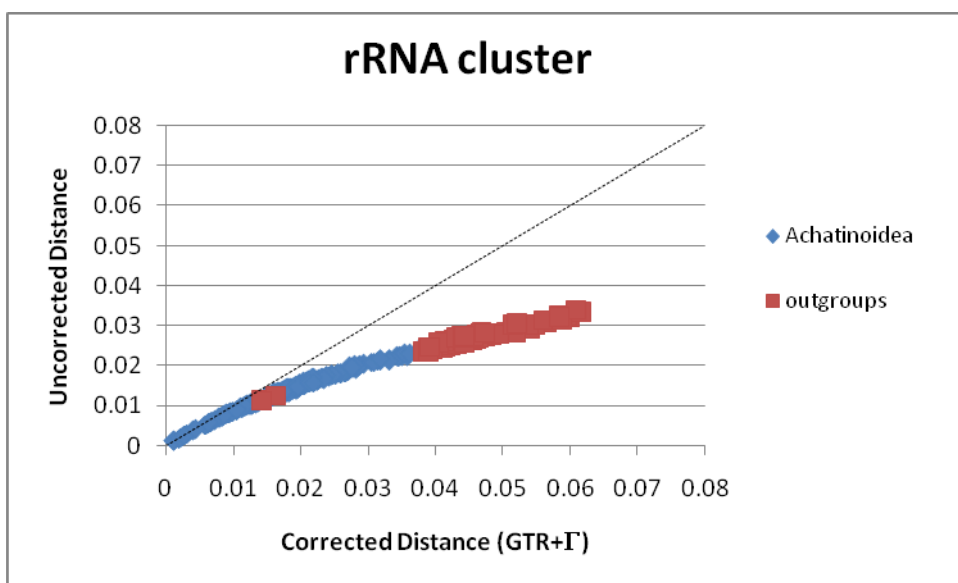
Table 3.3: Summary of molecular data across all genes used for the Achatinoidea and three streptaxid outgroup taxa (rRNA gene cluster, actin and 16S) and one streptaxid outgroup taxon (histone 3 and COI). A=Achatinoidea only; A+O=Achatinoidea and outgroup.

	Length (bp)	# of variable sites (%)		Range of distances (Uncorrected)		Optimal Model (α value for Γ distribution)	Range of distances (Corrected)		Average base frequencies								# of parsimony-informative sites	
									A		C		G		T			
									A	A+O	A	A+O	A	A+O	A	A+O		
rRNA cluster	3435	212 (6.2)	260 (7.6)	0.001-0.029	0.001-0.034	GTR+ Γ (0.049)	0.001-0.048	0.001-0.062	0.229	0.229	0.260	0.260	0.319	0.318	0.192	0.193	113	151
Actin combined	861	283 (32.9)	288 (33.5)	0.009-0.173	0.009-0.173	GTR+ Γ (0.190)	0.009-0.469	0.009-0.469	0.276	0.274	0.254	0.255	0.219	0.220	0.251	0.250	230	240
Actin 1 st and 2 nd codon positions	574	50 (8.7)	51 (8.9)	0-0.046	0-0.046	TN93+ Γ (0.059)	0-0.105	0-0.105	0.316	0.316	0.228	0.228	0.228	0.228	0.229	0.229	29	30
Actin 3 rd codon position	287	233 (81.2)	237 (82.6)	0.025-0.444	0.025-0.444	GTR+ Γ (1.339)	0.023-1.056	0.023-1.056	0.192	0.189	0.307	0.312	0.203	0.206	0.298	0.294	201	210
Histone3 combined	328	97 (29.6)	103 (31.4)	0-0.156	0-0.183	GTR+ Γ (0.164)	0-0.491	0-0.605	0.241	0.242	0.322	0.321	0.272	0.271	0.165	0.166	66	71
Histone 3 1 st and 2 nd codon positions	218	8 (3.7)	10 (4.6)	0-0.023	0-0.028	TN93 (N/A)	0-0.024	0-0.029	0.287	0.287	0.285	0.285	0.261	0.261	0.167	0.167	1	1
Histone 3 3 rd codon positions	110	89 (80.9)	93 (84.5)	0-0.449	0-0.520	GTR+ Γ (1.811)	0-1.289	0-1.452	0.145	0.147	0.397	0.395	0.297	0.295	0.161	0.163	65	70
COI combined	607	276 (45.5)	279 (46.0)	0.160-0.265	0.160-0.265	GTR+ Γ (0.077)	7.215-57.369	7.215-57.369	0.249	0.249	0.163	0.163	0.194	0.194	0.394	0.395	250	250
COI 1 st and 2 nd codon positions	404	75 (18.6)	78 (19.3)	0.032-0.119	0.032-0.119	GTR+ Γ (0.086)	0.046-0.459	0.046-0.459	0.205	0.205	0.199	0.199	0.235	0.235	0.361	0.361	62	62
COI 3 rd codon positions	203	201 (99.0)	201 (99.0)	0.365-0.631	0.365-0.631	GTR+ Γ (0.386)	299.377-5420.470	299.377-5420.470	0.335	0.336	0.091	0.090	0.114	0.113	0.459	0.462	188	188
16S rRNA	294	137 (46.6)	139 (47.3)	0.078-0.279	0.078-0.279	GTR+ Γ (0.233)	0.107-1.082	0.107-1.082	0.292	0.296	0.184	0.181	0.220	0.217	0.304	0.306	113	120

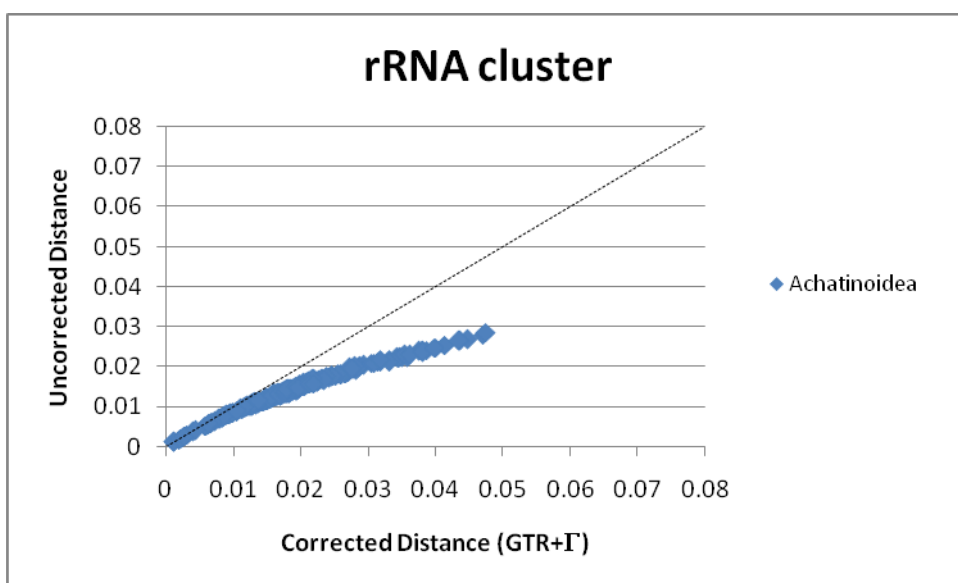
3.3.2. Sequence analyses

3.3.2.1. Evaluating for saturation and phylogenetic signal

For the rRNA cluster, the uncorrected versus corrected distances plots (Plot 1, Figure 3.2) demonstrated that the corrected distance based on the optimal GTR+ Γ model deviated from linearity starting at an uncorrected (p) distance of approximately 0.01 but no plateau was reached for the plots of either the Achatinoidea or the Achatinoidea plus outgroups. This suggested that the dataset for the rRNA cluster was far from being saturated and that the GTR+ Γ model was adequate at correcting the distances for multiple hits.



(A)



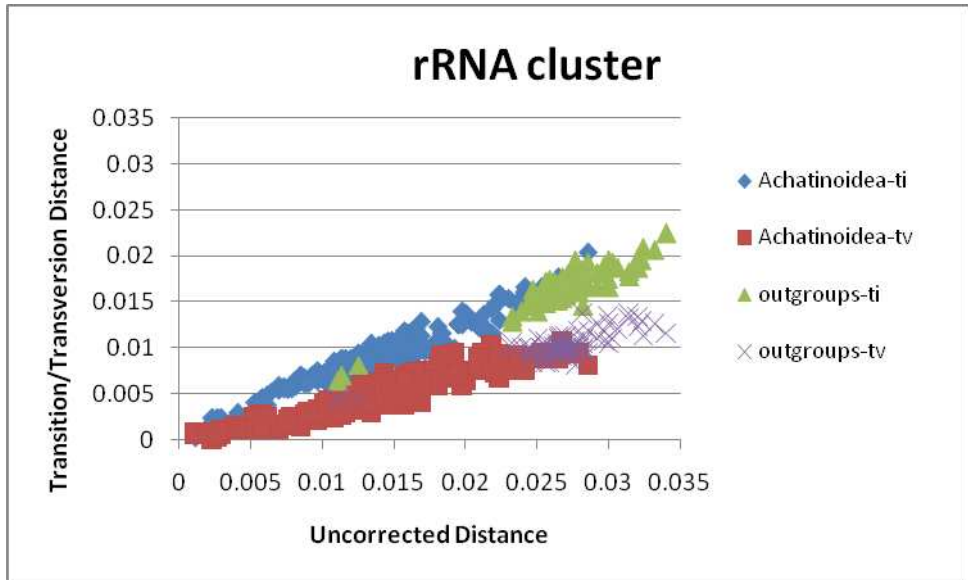
(B)

Figure 3.2: Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the rRNA cluster in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

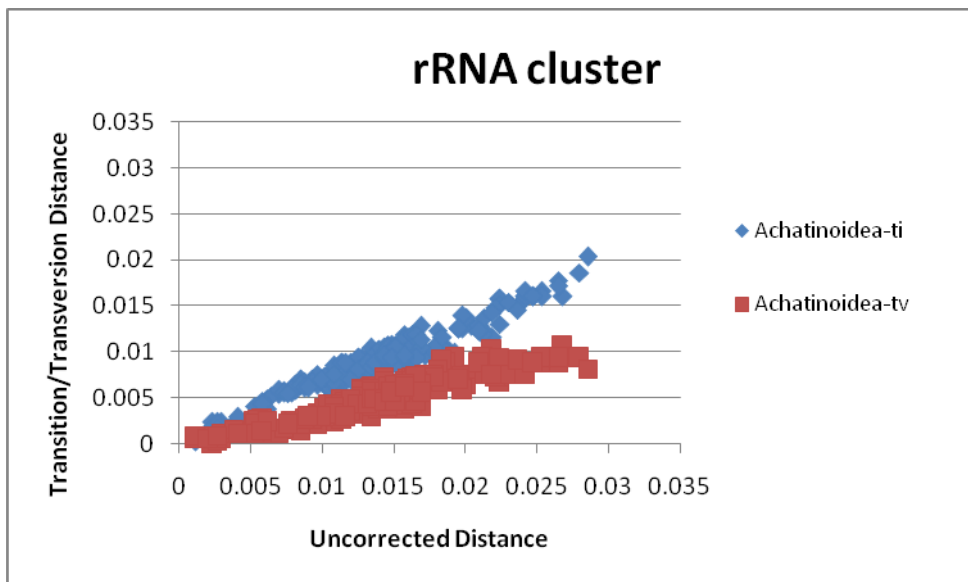
Plots for the transition and transversion rates versus uncorrected distances both with and without the outgroup taxa (Plot 2, Figure 3.3) showed transition distances that were higher than the transversion distances, with both types of substitutions increasing linearly. Plots for transitions against transversions both with and without the outgroup taxa (Plot 3, Figure 3.4) demonstrated that all transitions were higher than transversions. These suggested that neither transition nor transversion distances were saturated for the rRNA cluster.

A g_i value of -1.200 was obtained for the rRNA cluster with 10,000 replicates based on 27 taxa and 151 parsimony-informative characters. This value differed significantly from the critical g_i value of -0.1 at $P=0.05$ level of significance for 25 taxa and 100 parsimony-informative characters (Hillis & Huelsenbeck, 1992). This result was indicative of a strong phylogenetic signal.

The lack of substitution saturation and the presence of a strong phylogenetic signal suggested that the rRNA cluster was suitable for phylogenetic analyses of the Achatinoidea and outgroups.

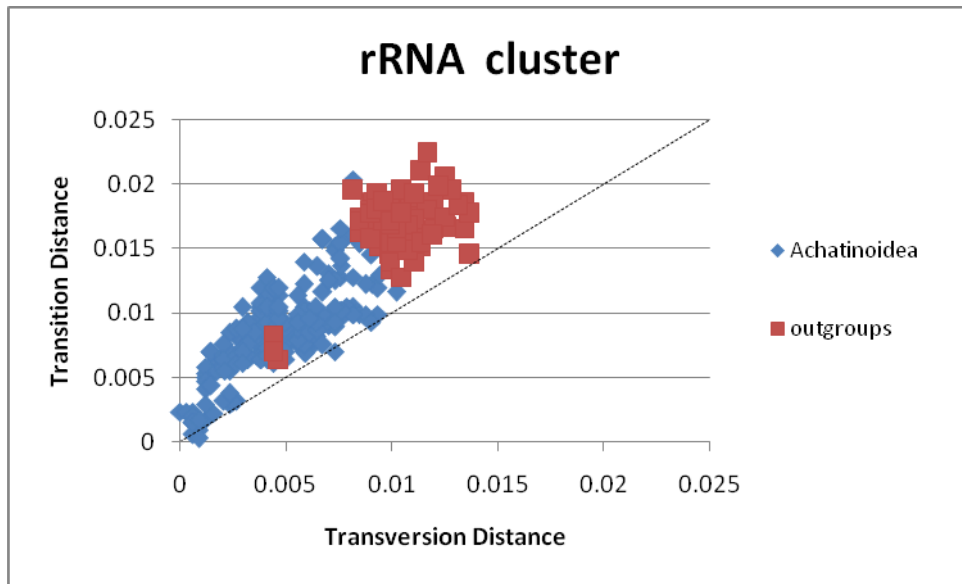


(A)

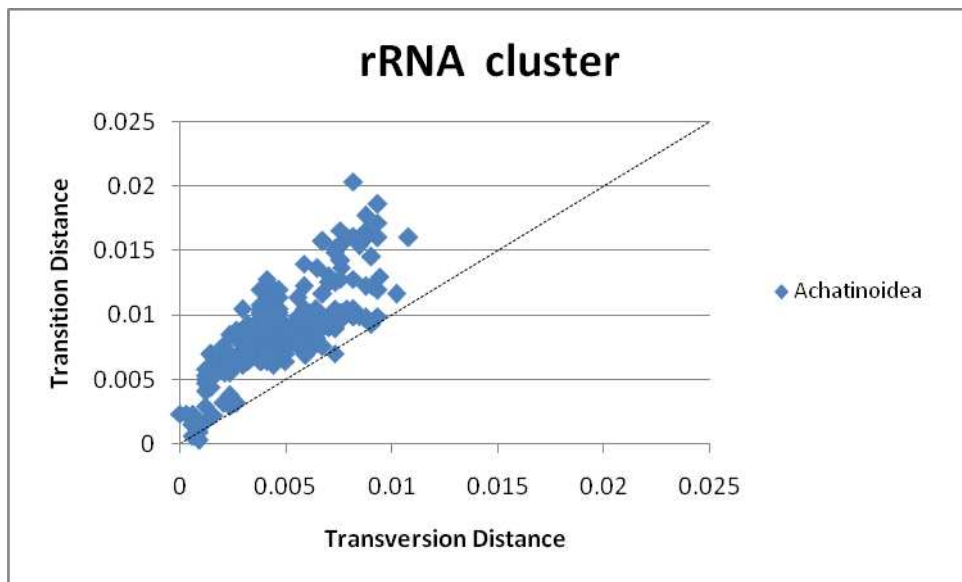


(B)

Figure 3.3: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the rRNA cluster in (A) the Achatinoidea and outgroup taxa and (B) the Achatinoidea only.



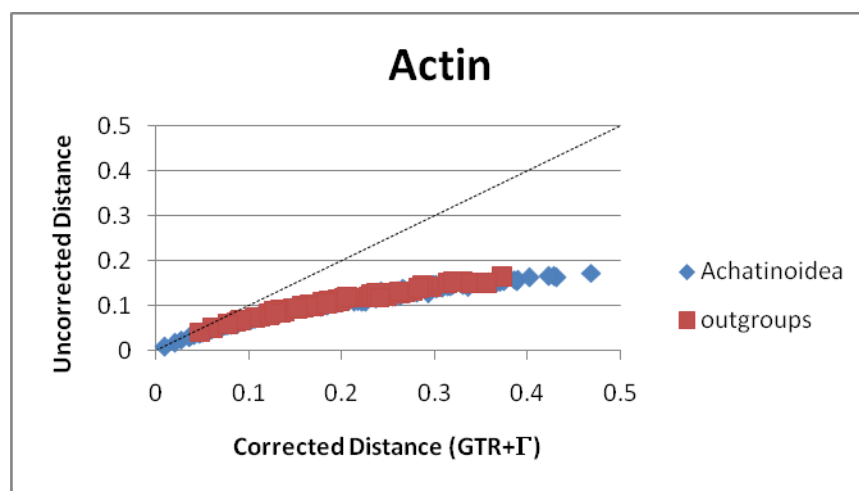
(A)



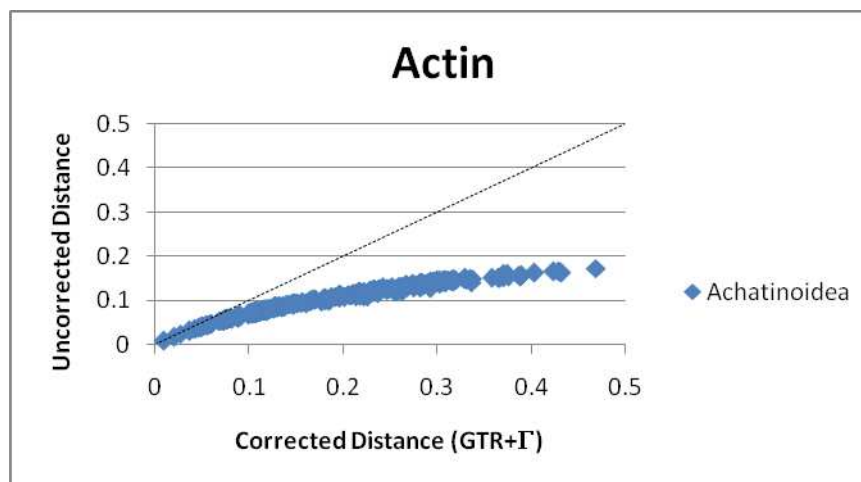
(B)

Figure 3.4: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the rRNA cluster in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

For the actin gene, Plot 1 (Figure 3.5) showed that the corrected distance, based on the optimal GTR+ Γ model, deviated from linearity starting at an uncorrected (p) distance of approximately 0.04 for both the Achatinoidea only and the Achatinoidea plus outgroup taxa. The plots were still increasing and had not reached a plateau, suggesting adequate correction of the optimal GTR+ Γ model and no saturation in the dataset.



(A)



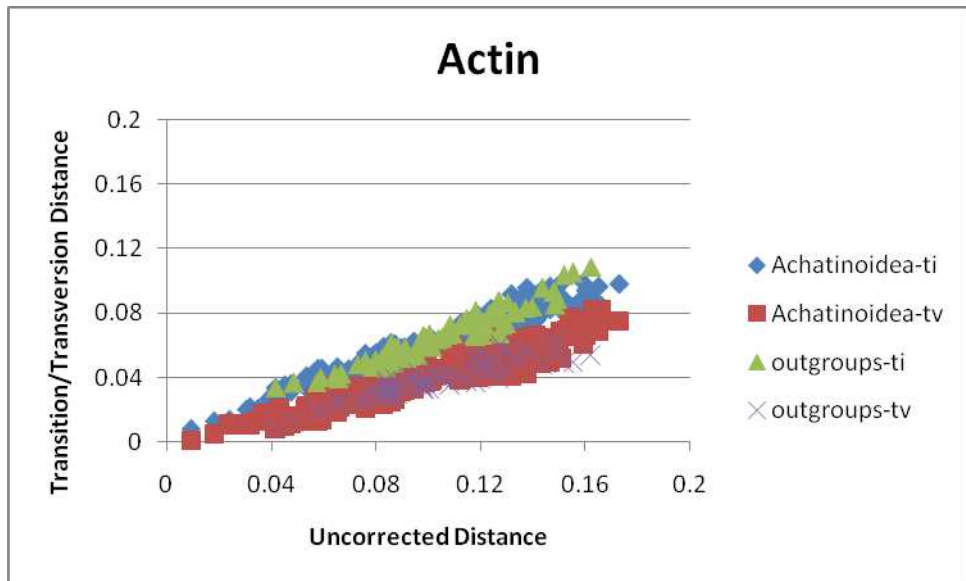
(B)

Figure 3.5: Plots of pairwise uncorrected distance against corrected distance for the actin gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

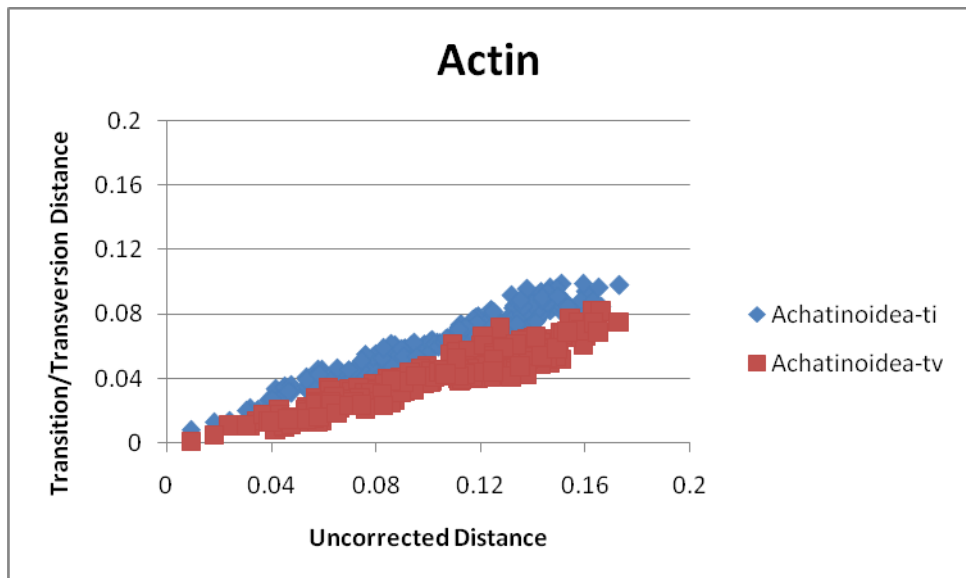
Plot 2 (Figure 3.6) for the actin gene, whether with or without outgroup taxa, exhibited a linear increase for both transitions and transversions. Plot 3 (Figure 3.7) showed that except for four points, all transitions were higher than transversions. These suggest that neither transition nor transversion distances were saturated for the actin gene.

A g_i value of -0.767 was computed for the actin gene based on 27 taxa and 245 parsimony-informative sites, much lower than the critical value of -0.1 at $p=0.05$ level of significance for 25 taxa and 100 parsimony-informative characters (Hillis & Huelsenbeck, 1992). The actin gene therefore exhibited strong phylogenetic signal.

The absence of substitution saturation and the presence of phylogenetic signal suggested that the actin gene was suitable for phylogenetic analyses of the Achatinoidea and outgroups.

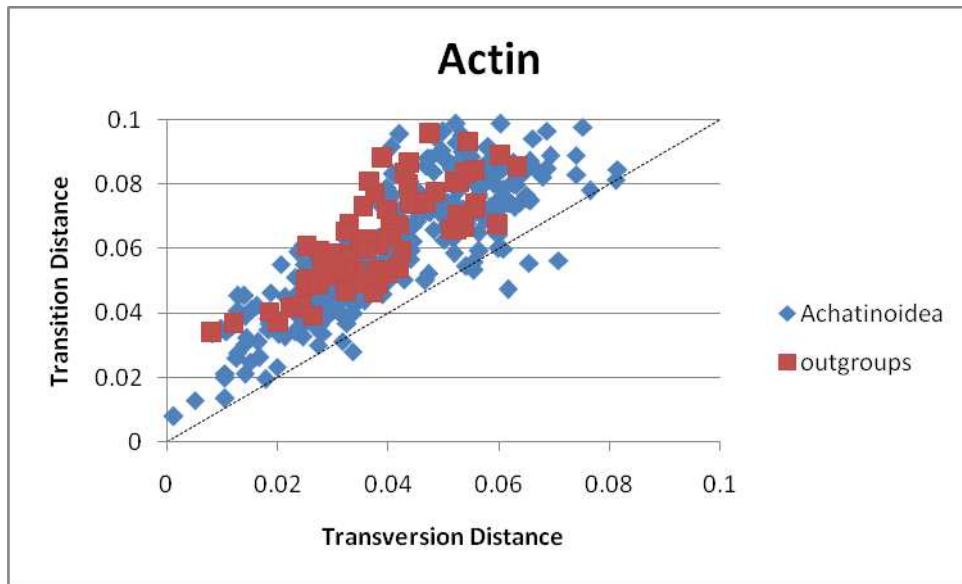


(A)

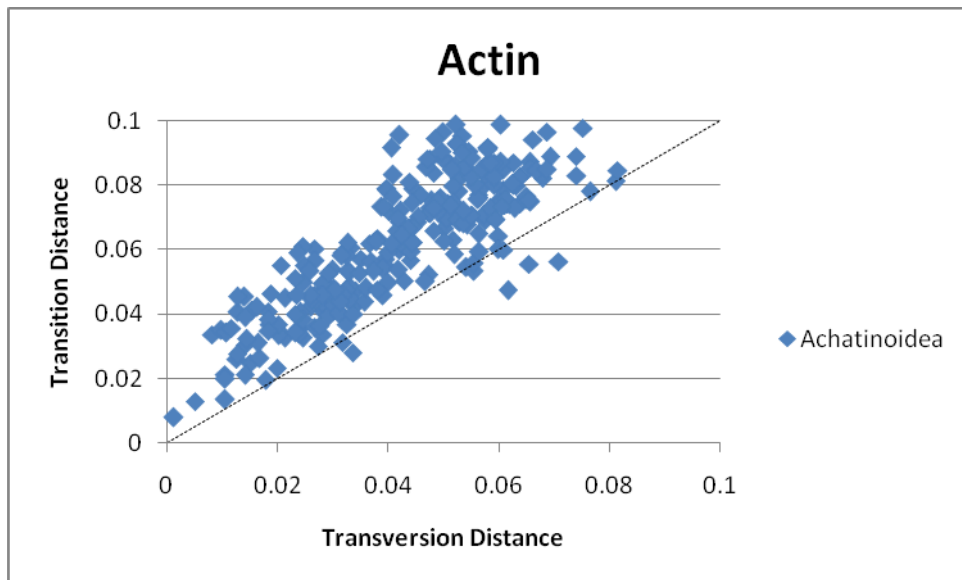


(B)

Figure 3.6: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the actin in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.



(A)



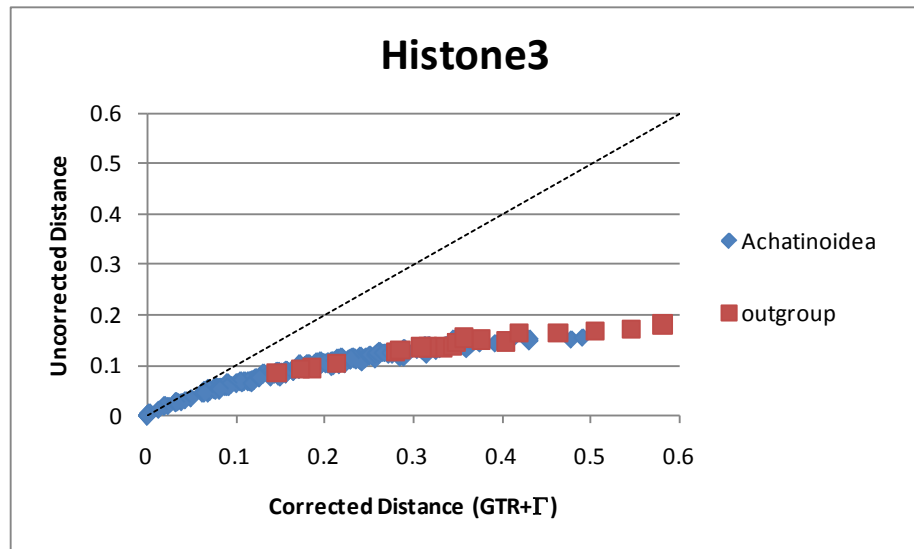
(B)

Figure 3.7: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the actin gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

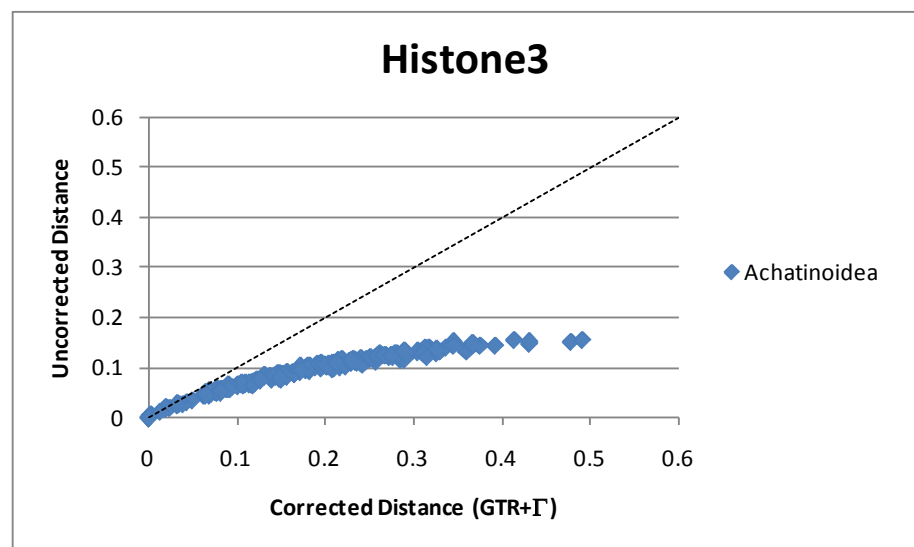
For the histone 3 gene, Plot 1 (Figure 3.8) revealed a curve for the corrected distances using the optimal GTR+ Γ model, with the deviation from a linear increase beginning at an uncorrected (p) distance of approximately 0.02 for both the

Achatinoidea only and including the streptaxid outgroup taxon *Gibbulinella dewinteri*.

The plots were still increasing and had not reached a plateau, implying that the histone 3 gene had not reached saturation.



(A)

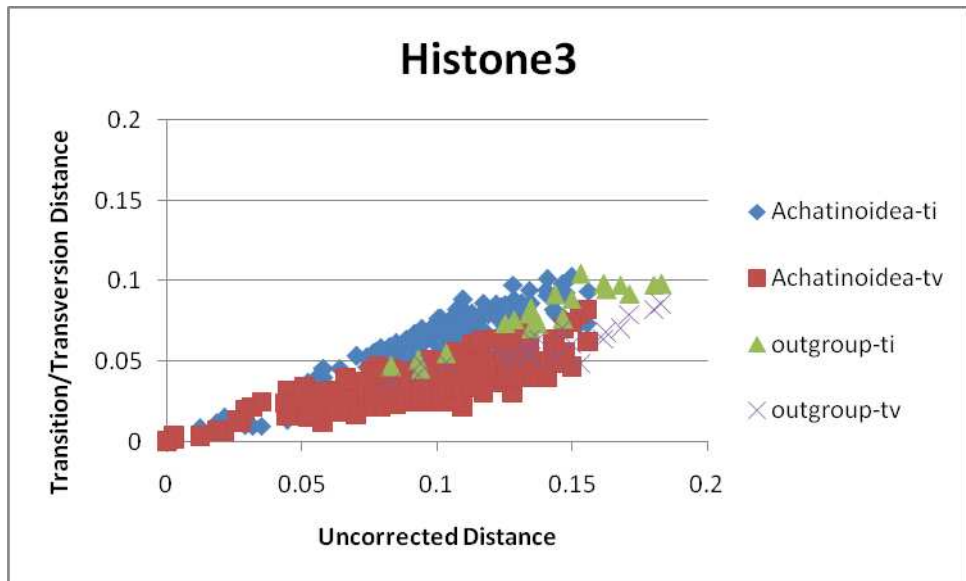


(B)

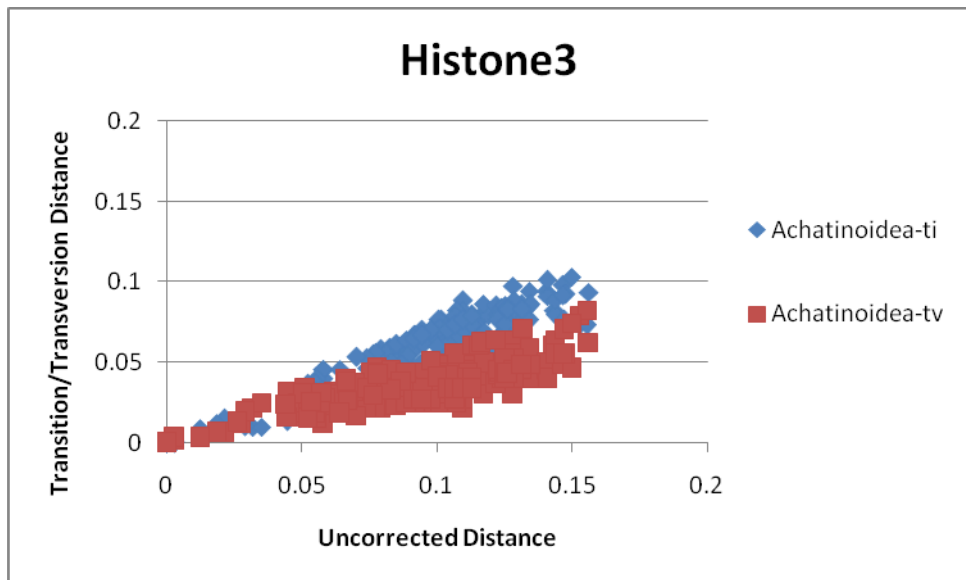
Figure 3.8: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.9) for histone 3 revealed a linear increase of both transitions and transversions for the Achatinoidea, but the transition line appeared to be beginning to curve when the outgroup was included. Although transitions were generally higher than transversions, some overlap was seen. Plot 3 (Figure 3.10) showed that the majority of transitions were higher than transversions. This suggested that the histone 3 dataset was just beginning to saturate, particularly if the outgroup taxon was included.

The g_1 value for histone 3 based on 21 taxa and 71 parsimony-informative sites was -0.539, which was significantly smaller than the critical value of -0.16 based on 15 taxa and 50 parsimony-informative sites. The histone 3 gene therefore exhibited phylogenetic signal.

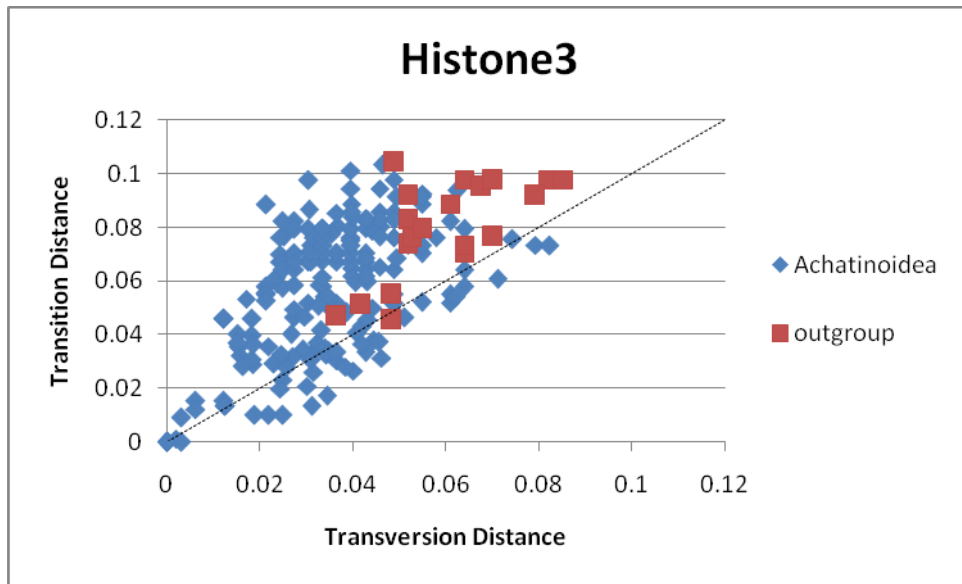


(A)

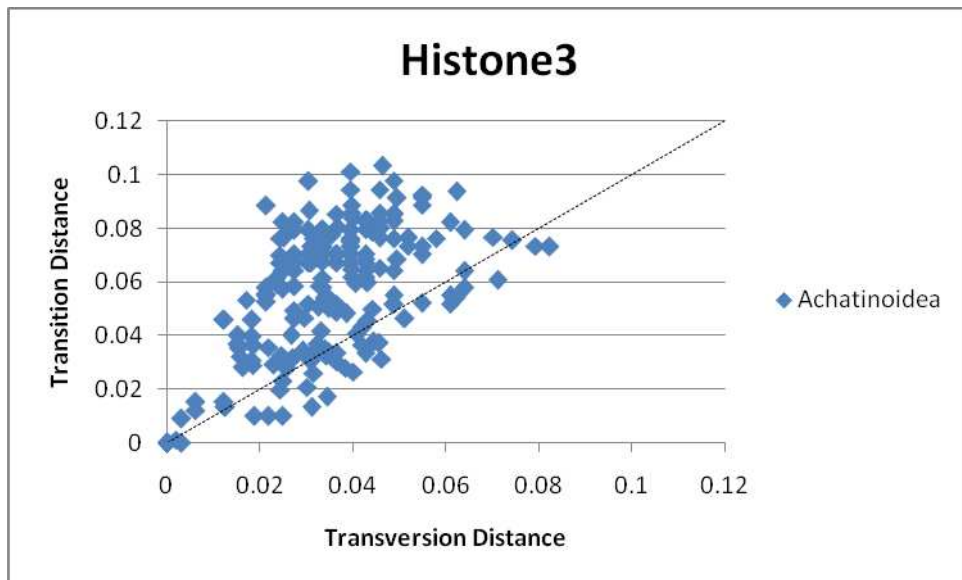


(B)

Figure 3.9: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

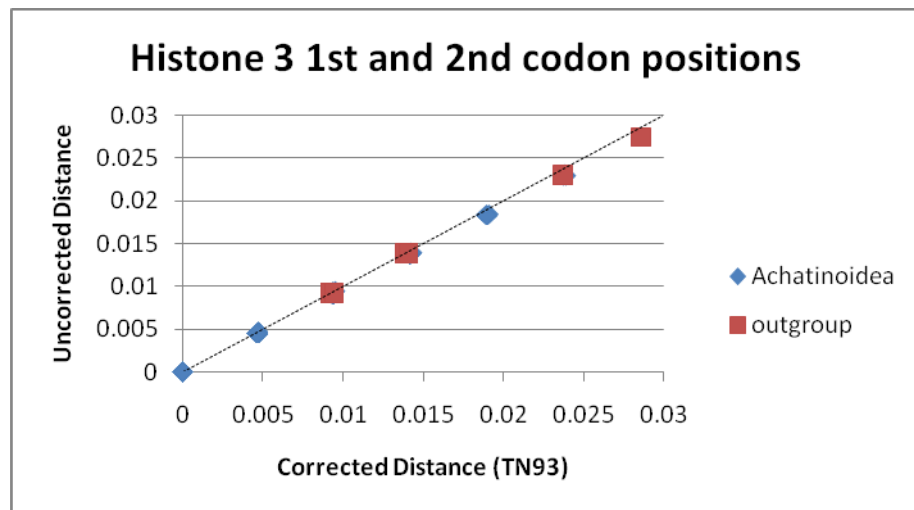


(B)

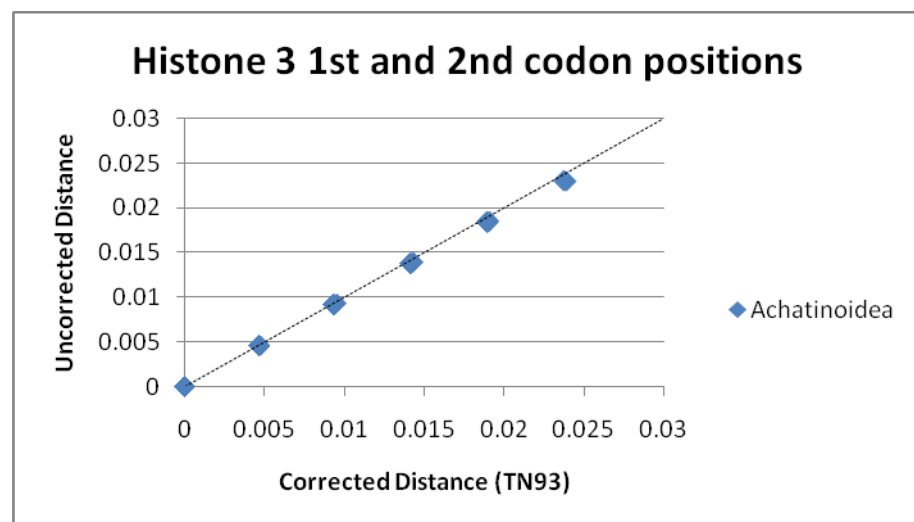
Figure 3.10: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

The analyses of the histone 3 gene as a whole (all codon positions) demonstrated that the dataset began to saturate when the outgroup taxon was included. The combined 1st and 2nd codon positions and the 3rd codon positions were therefore evaluated separately.

For the 1st and 2nd codon positions of the histone 3 gene, only a handful of sites varied (10 of 218 sites in the dataset that included the Achatinoidea and outgroup taxon). Plot 1 (Figure 3.11) showed a direct relationship between the corrected (based on the optimal TN93 model) and uncorrected distances, indicative of a highly conserved dataset with no saturation.



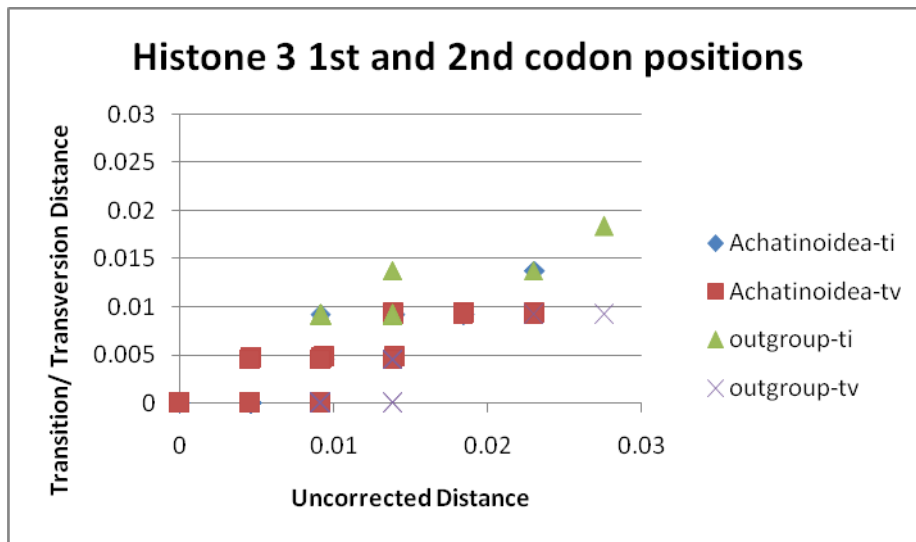
(A)



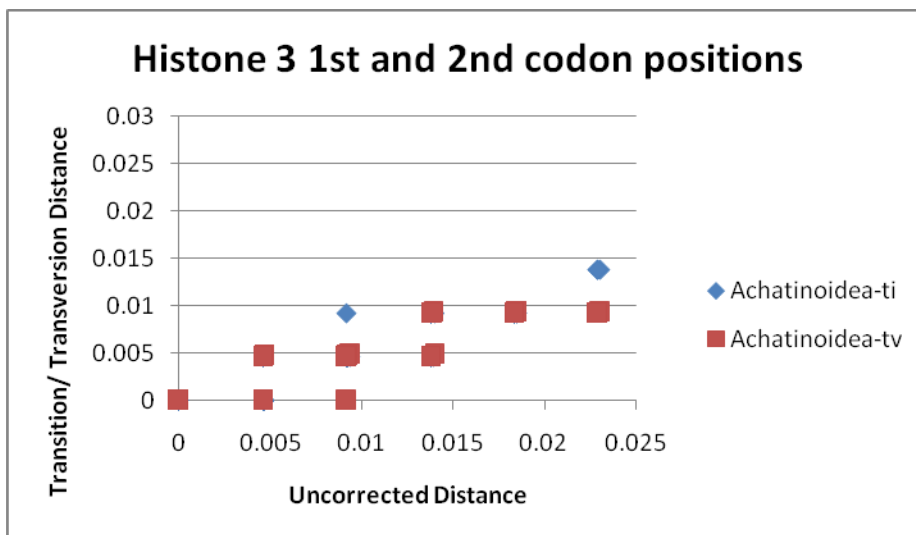
(B)

Figure 3.11: Plots of pairwise uncorrected distance versus corrected (TN93) distance for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plots 2 (Figure 3.12) and 3 (Figure 3.13) showed overlap between transitions and transversions, but curving due to saturation could not be assessed due to the small number of variable sites (10) evaluated.

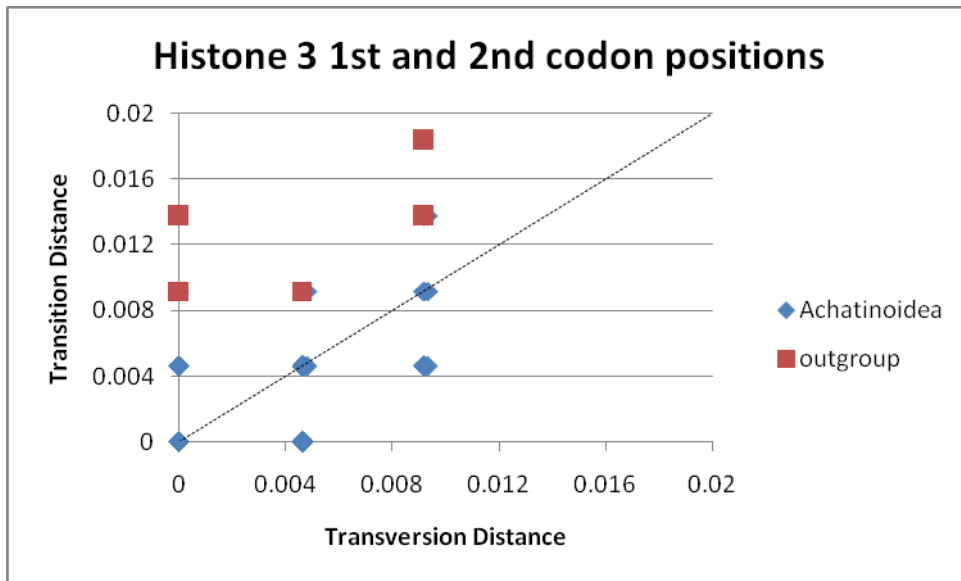


(A)

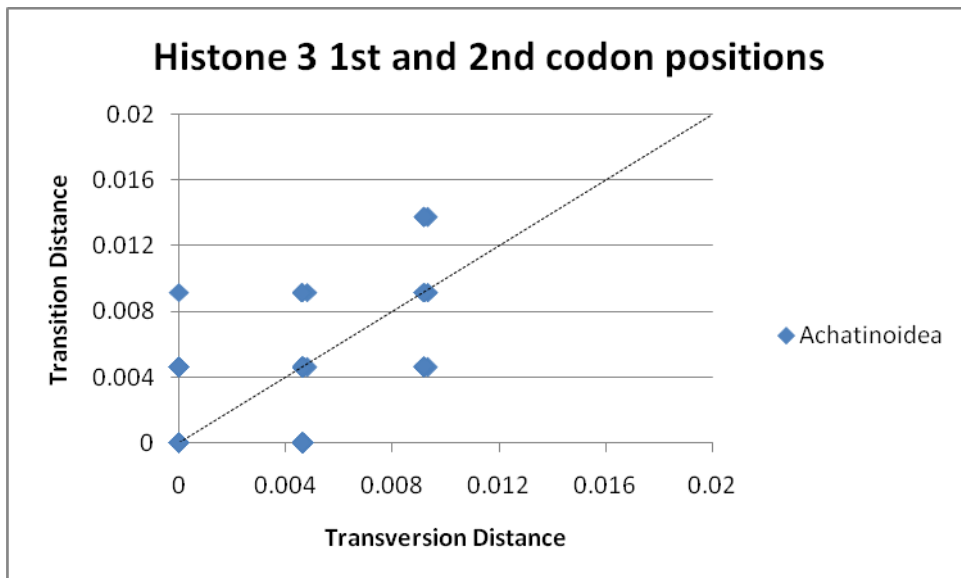


(B)

Figure 3.12: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only. Some transitions overlapped with transversions and were therefore not evident in the plots.



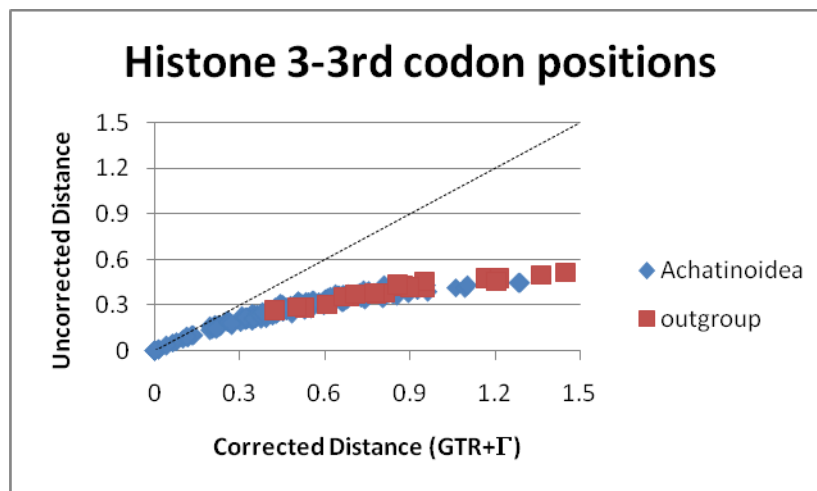
(A)



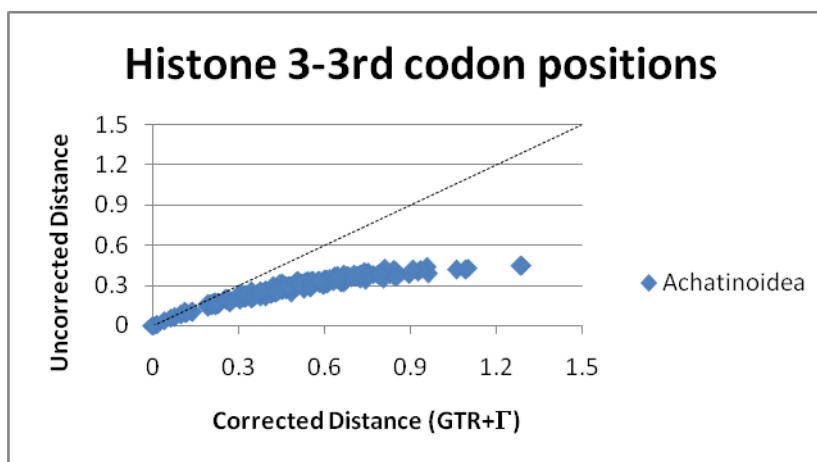
(B)

Figure 3.13: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

For the 3rd codon position, Plot 1 (Figure 3.14) revealed a curve for the corrected distances based on the optimal GTR+ Γ model that deviated from linearity at an uncorrected (p) distance of approximately 0.1. The corrected distances were still increasing and had not reached a plateau, implying that the dataset had not reached saturation.



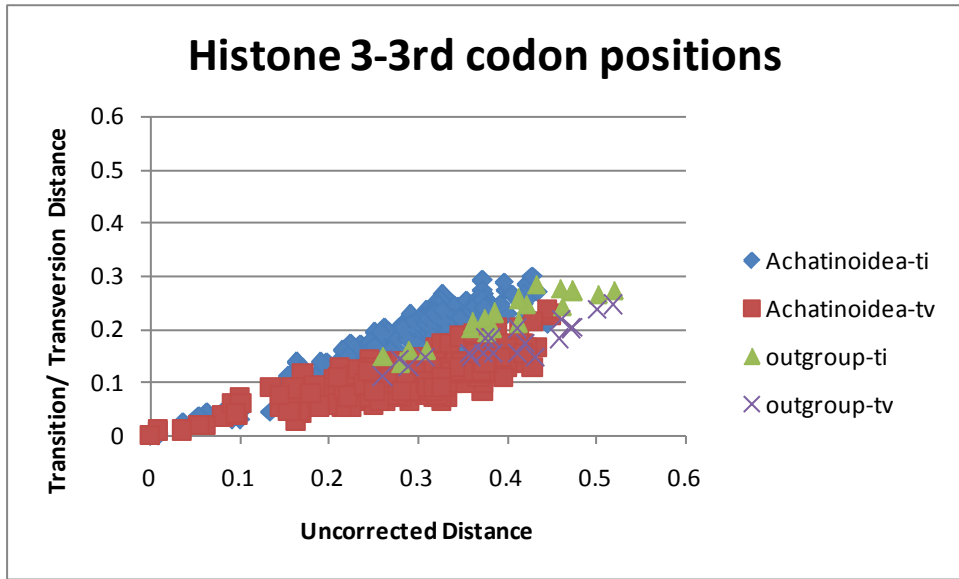
(A)



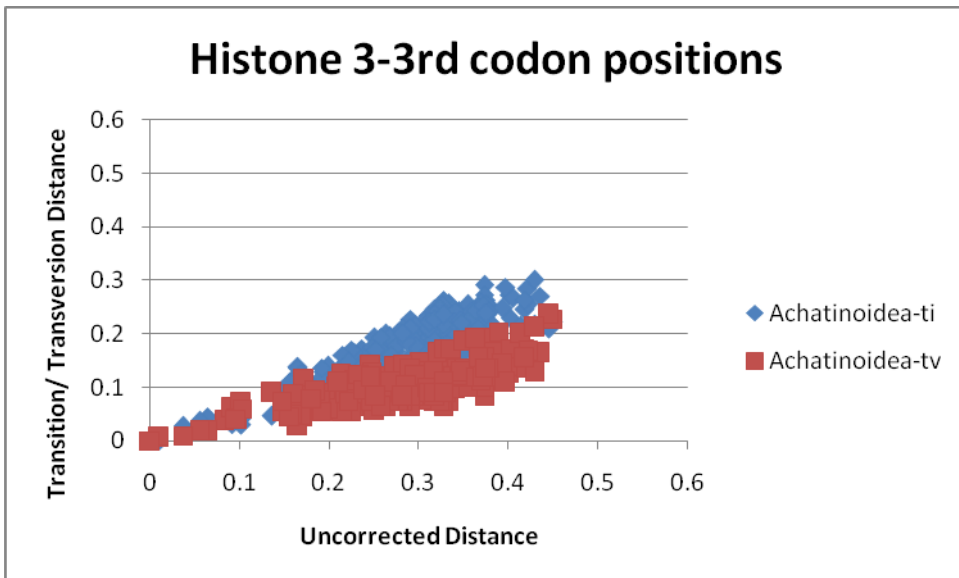
(B)

Figure 3.14: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 3rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.15) revealed a linear increase of both transitions and transversions, but the transition line appeared to be beginning to curve when the outgroup was included. Although transitions were generally higher than transversions, some overlap was seen. Plot 3 (Figure 3.16) demonstrated that majority of transitions were still higher than transversions. These results suggested that the 3rd codon positions of the histone 3 gene were just beginning to saturate, particularly as the outgroup taxon was included.

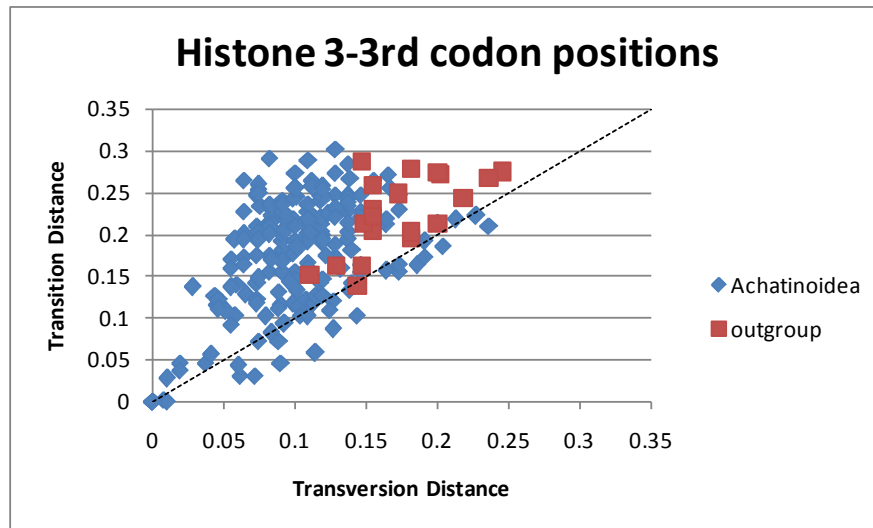


(A)

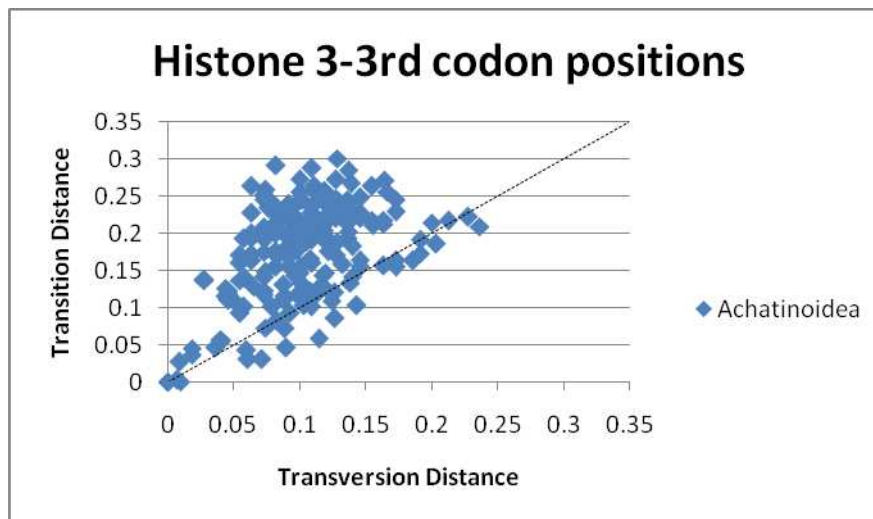


(B)

Figure 3.15: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

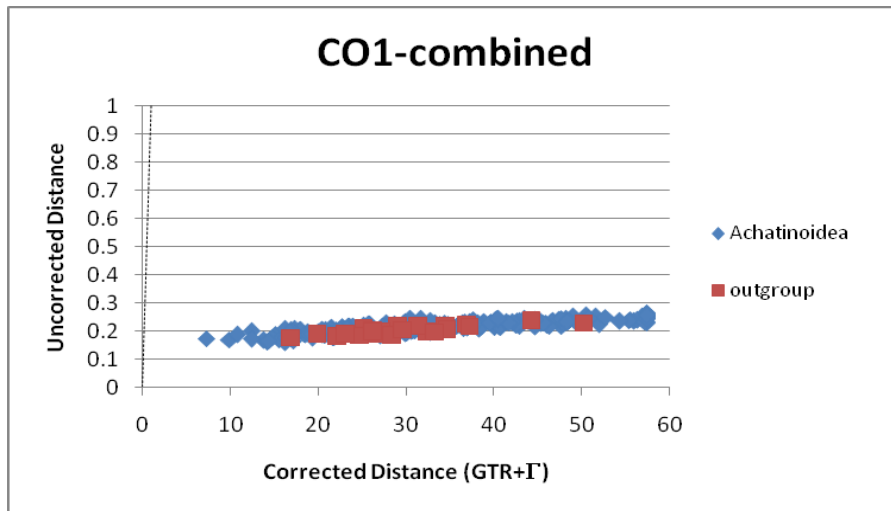


(B)

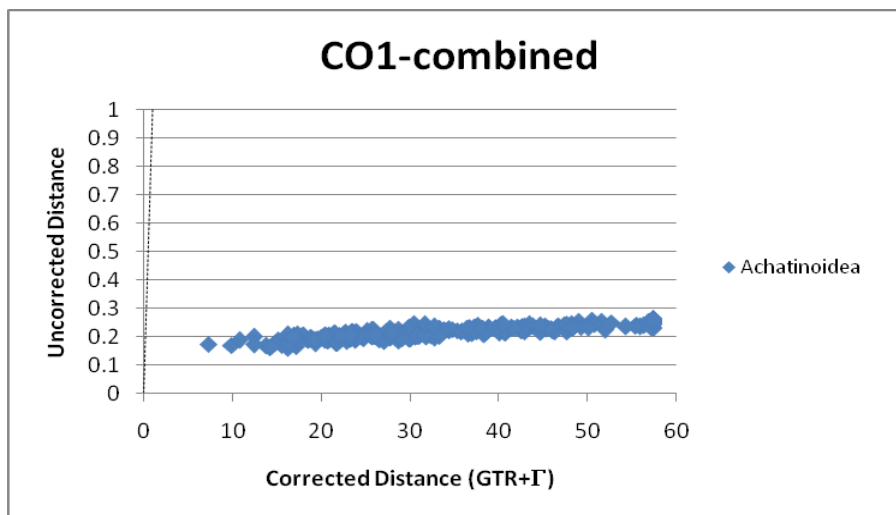
Figure 3.16: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3rd codon position of the histone 3 gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

The findings suggested that the histone 3 dataset could be employed in its entirety in phylogenetic analyses of the Achatinoidea though with caution, particularly when the outgroup taxon was included. Moreover, particular care should be taken over the interpretation of the findings of the non-model based maximum parsimony method.

For the COI gene, Plot 1 (Figure 3.17) yielded an almost horizontal trend for all taxa and for the Achatinoidea only. This, along with corrected distances in excess of 50, indicated severe saturation for the COI gene and the optimal GTR+ Γ model was unable to correct the dataset for multiple hits.



(A)

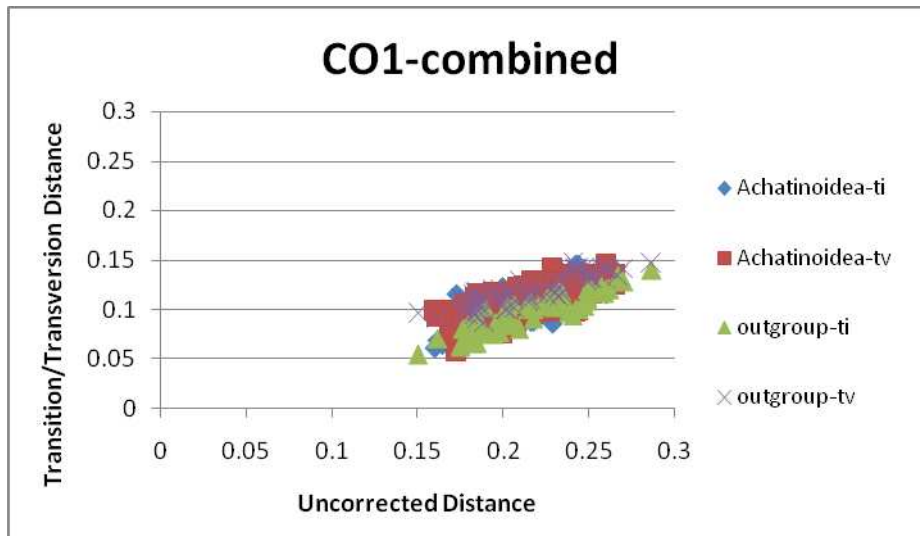


(B)

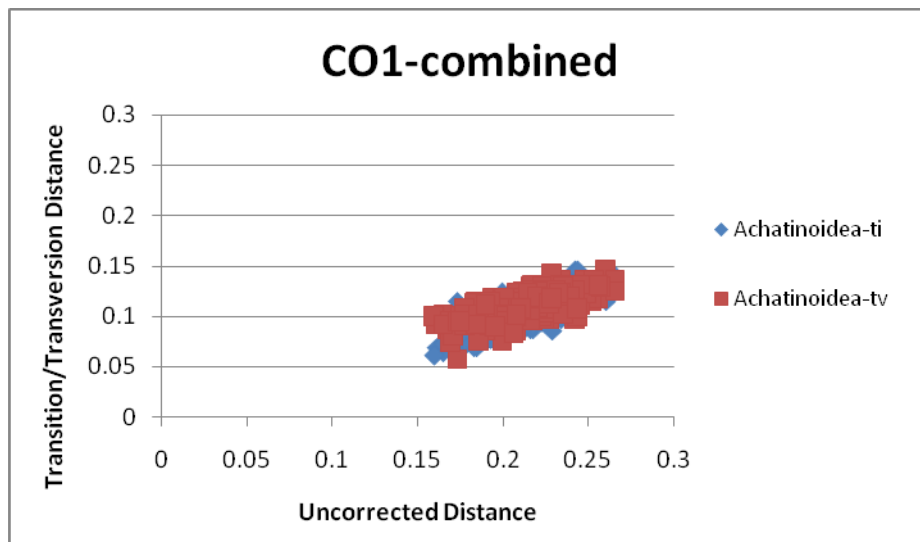
Figure 3.17: Plots of pairwise uncorrected distance against corrected distance for the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.18) revealed transversions overlapping with transitions in both the Achatinoidea only and the Achatinoidea plus outgroup datasets, while Plot 3 (Figure 3.19) revealed that the majority of the pairwise comparisons had higher rates of transversions than transitions.

The $g1$ test score for the COI gene based on 24 taxa and 250 parsimony-informative sites was -0.142, still significantly larger than the critical value of -0.16 for 15 taxa and 250 parsimony-informative sites. The $g1$ test therefore suggested that no phylogenetic information could be generated from the dataset.

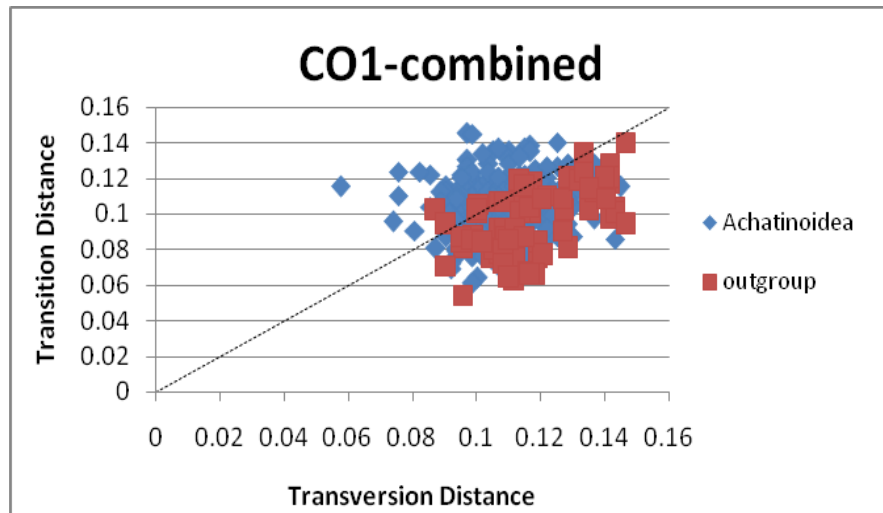


(A)

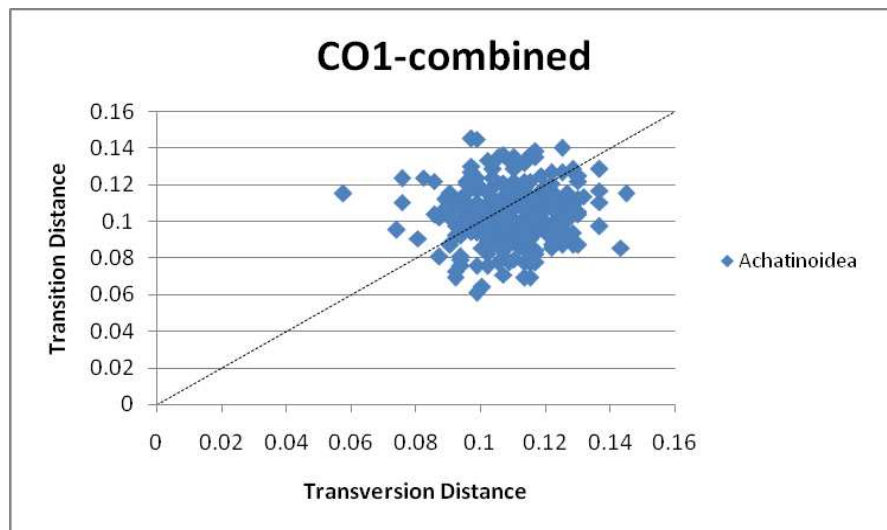


(B)

Figure 3.18: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

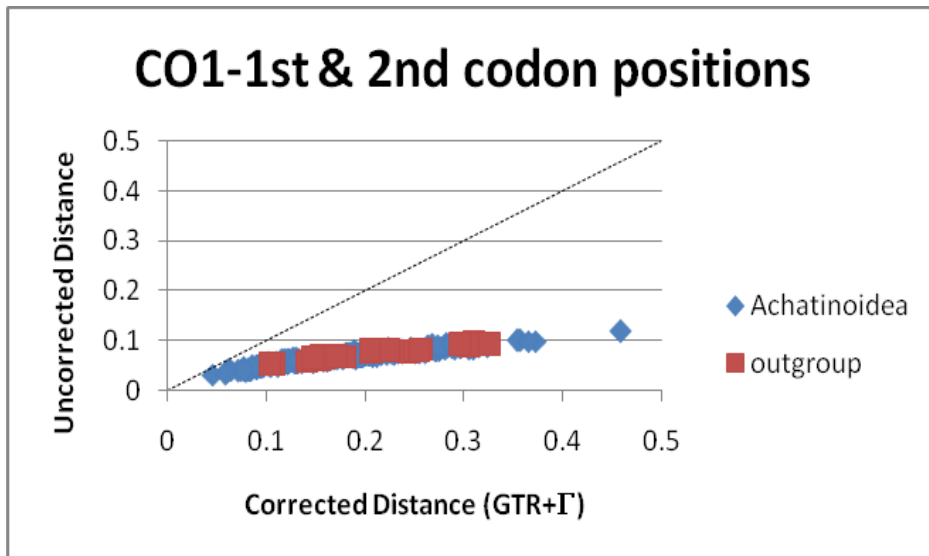


(B)

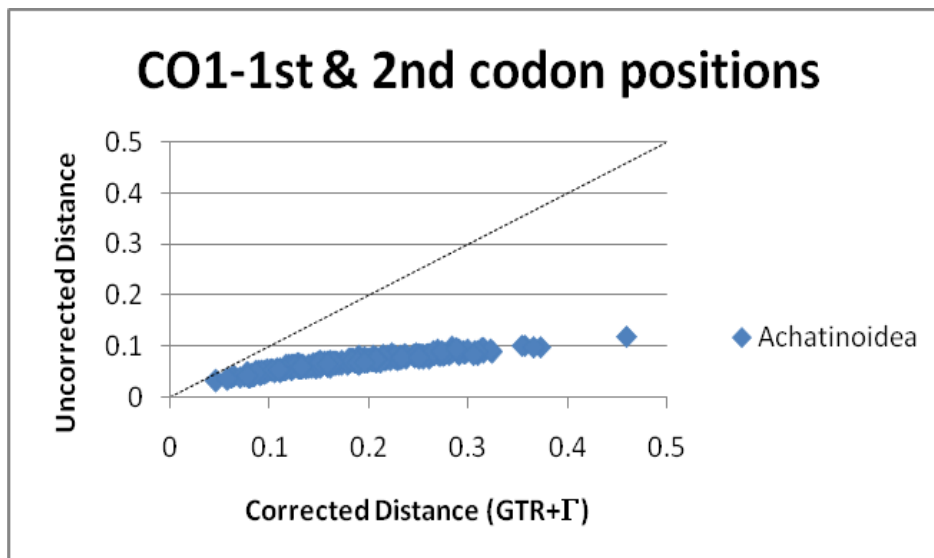
Figure 3.19: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the COI gene in **(A)** the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and **(B)** the Achatinoidea only.

The analyses of the COI gene as a whole (all codon positions) clearly demonstrated the existence of extensive saturation in the dataset, especially as shown by Plot 1. Its utility was further compromised by the absence of phylogenetic signal based on the g1 test. In order to evaluate whether this saturation was restricted to the 3rd codon position and whether some phylogenetic signal could be recovered from the 1st and 2nd codon positions, the combined 1st and 2nd codon positions and the 3rd codon position were evaluated separately.

For the 1st and 2nd codon positions of the COI gene, Plot 1 (Figure 3.20) revealed a curve for the corrected distances using the optimal GTR+ Γ model, with the deviation from linearity beginning at an uncorrected (p) distance of approximately 0.03 for both the Achatinoidea only and including the outgroup taxon. The plots were still increasing slightly and had not reached a plateau, implying that the 1st and 2nd codon positions of COI had not yet reached saturation.



(A)

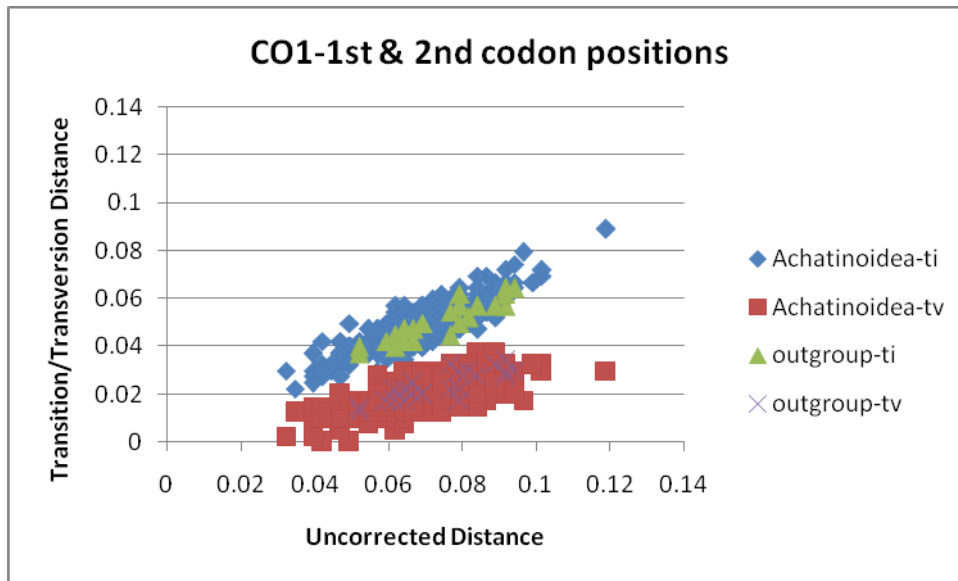


(B)

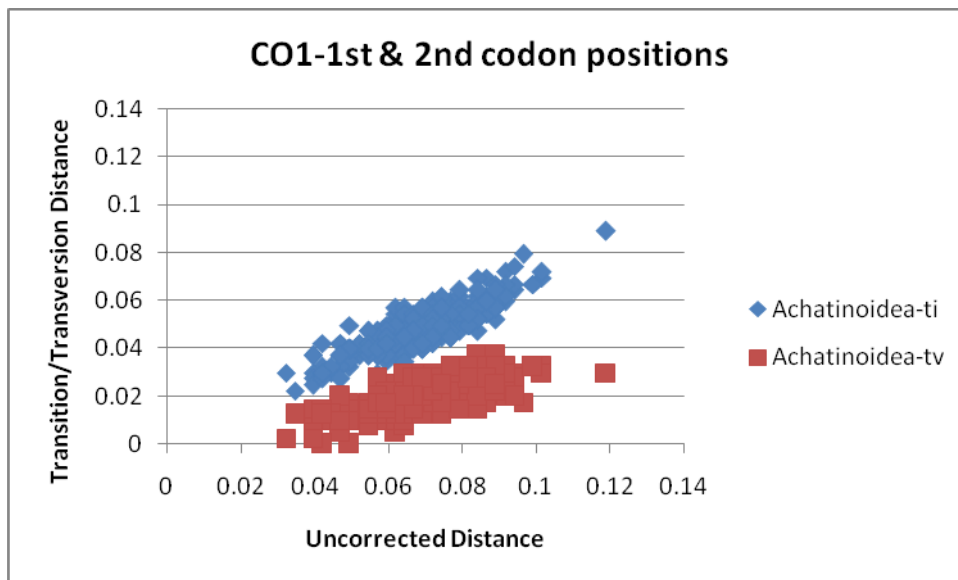
Figure 3.20: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 1st and 2nd codon positions of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.21) for the 1st and 2nd codon positions of the COI gene showed a linear increase of both transitions and transversions for the Achatinoidea and when the outgroup taxon was included, with the transitions being higher than the transversions. Plot 3 (Figure 3.22) also demonstrated the same conclusions as Plot 2. These findings suggested that the 1st and 2nd codon positions were not saturated.

The g1 value based on 24 taxa and 62 parsimony-informative sites was -0.283, which was significantly smaller than the critical value of -0.16 based on 15 taxa and 50 parsimony-informative sites. The 1st and 2nd codon positions of the COI gene therefore exhibited phylogenetic signal.

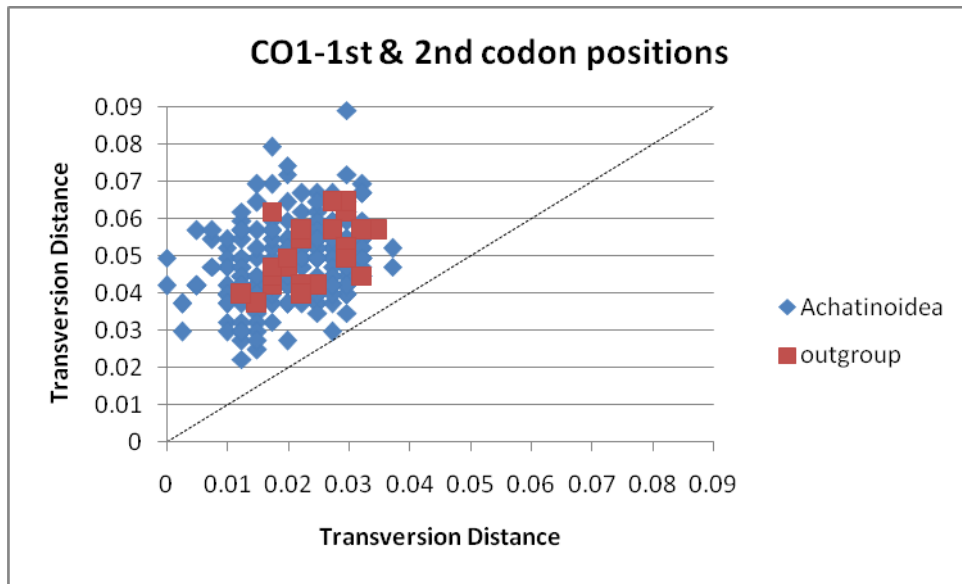


(A)

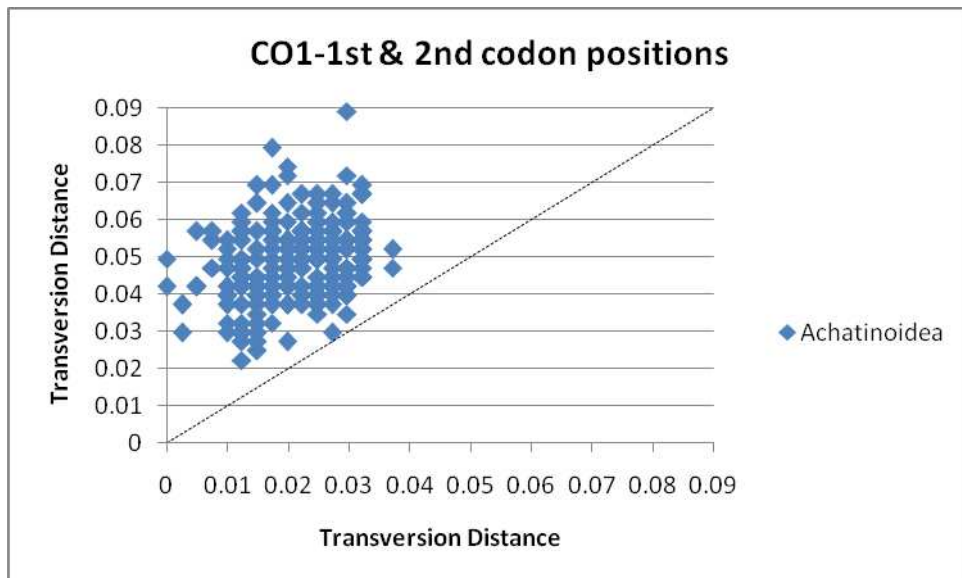


(B)

Figure 3.21: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1st and 2nd codon positions of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

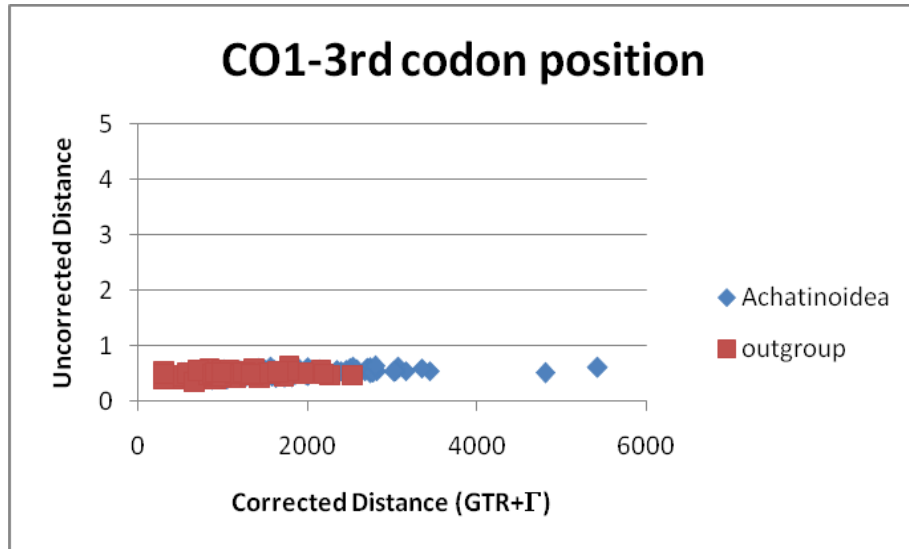


(B)

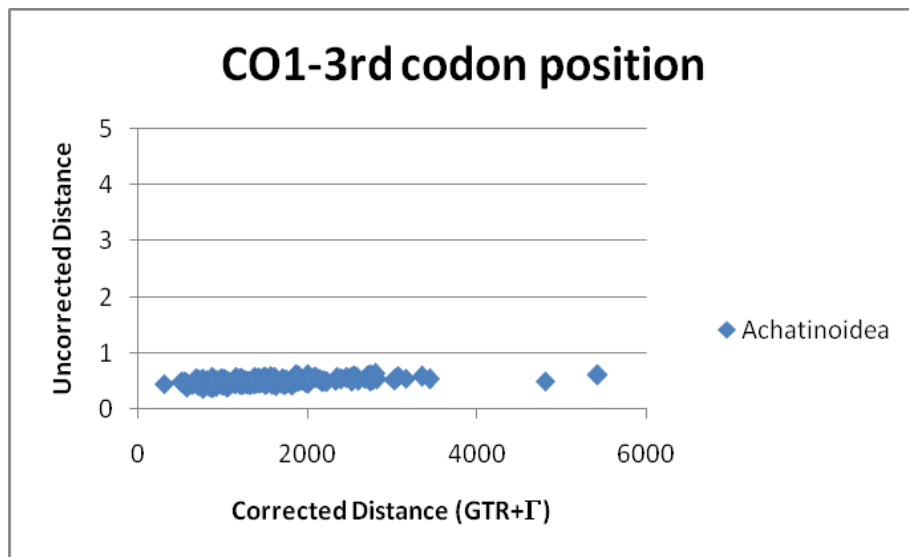
Figure 3.22: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1st and 2nd codon positions of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

For the 3rd codon position of the COI gene, Plot 1 (Figure 3.23) revealed that the optimal GTR+ Γ model was unable to correct the dataset for multiple hits as demonstrated by the nearly horizontal trend for both plots. The uncorrected distances

also ranged from 0.365 to 0.631 while corrected distances ranged from 299.377 to 5420.470 (see also Table 3.3). This indicated severe saturation for the 3rd codon position.



(A)

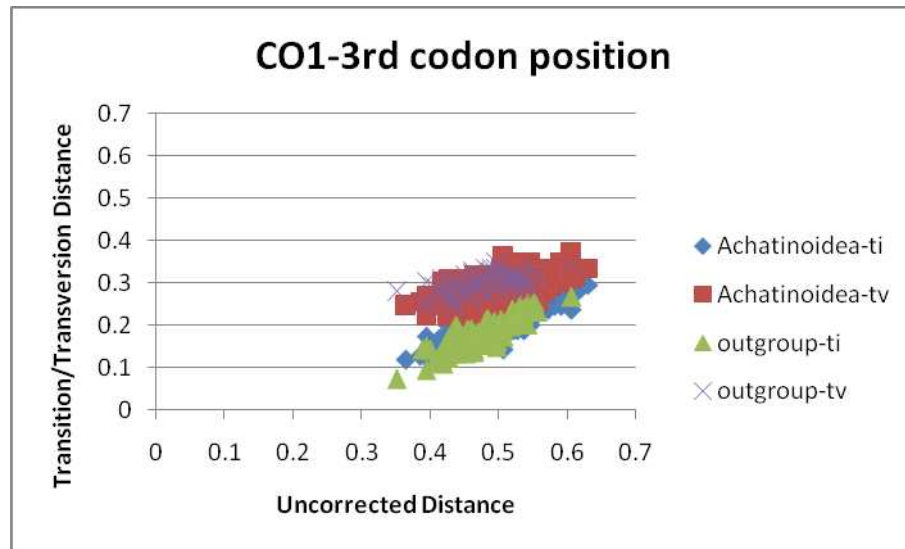


(B)

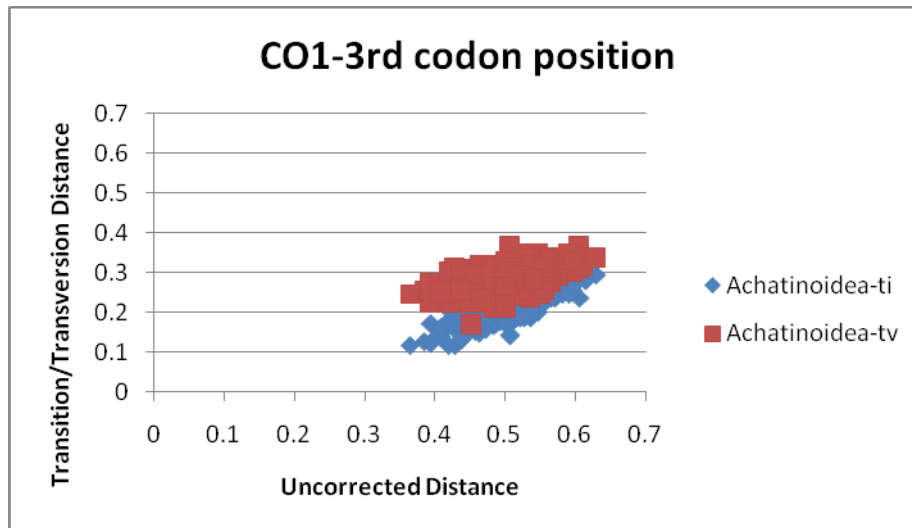
Figure 3.23: Plots of pairwise uncorrected distance against corrected distance for the 3rd codon position of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

Plot 2 (Figure 3.24) for all taxa as well as for the Achatinoidea only showed transversions overlapping with transitions, with the transversions generally higher than the transitions. Plot 3 (Figure 3.25) demonstrated that most of the pairwise comparisons had higher rates of transversions than transitions.

The $g1$ test score for the 3rd codon position of the COI gene based on 24 taxa and 188 parsimony-informative sites was -0.050, which was significantly larger than the critical value of -0.16 for 15 taxa and 100 parsimony-informative sites. The 3rd codon position therefore did not exhibit any phylogenetic signal.

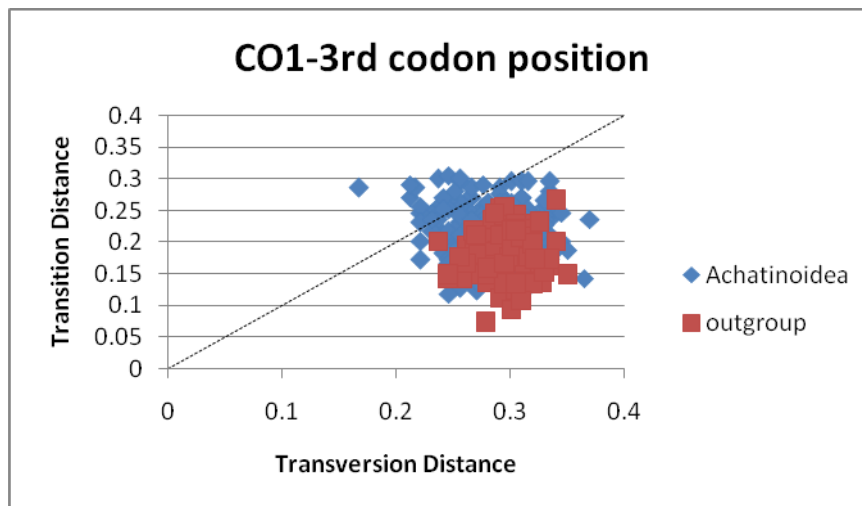


(A)

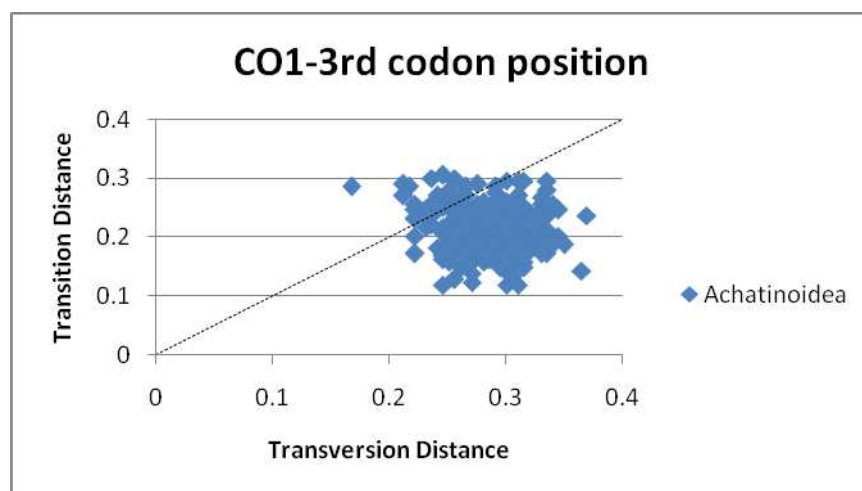


(B)

Figure 3.24: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3rd codon position of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.



(A)

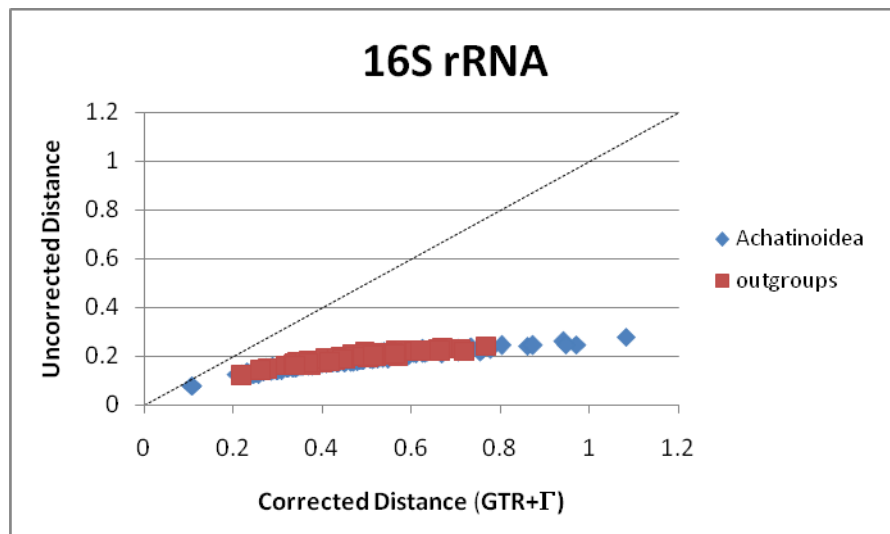


(B)

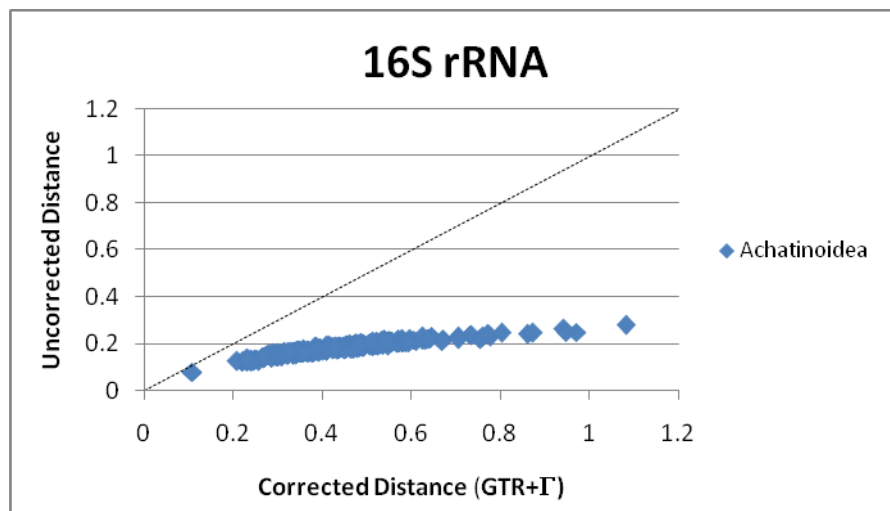
Figure 3.25: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3rd codon position of the COI gene in (A) the Achatinoidea and the streptaxid outgroup taxon *Gibbulinella dewinteri* and (B) the Achatinoidea only.

The lack of phylogenetic signal (as indicated by the g1 test) and the strong evidence for saturation of the entire COI gene suggested that the gene should not be used in its entirety for phylogenetic analyses. Further analysis revealed that the saturation is entirely due to the 3rd codon position. Phylogenetic analyses for the COI gene were therefore limited to the 1st and 2nd codon positions only.

For the 16S rRNA gene, Plot 1 (Figure 3.26) produced corrected distances based on the optimal GTR+ Γ model that curved from linearity at an uncorrected (p) distance of 0.08 or below and approached a plateau at an uncorrected distance of approximately 0.3. One corrected distance even exceeded the value of 1. These findings suggest the possibility of saturation in the 16S dataset.



(A)



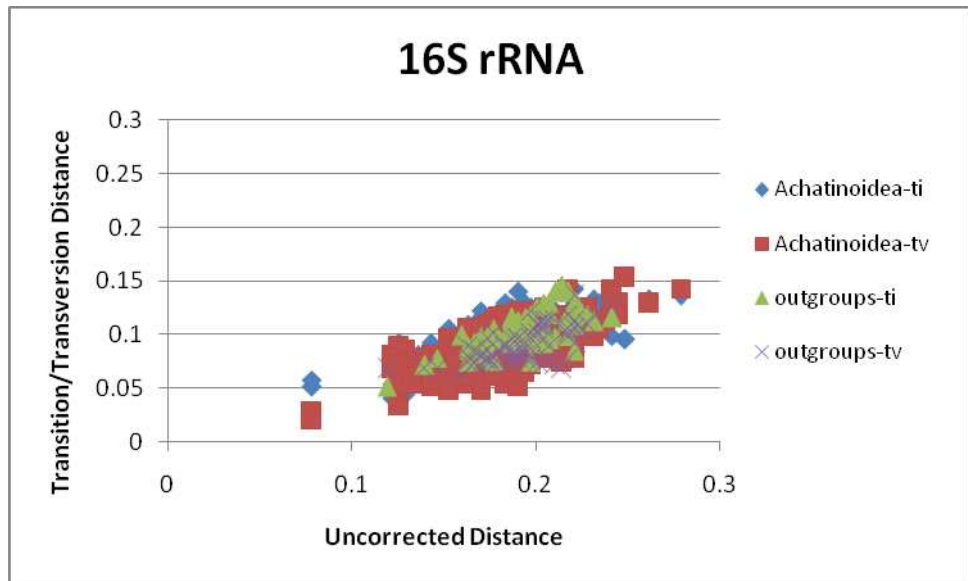
(B)

Figure 3.26: Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the 16S rRNA gene in (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

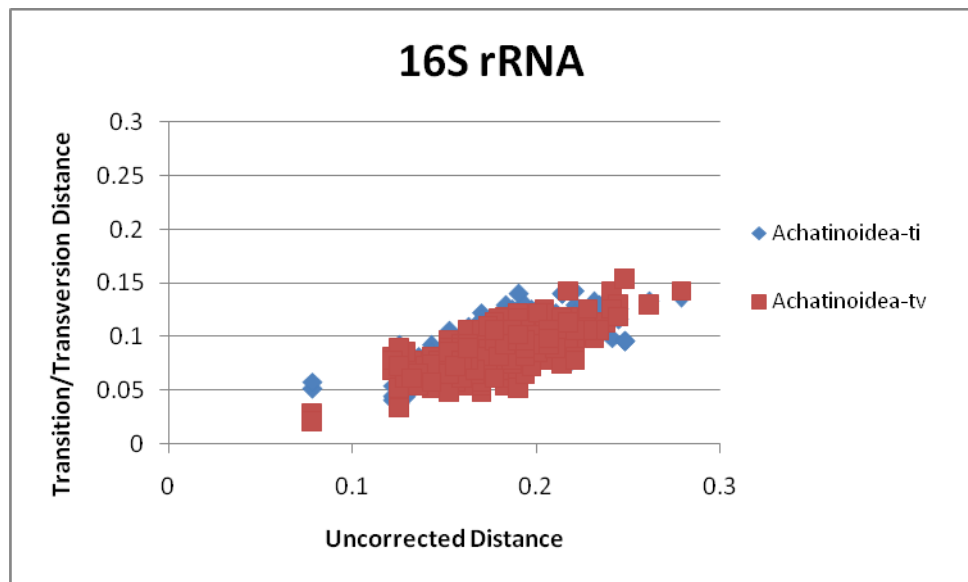
Plot 2 (Figure 3.27) yielded a transition line that began to curve at higher distances where it was overtaken by the transversion line that continued to increase, thus indicative of saturation. An unmistakable overlap was also seen between transitions and transversions. Plot 3 (Figure 3.28) revealed that approximately half of all pairwise comparisons had higher rates of transversions than transitions.

The $g1$ value based on 25 taxa and 120 parsimony-informative sites was -0.437, which was significantly smaller than the critical value of -0.1 at $p=0.05$ for 25 taxa and 100 parsimony-informative sites. Phylogenetic signal was therefore evident in the 16S rRNA gene.

Similarly to the COI gene, there was evidence of saturation in the 16S rRNA dataset. This suggested that the gene should not be used for phylogenetic analyses, despite apparently exhibiting some phylogenetic signal based on the $g1$ test.

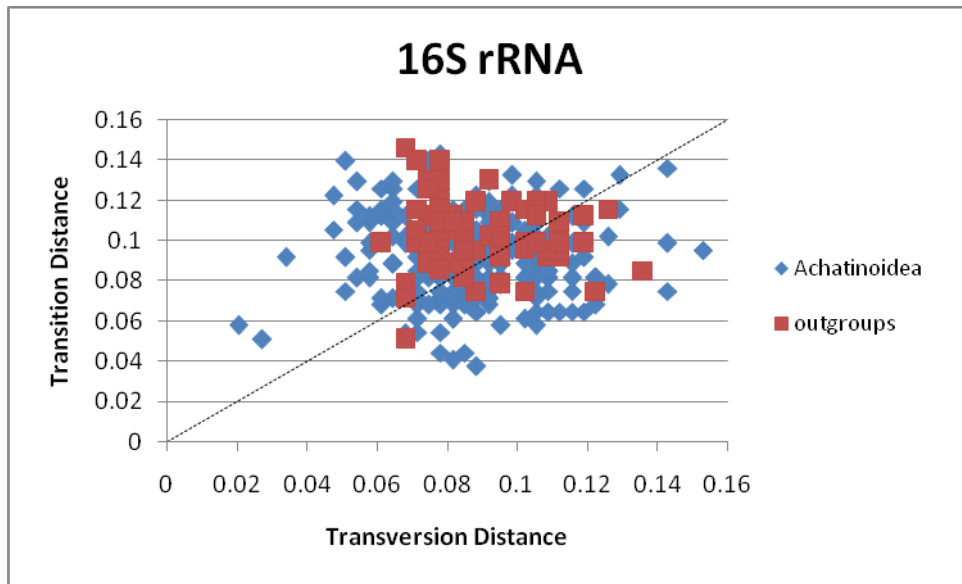


(A)

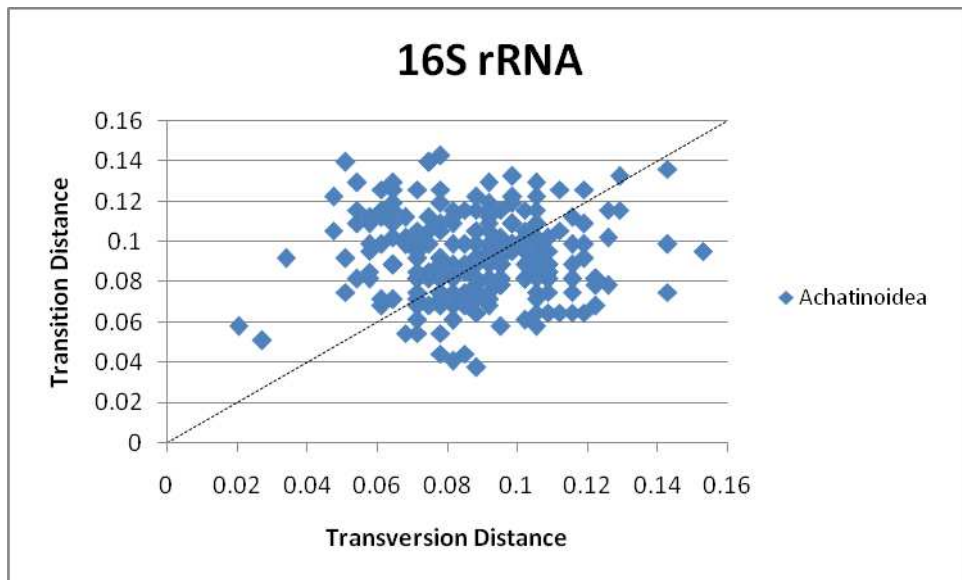


(B)

Figure 3.27: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 16S rRNA gene in **(A)** the Achatinoidea and the streptaxid outgroup taxa and **(B)** the Achatinoidea only.



(A)



(B)

Figure 3.28: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances (A) the Achatinoidea and the streptaxid outgroup taxa and (B) the Achatinoidea only.

3.3.2.2. Inferring the presence of paralogous sequences in the actin dataset and assessing its utility for phylogenetic analysis

To infer the presence of paralogous sequences, the actin sequences (obtained by direct sequencing of PCR products) were examined for the presence of ambiguous sites. Ambiguous sites were detected in 17 out of 24 achatinoid taxa and all three streptaxid outgroups (Appendix 3.7, p. 434), indicating the presence of more than one copy of the actin gene in the Achatinoidea and the Streptaxoidea.

Clones derived from three representative taxa, which had varied numbers of ambiguous sites in their direct sequences (*Coeliaxis blandii*, 21 ambiguities; *Cochlitoma granulata*, 9 ambiguities; *Leptinaria lamellata*, 0 ambiguities), were examined to assess the ability of the PCR-direct sequences to detect variation among actin copies as ambiguities in the direct sequence. (See details of actin clones in Appendix 3.8, pp. 435-437). The three taxa were selected to represent those with many, few and no ambiguous sites, respectively and were chosen from a batch of actin sequences processed early on, with their cloned sequences used as guides to assign ambiguities in direct sequences. Other actin sequences obtained later yielded considerably more ambiguous sites after subsequent processing, as in the case of *Bocageia* sp. with 101 ambiguous sites. Although the sequences of the clones would be subject to Taq error, a reasonably close correlation would be expected between the amount of variation detected among clones and the amount of ambiguity detected in the direct sequences. However, for *C. blandii*, a total of 163 variable sites were detected among the 4 clones obtained, as opposed to only 21 ambiguities in the direct sequence (Appendix 3.8A, p. 435). Of these variable sites, 132 were attributed to a single clone (Clone 3) that was clearly not picked up in the direct sequence (see Appendix 3.8B-1, p. 436). Likewise for *C. granulata*, 72 variable sites were detected among the 7 clones obtained as

opposed to only 9 ambiguities in the direct sequence (Appendix 3.8A). The majority of these variable sites (55) were attributed to Clones 3 and 6 that were not picked up in the direct sequence (see Appendix 3.8B-2, p. 437). For *L. lamellata*, six clones were obtained, with three clones (Clones 1, 3 and 5) bearing sequences identical to the direct sequence (Appendix 3.8A). Clones 2 and 4 had one variable site each while Clone 6 had two; none of the four variable sites were picked up in the direct sequence (see Appendix 3.8B-3, p. 437). The very small number of variable sites in the *L. lamellata* clones and their absence in the direct sequence would suggest that these variable sites were probably due to Taq sequencing error. The cloned sequences therefore confirmed the presence of multiple actin genes in at least some achatinoid taxa, with some clones exhibiting highly divergent sequences when compared to other clones derived from the same taxon. Moreover, not all of the variation observed in the clones was detected in the PCR-direct sequences. The reason for the failure of the direct sequences to identify all of the variation among the clones is not clear. One explanation may lie in the fact that the PCR-direct sequence is effectively a consensus of the different actin copies within an individual in which rare copies of the gene might reasonably be expected to be averaged out.

To assess the utility of the actin gene for phylogenetic analysis of the Achatinoidea, four approaches were undertaken to determine whether the observed paralogy in actin was likely to mislead phylogeny. First, a neighbor-joining tree was constructed for all actin sequences for the Achatinoidea as well as all clones obtained for *C. blandii* (4 clones), *C. granulata* (7 clones) and *L. lamellata* (6 clones) in order to determine whether the clones for each species were monophyletic. Monophyly would suggest that the gene duplication event that led to the actin paralogs detected was recent relative to the date of species divergence. Moreover, if all copies of the gene were

monophyletic, the organismal phylogeny would not be misled. However, monophyly was not observed for the clones of either *C. blandii* or *C. granulata*, with the most divergent clones (Clone 3 for *C. blandii* and Clones 3 and 6 for *C. granulata*) falling separately from both the PCR direct sequence and the other clones from the species (see Appendix 3.9, p. 438). This suggested a high probability that the organismal phylogeny may be misled. Nevertheless, the most divergent sequences for *C. blandii* and *C. granulata* (that fell separately from the other *C. blandii* and *C. granulata* sequences in the neighbor-joining tree) were not represented as ambiguities in the PCR-direct sequences and would therefore not affect the phylogenetic signal of the direct sequences.

Second, the number of ambiguous sites within a taxon was compared with the total number of nucleotide differences between the taxon and its closest relative (based on actin sequence identity). The number of ambiguous sites within a taxon should not exceed the number of differences between this taxon and its closest relative if the time of divergence between the two taxa were to be earlier than the evolution of the different actin genes within these taxa. With the exception of *Pyrgina umbilicata* and *Thyrophorella thomensis*, a lower number of ambiguous sites was observed within each taxon when compared to the number of nucleotide differences between the taxon and its closest relative. Thus, based on the number of ambiguous sites observed from the direct sequences, in most cases the divergence among taxa was deeper than the divergence among the different copies of the actin gene, suggesting that some phylogenetic signal could be derived from actin at and above the genus level (see Table 3.4). However, the presence of divergent actin gene sequences falling deeper than the divergence among taxa should not be ruled out, as some divergent sequences, which were not picked up as ambiguities in the direct sequences, were detected by cloning; these sequences did not

cluster with the other clones or the direct sequence from that species (i.e. *Coelioxys blandii* and *Cochlitoma granulata*). It should also be noted that since the closest relative was identified based on actin sequence identity, such a relationship was not always found to be associative. For instance, *Achatina achatina* is most similar to *A. fulica* with 65 nucleotide differences, and yet the latter is most similar to *C. ustulata* with only 46 differences.

Third, a partition homogeneity test (see Section 2.9.10, p. 86-88) was undertaken to determine whether or not the sequences from the actin gene exhibited a significant difference in terms of phylogenetic signal compared to the other genes being evaluated (see results in Section 3.3.2.3, p. 161-165). The test revealed that the actin dataset was not too divergent in terms of its evolutionary history relative to the other datasets and that the presence of multiple copies of the actin gene was not having a significant effect on phylogeny.

Lastly, the phylogenetic tree obtained from the actin dataset was checked for concordance with the phylogeny obtained from the rRNA cluster. The actin phylogeny showed concordance with the rRNA phylogeny with respect to many well-supported groups (see results in Section 3.3.3, pp. 162-163 and 169-171), suggesting that the presence of multiple copies of the actin gene was not having a significant effect on phylogeny.

Thus, despite serious reservations over the utility of the actin gene in phylogenetic analysis of the Achatinoidea, it seems that some useful phylogenetic signal could be gleaned from the gene. Actin was therefore utilised in phylogenetic analyses of the Achatinoidea though its shortcomings suggest that its findings should be interpreted with extreme caution.

Table 3.4. Comparison of the actin ambiguous sites and nucleotide differences for each achatinoid and its closest relative. The closest relative for each taxon is determined based on the actin sequence identity (least number of nucleotide differences). Note that in scoring for the total differences, comparison between an ambiguous site and an unambiguous site is counted as different even if one of the possible nucleotides for the ambiguous site is the same as that found in the unambiguous site being compared (e.g. A vs. R, which is either A or G). See also Table 2.8 of p. 73 for assignment of ambiguous positions.

Species (total ambiguous sites)	Closest relative (total ambiguous sites)	Total number of nucleotide differences between each species and its closest relative
ACHATINIDAE		
<i>Achatina achatina</i> (45)	<i>Achatina fulica</i> (18)	65
<i>Achatina fulica</i> (18)	<i>Cochlitoma ustulata</i> (0)	46
<i>Achatina stuhlmanni</i> (2)	<i>Cochlitoma ustulata</i> (0)	28
<i>Cochlitoma ustulata</i> (0)	<i>Achatina stuhlmanni</i> (2)	28
SUBULINIDAE		
<i>Allopeas clavulinum</i> (0)	<i>Eutomopeas layardi</i> (0)	36
<i>Bocageia</i> sp. (101)	<i>Rumina decollata</i> (29)	126
<i>Eutomopeas layardi</i> (0)	<i>Allopeas clavulinum</i> (0)	36
<i>Leptimnaria lamellata</i> (0)	<i>Allopeas clavulinum</i> (0)	104
<i>Paropeas clavulinum</i> (1)	<i>Allopeas clavulinum</i> (0)	46
<i>Riebeckia</i> sp. (76)	<i>Coelioxys blandii</i> (21)	107
<i>Rumina decollata</i> (29)	<i>Zootecus insularis</i> (20)	55
<i>Subulina octona</i> (0)	<i>Subulina striatella</i> (26)	63
<i>Subulina striatella</i> (26)	<i>Subulina octona</i> (0)	63
<i>Subulina vitrea</i> (37)	<i>Coelioxys blandii</i> (21)	87
<i>Subulona</i> sp. (0)	<i>Achatina stuhlmanni</i> (2) <i>Cochlitoma ustulata</i> (0)	117
<i>Tortaxis erectus</i> (32)	<i>Rumina decollata</i> (29) <i>Zootecus insularis</i> (20)	110
<i>Xerocerastus</i> sp. (0)	<i>Zootecus insularis</i> (20)	60
<i>Zootecus insularis</i> (20)	<i>Rumina decollata</i> (29)	55
COELIAXIDAE		
<i>Coelioxys blandii</i> (21)	<i>Zootecus insularis</i> (20)	82
<i>Pyrgina umbilicata</i> (88)	<i>Thyrophorella thomensis</i> (62)	77
THYROPHORELLIDAE		
<i>Thyrophorella thomensis</i> (62)	<i>Pyrgina umbilicata</i> (88)	77
GLESSULIDAE		
<i>Glessula ceylanica</i> (53)	<i>Cochlitoma ustulata</i> (0)	149
FERUSSACIIDAE		
<i>Ceciloides gokweanus</i> (4)	<i>Xerocerastus</i> sp. (0)	81
<i>Ferussacia folliculus</i> (4)	<i>Xerocerastus</i> sp. (0)	99
STREPTAXIDAE (OUTGROUP)		
<i>Gibbulinella dewinteri</i> (6)	<i>Gonaxis quadrilateralis</i> (55)	94
<i>Gonaxis quadrilateralis</i> (55)	<i>Gibbulinella dewinteri</i> (6)	94
<i>Gonospira</i> sp. (77)	<i>Gonaxis quadrilateralis</i> (55)	117

3.3.2.3. Partition homogeneity test for combining datasets

A partition homogeneity test of the rRNA cluster, actin and histone 3 datasets and the 1st and 2nd codon positions of the COI gene (5028 nucleotides total) for all taxa

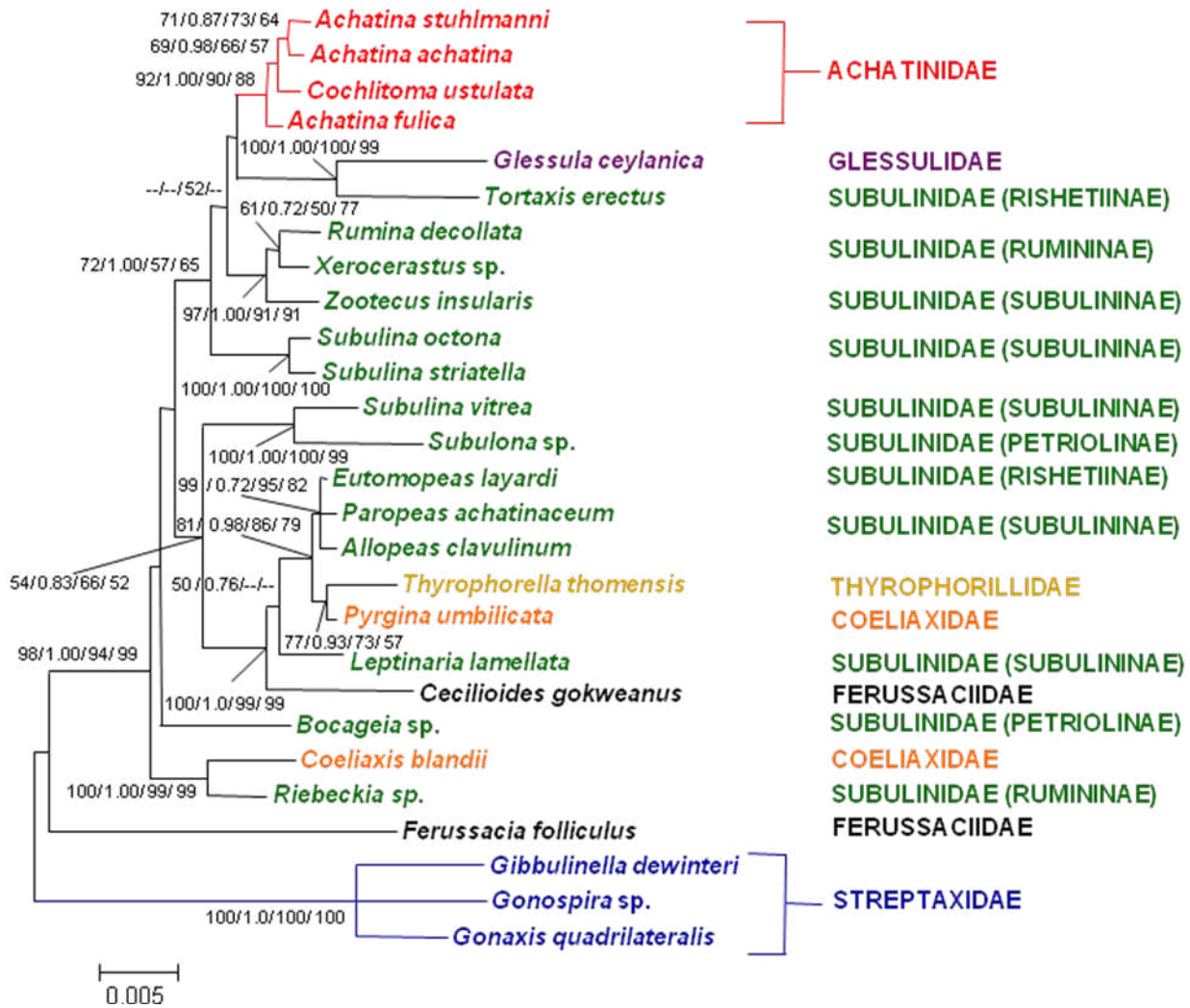
with complete DNA sequences yielded a *P* value of 0.01 (note that the 16S rRNA gene and the 3rd codon position of the COI gene were excluded due to saturation (see Section 3.3.2.1, pp. 138-154). This would indicate that the genes under consideration, including the actin, exhibited similar evolutionary histories (Cunningham, 1997). The datasets from the four genes could therefore be combined as a single dataset.

3.3.3. Molecular phylogeny

The maximum likelihood (ML) phylogenies for the rRNA cluster, actin, H3 and COI (1st and 2nd codon positions only) are shown in Figure 3.29 and those of the concatenated sequences from the four genes in Figure 3.30. Note that for the concatenated sequences, two sets of phylogenetic analyses were carried out. In the first, only taxa with sequences from all four genes were used in the phylogeny; thus, the achatinoids *Celilioides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp. were excluded (Figure 3.30A). In the second, all taxa were included in the analyses (Figure 3.30B), with missing data assigned in the analyses for taxa with incomplete datasets. Also shown in the phylogenies are the bootstrap support values for ML, neighbor-joining (NJ) and maximum parsimony (MP) analyses and the posterior probabilities for Bayesian inference (BI) analysis. The BI, NJ and MP phylogenies for the genes and the concatenated sequences are shown in Appendices 3.10-3.15, pp. 439-450.

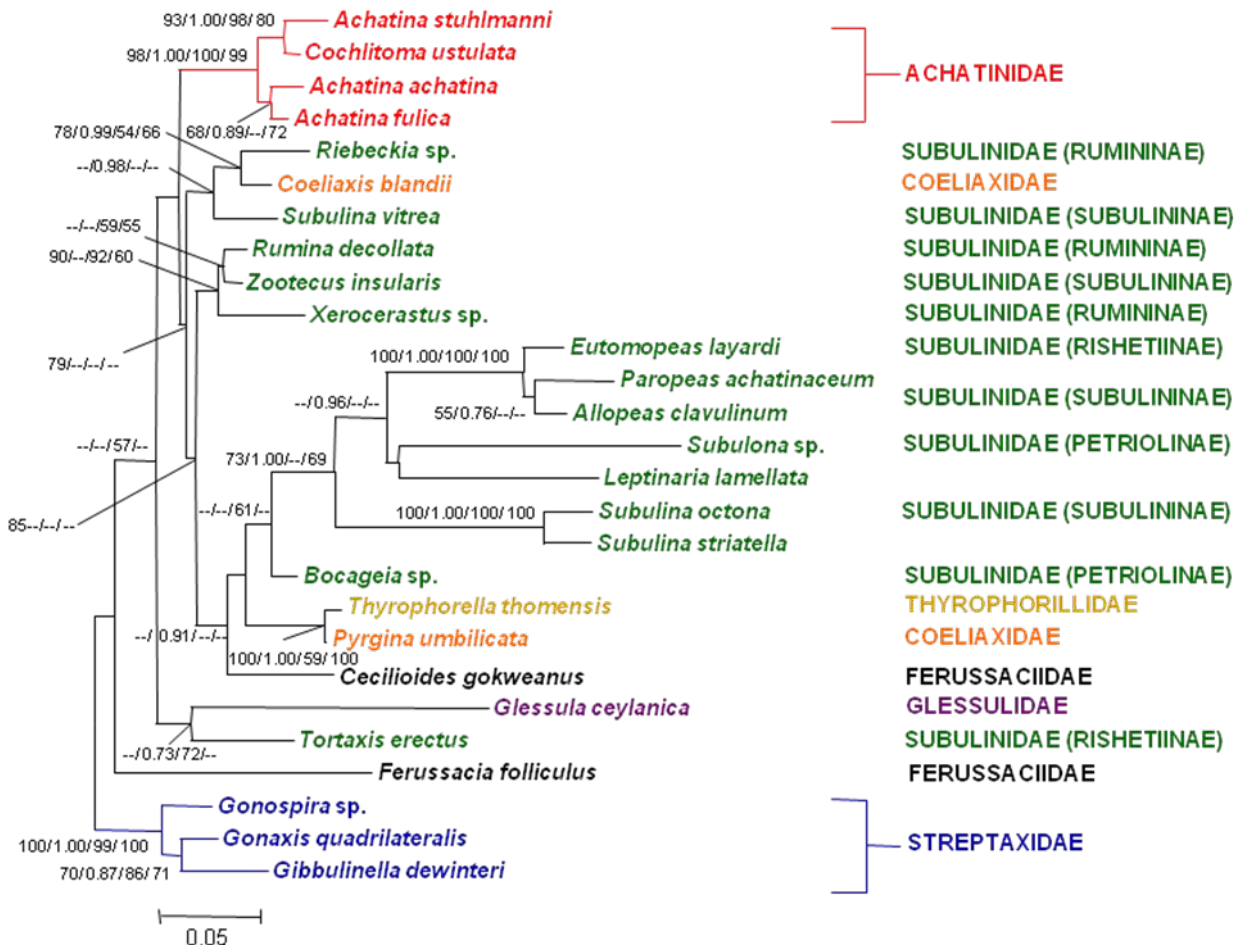
Figure 3.29: (See next four pages). Maximum likelihood phylogenetic trees of the Achatinoidea based on (A) the rRNA cluster (3435 nucleotides); (B) the actin gene (861 nucleotides); (C) the histone 3 gene (328 nucleotides) and (D) the 1st and 2nd codon positions of the COI gene (404 nucleotides). The phylogenies were rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp for the rRNA and actin datasets and *Gibulinella dewinteri* for the histone 3 and COI datasets. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap support (1000 replicates) for NJ and MP, respectively. Bootstrap support less than 50% and posterior probabilities less than 0.7 are not shown. For BI, the optimized number of generations to explore the tree space was 2,000,000 for all the genes; the optimized heating temperatures for each gene were as follows: rRNA cluster-0.1; actin-0.1; histone 3-0.1; 1st and 2nd codon positions of COI-0.075. The scale bar for (A) represents 5 substitutional changes per 1000 nucleotide positions; those of (B), (C) and (D) represent 5 substitutional changes per 100 nucleotide positions.

Figure 3.29 (contd.)



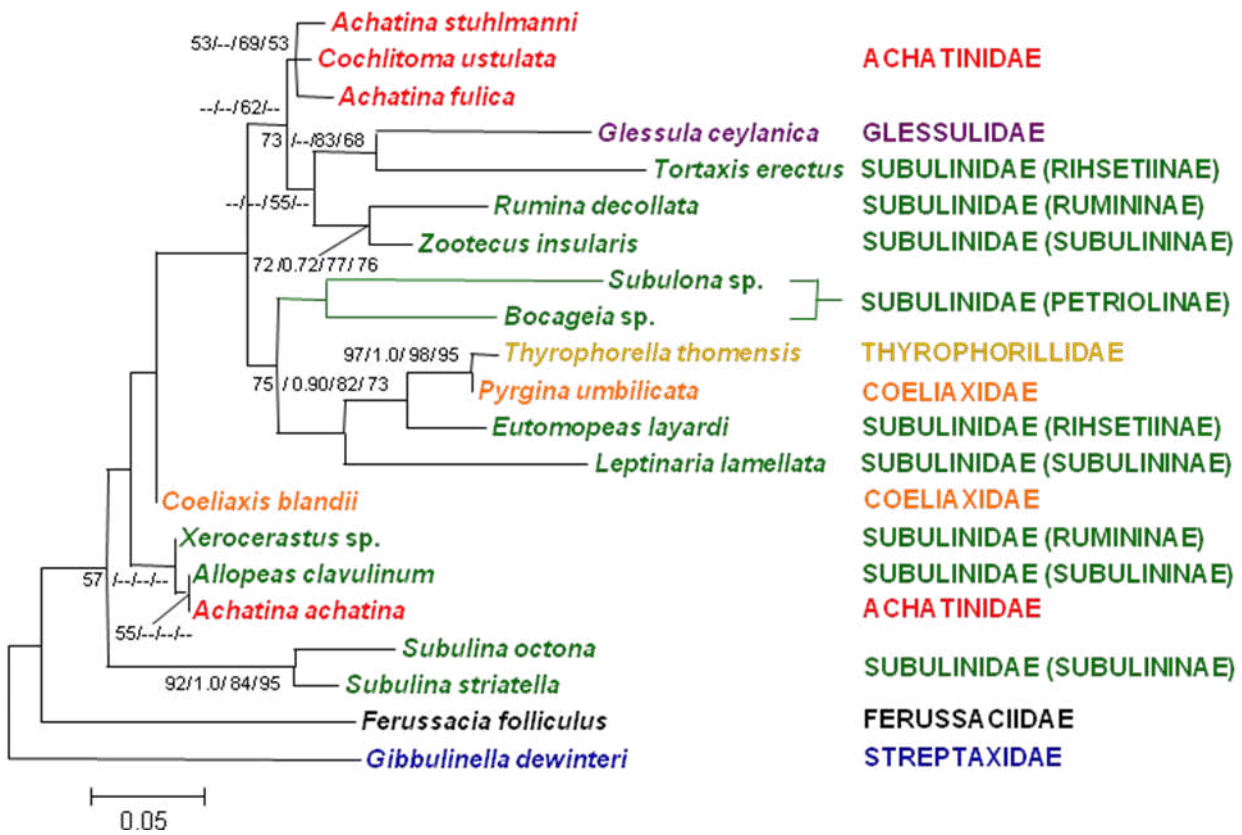
(A)– rRNA cluster

Figure 3.29 (contd.)



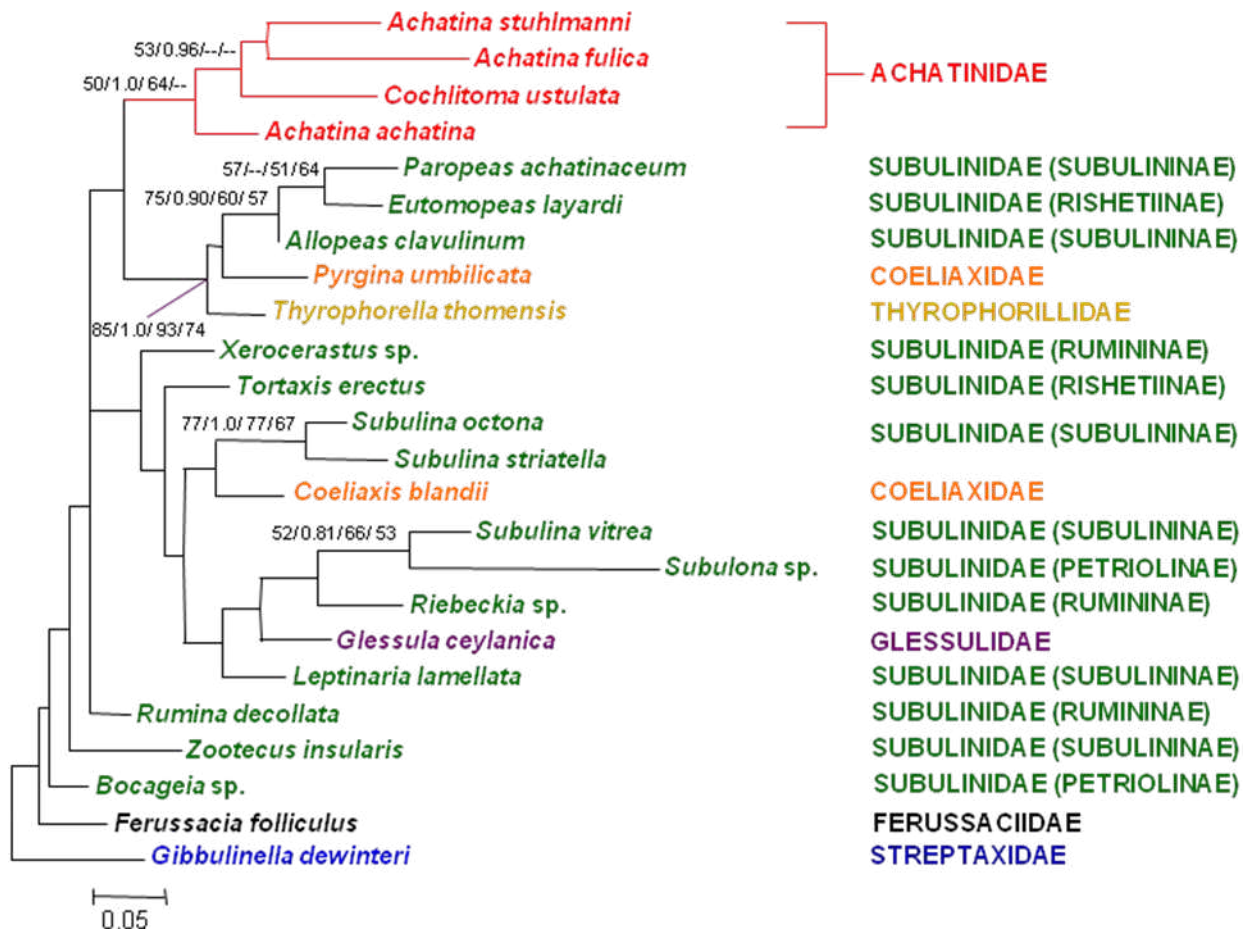
(B) – Actin

Figure 3.29 (contd.)



(C)–H3

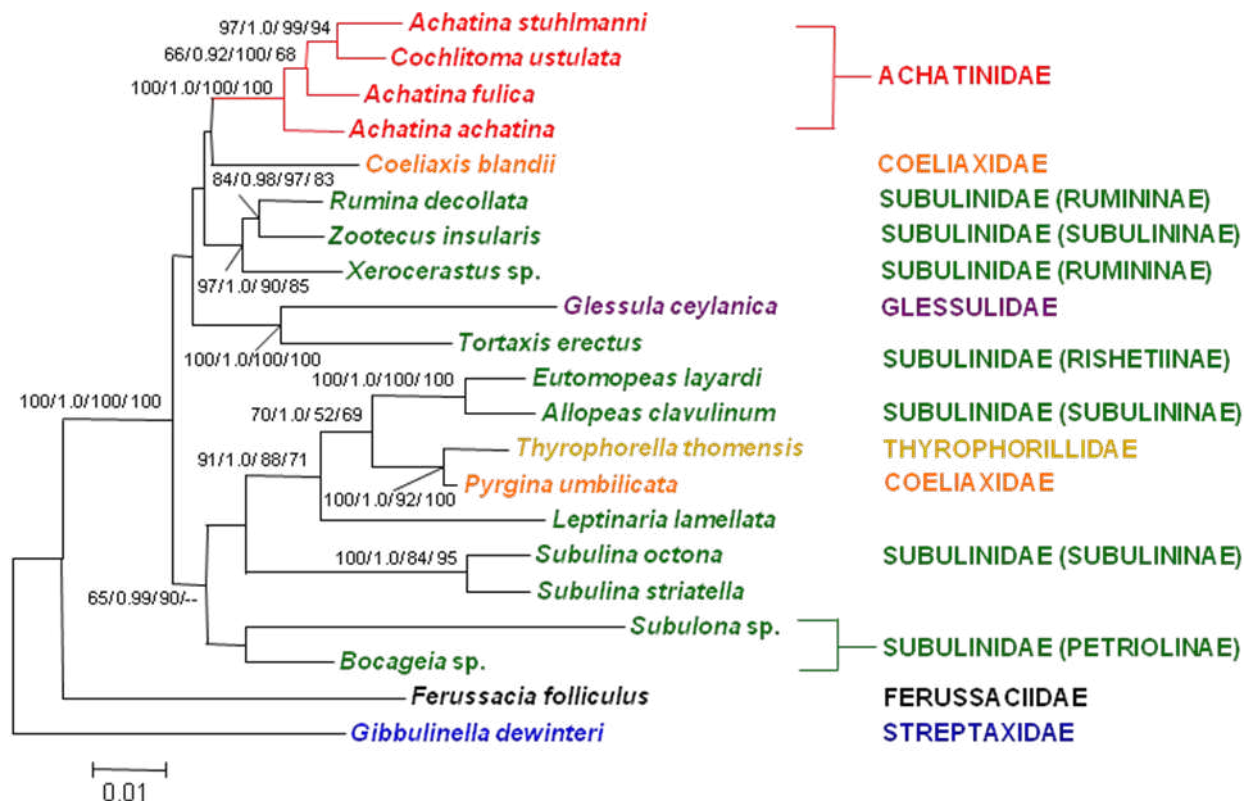
Figure 3.29 (contd.)



(D)– COI-1st and 2nd codon positions

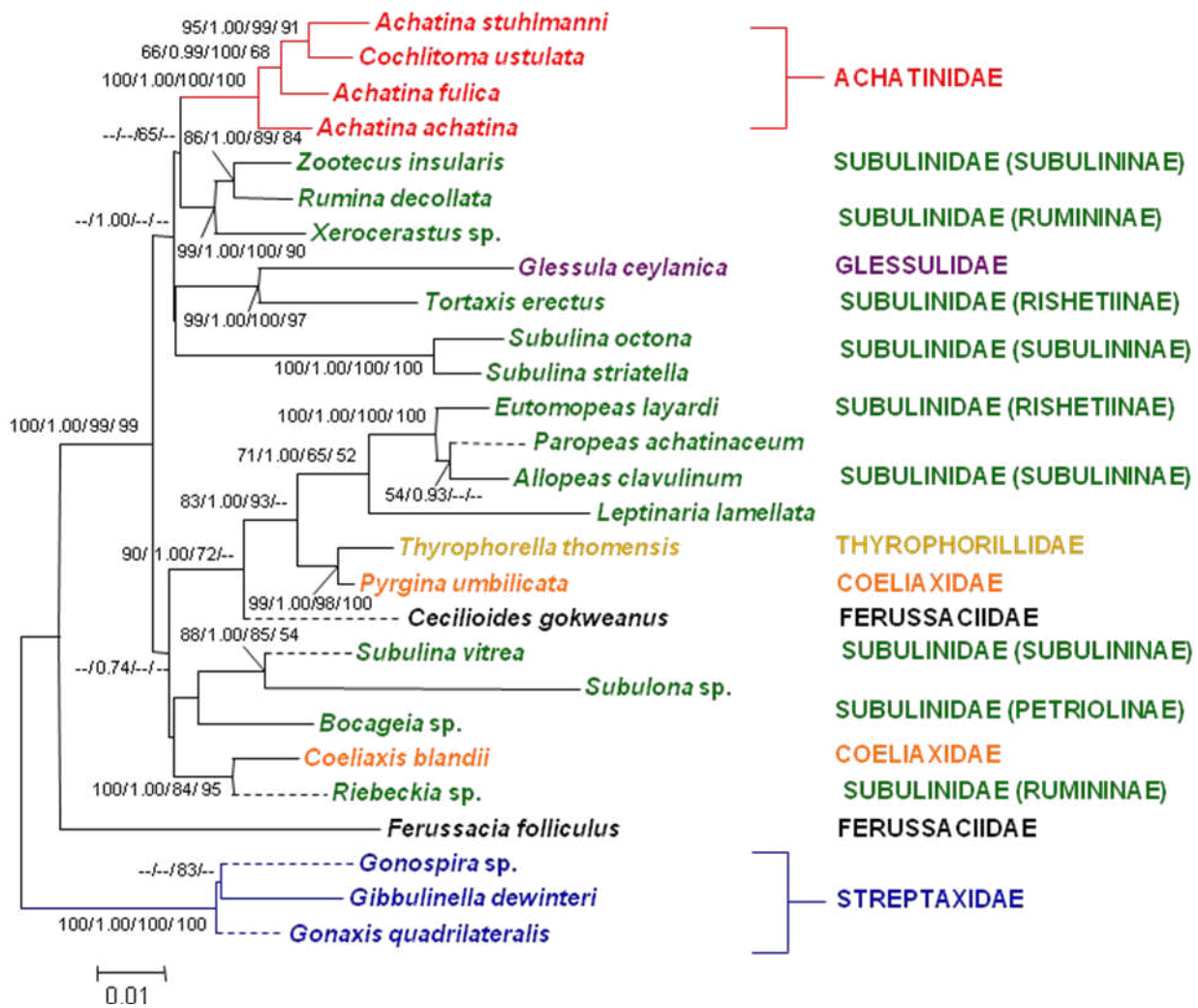
Figure 3.30: (See next two pages.) Maximum likelihood phylogenetic trees of the Achatinoidea based on a concatenated sequence of 5028 nucleotides derived from the combined dataset of the rRNA cluster, actin and H3 genes as well as the 1st and 2nd codon positions of the COI gene. The phylogeny in **(A)** includes only those taxa which have complete sequence data for all sequence regions while that in **(B)** includes all taxa including some with missing data. The following taxa had missing DNA sequence data: *Cecilioides gokweanus* (Histone 3 and COI), *Paropeas achatinaceum* (Histone 3), *Riebeckia* sp. (Histone 3), *Subulina vitrea* (Histone 3), *Gonaxis quadrilateralis* (Histone 3 and COI) and *Gonospira* sp. (Histone 3 and COI). The phylogeny in **(A)** is rooted on the streptaxid *Gibbulinella dewinteri* while that in **(B)** is rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap support (1000 replicates) for NJ and MP, respectively. Bootstrap support less than 50% and posterior probabilities less than 0.7 are not shown. For BI, the optimized number of generations to explore the tree space was 2,000,000 while the optimized heating temperature was 0.125 for both trees. The scale bars represent 1 substitutional change per 100 nucleotides. Branches for taxa with missing data are marked with dashes.

Figure 3.30 (contd.)



(A) – taxa with complete DNA sequence data

Figure 3.30 (contd.)



(B) - all taxa

3.3.3.1. Individual gene phylogenies

The phylogenies based on the four genes yielded several consistent groups. Even with the incorporation of new taxa and the addition of an extra 2612 nucleotide sites, the topology of the Achatinoidea based on the nuclear rRNA cluster (Figure 3.29A) remained similar to that obtained by Wade *et al.* (2006) based on a partial 823 bp fragment of the rRNA cluster. Several well-supported groups were recovered in the rRNA phylogeny. The Achatinidae, which included *Achatina fulica*, was recovered as monophyletic, with strong support with all tree methods (92% ML bootstraps, $P=1.0$ BI, 90% NJ bootstraps and 88% MP bootstraps). The other achatinoid families were not recovered in the tree as expected, but six groupings with strong support were resolved. The first group incorporated the coeliacid *Coeliaxis blandii* and the subulinid *Riebeckia* sp. (100 ML bootstraps, $P=1.00$, 99% NJ bootstraps and 99% MP bootstraps). The second group included the thyrophorellid *Thyrophorella thomensis*, the coeliacid *Pyrgina umbilicata*, the subulinids *Leptinaria lamellata*, *Eutomopeas layardi*, *Paropeas achatinaceum* and *Allopeas clavulinun* and the ferussaciid *Cecilioides gokweanus* (100% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 99% MP bootstraps). Within this group, the two São Thomé taxa, the ‘operculate’ *T. thomensis* and the coeliacid *P. umbilicata*, clustered together (77% ML bootstraps, $P=0.93$ BI, 73% NJ bootstraps and 57% MP bootstraps) while *E. layardi*, *P. achatinaceum* and *A. clavulinum* formed another cluster (99% ML bootstraps, $P=0.72$, 95% NJ bootstraps and 82% MP bootstraps). The third group included the subulinids *Subulina vitrea* and *Subulona* sp. (100% ML bootstraps, $P=1.0$, 100% NJ bootstraps and 99% MP bootstraps). The fourth group consisted of another pair of subulinids, *Subulina octona* and *Subulina striatella* (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). The fifth group incorporated the subulinids *Xerocerastus* sp., *Rumina decollata* and

Zootecus insularis (97% ML bootstraps, $P=1.0$ BI, 91% NJ bootstraps and 91% MP bootstraps). The sixth group clustered together the subulinid *Tortaxis erectus* and the glessulid *Glessula ceylanica* (100 ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 99% MP bootstraps). Finally, the ferussacid *Ferussaccia folliculus* fell at the base of the Achatinoidea and was separated from the other ferussacid in the study, *Cecilioides gokweanus*.

Despite the limitations of the nuclear actin gene for phylogenetic analysis due to the presence of multiple copies of the gene within an individual, the actin phylogeny was very similar to the rRNA tree, with many strongly supported groups in the actin tree also recovered in the rRNA tree. This suggests that the presence of multiple copies of the actin gene had a minimal effect on the achatinoid phylogeny. There was, however, less structure in the actin tree when compared to the rRNA tree as shown by the lower or equivocal support in many of the internal branches (Figure 3.29B). *Ferussaccia folliculus* again fell at the base of the tree; however, this was only moderately supported by NJ (57% bootstraps). Other groups that were consistent with the rRNA cluster phylogeny included the Achatinidae, which again formed a monophyletic group (98% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 99% MP bootstraps); the *C. blandii* and *Riebeckia* sp. group (78% ML bootstraps, $P=0.99$ BI, 54% NJ bootstraps and 66% bootstraps); the São Thomé taxa *T. thomensis* and *P. umbilicata* (100 ML bootstraps, $P=1.0$ BI, 59% NJ bootstraps and 100% MP bootstraps); the *E. layardi*, *P. achatinaceum* and *A. clavulinum* group (100 ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps); the *S. octona* and *S. striatella* group (100 ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps); the *Xerocerastus* sp., *R. decollata* and *Z. insularis* group (90% ML

bootstraps, $P < 0.7$ BI, 92% NJ bootstraps and 60% MP bootstraps); and the *T. erectus* and *G. ceylanica* group (with only $P = 0.73$ BI and 72% NJ bootstraps as support).

The phylogeny based on the nuclear histone 3 gene (Figure 3.29C), which excluded the achatinoids *Cecilioides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilaterlis* and *Gonospira* sp., revealed some groupings that were consistent with the rRNA phylogeny but showed less structure and a general reduction of support for most groups when compared to either the rRNA or the actin gene phylogenies (as shown in Figure 3.30C). For instance, *Achatina achatina* did not cluster with the rest of the Achatinidae and instead grouped with the subulinids *Xerocerastus* sp. and *A. clavulinum*, although it only received 57% ML bootstrap support. Some groups were recovered, however, such as *T. thomensis* and *P. umbilicata* (97% ML bootstraps, $P = 1.0$ BI, 98% NJ bootstraps and 95% MP bootstraps), *S. octona* and *S. striatella* (92% ML bootstraps, $P = 1.0$ BI, 84% NJ bootstraps and 95% MP bootstraps), and *T. erectus* and *G. ceylanica* (73% ML bootstraps, 83% NJ bootstraps and 68% MP bootstraps).

The mitochondrial COI gene phylogeny for codon positions 1 and 2 (Figure 3.29D) likewise revealed some groups that were consistent with the rRNA phylogeny but showed less structure and support (see Figure 3.30D). These were the Achatinidae (50% ML bootstraps, $P = 1.0$ BI and 64% NJ bootstraps), *S. vitrea* and *Subulona* sp. (52% ML bootstraps, $P = 0.81$ BI, 66% NJ bootstraps and 53% MP bootstraps), *S. octona* and *S. striatella* (77% ML bootstraps, $P = 1.0$ BI, 77% NJ bootstraps and 67% MP bootstraps), and *E. layardi*, *P. achatinaceum* and *A. clavulinum* (75% ML bootstraps, $P = 0.90$ BI, 60% NJ bootstraps and 57% MP bootstraps). It should be noted that *T. thomensis* and *P. umbilicata* also clustered with these three taxa with strong support

(85% ML bootstraps, $P=1.0$ BI, 93% NJ bootstraps and 74% MP bootstraps) as with the rRNA phylogeny.

3.3.3.2. Combined gene phylogenies

Combining datasets from genes with different evolutionary rates and mechanisms of inheritance provides information at different levels of phylogeny and therefore provides signals both for the internal branches and the terminal branches (Graybeal, 1994). The combined analysis for the rRNA cluster, actin, H3, and the 1st and 2nd codon positions of COI, but excluding the four taxa that did not yield either or both H3 and COI sequences, recovered most groups identified in the rRNA phylogeny and exhibited longer internal branches with similar or better support (Figure 3.30A). *Ferussacia folliculus* still fell at the base of the Achatinoidea while the other achatinoids clustered together but with improved support when compared to the rRNA tree (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). The monophyly of the Achatinidae was also recovered again with better support (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). Consistent with the rRNA tree was the group that consisted of *L. lamellata*, *T. thomensis*, *P. umbilicata*, *E. layardi* and *A. clavulinum* (91% ML bootstraps, $P=1.0$ BI, 88% NJ bootstraps and 71% MP bootstraps), with the São Thomé taxa *T. thomensis* and *P. umbilicata* again forming one pair and with increased support (100% ML bootstraps, $P=1.0$ BI, 92% NJ bootstraps and 100% MP bootstraps) and *E. layardi* and *A. clavulinum* forming a second pair likewise with increased support (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). Still other groups from the rRNA tree that were also recovered included *S. octona* and *S. striatella* (100% ML bootstraps, $P=1.0$ BI, 84% NJ bootstraps and 95% MP bootstraps), *Xerocerastus* sp., *R. decollata* and *Z.*

insularis (97% ML bootstraps, $P=1.0$ BI, 90% NJ bootstraps and 85% MP bootstraps) and *T. erectus* and *G. ceylanica* (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps).

When all taxa were included in the combined analysis (including those taxa for which there was missing data), all groups found in the rRNA phylogeny were recovered and in general received higher support (Figure 3.30B). *Ferussacia folliculus* fell at the base of the Achatinoidea while the remaining Achatinoidea formed a separate clade with increased support (100% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 99% MP bootstraps). Within this clade and consistent with the rRNA tree were the monophyletic Achatinidae with increased support (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps); *C. blandii* and *Riebeckia* sp. (100% ML bootstraps, $P=1.0$ BI, 84% NJ bootstraps and 95% MP bootstraps); *Subulina vitrea* and *Subulona* sp. (88% ML bootstraps, $P=1.0$ BI, 85% NJ bootstraps and 54% MP bootstraps); *S. octona* and *S. striatella* (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps); *G. ceylanica* and *T. erectus* (99% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 97% MP bootstraps); *Xeroxerastus* sp., *Rumina decollata* and *Zootecus insularis* with increased support (99% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 90% MP bootstraps); and *C. gokweanus*, *P. umbilicata*, *T. thomensis*, *L. lamellata*, *A. clavulinum*, *P. achatinaceum*, and *E. layardi* (90% ML: bootstraps, $P=1.0$ BI and 72% NJ bootstraps). Within the last group, *A. clavulinum*, *P. achatinaceum* and *E. layardi* clustered together with increased support (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps), and in turn clustered with *L. lamellata* (71% ML bootstraps, $P=1.0$ BI, 65% NJ bootstraps and 52% MP bootstraps) while the São Thomé group *T. thomensis* and *P. umbilicata* clustered

together with increased support (99% ML bootstraps, $P=1.0$ BI, 98% NJ bootstraps and 100% MP bootstraps).

3.3.3.3. Hypothesis testing of groups expected to be monophyletic based on taxonomy

Seven expected monophyletic groups (based on taxonomy) that were not recovered in either the single gene or combined gene phylogenies were subjected to hypothesis testing using the Shimodaira-Hasegawa (1999) test. These groups were the Coeliacidae, Ferussaciidae, and the Subulinidae; within the Subulinidae were the subfamilies Petriolinae, Rishetiinae, Rumininae and the Subulininae. The placement of *Zootecus insularis* in the Subulininae (according to Schileyko, 1999), in contrast to its placement in the Rumininae (according to Zilch, 1959), was also subjected to the SH test. A summary of the Shimodaira-Hasegawa tests are shown in Table 3.5. Constraining the NJ trees for the four single gene and combined gene phylogenies to show monophyly of the Coeliacidae produced likelihood scores that were significantly worse ($P<0.01$) than those of the optimal trees (see Table 3.5 for details). This confirmed the non-monophyly of the Coeliacidae. The monophyly of the Ferussaciidae was evaluated only for the rRNA cluster and actin gene phylogenies, for which both *Ferussacia folliculus* and *Cecilioides gokweanus* were successfully sequenced, as well as the combined gene phylogeny that included taxa with missing data. The constrained trees showing monophyly of the Ferussaciidae for the rRNA cluster and the combined genes that included all taxa were significantly worse than the optimal tree ($P<0.01$, Table 3.5), indicating that the family is not monophyletic. This was not the case for the actin gene where no significant difference was noted ($P=0.293$, Table 3.5). The monophyly of the Subulinidae was also rejected in the SH tests ($P<0.01$, Table 3.5) as

was the monophyly of two of its subfamilies, the Rishetiinae ($P < 0.03$, Table 3.5) and the Subulininae (with *Z. insularis*- $P < 0.01$; without *Z. insularis*- $P < 0.04$, Table 3.5). For the Petriolinae, the expected topology (Petriolinae monophyletic) was present in the actin NJ tree (see Appendix 3.10B, p. 440) while no significant difference was shown between the likelihood scores for the constrained trees in which the Petriolinae were monophyletic and the optimal NJ trees in the rRNA cluster ($P = 0.093$), histone 3 ($P = 0.184$), 1st and 2nd codon positions of COI ($P = 0.083$), combined genes but excluding taxa with missing data ($P = 0.433$) and combined genes with all taxa ($P = 0.150$). Support for the monophyly of the Petriolinae based on the molecular data is therefore equivocal. The constrained trees showing monophyly of the Rumininae yielded significantly worse likelihood scores than those of the optimal trees ($P \leq 0.001$ for both with and without *Z. insularis* for the rRNA cluster, actin and combined genes with all taxa, Table 3.5). *Riebeckia* sp. was not included in the datasets for histone 3, COI (1st and 2nd codon positions) and the combined genes that excluded taxa with missing data. Significant difference between the optimal tree and the constrained tree was observed in histone 3 ($P \leq 0.01$ for both with and without *Z. insularis*) but not in the 1st and 2nd codon positions of the COI ($P \leq 0.3$ for both with and without *Z. insularis*) and the combined genes but excluding taxa with missing data ($P = 0.370$ for without *Z. insularis* while expected NJ topology was generated when *Z. insularis* was included) (see Table 3.5). On the basis of the taxa examined, neither Shileyko's (1999) nor Zilch's (1959) Rumininae is monophyletic.

Table 3.5: Hypothesis testing for the monophyly of selected taxa of the Achatinoidea using the Shimodaira-Hasegawa test.

Gene	Likelihood Score		P for SH ^a Test (Conclusion ^b)
	Optimal NJ tree	Constrained NJ tree	
COELIAXIDAE			
rRNA cluster	7951.71731	8036.16984	<0.001 (Significant)
Actin	5881.25298	5972.47679	<0.001 (Significant)
Histone 3	1815.79561	1864.59737	<0.001 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1924.07354	0.006 (Significant)
Combined	16679.19409	16826.68130	<0.001 (Significant)
Combined-all taxa	18474.64048	18683.79407	<0.001 (Significant)
FERUSSACIIDAE			
rRNA cluster	7951.71731	7982.47034	0.010 (Significant)
Actin	5881.25298	5877.42426	0.293 (Not significant)
Histone 3	Not applicable; <i>Cecilioides gokweanus</i> not included		
COI (1 st and 2 nd codons)	Not applicable; <i>Cecilioides gokweanus</i> not included		
Combined	Not applicable; <i>Cecilioides gokweanus</i> not included		
Combined-all taxa	18474.64048	18518.72376	0.010 (Significant)
SUBULINIDAE			
rRNA cluster	7951.71731	8098.27402	<0.001 (Significant)
Actin	5881.25298	5922.88073	0.001 (Significant)
Histone 3	1815.79561	1874.36463	0.008 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1933.58362	0.005 (Significant)
Combined	16679.19409	16974.41302	0.005 (Significant)
Combined-all taxa	18474.64048	18638.18916	<0.001 (Significant)
PETRIOLINAE (SUBULINIDAE)			
rRNA cluster	7951.71731	7967.95035	0.093 (Not significant)
Actin	Not applicable; expected topology exists in optimal NJ tree		
Histone 3	1815.79561	1813.07222	0.184 (Not significant)
COI (1 st and 2 nd codons)	1885.86879	1911.13362	0.083 (Not significant)
Combined	16679.19409	16680.26472	0.433 (Not significant)
Combined-all taxa	18474.64048	18486.19798	0.150 (Not significant)
RISHETIINAE (SUBULINIDAE)			
rRNA cluster	7951.71731	8084.95708	<0.001 (Significant)
Actin	5881.25298	6059.01720	<0.001 (Significant)
Histone 3	1815.79561	1838.11663	0.026 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1924.68912	<0.001 (Significant)
Combined	16679.19409	16914.16222	<0.001 (Significant)
Combined-all taxa	18474.64048	18758.90087	<0.001 (Significant)
RUMININAE sensu Schileyko-without <i>Zootecus insularis</i> (SUBULINIDAE)			
rRNA cluster	7951.71731	8019.95480	<0.001 (Significant)
Actin	5881.25298	5911.57548	0.001 (Significant)
Histone 3	1815.79561	1837.83013	0.01 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1896.97810	0.226 (Not significant)
Combined	16679.19409	16681.37066	0.370 (Not significant)
Combined-all taxa	18474.64048	18578.49191	<0.001 (Significant)
RUMININAE sensu Zilch-with <i>Zootecus insularis</i> (SUBULINIDAE)			
rRNA cluster	7951.71731	8000.77016	0.001 (Significant)
Actin	5881.25298	6042.92852	<0.001 (Significant)
Histone 3	1815.79561	1835.88838	0.009 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1894.94276	0.244 (Not significant)
Combined	Not applicable; expected topology exists in optimal NJ tree		
Combined-all taxa	18474.64048	18542.21668	<0.001 (Significant)

SUBULININAE sensu Schileyko-with <i>Zootecus insularis</i> (SUBULINIDAE)			
rRNA cluster	7951.71731	8106.81200	<0.001 (Significant)
Actin	5881.25298	6067.38758	<0.001 (Significant)
Histone 3	1815.79561	1865.98883	0.002 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1940.19417	<0.003 (Significant)
Combined	16679.19409	16965.35856	<0.001 (Significant)
Combined-all taxa	18474.64048	18805.02183	<0.001 (Significant)
SUBULININAE sensu Zilch-without <i>Zootecus insularis</i> (SUBULINIDAE)			
rRNA cluster	7951.71731	8051.96505	<0.001 (Significant)
Actin	5881.25298	5898.27655	<0.028 (Significant)
Histone 3	1815.79561	1841.74798	0.034 (Significant)
COI (1 st and 2 nd codons)	1885.86879	1933.96199	<0.001 (Significant)
Combined	16679.19409	16825.89704	<0.001 (Significant)
Combined-all taxa	18474.64048	18706.81385	<0.001 (Significant)

^a SH test – Shimodaira-Hasegawa Test

^b Significant at $P < 0.05$

3.4. Discussion

This phylogenetic study provides the first systematic coverage of the Achatinoidea with over 24 species from 20 genera and 6 families (Achatinidae, Coeliacidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophorellidae), including 4 subfamilies from the Subulinidae (Petriolinae, Rishetiinae, Rumininae and Subulininae).

3.4.1. Single versus combined gene phylogenies

Three nuclear genes and two mitochondrial genes were used for this study. Among these genes, the nuclear rRNA showed the least saturation, and its phylogeny provided the most resolved structure. Many of the basal nodes were well supported, most notably the splitting of the Achatinoidea into *Ferussacia folliculus* and the rest of the achatinoids. Many terminal nodes likewise received support. Expanding the coverage of the rRNA cluster to include nearly the entire LSU when compared to Wade *et al.*'s (2001, 2006) studies led to an increase in support for the Achatinoidea (99% NJ bootstraps and $P=1.0$ BI for this study versus 93% NJ bootstraps and $P=0.98$ BI in Wade *et al.*'s (2006) study). Support for the Achatinidae increased significantly from

38% NJ bootstraps and $P=0.94$ BI to 90% NJ bootstraps and $P=1.0$ BI, although a different set of achatinid taxa (apart from *A. fulica*) were used when compared to the Wade *et al.* (2006) study. There was also a slight increase in support for the *Coeliaxis blandii* and *Riebeckia* sp. grouping (99% NJ bootstraps and $P=1.0$ BI for this study versus 92% NJ bootstraps and $P=1.0$ BI in Wade *et al.*'s (2006) study). However, a rather surprising decrease in support was noted for the *Pyrgina umbilicata* and *Thyrophorella thomensis* group (73% NJ bootstraps and $P=0.93$ BI for this study versus 100% NJ bootstraps and $P=1.0$ BI in Wade *et al.*'s (2006) study). The actin gene also recovered many strongly-supported groupings found in the rRNA cluster despite the presence of several copies of the actin gene within many taxa leading to ambiguous sites in the sequences. While many basal groups received less than 50% bootstrap support (ML, NJ and MP) and less than 0.7 posterior probabilities (BI), the splitting of the Achatinoidea into *F. thomensis* and the other achatinoids was still strongly supported with 100% bootstrap support (ML, NJ, MP) and a posterior probability of 1.0 (BI). Two terminal groups also received higher support in actin than in the rRNA cluster. For instance, the Achatinidae obtained 98% NJ bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 99% MP bootstraps. The second group, comprising *Pyrgina umbilicata* and *Thyrophorella thomensis*, received 100% ML bootstraps, $P=1.0$ BI, 59% NJ bootstraps and 100% MP bootstraps; however, *Pyrgina* exhibited more ambiguous sites (88) than the total number of nucleotide differences (77) between it and *Thyrophorella thomensis*. It is highly probable that the actin copies in *P. umbilicata* and *T. thomensis* diverged earlier than the two taxa, and interpretation of this grouping should be treated with caution. On the other hand, the histone 3 and COI (1st and 2nd codon position) gene phylogenies generally had less structure with little support for basal nodes in the tree. This could be attributed to the relatively smaller number of sites used, particularly in the

COI where the 3rd codon positions were excluded, as well as the possibility that both datasets were approaching saturation. In the COI, for example, the high levels of saturation of the 3rd codon position would imply the fast rate of evolution for this gene. Nevertheless, several groups in common with the rRNA and actin phylogenies were also recovered from the histone 3 and COI data. The 16S rRNA gene was excluded because it was found to be saturated and therefore not suitable for phylogenetic analyses at the level of the Achatinoidea.

The groups recovered in the rRNA tree also emerged in the phylogenies based on the concatenated sequence after the partition homogeneity test revealed that the individual datasets could be combined. The resolution within the internal branches and the statistical support for groupings in the tree clearly improved when the datasets were combined and analyzed under a single model (GTR+ Γ) for both the phylogeny that excluded taxa with missing datasets and for the phylogeny with all taxa. The combined phylogenies were dominated by the signal from the rRNA cluster as all groups from the rRNA phylogeny that did not appear in the other single gene phylogenies were recovered in the combined phylogenies.

3.4.2. Phylogenetic relationships within the Achatinoidea

3.4.2.1. The monophyly and phylogenetic position of the Achatinidae

The limited natural range of the Achatinidae to sub-Saharan Africa (Tillier, 1989, Raut & Barker, 2002) and the existence of fossils no earlier than the Pleistocene (0.01-1.8 MYA) (Zilch, 1959-1960; Solem, 1979) suggest that the achatinids are a relatively recent group (Tillier, 1989), although Raut & Barker (2002) believed them to be much older than the fossil record. The first molecular evidence for the monophyly of

the Achatinidae was provided by Wade *et al.* (2001) based on the analysis of a partial fragment of the rRNA cluster. Further molecular evidence for achatinid monophyly is provided in this study based on the analysis of an expanded rRNA dataset as well as the analysis of data from three other genes; actin, histone 3 and the 1st and 2nd codon positions of COI. High support for the monophyly of the family was also obtained when the genes were combined. The relative position of the Achatinidae within the Achatinoidea was not resolved.

3.4.2.2. The monophyly and phylogenetic position of the Ferussaciidae

This study included two representatives from the Ferussaciidae, *Ferussacia folliculus* and *Cecilioides gokweanus*, although the H3 and COI gene fragments failed to amplify in the latter. In the rRNA phylogeny, the two taxa did not cluster together as expected, and the early divergence of *F. folliculus* from the rest of the Achatinoidea, including *C. gokweanus*, was strongly supported (98% ML bootstraps, $P=1.00$ BI, 94% NJ bootstraps and 99% MP bootstraps). Furthermore, the Shimodaira-Hasegawa test conclusively rejected the monophyly of the Ferussaciidae. The actin gene also did not recover the Ferussaciidae as a monophyletic unit, though both bootstrap support/Bayesian posterior probabilities and the Shimodaira-Hasegawa test were equivocal. *Cecilioides gokweanus* has only been described conchologically; its internal anatomy has not been described, although that of *Cecilioides acicula* is already available. *Cecilioides acicula*, like *Ferussacia folliculus*, exhibits characteristics unique to Ferussaciidae such as the short but transversely elongated kidney (Watson, 1928; Tillier, 1989). However, the kidney of *C. acicula* is broader and curves gradually forward as it approaches the rectum whereas the kidney of *F. folliculus* bents abruptly where it meets the rectum. Furthermore, *C. acicula* has a simpler penis structure than *F.*

folliculus, as the former possesses an indistinct penis papilla (Watson, 1928). There is therefore no compelling evidence from morphological data to show that the Ferussaciidae are monophyletic, and the molecular data from this study confirm their polyphyly.

3.4.2.3. Evaluation of the taxonomic designation of the Subulinidae, Coeliacidae, Glessulidae and Thyrophorellidae

The single and combined phylogenies clearly showed that the Subulinidae (represented by eleven taxa in this study), are a polyphyletic group, with members of the Coeliacidae (two representatives), Glessulidae (one representative) and Thyrophorellidae (one representative) falling among the subulinids. Subulinid monophyly was also significantly rejected by the SH test. The phylogenies also revealed that the Coeliacidae, which are regarded as a subfamily, Coeliacinae, under the Subulinidae by Schileyko (1999), are likewise not monophyletic, as *Coeliaxis blandii* and *Pyrgina umbilicata* did not cluster together. Coeliacidae monophyly was also significantly rejected by the SH test.

The polyphyly of the subulinids is not surprising considering several taxa have not been described anatomically and are grouped together based only on their shell morphology and geographic distribution (Schileyko, 1999). However, some fascinating groupings have emerged. For instance, *Xerocerastus* sp. from southwest Africa (Schileyko, 1999), *Rumina decollata* from the Mediterranean (Schileyko, 1999) and *Zootecus insularis* from the Sahara, Arabia, India and Burma (Schileyko, 1999) formed one group despite having distinct geographic distributions. Most descriptions of *Xerocerastus* species are limited to the shell (Van Bruggen, 1970; Schileyko, 1999), but both *R. decollata* and *Z. insularis* have a very short to nearly absent oviduct (Schileyko,

1999). *Xerocerastus* should be evaluated anatomically to determine if it shares any morphological features with *R. decollata* and *Z. insularis*.

Two of the three *Subulina* species clustered together: *S. octona* from central and tropical South America (Schileyko, 1999) and *S. striatella* from tropical Africa (Jurickova, 2006). The third species, *S. vitrea*, which has a geographic distribution limited to within southwest Africa (Van Bruggen, 1970), clustered with the tropical African *Subulona* sp. The internal structures of *S. striatella* and *S. vitrea* have not been described, but *S. octona* and the type species of *Subulona*, *S. badia* (Martens), are distinct morphologically in that the former has a penis sheath while the latter does not (Schileyko, 1999). The reproductive tracts of *S. striatella* and *S. vitrea* are therefore worth investigating to determine if they are also distinct.

The subulinid *Allopeas clavulinum* (Subulininae), which has a New World distribution, clustered with another subulinid, *Eutomopeas layardi* (Rishetiinae), which is restricted to Sri Lanka and neighboring islands (Schileyko, 1999). Like most subulinid genera, which are classified according to their shell characteristics, both taxa can be distinguished from one another conchologically based on the shape of the apex, color pattern and the presence or absence of the umbilicus (Schileyko, 1999). However, *Allopeas* and *Eutomopeas* also share other shell characteristics, including shape, size and translucence (Schileyko, 1999). Cain (1977) observed that distinct taxa, such as the high spired *Cochlicella acuta* and *Cionella* (= *Cochlicopa*) *lubrica*, which live in separate but very similar environments and possibly exhibit similar feeding habits, may manifest strikingly similar shell morphology as a result of convergent evolution. Mead (1994) found the same observations for two species of the Achatinidae. This could also be the case for *A. clavulinum* and *E. layardi*. Only the internal anatomy of *A. clavulinum* has been described, so it is therefore worthwhile to note if the internal

structures of these taxa are also congruent with molecular data. If that is the case, then the shell characteristics used to distinguish the two taxa into separate subfamilies and even genera may not be valid.

The coeliacid *Coeliaxis blandii* from South Africa (Schileyko, 1999) grouped with the subulinid *Riebeckia* sp. found in Sokotra near the tip of Somalia (Schileyko, 1999). Only *C. blandii* has been described morphologically. Comparison of the two taxa based on internal structures is therefore not possible.

The other coeliacid, *Pyrgina umbilicata*, forms a group with the thyrophorellid *T. thomensis* that is consistent in all the single gene and combined gene phylogenies. This grouping is highly controversial in that, in contrast to *A. clavulinum* and *E. layardi*, these two taxa are very distinct from each other conchologically. Whereas *P. umbilicata* is dextral and turricate with 13 narrow whorls (Schileyko, 1999), *T. thomensis* is sinistral and low conic above and bulging below (Schileyko, 2001). Furthermore, the thyrophorellid bears a unique operculum-like structure (Schileyko, 2001). It is speculated that differences in shell morphology often reflect the differences of the feeding behaviour of the taxa being compared (Cain, 1977) and not necessarily their phylogenetic relationship. The only similarity exhibited by *P. umbilicata* and *T. thomensis* at present is their geographic distribution; both are restricted to São Thomé (Schileyko, 1999, 2001). It is not uncommon to have distinct morphological variation that arises between closely related taxa within a small geographic area as a result of adaptive radiation. A classic example of this is Darwin's finches in the Galapagos Islands, where different environmental conditions could have given rise to different feeding niches for these birds that could have led to the evolution of different beak forms (Whittaker & Fernandez-Palacios, 2007). Further examination of the internal

structures of *P. umbilicata* is imperative so that it can be compared with *T. thomensis*, which is already anatomically described (Schileyko, 2001).

Tortaxis erectus from Indochina and southern China (Schileyko, 1999) and the glessulid *Glessula ceylanica* from the Indian subcontinent and Sri Lanka (Schileyko, 1999) formed another group. Conchologically, the two species are somewhat similar in having glossy shells, although *T. erectus* has a turreted, cylindrical-turreted or subulate-turreted shell with 6-13 convex whorls while *Glessula* has an elongated-ovate to ovate-conic shell with 5-8 convex whorls (Schileyko, 1999). The anatomy of *T. erectus* is likewise unknown, so comparison of the two taxa based on internal structures is not possible. In addition to *G. ceylanica*, there are various other *Glessula* species that need to be characterized at the molecular level as they are very similar to each other conchologically (Beddome, 1906).

Based, therefore, on molecular data presented in this study, and in light of the fact that numerous subulinids are not described anatomically, revision of the Subulinidae is clearly needed as this no longer appears to be a valid group. The Coeliacidae must also be revised as its representatives in this study grouped with either another subulinid or a thyrophorellid. The designation of the monotypic Thyrophorellidae requires re-examination based on *Thyrophorella thomensis*' close association with the coeliacid *Pyrgina umbilicata*. Finally, the association of *Tortaxis erectus* with *Glessula ceylanica* raises the question as to whether the Glessulidae are really restricted to the Indian subcontinent.

3.4.2.4. Evaluation of the monophyly of the different subulinid subfamilies

Of the four subulinid subfamilies represented in this study, only the Petriolinae (represented by *Bocageia* sp. and *Riebeckia* sp.) were recovered in any of the gene

phylogenies, in this case the NJ tree based on the actin gene though bootstrap support was less than 50% (see Appendix 3.11B, p. 442). For the rRNA, H3, COI and combined gene phylogenies, the Shimodaira-Hasegawa test did not find any significant difference in the likelihood scores between the optimal NJ tree (in which the Petriolinae were not monophyletic) and the constrained tree showing monophyly of the Petriolinae. There is therefore no compelling evidence to rule out the monophyly of the Petriolinae but neither is there any convincing evidence in support of the subfamily. The Petriolinae is distinct from all the other subfamilies based on the presence of a well-developed epiphallus and a very large and fleshy stimulator in the penis. *Bocageia* sp. and *Subulona* sp. have not yet been described based on their reproductive structure and were only included in the subfamily based on their shell morphology (Schileyko, 1999). The monophyly of the other subulinid subfamilies, particularly Rumininae sensu Schileyko (without *Zootecus*) and Rumininae sensu Zilch (with *Zootecus*), were strongly rejected by the Shimodaira-Hasegawa Test, and a re-examination of their taxonomic position is warranted. As discussed in the previous section, many of these taxa that have been included in the subfamilies have not been described anatomically, particularly with regards to the reproductive structures. In fact, this situation applies to many subulinids in general, which leaves their taxonomic position questionable (Schileyko, 1999).

3.4.3. The next step

The monophyly of the Achatinidae using molecular data is based only on very limited taxa sampled to date. A comprehensive survey of taxa within the family will determine the extent of their monophyly and at the same time give a glimpse of the inter-relationships of the various groups in the family on the basis of molecular data. Mead (1991, 1995) has already described various groups within the Achatinidae using

the reproductive tract as a criterion, and it would be interesting to know if these groupings are supported by molecular data. The molecular data have revealed that the Ferusaciidae are not monophyletic; the inclusion of additional taxa from the family is therefore necessary in order to re-evaluate the relationships within the group. The Subulinidae are also determined to be non-monophyletic based on molecular data; this necessitates a re-examination of the different surprising groupings of these subulinids in the light of their association with the Coeliacidae (also non-monophyletic), Glessulidae and Thyrophorellidae by expanding the taxon coverage to include other members of the families mentioned. There is also a need to investigate the morphological features of many of the anatomically undescribed species to correlate the molecular data with morphological data. Within the Subulinidae, only the Petriolinae had equivocal support for its monophyly. Including other taxa from the Petriolinae could shed more light on its status as a valid group. Other subfamilies that were not represented in this study (Obeliscinae, Opeatinae, Perrieriinae and Tristaniinae) should likewise be surveyed to determine their validity in the light of the rejection of the monophyly of the Rishetiinae, Rumininae and Subulininae.

3.5. Summary

The study provides a phylogenetic analysis of the Achatinoidea based on 24 taxa representing six families using the nuclear rRNA, actin and histone 3 genes and the 1st and 2nd codon positions of the mitochondrial cytochrome c oxidase subunit I gene. The 16S rRNA gene and the 3rd codon position of the COI gene were severely saturated and were excluded from the phylogenetic analyses. Combined gene phylogenies recovered most of the groupings in the rRNA phylogeny with longer internal branches and similar or slightly better statistical support. The study provided further strong molecular support

for the monophyly of the Achatinidae. The Ferussaciidae did not form a monophyletic group within the molecular trees, with *Ferussacia folliculus* falling separately from *Cecilioides gokweanus* at the base of the Achatinoidea, thus warranting a re-evaluation of the Ferussaciidae as a valid taxonomic group. The Coelioxidae was not supported, with the two representatives sequenced to date falling separately in the molecular trees, thereby prompting a re-evaluation of the validity of the family. Finally, the Subulinidae were not supported in the molecular trees with members of the Coelioxidae, Glessulidae and Thyrophorellidae falling among the subulinids. As with the Ferussaciidae and the Coelioxidae, the molecular data prompt a re-evaluation of the Subulinidae as a valid taxonomic group.

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CHAPTER 4 - Phylogeny of the Achatinidae

4.1. Introduction

4.1.1. The Achatinidae

Other than as exotic alien species, the Achatinidae are confined to sub-Saharan Africa and the Atlantic Islands in the Gulf of Guinea, West Africa. *Achatina (Lissachatina) immaculata* Lamarck is considered to have been introduced into Madagascar (Raut & Barker, 2002), but the most notable colonizing achatinid is *Achatina (Lissachatina) fulica* Bowdich, which has been introduced by Man throughout much of the tropics where it is a major pest species (Mead, 1979a; Raut & Barker, 2002). Achatinids include some of the largest known terrestrial molluscs (Bequaert, 1950) and have been aptly called Giant African Land Snails. There are 13 genera and approximately 200 species (Raut & Barker, 2002) distributed across 3 subfamilies within the Achatinidae (Schileyko, 1999). With the exception of the sinistral *Columna columna* (Müller) and *Archachatina bicarinata* (Bruguière), the shells of the Achatinidae are dextral. In general, achatinid shells are higher than wide and are fusiform, ovoid or pillar-shaped (Schileyko, 1999). Although achatinids do not possess the wide range of anatomical accessory structures that are found in many other stylommatophoran groups, the morphology of the genital structures in achatinids is highly variable (Mead, 1991); all members of the Achatinidae possess a well-developed penis sheath (Schileyko, 1999).

4.1.2. Distribution and habitat of the Achatinidae

The Achatinidae are widely distributed in sub-Saharan Africa. They are present from Senegal to Guinea in the west, the Zaire Basin, the Lake District and Southern Sudan in the centre, Somalia and southern Ethiopia in the northeast, Kenya, Tanzania and Mozambique in the east, and to Zambia, Botswana, Zimbabwe, South Africa, Namibia and Angola in the south and southwest (Mead, 1991; Raut & Barker, 2002). The Atlantic Islands in the Gulf of Guinea, including São Thomé and Príncipe, also harbor certain achatinids such as *Columna columna* (Schileyko, 1999) and the monotypic *Atopocochlis exarata* (Müller) (Mead, 1991; Raut & Barker, 2002). Van Bruggen (1986) noted an unequal distribution of diversity of the Achatinidae within sub-Saharan Africa, with Western Africa exhibiting a high concentration of endemic species. Habitat preferences of the achatinids are also diverse. Members of the genus *Achatina* in Central Africa thrive in humid, tropical forest areas. Some achatinids such as *Achatina (Lissachatina) fulica* in Eastern Africa flourish in modified tropical forests and forest margins and even in highly disturbed areas such as gardens, intensively farmed crops and plantations. Others such as the lowland *Cochlitoma zebra* (Bruguière) and the montane *Cochlitoma omissa* (van Bruggen) live in temperate regions in Southern Africa. Still others can be found in less humid areas, for example *Achatina (Lissachatina) immaculata* Lamarck in savannah and *Achatina (Achatina) damarensis* (Pfeiffer) in deserts of south and southwest Africa (Raut & Barker, 2002).

4.1.3. The search for the phylogeny of the Achatinidae using morphological characters

Several studies have attempted to elucidate the phylogeny of the Achatinidae using morphological characters (Bequaert, 1950; Mead, 1950, 1979b, 1991, 1995).

Classification within the family is based mostly on shell form and structure as well as the anatomy of the genitalia. However, different forces of selection act to different extents on the two systems. Shell form is subjected to selection pressures associated with particular habitats and homoplasy in shell form is common (Cain, 1977; Mead, 1994); modifications to the genitalia are more likely to reflect evolutionary processes (Mead, 1994) partly in response to sexual selection and selection pressures associated with the need for compatibility of form within species and incompatibility of form acting as reproductive isolation mechanisms between species. It is therefore not unusual to have cases where there is no correspondence between shell structures and the genitalia (Mead, 1979b, 1991) where distinct taxa living in separate but very similar environments may undergo convergent evolution and manifest strikingly similar shell morphology (Mead, 1994). For instance, the western species *Achatina* (*Achatina*) *achatina* Linnaeus and the southern species *Cochlitoma zebra* (Bruguière) are very similar conchologically and were placed in the same genus, *Achatina*, by Bequaert (1950) when clearly they belong to different genera based on the differences in their reproductive organs (Mead, 1991).

A typical achatinid reproductive system based on *Archachatina* (*Calachatina*) *marginata* Swainson is shown in Figure 4.1. Achatinids, like all stylommatophorans, are hermaphroditic. Both the sperm and egg are produced in the ovotestis, though not at the same time, and pass through the hermaphrodite duct. During copulation, the sperm proceeds to the male portion of the common hermaphrodite duct and then to the vas deferens and the penis. The penis then protrudes through the genital atrium and the genital pore and inserts into the reproductive tract of the partner until it reaches the vagina where it injects the sperm into the spermatolytic sac through the spermatolytic duct. The penis then retracts through the penial retractor muscle. Most of the sperm injected

into the spermolytic sac is digested by proteolytic enzymes. Sperm that survive go through the spermolytic duct and pass through the oviduct into the female hermaphrodite duct (=uterus) before they are stored in the seminal receptacle (=carrefour). Fertilization also takes place in the seminal receptacle, after which the fertilized eggs or zygotes receive yolk from the albumin gland. The zygotes then pass through the female hermaphrodite duct where the yellow egg shells are added before the eggs are released through the genital pore (Segun, 1975; Egonmwan, 2007).

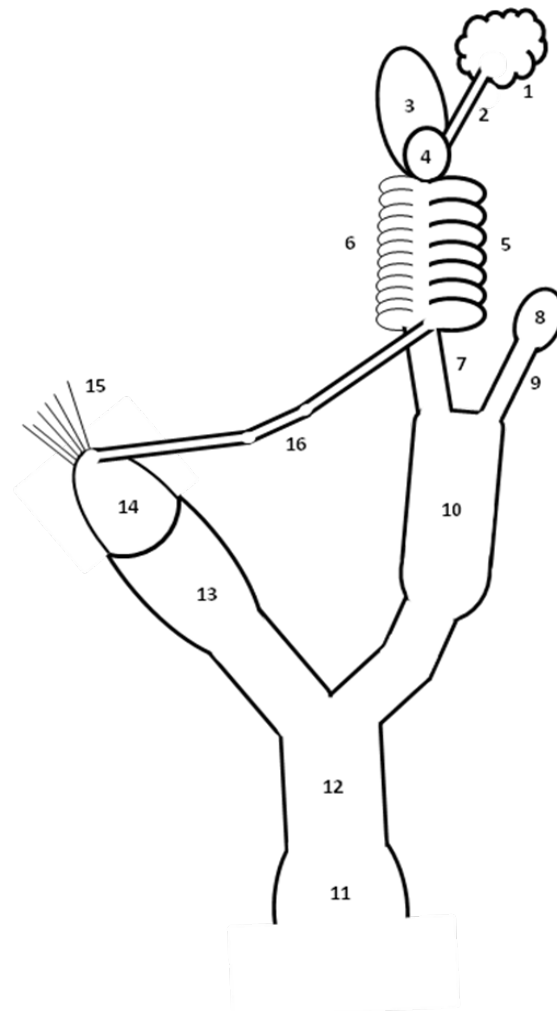


Figure 4.1: Generalized reproductive structure of achatinids. 1 – ovotestis; 2 – hermaphroditic duct; 3 – albumin gland; 4 – seminal receptacle; 5 - male part of common hermaphrodite duct; 6 – female part of common hermaphrodite duct (=uterus); 7 oviduct; 8 – spermolytic sac (=spermatheca); 9 – spermolytic duct; 10 – vagina; 11 – genital pore; 12 – genital atrium; 13 – penis sheath; 14 – penis; 15 – penial retractor muscle; 16 – vas deferens (Segun, 1975; Gomez, 2001; Egonmwan, 2007).

Morphological features of achatinid reproductive systems show a strong correlation with geographic distribution. For example, Mead (1991, 1994) hypothesized that members of the subfamily Callistoplepinae, which are found in Lower Guinea (Cameroon, Equatorial Guinea, Gabon and adjacent islands) in West Africa and include the genera *Callistoplepa* and *Leptocala*, probably gave rise to the other achatinids because they have the most 'primitive' genitalia in which the penis sheath does not enclose the vas deferens (Figure 4.2A). For the rest of the achatinids, the developing penis sheath envelopes the apical vas deferens to divide it into the apical and basal portions. Among these achatinids, the penis sheath either encloses the basal vas deferens but not the entire penis (microphallate type, Figure 4.2B) or it encloses both the basal part of the vas deferens as well as the entire penis (macrophallate type, Figure 4.2C). These two types can be found across different geographic regions of the Sub-Saharan Africa over several genera; for instance, the microphallate type is present in the West African *Achatina* (*Achatina*) *achatina*, *Archachatina marginata* and *Pseudachatina downesii* Sowerby and the Southern African *Metachatina kraussi* L. Pfeiffer while the macrophallate type is evident in the East African *Achatina* (*Lissachatina*) *fulica*, the Southern African *Cochlitoma* species and the East and West African *Limicolaria* species (Mead, 1979b; 1991). Apart from the penis sheath, other features also reflect groupings within the Achatinidae that have been thought to mirror their routes of dispersal. In the subfamily Limicolariinae, members are restricted to tropical regions of Africa and possess penial retractor muscles that originate from the diaphragm and are not enclosed by the penis sheath; a verge or penis papilla is also present at the base of the penis in this subfamily (Schileyko, 1999). Greater variation in the reproductive structure is exhibited in the subfamily Achatininae, members of which are distributed all over Sub-Saharan Africa (Schileyko, 1999), with some genera

possessing reproductive features that reflect their geographic distributions. For instance, extroversion muscles at the base of the penis sheath are very prominent in East African *Achatina* in the subgenus *Lissachatina* (Mead, 1979b, 1991), which include *A. (L.) fulica*, *A. (L.) immaculata*, *A. (L.) reticulata* Pfeiffer and *A. (L.) zanzibarica* Bourguignat. In Southern Africa, several species of *Archachatina* and *Achatina* exhibit a distinct folding of the penis, which prompted Mead (2004) to resurrect *Cochlitoma* Pilsbry. Based on these morphological features, it was proposed that the Achatinidae originated in Lower Guinea and dispersed through much of Sub-Saharan Africa (south of the Sahel) where they adapted to specific environmental conditions, from humid to arid, and tropical to temperate (Mead, 1991; Raut & Barker, 2002).

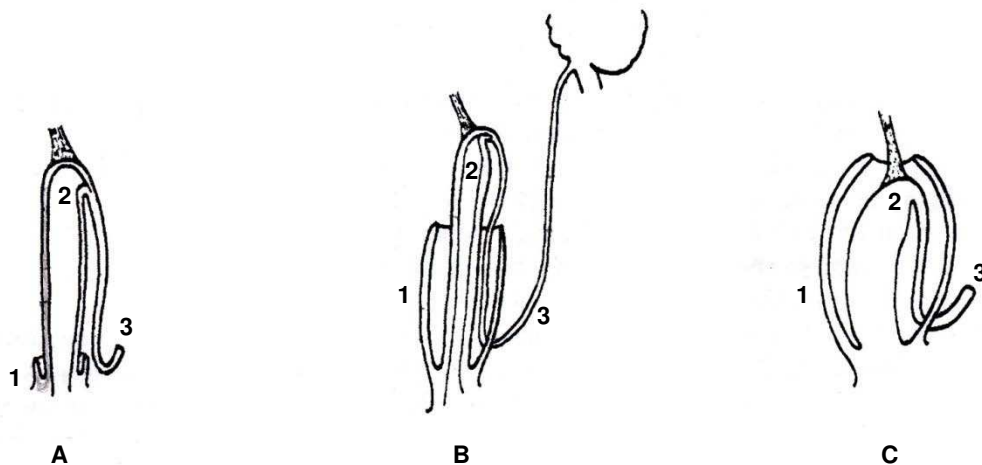


Figure 4.2: Schematic diagrams of how the penis sheath envelops certain parts of achatinid genitalia. 1 – penis sheath; 2 – penis; 3 – vas deferens. **A** – the penis sheath does not envelop the vas deferens, as in *Callistoplepa*. **B** – the penis sheath envelops the vas deferens and divides into the apical and basal portions, as in the West African *Archachatina*. **C** – the penis sheath envelops both the entire penis and the basal segment of the vas deferens, as in *Achatina (Lissachatina)*. (From Mead, 1991).

4.1.4. Taxonomic revision of the genus *Achatina* based on morphological characteristics

Bequaert (1950) used the appearance of the nepionic whorls of the shell to distinguish two subgenera of the genus *Achatina*, namely: (1) *Achatina* found in West and Central Africa; and (2) *Lissachatina* found in East Africa. He noted that the nepionic whorls in *Achatina* (*Achatina*) are sculptured whereas those of *Achatina* (*Lissachatina*) are either smooth or with faint vertical wrinkles. Mead (1995) further added that *Achatina* (*Achatina*) normally has a plateaued apex while *Achatina* (*Lissachatina*) exhibits a lopsided or half-dome apex. However, distinguishing these two subgenera using taxonomic characters based on the shell can be unreliable in the case of specimens with worn or weathered shells (Mead, 1995). The morphology of the reproductive organs is considered to provide more reliable characters for recognizing natural groups (Mead, 1991). For example, *Achatina* (*Achatina*) has a short spermolytic duct (=spermathecal duct *sensu* Mead), which causes the spermolytic sac (=spermatheca) to be located below the junction of the apical vas deferens and the uterine portion of the common hermaphrodite duct (=spermoviduct). In contrast, *Achatina* (*Lissachatina*) has a very long spermolytic duct, thus pushing the spermolytic sac way above the junction of the apical vas deferens and the uterine portion of the common hermaphrodite duct. Furthermore, *Achatina* (*Lissachatina*) tends to have a muscular, bulboid enlargement at the basal vagina (Fig. 4.3) in addition to the presence of extroversion muscles at the base of the penis sheath (Mead 1991; 1995).

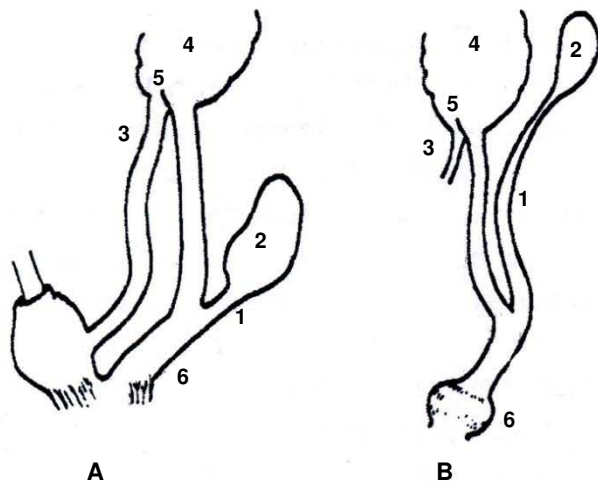


Figure 4.3: Schematic illustrations of the reproductive organs of the West and Central African *Achatina* (*Achatina*) (A) and the East African *Achatina* (*Lissachatina*) (B). 1 – spermolytic duct; 2 – spermolytic sac; 3 – apical vas deferens; 4 – uterine portion of the common hermaphrodite duct; 5 – junction of 3 and 4; 6 – basal vagina. Note that the spermolytic duct is longer in *Achatina* (*Lissachatina*), pushing the spermolytic sac above the junction of the vas deferens and the uterine portion of the common hermaphroditic duct. *Lissachatina* also tends to have a bulbous basal vagina. (From Mead, 1991)

4.1.5. Objective of the study

Results from Chapter 3 confirm the monophyly of the Achatinidae as represented by four taxa. However, no molecular based phylogenetic analyses have been carried out to determine the relationships within the Achatinidae. The current investigation aimed to address this issue by obtaining molecular data for the nuclear ribosomal rRNA cluster, actin and histone 3 genes as well as the mitochondrial cytochrome c oxidase subunit 1 and 16S rRNA genes to elucidate the phylogeny of the Achatinidae. The validity of the taxonomic designation of the East African *Lissachatina* and the West African *Achatina* was also evaluated using the molecular data.

4.2. Materials and methods

4.2.1. Taxa used

Twenty-nine taxa from the family Achatinidae were used to evaluate the phylogeny of the Achatinidae (see Table 4.1). Except for five taxa used by Wade *et al.* (2006), namely *Achatina (Lissachatina) fulica*, *Archachatina marginata*, *Atopocochlis exarata*, *Limicolaria kameul* and the subulinid outgroup taxon *Rumina decollata*, all other taxa have not been evaluated in previous studies. *Rumina decollata* was chosen as the outgroup because it fell immediately outside of the Achatinidae in the phylogenetic studies found in Chapter 3, and it was the taxon that allowed incorporation of the greatest number of unambiguously aligned nucleotide sites across the Achatinidae for the ITS 2 region. Three subgenera of *Achatina* were represented in this study, namely *Achatina* (represented by *A. achatina*, *A. biscalpta*, *A. damarensis* and *A. stuhlmanni*), *Euaethiopina* (*A. loveridgei*) and *Lissachatina* (*A. fulica*, *A. immaculata*, *A. reticulata* and *A. zanzibarica*).

Table 4.1: Taxa used for the phylogenetic study of the Achatinidae. With the exception of *Achatina (Lissachatina) fulica*, *Archachatina marginata*, *Atopocochlis exarata*, *Limicolaria kambeul* and *Rumina decollata* from Wade *et al.* (2006), all other taxa were not used in previous studies and were solicited through email by myself, C. Hudelot, F. Naggs and C. Wade from the collectors shown in the table. The DNA for the five taxa used by Wade *et al.* (2006) was provided by C. Wade.

Family	Species	Collection/Location	Collector/Provider
Achatinidae	<i>Achatina (Achatina) achatina</i> (Linnaeus, 1758)	Unknown (Zool. Soc. Lond. Colln.)	NHM*
	<i>Achatina (Achatina) bisculpa</i> Smith, 1878	Gauteng Prov., South Africa	K. du Preez
	<i>Achatina (Achatina) damarensis</i> (Pfeiffer, 1870)	Damaraland, Namibia	D. Herbert
	<i>Achatina (Achatina) stuhlmanni</i> von Martens, 1892	Semuliki National Park, Uganda	B. Rowson
	<i>Achatina (Lissachatina) fulica</i> Bowdich, 1822	Captive bred, unknown origin	NHM
	<i>Achatina (Lissachatina) immaculata</i> Lamarck, 1822	Cato Ridge, South Africa	D. Herbert
	<i>Achatina (Lissachatina) reticulata</i> Pfeiffer, 1845	Mkungwe Forest reserve, Uluguru Mts, Tanzania	B. Rowson
	<i>Achatina (Lissachatina) zanzibarica</i> Bourguignat, 1879	Lulanda Forest Reserve, Udzungwa Scarp, Tanzania	B. Rowson
	<i>Achatina (Euaethiopina) loveridgei</i> (Clench & Archer, 1930)	Uluguru Forest Reserve, Tanzania	B. Rowson
	<i>Archachatina marginata</i> (Swainson, 1821)	Purchased in Lagos Market, Nigeria	NHM
	<i>Atopocochlis exarata</i> (Müller, 1774)	Nr. Belem, São Thomé	A. Gascoigne
	<i>Cochlitoma churchilliana</i> (Melvill & Ponsonby, 1895)	KwaZulu-Natal, South Africa	D. Herbert
	<i>Cochlitoma dimidiata</i> (Smith, 1878)	Mpumalanga, South Africa	A. Moussalli & D. Stuart-Fox
	<i>Cochlitoma granulata</i> (Krauss, 1848)	KwaZulu-Natal, South Africa	A. Moussalli & D. Stuart-Fox
	<i>Cochlitoma kilburni</i> Mead, 2004	Eastern Cape Prov., South Africa	L. Davis & M. Burse
	<i>Cochlitoma marinae</i> (Sirgel, 1989)	Mossel Bay, Eastern Cape Prov., South Africa	K. Tolly
	<i>Cochlitoma montistempli</i> (van Bruggen, 1965)	Cathedral Peak area, Fern Forest, South Africa	D. Herbert
	<i>Cochlitoma omissa</i> (van Bruggen, 1965)	Monks Cowl area, South Africa	D. Herbert
	<i>Cochlitoma semidecussata</i> (Pfeiffer, 1846)	Durban, Pigeon Valley Park, South Africa	A. Moussalli & D. Stuart-Fox
	<i>Cochlitoma simplex</i> (Smith 1878)	Colenso area, Emaweni Game Ranch, near Brakfontein, South Africa	V. Albers
	<i>Cochlitoma</i> sp. cf <i>vestita</i> (Pfeiffer, 1861)	KwaZulu-Natal, South Africa	D. Herbert
	<i>Cochlitoma ustulata</i> (Lamarck, 1822)	Western Cape Prov., South Africa	A. Moussalli & D. Stuart-Fox
	<i>Cochlitoma varicosa</i> (Pfeiffer, 1861)	Transkei, South Africa	J. Marais
	<i>Cochlitoma zebra</i> (Bruguère, 1789)	W. Cape, Witsand, South Africa	A. Moussalli & D. Stuart-Fox
	<i>Limicolaria kambeul</i> (Bruguere, 1792)	Somalia (NHM collection)	M. Leng
	<i>Limicolaria martensii</i> (Smith, 1880)	Bulyanbulu, Tanzania	A. Moussalli
	<i>Limicolariaopsis ruwenzoriensis</i> Pilsbry, 1919	Rwenzori Mts, National Park, Uganda	B. Rowson
<i>Limicolariaopsis</i> sp.	Saimofores, Tupen Hills, Kenya	M. Pickford	
<i>Metachatina kraussi</i> Pilsbry, 1904	Zululand, South Africa	D. Herbert	
Subulinidae (outgroup)	<i>Rumina decollata</i> (Linnaeus, 1758)	Sicily	A. Davison

*NHM-Natural History Museum

4.2.2. DNA extraction, PCR amplification and sequencing

The CTAB method of DNA extraction (Section 2.1.1 of Chapter 2, pp. 37 – 39) was used on tissue slices (approximately 8 mm³) derived from the foot muscle of each snail. The DNA for *Achatina (Lissachatina) fulica*, *Archachatina marginata*, *Atopocochlis exarata*, *Limicolaria kambeul* and *Rumina decollata* was provided by C.M. Wade.

Amplification by PCR, gel migration and purification of PCR products from agarose gels were carried out as described in Sections 2.2 – 2.4 of Chapter 2, pp. 42 – 59. Approximately 4000 bp of the nuclear rRNA cluster (80 bp of the 5.8S rRNA, the full ITS2 and nearly the entire fragment of the LSU rRNA), 900 bp of the nuclear cytoplasmic actin gene, 328 bp of the nuclear histone 3 gene, 650 bp of the Folmer *et al.* (1994) mitochondrial COI fragment and 420-500 bp of the Palumbi *et al.* (1991) mitochondrial 16S (420-500 bp) fragment were amplified using the primers listed in Tables 2.1 – 2.5 of Chapter 2, pp. 47 – 53. Both sense and anti-sense strands were sequenced directly using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 terminator cycle sequencing chemistry (see Section 2.6 of Chapter 2, pp. 59 – 61 for details).

4.2.3. Sequence analysis

Sequences were assembled using the STADEN package version 1.5.3 (Staden *et al.*, 2000) and aligned manually within the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994). Ambiguous sites due to sequencing errors (for all genes) or the presence of multiple copies (for the actin gene) were assigned as described in Section 2.9.3 of Chapter 2, pp. 72-73 and using as a guide the three cloned sequences from representative taxa of the Achatinoidea in Chapter 3.

The average base frequencies and the numbers of variable and parsimony-informative sites for each gene fragment were determined (Section 2.9.4 of Chapter 2, p. 73). Corrected distances were computed after determination of the optimal model for DNA sequence evolution. Models evaluated by the Likelihood Ratio Test (LRT) were the JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P (Kimura, 1980), HKY85 (Hasegawa *et al.*, 1985), TN93 (Tamura & Nei, 1993) and the GTR (Rodriguez *et al.*, 1990) models as well as their variant that incorporated gamma distributed rates (Γ) (Yang, 1993) (Section 2.9.5 of Chapter 2, pp. 73-77). The sequences were also examined for evidence of saturation by plotting the pairwise corrected versus pairwise uncorrected distances (Plot 1), pairwise uncorrected transition and transversion distances versus pairwise uncorrected total distances (Plot 2), and pairwise uncorrected transition distances versus pairwise uncorrected transversion distances (Plot 3) (Section 2.9.6 of Chapter 2, pp. 77-81). Gene datasets that demonstrated saturation were excluded from subsequent phylogenetic analyses. The sequences were also subjected to the gI measure of skewness test based on 10,000 tree length replicates to determine if there is phylogenetic signal (Section 2.9.7 of Chapter 2, p. 81-82). The presence of paralogous sequences in actin that could affect phylogenetic analyses was detected (Section 2.9.12 of Chapter 2, pp. 89-90) and the utility for of the actin gene for phylogenetic studies assessed (Section 2.9.13 of Chapter 2, pp. 90-91).

Phylogenetic trees were generated from the aligned datasets using the model based maximum likelihood (ML), Bayesian inference (BI) and neighbor-joining (NJ) methods and the non-model based maximum parsimony (MP) method (Section 2.9.8 of Chapter 2, pp. 82-85). Bootstrap resampling (Felsenstein, 1985) with 1000 replicates for ML, MP and NJ were also carried out. Bayesian inference (BI) was performed using four chains of a Markov Chain Monte Carlo algorithm. The number of

generations to explore the tree space and the heating temperature used for each gene were optimised as described in the BI segment of Section 2.9.8 of Chapter 2, p. 83-84 and in Appendix 2.3.2, pp. 369-370. A consensus tree for each gene (where unsaturated) was constructed using the last 1000 trees.

A partition-homogeneity test (Swofford, 2002) was carried out to determine if the sequences from the different genes could be combined into one dataset (Section 2.9.10 of Chapter 2, pp. 86-88). Two sets of concatenated sequences were prepared; one set included only those taxa with complete sequences for all gene fragments while the other set included all taxa.

The Shimodaira-Hasegawa (1999) test was used to test the monophyly of taxonomic groups that were expected to be monophyletic based on taxonomy but did not cluster together in the optimal trees (Section 2.9.11 of Chapter 2, p. 89).

4.3. Results

4.3.1. Molecular data

Twenty nine achatinids and the subulinid *Rumina decollata* were examined for the nuclear rRNA cluster, actin and histone 3 genes as well as the mitochondrial 16S and cytochrome c oxidase I genes. Sequence alignments are presented in Appendices 4.1 – 4.5 on pp. 451-507. All sequences were checked for the presence of ambiguous positions in the direct sequences that might indicate the presence of multiple gene copies. (Ambiguities due to multiple gene copies were detected only in the actin gene; see Section 4.3.2.2, pp. 247-249 for details). A summary of the molecular data for all the sequences is shown in Table 4.1. For the protein-coding genes (actin, histone 3 and COI), information was obtained separately for the entire gene (all codon positions), the

combined 1st and 2nd codon positions and the 3rd codon position only. Uncorrected and corrected pairwise distances and their optimal models were determined for all five genes and are likewise summarized in Table 4.2. The following models were found to be the optimal models for the different datasets: GTR+ Γ for the 16S rRNA gene and the full (all codon positions) of the actin and COI genes as well as the 1st + 2nd codon position and the 3rd codon position datasets of COI and the 3rd codon position dataset of actin; TN93+ Γ for the actin 1st and 2nd codon positions; TN93 for the nuclear rRNA cluster; HKY for the histone 1st and 2nd codon positions; and HKY+ Γ for the histone 3 full and 3rd codon position datasets. (See Appendix 4.6, p. 508 for the summary of the LRT results).

Nearly the entire fragment of the LSU rRNA gene, the internal transcribed spacer 2 (ITS 2) and 80 nucleotides of the 5.8S rRNA gene were sequenced (approximately 4000 nucleotides) for the rRNA gene cluster, of which 3648 nucleotides (including 194 nucleotides of the ITS2) could be aligned unambiguously. A total of 105 (2.9%) variable sites was found, of which 40 were parsimony-informative. Pairwise distances across all taxa ranged from 0 to 0.014 (uncorrected) and 0 to 0.139 (corrected). *Cochlitoma montistempli* and *C. omissa* yielded identical sequences based on the 3648 unambiguously aligned sites, although variations were exhibited between these two taxa in the excluded nucleotides, particularly in the ITS2 region. The highest base frequency for the rRNA gene cluster dataset was obtained for G (0.320) followed by C (0.266), then by A (0.224) and finally by T (0.190). For the actin gene, 873 nucleotide positions were sequenced, of which all sites could be aligned unambiguously; of these, 185 (21.2%) were variable and 84 were parsimony-informative. Pairwise distances ranged from 0 to 0.147 (uncorrected) and 0 to 0.372 (corrected). Highest average base frequency for actin was computed for A (0.268)

followed by T (0.257) then by C (0.254) and finally by G (0.221). Looking at the different codon positions, the majority of the variable sites (161) and parsimony-informative sites (72) were at the 3rd codon position, while the combined 1st and 2nd codon positions only had 24 variable sites and 12 parsimony-informative sites. Amplification of the actin gene for *Limicolariopsis ruwenzoriensis* was unsuccessful. For the histone 3 gene, 328 nucleotides were sequenced and aligned unambiguously, with 61 (18.6%) variable positions and 27 parsimony-informative sites. Pairwise distances ranged from 0 to 0.090 (uncorrected) and 0 to 0.148 (corrected). Five sets of identical sequences were observed for the histone 3 gene; the first set included *Achatina (Lissachatina) fulica*, *A. (L.) reticulata* and *A. (L.) zanzibarica*; the second included *Achatina (Achatina) bisculpta*, *A. (A.) damarensis* and *A. (A.) stuhlmanni*; the third included *Limicolaria kambeul* and *Limicolaria martenssii*; the fourth included *Cochlitoma dimidiata*, *C. montistempli*, *C. omissa*, *C. semidecussata*, *C. simplex* and *C. varicosa*; and the fifth included *C. kilburni* and *C. sp. cf. vestita*. The presence of these sets of identical sequences and the low uncorrected distances demonstrate the conserved nature of the histone 3 gene fragment at the level of the Achatinidae. Average base frequencies for histone 3, in decreasing order, were as follows: C (0.320); G (0.275); A (0.243) and T (0.162). As with the actin gene, the 3rd codon position exhibited the most number of variable sites (57) as well as parsimony-informative sites (26) in comparison to the combined 1st and 2nd codon positions with only 4 and 1, respectively. Amplification of the histone 3 gene fragment for *Achatina (Euaethiopina) loveridgei* and *Limicolariopsis ruwenzoriensis* was unsuccessful. For the mitochondrial COI gene, approximately 650 nucleotides were sequenced, with the length varying depending on the primers used. A total of 641 unambiguously aligned nucleotides were used, of which 293 (45.7%) were variable and 276 were parsimony-informative. Pairwise

distances ranged from 0.067 to 0.264 (uncorrected) and 0.211 to 11.856 (corrected), making the COI gene fragment more variable than the nuclear genes above. No two sequences were found alike, and the smallest uncorrected distance is 0.067, way above the 0.03 sequence divergence threshold for any two species set by Hebert *et al.* (2003) for the COI.

Average base frequencies, in decreasing order, were as follows: T (0.376); A (0.239); G (0.204) and C (0.180), indicating a strong bias for T; this was heightened in the 3rd codon position where average frequency of T rose to 0.404. The 3rd codon position had 214 variable sites and 212 parsimony-informative sites, whereas the combined 1st and 2nd codon positions only had 79 and 64, respectively. Note that sequencing was unsuccessful for *A. (E.) loveridgei* and *Limicolariopsis ruwenzoriensis*. The mitochondrial 16S rRNA gene product ranged from 420-500 nucleotides, with 310 that were unambiguously aligned, 147 (47.4%) that were variable, and 110 that were parsimony-informative. Pairwise distances ranged from 0.019 to 0.242 (uncorrected) and 0.021 and 0.837 (corrected); thus, the gene is likewise more variable than the nuclear genes. As with the COI, no two sequences were found alike. Base frequencies were as follows: T (0.285); A (0.277); G (0.247) and C (0.191), showing bias for A and T. Sequencing of the 16S rRNA gene fragment for *L. ruwenzoriensis* was unsuccessful.

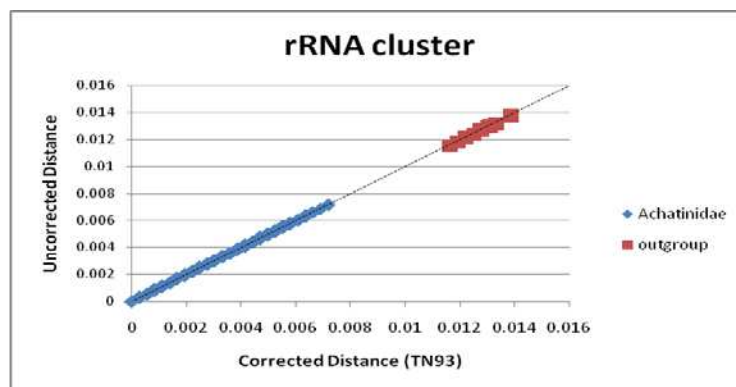
Table 4.2: Summary of molecular data across all genes used (rRNA gene cluster, actin, histone 3, COI and 16S) for the Achatinidae and the subulinid outgroup *Rumina decollata*. A=Achatinidae only; A+O=Achatinidae and outgroup.

	Length (bp)	# of variable sites (%)		Range of distances (Uncorrected)		Optimal Model (α value for Γ distribution)	Range of distances (Corrected)		Average base frequencies								# of parsimony-informative sites	
									A		C		G		T			
									A	A+O	A	A+O	A	A+O	A	A+O		
rRNA cluster	3648	77 (2.1)	105 (2.9)	0-0.007	0-0.014	TN93 (N/A)	0-0.007	0-0.139	0.224	0.224	0.266	0.266	0.320	0.320	0.190	0.190	37	40
Actin combined	873	176 (20.2)	185 (21.2)	0-0.147	0-0.147	GTR+ Γ (0.175)	0-0.372	0-0.372	0.268	0.268	0.254	0.254	0.221	0.221	0.257	0.257	71	84
Actin 1 st and 2 nd codon positions	582	23 (4.0)	24 (4.1)	0-0.031	0-0.031	TN93+ Γ (0.043)	0-0.053	0-0.053	0.311	0.311	0.229	0.229	0.227	0.227	0.233	0.233	11	12
Actin 3 rd codon position	291	153 (52.6)	161 (55.3)	0-0.628	0-0.384	GTR+ Γ (0.686)	0-0.628	0-0.628	0.180	0.179	0.303	0.304	0.211	0.211	0.306	0.306	60	72
Histone3 combined	328	53 (16.2)	61 (18.6)	0-0.090	0-0.090	HKY85+ Γ (0.256)	0-0.148	0-0.148	0.243	0.243	0.320	0.320	0.275	0.275	0.162	0.162	22	27
Histone 3 1 st and 2 nd codon positions	218	4 (1.8)	4 (1.8)	0-0.019	0-0.019	HKY85 (N/A)	0-0.019	0-0.019	0.288	0.288	0.285	0.285	0.261	0.261	0.166	0.166	1	1
Histone 3 3 rd codon positions	110	49 (44.5)	57 (51.8)	0-0.280	0-0.280	HKY85+ Γ (2.967)	0-0.460	0-0.460	0.150	0.151	0.392	0.391	0.305	0.305	0.154	0.154	21	26
COI combined	641	292 (40.9)	293 (45.7)	0.067-0.264	0.067-0.264	GTR+ Γ (0.106)	0.211- 11.856	0.211-11.856	0.239	0.239	0.181	0.180	0.205	0.204	0.376	0.376	275	276
COI 1 st and 2 nd codon positions	427	78 (18.3)	79 (18.5)	0.007-0.098	0.007-0.098	GTR+ Γ (0.072)	0.008- 0.389	0.008- 0.389	0.197	0.197	0.204	0.204	0.236	0.236	0.362	0.362	63	64
COI 3 rd codon positions	214	214 (100)	214 (100)	0.164-0.625	0.164-0.625	GTR+ Γ (0.659)	1.304-120.141	1.304-120.141	0.321	0.323	0.134	0.133	0.142	0.140	0.403	0.404	212	212
16S rRNA	310	145 (46.8)	147 (47.4)	0.019-0.242	0.019-0.242	GTR+ Γ (0.250)	0.021- 0.837	0.021- 0.837	0.277	0.277	0.191	0.191	0.247	0.247	0.285	0.285	107	110

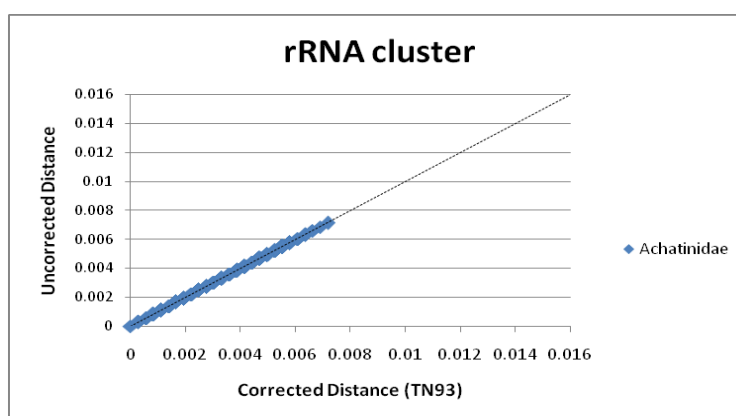
4.3.2. Sequence analyses

4.3.2.1. Evaluating for saturation and phylogenetic signal

For the rRNA cluster, the uncorrected versus corrected distances plots (Plot 1, Figure 4.4) demonstrated a linear relationship between the corrected distance (based on the optimal TN93 model) and the uncorrected (p) distance. This suggested that the dataset for the rRNA cluster was not saturated and that the TN93 model was adequate for correcting the distances for multiple hits.



(A)



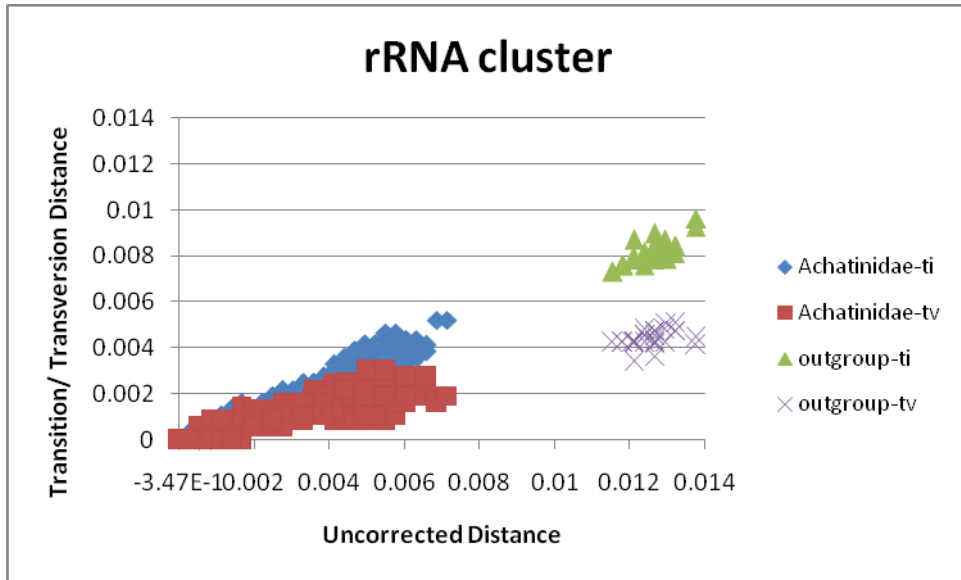
(B)

Figure 4.4: Plots of pairwise uncorrected distance against corrected (TN93) distance for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

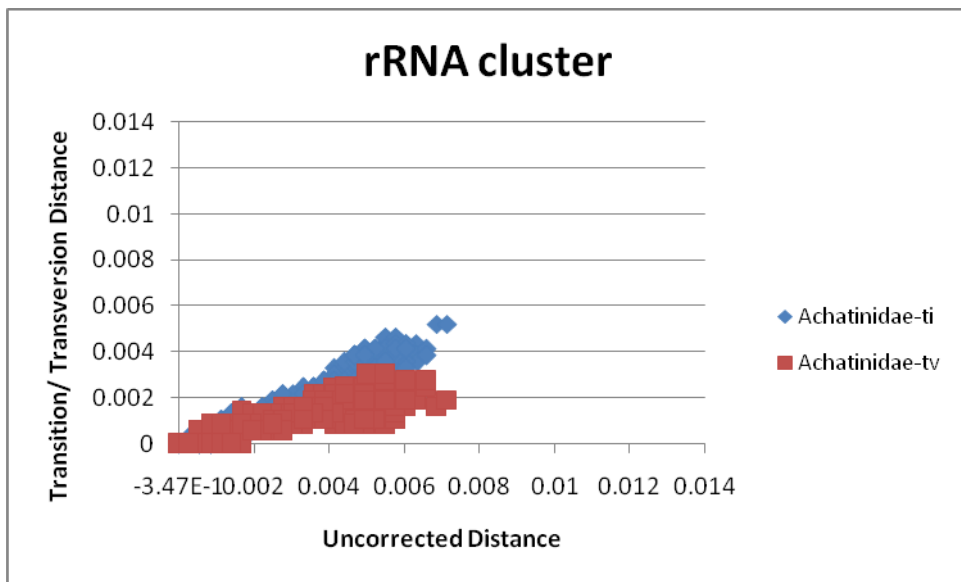
Plots for the transition and transversion rates versus uncorrected distances both with and without the outgroup taxon (Plot 2, Figure 4.5) showed that both transition and transversion substitutions were increasing linearly with transition distances higher than transversion distances. Plots for transitions against transversions both with and without the outgroup taxon (Plot 3, Figure 4.6) confirmed the results of Plot 2 where transition rates were higher than transversion rates. The results suggested that the rRNA cluster dataset was not saturated.

A g_1 value of -0.596 was obtained for the rRNA cluster with 10,000 replicates based on 30 taxa and 40 parsimony-informative characters. This value differed significantly from the critical g_1 value of -0.16 at $P=0.05$ level of significance for 25 taxa and 10 parsimony-informative characters (Hillis & Huelsenbeck, 1992). This result was indicative of a strong phylogenetic signal.

The absence of saturation and the presence of phylogenetic signal implied that the rRNA cluster was suitable for phylogenetic analyses of the Achatinidae.

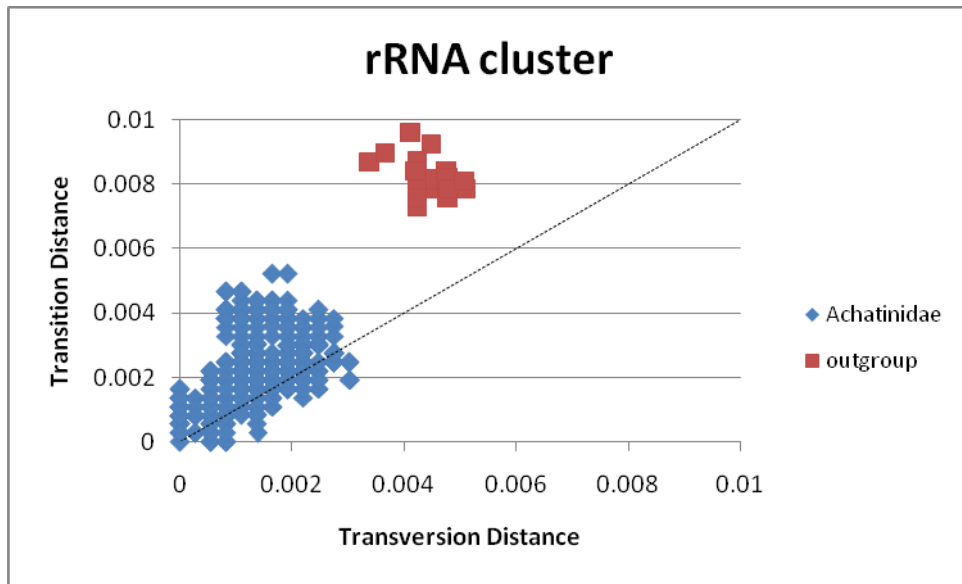


(A)

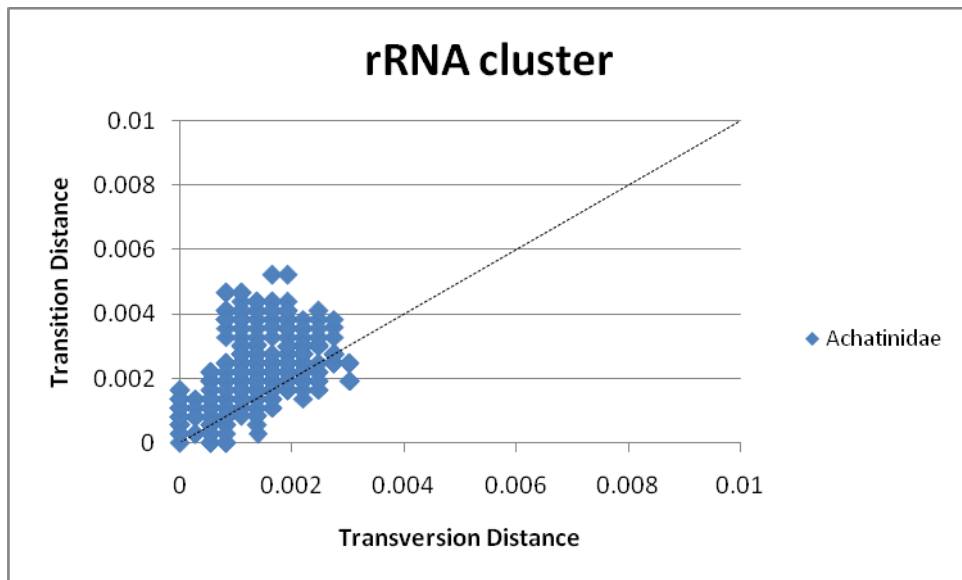


(B)

Figure 4.5: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



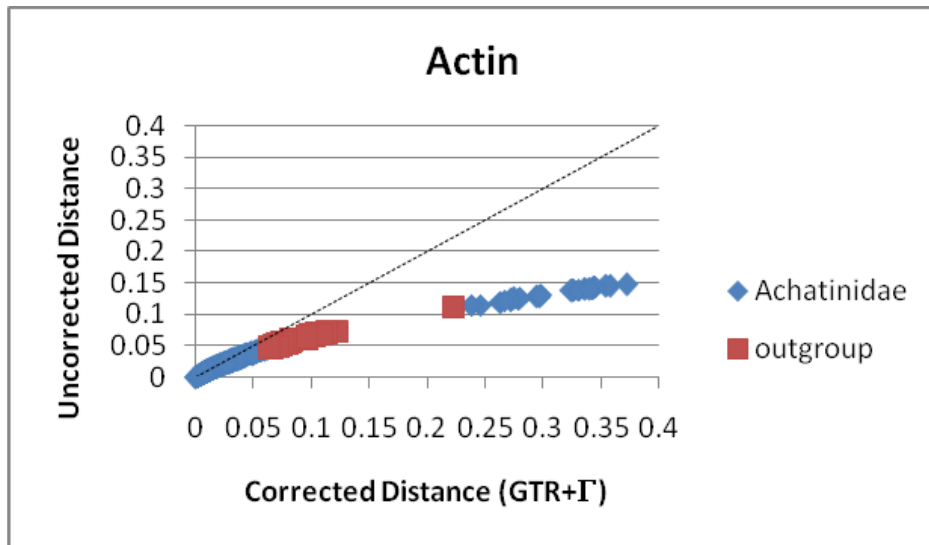
(A)



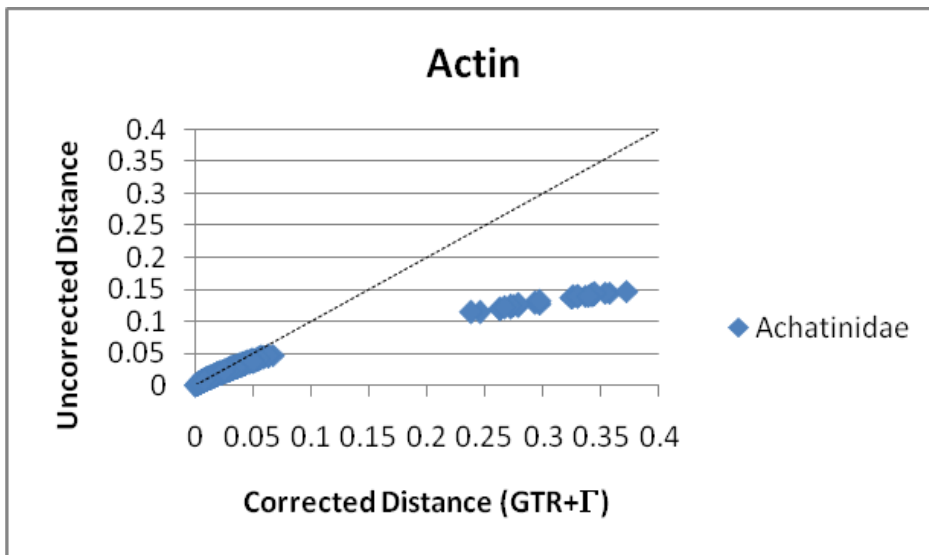
(B)

Figure 4.6: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the rRNA cluster in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

For the actin gene, Plot 1 (Figure 4.7) demonstrated that the corrected distance, based on the optimal GTR+ Γ model, deviated from linearity starting at an uncorrected (p) distance of approximately 0.05 for both the Achatinidae only and the Achatinidae plus outgroup taxon. A big gap was also shown in both plots; the points that came after the gap were attributed to the distance between the actin copy of *Cochlitoma marinae* and those of the other taxa. The presence of this highly divergent actin copy and its implications in the phylogenetic analysis of the Achatinidae using actin is discussed further in Section 4.3.2.2, pp. 247-249. Both plots were still increasing and had not reached a plateau, suggesting adequate correction of the optimal GTR+ Γ model and no saturation in the dataset.



(A)



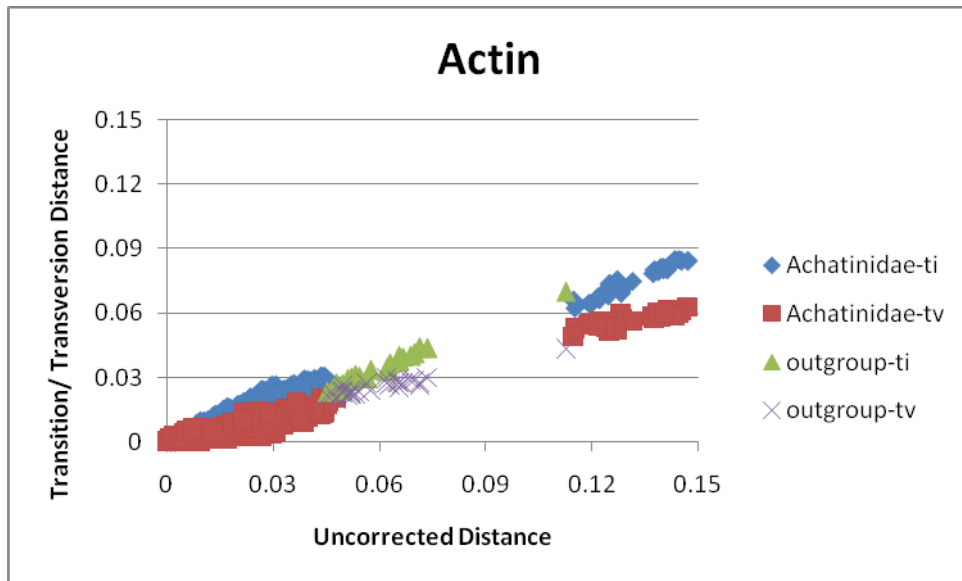
(B)

Figure 4.7: Plots of pairwise uncorrected distance against corrected distance for the actin gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only. Points after the gap were attributed to the divergent actin sequence of *Cochlitoma marinae*.

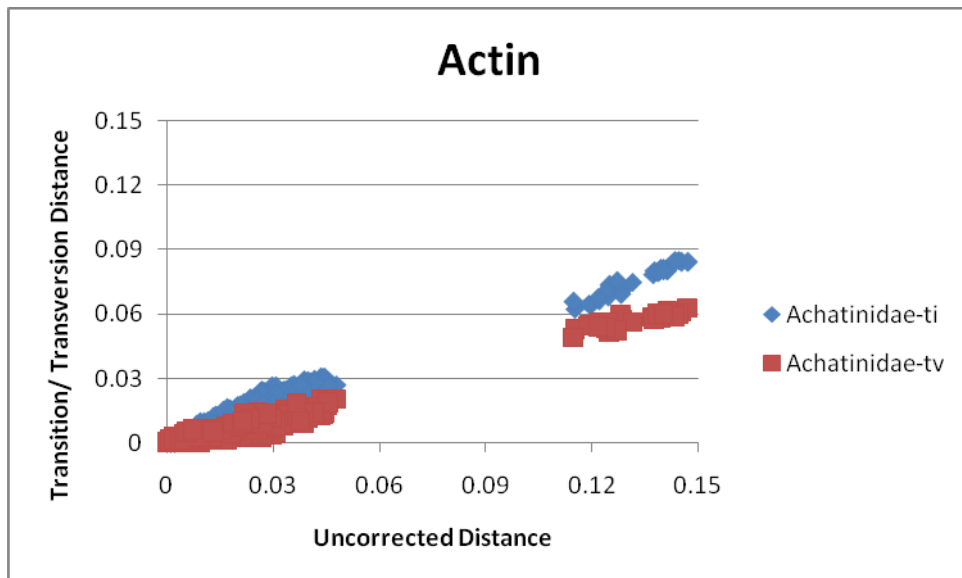
Plot 2 (Figure 4.8) for the actin gene, whether with or without outgroup taxa, exhibited a linear increase for both transitions and transversions, with transition distances higher than transversion distances. Plot 3 (Figure 4.9) confirmed the higher transition rates over the transversion rates. These results suggested that the actin gene was not saturated. Once again, a gap was observed in both plots due to the highly divergent *Cochlitoma marinae*.

A gI value of -0.475 was computed for the actin gene based on 29 taxa and 84 parsimony-informative sites, much lower than the critical value of -0.12 at $p=0.05$ level of significance for 25 taxa and 50 parsimony-informative characters (Hillis & Huelsenbeck, 1992). The actin gene therefore exhibited strong phylogenetic signal.

Lack of saturation and the presence of phylogenetic signal implied that the actin gene was suitable for phylogenetic analyses of the Achatinidae and the outgroup taxon. However, the observed gap in the plots due to the highly divergent sequence of *Cochlitoma marinae* suggested caution must be exercised in using this gene in phylogenetic analyses. These issues are discussed in section 4.3.2.2, pp. 247-249.

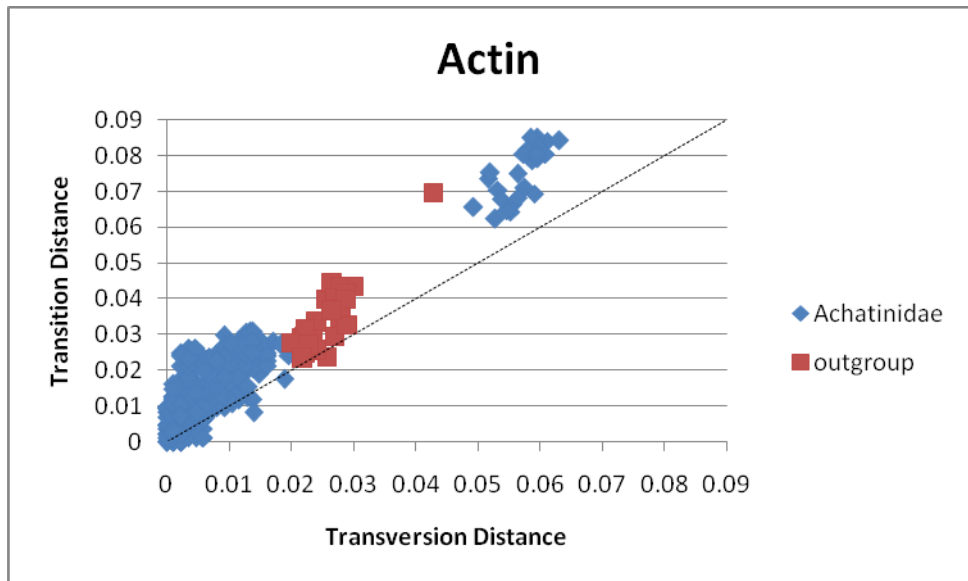


(A)

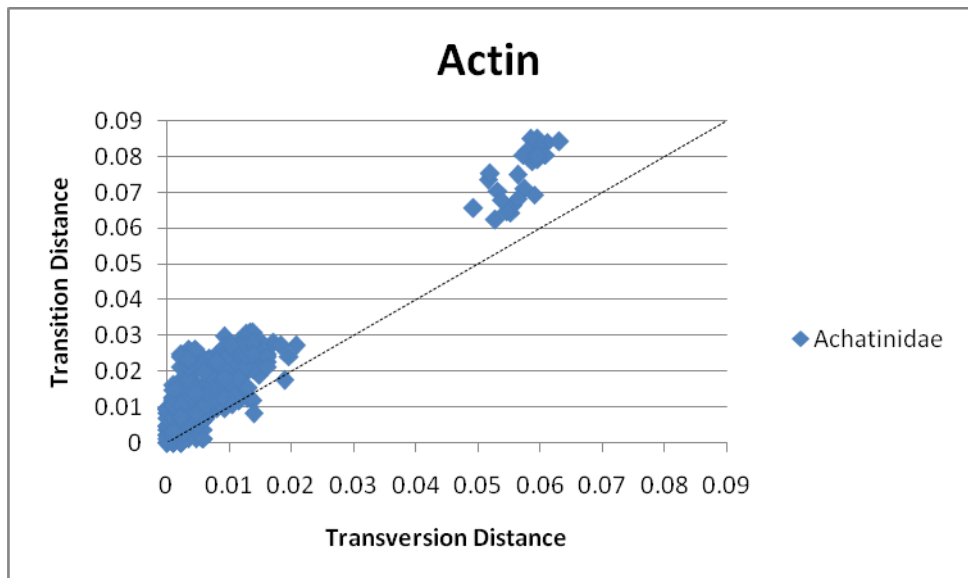


(B)

Figure 4.8: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the actin in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only. Points after the gap were attributed to the divergent actin sequence of *Cochlitoma marinae*.



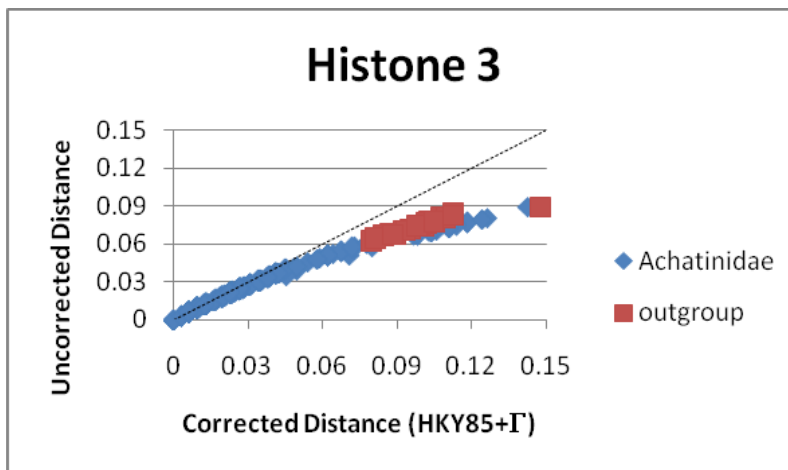
(A)



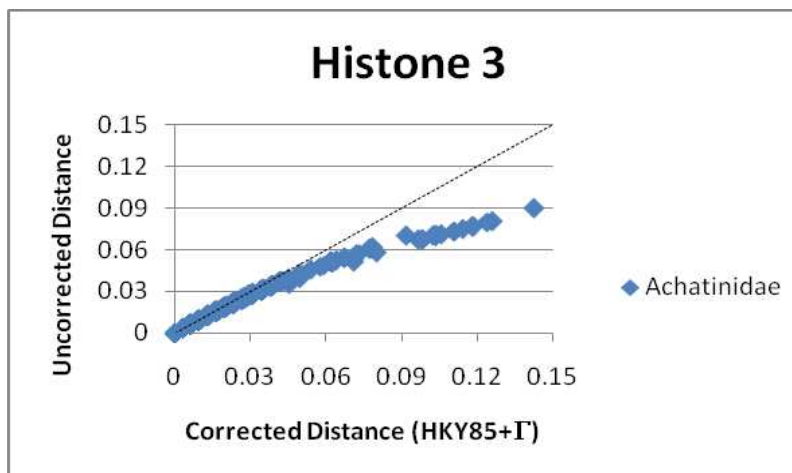
(B)

Figure 4.9: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the actin gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only. Points after the gap were attributed to the divergent actin sequence of *Cochlitoma marinae*.

For the histone 3 gene, Plot 1 (Figure 4.10) revealed a curve for the corrected distances using the optimal GTR+ Γ model, with the deviation from a linear increase beginning at an uncorrected (p) distance of approximately 0.03 for both the Achatinidae only and including the subulinid outgroup taxon. Both plots were still increasing and had not reached a plateau, implying that the histone 3 had not reached saturation.



(A)

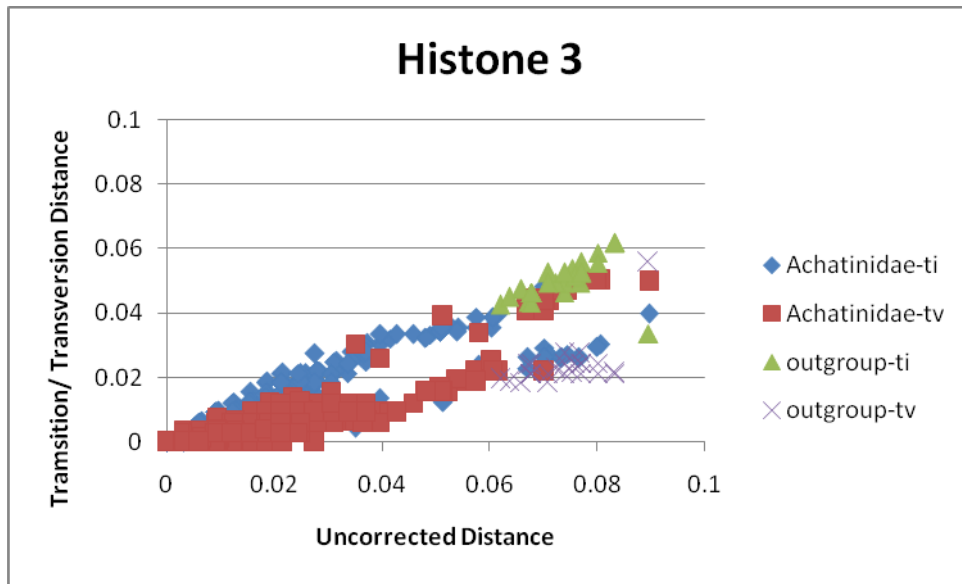


(B)

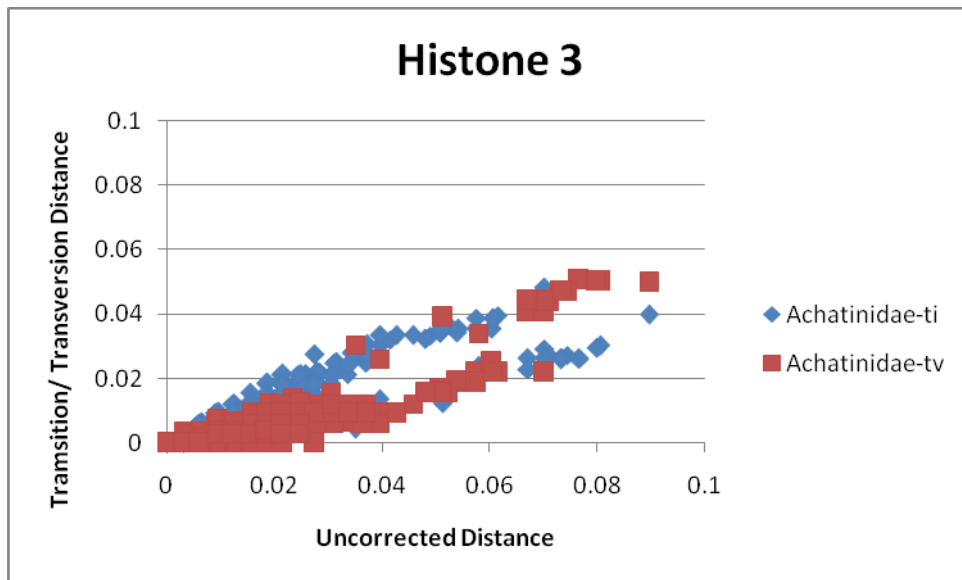
Figure 4.10: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

For plot 2, transitions and transversions were increasing and did not curve for both the Achatinidae only and including the outgroup taxon. Transitions were also generally higher than transversions. However, both the transition and transversion lines appeared to each form two separate lines beginning at approximately 0.035 uncorrected distance in both plots (Figure 4.11). Plot 3 (Figure 4.12) supported the observations from Plot 2, with several pairwise comparisons exhibiting higher transversions than transitions.

The g_1 value for the histone 3 gene based on 28 taxa and 27 parsimony-informative sites was -0.900, significantly smaller than the critical value of -0.16 based on 25 taxa and 10 parsimony-informative sites. The histone 3 gene therefore exhibited phylogenetic signal.

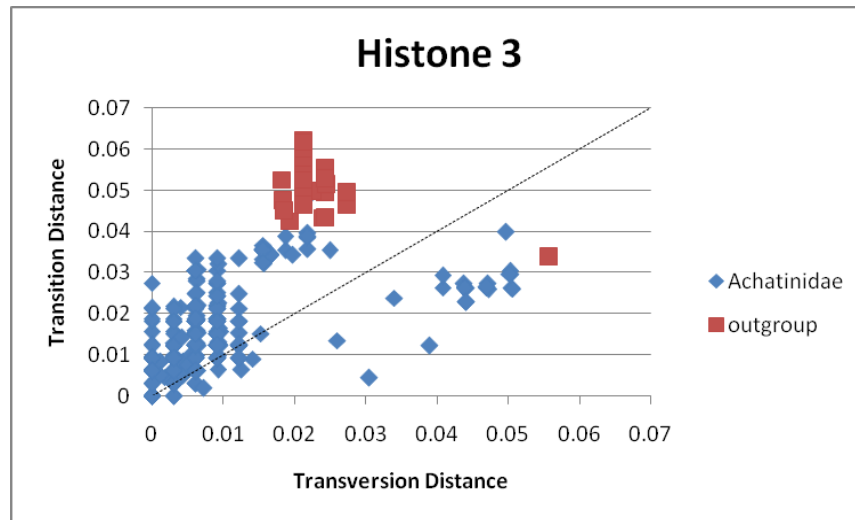


(A)

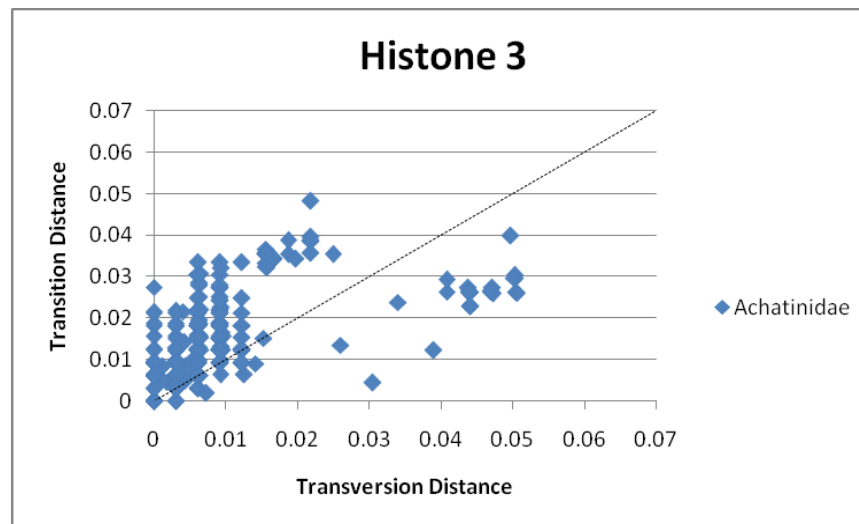


(B)

Figure 4.11: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



(A)

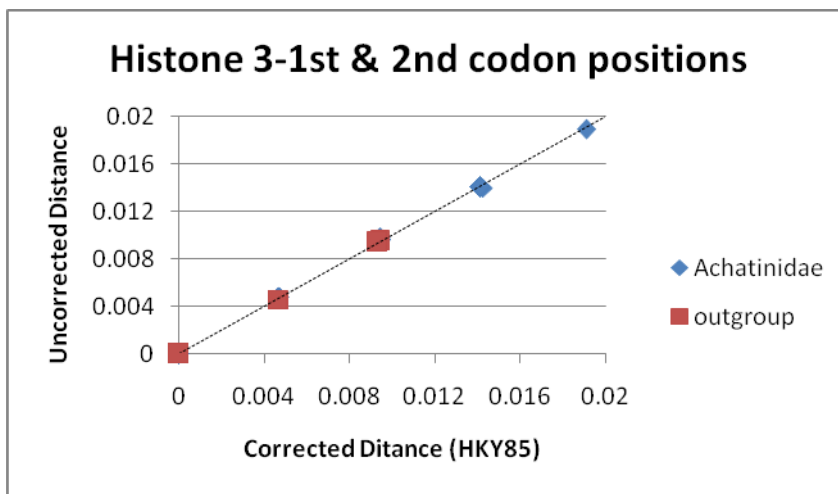


(B)

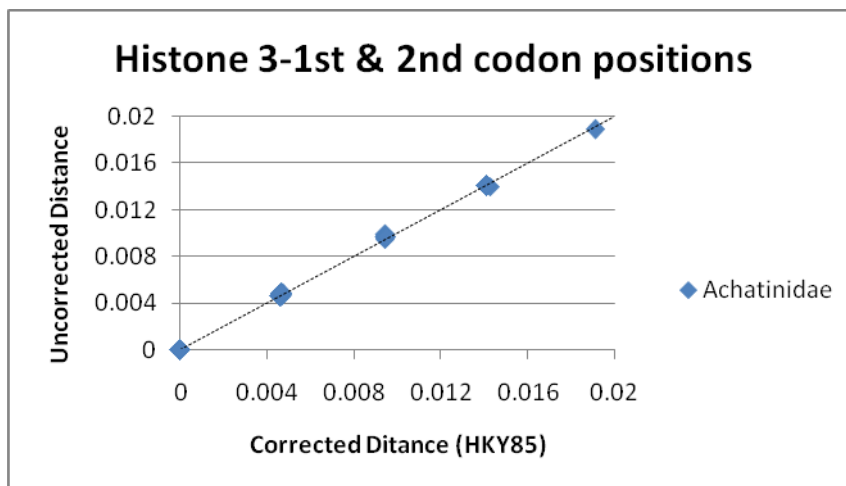
Figure 4.12: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

To account for the distinct lines that separated from the main transition and transversion lines observed in Figure 4.9, to determine if these observations were restricted to the 3rd codon positions, and whether such observations could be associated with saturation, separate plots were made for the combined 1st and 2nd codon positions as well as the 3rd codon positions.

For the 1st and 2nd codon positions, Plot 1 (Figure 4.13) showed a direct relationship between the corrected distances based on the optimal HKY85 model and the uncorrected distances, signifying adequate correction of the model and the absence of saturation. Note, however, that the distances were computed based only on four variable sites.



(A)

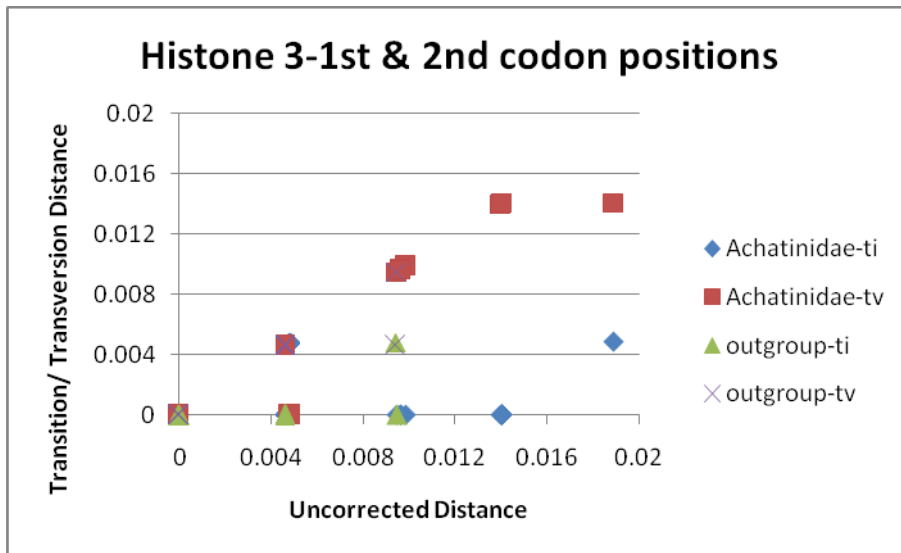


(B)

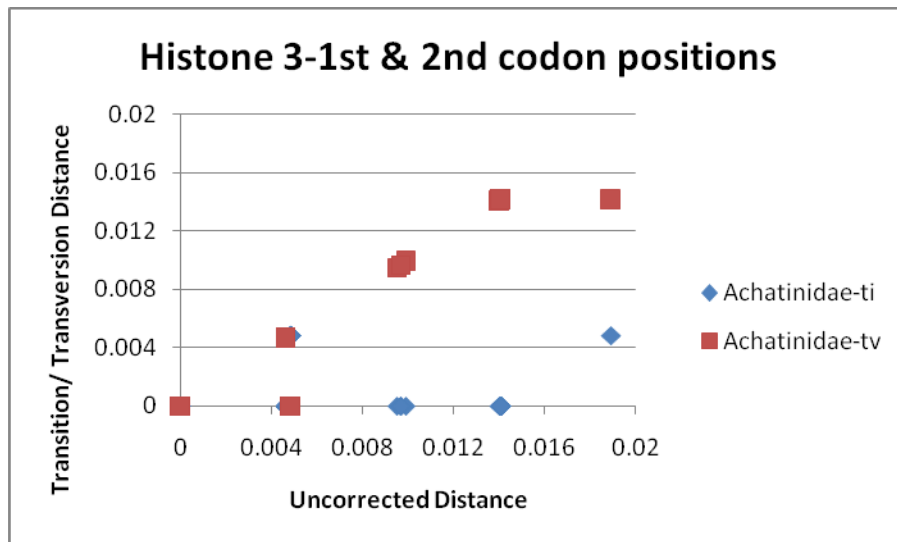
Figure 4.13: Plots of pairwise uncorrected distance versus corrected (HKY85) distance for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

For Plot 2 (Figure 4.14), transversion distances overtook the transition distances, and this was also evident in Plot 3 (Figure 4.15) where there were more transversions than transitions. However, the limited number of variable sites in the 1st and 2nd codon positions made it difficult to assess the presence of saturation through the curving of the plots.

The g1 value for the 1st and 2nd codon positions of histone 3 gene based on 28 taxa and one parsimony-informative site was -0.204. No critical value was available for 25 taxa and at least 2 parsimony-informative sites. The presence of phylogenetic signal could not be assessed based on one parsimony-informative site.

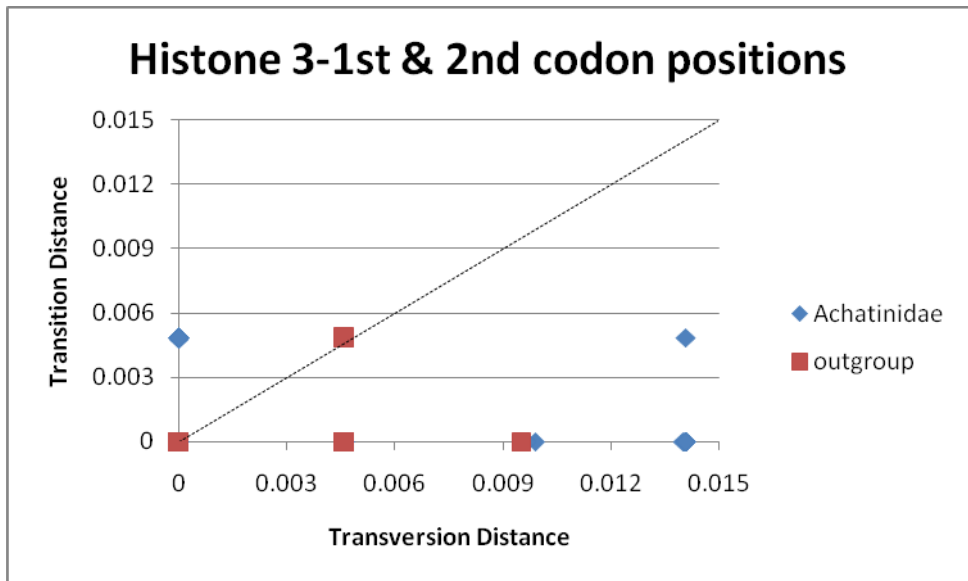


(A)

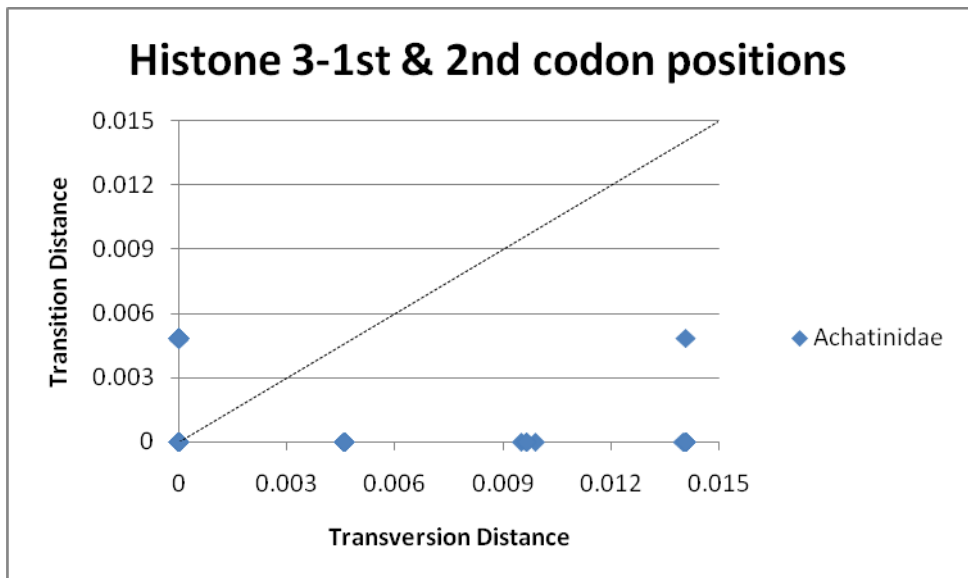


(B)

Figure 4.14: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



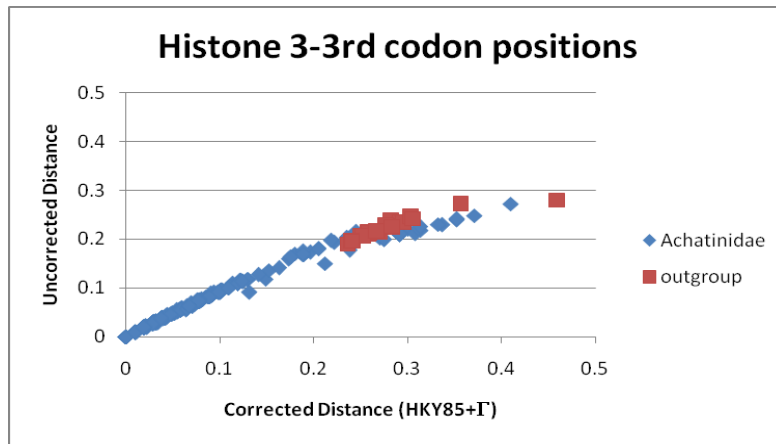
(A)



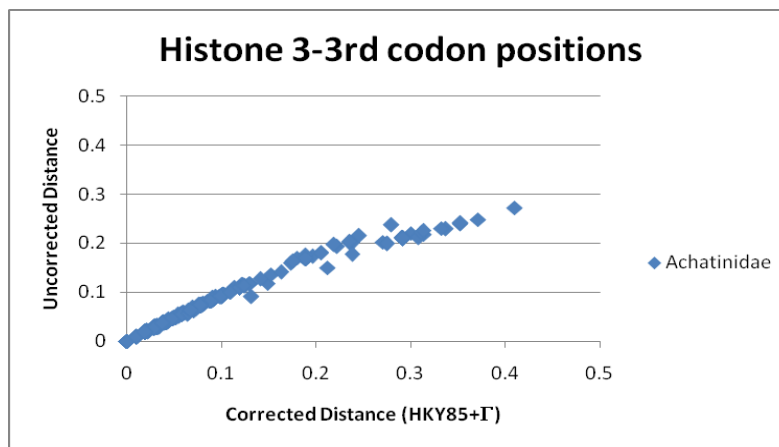
(B)

Figure 4.15: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1st and 2nd codon positions of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

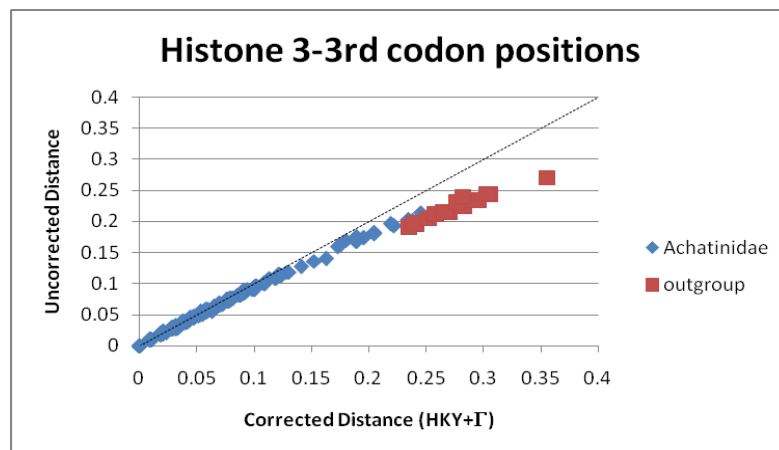
For the 3rd codon position of the histone 3 gene, Plot 1 (Figure 4.16) showed the corrected distances based on the optimal model HKY85+ Γ to be increasing but deviating from linearity at an uncorrected (p) distance of approximately 0.14. However, both plots showed an unmistakable second line starting at 0.13 corrected distance. This second line was attributed to the divergent sequence exhibited by *Archachatina marginata* as supported by the fact that the second line disappeared when *A. marginata* was removed from the plots (see Figure 4.16C). Nevertheless, no plateau was observed, suggesting that adequate correction was made by the optimal model and that the 3rd codon position was not saturated.



(A)



(B)

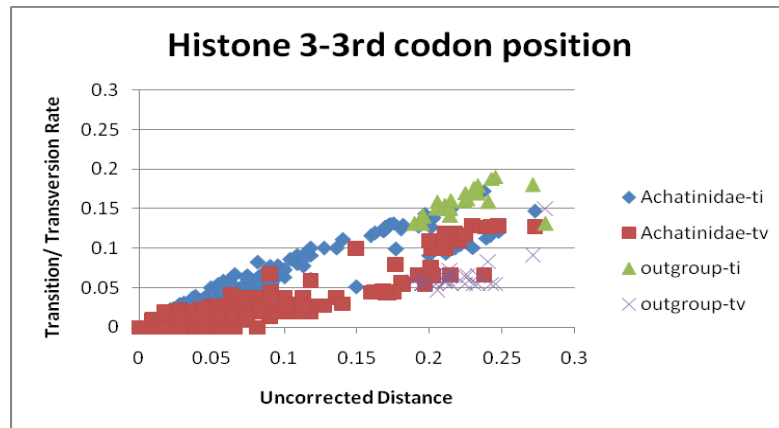


(C)

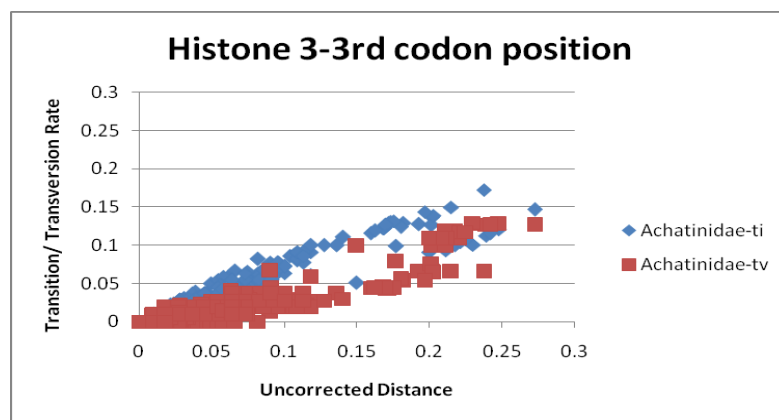
Figure 4.16: Plots of pairwise uncorrected distance versus corrected (HKY85+ Γ) distance for the 3rd codon position of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata*; (B) the Achatinidae only; and (C) Achatinidae and subulinid outgroup *Rumina decollata* but without *Archachatina marginata*.

For Plot 2 (Figure 4.17), the same observations were seen as in Figure 4.11 where both the transition and transversion lines each formed a distinct line beginning at approximately 0.09 uncorrected distance and resulting in a separate transversion line that was higher than the separate transition line in both the Achatinidae only and with the outgroup taxon. These separate lines were attributed to the divergent sequence of *Archachatina marginata*. When this sequence was excluded from the plots (see Figure 4.17C), it became clear that both the transition and transversion lines were increasing with no curving evident. Plot 3 (Figure 4.18) also showed majority of the pairwise comparisons with higher transitions than transversions. Those points with higher transversions than transitions involved *A. marginata* as these points disappeared when *A. marginata* was excluded from the plot (see Figure 4.18C). These results indicated that the 3rd codon position was not saturated.

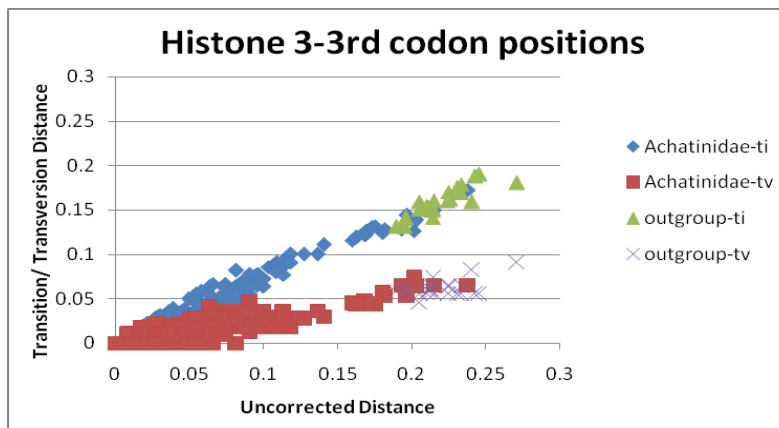
The g1 value for the 3rd codon position of the histone 3 gene based on 28 taxa and 26 parsimony-informative sites was -0.886, which was significantly smaller than the critical value of -0.16 based on 25 taxa and 10 parsimony-informative sites. The 3rd codon position of the histone 3 gene therefore exhibited phylogenetic signal.



(A)

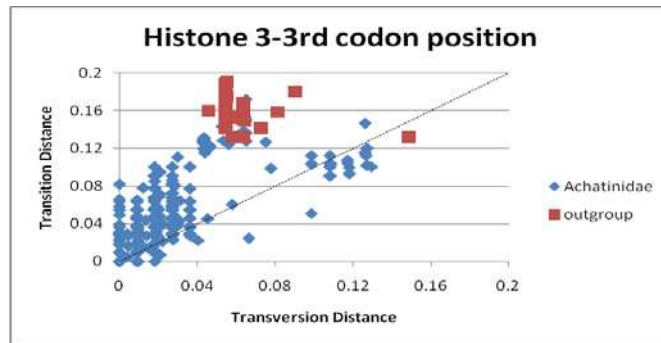


(B)

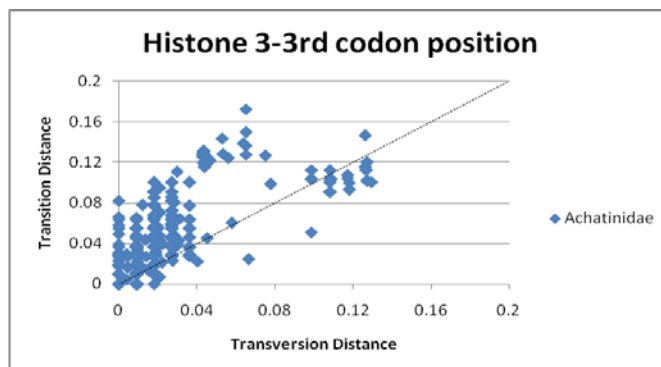


(C)

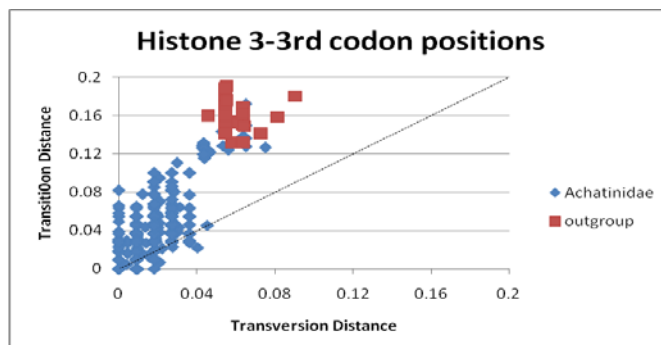
Figure 4.17: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3rd codon position of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata*; (B) the Achatinidae only; and (C) the Achatinidae and the subulinid outgroup *Rumina decollata* but excluding *Archachatina marginata*.



(A)



(B)

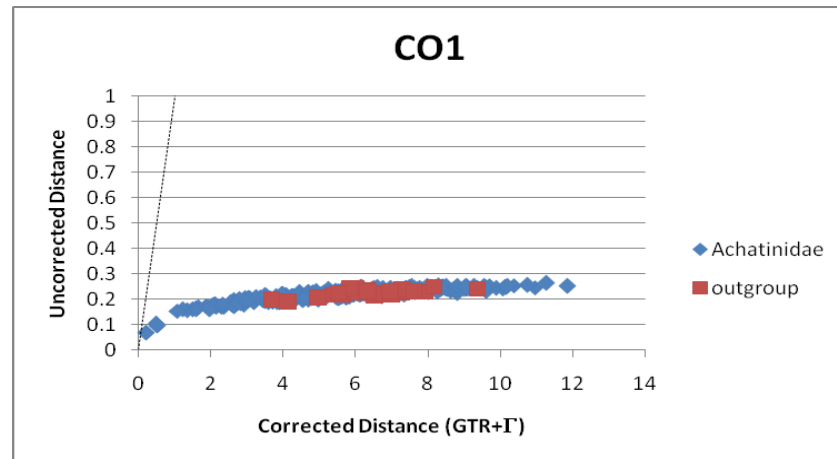


(C)

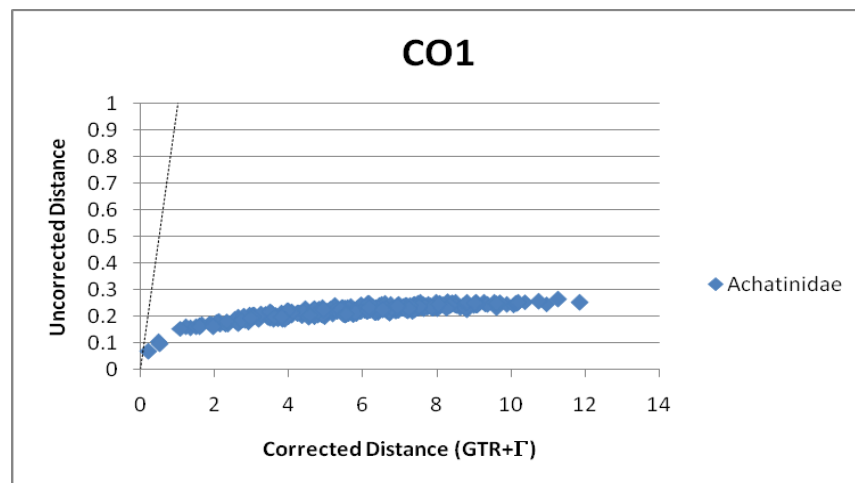
Figure 4.18: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3rd codon position of the histone 3 gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata*; (B) the Achatinidae only; and (C) the Achatinidae and the subulinid outgroup *Rumina decollata* but excluding *Archachatina marginata*.

The findings suggested that the entire histone 3 dataset could be employed for the phylogenetic analyses of the Achatinidae, although the divergent sequence of *Archachatina marginata* should be taken into account when interpreting the trees.

For the COI gene, Plot 1 (Figure 4.19) yielded an almost horizontal trend for all taxa and for the Achatinidae only. This, along with corrected distances of up to 12, indicated severe saturation for the COI gene and the optimal GTR+ Γ model was unable to correct the dataset for multiple hits.



(A)

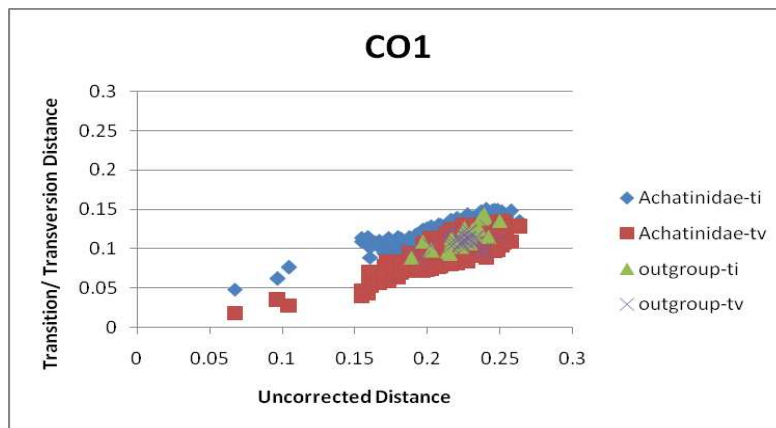


(B)

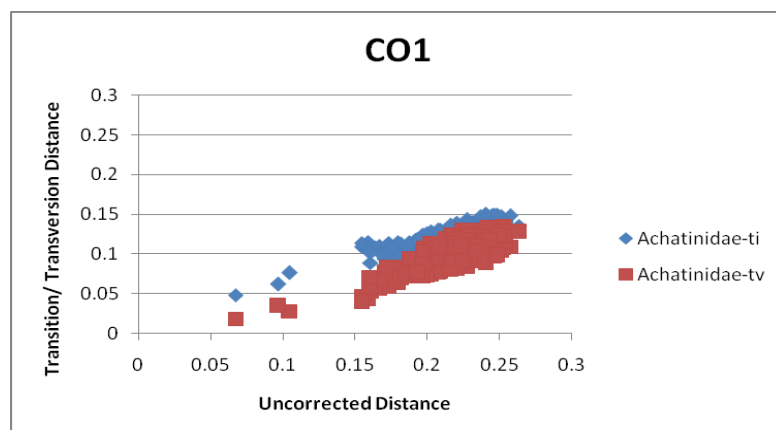
Figure 4.19: Plots of pairwise uncorrected distance against corrected distance for the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Plot 2 (Figure 4.20) for all taxa as well as the Achatinidae only revealed transversions overlapping with transitions. Plot 3 (Figure 4.21) showed approximately a quarter of all pairwise distances showing transversions that were higher than transitions.

The g1 test score for the COI gene based on 28 taxa and 276 parsimony-informative sites was -0.507, significantly smaller than the critical value of -0.08 for 25 taxa and 250 parsimony-informative sites. The g1 test suggested some phylogenetic information could be generated from the dataset.

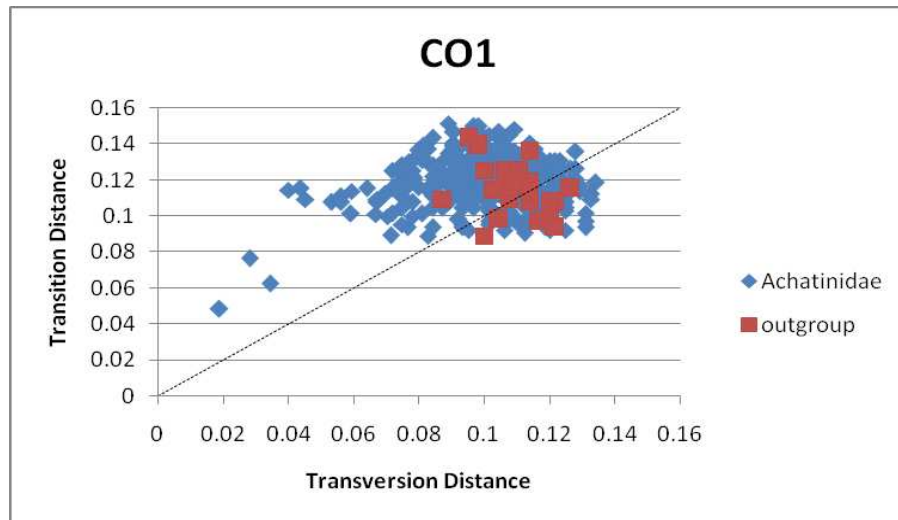


(A)

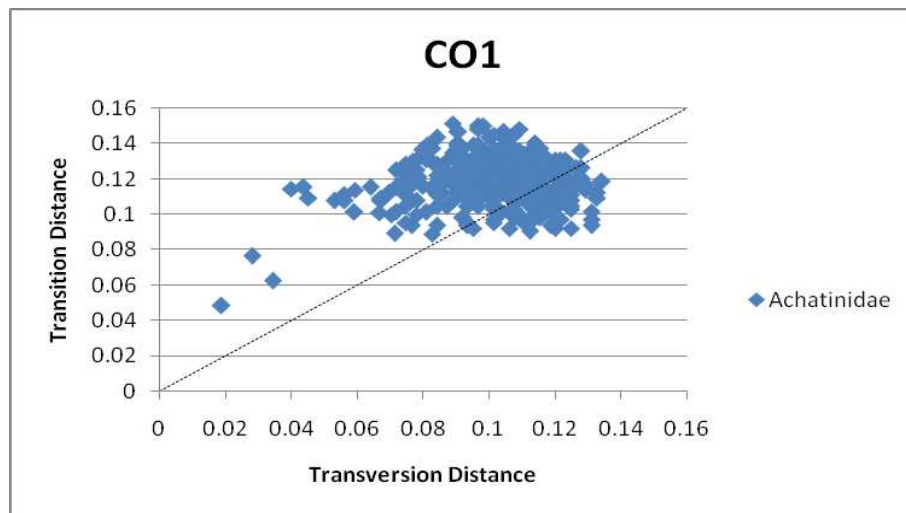


(B)

Figure 4.20: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



(A)

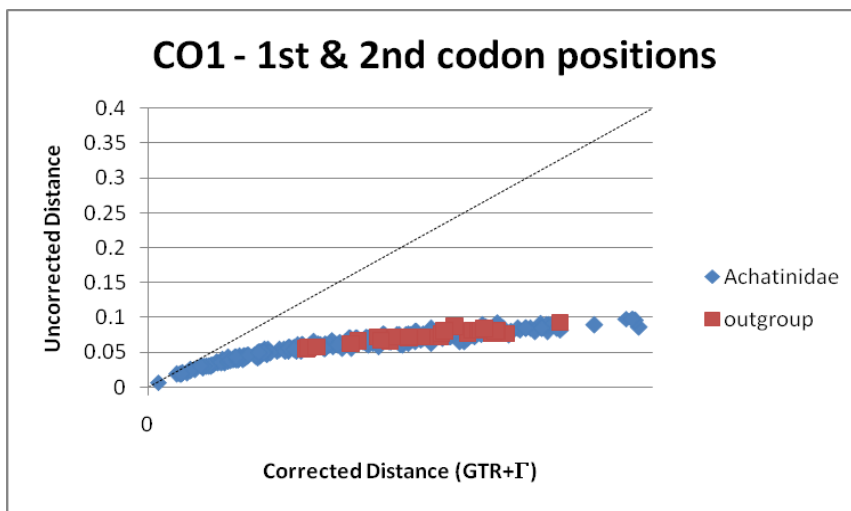


(B)

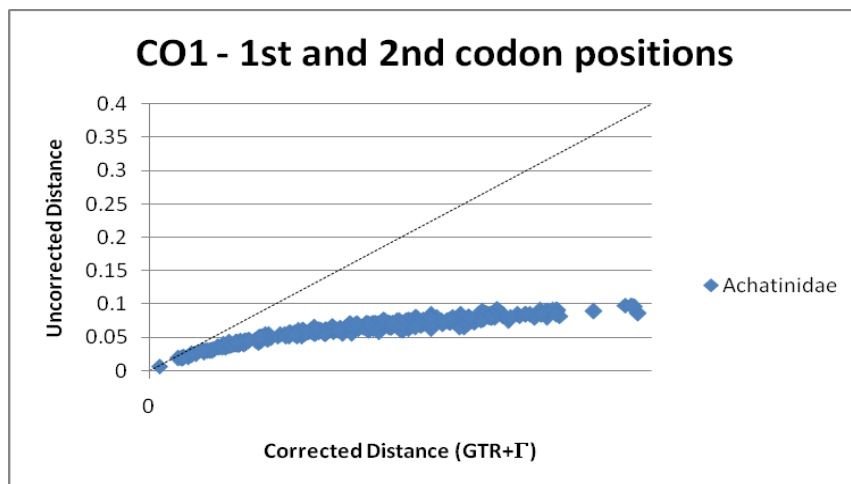
Figure 4.21: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Despite the presence of phylogenetic signal, the problem of saturation remained as shown in Plot 1. In order to evaluate whether saturation of the COI gene was restricted to 3rd codon positions, the combined 1st and 2nd codon positions and the 3rd codon positions were evaluated separately.

For the 1st and 2nd codon positions of the COI gene, Plot 1 (Figure 4.22) revealed a curve for the corrected distances using the optimal GTR+ Γ model, with the deviation from linearity beginning at an uncorrected (p) distance of approximately 0.03 for both the Achatinidae only and including the outgroup taxon. The plots were still increasing slightly and had not reached a plateau, implying that the 1st and 2nd codon positions of COI had not yet reached saturation.



(A)

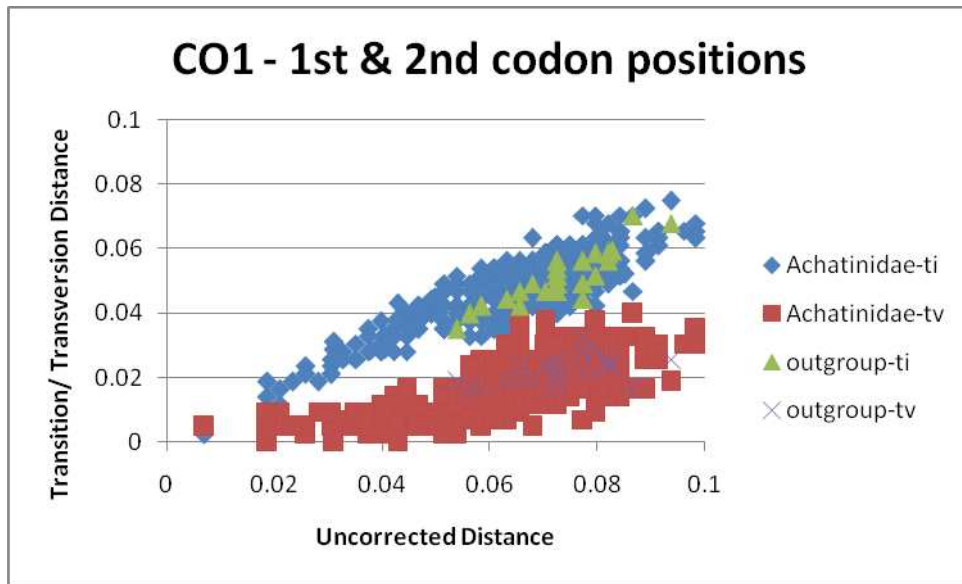


(B)

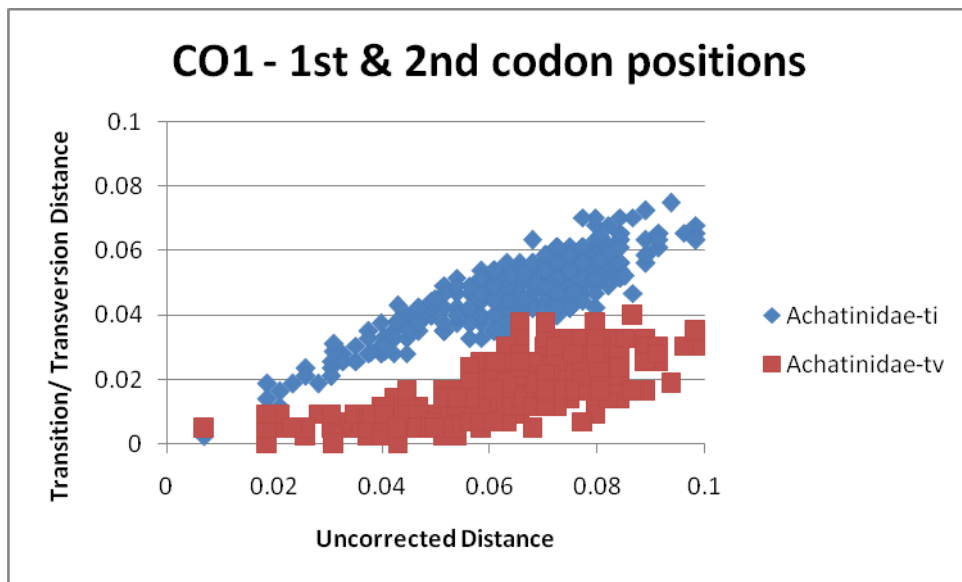
Figure 4.22: Plots of pairwise uncorrected distance versus corrected (GTR+ Γ) distance for the 1st and 2nd codon positions of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Plot 2 (Figure 4.23) for the 1st and 2nd codon positions of the COI gene revealed a linear increase of both transitions and transversions for the Achatinidae and when the outgroup taxon was included, with the transitions being higher than the transversions, while plot 3 (Figure 4.24) revealed that all but three pairwise comparisons had higher rates of transversions than transitions. These findings suggested that the 1st and 2nd codon positions were not saturated.

A g_1 value of -0.309 was obtained based on 28 taxa and 64 parsimony-informative sites, which was significantly smaller than the critical value of -0.12 based on 25 taxa and 50 parsimony-informative sites. The 1st and 2nd codon positions of the COI gene therefore exhibited phylogenetic signal.

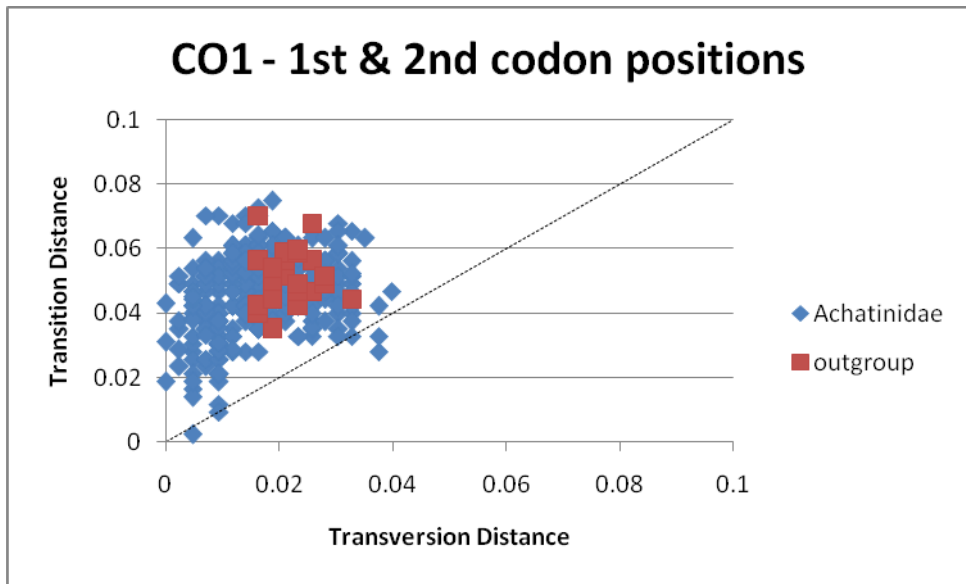


(A)

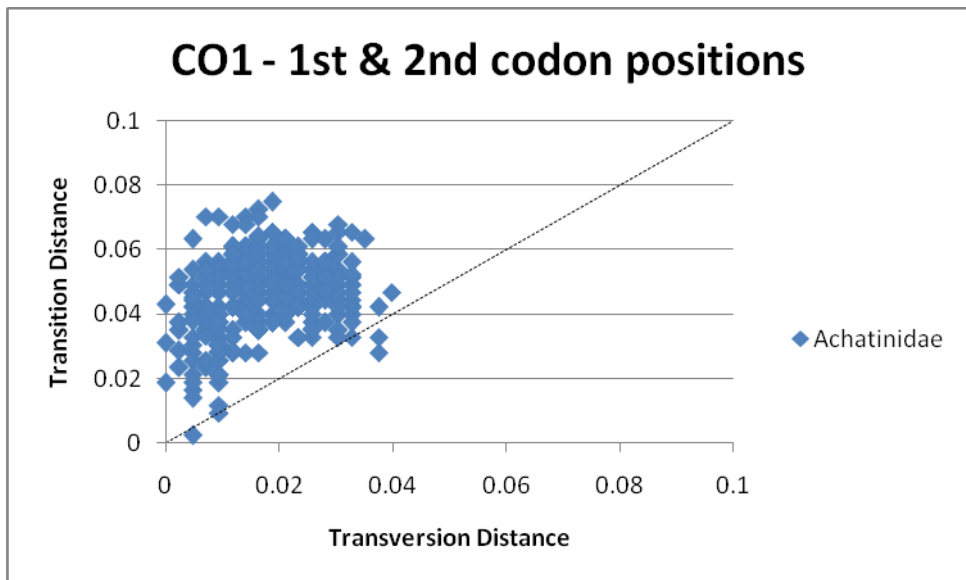


(B)

Figure 4.23: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 1st and 2nd codon positions of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



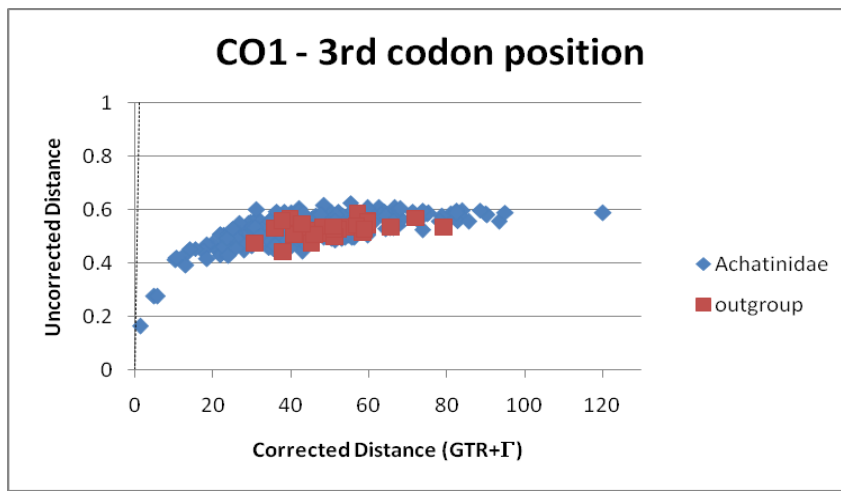
(A)



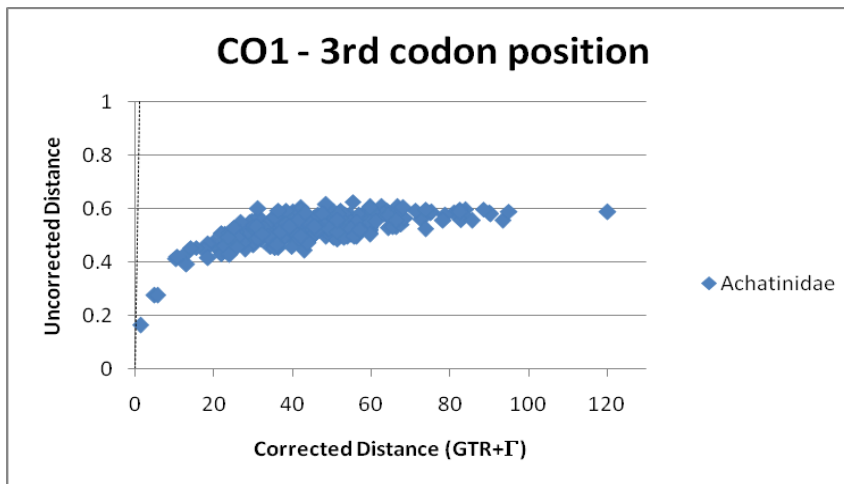
(B)

Figure 4.24: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 1st and 2nd codon positions of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

For the 3rd codon position of the COI gene, Plot 1 (Figure 4.25) showed that the optimal GTR+ Γ model was unable to correct the dataset for multiple hits as demonstrated by the nearly horizontal trend for both plots. The uncorrected distances also ranged from 0.164 to 0.625 while corrected distances ranged from 1.303 to 120.141 (see also Table 4.1). This indicated severe saturation for the 3rd codon position.



(A)

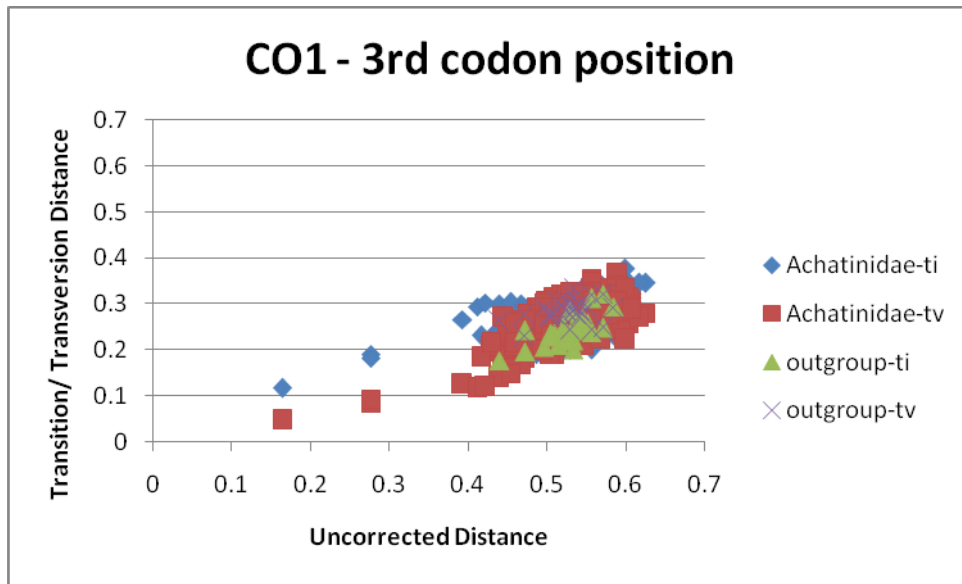


(B)

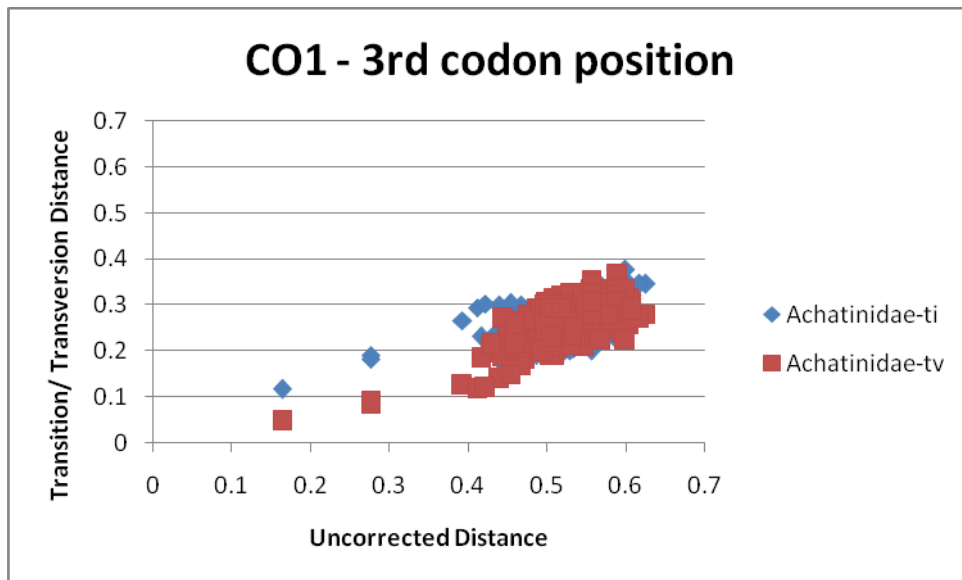
Figure 4.25: Plots of pairwise uncorrected distance against corrected distance for the 3rd codon position of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Plot 2 (Figure 4.26) for all taxa as well as for the Achatinidae only revealed transversions overlapping with transitions. Plot 3 (Figure 4.27) demonstrated that approximately half of the pairwise comparisons had higher rates of transversions than transitions.

The g_1 test score for the COI gene based on 28 taxa and 212 parsimony-informative sites was -0.457, significantly smaller than the critical value of -0.1 for 25 taxa and 100 parsimony-informative sites. The 3rd codon position therefore exhibited phylogenetic signal.

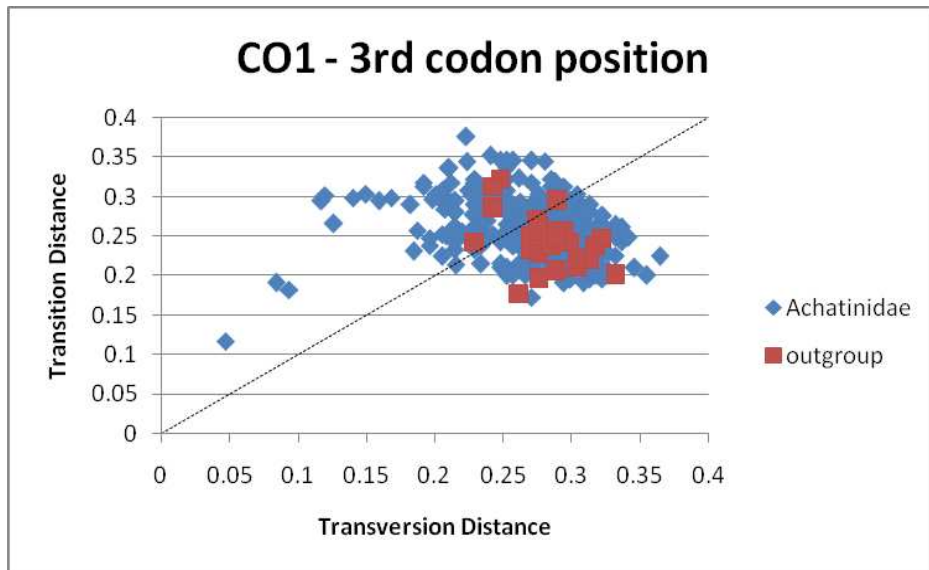


(A)

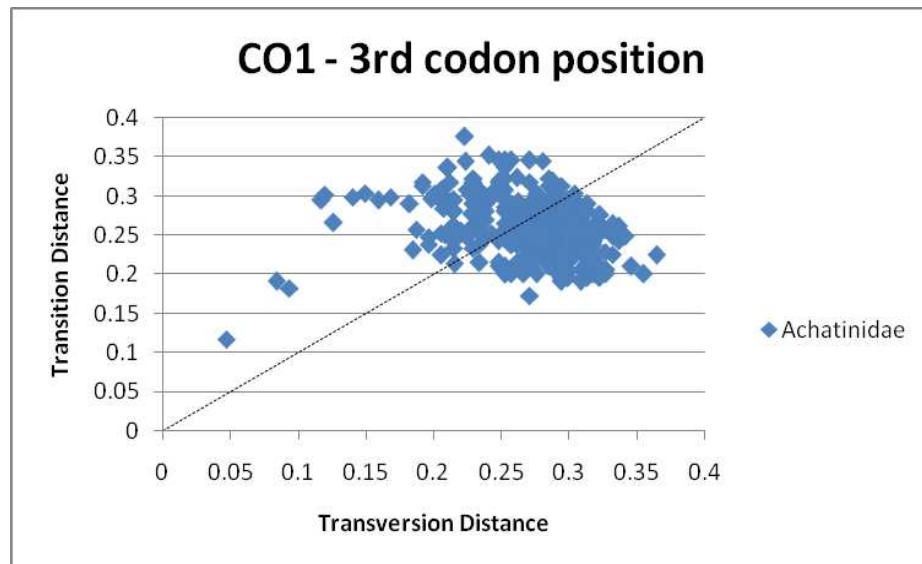


(B)

Figure 4.26: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 3rd codon position of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



(A)

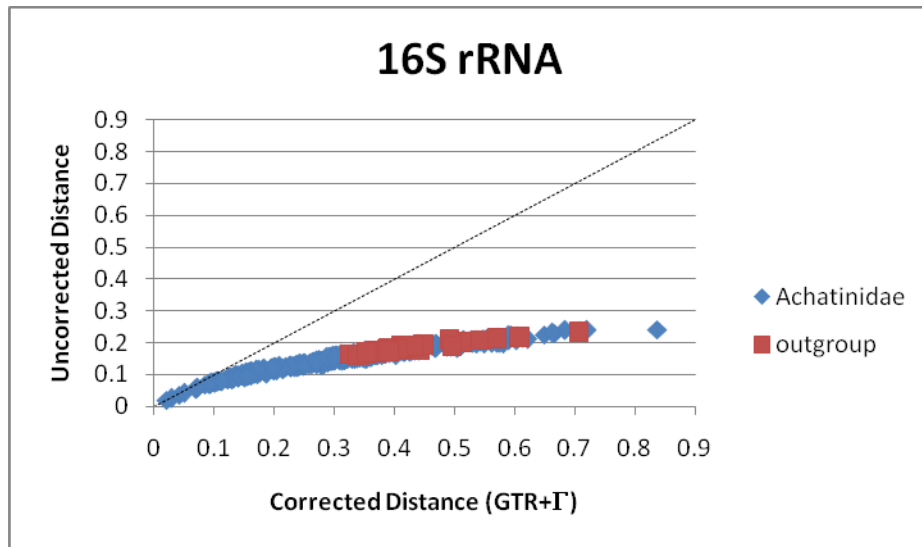


(B)

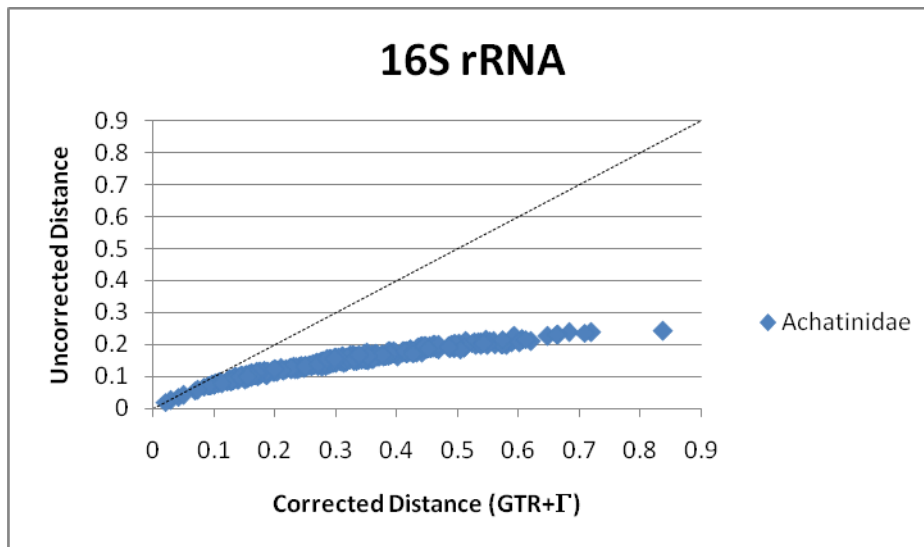
Figure 4.27: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances for the 3rd codon position of the COI gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

Despite the presence of phylogenetic signal as indicated by the g1 test, the strong evidence for saturation of the entire COI gene suggested that the gene should not be used in its entirety for phylogenetic analyses. The results also revealed that saturation was restricted to the 3rd codon position despite showing phylogenetic signal based on the g1 test. These contradictory results from the saturation tests and the g1 test are most likely due to the presence of taxa with very similar sequences that led to a strong left skew of tree length distribution based on parsimony in the g1 test. In these instances, the saturation tests took precedence over the g1 test in deciding whether or not datasets should be included. Phylogenetic analyses for the COI gene were therefore limited to the 1st and 2nd codon positions only.

For the 16S rRNA gene, Plot 1 (Figure 4.28) produced corrected distances based on the optimal GTR+ Γ model that curved from linearity at an uncorrected (p) distance of 0.04 but was still increasing and had not reached a plateau. These findings did not indicate any saturation.



(A)



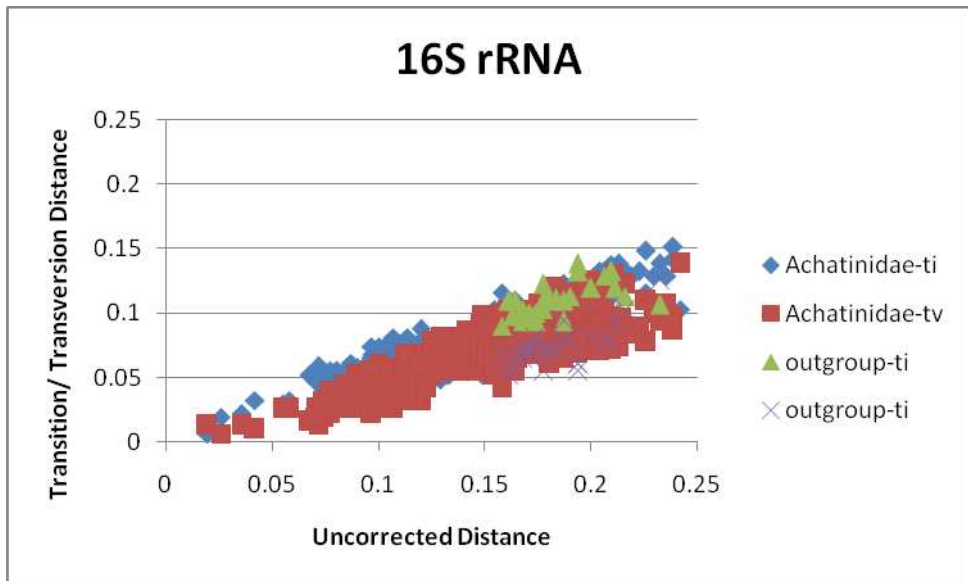
(B)

Figure 4.28: Plots of pairwise uncorrected distance against corrected (GTR+ Γ) distance for the 16S rRNA gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

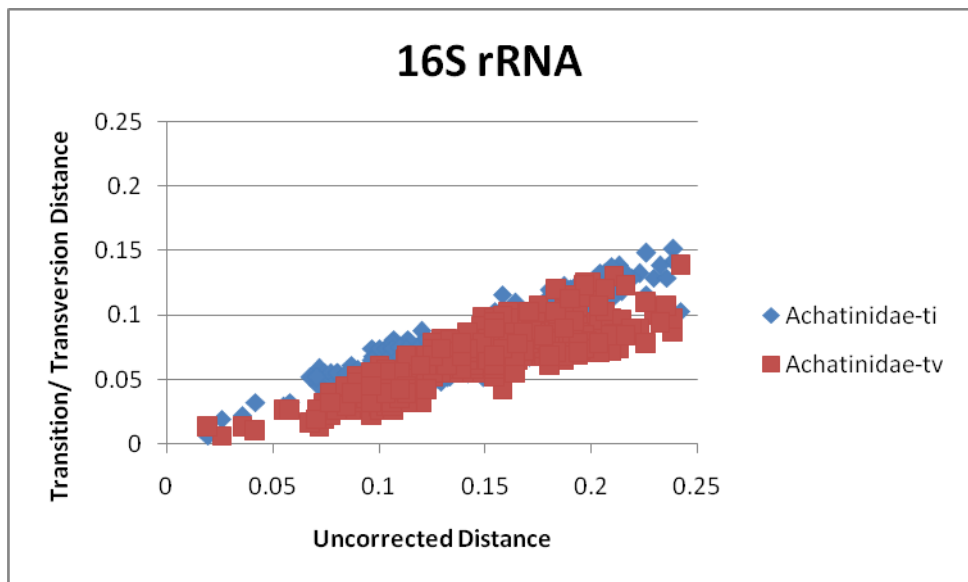
Plot 2 (Figure 4.29) showed an unmistakable overlap between transitions and transversions for the Achatinidae only and including the streptaxid outgroup. However, both plots were still increasing with no evidence of curving for either transitions or transversions. Plot 3 (Figure 4.30) revealed that many pairwise comparisons had higher rates of transversions than transitions, although more than half of the transitions were still higher than transversions.

A g_1 value of -0.507 was obtained based on 29 taxa and 110 parsimony-informative sites, which was significantly smaller than the critical value of -0.1 at $p=0.05$ for 25 taxa and 100 parsimony-informative sites. Phylogenetic signal was therefore evident in the 16S rRNA gene.

The lack of evidence for saturation and the presence of strong phylogenetic signal implied that the 16S rRNA gene could to infer the phylogenetic relationship of the Achatnidae.

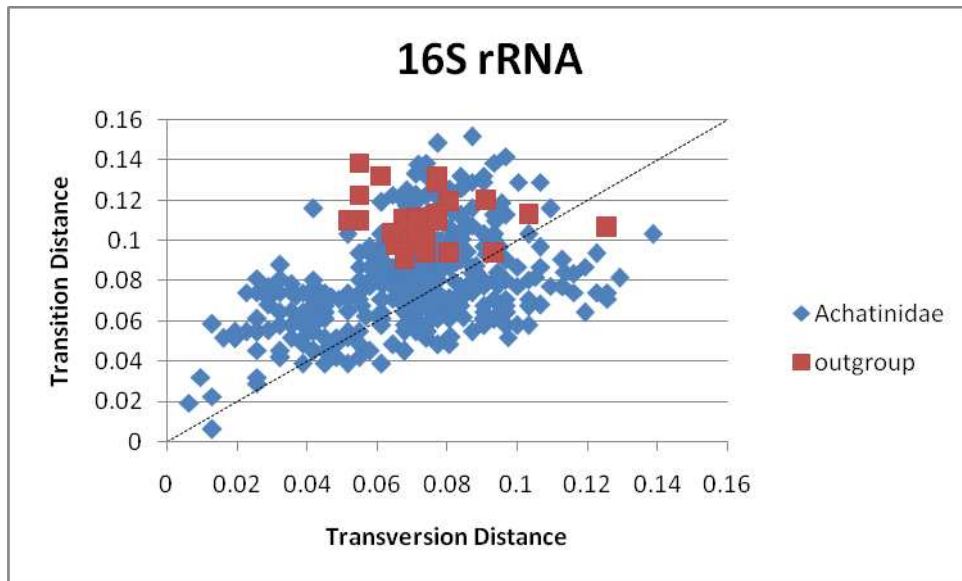


(A)

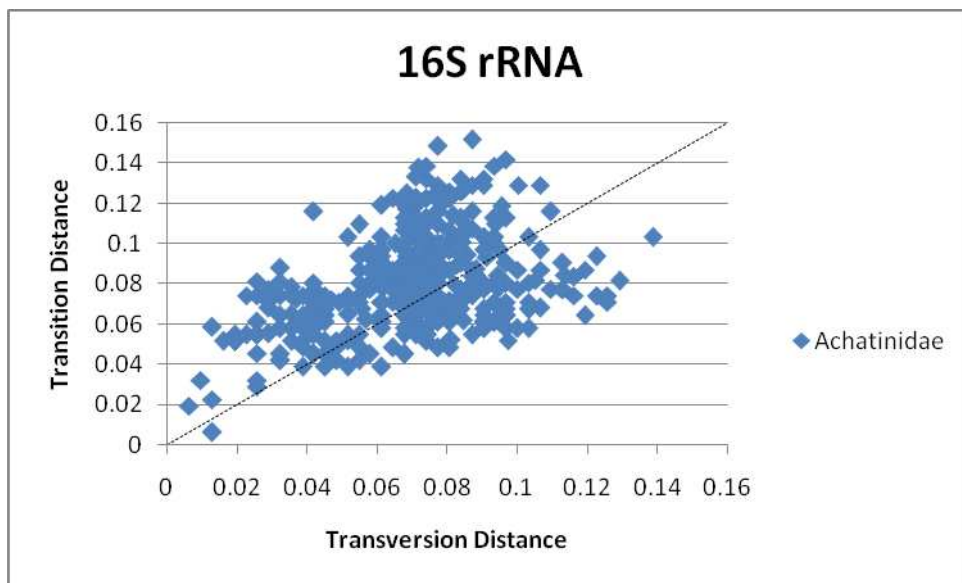


(B)

Figure 4.29: Plots of uncorrected pairwise transition (ti) and transversion (tv) distances against pairwise total uncorrected distances for the 16S rRNA gene in (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.



(A)



(B)

Figure 4.30: Plots of uncorrected pairwise transition (ti) distances against transversion (tv) distances (A) the Achatinidae and the subulinid outgroup *Rumina decollata* and (B) the Achatinidae only.

4.3.2.2. Inferring the presence of paralogous sequences in the actin dataset and assessing its utility for phylogenetic analyses

To infer the presence of paralogous sequences, the actin sequences (obtained by direct sequencing of PCR products) were examined for ambiguous sites. Ambiguous sites were detected in 22 out of 28 achatinid taxa (Appendix 4.7, p. 509), which implied the presence of more than one copy of the actin gene in the Achatinidae. The presence of multiple actin copies was further supported when seven distinct cloned sequences were observed for *Cochlitoma granulata* (see Appendix 3.8, pp. 435-437; note that the clone data for *Cochlitoma granulata* is also discussed as part of the discussion on paralogy of the actin gene in the Achatinoidea, Section 3.3.2.2, Chapter 3, pp. 155-159). A total of 72 variable sites were detected among the seven clones as opposed to only 9 ambiguities in the direct sequence. The majority of these variable sites (55) were attributed to Clones 3 and 6 that were not picked up by the direct sequence. As discussed in chapter 3, the reason for the failure of the direct sequences to identify as ambiguous sites all of the variation among the clones is not clear, but as the PCR-direct sequence is effectively a consensus of the different actin copies within an individual, it is possible that rare copies of the gene might be averaged out.

Four approaches were carried out to determine whether the observed paralogy in actin was likely to mislead phylogeny. First, a neighbor-joining tree of all the actin sequences and the clones of *C. granulata* was generated to determine if the clones were monophyletic. As discussed previously in Section 3.3.2.2 of Chapter 3, the results revealed that some clones (Clones 1, 2, 4 and 5) formed a monophyletic group together with the direct sequence for *C. granulata* while others (Clones 3 and 6) fell elsewhere in the tree (see also Appendix 3.9, p. 438). Second, pairwise comparison between closely related taxa across the Achatinidae based on actin sequence identity (i.e. least number of

nucleotide differences) showed that for some pairwise comparisons, the number of ambiguous sites for at least one taxon in the pair were either very close to the number of total nucleotide differences (i.e. *Achatina (Achatina) achatina* and *Archachatina marginata*; *Achatina (Achatina) damarensis* and *Metachatina kraussi*; and *Cochlitoma montistempli* and *C. omissa*) or were larger than the total differences (i.e. *Achatina (Achatina) damarensis* and *Atopocochlis exarata*; *C. dimidiata* and *Atopocochlis exarata*; *C. varicosa* and *C. sp. cf. vestita*; *Limicolaria kambeul* and *L. martenssii*; and *Limicolariopsis sp.* and *Achatina (Achatina) damarensis*). (See also Table 4.3.) These observations suggested that the divergence among the actin copies within some taxa were just as deep if not deeper than the divergence of the taxa themselves and that phylogenetic signal could be obscured at and above the genus level for these taxa. Third, a partition homogeneity test was supposed to evaluate whether or not the actin sequences exhibited significant difference in the phylogenetic signal with respect to the other genes. However, the test was not successfully carried out due to the polytomies exhibited by the actin gene using maximum parsimony. It was therefore not possible to assess the utility of the actin sequence for phylogenetic analyses in the Achatinidae using the partition homogeneity test. Last, the actin phylogeny was evaluated for concordance with the rRNA gene cluster phylogeny, and results showed little concordance. For example, *Cochlitoma marinae* had a very divergent sequence relative to the other achatinids, particularly the other Southern African *Cochlitoma* species; it fell at the base of the Achatinidae (see Appendix 4.8 on p. 510 for the actin phylogenies and Figure 4.31A on p. 252 for the rRNA gene phylogeny). Based on these results, the presence of multiple copies of the actin gene was having a significant effect on phylogeny; the actin gene was therefore excluded from further phylogenetic analyses.

Table 4.3. Comparison of the actin ambiguous sites and nucleotide differences for each achatinid and its closest relative. Closest relative for each taxon is determined based on the actin sequence identity (least number of nucleotide differences). Note that in scoring for the total differences, comparison between an ambiguous site and an unambiguous site is counted as different even if one of the possible nucleotides for the ambiguous site is the same as that found in the unambiguous site being compared (e.g. A vs. R, which is either A or G). See also Table 2.8 of p. 73 for assignment of ambiguous positions.

Species (total ambiguous sites)	Closest relative with least number of differences (total ambiguous sites)	Total number of nucleotide differences
WEST AFRICAN		
<i>Achatina (Achatina) achatina</i> (45)	<i>Archachatina marginata</i> (27)	48
<i>Archachatina marginata</i> (27)	<i>Achatina (Achatina) achatina</i> (45)	48
EAST AFRICAN		
<i>Achatina (Lissachatina) fulica</i> (18)	<i>Achatina (Lissachatina) immaculata</i> (15)	27
<i>Achatina (Lissachatina) immaculata</i> (15)	<i>Achatina (Lissachatina) fulica</i> (18)	27
<i>Achatina (Lissachatina) reticulata</i> (33)	<i>Achatina (Euaethiopina) loveridgei</i> (0)	39
<i>Achatina (Lissachatina) zanzibarica</i> (1)	<i>Achatina (Euaethiopina) loveridgei</i> (0)	13
<i>Achatina (Euaethiopina) loveridgei</i> (0)	<i>Achatina (Lissachatina) zanzibarica</i> (1)	13
OTHERS		
<i>Achatina (Achatina) bisculpta</i> (32)	<i>Limicolaria kambeul</i> (30)	49
<i>Achatina (Achatina) damarensis</i> (34)	<i>Atopocochlis exarata</i> (29)	33
<i>Achatina (Achatina) stuhlmanni</i> (2)	<i>Cochlitoma churchilliana</i> (0) <i>Cochlitoma montisptempli</i> (0) <i>Cochlitoma ustulata</i> (0) <i>Cochlitoma zebra</i> (0)	29
<i>Atopocochlis exarata</i> (29)	<i>Achatina (Achatina) damarensis</i> (34)	33
<i>Limicolaria kambeul</i> (30)	<i>Limicolaria martenssii</i> (33)	31
<i>Limicolaria martenssii</i> (33)	<i>Limicolaria kambeul</i> (30)	31
<i>Limicolariopsis</i> sp. (40)	<i>Achatina (Achatina) damarensis</i> (34)	42
SOUTHERN		
<i>Cochlitoma varicosa</i> (27)	<i>Cochlitoma</i> sp. cf. <i>vestita</i> (30)	18
<i>Cochlitoma dimidiata</i> (31)	<i>Atopocochlis exarata</i> (29)	33
<i>Cochlitoma</i> sp. cf. <i>vestita</i> (30)	<i>Cochlitoma varicosa</i> (27)	18
<i>Cochlitoma marinae</i> (4)	<i>Cochlitoma semidecussata</i> (0)	124
<i>Cochlitoma churchilliana</i> (0)	<i>Cochlitoma zebra</i> (0)	8
<i>Cochlitoma kilburni</i> (1)	<i>Cochlitoma zebra</i> (0)	5
<i>Cochlitoma granulata</i> (9)	<i>Cochlitoma omissa</i> (6) <i>Cochlitoma semidecussata</i> (0) <i>Cochlitoma ustulata</i> (0)	15
<i>Cochlitoma simplex</i> (2)	<i>Cochlitoma zebra</i> (0)	10
<i>Cochlitoma montisptempli</i> (0)	<i>Cochlitoma omissa</i> (6)	5
<i>Cochlitoma omissa</i> (6)	<i>Cochlitoma montisptempli</i> (0)	5
<i>Cochlitoma semidecussata</i> (0)	<i>Cochlitoma ustulata</i> (0)	11
<i>Cochlitoma ustulata</i> (0)	<i>Cochlitoma churchilliana</i> (0) <i>Cochlitoma semidecussata</i> (0) <i>Cochlitoma zebra</i> (0)	11
<i>Cochlitoma zebra</i> (0)	<i>Cochlitoma kilburni</i> (1)	5
<i>Metachatina kraussi</i> (52)	<i>Achatina (Achatina) damarensis</i> (34)	59
SUBULINIDAE (OUTGROUP)		
<i>Rumina decollata</i> (29)	<i>Cochlitoma churchilliana</i> (0)	82

4.3.2.3. Partition homogeneity test for combining datasets

A partition homogeneity test was done separately for the dataset incorporating the rRNA cluster, histone 3, the 1st and 2nd codon positions of the COI, and the 16S rRNA (5586 nucleotides total) for all taxa with complete DNA sequences as well as the dataset that excluded actin (4713 total). However, PAUP* version 4.0b10 was unable to compute the *P* value for 1000 tree replicates using maximum parsimony. This was due to the presence of polytomies in the topology of the trees generated by maximum parsimony that severely slowed down PAUP. It was therefore not possible to assess whether the gene sequences could be combined as a single dataset using the partition homogeneity test. The decision to combine, therefore, relied on whether the single gene phylogenies exhibited some degree of concordance with each other, which they did for some groups of taxa based on the results in Section 4.3.3.1, pp. 259-261. On this basis, the rRNA cluster, histone 3, the 1st and 2nd codon positions of the COI, and the 16S rRNA were combined into a single dataset.

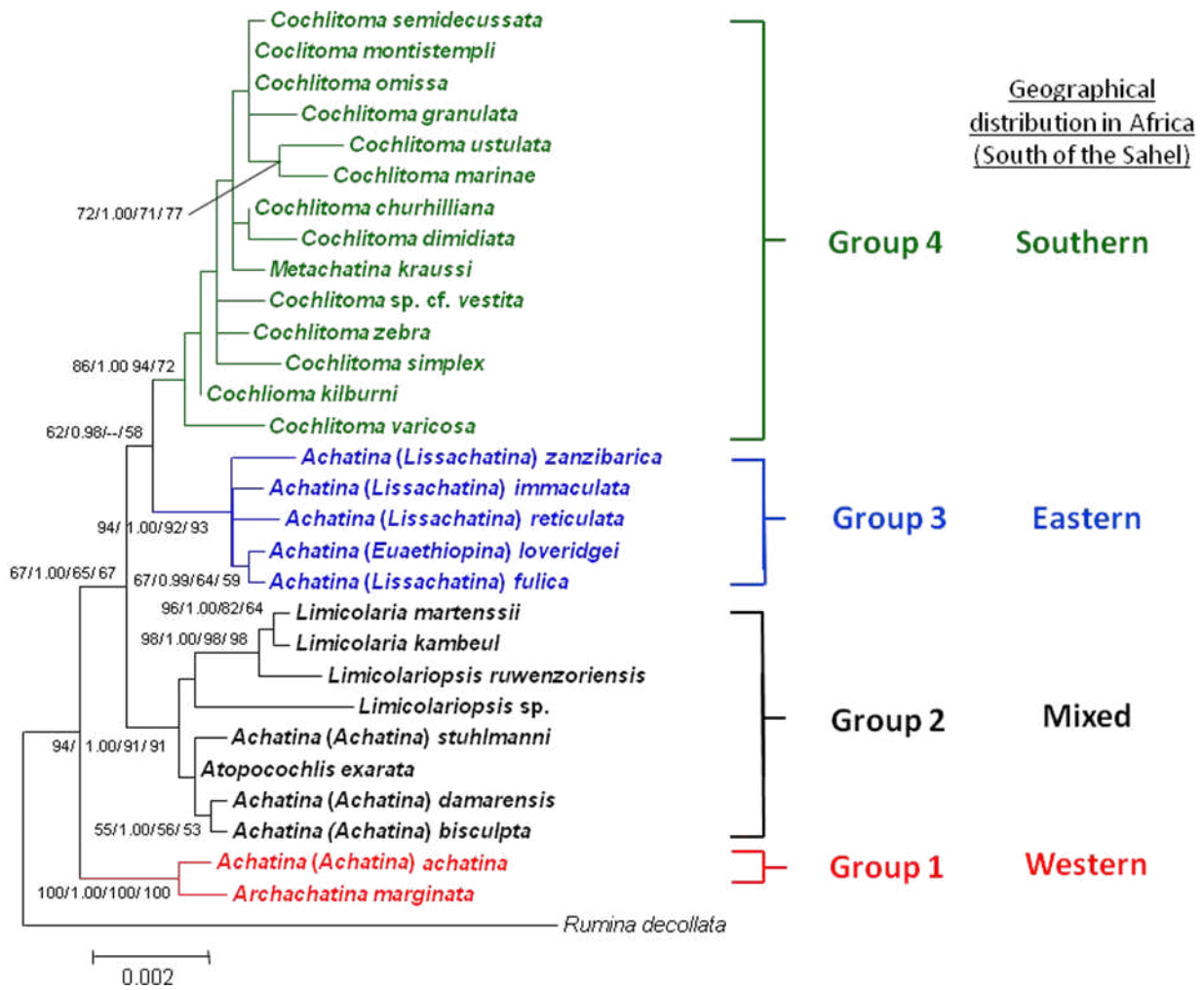
4.3.3. Molecular phylogeny

The maximum likelihood (ML) phylogenies for the rRNA cluster, histone 3, COI (1st and 2nd codon positions only) and 16S rRNA are shown in Figure 4.31 and those of the concatenated sequences from the four genes in Figure 4.32. For the concatenated sequences, two sets of phylogenetic analyses were done. In the first, only taxa with sequences from all four genes were used in the phylogeny; thus, *Achatina (Euaethiopina) loveridgei* (histone 3 and COI data missing) and *Limicolariopsis ruwenzoiensis* (histone 3, COI and 16S rRNA data missing) were excluded (Figure 4.30A). In the second, all taxa were included in the analyses (Figure 4.30B), with missing data assigned in the analyses for taxa with incomplete datasets. The bootstrap

support values for ML, neighbor-joining (NJ) and maximum parsimony (MP) analyses and the posterior probabilities for Bayesian inference (BI) analysis are also shown. For the BI, NJ and MP phylogenies for the genes and the concatenated sequences, see Appendices 4.9-4.14, pp. 511-522.

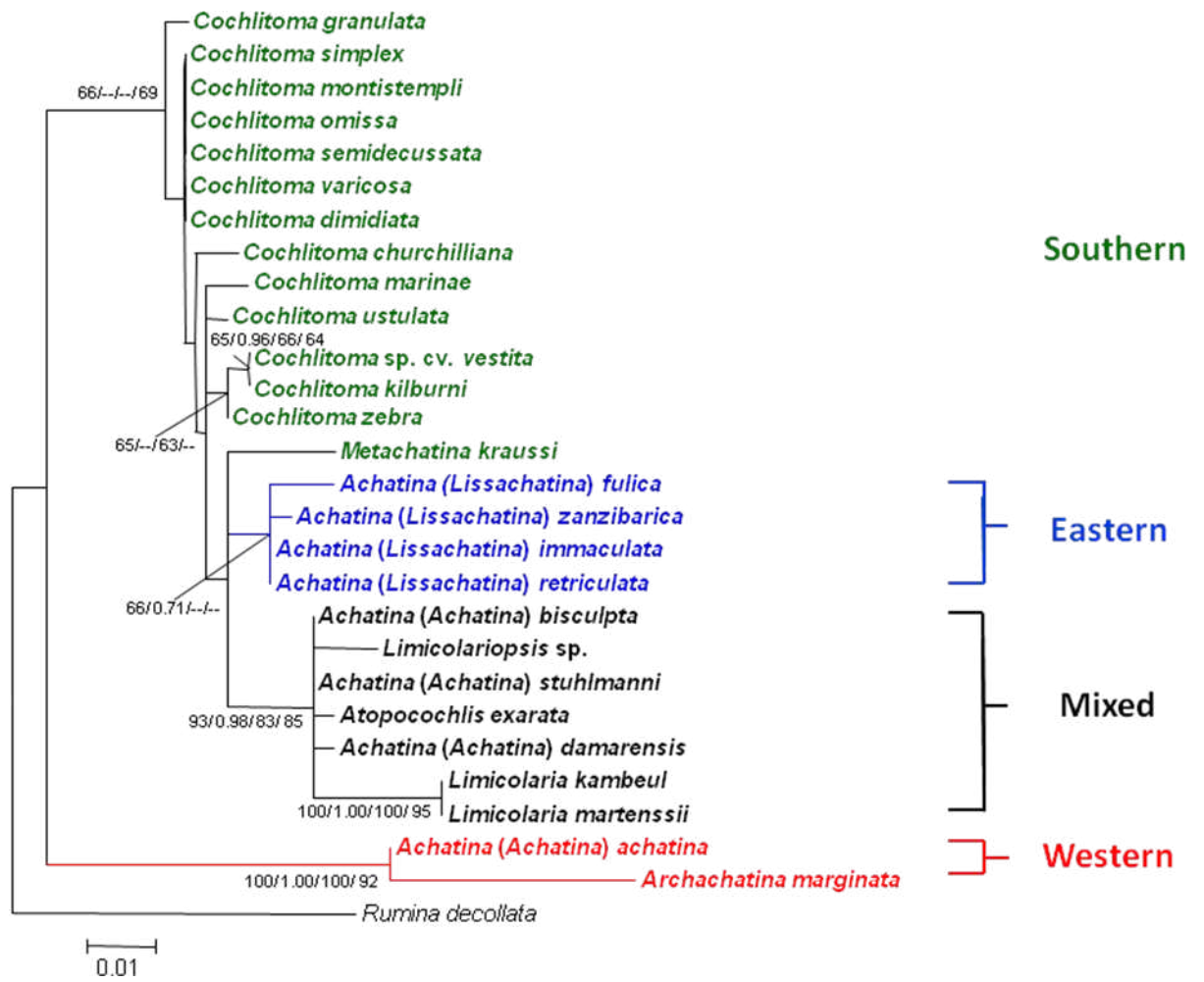
Figure 4.31: (See next four pages.) Maximum likelihood phylogenetic trees of the Achatinidae based on (A) the rRNA gene cluster (3648 nucleotides); (B) the histone 3 gene (328 nucleotides); (C) the COI gene (641 nucleotides) and (D) the 16S rRNA gene (310 nucleotides). The phylogenies were rooted on the subulinid *Rumina decollata*. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap support (1000 replicates) for NJ and MP, respectively. For BI, the optimized number of generations to explore the tree space was 2,000,000 for all the genes; the optimized heating temperatures for each gene were as follows: rRNA cluster-0.1; histone 3-0.075; 1st and 2nd codon positions of COI-0.075; 16S rRNA-0.1. The optimal model for the rRNA cluster was TN93; however, this model is not available in MrBayes, so the GTR model was instead used after determining that there was no significant difference between the TN93 and GTR models and that the GTR model was significantly better than HKY85 (see also Appendix 4.6, p. 508). The scale bar for (A) represents 2 substitutional changes per 1000 nucleotide positions; that of (B) represents 5 substitutional changes per 100 nucleotides; that of (C) represents 1 substitutional change per 100 nucleotides; and that of (D) represents 10 substitutional changes per 100 nucleotides.

Figure 4.31 (contd.)



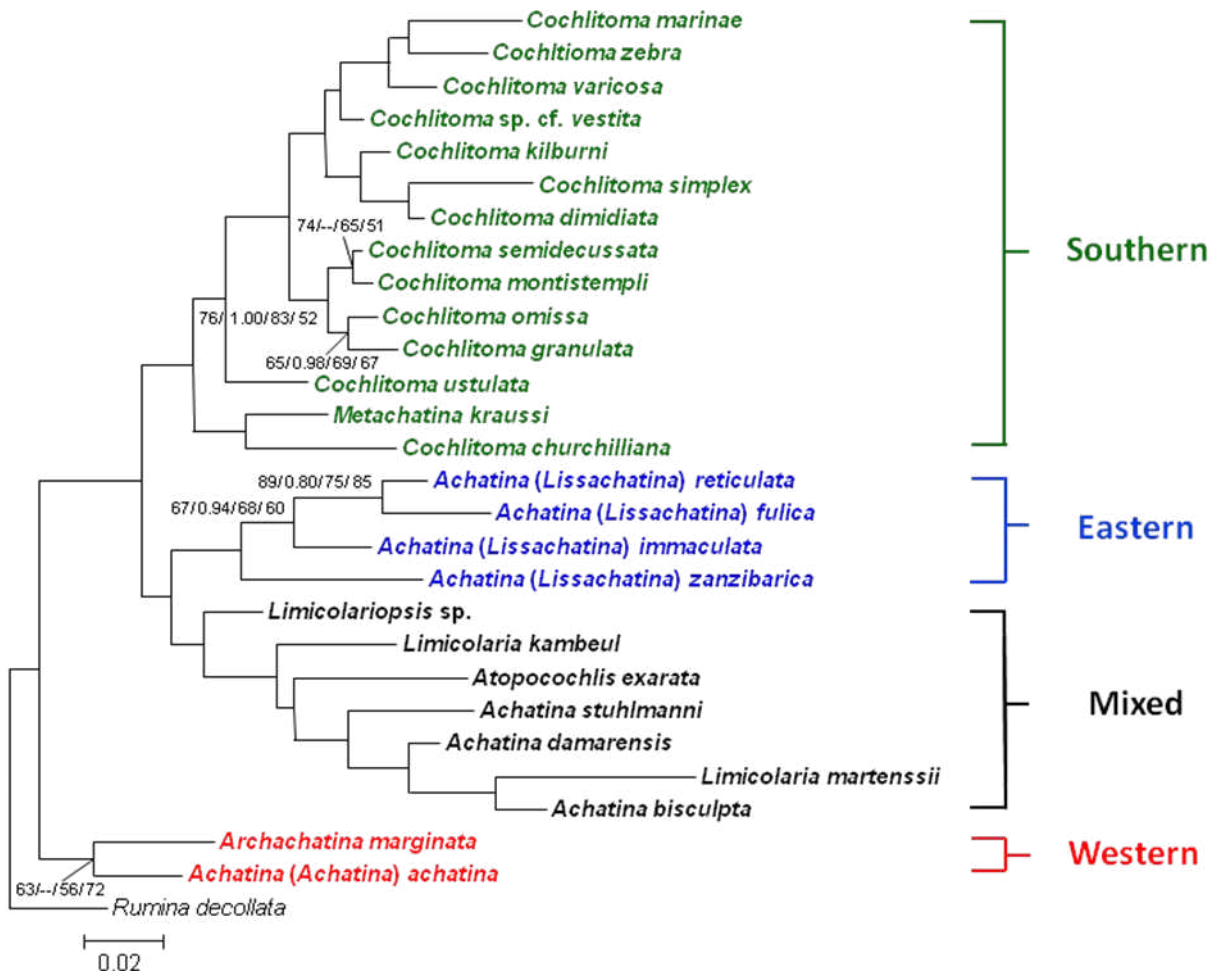
(A) rRNA gene cluster

Figure 4.31 (contd.)



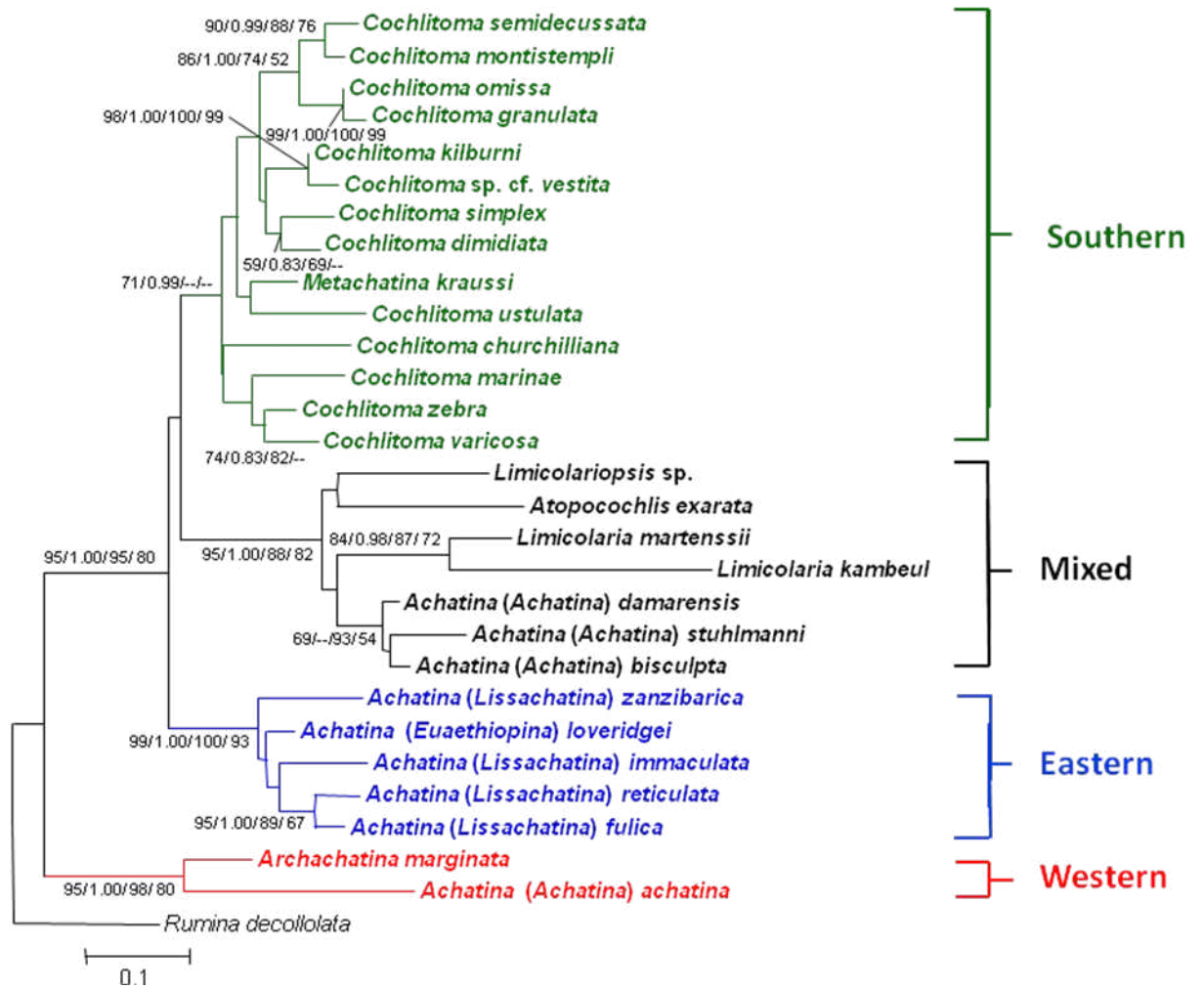
(B) Histone 3

Figure 4.31 (contd.)



(C) COI (1st and 2nd codon positions only)

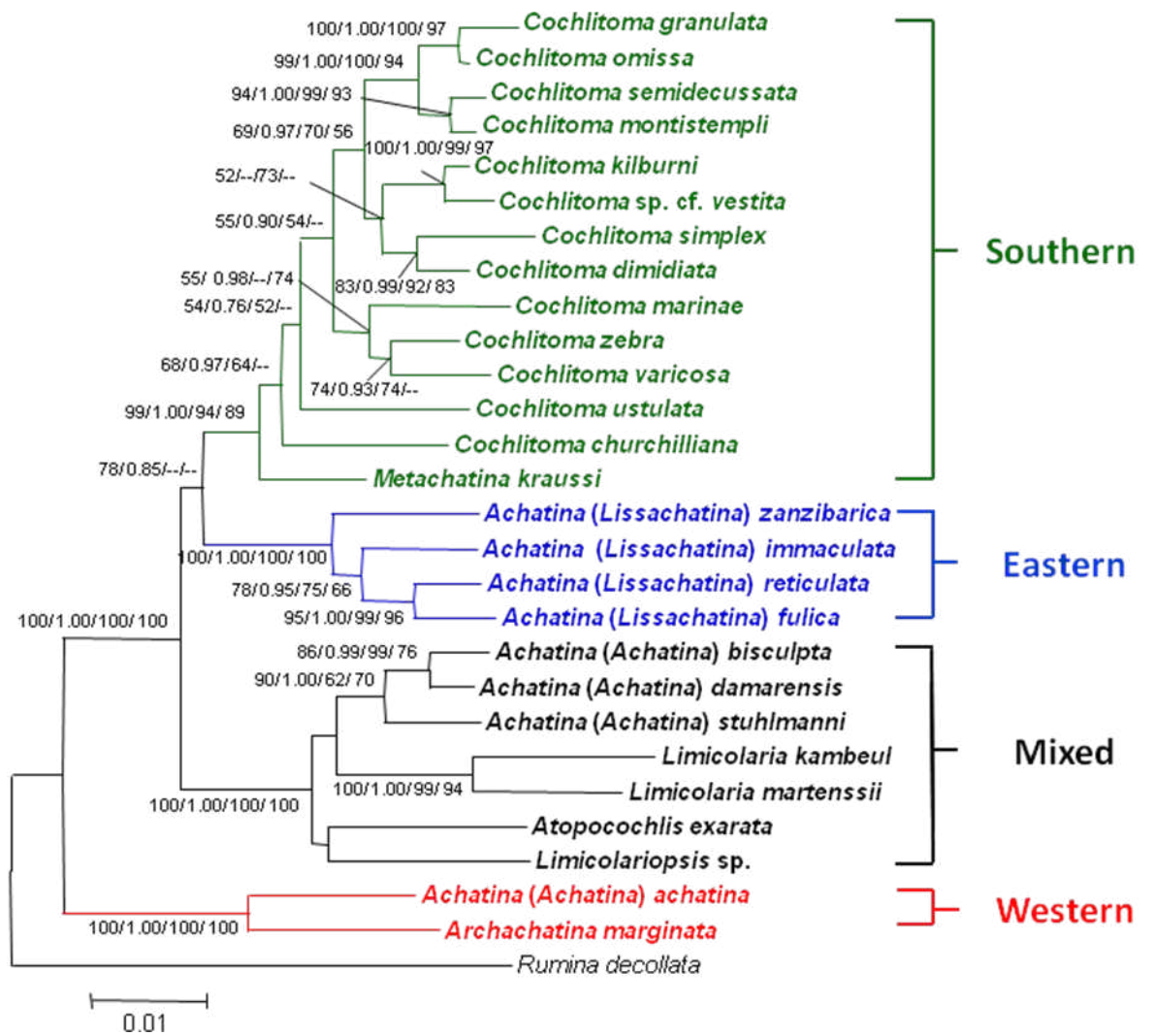
Figure 4.31 (contd.)



(D) 16S rRNA

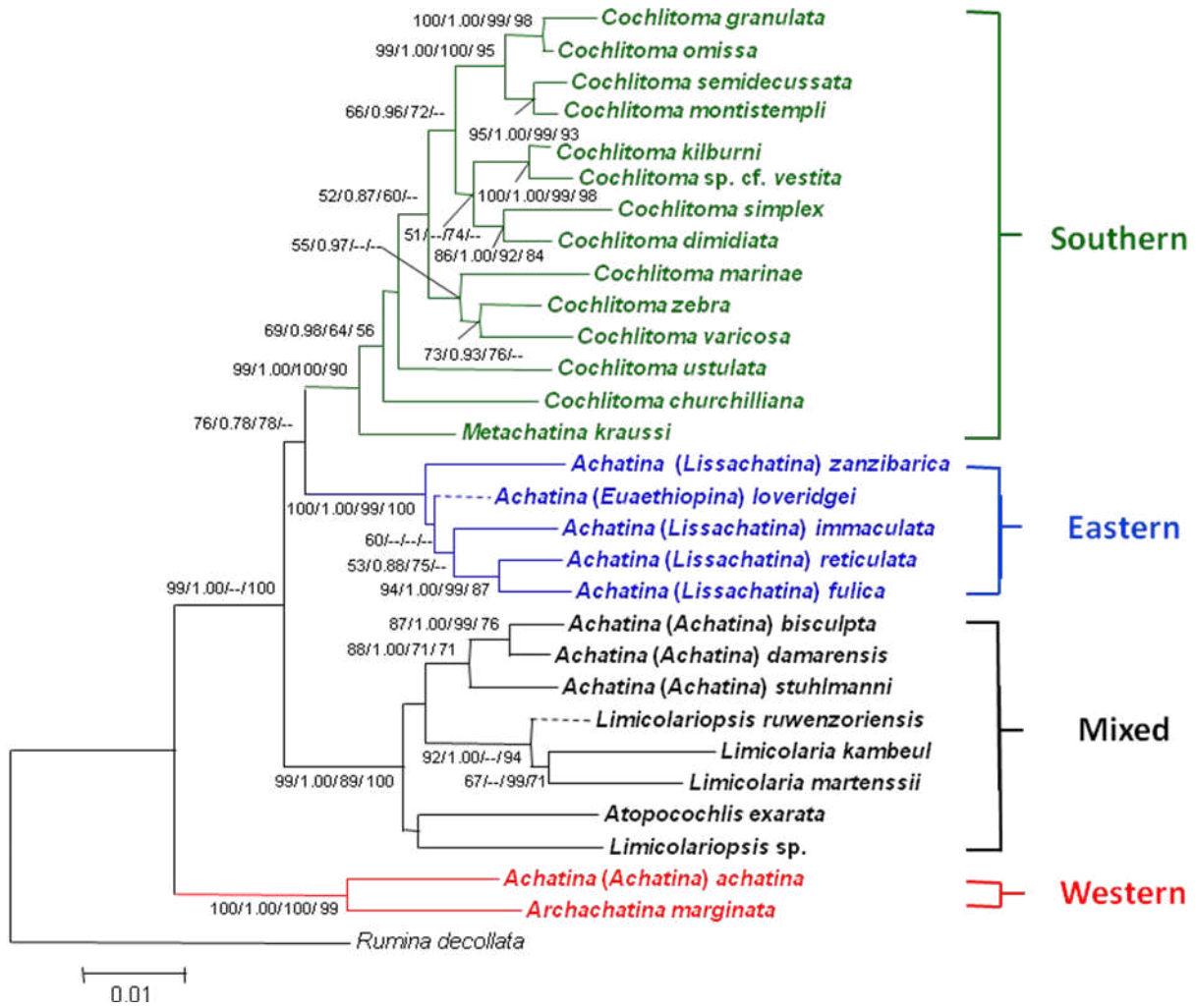
Figure 4.32: (See next two pages.) Maximum likelihood phylogenetic trees of the Achatinidae based on a concatenated sequence of 4403 nucleotides derived from the combined dataset of the rRNA cluster, H3 genes and the 1st and 2nd codon positions of the COI gene. The phylogeny in **(A)** includes only those taxa which have complete sequence data for all sequence regions while that in **(B)** includes all taxa including some with missing data. Both *Achatina (Euaethiopina) loveridgei* and *Limicolariopsis ruwenzoriensis* have missing datasets for the histone 3 and COI genes. Both phylogenies are rooted on the subulinid *Rumina decollata*. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap support (1000 replicates) for NJ and MP, respectively. Bootstrap support less than 50% and posterior probabilities less than 0.7 are not shown. For BI, the optimized number of generations to explore the tree space was 1,000,000 while the optimized heating temperature was 0.1 for both trees. The scale bars represent 1 substitutional change per 100 nucleotides. Branches for taxa with missing data are marked with dashes.

Figure 4.32 (contd.)



(A) Combined dataset (taxa with complete gene sequences only)

Figure 4.32 (contd.)



(B) combined dataset (all taxa)

4.3.3.1. Single gene phylogenies

For the rRNA gene cluster (Fig 4.31A), four major groups emerged that reflect certain geographical distributions. Group 1 at the base of the tree incorporated the two West African achatinids: *Achatina (Achatina) achatina* and *Archachatina marginata* (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). Group 2 incorporated several achatinids with mixed geographic distributions (94% ML bootstraps, $P=1.0$ BI, 91% NJ bootstraps and 91% MP bootstraps). These included the West African (São Thomé) *Atopocochlis exarata*, the Southern African *Achatina (Achatina) bisculpta*, the Southwestern African *Achatina (Achatina) damarensis*, the Central African *Achatina (Achatina) stuhlmanni* and *Limicolaria martenssii*, the tropical African (i.e. Uganda and Kenya) *Limicolariaopsis ruwenzoriensis* and *Limicolariaopsis* sp. and the widely distributed (from Senegal to Sudan and Somalia) *Limicolaria kambeul*. The other two groups formed sister clades. Group 3 incorporated the East African *Achatina (Lissachatina)* species, *A. (L.) fulica*, *A. (L.) immaculata*, *A. (L.) reticulata* and *A. (L.) zanzibarica*, as well as the East African *Achatina (Euaethiopina)* species, *A. (E.) loveridgei* (94% ML bootstraps, $P=1.00$ BI, 92% NJ bootstraps and 93% MP bootstraps). Group 4 included the Southern African *Cochlitoma* as well as *Metachatina kraussi* (86% ML bootstraps, $P=1.0$ BI, 94% NJ bootstraps and 72% MP bootstraps).

For the histone 3 gene (Figure 4.31B), three of the four groups identified in the rRNA gene phylogenies were recovered. These were the West African (Group 1) achatinids (100% ML bootstraps, $P=1.00$ BI, 100% NJ bootstraps and 92% MP bootstraps), the mixed group (Group 2) (93% ML bootstraps, $P=0.98$ BI, 83% NJ bootstraps and 85% MP bootstraps) and the East African group (Group 3), although support for this group was equivocal (66% ML bootstraps, $P=0.71$ BI and less than 50%

NJ and MP bootstraps). The Southern African group (group 4) resolved in the rRNA tree did not form a monophyletic unit in the H3 tree but instead was resolved as a paraphyletic group in that it included the Eastern (Group 3) and Mixed (Group 2) groups. In fact, *Metachatina kraussi*, a Southern African taxon, clustered with Groups 2 and 3, although this grouping is not well supported.

For the 1st and 2nd codon positions of the COI gene (Figure 4.31C), all four groups were recovered but there was generally less structure in the topology of the phylogeny and little support for groupings. Of the four groups, only the West African group (Group 1) received moderate support (63% ML bootstraps, $P < 0.7$ BI, 56% NJ bootstraps and 72% MP bootstraps).

For the 16S rRNA gene (Figure 4.31D), all four groups were recovered with good to moderate support: the West African group (Group 1) (95% ML bootstraps, $P = 1.00$ BI, 98% NJ bootstraps and 80% MP bootstraps); the mixed group (Group 2) (95% ML bootstraps, $P = 1.00$ BI, 88% NJ bootstraps and 82% MP bootstraps); the East African group (Group 3) (99% ML bootstraps, $P = 1.00$ BI, 100% NJ bootstraps and 93% MP bootstraps); and the Southern African group (Group 4) (71% ML bootstraps, $P = 0.99$ BI, and less than 50% NJ and MP bootstraps).

4.3.3.2. Combined gene phylogenies

The combined analysis for the rRNA cluster, H3, the 1st and 2nd codon positions of COI and 16S rRNA, but excluding *Achatina (Euaethiopina) loveridgei* and *Limicolariopsis ruwenzoriensis* as they had some gene sequences missing, recovered all four groups present in the rRNA phylogeny and exhibited longer internal branches with better support (Figure 4.32A). The West African group (Group 1) still fell at the base of the Achatinidae (100% ML bootstraps, $P = 1.00$ BI, 100% NJ and 100% MP bootstraps)

while the other achatinids clustered together (100% ML bootstraps, $P=1.0$ BI, 100% NJ bootstraps and 100% MP bootstraps). Within this cluster were the mixed group (Group 2) (100% ML bootstraps, $P=1.00$ BI, 100% NJ and 100% MP bootstraps), the East African group (Group 3) (100% ML bootstraps, $P=1.00$ BI, 100% NJ and 100% MP bootstraps) and the Southern African group (Group 4) (99% ML bootstraps, $P=1.00$ BI, 94% NJ and 89% MP bootstraps), with *Metachatina kraussi* falling at the base of the latter. Groups 3 and 4 clustered as sister clades within the tree.

When all taxa were included in the combined analysis (including the two taxa with missing data), all groups found in the rRNA phylogeny were recovered and in general received higher support (Figure 4.32B). The West African group (Group 1) again fell at the base of the Achatinidae (100% ML bootstraps, $P=1.0$ BI, 100% NJ and 99% MP bootstraps) while the remaining achatinids formed a separate clade with increased support in comparison to the rRNA cluster except for NJ (99% ML bootstraps, $P=1.0$ BI, less than 50% NJ bootstraps and 100% MP bootstraps). Within this clade and consistent with the rRNA tree were the mixed group (Group 2) with increased support (99% ML bootstraps, $P=1.0$ BI, 89% NJ and 100% MP bootstraps) and the clade incorporating the East African and Southern African groups with increased support except for MP (76% ML bootstraps, $P=0.78$ BI, 78% NJ and less than 50% MP bootstraps). The East African group (Group 3) also received increased support (100% ML bootstraps, $P=1.00$ BI, 99% NJ and 100% MP bootstraps) as did the Southern African group (Group 3) (99% ML bootstraps, $P=1.0$ BI, 100% NJ and 90% MP bootstraps). Some well-supported groupings were also observed within groups 2-4. Within the mixed group (Group 2), for instance, there was strong support for the clustering of *Achatina (Achatina) bisculpta*, *A. (A.) damarensis* and *A. (A.) stuhlmanni* (88% ML bootstraps, $P=1.0$ BI, 71% NJ bootstraps and 71% MP bootstraps) as well as

Limicolaria kambeul, *L. martensii* and *Limicolariopsis ruwenzoriensis* (92% ML bootstraps, $P=1.0$ BI, less than 50% NJ bootstraps and 94% MP bootstraps). Within the East African group (Group 3), *Achatina (Lissachatina) fulica* clustered with *A. (L.) immaculata* (94% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 87% MP bootstraps). Within the Southern African group (Group 4), *Metachatina kraussi* again fell at the base. Very strong support was also shown for several groups. These were *Cochlitoma omissa* and *C. granulata* (100% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 98% MP bootstraps); *C. montistempli* and *C. semidecussata* (95% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 93% MP bootstraps); *C. kilburni* and *C. sp. cf. vestita* (100% ML bootstraps, $P=1.0$ BI, 99% NJ bootstraps and 98% MP bootstraps); *C. dimidiata* and *C. simplex* (86% ML bootstraps, $P=1.0$ BI, 92% NJ bootstraps and 84% MP bootstraps); and *C. varicosa* and *C. zebra* (73% ML bootstraps, $P=0.93$ BI, 76% NJ bootstraps and less than 50% MP bootstraps). The geographic distributions of the members of the four groups are shown in Fig. 4.33.



Figure 4.33: Geographic distribution of the achatinid taxa used in this study (distribution data from Bequaert, 1950; Mead, 1950; Schileyko, 1999; and Raut & Barker, 2002 as well as the actual sites from which some of the taxa were collected).

3.3.3.3. Hypothesis testing of *Achatina* (*Achatina*) based on taxonomy

The four members of the subgenus *Achatina* (*A. achatina*, *A. bisculpta*, *A. damarensis* and *A. stuhlmanni*) were not recovered as monophyletic, with *Achatina* (*Achatina*) *achatina* clustering with *Archachatina marginata*; these taxa were subjected to hypothesis testing using the Shimodaira-Hasegawa (1999) test. Constraining the NJ trees for the four single gene and combined gene phylogenies to show monophyly of the subgenus *Achatina* produced likelihood scores that were significantly worse ($p < 0.01$) than those of the optimal trees except for the 1st and 2nd codon positions of the COI gene ($p = 0.177$) (see Table 4.4 for details). The results confirmed the non-monophyly of the subgenus *Achatina*.

Table 4.4: Hypothesis testing for the monophyly of *Achatina* (*Achatina*) using the Shimodaira-Hasegawa test.

Gene	Likelihood Score		<i>P</i> for SH ^a Test (Conclusion ^b)
	Optimal NJ tree	Constrained NJ tree	
rRNA cluster	6146.48364	6240.31789	0.002 (Significant)
Histone 3	909.37626	949.45797	0.001 (Significant)
COI (1 st and 2 nd codons)	1995.68125	2004.66467	0.177 (Not significant)
16S rRNA	2876.32254	2906.36582	0.008 (Significant)
Combined	12509.66094	12643.04203	<0.001 (Significant)
Combined-all taxa	12640.55433	12756.19951	<0.001 (Significant)

^a SH test – Shimodaira-Hasegawa Test

^b Significant at $P < 0.05$

4.4. Discussion

4.4.1. Single versus combined gene phylogenies

Among the five genes used for the Achatinidae, the nuclear rRNA cluster provided the most resolved structure of the internal branches with the four major groups based on biogeography receiving good support. The terminal nodes, however, were not well supported, suggesting that the gene is too conserved for that level. The nuclear histone 3 gene yielded even less structure with only three groups recovered and very few terminal nodes received support. The gene was also too conserved as evidenced by the presence of five sets of identical sequences among 16 taxa. On the other hand, the two mitochondrial genes were more variable and were therefore evolving faster than the two nuclear genes. Of the two, the 16S rRNA gene gave support to most terminal nodes while at the same time recovering the four major groups. The COI had less structure than 16S because the groups were not well supported. Fewer sites were used for COI as the third codon positions were excluded due to their saturated condition, which is indicative of their faster rate of evolution. The fifth gene, the nuclear actin, was also not included due to significant levels of paralogy that disrupted phylogenetic signal.

When the four genes were combined, both the terminal branches and the internal nodes received support based on ML, NJ and MP bootstraps as well as BI posterior probabilities. Underlying signal was also detected in the combined phylogenies with very good support that was not evident in the single gene phylogenies (i.e. the basal position of *Metachatina kraussi* in the Southern African group).

4.4.2. Evolutionary history of the Achatinidae based on molecular data

The existence of no fossils earlier than the Pleistocene (0.01-1.8 MYA) (Zilch, 1959-1960; Solem, 1979) suggests that the Achatinidae are a relatively recent group (Tillier, 1989), although Raut & Barker (2002) believed them to be much older than the fossil record. Mead (1991) suggested that the primitive achatinids occurred in the region of the Lower Guinea in West Africa, and he postulated that the achatinids radiated from there to the other parts of Africa south of the Sahel. Tillier (1989) described the achatinids as giant subulinids, although the taxonomy of the latter is still in a state of flux, and Tillier could not identify with certainty from which subulinid group the achatinids arose.

This study presents the first attempt to elucidate the phylogeny of the Achatinidae using molecular data. Phylogenies derived from the combined nuclear rRNA cluster and histone 3 genes and the mitochondrial cytochrome c oxidase 1 and 16S rRNA genes demonstrated groupings that roughly corresponded to the geographical distributions of the Achatinidae. These groups were the West African group (Group 1) found at the base of the tree followed by the mixed group (Group 2), the East African (Group 3) group and Southern African (Group 4) group, the last two of which clustering as sister clades. Group 2 included species found in the other three regions as well as some that are present in Central Africa.

The West African *Achatina (Achatina) achatina* (found in the region of Upper Guinea of West Africa; Bequaert, 1950) and *Archachatina marginata* (found in the region of West Africa east of the Dahomey gap, Bequaert, 1950) both fell at the base of the tree, which supports Mead's (1991) hypothesis that the achatinids originated in the west, particularly in the Gabon-Cameroon region. Both species share a penis sheath that only covers half of the penis (Mead, 1950). In fact, *Archachatina* Albers was erected as a distinct genus from *Achatina* purely on conchological criteria, primarily on the form of the shell apices. The large obtuse apex of *Archachatina* is a result of the large embryonic whorls that form in their large eggs (about 19 by 14.5 mm); *Achatina* possesses an acute apex as a result of its small eggs (Bequaert, 1950).

The mixed group includes several species with varying distribution patterns, including the São Thomé endemic *Atopocochlis exarata*. Mead (1991) included *Atopocochlis exarata* as one of the most primitive achatinids together with *Callistoplepa*, *Leptocala*, *Tripachatina* and *Columna*, none of which was sampled in this study due to inability to obtain fresh material. The reproductive structures of *A. exarata* include a very short penis sheath similar to that of *Achatina (Achatina) achatina* and *Archachatina marginata* and an enlarged vagina (Mead, 1950); the vas deferens is also enclosed by the penis sheath, unlike in the other “primitive” achatinids like *Callistoplepa* and *Leptocala* where the vas deferens is free (Schileyko, 1999). The molecular data as well as the morphological features of the genitalia puts into question the position of *Atopocochlis exarata* among the “primitive” achatinids as suggested by Mead (1991). Another group within the mixed group with high support is the grouping of the two *Limicolaria* species. Both have a distinct penis papilla (Mead, 1979b; 1991). Also within the mixed group are *Achatina (Achatina) bisculpta* and *Achatina (Achatina) damarensis*, which clustered together with strong support. In both species, the basal vas

deferens and penis are either intimately and tightly connected to the penial retractor muscles, as in the case *A. (A.) bisculpta* (Sirgel, 2000), or cocooned by them, as in the case of *A. (A.) damarensis* (Mead, 1950, 1991; Van Bruggen, 1970). These two taxa clustered with *A. stuhlmanni*, and all three belong to the subgenus *Achatina*, though these taxa did not cluster with another member of the subgenus, *A. achatina*, which grouped with *Archachatina marginata*. The non-monophyly of this subgenus was further supported by the Shimodaira-Hasegawa test. The validity of this taxon therefore warrants re-examination.

The Southern African and East African groups together form a sister clade with the mixed group according to the combined gene phylogenies. Indeed, both *Achatina (Lissachatina)* from East Africa and *Cochlitoma* of Southern Africa share a “long spermathecal [=gametolytic] duct [that] places the spermatheca [=gametolytic sac] on the uterine portion of the spermoviduct, well above the junction of” the apical vas deferens and the uterine portion of the spermoviduct (Mead, 1991, p. 553). Such a characteristic is probably derived as it is not found in the Western group, which falls at the base of the Achatinidae based on the single and combined gene phylogenies.

The presence of the penial groove in *Cochlitoma* is most likely a derived character as it is not present in *Metachatina kraussi* found at the base of the southern African clade in the combined gene phylogenies, thus lending support to the assumption of Mead (1991) that *Cochlitoma* is an ‘advanced’ genus. This topology was not evident in the single gene phylogenies and was only revealed as an underlying signal when the genes were combined, as shown by the higher bootstrap and posterior probability supports. The clustering of *Cochlitoma* species within the Southern African group based on the molecular phylogenies supports Mead’s (2004) recognition of *Cochlitoma*

as a distinct genus rather than a subgenus of Western African *Archachatina* even if they share similar conchological features.

The members of the East African group under *Achatina* (*Lissachatina*) have extroversion muscles as well as a muscular, bulboid enlargement of the basal vagina (Mead, 1991, 1995). The results from the molecular data in this study support Mead's view that *Lissachatina* be distinguished from the West and central African *Achatina* species. Furthermore, judging from the topology of the East African group in relation to other *Achatina* species in the molecular trees, coupled with the distinct morphological characteristics of both the male and female conduits of the reproductive tract, it is clear that there is no basis for including *Lissachatina* within *Achatina* and *Lissachatina* should be recognized as a distinct genus as with *Cochlitoma*. Within the East African group, *Achatina* (*Lissachatina*) *fulica* clustered with *A. (L.) reticulata*. They differ morphologically in that *A. reticulata* has a short penis sheath that does not cover the entire penis (Mead, 1950). As with *Atopocochlis exarata*, the presence of a short penis sheath in *A. reticulata* is hypothesized to be a derived state and does not indicate plesiomorphic condition or 'primitive' state *sensu* Mead. It can be noted that the Southern African *Metachatina kraussi* (Mead, 1950) and *Cochlitoma granulata* (Mead, 2004) also possess a short penis sheath. The subgenus *Euaethiopina*, represented by *A. (E.) loveridgei*, has a shell that is more elongated ovoid than that of *Lissachatina*, which is more broadly ovate to obesely fusiform; the two taxa also have distinct coloration (Bequaert, 1950; Schileyko, 1999). However, *Achatina* (*Euaethiopina*) *loveridgei* is very similar to *Achatina* (*Lissachatina*) *zanzibarica* with respect to its reproductive anatomy (Mead, 1950). Its placement within *Lissachatina* based on molecular data, coupled with the evidence based on reproductive anatomy, would suggest that *A. loveridgei* should be moved to *Lissachatina*.

All these results support the hypothesis that the achatinids migrated to the central region from the west, then radiated to the east and south. As further support to this hypothesis, van Bruggen (1986) noted that terrestrial snails and slugs of the Afrotropical region (south of the Sahel) occur in four major centres of endemism: 1) Southern Africa; 2) East Africa; 3) North-East Africa; and 4) Central/West Africa. He also observed the highest taxon density for the Achatinidae in West Africa that extended to Central Africa, followed by East and North-East Africa and finally Southern Africa.

Aside from the Achatinidae, there are 33 other families of terrestrial gastropods in the Afrotropical region, of which eight are endemic. Of these, three families (Maizaniidae, Gymnarionidae and Urocyclidae) are found in all four major centres like the Achatinidae; one family (Aillyidae) is found only in West Africa and another (Thyrophorellidae) is restricted to São Thomé off the coast of West Africa. The remaining endemic families (Aperidae, Dorcasiidae and Prestonellidae) are distributed in Southern Africa (van Bruggen, 1986). Diamond and Hamilton (1980) also found that forest passerine birds roughly occur in the four endemic centres, either limited to one or a few of these centres or occurring in all. These centres most likely served as refugia for forest-dependent animals, such as terrestrial gastropods and passerine birds, when forests contracted due to changing rainfall patterns as a result of alternating glacial and interglacial periods during the Quaternary, particularly the Holocene (18,000 years ago), which in turn gave opportunities for vicariant speciation to occur (van Bruggen, 1986). However, Raut & Barker (2002) acknowledged that much of the speciation process, particularly for the terrestrial gastropods, could have predated the changing forest cover of the Holocene.

4.4.3. Limitations of the study and the next step

A number of achatinid groups were not available for this study. Among these are genera found in West Africa considered primitive by Mead (1991) in that the penis sheath is very short and does not enclose the vas deferens. These include *Leptocala* and *Callistoplepa*. If Mead's hypothesis is correct, then these genera should fall at the base of the tree. Furthermore, as West Africa harbors a high number of endemic species of achatinids (van Bruggen, 1986), it is desirable to include as many of these species as possible in future investigations. Of particular interest are the sinistral *Columna columna* and *Archachatina bicarinata*. Equally interesting is the relationship of the East African *Bequaertina* with *Lissachatina*.

4.5. Summary

This study is the first attempt to reconstruct the phylogeny of the Achatinidae using molecular data derived from the nuclear rRNA cluster, actin and histone 3 genes and the mitochondrial cytochrome c oxidase subunit 1 and 16S rRNA genes. Results demonstrated four distinct groups that to a large extent follow the geographical distribution of the sampled taxa and validate groupings previously erected on the basis of reproductive morphology. At the base of the trees are West African species followed by a group which includes taxa with varying geographical distributions ranging from the western, central, eastern and southern regions. Two other groups probably arose from the mixed group and form sister clades: one is the East African group that includes *Achatina (Lissachatina) fulica*; the other group encompasses most of the Southern African species. The results support Mead's recognition of Southern African *Cochlitoma* as a genus separate from the West African *Archachatina*. The results likewise support Mead's (1950) distinction of the Eastern subgenus *Lissachatina* from

the western and central *Achatina* and demonstrate that there is no basis for continuing to include *Lissachatina* as a subgenus of *Achatina*.

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CHAPTER 5 - Genetic variation in global populations of *Achatina fulica*

5.1. Introduction

The Giant African Land Snail, *Achatina (Lissachatina) fulica* Bowdich, is a tropical species with a widespread distribution across East Africa, the Indo-Pacific and the Caribbean. Its natural range is East Africa, but its current global distribution is primarily due to human intervention (Raut & Barker, 2002). It is a serious crop pest that can easily spread into new areas where it causes significant damage to vegetables and other food crops (Mead, 1979; Raut & Barker, 2002). The World Conservation Union (IUCN) has listed *A. fulica* in its 100 most invasive species (Lowe *et al.*, 2000), and among the land snails in the world, it is probably the most invasive (Raut & Barker, 2002). Its success as an invasive species is probably attributable to its high reproductive capacity and its generalist food behavior, feeding on a wide range of plants and detritus (Raut & Barker, 2002). The tendency for people to transport the snails and release them into the wild either intentionally or inadvertently also helps to spread them further (Thiengo *et al.*, 2007). The snail also serves as the intermediate host of the rat lungworm *Angiostrongylus cantonensis* that causes eosinophilic meningitis in humans (Marquardt *et al.*, 2000).

5.1.1. The Dispersal of *Achatina fulica* from East Africa

The ease with which *Achatina fulica* spreads into new areas is attributed to the availability of several pathways of dispersal for the snail. A pathway, which pertains to an activity that involves the introduction of a species, can be accidental or deliberate (Cowie & Robinson, 2003). *Achatina fulica* is deliberately introduced into new areas for the purposes of consumption, as pets, aesthetics or ornaments, religious symbols, or

for medicinal or research use (Mead, 1979; Kliks & Palumbo, 1992; Cowie & Robinson, 2003; Budha & Naggs, 2008). On the other hand, *A. fulica* is accidentally introduced when it “hitchhikes” with agricultural and horticultural products, the packaging or containers of any commodity, or even on vehicles such as cars and trucks (Cowie & Robinson, 2003). *Achatina fulica* can also escape from local farm operations where they are bred for their meat (Paiva, 1999).

Achatina fulica has a pan-tropical distribution brought about within the last 200 years by its human-aided dispersal (Fig. 5.1). The snail is indigenous to the coast of East Africa (probably originating in Kenya and Tanzania) but was introduced into the nearby islands of Madagascar, Mauritius, the Comoros, Mayotte and Reunion prior to 1800 (Bequaert, 1950). The snail was then brought to India and Sri Lanka by naturalists in the early 1900s (Naggs, 1997; Raut & Barker, 2002) and spread to Nepal to the north (Budha & Naggs, 2008) and to the Malay Peninsula to the southeast via sea routes in cargo vessels (Mead, 1961). Immediately before and during the Second World War, Japanese merchants and soldiers spread *A. fulica* further into Southeast Asia, Taiwan, and many islands of the Pacific as they were used for food and medicinal purposes and sometimes as pets (Kliks & Palumbo, 1992). By the late 1980’s, *A. fulica* had been reported in the Caribbean, particularly in Guadeloupe and Martinique (Schotman, 1989) and later on in St. Lucia and Barbados by 2000 (Fields *et al.*, 2006) and in Antigua by 2008 (NAPPO-PAS, 2008). By the 1990’s, *A. fulica* had already established itself in South America, particularly Brazil (Paiva, 1999; Carvalho de Vasconcellos & Pile, 2001; Thiengo *et al.*, 2007), Colombia, Ecuador and Peru (Borrero *et al.*, 2009). Recently, the snail has been observed in the Ivory Coast, Ghana and Morocco in West and Northwest Africa (Raut & Barker, 2002). Although *A. fulica* has reached Australia, Japan, and the United States mainland in the past, authorities in these countries have

successfully eradicated *A. fulica* for fear of its potential for causing damage to agriculture (Mead, 1961; Kliks & Palumbo, 1992). As an exotic pet (Kliks & Palumbo, 1992), *A. fulica* is now present in temperate countries such as the UK and France.

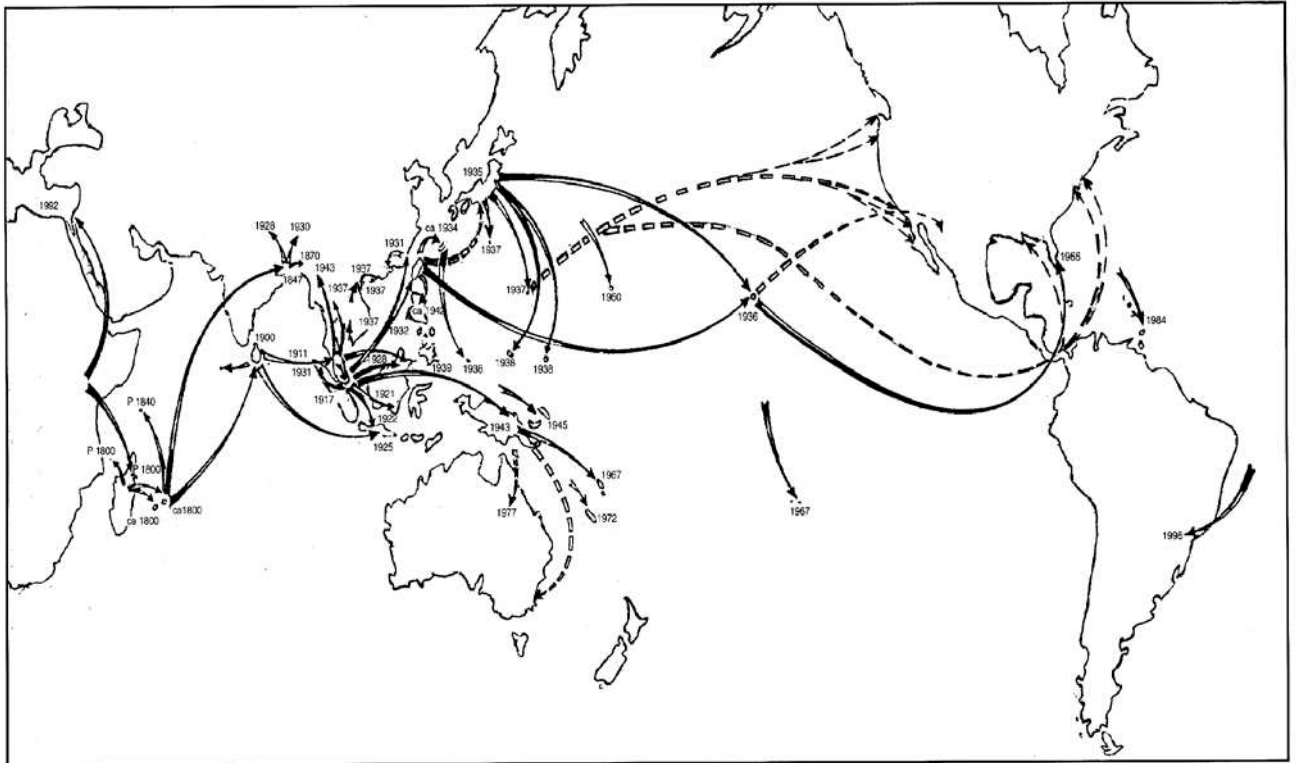


Figure 5.1: Dispersal route of *Achatina fulica* from East Africa. Years indicate approximate dates when the snail was first observed in each region. Solid lines denote successful introductions while broken lines denote introductions that were intercepted. (From Raut & Barker, 2002).

5.1.2. Genetic variation in introduced species

The genetic composition of the population of an introduced species in a new area may affect the success of that population. Invasion may involve founder events in which a population of a few individuals carries a reduced amount of genetic variation in comparison to the source population. Over time, this leads to a population bottleneck in which individuals suffer from a lack of genetic variation and where massive shifts in

allele frequencies are observed (Dlugosch & Parker, 2008). This becomes particularly problematic to the invasive species if the loss of variation includes adaptive traits that could have improved their chance of establishing in a new habitat, preventing extinction and increasing population growth (Kolbe *et al.*, 2007). Although some data do show lack of genetic diversity in introduced species, as seen in the apple snails *Pila conica* and *Pomacea canalicuata* in Hawaii (Tran Chuong *et al.*, 2008), other data on invasive species reveal that many populations do not exhibit such lack of diversity. One possible explanation is the role of multiple introductions in augmenting the genetic diversity of the invasive species. Multiple introductions may involve separate dispersal events over a long period of time in which new genes from recently introduced populations are spread among the individuals in the established population (Dlugosch & Parker, 2008). This was evident in the cheatgrass, *Bromus tectorum*, in which multiple introductions from Europe and Southwest Asia counteracted the effect of bottlenecking in introduced populations in western North America (Novack & Mack, 1993). Kolbe *et al.* (2007) also noted the same pattern in eight species of *Anolis* lizards introduced in Florida and the Dominican Republic from the northern Caribbean using the ND2, tRNA-Trp and tRNA-Ala genes of the mitochondrial DNA. Their results revealed that 80% of introductions were derived from multiple native sources. Furthermore, haplotypes that were found in distinct geographic locations in the native range occurred as variants within the introduced populations. They proposed a two-step process in introductions marked by the initial decrease in genetic variation as a result of founder events and bottlenecking followed by an increase in genetic variation if multiple introductions from different sources take place.

Despite the detailed historical and anecdotal information concerning the spread of *Achatina fulica*, no systematic evaluation of its genetic variation across global

populations has been undertaken. It is therefore not known whether multiple introductions of *A. fulica* from different sources have occurred in any part of its new range or whether the introduced populations are undergoing bottlenecks. In either case, these factors could have a profound impact on the success of *A. fulica* as an introduced species.

5.1.3. Mitochondrial molecular markers for evaluating genetic variation in snail populations and elucidating their population history

Data derived from molecular genetic variation in snail populations can yield useful information about the routes of introduction or dispersal and colonization of a particular snail species into new areas (Davison, 2000; Gittenberger *et al.*, 2004; Pinceel *et al.*, 2005; Rawlings *et al.*, 2007). Mitochondrial genes have been used as the markers of choice to assess variation within and between snail populations because they generally evolve faster than nuclear genes and are maternally inherited and therefore not subject to recombination (Brown, 1985; Avise *et al.*, 1987). For example, Davison (2000) analyzed a fragment of the mitochondrial 16S rRNA gene using single-stranded conformational polymorphism (SSCP) analysis and DNA sequencing to identify different haplotypes from populations of the banded grove snail *Cepaea nemoralis* from Britain and Ireland. The fragment he used for the SSCP analysis corresponded to the variable domain IV segment of the secondary structure of the post-transcribed 16S rRNA gene (Lydeard *et al.*, 2000) for which Palumbi (1996) designed universal primers. Davison's work revealed two mitochondrial lineages that had an east-west distribution, suggesting two possible routes of colonization of Britain after the last ice age. Pinceel *et al.* (2005) also used the same variable domain IV segment of the 16S rRNA gene as part of their population genetic survey of the terrestrial slug *Arion*

subfuscus in northeastern USA and Europe. They likewise used SSCP and DNA sequencing. Their results revealed the existence of two distinct 16S rRNA haplotypes in the USA representing at least two independent introductions from the British Isles on the one hand and mainland Europe on the other. In another study, Gittenberger *et al.* (2004) sequenced a fragment of the cytochrome c oxidase subunit I (COI) gene for several populations of the snail *Arianta arbustorum* and three other species of *Arianta* to elucidate the evolutionary history of the two shell forms of *A. arbustorum*.

5.1.5. Objective of the study

This study aimed to assess the genetic variation in global populations of *Achatina fulica*. Comparison of recently dispersed *A. fulica* populations from across the globe with those from East Africa and adjacent islands were undertaken to determine where the recently introduced global populations of *A. fulica* originated and whether they represent single or multiple introductions. To this effect, the genetic diversity of *A. fulica* populations from East Africa, the Indian subcontinent, Southeast Asia, the Pacific and the Caribbean were evaluated using single stranded conformational polymorphism (SSCP) analysis of the 16S ribosomal (r) RNA gene.

5.2. Materials and methods

5.2.1. Sample collection

In total, 382 individuals of *Achatina fulica* from 19 populations across the globe were sampled. Two populations were obtained from East Africa: Kampala, Uganda (5 snails) and Dar Es Salaam, Tanzania (38 snails). Three populations were from Indian Ocean islands near East Africa: Mayotte (50 snails) off the coast of Mozambique;

Souillac, Mauritius (45 snails) east of Madagascar; and Mahe, Seychelles (2 snails) northeast of Madagascar. Samples from Nepal (22 snails) represented the Indian subcontinent while those from Southeast Asia included Myanmar/ Burma (20 snails), Thailand (20 snails), the Philippines (2 population samples with 20 snails each), Malaysia (20 snails) and Singapore (20 snails). The Pacific was represented by Ogasawara/ Bonin (12 snails), New Caledonia (21 snails), Hawaii (20 snails), Moorea (10 snails) and Tahiti (5 snails). Two populations were obtained from the Caribbean: Martinique (20 snails) and Barbados (12 snails). The Martinique specimens were a captive bred population of F1 offspring bred from Martinique parents by F. Adnai. See also Table 5.1.

Table 5.1: Locality, collector and sample size of global populations of *Achatina fulica* used in the study

Locality	Collector	Sample size
Kampala, Uganda	B. Rowson	5
Dar Es Salaam, Tanzania	C. Ngereza	38
Mayotte	F. Barthelat	50
Souillac, Mauritius	O. Griffiths	45
Mahe, Seychelles	J. Gerlach	2
Bharatpur, Nepal	P. Budha	22
Yangon, Myanmar/ Burma	F. Naggs	20
Trok Nong Area, Chantaburi, Thailand	S. Panha & C. Sutcharit	20
Hahajima, Ogasawara/ Bonin Islands	A. Davison	12
University of the Philippines, Diliman, Quezon City, Philippines	I. Fontanilla	20
Los Baños, Philippines	M. Carandang	20
Singapore	M. Posa	20
Kota Kinabalu, Sabah, Malaysia	M. Schilthuizen & T. Liew	20
Noumea, New Caledonia	C. Wade	21
Moaroa Valley, Tahiti, French Polynesia	T. Coote	5
Haapiti Valley, Moorea, French Polynesia	T. Coote	10
Kaneohe, Oahu, Hawaii	K. Hayes	20
Martinique (captive bred F1 population, Nancy, Lorraine, France)	F. Adnai	20
Barbados	A. Norville	12

5.2.2. PCR and SSCP analysis using the 16S rRNA gene

Small tissue slices (approximately 8 mm³) were cut from the foot muscle of each snail and subjected to the NaOH direct lysis DNA extraction method for snails (Section 2.1.2 of Chapter 2, pp 39-40).

A 400 bp PCR fragment used for SSCP analysis can only detect 80% of single nucleotide differences whereas smaller fragments have an increased probability of the detection of differences (Sunnucks *et al.*, 2000). For the 16S rRNA gene, a primer pair, 16S1i and 16S_SSCP2i, was designed to produce a 293 bp fragment of domain IV of the post-transcribed 16S rRNA (Lydeard *et al.*, 2000). Note that primers 16S1i and 16S_SSCP2i bind to positions 781-802 and 1049-1068, respectively, of the 16S rRNA gene of *Cepaea nemoralis* (GenBank NC_001816). Details of the primer pair are shown in Table 2.5 of Chapter 2, p. 53.

The amplified PCR products were then subjected to SSCP analysis (Section 2.8 of Chapter 2, pp. 66-70). The bands in the native acrylamide gel were visualized using silver staining, after which haplotypes were identified and scored. Representative samples bearing unique haplotype gel profiles were checked for single nucleotide differences by re-amplifying the PCR product from the DNA sample and sequencing the amplified fragment. Both sense and anti-sense strands were sequenced directly using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 termination cycle sequencing chemistry (Section 2.6 of Chapter 2, pp. 59-61). The sequences were subsequently assembled in the STADEN package version 1.5.3 (Staden *et al.*, 2000) and aligned manually within GDE Version 2.2 (Smith *et al.*, 1994).

5.2.3. Phylogenetic and network analyses of the 16S rRNA haplotypes

To determine the evolutionary relationships of the different 16S haplotypes, the best model for sequence evolution with optimized parameters was determined and an NJ tree was generated in PAUP* Version 4.0b10 package (Swofford, 2002) (see Section 2.9.8 of Chapter 2, p. 84). *Achatina reticulata* was used as an outgroup as this species had the smallest uncorrected distance to *A. fulica* based on the same 16S rRNA SSCP fragment (see Appendix 5.1, p. 523). A median joining network of the haplotypes (Bandelt *et al.*, 1999) was also drawn using the Network Version 4.502 program (<http://www.fluxus-engineering.com>).

5.3. Results

5.3.1. PCR and SSCP analysis of the 16S rRNA gene

PCR and SSCP analysis carried out on 382 snails from 19 populations yielded 15 unique 16S rRNA haplotypes (Figure 5.2).

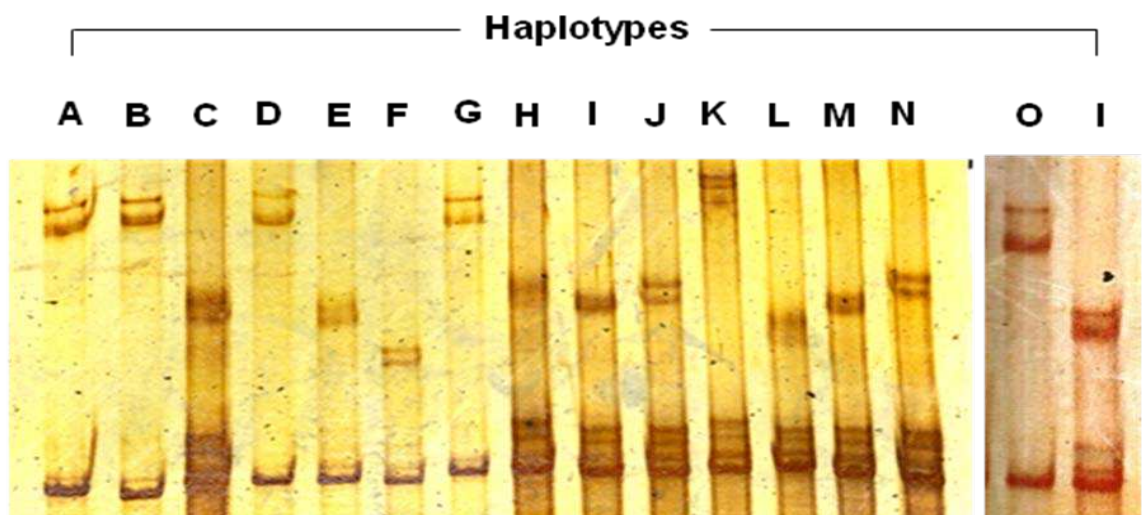


Figure 5.2: Gel profiles of 15 haplotypes (labeled A – O) based on a 293 bp fragment in domain IV of the 16S rRNA gene. (Haplotype O was run on a separate gel alongside haplotype I, which was used as basis of comparison.) Samples were run on a 50% native polyacrylamide gel at 180 V for 24 hours. The bands were visualized by silver staining.

PCR-direct sequencing of the haplotypes revealed 17 variable sites (Figure 5.3).

Two of these sites (pos. 285-286) were found to be indels.

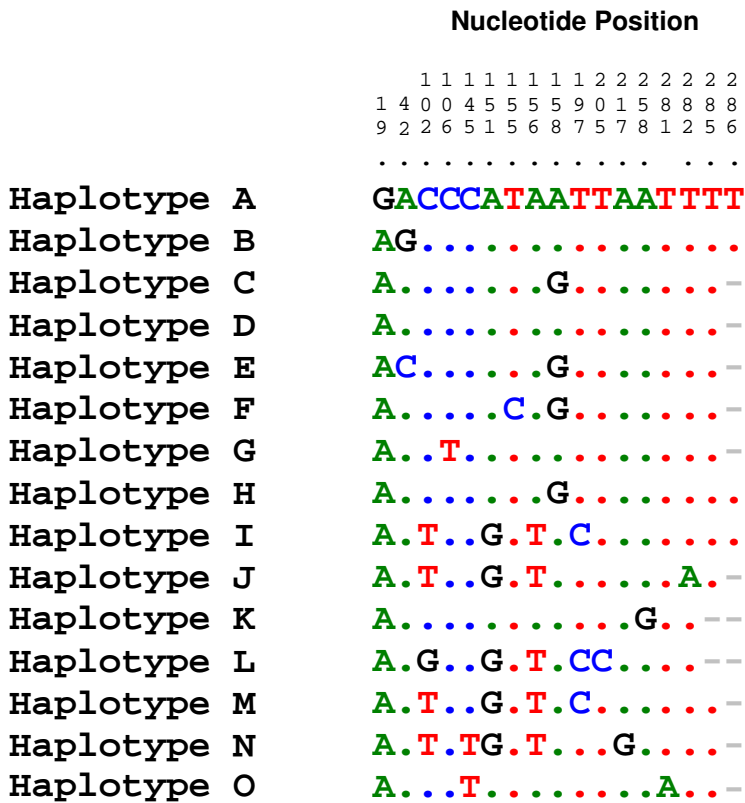


Figure 5.3: Seventeen variable sites across the 15 haplotypes. The number shown above each site refers to the variable site based on the 16S rRNA SSCP fragment.

A summary of the frequency and distribution of these haplotypes is shown in Table 5.2 and Figure 5.4. Seven haplotypes were found in the two East African populations: Uganda with one haplotype (O) and Tanzania with six haplotypes (I, J, K, L, M, and N). Among the Indian Ocean islands, Mayotte yielded the highest number of haplotypes with six (A, B, C, D, G and H), three of which were also found in Mauritius (C, D and H) and one in Seychelles (C). However, of the haplotypes found in Africa and on the Indian Ocean islands, only haplotype C was found in the populations surveyed outside East Africa and the Indian Ocean islands, thus making type C the only

pan-tropical haplotype. There was also one unique haplotype (E) detected in the Los Baños site in the Philippines where it was found in conjunction with haplotype C. Another unique haplotype (F) was found in New Caledonia in the Pacific and Barbados in the Caribbean where it was the only haplotype identified for both populations.

Table 5.2: Summary of the geographic distribution and frequency of the 16S rRNA haplotypes from *Achatina fulica* populations as evaluated by SSCP analysis.

Locality	Sample size (& no. of haplotypes)	Haplotype name (& no. of individuals)
Kampala, Uganda	5 (1)	O (5)
Dar Es Salaam, Tanzania	38 (6)	I (11) J (1) K (12) L (10) M (2) N (2)
Mayotte	50 (6)	A (1) B (1) C (6) D (40) G (1) H (1)
Souillac, Mauritius	45 (3)	C (34) D (9) H (2)
Mahe, Seychelles	2 (1)	C (2)
Bharatpur, Nepal	22 (1)	C (22)
Yangon, Myanmar/ Burma	20 (1)	C (20)
Trok Nong Area, Chantaburi, Thailand	20 (1)	C (20)
Hahasima, Ogasawara/ Bonin	12 (1)	C (12)
Quezon City, Philippines	20 (1)	C (20)
Los Baños, Philippines	20 (2)	C (17) E (3)
Singapore	20 (1)	C (20)
Kota Kinabalu, Sabah, Malaysia	20 (1)	C (20)
Noumea, New Caledonia	21 (1)	F (21)
Moaroa Valley, Tahiti, French Polynesia	5 (1)	C (5)
Haapiti Valley, Moorea, French Polynesia	10 (1)	C (10)
Kaneohe, Oahu, Hawaii	20 (1)	C (20)
Martinique (captive bred F1 population, Nancy, Lorraine, France)	20 (1)	C (20)
Barbados	12 (1)	F (12)
TOTAL	382 (15)	A (1) I (11) B (1) J (1) C (248) K (12) D (49) L (10) E (3) M (2) F (33) N (2) G (1) O (5) H (3)

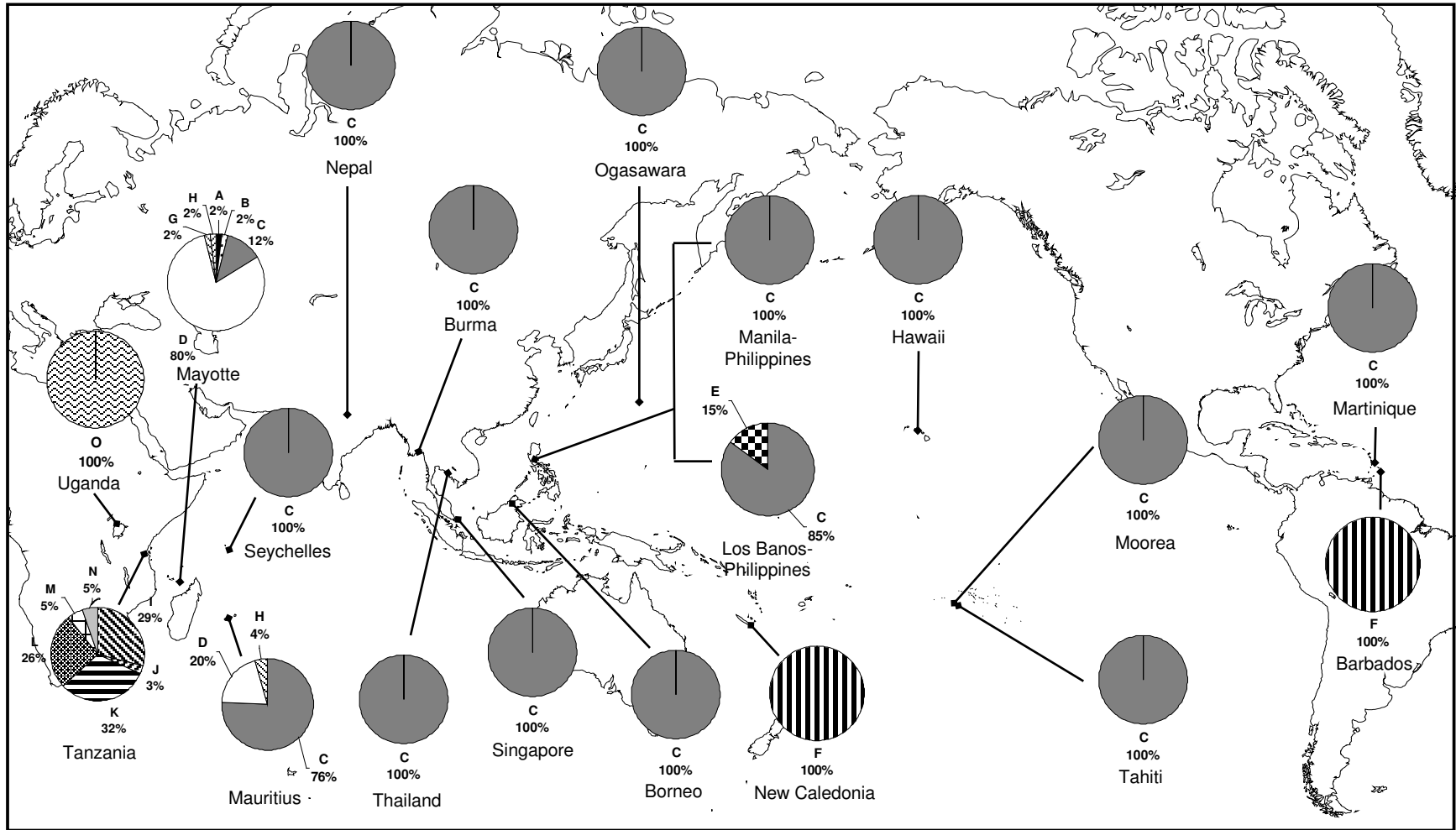


Figure 5.4: Distribution map of the 15 *Lissachatina fulica* populations and their 16S rRNA haplotypes.

5.3.2. Phylogenetic and network analyses of the 16S rRNA haplotypes

A neighbor-joining phylogenetic tree showing the interrelationships among the 15 haplotypes is shown in Figure 5.5 (see also alignment in Appendix 5.2, pp. 523-524). The tree was built using the optimal HKY85 model of DNA sequence evolution (see Appendix 5.3 for the LRT results, p. 525) and is rooted on *A. reticulata*. In general, there is very little phylogenetic structure within the tree and the interrelationships among the haplotypes remain largely unresolved. Nevertheless, two clades are apparent. The first comprises five of the six Tanzanian haplotypes (I, J, L, M and N) and is supported in 96% NJ bootstraps. The second comprises the pan-tropical (Mayotte) haplotype C, the East African (Mayotte) haplotype H, the Philippine haplotype E and the New Caledonian/Barbados haplotype F, albeit with only 54% NJ bootstrap support.

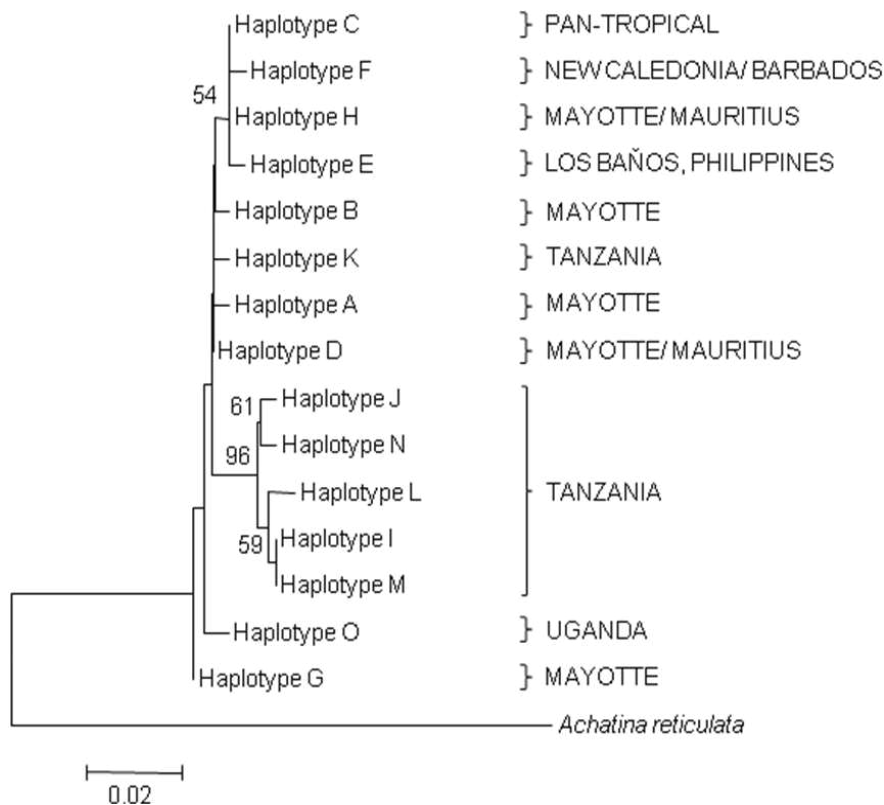


Figure 5.5: Neighbor-joining tree of the 15 *Achatina fulica* 16S rRNA haplotypes based on 293 unambiguously aligned nucleotide sites and using the optimal HKY model. The tree was rooted on the East African *Achatina reticulata*. Bootstrap values indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar represents two substitutional changes per 100 nucleotide positions.

Median network analysis (Figure 5.6) showed that the Tanzanian haplotypes (I, J, L, M and N) that formed a clade in the NJ tree were also grouped together in the network analysis where they were linked to the Mayotte haplotype D with three substitutions. The Tanzanian haplotype K was also linked to haplotype D by two substitutional steps but was separated from the other Tanzanian haplotypes. Other haplotypes linked to the Mayotte haplotype D were the Ugandan haplotype O, Mayotte haplotype G and the pantropical (Mayotte) haplotype C with one substitutional step each. Mayotte haplotype H, in turn, was linked to haplotype C by one substitution. Mayotte haplotypes A and B probably originated independently by one substitutional

step from a hypothetical haplotype, which, in turn, was probably only one substitutional step away from either D or H. The non-African haplotypes (E and F) arose through a single mutation each from the pantropical (Mayotte) haplotype C.

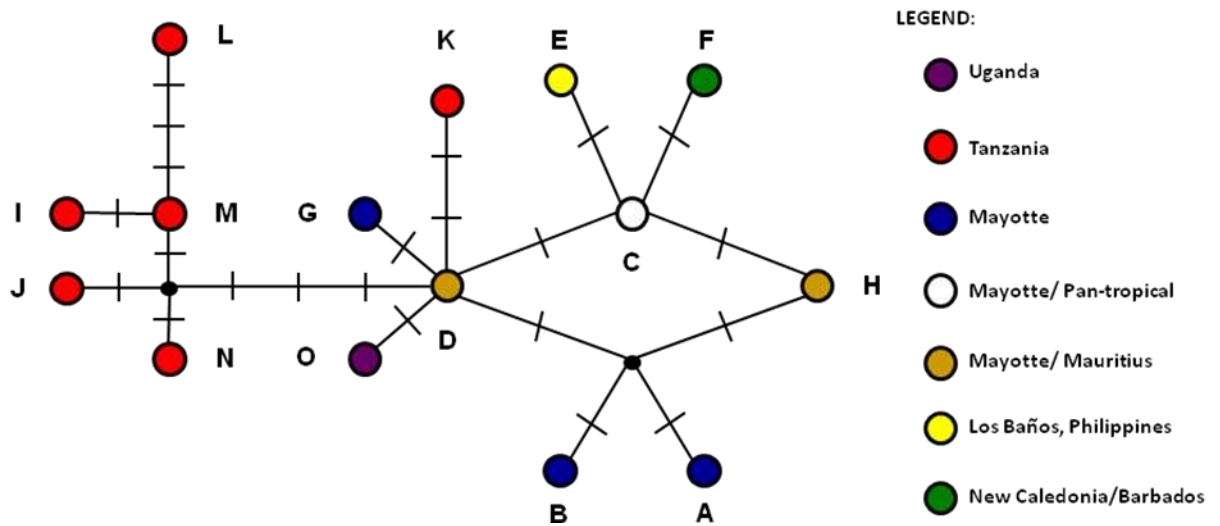


Figure 5.6: Median-joining network of the 15 *Achatina fulica* 16S haplotypes. Each mutation between haplotypes is represented by a bar. Unsampled putative haplotypes are represented by black dots.

5.4. Discussion

Results of the SSCP analysis, NJ tree and median-joining network indicated that all *Achatina fulica* populations outside East Africa and adjacent islands descended from haplotype C, presumably from a few individuals, as there is a complete absence of any other East African haplotypes among the non-African populations. Multiple introductions that could offset the lack of genetic variation as a consequence of founder events were therefore not detected. In India in particular, historical records suggest that the prevalence of *Achatina fulica* could be traced back to the introduction of two individuals from Mauritius to Calcutta by the malacologist William Benson in 1847 (Naggs, 1997). These snails were released by Benson's friend into his garden (Naggs,

1997) and then began to proliferate and slowly spread to the rest of India and later to Nepal (Raut & Barker, 2002; Budha & Naggs, 2008). Though samples were not available from India, particularly from Calcutta, they are expected to bear haplotype C because the population from Nepal also harbored this haplotype. Haplotype C was also the most common haplotype found in Mauritius, the source of Benson's *A. fulica*. The Giant African Land Snail was introduced separately to Sri Lanka from India and Mauritius by Oliver Collett sometime in 1900 (Mead, 1961; Naggs, 1997), and it would be worthwhile to investigate if *A. fulica* individuals in that island are also of the C haplotype or if they harbor other African haplotypes. It is highly likely, however, that the Sri Lankan *A. fulica* would carry the C haplotype as the snail populations from Peninsular Malaysia were derived from Sri Lanka, and Peninsular Malaysia, in turn, became the source of the snail for Burma, Singapore and Thailand (Mead, 1961), all of which bear the C haplotype; Singapore also became the source of the snail for Borneo (Jarrett, 1931), and the population there also carried the C haplotype. A similar case to India also took place in Hawaii where the snails descended from two individuals introduced there from Taiwan (Formosa) in 1936 (Pemberton, 1938), which in turn got the snail from Singapore after 1917 (Mead, 1961). As the *A. fulica* populations in Hawaii and Singapore are of the C haplotype, *A. fulica* populations in Taiwan would therefore also be expected to harbor the C haplotype.

The median-joining network demonstrated that the non-African haplotypes E (Philippines) and F (New Caledonia and Barbados) were probably recent mutations and were derived from haplotype C. To date, haplotype E has only been found in the Philippines where it is present alongside the pantropical haplotype C. It is therefore likely that haplotype E originated in the Philippines and since we know that *Achatina fulica* was introduced in the Philippines in 1931 (Muniappan *et al.*, 1986) this would

suggest that the single nucleotide substitution from C to E took place within the last 70 years or so.

The presence of F in areas that are far apart from each other such as New Caledonia in the Pacific and Barbados in the Caribbean would indicate that either both areas had the same source or that the snails from Barbados originated from New Caledonia as the snail was introduced earlier in New Caledonia (Lambert, 1974). A less likely possibility is that haplotype F arose independently by mutation in both places. Further sampling in New Caledonia would determine if the mutation from C to F took place there or elsewhere. To date, only haplotype F has been found in New Caledonia but so far sampling has been restricted to a single site in Noumea.

The relationship of the East African haplotypes from Tanzania and Uganda with Mayotte is less clear due to limited sampling in the region. It is believed that the Giant African Land Snail was introduced first in Madagascar from Kenya prior to 1800, and it is these Madagascan *A. fulica* snails that were eventually introduced to nearby islands (Bequaert, 1950; Raut & Barker, 2002) such as Mayotte. This raises some very important questions. Is haplotype C found in Madagascar and mainland East Africa? If so, then this would suggest that C was originally present in East Africa and was brought to nearby islands such as Mayotte. If C is *not* present in mainland East Africa, then this would indicate that this haplotype originated on islands off East Africa, possibly on Madagascar or indeed even on Mayotte itself. This question also applies to the other Mayotte haplotypes. Are they also found in Madagascar and mainland East Africa? Clearly, sampling from East Africa and Madagascar is necessary in order to answer these questions and get a clearer picture of the pattern of genetic variation in *A. fulica* populations in East Africa.

The use in this study of a single mitochondrial gene marker such as the 16S rRNA was sufficient to show that all the non-African *Achatina fulica* populations were very closely related and are undergoing bottlenecks as measured by the reduction or loss of genetic variation due to founder events. Whether or not this observation extends to other genes warrants further investigation. Using different markers to measure genetic variation can lead to conflicting results because these markers evolve independently of each other or may have varying degrees of sensitivity. This was proven to be the case when the effectiveness of various molecular markers on native and introduced North American house finches (*Carpodacus mexicanus*) were compared. Hawley *et al.* (2008) found that AFLP, mitochondrial DNA RFLP and allozyme markers did not show statistically significant difference in the degree of variability between introduced and native finches whereas 12 microsatellite loci and ND2 mitochondrial sequences did. They recommended that rapidly evolving single-locus haploid mitochondrial markers be used in conjunction with multi-locus variable nuclear microsatellite markers to detect the presence of recent bottlenecks.

Whether the lack of genetic variation has any effect on the overall fitness of the *A. fulica* populations is another matter altogether. Mead (1979) noted that the snail frequently undergoes three phases after establishment in a new area: (1) an exponential increase characterized by large individuals; (2) a stable phase of variable duration; and (3) decline exemplified by small individuals. Civeyrel and Simberloff (1996) also cited similar patterns in other introduced species; these include the cane toad (*Bufo marinus*) in Australia, the red-billed leiothrix (*Leiothrix lutea*) in Oahu, Hawaii and the weasel (*Mustela nivalis*) in New Zealand. Mead (1979) speculated that the decline in *A. fulica* could be attributed to the snail's susceptibility to the bacterium *Aeromonas hydrophila*, which causes leucodermic lesions. He based his theory on a study by Dean *et al.* (1970)

in which the bacterium, then called *Aeromonas liquefaciens* but was later synonymised with *Aeromonas hydrophila* by Ewing & Hugh (1974), was isolated from the snails and a positive correlation was detected between the presence of the bacteria and the manifestation of leucodermic lesions. What leads to the susceptibility is unknown, but Raut & Barker (2002) cited two possible explanations. One is the breakdown of natural resistance caused by stress in a densely populated environment (Mead, 1979) and the other is the increased chance of transmission of the bacterium due to crowding (Civeyrel & Simberloff, 1996). An alternative explanation is the depletion of food supply as a result of high population density, leading to starvation of the snails and making them more susceptible to bacterial infection (Civeyrel & Simberloff, 1996) or to any other disease-causing vector. Cowie (pers. comm.) offered yet another explanation in which he speculated that native and genetically diverse populations of *A. fulica* were naturally neither abundant nor large because they harboured the bacterium (or other disease agent). When the introduced populations were released from the disease agent pressure, it allowed them to become abundant and larger; eventually, however, the disease made its way to the new region and re-associated with the snails, reducing population density and snail size.

Whether or not the decline phenomenon observed in introduced *Achatina fulica* populations in many areas across the globe is a consequence of founder events is difficult to say. It should be noted that in general, the loss of genetic variation (i.e. heterozygosity and presence of rare alleles) may seriously affect the ability of future generations to adapt to changing environments and subsequently trigger a decline (Backeljau *et al.*, 2001). This was seen in the house finch, *Carpodacus mexicanus*, in which resistance to experimentally infected *Mycoplasma gallisepticum* is associated with increasing multilocus heterozygosity in 12 microsatellites (Hawley *et al.*, 2005). This

led Hawley *et al.* (2008) to speculate that the susceptibility of the house finch to the bacterium in its introduced range in eastern North America, the site of an epidemic in 1994, could be attributed to the reduced genetic variation of the introduced populations as a consequence of founder events. Indeed, their study (2008) revealed reduced genetic variability among the introduced populations of *C. mexicanus* as demonstrated by microsatellite and mitochondrial DNA data. A similar situation could also be taking place among global populations of *A. fulica* in which reduced heterozygosity or variability due to founder events could lead to susceptibility to bacteria or other disease-causing agents in the Giant African Land Snail.

As already mentioned in previously, this study is limited by the availability of sampling, and many crucial areas were not covered, particularly from East Africa (i.e. Kenya and Madagascar), the Indian subcontinent (Calcutta in India and Sri Lanka) and even in the Caribbean and South America (Brazil). The logical next step is to gather samples from these localities. Sampling from Kenya and Madagascar will most likely provide additional haplotypes not found in Uganda, Tanzania and Mayotte, which could give a clearer picture of the pattern of genetic variation in East Africa. Samples from Calcutta in India would determine if indeed the snails in India descended from two haplotype C individuals (if only C are found) or if multiple introductions took place (if other haplotypes are found). Likewise, samples from Sri Lanka would determine whether other haplotypes are present on the island, which could indicate more than one introduction. Sri Lanka is crucial as it served as a source of the Giant African Land Snail for some countries like Malaysia that ultimately became the source for many other countries. Additional sampling from the other Caribbean islands such as Guadeloupe could show if other areas also harbor haplotype F and if introduction occurred from different sources. The presence of *A. fulica* in Brazil is a result of recent introduction to

raise snails for food (Paiva, 1999; Carvalho de Vasconcellos & Pile, 2001; Thiengo *et al.*, 2007) and was probably derived from Indonesia (Thiengo *et al.*, 2007). Sampling from either Brazil or Indonesia will determine whether the pantropical haplotype C is also present in these areas or whether the populations in these areas comprise other haplotypes. Other South American countries such as Colombia, Ecuador and Peru are also worth investigating to track the spread of the snail in the continent.

5.5. Summary

The extent of genetic diversity in *Achatina fulica* populations was investigated using single-stranded conformational polymorphism (SSCP) analysis. A fragment of the 16S rRNA gene found within the variable domain IV was used as marker for the SSCP analysis. Fifteen haplotypes were identified from populations of *A. fulica*, but only haplotype C emerged to spread across the globe. Two non-African haplotypes (E from the Philippines and F from New Caledonia and Barbados) were recent mutations derived from haplotype C as shown by the neighbor-joining phylogenetic and median-joining network analyses. Non-East African populations of *A. fulica* therefore exhibited a lack of genetic variation based on the 16S rRNA marker, implying bottlenecking has taken place in these populations.

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CHAPTER 6 - The SSU rRNA gene as a genetic marker for identifying infective 3rd juvenile stage *Angiostrongylus cantonensis* from known gastropod intermediate hosts¹

6.1. Introduction

The rat lungworm, *Angiostrongylus cantonensis* (Chen), is a nematode parasite of murid rodents (Ubelaker, 1986; Marquardt *et al.*, 2000). Its life cycle requires invertebrate intermediate hosts such as snails and slugs (Roberts & Janovy, 2005). The lungworm can also accidentally infect humans where it causes eosinophilic meningoencephalitis (EME) or neurologic angiostrongyliasis, a disease of the central nervous system that leads to headache, vomiting, nausea, neck stiffness and photophobia (Kliks & Palumbo, 1992; Senanayake *et al.*, 2003; see also Section 1.8.5 of Chapter 1, pp. 26-28). Humans become infected with the 3rd juvenile stage *A. cantonensis* by handling infected snails (Wan & Weng, 2004) or by eating raw/undercooked infected snails or contaminated salad crops (Marquardt *et al.*, 2000). Reported cases of gastropod, rat and human infection reflect the wide geographic distribution of *A. cantonensis*, from East Africa and Madagascar to the Indian subcontinent, East and Southeast Asia, Australia, most islands in the Pacific (Kliks & Palumbo, 1992; Marquardt *et al.*, 2000) and the Caribbean (Raccurt *et al.*, 2003; Waugh *et al.*, 2005).

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6.1.1. Problems with identifying nematodes: the angiostrongylids as examples

Accurately identifying medically important nematodes such as *A. cantonensis* is crucial in diagnosing and controlling the diseases that they cause (Gasser & Newton, 2000). However, it is difficult to identify worms to species level due to a lack of suitable morphological characters in both the adult (Newton *et al.*, 1998a; Roberts & Janovy, 2005) and, more importantly, the infective juvenile stages (Newton *et al.*, 1998a). Some examples of these medically important nematodes include various species of the hookworm *Ancylostoma* as well as the other hookworm *Necator americanus*, in which the juveniles of these worms, which are difficult to distinguish from one another, infect humans either through the oral route or by penetration of the skin and cause hookworm disease. Other examples include different species of *Strongyloides*, most notably *S. stercoralis*, whose juveniles infect humans by contact through contaminated water or soil and cause strongyloidiasis, leading to damage of the skin, lungs and intestines; juveniles of *Strongyloides* are also difficult to detect and identify (Roberts & Janovy, 2005).

Identification of members of the order Strongylida such as the angiostrongylids relies heavily on expert knowledge of the morphological characteristics of the copulatory bursa found only in adult males, which it uses to wrap around the body of the female during copulation (Newton *et al.*, 1998a; Roberts & Janovy, 2005). For example, adult *A. cantonensis* can only be distinguished from the closely related species *A. vasorum* (Baillet) by differences in the copulatory bursa; in *A. cantonensis* the lateral rays arise from a single trunk whereas in *A. vasorum* the lateral rays arise separately (see Fig. 1.3 in Chapter 1, p. 23) (Ubelaker, 1986). Complicating the identification of nematodes is the ambiguity of morphological characters of the juvenile worms (Newton *et al.*, 1998a). For instance, the infective 3rd juvenile stages of *A. cantonensis* and *A.*

vasorum are differentiated only through the fine point termination of the tail of the former and the digitiform termination of the tail in the latter (Ash, 1970, see Fig. 6.1). Whether juveniles of other *Angiostrongylus* species can also be distinguished based on differences in the tail is not known, and some of these species may possibly exhibit similar tail terminations to those of either *A. cantonensis* or *A. vasorum*. In order to identify these juvenile angiostrongylids with certainty, they must be fed to laboratory mice until such time the adults can be observed (Wallace & Rosen, 1969). However, this process takes time, is tedious and does not guarantee that all the juveniles will mature into adults. A viable alternative is clearly needed to identify the 3rd juvenile stage *A. cantonensis*.

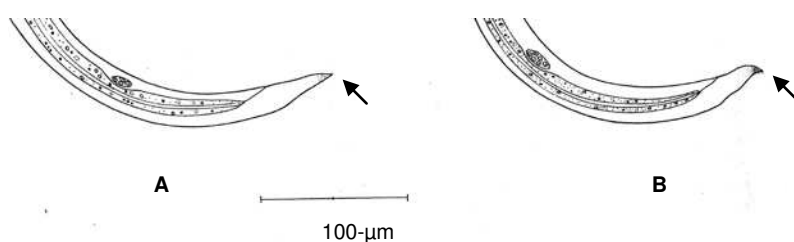


Figure 6.1: Posterior portion of the infective third juvenile stage of (A) *Angiostrongylus cantonensis* and (B) *Angiostrongylus vasorum*. Arrows indicate the type of termination of the tail for each nematode (e.g. fine point for *A. cantonensis* and digitiform, not fine point for *A. vasorum*). Taken from Ash (1970).

6.1.2. Identifying nematodes using molecular markers

In lieu of morphological expertise, the problem of identifying nematodes accurately and rapidly can be addressed by using various molecular genetic markers that are applicable at any stage of a nematode's development. Examples of these molecular markers are nuclear genes such as the ribosomal RNA genes and their internal transcribed spacers (ITS) (Newton *et al.*, 1998a; Gasser, 1999; Floyd *et al.*, 2002; Blaxter *et al.*, 2005) and mitochondrial genes like cytochrome c oxidase I (Caldeira *et*

al., 2003; Hebert *et al.*, 2003; Blaxter *et al.*, 2005). Using these markers requires PCR-based techniques that include polymerase chain reaction-restriction fragment length polymorphisms (PCR-RFLP) analysis and PCR-direct sequencing (Gasser, 1999).

In PCR-RFLP, PCR products from DNA target regions are digested by restriction enzymes. Depending on the sequence variations within the restriction sites that may result in either their recognition or non-recognition by restriction enzymes, fragments of varying lengths will be produced among different species. Through gel electrophoresis, each species can potentially generate a unique banding profile based on these fragments (Gasser, 1999). Newton *et al.* (1998a) employed this technique on the ITS-2 region between the 5.8S and the large subunit (LSU) ribosomal (r) RNA genes to characterize 24 previously described nematode species in the order Strongylida that parasitise livestock. The banding patterns they obtained from both adults and 3rd stage juveniles were unique for each of the species they surveyed except for *Cooperia oncophora* and *C. surnabada*. The ITS-2 can therefore be used to identify most of these parasites at specific developmental stages at which morphological characters are ambiguous. In another study, Caldeira *et al.* (2003) also used PCR-RFLP on the ITS2 as well as the mitochondrial cytochrome c oxidase subunit 1 to distinguish closely related taxa in the Angiostrongylidae such as *A. cantonensis*, *A. costaricensis* and *A. vasorum* using 3rd stage juveniles and adults. However, this technique cannot detect nucleotide substitutions outside the restriction site. Closely related species may share restriction sites but differ outside these restriction sites, thereby producing identical banding patterns after gel electrophoresis.

A more reliable molecular method of identifying nematodes is direct sequencing (Bhadury *et al.*, 2006). The ribosomal genes, which are advantageous for several reasons, have often been used for this purpose. First, they consist of highly variable and

phylogenetically informative regions interspersed with conserved regions that are suitable for primer-binding sites (Palumbi, 1996). Second, they are present as multiple copies of the same sequence (~50-100 copies in nematodes) arranged in tandem arrays across the genome, thus making them easier to PCR amplify (Floyd *et al.*, 2002). Finally, they show very little variation within a species due to concerted evolution (Gasser & Newton, 2000; Floyd *et al.*, 2002). When Newton *et al.* (1998a) generated the same banding profile for *Cooperia oncophora* and *C. surnabada* using PCR-RFLP of the ITS-2, they sequenced this region and discovered that these two taxa have identical sequences, which supports their hypothesis that they may in fact belong to the same species (Newton *et al.*, 1998b). In another example, Bhadury *et al.* (2006) employed a marker near the 5' end of the SSU rRNA gene to rapidly identify marine nematodes collected from southwest England. When they compared their sequences to public databases and conducted phylogenetic analyses, over 97% of their sequences corresponded with the correct species designation based on their morphological identification, and they concluded that amplifying and sequencing a specific region of the SSU rRNA gene could be utilized to rapidly identify nematodes. They accounted for the 3% misidentification either as DNA contamination or distortion of morphological features during ethanol preservation of the nematodes. In another study, Santos *et al.* (2006) successfully employed direct sequencing of the SSU rRNA to detect and identify nematode species in the family Anisakidae (i.e. *Anisakis simplex*, *Contracaecum maculates*, *Pseudoterranova decipiens* and *Hysterothylacium aduncus*) that parasitise the commercially important sardine *Sardina pilchardus*. Humans get infected by the juvenile stages of these nematodes by eating raw or inadequately processed fish. Still other studies by Qvarnstrom *et al.* (2007) and Hollingsworth *et al.* (2007), which were done concurrently with this study, used a 1134 bp fragment of the

SSU to successfully detect by direct sequencing the presence of *A. cantonensis* from tissue and mucus secretions from gastropod intermediate hosts, primarily the semi-slug *Parmarion cf. martensi* in Hawaii. However, only *A. cantonensis* (Carreno & Nadler, 2003) and *A. vasorum* (Chilton *et al.*, 2006) SSU sequences were available to Qvarnstrom *et al.* (2007) and Hollingsworth *et al.* (2007), and their studies were therefore unable to exclude the possibility of detecting other closely related *Angiostrongylus* species. Moreover, sequencing the entire SSU rRNA gene to identify nematodes is not practical if a shorter fragment will suffice. Floyd *et al.* (2002) noted that 50% of the variations found in the SSU rRNA gene of nematodes occur in the first one third from the 5' end (approximately 450 bp), and sequences available for this region for many nematodes in GenBank could distinguish different species. They also considered individuals to belong to the same species if their sequences of the 5' end of the SSU rRNA gene are more than 99.5% identical.

6.1.3. Objectives of the study

The main objective of this study was to develop a quick and reliable molecular method for the identification of infective 3rd juvenile stage *Angiostrongylus cantonensis* based on PCR-direct sequencing a fragment of the small subunit (SSU) ribosomal (r) RNA gene. The SSU rRNA gene was chosen because it is well represented in nematodes in public databases (Floyd *et al.*, 2002) and is variable enough to permit differentiation of closely related nematode species (Gasser & Newton, 2000). The SSU sequence of *A. cantonensis* (voucher specimen no. 92610 of the US National Parasite Collection; identified using published descriptions) became available in Genbank during the course of my PhD studies (Accession AY295804, Carreno & Nadler, 2003) while that of *A. vasorum* (Accession AJ920365, Chilton *et al.*, 2006)

became available after I had already sequenced another individual from the same species (see below). However, there are no full length SSU sequences for many other closely related species that could be mistaken for *A. cantonensis*. To ascertain whether the SSU rRNA gene is indeed suitable to distinguish *A. cantonensis*, the full length SSU for several species of *Angiostrongylus* was sequenced and a suitable variable marker region within the gene determined. In particular, species closely related to *A. cantonensis* were chosen, such as those grouped together by Ubelaker (1986) under *Parastrongylus* based on the morphological characteristics of the male copulatory bursa and host specificity. To test the efficacy of the marker for identifying the 3rd juvenile stage *A. cantonensis*, worms were extracted and identified from gastropod intermediate hosts from a trial area, the Philippines, particularly in Manila, which is known for its prevalence of the parasite (De Leon & Saulog, 1965; Nishimura & Yogore, 1965) and its suburb Quezon City (Salazar & Cabrera, 1969). Furthermore, Salazar & Cabrera (1969) positively identified 3rd juvenile stage *A. cantonensis* in several gastropod intermediate hosts such as the widespread giant African land snail, *Achatina fulica* (with an infection rate of 4.52%), and the common black slug *Laevicaulis alte* (18.18% infected).

As this study resulted in new SSU sequences for several *Angiostrongylus* species, and with two other angiostrongylid SSU sequences already available in GenBank (Carreno & Nadler, 2003; Chilton *et al.*, 2006), a second objective was to conduct phylogenetic analyses of the Angiostrongylidae. These analyses had a dual purpose: to test the monophyly of the group as well as to test Ubelaker's (1986) hypothesis that some members of the genus *Angiostrongylus*, including *A. cantonensis*, are distinct and should be reclassified under the genus *Parastrongylus*.

6.2. Materials and methods

6.2.1. Sequence variation in angiostrongylid worms

Nearly the entire length of the SSU rRNA gene was obtained for four species of Angiostrongylidae: *Angiostrongylus costaricensis*, *A. dujardini*, *A. malaysiensis* and *A. vasorum*. These species were identified, based on published descriptions of the morphological characters of the adult and their definitive hosts, by the taxonomic experts listed in Table 6.1

Table 6.1: Detailed information of the angiostrongylid worms used in this study.

Specimen	Host	Location	Collector/Provider
<i>Angiostrongylus costaricensis</i>	<i>Sigmodon hispidus</i> (cotton rat) & <i>Biomphalaria glabrata</i> (blood fluke planorbid snail)	Maintained in the Department of Parasitology, Hamamatsu University School of Medicine, Japan	Akira Ishih
<i>Angiostrongylus dujardini</i>	<i>Clethrionomys glareolus</i> (red-backed mouse)	Prades, Pyrenees- Orientales, France	Serge Morand & Carlos Feliu
<i>Angiostrongylus malaysiensis</i>	<i>Rattus tiomanicus</i> (Malaysian field rat)	Endau-Rompin National Park, Johor, Malaysia	Siti Nursheena Mohd Zain
<i>Angiostrongylus vasorum</i>	<i>Vulpes vulpes</i> (common fox)	Swansea, Great Britain	Eric Morgan

With the exception of *A. vasorum*, the DNA of which was provided by Eric Morgan of the University of Bristol School of Biological Sciences, all other samples were subjected to DNA extraction following the NaOH direct lysis protocol of Floyd *et al.* (2002) (see Section 2.1.4 of Chapter 2, pp. 41-42).

For the PCR, the 4 overlapping primer sets (see Section 2.2.1.2 of Chapter 2, pp. 53-54) of Blaxter *et al.* (1998) were used to amplify nearly the entire fragment (approximately 1670 nucleotides) of the SSU rDNA gene. PCR amplification, gel

migration and purification of PCR products from agarose gels were done as previously described (see Sections 2.2.1.2 – 2.4 of Chapter 2, pp. 53-59). Both sense and anti-sense strands were sequenced directly using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 termination cycle sequencing chemistry (see Section 2.6 of Chapter 2, pp. 59-61).

Sequences were assembled using the STADEN package version 1.5.3 (Staden *et al.*, 2000) and aligned manually within the Genetic Data Environment (GDE) Version 2.2 (Smith *et al.*, 1994) together with the previously published SSU sequence of *Angiostrongylus cantonensis* (Carreno & Nadler, 2003). The primer set that amplified the most variable region that is also unique to *A. cantonensis* was chosen to identify the parasite.

6.2.2. Using the SSU rRNA gene to identify *Angiostrongylus cantonensis* in a trial population

The widely distributed Giant African Land Snail, *Achatina fulica*, and the black slug *Laevicaulis alte* were sampled for *A. cantonensis* from a grassy area within the College of Science complex at the University of the Philippines, Diliman campus in Quezon City, Philippines (samples collected by I. Fontanilla). *Angiostrongylus cantonensis* is common in Manila and its suburb Quezon City. Moreover, as rats frequent the Diliman collection site, snails and slugs were highly likely to be infected with *A. cantonensis*.

Snails and slugs were cut into small pieces and digested overnight in a Petri dish containing Ash's (1970) digestive fluid (0.7% pepsin in 0.5% HCl). Individual nematodes were collected under a dissecting microscope using a yellow-tipped pipette

(20-200- μ l capacity) and transferred into wells containing 100- μ l TE buffer in a 96-well microtitre plate. The worms were then stored in a -20 °C freezer until use.

DNA extraction, PCR amplification and direct sequencing were undertaken for each nematode using the same protocols described in Section 6.2.1 except that only the primer set that amplified the most variable region among the *Angiostrongylus* species was used (Primer Set A – see results). In cases in which the PCR amplifies products from both the nematode and the snail or slug host, the nematode band can be clearly distinguished as it is smaller (~480 bp) than the snail/slug host band (~510 bp). Once the purified PCR products were obtained, only the antisense strand (amplified by SSU_R09 of Set A) was directly sequenced.

Nematode sequences were processed using the STADEN package, after which distinct nematode sequences were identified and a BLAST search in GenBank was carried out to determine an exact match or identify the closest similarity for each sequence. To complement the BLAST results, a neighbor-joining (NJ) tree based on 376 unambiguously aligned nucleotide sites was generated in order to illustrate the phylogenetic position of the nematode sequences relative to all known nematode sequences in GenBank (GB Release No. 157, see Appendix 6.1, pp 526-536) as well as the new *Angiostrongylus* sequences obtained in this study. The phylogeny was rooted on 4 non-nematode taxa: *Chordodes morgani* (AF036639, Blaxter *et al.*, 1998) and *Gordius aquaticus* (X87985, Winnepenninckx *et al.*, 1995) of Phylum Nematomorpha, *Priapulius caudatus* (Z38009, Aleshin *et al.*, 1998) of Phylum Priapulida, and *Brachionus plicatis* (U49911, Aguinaldo *et al.*, 1997) of Phylum Rotifera. (See Appendix 6.2, pp 537-603 for the Nematoda alignment). The optimal model for DNA sequence evolution was determined using likelihood by estimating the log likelihood scores in PAUP* for the JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P

(Kimura, 1980), HKY85 (Hasegawa *et al.*, 1985), TN93 (Tamura & Nei, 1993) and the GTR (Rodriguez *et al.*, 1990) models as well as their variants that incorporated gamma distributed rates (Γ) (Yang, 1993) followed by comparing these scores for significant differences using the Likelihood Ratio Test (LRT) as described in Section 2.9.5 of Chapter 2, pp. 73-77). Bootstrap resampling (Felsenstein, 1985) with 1000 replicates was carried out. Once the phylogenetic positions of the nematode sequences were identified with specific clades, other nematode taxa that do not belong to these clades were removed in order to construct a smaller NJ tree based on the same fragment (376 sites) and using the best model with optimised parameters; however, a suitable outgroup [*Plectus acuminatus* (AF037628, Blaxter *et al.*, 1998), which falls immediately outside of the Rhabditida clade of Meldal *et al.* (2006) –see results] was retained.

6.2.3. Phylogeny of the Angiostrongylidae

In order to investigate relationships among the Angiostrongylidae, the full length SSU rRNA of all *Angiostrongylus* species that were not sequenced prior to this study as well as *A. cantonensis* from Carreno & Nadler (2003) were aligned with the rest of the sub-order Metastrongylina/ superfamily Metastrongyloidea and phylogenetic trees constructed. Two taxa from the Trichostrongylina, which falls immediately outside of the Metastrongylina based on phylogenetic studies of the Metastrongyloidea (Carreno & Nadler, 2003) and the Strongylida (Chilton *et al.*, 2006), were included in the alignment to serve as outgroups (Table 6.2). Phylogenies were constructed from the aligned dataset using the model based maximum likelihood (ML), Bayesian inference (BI) and neighbor-joining (NJ) methods as well as the non-model based maximum parsimony (MP) method (Section 2.9.8 of Chapter 2, pp. 82-85). A total of 1628 unambiguously aligned nucleotide sites were used for the phylogenies. The sequences were first

checked for base composition bias using the Chi square test in PAUP* (version 4.0b10) program (Swofford, 2002). The optimal model for DNA sequence evolution was then determined for the model-based methods. Bootstrap resampling (Felsenstein, 1985) with 1000 replicates for MP and NJ and 100 replicates for ML was also carried out. Bayesian inference (BI) was performed on the MrBayes (version 3.1.2) package (Ronquist & Huelsenbeck, 2003) using four chains of Markov Chain Monte Carlo algorithm to explore the tree space for 2 million generations with sampling at every 100 generations. The heating parameter was set to 0.125 to ensure adequate chain swapping. A consensus tree was constructed using the last 1000 trees (burnin=19001 samples).

Table 6.2: Taxa used for the sequence analysis of the angiostrongylid worms.

Sub-order	Family	Species	Reference	GenBank Accession #	
Metastrongylina (Superfamily Metastrongyloidea)	Angiostrongylidae	<i>Angiostrongylus costaricensis</i> (Morera & Cespedes, 1971)	This study	EF514913	
		<i>Angiostrongylus dujardini</i> (Drozdz & Doby, 1969)	This study	EF514915	
		<i>Angiostrongylus malaysiensis</i> (Bhaibulaya & Cross, 1971)	This study	EF514914	
		<i>Angiostrongylus cantonensis</i> (Chen, 1935)	Carreno & Nadler (2003)	AY295804	
		<i>Angiostrongylus vasorum</i> (Baillet, 1866)	This study	EF514916	
		<i>Aelurostrongylus abstrusus</i> (Raillet, 1898)	Chilton <i>et al.</i> (2006)	AJ920366	
		<i>Didelphostrongylus hayesi</i> Prestwood, 1976	Carreno & Nadler (2003)	AY295806	
	Crenosomatidae	<i>Crenosoma mephitidis</i> Hobmaier, 1941	Carreno & Nadler (2003)	AY295805	
		<i>Otostrongylus circumlitus</i> (Railliet, 1899)	Carreno & Nadler (2003)	AY295813	
		<i>Troglostrongylus wilsoni</i> (Stough, 1953)	Carreno & Nadler (2003)	AY295820	
	Filaroididae	<i>Filaroides martis</i> (Werner, 1782)	Carreno & Nadler (2003)	AY295807	
		<i>Ostlerus osleri</i> (Cobbold, 1889)	Carreno & Nadler (2003)	AY295812	
		<i>Parafilaroides decorus</i> Dougherty & Herman, 1947	Carreno & Nadler (2003)	AY295814	
	Metastrongylidae	<i>Metastrongylus salmi</i> (Gedoelst, 1823)	Carreno & Nadler (2003)	AY295809	
	Protostrongylidae	<i>Muellerius capillaris</i> (Müller, 1889)	Carreno & Nadler (2003)	AY295810	
		<i>Parelaphostrongylus odocoilei</i> (Hobmaier & Hobmaier, 1934)	Carreno & Nadler (2003)	AY295815	
	Pseudaliidae	<i>Halocercus invaginatus</i> (Quekett, 1841)	Carreno & Nadler (2003)	AY295808	
		<i>Pseudalius inflexus</i> (Rudolphi, 1808)	Carreno & Nadler (2003)	AY295816	
		<i>Stenurus minor</i> (Kühn, 1829)	Carreno & Nadler (2003)	AY295817	
		<i>Torynurus convolutus</i> (Kühn, 1829)	Carreno & Nadler (2003)	AY295818	
	Skrjabingylidae	<i>Skrjabingylus chitwoodrum</i> Hill, 1939	Carreno & Nadler (2003)	AY295819	
	Trichostrongylina (Superfamily Trichostrongylina) --OUTGROUP--	Molineidae	<i>Nematodirus battus</i> (Crofton and Thomas, 1951)	Zarlenga <i>et al.</i> (1994)	U01230
		Heligmonellidae	<i>Nippostrongylus brasiliensis</i> (Travassos, 1914)	Blaxter <i>et al.</i> (1998)	AF036597

6.3. Results

6.3.1. Sequence variation in the angiostrongylid worms

When the sequences of all five *Angiostrongylus* species available to date (including the four new *Angiostrongylus* sequences obtained in this study) were compared, a total of 31 variable sites were found, including an indel at position 1323 (Fig. 6.2). A pairwise comparison of the number of differences between any two *Angiostrongylus* species (Table 6.3) reveals that the least number of differences (9) was found between *A. cantonensis* and *A. malaysiensis* whereas the greatest number of differences (22) was seen between *A. malaysiensis* and *A. dujardini*. The sequences of the SSU rRNA clearly demonstrate that *A. cantonensis* has a distinct sequence that could be used to distinguish it from the other angiostrongylid worms in this study. Furthermore, 12 out of the 31 sites (or more than a third of the total) were within the first 480 bp from the 5' end. Moreover, *A. cantonensis* is distinct from all other *Angiostrongylus* species in this 480 bp 5' region. These findings demonstrate that the first 480 bp fragment of the SSU rRNA gene, which is amplified by primer Set A (see Section 2.2.1.2, pp. 53-54, and Table 2.6, p. 54 of Chapter 2), is a suitable marker to identify *A. cantonensis* and set it apart from other *Angiostrongylus* species sequenced to date, including three of its closest relatives, *A. costaricensis*, *A. dujardini* and *A. malaysiensis*, based on the morphology of the copulatory bursa (Ubelaker, 1986).

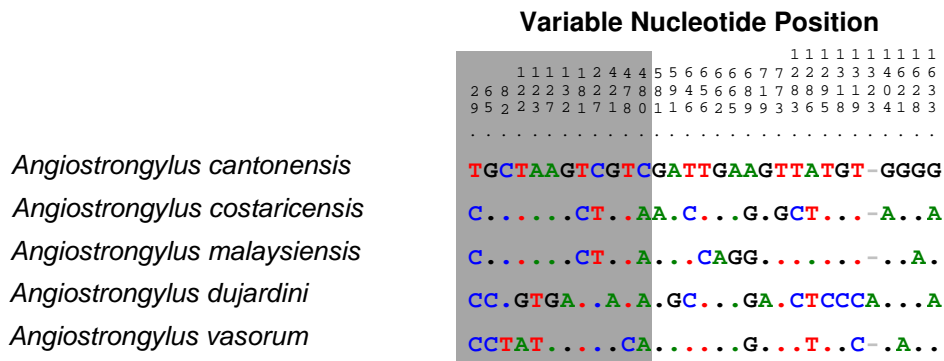


Figure 6.2: Variable sites across the SSU rDNA region for the previously sequenced *Angiostrongylus cantonensis* (Carreno & Nadler, 2003) and the four other angiostrongylid worms sequenced in this study. The number shown directly above each site pertains to the variable site relative to the SSU rRNA sequence of *A. cantonensis* (GenBank AY295804). The grey area corresponds to the 480 bp marker used for this study.

Table 6.3: Matrix showing the number of differences between any two *Angiostrongylus* species.

	<i>A. cantonensis</i>	<i>A. costaricensis</i>	<i>A. malaysiensis</i>	<i>A. dujardini</i>
<i>A. costaricensis</i>	12			
<i>A. malaysiensis</i>	9	11		
<i>A. dujardini</i>	19	16	22	
<i>A. vasorum</i>	11	15	14	16

6.3.2. Using the SSU rRNA gene to identify *Angiostrongylus cantonensis* in a trial population

In total, 27 nematodes were extracted from 15 *Achatina fulica* individuals (with 6 nematodes) and 19 *Laevicaulis alte* individuals (with 21 nematodes) from the University of the Philippines, Diliman campus in Quezon City. The 480 bp fragment at the 5' end of the SSU rRNA gene identified as a suitable marker for the discrimination of *A. cantonensis* (see Section 6.3.2, fragment amplified by Primer Set A) was then amplified for each nematode extracted. The results yielded two distinct nematode sequences (Philippine Sequences 1 and 2). Table 6.4 lists the distribution of these nematode sequences.

Table 6.4: Summary of the distribution of the two Philippine nematode sequences from the University of the Philippines, Diliman campus in Quezon City.

Nematode sequence	Gastropod host	Number of nematodes
Philippine Sequence 1	<i>Achatina fulica</i>	1
	<i>Laevicaulis alte</i>	20
Philippine Sequence 2	<i>Achatina fulica</i>	5
	<i>Laevicaulis alte</i>	1

The two Philippine nematode sequences were then subjected to a BLAST search in GenBank (GB Release No. 157, 22 December 2006), to identify an exact match or closest identity to a named nematode species. BLAST results showed Philippine Sequence 1 is identical to *Angiostrongylus cantonensis*. On the other hand, Philippine Sequence 2 did not match any named taxon in GenBank based on BLAST results but can only be assigned to its closest sequence identity to a named nematode species, which is *Oslerus osleri*, a dog lungworm, at 99% BLAST identity; since the value is less than 99.5% to be considered the same species as *O. osleri* based on Floyd *et al.* (2002), the exact species identification of Philippine Sequence 2 could therefore not be known as of GB Release No. 157.

To complement the BLAST results, a neighbor-joining tree of the Nematoda was generated in order to illustrate the phylogenetic position of the new nematode sequences relative to all 349 nematode taxa having at least the first 480 nucleotides of the 5' end of the SSU rRNA in GenBank (GB Release No. 157) as well as the new *Angiostrongylus* sequences obtained in this study. A GTR+ Γ model was used to correct distances for multiple hits after it was determined to be the best model (see Appendix 6.3A, p. 604). As this phylogeny is based on a small marker intended for barcoding purposes, it should not be expected to accurately show the deep-level evolutionary relationships amongst the nematodes (see Hajibabaei *et al.*, 2007 for a discussion of the limitations of “barcode” markers in phylogenetic studies). Nevertheless, the phylogeny

clearly demonstrates that the two Philippine sequences and all the *Angiostrongylus* taxa fall within the Rhabditida clade (as defined by Meldal *et al.*, 2006). Another NJ phylogeny of the same gene fragment (376 nucleotide sites) was then constructed (Fig. 6.3B) using 229 rhabditid taxa and rooted on the plectid nematode *Plectus acuminatus*. A GTR+ Γ model was determined to be the best model and was subsequently used for this analysis (see Appendix 6.3B, p. 604). The two Philippine nematode sequences and the *Angiostrongylus* taxa all fall within the Strongyloidea. Philippine Sequence 1 clusters with *Angiostrongylus cantonensis* (AY295804, Carreno & Nadler, 2003) with 100% BLAST identity. Philippine Sequence 2 clusters with *Oslerus osleri* (AY295812, Carreno & Nadler, 2003) with 99% BLAST identity.

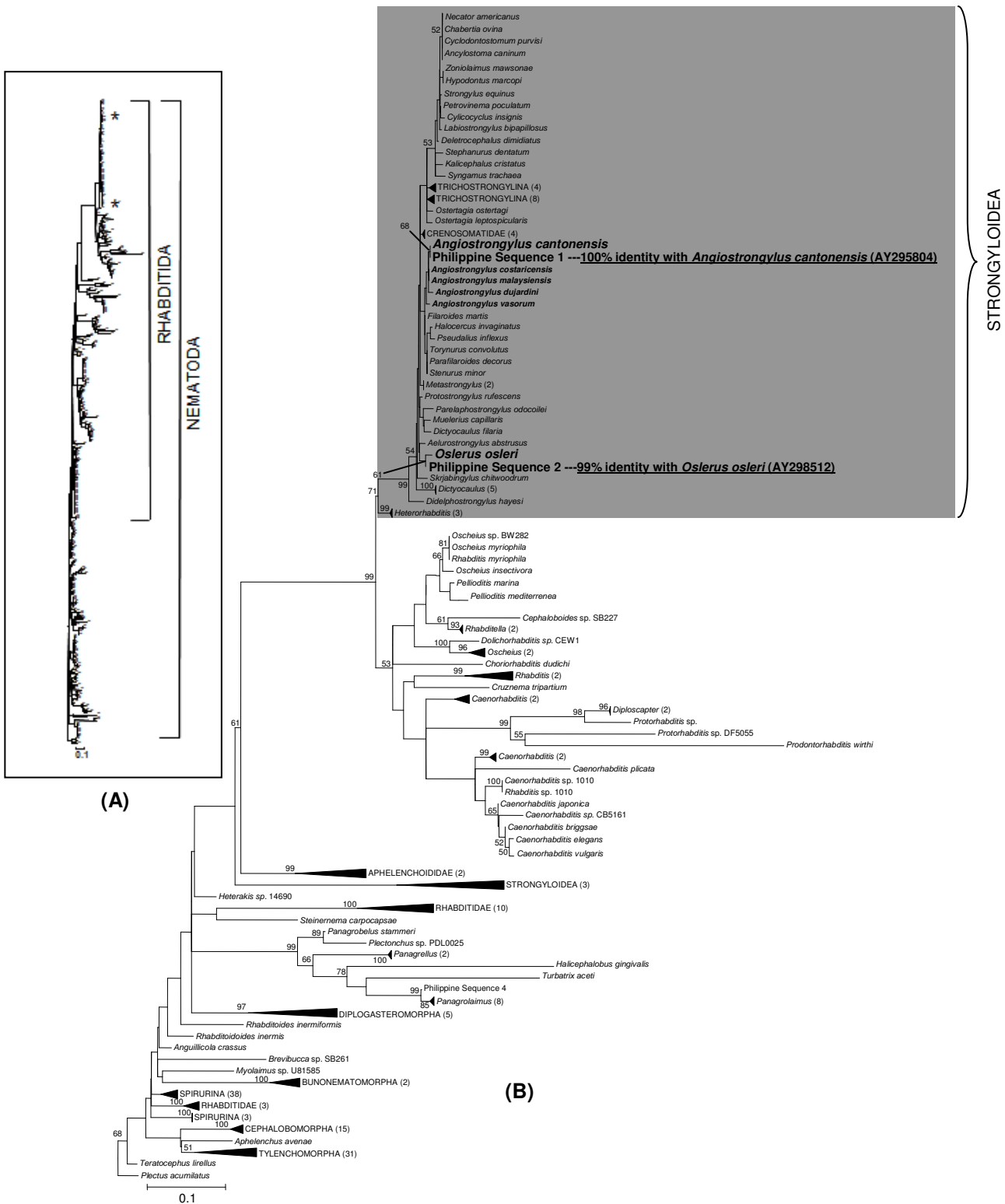


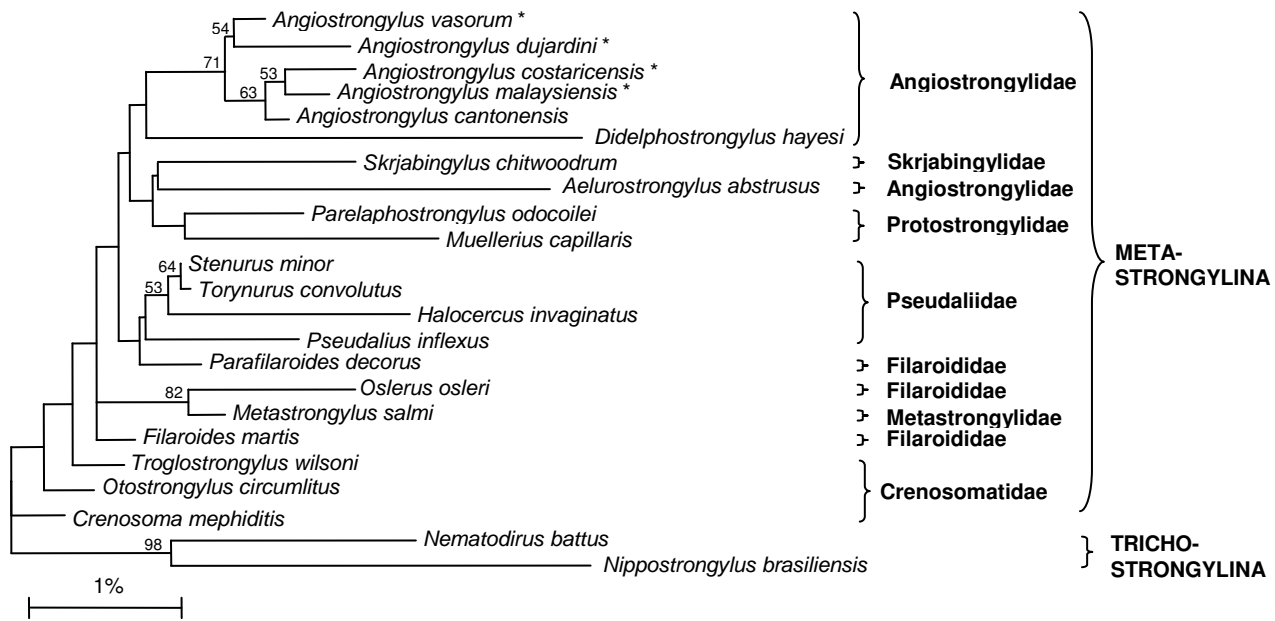
Figure 6.3: Neighbor-joining (NJ) phylogenetic trees based on 376 unambiguously aligned nucleotide sites of the SSU rRNA gene. **(A)** NJ tree of the Nematoda (349 taxa excluding the new nematode sequences from the Philippines) rooted on 4 non-nematode taxa: *Chordodes morgani* (Nematomorpha), *Gordius aquaticus* (Nematomorpha), *Priapulius caudatus* (Priapulida) and *Brachionus plicatis* (Rotifera). Both Philippine nematode sequences, as indicated by (*), fall within the Rhabditida clade. The scale bar shows one nucleotide change per ten nucleotide positions. **(B)** NJ tree of the Rhabditida (229 taxa excluding Philippine sequences) from (A), which is rooted on the nematode plectid worm *Plectus acuminatus*, and showing the positions (indicated by lines) of the 7 Philippine nematode sequences as well as their GenBank BLAST results from Table 6.6. Bootstrap values indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. Numbers in parentheses denote the number of species within a clade.

6.3.3. Phylogeny of the Angiostrongylidae

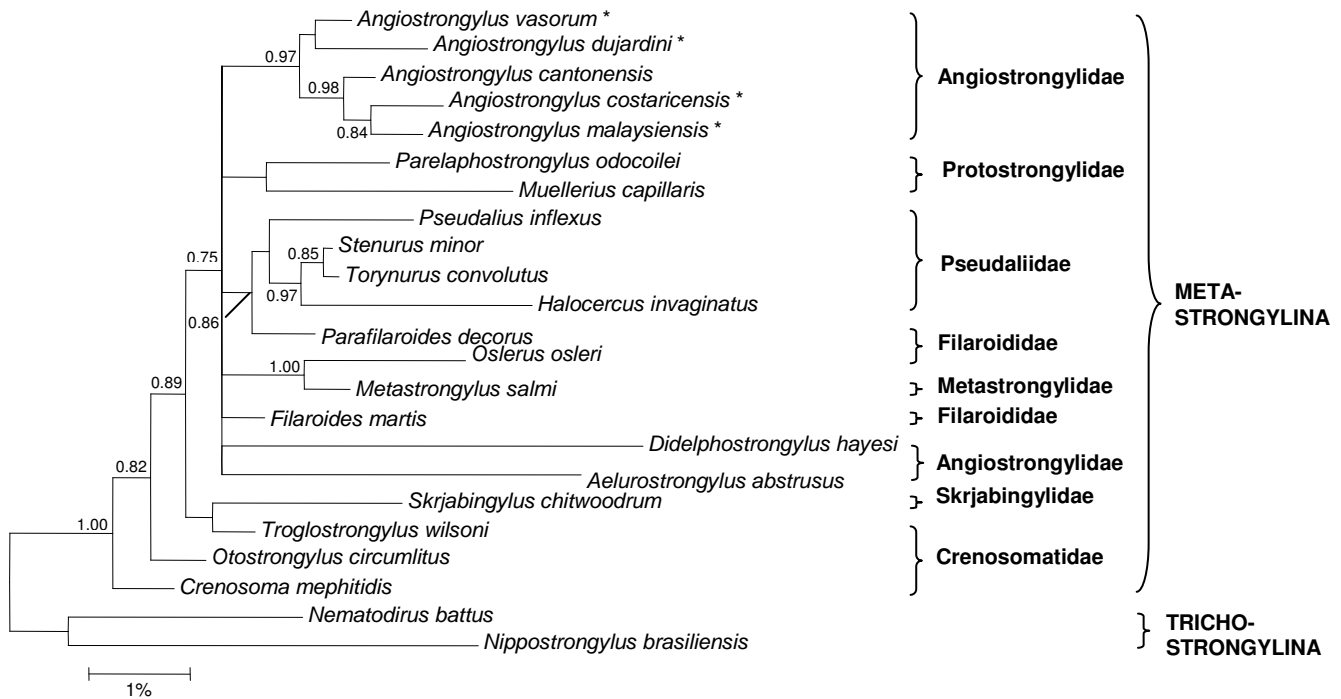
Phylogenetic analyses were undertaken to determine the relationships among the *Angiostrongylus* species sequenced to date (including the four angiostrongylid species sequenced for the first time in this study) and to determine the phylogenetic position of the Angiostrongylidae within the Sub-order Metastrongylina/ Superfamily Metastrongyloidea. (See alignment on Appendix 6.4, pp. 605-615.) Tests for base composition biases using the Chi square test in PAUP* showed no composition bias ($P=1.0$). GTR+ Γ was determined to be the best model (Appendix 6.5, p. 616) and was subsequently used for tree building in the model-based tree construction methods (ML, BI and NJ).

The maximum likelihood (ML), Bayesian inference (BI), neighbor-joining (NJ) and maximum parsimony (MP) phylogenies are shown in Figure 6.4. The trees were constructed using 1628 unambiguously aligned nucleotide sites and were rooted on the trichostrongylids *Nematodirus battus* (GenBank U01230, Zarlenga *et al.*, 1994) and *Nippostrongylus brasiliensis* (Genbank AF036597, Blaxter *et al.*, 1998).

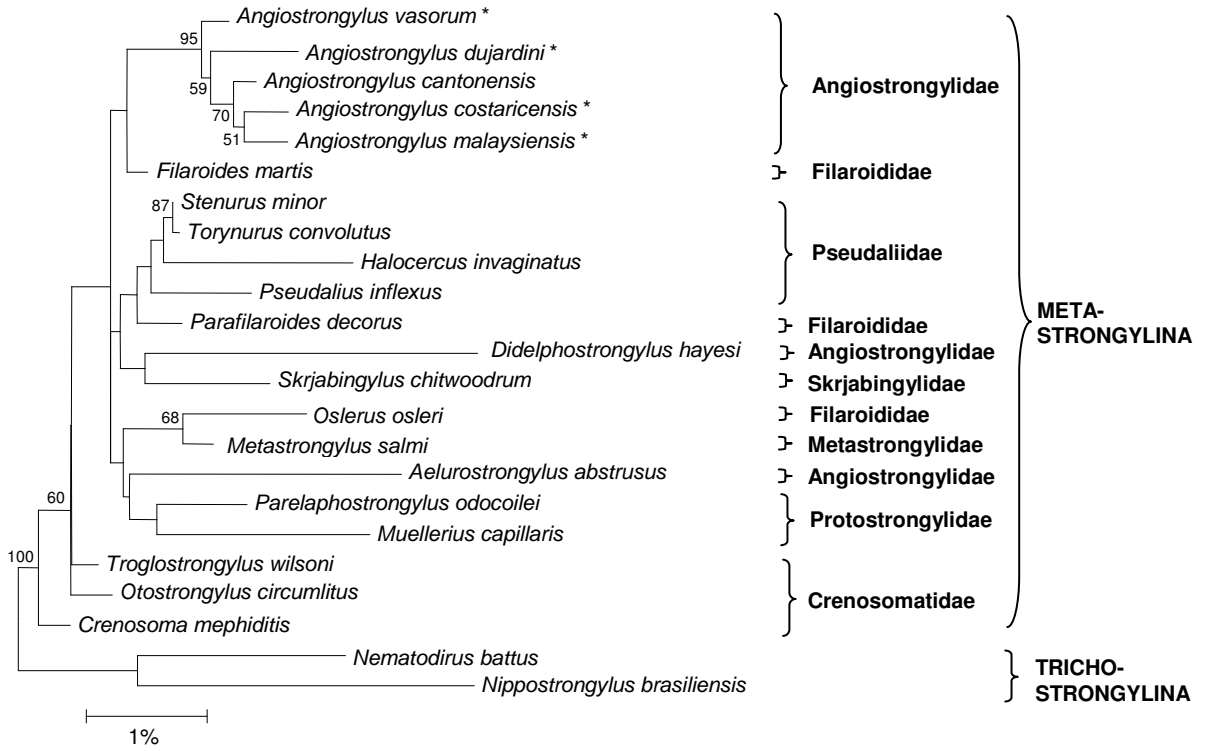
Figure 6.4: (See next two pages.) Phylogenetic trees of the angiostrongylids and worms in other families of the sub-order Metastrongylina based on the SSU rRNA gene using (A) maximum likelihood, (B) Bayesian analysis, (C) neighbor-joining and (D) maximum parsimony based on majority rule consensus of 3 equally parsimonious trees. The phylogenies are constructed from 1628 unambiguously aligned nucleotide sites and are rooted on the trichostrongylid worms *Nematodirus battus* (GenBank U01230) and *Nippostrongylus brasiliensis* (Genbank AF036597). Bootstrap values in (A) indicate the percentage support for individual branches based on 100 replicates. Bootstrap values lower than 50% are not shown. Bayesian posterior probabilities in (B) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (C) and (D) indicate the percentage support for individual branches based on 1000 replicates. The scale bars for (A), (B) and (C) represent one substitutional change per 100 nucleotide positions. *Angiostrongylus* species not sequenced in previous studies are highlighted with (*).



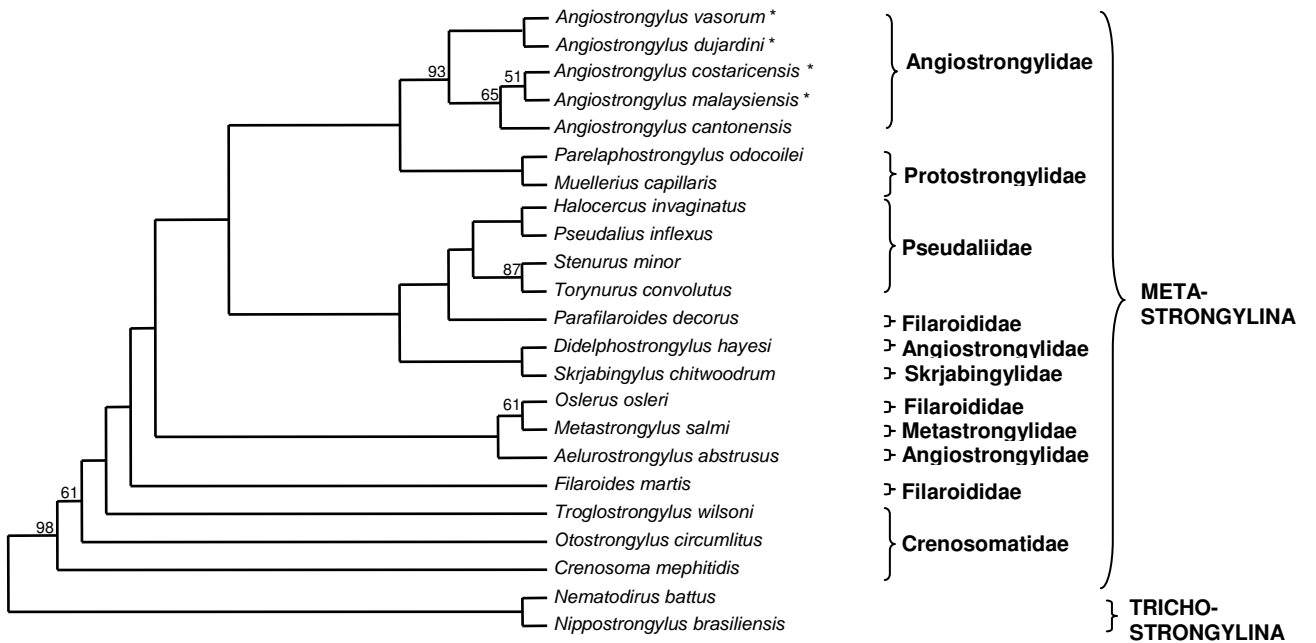
(A) - ML



(B) - BI



(C) - NJ



(D) - MP

The four phylogenies reveal several important points. First, monophyly of the Angiostrongylidae is neither conclusively supported nor rejected. For instance, in the ML tree (Fig. 6.4A) *Aelurostrongylus abstrusus* fell outside the main angiostrongylid group, while in Bayesian (Fig. 6.4B), NJ (Fig. 6.4C) and MP (Fig. 6.4D) analyses, both *Aelurostrongylus abstrusus* and *Didelphostrongylus hayesi* fell outside the main angiostrongylid group, though bootstrap support and posterior probabilities for these branches in the four trees were less than 50% and 0.7, respectively. Second, the phylogenies did not seem to support the distinction between *Angiostrongylus* and *Parastrongylus* as proposed by Ubelaker (1986) based on differences in morphology of the male bursa and the mammalian final host. The grouping of *A. vasorum*, [retained in the genus *Angiostrongylus* by Ubelaker (1986)] with *A. dujardini* [included in the genus *Parastrongylus* (Ubelaker, 1986)], was equivocal in these analyses ($P=0.67$ BI, 54% ML bootstraps, 30.2% NJ bootstraps and 43.9% MP bootstraps). Lastly, *Angiostrongylus cantonensis* was distinct from all other angiostrongylids and clustered with the *A. costaricensis* and *A. malaysiensis* group ($P=0.98$ BI, 68% ML bootstraps, 70% NJ bootstraps and 65% MP bootstraps).

6.4. Discussion

6.4.1. Suitability of the 5' end of the SSU rRNA gene as a molecular marker for *Angiostrongylus cantonensis*

A total of five *Angiostrongylus* species (including four from this study) have now been sequenced for almost the entire region (approximately 1670 nucleotides) of the SSU rRNA gene. All sequences are distinct and can therefore distinguish the *Angiostrongylus* species. However, this almost complete SSU rRNA gene region is too

long to be used as a molecular marker to rapidly and accurately identify specific taxa. When the first 480 bp of the 5' end of the gene was compared across the different *Angiostrongylus* taxa, 12 variable sites (more than a third of the total number of variations for the entire gene) were found. Furthermore, *A. cantonensis* exhibited a unique sequence for this region even when compared to its closest relatives *A. costaricensis*, *A. dujardini* and *A. malaysiensis* (based on morphology and host specificity). Furthermore, this segment of the SSU is available for the majority of the named nematodes in GenBank and encompasses the segment Bhadury *et al.* (2006) used to identify their marine nematodes. These factors make the first 480 bp region eminently suitable as a molecular marker to identify *A. cantonensis* in nematode surveys.

6.4.2. Molecular identification of nematodes from a trial population of *Achatina fulica* and *Laevicaulis alte* in the Philippines

Two distinct nematode sequences were detected by the SSU marker when applied to a trial population of intermediate hosts. Of these, Philippine Sequence 1 was identified as *A. cantonensis* on account of its 100% identity based on GenBank BLAST results of the variable region at the 5' end of the SSU rRNA gene. One hundred percent sequence identity could either mean 'the same taxon' or that 'there is insufficient variation to define taxa' (Floyd *et al.*, 2002). In the case of Philippine Sequence 1, its designation as *A. cantonensis* is valid as the current study already ascertained the uniqueness of the segment of the gene surveyed for this species in comparison to other closely related *Angiostrongylus* species (Figure 6.3).

This study clearly demonstrates the possibility of identifying infective 3rd juvenile stage *A. cantonensis* isolated from slug and snail intermediate hosts using the

5' end of the rRNA gene. This molecular approach to identification is a reliable alternative to morphological identification of nematode samples, especially in cases in which morphological characters are ambiguous in juvenile stages, thus rendering positive identification difficult.

6.4.3. Phylogenetic position of the Angiostrongylidae and the *Angiostrongylus* species within the Metastrongylina

Members of the Angiostrongylidae share certain features such as a posterior vulva but no buccal cavity or lips and the ability to infect a wide range of mammals as final hosts, including marsupials, insectivores, carnivores and rodents (Anderson, 2000; Roberts & Janovy, 2005). However, results from this study based on the SSU rRNA gene do not strongly support the monophyly of this group and show that only the *Angiostrongylus* species form a distinct group. Carreno and Nadler (2003) have already hinted that there is wide morphological diversity within the Angiostrongylidae. A further examination of the Angiostrongylidae that includes a broader taxonomic coverage of the group is required to determine its validity.

Ubelaker (1986) moved certain *Angiostrongylus* species, such as *A. cantonensis*, *A. costaricensis*, *A. dujardini*, *A. mackerrrasae*, *A. malaysiensis*, *A. petrowi*, *A. ryjikovi*, *A. sandarsae*, *A. schmidtii*, and *A. siamensis* to the genus *Parastrongylus*, which was first described by Baylis (1928) based on *Parastrongylus tateronae* as the type species. This is to distinguish *Parastrongylus* from *Angiostrongylus*, first described by Baillet (1866) based on *A. vasorum* as the type species and includes *A. chabaudi* and *A. railletii*. This distinction was based on differences of the morphology of the male bursa and the mammalian final host. However, this distinction is not widely accepted, and

molecular data from this study based on five *Angiostrongylus* species neither conclusively support nor reject this distinction.

6.5. Conclusion

An alternative approach to morphological identification of infective 3rd juvenile stage *Angiostrongylus cantonensis* was developed and successfully employed using the 5' end of the SSU rRNA gene (~480 bp). Sequences of other closely related *Angiostrongylus* species were obtained for this region to ascertain the uniqueness of the marker for *A. cantonensis*. When the SSU rRNA marker was used on nematode populations extracted from the giant African land snail, *Achatina fulica*, and the black slug, *Laevicaulis alte*, from the Philippines, the infective 3rd juvenile stage *A. cantonensis* was detected without difficulty. The technique developed in this study therefore enables the rapid and accurate identification of medically important nematode taxa when morphological analysis proves difficult or inadequate.

Phylogenetic analyses based on the full length SSU rRNA of the Metastrongylina with 5 *Angiostrongylus* species and 2 other angiostrongylids do not support the monophyly of the Angiostrongylidae. The same data neither support nor reject Ubelaker's distinction of *Parastrongylus* from *Angiostrongylus*.

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CHAPTER 7 - Survey of *Angiostrongylus cantonensis* (and other nematodes) from global populations of the giant African snail, *Achatina fulica* (and other snails)

7.1. Introduction

Angiostrongylus cantonensis, like one of its gastropod intermediate hosts *Achatina fulica*, has a global distribution (Marquardt *et al.*, 2000) and is a major concern from a human health perspective as it causes the food-borne disease eosinophilic meningoencephalitis (EME) or angiostrongyliasis (Kliks & Palumbo, 1992; Prociv *et al.*, 2000). The presence of *A. cantonensis* in rats in New Orleans, USA (Campbell & Little, 1988) and the recent outbreak of EME among medical students visiting Jamaica (Waugh *et al.*, 2005) illustrate the expanding distribution of the parasite. The life cycle of *A. cantonensis* involves a gastropod intermediate host and a murid rodent definitive host (Fig. 1.4, Chapter 1, p. 25); interestingly, the parasite is not highly specific to either its intermediate or its definitive host, which could help to explain its global distribution (Prociv *et al.*, 2000). The route for human infection by *A. cantonensis* is mostly through the gastropod intermediate host. The consumption of contaminated salad crops or raw/undercooked snails (Marquardt *et al.*, 2000) either as an alternative food source during famine (Kliks & Palumbo, 1992) or as delicacies (Chau *et al.*, 2003), their use as medicine (Kliks & Palumbo, 1992) and the predilection of humans to keep gastropods as pets (Wan & Weng, 2004) has led to sudden outbreaks or isolated cases of eosinophilic meningoencephalitis due to 3rd juvenile stage *A. cantonensis* infection (Kliks & Palumbo, 1992). Some of these outbreaks were attributed to the Giant African Land Snail, *Achatina fulica* (Kliks & Palumbo, 1992), which Alicata (1966) suggested as primarily responsible for the spread of the parasite to Southeast Asia and the Pacific.

7.1.1. Dispersal of *Angiostrongylus cantonensis* from East Africa: Alicata's hypothesis

First reported in rats in Canton, China in 1933 (Chen, 1935), *Angiostrongylus cantonensis* has a wide distribution and infects a range of intermediate and final hosts (Alicata, 1966, Fig. 7.1). The parasite was thought to have spread from East Africa towards the Indian subcontinent until it reached the Malay Peninsula (Alicata, 1966; Marquardt *et al.*, 2000). From there it migrated northwards to Indochina, southern China, Taiwan, and Okinawa, and eastwards to the Philippines, New Guinea, Australia, New Zealand, and several islands and island groups of the Pacific including Palau, Guam, Pohnpei, Pingelap, Moen, Ponape, Mariana, Wallis, Solomon, New Hebrides, New Guinea, Fiji, Loyalty, New Caledonia, Tahiti, Tonga, Rarotonga, Raiatea, Samoa, Tokelau and Hawaii (Kliks & Palumbo, 1992). *Angiostrongylus cantonensis* has also been reported in Western Africa (Ivory Coast, Liberia and Nigeria) (Kliks & Palumbo, 1992; Marquardt *et al.*, 2000), the Middle East (Egypt) (Marquardt *et al.*, 2000), North America (Campbell & Little, 1988; New *et al.*, 1995), and the Caribbean (Cuba, Puerto Rico, Jamaica and Haiti) (Pascual *et al.*, 1981; Andersen *et al.*, 1986; Lindo *et al.*, 2002; Raccurt *et al.*, 2003). Prociv *et al.* (2000) observed a general pattern in the recognition of *A. cantonensis* in new locations that begins with the initial detection in rats and occasionally in captive primates, followed by reported human cases of EME, and finally by the recovery of worms from some patients. Subsequent surveys of gastropod intermediate hosts in these new locations oftentimes lead to the detection of the parasite following confirmation by infecting the juveniles in laboratory rats, as in the study of Wallace & Rosen (1969).

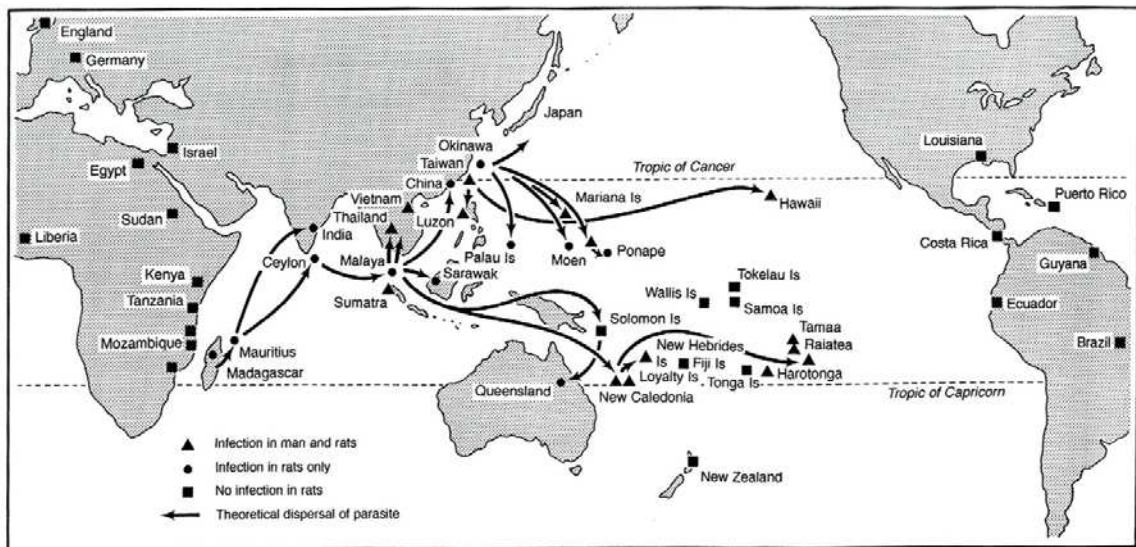


Figure 7.1: Probable dispersal route of *Angiostrongylus cantonensis*. (From Marquardt *et al.*, 2000.)

Alicata's (1966) hypothesis that the worm most probably originated from East Africa and migrated to East Asia and the Pacific is based on three observations. First, *A. cantonensis* is not found in all major Pacific islands despite the prevailing similar environmental conditions. Second, reported cases of eosinophilic meningoencephalitis in the Pacific are relatively recent, from just after the Second World War. Third, the parasite was only reported during the latter half of the 20th century in East Asia, the Pacific and Australia. This hypothesis was further supported by the total absence of the worm in 1967 in Central America (Costa Rica and Puerto Rico) and South America (Brazil, Ecuador and Guyana) based on a survey of 1810 rats (Alicata, 1967) only to appear later on in Cuba in 1973 (Pascual *et al.*, 1981), Puerto Rico in 1984 (Andersen *et al.*, 1986), Jamaica in 2000 (Waugh *et al.*, 2005) and Haiti in 2002 (Raccurt *et al.*, 2003; Marquardt *et al.*, 2000). The parasite was also reported for the first time in North America in New Orleans in 1986 (Campbell & Little, 1988).

It is not yet clear how *A. cantonensis* spread from the Pacific Basin to the Caribbean and beyond, although Kliks and Palumbo (1992) pointed to rats as culprits.

They suspected that infected rats from Indochina were accidentally brought to Cuba between 1966 and 1975 during the Vietnam War, which led to cases of EME beginning in 1973. From there it spread to Puerto Rico where the worms were detected in adult rats in 1981. How the worms got to West Africa, particularly the Ivory Coast and Nigeria is less clear. The first and only case of eosinophilic meningoencephalitis in the Ivory Coast was reported in 1979 (Nozais *et al.*, 1980). In Nigeria, adult worms were also recovered from rats in Port Harcourt but not in Ibadan (Kliks & Palumbo, 1992). Nematodes were also found in Giant African Land Snails in Port Harcourt, reported by Kliks and Palumbo as *Achatina fulica*, though they are more likely to be *Achatina achatina* based on the prevalence of this snail in that area and the fact that *A. fulica* has not yet been officially recorded there.

7.1.2. Perceived role of *Achatina fulica* in the spread of *A. cantonensis*

Alicata (1966) pinned his East Africa origin for *A. cantonensis* on the nearly parallel distribution of the parasite with one of its intermediate hosts, *Achatina fulica*, particularly in Southeast Asia and the Pacific. He based his hypothesis on the observations he made when he surveyed 8 areas from East Africa to the Western Pacific, in which he checked for the presence of *A. cantonensis* in adult rats. He found the parasite in Madagascar, Mauritius, Sri Lanka and Borneo but not in Mozambique, Kenya, Tanzania and India. By combining his findings with what was known at that time regarding the dispersal route of *A. fulica* and the reported cases of rat and human infections of *A. cantonensis*, he concluded that the rapid spread of the snail led to the spread of the parasite based on the following reasons. First, he observed *A. cantonensis* along the dispersal route of the snail. Second, he found the parasite in rats in places where *A. fulica* was introduced. Last, he also noted the close time relationship between

the introduction of the snail and the first reported case of EME in several Pacific islands. In the case of West Africa, however, the presence of *A. cantonensis* there can not be pinned on *A. fulica*. Although Kliks and Palumbo (1992) reported the presence of the parasite in *A. fulica*, this was probably a result of a misidentification of another giant African snail, probably *Achatina achatina*.

7.1.3. Dispersal of *Angiostrongylus cantonensis* from South and Southeast Asia: Drozd et al.'s alternative hypothesis

An alternative hypothesis for the dispersal of *A. cantonensis* was proposed by Drozd *et al.* (1975) in which they postulated that the parasite originated in South and Southeast Asia rather than East Africa and that the murid rodent definitive hosts and not *Achatina fulica* were responsible for the current distribution of *A. cantonensis*. First, they argued that the occurrence within close geographical regions (Indo-Malayan region and Australia) of closely related *Angiostrongylus* species and their respective *Rattus* final hosts would indicate a very long host-parasite association that probably encompassed the radiation of murid rodents in Asia. Such is probably the case for *A. cantonensis* (China, Southeast Asia) in *R. rattus* and *R. norvegicus*, *A. malaysiensis* (Malaysia) in *R. jalorensis*, and *A. mackerrasae* (Australia) in *R. fuscipes*. Second, they believed that the adaptation of rat hosts such as *Rattus rattus* and *R. norvegicus*, which originated in South and Southeast Asia, to survive high dosage infections (150 parasites per individual) would have taken a long time to evolve, which could not have happened had the parasite originated in East Africa and spread to Asia in a short period of time as postulated by Alicata (1966). They therefore concluded that *Achatina fulica* is a recent introduction in the parasite's life cycle. As further support of this hypothesis, *A. cantonensis* has so far only been found in Madagascar in *Rattus rattus*, a recent human

introduction to the island (Drodz *et al.*, 1975; Prociv *et al.*, 2000); no other endemic murid rodent is found in Madagascar (Drodz *et al.*, 1975). Prociv *et al.* (2000) believed that the role of *A. fulica* in the dispersal of *A. cantonensis* was over-emphasised as the parasite was more likely to have been introduced into new areas by rats, owing to their cosmopolitan distribution, and that local snails and slugs had just as easily assumed the role of intermediate hosts. Furthermore, Civeyrel & Simberloff (1996) pointed the possibility of other introduced snails, such as *Euglandina*, *Edentulina* and *Gonaxis*, in spreading the parasite.

7.1.4. Other nematodes in *Achatina fulica*

In addition to *Angiostrongylus cantonensis*, other nematodes may also be present in *Achatina fulica*. Indeed, land snails and slugs in general have often been used by nematodes either as intermediate hosts or as definitive hosts (Grewal *et al.*, 2003). There are 61 known nematode species that utilise molluscs as intermediate hosts, with 49 belonging to the superfamily Metastrongyloidea; 47 known nematode species use molluscs as definitive hosts, 33 of which belong to the order Rhabditida (Grewal *et al.*, 2003). Both groups are included in the Rhabditida clade by Meldal *et al.* (2006) based on phylogenetic analyses using the small subunit (SSU) ribosomal (r) RNA gene. In the case of *Achatina fulica*, previous studies have already demonstrated the presence of nematodes. For instance, the metastrongyloid lungworm *Oslerus ostratus* (= *Anafilaroides rostratus*), which parasitises cats as the final host and is closely related to the dog lungworm *Oslerus osleri*, can also infect *Achatina fulica* as an intermediate host (Grewal *et al.*, 2003). Another nematode, *Rhabditis* sp., was found in *A. fulica* populations in Thailand in which the parasite utilised the snail as a definitive host but without causing any noticeable damage (Viyada, 2005). Juvenile nematodes were also

noted in the mucus of *A. fulica* in Brazil though their identities were not reported (Fonseca & Nascimento, 2004).

7.1.5. Objectives of the study

Alicata's (1966) hypothesis states that the Giant African Land Snail, *Achatina fulica*, caused the spread of *Angiostrongylus cantonensis*, particularly in Southeast Asia and the Pacific, whereas Drozd *et al.*'s (1975) hypothesis points to murid rodents. The objectives of this study were to (1) survey global populations of *Achatina fulica* for the presence of *Angiostrongylus cantonensis* using the technique developed in Chapter 6 that employed the 5' end of the small subunit rRNA gene for rapid identification of the parasite, and (2) investigate the role of *A. fulica* in the dispersal of *A. cantonensis* (as proposed by Alicata, 1966) by comparing the movement of the snail through population genetic analysis with the movement of the parasite also through population genetic analysis using a suitable molecular marker. Both objectives, however, were dependent on the availability of sufficient population samples of both the parasite and the snail. While the focus was on *A. cantonensis* infection in *A. fulica*, where samples of other snails and slugs were obtained within the distribution range of the parasite (in this case two other species, *Laevicaulis alte* and *Achatina* sp.), they were also surveyed for *A. cantonensis* infection. This is the first systematic attempt to survey *A. cantonensis* from global populations of a known intermediate host using molecular identification. If Alicata's hypothesis is correct, then the nematode parasite should be present within the dispersal route of the snail intermediate host, and the dispersal patterns of the snail and the parasite based on population genetic analysis should also be similar.

7.2. Materials and methods

7.2.1. Samples and localities

Ten global populations of *Achatina fulica*, two West African populations of *Achatina* sp. and two Philippine populations of *Laevicaulis alte* were sampled for *Angiostrongylus cantonensis*. The *Achatina fulica* and *Laevicaulis alte* sample populations from UP Diliman used in Chapter 6 were also included in this survey. For the Ivory Coast *Achatina* sp. samples, the exact location of the sampling site was not identified as the snails were confiscated by customs officials at Waterloo Station in London from a man trying to smuggle the snails in October 2005. Details of the samples, their sampling localities and collectors are provided in Table 7.1.

Table 7.1: Snail samples, collectors and localities

Locality	Species	Collector
Bharatpur, Nepal	<i>Achatina fulica</i>	P. Budha
Yangon, Myanmar	<i>Achatina fulica</i>	F. Naggs
Trok Nong Area, near Phlieu Waterfall National Park, Chantaburi, Eastern Thailand near Cambodia	<i>Achatina fulica</i>	S. Panha & C. Sutcharit
Hahajima, Ogasawara, Japan	<i>Achatina fulica</i>	A. Davison
UP Diliman Campus, Quezon City, Philippines	<i>Achatina fulica</i>	I. Fontanilla
Batasan Hills, Quezon City, Philippines	<i>Achatina fulica</i>	E. Fontanilla & I. Fontanilla
Subic Bay Forest Reserve, Zambales, Philippines	<i>Achatina fulica</i>	M.R. Posa
Singapore	<i>Achatina fulica</i>	M.R. Posa
Haapiti Valley, Moorea, French Polynesia	<i>Achatina fulica</i>	T. Coote
Moaroa Valley, Tahiti, French Polynesia	<i>Achatina fulica</i>	T. Coote
Ivory Coast	<i>Achatina</i> sp.	Intercepted by customs officials at Waterloo Station in London from a man from the Ivory Coast (October 2005)
Ibadan, Nigeria	<i>Achatina</i> sp.	F. Opeyemi
UP Diliman Campus, Quezon City, Philippines	<i>Laevicaulis alte</i>	I. Fontanilla
Batasan Hills, Quezon City, Philippines	<i>Laevicaulis alte</i>	E. Fontanilla & I. Fontanilla

7.2.2. DNA extraction, PCR amplification and sequencing

The protocols in this section follow those developed in Chapter 6. The snails were cut into small pieces and digested overnight in Ash's digestive fluid, after which the worms were individually collected using yellow-tipped pipettes and stored at -20°C in TE buffer until use (see Section 6.2.2 of Chapter 6, pp. 305-307). DNA extraction, PCR amplification, gel extraction and purification were carried out for each nematode using the protocols in Chapter 2, Section 2.1.4, pp. 41-42 and Sections 2.2 - 2.4, pp. 42-59, except that only Primer Set A (SSU_F07 and SSU_R09) for the 5' end of the SSU rRNA was used to amplify an approximately 480 bp PCR product. The purified PCR products were subsequently sequenced using the SSU_R09 primer and the peaks visualised using an Applied Biosystems 3730 DNA sequencer and BigDye version 3.1 termination cycle sequencing chemistry (Section 2.6 of Chapter 2, pp. 59-61). PCR amplification and sequencing of a second molecular marker for the population genetic analyses of the snail and parasite samples (second objective) were no longer carried out due to the limited and patchy sampling of the parasite and the difficulty of obtaining live snail intermediate host (see results and discussion).

7.2.3. Sequence analyses

Individual nematode sequences were processed using the STADEN package (Staden *et al.*, 2000) and the presence of *Angiostrongylus cantonensis* was determined by BLAST search in GenBank based on the 5' end of the SSU rRNA gene. Where possible, the non-angiostrongylid nematodes were also identified based on the closest sequence match to a named nematode species. If a 100% BLAST sequence identity to a named nematode species was obtained, it could mean the same taxon or that there is insufficient variation to distinguish taxa (Floyd *et al.*, 2002). In the case of

Angiostrongylus, results in Chapter 6 have already shown that the five closely related species of *Angiostrongylus*, which include *A. cantonensis*, can be distinguished from each other based on the 5' end of the SSU rRNA gene. To complement the BLAST results, a neighbor-joining phylogeny was then constructed to visualise the relationship of the nematode sequences against all known nematode taxa in Genbank as well as the *Angiostrongylus* species obtained in Chapter 6. The phylogeny was based on 376 unambiguously aligned nucleotide sites and rooted on 4 non-nematode taxa: *Chordodes morgani* (AF036639, Blaxter *et al.*, 1998) and *Gordius aquaticus* (X87985, Winnepenninckx *et al.*, 1995) of Phylum Nematomorpha, *Priapulid caudatus* (Z38009, Aleshin *et al.*, 1998) of Phylum Priapulida, and *Brachionus plicatis* (U49911, Aguinaldo *et al.*, 1997) of Phylum Rotifera (for alignment, see Appendix 6.2, pp. 537-603). The sequences were checked for base composition bias using the Chi square test in PAUP* (version 4.0b10) (Swofford, 2002), after which the optimal model for DNA sequence evolution was determined by comparing, through the Likelihood Ratio Test (LRT), the likelihood scores of the following: JC69 (Jukes & Cantor, 1969), F81 (Felsenstein, 1981), K2P (Kimura, 1980), HKY85 (Hasegawa *et al.*, 1985), TN93 (Tamura & Nei, 1993) and the GTR (Rodriguez *et al.*, 1990) models as well as their variants that incorporated gamma distributed rates (Γ) (Yang, 1993). Bootstrap resampling (Felsenstein, 1985) with 1000 replicates was then carried out (see Section 2.9.9 of Chapter 2, pp. 85-86). Once the clades to which the nematode sequences belong were identified, a second NJ tree based on the same gene fragment (376 sites) with optimised parameters was constructed but this time excluding the taxa that do not belong to these clades.

7.3. Results

A total of 191 *Achatina fulica* from 10 global populations, 25 *Achatina* sp. from 2 West African populations and 39 *Laevicaulis alte* from 2 Philippine populations were surveyed for *A. cantonensis* infections. Of the 10 global populations of *Achatina fulica*, 6 yielded nematodes; these were Thailand (20 snails sampled), Ogasawara (12 snails), UP Diliman-Philippines (15 snails), Batasan Hills-Philippines (50 snails), Singapore (33 snails) and Tahiti (5 snails). Population samples from Ogasawara and Tahiti had a 100% infection rate. Population samples from Myanmar (20 snails), Nepal (22 snails), Zambales-Philippines (4 snails) and Moorea (10 snails) had a 0% infection rate. The two *Achatina* sp. populations from West Africa, namely the Ivory Coast (22 snails) and Nigeria (3 snails), as well as the two Philippine populations of *Laevicaulis alte*, UP Diliman (19 slugs) and Batasan Hills (20 slugs), also manifested nematode infection, with the population sample from Nigeria exhibiting a 100% infection rate. See also Table 7.2.

Table 7.2: Population sizes and percentage of nematode infection

Locality	Species	Sample Size	% Infected
Bharatpur, Nepal	<i>Achatina fulica</i>	22	0
Yangon, Myanmar	<i>Achatina fulica</i>	20	0
Chantaburi, Eastern Thailand	<i>Achatina fulica</i>	20	5.0%
Hahajima, Ogasawara, Japan	<i>Achatina fulica</i>	12	100.0%
UP Diliman Campus, Philippines	<i>Achatina fulica</i>	15	6.7%
Batasan Hills, Philippines	<i>Achatina fulica</i>	50	14.0%
Subic Bay Forest Reserve, Philippines	<i>Achatina fulica</i>	4	0
Singapore	<i>Achatina fulica</i>	33	15.2%
Haapiti Valley, Moorea	<i>Achatina fulica</i>	10	0
Moaroa Valley, Tahiti	<i>Achatina fulica</i>	5	100.0%
Ivory Coast	<i>Achatina</i> sp.	22	18.2%
Ibadan, Nigeria	<i>Achatina</i> sp.	3	100.0%
UP Diliman Campus, Philippines	<i>Laevicaulis alte</i>	19	21.1%
Batasan Hills, Philippines	<i>Laevicaulis alte</i>	20	30.0%

A total of 25 distinct nematode sequences were identified from the snail and slug populations. For the *Achatina fulica* populations, a single unique nematode sequence was found in the Thailand population; two nematode sequences were each found in the UP Diliman-Philippines, Ogasawara and Tahiti populations; three were found in the Batasan-Philippines population, and four in the Singapore population. For the West African *Achatina* sp. populations, three unique nematode sequences were found in Nigeria and seven in the Ivory Coast. For the Philippine *Laevicaulis alte* populations, the same two nematode sequences found in *A. fulica* were also found in the UP Diliman *L. alte* population, whereas two unique sequences were found in the Batasan Hills population in addition to a nematode sequence that it shares with the *A. fulica* population from that site. One of the Tahiti sequences (Tahiti Sequence 1) is identical to one of the Philippines sequences (Philippines Sequence 1), which was present in the UP Diliman population of *Achatina fulica* and in both UP Diliman and Batasan Hills populations of *Laevicaulis alte*. A summary of the frequency and distribution of these nematode sequences is given in Table 7.3.

Table 7.3: Distribution and frequencies of the nematode sequences across global populations of *Achatina fulica*, West African populations of *Achatina* sp. and Philippine populations of *Laevicaulis alte*. Note that Philippines Sequence 1 and Tahiti Sequence 1 are 100% identical.

Locality	Host	Nematode Sequences	Number (Frequency relative to the total nematodes found in the whole snail population)
Nepal	<i>Achatina fulica</i>	None	None
Myanmar	<i>Achatina fulica</i>	None	None
Thailand	<i>Achatina fulica</i>	Thailand Sequence 1	1 (100.0%)
Ogasawara, Japan	<i>Achatina fulica</i>	Ogasawara Sequence 1	2 (2.3%)
		Ogasawara Sequence 2	86 (97.7%)
UP Diliman, Philippines	<i>Achatina fulica</i>	Philippines Sequence 1 (=Tahiti Sequence 1)	1 (16.7%)
		Philippines Sequence 2	5 (83.3%)
Batasan, Philippines	<i>Achatina fulica</i>	Philippines Sequence 5	6 (33.3%)
		Philippines Sequence 6	1 (5.6%)
		Philippines Sequence 7	11 (61.1%)
Subic Forest Reserve, Philippines	<i>Achatina fulica</i>	None	None
Singapore	<i>Achatina fulica</i>	Singapore Sequence 1	6 (9.7%)
		Singapore Sequence 2	1 (1.6%)
		Singapore Sequence 3	1 (1.6%)
		Singapore Sequence 4	54 (87.1%)
Tahiti	<i>Achatina fulica</i>	Tahiti Sequence 1 (=Philippines Sequence 1)	91 (98.9%)
		Tahiti Sequence 2	1 (1.1%)
Moorea	<i>Achatina fulica</i>	None	None
Ivory Coast	<i>Achatina</i> sp.	Ivory Coast Sequence 1	2 (8.7%)
		Ivory Coast Sequence 2	11 (47.8%)
		Ivory Coast Sequence 3	1 (4.3%)
		Ivory Coast Sequence 4	1 (4.3%)
		Ivory Coast Sequence 5	5 (21.7%)
		Ivory Coast Sequence 6	1 (4.3%)
		Ivory Coast Sequence 7	2 (8.7%)
Nigeria	<i>Achatina</i> sp.	Nigeria Sequence 1	1 (1.8%)
		Nigeria Sequence 2	1 (1.8%)
		Nigeria Sequence 3	54 (96.4%)
UP Diliman, Philippines	<i>Laevicaulis alte</i>	Philippines Sequence 1 (=Tahiti Sequence 1)	20 (95.2%)
		Philippines Sequence 2	1 (4.8%)
Batasan, Philippines	<i>Laevicaulis alte</i>	Philippines Sequence 1 (=Tahiti Sequence 1)	8 (80%)
		Philippines Sequence 3	1 (10%)
		Philippines Sequence 4	1 (10%)

Nine snails from six localities (UP Diliman-Philippines, Batasan Hills-Philippines, Ogasawara, Singapore, Tahiti and Ivory Coast) were infected by more than one type of worm based on the SSU rRNA sequences. Table 7.4 summarises the multiple infections of these snails.

Table 7.4: Snails infected with more than one type of worm based on the sequence of the 5' end of the SSU rRNA gene.

Locality	Host #	Nematode Sequence	Number (Frequency*)
UP Diliman, Philippines	<i>Achatina fulica</i> D51	Philippines Sequence 1	1 (16.7%)
		Philippines Sequence 2	5 (83.3%)
Batasan Hills, Philippines	<i>Achatina fulica</i> B42	Philippines Sequence 5	1 (33.3%)
		Philippines Sequence 7	2 (66.67%)
	<i>Achatina fulica</i> B43	Philippines Sequence 5	1 (50%)
		Philippines Sequence 7	1 (50%)
Ogasawara, Japan	<i>Achatina fulica</i> OJ2	Ogasawara Sequence 1	1 (8.3%)
		Ogasawara Sequence 2	11 (91.7%)
	<i>Achatina fulica</i> OJ3	Ogasawara Sequence 1	1 (14.3%)
		Ogasawara Sequence 2	6 (85.7%)
Singapore	<i>Achatina fulica</i> Si63	Singapore Sequence 1	4 (80.0%)
		Singapore Sequence 2	1 (8.3%)
Tahiti	<i>Achatina fulica</i> T1	Tahiti Sequence 1	11 (91.7%)
		Tahiti Sequence 2	1 (1.1%)
Ivory Coast	<i>Achatina</i> sp. IVC19	Ivory Coast Sequence 2	11 (84.6%)
		Ivory Coast Sequence 3	1 (7.7%)
		Ivory Coast Sequence 4	1 (7.7%)
	<i>Achatina</i> sp. IVC22	Ivory Coast Sequence 6	2 (66.7%)
		Ivory Coast Sequence 7	1 (33.3%)

*Frequency is relative to the total nematodes found per snail.

The 25 nematode sequences were then subjected to a BLAST search in GenBank (GB Release No. 158, 15 February 2007) to identify an exact match or closest identity to named nematode species. Two of these sequences, namely Tahiti Sequence 1/ Philippines Sequence 1 and Tahiti Sequence 2, are angiostrongylids based on the BLAST results. However, only Tahiti Sequence 1/Philippines Sequence 1 has a 100% match with *Angiostrongylus cantonensis* (AY295804, Carreno & Nadler, 2003). Among the non-angiostrongylids, Philippines Sequence 5 is the only other nematode sequence to have a 100% BLAST identity to a named species in GenBank. This

sequence is identical to *Ancylostoma caninum* (AJ920347, Chilton *et al.*, 2006). As the other sequences did not match any named taxon in GenBank based on BLAST results, only the closest sequence identity to a named nematode species could be assigned; their exact species identification based on the BLAST results could therefore not be known as of GB Release No. 158. For instance, the second highest sequence similarity attained was 99%; this was the case for Philippines Sequence 2 (for *Oslerus osleri*), Singapore Sequence 4 (for *Oslerus osleri*), Tahiti Sequence 2 (for *Angiostromgylus cantonensis*), Ivory Coast Sequence 1 (for *Metastrongylus elongatus* and *M. salmi*) and Nigeria Sequence 2 (for *Ancylostoma caninum*). On the other hand, Singapore Sequence 1 and Philippines Sequence 3 have the lowest sequence identity (87% BLAST identity for both with *Pellioiditis marina*, a marine nematode, which, being terrestrial nematodes, Singapore Sequence 1 and Philippines Sequence 3 clearly are not). A summary of the BLAST results is shown in Table 7.5.

Table 7.5: GenBank BLAST results for the nematode sequences

Sequence	BLAST Result
Thailand Sequence 1	96% identity with <i>Heterakis</i> sp. 14690 (AF083003) [Ascaridida]
Ogasawara Sequence 1	93% identity with <i>Panagrolaimus subelongatus</i> (AY284681) [Panagrolaimidae]
Ogasawara Sequence 2	93% identity with <i>Phasmarhabditis hermaphrodita</i> (DQ639981) [Rhabditida]
Philippines Sequence 1 (=Tahiti Sequence 1)	100% identity with <i>Angiostrongylus cantonensis</i> (AY295804) [Strongyloidea]
Philippines Sequence 2	99% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea] (but different from Singapore Sequence 4)
Philippines Sequence 3	87% identity with <i>Pellioditis marina</i> (AF038021) [Rhabditidae] (but different from Singapore Sequence 1)
Philippines Sequence 4	97% identity with <i>Panagrolaimus</i> c.f. <i>rigidus</i> (DQ285636) [Panagrolaimidae]
Philippines Sequence 5	100% identity with <i>Ancylostoma caninum</i> (AJ920347) [Strongyloidea]
Philippines Sequence 6	93% identity with <i>Pellioditis marina</i> (AF038021) [Rhabditidae]
Philippines Sequence 7	97% identity with <i>Caenorhabditis vulgaris</i> (U13931) [Rhabditidae]
Singapore Sequence 1	87% identity with <i>Pellioditis marina</i> (AF038021) [Rhabditidae]
Singapore Sequence 2	95% identity with <i>Heterakis</i> sp. 14690 (AF083003) [Ascaridida]
Singapore Sequence 3	98% identity with <i>Caenorhabditis briggsae</i> (U13929) [Rhabditidae]
Singapore Sequence 4	99% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea] (but different from Singapore Sequence 2)
Tahiti Sequence 1 (=Philippines Sequence 1)	100% identity with <i>Angiostrongylus cantonensis</i> (AY295804) [Strongyloidea]
Tahiti Sequence 2	97% identity with <i>Angiostrongylus cantonensis</i> (AY295804) [Strongyloidea]
Ivory Coast Sequence 1	99% identity with <i>Metastrongylus elongatus</i> (AJ920363) [Strongyloidea] 99% identity with <i>Metastrongylus salmi</i> (AY295801) [Strongyloidea]
Ivory Coast Sequence 2	98% identity with <i>Metastrongylus elongatus</i> (AJ920363) [Strongyloidea] 98% identity with <i>Metastrongylus salmi</i> (AY295801) [Strongyloidea]
Ivory Coast Sequence 3	98% identity with <i>Protostrongylus rufescens</i> (AJ920364) [Strongyloidea]
Ivory Coast Sequence 4	98% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea] (but different from Ivory Coast sequence 6)
Ivory Coast Sequence 5	97% identity with <i>Heterakis gallinarum</i> (DQ503462) [Ascaridida]
Ivory Coast Sequence 6	98% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea] (but different from Ivory Coast sequence 4)
Ivory Coast Sequence 7	97% identity with <i>Oslerus osleri</i> (AY295812) [Strongyloidea]
Nigeria Sequence 1	97% identity with <i>Protostrongylus rufescens</i> (AJ920364) [Strongyloidea]
Nigeria Sequence 2	99% identity with <i>Ancylostoma caninum</i> (AJ920347) [Strongyloidea]
Nigeria Sequence 3	97% identity with <i>Metastrongylus elongatus</i> (AJ920363) [Strongyloidea] 97% identity with <i>Metastrongylus salmi</i> (AY295801) [Strongyloidea]

To complement the BLAST results, a neighbor-joining tree of the Nematoda based on 376 unambiguously aligned nucleotide sites of the 5' end of the SSU rRNA gene (Fig. 7.2A) was then generated to visualise the relationships of the 25 nematode sequences with all 361 nematode taxa having at least the first 480 nucleotides of the 5' end of the SSU rRNA in GenBank (GB Release No. 158, see Appendix 6.2) as well as the new sequences from the four *Angiostrongylus* species in Chapter 6. The GTR+ Γ model was used to construct the tree after it was determined to be the best model

(Appendix 7.1A, p. 617). According to this phylogeny, all 25 nematode sequences fall within the Rhabditida clade (Meldal *et al.*, 2006). A smaller NJ phylogeny of the same gene fragment (376 nucleotide sites) was constructed (Fig. 7.2B) for the Rhabditida clade using all 241 rhabditid SSU sequences available in GenBank (GB Release No. 158) as well as the other *Angiostrongylus* sequences and rooted on the plectid nematode *Plectus acuminatus* (AF037628, Blaxter *et al.*, 1998). The GTR+ Γ model was determined to be the best model and was subsequently used for this analysis (Appendix 7.1B, p. 617). This NJ phylogeny shows that 22 out of the 25 nematode sequences, including the two angiostrongylid sequences (Philippines Sequence 1/ Tahiti Sequence 1 and Tahiti Sequence 2), are distributed across three major clades in the tree: the Strongyloidea, the Rhabditidae and the Panagrolaimidae. The three remaining nematode sequences (Ivory Sequence 5, Singapore Sequence 2 and Thailand Sequence 1) group with the two *Heterakis* species found outside the three aforementioned clades. The groupings of the nematode sequences to the different clades correspond to the taxonomic grouping of the closest match(es) of each of the nematode sequences according to BLAST (see also Table 7.5).

The Strongyloidea clade includes the 2 angiostrongylid sequences: Tahiti Sequence 1/ Philippines Sequence 1 and Tahiti Sequence 2. Tahiti Sequence 1/ Philippines Sequence 1 has 100% BLAST identity to the sequence of *Angiostrongylus cantonensis* on Genbank (AY295804, Carreno & Nadler, 2003) and clusters with *A. cantonensis* in the tree, while the sequences from the other *Angiostrongylus* species sequenced in Chapter 6 fall outside this cluster. Tahiti Sequence 2 also clusters with *A. cantonensis* in the tree but has a BLAST identity of 97% with *A. cantonensis*.

There are 12 non-angiostrongylid sequences that are also included in the Strongyloidea; these are the 3 Nigeria sequences, 6 out of 7 Ivory Coast sequences, 2

Philippines sequences and Singapore Sequence 4. Nigeria Sequence 1 clusters with the deer muscieworm *Parelaphostrongylus odocoilei*, the hair lungworm *Muellerius capillaris*, and the sheep lungworm *Dictyocaulus filaria*, though it has a 97% BLAST identity with the goat and sheep lungworm *Protostrongylus rufescens* (AJ920364, Chilton *et al.*, 2006) (see Table 7.5). Nigeria Sequence 2 and Philippines Sequence 5 cluster with the dog hookworms *Ancylostoma caninum* and *Necator americanus*; both sequences are most similar to *Ancylostoma caninum* (AJ920347, Chilton *et al.*, 2006) with 99% and 100% BLAST identities, respectively. Nigeria Sequence 3 clusters with Ivory Coast Sequence 7, though they have different BLAST identities; Nigeria Sequence 3 has a 97% BLAST identity with both pig lungworms *Metastrongylus elongatus* (AJ920363, Chilton *et al.*, 2006) and *M. salmi* (AY295609, Carreno & Nadler, 2003) while Ivory Coast Sequence 7 has a 97% BLAST identity with the filaroid dog lungworm *Oslerus osleri* (AY295812, Carreno & Nadler, 2003). Ivory Coast Sequence 1 groups with *Metastrongylus elongatus* (AJ920363) and *M. salmi* (AY295609) with 99% BLAST identity for both taxa. Ivory Coast Sequence 2 clusters with the cat lungworm *Aelurostrongylus abstrusus* (Chilton *et al.*, 2006), though it has a 98% BLAST identity with both *Metastrongylus elongatus* (AJ920363) and *M. salmi* (AY295609) (see Table 7.5). Ivory Coast Sequence 3 clusters with *Protostrongylus rufescens* (AJ920364) with 98% BLAST identity. Ivory Coast Sequences 4 and 6 cluster together and both have a 98% BLAST identity with *Oslerus osleri* (AY295812) though the two sequence types are not identical with each other (see Table 7.5). Both Philippines Sequence 2 and Singapore Sequence 4 cluster with *Oslerus osleri* (AY295812) with 99% BLAST identity though they are not identical with each other (see Table 7.5).

The Rhabditidae clade includes six nematode sequences: Singapore Sequences 1 and 3, Philippines Sequences 3, 6 and 7 and Ogasawara Sequence 2. Singapore Sequence 1 and Philippines Sequence 3 cluster together, and in turn group with the free-living marine nematode *Pellioditis mediterranea*, though both have an 87% BLAST identity with *Pellioditis marina* (AF038021, Fitch, 2000). Philippines Sequence 6 forms a sister group taxon with a group of rhabditids that include the *Pellioditis* species and has a 93% BLAST identity with *Pellioditis marina* (AF038021). Singapore Sequence 3 clusters with *Caenorhabditis* spp., free-living soil nematodes, along with Philippines Sequence 7; Singapore Sequence 3 is 98% identical with *Caenorhabditis briggsae* (U13929, Fitch *et al.*, 1995) while Philippines Sequence 7 is 97% identical with *Caenorhabditis vulgaris* (U13931) based on BLAST results. Ogasawara Sequence 2 groups with the two *Phasmarhabditis* taxa and has a 93% BLAST identity with *Phasmarhabditis hermaphrodita*.

Two nematode sequences, Ogasawara Sequence 1 and Philippines Sequence 4, are included in the Panagrolaimidae clade; the former has a 93% BLAST identity with the soil nematode *Panagrolaimus subelongatus* (AY284681, Holterman *et al.*, 2006) while the latter has a 97% identity with *Panagrolaimus* c.f. *rigidus* (DQ285636, Houthoofd *et al.*, 2006).

The three remaining nematode sequences, Singapore Sequence 2, Thailand Sequence 1 and Ivory Coast Sequence 5, all cluster with the bird parasites in the genus *Heterakis*; Singapore Sequence 2 and Thailand Sequence 1 are 95% and 96% identical, respectively, with *Heterakis* sp. isolate 14690 (AF083003, Fitch, 2000) based on BLAST results whereas Ivory Coast Sequence 5 has a 97% BLAST identity with *Heterakis galinarum* (DQ503462, Smythe *et al.*, 2006).

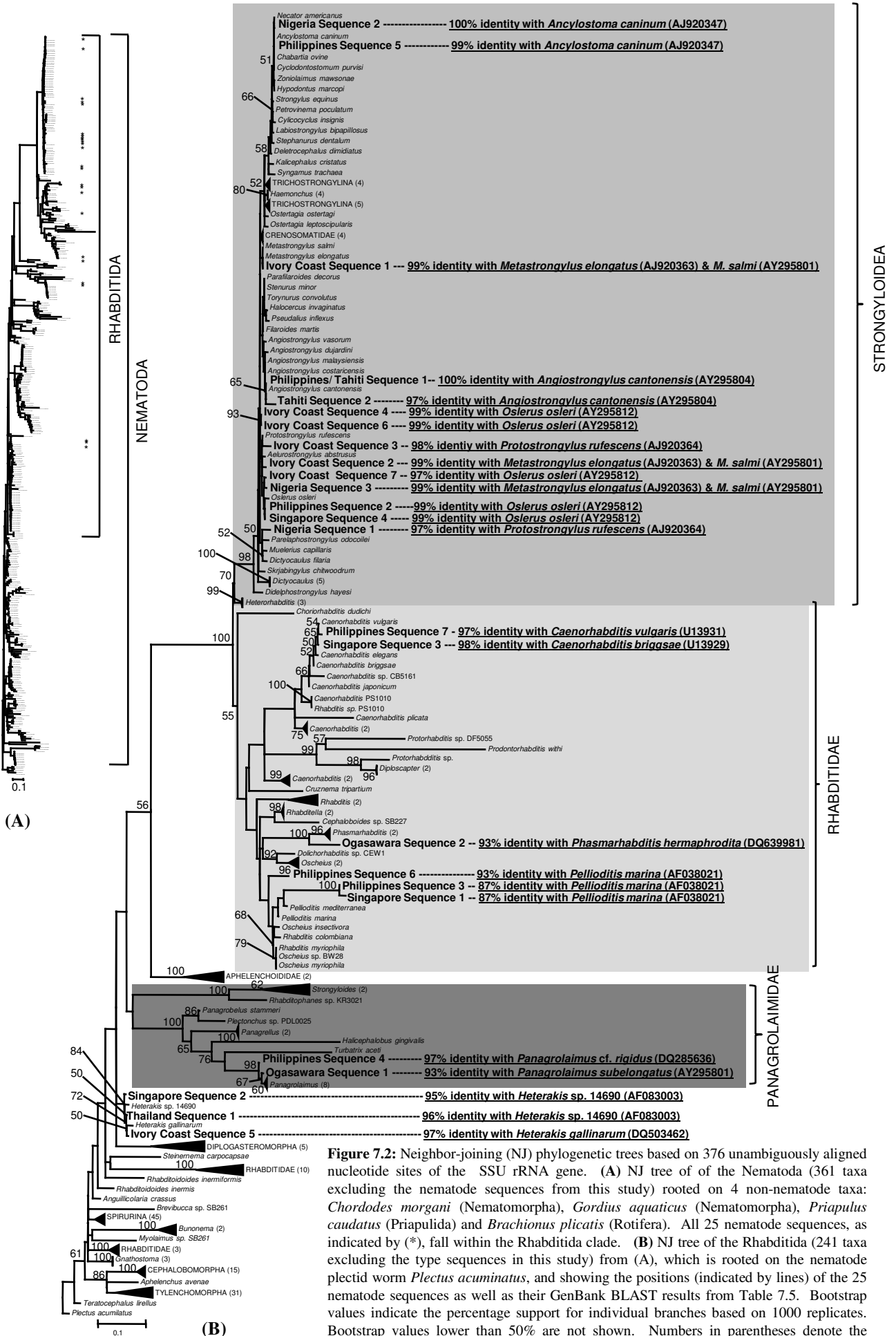


Figure 7.2: Neighbor-joining (NJ) phylogenetic trees based on 376 unambiguously aligned nucleotide sites of the SSU rRNA gene. **(A)** NJ tree of the Nematoda (361 taxa) excluding the nematode sequences from this study) rooted on 4 non-nematode taxa: *Chordodes morgani* (Nematomorpha), *Gordius aquaticus* (Nematomorpha), *Priapulius caudatus* (Priapulida) and *Brachionus plicatis* (Rotifera). All 25 nematode sequences, as indicated by (*), fall within the Rhabditida clade. **(B)** NJ tree of the Rhabditida (241 taxa) excluding the type sequences in this study) from (A), which is rooted on the nematode plectid worm *Plectus acuminatus*, and showing the positions (indicated by lines) of the 25 nematode sequences as well as their GenBank BLAST results from Table 7.5. Bootstrap values indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. Numbers in parentheses denote the number of species within a clade. Scale bar for both trees shows one nucleotide change per ten nucleotide positions.

7.4. Discussion

7.4.1. *Angiostrongylus cantonensis* infection in snail and slug populations

The results of this study demonstrate the presence of *Angiostrongylus cantonensis* from *Achatina fulica* populations in the Philippines and Tahiti among the areas within the route of dispersal of *Achatina fulica*; in the case of the Philippines, the worm was also detected in the black slug *Laevicaulis alte*. This was demonstrated by the 100% BLAST identity of Philippines Sequence 1/Tahiti 1 with *A. cantonensis* based on the 5' end of the SSU rRNA gene and its distinction from all other *Angiostrongylus* species sequenced to date. Historically, cases of rat and mollusc infections in the Philippines (Garcia, 1979) and rat, gastropod and human infections in Tahiti (Alicata, 1966; Wallace & Rosen, 1969; Bronstein *et al.*, 1977) have been recorded. Although Wallace and Rosen (1969) surveyed Tahiti for the presence of *Angiostrongylus cantonensis* in gastropod intermediate hosts between 1965 and 1966 and found 3rd stage juveniles in areas where the rats were known to be infected with the parasite, they did not confirm the identity of the juveniles by feeding them to laboratory rats. Their survey did not include *A. fulica* among the gastropods they examined as they did not encounter the snail, noting that it was only reported in Tahiti in 1967 (Raut & Barker, 2002). This study therefore provides the first ever reported case of *A. cantonensis* infection in *A. fulica* in Tahiti. The first recorded case of human infection in Tahiti by the nematode parasite occurred in 1957 and rat infection in 1961 (Alicata, 1966), several years before *A. fulica* was supposedly introduced. In the case of Tahiti, at least, Alicata (1966) hypothesised that *A. cantonensis* was not brought there by *A. fulica*. Nevertheless, the results show that the Tahitian *Achatina fulica* is used as an intermediate host of *A. cantonensis*. Alicata (1966) also found *A. fulica* in Hawaii to be

infected with the parasite, and he concluded that if the snails and the worms are found in the same area, then it is highly likely that the majority of the snails in the population will be infected.

The other *A. fulica* populations surveyed in the current study that are within the dispersal route of *A. fulica* proved negative for *A. cantonensis*. Among these areas, Thailand, Japan and Singapore had prior reported cases of *A. cantonensis*. In the case of Thailand, *A. cantonensis* was “found” in the edible land snail, *Hemiplecta distincta*, from 16 provinces where a total of 467 snails were surveyed (Panha, 1988). However, it was not definitively identified by feeding the nematodes to laboratory rats; instead, identification relied more on the morphological features of the juveniles. The sampling site in Thailand for this study (with only 20 snails surveyed but with no *A. cantonensis* infection) is also near the border with Cambodia where the parasite was detected in both humans and rats (Brumpt *et al.*, 1968). In Japan, *A. cantonensis* was reported from 27 cases of human infections, the majority of which are from the Okinawa Prefecture in the Pacific south of the Japanese main islands (Nakazawa *et al.*, 1992). In Ogasawara (with 12 snails surveyed in this study), there are no reported cases of *A. cantonensis* to date; Ogasawara is another group of islands in the Pacific southeast of the main Japanese islands. In Singapore (33 snails surveyed), there is one previous case of eosinophilic meningoencephalitis in a woman who was suspected of acquiring the parasite from eating contaminated raw vegetables, though it was more likely that these vegetables were imported from a nearby country where these worms are found (Lim *et al.*, 2004). For the remaining areas, Nepal, Myanmar and Moorea, there are no known reported cases to date that demonstrate the presence of *Angiostrongylus cantonensis*, though no systematic attempts to survey the snails and rats for worms in these areas have been done so far.

For West Africa, neither the *Achatina* sp. sample population from Ibadan, Nigeria (3 snails surveyed) nor that of the Ivory Coast (22 snails surveyed) yielded *A. cantonensis*. However, both countries had prior reported cases of *A. cantonensis* infections. A previous survey conducted on 22 rats in Ibadan in 1989 also failed to recover the parasite (Kliks & Palumbo, 1992), although it was detected in rats in Port Harcourt 450 km to the southeast (Udonsi, 1989; Kliks & Palumbo, 1992). In the Ivory Coast, the only recorded incidence was based on a single patient in 1979 that manifested the disease (Nozais *et al.*, 1980).

Angiostrongylus cantonensis was found in only two of the global populations of *A. fulica* sampled in this study. These results are therefore insufficient to permit any consideration of the role of *A. fulica* in the dispersal of *A. cantonensis*. The low success in finding the parasite in the snail intermediate host could have been caused by several factors. For one, the absence of *A. cantonensis* in *A. fulica* populations within the snail's route of dispersal, with the exception of the Philippines and Tahiti, may be due to the patchy distribution of the parasite. This was shown in a previous study by Bisseru (1971) in which he surveyed the presence of *A. cantonensis* in *A. fulica* populations from 27 sites in West Malaysia and found two sites with no parasite in any of the snails. One of these sites, Padang Besar, Perlis, with 100 snails sampled, is less than 65 km from Alor Star, Kedah, with 70 snails sampled and with an infection rate of 28.5%. Further to the patchy distribution of the parasite, the snail intermediate host itself could also have a patchy distribution and be uncommon in some of the sampling sites, thus reducing the probability of finding the parasites. For example, the Subic Bay Forest Reserve in the Philippines only yielded four individuals despite rigorous searching. In addition to the issue of patchy distribution, this study was also affected by sampling limitation; in particular, sampling was severely hampered by the availability

of live snail samples that could be brought out of the sampling sites. For instance, governmental restrictions in India and Sri Lanka prevent the export of live snails from these countries. Based on these factors, a comprehensive global survey of *A. fulica* populations was not possible.

Angiostrongylus cantonensis is not stringent as to its choice of intermediate and definitive hosts (Prociv *et al.*, 2000); therefore, the current distribution range of the parasite could be attributed to more than just one host. Indeed, the parasite is known to have been established in many localities where *A. fulica* is absent or has arrived after *A. cantonensis*. Already mentioned is Tahiti, to which the parasite could have been brought either by contaminated rats or by molluscs other than *A. fulica* that were probably imported by labourers from Indochina and Indonesia in the 1950s (Alicata, 1966). Other examples include Brisbane, Australia, where the parasite was found in the exotic rats *R. norvegicus* and *R. rattus* (Yong *et al.*, 1981), and New Orleans, USA where the worm was recovered from *R. rattus* (Campbell & Little, 1988). In both cases, local snails and slugs were also found to harbor *A. cantonensis* either in the wild (Yong *et al.*, 1981) or experimentally (Campbell & Little, 1988). Furthermore, other intermediate hosts with similar routes of dispersal to that of *A. fulica* could just as easily have aided the dispersal of *A. cantonensis*. One candidate is the black slug, *Laevicaulis alte*, which originated from Africa (Solem, 1964) and has spread across the globe in association with humans (Hoffman, 1925); the slug is also known to harbor *A. cantonensis* as demonstrated by Salazar and Cabrera (1969) and by the results from this study. It is therefore highly likely that the dispersal of the nematode is attributed to a complex combination of rats and mollusc intermediate hosts, and a systematic survey of all possible taxa that could harbour *A. cantonensis* as well as the population genetic analysis of global populations of the parasite and its hosts, juxtaposed with the

information regarding the first reported cases of infection in new areas, is perhaps the only way to trace the dispersal of *A. cantonensis*.

7.4.2. The presence of other worms in snail and slug populations

Nematodes other than *Angiostrongylus cantonensis* can likewise infect terrestrial snails and slugs such as *Achatina fulica* and *Laevicaulis alte*. The BLAST results and the phylogenetic analysis in this study clearly revealed 23 unique nematode species from the snails and slugs that fall within the Rhabditida clade and which have not yet been characterised molecularly using the 18S rRNA gene as they did not have a match with any named nematode taxa in GenBank. Some of these nematode species were also found to infect the same snail, as in the case of two *Achatina* sp. from the Ivory Coast, three *Achatina fulica* from the Philippines and one *A. fulica* each from Singapore, Ogasawara and Tahiti. Infection in gastropods by more than one species of nematodes is not uncommon, although localization of these parasites to specific organs does occur (Morand *et al.*, 2004). In another study, Viyada (2005) surveyed 200 adult *A. fulica* from two provinces in Thailand in which he found *Rhabditis* sp. in the alimentary tract. He suggested that this species could have been an accidental parasite of the snail as no noticeable damage to the host was observed.

Some species of nematodes pass through a dauer stage, a non-feeding alternative 3rd juvenile stage that is resistant to stress and can survive for several months without feeding. In free living soil nematodes such as *Caenorhabditis elegans*, the dauer stage is utilised for dispersal and forms associations with invertebrate hosts or even feeds on carcasses of animals. For instance, *C. elegans* was found in snails (*Helix*, *Oxychilus* and *Pomatias*), isopods (*Oniscus asellus*) and a glomeris myriapod, indicating a wide host specificity for a nematode originally thought to be free-living (Barriere & Felix,

2005). Some of the nematode infections in this study could have been attributed to associations with the dauer stage; for example, Singapore Sequence 3 has a 98% identity with the free-living soil nematode *Caenorhabditis briggsae* while Philippines Sequence 7 has a 97% identity with *C. vulgaris*.

Grewal *et al.* (2003) listed 108 nematodes species that are known to be parasitic on gastropods and divided them into those that use gastropods as intermediate hosts and those that use gastropods as definitive hosts. For the first group of gastropod-parasitizing nematodes, the majority belong to the order Strongylida while the rest are from the Ascaridia, Spirurida and Rhabditida. Based on the results of this study, 13 nematode sequences fall within the Strongyloidea: Tahiti Sequence 2, Nigeria Sequences 1, 2 and 3, Ivory Coast Sequences 1, 2, 3, 4, 6 and 7, Philippines Sequences 2 and 5 and Singapore Sequence 4. A further three sequences cluster with the *Heterakis* species from the Ascaridia: Singapore Sequence 2, Thailand Sequence 1 and Ivory Coast Sequence 5. Based on their phylogenetic positions, these nematodes were probably juveniles and most likely used *Achatina fulica* and *Achatina* sp. as intermediate hosts. For the second group of gastropod-parasitizing nematodes, the majority belong to the Rhabditida (Grewal *et al.*, 2003). Six other sequences in this study fall within the Rhabditidae: Singapore Sequences 1 and 3, Philippines Sequences 3, 6 and 7, and Ogasawara Sequence 2. These nematodes were probably adults that used the snails and slugs as definitive hosts. On the other hand, Ogasawara Sequence 1 and Philippines Sequence 4 did not fall in either group described by Grewal *et al.* (2003) as they cluster with the Panagrolaimidae, a group of bacteria-feeding soil nematodes. It is highly likely that the *A. fulica* samples from Ogasawara and the Philippines *L. alte* that was infected with the nematode bearing Philippines Sequence 4 became accidental hosts of these probable free-living soil nematodes.

There were instances in the study when a nematode sequence clustered with a particular taxon or group of taxa in the phylogenetic analysis and yet was most similar to another species based on the BLAST result. This was true for Nigeria Sequences 1 and 3 and Ivory Coast Sequence 2. It should be noted that the BLAST results were based on approximately 480 nucleotide sites whereas the phylogenetic analysis utilised only 376 sites as the rest could not be unambiguously aligned and that the clades generated were not well supported at the tips. For this reason, BLAST results take precedence as they identify the closest sequence match to named nematode taxa found in GenBank using all available nucleotide sites obtained while the phylogenetic analyses only determine which clades the unknown sequence types belong to based on a subset of the sequence information obtained.

7.4.3. Limitations and the next step

This study was heavily dependent on the availability of live samples of *Achatina fulica* and other gastropod intermediate hosts. Alicata's hypothesis was not sufficiently tested precisely because of the patchy and limited sampling of live snails and parasites, and the second objective (comparison of the population genetics of the snail and the parasite to check for similarity in their dispersal) was therefore not met as only the population genetics of global samples of *Achatina fulica* based on the 16S rRNA gene was available (see Chapter 5). Many areas were not covered because of governmental restrictions on the export of live snails, the difficulty of finding these snails by collectors and keeping them alive whilst in transit, the unwillingness of some couriers to ship live specimens, and the UK government's restrictions on the importation of such live snails from different countries. One practical solution will be to collect the samples and extract the worms in the field. Another is to develop a technique that will detect the

presence of *A. cantonensis* through PCR in ethanol-preserved specimens, though this technique will not be able to determine the parasite load (number of parasites present) per individual snail.

Owing to the fact that *Angiostrongylus cantonensis* has a patchy distribution as demonstrated by the survey from three sampling sites in the Philippines, several sampling sites per locality should also be covered. In addition, other gastropod intermediate hosts that could have had a role in the dispersal of *A. cantonensis* should be surveyed alongside *A. fulica*.

7.5. Summary

Within the route of dispersal of *Achatina fulica*, *Angiostrongylus cantonensis* was only found in *A. fulica* populations in the Philippines and Tahiti as well as the *Laevicaulis alte* populations in the Philippines. The results for Tahiti provide the first recorded case of *A. cantonensis* infection for *A. fulica* in that island. Due to the patchy distribution of *A. cantonensis* and the limited sampling of the snail intermediate host, the results from this study are insufficient to test Alicata's hypothesis that the Giant African Land Snail is primarily responsible for the spread of the parasite. Further sampling is therefore required. *Angiostrongylus cantonensis* has a wide range of suitable intermediate hosts, and gastropods such as the black slug *Laevicaulis alte*, which this study clearly demonstrated to harbor the parasite, should also be considered in future studies to determine their role in the dispersal of the parasite. Other worms found in *Achatina fulica*, *Achatina* sp. and *Laevicaulis alte* include 12 strongyloid species and 3 ascarid species that probably used the snails as intermediate hosts and 5 rhabditid species that probably used the snails as definitive hosts. One panagrolaimid

nematode species each from Ogasawara and the Philippines most likely used *A. fulica* and *L. alte*, respectively, as accidental hosts.

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CHAPTER 8 – Summary

The Giant African Snail *Achatina (Lissachatina) fulica* is a pest species that originated in East Africa but currently has a pantropical distribution brought about by human activities. It is also a known intermediate host of the rat lungworm, *Angiostrongylus cantonensis*, which can infect humans and cause eosinophilic meningoencephalitis.

Achatina fulica belongs to the Stylommatophora, a group of land snails and slugs that has two pairs of caudal tentacles, a contractile pneumostome and no operculum. Recent molecular phylogenetic study based on the rRNA gene cluster has shown a clear dichotomy of the Stylommatophora into the ‘non-achatinoidean’ clade and the ‘achatinoidean’ clade, which includes *A. fulica*. Within the ‘achatinoidean clade’ is the Achatinoidea, which includes the Achatinidae, Coeliacidae, Ferussaciidae, Glessulidae, Subulinidae and Thyrophorellidae. The phylogeny of the Achatinoidea was investigated further using 24 taxa across 6 families based on 3 nuclear genes (rRNA cluster, actin and histone 3) and 2 mitochondrial genes (CO1 and 16S rRNA). The 3rd codon position of the CO1 gene and the 16S rRNA were later excluded as they were found to be saturated. All strongly-supported groups in the rRNA phylogeny were recovered in the combined gene phylogenies with longer internal branches and receiving similar or slightly better statistical support. The study demonstrated the monophyly of the Achatinidae but not the Ferussaciidae, Coeliacidae and the Subulinidae. These results suggest the need to re-evaluate of the Ferussaciidae, Coeliacidae and Subulinidae as valid groupings.

The first systematic attempt to reconstruct the phylogeny of the Achatinidae was also conducted using 26 species from 7 genera based on the same genes used to evaluate the relationships of the Achatinoidea. The nuclear actin gene and the 3rd codon position of the mitochondrial CO1 gene were subsequently excluded due to the presence of paralogous sequences in the former that affected the phylogeny and the severe saturation of the latter. Results showed four distinct groups that to a large extent follow the geographical distributions of the taxa used in the survey. These were the West African group that fell at the base of the tree, a mixed group consisting of members with distributions ranging from the western, central, eastern and southern regions, and two groups that formed sister clades and which probably arose from the mixed group: the Southern African group and the East African group, which included *A. fulica*. The East African group incorporated all members of the subgenus *Lissachatina* of *Achatina*, which also exhibit distinct features of the reproductive organ, as well as the subgenus *Euaethiopina* of *Achatina*. Evidence from both molecular and morphological data would suggest that *Lissachatina* should be elevated to genus status.

The extent of genetic variation among global populations of *A. fulica* was also investigated using a fragment of the 16S rRNA gene to determine how many genetic types emerged from East Africa. Of the 15 distinct haplotypes observed, 13 were found in East Africa and nearby Indian Ocean islands while the rest of the global populations shared only haplotype C that came out of East Africa. Two non-African haplotypes (E from the Philippines and F from New Caledonia and Barbados) were probably recent mutations derived from C. This would indicate that the global populations of *A. fulica* descended

from a small group of individuals that shared the same haplotype and that these populations could be experiencing genetic bottlenecks as a consequence of lack of variation.

The role of *A. fulica* in spreading the rat lungworm *Angiostrongylus cantonensis* was evaluated. It is believed that the rat lungworm, like *A. fulica*, also originated in East Africa, and the nearly parallel distribution of the snail intermediate host and the parasite is noted. A molecular marker was first developed using a small segment of the SSU rRNA gene to rapidly identify the worm from snail tissues. This marker was also found to discriminate between closely related species of *Angiostrongylus*. Using this molecular marker, global populations of *A. fulica* were surveyed to detect the presence of *A. cantonensis* within the route of dispersal of *A. fulica*. The parasite was only found in the Philippines and Tahiti. The results for Tahiti provide the first recorded case of *A. cantonensis* infection for *Achatina fulica* in that country. The limited sampling of *A. fulica* and the patchy distribution of *A. cantonensis* could explain the lack of *A. cantonensis* among the populations sampled, with the exception of the Philippines and Tahiti. It is therefore difficult to assess if the snail has a role in the spread of the parasite, and the possibility of other gastropods such as *Laevicaulis alte* contributing to the spread of *A. cantonensis* is not discounted.

APPENDIX

Appendix 2.1: Command lines for generating likelihood scores for the 12 models in PAUP*

2.1.1. JC69

1. paup> log file=filename.JC69models.paupout (this saves an output file)
2. paup> execute dataset.nex
3. paup> set criterion=distance (sets the optimality criterion to distance)
4. paup> dset distance=ml (specifies the distance measure to maximum likelihood for the estimation of the likelihood score for the model, which will be used for the LRT)
5. paup> lset nst=1 basefreq=equal rates=equal pinvar=0 (this specifies the assumptions of the JC69 model, which are equal frequency for all bases and equal rate of substitutions)
6. paup> nj (this generates a neighbor-joining tree based on the assumptions of the model)
7. paup> likelihoods /basefreq=equal rates=equal pinvar=0 (this estimates a likelihood score based on the model)

2.1.2. JC69+ Γ (continued from JC69)

8. paup> likelihoods /basefreq=equal rates=gamma shape=estimate ncat=16 pinvar=0 (this adds the among site gamma rate variation with 16 categories dividing the discrete approximation of the gamma distribution)

9. paup> lset nst=1 basefreq=equal rates=gamma shape=previous ncat=16
pinvar=0
10. paup> nj
11. repeat 8-10 until the parameters and likelihood score do not change; note the likelihood score.

2.1.3. F81

1. paup> log file=filename.F81models.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml
5. paup> lset nst=1 basefreq=estimate rates=equal pinvar=0 (this specifies the assumptions of the F81 model, which are unequal base frequencies and equal rate of substitutions)
6. paup> nj
7. paup> likelihoods /basefreq=estimate rates=equal pinvar=0
8. paup> lset nst=1 basefreq=previous rates=equal pinvar=0
9. nj
10. repeat 7-9 until parameters and likelihood score do not change; note the likelihood score.

2.1.4. F81+ Γ (continued from F81)

11. paup> likelihoods /basefreq=estimate rates=gamma shape=estimate ncat=16
pinvar=0
12. paup> lset nst=1 basefreq=previous rates=gamma shape=previous ncat=16
pinvar=0
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the likelihood score.

2.1.5. K2P

1. paup> log file=filename.K2Pmodels.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml
5. paup> lset nst=2 basefreq=equal tratio=estimate rates=equal pinvar=0 (this specifies the assumptions of the K2P model, which are equal frequency of bases and different substitution rates for transtitions and transversions)
6. paup> nj
7. paup> likelihoods /basefreq=equal tratio=estimate rates=equal pinvar=0
8. paup> lset nst=2 basefreq=equal tratio=previous rates=equal pinvar=0
9. paup> nj
10. repeat 7-9 until parameters and likelihood score do not change; note the likelihood score.

2.1.6. K2P+ Γ (continued from K2P)

11. paup> likelihoods /basefreq=equal tratio=estimate rates=gamma
shape=estimate ncat=16 pinvar=0
12. paup> lset nst=2 basefreq=equal tratio=previous rates=gamma shape=previous
ncat=16 pinvar=0
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the
likelihood score.

2.1.7. HKY85

1. paup> log file=filename.HKY85models.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml
5. paup> lset nst=2 basefreq=estimate tratio=estimate rates=equal pinvar=0 (this
specifies the assumptions of the HKY85 model, which are unequal base
frequencies and different substitution rates for transitions and transversions)
6. paup> nj
7. paup> likelihoods /basefreq=estimate tratio=estimate rates=equal pinvar=0
8. paup> lset nst=2 basefreq=previous tratio=previous rates=equal pinvar=0
9. paup> nj

10. repeat 7-9 until parameters and likelihood score do not change; note the likelihood score.

2.1.8. HKY85+ Γ (continued from HKY85)

11. paup> likelihoods /basefreq=estimate tratio=estimate rates=gamma
shape=estimate ncat=16 pinvar=0
12. paup> lset nst=2 basefreq=previous tratio=previous rates=gamma
shape=previous ncat=16 pinvar=0
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the likelihood score.

2.1.9. TN93

1. paup> log file=filename.TN93models.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml
5. paup> lset nst=6 basefreq=estimate rmatrix=estimate rates=equal pinvar=0
rclass=(a b a a e a) (this specifies the assumptions of the TN93 model, which are unequal base frequencies and different substitution rates for transitions in purines and pyrimidines and transversions)
6. paup> nj

7. paup> likelihoods /basefreq=estimate rmatrix=estimate rates=equal pinvar=0
rclass=(a b a a e a)
8. paup> lset nst=6 basefreq=previous rmatrix=previous rates=equal pinvar=0
rclass=(a b a a e a)
9. paup> nj
10. repeat 7-9 until parameters and likelihood score do not change; note the
likelihood score.

2.1.10. TN93+ Γ (continued from TN93)

11. paup> likelihoods /basefreq=estimate rmatrix=estimate rates=gamma
shape=estimate ncat=16 pinvar=0 rclass=(a b a a e a)
12. paup> lset nst=6 basefreq=previous rmatrix=previous rates=gamma
shape=previous ncat=16 pinvar=0 rclass=(a b a a e a)
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the
likelihood score.

2.1.11. GTR

1. paup> log file=filename.GTRmodels.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup> dset distance=ml

5. paup> lset nst=6 basefreq=estimate rmatrix=estimate rates=equal pinvar=0 (this specifies the assumptions of the GTR model, which are unequal base frequencies and different substitution rates)
6. paup> nj
7. paup> likelihoods /basefreq=estimate rmatrix=estimate rates=equal pinvar=0
8. paup> lset nst=6 basefreq=previous rmatrix=previous rates=equal pinvar=0
9. paup> nj
10. repeat 7-9 until parameters and likelihood score do not change; note the likelihood score.

2.1.12. GTR+ Γ (continued from GTR)

11. paup> likelihoods /basefreq=estimate rmatrix=estimate rates=gamma shape=estimate ncat=16 pinvar=0
12. paup> lset nst=6 basefreq=previous rmatrix=previous rates=gamma shape=previous ncat=16 pinvar=0
13. paup> nj
14. repeat 11-13 until the parameters and likelihood score do not change; note the likelihood score.

Appendix 2.2: Command lines in PAUP* for estimating the uncorrected and corrected distances as well as the transition and transversion values

1. paup> log file=filename.distances.paupout
2. paup> execute dataset.nex
3. paup> set criterion=distance
4. paup>dset distance=p
5. paup> showdist (this generates the uncorrected total pairwise distances)
6. paup> savedist /format=onecolumn file=filename.distances.p.1col (this saves the pairwise uncorrected total distances in one column)
7. paup> dset subst=ti (this specifies that the transition distances will be computed)
8. showdist (this generates the transition distances)
9. paup>savedist /format=onecolumn file=filename.distances.p.ti.1col
10. paup> dset subst=tv (this specifies that the transversion distances will be computed)
11. showdist (this generates the transversion distances)
12. paup>savedist /format=onecolumn file=filename.distances.p.tv.1col
13. paup> dset subst=all (this restores the setting to pertain to total distances)
14. paup> set criterion=distance
15. paup> dset distance=ml
16. set the predetermined optimal criterion using the lset command
17. paup> showdist (this generates the corrected total distances)
18. paup> savedist /format=onecolumn file=filename.distances.ml.model.1col

Appendix 2.3: Command lines for the different tree-construction methods

2.3.1. Maximum likelihood (ML) method

The ML method was done in PHYML Version 2.4.4 package (Guindon *et al.*, 2005). The parameters of the optimal model was specified when the dataset analyzed.

Enter the sequence file name > sequence.file.phy
er the sequence file name > sequence.file.phy

Settings:

D:	Data type (DNA/AA)	DNA
I:	Input sequences interleaved (or sequential)	interleaved
S:	Analyze multiple data sets	no
B:	Non parametric bootstrap analysis	yes
	Number of replicates >	1000
	Print bootstrap trees (and statistics) ? [Y/n]	y
M:	Model of nucleotide substitution	(depending on optimal model)
E:	Base frequency estimates (empirical/ML)	ML
V:	Proportion of invariable sites (fixed/estimated)	fixed (p-invar = 0.00)
R:	One category of substitution rate [yes/no]	no (if with gamma distribution)
C:	Number of substitution rate categories	16 (if with gamma distribution)

A:	Gamma distribution parameter (fixed/estimated)	estimated (if with gamma distribution)
	Optimize alpha ? [Y/n]	Y
U:	Input tree (BIONJ/user tree)	BIONJ
O:	Optimize tree topology	yes

2.3.2. Bayesian inference

Bayesian inference (BI) analysis was carried out in the MrBayes Version 3.1.2 package (Ronquist & Huelsenbeck, 2003). The program was set to run with four independent chains, each of which aimed to converge on a stable likelihood value from a number of generations. Below are the command lines for approximating the posterior probabilities.

1. MrBayes > log start file=filename.mbout (this saves an output file)
2. MrBayes > execute filename
3. MrBayes > lset nst=1/2/6 rates=equal/gamma Ngammacat=16 (only for gamma distributed rates) (this states the model in which the tree will be based)
4. MrBayes > showmodel (this is to show the model settings)
5. MrBayes > help mcmcp (this is to show default settings of the Markov Chain Monte Carlo analysis)
6. MrBayes > mcmcp Nruns=1 Ngen=2000000 Temp=0.075 or 0.1 or 0.125 (the number of generations to be run as defined by Ngen and the temperature to heat the chains as defined by Temp need to be optimized as described below;

initially, values of 2000000 for Ngen and 0.1 for Temp are used and later on changed if necessary)

7. MrBayes > mcmc (this is to run the analysis)

At the end of the run, the program asked if the analysis should be continued. The word 'yes' was typed when the values from the four columns did not converge to similar likelihood scores. The number of additional generations (e.g. 1000000) was indicated to resume the analysis. Otherwise, the word 'no' was typed. When the cold chain (value enclosed with bracket) did not switch randomly across the four chains, the analysis was repeated but the Temp setting was changed.

When the analysis was completed, the state exchange information was displayed. As a rule of thumb, the values found on the upper diagonal, which expressed the acceptance rates of the Metropolis-Hastings mcmc sampler, should range between 0.1 and 0.7. When the acceptance rates were too low, the analysis was repeated but using a lower Temp. When the rates were too high, the analysis was repeated with a higher Temp.

8. MrBayes > sump burnin=value (e.g. 19001 if 2000000 generations were made; this summarizes the samples of model parameters)

Note: The value given indicated the number of samples to be discarded. A sample contained 100 generations; however, the first generation is always sampled and should be included in the samples to be discarded.

9. MrBayes > sumt burnin =value (e.g. 19001 if 2000000 generations were made; this summarized the samples of trees and branch lengths)

2.3.3. Neighbor-joining (NJ) method

The NJ method was done in PAUP* by starting out with the same parameters set by the identified optimal model for DNA sequence evolution, which was then followed by the commands listed below.

1. paup> nj brlens=yes (to display the table of branch lengths)
2. paup> savetrees /format=phylip brlens file=filename.nj.tree.phy (to save the tree in phylip format)
3. paup> savetrees /format=nexus brlens file=filename.nj.tree.nex (to save the tree in nexus format)
4. paup>bootstrap nreps=1000 method=nj keepall=yes
treefile=filename.nj.boottree (to calculate and save the bootstrap values for 1000 replicates)
5. paup>quit

2.3.4. Maximum parsimony (MP) method

The MP method was also carried out in Paup*; this did not employ any models of DNA sequence evolution.

1. paup>set maxtrees=100 increase=auto
2. paup> set criterion=parsimony
3. paup> hsearch swap=tbr addseq=random nreps=10 start=stepwise (this is to make a heuristic or non-exhaustive search of the most parsimonious trees)
4. paup> showtrees all
5. paup> savetrees /fmt=phylip brlens=yes file=filename.mp.phy

6. paup> savetrees /fmt=nexus brlens=yes file=filename.mp.nex
7. paup> contree /majrule=yes file=filename.mp.constree (this generates a consensus tree of all the equally most parsimonious trees found)
8. paup> bootstrap nreps=1000 search=heuristic keepall=yes
file=filename.mpboot.treefile (to calculate and save the bootstrap values for 1000 replicates)

Appendix 2.4: Command lines for the partition homogeneity test in PAUP* to determine if individual genes could be combined as a single dataset.

1. paup> execute concatenated.file.nex
2. paup> charpartition gene1:1-3435, gene2:3436-4296, gene3:4297-4624. (this is to specify which segment of the concatenated sequence belongs to which gene)
3. paup> hompart partition=genes nreps=1000 seed=123 search=heuristic (this is to perform a homogeneity test with 1000 homogeneity replicates searched heuristically using a random seed number of 123)

Appendix 2.5: The command lines to run the Shimodaira-Hasegawa test in PAUP* to compare for significant difference between optimal and constrained trees

1. paup> execute concatenated.file.nex
2. set the parameters of the optimal model
3. paup> constraints taxonmonophyly (MONOPHYLY) = ((1,2)) (where numbers refer to the taxon numbers; this specifies which taxa were constrained to form a clade based on an *a priori* hypothesis that they are monophyletic)
4. paup> nj brlens=yes enforce=yes constraints=taxonmonophyly (this generated a constrained neighbor-joining tree)
5. paup> savetrees /format=nexus brlens file=taxonmonophyly.nex (this saved the constrained neighbor-joining tree)
6. execute trees.file (this file should include both the constrained tree from no. 5 as well as the optimal neighbor-joining tree)
7. paup> showtrees all (this displayed both trees)
8. paup> lsocres all/shtest=rell (this computed for the likelihood scores of the two trees based on the estimated likelihood method and compared for significant difference between the two likelihood scores at $P=0.05$ level of significance)

Appendix 3.1: Alignment of the LSU rRNA gene for the Achatinoidea and three streptaxid outgroup taxa. Positions included for the phylogenetic analyses were marked by “m” within “NUCEOTIDES INCLUDED”.

	10	20	30	40	50	60
<i>Achatina fulica</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Achatina achatina</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Achatina stuhlmanni</i>	-----AGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Cochlitoma ustulata</i>	-----	---TTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Allopeas clavulinum</i>	-----AGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-GCCATCC
<i>Bocageia</i> sp.	ATTGCAGAAC	ACATTGAACA	TCGATACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Eutomopeas layardi</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Leptinaria lamellata</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-GCCATCC
<i>Paropeas achatinaceum</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-GCCATCC
<i>Riebeckia</i> sp.	-----AAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Rumina decollata</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Subulina octona</i>	-----	-----	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Subulina striatella</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Subulina vitrea</i>	-----	-----ACA	TCGATACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Subulona</i> sp.	-----	-----	-----CCTT	GAACGCATAT	GGCGGCCTCG	GGTCCATCC
<i>Tortaxis erectus</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Xerocerastus</i> sp.	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Zootecus insularis</i>	ATTGCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Coeliaxis blandii</i>	-----AAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Pyrgina umbilicata</i>	-----AAC	ACATTGAACA	TCGACATCTT	GAACGCACAT	GGCGGCCTCG	GG-GCCATCC
<i>Glessula ceylanica</i>	ATTGCAGAAC	ACATTGAACA	TCGATACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Cecilioides gokweanus</i>	-----	-----	---ACATCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Ferussacia folliculus</i>	---GCAGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Thyrophorella thomensis</i>	ATTGCAGAAC	ACATTGAACA	TcGACATCTT	GAACGCACAT	GGCGGCCTCG	GG-GCCATCC
<i>Gibbulinella dewinteri</i>	-----	-----	-----NCTT	GNACGCANAT	GGCGGCCTCG	GG-TCCATCC
<i>Gonaxis quadrilateralis</i>	-----AGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TCCATCC
<i>Gonospira</i> sp.	-----AGAAC	ACATTGAACA	TCGACACCTT	GAACGCACAT	GGCGGCCTCG	GG-TAAATCC
NUCLEOTIDES INCLUDED	-----	-----mmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mm-mmmmmmm

	70	80	90	100	110	120
<i>Achatina fulica</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---TACAA	AGCCTTCGCT	TCGTATGAGG
<i>Achatina achatina</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---TACAA	AGCCTTCGCT	TCGTATGAGG
<i>Achatina stuhlmanni</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---TACAA	AGCCTTCGCT	TCGTATGAGG
<i>Cochlitoma ustulata</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---TACAA	AGCCTTCGCT	TCGTATGAGG
<i>Allopeas clavulinum</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---AACAA	AACCATCGCT	TCCTTTCTTC
<i>Bocageia</i> sp.	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---TCAA	AGCAATCGCT	TCCTGTTTGG
<i>Eutomopeas layardi</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---ATCA	AACCATCGCT	CCTCTT-TTC
<i>Leptinaria lamellata</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---ACTA	AACCAATCGCT	TCAGTT-TTT
<i>Paropeas achatinaceum</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---ACCA	AACCATCGCT	TCCTTTTTTT
<i>Riebeckia</i> sp.	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---TCAA	AGCAATCGCT	TCGAGATGGC
<i>Rumina decollata</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---TCAA	AGCCTTCGCT	TCGCTTCTGG
<i>Subulina octona</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---TCGAA	AGCAATCGCT	TCGTTTGTGT
<i>Subulina striatella</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---TCTCA	AGCAATCGCT	TCGTTTTATA
<i>Subulina vitrea</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---TCAA	AGCAATCGCT	TCTATTTTGG
<i>Subulona</i> sp.	CGGNGCCACG	CCCCTCTGAG	GGTCGGCGAG	AATCTTAATA	AGCAATCGCT	TCTTATCGGA
<i>Tortaxis erectus</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---TATCA	AGCAATCGCT	TCGCACTAAA
<i>Xerocerastus</i> sp.	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AR---TCAA	AGCAATCGCT	TCGCTTTTTT
<i>Zootecus insularis</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG---ACAA	AGCAATCGCT	TCGCTTTTTT
<i>Coeliaxis blandii</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---TCAA	AGCAATCGCT	TCGAGCTGGG
<i>Pyrgina umbilicata</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---ATCA	AACCAATCGCT	TCTCCTCAGA
<i>Glessula ceylanica</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---GTCAA	AGCAATCGCT	TCGAATTTGA
<i>Cecilioides gokweanus</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AA---ACTA	CACCAATCGCT	TCGCTTTATG
<i>Ferussacia folliculus</i>	CGAGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AG-----AA	ACTCATCGCT	TTTCATTTGC
<i>Thyrophorella thomensis</i>	CGGGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AT---ATCA	AACAATCGCT	TCCTTTTTCG
<i>Gibbulinella dewinteri</i>	CGAGGCCACG	CCCCTCTGAG	GGTCGGCGAG	GGTGGGGAGT	CTTAAAAAAA	GCCGTTTCGCT
<i>Gonaxis quadrilateralis</i>	CGAGGCCACG	CCCCTCTGAG	GGTCGGCGAG	AC---TTATA	AGCAATCGCT	TTTTCCGTTG
<i>Gonospira</i> sp.	CGAGGCCACG	CCCCTCTGAG	GGTCGGCGAG	TC---TAAAA	GCCGTTTCGCT	TTCTGCTGAC
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	-----	-----	-----


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          130      140      150      160      170      180
Achatina fulica      -CAGCAGGTC TCGCTC---A ATTTATTCCG AGATCCGTCC AGCTCTTC-- ---CTCTATC
Achatina achatina   GCAGCAGGTC TCG----- ATTTATTCCG AGATCCGTCT A-TACTTC-- NTGNTCTANT
Achatina stuhlmanni -CAGCAGGTC TCGCTT--AA ATTTCCGTCC AGATCTTC-- ----- -CTCTATC
Cochlitoma ustulata -CAGCAGGTC TCGCTC---A ATTTATTCCA AGATCCGTCT AGCTCTTC-- ---CTCTATC
Allopeas clavulinum TTACCAGACC AAAGGCCG-C ACGCCGTCGG GGTCTTTTCG TCTTCCGAAG CTCGAAGGGA
Bocageia sp.        CTCGGGGTTC TAACTAAACC CGTTCGCGCG CGTCCTTCTT CTTTTCCCG AGCTCGTGCG
Eutomopeas layardi  TATGTGACCA AAGGCCACCA CGCCGTCGTG AGACAGGTCT TTCGCTCATC CGAAGCTCGA
Leptinaria lamellata ATGACAACCTG GCATTCCCGT TTCTTCCGAG CTCGAGGAAG AGCGGGAGAG CCGCCGAAGC
Paropeas achatinaceum CTTATCAGAC CAAAGGCAGC GGCCTGTCGG TGT--ATCGT TCTTCCGAAG CTCGAAGGGA
Riebeckia sp.       GGTAAACAGGT CGCCTC-AAA CAAGCTGCTC CGATCCGATC GCCGCCGTTT CTTCCCGGGC
Rumina decollata    -CAGCTGGTC CGGTTT--- ----- -TA
Subulina octona     -CTCGACAGC GGGTCTCCAG CCTCCAGAGA AGGCTT---- -----
Subulina striatella -GTGACGGCG GGTCTCCAGC TTCCGATCGT CCCACAACCT TCCTTCGGCG TCGGAGGGA
Subulina vitrea     CACAGGGGTC TCCCGTTGGG GGGTTGGCTG GCAAGGCAAG GCAGGCATAT TTTCTTTCTC
Subulona sp.        GAA----- NNNNNNNNNN NNNNN-----
Tortaxis erectus    -CCGACGAGT GGTCTGCTT CCCCACCTTG GCTCTGAGTT TGCGGGCGGG CAGGCCGACC
Xerocerastus sp.    GGTNAACGGG TCCNCCTTGN TCAGNNAATT CCGTTC-CAT TNGNCTCGCG GGAAACAAAA
Zootecus insularis TTTTTYCTTT AAAAAAAAAA ATGCACCGGG TTTTGCTTAT TCCCACCATG TCGTTCTCCG
Coeliaxis blandii   TTGGGTCGTC TCGAAGTCCG ATCAGCTCAG TCCGTTCCCT CGCGCTCATC GTGGGAGGAT
Pyrgina umbilicata  GACCAAAGGC ACTCGCCGTC CGTCCGTCGG CCTTCTTAAG CTCGAAGGAA GCGCTTGATG
Glessula ceylanica CATGGGTGAC ACAATTCNA ANANACTTCN AANAATNCTT TTTTNTCCCC GCTTTTGAAA
Cecilioides gokweanus GCAACAGGTT TCCCCG-TCT TCTTCCGAGC T-----
Ferussacia folliculus CCTAAGGATT CGACCTTGCC ACCTCTGTGT TCGTGTTAGA GGGAGAAGGT GCAGAATTGC
Thyrophorella thomensis -ATCAAAGGC ACTCTCGC-C CGTCCGTCGG CCTTCTTAAG CTCGAAGGAA AGGCTTGATG
Gibbulinella dewinteri ATCGGTAGGC CTACGCGGCG CGTCCGCGAG AGGTTTGGCT TTTTCGGGAG GGCATATTTT
Gonaxis quadrilateralis GACTGCTGCT CGGGCCACCG TCTGTCTGGG TGANGAGGGG NATTGACGCN TCAACTCGTT
Gonospira sp.       TGGAAGGCAA CCGTCCGTCT GTACACAGCG CGGAGGAAAG TCCCTCTCGT GTTGCACGCT
NUCLEOTIDES INCLUDED -----

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          190      200      210      220      230      240
Achatina fulica      C-ATCCGCGG -CTCGTCCGG A--GGGGTTA CAGAGAGGAA GAAT-CGGGC G-----
Achatina achatina   G-NGTCCCG CTGCTCGTGC T---GGGACA CGGAAAAGAA GAATTGACN N-----
Achatina stuhlmanni T--TCCGCGG -CTCGTCCGG -----GGA CAGAAAGAAA GAAT-CTGTC G-----
Cochlitoma ustulata T-ATCCGCGG -CTCGTCCGG T---GGAGGA CAGAAAGGCG -----
Allopeas clavulinum GAGCGCAGAC ATGGTGGCGA TGGGCCGCGAG CAGAAGCGCT TTGGGCTCTC GTAGGTCGAT
Bocageia sp.        GTTGAGA--
Eutomopeas layardi  AGGGAGGGCG AAGACCGCGG CACTGCGAT GGGCCGCGC AGGAGCGCGT TGGGCTCTCG
Leptinaria lamellata GCATTGGGCT TTCGCGGGAC GCGCAGGGCA CGTGCTCGTC CGTCCGCGCT TCGTACCGCG
Paropeas achatinaceum GGGCGAAGAT ACGATGACGT CGGGCCGCGG CAGAAGCGCG TTGGGCTGTC GTAGGTCGAT
Riebeckia sp.       TCGTGGGAGG ATTCAAGTGG TTAGTTAGGA GGATGCTTTC GAGAGGTGAC CGTCAAGCGG
Rumina decollata    CCACGTCTGT TTTCACTCCG TGCTCTGTGG AGGGAGAAGC TGCGTCTGTT ATGA-----
Subulina octona     -----
Subulina striatella GGGTTTGGGT GACGTTT--- -----
Subulina vitrea     CTAATTTTTT TCATTTTCTT CCGCCCGGTG CGCTCTGACG GCGCGGGGTA GAGAGTGAAA
Subulona sp.        -----
Tortaxis erectus    GTCGAAGCGC GCTGGGCATT CCGAGGATGA GTGCTCCTCC ACATGTCATC CCCG-----
Xerocerastus sp.    A----- -NNNNNNNNN N-----
Zootecus insularis TCCGCCGTTT TTGGGATGAA AAGCGGCTCG TGGAGTGGCC GAA-----
Coeliaxis blandii   ACAAAGGCG GGGCGCGACT NAAATGNAC CGTACAATCG CACTGGGCT- ---- -NNNNNN
Pyrgina umbilicata  ATGACGACGG GCTTGTCTGCC GCAGAAGCGC TTTGAG-CTG TCGTGGGTCG TCAGTGACCC
Glessula ceylanica  AAAAA----- NNNNNNNNN- -----
Cecilioides gokweanus -----
Ferussacia folliculus CGGAAGCGCA CTGGGCTTTC GAGAATCACA CCATTCTCGC TCGAAAATAA AGAATGAAGT
Thyrophorella thomensis ACGACG-GGC TGCTGCTGCC GCAGAAGCGC TTTGGGCTT TCGTGGGTCG TCAGCGACCC
Gibbulinella dewinteri TTTTTTGCC ACTCGGTTA CCAGCGGCGT CCGTGAAGAC CGAAGAAAGC GCTTCTGAGC
Gonaxis quadrilateralis ACGGTGTGGT TGTCCCGAGG AGGAGGAAAA GCGCTCCGGG CTTTCGCGGN TGGGCAAAAA
Gonospira sp.       CCGGTTGCCG AGAAAGCGGT ATGGGCCTTC GCGGATGATG ACGTGCATCC CCGCGCCCTC
NUCLEOTIDES INCLUDED -----

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	250	260	270	280	290	300
Achatina fulica	-----	-----	-----	-----	-----	-----	-----
Achatina achatina	-----	-----	-----	-----	-----	-----	-----
Achatina stuhlmanni	-----	-----	-----	-----	-----	-----	-----
Cochlitoma ustulata	-----	-----	-----	-----	-----	-----	-----GC
Allopeas clavulinum	TTCAACGACC	ACCGTGGCCT	CAAGTACATC	GTGCGCCTGC	CCCGTCCGTT	TT--AATCGG	-----
Bocageia sp.	-----	-----	-----	-----	-----	-----	-----TTAA
Eutomopeas layardi	TAGGTCGATT	TCTACGACCG	CCGTGGCCTC	AAGTACATCG	TGCGCCTGCC	CCTGTCCGTT	-----
Leptinaria lamellata	TTGCCTCAAG	TACGTAGTGC	GCCCCCCTG	TCCGATCAAT	CGGTCCGTTG	TGCTCTCGAC	-----
Paropeas achatinaceum	TTCAACGACC	CCCGTAGCCT	CAAGTACATC	GTGCGCCTGC	CCTGTCCGTT	TTTTAATCGG	-----
Riebeckia sp.	CAATGGGCTT	TCGAGGATGC	TTGCTGCCTA	TNATGGANGG	AAAAAAGGAT	GGCTCGCTCC	-----
Rumina decollata	-----	-----	-----	-----	-----	-----	-----
Subulina octona	-----	-----	-----	-----	-----	-----	-----
Subulina striatella	-----	-----	-----	-----	-----	-----	-----
Subulina vitrea	ATACAGGAAA	AGAAGAAGAG	CCGCGGCCGC	CGTCGCCGCA	CG-----CCA	TTTTACTTTT	-----
Subulona sp.	-----	-----NNNNN	NNNNNNNNNN	NNNNNNNNNN	-----TTTTTTT	TTTTTGTTA-	-----
Tortaxis erectus	-----	-----	-----	-----	-----	-----	-----
Xerocerastus sp.	-----	-----	-----	-----	-----	-----	-----
Zootecus insularis	-----	-----	-----	-----	-----	-----	-----
Coeliaxis blandii	NNNN-----	-----	-----	-----	-----	-----	-----
Pyrgina umbilicata	TCGTGGCTTC	AAGTACAACG	TGCGCCTGCC	CCTGTCCGTT	TTAGTCGGTC	TCGTTAGCTC	-----
Glessula ceylanica	-----	-----	-----	-----	-----	-----	-----
Ceciloides gokweanus	-----	-----	-----	-----	-----	-----	-----
Ferussacia folliculus	GGTTTCCGTT	GCCTGAAGTA	CTGAAAAAAA	CTGCGTCCTC	TGTCCGTTCT	AGCTAGTCTA	-----
Thyrophorella thomensis	TCGTGGCCTC	AAGTACAACG	TGCGCCTGCC	CCTGTCCGTT	TTGGTCGGTC	TCGTTAGCTC	-----
Gibbulinella dewinteri	CTTCGCGGAT	CGCATGATCC	CCGTGGCTTC	AAGTGTACTC	GATGCGCGCG	CGCCCTTTGC	-----
Gonaxis quadrilateralis	TCANCCCCGT	GGCCACAAGT	GTAAGAAGATG	CGCGCGTGTG	TGCTGANCTG	CCGCCTGCCT	-----
Gonospora sp.	AAGTGCAAGC	GTTGAGGGAG	CGTCTTAGCT	GCTGCTTCTG	TCCCCGAC	TAAACGTGGA	-----
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	-----

	310	320	330	340	350	360
Achatina fulica	-GAGAAGGAA	GAAGAAGAGA	CCG-----	TCGA-AGCGC	TGTGGGCCGT	--CGTGGGAG	-----
Achatina achatina	ATANNGAAGG	G-AAGAGAGG	CCG-----	TCGA-AGCGC	TCTGGGCCGT	--CGTGGGAG	-----
Achatina stuhlmanni	-----GAG	AAGAAAGAGA	CCG-----	TCGA-AGCGC	TGTGGGCCGT	--CGTGGGAT	-----
Cochlitoma ustulata	GAAGGAAGAA	GAAGAAGAGA	CCG-----	TCGA-AGCGC	TGTGGGCCGT	--CGTGGGAG	-----
Allopeas clavulinum	TCTCGATGGT	TCTC-----	--TAACCGTC	GGAGCGCTCG	CTCGCTCGCC	GTCCGTCTGG	-----
Bocageia sp.	TAGGGCCGTG	CTGATG----	---AAGA-AG	CGCACTGGGC	TCT--CGAGG	AGAGAAGAAC	-----
Eutomopeas layardi	TTAATCGGTG	TCGATGGCTC	TCTAGCCGTC	TGAGCGCTCG	CTCGCTCGCC	GTCCGTCTGG	-----
Leptinaria lamellata	CGGAGCGCTC	GCCGTCAGTC	TGGCAGGACT	CGGCTCGCAT	AAAAGCATGA	AAATCGAGCCT	-----
Paropeas achatinaceum	TCTC-----	-----	---TACCGTC	GGAGCGCTCG	CTCGCTCGCC	GTCCGTCTGG	-----
Riebeckia sp.	TTTTTTTTTT	-----	-----N	NNNNNNNNNN	-----	-----	-----
Rumina decollata	-----	---TGAAGA	CCG-----	TCGA-AGCGC	ACTGGGCTTT	--CGAGGACG	-----
Subulina octona	-----	---GGTGA	CCG-----	TAGA-AGCGC	CCTGGGCTCT	--CGAGGAGG	-----
Subulina striatella	-----GAAGA	GGCTTGGCGA	CCG-----	TGGA-AGCGC	CCTGGGCTGT	--CGAGGAGG	-----
Subulina vitrea	AACGTGAGAG	GACCCG----	---GAGAAG	CGCACTGGGC	TTT--CGAGC	TAAAGAAACA	-----
Subulona sp.	-----	-----	-----	-----	-----	-----	-----
Tortaxis erectus	-----	---TGGC	CTGAAGTACA	AAATTTATGC	GCCGTCCCCT	CCAACCGAGC	-----
Xerocerastus sp.	-----	-----	-----N	NNNNNNNNNN	-----	-----	-----
Zootecus insularis	-----	---GACG---	---TCGAAG	CGTTCTGGCC	TT---CGAGG	ACGAGGAGGA	-----
Coeliaxis blandii	-----	-----	-----	NNNNNNNNNN	-----	-----	-----
Pyrgina umbilicata	TCGATGATCG	TCTCCGAGCG	CTAGCTAGCT	CGCTCGCTCG	CCGTCCGTCT	GGCAGGACTC	-----
Glessula ceylanica	-----	-----	-----NN	NNNNNNNNNN	-----	-----	-----
Ceciloides gokweanus	-----C	GGTTGG----	---ATGA-AG	CGAGGTTAGC	CGC--CGAAG	CGCCTTGGGT	-----
Ferussacia folliculus	CGAGTACGCC	ATCTCGTCTA	CTTGCCGTTA	TGTCTGGCAG	GACTCGGCTC	GCTAATTCTGA	-----
Thyrophorella thomensis	TCGATGATCG	TCT--GAGCG	ATAGC-----	---TCGCTCG	CCGTCCGTCT	GGCAGGACTC	-----
Gibbulinella dewinteri	GCGACCCTTC	CCGCCATTTA	GCGGCCCGCC	GAACCGCGGC	AGGACTCGGC	TCGTGATGAC	-----
Gonaxis quadrilateralis	CTCGGGGAAT	TGTTTGNTT	CCCAGCCGNC	CNCACGAAG	AAACCTTTCT	GCATCCNCCG	-----
Gonospora sp.	CAGGGCGGAG	AGCACACAAC	ATCTCCTTTT	TTTCTGTGCA	TCCGCCGGAT	CGTGGCGGGA	-----
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	-----

	370	380	390	400	410	420	
<i>Achatina fulica</i>	GAGGAAGAA-	---GGGTTT	T-----	---TTTCCAC	CC-TT-CCCC	CGTGGCCTCA	
<i>Achatina achatina</i>	AGGAGGAGAG	G-----	GATTTTGGAT	TTTTTCTTCTT	TC-TTCCCC	CGTGGCCTCA	
<i>Achatina stuhlmanni</i>	GAGGAATAAC	T---GGGTTT	TTCCCAG---	--TTTTTTTC	TC-TT-CCCC	CGTGGCCTCA	
<i>Cochlitoma ustulata</i>	GAGGAAGAAG	T---GCTTTT	TTTT-----	-----TCTC	TC-TT-TCCC	CGGGGCTTCC	
<i>Allopeas clavulinum</i>	CAGGACTCGG	CTCG---CTT	TTTCGTGCAT	CGAGCCTACC	AGACCACGTC	CGAA---GAA	
<i>Bocageia</i> sp.	A-CA---CTT	TCCCCGTGGC	CTGAAGTACA	CGTTGCGTCG	TCGTTTGTC	TTTGAGCGGC	
<i>Eutomopeas layardi</i>	CAGG---ACT	CGGCTCGCTT	TTTCTTGCAT	CGAGCCTGCC	AGACCCCGT	CCGAAGAAG	
<i>Leptinaria lamellata</i>	GTTCG---ACC	GTCCGAAGAA	GCGGCCTGCG	CGCAAAATAT	CCGCTAGAGA	GAGAAGGACC	
<i>Paropeas achatinaceum</i>	CAGGACTCGG	CTCG---CTC	AATCTTGCAT	CGAGCCTGCC	AGACCACGTC	CCAA---GAA	
<i>Riebeckia</i> sp.	-----	-----N	NNNNNNNNN-	-----	-----	-----NN	
<i>Rumina decollata</i>	AGGAGGAT-C	G---AGTTTT	CTCT-----	-----TTCCA	CC-TT-CCCT	CGTAGCCTCA	
<i>Subulina octona</i>	A-----	-----	-----	-----TGGCTC	TTTTCTTCCC	CGTGGCTTCA	
<i>Subulina striatella</i>	AGGAGACGTA	TG-----	---TATGCTC	TGTCGTGCTC	GTCTCTTCCC	CGTGGCTTCA	
<i>Subulina vitrea</i>	AAACCGAAGT	TAGCCCGAT-	--GCCTGAAG	TACACGTTCG	GTCGTACAGT	CTTTGAGCGG	
<i>Subulona</i> sp.	-----	-----	-----	-----	-----	-----	
<i>Tortaxis erectus</i>	GCCCACGC--	-CCAACACCA	CGCTCCGCTC	GCCGTCCGTC	TGGCAGGACT	CGGCTCGCTT	
<i>Xerocerastus</i> sp.	-----	-----N	NNNNNNNNN-	-----	-----	-----NN	
<i>Zootecus insularis</i>	T-GC-----	-----	---TTGCT	TTATTTTCTT	CCTTCTTCCC	CGTGGCCTCA	
<i>Coeliaxis blandii</i>	-----	-----N	NNNNNNNNN-	-----	-----	-----NN	
<i>Pyrgina umbilicata</i>	GGCT---CGC	TTTATATTAT	CGCCTCGAGC	CTGCAGACTC	CGTCCAAAGA	AGAAGCGCGC	
<i>Glessula ceylanica</i>	-----	---NNN	NNNNNNNNN-	-----	-----	---NNNN	
<i>Cecilioides gokweanus</i>	CT-----TCG	CGGAGGACGC	GGATACTGTG	CTCGTTATGC	GTATCTGAAA	CCTCCCCGTG	
<i>Ferussacia folliculus</i>	GCCT---GTC	AAAGACAAC	TGTCCGTAAG	CGGCTCGTGT	CGTGACGATG	ATAGTGCCG	
<i>Thyrophorella thomensis</i>	GGCT---CGC	ATTATAATAT	CGCCTCGAGC	CTGCAGACTC	GGTCC---GA	AGAAGCGCGC	
<i>Gibbulinella dewinteri</i>	GCTA---TCG	AGCCTGCCCG	GTCCGTCGAG	CGTTGCGGTG	CCTAACCGCG	GAGGAGAGT	
<i>Gonaxis quadrilateralis</i>	AACC---GTT	GCAGGACTCG	GCTCGTGGTT	GTGTGGGTGG	CGGCGGCGAC	GANGCTGTCC	
<i>Gonospira</i> sp.	CTCG---GCT	CGCGGTGACC	AGAACATCGA	GCCTGCTCGG	TCAGTCTTGT	ACTACGCGCG	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	430	440	450	460	470	480	
<i>Achatina fulica</i>	AGTACACGCA	TGCGCCGTCC	TCGTCATTTC	AGCGACTCTT	TCGCT---CG	CCCAGGAGCC	
<i>Achatina achatina</i>	AGTGCAC-GG	TGCGCCGTCC	-CGTCAGTTT	AGCGACTCTT	TCGCT---TG	CCCAGGAGCC	
<i>Achatina stuhlmanni</i>	AGTACACGCA	TGCGCCGTCC	TCGTCATTTT	AGCGACTCTT	TCGCT---CG	CCCAGGAGCC	
<i>Cochlitoma ustulata</i>	AGTACACCCA	TGCGCCGTCC	TCGGCATTTT	AGCAACTCCT	TCGCTGGTTC	CCCAGGAGCC	
<i>Allopeas clavulinum</i>	GGCGGCTGCG	TTCCATGTCC	TTCCGATCGC	GGTTG-----	-----GAGA	GTGGAGCCGA	
<i>Bocageia</i> sp.	CGGTCTTCGC	TCATCTCATC	CGAAACGCTC	GTGATGTTTC	TTGCTCTGCC	GTCCGTCTGG	
<i>Eutomopeas layardi</i>	CGGCTTGCTT	GCTTGCCTTG	CGTCTGATG	GCGGATGGAG	GGCCGAGATC	GACTTTGGCT	
<i>Leptinaria lamellata</i>	GATGGCGGCG	TCGCCAGGAG	CGCAGGGAAG	AGAGTAAGCT	TTGCTCTCTT	TCCTCTCTCC	
<i>Paropeas achatinaceum</i>	GGCGGCTTGC	TTGCACCGCG	TTCCGATGAG	TGGAT-----	---TATGAGAC	CGGTTTCGCG	
<i>Riebeckia</i> sp.	NNNNNNNN--	-----	-----	-----	-----	-----	
<i>Rumina decollata</i>	AGTACACGAA	TGCGCCGTCC	CCGTCTCTTG	AGCA-----	---TCGCTCT	CTCGAAGCGC	
<i>Subulina octona</i>	AGTACTCGCG	ATGCGT---	-----	---CGCCCA	GTCAATTTT	GTTTGCGGCA	
<i>Subulina striatella</i>	AGTGCTCGAT	GCGTCGT---	-----	-CCAGTCAAT	TTCTTTTTCG	GCGGCACGCT	
<i>Subulina vitrea</i>	TC-----	TCCTTCGCTC	TTCTACGAAG	CCATTCTTCG	TTGCTTGCC	-GTCGCTCT-	
<i>Subulona</i> sp.	-----	-----	-----	-----	-----	-----	
<i>Tortaxis erectus</i>	ACTCGAAACC	AAATC-----	-----	---GAGCCT	GTCGGACCAG	CCCGAATCGG	
<i>Xerocerastus</i> sp.	NNNNNNNN--	-----	---TTGGAAG	ACGAGGACCT	TTGAACCGCC	TTGGCATTGG	
<i>Zootecus insularis</i>	AGTACATGTA	TGCGTCGTCC	CGTCATCTGA	GCGGCTACAT	CGCTCTCTCG	GAGCGCTCAT	
<i>Coeliaxis blandii</i>	NNNNNNNN--	-----	-----	-----	---NNNNNNN	NNN---AAAA	
<i>Pyrgina umbilicata</i>	TTGCTTGCTT	GCTGGCTGTG	CTCGTTACGA	TTGGATTGAG	GGTGAGATCG	ATTTCCGCGG	
<i>Glessula ceylanica</i>	NNNNNN--	-----	-----	-----	-----	-----	
<i>Cecilioides gokweanus</i>	GCCTGAAGTA	CGCCTATGCG	TCGCTTGCCC	GGAGAGCAGC	TCACTATCGT	CCGCTCGCCG	
<i>Ferussacia folliculus</i>	TGCAGTGGGG	TCTGGCTTTG	AGGATGCCAG	GAAGATGTGA	GATGCCACCG	TTTGGCGGCT	
<i>Thyrophorella thomensis</i>	TTGCTTGCTT	TGCTCGATTA	CGGTCG---	-----GAG	GGTGAGATCG	ATTTCCGCGG	
<i>Gibbulinella dewinteri</i>	GGCCTTGCTT	GGAGGCTGCG	CGGTCGCGCA	GCTGCAGAGC	AGCAGCAGAA	TCGTACGATG	
<i>Gonaxis quadrilateralis</i>	CTTCCCATT	NAGCNFTGCT	GGTCGCNNTG	CTGAAANAAG	GGTTNTTTCG	GACAAACGTG	
<i>Gonospira</i> sp.	TTCTTGAAAA	ACGAGAGCTT	GTTTTGTGCG	GTGAGGCGCC	TCCTCTGGAT	TCCTACAGAG	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	

	490 500 510 520 530 540
Achatina fulica	T-CATCTTCG TTT-GTTTCG TCGCCGTCCG TCCGGCAGGA CTCGGGTCGC TTCTTT----
Achatina achatina	C-CGTCTTCG TTC-GTTTCG ----CGTCCG TCCGGCAGGA CTCGGGTCGC TT-----
Achatina stuhlmanni	TCCATCTTCG TTT-GTTTCG TCGCCGTCCG TCCGGCAGGA CTCGGGTCGC TTCTTTTTTT-
Cochlitoma ustulata	T-CATCTTCG TTT-GTTTCG TCGCCGKCCG TCCGGCAGGA CTCGGGTCGC TTCTTTTTTTT
Allopeas clavulinum	TTTCGGCGGC GCCCAGCAAC ATTCATCAAT TTTCAA----
Bocageia sp.	CAGGACTCGG CTCGCTTAAT GACTCAAATC GAGCCTGCTG GCGCGTCTGA AAACGGCACC
Eutomoepas layardi	GCGCCAGAAA CATATGTATT TTTAA-----
Leptinaria lamellata	GTATTCTTTT GATTTTCTCT TCCA-----
Paropeas achatinaceum	AGGCCAGAAA TTTTTTTTGT TTTTACACTT TTTCAA----
Riebeckia sp.	-----
Rumina decollata	T-CATCTTCG TTC-GCTCGC CGTCCGTCTG ---GCAGGA CTCGGGTCGC TGTG-----
Subulina octona	CGCTCTTTCG TTCGCTCGCC CGTCCGTCT- ---GGCAGGA CTCGGGTCGC TTAGTTTT--
Subulina striatella	CTTTCGTTTCG CTCGCCGTC CGTCCGTCC- ---GGCAGGA CTCGGGTCGC TTTATTTT--
Subulina vitrea	---GGCAGGA CTCGGCTCGC TGTGAGACAG ACAAACACTGT ATCGAGCCTG CCGGTGCAAT
Subulona sp.	-----
Tortaxis erectus	CTGTGCTGTG TGGTGTATCA AGGAGTGC GG CCTGTCCGC GCGGTGCCA AAATTTATCC
Xerocerastus sp.	AGGATTTGAG GGAAGGATTA ATTTCTCCTC CTCATTTCCC CGTTGCCTCA AGTGCACGCA
Zootecus insularis	CTTCGTTCGC TCGCCGTC CG TCTGGCAGGA CTCGGCTCGC TGTGGGAACA TAAGCACAAA
Coelioxaxis blandii	AAACGGCTTG CGGATGCAAT GATGTTAAAA AGGCCTGCGA CGGTAAGGGT CCGGCCCCAA
Pyrgina umbilicata	CGCTCTCATT TTTAAAAATAT CAA-----
Glessula ceylanica	-----GAG GGAAATAAAA GCGTTTGCCG
Cecilioides gokweanus	TCCGTCTGGC CGGACTCGGC TCGCTTCGTA AACGAACACA ATCGAGCCGG CCTGACAGTC
Ferussacia folliculus	TCGTT-----
Thyrophorella thomensis	TGCTCATTAT TGTGTCCTCT TCTTTCAA--
Gibbulinella dewinteri	AT-----
Gonaxis quadrilateralis	CGTGCGCCCA CAACAAGGTG GCAGAGGCAC AAAAGTGNNT GCTTGA----
Gonospora sp.	AA-----
NUCLEOTIDES INCLUDED	-----

	550 560 570 580 590 600
Achatina fulica	-----AACAA ATCGAGCCTG CCAGAC---C GTCCGAAGCG GCTAGGG-AT TG-CGAAGTG
Achatina achatina	-----TT ATCGAGCCTG CCAGAC---T GTCCGAAGCG GCTAGCG-GT TG-CGAAGTG
Achatina stuhlmanni	-----A ATCGAGCCTG CCAGAC---C GTCCGAAGCG GCTAGCG-AT TG-CGAAGTG
Cochlitoma ustulata	T-----A ATCGAGCCTG CCAGAC---C GTCCGAAGCG GCTAGCGGAT TG-CGAAGTG
Allopeas clavulinum	-----
Bocageia sp.	CGGTATGAAC AGTGAGGAGA GAGATGATTA GCGCTGTTGC CCGCTGCGAC GCGCCAGGC
Eutomoepas layardi	-----
Leptinaria lamellata	-----
Paropeas achatinaceum	-----
Riebeckia sp.	-----
Rumina decollata	-----AATAA AACGAGCCTG CCGAC---C GTCCGAAGTG GCATGCGGTT TCGAAGTGTG
Subulina octona	-----CA CTCGAGCCTG TCAGTA--CT GTCCGAAACG GCGA----- --AGGACGG
Subulina striatella	----- CTCGAGCCTG TCGGTAC--T GTCCGAAACG GCGAGTGGGG TGGGTGACGG
Subulina vitrea	GTCCGAAGCG GCTAGCGCTT ATTTGATGAG GTTATGGCCG AGAGAAAGAA GTGTGTGGGA
Subulona sp.	-----
Tortaxis erectus	ATTTGAAC--
Xerocerastus sp.	TGGTCGTCCC GTCATCTGAG CCGCTACATC GCTCTCTCTC GGAGCGCTCA TCTTCGTTCCG
Zootecus insularis	TCGAGCCTGC CGACCGTCCG AAGCGGCTTG CTGTATCGAG GAGAGCGTGA GAGGGCGGGA
Coelioxaxis blandii	GGAGGGCAAG ACTTCGTCTC GTTTTCATA--
Pyrgina umbilicata	-----
Glessula ceylanica	CGACGGTGCC CACCAAGTTT TTATTCACTA
Cecilioides gokweanus	CGTGAACGGC TTGCGGCGTT CGAGTTGCCA ATCGACGCCA CTAATCCGCA GCAAATTTTG
Ferussacia folliculus	-----
Thyrophorella thomensis	-----
Gibbulinella dewinteri	-----
Gonaxis quadrilateralis	-----
Gonospora sp.	-----
NUCLEOTIDES INCLUDED	-----

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      ....|.....|.....|.....|.....|.....|.....|.....|.....|.....|
      610      620      630      640      650      660
Achatina fulica      GGG-----CG CGCAAGCGCA TGGG----TC TGCTGCGGCG GCGCCA----
Achatina achatina    GGT---GGCG GGCAAGCG-A TCGG---TC T---GCGGCG GCGCCAGCA-
Achatina stuhlmanni  GGG---CGCC CGCGAGCGGA TTGGGGGGTC TGCTGTGGCG GCGCCA----
Cochlitoma ustulata  GGG--CGCGC GAGCGCA--- TGGG----TC TGTGCGGCG GCGCCA----
Allopeas clavulinum  -----
Bocageia sp.         AGTTTTTGTA TTTTTCATA -----
Eutomopeas layardi   -----
Leptinaria lamellata -----
Paropeas achatinaceum -----
Riebeckia sp.        TACTACATAA ACCGTTTCGC A-----
Rumina decollata     AGAGGGCGAT TGCCGCGGCG GCGCCAATTT TTTTGTGTC GTTCTAAA--
Subulina octona      AGGCAGTCGA GAGCCGCAAC GACGACGGCG CCCCCTGGT- -AATGTGTCA
Subulina striatella  AAGCAGTCGA TGGCTAGCGA AAGCGACGGC GCCCGTGGTC TATTACCGT ACGTCGTCGT
Subulina vitrea      CCGCGACGTC GCCTATTTGT AAAAAAATT TGTATTATT TCTTTAC--
Subulona sp.         -----
Tortaxis erectus     -----
Xerocerastus sp.     CTCGCCGTCC GTCCGGCAGG ACTCGGCTCG CTGTGAAAAC AAAGCACACT ATTCGAGCCT
Zootecus insularis  TTGCTGCGGC GCGCCAACT TTTGTTTTT CTTCTAAA-
Coeliaxis blandii    -----
Pyrgina umbilicata   -----
Glessula ceylanica   -----
Cecilioides gokweanus -----
Ferussacia folliculus -----
Thyrophorella thomensis -----
Gibbulinella dewinteri -----
Gonaxis quadrilateralis -----
Gonospira sp.        -----
NUCLEOTIDES INCLUDED -----

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      ....|.....|.....|.....|.....|.....|.....|.....|.....|.....|
      670      680      690      700      710      720
Achatina fulica      GTCTTTCT-- -----AAA ATCTTTCTTT A-----
Achatina achatina    GTCTCTATAA CGACGACGTT TNNTTTAAAA TTTTA----
Achatina stuhlmanni  GTCTTTCT-- -----AAA ATTTTCTTTT A-----
Cochlitoma ustulata  GTCTTTCT-- ----- TTCTTTCTTT A-----
Allopeas clavulinum  -----
Bocageia sp.         -----
Eutomopeas layardi   -----
Leptinaria lamellata -----
Paropeas achatinaceum -----
Riebeckia sp.        -----
Rumina decollata     -----
Subulina octona      TACTATTATT T-----
Subulina striatella  ACGAATCATT T-----
Subulina vitrea      -----
Subulona sp.         -----
Tortaxis erectus     -----
Xerocerastus sp.     GCC-GACCCC GTCCGAAGCG GCTTGCGGTT TTGAAGTTG AGGCGGCGC CGCCCCGCTC
Zootecus insularis  -----
Coeliaxis blandii    -----
Pyrgina umbilicata   -----
Glessula ceylanica   -----
Cecilioides gokweanus -----
Ferussacia folliculus -----
Thyrophorella thomensis -----
Gibbulinella dewinteri -----
Gonaxis quadrilateralis -----
Gonospira sp.        -----
NUCLEOTIDES INCLUDED -----

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	730	740	750	760	770	780
Achatina fulica	-----	-----	-----	-----	-----	-----T-
Achatina achatina	-----	-----	-----	-----	-----	-----T-
Achatina stuhlmanni	-----	-----	-----	-----	-----	-----T-
Cochlitoma ustulata	-----	-----	-----	-----	-----	-----T-
Allopeas clavulinum	-----	-----	-----	-----	-----	-----T-
Bocageia sp.	-----	-----	-----	-----	-----	-----T-
Eutomopeas layardi	-----	-----	-----	-----	-----	-----T-
Leptinaria lamellata	-----	-----	-----	-----	-----	-----T-
Paropeas achatinaceum	-----	-----	-----	-----	-----	-----T-
Riebeckia sp.	-----	-----	-----	-----	-----	-----T-
Rumina decollata	-----	-----	-----	-----	-----	-----T-
Subulina octona	-----	-----	-----	-----	-----	-----C-
Subulina striatella	-----	-----	-----	-----	-----	-----C-
Subulina vitrea	-----	-----	-----	-----	-----	-----T-
Subulona sp.	-----	-----	-----	-----	-----	-----T-
Tortaxis erectus	-----	-----	-----	-----	-----	-----T-
Xerocerastus sp.	GAGAGGTGGA	GGAGGGCGAT	TGCTGCCCGG	GAGGCGCCAG	TATTTANTAT	TTTCTAAAT-
Zootecus insularis	-----	-----	-----	-----	-----	-----T-
Coelioxaxis blandii	-----	-----	-----	-----	-----	-----T-
Pyrgina umbilicata	-----	-----	-----	-----	-----	-----T-
Glessula ceylanica	-----	-----	-----	-----	-----	-----T-
Cecilioides gokweanus	-----	-----	-----	-----	-----	-----T-
Ferussacia folliculus	-----	-----	-----	-----	-----	-----T-
Thyrophorella thomensis	-----	-----	-----	-----	-----	-----T-
Gibbulinella dewinteri	-----	-----	-----	-----	-----	-----T-
Gonaxis quadrilateralis	-----	-----	-----	-----	-----	-----TN
Gonospira sp.	-----	-----	-----	-----	-----	-----T-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----m-

					
	790	800	810	820	830	840
Achatina fulica	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Achatina achatina	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Achatina stuhlmanni	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Cochlitoma ustulata	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Allopeas clavulinum	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Bocageia sp.	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Eutomopeas layardi	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Leptinaria lamellata	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Paropeas achatinaceum	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Riebeckia sp.	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Rumina decollata	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Subulina octona	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Subulina striatella	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Subulina vitrea	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Subulona sp.	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Tortaxis erectus	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Xerocerastus sp.	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Zootecus insularis	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Coelioxaxis blandii	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Pyrgina umbilicata	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Glessula ceylanica	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Cecilioides gokweanus	CCGACCTCAG	TTCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Ferussacia folliculus	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Thyrophorella thomensis	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Gibbulinella dewinteri	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Gonaxis quadrilateralis	CCGACCTCAG	ATCGGACGAG	ATTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
Gonospira sp.	CCGACCTCAG	ATCGGACGAG	ACTACCCGCT	GAATTTAAGC	ATATAACTAA	GCGGAGGAAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	850	860	870	880	890	900
<i>Achatina fulica</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Achatina achatina</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Achatina stuhlmanni</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Cochlitoma ustulata</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Allopeas clavulinum</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Bocageia</i> sp.	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Eutomopeas layardi</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Leptinaria lamellata</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Paropeas achatinaceum</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Riebeckia</i> sp.	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Rumina decollata</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Subulina octona</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Subulina striatella</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Subulina vitrea</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Subulona</i> sp.	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Tortaxis erectus</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Xerocerastus</i> sp.	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Zootecus insularis</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Coeliaxis blandii</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Pyrgina umbilicata</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Glessula ceylanica</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Cecilioides gokweanus</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Ferussacia folliculus</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Thyrophorella thomensis</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAATAGCC	CAGCACCGAA
<i>Gibbulinella dewinteri</i>	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAAGAGCC	CAGCACCGAA
<i>Gonaxis quadrilateralis</i>	AGAAACTAAC	AAGGATTTCC	CCAATAACGG	CGAGTGAAGC	GGGATAGGCC	CAGCACTGAA
<i>Gonospira</i> sp.	AGAAACTAAC	AAGGATTTCC	CCAGTAACGG	CGAGTGAAGC	GGGAACGGCC	CAGCACCGAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	910	920	930	940	950	960
<i>Achatina fulica</i>	TCCCTCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Achatina achatina</i>	TCCCTCAGTG	TCACGCTGAC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Achatina stuhlmanni</i>	TCCCTCAGTG	TCACGCTGAC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Cochlitoma ustulata</i>	TCCCTCAGTG	TCACGCTGAC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Allopeas clavulinum</i>	TCCCCAGTG	TCACGCTGGC	GGGAAATGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Bocageia</i> sp.	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Eutomopeas layardi</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Leptinaria lamellata</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Paropeas achatinaceum</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Riebeckia</i> sp.	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Rumina decollata</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Subulina octona</i>	TCCCTCAGTG	TCACGCTGAC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Subulina striatella</i>	TCCCTCAGTG	TCACGCTGAC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Subulina vitrea</i>	TCCCTCAGTG	TCACGCTGAC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Subulona</i> sp.	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Tortaxis erectus</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Xerocerastus</i> sp.	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Zootecus insularis</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Coeliaxis blandii</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Pyrgina umbilicata</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Glessula ceylanica</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Cecilioides gokweanus</i>	TCCCTCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Ferussacia folliculus</i>	TCCCTCAGTG	TCACGCTGAC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Thyrophorella thomensis</i>	TCCCCAGTG	TCACGCTGGC	GGGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Gibbulinella dewinteri</i>	TCCCTCAGTG	TCACGCTGAC	GGGAAATGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Gonaxis quadrilateralis</i>	TCCCTCAGTG	TCACGCTGAC	GGGAACTTTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
<i>Gonospira</i> sp.	TCCCTCAGTG	TCACGCTGAC	GGAACTGTG	GTGTGTGGGA	CGCCACCAGT	CGCATCAGAG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	970	980	990	1000	1010	1020		
<i>Achatina fulica</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Achatina achatina</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Achatina stuhlmanni</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Cochlitoma ustulata</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Allopeas clavulinum</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Bocageia sp.</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Eutomopeas layardi</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Leptinaria lamellata</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Paropeas achatinaceum</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Riebeckia sp.</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Rumina decollata</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Subulina octona</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Subulina striatella</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Subulina vitrea</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Subulona sp.</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Tortaxis erectus</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Xerocerastus sp.</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Zootecus insularis</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Coelioxaxis blandii</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Pyrquina umbilicata</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Glessula ceylanica</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Ceciloides gokweanus</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Ferussacia folliculus</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Thyrophorella thomensis</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Gibbulinella dewinteri</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Gonaxis quadrilateralis</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
<i>Gonospira sp.</i>	GGCGTCGAAG	TCCTCCTGAT	CGGGGCTTCA	CCCAGAGCGG	GTGTAAGGCC	TTTGCAGGCC		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	1030	1040	1050	1060	1070	1080		
<i>Achatina fulica</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Achatina achatina</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Achatina stuhlmanni</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Cochlitoma ustulata</i>	CCACTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Allopeas clavulinum</i>	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Bocageia sp.</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Eutomopeas layardi</i>	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Leptinaria lamellata</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Paropeas achatinaceum</i>	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Riebeckia sp.</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Rumina decollata</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Subulina octona</i>	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Subulina striatella</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Subulina vitrea</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Subulona sp.</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Tortaxis erectus</i>	CCACTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Xerocerastus sp.</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Zootecus insularis</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Coelioxaxis blandii</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Pyrquina umbilicata</i>	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Glessula ceylanica</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Ceciloides gokweanus</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Ferussacia folliculus</i>	CCTCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Thyrophorella thomensis</i>	CCCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Gibbulinella dewinteri</i>	CTCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Gonaxis quadrilateralis</i>	CTCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
<i>Gonospira sp.</i>	CTCCTCTGTG	CGGCCGCGAG	CGTCTCAGGA	GTCGGGTTGT	TTGGGAATGC	AGCCCAAAGC		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm		

	1090	1100	1110	1120	1130	1140
Achatina fulica	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
Achatina achatina	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
Achatina stuhlmanni	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
Cochlitoma ustulata	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
Allopeas clavulinum	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Bocageia sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
Eutomopeas layardi	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Leptinaria lamellata	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Paropeas achatinaceum	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Riebeckia sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Rumina decollata	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-TGCACGAG	TCCGATAGCG	GACAAGTACC
Subulina octona	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Subulina striatella	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Subulina vitrea	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Subulona sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Tortaxis erectus	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	G-TGCACGAG	TCCGATAGCG	GACAAGTACC
Xerocerastus sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
Zootecus insularis	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
Coeliaxis blandii	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Pyrgina umbilicata	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Glessula ceylanica	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	G-TGCACGAG	TCCGATAGCG	GACAAGTACC
Cecilioides gokweanus	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Ferussacia folliculus	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	G-TGCACGAG	TCCGATAGCG	GACAAGTACC
Thyrophorella thomensis	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	C-GGCACGAG	TCCGATAGCG	GACAAGTACC
Gibbulinella dewinteri	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	TTTGCACGAG	TCCGATAGCG	GACAAGTACC
Gonaxis quadrilateralis	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
Gonospira sp.	GGGTGGTAAA	CTCCATCTAA	GGCTAAATAC	T-GGCACGAG	TCCGATAGCG	GACAAGTACC
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	m-mmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1150	1160	1170	1180	1190	1200
Achatina fulica	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Achatina achatina	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Achatina stuhlmanni	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Cochlitoma ustulata	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Allopeas clavulinum	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Bocageia sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Eutomopeas layardi	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Leptinaria lamellata	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Paropeas achatinaceum	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Riebeckia sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Rumina decollata	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Subulina octona	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Subulina striatella	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Subulina vitrea	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Subulona sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Tortaxis erectus	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Xerocerastus sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Zootecus insularis	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Coeliaxis blandii	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Pyrgina umbilicata	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Glessula ceylanica	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Cecilioides gokweanus	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Ferussacia folliculus	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Thyrophorella thomensis	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCTCAG
Gibbulinella dewinteri	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Gonaxis quadrilateralis	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
Gonospira sp.	GTGAGGGAAA	GTTGAAAAGA	ACTTTGAAGA	GAGAGTTCAA	GAGTACGTGA	AACCGCCCAG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1210 1220 1230 1240 1250 1260
<i>Achatina fulica</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCTCGG
<i>Achatina achatina</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCTCGG
<i>Achatina stuhlmanni</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Cochlitoma ustulata</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCTCGG
<i>Allopeas clavulinum</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Bocageia sp.</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Eutomopeas layardi</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Leptinaria lamellata</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Paropeas achatinaceum</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Riebeckia sp.</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Rumina decollata</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Subulina octona</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Subulina striatella</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Subulina vitrea</i>	AGGTAAACGG GTGGATCCGC AAATTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Subulona sp.</i>	AGGTAAACGG GTGGATCCGC AAATTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Tortaxis erectus</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCT CGCGGAATTC AGCGCGGCGC GCGGCCTGTG
<i>Xerocerastus sp.</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Zootecus insularis</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Coeliaxis blandii</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Pyrquina umbilicata</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Glessula ceylanica</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCT CGCGGAATTC AGCGCGGCGC GCGATCTGTG
<i>Ceciloides gokweanus</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCT CGCGGAATTC AGCGCGGCGC GCGGTTT-GG
<i>Ferussacia folliculus</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Thyrophorella thomensis</i>	AGGTAAACGG GTGGATCCGC AAAGTCGGCC CGCGGAATTC AGCGCGGCGC GCGGCCT-GG
<i>Gibbulinella dewinteri</i>	AGGTAAACGG GTGGATCCGC AATGTCGGCC CGCGGAATTC AGCGCGGCGT GCGGCCT-GG
<i>Gonaxis quadrilateralis</i>	AGGTAAACGG GTGGATCCGC AATGTCGGCC CGCGGAATTC AGCGCGGCGT GCGGCCT-GG
<i>Gonospira sp.</i>	AGGCAAACGG GTGGATCCGC AATGTCGGCC CGCGGAATTC AGCGCGGCGT GCGGCTC-GG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmm-m

	1270 1280 1290 1300 1310 1320
<i>Achatina fulica</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Achatina achatina</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Achatina stuhlmanni</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Cochlitoma ustulata</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Allopeas clavulinum</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Bocageia sp.</i>	GGCGACTGCG GCCGGGGATC CCTGGGACCC CGCTGCGGTG TCGATCCGGG TTCCGCCCGG
<i>Eutomopeas layardi</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Leptinaria lamellata</i>	GGCGATTGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Paropeas achatinaceum</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Riebeckia sp.</i>	GACTATCGCA GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Rumina decollata</i>	GGCTATTGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Subulina octona</i>	GGCGATCGCT GCCGGGGATC CCTGGGACCC CGCTGTGGTG TCGATCCGGG CTCCGCCCGG
<i>Subulina striatella</i>	GGCGATCGCT GCCGGGGATC CCTGGGACCC CGCTGTGGTG TCGATCCGGG CTCCGCCCGG
<i>Subulina vitrea</i>	GGCGACTGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Subulona sp.</i>	GGCGACTGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Tortaxis erectus</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Xerocerastus sp.</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Zootecus insularis</i>	GGCGATCGCG GCCGGGGATC CCTGGGACT CGCCGTGGTG TCGAACCAGG CTCCGCCCGG
<i>Coeliaxis blandii</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGTGC TCGATCCGGG CTCCGCCCGG
<i>Pyrquina umbilicata</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Glessula ceylanica</i>	GGCGATTGCG GCCGGGGATC CCTGGGACCC CGCCGTGGTG TCGAACCAGG TTCCGCCCGG
<i>Ceciloides gokweanus</i>	GGCGATTGCG GCCGGGGATC CCTGGGACCC CGCCGTGGTG TCGAACCAGG CTCCGCCCGG
<i>Ferussacia folliculus</i>	GACAATCGTG GCCGGGGATT CTTGGGACCC CGCTTCGGTG TCGAGCCGGG CTCCGCCCGG
<i>Thyrophorella thomensis</i>	GGCGATCGCG GCCGGGGATC CCTGGGACCC CGCCGCGGTG TCGATCCGGG CTCCGCCCGG
<i>Gibbulinella dewinteri</i>	GGCCGTGCGT GCAAGGGATC CCTGGGACCC TGCCGCGATG TCGAGCCCGG CCCC GCCCGG
<i>Gonaxis quadrilateralis</i>	GACCGTGTG GCCAGGGATC CGTGGGACCC TGCCGCGATG TCGAGCTGGG CCCC GCCCGG
<i>Gonospira sp.</i>	GGACGTGCGT GCCAGGGATC CCTGGGACCC TGCCGCGATG TCGAGCTGGG CCCC GCCCGG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	1330	1340	1350	1360	1370	1380		
Achatina fulica	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--GCCGTC	AGAGGCCGGG		
Achatina achatina	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--GCCGTC	AGAGGCCGGG		
Achatina stuhlmanni	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--GCCGTC	AGAGGCCGGG		
Cochlitoma ustulata	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--GCCGTC	AGAGGCCGGG		
Allopeas clavulinum	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GTCGTC	AGAGGCCGGG		
Bocageia sp.	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	G---GCCGTC	AGAGGCCGGG		
Eutomopeas layardi	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GTCGTC	AGAGGCCGGG		
Leptinaria lamellata	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GCCGTC	AGAGGCCGGG		
Paropeas achatinaceum	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GTCGTC	AGAGGCCGGG		
Riebeckia sp.	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----TCCGTC	AGAGGCCGGG		
Rumina decollata	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--GCCGTC	AGAGGCCGGG		
Subulina octona	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GT--GCCGTC	AGAGGCCGGG		
Subulina striatella	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GT--GTCGTC	AGAGGCCGGG		
Subulina vitrea	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GG--GCCGTC	AGATGCCGGG		
Subulona sp.	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GG--GCCGTC	AGATGCCGGG		
Tortaxis erectus	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--GCCGTC	AGCAGCCGGG		
Xerocerastus sp.	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--GCCGTC	AGAGGCCGGG		
Zootecus insularis	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--GCCGTC	AGAGGCCGGG		
Coeliaxis blandii	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----TCTGTC	AGAGGCCGGG		
Pyrgina umbilicata	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GTCGTC	AGAGGCCGGG		
Glessula ceylanica	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GT--GCCGTC	AGAGGCCGGG		
Cecilioides gokweanus	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GCCGCC	AGAGGCCGGG		
Ferussacia folliculus	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GA--ACGCTC	AGAAGTCCGG		
Thyrophorella thomensis	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GTCGTC	AGAGGCCGGG		
Gibbulinella dewinteri	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GCCGTC	AAAAGCCGGG		
Gonaxis quadrilateralis	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	GAAGGCCGTC	AAAAGCCGGG		
Gonospira sp.	TGCACTTTCC	GCGGGCAGAG	TGCCACAACC	GGTTCGCTG	----GCCGTC	AAAAGCCGGG		
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	----mmmmmm	mmmmmmmmmm		

	1390	1400	1410	1420	1430	1440	
Achatina fulica	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Achatina achatina	C-GAGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Achatina stuhlmanni	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Cochlitoma ustulata	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Allopeas clavulinum	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Bocageia sp.	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Eutomopeas layardi	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Leptinaria lamellata	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Paropeas achatinaceum	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Riebeckia sp.	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Rumina decollata	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Subulina octona	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Subulina striatella	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Subulina vitrea	T-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Subulona sp.	AAGGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---AACTCGG	CGGACGGCCT	
Tortaxis erectus	T-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCAACGG	CGGACGGCCT	
Xerocerastus sp.	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Zootecus insularis	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Coeliaxis blandii	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	GCCGCCCGG	CGGACGGCCT	
Pyrgina umbilicata	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Glessula ceylanica	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GTTACGG	CGGACGGCCT	
Cecilioides gokweanus	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Ferussacia folliculus	T-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCTGAGG	CGGACGGCCT	
Thyrophorella thomensis	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Gibbulinella dewinteri	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Gonaxis quadrilateralis	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---GCCCCGG	CGGACGGCCT	
Gonospira sp.	C-GGGTTGTC	GGTGGGGGCG	CTTGC-GTCC	CTACCAGCCC	---TCCCCGG	CGGACGGCCT	
NUCLEOTIDES INCLUDED	m-mmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	---mmmmmm	mmmmmmmm	

	1450 1460 1470 1480 1490 1500
<i>Achatina fulica</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTACCCG GCCCCTCTGC GCGAGTCCGA
<i>Achatina achatina</i>	-CGGGACCGA GGGACCGCCG -CGCGCTTCG AGGCTACCCG ACCCCTCCGC GCGAGTCCGA
<i>Achatina stuhlmanni</i>	-CGGGACCGA GGAACTGCCG -CGCGCTTCG AGGCTACCCG ACCCCTCCGC GCGAGTCCGA
<i>Cochlitoma ustulata</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTACCCG ACCCCTCCGC GCGAGTCCGA
<i>Allopeas clavulinum</i>	-CGGGACCGA GGAACCGCCG -CGCACTTCG AGGCCGCCT- GCCCCTCCGC GCGAGTTCGA
<i>Bocageia sp.</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTACCT- GCCCCTCCGC GCGAGTTCGA
<i>Eutomopeas layardi</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCCGCCT- GCCCCTCCGC GCGAGTTCGA
<i>Leptinaria lamellata</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCCGCCT- GCCCCTCCGC GCGAGTTCGA
<i>Paropeas achatinaceum</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCCGCCT- GCCCCTCCGC GCGAGTTCGA
<i>Riebeckia sp.</i>	-TGGGATCGA GGAACCGCCG -CGCGCTTCG AGGCTACCT- GTCCTCCGC GCGAGTTCGA
<i>Rumina decollata</i>	-CGGGACCGA GGAACCGCCG -CACGCTTCG AGGCTACCT- GTCCTCCGC GCGAGTTCGA
<i>Subulina octona</i>	-CGGGACCGA GGAACTGCCG -CGCGCTTCG AGGCTACCC- GTCCTCCGC GCGAGTTCGA
<i>Subulina striatella</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTACCC- GTCCTCCGC GCGAGTTCGA
<i>Subulina vitrea</i>	-CGGGACCGA GGAACCGCCG -CGTGCTTCG AGGCTACTC- GTCCTCCGC GTGAGTTCGA
<i>Subulona sp.</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTGCTC- GTCCTCCGC GCGAGTTCGA
<i>Tortaxis erectus</i>	-CGGGACCGA GGAACTGCCG -CGCGCTTCG AGGCTACCT- GCCCCTCCGC ACGAGTTCGA
<i>Xerocerastus sp.</i>	-CGGGACCGA GGGACCGCCG -CACGCTTCG AGGCTACCT- GTCCTCAGC GCGAGTCCGA
<i>Zootecus insularis</i>	-CGGGACCGA GGGACCGCCG -CGCGCTTCG AGGCTACCT- GTCCTCCGC GCGAGTTCGA
<i>Coeliaxis blandii</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTACCT- GTCCTCCGC GCGAGTCCGA
<i>Pyrquina umbilicata</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTACCT- GTCCTCCGC GCGAGTTCGA
<i>Glessula ceylanica</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTACCT- GTCCTCCGC ACGAGTTCGA
<i>Ceciloides gokweanus</i>	-CGGGACCGA GGAACAGCCG -CGCGCTTCG AGGCCACAT- GCCCCTCCGT GCGAGTCCGA
<i>Ferussacia folliculus</i>	-TGGGACCGA GGAACAGCCG TCGCGCTTCG AGGCTACCT- GTCCTCCGT GTGAGTCAA
<i>Thyrophorella thomensis</i>	-CGGGACCGA GGAACCGCCG -CGCGCTTCG AGGCTGCTC- GCCCCTCCGC GCGAGTTCGA
<i>Gibbulinella dewinteri</i>	GCCAGACCGA GGAACCGCCG -CTCGCTTCG AGGCTACCC- GTCCTCAGG GCGAGTTCGA
<i>Gonaxis quadrilateralis</i>	-CCGGACCGA GGAACCGCCG -CTCGCTTCG AGGCTACCC- GTCCTCCGC GTGAGTTCGA
<i>Gonospira sp.</i>	-CCGGACCGA GGAACCGCCG -CTCGCTTCG AGGCTGCTC- GTCCTCAGG GTGAGTTCGA
NUCLEOTIDES INCLUDED	-mmmmmmmmmm mmmmmmmmmmm -mmmmmmmmmm mmmmmmmmmmm -mmmmmmmmmm mmmmmmmmmmm

	1510 1520 1530 1540 1550 1560
<i>Achatina fulica</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGCGACCGGG CC-GGGCTAG
<i>Achatina achatina</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG TGCGACCGGG CC-GGGCTAG
<i>Achatina stuhlmanni</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CAAGACCGGG CC-GGGCTAG
<i>Cochlitoma ustulata</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGCGACCGGG CC-GGGCTAG
<i>Allopeas clavulinum</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG TGCGACCGGG ATCGGGCCGG
<i>Bocageia sp.</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGTGATCGGG TT-GGGCTAG
<i>Eutomopeas layardi</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG TGCGACCGGG ATCGGGCCGG
<i>Leptinaria lamellata</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG -CTGACAGGG ATCGGGCCGG
<i>Paropeas achatinaceum</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG TGCGACCGGG ATCGGGCCGG
<i>Riebeckia sp.</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCT CGCGATCGGG TTGGGGCTAG
<i>Rumina decollata</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGCGACCGGG CC-GGGCTAG
<i>Subulina octona</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG TGCGACCGGG CC-GGGATAG
<i>Subulina striatella</i>	CTGGGAGAGA CCGGGCAACC GTGTCTTCGG ACCGCTCGCG TGTGATCGGG CC-GGGATAG
<i>Subulina vitrea</i>	CCGGGAGAGA CTGGGCAACC GTGTCTTCGG ACTGCTCGCG CGCGACCGGG TT-GGGCTAG
<i>Subulona sp.</i>	CCGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGCGATTGGG TT-GGGCTAG
<i>Tortaxis erectus</i>	CTGGAAGAGA CTGGGTAACC GTGGCTTCGG AATGCTCGAG CGCGATCGGG CC-GGGCTAG
<i>Xerocerastus sp.</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGCGACCGAG CC-GGGCTAG
<i>Zootecus insularis</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGCGACCGGG CC-GGGCTAG
<i>Coeliaxis blandii</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCT CGCGATCGGG TTGGGGTTAG
<i>Pyrquina umbilicata</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG TGCGACCGGG ATCGGGCCGG
<i>Glessula ceylanica</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG AATGCTCGAG CGTGACTGGG TC-GGGCTAA
<i>Ceciloides gokweanus</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGCGACCGGG ATCGTGTCCG
<i>Ferussacia folliculus</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCA CGCGACCGGG TC-GGGCTAG
<i>Thyrophorella thomensis</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCGCG CGCGACCGGG ATCGGGCCGG
<i>Gibbulinella dewinteri</i>	CTGGAAGAGA CTGGGCAACC GTGACTTCGG ACCGCTCATC CTCGATCGGG CC-GGGCTAG
<i>Gonaxis quadrilateralis</i>	CAGGAAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCATC CTCGACCGGG CC-GGGCTAG
<i>Gonospira sp.</i>	CTGGGAGAGA CTGGGCAACC GTGTCTTCGG ACCGCTCATC CTCGATCGGG CC-GGGCCAG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mm-mmmmmmm

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1570      1580      1590      1600      1610      1620
Achatina fulica      CCGGGCGT-C GCG----ACT GTAA---CAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Achatina achatina   CCGGGCGC-- G-----ACT GAAG---CAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Achatina stuhlmanni CCGGGCGC-C GCG----ACT GTAA---CAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Cochlitoma ustulata CCGGGCGT-C GCG----ACT GTAA---CAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Allopeas clavulinum CCGGGTGC-C GCTTGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Bocageia sp.        CCGGGTGC-- GTTAGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Eutomopeas layardi  CCGGGCGC-C GCTTGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Leptinaria lamellata CCGGGCGC-- GCACGC-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Paropeas achatinaceum CCGGGCGC-C GCTTGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Riebeckia sp.       CCGGGCGC-C GCCAGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Rumina decollata    CCGGGCGT-- GCACA-----CGG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Subulina octona     CCGGGCGCGC GCACGCA-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Subulina striatella CCGGGCGCGC GCATGCAG-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Subulina vitrea     CCGGGCAC-- GCCCGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Subulona sp.        CCGGGCGC-- GCCCGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Tortaxis erectus    CCGGGCGC-- GCAACGCTT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Xerocerastus sp.    CCGGGCGT-- GCACA-----CTG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Zootecus insularis  CCGGGCGT-- GCACA-----CGG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Coeliaxis blandii   CCGGGCGC-C GCCAGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Pyrgina umbilicata  CCGGGCGC-C GCTCGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Glessula ceylanica  CCGGGCGT-- GCATTT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Cecilioides gokweanus CCGGGTGC-- GTTCGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Ferussacia folliculus CCGGGCGC-- GCGTCAGT-----GAG GGTGGTGGC AAGTCTGTCTG GCATTCCACC
Thyrophorella thomensis CCGGGCGC-C GCTCGT-----GAG GGTGGTGGC GAGTCCGTCG GCATTCCACC
Gibbulinella dewinteri CCGGGCGA-- GCCGATTTG-----GAG GGTGGTGGC GAATCTGTCTG GCATTCCACC
Gonaxis quadrilateralis CCGGGCGA-- GCCTGTCTG-----GAG GGTGGTGGC GAATCTGTCTG GCATTCCACC
Gonospira sp.       CCGGGCGA-- GCCGATTTG-----GAG GGTGGTGGC GAATCTGTCTG GCATTCCACC
NUCLEOTIDES INCLUDED mmmmmmmmm-- mm----- mmm mmmmmmmmm mmmmmmmmm mmmmmmmmmmm

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1630      1640      1650      1660      1670      1680
Achatina fulica      CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Achatina achatina   CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Achatina stuhlmanni CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Cochlitoma ustulata CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Allopeas clavulinum CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Bocageia sp.        CGACCCGTCT TGA AACACCGG GACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Eutomopeas layardi  CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Leptinaria lamellata CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Paropeas achatinaceum CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Riebeckia sp.       CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Rumina decollata    CGACCCGTCT TGA AANNNNN -NCCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Subulina octona     CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Subulina striatella CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Subulina vitrea     CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Subulona sp.        CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Tortaxis erectus    CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Xerocerastus sp.    CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Zootecus insularis  CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Coeliaxis blandii   CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Pyrgina umbilicata  CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Glessula ceylanica  CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Cecilioides gokweanus CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Ferussacia folliculus CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Thyrophorella thomensis CGACCCGTCT TGA AANNNNN -NCCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Gibbulinella dewinteri CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Gonaxis quadrilateralis CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
Gonospira sp.       CGACCCGTCT TGA AACACCGG -ACCAAGGAG TCTAACATGC GCGCGAGTCA TTGGGCGGTA
NUCLEOTIDES INCLUDED mmmmmmmmm mmmmmmmmm -mmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

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1690      1700      1710      1720      1730      1740
Achatina fulica      CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Achatina achatina    CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Achatina stuhlmanni  CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Cochlitoma ustulata  CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Allopeas clavulinum  CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Bocageia sp.         CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Eutomepeas layardi   CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Leptinaria lamellata CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Paropeas achatinaceum CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Riebeckia sp.        CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Rumina decollata     CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Subulina octona      CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Subulina striatella  CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Subulina vitrea      CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Subulona sp.         CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Tortaxis erectus     CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Xerocerastus sp.    CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Zootecus insularis   CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Coeliaxis blandii    CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Pyrgina umbilicata   CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Glessula ceylanica   CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Cecilioides gokweanus CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Ferussacia folliculus CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Thyrophorella thomensis CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Gibbulinella dewinteri CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Gonaxis quadrilateralis CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
Gonospora sp.        CGAAACCCAA AGGCGCAGTG AAAGCGAGGG TC-GTCT--- ----CGCGC GACCCAGGTG
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mm----- -----nn nnnnnnnnnnnnn

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1750      1760      1770      1780      1790      1800
Achatina fulica      GGATCCT-CC GCTCCT--- -----TC G---CGGGAG CGGT-GGCCG
Achatina achatina    GGATCCT-CC GCTCCT--- -----TC G---CGGGAG CGGT-GGCCG
Achatina stuhlmanni  GGATCCT-CC GCTCCT--- -----TC G---CGGGAG CGGT-GGCCG
Cochlitoma ustulata  GGATCCT-CC GCTCCT--- -----TC G---CGGGAG CGGT-GGCCG
Allopeas clavulinum  GGATCCT-CC CTACCCC--G CGTTATCGCG CGGGT----- ----GCGT GGGT-GGCCG
Bocageia sp.         GGATCCA-TC CTCGCT--- -----TC G---CGGGAG CGGT-GGCCG
Eutomepeas layardi   GGATCCT-CC CTACCCC--G CGTTATCGCG CGGGT----- ----GCGT GGGT-GGCCG
Leptinaria lamellata GGATCCT-CC CTACCCC--G CGTTATCGCG CGGGT----- ----GCGT GGGT-GGCCG
Paropeas achatinaceum GGATCCT-CC CTACCCC--G CGTTATCGCG CGGGT----- ----GCGT GGGT-GGCCG
Riebeckia sp.        GGATCCT-CC TCCTCCTCCT CCTTCCCCTC --GCGGGGGT G---GTGGTG GGGT-GGCCG
Rumina decollata     GGATCCT-CC ACCCTCTCG -----TC G---CGGGAG CGGT-GGCCG
Subulina octona      GGATCCT-CC ACTTC----- -----TC G---CGGGAG CGGT-GGCCG
Subulina striatella  GGATCCT-CC ACTTC----- -----TC G---CGGGAG CGGT-GGCCG
Subulina vitrea      GGATCCATCC CTCGCC--- ----GCGAGC GCAAGCCCGC G---CGTCGG GGGT-GGCCG
Subulona sp.         GGATCCATCC CTCCTGTCTG CGCGAGCGCA AGCCCGCGTC GGCGGTCTGG GGGT-GGCCG
Tortaxis erectus     GGATCCT-CC GCTCTCTC--- -----TC G---CGGGAG CGGT-GGCCG
Xerocerastus sp.    GGATCCT-CC ACCCTCTC--- -----TC G---CGGGAG CGGT-GGCCG
Zootecus insularis   GGATCCT-CC ACTCCTC--- -----TC G---CGGGAG CGGT-GGCCG
Coeliaxis blandii    GGATCCT-CC T-----TC G---CGGGAG CGGT-GGCCG
Pyrgina umbilicata   GGATCCT-CC CTACCCC--G CGTTTCTCGCG CGGGT----- ----GCGT GGGT-GGCCG
Glessula ceylanica   GGATCCT-CC GCTCTC--- -----TC G---CGGGAG CGGT-GGCCG
Cecilioides gokweanus GGATCCT-CC CTGCGCG--- CCCGTTTACG CGGGT----- ----GCGT GGGT-GGCCG
Ferussacia folliculus GGATCCT-CC CCGTCTC--- -----TC G---CGGGAG CGGT-GGCCG
Thyrophorella thomensis GGATCCT-CC CTACCCC--G CGTTTCTCGCG CGGGT----- ----GCGT GGGT-GGCCG
Gibbulinella dewinteri GGATCCT-CC TCTCCCACGC CTTCT-GGCG CG----- ----GCGT GGGT-GGCCG
Gonaxis quadrilateralis GGATCCT-TC CCTCCCCTC ACATTGGCG C-----TC G---CGGGAG CGGT-GGCCG
Gonospora sp.        GGATCCT-GT CCTTCGCGCC ACATTGGCG CG----- ----GCGT GGGT-GGCCG
NUCLEOTIDES INCLUDED mmmmmmmmmmm -----TC G---CGGGAG CGGT-GGCCG

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	1810	1820	1830	1840	1850	1860
Achatina fulica	ACCACCGGCC	CGTCCCGTCC	GCGTCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Achatina achatina	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Achatina stuhlmanni	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Cochlitoma ustulata	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Allopeas clavulinum	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Bocageia sp.	ACCACCGGCC	CGTCCCATCC	GCGCCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Eutomopeas layardi	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Leptinaria lamellata	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Paropeas achatinaceum	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Riebeckia sp.	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Rumina decollata	ACCACCGGCC	CGTCCCGTCC	GCGYCYGYCYG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Subulina octona	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Subulina striatella	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Subulina vitrea	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Subulona sp.	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Tortaxis erectus	ACCACCGGCC	CGTCCCGTCC	GCGCTGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Xerocerastus sp.	ACCACCGGCC	CGTCCCGTCC	GCGCCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Zootecus insularis	ACCACCGGCC	CGTCCCGTCC	GCGCTGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Coeliaxis blandii	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Pyrgina umbilicata	ACCACCGGCC	CGTCCCGTCC	GCGCCGGCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Glessula ceylanica	ACCACCGGCC	CGTCCCGTCC	GCGTCGTCGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Cecilioides gokweanus	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Ferussacia folliculus	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGAG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Thyrophorella thomensis	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGGG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Gibbulinella dewinteri	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGAG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Gonaxis quadrilateralis	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGAG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
Gonospira sp.	ACCACCGGCC	CGTCCCGTCC	GCGTCGCGAG	TGGGGCGGAG	CAAGAGCGTG	CACGCTGGGA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1870	1880	1890	1900	1910	1920
Achatina fulica	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Achatina achatina	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Achatina stuhlmanni	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Cochlitoma ustulata	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Allopeas clavulinum	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Bocageia sp.	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Eutomopeas layardi	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Leptinaria lamellata	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Paropeas achatinaceum	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Riebeckia sp.	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Rumina decollata	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Subulina octona	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Subulina striatella	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Subulina vitrea	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Subulona sp.	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Tortaxis erectus	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Xerocerastus sp.	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Zootecus insularis	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Coeliaxis blandii	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Pyrgina umbilicata	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Glessula ceylanica	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Cecilioides gokweanus	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Ferussacia folliculus	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Thyrophorella thomensis	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Gibbulinella dewinteri	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Gonaxis quadrilateralis	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
Gonospira sp.	CCCgAAAGAT	GGTgAACTAT	GCCTGAGTAG	AACGAAGCCA	GAGGAAACTC	TGgTGGAGGT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1930	1940	1950	1960	1970	1980
Achatina fulica	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Achatina achatina	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Achatina stuhlmanni	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Cochlitoma ustulata	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Allopeas clavulinum	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Bocageia sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Eutomopeas layardi	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Leptinaria lamellata	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Paropeas achatinaceum	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Riebeckia sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Rumina decollata	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Subulina octona	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Subulina striatella	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Subulina vitrea	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Subulona sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Tortaxis erectus	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Xerocerastus sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Zootecus insularis	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Coeliaxis blandii	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Pyrgina umbilicata	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Glessula ceylanica	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Ceciloides gokweanus	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Ferussacia folliculus	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Thyrophorella thomensis	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Gibbulinella dewinteri	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Gonaxis quadrilateralis	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
Gonospira sp.	TCGTAGCGAT	TCTGACGTGC	AAATCGATCG	TCAAACCTGG	GTATAGGGGC	GAAAGACTAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1990	2000	2010	2020	2030	2040
Achatina fulica	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Achatina achatina	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Achatina stuhlmanni	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Cochlitoma ustulata	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Allopeas clavulinum	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Bocageia sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Eutomopeas layardi	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Leptinaria lamellata	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Paropeas achatinaceum	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Riebeckia sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Rumina decollata	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Subulina octona	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Subulina striatella	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Subulina vitrea	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Subulona sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Tortaxis erectus	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Xerocerastus sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Zootecus insularis	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Coeliaxis blandii	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Pyrgina umbilicata	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Glessula ceylanica	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Ceciloides gokweanus	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Ferussacia folliculus	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Thyrophorella thomensis	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Gibbulinella dewinteri	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Gonaxis quadrilateralis	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
Gonospira sp.	TCGAACCATC	TAGTAGCTGG	TTCCCTCCGA	AGTTTCCCTC	AGGATAGCTG	GCGCTCGATC
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	2050	2060	2070	2080	2090	2100	
Achatina fulica	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Achatina achatina	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Achatina stuhlmanni	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Cochlitoma ustulata	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Allopeas clavulinum	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Bocageia sp.	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Eutomopeas layardi	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Leptinaria lamellata	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Paropeas achatinaceum	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Riebeckia sp.	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Rumina decollata	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Subulina octona	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Subulina striatella	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Subulina vitrea	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Subulona sp.	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Tortaxis erectus	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Xerocerastus sp.	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Zootecus insularis	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Coeliaxis blandii	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Pyrgina umbilicata	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Glessula ceylanica	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Cecilioides gokweanus	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Ferussacia folliculus	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Thyrophorella thomensis	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Gibbulinella dewinteri	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Gonaxis quadrilateralis	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
Gonospira sp.	GCAGTTTTAT	CCGGTAAAGC	GAATGATTAG	AGGTCTTGGG	GACGAAACGA	CCTCAACCTA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2110	2120	2130	2140	2150	2160	
Achatina fulica	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGT---	
Achatina achatina	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGT---	
Achatina stuhlmanni	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGT---	
Cochlitoma ustulata	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGT---	
Allopeas clavulinum	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Bocageia sp.	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGT---	
Eutomopeas layardi	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Leptinaria lamellata	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Paropeas achatinaceum	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	AT-GGAGCCG	GGCGCGT---	
Riebeckia sp.	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGTC----	
Rumina decollata	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	YT-GGAGCCG	GGCGCGT---	
Subulina octona	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Subulina striatella	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Subulina vitrea	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TTTGAGCCG	GGCGCGT--G	
Subulona sp.	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGTTTG	
Tortaxis erectus	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGT---	
Xerocerastus sp.	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Zootecus insularis	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Coeliaxis blandii	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCG-----	
Pyrgina umbilicata	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Glessula ceylanica	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGT---	
Cecilioides gokweanus	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT--G	
Ferussacia folliculus	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	TT-GGAGCCG	GGCGCGT---	
Thyrophorella thomensis	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	CT-GGAGCCG	GGCGCGT---	
Gibbulinella dewinteri	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	AT-GGAGCCG	GGCGCGT---	
Gonaxis quadrilateralis	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	AT-GGAGCCG	GGCGCGT---	
Gonospira sp.	TTCTCAAAC	TAAATGGGT	AAGAAGTCCG	GCTCGCTCGA	AT-GGAGCCG	GGCGTGT---	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mm-mmmmmmm	mmmm-----	

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2170      2180      2190      2200      2210      2220
Achatina fulica      -NTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Achatina achatina   -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Achatina stuhlmanni -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Cochlitoma ustulata -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Allopeas clavulinum GTTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Bocageia sp.        --TCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Eutomepeas layardi  GTTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Leptinaria lamellata GTTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Paropeas achatinaceum GTTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Riebeckia sp.       -GTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Rumina decollata    -NTCGAATGC NTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Subulina octona     -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Subulina striatella -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Subulina vitrea     TGTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Subulona sp.        -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Tortaxis erectus    GTACGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Xerocerastus sp.   -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Zootecus insularis -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Coeliaxis blandii  --TCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Pyrgina umbilicata GTTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Glessula ceylanica GTCCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Cecilioides gokweanus TGTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Ferussacia folliculus -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Thyrophorella thomensis GTTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Gibbulinella dewinteri -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Gonaxis quadrilateralis -TTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
Gonospira sp.      -CTCGAATGC GTGTGCCAAG TGGGCCACTT TTGGTAAGCA GAACTGGCGC TGTGGGATGA
NUCLEOTIDES INCLUDED ---mmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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2230      2240      2250      2260      2270      2280
Achatina fulica      ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Achatina achatina   ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Achatina stuhlmanni ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Cochlitoma ustulata ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Allopeas clavulinum ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Bocageia sp.        ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Eutomepeas layardi  ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Leptinaria lamellata ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Paropeas achatinaceum ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Riebeckia sp.       ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Rumina decollata    ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Subulina octona     ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Subulina striatella ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Subulina vitrea     ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Subulona sp.        ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Tortaxis erectus    ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Xerocerastus sp.   ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Zootecus insularis ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Coeliaxis blandii  ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Pyrgina umbilicata ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Glessula ceylanica ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Cecilioides gokweanus ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Ferussacia folliculus ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Thyrophorella thomensis ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Gibbulinella dewinteri ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Gonaxis quadrilateralis ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
Gonospira sp.      ACCAAACGCC CGGTTAAGGT GCCAAACGCT GACGCTCATC AGACACCATA AAAGGTGTTG
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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	2290 2300 2310 2320 2330 2340
<i>Achatina fulica</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Achatina achatina</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Achatina stuhlmanni</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Cochlitoma ustulata</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Allopeas clavulinum</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Bocageia</i> sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Eutomopeas layardi</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Leptinaria lamellata</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Paropeas achatinaceum</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Riebeckia</i> sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Rumina decollata</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Subulina octona</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Subulina striatella</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Subulina vitrea</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Subulona</i> sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Tortaxis erectus</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Xerocerastus</i> sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Zootecus insularis</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Coeliaxis blandii</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Pyrgina umbilicata</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Glessula ceylanica</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Cecilioides gokweanus</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Ferussacia folliculus</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Thyrophorella thomensis</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Gibbulinella dewinteri</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Gonaxis quadrilateralis</i>	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
<i>Gonospira</i> sp.	GTTGATATAG ACAGCAGGAC GGTGGCCATG GAAGTCGGAA CCCGCTAAGG AGTGTGTAAC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2350 2360 2370 2380 2390 2400
<i>Achatina fulica</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Achatina achatina</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Achatina stuhlmanni</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Cochlitoma ustulata</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Allopeas clavulinum</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Bocageia</i> sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Eutomopeas layardi</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Leptinaria lamellata</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Paropeas achatinaceum</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Riebeckia</i> sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Rumina decollata</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Subulina octona</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Subulina striatella</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Subulina vitrea</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Subulona</i> sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Tortaxis erectus</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Xerocerastus</i> sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Zootecus insularis</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Coeliaxis blandii</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Pyrgina umbilicata</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Glessula ceylanica</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Cecilioides gokweanus</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Ferussacia folliculus</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Thyrophorella thomensis</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Gibbulinella dewinteri</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Gonaxis quadrilateralis</i>	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
<i>Gonospira</i> sp.	AACTCACCTG CCGAATCAAC CAGCCCTGAA AATGGATGGC GCTAGAGCGT CGGACCCATA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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2410      2420      2430      2440      2450      2460
Achatina fulica      CCGGGCCGTC TCGGCAATGG GCCCTTCCAT GGGG-CCGGA AGCGAGGCCG AGACGAGTAG
Achatina achatina   CCGGGCCGTC TCGGCAATAG GCC---TTCT TCCG-CCGGA AGCGAGGCCG AGACGAGTAG
Achatina stuhlmanni CCGGGCCGTC TCGGCAATGG GCCCTTTCAT GGGG-CCGGA AGCGAGGCCG AGACGAGTAG
Cochlitoma ustulata CCGGGCCGTC TCGGCAATGG GCCC-TTCCA CGGG-CCGGA AGCGAGGCCG AGACGAGTAG
Allopeas clavulinum CCGGGCCGTC TCGGCAATCG GAAACAA--- -----G CGCGAGGCCG AGACGAGTAG
Bocageia sp.        CCGGGCCGTC TCGGCAATCA GAAAGCA--- -----A ATCGAGGCCG AGACGAGTAG
Eutomopeas layardi  CCGGGCCGTC TCGGCAATCG GAAACAA--- -----G CGCGAGGCCG AGACGAGTAG
Leptinaria lamellata CCGGGCCGTC TCGGCAATCG TATGCAA--- -----G CGCGAGGCCG AGACGAGTAG
Paropeas achatinaceum CCGGGCCGTC TCGGCAATCG GAAACAA--- -----G CGAGAGGCCG AGACGAGTAG
Riebeckia sp.       CCGGGCCGTC TCGGCAATCG AATAAAGCCG CA-----A AGCGAGGCCG AGACGAGTAG
Rumina decollata    CCGGGCCGTC TCGGCAATGG CGATAGCCGC G-----A ATCGAGGCCG AGACGAGTAG
Subulina octona     CCGGGCCGTC TCGGCAATTG CCCCC----- -----A AGCGAGGCCG AGACGAGTAG
Subulina striatella CCGGGCCGTC TCGGCAATTG TTGCCTC--- -----A AGCGAGGCCG AGACGAGTAG
Subulina vitrea     CCGGGCCGTC TCGGCAATCG GATCTAACCG AGAGA----G AGAGAGGCCG AGACGAGTAG
Subulona sp.        CCGGGCCGTC TCGGCAATGG TTG-----C TGAGAGGCCG AGACGAGTAG
Tortaxis erectus    CCGGGCCGTC TCGGCAATGG CCTTTTACTC A-----A AGCGAGGCCG AGACGAGTAG
Xerocerastus sp.   CCGGGCCGTC TCGGCAATGG CTTTCGTTGCC GCG-----A ATCGAGGCCG AGACGAGTAG
Zootecus insularis  CCGGGCCGTC TCGGCAATGG A-----A ATCGAGGCCG AGACGAGTAG
Coeliaxis blandii   CCGGGCCGTC TCGGCAATCG AAACATAAAG GC-----A AGCGAGGCCG AGACGAGTAG
Pyrquina umbilicata CCGGGCCGTC TCGGCAATCG GAAACAA--- -----A CGCGAGGCCG AGACGAGTAG
Glessula ceylanica  CCGGGCCGTC TCGGCAATGG TCACGCCATT -----A AGCGAGGCCG AGACGAGTAG
Cecilioides gokweanus CCGGGCCGTC TCGGCAATCG ATCTTCGGTA -----A CGCGAGGCCG AGACGAGTAG
Ferussacia folliculus CCGGGCCGTC TCGGCAATAC GTGCA-----A AAAAAAGGCCG AGACGAGTAG
Thyrophorella thomensis CCGGGCCGTC TCGGCAATCG GAAACAA--- -----A CGCGAGGCCG AGACGAGTAG
Gibbulinella dewinteri CCGGGCCGTC TCGGCAATGG CCGCGCCGA- -----A CGCAAGGCCG AGACGAGTAG
Gonaxis quadrilateralis CCGGGCCGTC TCGGCAATCT GGCCGA-----A GGCAAGGCCG AGACGAGTAG
Gonospira sp.       CCGGGCCGTC TCGGCAATCG GTGCCGCA-- -----A GGCGAGGCCG AGACGAGTAG
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm-----m mmmmmmmmmmm mmmmmmmmmmm

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2470      2480      2490      2500      2510      2520
Achatina fulica      GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Achatina achatina   GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Achatina stuhlmanni GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Cochlitoma ustulata GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Allopeas clavulinum GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Bocageia sp.        GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Eutomopeas layardi  GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Leptinaria lamellata GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Paropeas achatinaceum GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Riebeckia sp.       GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Rumina decollata    GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Subulina octona     GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Subulina striatella GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Subulina vitrea     GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Subulona sp.        GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Tortaxis erectus    GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Xerocerastus sp.   GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Zootecus insularis  GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Coeliaxis blandii   GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Pyrquina umbilicata GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Glessula ceylanica  GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Cecilioides gokweanus GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Ferussacia folliculus GAGGGCCGTC GAGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCTCGGG
Thyrophorella thomensis GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Gibbulinella dewinteri GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Gonaxis quadrilateralis GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
Gonospira sp.       GAGGGCCGTC GGGGTGAGCG TGGAAAGCCTG GGGAGCGATC CTGGGTGGAG CCGCCCCGGG
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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	2530	2540	2550	2560	2570	2580	
<i>Achatina fulica</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Achatina achatina</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Achatina stuhlmanni</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Cochlitoma ustulata</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Allopeas clavulinum</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Bocageia</i> sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Eutomopeas layardi</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Leptinaria lamellata</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Paropeas achatinaceum</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Riebeckia</i> sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Rumina decollata</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Subulina octona</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Subulina striatella</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Subulina vitrea</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Subulona</i> sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Tortaxis erectus</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Xerocerastus</i> sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Zootecus insularis</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Coeliaxis blandii</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Pyrgina umbilicata</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Glessula ceylanica</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Cecilioides gokweanus</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Ferussacia folliculus</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Thyrophorella thomensis</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Gibbulinella dewinteri</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Gonaxis quadrilateralis</i>	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
<i>Gonospira</i> sp.	TGCAGATCTT	GGTGGTAGTA	GCAAATATTC	AAACGAGAAC	TTTGAAGACT	GAAGTGGAGA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2590	2600	2610	2620	2630	2640
<i>Achatina fulica</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Achatina achatina</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Achatina stuhlmanni</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Cochlitoma ustulata</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Allopeas clavulinum</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Bocageia</i> sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Eutomopeas layardi</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Leptinaria lamellata</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Paropeas achatinaceum</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Riebeckia</i> sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Rumina decollata</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Subulina octona</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Subulina striatella</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Subulina vitrea</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Subulona</i> sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Tortaxis erectus</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Xerocerastus</i> sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Zootecus insularis</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Coeliaxis blandii</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Pyrgina umbilicata</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Glessula ceylanica</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Cecilioides gokweanus</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Ferussacia folliculus</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Thyrophorella thomensis</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Gibbulinella dewinteri</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Gonaxis quadrilateralis</i>	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
<i>Gonospira</i> sp.	AGGGTTCCAT	GTGAACAGCA	GTTGAACATG	GGTCAGTCGG	TCCTAAGAGA	TAGGAAAAC
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	2650 2660 2670 2680 2690 2700
<i>Achatina fulica</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT CTTTCT---- -----AGAAGA
<i>Achatina achatina</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTGTC GCAGT----- -----CAA
<i>Achatina stuhlmanni</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTATAA ---TTCGTTA TT----- -----AGA
<i>Cochlitoma ustulata</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCGTG ----- -----ATTAAA
<i>Allopeas clavulinum</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT CTTTCTTCT- -----CGAA GCCAGAAGTA
<i>Bocageia</i> sp.	CCGTTCTGAC CCCGGGGC-A TTCTTTACTA ACA----- -----AAA AGGAAGAAAA
<i>Eutomopeas layardi</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT CTTCTC---- ----- --GAAGCAGA
<i>Leptinaria lamellata</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTG- ----- -----AATGAGTA
<i>Paropeas achatinaceum</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTA CTTCTTCTTC T--CGCGAAC GAAGAGCAGA
<i>Riebeckia</i> sp.	CCGTTCTGAC CCCGGGGCAA TTCTTTCTTT CTTTCT---- -----CGAAC GAACGATCGA
<i>Rumina decollata</i>	CCGTTCTGAC CCCGGGGC-A ATCTTTCTCG AGCTA----- -----AGA AGAAAAAGAA
<i>Subulina octona</i>	CCGTTCTGAC CCCGGGGC-A ATCTTTTCTT C----- ----- -----AAGA
<i>Subulina striatella</i>	CCGTTCTGAC CCCGGGGC-A ATCTTCATTT TT----- ----- -----GA
<i>Subulina vitrea</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT AATCTTTT-- ----- -----GCAGA
<i>Subulona</i> sp.	CCGTTCTGAC CCCGGGGC-G TTTTTTTGTA ATAATCTT-- ----- -----A
<i>Tortaxis erectus</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTC- ----- ----- --AGCGAATA
<i>Xerocerastus</i> sp.	CCGTTCTGAC CCCGGGGC-A CTCFTT---- ----- ---CTTGACG ATTCAATTGA
<i>Zootecus insularis</i>	CCGTTCTGAC CCCGGGGC-A ATCTTTCTTT CTTTCTC-- ----- ---G AGAAGAAGAA
<i>Coeliaxis blandii</i>	CCGTTCTGAC CCCGGGGCAA TTCFTT---- ----- ----- ---CGTTGA
<i>Pyrgina umbilicata</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTTT CACT----- -----CGA AGAAGCAAGA
<i>Glessula ceylanica</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTAG- ----- ----- -----AAAA
<i>Ceciloides gokweanus</i>	CCGTTCTGAC CCCGGGGCAA TTCTTTCTTT CTGTTT---- ----- -----GA
<i>Ferussacia folliculus</i>	CCGTTCTGAC CCCGGGGC-A CTCFTT---- ----- -----ACGT CGGTAAACGA
<i>Thyrophorella thomensis</i>	CCGTTCTGAC CCCGGGGC-A TTCTTTCTT- ----- ----- ---ACTCGAAGCA
<i>Gibbulinella dewinteri</i>	CCGTTCTGAC CCCGGGGC-G CTCFTTACG- ----- ----- ---C AACGTTTCGA
<i>Gonaxis quadrilateralis</i>	CCGTTCTGAC CCCGGGGC-G CTCFTTTTTT- ----- ----- ---GTTGATCGA
<i>Gonospora</i> sp.	CCGTTCTGAC CCCGGGGC-G ATCTTTTACA- -----CAAC AACACGCGAG TTGTCTGCGA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmm-- ----- ----- -----m

	2710 2720 2730 2740 2750 2760
<i>Achatina fulica</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Achatina achatina</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Achatina stuhlmanni</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Cochlitoma ustulata</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Allopeas clavulinum</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Bocageia</i> sp.	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Eutomopeas layardi</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Leptinaria lamellata</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Paropeas achatinaceum</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Riebeckia</i> sp.	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Rumina decollata</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Subulina octona</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Subulina striatella</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Subulina vitrea</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Subulona</i> sp.	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Tortaxis erectus</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Xerocerastus</i> sp.	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Zootecus insularis</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Coeliaxis blandii</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Pyrgina umbilicata</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Glessula ceylanica</i>	G-TCTGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Ceciloides gokweanus</i>	A-TCAGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Ferussacia folliculus</i>	GCTCGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Thyrophorella thomensis</i>	GNGTCGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Gibbulinella dewinteri</i>	G-TACGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Gonaxis quadrilateralis</i>	G-CACGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
<i>Gonospora</i> sp.	G-CACGCCCG CAGCCTATCG AAAGGGAATC GGGTTAATAT TCCCGAACCT GGACACGGAG
NUCLEOTIDES INCLUDED	m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2770	2780	2790	2800	2810	2820	
Achatina fulica	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Achatina achatina	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Achatina stuhlmanni	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Cochlitoma ustulata	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Allopeas clavulinum	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Bocageia sp.	ATTGGTCCTC	TGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGAGAGCC	
Eutomopeas layardi	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Leptinaria lamellata	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Paropeas achatinaceum	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Riebeckia sp.	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Rumina decollata	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Subulina octona	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Subulina striatella	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Subulina vitrea	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Subulona sp.	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Tortaxis erectus	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGAGGCC	
Xerocerastus sp.	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Zootecus insularis	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Coeliaxis blandii	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Pyrgina umbilicata	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Glessula ceylanica	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Cecilioides gokweanus	ATTGGTCCTC	TGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Ferussacia folliculus	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Thyrophorella thomensis	ATTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Gibbulinella dewinteri	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Gonaxis quadrilateralis	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
Gonospira sp.	ACTGGTCCTC	AGGGGCCACG	TGCGGCAACG	CAAACGAAGT	GGGGGACGTC	GGCGGGAGCC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2830	2840	2850	2860	2870	2880	
Achatina fulica	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Achatina achatina	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Achatina stuhlmanni	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Cochlitoma ustulata	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Allopeas clavulinum	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Bocageia sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Eutomopeas layardi	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Leptinaria lamellata	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Paropeas achatinaceum	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Riebeckia sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Rumina decollata	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Subulina octona	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Subulina striatella	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Subulina vitrea	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Subulona sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Tortaxis erectus	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Xerocerastus sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Zootecus insularis	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Coeliaxis blandii	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Pyrgina umbilicata	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Glessula ceylanica	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Cecilioides gokweanus	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Ferussacia folliculus	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Thyrophorella thomensis	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Gibbulinella dewinteri	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Gonaxis quadrilateralis	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
Gonospira sp.	CCGGGAAGAG	TTCTCTTTTC	TTTGTAAGGA	GCCACATCCC	TGGAATCGGC	TTGCCCGGAG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2890 2900 2910 2920 2930 2940
<i>Achatina fulica</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Achatina achatina</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Achatina stuhlmanni</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Cochlitoma ustulata</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Allopeas clavulinum</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Bocageia</i> sp.	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Eutomopeas layardi</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Leptinaria lamellata</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Paropeas achatinaceum</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Riebeckia</i> sp.	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Rumina decollata</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Subulina octona</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Subulina striatella</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Subulina vitrea</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Subulona</i> sp.	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Tortaxis erectus</i>	ATAGGGACCG CGGCTCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Xerocerastus</i> sp.	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Zootecus insularis</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Coeliaxis blandii</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Pyrquina umbilicata</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Glessula ceylanica</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Ceciloides gokweanus</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Ferussacia folliculus</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Thyrophorella thomensis</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Gibbulinella dewinteri</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Gonaxis quadrilateralis</i>	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
<i>Gonospira</i> sp.	ATAGGGACCG CGGCCCGTA AAGCACCGCG GCTCTGCGG TGTCCGGTGC GCTCCCGTCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2950 2960 2970 2980 2990 3000
<i>Achatina fulica</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Achatina achatina</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Achatina stuhlmanni</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Cochlitoma ustulata</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Allopeas clavulinum</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Bocageia</i> sp.	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Eutomopeas layardi</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Leptinaria lamellata</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Paropeas achatinaceum</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Riebeckia</i> sp.	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Rumina decollata</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Subulina octona</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Subulina striatella</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Subulina vitrea</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Subulona</i> sp.	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Tortaxis erectus</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Xerocerastus</i> sp.	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Zootecus insularis</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Coeliaxis blandii</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Pyrquina umbilicata</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Glessula ceylanica</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Ceciloides gokweanus</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Ferussacia folliculus</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Thyrophorella thomensis</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Gibbulinella dewinteri</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Gonaxis quadrilateralis</i>	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
<i>Gonospira</i> sp.	GCCCTTGAAA ACCCCACGGA GACGGTGTGA ATTTTCGTGCC AGGCCGTACC CATATCCGCA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3010 3020 3030 3040 3050 3060
<i>Achatina fulica</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Achatina achatina</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Achatina stuhlmanni</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Cochlitoma ustulata</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Allopeas clavulinum</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Bocageia</i> sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Eutomopeas layardi</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Leptinaria lamellata</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Paropeas achatinaceum</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Riebeckia</i> sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Rumina decollata</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Subulina octona</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Subulina striatella</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Subulina vitrea</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Subulona</i> sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Tortaxis erectus</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Xerocerastus</i> sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Zootecus insularis</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Coeliaxis blandii</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Pyrgina umbilicata</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Glessula ceylanica</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Cecilioides gokweanus</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Ferussacia folliculus</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Thyrophorella thomensis</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Gibbulinella dewinteri</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Gonaxis quadrilateralis</i>	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
<i>Gonospira</i> sp.	GCAGGTCTCC GAGGTGCACA GCCTCTAGTC GATAGAACAA TGTTAGGTAAG GGAAGTCGGC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3070 3080 3090 3100 3110 3120
<i>Achatina fulica</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Achatina achatina</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Achatina stuhlmanni</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Cochlitoma ustulata</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Allopeas clavulinum</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Bocageia</i> sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Eutomopeas layardi</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Leptinaria lamellata</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Paropeas achatinaceum</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Riebeckia</i> sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Rumina decollata</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Subulina octona</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Subulina striatella</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Subulina vitrea</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Subulona</i> sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Tortaxis erectus</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Xerocerastus</i> sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Zootecus insularis</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Coeliaxis blandii</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Pyrgina umbilicata</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Glessula ceylanica</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Cecilioides gokweanus</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Ferussacia folliculus</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Thyrophorella thomensis</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Gibbulinella dewinteri</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Gonaxis quadrilateralis</i>	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
<i>Gonospira</i> sp.	AAATTGGATC CGTAACCTTCG GGAAAAGGAT TGGCTCTGAG GGCTGGGTCA GTCGGGCCGG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3130 3140 3150 3160 3170 3180
Achatina fulica	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Achatina achatina	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Achatina stuhlmanni	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Cochlitoma ustulata	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Allopeas clavulinum	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Bocageia sp.	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
Eutomopeas layardi	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Leptinaria lamellata	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Paropeas achatinaceum	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Riebeckia sp.	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Rumina decollata	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
Subulina octona	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Subulina striatella	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Subulina vitrea	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Subulona sp.	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Tortaxis erectus	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
Xerocerastus sp.	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
Zootecus insularis	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
Coeliaxis blandii	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Pyrquina umbilicata	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Glessula ceylanica	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
Ceciloides gokweanus	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Ferussacia folliculus	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TCCTCCGGCG
Thyrophorella thomensis	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGCCGGCGG
Gibbulinella dewinteri	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
Gonaxis quadrilateralis	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
Gonospira sp.	GGTACGAAGC GGGACTGGGA TGGGCCCGGG CTGGGCGAGG CCGCCGCCGC TAGTCCGGCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3190 3200 3210 3220 3230 3240
Achatina fulica	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Achatina achatina	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Achatina stuhlmanni	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Cochlitoma ustulata	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Allopeas clavulinum	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Bocageia sp.	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
Eutomopeas layardi	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Leptinaria lamellata	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Paropeas achatinaceum	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Riebeckia sp.	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Rumina decollata	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Subulina octona	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Subulina striatella	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Subulina vitrea	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Subulona sp.	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Tortaxis erectus	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
Xerocerastus sp.	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
Zootecus insularis	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
Coeliaxis blandii	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Pyrquina umbilicata	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Glessula ceylanica	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
Ceciloides gokweanus	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Ferussacia folliculus	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
Thyrophorella thomensis	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGCCGGCGG
Gibbulinella dewinteri	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
Gonaxis quadrilateralis	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
Gonospira sp.	GCCGGCCGAG CTCGGAACGC GGCTGCAACC TTCCCGTGGA CCGCCCCAGC TAGTCCGGCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3250	3260	3270	3280	3290	3300			
<i>Achatina fulica</i>	CGCCT-----	-CCCCGGCGT	CGTTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Achatina achatina</i>	CGCCT-----	-CCCCGGCGT	CGTTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Achatina stuhlmanni</i>	CGCCT-----	-CCCCGGCGT	CGTTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Cochlitoma ustulata</i>	CGCCT-----	-CCCCGGCGT	CGTTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Allopeas clavulinum</i>	CGCCT-----	TCCCCGGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Bocageia</i> sp.	CGCCT-----	-CCCCGGCGT	CGTTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Eutomopeas layardi</i>	CGCCT-----	TCCCCGGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Leptinaria lamellata</i>	CGCCC-----	TCCCCGGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Paropeas achatinaceum</i>	CGCCT-----	TCCCCGGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Riebeckia</i> sp.	CGCCT-----	-CCCCGGCGT	CGTTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Rumina decollata</i>	CGCCT-----	--CCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Subulina octona</i>	CGCCT-----	-CCCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Subulina striatella</i>	CGCCT-----	-CCCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Subulina vitrea</i>	CGCCT-----	TCCCCGGGCGT	CGTTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Subulona</i> sp.	CGCCT-----	TCCACGGGCGT	CGTTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Tortaxis erectus</i>	CGCCT-----	TCCCAGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Xerocerastus</i> sp.	CGCCT-----	-CCCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Zootecus insularis</i>	CGCCT-----	-CCCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Coeliaxis blandii</i>	CGCCT-----	-CCCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Pyrgina umbilicata</i>	CGCCT-----	TCCCCGGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Glessula ceylanica</i>	CACCG-----	TCCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Cecilioides gokweanus</i>	CGCCT-----	TCCCCGGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Ferussacia folliculus</i>	CGCCTCCTCC	CGCGGGGTGW	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Thyrophorella thomensis</i>	CGCCT-----	TCCCCGGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Gibbulinella dewinteri</i>	CGCCT-----	--CCCGGCGT	TGTCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Gonaxis quadrilateralis</i>	CGCCT-----	--CCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
<i>Gonospira</i> sp.	CGCCT-----	--CCCGGCGT	CGTCCGCGTC	GGCTGGCATT	CAACAGCCAA	CTCAGAACTG			
NUCLEOTIDES INCLUDED	mmmmmm-----	--mmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm			

	3310	3320	3330	3340	3350	3360			
<i>Achatina fulica</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Achatina achatina</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Achatina stuhlmanni</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Cochlitoma ustulata</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Allopeas clavulinum</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Bocageia</i> sp.	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Eutomopeas layardi</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Leptinaria lamellata</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Paropeas achatinaceum</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Riebeckia</i> sp.	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Rumina decollata</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Subulina octona</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Subulina striatella</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Subulina vitrea</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Subulona</i> sp.	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Tortaxis erectus</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Xerocerastus</i> sp.	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Zootecus insularis</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Coeliaxis blandii</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Pyrgina umbilicata</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Glessula ceylanica</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Cecilioides gokweanus</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Ferussacia folliculus</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Thyrophorella thomensis</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Gibbulinella dewinteri</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Gonaxis quadrilateralis</i>	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
<i>Gonospira</i> sp.	GTACGGACCA	GGGGAATCCG	ACTGTCTAAT	TAAAACAAAG	CATTGCGACG	GCCGTCACCC			
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm			

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          3370      3380      3390      3400      3410      3420
Achatina fulica      GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Achatina achatina   GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Achatina stuhlmanni GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Cochlitoma ustulata GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Allopeas clavulinum GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Bocageia sp.        GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Eutomopeas layardi  GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Leptinaria lamellata GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Paropeas achatinaceum GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Riebeckia sp.       GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Rumina decollata    GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Subulina octona     GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Subulina striatella GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Subulina vitrea     GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Subulona sp.        GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Tortaxis erectus    GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Xerocerastus sp.   GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Zootecus insularis  GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Coeliaxis blandii   GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Pyrgina umbilicata  GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Glessula ceylanica  GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Cecilioides gokweanus GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Ferussacia folliculus GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Thyrophorella thomensis GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Gibbulinella dewinteri GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Gonaxis quadrilateralis GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
Gonospira sp.       GGTGTTGACG CAATGTGATT TCTGCCCAGT GCTCTGAATG TCAAAGTGAA GAAATTCAAC
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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          3430      3440      3450      3460      3470      3480
Achatina fulica      CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Achatina achatina   CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Achatina stuhlmanni CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Cochlitoma ustulata CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Allopeas clavulinum CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Bocageia sp.        CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Eutomopeas layardi  CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Leptinaria lamellata CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Paropeas achatinaceum CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Riebeckia sp.       CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Rumina decollata    CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Subulina octona     CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Subulina striatella CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Subulina vitrea     CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Subulona sp.        CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Tortaxis erectus    CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Xerocerastus sp.   CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Zootecus insularis  CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Coeliaxis blandii   CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Pyrgina umbilicata  CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Glessula ceylanica  CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Cecilioides gokweanus CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Ferussacia folliculus CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Thyrophorella thomensis CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Gibbulinella dewinteri CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Gonaxis quadrilateralis CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
Gonospira sp.       CAAGCGCGGG TAAACGGCGG GAGTAACTAT GACTCTCTTA AGGTAGCCAA ATGCCTCGTC
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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	3490 3500 3510 3520 3530 3540
<i>Achatina fulica</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Achatina achatina</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Achatina stuhlmanni</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Cochlitoma ustulata</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Allopeas clavulinum</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Bocageia</i> sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Eutomopeas layardi</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Leptinaria lamellata</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Paropeas achatinaceum</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Riebeckia</i> sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Rumina decollata</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Subulina octona</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Subulina striatella</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Subulina vitrea</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Subulona</i> sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Tortaxis erectus</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Xerocerastus</i> sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Zootecus insularis</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Coeliaxis blandii</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Pyrgina umbilicata</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Glessula ceylanica</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Cecilioides gokweanus</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Ferussacia folliculus</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Thyrophorella thomensis</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Gibbulinella dewinteri</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Gonaxis quadrilateralis</i>	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
<i>Gonospira</i> sp.	ATCTAATTAG TGACGCGCAT GAATGGATTA ACGAGATTCC CACTGTCCCT ATCTACTATC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3550 3560 3570 3580 3590 3600
<i>Achatina fulica</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Achatina achatina</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Achatina stuhlmanni</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Cochlitoma ustulata</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Allopeas clavulinum</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Bocageia</i> sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Eutomopeas layardi</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Leptinaria lamellata</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Paropeas achatinaceum</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Riebeckia</i> sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Rumina decollata</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Subulina octona</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Subulina striatella</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Subulina vitrea</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Subulona</i> sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Tortaxis erectus</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Xerocerastus</i> sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Zootecus insularis</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Coeliaxis blandii</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Pyrgina umbilicata</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Glessula ceylanica</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Cecilioides gokweanus</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Ferussacia folliculus</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Thyrophorella thomensis</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Gibbulinella dewinteri</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Gonaxis quadrilateralis</i>	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
<i>Gonospira</i> sp.	TAGCGAAACC ACAGCCAAGG GAACGGGCTT GGTAGAATCA GCGGGGAAAG AAGACCCTGT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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3610      3620      3630      3640      3650      3660
Achatina fulica      TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Achatina achatina   TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Achatina stuhlmanni TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Cochlitoma ustulata TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Allopeas clavulinum TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Bocageia sp.        TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Eutomoepas layardi  TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Leptinaria lamellata TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Paropeas achatinaceum TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Riebeckia sp.       TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Rumina decollata    TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Subulina octona     TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Subulina striatella TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Subulina vitrea     TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Subulona sp.        TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Tortaxis erectus    TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Xerocerastus sp.   TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Zootecus insularis TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Coeliaxis blandii  TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Pyrgina umbilicata TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Glessula ceylanica TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Cecilioides gokweanus TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Ferussacia folliculus TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Thyrophorella thomensis TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Gibbulinella dewinteri TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Gonaxis quadrilateralis TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
Gonospira sp.       TGAGCTTGAC TCTAGTCCGA CTTTGTGAAG ACACATGAAG GGTGTAGCAT AGGTGGGAGC
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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3670      3680      3690      3700      3710      3720
Achatina fulica      GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Achatina achatina   GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Achatina stuhlmanni GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Cochlitoma ustulata GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Allopeas clavulinum GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Bocageia sp.        GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Eutomoepas layardi  GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Leptinaria lamellata GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Paropeas achatinaceum GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Riebeckia sp.       GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Rumina decollata    GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Subulina octona     GCGAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Subulina striatella GCGAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Subulina vitrea     GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Subulona sp.        GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Tortaxis erectus    GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Xerocerastus sp.   GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Zootecus insularis GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Coeliaxis blandii  GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Pyrgina umbilicata GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Glessula ceylanica GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Cecilioides gokweanus GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Ferussacia folliculus GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Thyrophorella thomensis GCAAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Gibbulinella dewinteri GCGAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Gonaxis quadrilateralis GCGAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
Gonospira sp.       GCGAGCGCAA TTGAAATACC ACTACTTTTA TCGTTTCTTT ACTTATTCAG TCAAGCGGAG
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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	3730 3740 3750 3760 3770 3780
Achatina fulica	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Achatina achatina	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Achatina stuhlmanni	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Cochlitoma ustulata	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA TCGGCGCTCG TCGTCGGCCG
Allopeas clavulinum	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Bocageia sp.	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Eutomopeas layardi	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Leptinaria lamellata	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Paropeas achatinaceum	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Riebeckia sp.	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Rumina decollata	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Subulina octona	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Subulina striatella	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Subulina vitrea	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Subulona sp.	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA TCGGCGCTCG TCGTCGGCCG
Tortaxis erectus	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Xerocerastus sp.	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Zootecus insularis	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Coeliaxis blandii	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Pyrgina umbilicata	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Glessula ceylanica	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCTCG TCGTCGGCCG
Cecilioides gokweanus	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA TCGGCGCTCG TCGTCGGCCG
Ferussacia folliculus	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAC TCGGCGCTCG TCGTCGGCCG
Thyrophorella thomensis	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA CCGGCGCCCG TCGCCGGCCG
Gibbulinella dewinteri	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA TCGGCGCTCG TCGTCGGCCG
Gonaxis quadrilateralis	AGCGGGGCGA AAGCCCTCG CTTCTGGAGT TAAGCGGCAA TCGGCGCTCG TCGTCGGCCG
Gonospira sp.	AGCGGGGCGC AAGCCCTCG CTTCTGGAGT TAAGCGGCAA TCGGCGCTCG TCGTCGGCCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3790 3800 3810 3820 3830 3840
Achatina fulica	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Achatina achatina	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Achatina stuhlmanni	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Cochlitoma ustulata	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Allopeas clavulinum	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Bocageia sp.	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Eutomopeas layardi	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Leptinaria lamellata	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Paropeas achatinaceum	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Riebeckia sp.	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Rumina decollata	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Subulina octona	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Subulina striatella	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Subulina vitrea	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Subulona sp.	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Tortaxis erectus	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Xerocerastus sp.	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Zootecus insularis	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Coeliaxis blandii	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Pyrgina umbilicata	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Glessula ceylanica	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Cecilioides gokweanus	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Ferussacia folliculus	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Thyrophorella thomensis	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Gibbulinella dewinteri	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Gonaxis quadrilateralis	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
Gonospira sp.	CGATCCGCTC TGAAGACAGT GTCAGGCGGG GAGTTTGACT GGGGCGGTAC ATCTGTCAAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3850 3860 3870 3880 3890 3900
<i>Achatina fulica</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Achatina achatina</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Achatina stuhlmanni</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Cochlitoma ustulata</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Allopeas clavulinum</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Bocageia</i> sp.	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Eutomopeas layardi</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Leptinaria lamellata</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Paropeas achatinaceum</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Riebeckia</i> sp.	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Rumina decollata</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Subulina octona</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Subulina striatella</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Subulina vitrea</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Subulona sp.</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Tortaxis erectus</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Xerocerastus</i> sp.	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Zootecus insularis</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Coeliaxis blandii</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Pyrgina umbilicata</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Glessula ceylanica</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Ceciloides gokweanus</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Ferussacia folliculus</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Thyrophorella thomensis</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Gibbulinella dewinteri</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Gonaxis quadrilateralis</i>	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
<i>Gonospira</i> sp.	AGGTAACGCA GGTGTCCTAA GCGGAGCTCA GCGAGGACGG AAACCTCGCG TAGAGCAAAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3910 3920 3930 3940 3950 3960
<i>Achatina fulica</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Achatina achatina</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Achatina stuhlmanni</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Cochlitoma ustulata</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Allopeas clavulinum</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Bocageia</i> sp.	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Eutomopeas layardi</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Leptinaria lamellata</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Paropeas achatinaceum</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Riebeckia</i> sp.	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Rumina decollata</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Subulina octona</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Subulina striatella</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Subulina vitrea</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Subulona sp.</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Tortaxis erectus</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Xerocerastus</i> sp.	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Zootecus insularis</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Coeliaxis blandii</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Pyrgina umbilicata</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Glessula ceylanica</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Ceciloides gokweanus</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Ferussacia folliculus</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Thyrophorella thomensis</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Gibbulinella dewinteri</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Gonaxis quadrilateralis</i>	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
<i>Gonospira</i> sp.	GGGCAAAAGC TCGCTTGATT TTGATTTTCA GTACGAATAC AGACCGTGAA AGCGTGGCCT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3970 3980 3990 4000 4010 4020
<i>Achatina fulica</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Achatina achatina</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Achatina stuhlmanni</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Cochlitoma ustulata</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Allopeas clavulinum</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Bocageia</i> sp.	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Eutomopeas layardi</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Leptinaria lamellata</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Paropeas achatinaceum</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Riebeckia</i> sp.	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Rumina decollata</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Subulina octona</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Subulina striatella</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Subulina vitrea</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Subulona</i> sp.	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Tortaxis erectus</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Xerocerastus</i> sp.	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Zootecus insularis</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Coeliaxis blandii</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Pyrgina umbilicata</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Glessula ceylanica</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Cecilioides gokweanus</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Ferussacia folliculus</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Thyrophorella thomensis</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Gibbulinella dewinteri</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Gonaxis quadrilateralis</i>	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
<i>Gonospira</i> sp.	ATCGATCCTT TTGACTTTAA GAGTTTTAAG CAAGAGGTGT CAGAAAAGTT ACCACAGGGA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	4030 4040 4050 4060 4070 4080
<i>Achatina fulica</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Achatina achatina</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Achatina stuhlmanni</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Cochlitoma ustulata</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Allopeas clavulinum</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Bocageia</i> sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Eutomopeas layardi</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Leptinaria lamellata</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Paropeas achatinaceum</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Riebeckia</i> sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Rumina decollata</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Subulina octona</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Subulina striatella</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Subulina vitrea</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Subulona</i> sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Tortaxis erectus</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Xerocerastus</i> sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Zootecus insularis</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Coeliaxis blandii</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Pyrgina umbilicata</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Glessula ceylanica</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Cecilioides gokweanus</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Ferussacia folliculus</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Thyrophorella thomensis</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Gibbulinella dewinteri</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Gonaxis quadrilateralis</i>	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
<i>Gonospira</i> sp.	TAACTGGCTT GTGGCAGCCA AGCGTTCATA GCGACGTTGC TTTTGTGATCC TTCGATGTCC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	4090	4100	4110	4120	4130	4140	
Achatina fulica	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Achatina achatina	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Achatina stuhlmanni	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Cochlitoma ustulata	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Allopeas clavulinum	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Bocageia sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Eutomopeas layardi	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Leptinaria lamellata	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Paropeas achatinaceum	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Riebeckia sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Rumina decollata	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Subulina octona	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Subulina striatella	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Subulina vitrea	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Subulona sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Tortaxis erectus	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Xerocerastus sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Zootecus insularis	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Coeliaxis blandii	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Pyrgina umbilicata	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Glessula ceylanica	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Ceciloides gokweanus	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Ferussacia folliculus	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Thyrophorella thomensis	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Gibbulinella dewinteri	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Gonaxis quadrilateralis	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
Gonospira sp.	GCTCTTCCTA	TCATTGCGAA	GCAGAATTCCG	CCAAGCGTTG	GATTGTTTCAC	CCACTAATAG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	4150	4160	4170	4180	4190	4200	
Achatina fulica	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Achatina achatina	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Achatina stuhlmanni	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Cochlitoma ustulata	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Allopeas clavulinum	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Bocageia sp.	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Eutomopeas layardi	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Leptinaria lamellata	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Paropeas achatinaceum	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Riebeckia sp.	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Rumina decollata	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA	
Subulina octona	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Subulina striatella	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Subulina vitrea	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Subulona sp.	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Tortaxis erectus	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Xerocerastus sp.	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA	
Zootecus insularis	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA	
Coeliaxis blandii	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Pyrgina umbilicata	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Glessula ceylanica	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Ceciloides gokweanus	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Ferussacia folliculus	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA	
Thyrophorella thomensis	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Gibbulinella dewinteri	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAA	
Gonaxis quadrilateralis	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
Gonospira sp.	GGAACGTGAG	CTGGGTTTAT	ACCGTCGTGA	GACAGGTTAG	TTTTACCCTA	CTGATGACAG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	4210	4220	4230	4240	4250	4260	
<i>Achatina fulica</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Achatina achatina</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Achatina stuhlmanni</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Cochlitoma ustulata</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Allopeas clavulinum</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Bocageia</i> sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Eutomopeas layardi</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Leptinaria lamellata</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Paropeas achatinaceum</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Riebeckia</i> sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Rumina decollata</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Subulina octona</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Subulina striatella</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Subulina vitrea</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Subulona</i> sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Tortaxis erectus</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Xerocerastus</i> sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Zootecus insularis</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Coeliaxis blandii</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Pyrgina umbilicata</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Glessula ceylanica</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Cecilioides gokweanus</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Ferussacia folliculus</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Thyrophorella thomensis</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Gibbulinella dewinteri</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Gonaxis quadrilateralis</i>	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
<i>Gonospira</i> sp.	GTCGTTGCTA	CGGTAATCCT	GCTCAGTACG	AGAGGAACCG	CAGGTTTCAGA	CATTTGGTTC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	4270	4280	4290	4300	4310		
<i>Achatina fulica</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Achatina achatina</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Achatina stuhlmanni</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Cochlitoma ustulata</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Allopeas clavulinum</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Bocageia</i> sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Eutomopeas layardi</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Leptinaria lamellata</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Paropeas achatinaceum</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Riebeckia</i> sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Rumina decollata</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Subulina octona</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Subulina striatella</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Subulina vitrea</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Subulona</i> sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Tortaxis erectus</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Xerocerastus</i> sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Zootecus insularis</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Coeliaxis blandii</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Pyrgina umbilicata</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Glessula ceylanica</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Cecilioides gokweanus</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Ferussacia folliculus</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Thyrophorella thomensis</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Gibbulinella dewinteri</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Gonaxis quadrilateralis</i>	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
<i>Gonospira</i> sp.	ACGTGCTTGG	CTGATAAGCC	AATGGTGCGA	GGCTACCATC	TGAGGGATTA	TGG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

Appendix 3.2: Alignment of the actin gene for the Achatinoidea and three streptaxid outgroup taxa.

	10	20	30	40	50	60
<i>Achatina achatina</i>	TCCAAGAGAG	GTATYCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Achatina fulica</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Achatina stuhlmanni</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Cochlitoma ustulata</i>	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Allopeas clavulinum</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGAATTGT	CACCAACTGG
<i>Bocageia</i> sp.	TCCAAGAGAG	GTATCCTCAC	ACTCAAGTAY	CCCATTGARC	ATGGTATTGT	CACCAACTGG
<i>Eutomopeas layardi</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGAATTGT	CACCAACTGG
<i>Leptinaria lamellata</i>	TCCAAGAGAG	GTATCCTAAC	TCTCAAGTAT	CCCATTGAGC	ATGGCATTGT	CACCAACTGG
<i>Paropeas achatinaceum</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGAATTGT	CACCAACTGG
<i>Riebeckia</i> sp.	TCYAAGAGAA	GKATCCTCAC	CYTCAAGTAC	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Rumina decollata</i>	TCYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Subulina octona</i>	TCTAAGAGAG	GTATACTTAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Subulina striatella</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	AYGGTATTGT	CACCAACTGG
<i>Subulina vitrea</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAY	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Subulona</i> sp.	TCCAAGAGAG	GTATCCTCAC	TCTAAAGTAC	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Tortaxis erectus</i>	TCTAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGARC	ATGGTATTGT	CACCAACTGG
<i>Xerocerastus</i> sp.	TCCAAGAGAG	GTATCCTCAC	CCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Zootecus insularis</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Coeliaxis blandii</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Pyrgina umbilicata</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGWATTGT	CACCAACTGG
<i>Glessula ceylanica</i>	TCCAAGAGAG	GTATCCTCAC	TCTYAAGTAT	CCCATTGAGC	ATGGTRTTGT	CACCAACTGG
<i>Ceciloides gokweanus</i>	TCCAAGAGAG	GTATCCTCAC	ACTCAAGTAT	CCCATTGAGC	ACGGYATTGT	CACCAACTGG
<i>Ferussacia folliculus</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAC	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Thyrophorella thomensis</i>	TCYAAGAGAG	GTATCCTTAC	TCTCAARTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
<i>Gibbulinella dewinteri</i>	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAC	CCTATTGAAC	ATGGCATTGT	CACCAACTGG
<i>Gonaxis quadrilateralis</i>	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAC	CCCATTGAGC	ACGGCATTGT	CACCAACTGG
<i>Gonospira</i> sp.	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAC	CCCATTGAGC	ATGGCATTGT	CACCAACTGG

	70	80	90	100	110	120
<i>Achatina achatina</i>	GATGATATGG	AGAAGATCTG	GCATCACACY	TTCTACAATG	AGTTGAGAGT	TGCWCCAGAA
<i>Achatina fulica</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGTTSAGAGT	TGCACCAGAA
<i>Achatina stuhlmanni</i>	GATGATATGG	AGAAAATATG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
<i>Cochlitoma ustulata</i>	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
<i>Allopeas clavulinum</i>	GACGATATGG	AGAAGATCTG	GCATCACACT	TTCTACAACG	AGCTCAGAGT	TGCTCCTGAA
<i>Bocageia</i> sp.	GAYGATATGG	AGAAGATYTG	GCAAYCAYCY	TTCTACAAYG	AGCTKAGAGT	TGCWCCTGAA
<i>Eutomopeas layardi</i>	GACGATATGG	AGAAGATCTG	GCATCACACT	TTCTACAACG	AGCTCAGAGT	TGCTCCTGAA
<i>Leptinaria lamellata</i>	GACGATATGG	AGAAGATCTG	GCATCACACT	TTCTACAACG	AGCTCAGAGT	TGCTCCTGAA
<i>Paropeas achatinaceum</i>	GATGATATGG	AAAAGATCTG	GCATCACACT	TTCTACAACG	AGCTCAGAGT	TGCTCCTGAA
<i>Riebeckia</i> sp.	GAYGAYATGG	AGAAGATCTG	GCATCACACY	TTCTACAACG	AGCTGAGAGT	TGCWCCTGAA
<i>Rumina decollata</i>	GAYGATATGG	AGAAGATCTG	GCATCACACY	TTCTACAATG	AGCTGAGAGT	TGCWCCTGAA
<i>Subulina octona</i>	GATGATATGG	AAAAGATCTG	GCATCACACT	TTCTACAATG	AGCTGAGAGT	TGCTCCTGAA
<i>Subulina striatella</i>	GATGATATGG	AGAAGATCTG	GCATCACACT	TTCTACAATG	ARCTGAGAGT	TGCTCCTGAA
<i>Subulina vitrea</i>	GATGATATGG	AGAAGATCTG	GCATCACACY	TTCTACAATG	ARCTGAGAGT	TGCCCCAGAA
<i>Subulona</i> sp.	GACGATATGG	AGAAGATCTG	GCACCATACT	TTCTACAACG	AGCTTAGAGT	TGCTCCAGAA
<i>Tortaxis erectus</i>	GATGACATGG	AGAAAATTG	GCATCACACT	TTCTACAATG	AGCTCAGAGT	TGCACCTGAA
<i>Xerocerastus</i> sp.	GATGATATGG	AGAAGATCTG	GCATCATACC	TTCTACAATG	AGCTGAGAGT	TGCCCCTGAG
<i>Zootecus insularis</i>	GATGATATGG	AGAAGATCTG	GCATCACACY	TTCTACAATG	AGCTGAGAGT	TGCCCCTGAA
<i>Coeliaxis blandii</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCCCCTGAA
<i>Pyrgina umbilicata</i>	GAYGAYATGG	AGAAGATCTG	GCAAYAYCY	TTCTACAACG	AGCTSAGAGT	TGCYCCTGAA
<i>Glessula ceylanica</i>	GATGACATGG	AGAAGATCTG	GCACCATACT	TTCTACAATG	AGCTCAGAGT	TGCCCCTGAA
<i>Ceciloides gokweanus</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAACG	AGCTGAGAGT	TGCCCCAGAA
<i>Ferussacia folliculus</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTCAGAGT	TGCCCCAGAG
<i>Thyrophorella thomensis</i>	GAYGAYATGG	AGAAGATCTG	GCATCAYCY	TTCTACAACG	AGCTSAGAGT	TGCCCCTGAA
<i>Gibbulinella dewinteri</i>	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAACG	AGCTGAGAGT	TGCCCCTGAG
<i>Gonaxis quadrilateralis</i>	GATGACATGG	AGAAGATCTG	GCATCACACC	TTCTACAACG	AGCTGAGAGT	TGCCCCTGAG
<i>Gonospira</i> sp.	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAACG	AGTTGAGAGT	TGCCCCTGAG

	130	140	150	160	170	180	
<i>Achatina achatina</i>	GAGCATCCAG	TTCTRCTKAC	AGAGGCTCCA	CTCAATCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Achatina fulica</i>	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Achatina stuhlmanni</i>	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Cochlitoma ustulata</i>	GAGCATCCAG	TTCTGCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Allopeas clavulinum</i>	GAACATCCAG	TTTTACTTAC	AGAAGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Bocageia sp.</i>	GARCAVCCAG	TWYTKYTKAC	AGAGGCTCCA	CTCAAYCCMA	AGGCCAACAG	AGARAAGATG	
<i>Eutomopeas layardi</i>	GAGCATCCAG	TTTTACTCAC	AGAAGCTCCA	CTCAACCCCA	AGGCCAATAG	AGAGAAGATG	
<i>Leptinaria lamellata</i>	GAGCATCCAG	TTTTACTCAC	AGAGGCTCCA	CTGAATCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Paropeas achatinaceum</i>	GAGCATCCAG	TTTTACTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAATCG	AGAAAAGATG	
<i>Riebeckia sp.</i>	GARCAVCCAG	TAYTTCGTAC	AGAGGCTCCM	CTCAATCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Rumina decollata</i>	GAGCACCAG	TYTSCCTKAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Subulina octona</i>	GAGCATCCAG	TTTTGCTGAC	AGAAGCACCA	CTCAACCCCA	AGGCCAATAG	AGAGAAGATG	
<i>Subulina striatella</i>	GAGCATCCAG	TTTTGCTGAC	AGAAGCACCA	CTCAACCCCA	AGGCCYAACAG	AGAGAARATG	
<i>Subulina vitrea</i>	GAGCATCCAG	TTTTGCTGAC	AGAAGCACCA	CTCAACCCCA	AGGCCYAACAG	AGAGAAGATG	
<i>Subulona sp.</i>	GAGCATCCAG	TTTTGCTGAC	AGAAGCACCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Tortaxis erectus</i>	GAACATCCAG	TTCTCCTTAC	AGAGGCTCCA	CTCAATCCCA	AGGCCAACAG	AGAAAAGATG	
<i>Xerocerastus sp.</i>	GAGCACCAG	TCCTCCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Zootecus insularis</i>	GAGCACCAG	TCCTCCTGAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Coelioxaxis blandii</i>	GAGCACCAG	TACTTCTGAC	AGAGGCTCCA	CTCAATCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Pyrgina umbilicata</i>	GARCAVCCAG	TWYTMCTCAC	AGAGGCTCCW	CTTAACCCCA	AAGCCAAYAG	AGARAAGATG	
<i>Glessula ceylanica</i>	GAACAYCCAG	TYTTCCTYAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Cecilioides gokweanus</i>	GAGCACCAG	TACTCCTTAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Ferussacia folliculus</i>	GAGCACCAG	TCCTCCTCAC	AGAGGCTCCA	CTCAACCCCA	AGGCCAACAG	AGAGAAGATG	
<i>Thyrophorella thomensis</i>	GAGCAYCCAG	TWTCCTCAC	AGAGGCTCCA	CTTAACCCCA	AAGCTAAYAG	AGAAAAGATG	
<i>Gibbulinella dewinteri</i>	GAACACCAG	TCCTACTCAC	AGAGGCTCCA	CTCAACCCCA	AAGCYAACAG	AGAGAAGATG	
<i>Gonaxis quadrilateralis</i>	GAACACCAG	TYCTACTCAC	AGAGGCTCCW	CTCAATCCCA	AAGCCAACAG	AGAGAAGATG	
<i>Gonospira sp.</i>	GAACACCAG	TCCTACTCAC	AGAGGCTCCW	CTCAATCCCA	AAGCCAAYAG	AGAGAAGATG	

	190	200	210	220	230	240	
<i>Achatina achatina</i>	ACCCAGATCA	TGTTTGAGAC	YTTCAAWSY	CCAGCYATGT	ATGTYGCIAT	TCAAGCNGTG	
<i>Achatina fulica</i>	ACCCAGATCA	TGTTTGAAAC	YTTCAAAYTCT	CCAGCCATGT	ATGTCGCCAT	YCAAGCYGTG	
<i>Achatina stuhlmanni</i>	ACCCAGATCA	TGTTTGAAAC	YTTCAAAYTCT	CCAGCCATGT	ATGTTGCCAT	TCAAGCAGTC	
<i>Cochlitoma ustulata</i>	ACCCAGATCA	TGTTTGAGAC	CTTCAACTCT	CCAGCCATGT	ATGTCGCCAT	TCAAGCCGTG	
<i>Allopeas clavulinum</i>	ACACAAATTA	TGTTTGAAAC	CTTCAACTACT	CCAGCCATGT	ATGTCGCCAT	CCAAGCCGTT	
<i>Bocageia sp.</i>	ACCCAGATCA	TGTTTGAGAC	YTTCAAAYACY	CCAGCYATGT	ACGTSKCIAT	CCARGCTGTS	
<i>Eutomopeas layardi</i>	ACACAGATTA	TGTTTGAAAC	CTTCAACTACT	CCAGCCATGT	ATGTCGCCAT	CCAAGCCGTT	
<i>Leptinaria lamellata</i>	ACCCAGATTA	TGTTTGAGAC	CTTCAACTACT	CCAGCAATGT	ACGTCGCCAT	TCAAGCTGTG	
<i>Paropeas achatinaceum</i>	ACACAGATTA	TGTTTGAAAC	CTTCAACTACT	CCAGCCATGT	ATGTCGCCAT	CCAAGCCGTT	
<i>Riebeckia sp.</i>	ACCCAGATCA	TGTTTGAGAC	YTTCAAACWCY	CCAGCCATGT	AYGTCGCIAT	YCARGCWGTY	
<i>Rumina decollata</i>	ACACAGATCA	TGTTTGAGAC	CTTCAAAYACC	CCAGCNATGT	AYGTCGCIAT	YCARGCCGTG	
<i>Subulina octona</i>	ACACAGATCA	TGTTTGAGAC	ATTCAAACACC	CCAGCTATGT	ATGTTGCTAT	CCAGGCTGTA	
<i>Subulina striatella</i>	ACACAGATCA	TGTTTGAGAC	ATTCAAAYTCY	CCAGCCATGT	ATGTTGCTAT	YCARGCTGTS	
<i>Subulina vitrea</i>	ACCCAGATCA	TGTTTGAGAC	CTTCAAACACA	CCAGCCATGT	ATGTTGCIAT	CCAGGCCGTA	
<i>Subulona sp.</i>	ACCCAGATCA	TGTTTGAGAC	CTTCAACTCT	CCAGCCATGT	ACGTTGCAAT	TCAAGCCGTG	
<i>Tortaxis erectus</i>	ACCCAGATAA	TGTTTGAGAC	CTTCAAAYWCT	CCAGCGATGT	ATGTTGCMAT	CCAGGCAGTT	
<i>Xerocerastus sp.</i>	ACACAGATCA	TGTTTGAAAC	CTTCAACTCT	CCAGCCATGT	ATGTTGCAAT	TCAAGCAGTG	
<i>Zootecus insularis</i>	ACACAGATCA	TGTTTGAAAC	CTTCAACTACT	CCAGCAATGT	ATGTTGCCAT	CCAGGNCNGTG	
<i>Coelioxaxis blandii</i>	ACCCAGATCA	TGTTTGAGAC	CTTCAAACACC	CCAGCCATGT	ACGTCGCCAT	CCAGGCCGTA	
<i>Pyrgina umbilicata</i>	ACCCAGATTA	TGTTTGAGAC	CTTCAAAYWCY	CCAGCCATGT	ACGTTGCCAT	CCAAGCTGTG	
<i>Glessula ceylanica</i>	ACCCAGATMA	TGTTTGAGAC	CTTCAACTCT	CCAGCMATGT	WTGTTGCMAT	TCAAGCCGTA	
<i>Cecilioides gokweanus</i>	ACCCAGATCA	TGTTTGAGAC	CTTCAACTACT	CCAGCCATGT	ACGTTGCCAT	CCAGGCCGTA	
<i>Ferussacia folliculus</i>	ACACAGATCA	TGTTTGAAAC	CTTCAACTCT	CCAGCAATGT	ACGTCGCCAT	TCAAGCCGTA	
<i>Thyrophorella thomensis</i>	ACCCAGATTA	TGTTTGAGAC	YTTCAAAYWCT	CCAGCYATGT	AYGTCGCTAT	CCAAGCTGTC	
<i>Gibbulinella dewinteri</i>	ACTCAGATCA	TGTTTGAGAC	ATTCAAACTACT	CCAGCCATGT	ATGTCGCCAT	CCAGGCCGTA	
<i>Gonaxis quadrilateralis</i>	ACCCAGATCA	TGTTTGAAAC	ATTCAAACTCW	CCAGCCATGT	ATGTCGCIAT	CCAGGCAGTY	
<i>Gonospira sp.</i>	ACCCAGATCA	TGTTTGAGAC	ATTYAAACWCT	CCAGCCATGT	ATGTYGCSAT	YCAGGNCNGTT	

	250	260	270	280	290	300	
<i>Achatina achatina</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	WGATGGTGM	
<i>Achatina fulica</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	TGATGGTGM	
<i>Achatina stuhlmanni</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	TGATGGTGM	
<i>Cochlitoma ustulata</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	TGATGGTGM	
<i>Allopeas clavulinum</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Bocageia sp.</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Eutomopeas layardi</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Leptinaria lamellata</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Paropeas achatinaceum</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Riebeckia sp.</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Rumina decollata</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Subulina octona</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Subulina striatella</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Subulina vitrea</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Subulona sp.</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Tortaxis erectus</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Xerocerastus sp.</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Zootecus insularis</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Coelioxaxis blandii</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Pyrquina umbilicata</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Glessula ceylanica</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Ceciloides gokweanus</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Ferussacia folliculus</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Thyrophorella thomensis</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Gibbulinella dewinteri</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Gonaxis quadrilateralis</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	
<i>Gonospora sp.</i>	CTTTCYTRT	ATGCTTCAGG	TCGTACAAC	GGTATTGK	TGGATTCTG	AGATGGTGM	

	310	320	330	340	350	360	
<i>Achatina achatina</i>	ACYCAYACTG	TNCCAATMTA	TGAAGGTTAT	GCTCTCCY	AYGCCATCAT	GAGACTGGAY	
<i>Achatina fulica</i>	ACCCACACTG	TCCCAATTTA	TGAAGGTTAT	GCTCTACCT	ATGCCATCAT	GAGACTGGAC	
<i>Achatina stuhlmanni</i>	ACCCACACTG	TCCCAATTTA	TGAAGGTTAT	GCTCTACCT	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma ustulata</i>	ACTCACACTG	TCCCAATTTA	TGAAGGTTAT	GCTCTACCT	ATGCCATCAT	GAGACTGGAC	
<i>Allopeas clavulinum</i>	ACTCACACTG	TCCCAATTTA	TGAAGGTTAT	GCTCTACCT	ATGCCATCAT	GAGACTGGAC	
<i>Bocageia sp.</i>	WYCYAYACWG	TSCCYATMTA	TGAAGGTTAT	GCCCTTCCT	ATGCCATCAT	GAGRYTKGAC	
<i>Eutomopeas layardi</i>	ACTCATACTG	TCCCATCTA	TGAAGGTTAT	GCTCTCCAC	ACGCCATCAT	GAGATTAGAC	
<i>Leptinaria lamellata</i>	ACTCATACTG	TCCCATCTA	TGAAGGTTAT	GCCCTTCCT	ATGCTATAT	GAGACTGGAC	
<i>Paropeas achatinaceum</i>	ACTCATACTG	TCCCATCTA	TGAAGGTTAT	GCTCTCCAC	ATGCCATCAT	GAGATTAGAC	
<i>Riebeckia sp.</i>	ACACACACTG	TCCCATCTA	TGAAGGTTAT	GCMTTCCCC	AYGCCATCAT	GAGACTGGAT	
<i>Rumina decollata</i>	ACCCAYACTG	TCCCATCTA	TGAAGGTTAT	GCCCTTCCT	ATGCCATCAT	GAGAYTGGAC	
<i>Subulina octona</i>	ACCCACACTG	TCCCATCTA	TGAAGGTTAT	GCTCTCCCT	ATGCCATCAT	GAGACTGGAC	
<i>Subulina striatella</i>	ACCCACACTG	TCCCATCTA	TGAAGGTTAT	GCTCTCCCT	ACGCCATCAT	GAGACTGGAC	
<i>Subulina vitrea</i>	ACTCACACTG	TYCCATCTA	TGAAGGTTAT	GCTCTCCCT	ATGCCATCAT	GAGACTGGAT	
<i>Subulona sp.</i>	ACCCATACTG	TCCCATCTA	TGAAGGTTAT	GCTCTCCAC	ATGCCATCAT	GAGACTGGAC	
<i>Tortaxis erectus</i>	ACCCACACTG	TCCCATCTA	TGAAGGTTAT	GCCCTTCCT	ATGCCATCAT	GAGACTGGAC	
<i>Xerocerastus sp.</i>	ACCCATACTG	TCCCATATA	TGAAGGTTAT	GCCCTTCCT	ATGCCATCAT	GAGACTGGAC	
<i>Zootecus insularis</i>	ACCCACACTG	TCCCYATCTA	TGAAGGTTAT	GCCCTTCCT	ATGCCATCAT	GAGACTGGAY	
<i>Coelioxaxis blandii</i>	ACWCACACTG	TCCCATCTA	TGAAGGTTAT	GCCCTTCCT	ATGCCATCAT	GAGACTGGAT	
<i>Pyrquina umbilicata</i>	ACTCAYACWG	TCCCYATCTA	TGAAGGTTAT	GCTCTCCCT	ACGCCATYAT	GAGACTRGAT	
<i>Glessula ceylanica</i>	ACCCACACTG	TCCCATYTW	TGAAGGTTAT	GCCCTTCCT	ATGCCATCAC	GAGATTGGAC	
<i>Ceciloides gokweanus</i>	ACTCACACTG	TCCCATCTA	TGAAGGTTAT	GCTCTCCCT	ATGCCATCAT	GAGACTGGAC	
<i>Ferussacia folliculus</i>	ACCCACACTG	TCCCATCTA	TGAAGGTTAT	GCTTTGCCC	ACGCCATCAT	GAGACTGGAT	
<i>Thyrophorella thomensis</i>	ACTCAYACWG	TCCCYATMTA	TGAAGGTTAT	GCTCTCCCT	ACGCCATYAT	GAGAYTGGAT	
<i>Gibbulinella dewinteri</i>	ACCCACACTG	TCCCATCTA	TGAAGGTTAT	GCTCTCCCT	ACGCCATCAT	GAGACTGGAT	
<i>Gonaxis quadrilateralis</i>	ACCCACACTG	TCCCYATMTA	TGAAGGTTAT	GCTCTCCCT	AYGCCATCAT	GAGACTGGAY	
<i>Gonospora sp.</i>	ACCCAYACAG	TCCCYATCTA	TGAAGGTTAT	GCCCTTCCT	AYGCCATYAT	GAGATTGGAT	

	370 380 390 400 410 420
<i>Achatina achatina</i>	TTGGCTGGTC GTGACCTCAC AGATTATCTC ATGAARATCC TCACAGAGAG AGGTTACAGC
<i>Achatina fulica</i>	TTGGCTGGTC GTGACCTCAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC
<i>Achatina stuhlmanni</i>	TTGGCTGGTC GTGACCTCAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC
<i>Cochlitoma ustulata</i>	TTGGCTGGTC GTGACCTCAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC
<i>Allopeas clavulinum</i>	TTGGCTGGCC GTGACCTCAC CGACTACCTC ATGAAGATTC TGAAGATCC AGGATAACAGC
<i>Bocageia sp.</i>	YTGGCTGGYC GTGACCTTAC AGAYTACCTS ATGAAGATYC TSACWGAGAG AGGCTACAGC
<i>Eutomopeas layardi</i>	TTGGCTGGCC GTGACCTCAC TGACTACCTC ATGAAGATCC TCACTGAGCG AGGATAACAGC
<i>Leptinaria lamellata</i>	TTGGCTGGTC GTGATCTCAC AGATTATTTG ATGAAGATTC TAACAGAGAG AGGCTACAGC
<i>Paropeas achatinaceum</i>	TTGGCTGGCC GTGACCTCAC TGACTACCTC ATGAAGATTC TCACTGAGCG AGGCTACAGC
<i>Riebeckia sp.</i>	CTGGCYGGCC GTGACCTCAC AGACTACCTC ATGAAGATCC TCACAGAGAG GGGCTACAGC
<i>Rumina decollata</i>	TTGGCTGGTC GTGACCTYAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC
<i>Subulina octona</i>	TTGGCTGGTC GCGACCTGAC AGATTATCTA ATGAAGATTC TCACGGAGAG AGGTTACAGC
<i>Subulina striatella</i>	TTGGCTGGTC GAGACCTGAC AGATTATCTG ATGAAGATTC TCACAGAGAG AGGTTACAGC
<i>Subulina vitrea</i>	CTGGCTGGCC GTGACCTCAC AGACTAYCTC ATGAAGATCC TGACAGAGAG AGGCYACAGC
<i>Subulona sp.</i>	TTGGCTGGTC GAGACCTCAC AGACTACCTC ATGAAGATCC TCACTGAGAG AGGTTACAGC
<i>Tortaxis erectus</i>	TTGGCTGGCC GTGACCTCAC AGATTACCTC ATGAARATCC TCACAGAGAG AGGTTACAGC
<i>Xerocerastus sp.</i>	TTGGCTGGTC GTGACCTCAC AGATTACCTC ATGAAAATCC TCACAGAGAG AGGCTACAGC
<i>Zootecus insularis</i>	TTGGCTGGTC GTGACCTCAC AGATTAYCTC ATGAAGATCC TCACAGAGAG AGGCTAYAGY
<i>Coeliaxis blandii</i>	CTGGCTGGCC GTGACCTCAC AGACTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC
<i>Pyrgina umbilicata</i>	CTGGCTGGCC GTGACCTCAC AGAYTAYCTC ATGAAGATCC TMACAGAGMG AGGMTACAGC
<i>Glessula ceylanica</i>	CTGGCTGGAC GAGACCTCAC AGATTATCTC ATGAAGATTC TCACAGAGAG AGGCTACCCC
<i>Cecilioides gokweanus</i>	TTGGCTGGCC GTGACCTTAC AGATTACCTC ATGAAGATCC TCACAGAGAG AGGCTACAGC
<i>Ferussacia folliculus</i>	CTGGCTGGTC GTGATCTCAC AGATTACCTC ATGAAGATCC TYACAGAGAG AGGCTACAGC
<i>Thyrophorella thomensis</i>	CTRGCTGGCC GTGAYCTCAC AGATTAYCTC ATGAAGATCC TCACAGAGAG AGGATAACAGC
<i>Gibbulinella dewinteri</i>	CTGGCTGGCC GTGACCTTAC AGATTAYCTG ATGAAGATCC TCACAGAGAG AGGCTACAGT
<i>Gonaxis quadrilateralis</i>	TTGGCYGGGC GTGACCTTAC AGATTACCTG ATGAAGATCC TCACAGAGAG AGGCTACAGC
<i>Gonospira sp.</i>	TTGGCTGGAC GTGACCTKAC AGATTAYYTG ATGAAGATCC TCACAGAGAG AGGCTAYAGC

	430 440 450 460 470 480
<i>Achatina achatina</i>	TTCACCACCA CAGCTGAGAG AGAAATTGTT CGAGAYATCA ARGAGAAGCT GTGCTATGTG
<i>Achatina fulica</i>	TTCACCACCA CAGCTGAGAG AGAAATTGTT CGAGACATCA AAGAGAAGCT TTGCTATGTG
<i>Achatina stuhlmanni</i>	TTCACCACCA CAGCTGAGAG AGAAATTGTT CGAGACATCA AAGAGAAGCT TTGCTATGTG
<i>Cochlitoma ustulata</i>	TTCACCACCA CAGCTGAGAG AGAAATTGTT CGAGACATCA AAGAGAAGCT TTGCTATGTG
<i>Allopeas clavulinum</i>	TTCACTACAA CAGCCGAGAG AGAAATTGTA CGTGACATCA AGGAGAAACT TTGCTACGTT
<i>Bocageia sp.</i>	TTCACYACMA CWCWGAGAG AGAAATTGTT CGAGAYATCA AGGAAAAACT TTSCTATGTY
<i>Eutomopeas layardi</i>	TTCACTACAA CAGCCGAGAG AGAAATTGTA CGCGACATCA AGGAGAAACT TTGCTACGTT
<i>Leptinaria lamellata</i>	TTCACAACAA CCGCTGAGAG AGAAATTGTA CGTGACATCA AGGAGAAACT TTGCTATGTT
<i>Paropeas achatinaceum</i>	TTCACAACAA CAGCCGAGAG AGAAATTGTA CGTGACATCA AAGAGAAACT TTGCTACGTT
<i>Riebeckia sp.</i>	TTCACCACCA CTGCTGAGAG AGAAATTGTT CGAGACATCA AAGARAAGCT TTSYATYGR
<i>Rumina decollata</i>	TTCACCACCA CTGCTGAGAG AGAAATTGTT CGAGACATCA AGGAGAAGCT GTSCTATGTT
<i>Subulina octona</i>	TTCACAACCA CAGCTGAGAG AGAAATTGTT CGAGACATCA AGGAGAAACT TTGCTACGTC
<i>Subulina striatella</i>	TTCACCACCA CAGCYGAGAG AGAAATTGTY CGAGACATCA AAGAGAAGCT TTGCTACGTC
<i>Subulina vitrea</i>	TTCACCACCA CWCCTGAGAG AGAAATTGTT CGAGACATCA AGGAGAAACT CTGYTATGTT
<i>Subulona sp.</i>	TTCACCACCA CCGCCGAGAG AGAAATTGTT CGAGACATCA AGGAGAAACT CTGCTACGTC
<i>Tortaxis erectus</i>	TTCACYACYA CAGCYGAGAG AGAAATTGTT CGAGAYATCA AGGAGAAGCT KTSCTAYGTK
<i>Xerocerastus sp.</i>	TTCACCACCA CCGCTGAGAG AGAAATTGTT CGAGACATCA AGGAGAAGCT TTGCTATGTT
<i>Zootecus insularis</i>	TTCACCACCA CCGCTGAGAG AGAAATTGTT CGAGACATCA AGGARAAGCT TTSCTATGTT
<i>Coeliaxis blandii</i>	TTCACCACCA CTGCTGAGAG AGAAATYGTG MGAGACATCA ARGAGAAGCT TTSCTATGTT
<i>Pyrgina umbilicata</i>	TTYACTACCA CAGCYGAGAG AGAAATTGTT MWGACATCA ARGAGAARCT KTYTAYGTK
<i>Glessula ceylanica</i>	TTCACYACMA CAGCCGAGAG GRAAATYGTK CGAGACATGA AAGAGAAGCT TGCGTATGTT
<i>Cecilioides gokweanus</i>	TTCACCACCA CAGCCGAGAG GGAATGTC CGAGACATCA AGGAGAAGCT TTCATACGTT
<i>Ferussacia folliculus</i>	TTCACCACCA CTGCTGAGAG AGAAATGTS AGGGAYATCA AGGAGAAGCT CTGTTATGTT
<i>Thyrophorella thomensis</i>	TTYACTACCA CAGCYGAGAG AGAAATGTT MGRGAYATCA ARGAGAARCT NTSYATACGTK
<i>Gibbulinella dewinteri</i>	TTCACCACCA CTGCTGAGAG AGAGATTGTA CGAGACATCA AGGAGAAGCT CTGCTACGTT
<i>Gonaxis quadrilateralis</i>	TTCACCACCA CTGCTGAGAG AGAAATTGTA MGAGACATCA AGGAGAARCT CTGCTACGTT
<i>Gonospira sp.</i>	TTCACCACCA CTGCGWAGAG AGAGATYGTA CGAGACATMA ARGARAAGCT MTGYTATGTT

	490	500	510	520	530	540	
<i>Achatina achatina</i>	GCYCTTGACT	TTGAGCAAGA	GATGTCRACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGY	
<i>Achatina fulica</i>	GCCCTTGACT	TTGAGCAAGA	RATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Achatina stuhlmanni</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTGCCACAT	CATCTTYATT	AGAGAAGAGT	
<i>Cochlitoma ustulata</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Allopeas clavulinum</i>	GCCCTAGATT	TTGAGCAGGA	GATGGGAACA	GCTGCTACAT	CGTCATCTTT	GGAGAAGAGC	
<i>Bocageia sp.</i>	GCTCTTGACT	TTGAACAGGA	AATGSACACA	GCTGCCWCAT	CATCTTCYCT	RGAGAAGAGC	
<i>Eutomopeas layardi</i>	GCTCTGGACT	TTGAGCAGGA	GATGGGAACA	GCTGCTACAT	CATCATCGTT	GGAAAAGAGC	
<i>Leptinaria lamellata</i>	GCTCTTGATT	TTGAGCAGGA	GATGGGTACA	GCTGCAACGT	CATCATCATT	AGAGAAGAGC	
<i>Paropeas achatinaceum</i>	GCCCTAGATT	TCGAACAGGA	GATGGGAACA	GCTGCCACAT	CATCATCTTT	GGAGAAGAGC	
<i>Riebeckia sp.</i>	GCTCTYGACT	TTGAGCAAGA	AAWGGSTACG	GCTGCCACAT	CWTCCCTCAYT	AGAGAAGAGT	
<i>Rumina decollata</i>	GCTCTTGACT	TTGAACAAGA	AATGGCAACA	GCTGCCACTT	CATCTTCWCT	AGAGAAGAGT	
<i>Subulina octona</i>	GCCCTTGACT	TTGAACAAGA	AATGGGTACA	GCTGCCACAT	CGTCATCTTT	AGAGAAGAGC	
<i>Subulina striatella</i>	GCCCTTGACT	TCGAACAAGA	AATGGGTACA	GCTGCCACAT	CCTCATCTTT	GGAGAAGAGT	
<i>Subulina vitrea</i>	GCTCTTGACT	TCGARCAAGA	AATGTCTACA	GCTGCCACRT	CWTCTTCATT	RGAGAAGAGT	
<i>Subulona sp.</i>	GCACTTGACT	TTGAGCAGGA	GATGCAGACA	GCTGCTACAT	CATCATCTTT	GGAGAAGAGC	
<i>Tortaxis erectus</i>	GCTCTAGACT	TTGAGCAGGA	GATGGCAACA	GCTGCCACTT	CATCTWCTCT	AGAGAAAAGT	
<i>Xerocerastus sp.</i>	GCTCTTGACT	TTGAACAAGA	AATGGCAACA	GCTGCCACTT	CATCTTCACT	AGAGAAGAGT	
<i>Zootecus insularis</i>	GCTCTTGAYT	TTGAACAAGA	AATGGCAACA	GCTGCCACAT	CATCTTCACT	AGAGAAGAGT	
<i>Coelioxaxis blandii</i>	GCTCTTGACT	TTGAGCAAGA	AATGGTACA	GCTGCYACAT	CATCCTCATT	AGAGAAGAGT	
<i>Pyrquina umbilicata</i>	GCTCTRGACT	TTGARCAAGA	RATGGCRACA	GCWGCYWCTT	CATCYWCYCT	NGAGAAGAGC	
<i>Glessula ceylanica</i>	GCTCTGGACT	TTGAMCARGA	GATGCAGWTA	TCATCCAGYK	CCTCAWCAAT	TGAARAAAGT	
<i>Ceciloides gokweanus</i>	GCWCTAGACT	TTGAGCAAGA	AATGGCAACA	GCTGCAACTT	CATCTTCACT	RGAGAAGAGC	
<i>Ferussacia folliculus</i>	GCTCTGGATT	TTGAACAGGA	GATGGCTACA	GCTGCAACTT	CCTCCTCCCT	GGAGAAGAGC	
<i>Thyrophorella thomensis</i>	GCTCTRGACT	TTGAGCAAGA	AATGGCAACA	GCTGCYACTT	CATCYTCYCT	GGAGAAGAGY	
<i>Gibbulinella dewinteri</i>	GCTCTGGACT	TTGAGCAGGA	GATGGCAACA	GCTGCTACAT	CATCATCATT	GGAGAAGAGC	
<i>Gonaxis quadrilateralis</i>	GCTCTGGACT	TTGAGCARGA	GATGGCMACA	GCTGCTACAT	CATCMTCCCT	TGAGAAGAGY	
<i>Gonospora sp.</i>	GCTCTGGACT	TTGAACARGA	GATGGSWACW	GCWGCWVCAT	CATCWTCCCT	TGAGAAGAGC	

	550	560	570	580	590	600	
<i>Achatina achatina</i>	TATGAATTGC	CTGATGGACA	RGTCATYACT	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA	
<i>Achatina fulica</i>	TATGAATTRC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	ATGTCCAGAA	
<i>Achatina stuhlmanni</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGATTTCAG	GTGCCAGAA	
<i>Cochlitoma ustulata</i>	TATGAATTAC	CTGATGGACA	GGTCATAACT	ATTGGTAATG	AGCGCTTCAG	GTGCCAGAA	
<i>Allopeas clavulinum</i>	TACGAACTTC	CAGATGGACA	GGTCATCACC	ATTGGCAACG	AGCGCTTCAG	ATGCCCTGAG	
<i>Bocageia sp.</i>	TAYGAACTTC	CTGATGGACA	GGTCATCACC	ATTGGCAATG	ARCGWTTTCAG	ATGTCCWGAR	
<i>Eutomopeas layardi</i>	TACGAGCTTC	CAGATGGACA	GGTCATCACA	ATCGGCAATG	AGCGCTTCAG	ATGCCCCGAG	
<i>Leptinaria lamellata</i>	TATGAGCTGC	CTGATGGACA	AGTCATTACC	ATTGGAAATG	AGCGCTTCAG	ATGCCCCGAG	
<i>Paropeas achatinaceum</i>	TACGAACTTC	CAGATGGACA	AGTTATCACA	ATTGGCAATG	AGCGCTTCAG	ATGCCCCGAA	
<i>Riebeckia sp.</i>	TAYGAAATGC	CTGATGGACA	AGTCATCACC	ATTGGYAAAG	ARCGKTTTCAG	GWGYCCAGAA	
<i>Rumina decollata</i>	TATGAATTGC	CTGAYGGACA	GGTCATCACC	ATCGGCAAYG	AGCGTTTCAG	ATGTCCAGAA	
<i>Subulina octona</i>	TACGAACTTC	CAGATGGACA	AGTGATTACC	ATTGGCAATG	AACGTTTCAG	ATGCCCTGAG	
<i>Subulina striatella</i>	TACGAACTTC	CAGATGGACA	AGTTATTACC	ATTGGCAATG	AACGTTTYAG	ATGCCCTGAG	
<i>Subulina vitrea</i>	TACGAGYTCG	CTGATGGACA	GGTCATCACC	ATTGGCAAYG	AGCGTTTCAG	ATGYCCWGAA	
<i>Subulona sp.</i>	TACGAACTGC	CAGATGGACA	AGTCATTACT	ATTGGTAACG	AGCGATTTCAG	ATGTCCCAGAA	
<i>Tortaxis erectus</i>	TATGAACTRC	CTGATGGACA	GGTCATCACC	ATTGGCAAYG	AAAGATTTCAG	RTGNCCAGAA	
<i>Xerocerastus sp.</i>	TACGAACTGC	CTGATGGACA	GGTCATCACC	ATTGGCAATG	AGCGCTTCAG	GTGCCAGAA	
<i>Zootecus insularis</i>	TATGAATTGC	CTGAYGGACA	GGTCATCACC	ATTGGCAATG	AGCGTTTCAG	RTGTCCAGAA	
<i>Coelioxaxis blandii</i>	TATGAATTGC	CTGATGGACA	AGTCATCACC	ATCGGTAAYG	AGCGTTTCAG	ATGCCCWGAA	
<i>Pyrquina umbilicata</i>	TACGARCTKC	CWGTGGACA	GGTCATCACC	ATTGGNAAAG	AGCGYTTTCAG	RTGTCCWGAR	
<i>Glessula ceylanica</i>	TACGARCTKC	CGGATGGAMA	SRTMATYACM	ATWGGAAACG	AGCGATTTCAG	RTGTCCCAGAA	
<i>Ceciloides gokweanus</i>	TACGAACTTC	CCGATGGTCA	AGTCATCACC	ATTGGCAACG	AGCGTTTCAG	ATGTCCAGAA	
<i>Ferussacia folliculus</i>	TACGAACTGC	CTGACGGACA	GGTCATCACC	ATTGGCAACG	AGCGATTTCAG	RGTCCCGAA	
<i>Thyrophorella thomensis</i>	TACGARCTNC	CWGTGGACA	GGTYATCACC	ATTGGWAAAG	AGCGTTTCAG	ATGTCCCAGAA	
<i>Gibbulinella dewinteri</i>	TAYGAACTTC	CAGACGGACA	GGTCATCACC	ATTGGCAATG	ARCGATTTCAG	GTGCCAGAA	
<i>Gonaxis quadrilateralis</i>	TATGAACTRC	CWYAYGGACA	GGTCATYACY	ATTGGMAACG	ARCGWTTTYAG	RTGCCAGAA	
<i>Gonospora sp.</i>	TATGAACTTC	CTGAYGGACA	GGTCATYACC	ATTGGCAAYG	ARCGWTTTYAG	GWSWCTGAA	

	610	620	630	640	650	660	
<i>Achatina achatina</i>	GCCATGTTCC	AGCCATCTTT	CCTTGGTATG	GARWCTGCAG	GTATTCATGA	AACCACCTAC	
<i>Achatina fulica</i>	GCCATGTTCC	AGCCWTCTTT	CCTTGGTATG	GARWCCGCAG	GTATTCATGA	RACCACYTAC	
<i>Achatina stuhlmanni</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCACGA	GACCACCTAC	
<i>Cochlitoma ustulata</i>	GCAGAATTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCAG	GTATTCATGA	GACCACCTAC	
<i>Allopeas clavulinum</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTACCCATGA	AACAACGTAC	
<i>Bocageia sp.</i>	GCAATGTTYC	AGCCATCTTT	CCTKGGAAATG	GAGTCTGCWG	GTATTCATGA	GACYAYCTAC	
<i>Eutomopeas layardi</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTACCCATGA	AACAACGTAC	
<i>Leptinaria lamellata</i>	GCAATGTTCC	AACCATCTTT	TCTTGGCATG	GAATCTGCTG	GCACACATGA	AACAACATAC	
<i>Paropeas achatinaceum</i>	GCAATGTTCC	AGCCATCATT	CCTTGGTATG	GAGTCTGCAG	GTACCCATGA	AACAACATAC	
<i>Riebeckia sp.</i>	GCAATGTTCC	AGCCATCTTT	YCTTGGYATG	GAGTCTGCTG	GTATTCAYGA	RACCACYTAC	
<i>Rumina decollata</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATTCATGA	GACCACCTAC	
<i>Subulina octona</i>	GCAATGTTTC	AGCCATCTTT	TCTGGGCATG	GAATCTGCAG	GTACTCATGA	AACAACGTAC	
<i>Subulina striatella</i>	GCAATGTTYC	AGCCATCTTT	TCTGGGCATG	GAATCTGCAG	GTACTCACGA	AACAACATAC	
<i>Subulina vitrea</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GARTCWGCTG	GTAYTCACGA	GACCACYTAC	
<i>Subulona sp.</i>	GCTGAGTTCC	AACCATCTTT	CCTGGGATG	GAGTCAGCTG	GCATTACCGA	AACTACCTAT	
<i>Tortaxis erectus</i>	GCWATGTTNC	ARCCATCTTT	TCTTGGTATG	GAATCTGCTG	GWATCCATGA	GACCACATAC	
<i>Xerocerastus sp.</i>	GCAGAATTCC	AACCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATTCATGA	GACCACCTAC	
<i>Zootecus insularis</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATYCATGA	GACCACCTAC	
<i>Coeliaxis blandii</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGWATG	GAATCTGCTG	GTATTCACGA	GACCACCTAC	
<i>Pyrgina umbilicata</i>	GCAATRTTCC	ARCCATCWTT	CCTTGGYATG	GARTCTGCTG	GTATTCATGA	AACCACCTAC	
<i>Glessula ceylanica</i>	GCTTTGTTCC	AGCCATCTTT	TCTAGGTMTA	GAGATCRSTG	GTATTCACGA	AACCACCTAC	
<i>Cecilioides gokweanus</i>	GCAATGTTCC	AGCCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATTCACGA	GACCACATAC	
<i>Ferussacia folliculus</i>	GCAGAATTTT	AGCCATCTTT	CCTTGGCATG	GAATCTGCCG	GTATTCACGA	AACAACCTAC	
<i>Thyrophorella thomensis</i>	GCAATRTTCC	ARCCATCNTT	CCTTGGTATG	GAGTCWGCTG	GTATTCATGA	AACCACCTAC	
<i>Gibbulinella dewinteri</i>	GCAATGTTCC	AACCATCTTT	CCTTGGTATG	GAGTCTGCTG	GTATCCACGA	GACCACCTAC	
<i>Gonaxis quadrilateralis</i>	GCAATGTTYC	AGCCATCWTT	CYTTGGTATG	GARTCTGCTG	GYRTCCAYGA	GACMACATAC	
<i>Gonospira sp.</i>	GCAATGTTYC	AGCCATCYTT	CYTSGGYATG	GAGTCTGCTG	GTRTYCAYGA	GACCACATAC	

	670	680	690	700	710	720	
<i>Achatina achatina</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTA	
<i>Achatina fulica</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Achatina stuhlmanni</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTACTT	
<i>Cochlitoma ustulata</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Allopeas clavulinum</i>	AATTCATCA	TGAAGTGTGA	TGTTGACATC	AGAAAAGACC	TCTATGCCAA	CACTGTCTGT	
<i>Bocageia sp.</i>	AAATCYATCA	TGAAGTGTGA	TGTYGAYATC	CGTAAAGAYY	TGTAYGCCAA	YAYTGTACTG	
<i>Eutomopeas layardi</i>	AATTCATCA	TGAAAATGCGA	TGTTGATATC	AGAAAAGACC	TCTATGCCAA	CACTGTCTGT	
<i>Leptinaria lamellata</i>	AATTCATTA	TGAAGTGTGA	TGTTGATATC	AGAAAAGACC	TCTATGCCAA	CACTGTACTA	
<i>Paropeas achatinaceum</i>	AATTCATTA	TGAAGTGTGA	TGTTGATATC	AGAAAAGACC	TTTATGCTAA	CACTGTATTG	
<i>Riebeckia sp.</i>	AAATCCWTYA	TGAARTGTGA	YGTYGACATY	CGTAAAGACT	TGTATGCCAA	CWCCGTCTTG	
<i>Rumina decollata</i>	AATTCATCA	TGAARTGTGA	TGTYGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTCTTG	
<i>Subulina octona</i>	AATTCATCA	TGAAGTGTGA	CGTTGATATC	CGAAAAGACC	TTTATGCTAA	CACTGTACTG	
<i>Subulina striatella</i>	AATTCATCA	TGAAAATGCGA	CGTCGATATC	CGAAAAGACC	TATATGCTAA	CACTGTACTG	
<i>Subulina vitrea</i>	AACTCCATCA	TGAAGTGTGA	TGTYGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTAYTG	
<i>Subulona sp.</i>	AACTCCATCA	TGAAGTGTGA	TGTTGACATC	AGAAAAGACC	TTTACGCCAA	TACTGTGTTG	
<i>Tortaxis erectus</i>	AACTCCATCA	TGAARTGTGA	TGTTGACATC	CGTAAAGACT	TGTATGCAAA	YACAGTCTGT	
<i>Xerocerastus sp.</i>	AACTCCATCA	TGAAGTGTGA	TGTTGACATT	CGTAAAGACT	TGTATGCCAA	CACTGTGTTG	
<i>Zootecus insularis</i>	AACTCCATCA	TGAARTGCGA	TGTTGACATY	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Coeliaxis blandii</i>	AACTCCATCA	TGAAGTGTGA	CGTAGACATY	CGTAAAGACT	TGTACKCCAA	CAYCGTATTG	
<i>Pyrgina umbilicata</i>	AACTCCATCA	TGAAAATGYGA	YGTYGACATY	CGTAAAGAYT	TGTATGCCAA	CACTGTATTG	
<i>Glessula ceylanica</i>	AACTCCATMA	TSAAAATGTGA	CGTCGACATC	CGTAAAGACC	TGTAYGCMMA	CACWGTCTYTR	
<i>Cecilioides gokweanus</i>	AACTCCATCA	TGAAGTGTGA	CGTTGACATC	CGTAAAGACT	TGTATGCCAA	CACTGTATTG	
<i>Ferussacia folliculus</i>	AACTCCATCA	TGAAGTGTGA	CGTCGACATC	CGTAAAGACT	TGTACGCCAA	CACTGTCTCT	
<i>Thyrophorella thomensis</i>	AAATCYATCA	TGAAAATGCGA	CGTYGAYATY	CGWAAAAGACY	TGTATGCCAA	CACTGTATTG	
<i>Gibbulinella dewinteri</i>	AACTCCATCA	TGAAGTGTGA	CGTTGACATC	CGTAAAGACT	TGTACGCCAA	CACCGTGCTG	
<i>Gonaxis quadrilateralis</i>	AACTCCATCA	TGAARTGCGA	YGTTGATATC	CGTAAAGAYT	TGTACKCCAA	CACYGNTTGT	
<i>Gonospira sp.</i>	AACTCCATCA	TGAARTGTGA	TGTRGAYATC	CGMAAAGACT	TGTATGCCAA	CACTGTCTGT	

	730 740 750 760 770 780
<i>Achatina achatina</i>	TCTGGTGGAW CTACCATGTA TCCTGGCATT GCTGACCGCA TGCAGAAGGA AATACNAAY
<i>Achatina fulica</i>	TCTGGTGGAW CTACCATGTA TCCAGGCATT GCTGACCGCA TGCAGAAGGA AATYGYCAAC
<i>Achatina stuhlmanni</i>	TCTGGTGGCT CCACAATGTT TCCAGGCATC GCTGACCGCA TGCAGAAGGA AATCACCAGC
<i>Cochlitoma ustulata</i>	TCTGGTGGCT CCACAATGTT CCCTGGCATT GCTGACCGCA TGCAGAAGGA AATCACCAGC
<i>Allopeas clavulinum</i>	TCTGGTGGTT CCACAATGTT CCCTGGCATT GCTGACCGCA TGCAGAAAGA GATAACCCTG
<i>Bocageia sp.</i>	TCTGGMGGTT CYACAATGTT YCCTGGMATY GCTGACCGCA TGCAGAAARGA AATSACCTCW
<i>Eutomopeas layardi</i>	TCTGGTGGTT CCACAATGTT CCCTGGCATT GCTGACCGCA TGCAGAAAGA GATAACCCTG
<i>Leptinaria lamellata</i>	TCTGGTGGTT CCACTATGTT CCCTGGCATT GCTGATCGTA TGCAGAAAGA AATCACTGCT
<i>Paropeas achatinaceum</i>	TCTGGTGGGT CCACAATGTT CCCAGGCATA GCTGACCGCA TGCAGAAAGA GATAACTGCT
<i>Riebeckia sp.</i>	TCYGGWGGST CCACCATGTT CCCTGGSATC GCTGACCGCA TGCAGAAGGA AATCACYKCT
<i>Rumina decollata</i>	TCTGGTGGNT CAACAATGTT CCCAGGCATC GCTGACCGCA TGCAGAAAGA AATCACCAMT
<i>Subulina octona</i>	TCTGGCGGTT CTACAATGTT CCCTGGCATT GCCGATCGCA TGCAGAAAGA AATCACCCTG
<i>Subulina striatella</i>	TCTGGCGGTT CWACAATGTT CCCTGGCATT GCCGACCGCA TGCAGAAAGA AATCACCAGT
<i>Subulina vitrea</i>	TCTGGTGGTT CYACAATGTT YCCTGGCATT GCCGACCGCA TGCAGAAAGA AATYRTCAAT
<i>Subulona sp.</i>	TCTGGAGGAT CCACAATGTT CCCAGGCATC GCTGACCGCA TGCAGAAAGA AATTACTTCT
<i>Tortaxis erectus</i>	TCTGGWGGTT CTACCATGTW CCCTGGARTT GCTGAYCGCA TGCAGAAAGA AATCAGGMT
<i>Xerocerastus sp.</i>	TCTGGTGGCT CAACAATGTT CCCAGGCATC GCTGACCGCA TGCAGAAAGA AATCACCAGC
<i>Zootecus insularis</i>	TCTGGTGGTT CAACAATGTT CCCTGGCATT GCTGACCGCA TGCAGAAAGA AATCACCAMT
<i>Coelioxys blandii</i>	TCTGGTGGTW CCWCAATGTT CCCYGGTATY GCTGACCGCA TGCAGARGGA AATCGTGAAC
<i>Pyrquina umbilicata</i>	TCTGGWGGTT CCACAATGTT CCCTGGAATC GCTGACCGCA TGCAGAAAGA AATCACTKCY
<i>Glessula ceylanica</i>	TCTGGAGGAT CAACAATGTT TCCTGGAATT GCTGACCGCA TGCAGAAAGA AATCACCAMT
<i>Ceciloides gokweanus</i>	TCTGGTGGCT CCACCATGTT CCCCAGGCATC GCTGACCGCA TGCAGAAAGA AATTAACCTGC
<i>Ferussacia folliculus</i>	TCTGGAGGTT CAACAATGTT CCCTGGCATT GCTGACCGCA TGCAGAAAGA GATCACGCTT
<i>Thyrophorella thomensis</i>	TCTGGWGGTT CCACAATGTT CCCTGGAATC GCTGACCGCA TGCAGAAAGA AATCACTKCT
<i>Gibbulinella dewinteri</i>	TCTGGTGGTT CTACCATGTA CCCAGGCATC GCCGACCGTA TGCAGAAAGA AATTACTTCT
<i>Gonaxis quadrilateralis</i>	TCTGGTGGTT CCACCATGTA YCCCAGGCATT GCTGACCGCA TGCAGAAAGA AATYACYTCC
<i>Gonospora sp.</i>	TCTGGAGGTT CTACMATGTT CCCWGSATT GCWGAYCGCA TGCAGAAARGA AATYACKKSY

	790 800 810 820 830 840
<i>Achatina achatina</i>	CTGGCTCCAS CCACAATGAA GATCAAGATC ATTGCTCCAC CAGAACGTAA ATACTCTGTC
<i>Achatina fulica</i>	CTGGCTCCTT CCACAATGAA GATCAAGATC ATTGCTCCAC CTGAGCGTAA ATACTCTGTC
<i>Achatina stuhlmanni</i>	CTGGCTCCTG CCACAATGAA GATCAAGATC ATTGCTCCAC CTGAACGTAA ATACTCTGTC
<i>Cochlitoma ustulata</i>	CTGGCTCCTG CCACAATGAA GATCAAGATC ATTGCTCCAC CTGAGCGTAA ATACTCTGTC
<i>Allopeas clavulinum</i>	TTGGCTCCAC CAACAATGAA GATCAAGATC ATTGCCCCAC CAGAAAGGAA GTACTCAGTC
<i>Bocageia sp.</i>	CTTGCTCCAY CCACAATGAA GATCAAGATC ATTGCTCCTC CYGAGMGGAA ATACTCAGTC
<i>Eutomopeas layardi</i>	TTGGCTCCAC CAACGATGAA GATCAAGATC ATCGTCCAC CAGAGAGGAA GTACTCGGTC
<i>Leptinaria lamellata</i>	CTCGCACCAC CAACAATGAA AATCAAAATC ATCGTCCAC CTGAGAGAAA GTACTCCGTA
<i>Paropeas achatinaceum</i>	TTGGCTCCAC CAACAATGAA AATCAAGATC ATTGCCCCAC CAGAGAGAAA GTACTCAGTC
<i>Riebeckia sp.</i>	CTRGCTCCAM CCACAATGAA GATCAAGATC ATTGCTCCAC CTGAAMGCAA ATACTCWGTC
<i>Rumina decollata</i>	CTGGCTCCAC CCACAATGAA GATCAAAATC ATTGCTCCTC CTGAGAGAAA ATACTCTGTC
<i>Subulina octona</i>	CTAGCTCCAC CCACAATGAA AATCAAGATA ATTGCGCCAC CTGAAAGGAA ATATTCTGTC
<i>Subulina striatella</i>	CTAGCTCCAC CCACAATGAA AATMAAAATA ATTGACCAC CTGAACGGAA ATATTCTGTT
<i>Subulina vitrea</i>	CTTGCTCCAC CMACAATGAA GATCAAAATC ATCGCACCCC CAGAACGYAA ATACTCTGTC
<i>Subulona sp.</i>	CTTGCTCCAG CAACAATGAA GATCAAGATA ATAGTCCCC CTGAGAGGAA GTACTCTGTT
<i>Tortaxis erectus</i>	CTRGCTCCAC CCACAATGAA GATCAAGATC ATAGTCCCTC CTGAACGCAA GTACTCTGTC
<i>Xerocerastus sp.</i>	CTGGCTCCAG CCACAATGAA GATCAAGATC ATTGCTCCTC CTGAGAGGAA ATACTCTGTC
<i>Zootecus insularis</i>	CTGGCYCCAC CCACGATGAA RATYAAAATC ATTGCTCCTC CTGAGAGAAA ATACTCTGTC
<i>Coelioxys blandii</i>	CTTGCTCCAC CCACAATGAA GATCAAAATC ATTGCTCCCC CTGAACGTAA ATACTCAGTC
<i>Pyrquina umbilicata</i>	CTTGCTCCAA CAACRATGAA GATCAAGATC ATYGCTCCAC CWGARAGRAA GTACTCAGTC
<i>Glessula ceylanica</i>	CTGGCWCAA SCACCATGAA AATCAAGATC ATTGCTCCCC CAGARCGGAA GTACTCTGTC
<i>Ceciloides gokweanus</i>	TTGGCTCCAC CCACAATGAA GATCAAGATC ATCGTCCCC CAGAGAGGAA ATACTCAGTC
<i>Ferussacia folliculus</i>	TTAGCCCCTG CCACCATGAA AATCAAGATC ATCGTCCAC CAGAACGTAA ATACTCCGTC
<i>Thyrophorella thomensis</i>	CTTGCTCCAG CAACAATGAA GATCAAGATC WTYGCTCCWC CTGAARRRAA GTACTCAGTC
<i>Gibbulinella dewinteri</i>	CTCGCTCCAC CCACAATGAA GATCAAGATC ATTGCTCCCC CCGAGCGTAA ATACTCTGTA
<i>Gonaxis quadrilateralis</i>	CTKGCTCCAM STACAATGAA GATCAAGATY ATTGCTCCYC CWGAGCGNAA ATACTCTGTA
<i>Gonospora sp.</i>	YTKGCTCCAC CCACTATGAA RATCAAGATC ATTGCTCCCC CWGAGCGWAA GTAYTCTGTC

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      ....|....| ....|....| .
            850      860
Achatina achatina      TGGATTGGAG GTTCCATTCT G
Achatina fulica        TGGATTGGAG GATCCATCCT G
Achatina stuhlmanni    TGGATTGGAG GATCCATCCT G
Cochlitoma ustulata    TGGATTGGAG GATCCATCCT G
Allopeas clavulinum    TGGATTGGTG GCTCCATCCT G
Bocageia sp.           TGGATTGGAG GCTCCATCCT G
Eutomopeas layardi     TGGATCGGTG GCTCCATCCT G
Leptinaria lamellata   TGGATTGGTG GTTCAATCCT A
Paropeas achatinaceum TGGATCGGTG GCTCCATCCT G
Riebeckia sp.          TGGATTGGAG GCTCCATCCT G
Rumina decollata       TGGATTGGAG GTTCCATCCT G
Subulina octona        TGGATTGGAG GCTCCATTTT G
Subulina striatella    TGGATTGGAG GCTCTATTTT G
Subulina vitrea        TGGATTGGAG GTTCCATCCT G
Subulona sp.           TGGATTGGTG GCTCCATCCT G
Tortaxis erectus       TGGATTGGAG GTTCCATCCT G
Xerocerastus sp.      TGGATTGGAG GTTCCATCCT G
Zootecus insularis    TGGATTGGAG GTTCCATCCT G
Coeliaxis blandii      TGGATTGGAG GTTCCATCCT G
Pyrgina umbilicata     TGGATCGGAG GTCCCATYTT G
Glessula ceylanica     TGGATTGGAG GTTCCATCCT G
Cecilioides gokweanus  TGGATCGGAG GCTCAATCCT G
Ferussacia folliculus  TGGATTGGTG GCTCCATCCT G
Thyrophorella thomensis TGGATCGGAG GTTCCATCCT G
Gibbulinella dewinteri TGGATCGGAG GCTCCATCCT G
Gonaxis quadrilateralis TGGATCGGAG GCTCCATCCT G
Gonospira sp.          TGGATCGGAG GTTCCATTTT G

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Appendix 3.3: Alignment of the histone 3 gene for the Achatinoidea and three streptaxid outgroup taxa. Note that sequencing was unsuccessful for the achatinoids *Ceciloides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp.

	10 20 30 40 50 60
<i>Achatina achatina</i>	TCGTAAATCC ACCGGAGGCA AGGCTCCCCG CAAGCARCTG GCCACCAAGG CKGCCAGRAA
<i>Achatina fulica</i>	TCGTAAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CCGCTAGAAA
<i>Achatina stuhlmanni</i>	TCGCAAATCC ACCGGAGGTA AGGCTCCTCG CAAACAGCTT GCCACCAAGG CCGCTAGAAA
<i>Cochlitoma ustulata</i>	TCGCAAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CCGCTAGGAA
<i>Allopeas clavulinum</i>	TCGTAAATCY ACCGGAGGCA ARGCTCCCCG CAARCARCTG GCCACCAAGG CNGCCAGGAA
<i>Bocageia</i> sp.	TCGCAAGTCC ACCGGTGGTA AAGCACCCCG CAAACAGCTT GCCACAAAGG CCGCTAGAAA
<i>Eutomopeas layardi</i>	TCGCAAATCT ACCGGAGGCA AAGCACCCCG CAAGCAGCTT GCCACCAAGG CCGCTAGGAA
<i>Leptinaria lamellata</i>	CCGAAAATCT ACTGGTGGTA AAGCACCCCG CAAGCAACTT GCCACCAAGG CTGCTAGAAA
<i>Rumina decollata</i>	TCGCAAATCC ACAGGAGGAA AAGCTCCCCG CAAACAGCTT GCCACTAAGG CCGCTAGAAA
<i>Subulina octona</i>	CCGTAAATCA ACTGGAGGAA AGGCACCACG CAAACAAGT GCCACTAAGG CAGCCAGAAA
<i>Subulina striatella</i>	CCGTAAATCA ACTGGAGGCA AGGCACCACG CAAACAGCTG GCCACTAAGG CCGCTAGAAA
<i>Subulona</i> sp.	GCGAAAGTCT ACCGGTGGCA AAGCTCCCCG CAAGCAGCTA GCCACCAAGG CCGCTAGAAA
<i>Tortaxis erectus</i>	CCGCAAATCT ACCGGAGGAA AGGCACCCTG TAAACAGCTT GCCACCAAGG CTGCTAGAAA
<i>Xerocerastus</i> sp.	TCGYAAATCY ACCGGAGGYA ARGCTCCCCG CAARCARCTK GCCACCAARG CRGCYAGRAA
<i>Zootecus insularis</i>	CCGCAAAGTCC ACCGGAGGCA AGGCTCCACG TAAACAGCTT GCCACCAAGG CCGCTAGAAA
<i>Coelioxaxis blandii</i>	TCGYAAATYY ACCGGAGGCA AGGCTCCYCG CAARCARCTK GCCACCAARG CCGCYAGRAA
<i>Pyrgina umbilicata</i>	TCGTAAATCT ACCGGAGGCA AAGCACCCCG AAAGCAGCTT GCCACCAARG CCGCTAGGAA
<i>Glessula ceylanica</i>	CAGGAAATCA ACCGGAGGCA AGGCTCCGCG TAAACAGCTT GCCACCAAAG CTGCCAGAAA
<i>Ferussacia folliculus</i>	TCGTAAATCT ACTGGTGGTA AGGCACCACG CAAACAGTTG GCCACCAAAG CAGCCAGGAA
<i>Thyrophorella thomensis</i>	TCGTAAATCT ACCGGAGGCA AAGCACCCCG AAAGCAGCTT GCCACCAAAG CCGCTAGGAA
<i>Gibbulinella dewinteri</i>	ACGTAAATCC ACCGGAGGCA AGGCTCCACG AAAGCAGCTG GCCACCAAGG CAGCAAGGAA

	70 80 90 100 110 120
<i>Achatina achatina</i>	RTCGGCCCCR GCCACMGGRG GTGTSAAAGAA RCCCCATCGW TACAGGCCCG GCACNGTSGC
<i>Achatina fulica</i>	GTCCGGCCCCG GCCACAGGAG GTGTCAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCGC
<i>Achatina stuhlmanni</i>	GTCCGGCCCCA GCCACAGGAG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACAGTCGC
<i>Cochlitoma ustulata</i>	GTCCGGCCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCGC
<i>Allopeas clavulinum</i>	GTCCGGCCCCR GCCACCGGWG GTGTSAAAGAA ACCCCATCGT TACAGGCCCG GCACCGTGGC
<i>Bocageia</i> sp.	ATCCGGCCCCA GCCACAGGAG GTGTGAAGAA ACCTCACCGA TACAGGCCCG GTACAGTGGC
<i>Eutomopeas layardi</i>	GTCCGGCCCCA GCTACCGGAG GTGTCAAGAA ACCTCACAGA TACAGGCCCG GCACAGTCGC
<i>Leptinaria lamellata</i>	GTCCGGCCCCG GCTACTGGGG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACTGTTCGC
<i>Rumina decollata</i>	GTCCGGCCCCG GCCACAGGAG GCGTGAAGAA ACCCCAYCGA TACAGGCCTG GCACGGTTCGC
<i>Subulina octona</i>	GTCTGCCCCA GCTACAGGAG GTGTCAAGAA RCCGCATCGT TACAGGCCCG GCACTGTTCGC
<i>Subulina striatella</i>	GTCCGGCCCCG GCCACAGGGG GTGTCAAGAA GCCCCATCGT TACAGGCCCG GCACTGTTCGC
<i>Subulona</i> sp.	GTCCGGCCCCA GCGACTGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCAG GCACCGTGGC
<i>Tortaxis erectus</i>	GTCCGGTCCCG GCCACAGGAG GTGTGAAGAA GCCCCACAGG TACAGGCCCG GTACAGTCGC
<i>Xerocerastus</i> sp.	RTCKGCCCCR GCCACCGGWG GTGTSAAAGAA ACCCCAYCGW TACAGGCCCG GCACCGTSGC
<i>Zootecus insularis</i>	GTCCGGCCCCG GCCACAGGAG GTGTGAAGAA ACCCCACCGA TACAGACCTG GCACGGTTCGC
<i>Coelioxaxis blandii</i>	GTCKGCCCCR GCCACCGGWG GTGTSAAAGAA RCCYCAYMGA TACMGGCCSG GCACNGTSGC
<i>Pyrgina umbilicata</i>	ATCCGCCCCA GCCACCGGAG GTGTCAAGAA ACCTCACAGA TACAGGCCCG GCACTGTTCGC
<i>Glessula ceylanica</i>	GTCCGGCCCCG GCCACAGGAG GCGTGAAGAA ACCTCACAGR TACAGGCCCG GGACAGTCGC
<i>Ferussacia folliculus</i>	GTCCGGCCCCA GCCACTGGTG GTGTGAAGAA ACCTCACCGT TACAGGCCTG GTACAGTTGC
<i>Thyrophorella thomensis</i>	ATCCGCCCCA GCCACCGGAG GTGTCAAGAA ACCTCACAGA TACAGGCCCG GCACTGTTCGC
<i>Gibbulinella dewinteri</i>	ATCTGCCCCA GCAACCGGTG GTGTGAAGAA GCCCCATCGC TACAGGCCAG GCACTGTTCGC

	130	140	150	160	170	180	
<i>Achatina achatina</i>	TCTCKGAGAR	ATCCGTCGTT	ACCAGAAGAG	CACKGAGCTC	CTCATCCGCA	AGCTGCCTTT	
<i>Achatina fulica</i>	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACCTGAGCTT	CTCATCCGCA	AACTGCCTTT	
<i>Achatina stuhlmanni</i>	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AACTGCCTTT	
<i>Cochlitoma ustulata</i>	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATTCGCA	AACTGCCTTT	
<i>Allopeas clavulinum</i>	TCTGCGAGAR	ATCCGTCGTT	ACCAGAAGAG	CACCTGAGCTS	CTCATCCGCA	AGCTGCCTTT	
<i>Bocageia sp.</i>	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTC	CTCATCCGTA	AACTGCCTTT	
<i>Eutomopeas layardi</i>	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTC	CTCATCCGCA	AGCTGCCTTT	
<i>Leptinaria lamellata</i>	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAACCT	CTCATCCGCA	AGCTGCCTTT	
<i>Rumina decollata</i>	GCTTCGAGAG	ATCCGACGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	AGCTGCCTTT	
<i>Subulina octona</i>	ACTTCGAGAG	ATCCGCCGTT	ATCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AGCTGCCTTT	
<i>Subulina striatella</i>	ACTTCGAGAG	ATCCGCCGTT	ACCAGAAGAG	CACGGAGCTC	CTCATCCGCA	AGCTGCCTTT	
<i>Subulona sp.</i>	ACTCCGAGAG	ATCCGCCGTT	ACCAGAAGAG	CACCGAGCTT	CTCATCCGCA	AACTGCCTTT	
<i>Tortaxis erectus</i>	ACTCAGAGAG	ATCCGCCGTT	ACCAGAAGAG	CACAGAGCTG	CTCATCCGCA	AATTGCCTTT	
<i>Xerocerastus sp.</i>	TCTGCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACCTGAGCTK	CTCATCCGCA	AGCTGCCTTT	
<i>Zootecus insularis</i>	ACTTCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTT	CTCATCCGCA	AACTGCCTTT	
<i>Coelioxaxis blandii</i>	TCTCKGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACKGAGCTN	CTCATCCGCA	ARCTGCCTTT	
<i>Pyrgina umbilicata</i>	ACTCCGAGAG	ATCCGTCGTT	ACCARAAGAG	CACGGAGCTM	CTCATCCGCA	AGCTGCCTTT	
<i>Glessula ceylanica</i>	ACTTCGAGAG	ATCCGCCGTT	ACCAGAAGAG	CACGGAGCTG	CTCATCCGCA	AACTGCCTTT	
<i>Ferussacia folliculus</i>	ACTCCGTGAG	ATCCGTCGTT	ACCAGAAGAG	CACAGAGCTG	CTCATCCGCA	AACTGCCTTT	
<i>Thyrophorella thomensis</i>	ACTCCGAGAG	ATCCGTCGTT	ACCAGAAGAG	CACGGAGCTT	CTCATCCGCA	AGCTGCCTTT	
<i>Gibbulinella dewinteri</i>	TCTTCGTGAG	ATTCGTCGTT	ATCAGAAAAG	CACGGAACCT	CTCATCCGCA	AACTGCCTTT	

	190	200	210	220	230	240	
<i>Achatina achatina</i>	CCAGCGCCTG	GTCAGAGAAA	TGCGCCAGGA	CTTCAAGACY	GACCTGCGCT	TCCAGAGCTC	
<i>Achatina fulica</i>	CCAGCGGCTG	GTCAGAGAGA	TGCGCCAGGA	CTTTAAGACC	GATCTGCGTT	TCCAGAGCTC	
<i>Achatina stuhlmanni</i>	CCAGCGGCTG	GTCAGAGAGA	TGCGCACAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
<i>Cochlitoma ustulata</i>	CCAGCGGCTG	GTCAGAGAGA	TGCGCCAGGA	CTTCAAGACC	GATCTGCGTT	TCCAGAGCTC	
<i>Allopeas clavulinum</i>	CCAGCGCCTG	GTSMGAGAAA	TYGCYCAGGA	CTTCAAGACY	GACCTGCGYT	TCCAGAGCTC	
<i>Bocageia sp.</i>	CCAGCGCCTG	GTCAGAGAAA	TGCGCCAGGA	TTTTAAAACC	GACCTGCGTT	TCCAGAGCTC	
<i>Eutomopeas layardi</i>	CCAGCGCTG	GTCAGAGAGA	TGCGCCAGGA	CTTTAAGACA	GATCTGCGTT	TCCAGAGCTC	
<i>Leptinaria lamellata</i>	CCAGCGCCTG	GTCAGAGAAA	TGCGTCAGGA	CTTTAAAACA	GATCTGCGTT	TCCAAAGTTC	
<i>Rumina decollata</i>	CCAGCGCCTT	GTCAGAGAGA	TGCGCCAGGA	TTTTAAGACT	GACCTGCGTT	TTCAGAGCTC	
<i>Subulina octona</i>	CCAGCGCCTG	GTCAGAGAAA	TGCGCCAGGA	TTTTAAGACG	GACCTGCGCT	TCCAGAGCTC	
<i>Subulina striatella</i>	CCAGCGACTG	GTCAGAGAAA	TGCGCCAGGA	TTTTAAGACC	GACCTGCGCT	TCCAGAGCTC	
<i>Subulona sp.</i>	CCAGCGACTG	GTCAGAGAAA	TGCGCCAGGA	CTTCAAGACC	GATCTTCGAT	TCCAGAGCTC	
<i>Tortaxis erectus</i>	CCAGCGCCTT	GTCAGGAGAA	TGCGCACAGGA	CTTTAAGACT	GACCTGCGTT	TCCAGAGCTC	
<i>Xerocerastus sp.</i>	CCAGCGCCTG	GTSMGAGARA	TYGCYCAGGA	YTTCAAGACY	GAYCTGCGYT	TCCAGAGCTC	
<i>Zootecus insularis</i>	CCAGCGCCTG	GTCAGAGAGA	TGCGCCAGGA	TTTTAAGACT	GATCTGCGCT	TCCAGAGCTC	
<i>Coelioxaxis blandii</i>	CCARCGCCTG	GTSMGAGAAA	TGCGYCAGGA	YTTCAAGACY	GAYCTGCGYT	TCCAGAGCTC	
<i>Pyrgina umbilicata</i>	CCAGCGCCTG	GTCAGAGAAA	TGCGTCAGGA	CTTCAAGACC	GATCTGCGCT	TCCAGAGCTC	
<i>Glessula ceylanica</i>	CCAGCGCCTG	GTCAGAGAGA	TTGCCCAGGA	CTTCAAGACC	GACCTGCGTT	TCCAGAGCTC	
<i>Ferussacia folliculus</i>	CCAGCGACTG	GTCAGAGAAA	TGCGCACAGGA	CTTCAAGACT	GACCTGCGCT	TCCAAAGCTC	
<i>Thyrophorella thomensis</i>	CCAGCGCCTG	GTCAGAGAAA	TGCGTCAGGA	CTTCAAGACC	GATCTGCGCT	TCCAAAGCTC	
<i>Gibbulinella dewinteri</i>	CCAACGCTTG	GTAAGAGAAA	TTGCCCAGGA	CTTCAAGACA	GATCTGCGCT	TCCAGAGCTC	

	250	260	270	280	290	300	
<i>Achatina achatina</i>	GGCYGTCATG	GCCCTGCAAG	AGGCYAGCGA	RGCCTACCTG	GTGGGTCTGT	TYGAGGACAC	
<i>Achatina fulica</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Achatina stuhlmanni</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma ustulata</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Allopeas clavulinum</i>	SGCTGTCATG	GCYCTGCARG	AGGCCAGCGA	RGCCTACCTG	GTSGGYCTST	TTGAGGACAC	
<i>Bocageia sp.</i>	GGCTGTCATG	GCTCTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTCGGTCTGT	TTGAGGACAC	
<i>Eutomopeas layardi</i>	GGCTGTCATG	GCACTGCAGG	AAGCCAGCGA	GGCCTACCTC	GTAGGTCTGT	TTGAAGACAC	
<i>Leptinaria lamellata</i>	AGCTGTCATG	GCTCTGCAGG	AGGCTAGCGA	GGCCTACCTT	GTGGGTCTGT	TGGAAGACAC	
<i>Rumina decollata</i>	RGCYGTCATG	GCACTGCAGG	AGGCTAGCGA	GGCMTACYTR	GTGGGCCTGT	TTGAGGACAC	
<i>Subulina octona</i>	GGCTGTCATG	GCGCTGCAGG	AGGCCAGCGA	GGCCTACCTC	GTAGGTCTGT	TTGAGGACAC	
<i>Subulina striatella</i>	GGCTGTCATG	GCGCTGCAGG	AGGCCAGCGA	GGCCTACCTG	GTAGGTCTGT	TTGAGGACAC	
<i>Subulona sp.</i>	TGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCTTACCTG	GTCGGCCTGT	TGAGGACAC	
<i>Tortaxis erectus</i>	GGCCGTCATG	GCTCTGCAGG	AAGCAAGCGA	GGCCTATCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Xerocerastus sp.</i>	SGCTGTCATG	GCYCTGCARG	AGGCYAGCGA	RGCCTACCTG	GTSGGYCTST	TTGAGGACAC	
<i>Zootecus insularis</i>	GGCTGTCATG	GCACTGCAGG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Coelioxaxis blandii</i>	SGCTGTCATG	GCYCTGCAGG	AGGCYAGCGA	RGCCTACCTG	GTSGGTCTST	TTGAGGACAC	
<i>Pyrgina umbilicata</i>	GGCTGTCATG	GCACTGCAGG	AAGCCAGCGA	GGCCTACCTC	GTAGGTCTGT	TTGAAGACAC	
<i>Glessula ceylanica</i>	AGCTGTCATG	GCACTGCAGG	AAGCCAGCGA	AGCCTAYTTG	GTGGGTCTGT	TGAGGACAC	
<i>Ferussacia folliculus</i>	SGCCGTCATG	GCCTGCAAG	AGGCAAGYGA	GGCCTACCTT	GTGGGTCTGT	TTGAAGACAC	
<i>Thyrophorella thomensis</i>	GGCTGTCATG	GCACTGCAGG	AAGCCAGCGA	GGCCTACCTC	GTAGGTCTGT	TTGAAGACAC	
<i>Gibbulinella dewinteri</i>	RGCCGTCATG	GCAATTGCAGG	AGGCCAGCGA	GGCCTACCTC	GTGGGTCTCT	TTGAGGACAC	

	310	320	
<i>Achatina achatina</i>	YAACYTGTGC	GCYATCCACG	CCAARCGT
<i>Achatina fulica</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Achatina stuhlmanni</i>	CAACCTGTGC	GCCATYCACG	CCAAACGT
<i>Cochlitoma ustulata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Allopeas clavulinum</i>	CAACYTGTGC	GCCATCCAYG	CCAAGCGT
<i>Bocageia sp.</i>	CAACCTGTGC	GCCATCCATG	CCAAGCGG
<i>Eutomopeas layardi</i>	CAATCTGTGC	GCAATTCACG	CCAAGCGT
<i>Leptinaria lamellata</i>	CAACCTGTGC	GCCATCCATG	CCAAACGT
<i>Rumina decollata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Subulina octona</i>	CAACCTGTGC	GCCATCCACG	CCAAGCGT
<i>Subulina striatella</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Subulona sp.</i>	CAACCTGTGC	GCCATCCATG	CCAAGCGC
<i>Tortaxis erectus</i>	AAACCTGTGC	GCCATCCACG	CCAAGCGA
<i>Xerocerastus sp.</i>	CAACTTGTGC	GCCATCCAYG	CCAARCGT
<i>Zootecus insularis</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Coelioxys blandii</i>	CAACYTGTGC	GCYATCCAYG	CYAAGCGY
<i>Pyrgina umbilicata</i>	CAACCTGTGC	GCCATCCATG	CCAAGCGT
<i>Glessula ceylanica</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Ferussacia folliculus</i>	CAACCTGTGC	GCCATCCACG	CCAAGCGT
<i>Thyrophorella thomensis</i>	TAACCTGTGC	GCCATCCATG	CCAAGCGT
<i>Gibbulinella dewinteri</i>	CAACCTGTGC	GCTATCCACG	CCAAGCGT

Appendix 3.4: Alignment of the CO1 gene for the Achatinoidea and three streptaxid outgroup taxa. Note that sequencing was unsuccessful for *Cecilioides gokweanus* as well as the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp.

	10 20 30 40 50 60
Achatina achatina	AGGTACAGGC CTATCATTGT TAATTCGATT AGAATTAGGG ACAGCTGGGG TTTTAACCGA
Achatina fulica	TGGCACAGGC TTGTCACTCT TAATTCGGTT AGAGCTTGGG ACAGTGGGAA CCTTAACTGA
Achatina stuhlmanni	TGGTACCGGC TTGTCCTTAC TTATCCGGCT AGAACTAGGG AGTATTGGAA CACTGACTGA
Cochlitoma ustulata	GGGCACAGGT CTATCTTTAT TGATCCGACT AGAGCTAGGA ACGACCTCTA CTTTAACTGA
Coeliaxis blandii	TGGGACTGGG TTATCTTTAC TTATTCGTTT AGAATTAGGT ACTGTGGAG TACTTAACTGA
Pyrgina umbilicata	CGGGACTGGT CTATCTTTAT TAATCCGTTT AGAACTCGGA ATAGGGGGCG TGTTAATTGA
Glessula ceylanica	CGGTACAGGA TTATCATTAT TAATTCGGTT AGAATTAGGA ACAACTGGTG TATTGACGGA
Ferussacia folliculus	TGGAACAGGT TTATCTTTGC TTATTCGTTT AGAATTAGGT ACATCGGGCG TCTTAACAGA
Thyrophorella thomensis	TGGAACAGGT TTGTCATTAC TGATCCGCTC GGAGTTAGGA ACAGCCGGTG TACTTAACTGA
Allopeas clavulinum	TGGGACAGGA TTATCTCTTC TCATTCGTTT TGAGTTAGGT ACTAGTGGGG TGTTAACCGA
Bocageia sp.	AGGTACCGGC TTATCCTTAT TAATTCGCTC GGAATTAGGA ACCTCTGGTG TTTTGACTGA
Eutomopeas layardi	GGGGACAGGT TTATCTTTAC TAATCCGTTT AGAATTAGGA ACTAGCGGAG TTTTAACTGA
Leptinaria lamellata	AGGTACAGGT TTATCGTTAT TAATTCGTTT AGAATTAGGG ACAATGGAAG TGTTAACAGA
Paropeas achatinaceum	TGGGACTGGT TTATCTTTAC TAATTCGCTC TGAGTTAGGA ACTAGGGGAG TGTTAACCGA
Riebeckia sp.	TGGGACTGGT TTATCTTTAT TAATTCGATT AGAGTTAGGT ACTTCAGGTG TATTAACTGA
Rumina decollata	TGGTACTGGT TTATCTCTTC TTATTCGTTT AGAACTAGGT ACAGCAGGAG TGTTAACAGA
Subulina octona	TGGGACTGGT TTATCATTAT TAATTCGTTT AGAGCTGGG ACGGCTGGAG TGTTAACTGA
Subulina striatella	TGGGACGGGT TTGTCATTAT TAATTCGACT TGAGTTAGGT ACTGCTGGTG TTTTAACCGA
Subulina vitrea	TGGTACAGG CTTCCCTTT TAATTCGTAT TGAGCTTGGG ACTGCTGGGG TTCTTAACTGA
Subulona sp.	AGGGACAGGA TTATCTTTAT TAATTAGTTT AGAATTAGGG ACTGTAAGGG TATTAGTAGA
Tortaxis erectus	TGGGACAGGG TTGTCCTTAT TGATTCGCTC TGAACTAGGT ACTTCTGGTG TATTAACAGA
Xerocerastus sp.	TGGTACTGGA CTATCTTTAT TAATTCGGCT TGAGTTAGGA ACAGCTGGTG TTTTAACTGA
Zootecus insularis	TGGGACAGGG TTGTCGTTAT TAATTCGGCT TGAATTAGGT ACCTCAGGTG TGTTAACAGA
Gibbulinella dewinteri	TGGGACCGGT TTGTCCTTAT TAATTCGGCT AGAATTAGGT ACAGCTGGGG TTCTTATTGA

	70 80 90 100 110 120
Achatina achatina	TGATCATTTC TTTAATGTAG TGGTGACAGC CCATGCTTTT GTTATAAATT TTTTATAGT
Achatina fulica	TGATCACTTT TTCAACGTGG TTGTAACCTGC GCATGCTTTT GTCATAAATT TTTTATGGT
Achatina stuhlmanni	CGATCATTTC TTTAATGTTG TAGTTACTGC TCATGCTTTT GTGATAAATT TTTTATAGT
Cochlitoma ustulata	TGATCATTTC TTTAATGTTA TCGTGACAGC GCACGCTTTT GTAATAAATT TTTTCATAGT
Coeliaxis blandii	TGATCATTTC TTTAATGTGG TTGTAACCTGC TCATGCTTTT GTAATAAATT TTTTATAGT
Pyrgina umbilicata	TGACCACTTT TTCAATGTTG TTGTAACAGC TCATGCGTTT GTAATAAATT TTTTATAGT
Glessula ceylanica	CGATCATTTC TTTAATGTAA TTGTTACCGC TCATGCTTTT GTTATAAATT TTTTATAGT
Ferussacia folliculus	TGATCACTTT TTTAATGTTA TTGTTACTGC ACACGCTTTT GTTATGATTT TTTTATGGT
Thyrophorella thomensis	TGATCACTTT TTTAATGTGG TCGTCACAGC CCACGCTTTT GTTATAAATT TTTTATAGT
Allopeas clavulinum	TGACCACTTT TTTAATGTTG TAGTTACAGC TCATGCTTTC GTAATAAATT TTTTATAGT
Bocageia sp.	CGATCACTTT TTTAACGTTG TAGTAAACAGC TCATGCGTTT GTTATAAATT TCTTTATAGT
Eutomopeas layardi	TGATCACTTT TTTAATGTTG TAGTAAACAGC TCACGCTTTC GTGATGATTT TTTTATAGT
Leptinaria lamellata	TGATCATTTC TTTAACGTAG TGGTTACTGC TCACGCTTTT GTAATAAATT TTTTATAGT
Paropeas achatinaceum	TGATCATTTC TTTAATGTTG TAGTAACTGC TCATGCATTT GTAATAAATT TTTTATAGT
Riebeckia sp.	TGATCACTTT TTTAATGTAA TTGTAACCGC ACATGCTTTT GTTATAAATT TTTTATAGT
Rumina decollata	TGACCACTTT TTTAATGTGG TAGTTACTGC CCATGCATTT GTGATAAATT TTTTATAGT
Subulina octona	TGATCATTTC TTTAATGTTA TTGTAACCTGC TCATGCATTT GTTATGATTT TTTTATGGT
Subulina striatella	TGACCACTTT TTTAATGTAA TTGTGACAGC TCATGCTTTT GTTATAAATT TTTTATAGT
Subulina vitrea	CGATCATTTC TTTAATGTAA TTGTGACTGC TCATGCTTTT GTTATAAATT TTTTATAGT
Subulona sp.	CGATCATTTC TTTAACGTAA TTGTTACTGC TCATGCTTTT GTAATAAATT TCTTTATAGT
Tortaxis erectus	TGATCATTTC TTTAATGTTG TAGTTACAGC CCATGCTTTT GTTATGATTT TTTTATAGT
Xerocerastus sp.	TGATCACTTT TATAATGTTG TTGTTACGGC TCATGCTTTT GTAATAAATT TTTTATAGT
Zootecus insularis	CGACCACTTC TTTAATGTTA TTGTAACAGC TCATGCATTC GTAATAAATT TTTTATAGT
Gibbulinella dewinteri	TGATCATTTC TTTAAWGTTA TTGTTACWGC TCACGCTTTT GTTATAAATT TTTTATAGT

	130	140	150	160	170	180	
<i>Achatina achatina</i>	CATACCAATT	ATGATTGGGG	GTTTTGGTAA	TTGAATAGTC	CCACTATTAA	TTGGGGCTCC	
<i>Achatina fulica</i>	TATACCAATT	ATAATTGGCG	GATTTGGAAA	CTGGATGGTC	CCAATACTTA	TTGGTGCTCC	
<i>Achatina stuhlmanni</i>	GATACCAATT	ATAATTGGTG	GTTTTGGAAA	CTGGATGGTC	CCATTACTGA	TTGGTGCTCC	
<i>Cochlitoma ustulata</i>	AATACCCATT	ATAATTGGGG	GTTTTGGTAA	TTGAATAACT	CCCTTTTAA	TCGGGGCTCC	
<i>Coelioxaxis blandii</i>	TATACCTATT	ATAATTGGGG	GATTTGGAAA	TTGAATGGTT	CCATTATTAA	TTGGAGCTCC	
<i>Pyrgina umbilicata</i>	TATACCAATT	ATGATTGGTG	GTTTTGGGAA	TTGAATAGTG	CCGCTTTTAA	TTGGTGCTCC	
<i>Glessula ceylanica</i>	TATGCCTATT	ATAATTGGAG	GATTTGGTAA	TTGGATAGTT	CCGTTATTAA	TTGGTGCTCC	
<i>Ferussacia folliculus</i>	TATACCTATT	ATAATTGGAG	GTTTTGGAAA	TTGAATGGTG	CCCTTTTGA	TTGGAGCTCC	
<i>Thyrophorella thomensis</i>	TATACCAATT	ATAATTGGTG	GTTTTGGTAA	TTGAATAGTA	CCACTTTTAA	TTGGTGCTCC	
<i>Allopeas clavulinum</i>	TATGCCTATT	ATAATTGGTG	GCTTTGGGAA	CTGAATGGTC	CCACTATTAA	TTGGGGCTCC	
<i>Bocageia sp.</i>	AATACCTATC	ATAATTGGTG	GTTTTGGGAA	CTGAATAGTT	CCACTATTAA	TTGGAGCTCC	
<i>Eutomopeas layardi</i>	TATACCAATC	ATGATTGGGG	GTTTTGGAAA	CTGAATAGTC	CCATTACTGA	TTGGGGCCCC	
<i>Leptinaria lamellata</i>	TATACCTATT	ATAATTGGGG	GATTTGGAAA	TTGAATAGTG	CCTTTACTAA	TTGGTGACCC	
<i>Paropeas achatinaceum</i>	TATACCAATT	ATAATTGGAG	GTTTTGGGAA	TTGGATAGTC	CCCTTATTAA	TTGGTGCTCC	
<i>Riebeckia sp.</i>	TATACCAATT	ATAATTGGAG	GATTTGGAAA	TTGAATAGTC	CCTTTACTAA	TTGGGGCCCC	
<i>Rumina decollata</i>	AATGCCTATT	ATAATCGGTG	GTTTTGGAAA	TTGAATAGTG	CCCTTATTAA	TTGGGGCACC	
<i>Subulina octona</i>	AATACCTATT	ATAATTGGAG	GATTTGGGAA	TTGAATAGTT	CCTTTGCTTA	TTGGGGCTCC	
<i>Subulina striatella</i>	CATACCTATT	ATGATTGGAG	GTTTTGGGAA	TTGGATAGTT	CCTTTACTGA	TTGGTGCTCC	
<i>Subulina vitrea</i>	TATACCTATT	ATAATTGGGG	GTTTTGGGAA	TTGGATAGTA	CCACTACTAA	TTGGTGCGCC	
<i>Subulona sp.</i>	AATACCTATT	ATGATTGGGG	GATTTGGAAA	TTGAATAGTG	CCTTTACTTA	TTGGGGCTCC	
<i>Tortaxis erectus</i>	AATGCCTATT	ATAATTGGTG	GTTTTGGTAA	TTGAATAGTT	CCACTTTTAA	TTGGTGCTCC	
<i>Xerocerastus sp.</i>	TATACCAATT	ATAATTGGTG	GATTTGGTAA	TTGAATAGTT	CCATTATTAA	TTGGTGCTCC	
<i>Zootecus insularis</i>	TATGCCCATC	ATAATTGGCG	GCTTCGGAAA	TTGAATGGTC	CCACTATTAA	TTGGTGCTCC	
<i>Gibbulinella dewinteri</i>	TATACCACTA	ATAATTGGAG	GTTTTGGAAA	TTGAATAGTA	CCTCTTCTTA	TTGGGGCTCC	

	190	200	210	220	230	240
<i>Achatina achatina</i>	AGATATAAGA	TTCCCTCGGA	TGAATAATAT	AAGGTTCTGA	TTATTACCAC	CTTCATTTAC
<i>Achatina fulica</i>	TGATATAAGA	TTTCCACGAA	TAAATAATAT	AAGGTTTGA	CTTTTACCAC	CTTCATTTAC
<i>Achatina stuhlmanni</i>	TGATATAAGG	TTTCCGCGGA	TAAATAACAT	AAGATTTGA	CTACTTCCC	CATCATTTTC
<i>Cochlitoma ustulata</i>	TGATATAAGC	TTTCTCGTA	TAAATAACAT	GAGCTTTGA	TTGTTACCCC	CTTCTTTTAC
<i>Coelioxaxis blandii</i>	TGATATAAGA	TTTCCCCGAA	TAAATAATAT	AAGATTTGG	CTTTTACCTC	CATCTTTTAT
<i>Pyrgina umbilicata</i>	TGATATAAGT	TTCCACGAA	TAAATAATAT	AAGATTTGA	CTTTTACCTC	CATCTTTTAT
<i>Glessula ceylanica</i>	AGATATAAGA	TTCCCCGAA	TAAATAATAT	AAGATTTGG	TTATTACCAC	CATCTTTTGT
<i>Ferussacia folliculus</i>	TGATATAAGG	TTTCTCGTA	TAAATAATAT	GAGGTTTGA	CTTCTCCCGC	CTTCTTTTAT
<i>Thyrophorella thomensis</i>	TGATATGAGT	TTTCCCGTA	TAAACAACAT	AAGCTTCTGG	CTCCTTCCAC	CATCATTTAT
<i>Allopeas clavulinum</i>	AGATATAAGG	TTTCTCGAA	TAAATAATAT	AAGTTTTGG	CTTCTTCCAC	CATCTTTTAT
<i>Bocageia sp.</i>	GGACATAAGT	TTTCTCGTA	TAAATAATAT	AAGATTTGA	CTGCTTCCCTC	CGTCATTTAT
<i>Eutomopeas layardi</i>	GGATATGAGA	TTTCCCCGGA	TAAATAATAT	AAGTTTCTGG	TTACTTCCAC	CATCTTTCGT
<i>Leptinaria lamellata</i>	AGATATAAGA	TTTCTCGAA	TAAATAATAT	AAGATTTGG	TTATTGCCGC	CTTCTTTTAT
<i>Paropeas achatinaceum</i>	TGATATAAGA	TTCCCTCGTA	TAAATAATAT	AAGTTTTGA	TTACTTCCGC	CATCTTTTGT
<i>Riebeckia sp.</i>	AGATATAAGA	TTTCTCGAA	TAAATAATAT	AAGATTTGA	CTTTTACCTC	CATCATTTAT
<i>Rumina decollata</i>	AGATATAAGG	TTTCTCGAA	TAAATAATAT	GAGATTTGA	CTACTTCCCTC	CTTCTTTTAT
<i>Subulina octona</i>	TGATATAAGA	TTTCCGCGTA	TGAATAATAT	AAGGTTTGG	TTGTTACCTC	CTTCATTTTT
<i>Subulina striatella</i>	TGACATAAGT	TTTCCCCGTA	TAAATAATAT	AAGATTTGG	TTGCTACCAC	CTTCTTTTAT
<i>Subulina vitrea</i>	TGACATAAGA	TTTCCACGGA	TAAATAATAT	AAGATTTGG	CTTCTACCAC	CCTCATTTAT
<i>Subulona sp.</i>	AGATATAAGT	TTTCCACGAA	TAAATAATAT	AAGGTTTGA	TTATTGCCTC	CTTCTTTTAT
<i>Tortaxis erectus</i>	TGATATAAGG	TTTCCACGAA	TAAATAATAT	AAGATTTGA	TTATTACCTC	CATCTTTTAT
<i>Xerocerastus sp.</i>	CGATATAAGA	TTTCTCGGA	TAAATAACAT	AAGATTTGA	TTGTTACCAC	CATCATTTAT
<i>Zootecus insularis</i>	GGATATAAGT	TTTCTCGAA	TAAATAATAT	AAGATTTGG	CTTCTTCCGC	CTTCTTTTAT
<i>Gibbulinella dewinteri</i>	AGATATAAGT	TTTCCACGCA	TAAATAATAT	AAGATTTGA	TTATTACCCC	CTTCTTTTGT

	250 260 270 280 290 300
<i>Achatina achatina</i>	CTTACTTATT CTTTCTAGTA TAGTTGAGGG AGGAGCTGGT ACTGGGTGAA CAGTTTATCC
<i>Achatina fulica</i>	TTTGTTAATC TGTTCAAGTA TAGTGAAGG AGGGGCTGGA ACTGGGTGGA CTGTGTACCC
<i>Achatina stuhlmanni</i>	TTTGCTAATF CTCTCAAGTA TAGTAGAAGG TGGGGCTGGG ACAGGATGGA CAGTTTACCC
<i>Cochlitoma ustulata</i>	ACTTCTTATC TTGTCAAGAA TAGTGAAGG GGGTGCAGGT ACAGGTTGAA CTGTGTATCC
<i>Coelioxys blandii</i>	TTTTTTAATT GTCTCGAGAA TGGTAGAAGG GGGAGCCGGG ACTGGGTGAA CTGTTTACCC
<i>Pyrgina umbilicata</i>	TTTCCTTATT ATGTCTAGTA TAATAGAGGG TGGAGCTGGT ACAGGTTGGA CAGTTTATCC
<i>Glessula ceylanica</i>	TCTTTTAATT GTATCTAGTA TAATTGAAGG TGGTGCTGGT ACAGGTTGAA CTGTTTATCC
<i>Ferussacia folliculus</i>	TCTTTTAATT ACATCTAGTT TGGTAGAAGG TGGTGCAGGG ACAGGATGGA CAGTTTACCC
<i>Thyrophorella thomensis</i>	TTTCTTGATC ACGTCAAGGT TAATAGAAGG CGGAGCTGGC ACAGGTTGGA CTGTATACCC
<i>Allopeas clavulinum</i>	TTTTTTAATT ATGTCAAGAA TAATAGAAGG AGGAGCAGGA ACAGGTTGAA CCGTTTATCC
<i>Bocageia sp.</i>	TTTTTTAATT ATGTCTAGAA TAGTTGAAGG TGGGGCTGGA ACTGGTTGAA CTGTGTACCC
<i>Eutomopeas layardi</i>	TTTTTTAATT ATGTCTAGTA TAATAGAGGG GGGGGCTGGT ACAGGCTGAA CTGTTTACCC
<i>Leptinaria lamellata</i>	ATTTTTAATT ATTTCTAGTT TAGTGAAGG GGGTGCTGGA ACAGGTTGAA CAGTCTACCC
<i>Paropeas achatinaceum</i>	TTTTCTAATT GTTTC AAGGA TAATAGAGGG GGGAGCCGGA ACTGGTTGAA CTGTCTACCC
<i>Riebeckia sp.</i>	TCTACTTATT GTATCAAGTA TAGTAGAAGG AGGTGTAGGA ACAGGTTGAA CTGTTTACCC
<i>Rumina decollata</i>	TTTATTAATT ATATCTAGGA TAGTAGAAGG TGGGGCAGGA ACTGGTTGAA CAGTGTATCC
<i>Subulina octona</i>	ATTATTAATT TTATCTAGAA TAGTGAAGG TGGTGCTGGT ACTGGTTGAA CAGTTTATCC
<i>Subulina striatella</i>	TTTATTAATT TTATCAAGCA TGGTARAAGG AGGTGCAGGG ACAGGTTGAA CCGTTTATCC
<i>Subulina vitrea</i>	TTTTTTAATT GTCTCTAGGA TAGTGAAGG AGGGGCTGGA ACAGGTTGAA CAGTTTACCC
<i>Subulona sp.</i>	TTTTTTAATT ATTTCTAGGT TGATTGAAGG TGGAGCCGGA ACTGGTTGAA CTGTTTACCC
<i>Tortaxis erectus</i>	TTTATTAATT TTATCTAGAA TAGTTGAAGG TGGGCTGGG ACTGGTTGGA CTGTTTACCC
<i>Xerocerastus sp.</i>	TTTATTAATT ATATCGAGGA TAGTTGAAGG GGGTGCTGGT ACTGGTTGAA CTGTTTACCC
<i>Zootecus insularis</i>	TTTACTAATT GTTTC AAGTT TAGTTGAGGG GGGTGCAGGG ACAGGCTGAA CCGTATATCC
<i>Gibbulinella dewinteri</i>	TTTTTTAATT ATTTCTAGAA TAGTAGAAGG TGGAGCTGGA ACAGGTTGAA CCGTATACCC

	310 320 330 340 350 360
<i>Achatina achatina</i>	ACCTTTAAGA TCTACTTTAG GTCATAGAGG TGCCTCGGTA GATTTAGCGA TTTTTTCATT
<i>Achatina fulica</i>	GCCCTTAAGT TCTTGCTTAG GACACAGAGG GGCTTCAGTT GATTTAGCTA TTTTTTCTTT
<i>Achatina stuhlmanni</i>	TCTCTTAAGT TCATTAATTG GCCATAGTGG AGCATCAGTG GACTTGGCAA TTTTTTCTCT
<i>Cochlitoma ustulata</i>	ACCTTTAAGG TCTATCATAG GTCATAGGGG AGCATCAGTG GATTTGGCTA TCTTCTCTCT
<i>Coelioxys blandii</i>	GCCCTTAAGT GGTATTATTG GGCACAGCGG TGCCTCAGTT GATCTGGCTA TTTTTTCTTT
<i>Pyrgina umbilicata</i>	ACCCCTTAGC TCAATAATTG GCCACAGAGG TCCATCTGTT GATTTGGCAA TTTTCTCCCT
<i>Glessula ceylanica</i>	ACCATTAAGA AGTATTGTTG GACATAGCGG TCGCTCTGTT GACTTAGCTA TTTTTTCACT
<i>Ferussacia folliculus</i>	CCCTTTAAGC AGATTGGTAG GCCATAGAGG TGCTTCTGTA GATTTGGCGA TTTTCTCTCT
<i>Thyrophorella thomensis</i>	CCCCCTTAGA TCAATGATCG GTCATAGTGG TCCGTCTGTT GATTTAGCAA TCTTCTCTCT
<i>Allopeas clavulinum</i>	ACCATTAAGA TCAATATTAG GGCACGGAGG ACCTTCTGTA GACTTAGCTA TTTTTTCTCT
<i>Bocageia sp.</i>	TCCTTTAAGA AGTCTGATAG GTCATAGAGG TGCCTCCGTG GATTTGGCTA TTTTTTCACT
<i>Eutomopeas layardi</i>	ACCTTTAAGA TCAATATTAG GGCACGGTGG GCCCTCAGTA GACTTAGCTA TTTTTTCTCT
<i>Leptinaria lamellata</i>	ACCTTTAAGT GGGATTTTAG GTCATAGGGG CGCTTCTGTC GACTTAGCTA TTTTTTCTCT
<i>Paropeas achatinaceum</i>	ACCATTAAGG TCAATACTAG GTCATAGAGG CCCATCAGTA GACTTAGCTA TTTTTTCTCT
<i>Riebeckia sp.</i>	TCCTTTAAGA GGTATTTTAG GACATAGGGG AGCATCTGTT GATTTAGCTA TCTTTTCTCT
<i>Rumina decollata</i>	ACCCCTAAGC AGAATCATAG GACATAGTGG GGCATCTGTT GACTTAGCTA TTTTCTCTCT
<i>Subulina octona</i>	ACCTTTAAGC AGAATTATAG GTCATGCTGG GGCTTCAGTT GATCTTGCTA TTTTTTCTTT
<i>Subulina striatella</i>	CCCTCTTAGA GGAATTTTAG GACATGCTGG TCGCTCAGTG GATCTAGCTA TTTTTTCTTT
<i>Subulina vitrea</i>	CCCCCTAAGA GGGATCTTAG GCCATAGTGG CGCTTCTGTT GATTTAGCCA TTTTTTCTTT
<i>Subulona sp.</i>	TCCATTAAGT GGGATTAGAG GTCATGGTGG AGCTTCTGTT GACTTAGCAA TTTTTTCTTT
<i>Tortaxis erectus</i>	TCCTTTAAGT GGAATTATGG GCCATAGTGG TGCTTCAGTT GACTTAGCTA TTTTTTCTTT
<i>Xerocerastus sp.</i>	CCCATTAAGT AGTATTATTG GACACAGTGG TGCTTCTGTA GATTTAGCTA TCTTCTCTCT
<i>Zootecus insularis</i>	ACCTCTAAGA AGAAGCTTGG GACATAGTGG AGCTTCTGTC GATCTGGCTA TTTTCTCACT
<i>Gibbulinella dewinteri</i>	ACCATTAAGT AGCCTATTAG GACATGGAGG AGCTTCTGTA GATTTAGTTA TTTTTTCTTT

	370 380 390 400 410 420
<i>Achatina achatina</i>	ACATTTGGCT GGTATATCAT CTATCTTAGG GGCTATTAAC TTTATTACTA CAATTTTTAA
<i>Achatina fulica</i>	ACATTTGGCT GGTGTATCCT CAATTTTAGG GGCAATCAAT TTTATTACGA CCGTGTATAA
<i>Achatina stuhlmanni</i>	ACACTTAGCC GGAATATCCT CTATTCTCGG AGCTATCAAC TTTATTACTA CAATTTTTAA
<i>Cochlitoma ustulata</i>	ACACTTAGCT GGTATATCCT CTATTCTCGG GGCTATTAAC TTTATCACAA CTATCTTTAA
<i>Coelioxaxis blandii</i>	ACATTTAGCT GGTATGTCTT CAATTTTAGG TGCTATTAAT TTTATTACAA CTATTTTTAA
<i>Pyrgina umbilicata</i>	CCATTTAGCA GGAATTTCTT CAATCTTAGG GGCAATTAAC TTTATTACCA CTATTTTTAA
<i>Glessula ceylanica</i>	TCACCTAGCA GGAATATCAT CAATTTTAGG TGCTATCAAT TTTATTACAA CTATTTTTAA
<i>Ferussacia folliculus</i>	ACATCTTGCT GGAATATCCT CAATTTTAGG GGCAATTAAT TTTATTACTA CTATCTTTAA
<i>Thyrophorella thomensis</i>	TCACTTAGCA GGGATATCCT CTATTTTGGG TGCAATTAAC TTTATTACAA CAATCTTTAA
<i>Allopeas clavulinum</i>	TCATTTAGCT GGGATGTCTT CTATTCTTGG TGCTATCAAT TTTATTACTA CGATTTTTAA
<i>Bocageia sp.</i>	TCATCTTGCG GGTATATCCT CAATCTTAGG TGCCATCAAT TTTATTACTA CTATTTTTAA
<i>Eutomopeas layardi</i>	TCACTTAGCG GGGATATCCT CAATTTTAGG GGCTATCAAT TTTATTACCA CAATTTTTAA
<i>Leptinaria lamellata</i>	CCATTTAGCT GGAATATCCT CAATTTTAGG TGCAATTAAT TTTATTACTA CAATTTTTAA
<i>Paropeas achatinaceum</i>	CCATTTGGCT GGGATATCCT CAATTTTAGG GGCTATTAAC TTTATTACAA CAATCTTTAA
<i>Riebeckia sp.</i>	TCATTTAGCA GGAATATCCT CAATCTTAGG GGCTATTAAT TTTATTACAA CTATTTTTAA
<i>Rumina decollata</i>	TCATTTAGCA GGGATATCCT CTATTTTAGG TGCAATTAAT TTTATTACTA CTATTTATAA
<i>Subulina octona</i>	GCATTTAGCA GGAATGTCTT CTATTTTAGG TGCAATTAAT TTTATTACTA CTGATTTTAA
<i>Subulina striatella</i>	ACATTTGGCT GGTATGTCTT CAATTTTAGG TGCAATTAAT TTTATTACGA CTATTTTTAA
<i>Subulina vitrea</i>	ACATTTAGCA GGAATATCCT CTATTTTAGG GGCCATTAAT TTTATTACTA CAATTTATAA
<i>Subulona sp.</i>	ACATCTAGCT GGTATATCCT CCATTTTAGG TGCAATTAAC TTTATTACAA CTATTTATAA
<i>Tortaxis erectus</i>	ACATTTAGCT GGACTTTCTT CTATTTTAGG GGCTATTAAC TTTATTACAA CTATTTTTAA
<i>Xerocerastus sp.</i>	CCACTTAGCC GGGATGTCTT CTATCTTGGG GGCTATTAAT TTTATTACTA CCATTTTTAA
<i>Zootecus insularis</i>	TCACTTGCC GGTATATCCT CAATCTTGGG AGCTATTAAT TTTATTACTA CTATCTTTAA
<i>Gibbulinella dewinteri</i>	ACATCTAGCA GGTATATCCT CTATTTTAGG GGCTATTAAT TTTATTACTA CTATCTTTAA

	430 440 450 460 470 480
<i>Achatina achatina</i>	TATACGAGCA TCAGGGATAT CTATAGAACG GGTTAGACTG TTTGTATGAT CTATTTTAGT
<i>Achatina fulica</i>	TATACGAGCC TCTGGGCTAA CTATAGAACG GGTAAGCTTA TTCGTGIGGT CAATTTTAGT
<i>Achatina stuhlmanni</i>	CATACGAGTA TCTGGACTAT CTATGGAACG GGTGAGGTTA TTTGTATGAT CAATCTTTGT
<i>Cochlitoma ustulata</i>	TATACGAGCT CAAGGCTTAT CTATGGAGCG TATAAGATTA TTTGTATGAT CTATTTTAGT
<i>Coelioxaxis blandii</i>	TATACGTGCT TCCGGATTAA CAATAGAACG TGTAAGTCTT TTTGTATGAT CTATTTTTAAT
<i>Pyrgina umbilicata</i>	TATACGTTCG TCAGGAGTTA AATATGAACG GTTAAGCTTG TTTGTTTGTAT CAATTTTAGT
<i>Glessula ceylanica</i>	CATACGAGCT CCTGGTCTTA CCATAGAACG AGTGAGGTTG TTTGTATGAT CTATTTTAGT
<i>Ferussacia folliculus</i>	TATACGATCC CCAGGTATAA CTATGGAACG GGTTAATTTA TTTGTGTTGT CTATTTTGGT
<i>Thyrophorella thomensis</i>	CATACGTTCA TCTGGAATAA GTTATGAGCG TCTAAGATTA TTTGTGTGAT CAATTTTAGT
<i>Allopeas clavulinum</i>	TATACGATCG TACAGGAATA AGTACGAACG AATAAGGTTA TTTGTTTGTAT CTATTTTGGT
<i>Bocageia sp.</i>	TATACGATCA CCTGGTATAA CAATAGAACG AACAAGATTA TTTGTATGAT CTATCTTAGT
<i>Eutomopeas layardi</i>	TATACGTTCA TCTGGGTTGA AATATGAGCG AATAAGATTA TTTGTTTGGT CTATTTTAGT
<i>Leptinaria lamellata</i>	TATACGATCA CCTGGATTAA AAATAGAACG AGTAAGATTA TTTGTTTGTAT CAATTTTAGT
<i>Paropeas achatinaceum</i>	TATACGCTCA TCAGGGATGA AGTATGAACG TATAAGTCTA TTTGTGIGGT CTATTTTTAAT
<i>Riebeckia sp.</i>	CATACGTGCC TACAGATTAA CATGAGAACG TACTAGTTTA TTTGTTTGTAT CAATTTTAGT
<i>Rumina decollata</i>	TATACGTGCC CCAGGTCTTT CAATAGAACG GGTAAGCCTA TTTGTGTGAT CTATTTTAGT
<i>Subulina octona</i>	TATACGTCTT TCTGGTTTAT CAATAGAACG AGTTAGATTG TTTGTTTGGT CAATTTTTAAT
<i>Subulina striatella</i>	TATACGATCT TCCGGTTTAA GTTTTGGAGCG GGTGAGTTTA TTTGTATGAT CTATTTTAGT
<i>Subulina vitrea</i>	TATACGAGCC CCTGGTATAA CCCCAGAACG AATTAGGCTA TTTGTTTGTAT CAATTTTAGT
<i>Subulona sp.</i>	CATACGAGCA CCAGGCTTAA CTCCAGAACG AACTAGACTA TTTGTTTGGT CTATTTTAGT
<i>Tortaxis erectus</i>	TATACGTCTT TCAGGAATAT CTATGGAACG GGTAAGTCTA TTTGTTTGTAT CTATTTTAGT
<i>Xerocerastus sp.</i>	TATACGATCC TCTGGAATAA GTATGGAACG TGTGAGGTTG TTTGTATGAT CAATCTGTTG
<i>Zootecus insularis</i>	TATGCGCTCT CCCGGATTAA CTATGGAACG GGTGAGCTTA TTTGTATGAT CTATTTTGGT
<i>Gibbulinella dewinteri</i>	TATACGATCG CCTGGAATAA CAATAGAACG GGTGCTTTTA TTTGTTTGGT CTATTTTAGT

	490	500	510	520	530	540		
<i>Achatina achatina</i>	TACAGTATTT	TTACTGCTTT	TATCTCTCCC	AGTATTAGCG	GGTGCAATTA	CTATACTTCT		
<i>Achatina fulica</i>	TACAGTGTTT	CTACTATTGT	TATCATTGCC	AGTTCTAGCG	GGGGCAATTA	CTATACTATT		
<i>Achatina stuhlmanni</i>	TACTGTTTTC	CTTTTACTTT	TATCTCTACC	AGTCTTAGCT	GGGGCGATTA	CTATACTATT		
<i>Cochlitoma ustulata</i>	TACTGTTTTC	CTACTGCTCT	TATCTCTACC	AGTACTAGCA	GGGGCTATTA	CAATACTCCT		
<i>Coelioxaxis blandii</i>	TACAGTATTT	TTATTACTCC	TATCTTTACC	AGTCTTGCCA	GGGGCAATTA	CTATGTTATT		
<i>Pyrgina umbilicata</i>	TACTGTATTT	CTTCTCCTTC	TTTCCCTTCC	TGTATTAGCT	GGTGCAATTA	CCATATTATT		
<i>Glessula ceylanica</i>	TACAGTTTTT	TTACTTCTTC	TTTCATTACC	TGTGTTAGCT	GGAGCTATTA	CTATATTATT		
<i>Ferussacia folliculus</i>	TACAGTTTTC	CTCTTATTAT	TATCACTCCC	TGTACTTGCG	GGGGCTATTA	CTATATTATT		
<i>Thyrophorella thomensis</i>	CACTGTTTTT	CTTCTTCTCT	TATCTCTTCC	TGTTCTAGCT	GGGGCTATTA	CTATACTCTT		
<i>Allopeas clavulinum</i>	GACAGTTTTT	TTACTTCTTT	TATCTCTTCC	AGTTCTTGCC	GGTGTATCA	CTATACTTTT		
<i>Bocageia sp.</i>	CACAGTATTT	CTATTACTTT	TATCATTACC	TGTTTTAGCT	GGTGTATTA	CTATGCTTCT		
<i>Eutomopeas layardi</i>	AACAGTTTTT	TTGCTTTTAC	TATCTCTTCC	TGTTCTAGCT	GGCGCGATTA	CGATACTTTT		
<i>Leptinaria lamellata</i>	AACTGTTTTT	TTACTTCTCC	TATCGCTTCC	TGTTTTAGCT	GGGGCAATTA	CAATATTATT		
<i>Paropeas achatinaceum</i>	CACAGTTTTC	TTATTACTTC	TTTCGTTACC	TGTTCTTGCT	GGTGAATCA	CTATACTTCT		
<i>Riebeckia sp.</i>	AACAGTTTTT	TTACTTCTTT	TATCCCTTCC	TGTTTTAGCT	GGAGCAATTA	CTATACTTTT		
<i>Rumina decollata</i>	AACCGTTTTT	CTATTGCTTT	TATCCCTACC	AGTACTAGCA	GGAGCTATTA	CAATACTACT		
<i>Subulina octona</i>	TACTGTTTTT	TTATTATTAC	TATCATTGCC	TGTATTGGCT	GGTGAATTA	CAATATTATT		
<i>Subulina striatella</i>	AACGGTGTTT	TTACTATTAT	TGCTTTTACC	TGTATTGGCT	GGGGCAATCA	CTATGTTATT		
<i>Subulina vitrea</i>	AACAGTATTT	TTATTACTTT	TATCTTTACC	GGTTTTAGCT	GGGGCTATCA	CAATGTTATT		
<i>Subulona sp.</i>	TACTGTGTTT	TTATTATTGT	TATCATTACC	AGTACTAGCT	GGCGCAATTA	CTATACTACT		
<i>Tortaxis erectus</i>	TACAGTTTTC	TTGTTACTTC	TTTCACTTCC	TGTTTTAGCA	GGTGTATTA	CTATGTTATT		
<i>Xerocerastus sp.</i>	TACTGTATTC	CTTTTATTAC	TTTCACTCCC	AGTATTAGCT	GGTGTATTA	CTATATTATT		
<i>Zootecus insularis</i>	TACTGTATTT	CTATTACTGC	TATCATTACC	CGTTTTAGCG	GGTGAATTA	CTATGCTATT		
<i>Gibbulinella dewinteri</i>	TACTGTTTTC	CTCTTACTTT	TATCACTTCC	AGTTCTTGCT	GGAGCAATTA	CTATATTGTT		

	550	560	570	580	590	600		
<i>Achatina achatina</i>	GACCGATCGT	AACTTTAATA	CATCTTTTTT	TGATCCAGCT	GGAGGTGGTG	ATCCAGTGCT		
<i>Achatina fulica</i>	AACGGATCGT	AACTTTAACA	CATCTTTTTT	TGACCCAGCT	GGAGGGGGGG	ACCCAGTCTT		
<i>Achatina stuhlmanni</i>	AACTGATCGA	AATTTTAATA	CATCTTTTTT	TGATCCGGCA	GGGGGTGGTG	ATCCAATCCT		
<i>Cochlitoma ustulata</i>	TACTGATCGT	AATTTTAACA	CATCCTTTTT	CGACCCGGCA	GGGGGAGGAG	ATCCTATCTT		
<i>Coelioxaxis blandii</i>	AACAGACCGG	AACTTTAATA	CGTCATTTTT	TGACCCGGCC	GGGGGCGGAG	ATCCTGTCTT		
<i>Pyrgina umbilicata</i>	AACTGATCGT	AATTTTAATA	CTTCTTTTTT	TGATCCAGCT	GGGGGTGGTG	ATCCGTTGCT		
<i>Glessula ceylanica</i>	GACAGATCGA	AATTTTAATA	CATCTTTTTT	TGACCCAGCT	GGAGGTGGTG	ACCCAATTTT		
<i>Ferussacia folliculus</i>	AACGGATCGT	AATTTTAATA	CGTCTTTTTT	TGATCCAGCA	GGAGGTGGAG	ACCCAATTTT		
<i>Thyrophorella thomensis</i>	AACTGATCGA	AATTTTAATA	CTTCTTTCTT	TGACCCTGCG	GGAGGGGGCG	ATCCGTTGTT		
<i>Allopeas clavulinum</i>	AACTGATCGG	AATTTTAATA	CATCTTTTTT	TGATCCTGCT	GGTGGAGGGG	ACCCAGTCTT		
<i>Bocageia sp.</i>	TACAGATCGT	AATTTTAATA	CATCCTTTTT	TGATCCGGCT	GGGGGAGGGG	ATCCTATTTT		
<i>Eutomopeas layardi</i>	AACTGATCGA	AACTTTAACA	CATCTTTTTT	TGATCCGGCT	GGAGGGGGGG	ATCCTATCTT		
<i>Leptinaria lamellata</i>	AACAGATCGA	AATTTTAATA	CATCATTCTT	TGATCCAGCA	GGTGGTGGAG	ATCCAATTTT		
<i>Paropeas achatinaceum</i>	TACAGACCGT	AATTTTAATA	CATCTTTTTT	TGACCCCGCT	GGTGGCGGTG	ACCCTGTGTT		
<i>Riebeckia sp.</i>	AACAGACCGA	AATTTTAATA	CATCATTCTT	TGACCCAGCT	GGAGGAGGTG	ATCCTATTTT		
<i>Rumina decollata</i>	AACTGACCGT	AACTTTAACA	CTTCACTCTT	TGATCCTGCA	GGGGGAGGTG	ATCCAATCTT		
<i>Subulina octona</i>	AACTGATCGT	AATTTTAATA	CATCTTTTTT	TGATCCAGCT	GGTGGGGGGG	ACCCTATTCT		
<i>Subulina striatella</i>	AACTGATCGT	AATTTTAATA	CATCTTTTTT	TGACCCAGCA	GGTGGAGGGG	ACCCTATCTT		
<i>Subulina vitrea</i>	AACCGATCGT	AATTTTAACA	CATCTTTCTT	TGATCCGGCT	GGAGGGGGAG	ACCCTGTTTT		
<i>Subulona sp.</i>	GACGGACCGT	AATTTTAATA	CATCATTCTT	TGATCCTGCT	GGGGGAGGAG	ATCCAGTCTT		
<i>Tortaxis erectus</i>	GACTGATCGA	AATTTTAATA	CATCATTCTT	TGACCCAGCT	GGTGGTGGTG	ATCCAATCTT		
<i>Xerocerastus sp.</i>	AACTGATCGT	AATTTCAATA	CTTCACTTTT	TGATCCAGCA	GGCGGAGGTG	ACCCTGTACT		
<i>Zootecus insularis</i>	GACTGATCGA	AATTTTAACA	CGTCGTTTTT	TGATCCAGCT	GGGGGTGGTG	ATCCTATTTT		
<i>Gibbulinella dewinteri</i>	AACTGATCGG	AATTTTAATA	CGTCTTTTTT	TGATCCTGCT	GGGGGTGGGG	ATCCTATTTT		

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<i>Achatina achatina</i>	TTACCAG
<i>Achatina fulica</i>	GTACCAG
<i>Achatina stuhlmanni</i>	ATACCAG
<i>Cochlitoma ustulata</i>	GTATCAG
<i>Coelioxys blandii</i>	TTACCAA
<i>Pyrgina umbilicata</i>	ATATCAA
<i>Glessula ceylanica</i>	ATATCAG
<i>Ferussacia folliculus</i>	ATACCAA
<i>Thyrophorella thomensis</i>	ATATCAA
<i>Allopeas clavulinum</i>	ATATCAG
<i>Bocageia</i> sp.	ATATCAA
<i>Eutomopeas layardi</i>	ATACCAA
<i>Leptinaria lamellata</i>	ATATCAA
<i>Paropeas achatinaceum</i>	GTATCAA
<i>Riebeckia</i> sp.	GTATCAA
<i>Rumina decollata</i>	ATATCAG
<i>Subulina octona</i>	TTATCAG
<i>Subulina striatella</i>	ATATCAG
<i>Subulina vitrea</i>	ATACCAA
<i>Subulona</i> sp.	ATATCAA
<i>Tortaxis erectus</i>	TTATCAA
<i>Xerocerastus</i> sp.	TTATCAA
<i>Zootecus insularis</i>	GTACCAA
<i>Gibbulinella dewinteri</i>	ATATCAA
<i>Gonaxis quadrilateralis</i>	ATATCAA
<i>Gonospira</i> sp.	GTATCAA

Appendix 3.5: Alignment of the 16S rRNA gene for the Achatinoidea and three streptaxid outgroup taxa. Positions included for the phylogeny analyses were marked by “m” within “NUCEOTIDES INCLUDED”.

	10 20 30 40 50 60
Achatina fulica	TAGCTTAAAG A-----GTGC GTAACACAGA TTTTAAGTAA CCCCTGCCCG
Achatina achatina	TAGCTTTAGG A-----TTTAAA TCTTTAGTAC CTCTGCCCCA
Achatina stuhlmanni	TAGCTAATAG G-----CT CTTTCCATAA TTATTAGTGG ACCCTGCCCG
Cochlitoma ustulata	TAGCCTGCTG T-----AGATTTA TTACAGGTGA TCCTGCCCCG
Allopeas clavulinum	-----AATTAA TATAAGGTTA GACCTGCTCA
Bocageia sp.	-----AGATAT ATTCTGCCCA
Leptinaria lamellata	TTTCCTAAAG A-----TTTTTA TTTTAGGTTC AGCCTGCCCA
Paropeas clavulinum	TAGCCTTAAG G-----TATTA TTTAAGGTAA AGCCTGCTCA
Riebeckia sp.	-----GATGGCTA TACTTAGCAT CCCTGCCCCA
Rumina decollata	-----TTTAGGTAA CCCCTGCCCA
Subulina octona	TAGCTTGAAG A-----AAATTGG TTTTAGGTTA TTTCTGCCCA
Subulina striatella	-----TACTA TTTTAAGTGA CTTCTGCCCG
Subulina vitrea	TAGCTTAAAG A-----GTATTTTTA TTTTAAGTAC CTTCTGCCCA
Subulona sp.	TKRCTTAACG G-----ATATDGKGRA TGTAAAGCAT SSCTGCCCCA
Tortaxis erectus	-----CTTTATGTAA CTTCTGCCCG
Xerocerastus sp.	-----TTAGGTGA TTTCTGCCCG
Zootecus insularis	-----CATATTTTA TTAATTTGTA AGTTTATTTA TTCAAGCTT CTCCTGCCCA
Coeliaxis blandii	-----TTATAGATTA ATCCTGCCCA
Pyrgina umbilicata	-----AGCCTGCTCA
Cecilioides gokweanus	TAACCTAAAG A-----AATTTA TTTTAAGTAT ACCCTGCCCG
Ferussacia folliculus	-----A CTCTGCCCCG
Thyrophorella thomensis	-----AGCCTGCTCA
Gibbulinella dewinteri	TAGCCTAGTG A-----GTTTTA TATTAGGTTG CTTCTGCCCA
Gonaxis quadrilateralis	-----AGTGA CTTCTGCCCA
Gonospira sp.	-----AGTGT TGTCTGCCCA
NUCLEOTIDES INCLUDED	-----mmmmmmmmmm

	70 80 90 100 110 120
Achatina fulica	GTGAGTTATA GA---AACTT TAACGGCCCG AGTACCTTGA CTGTGCAAAG GTAGCATAAT
Achatina achatina	GTGATAGGTT AC---CTATT TAACGGCCCG AGTACCTTGA CTGTGCAAAG GTAGCATAAT
Achatina stuhlmanni	GTGAAGTTAA ----CCTTT TAACGGCCCG AGTACCCTGA CTGTGCTAAG GTAGCATAAT
Cochlitoma ustulata	GTGGGGCCTA TT---ACCTT AAACGGCCCG AGTACCTTGA CTGTGCAAAG GTAGCATAAT
Allopeas clavulinum	ATGAAAA--- ----TGTTT TAACAGCCCG AGTACCCTGA CTGTGCTAAG GTAGCATAAT
Bocageia sp.	ATGAACCCTA ----AGTTT TAATGGCCCG AGTACCTTGA CTGTGCAAAG GTAGCATAAT
Leptinaria lamellata	ATGATA---- ----TATTT TAATGGCCCG AGTACCCTGA CTGTGCTAAG GTAGCATAAT
Paropeas clavulinum	ATGAAT---- ----AATTT TAATAGCCCG AGTACCCTGA CTGTGCTAAG GTAGCATAAT
Riebeckia sp.	GTGAGATATA ----TTCTT AAATGGCCCG GGTACTTTGA CCGTGCTAAG GTAGCATAAT
Rumina decollata	GCGATATAT- ----AAGTT TAGCGCCCG AGTACTTTGA CTGTGCTAAG GTAGCATAAT
Subulina octona	GTGACGGTAA A---AAGTT AAATGGCCCG AGTACATTGA CTGTGCTAAG GTAGCATAAT
Subulina striatella	GTGAATAG-- ----AAATT TAACGGCCCG AGTACATTGA CTGTGCTAAG GTAGCATAAT
Subulina vitrea	ATGAAATT-- ----AATTT TAATGGCCCG AGTACTTTGA CTGTGCAAAG GTAGCATAAT
Subulona sp.	AWGAATAATT T---ATTKT TAATGGCCCG AGTACCTTGA CTGTRCTAAG GTAGCATAAT
Tortaxis erectus	GTGATT---- ----TTTTT TAACGGCCCG AGTACCCTGA CTGTGCTAAG GTAGCATAAT
Xerocerastus sp.	GTGATA---- ----TATTT TAACGGCCCG AGTACCTTGA CTGTGCTAAG GTAGCGTAAT
Zootecus insularis	GTGATTTT- ----ATATT TAACGGCCCG AGTACCCTGA CTGTGCTAAG GTAGCGTAAT
Coeliaxis blandii	GTGAATACTT GTGAAAATTT TAACGGCCCG GGTACCTTGA CCGTGCTAAG GTAGCATAAT
Pyrgina umbilicata	ATGAATAAAA A---ACTTT TAATAGCCCG AGTACCCTGA CTGTGCTAAG GTAGCATAAT
Cecilioides gokweanus	GTGAGTTATT T---TTCTT TAACGGCCCG AGTACCTTGA CTGTGCAAAG GTAGCATAAT
Ferussacia folliculus	GTGATGGT-- ----TTATT TAACGGCCCG AGTACCCTGA CTGTGCAAAG GTAGCGTAAT
Thyrophorella thomensis	ATGAA----- ----TATT TAATAGCCCG AGTACCCTGA CTGTGCTAAG GTAGCATAAT
Gibbulinella dewinteri	CTGATTAGTT AACT-AACTT TAACGGCCCG AGTACTTCTGA CTGTGCTAAG GTAGCATAAT
Gonaxis quadrilateralis	GTGAA----- ----AATTT TAACGGCCCG AGTACCCTGA CTGTGCTAAG GTAGCATAAT
Gonospira sp.	GTGATGT--- ----TTATT CAACGGCCCG AGTACTTTGA CTGTGCTAAG GTAGCATAAT
NUCLEOTIDES INCLUDED	mmmm-----mm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

	130	140	150	160	170	180
<i>Achatina fulica</i>	AATTTGTCTT	CTAATTAAGG	TCTGGAATGA	AGGGGGACA-	-CAGGGGAGA	GCTGTCTCCA
<i>Achatina achatina</i>	AATTTGTCCC	TTAATTAGGG	TCTGGTATGA	AAGGATTTT-	-CGGGAGTTT	ACTGTCTTAG
<i>Achatina stuhlmanni</i>	AATTTGTCCCT	TTAATTAAGG	TCTTGCAATGA	ATGGGTGTT-	-CGAGGAGTC	GCTGTCTTTA
<i>Cochlitoma ustulata</i>	CACCTGTCTCT	TTAATTGAGG	ACTGGAATGA	AAGGGAAGAA	ACGGGTATGA	GCTGTCTCTA
<i>Allopeas clavulinum</i>	AATTTGTCTT	TTAATTGGAG	TCTGGAATGA	ATGGATTAA-	-TGGGGGATA	ACTGTCTCTT
<i>Bocageia sp.</i>	AATTTGTCCCT	TTAATTAGGG	TCTAGAATGA	ATGAAAAATT	TTGGAAAAAC	TCTGTCTCAT
<i>Leptinaria lamellata</i>	AATTAGTCCT	TTAATTGAGG	TCTGGAATAA	ATGGTATAA-	-CGGATAATA	ACTGTCTCAT
<i>Paropeas clavulinum</i>	AATTTGTCTT	TTAATTGAGG	TCTAGAATGA	ATGGTTTAT-	-TGGGGAATA	ACTGTCTTAA
<i>Riebeckia sp.</i>	AATTTGTCCCT	TTAATTAAGG	TCTAGAATGA	ATGGGGTTA-	-CTGGAAAGT	ATTGTCTCTG
<i>Rumina decollata</i>	AACTTGTCCCT	TTAATTGAGG	TCTTGAATGA	ACGGGTTTA-	-CGAAAAATT	TCTGTCTCCT
<i>Subulina octona</i>	AATTTGTCTT	TTAATTGAGG	TCTAGAATGA	ATGAAATTA-	-TGGGGAATA	GCTGTCTCAA
<i>Subulina striatella</i>	AAATTGTCTC	TTAATTGGGG	TCTAGAATTA	ACGGAGTTA-	-TGGGGGATA	ACTGTCTCTA
<i>Subulina vitrea</i>	AATTTGTCTT	TTAATTGGGG	ACTAGAATGA	ATGAAAAAA-	-CGGAGGAAA	GCTGTCTCTT
<i>Subulona sp.</i>	CACCTGTCTT	TTAATTGAGG	ACTAGAATGA	ATGGGGGCA-	-CGGGAACCR	GCTGTCTGTA
<i>Tortaxis erectus</i>	AAATTTGCTCT	CTAATTAGGG	TCTAGAATGA	AAGAAGATT-	-TAGGGATAA	ACTGTCTCTG
<i>Xerocerastus sp.</i>	AATTTGGCTT	TTAATTGAGG	TCTTGAATGA	AAGGGATAA-	-CGTGGGGGA	ATTGTCTCTT
<i>Zootecus insularis</i>	AATTTGTCTT	TTAATTGGGG	TCAGGTATGA	ACGGAGATCA	-CACATAGCC	GCTGTCTCTT
<i>Coelioxix blandii</i>	AATTTGTCTT	TTAATTGAGG	TCTAGAATGA	ACGGATATT-	-CTGAAATTA	GCTGTCTTAA
<i>Pyrgina umbilicata</i>	AATTTGTCTT	TTAATTGAGG	TCTAGAATGA	ATGGCACCT-	-TGGGGTGAA	GCTGTCTTAA
<i>Ceciloides gokweanus</i>	AATTTGTCTT	TTAATTGGAG	TCTTGAATAA	ATGGGAATTT	-GGGGAGG-G	GCTGTCTCTT
<i>Ferussacia folliculus</i>	AATTTGTCTT	TTAATTGGGG	TCTAGAATGA	AAGGAATTTT	-GGGAAGA-A	GGTGTCTCAT
<i>Thyrophorella thomensis</i>	AACTTGTCTC	CTAATTAGGG	TCTAGAATAA	ACGGCATCTA	-GAAGAGA-A	TCTGTCTTGG
<i>Gibbulinella dewinteri</i>	AAATTGATTT	TTAATTGGAG	TCTAGAATGA	AAGGGATTA-	-TGGGGGGAT	ACTGTCTCTA
<i>Gonaxis quadrilateralis</i>	AAATTGATTT	TTAATTGAAG	TCTAGAATGA	ATGGAATTT-	-TGGAAAAAA	TCTGTCTTAA
<i>Gonospira sp.</i>	AAATTTGCTT	ATAATTAAGG	ACTAGAATCA	AAGAATTTA-	-TGGAGTTTA	ACTGTCTTAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	190	200	210	220	230	240
<i>Achatina fulica</i>	ATAAGGTAA	TT--TAACCT	TC-TTATCAG	GTGAAAAATTC	CTGAGCCT--	--CCGATGAA
<i>Achatina achatina</i>	TGAGGTAAAT	AT--GAATTT	ACTCAATAAG	GTGAAAAATTC	CTTAAAAAT--	--TAGATAAA
<i>Achatina stuhlmanni</i>	TTAGGGTTTA	ATT-AAATTT	AC-TCAAGAG	GTGAAAAATTC	CTCTAAATTT--	--AAGATAAA
<i>Cochlitoma ustulata</i>	GTTAGCTCTT	TT--AAATTT	AA-TTGTAG	GTGAAAAATTC	CTATACCT--	--TTAACAGA
<i>Allopeas clavulinum</i>	TAAGATTAAT	TT--TAACCT	GT-TTAAAAG	GTGAAAAAAC	CTTTGAAAA--	--CAGAAAAA
<i>Bocageia sp.</i>	TTAGAAGTTT	TG--TAAGTT	TC-TAATAAG	GTGAAAAATTC	CTTAGTGG--	--CCGAAGAA
<i>Leptinaria lamellata</i>	TGATTAATTT	TAATTAACCT	AC-TAATAAG	GTGAAAAATTC	CTTAAAA--	--TTGAAAAA
<i>Paropeas clavulinum</i>	TTCAATTAAC	TT--TAATTT	AC-TTAGAAG	GTGAAAAAAC	CTTCAAAAT--	--TAGAAAAA
<i>Riebeckia sp.</i>	CTTTATTACT	T--GAAATTT	TC-TTATCAG	GTGAAAAATTC	CTGAATTT--	--TTGAAAAA
<i>Rumina decollata</i>	GTACAAAATTT	TACTGAACCT	AC-TTAAGAG	GTGAAAAATTC	CTTTAGTA--	--ATGAAAAA
<i>Subulina octona</i>	TAAGAATTTA	TAT-GAATTT	AT-TAATAAG	GTGAAAAATTC	CTTAACT--	--TTGAAAAA
<i>Subulina striatella</i>	TAGGGTTTAT	AT--GAAGTT	AA-TTATTAG	GTGCAAAAAGC	CTAATTGT--	--TAGAAAAA
<i>Subulina vitrea</i>	GTAAGTTAAC	TT--TAATTT	AC-TTAAAAG	GTGAAAAATTC	CTTTATTT--	--AACATGAA
<i>Subulona sp.</i>	TTATATTATC	TA--TAATTT	GC-TAAGGAG	GTGAAAAATTC	CTCTAGC--	--TATAAGGT
<i>Tortaxis erectus</i>	TAGAGTTTAA	TT--GAATTT	CC-TTAAAAG	GTGAAAAATTC	CTTTATAA--	--ATGAAGTT
<i>Xerocerastus sp.</i>	TAAAAGTTTG	TA--GAATTT	GT-TTATTAG	GTGAAAAATTC	CTATATTT--	--AAATATAT
<i>Zootecus insularis</i>	AGAAGTTTTT	GT--GAAGTT	AC-TTATAAG	GTGAAAAATTC	CTTTGAGCT--	--TAGATAAA
<i>Coelioxix blandii</i>	TTTAATTTAA	TT--TAAGTT	CC-TAATAAG	GTGCAAAATTC	CTTAAATTTG	TTATGAAAAAT
<i>Pyrgina umbilicata</i>	TTGGGGATTA	TTT-TAACCT	ACTTTATAGG	GTGAAAAATTC	CCTTAAAT--	--ATTTATTT
<i>Ceciloides gokweanus</i>	ATAAATTTTT	TT--TAACCT	AC-TAAGAG	GTGAAAAATTC	CTCTGGTTA--	--TTTTTAAA
<i>Ferussacia folliculus</i>	TTAAACTTTT	TT--TAATTT	TC-TTAGGGT	GTGAAAAATTC	ACCTATTAAC	T-TTTAAGAA
<i>Thyrophorella thomensis</i>	GTAGGTAAAT	AC--TAACCT	AC-TTAAAAG	GTGAAAAATTC	CCTTGAAA--	--CTTTATTT
<i>Gibbulinella dewinteri</i>	GTTTCGATTT	ATTTGAACCT	AT-ATTTTAG	GTGAAAAATTC	CTAAGAATTT	--TTAATAAT
<i>Gonaxis quadrilateralis</i>	ATTAATTATT	TT--TAAATTT	AC-TTATAAG	GTGAAAAATTC	CTTAAATTT--	--TTAATTTA
<i>Gonospira sp.</i>	TTTAATTGTT	TT--GAATTT	AC-TTAAAAG	GTGAAAAATTC	CTTTATTA--	--AAAAATAT
NUCLEOTIDES INCLUDED	-----	-----mmmmmm	mm-mmmmmmm	mmmmmmmmmm	mmmmmmmmmm	-----mmmmmm

	250	260	270	280	290	300
Achatina fulica	AGACGAGAAG	ACCCTTAGAG	TTTTTATTAT	AACTTGATA-	-----	-----	TA
Achatina achatina	AGACGAGAAG	ACCCTTAGAG	TTTTTATTAT	ATAGATAGA-	-----	-----	
Achatina stuhlmanni	AGACGAGAAG	ACCCTTAGAG	TTTTTAATGAA	AAGTAAAC--	-----	-----	
Cochlitoma ustulata	AGACGAGAAG	ACCCTTAGAG	TTTTTATGAT	AAGCATTCT-	-----	-----	
Allopeas clavulinum	AGACGAGAAG	ACCCTTAGAG	TTTTTAATTA	ATTGAATTT-	-----	-----	
Bocageia sp.	AGACGAGAAG	ACCCTAAGAG	TTTTTATAAAA	CTTCTACAT-	-----	-----	
Leptinaria lamellata	AGACGAGAAG	ACCCTAAGAA	TTTTTATGAA	AAAG-----	-----	-----	
Paropeas clavulinum	AGACGAGAAG	ACCCTTAGAG	TTTTAATAAAA	ATGTTAATT-	-----	-----	
Riebeckia sp.	AGACGAGAAG	ACCCTTAGAA	TTTTTATAAAT	ACCTTAC---	-----	-----	
Rumina decollata	AGACGAGAAG	ACCCTTAGAA	TTTTAATAAAA	AACTGTCAT-	-----	-----	
Subulina octona	AGACGAGAAG	ACCCTAAGAG	TTTTTATAAAG	TAGTAAAAA-	-----	-----	T
Subulina striatella	AGACGAGAAG	ACCCTATGAA	TTTTAATTA	AATATTTGA-	-----	-----	
Subulina vitrea	AGACGAGAAG	ACCCTTAGAA	TTTTTATAAAA	CATAATTTTC-	-----	-----	
Subulona sp.	AGACGAGAAG	ACCCTAAGAA	TTTTTACACCT	GACCAGCGC-	-----	-----	
Tortaxis erectus	AGACGAGAAG	ACCCTTAGAA	TTTTTATAAAT	GTAACCTCTT-	-----	-----	TATAATA
Xerocerastus sp.	AGACGAGAAG	ACCCTTAGAA	TTTTAATTAAT	TG-----	-----	-----	
Zootecus insularis	AGACGAGAAG	ACCCTTAGAA	TTTTTATAAAT	AGGTAAGAA-	-----	-----	
Coelioxix blandii	AGACGAGAAG	ACCCTTAGAA	TTTTTATTTT	ATTTAAATA-	-----	-----	
Pyrgina umbilicata	AGACGAGAAG	ACCCTTAGAA	TTTTTAATTA	AAGAGG---	-----	-----	
Cecilioides gokweanus	AGACGAGAAG	ACCCTTAGAG	TTTTAATAAAA	ATTTAATTTTC	ATATAATTTA	AGTAAGAATT	
Ferussacia folliculus	AGACGAGAAG	ACCCTTAGAG	TTTTAATCGTG	TGAGAGTAT-	-----	-----	
Thyrophorella thomensis	AGACGAGAAG	ACCCTCAGAA	CTTTAACTAA	TGAAGG---	-----	-----	
Gibbulinella dewinteri	AGACGAGAAG	ACCCTTAGAA	TTTTTAAAAAT	AAATAAAA--	-----	-----	
Gonaxis quadrilateralis	AGACGAGAAG	ACCCTTAGAG	TTTTTATAAAA	AGG-----	-----	-----	
Gonospira sp.	AGACGAGAAG	ACCCTATGAA	TTTTTATAAAT	TGTTAGA---	-----	-----	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmm---	-----	-----	-----	

	310	320	330	340	350	360
Achatina fulica	TTAAGATTCT	TGTTCTGTTT	TTGTTGGGGC	GACAGGGTTA	CA-----	-----	A
Achatina achatina	-----TT	CTATTCATTT	TCGCTGGGGC	AGCGAGATTA	CA-----	-----	
Achatina stuhlmanni	-----TTT	ACTTAAGTTT	TTGTTGGGGC	AACAAGATGG	CA--AAAATT	AGTAATACTT	
Cochlitoma ustulata	---ACATATG	CTGTCAGTTT	TCGTTGGGGC	AACGAGATTG	CA-----	-----	TG
Allopeas clavulinum	--TTTTAATG	TTTATTTTTT	TTGTTGGGGC	GACATAGTAA	CA-----	-----	
Bocageia sp.	CAACTTTAGA	ATAGGCGTTT	TCGTTGGGGC	GACGAAATTA	CA-----	-----	ATAGTA
Leptinaria lamellata	-----TTAA	ACTTAAGTTT	TCGTTGGGGC	GACGGGTTTT	CA-----	-----	
Paropeas clavulinum	-----ATT	TTCTTTTTTT	TTGTTGGGGC	GACAAAGTAA	CA-----	-----	
Riebeckia sp.	-----TAAA	GGGAAAGTTT	TTGTTGGGGC	GACAAAGTTA	CA-----	-----	AG
Rumina decollata	--TGCCTGAT	TGTTTAGTTT	TCGTTGGGGC	GACGAGGTAG	CA-----	-----	AT
Subulina octona	TATTAGATTT	ACATAAGTTT	TCGTTGGGGC	GACGGGATTA	CA-----	-----	
Subulina striatella	-GAAGCAAAG	TATTAGATTT	TCGTTGGGGC	GACGGAATTA	CA-----	-----	TAAAGAGTT
Subulina vitrea	---AACTTAT	TATTGTATTT	TCGTTGGGGC	AACGGGCTGA	CA-----	-----	CG
Subulona sp.	-AACTCGTCT	GWATAGTTT	TCGTTGGGGC	GACGGATTA	CA-----	-----	ACTC
Tortaxis erectus	TTATTGTAGG	TTAATTTTTT	TCGTTGGGGC	GACGAAATTA	CA-----	-----	AAAATTTAT
Xerocerastus sp.	--TTTTAAAAG	TATAACATTT	TTGTTGGGGC	GACAAAGTTA	CA-----	-----	
Zootecus insularis	-----CTTTT	ACTTTTGTTT	TCGTTGGGGC	GACGAGGTGA	CATGTTGCTT	TTAGGTGGAC	
Coelioxix blandii	ATTAATTTTA	ATTAATGTTT	TCGTTGGGGC	GACGGGTTTA	CA-----	-----	
Pyrgina umbilicata	---TTACTCA	CTTTCATTTT	TCATTGGGGC	GATGAATTA	CA-----	-----	
Cecilioides gokweanus	ATTATATTTT	AAATTAGTTT	TCGTTGGGGC	GACGTAGTTA	CA-----	-----	
Ferussacia folliculus	---GTCTCTT	CTTGACGATT	TTGTTGGGGC	GACAAAGTTT	CA-----	-----	
Thyrophorella thomensis	--TTATATAT	CTTCTGTTTT	TTGTTGGGGC	GACAAAGTTA	CA-----	-----	
Gibbulinella dewinteri	--ATTATTTT	ATGAGTTTTT	TTGTTGGGGT	GACAAAATAA	CA-----	-----	GGTT
Gonaxis quadrilateralis	---TAATTA	ACTTTTTTTT	TTGTTGGGGT	GACAAAGTAR	AA-----	-----	A
Gonospira sp.	-----TTTTTC	TAATTTATTT	TTGTTGGGGC	AACAAGATAG	CA-----	-----	
NUCLEOTIDES INCLUDED	-----	-----mm	mmmmmmmmmm	mmmmmmmmmm	mm-----	-----	

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          370          380          390          400          410          420
Achatina fulica TAGATAA CACTT ACCCTACCAT ATTTTTATAT GGCGATCATT ATACTTT---
Achatina achatina ----TCTTTA CTTTTAATTA ATCTTTACTT ATCTAAACAT TTGCCAAGTA TTTATCCTAA
Achatina stuhlmanni AAACCCATCT AATTTTCTAA AGTGAATCAG TCAATTATTA TAAGAA---
Cochlitoma ustulata AAATAAACCA ATCATACGTT AATAGTAGAT AAGTCAATCT TAATAGGTA-
Allopeas clavulinum AAATTAACCT ACTATATTAT TTTATTTTAC ACAACGCTTA AGCTATA---
Bocageia sp. ACCATAACTA ATTAAGATTA ATCTTAAACA AATCAAAAGA ATTTTAACT---
Leptinaria lamellata --CATACTAT AACTTTACCT GATAATGATT ATTTTACTAA CCAAGACAAT TATTTATTA-
Paropeas clavulinum ----TATT ACTTATAACT TACTTTTTTA TTAGTTTATC TGACATCTTT ATTGAG---
Riebeckia sp. TTATATAATA ACTAACTATA AATAATTATT TTTGCCATAG TTTTATTGTG AT-----
Rumina decollata CAAATAACCT ACCTATTAAT ATTTAATACA CCGAAATCTA TTAAACA---
Subulina octona -TATAATTTA AACTTTTAA TTATAGAATA GGGATATTTT CCAATTTTGT TAGAA----
Subulina striatella AAAGAATATA ACTTTGTAA ACTTTTAA TTATTGAGTT TGTATATACC AGGTGATACC
Subulina vitrea GATTAATTC GGCCAAAAAG ATTTATATAT AAACCTATAT TTTCAAGAG-
Subulona sp. TACATAACTT AGCTGATAG AGTAAATTA YTATATARAT CTTGTTTTRTA AATGA-----
Tortaxis erectus GAACCTATTG TTTTGTGTTT TT----- -ATGTGCCGA ATCATATTAA GAA-----
Xerocerastus sp. ----TFTA ACTTACTATA TTATTTAAGC TACTAGCCGA GTTAATTATG AG-----
Zootecus insularis CCTGAAACTC ACCTTTAATT TTAA-AATTT TATAAGCCGA ATTTTATTAT GTA-----
Coelioxix blandii -TTCTGCATA AACTAATCTA TAAATTTTTC TTAATAAACCC GAGGATTTT ATCTT----
Pyrgina umbilicata ----TTTTT AGGTTTAACT TAATTTTTTT GTTTAGTACA AGACACTTTC TACAA----
Cecilioides gokweanus --TGATGTGA ACTAACTTAA TGAATTATTG TATTTTAAATA AGTTTATTT AATTAATTT
Ferussacia folliculus TAAATAACAA ACCTTTATGA AGTTTTTCAA GACGATTATA TTATGGT---
Thyrophorella thomensis GCAATAACTT AACTTTAAGT TCATTTTACA AGGTATCGTC TTAGGA----
Gibbulinella dewinteri AAATCAACTT ATTTTGGTA TTAGTTTAC AAGAAAGTTT TAATAACAGG AA-----
Gonaxis quadrilateralis AAATTAACCT ACATTATAAT TCAAGCCGAG TTATACTATG AA-----
Gonospira sp. TAAAAAATCT ATCTTAAAAAT GATTTACTAC ATTTCAAAGT TTTTATAA-
NUCLEOTIDES INCLUDED -----mm mmm-----

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          430          440          450          460          470          480
Achatina fulica -----TA ATTAAGCTAC CTAAGGGATA ACAGCGTAAT CTTTTTTTTT
Achatina achatina GAA-----AA ACTAAACTAC CCAAGGGATA ACAGCATAAT TCCTT-AGGG
Achatina stuhlmanni -----CA GAAAAACTAC CTAAGGGATA ACAGCGTAAT CTAAT-ATTA
Cochlitoma ustulata -----TA ATTAAACTAC CTAAGGGATA ACAGCGTAAT CTTAT-TTAT
Allopeas clavulinum -----AG AAGAAACTAC CTTAGGGATA ACAGCATAAT TTTTT-AAAA
Bocageia sp. -----TG AATAAACTAC CTTAGGGATA ACAGCATAAT CTATT-TTTA
Leptinaria lamellata -----AG ATTAAATTAC CTTAGGGATA ACAGCATAAT ATTTT-ATAA
Paropeas clavulinum -----AA ATTAAACTAC CTTAGGGATA ACAGCATAAT TTTA--ATCA
Riebeckia sp. -----TA AAAAAATTAC CTAAGGGATA ACAGCATAAT CTTAA-ATAA
Rumina decollata -----TA ATTAAATTAC CTAAGGGATA ACAGCATAAT ACTAA-GTAG
Subulina octona -----TA AGTAAACTAC CTTAGGGATA ACAGCATAAT ACTT--TTAG
Subulina striatella TTTTA-----TA AGAAAATTAC CTTAGGGATA ACAGCATAAT ACTT--TTAG
Subulina vitrea -----CA TTTAAATTAC CTAAGGGATA ACAGCATAAT TTTTT-AAAA
Subulona sp. -----TA ATTAAATTAC CTTAGGGATA ACAGCATAAT CTTTW-ATAA
Tortaxis erectus -----AA TTTAAATTAC CTAAGGGATA ACAGCATAAT ATTTT-TTTA
Xerocerastus sp. -----TG ATAAAATTAC CTGAGGGATA ACAGCATAAT TTAT--TTTA
Zootecus insularis -----AA ATTAAATTAC CTGAGGGATA ACAGCATAAT ATCCT-TTGA
Coelioxix blandii -----TA TATAAATTAC CTAAGGGATA ACAGCATAAT CTTAG-ATAA
Pyrgina umbilicata -----AG GAGAAGTTAC CTGAGGGATA ACAGCATTAT TTATT-TTTA
Cecilioides gokweanus AACGAGAAAT TTTAATTTTA ATTAAGCTAC CTTAGGGATA ACAGCATAAT ATTT--TTAA
Ferussacia folliculus -----TG AGAAAACTAC CTAAGGGATA ACAGCATAAT TCTAT-AAAG
Thyrophorella thomensis -----AG ATTAAGTTAC CTGAGGGATA ACAGCATTAT TTATT-AGTA
Gibbulinella dewinteri -----AG ATTAAATTAC CTTAGGGATA ACAGCATAAT TCTTT-TACG
Gonaxis quadrilateralis -----AG AAAAAACTAC CTTAGGGATA ACAGCATAAT TTTTT-TTAA
Gonospira sp. -----AG GAGAAATTAC CTTAGGGATA ACAGCGTAAT TTCCTTTGA
NUCLEOTIDES INCLUDED -----mmmmmmmm mmmmmmmmm mmmmmmmmm mmm-----mm

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      ....|....| ....|....| ....|....| ....|....| ....|....|
      490      500      510      520      530      540
Achatina fulica      GGTTTGCGAC  CTCGATGTTG  GAC-TAGGGG  CC---TACTG  GTTAGTTACC  ATTA-----T
Achatina achatina   AGATTGTGAC  CTCGATGTTG  GAC-TAGGGC  TC---TATTG  GTTAACCTACC AAGA-----A
Achatina stuhlmanni GGATTGCGAC  CTCGATGTTG  GAC-TAGGGA  CC---TAATG  ATGAGATATC  AAGGTGACTA
Cochlitoma ustulata GGATTGCGAC  CTCGATGTTG  GAC-TAGGGA  CC---CAGTG  GTGAGAAACC  AGCT-----
Allopeas clavulinum AGATTGTGAC  CTCGATGTTG  GAC-TAGGGA  CT---TTATG  GTTAACCACC  TTAA-----
Bocageia sp.        GGATTGTGAC  CTCGATGTTG  GAC-TAGGGT  CA---GAAAG  GTAAGCAATC  AAAA-----
Leptinaria lamellata AGATTGTGAC  CTCGATGTTG  GAC-TAGGGA  CC---TATAG  GTTAACCACC  AATA-----
Paropeas clavulinum AGATTGTGAC  CTCGATGTTG  GAC-TAGGGA  CT---TTATG  GTTAACCTACC TGAT-----
Riebeckia sp.       GGTTTGTGAC  CTCGATGTTG  GAT-TAGGAA  CA---TAATG  ATTAACAATC  AGCT-----
Rumina decollata    TGTTTGTGAC  CTCGATGTTG  GAC-TAGGGA  CT---AATTG  GTTAACCACC  AGGA-----
Subulina octona     TGATTGTGAC  CTCGATGTTG  GAC-TAGGGA  CAAATGAATG  GTTAGATGCC  TGTT-----
Subulina striatella TGTTTGTGAC  CTCGATGTTG  GAC-TAGGGA  CA--TTAGTG  GCGAGATGTC  AAAT-----
Subulina vitrea     AGATTGTGAC  CTCGATGTTG  GAT-TAGGAA  CT---TAATG  GTTAGCCACC  AAGT-----
Subulona sp.        GGATTRTGAC  CTCGAWGTTG  GAT-TAGGGA  TT--AGATAG  GTTAGCCATC  TTTT-----T
Tortaxis erectus    TGATTGTGAC  CTCGATGTTG  GAC-TAGGGA  CT---TAATG  GTTAATAACT  AAAA-----
Xerocerastus sp.   AGTTTGTGAC  CTCGATGTTG  GAT-TAGGGA  AA--ATAAAG  GTTAGTAATT  TTTA-----
Zootecus insularis TGATTGTGAC  CTCGATGTTG  GACTTAGGGC  CC---AAATG  GTTAGCCACC  AAGC-----
Coelioxix blandii  TGATTGTGAC  CTCGATGTTG  GAT-TAGGGC  CT---AAAAG  GTAAGTAACC  TAGT-----
Pyrgina umbilicata AGATTGTGAC  CTCGATGTTG  GAC-TAGGGC  CC---TGAGG  GTTAACCACC  CAAA-----
Cecilioides gokweanus TGATTGTGAC  CTCGATGTTG  GAC-TAGGGA  CT---TTCTG  ATTAACCATC  AAAC-----
Ferussacia folliculus AGATTATGAC  CTCGATGTTG  GAC-TAGGAA  AC---TTATG  GTTAGACACC  AAAA-----
Thyrophorella thomensis AGATTGTGAC  CTCGATGTTG  GAC-TAGGGC  CC--ATT-AG  GTTAACCACC  TTAA-----
Gibbulinella dewinteri AGTTTGTGAC  CTCGATGTTG  GAC-TAGGAA  CC---TAATA  ATAAGCAATT  AAAT-----
Gonaxis quadrilateralis AGATTGTGAC  CTCGATGTTG  GAC-TAGGAA  CT---TAATA  ATTAGTAATT  AAAA-----
Gonospira sp.      AGATTGCGAC  CTCGATGTTG  GAC-TAGGAA  CC---TTATG  ACTAAAAGTC  TAAA-----
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmm--mmmmmm mmm---mm--mm mmmmmmmmmmm mmmmm-----

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      ....|....| ....|....| ....|....| ....
      550      560      570
Achatina fulica      GGG-GCGGTT  CTGTTCGAAC  TTTTCCTACC  CTAC
Achatina achatina   TGT-AAGGTT  CTGTTCGAAC  TTT-TTATCC  CTAC
Achatina stuhlmanni TGG-ATTGTT  CTGTTCGAAC  AC--TAGACC  CTAC
Cochlitoma ustulata AGG-CAAGTT  CTGTTCGAAC  TTT-CCACC  CTAC
Allopeas clavulinum TAGATTTGCT  CTGTTCGAGC  TT--TAACCC  CTAC
Bocageia sp.        CTG-GATGTT  CTGTTCGAAC  ATT-TTACCC  CTAC
Leptinaria lamellata AGG-ATAGTT  CTGTTCGAAC  TC--TGTACC  CTAC
Paropeas clavulinum TAG-ATTGCT  CTGTTCGAGC  AC--TTGTTC  CTAC
Riebeckia sp.       TTG-TAAGTT  CTGTTCGAAC  TTT-TACTTC  CTAC
Rumina decollata    TAG-ATGATT  CTGTTCGAAT  CC--TTCACC  CTAC
Subulina octona     TTG-CTGGTT  CTGTTCGAAC  TT--AAAACC  CTAC
Subulina striatella TTG-CAGGTT  CTGTTCGAAC  AT--ATAACC  CTAC
Subulina vitrea     AAG-TTAGTT  CTGTTCGAAC  CC--TTCCTC  CTAC
Subulona sp.        TAA-TGAAYT  CTGTTCTRAAY TWA-ATCNCC  CTAC
Tortaxis erectus    AAG-ATTGTT  CTGTTCGAAC  ATA-ACAACC  CTAC
Xerocerastus sp.   -TA-TAAGTT  CTGTTCGAAC  ATT-TTTACC  CTAC
Zootecus insularis TGG-ATGGTT  CTGTTCGAAC  CG--TTTACC  CTAC
Coelioxix blandii  TAG-ATAGTT  CTGTTCGAAC  TT--TTTACC  CTAC
Pyrgina umbilicata TGG-ACTGCT  CTGTTCGAGC  AT--TGTTC  CTAC
Cecilioides gokweanus TAGATTTGTT  CTGTTCGAAC  TTT-TTTACC  CTAC
Ferussacia folliculus TGT-TATGTT  CTGTTCGAAC  ATT-AAACTC  CTAC
Thyrophorella thomensis CGG-ATAGCT  CTGTTCGAGC  TT--CTTACC  CTAC
Gibbulinella dewinteri TGG-AAAGTT  CTGTTCGAAC  AA--ATTATC  CTAC
Gonaxis quadrilateralis TTG-ATGGTT  CTGTTCGAAC  TT--ATTTTC  CTAC
Gonospira sp.      AGG-AGTGTT  CTGTTCGAAC  AAT-TATTTT  CTAC
NUCLEOTIDES INCLUDED -mm-----mmm mmmmmmmmmmm -----mm mmmmm

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Appendix 3.6: Log likelihood scores of the different models used for the Achatinoidea using (A) 3435 unambiguously aligned nucleotides from the rRNA gene cluster; (B) 861 unambiguously aligned nucleotides of actin; (B.1) 574 unambiguously aligned nucleotides of the 1st and 2nd codon positions of actin; (B.2) 287 unambiguously aligned nucleotides of the 3rd codon position of actin; (C) 328 unambiguously aligned nucleotides of histone 3; (C.1) 218 unambiguously aligned nucleotides of the 1st and 2nd codon positions of histone 3; (C.2) 110 unambiguously aligned nucleotides of the 3rd codon position of histone 3; (D) 607 unambiguously aligned nucleotides of CO1; (D.1) 404 unambiguously aligned nucleotides of the 1st and 2nd codon positions of CO1; (D.2) 203 unambiguously aligned nucleotides of the 3rd codon position of CO1; and (E) 294 unambiguously aligned nucleotides of the 16S rRNA gene. Scores in **bold** belong to optimal models as determined by Likelihood Ratio Test. Note that the K2P and F81 models are non-nested and therefore cannot be compared for significant difference using the LRT.

	(A)	(B)	(B.1)	(B.2)	(C)	(C.1)	(C.2)	(D)	(D.1)	(D.2)	(E)
Model	rRNA gene cluster	Actin-combined	Actin-1 st +2 nd codons	Actin 3 rd codons	Histone 3	Histone 3 1 st +2 nd codons	Histone 3 3 rd codons	CO1	CO1 1 st +2 nd codons	CO1 3 rd codons	16S rRNA
JC69	8722.54544	7034.70607	1582.96160	4551.89700	2263.05590	387.26745	1519.62452	9562.61908	2707.77230	5655.47905	3720.10115
JC69+Γ	8166.65498	6318.61908	1457.51470	4364.19564	2055.16076	387.26745	1503.87233	8204.89347	2288.23519	5546.19261	3285.58073
F81	8696.59840	6997.51585	1577.09006	4404.85550	2259.84534	384.18591	1516.51436	9502.53463	2670.20603	5476.35282	3669.15845
F81+Γ	8139.63345	6263.18854	1452.88144	4302.40699	2050.07377	384.18591	1499.41097	8005.17426	2231.16774	5356.26679	3214.09399
K2P	8604.34705	6815.80245	1569.84105	4225.33326	2188.48052	385.69744	1444.53135	9484.47349	2553.77652	5631.30210	3670.91274
K2P+Γ	8038.52028	6058.78021	1439.71055	4120.55651	1977.17697	385.69744	1425.02655	8027.24407	2125.16439	5511.17394	3223.49673
HKY85	8577.13470	6776.05418	1567.69219	4205.14693	2183.03436	382.58884	1438.38403	9391.53749	2525.16880	5283.03501	3610.74395
HKY85+Γ	8000.79127	6026.10365	1439.81885	4092.36162	1969.91019	382.58884	1420.79835	7546.29497	2090.98831	4997.88304	3103.10529
TrN93	8524.93663	6730.23510	1540.64290	4201.59908	2172.17869	376.07479	1444.42135	9390.78281	2515.71421	5278.46766	3609.31276
TrN93+Γ	7970.59211	6024.01007	1420.57243	4092.24782	1965.63536	376.07479	1420.52557	7522.62965	2083.36045	4997.26819	3102.99238
GTR	8515.61458	6707.83141	1539.76409	4145.18258	2167.71615	370.44371	1411.10672	9096.60454	2506.42682	5222.31450	3536.04532
GTR+Γ	7951.71731	5993.90425	1420.05757	4051.76435	1960.84472	370.44371	1408.67625	7507.55611	2069.51524	4993.80631	3089.76804

Appendix 3.7: Summary of ambiguous sites for the actin gene for the Achatinoidea and the three streptaxid outgroup taxa

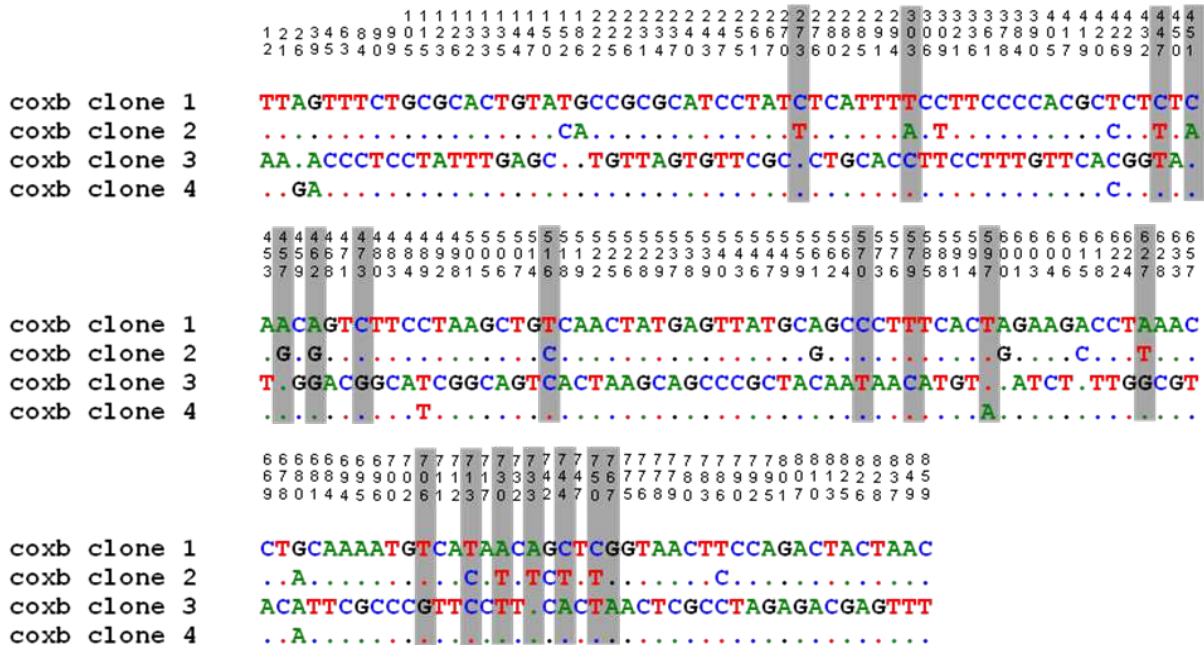
Species	Total ambiguous sites	% ambiguous sites (out of 873 total; 861 for subv)	Total ambiguous 3 rd codon positions	% ambiguous 3 rd codon positions relative to all ambiguous sites	Number of RY sites at ambiguous 3 rd codon positions	% RY sites relative to all ambiguous 3 rd codon positions	Number of ambiguous amino acids	% ambiguous amino acids (out of 291 total; 287 for subv)
ACHATINIDAE								
<i>Achatina achatina</i>	45	5.15	40	88.89	31	77.50	4	1.37
<i>Achatina fulica</i>	18	2.06	16	88.89	12	75.00	2	0.69
<i>Achatina stuhlmanni</i>	2	0.23	1	50.00	1	100.00	1	0.34
<i>Cochlitoma ustulata</i>	0	0	0	0	0	0	0	0
SUBULINIDAE								
<i>Allopeas clavulinum</i>	0	0	0	0	0	0	0	0
<i>Bocageia</i> sp.	101	11.57	85	84.16	51	60.00	14	4.81
<i>Eutomopeas layardi</i>	0	0	0	0	0	0	0	0
<i>Leptinaria lamellata</i>	0	0	0	0	0	0	0	0
<i>Paropeas achatinaceum</i>	1	0.11	0	0	0	0	1	0.34
<i>Riebeckia</i> sp.	76	8.71	59	77.63	44	74.58	15	5.15
<i>Rumina decollata</i>	29	3.32	24	82.76	16	66.67	3	1.03
<i>Subulina octona</i>	0	0	0	0	0	0	0	0
<i>Subulina striatella</i>	26	2.98	25	96.15	15	60.00	0	0
<i>Subulina vitrea</i>	37	4.30	32	86.49	26	81.25	4	1.39
<i>Subulona</i> sp.	0	0	0	0	0	0	0	0
<i>Tortaxis erectus</i>	32	3.67	26	81.25	20	76.92	7	2.41
<i>Xerocerastus</i> sp.	0	0	0	0	0	0	0	0
<i>Zootecus insularis</i>	20	2.29	18	90.00	16	88.89	1	0.34
COELIAXIDAE								
<i>Coeliaxis blandii</i>	21	2.41	13	61.90	9	69.23	7	2.41
<i>Pyrgina umbilicata</i>	88	10.08	78	88.64	55	70.51	7	2.41
THYROPHORELLIDAE								
<i>Thyrophorella thomensis</i>	62	7.10	51	82.26	37	72.55	7	2.41
GLESSULIDAE								
<i>Glessula ceylanica</i>	53	6.07	34	64.15	16	47.06	19	6.53
FERRUSSACIIDAE								
<i>Cecilioides gokweanus</i>	4	0.46	4	100.00	3	75.00	0	0
<i>Ferussacia folliculus</i>	4	0.46	4	100.00	4	100.00	0	0
STREPTAXIDAE (Outgroup)								
<i>Gibbulinella dewinteri</i>	6	0.69	6	100.00	4	66.67	0	0
<i>Gonaxis quadrilateralis</i>	55	6.30	49	89.09	34	69.39	4	1.37
<i>Gonospira</i> sp.	77	8.82	64	83.12	37	57.81	9	3.09

Appendix 3.8A: Summary of cloned actin sequences from three achatinoid species.

Species cloned	Total ambiguous sites	Total variable sites found in clones	Clone #	Sequence type*
<i>Coelioxys blandii</i>	29	163	coxb clone 1	a
			coxb clone 2	b
			coxb clone 3	c
			coxb clone 4	d
<i>Cochlitoma granulata</i>	9	72	coig clone 1	e
			coig clone 2	f
			coig clone 3	g
			coig clone 4	h
			coig clone 5	i
			coig clone 6	j
			coig clone 7	k
<i>Leptinaria lamellata</i>	0	4	letl clone 1	l
			letl clone 2	m
			letl clone 3	l
			letl clone 4	n
			letl clone 5	l
			letl clone 6	o

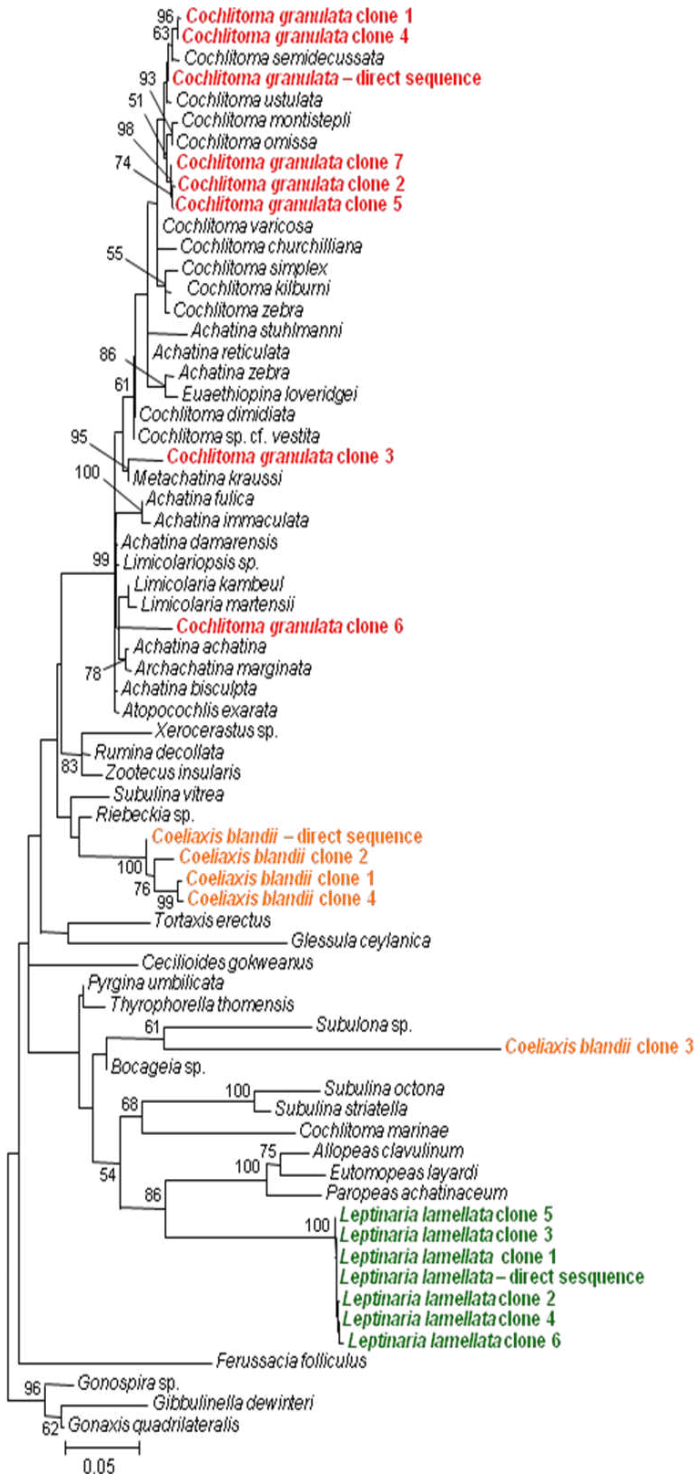
* NOTE: Clones with the same letter have identical actin sequences.

Appendix 3.8B: Variable sites across the actin sequence for the different clones of (1) *Coelioxys blandii*, (2) *Cochlitoma granulata* and (3) *Leptinaria lamellata*. The number shown directly above each site pertains to a variable site. Shaded positions refer to ambiguous sites found in the direct cycle sequences.

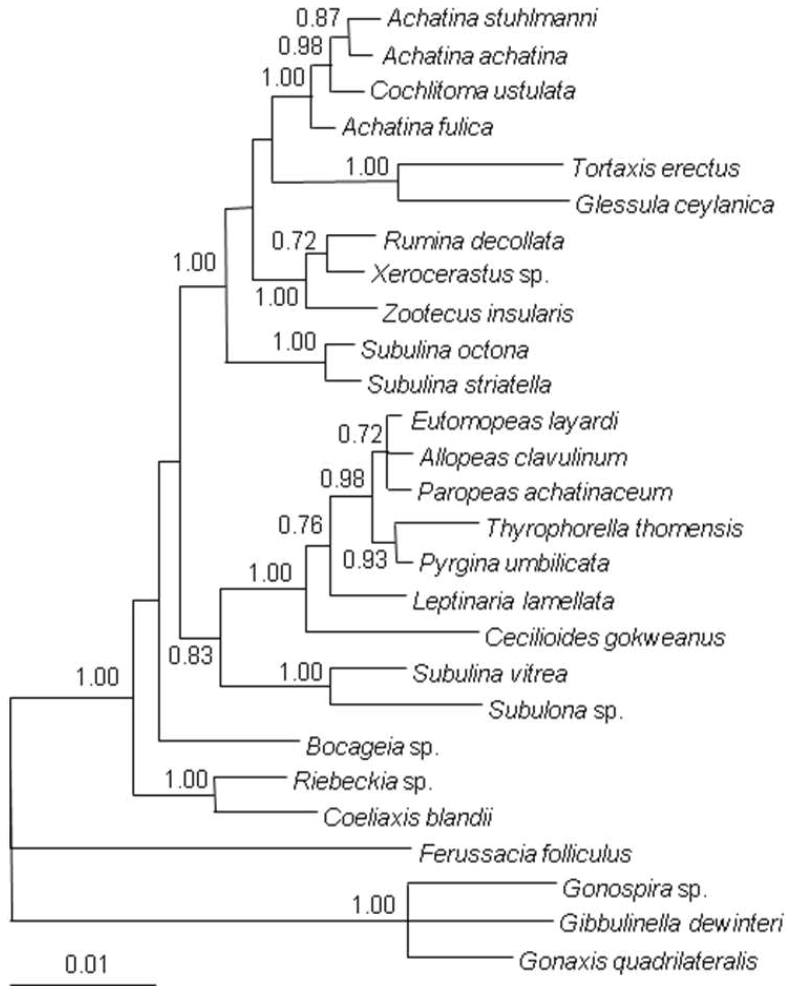


(1) *Coelioxys blandii* (coxb)

NOTE: A total of 163 variable positions were detected from the four clones, with 132 positions alone attributable to Clone 3; these positions were not picked up as ambiguities in the direct sequence.



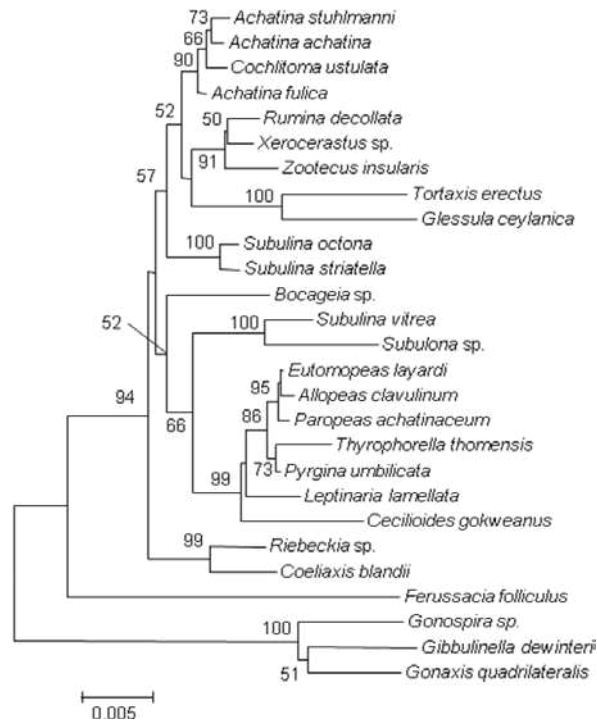
Appendix 3.9: Neighbor-joining phylogenetic tree of the Achatinoidea (including taxa from the Achatinidae) based on the actin and showing the clones of the three representative taxa (*Cochlitoma granulata*, *Coeliaxis blandii* and *Leptinaria lamellata*). The tree was constructed using 861 unambiguously aligned nucleotide sites and rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp. using the GTR+ Γ model of DNA sequence evolution. Bootstrap values lower than 50% are not shown. The scale bar represents 5 substitutional changes per 100 nucleotides.



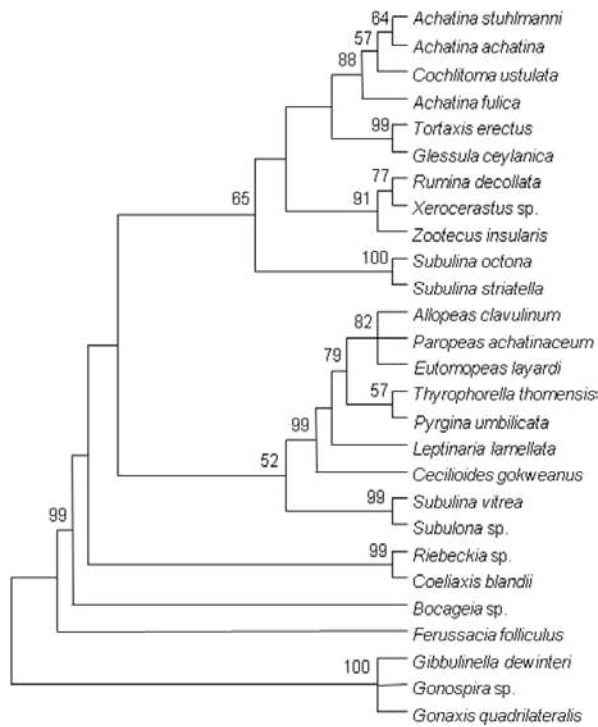
(A) BI (LSU rRNA)

Appendix 3.10: Phylogenetic trees of the Achatinoidea based on the large subunit rRNA gene using (A) Bayesian analysis, (B) neighbor-joining (p. 440) and (C) maximum parsimony based on two equally parsimonious trees (p. 440). The phylogenies were constructed from 3435 unambiguously aligned nucleotide sites and were rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira sp.* Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 100 nucleotides and that of (B) represents 5 substitutional changes per 1000 nucleotides.

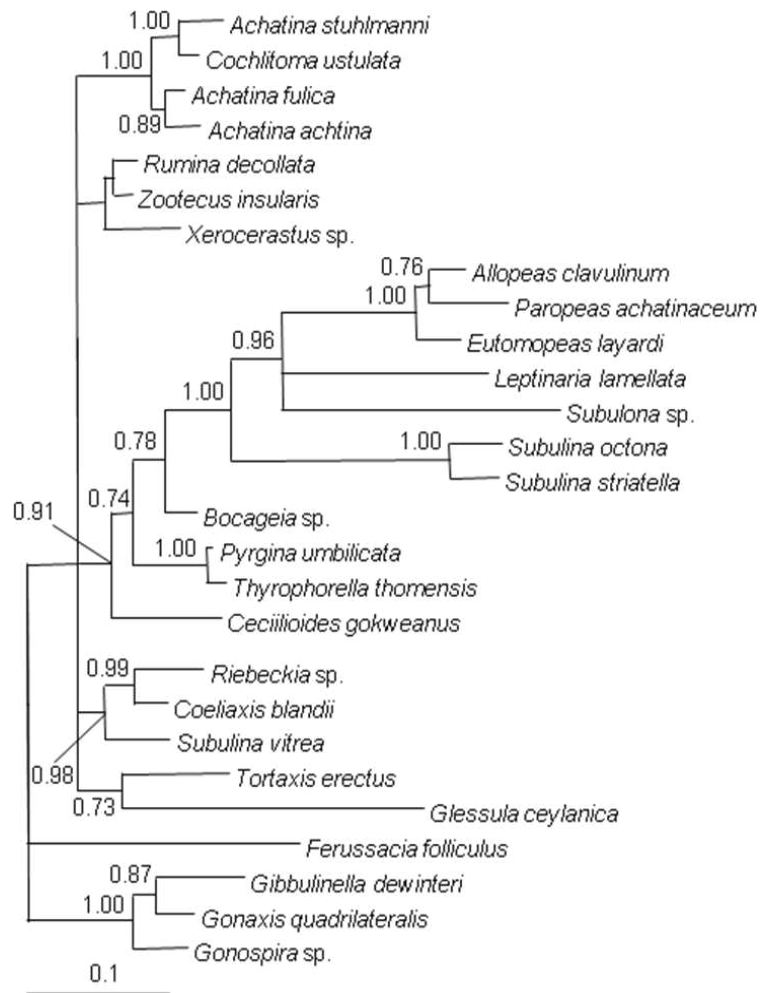
Appendix 3.10 (contd.)



(B) NJ (LSU rRNA)



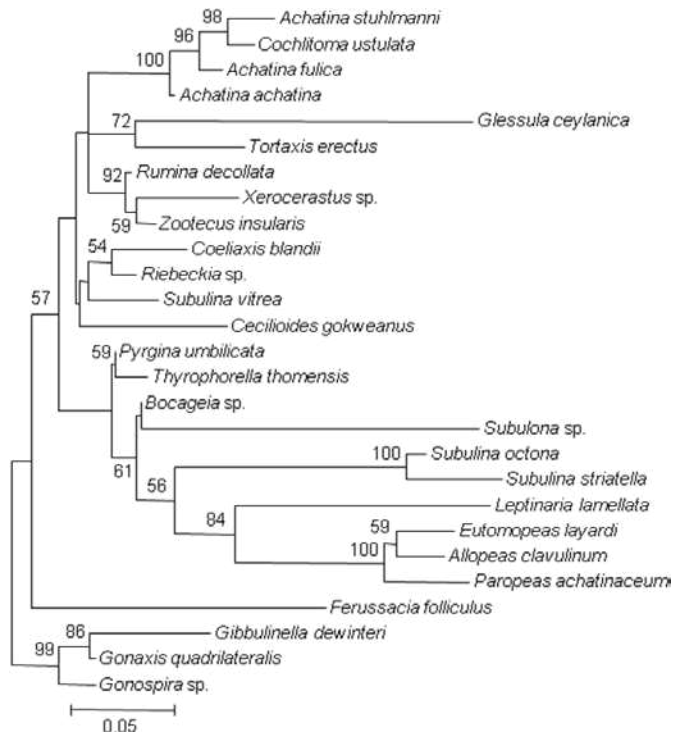
(C) MP (LSU rRNA)



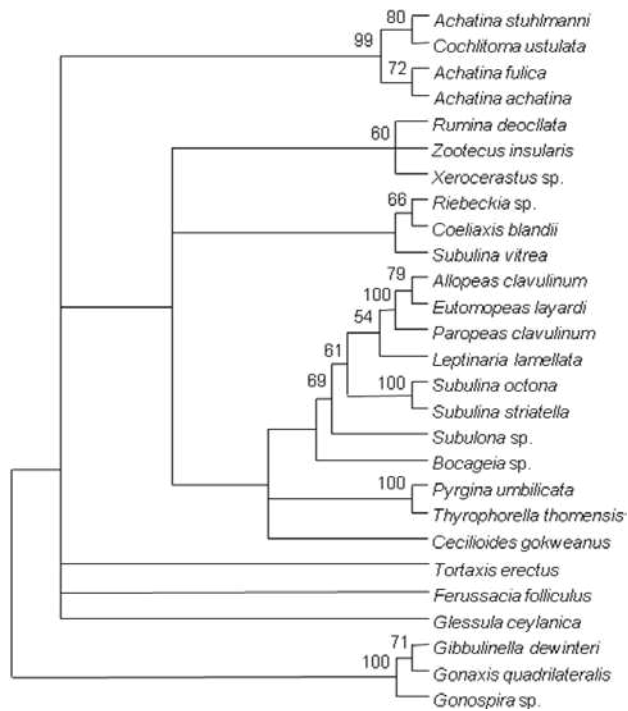
(A) BI (actin)

Appendix 3.11: Phylogenetic trees of the Achatinoidea based on the actin gene using (A) Bayesian analysis, (B) neighbor-joining (p. 442) and (C) maximum parsimony based on seven equally parsimonious trees (p. 442). The phylogenies were constructed from 861 unambiguously aligned nucleotide sites and were rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 10 nucleotides and that of (B) represents 5 substitutional changes per 100 nucleotides.

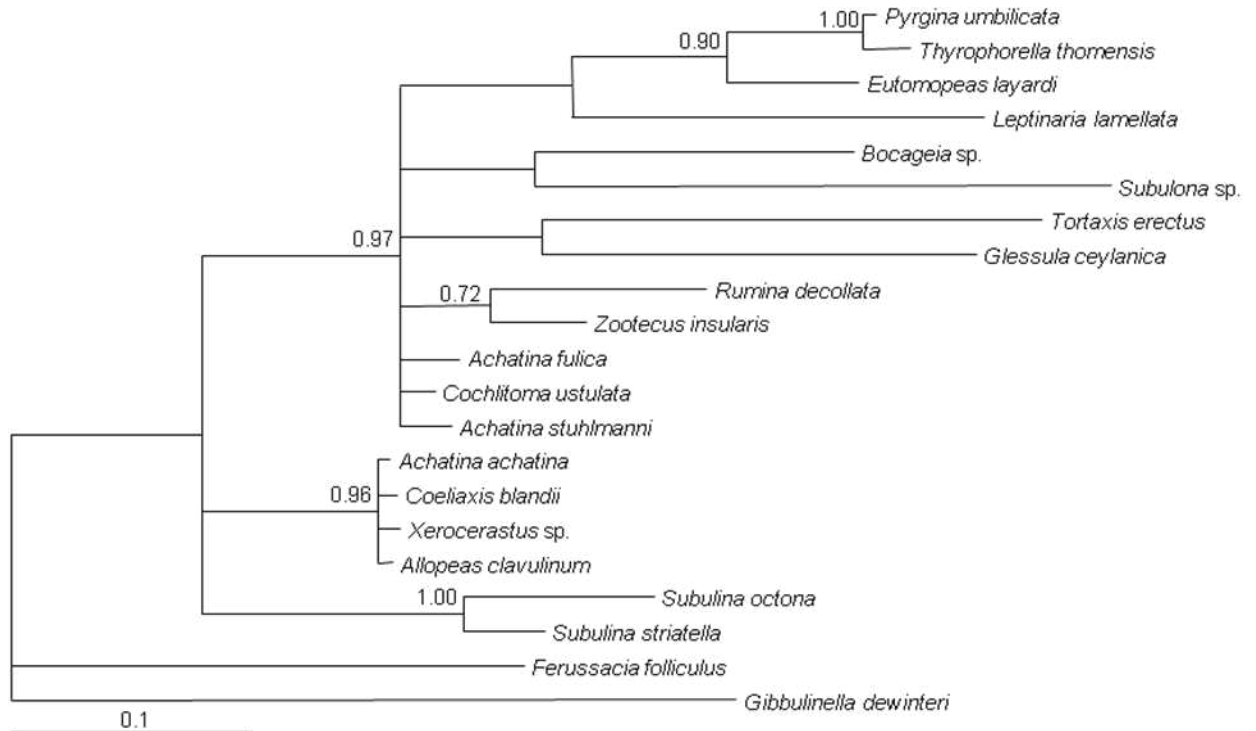
Appendix 3.11 (contd.)



(B) NJ (actin)



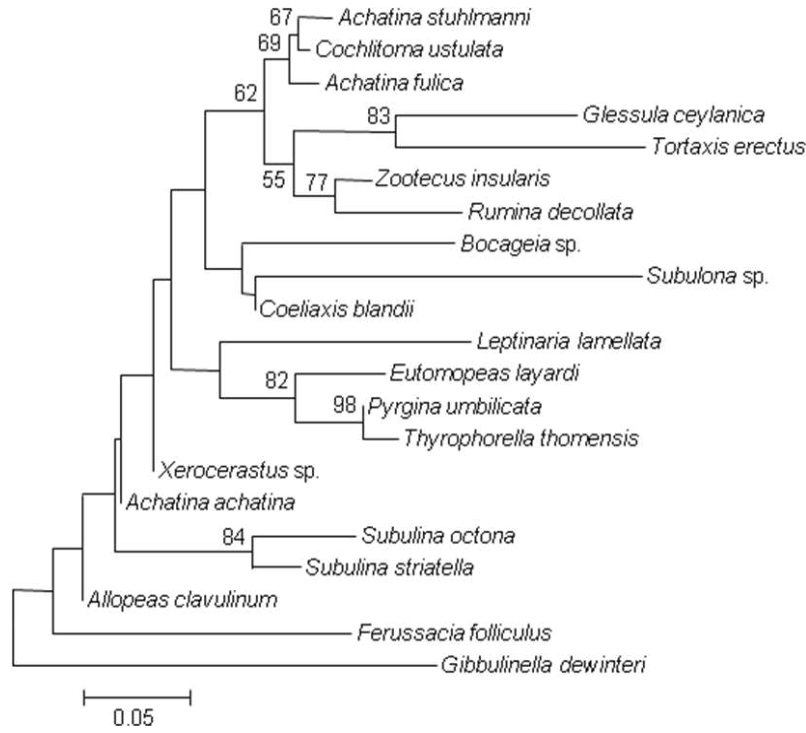
(C) MP (actin)



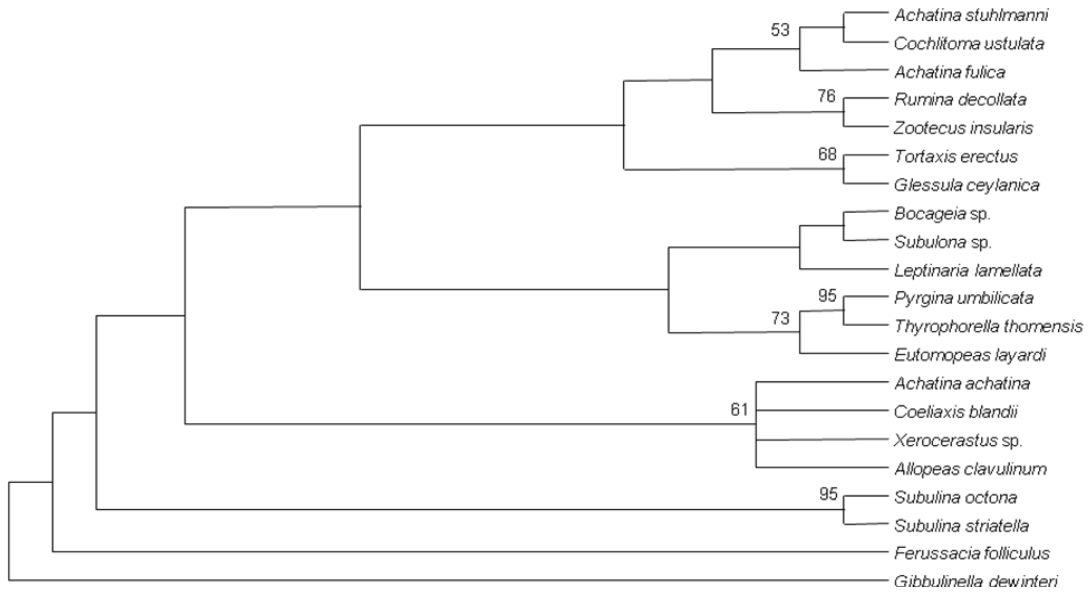
(A) BI (histone 3)

Appendix 3.12: Phylogenetic trees of the Achatinoidea based on the histone 3 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 444) and (C) maximum parsimony based on five equally parsimonious trees (p. 444). The phylogenies were constructed from 328 unambiguously aligned nucleotide sites and were rooted on the streptaxid *Gibbulinella dewinteri*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 10 nucleotides and that of (B) represents 5 substitutional changes per 100 nucleotides. Note that the achatinoids *Cecilioides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp. were not included in the phylogenies as they were not successfully sequenced.

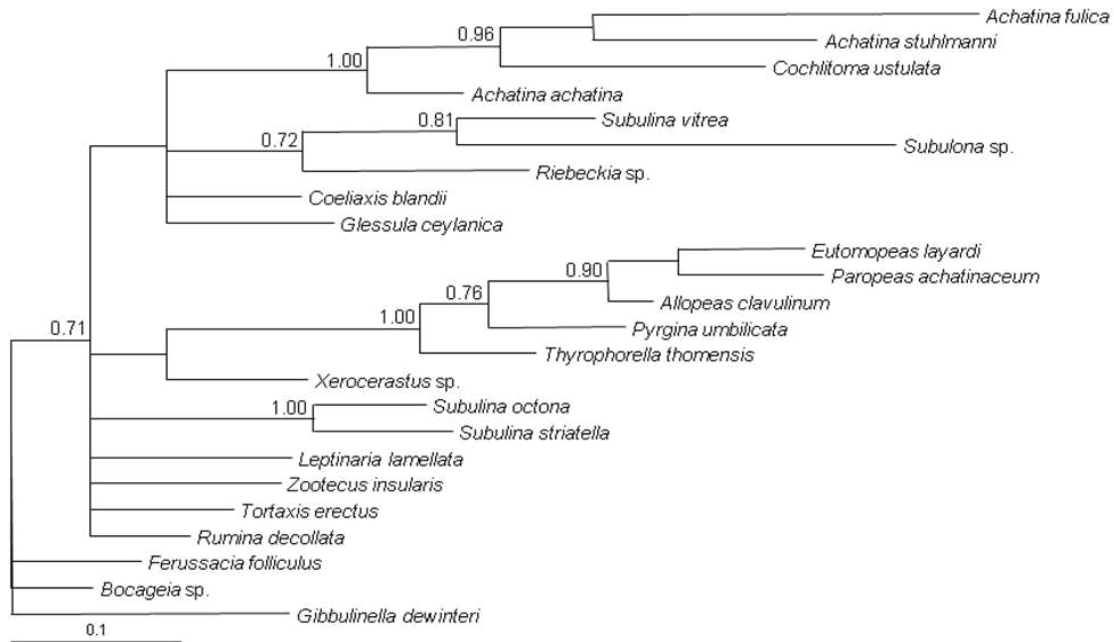
Appendix 3.12 (contd.)



(B) NJ (histone 3)



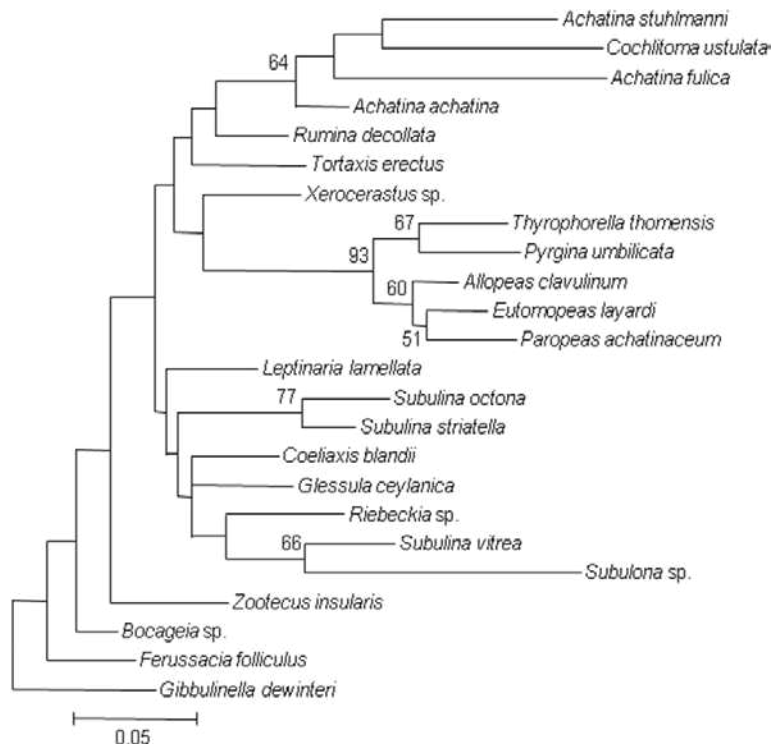
(C) MP (histone 3)



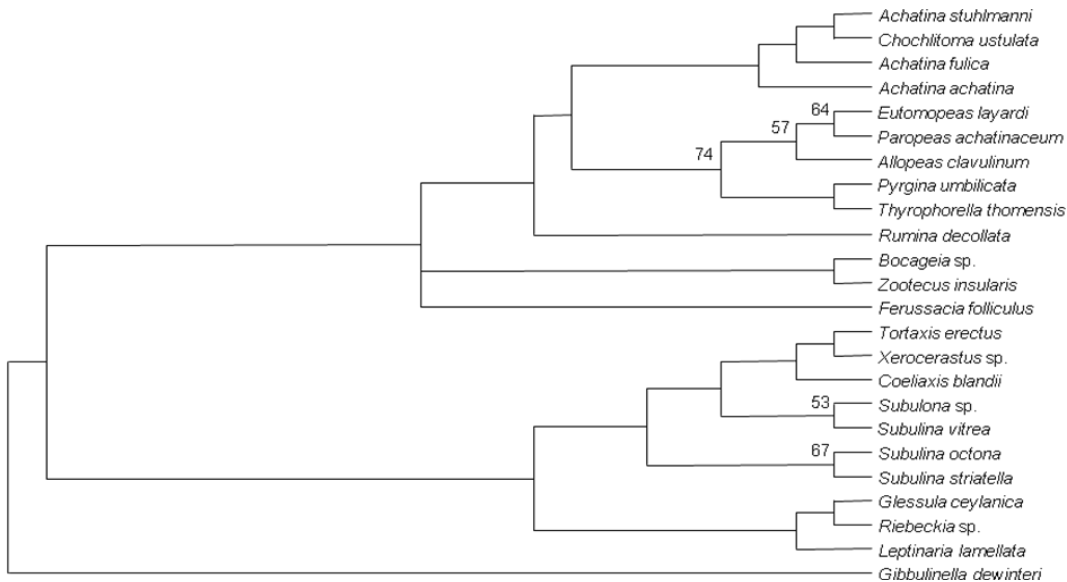
(A) BI (1st and 2nd codon positions of CO1)

Appendix 3.13: Phylogenetic trees of the Achatinoidea based on the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 446) and (C) maximum parsimony based on two equally parsimonious trees (p. 446). The phylogenies were constructed from 404 unambiguously aligned nucleotide sites and were rooted on the streptaxids *Gibbulinella dewinteri*, *Gonaxis quadrilateralis* and *Gonospira* sp. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. . The scale bar for (A) represents 1 substitutional change per 10 nucleotides and that of (B) represents 5 substitutional changes per 100 nucleotides. Note that *Ceceilioides gokweanus* and the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp. were excluded in the analyses as they were not successfully sequenced.

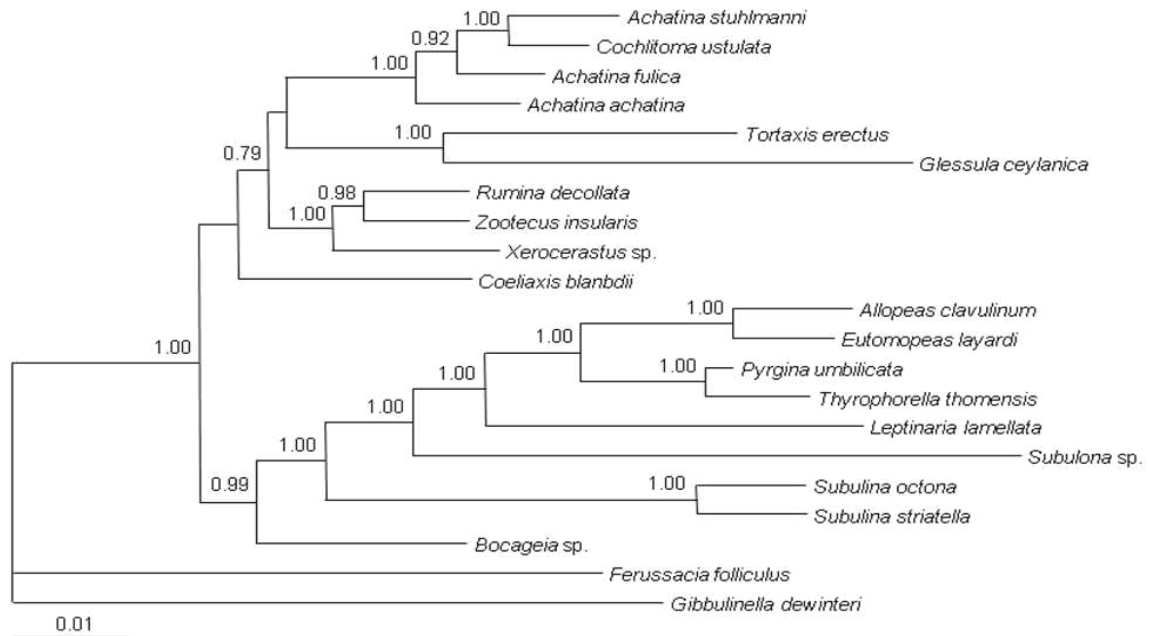
Appendix 3.13 (contd.)



(B) NJ (1st and 2nd codon positions of the CO1 gene)



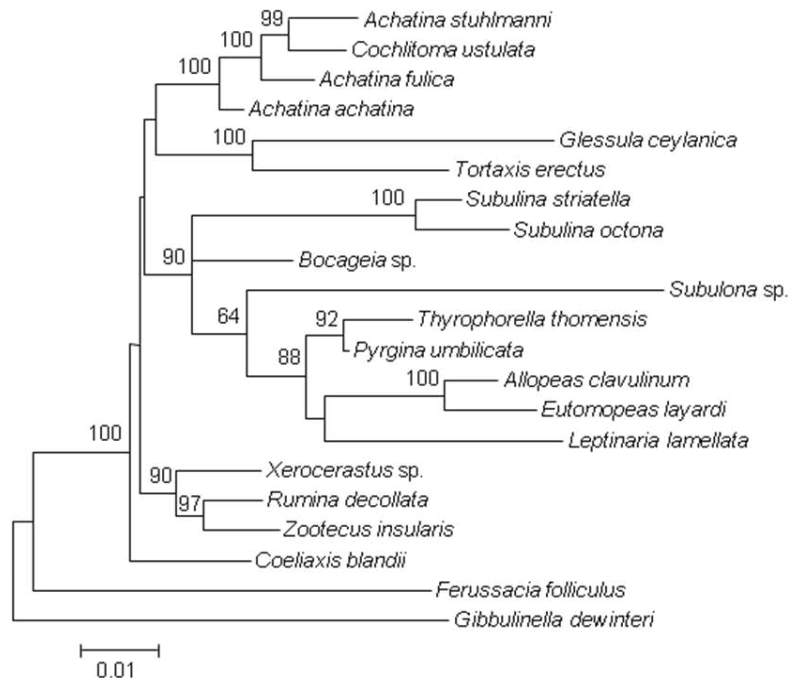
(C) MP (1st and 2nd codon positions of the CO1 gene)



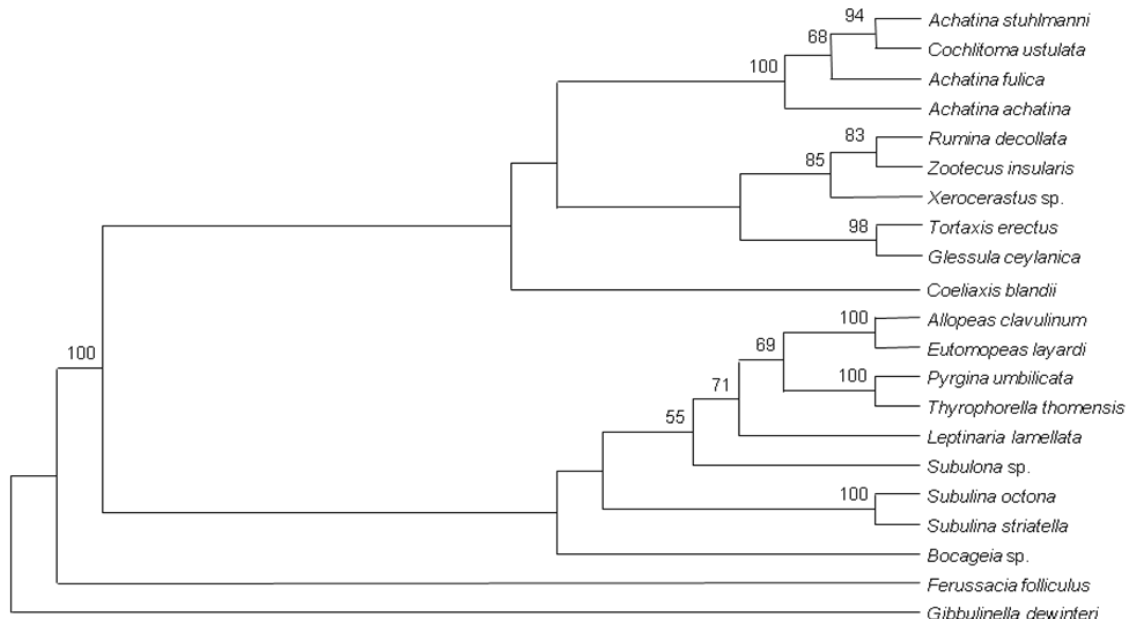
(A) BI (concatenated sequence – taxa with complete datasets only)

Appendix 3.14: Phylogenetic trees of the Achatinoidea (taxa with complete datasets only) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 448) and (C) maximum parsimony based on one most parsimonious tree (p. 448). The phylogenies were constructed from a concatenated sequence of 5028 nucleotides and were rooted on the streptaxid *Gibbulinella dewinteri*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bars for (A) and (B) represent 1 substitutional change per 100 nucleotides. Note that the achatinoid *Cecilioides gokweanus*, *Paropeas achatinaceum*, *Riebeckia* sp. and *Subulina vitrea* and the streptaxids *Gonaxis quadrilateralis* and *Gonospira* sp. were not included in the phylogenies as they lack at least one of the genes used for the combined analyses.

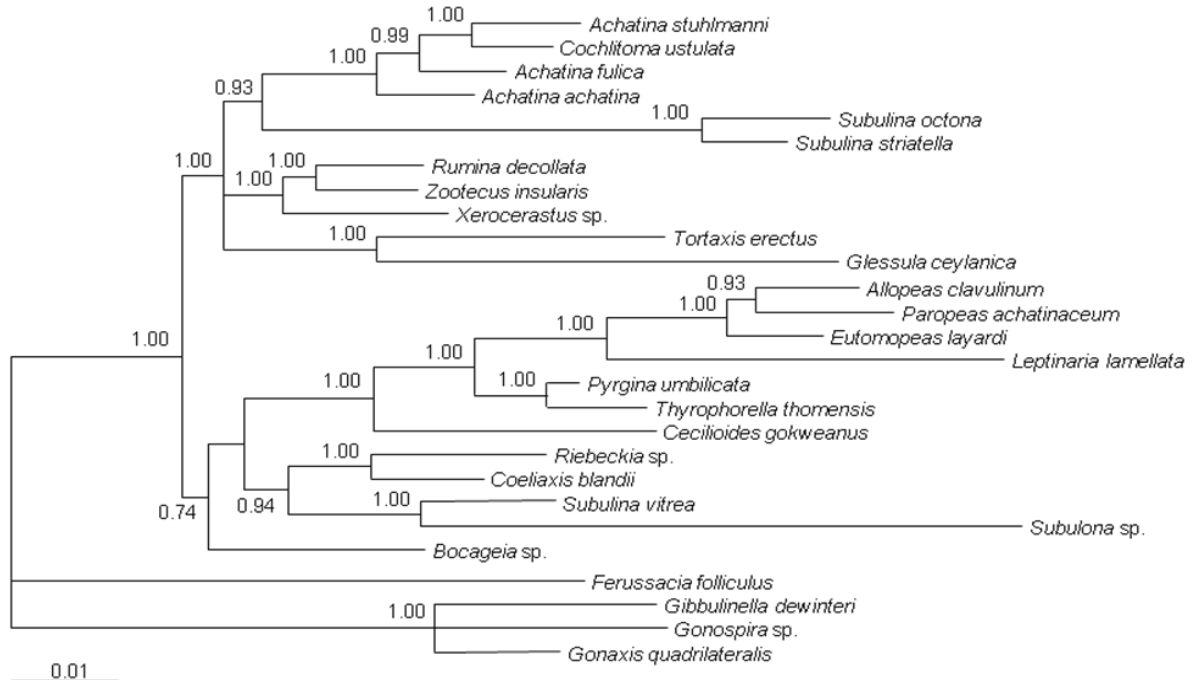
Appendix 3.14 (contd.)



(B) NJ (concatenated sequence – taxa with complete datasets only)



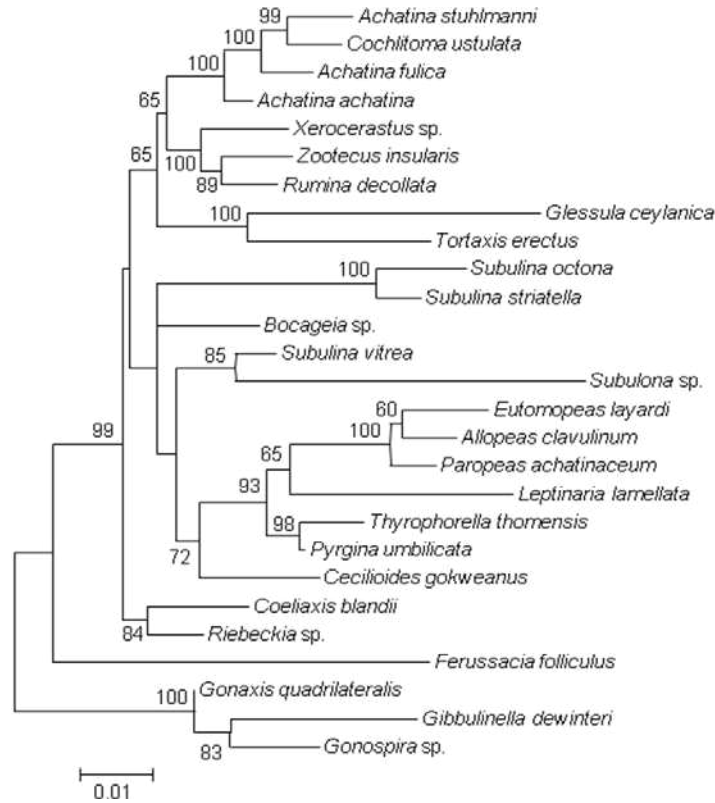
(C) MP (concatenated sequence – taxa with complete datasets only)



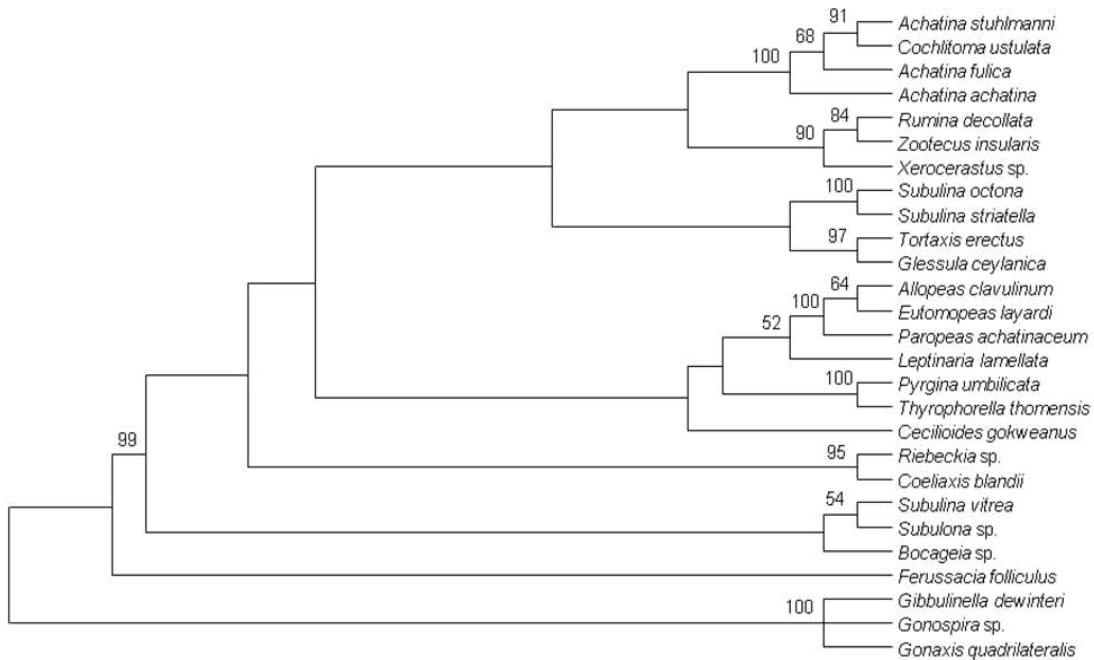
(A) BI (concatenated sequence – all taxa)

Appendix 3.15: Phylogenetic trees of the Achatinoidea (all taxa) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 450) and (C) maximum parsimony based on two equally parsimonious trees (p. 450). The phylogenies were constructed from a concatenated sequence of 5028 nucleotides and were rooted on the streptaxid *Gibbulinella dewinteri*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bars for (A) and (B) represent 1 substitutional change per 100 nucleotides.

Appendix 3.15 (contd.)



(B) NJ (concatenated sequence – all taxa)



(C) MP (concatenated sequence – all taxa)

	130	140	150	160	170	180
<i>Achatina fulica</i>	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCGC	TC---AATTT	ATTCCGAGAT	CCGTCCAGCT
<i>Achatina immaculata</i>	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCGC	TC---AATTT	ATTCCAGAT	CCGTCCAGCT
<i>Achatina reticulata</i>	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCAC	TC---AATTT	CTTCCGAGAT	CCGTCCAGCT
<i>Achatina zanzibarica</i>	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCGC	TC---AATTT	ATTCCGAGAT	CCGTCCAGCT
<i>Euaethiopina loveridgei</i>	TTCGCTTCGT	ATGAGG-CAG	CAGGTCTCGC	TC---AATTT	ATTCCGAGAT	CCGTCCAGCT
<i>Achatina achatina</i>	TTCGCTTCGC	ATGTGG-CAG	CAGGTCTTG-	-----ATTT	ATTCCGAGAT	CCGTCTAAAA
<i>Archachatina marginata</i>	TTCGCTTCGC	ATGTGGGCAG	CAGGTCTCG-	-----ATTT	ATTCCGAGAT	CCGTCTA-TA
<i>Achatina bisculpta</i>	TTCGCTTCGT	ATGCGG-CAG	CAGGTCTCTC	TC--AAATTT	CCGTCCAGAT	CTTCCAC---
<i>Achatina damarensis</i>	TTCGCTTCGT	ATGCGG-CAG	CAGGTCTCTC	TC--AAATTT	CCGTCCAGAT	CTTCC---
<i>Achatina stuhlmanni</i>	TTCGCTTCGT	ATGCGG-CAG	CAGGTCTCGC	TT--AAATTT	CCGTCCAGAT	CTTC-----
<i>Atopocochlis exarata</i>	TTCGCTTCGT	ATGCGG-CAG	CAGGTCTCGC	TCC--AGCTT	CCTTCCAGCT	CTTAATTC--
<i>Limicolaria kambeul</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AAATTT	CCGTCCAGCT	CTTCTTCCCT
<i>Limicolaria martenssii</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AAATTT	CCGTCCAGCT	CTTCTTCCCT
<i>Limicolariopsis sp.</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCAC	TC--AAATTT	CCGTCCAGCT	CTTCTTCCCT
<i>Limicolariopsis ruwenzori</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AAATTT	CCGTCCAGCT	CTTCTTCCCT
<i>Cochlitoma varicosa</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCCGAGAT	CCGTCTAGCT
<i>Cochlitoma dimidiata</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCCGAGAT	CCGTCTAGCT
<i>Cochlitoma sp. cf. vestita</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCCAATAT	CCGTCTAGCT
<i>Cochlitoma marinae</i>	CTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AGTTT	ATTCCAAGAT	CCGTCCAGCT
<i>Cochlitoma churchilliana</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TCCAAGATTT	ATTCCAAGAT	CCGTCCAGCT
<i>Cochlitoma granulata</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGGCTCGC	TCC---ATTT	ATTCCAGAA	CCGGCTAGCT
<i>Cochlitoma simplex</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCC---	--GTCTAGCT
<i>Cochlitoma kilburni</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCCAGAT	CCGTCTAGCT
<i>Cochlitoma montistempli</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCCAAGAT	CCGTCTAGCT
<i>Cochlitoma omissa</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCCAAGAT	CCGTCTAGCT
<i>Cochlitoma semidecussata</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCCAAGAT	CCGTCTAGCT
<i>Cochlitoma ustulata</i>	CTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTT	ATTCCAAGAT	CCGTCTAGCT
<i>Cochlitoma zebra</i>	TTCGCTTCGT	ATGTGGGCAG	CAGGTCTCGC	TC--AATTT	ATTCCAAGAT	CCGTCTAGCT
<i>Metachatina kraussi</i>	TTCGCTTCGT	ATGTGG-CAG	CAGGTCTCGC	TC--AATTTT	ATTCCAAGAT	CCGTCTAGCT
<i>Rumina decollata</i>	TTCGCTTCGC	TTCTGG-CAG	CTGGTCCGGT	TC-----	-----	-----
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmm-mmm	mmmmmmmmmm-	-----	-----	-----

	190	200	210	220	230	240
<i>Achatina fulica</i>	CTTC-----C	TCTATCC-AT	CCGCGG-CTC	GTGCGGA---	-----GGGG	TTACAGAGAG
<i>Achatina immaculata</i>	CTTCTCTTCC	TCTATCC-AT	CCGCGG-CTC	GTGCGGG---	-----GGGG	G-ACAGAAAAG
<i>Achatina reticulata</i>	CTTC--AACC	TCTATCC-AT	CCGCGG-CTC	GTGCGGG---	-----GGG-	TTACAGAGAG
<i>Achatina zanzibarica</i>	CTTC-----C	TCTATCC-AC	CCGCGG-CTC	GTGCGGG---	-----GGGG	TTACAGAGAT
<i>Euaethiopina loveridgei</i>	CTTC-----C	TCTATCC-AT	CCGCGG-CTC	GCGCGGA---	-----GGGG	TTACAGAGAG
<i>Achatina achatina</i>	CTTC---TTG	CTCTGCCGYN	CCGCTG-CTC	GTGCTGG---	-----GG	ACACCGAAAA
<i>Archachatina marginata</i>	CTTCNTGNTC	TANTGNGCTC	CCGCTG-CTC	GTGCTGG---	-----G	ACACGGAAAA
<i>Achatina bisculpta</i>	-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAAG
<i>Achatina damarensis</i>	-----C	TAAATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAAG
<i>Achatina stuhlmanni</i>	-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAAG
<i>Atopocochlis exarata</i>	-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAAG
<i>Limicolaria kambeul</i>	G-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAAG
<i>Limicolaria martenssii</i>	G-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACWAAAAG
<i>Limicolariopsis sp.</i>	-----T	CGAATCT--T	CCGCGG-CTC	GTGCGG---	-----G	GGACAGAAAAG
<i>Limicolariopsis ruwenzori</i>	G-----C	TCTATCT--T	CCGCGG-CTC	GTGCGG---	-----	GGACAGAAAAG
<i>Cochlitoma varicosa</i>	CTTC-----C	TCTATCTTAT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAAG
<i>Cochlitoma dimidiata</i>	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAAG
<i>Cochlitoma sp. cf. vestita</i>	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAAG
<i>Cochlitoma marinae</i>	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGAAAGGAAG
<i>Cochlitoma churchilliana</i>	CTTC-----	-----TTCCT	CCGCGG-CTC	GTGCGGT---	-----GG	GGACAGAAAAG
<i>Cochlitoma granulata</i>	CTTC-----C	TCTATCT-AT	CCGCGGGCTC	GGGGGGG---	-----GGA	GGACAGAAAAG
<i>Cochlitoma simplex</i>	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAAG
<i>Cochlitoma kilburni</i>	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAAG
<i>Cochlitoma montistempli</i>	CTTC-----C	TCYATCT-AT	CCGCGGGCTC	GTGCGGTGGA	GAAAGGAAGA	AGACAGAAAAG
<i>Cochlitoma omissa</i>	CTTC-----C	TCTATCT-AT	CCGCGGGCTC	GTGCGGT---	-----GGA	GGACAGAAAAG
<i>Cochlitoma semidecussata</i>	CTTC-----C	TCTATCT-AA	CCGCGGGCTC	GTGCGGT---	-----GGA	GGACAGAGAG
<i>Cochlitoma ustulata</i>	CTTC-----C	TCTATCT-AT	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAAG
<i>Cochlitoma zebra</i>	CTTC-----C	-----T	CCGCGG-CTC	GTGCGGT---	-----GGA	GGACAGAAAAG
<i>Metachatina kraussi</i>	CTTCCCT--C	TCTATCTTAT	CTGCGG-CTC	GTGCGGT---	----TT----	GGACAGAAAAG
<i>Rumina decollata</i>	-TACCACGTC	GTTTTTCACT	CCGTG--CTC	GTGCGA---	-----	---GGGAGAAAG
NUCLEOTIDES INCLUDED	-----	-----	-----mmm	mmmm-----	-----	-----

	250	260	270	280	290	300
Achatina fulica	GAAGAAT-CG	GG-CG----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT
Achatina immaculata	GAAGAAT-CG	GG-CG----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT
Achatina reticulata	GAAGAAT-CG	GG-CG----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT
Achatina zanzibarica	GAAGAAT-CG	GG-CG----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT
Euaethiopina loveridgei	GAAGAAT-CG	GG-CG----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT
Achatina achatina	GAAGAAT-TG	GGCG----	-----GATA	AGGAAGGGGA	AGAGAGACCG	TCGAAGCGCT
Archachatina marginata	GAAGAAT-TG	GA-CNN----	-----ATA	NNGAAGGG-A	AGAGAGGCCG	TCGAAGCGCT
Achatina bisculpta	AAAGAAT-CT	GG-CG----	-----GAGAA	GC GGAGACCG	TCGAAGCGCT	
Achatina damarensis	AAAGAAT-CT	GG-CG----	-----GAGAA	GC GGAGACCG	TCGAAGCGCT	
Achatina stuhlmanni	AAAGAAT-CT	GT-CG----	-----GAGAA	AAAGAGACCG	TCGAAGCGCT	
Atopocochlis exarata	GAAGAAT-CT	GG-CG----	-----GAGA	AGAGAGACCG	TCGAAGCGCT	
Limicolaria kambeul	GAAGAAT-CT	GG-CG----	-----GA	GAAGAGACCG	TCGAAGCGCT	
Limicolaria martenssii	GAAGAAT-CT	GG-CG----	-----GA	GAAGAGACCG	TCGAAGCGCT	
Limicolariopsis sp.	GAAGAAT-CT	GG-CG----	-----GA	GAAGAGACCG	TCGAAGCGCT	
Limicolariopsis ruwenzori	GAAGAAT-CT	GG-CG----	-----GA	GAAGAGGCCG	TCGAAGCGCT	
Cochlitoma varicosa	GCGGAGAA--	-----	-----GGAA	GAAGAGACCG	TCGAAGCGCT	
Cochlitoma dimidiata	GCGGAGAA--	-----GGA	AGAAGAAGAA	GAAGNGNCCG	TCGAAGCGCT	
Cochlitoma sp. cf. vestita	GCGGAGAA--	-----	-----GGAAGAA	GAAGAGACCG	TCGAAGCGCT	
Cochlitoma marinae	GCG-----	-----G	AGAAGGAAGA	AGAAGAAGAA	GAAGAGACCG	TCGAAGCGCT
Cochlitoma churchilliana	GAAGAA-TCG	GGCG-----	--GAGAAGGG	ACTAAGAAAG	GAAGAGACCG	TCGAAGCGCT
Cochlitoma granulata	GCG-----	-----	-----GA	NAAGAAGAA	GAAGANACCG	TCGAAGCGCT
Cochlitoma simplex	GCG-----	-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT
Cochlitoma kilburni	GCG-----	-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT
Cochlitoma montistempli	GCG-----	-----	-----GAGAA	GGAAGAAGAA	GAAGAGACCG	TCGAAGCGCT
Cochlitoma omissa	GCG-----	-----	-----GAGAA	GGAAGAAGAA	GAAGAGACCG	TCGAAGCGCT
Cochlitoma semidecussata	GCG-----	-----G	AGAAGGAAGA	AGAAGAAGAA	GAAGAGACCG	CCGAAGCGCT
Cochlitoma ustulata	GCG-----	-----	-----GCGAA	GGAAGAAGAA	GAAGAGACCG	TCGAAGCGCT
Cochlitoma zebra	GCG-----	-----	-----GA	GAAGGAAGAA	GAAGAGACCG	TCGAAGCGCT
Metachatina kraussi	GAAGAA-TCG	GGCG--GAGA	AGGAAGAAGA	AGAAGAAGAA	GCAGAGACCG	TCGAAGCGCT
Rumina decollata	CTGCGTCGTG	GATGA----	-----	-----	-TGAAGACCG	TCGAAGCGCA
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	----mnnnnnn	nnnnnnnnnnnn

	310	320	330	340	350	360
Achatina fulica	GTGGGCCGTC	GTGGGAGGAG	GAAGAA----	GGGTTTT--	-----T	TTCCACCTTT
Achatina immaculata	GTGGGCCGTC	GTGGGAGGAG	GAGGAAGA-A	GGGTTTTACC	CCCCTTTTTT	--ACACTCTT
Achatina reticulata	GTGGGCCGTC	GTGGGCCGAG	GAGGAAGA-A	GGGTCTCACC	CCCCTTTTTT	CCCCTACTTT
Achatina zanzibarica	GTGGGCCGTC	GTGGGAGGAG	GAAGAA----	GGGTTTCACC	CCCCTTTTTT	CC-ACTCTT
Euaethiopina loveridgei	GTGGGCCGTC	GTGGGAGGAG	GAAGAA----	GGGTTTCACC	CCCCTTTTTT	TTCCACTCTT
Achatina achatina	CTGGGCCGTC	GTGGGAGAGA	GG-AGGGA--	-----	--TTGTTTTT	TTTTTTTCTT
Archachatina marginata	GTGGGCCGTC	GTGGGAGGAG	AGGAGAGG--	-----GATT	TTGGATTTTT	TTTTTTTCTT
Achatina bisculpta	GTGGGCCGTC	GTGGGATGAG	GAAGAACT--	GGGTTTTTCC	CAG---TTTT	TT-CCCTCTT
Achatina damarensis	GTGGGCCGTC	GTGGGATGAG	GGTGAAGA--	-----ACC	CAG---TTTT	TTTTCTCTCT
Achatina stuhlmanni	GTGGGCCGTC	GTGGGATGAG	GAATAACT--	GGGTTTTTCC	CAG---TT	TTTTCTCTCT
Atopocochlis exarata	GTGGGCCGTC	GCGGGATGAG	GAAGAACT--	GGGTTTTCCC	CAG---TTT	TTTTTCTCTT
Limicolaria kambeul	GTGGGCCGTC	GTGGGATGAG	GAAGAACT--	GGGT-----	-----T	TTCCCTCTTT
Limicolaria martenssii	GTGGGCCGTC	GTGGGATGAG	GAAGAACT--	GGGT-----	-----	TTTCCCTCTT
Limicolariopsis sp.	GTGGGCCGTC	GTGGGATGAG	GAATTACT--	GGGTTT-ACC	CAG-----TT	TTTTTCTATT
Limicolariopsis ruwenzori	GTGGGCCGTC	GTGGGATGAG	GAAGAACT--	GGCT-----	-----TT	TTCCCTCTTT
Cochlitoma varicosa	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	-GCTTTTTTCT	TTT-----	--CCTCTCTT
Cochlitoma dimidiata	GTGGGCCGTC	GTGGGAGGAG	GAGGAAAA--	AGTTTTT--	-----	--CCTCTCTT
Cochlitoma sp. cf. vestita	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	-GCTTTTTTTT	TT-----	--CCTCTCTT
Cochlitoma marinae	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	-GCTTTTTTTT	T-----	--TCTCTCTT
Cochlitoma churchilliana	GTGGGCCGTC	GTGGGAGGGA	GGAAGAAG--	TGCTTTTTTT	CCC-----	--CCTGTCTT
Cochlitoma granulata	GTGGGCCGTC	GNGGGANNAC	GAAGAAGT--	-GNTTTTTTT	TCT-----	--NCTCTCTT
Cochlitoma simplex	GTGGGCCGTC	GTGGGAGGAG	GAGGAAGAAG	TGCTCTTATG	TTTTT----	CCCCCTCTTT
Cochlitoma kilburni	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	GCTTTTTTTT	TCTTT-----	--CCTCTCTT
Cochlitoma montistempli	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	GCTTTTTTTT	-----	CTCCTCTCTT
Cochlitoma omissa	GTGGGCCGTC	GTGGGAGGAG	GAAGAATT--	GCTTTTTTTT	-----	CTCCTCTCTT
Cochlitoma semidecussata	GTGGGCCGTC	GTGGGAGGAG	GAAGAAG--T	GCTTTTTTTT	T-----	--CCTCTCTT
Cochlitoma ustulata	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	GCTTTTTTTT	-----	--TCTCTCTT
Cochlitoma zebra	GTGGGCCGTC	GTGGGAGGAG	GAAGAAGT--	GCTTTTTTTT	CC-----	--CCTCTCTT
Metachatina kraussi	GTGGGCCGTC	GTGGGAGGAG	GAAGAGTG--	CTTTTTTTTT	-----	--CCCCTCTT
Rumina decollata	CTGGGCTTTC	G-AGGACGAG	GAGGATCG-A	GTTTTTCTCT-	-----	-TTCCACCTT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	m-mmmmm	-----	-----	-----	-----

	370 380 390 400 410 420
<i>Achatina fulica</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Achatina immaculata</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Achatina reticulata</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Achatina zanzibarica</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Euaethiopina loveridgei</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Achatina achatina</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Archachatina marginata</i>	CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Achatina bisculpta</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Achatina damarensis</i>	-TCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Achatina stuhlmanni</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Atopocochlis exarata</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTC---GCT
<i>Limicolaria kambeul</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Limicolaria martenssii</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Limicolariopsis sp.</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Limicolariopsis ruwenzori</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Cochlitoma varicosa</i>	-CCCCCGTGG CCTCAAGTAA ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Cochlitoma dimidiata</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGNGA CTCTTTTCGCT
<i>Cochlitoma sp. cf. vestita</i>	-CCCCCGTGG TCTCAAGTAT ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCCTTTTCGCT
<i>Cochlitoma marinae</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGAGA CTCTTTTCGCT
<i>Cochlitoma churchilliana</i>	TCCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Cochlitoma granulata</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCNTTTTCGCT
<i>Cochlitoma simplex</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Cochlitoma kilburni</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Cochlitoma montistempli</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Cochlitoma omissa</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Cochlitoma semidecussata</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Cochlitoma ustulata</i>	-TCCCCGGGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCAA CTCCTTTTCGCT
<i>Cochlitoma zebra</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Metachatina kraussi</i>	-CCCCCGTGG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATTTTCAGCGA CTCTTTTCGCT
<i>Rumina decollata</i>	-CCCTCTAG CCTCAAGTAC ACGCATGCGC CGTCCTCGTC ATCTGAGCA- -----TC
NUCLEOTIDES INCLUDED	--mmmmmmmm mmmmmmmmm mm---mmmm mmmmm-mmmmm m-----mmm- -----

	430 440 450 460 470 480
<i>Achatina fulica</i>	---CGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Achatina immaculata</i>	---CGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Achatina reticulata</i>	---CGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Achatina zanzibarica</i>	---CGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Euaethiopina loveridgei</i>	---CGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Achatina achatina</i>	---TGCCCC AGCGCC-CGT CTTCGTTT-G TTCGC----C GTCCGTCCGG CAGGACTCGG
<i>Archachatina marginata</i>	---TGCCCC AGCGCC-CGT CTTCGTTT-G TTCGC----C GTCCGTCCGG CAGGACTCGG
<i>Achatina bisculpta</i>	---CGCCAG AGCGCTCCAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Achatina damarensis</i>	---CGCCCG AGCGCTCCAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Achatina stuhlmanni</i>	---CGCCCG AGCGCTCCAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Atopocochlis exarata</i>	---CGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Limicolaria kambeul</i>	----- -CMT GTTCG---- -CTCGCC GTCCGTCCGG CAGGACTCGG
<i>Limicolaria martenssii</i>	----- -CAT CTTCGTTT-G TTCGTTTCGCC GTCCGTCCGG CAGGACTCGG
<i>Limicolariopsis sp.</i>	---CGCCCG AGCACT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Limicolariopsis ruwenzori</i>	----- -CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCAG CAGGACTCGG
<i>Cochlitoma varicosa</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma dimidiata</i>	GCTCNCNCG ANNGNT-CAT NTNNNTTTG TTCGTTTNC GTCCNTCCGN CAGGACTCGG
<i>Cochlitoma sp. cf. vestita</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma marinae</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma churchilliana</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma granulata</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma simplex</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma kilburni</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma montistempli</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma omissa</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma semidecussata</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Cochlitoma ustulata</i>	GTCGCCCCG AGCGT-CAT CTTCGTTT-G TTCGCTCGCC GKCCGTCCGG CAGGACTCGG
<i>Cochlitoma zebra</i>	GCTCGCCCG AGCGCT-CGT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Metachatina kraussi</i>	GCTCGCCCG AGCGCT-CAT CTTCGTTT-G TTCGCTCGCC GTCCGTCCGG CAGGACTCGG
<i>Rumina decollata</i>	GCTCTCTGA AGCGCT-CAT CTTCGTTT-G CTCGC----C GTCCGTCTGG CAGGACTCGG
NUCLEOTIDES INCLUDED	-----mmmm mmmmm----- -mmmmmmmm mmmmmmmmm

	490 500 510 520 530 540
<i>Achatina fulica</i>	CTCGCTTCTT T----- AACAAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Achatina immaculata</i>	CTCGCTTCTT T-----G AAAAAATCGA GCCTGCCAGA C---CGTCCA AAGCGGCTAG
<i>Achatina reticulata</i>	CTCGCTTCTT T----- AACAAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Achatina zanzibarica</i>	CTCGCTTTTT TTT----- AACAAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Euaethiopina loveridgei</i>	CTCGCTTCTT T----- AACAAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Achatina achatina</i>	CTCGCTT--- ------ --TTATCGA GCCTGCCAGA CGACTGTCCG AAGCGGCTAG
<i>Archachatina marginata</i>	GTCGCTT--- ------ --TTATCGA GCCTGCCAGA C---TGTCGG AAGCGGCTAG
<i>Achatina bisculpta</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Achatina damarensis</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Achatina stuhlmanni</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Atopocochlis exarata</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Limicolaria kambeul</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Limicolaria martenssii</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Limicolariopsis sp.</i>	CTCGCTTCTT TTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Limicolariopsis ruwenzori</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma varicosa</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma dimidiata</i>	CTCGNTTNTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AATCGGCTAG
<i>Cochlitoma sp. cf. vestita</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma marinae</i>	CTCGCTTCTT TTTGTTT--- --AATCGA GCCTGCCAGA CAC-CGTCCG AAGCGGCTAG
<i>Cochlitoma churchilliana</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma granulata</i>	CTCGCTTCTT TTTTNTT--- --ANTCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma simplex</i>	CTCGCTTCTT TTTTNTNNTT TTTTAATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma kilburni</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma montistempli</i>	CTCGCTTCAT TTTTATT--- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma omissa</i>	CTCGCTTCAT TTTTATT--- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma semidecussata</i>	CTCGCTTCTT TTT-ATT--- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma ustulata</i>	CTCGCTTCTT TTTT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Cochlitoma zebra</i>	CTCGCTTCTT TTTTNTT--- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Metachatina kraussi</i>	CTCGCTTCTT TTTAT----- --AATCGA GCCTGCCAGA C---CGTCCG AAGCGGCTAG
<i>Rumina decollata</i>	CTCGCTGTG- ------ AATAAAACGA GCCTGCC- GA C---CGTCCG AAGTGGCATG
NUCLEOTIDES INCLUDED	mmmmmm- --- ------ -m mmmmmmm- mm m---mmmmmm mmmmmmm- -m

	550 560 570 580 590 600
<i>Achatina fulica</i>	GG-ATTG-CG AAGTGGGG-- ---CGCGCAA GCGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Achatina immaculata</i>	GG-ATTG-CG AAGTGGGG-- ---CGCGCAA GCGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Achatina reticulata</i>	GG-ATTG-CG AAGTGGGG-- ---CGCGCAA GCGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Achatina zanzibarica</i>	GG-ATTG-CG AAGTGGGG-- ---CGCGCAA GCGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Euaethiopina loveridgei</i>	GG-ATTG-CG AAGTGGGG-- ---CGCGCAA GCGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Achatina achatina</i>	CG-GTTG-CG AAGCGGGT-- -GGCGCGTAA GCG-ATCGG- ---TCT---G CGGCGGCGCC
<i>Archachatina marginata</i>	CG-GTTG-CG AAGTGGGT-- -GGCGGGCAA GCG-ATCGG- ---TCT---G CGGCGGCGCC
<i>Achatina bisculpta</i>	CG-ATTG-CG AAGTGGAG-- ------ -GGG- ---TCTGCTG CGGCGGCGCC
<i>Achatina damarensis</i>	CG-ATTG-CG AAGTGGGG-C GCGTGAGCGC TGGCATGTTG GGGTCTGCTG CGGCGGCGCC
<i>Achatina stuhlmanni</i>	CG-ATTG-CG AAGTGGGG-- -CGCCCGCA GCGGATTGGG GGGTCTGCTG TGCGGCGCC
<i>Atopocochlis exarata</i>	CG-AATG-CG AAGTGGGG-- -CCCAGGAG CGGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Limicolaria kambeul</i>	CG-ATTGGCG AAGTG- --- ------ -CATGGG- ---TCTGCTG AGGCGGCGCC
<i>Limicolaria martenssii</i>	CG-ATTGGCG AAGTG- --- ------ -CATGGG- ---TCAGTG AGGCGGCGCC
<i>Limicolariopsis sp.</i>	TG-ATTG-CG AAGTGGGT-- ---CGCGGGT GCGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Limicolariopsis ruwenzori</i>	CG-ATTGGCG AAGTG- --- ------ -CATGGG- ---TCTGCTG CGGCGGCGCC
<i>Cochlitoma varicosa</i>	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAGCATGGG- --GTCTGCTG CGGCGGCGCC
<i>Cochlitoma dimidiata</i>	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAGCATGGG- GAGTCTGCTG CGGCGGCGCC
<i>Cochlitoma sp. cf. vestita</i>	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Cochlitoma marinae</i>	CGGATTG-CG AAGTGGGG-- ---CGCGCGA GCGCATGGG- ---TCTGTTG CGGCGGCGCC
<i>Cochlitoma churchilliana</i>	CG-AGTG-CG AAGTGGGG-- -GGCGCGCA GCGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Cochlitoma granulata</i>	CGGATTG-NG AAGNNGGG-- CGCGCGAGCG CAACATGGGG G--TCTGNTG CGGCGGCGCC
<i>Cochlitoma simplex</i>	CGGATTG-CG AAGTGGGGCG CACGCGAGCG CAGCATAGG- ---TCTGTTG CGGCGGCGCC
<i>Cochlitoma kilburni</i>	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAGCATGGG- ---TCTGCTG CGGCGGCGCC
<i>Cochlitoma montistempli</i>	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAACATGGGG GG-TCTGCTG CGGCGGCGCC
<i>Cochlitoma omissa</i>	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAACATGGGG G--TCTGCTG CGGCGGCGCC
<i>Cochlitoma semidecussata</i>	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CAACATGGGG G--TCTGCTG CGGCGGCGCC
<i>Cochlitoma ustulata</i>	CGGATTG-CG AAGTGGGG-- CGCGCGAGCG CA---TGGG- ---TCTGTTG CGGCGGCGCC
<i>Cochlitoma zebra</i>	CGGATTG-CG AAGTGGGT-- CGCGCGAGCG CAGCATGGG- ---TCTGTTG CGGCGGCGCC
<i>Metachatina kraussi</i>	CG-ATTG-CG AAGTGGGG-- CGCGCGAGCG CA---TGGG- ---TCTGCTG CGGCGGCGCC
<i>Rumina decollata</i>	CG-GTTT-CG AAGTGTGA- ------ -GAGG GCGATTGCCG CGGCGGCGCC
NUCLEOTIDES INCLUDED	mm- --- -mm mmmmm- --- ------ -m mmmmmmmmmmm

	730	740	750	760	770	780	
<i>Achatina fulica</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Achatina immaculata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Achatina reticulata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Achatina zanzibarica</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Euaethiopina loveridgei</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Achatina achatina</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Archachatina marginata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Achatina bisculpta</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Achatina damarensis</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Achatina stuhlmanni</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Atopocochlis exarata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Limicolaria kambeul</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Limicolaria martenssii</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Limicolariopsis sp.</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Limicolariopsis ruwenzori</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma varicosa</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma dimidiata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma sp. cf. vestita</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma marinae</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma churchilliana</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma granulata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma simplex</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma kilburni</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma montistempli</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma omissa</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma semidecussata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma ustulata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Cochlitoma zebra</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Metachatina kraussi</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAATAG	CCCAGCACCG	AATCCCTCAG	
<i>Rumina decollata</i>	ACAAGGATTT	CCCCAGTAAC	GGCGAGTGAA	GCGGGAAGAG	CCCAGCACCG	AATCCCTCAG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	790	800	810	820	830	840	
<i>Achatina fulica</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Achatina immaculata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Achatina reticulata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Achatina zanzibarica</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Euaethiopina loveridgei</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Achatina achatina</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Archachatina marginata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Achatina bisculpta</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Achatina damarensis</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Achatina stuhlmanni</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Atopocochlis exarata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Limicolaria kambeul</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Limicolaria martenssii</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Limicolariopsis sp.</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Limicolariopsis ruwenzori</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma varicosa</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma dimidiata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma sp. cf. vestita</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma marinae</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma churchilliana</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma granulata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma simplex</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma kilburni</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma montistempli</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma omissa</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma semidecussata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma ustulata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Cochlitoma zebra</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Metachatina kraussi</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
<i>Rumina decollata</i>	TGTCACGCTG	ACGGGAACTG	TGGTGTGTGG	GACGCCACCA	GTTCGCATCAG	AGGGCGTCTGA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	850	860	870	880	890	900	
<i>Achatina fulica</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Achatina immaculata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Achatina reticulata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Achatina zanzibarica</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Euaethiopina loveridgei</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Achatina achatina</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Archachatina marginata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Achatina bisculpta</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Achatina damarensis</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Achatina stuhlmanni</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Atopocochlis exarata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Limicolaria kambeul</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Limicolaria martenssii</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Limicolariopsis sp.</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Limicolariopsis ruwenzori</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma varicosa</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma dimidiata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma sp. cf. vestita</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma marinae</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma churchilliana</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma granulata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma simplex</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma kilburni</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma montistempli</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma omissa</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma semidecussata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma ustulata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Cochlitoma zebra</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Metachatina kraussi</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
<i>Rumina decollata</i>	AGTCCTCCTG	ATCGGGGCTT	CACCCAGAGC	GGGTGTAAGG	CCTTTGCAGG	CGCCTCTCTG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	910	920	930	940	950	960
<i>Achatina fulica</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Achatina immaculata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Achatina reticulata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Achatina zanzibarica</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Euaethiopina loveridgei</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Achatina achatina</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Archachatina marginata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Achatina bisculpta</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Achatina damarensis</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Achatina stuhlmanni</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Atopocochlis exarata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Limicolaria kambeul</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Limicolaria martenssii</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Limicolariopsis sp.</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Limicolariopsis ruwenzori</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma varicosa</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma dimidiata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma sp. cf. vestita</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma marinae</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma churchilliana</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma granulata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma simplex</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma kilburni</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma montistempli</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma omissa</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma semidecussata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma ustulata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Cochlitoma zebra</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Metachatina kraussi</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
<i>Rumina decollata</i>	TGCGGCCGCG	AGCGTCTCAG	GAGTCGGGTT	GTTTGGGAAT	GCAGCCCAA	GCGGGTGTTA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	970 980 990 1000 1010 1020
<i>Achatina fulica</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Achatina immaculata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Achatina reticulata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Achatina zanzibarica</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Euaethiopina loveridgei</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Achatina achatina</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Archachatina marginata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Achatina bisculpta</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Achatina damarensis</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Achatina stuhlmanni</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Atopocochlis exarata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Limicolaria kambeul</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Limicolaria martenssii</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Limicolariopsis sp.</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Limicolariopsis ruwenzori</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma varicosa</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma dimidiata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma sp. cf. vestita</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma marinae</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma churchilliana</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma granulata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma simplex</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma kilburni</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma montistempli</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma omissa</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma semidecussata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma ustulata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Cochlitoma zebra</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Metachatina kraussi</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
<i>Rumina decollata</i>	AACTCCATCT AAGGCTAAAT ACTGGCACGA GTCCGATAGC GGACAAGTAC CGTGAGGGAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	1030 1040 1050 1060 1070 1080
<i>Achatina fulica</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Achatina immaculata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Achatina reticulata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Achatina zanzibarica</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Euaethiopina loveridgei</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Achatina achatina</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Archachatina marginata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Achatina bisculpta</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Achatina damarensis</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Achatina stuhlmanni</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Atopocochlis exarata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Limicolaria kambeul</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Limicolaria martenssii</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Limicolariopsis sp.</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Limicolariopsis ruwenzori</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma varicosa</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma dimidiata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma sp. cf. vestita</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma marinae</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma churchilliana</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma granulata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma simplex</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma kilburni</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma montistempli</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma omissa</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma semidecussata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma ustulata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Cochlitoma zebra</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Metachatina kraussi</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
<i>Rumina decollata</i>	AGTTGAAAAG AACTTTGAAG AGAGAGTTCA AGAGTACGTG AAACCGCCCA GAGGTAAACG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	1090	1100	1110	1120	1130	1140
Achatina fulica	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Achatina immaculata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Achatina reticulata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Achatina zanzibarica	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Euaethiopina loveridgei	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Achatina achatina	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Archachatina marginata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Achatina bisculpta	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Achatina damarensis	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Achatina stuhlmanni	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Atopocochlis exarata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Limicolaria kambeul	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Limicolaria martenssii	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Limicolariopsis sp.	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Limicolariopsis ruwenzori	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma varicosa	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma dimidiata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma sp. cf. vestita	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma marinae	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma churchilliana	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma granulata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma simplex	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma kilburni	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma montistempli	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma omissa	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma semidecussata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma ustulata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Cochlitoma zebra	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Metachatina kraussi	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
Rumina decollata	GGTGGATCCG	CAAAGTCGGC	CCGCGGAATT	CAGCGCGGCG	CGCGGCCTCG	GGGCGATCGC
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1150	1160	1170	1180	1190	1200
Achatina fulica	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Achatina immaculata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Achatina reticulata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Achatina zanzibarica	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Euaethiopina loveridgei	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Achatina achatina	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Archachatina marginata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Achatina bisculpta	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Achatina damarensis	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Achatina stuhlmanni	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Atopocochlis exarata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Limicolaria kambeul	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Limicolaria martenssii	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Limicolariopsis sp.	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Limicolariopsis ruwenzori	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma varicosa	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma dimidiata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma sp. cf. vestita	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma marinae	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma churchilliana	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma granulata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma simplex	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma kilburni	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma montistempli	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma omissa	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma semidecussata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma ustulata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Cochlitoma zebra	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Metachatina kraussi	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
Rumina decollata	GGCCGGGGAT	CCCTGGGACC	CCGCCGCGGT	GTCGATCCGG	GCTCCGCCGC	GTGCACTTTC
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1210	1220	1230	1240	1250	1260
<i>Achatina fulica</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Achatina immaculata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Achatina reticulata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Achatina zanzibarica</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Euaethiopina loveridgei</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Achatina achatina</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GAGTTGTCCG
<i>Archachatina marginata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GAGTTGTCCG
<i>Achatina bisculpta</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Achatina damarensis</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Achatina stuhlmanni</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Atopocochlis exarata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Limicolaria kambeul</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Limicolaria martenssii</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Limicolariopsis sp.</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Limicolariopsis ruwenzori</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GTGTTGTCCG
<i>Cochlitoma varicosa</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma dimidiata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma sp. cf. vestita</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma marinae</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma churchilliana</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma granulata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma simplex</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma kilburni</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma montistempli</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma omissa</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma semidecussata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma ustulata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Cochlitoma zebra</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Metachatina kraussi</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
<i>Rumina decollata</i>	CGCGGGCAGA	GTGCCACAAC	CGGTTCCGCT	GGAGCCGTCA	GAGGCCGGGC	GGGTTGTCCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1270	1280	1290	1300	1310	1320
<i>Achatina fulica</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Achatina immaculata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Achatina reticulata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Achatina zanzibarica</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Euaethiopina loveridgei</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Achatina achatina</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGGACCG
<i>Archachatina marginata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGGACCG
<i>Achatina bisculpta</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Achatina damarensis</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Achatina stuhlmanni</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Atopocochlis exarata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Limicolaria kambeul</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Limicolaria martenssii</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Limicolariopsis sp.</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Limicolariopsis ruwenzori</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma varicosa</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma dimidiata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma sp. cf. vestita</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma marinae</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma churchilliana</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma granulata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma simplex</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma kilburni</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma montistempli</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma omissa</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma semidecussata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma ustulata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Cochlitoma zebra</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Metachatina kraussi</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGAACCG
<i>Rumina decollata</i>	TGGGGGCGCT	TGCGTCCCTA	CCAGCCCGCC	CCGGCGGACG	GCCTCGGGAC	CGAGGGACCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1330	1340	1350	1360	1370	1380	
Achatina fulica	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina immaculata	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina reticulata	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina zanzibarica	CCGCGCGCTT	CGAGGCTACC	CGGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Euaethiopina loveridgei	CCGCGCGCTT	CGAGGCTACC	CAGCCCTCT	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina achatina	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Archachatina marginata	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina bisculpta	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina damarensis	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Achatina stuhlmanni	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Atopocochlis exarata	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Limicolaria kambeul	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Limicolaria martenssii	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Limicolariopsis sp.	CCGCGCGCTT	CGAGGCTACC	CGATCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Limicolariopsis ruwenzori	CCGCGCGCTT	CGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma varicosa	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma dimidiata	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma sp. cf. vestita	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma marinae	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma churchilliana	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma granulata	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma simplex	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma kilburni	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma montistempli	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma omisa	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma semidecussata	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma ustulata	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GTGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Cochlitoma zebra	CCGCGCGCTT	GGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Metachatina kraussi	CCGCGCGCTT	AGAGGCTACC	CGACCCCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
Rumina decollata	CCGCACGCTT	CGAGGCTACC	T-GCTCTCC	GCGCGAGTCC	GACTGGGAGA	GACTGGGCAA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	m-mmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1390	1400	1410	1420	1430	1440	
Achatina fulica	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina immaculata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina reticulata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina zanzibarica	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Euaethiopina loveridgei	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina achatina	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Archachatina marginata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina bisculpta	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina damarensis	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Achatina stuhlmanni	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Atopocochlis exarata	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Limicolaria kambeul	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Limicolaria martenssii	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Limicolariopsis sp.	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Limicolariopsis ruwenzori	CCGTGTCTTC	CGACCGCTCG	CGCAAGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma varicosa	CCGTGTCTTC	CGACCGCTCG	TCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma dimidiata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma sp. cf. vestita	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma marinae	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma churchilliana	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma granulata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma simplex	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma kilburni	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma montistempli	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma omisa	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma semidecussata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma ustulata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Cochlitoma zebra	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Metachatina kraussi	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GCGACTGTAA	
Rumina decollata	CCGTGTCTTC	CGACCGCTCG	CGCGCGACCG	GGCCGGGCTA	GCCGGGCGTC	GC-AC---A-	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	m---mm----	

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1450      1460      1470      1480      1490      1500
Achatina fulica      CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Achatina immaculata CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Achatina reticulata CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Achatina zanzibarica CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Euaethiopia loveridgei CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Achatina achatina    CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Archachatina marginata CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Achatina bisculpta   CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Achatina damarensis  CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Achatina stuhlmanni  CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Atopocochlis exarata CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Limicolaria kambeul  CAGGGTTAGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Limicolaria martenssii CAGGGTTAGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Limicolariopsis sp. CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Limicolariopsis ruwenzori CAGGGTTAGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma varicosa  CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma dimidiata CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma sp. cf. vestita CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma marinae  CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma churchilliana CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma granulata CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma simplex   CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma kilburni  CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma montistempli CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma omissa    CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma semidecussata CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma ustulata  CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Cochlitoma zebra     CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Metachatina kraussi  CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
Rumina decollata    CAGGGTTGGT GGCAGTCCG TCGGCATTCC ACCCGACCCG TCTTGAACA CGGACCAAGG
NUCLEOTIDES INCLUDED mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

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1510      1520      1530      1540      1550      1560
Achatina fulica      AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Achatina immaculata AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Achatina reticulata AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Achatina zanzibarica AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Euaethiopia loveridgei AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Achatina achatina    AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Archachatina marginata AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Achatina bisculpta   AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Achatina damarensis  AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Achatina stuhlmanni  AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Atopocochlis exarata AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Limicolaria kambeul  AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Limicolaria martenssii AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Limicolariopsis sp. AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Limicolariopsis ruwenzori AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma varicosa  AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma dimidiata AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma sp. cf. vestita AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma marinae  AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma churchilliana AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma granulata AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma simplex   AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma kilburni  AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma montistempli AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma omissa    AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma semidecussata AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma ustulata  AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Cochlitoma zebra     AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Metachatina kraussi  AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
Rumina decollata    AGTCTAACAT GCGCGGAGT CATTGGGCGG TACGAAACCC AAAGGCGCAG TGAAGCGGAG
NUCLEOTIDES INCLUDED mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

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	1570	1580	1590	1600	1610	1620
<i>Achatina fulica</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Achatina immaculata</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Achatina reticulata</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Achatina zanzibarica</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Euaethiopina loveridgei</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Achatina achatina</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Archachatina marginata</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Achatina bisculpta</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Achatina damarensis</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Achatina stuhlmanni</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Atopocochlis exarata</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Limicolaria kambeul</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Limicolaria martenssii</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Limicolariopsis sp.</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Limicolariopsis ruwenzori</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma varicosa</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma dimidiata</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	AGGCGCACCA
<i>Cochlitoma sp. cf. vestita</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma marinae</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma churchilliana</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma granulata</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma simplex</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma kilburni</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma montistempli</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma omisa</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma semidecussata</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma ustulata</i>	GGTCGTCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Cochlitoma zebra</i>	GGTCGCCTCG	CGCGATCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Metachatina kraussi</i>	GGTCGCCTCG	CGCGACCCAG	GTGGGATCCT	CCGCTCCTTC	GCGGGAGCGG	TGGCGCACCA
<i>Rumina decollata</i>	GGTCGTCTCN	CGCGACCCAG	GTGGGATCCT	CCACCCCTTC	G-GGGGCGG	TGGCGCACCA
NUCLEOTIDES INCLUDED	mmmmmmmm--	-mmmmmmmm	mmmmmmmmmm	-----	--mmmmmmmm	mmmmmmmmmm

	1630	1640	1650	1660	1670	1680
<i>Achatina fulica</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Achatina immaculata</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Achatina reticulata</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Achatina zanzibarica</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Euaethiopina loveridgei</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Achatina achatina</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Archachatina marginata</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Achatina bisculpta</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Achatina damarensis</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Achatina stuhlmanni</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Atopocochlis exarata</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Limicolaria kambeul</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Limicolaria martenssii</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Limicolariopsis sp.</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Limicolariopsis ruwenzori</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma varicosa</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma dimidiata</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma sp. cf. vestita</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma marinae</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma churchilliana</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma granulata</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma simplex</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma kilburni</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma montistempli</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma omisa</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma semidecussata</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma ustulata</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Cochlitoma zebra</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Metachatina kraussi</i>	CCGGCCCGTC	CCGTCGCGCT	CGTCGGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
<i>Rumina decollata</i>	CCGGCCCGTC	CCGTCYGCY	CGYCYGTGGG	GCGGAGCAAG	AGCGTGCACG	CTGGGACCCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1690	1700	1710	1720	1730	1740
<i>Achatina fulica</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Achatina immaculata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Achatina reticulata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Achatina zanzibarica</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Euaethiopina loveridgei</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Achatina achatina</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Archachatina marginata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Achatina bisculpta</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Achatina damarensis</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Achatina stuhlmanni</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Atopocochlis exarata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Limicolaria kambeul</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Limicolaria martenssii</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Limicolariopsis sp.</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Limicolariopsis ruwenzori</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma varicosa</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma dimidiata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma sp. cf. vestita</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma marinae</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma churchilliana</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma granulata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma simplex</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma kilburni</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma montistempli</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma omissa</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma semidecussata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma ustulata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Cochlitoma zebra</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Metachatina kraussi</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
<i>Rumina decollata</i>	AAAGATGGTG	AACTATGCCT	GAGTAGAACG	AAGCCAGAGG	AAACTCTGGT	GGAGGTTTCGT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1750	1760	1770	1780	1790	1800
<i>Achatina fulica</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Achatina immaculata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Achatina reticulata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Achatina zanzibarica</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Euaethiopina loveridgei</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Achatina achatina</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Archachatina marginata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Achatina bisculpta</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Achatina damarensis</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Achatina stuhlmanni</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Atopocochlis exarata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Limicolaria kambeul</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Limicolaria martenssii</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Limicolariopsis sp.</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Limicolariopsis ruwenzori</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma varicosa</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma dimidiata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma sp. cf. vestita</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma marinae</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma churchilliana</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma granulata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma simplex</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma kilburni</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma montistempli</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma omissa</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma semidecussata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma ustulata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Cochlitoma zebra</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Metachatina kraussi</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
<i>Rumina decollata</i>	AGCGATTCTG	ACGTGCAAAT	CGATCGTCAA	ACTTGGGTAT	AGGGGCGAAA	GACTAATCGA
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1810	1820	1830	1840	1850	1860	
<i>Achatina fulica</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Achatina immaculata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Achatina reticulata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Achatina zanzibarica</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Euaethiopina loveridgei</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Achatina achatina</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Archachatina marginata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Achatina bisculpta</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Achatina damarensis</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Achatina stuhlmanni</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Atopocochlis exarata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Limicolaria kambeul</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Limicolaria martenssii</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Limicolariopsis sp.</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Limicolariopsis ruwenzori</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma varicosa</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma dimidiata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma sp. cf. vestita</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma marinae</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma churchilliana</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma granulata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma simplex</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma kilburni</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma montistempli</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma omisa</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma semidecussata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma ustulata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Cochlitoma zebra</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Metachatina kraussi</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
<i>Rumina decollata</i>	ACCATCTAGT	AGCTGGTTCC	CTCCGAAGTT	TCCCTCAGGA	TAGCTGGCGC	TCGATCGCAG	
NUCLEOTIDES INCLUDED	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	

	1870	1880	1890	1900	1910	1920	
<i>Achatina fulica</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Achatina immaculata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Achatina reticulata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Achatina zanzibarica</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Euaethiopina loveridgei</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Achatina achatina</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Archachatina marginata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Achatina bisculpta</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Achatina damarensis</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Achatina stuhlmanni</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Atopocochlis exarata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Limicolaria kambeul</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Limicolaria martenssii</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Limicolariopsis sp.</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTTT	
<i>Limicolariopsis ruwenzori</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma varicosa</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma dimidiata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma sp. cf. vestita</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma marinae</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma churchilliana</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma granulata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma simplex</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma kilburni</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma montistempli</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma omisa</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma semidecussata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma ustulata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Cochlitoma zebra</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Metachatina kraussi</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
<i>Rumina decollata</i>	TTTTATCCGG	TAAAGCGAAT	GATTAGAGGT	CTTGGGGACG	AAACGACCTC	AACCTATTCT	
NUCLEOTIDES INCLUDED	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	

	1930	1940	1950	1960	1970	1980
<i>Achatina fulica</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTNTC GAATG
<i>Achatina immaculata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Achatina reticulata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Achatina zanzibarica</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Euaethiopina loveridgei</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Achatina achatina</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Archachatina marginata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Achatina bisculpta</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Achatina damarensis</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Achatina stuhlmanni</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Atopocochlis exarata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Limicolaria kambeul</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Limicolaria martenssii</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Limicolariopsis sp.</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AACCC GGC	GTTTT GAATG
<i>Limicolariopsis ruwenzori</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma varicosa</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma dimidiata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma sp. cf. vestita</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma marinae</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma churchilliana</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma granulata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma simplex</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma kilburni</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma montistempli</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma omissa</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma semidecussata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma ustulata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Cochlitoma zebra</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Metachatina kraussi</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTTTC GAATG
<i>Rumina decollata</i>	CAAAC TTAA	ATGGG TAAGA	AGTCC GGCCTC	GCTCG ATTGG	AGCCG GGC	GTNTC GAATG
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	1990	2000	2010	2020	2030	2040
<i>Achatina fulica</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Achatina immaculata</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Achatina reticulata</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Achatina zanzibarica</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Euaethiopina loveridgei</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Achatina achatina</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Archachatina marginata</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Achatina bisculpta</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Achatina damarensis</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Achatina stuhlmanni</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Atopocochlis exarata</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Limicolaria kambeul</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Limicolaria martenssii</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Limicolariopsis sp.</i>	CGTGTGCCAA	GTGGGCCCT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Limicolariopsis ruwenzori</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma varicosa</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma dimidiata</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma sp. cf. vestita</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma marinae</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma churchilliana</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma granulata</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma simplex</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma kilburni</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma montistempli</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma omissa</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma semidecussata</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma ustulata</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Cochlitoma zebra</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Metachatina kraussi</i>	CGTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
<i>Rumina decollata</i>	CNTGTGCCAA	GTGGGCCACT	TTTGGTAAGC	AGAACTGGCG	CTGTGGGATG	AACCAAACGC
NUCLEOTIDES INCLUDED	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm	mmmmmmmm

	2050	2060	2070	2080	2090	2100	
Achatina fulica	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achtina immaculata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina reticulata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina zanzibarica	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Euaethiopina loveridgei	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina achatina	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Archachatina marginata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina bisculpta	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina damarensis	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Achatina stuhlmanni	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Atopocochlis exarata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Limicolaria kambeul	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Limicolaria martenssii	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Limicolariopsis sp.	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Limicolariopsis ruwenzori	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma varicosa	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma dimidiata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma sp. cf. vestita	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma marinae	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma churchilliana	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma granulata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma simplex	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma kilburni	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma montistempli	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma omissa	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma semidecussata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma ustulata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Cochlitoma zebra	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Metachatina kraussi	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
Rumina decollata	CCGGTTAAGG	TGCCAAACGC	TGACGCTCAT	CAGACACCAT	AAAAGGTGTT	GGTTGATATA	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2110	2120	2130	2140	2150	2160	
Achatina fulica	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achtina immaculata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina reticulata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina zanzibarica	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Euaethiopina loveridgei	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina achatina	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Archachatina marginata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina bisculpta	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina damarensis	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Achatina stuhlmanni	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Atopocochlis exarata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Limicolaria kambeul	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Limicolaria martenssii	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Limicolariopsis sp.	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Limicolariopsis ruwenzori	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma varicosa	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma dimidiata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma sp. cf. vestita	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma marinae	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma churchilliana	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma granulata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma simplex	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma kilburni	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma montistempli	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma omissa	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma semidecussata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma ustulata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Cochlitoma zebra	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Metachatina kraussi	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
Rumina decollata	GACAGCAGGA	CGGTGGCCAT	GGAAGTCGGA	ACCCGCTAAG	GAGTGTGTAA	CAACTCACCT	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2170	2180	2190	2200	2210	2220	
<i>Achatina fulica</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Achatina immaculata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Achatina reticulata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Achatina zanzibarica</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Euaethiopina loveridgei</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Achatina achatina</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Archachatina marginata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Achatina bisculpta</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Achatina damarensis</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Achatina stuhlmanni</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Atopocochlis exarata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Limicolaria kambeul</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Limicolaria martenssii</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Limicolariopsis sp.</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Limicolariopsis ruwenzori</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma varicosa</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma dimidiata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma sp. cf. vestita</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma marinae</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma churchilliana</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma granulata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma simplex</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma kilburni</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma montistempli</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma omissa</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma semidecussata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma ustulata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Cochlitoma zebra</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Metachatina kraussi</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
<i>Rumina decollata</i>	GCCGAATCAA	CCAGCCCTGA	AAATGGATGG	CGCTAGAGCG	TCGGACCCAT	ACCGGGCCGT	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2230	2240	2250	2260	2270	2280	
<i>Achatina fulica</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Achatina immaculata</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	NAAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Achatina reticulata</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Achatina zanzibarica</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Euaethiopina loveridgei</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Achatina achatina</i>	CTCGGCAATA	GGCC-----	TTCCG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Archachatina marginata</i>	CTCGGCAATA	GGCC---TTC	TTCCG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Achatina bisculpta</i>	CTCGGCAATG	GGCTCTTTCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Achatina damarensis</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Achatina stuhlmanni</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Atopocochlis exarata</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Limicolaria kambeul</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Limicolaria martenssii</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Limicolariopsis sp.</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Limicolariopsis ruwenzori</i>	CTCGGCAATG	GGCCCTTCCA	TGGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma varicosa</i>	CTCGGCAATG	GGCCCTTCCA	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma dimidiata</i>	CTCGGCAATG	GGCCCTTCCA	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma sp. cf. vestita</i>	CTCGGCAATG	GGCCCTTCCA	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma marinae</i>	CTCGGCAATG	GGCC--TTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma churchilliana</i>	CTCGGCAATG	GGCC--TTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma granulata</i>	CTCGGCAATG	GGCCCTTCCA	ACGGGGCCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma simplex</i>	CTCGGCAATG	GGCCCTTCCA	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma kilburni</i>	CTCGGCAATG	GGCCCTTCCA	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma montistempli</i>	CTCGGCAATG	GGCCCTTCCA	ACGGGGCCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma omissa</i>	CTCGGCAATG	GGCCCTTCCA	ACGGGGCCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma semidecussata</i>	CTCGGCAATG	GGCCCTTCCA	ACGGGGCCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma ustulata</i>	CTCGGCAATG	GGCC--TTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Cochlitoma zebra</i>	CTCGGCAATG	GGCCCTTCCA	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Metachatina kraussi</i>	CTCGGCAATG	GGCC--TTCC	ACGGG-CCGG	-AAGCGAGGC	CGAGACGAGT	AGGAGGGCCG	
<i>Rumina decollata</i>	CTCGGCAGTG	GCGATAGCCG	CG-----	-AATCGAGGC	CGAGACGAGT	AGGAGGGCCG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	m-----	-----mmmm	-mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2290	2300	2310	2320	2330	2340	
<i>Achatina fulica</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Achatina immaculata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Achatina reticulata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Achatina zanzibarica</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Euaethiopina loveridgei</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Achatina achatina</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Archachatina marginata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Achatina bisculpta</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Achatina damarensis</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Achatina stuhlmanni</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Atopocochlis exarata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Limicolaria kambeul</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Limicolaria martenssii</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Limicolariopsis sp.</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Limicolariopsis ruwenzori</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma varicosa</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma dimidiata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma sp. cf. vestita</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma marinae</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma churchilliana</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma granulata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma simplex</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma kilburni</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma montistempli</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma omissa</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma semidecussata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma ustulata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Cochlitoma zebra</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Metachatina kraussi</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
<i>Rumina decollata</i>	TCGGGGTGAG	CGTGGGAAGCC	TGGGGAGCGA	TCCTGGGTGG	AGCCGCCCCG	GGTGCAGATC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2350	2360	2370	2380	2390	2400	
<i>Achatina fulica</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Achatina immaculata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Achatina reticulata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Achatina zanzibarica</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Euaethiopina loveridgei</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Achatina achatina</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Archachatina marginata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Achatina bisculpta</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Achatina damarensis</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Achatina stuhlmanni</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Atopocochlis exarata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Limicolaria kambeul</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Limicolaria martenssii</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Limicolariopsis sp.</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Limicolariopsis ruwenzori</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma varicosa</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma dimidiata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma sp. cf. vestita</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma marinae</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma churchilliana</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma granulata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma simplex</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma kilburni</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma montistempli</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma omissa</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma semidecussata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma ustulata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Cochlitoma zebra</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Metachatina kraussi</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
<i>Rumina decollata</i>	TTGGTGGTAG	TAGCAAATAT	TCAAACGAGA	ACTTTGAAGA	CTGAAGTGGG	GAAGGGTTCC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2410 2420 2430 2440 2450 2460
<i>Achatina fulica</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Achatina immaculata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Achatina reticulata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Achatina zanzibarica</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Euaethiopina loveridgei</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Achatina achatina</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Archachatina marginata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Achatina bisculpta</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Achatina damarensis</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Achatina stuhlmanni</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Atopocochlis exarata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Limicolaria kambeul</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Limicolaria martenssii</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Limicolariaopsis sp.</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Limicolariaopsis ruwenzori</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma varicosa</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma dimidiata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma sp. cf. vestita</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma marinae</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma churchilliana</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma granulata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma simplex</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma kilburni</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma montistempli</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma omissa</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma semidecussata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma ustulata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Cochlitoma zebra</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Metachatina kraussi</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
<i>Rumina decollata</i>	ATGTGAACAG CAGTTGAACA TGGGTCAGTC GGTCCCTAAGA GATAGGAAAA CTC CGTTCTG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2470 2480 2490 2500 2510 2520
<i>Achatina fulica</i>	ACCCCGGGGC ATTCTTTCTT TCTTTCT--- -----AGAA GA GTCTGCCCGC
<i>Achatina immaculata</i>	ACCCCGGGGC ATTCTTTCTT TCTTTCT--- -----AGAA GA GTCTGCCCGC
<i>Achatina reticulata</i>	ACCCCGGGGC ATTCTTTCTT TCTTTCT--- -----AGAA GA GTCTGCCCGC
<i>Achatina zanzibarica</i>	ACCCCGGGGC ATTCTTTCTT TCTTTCT--- -----AGAA GA GTCTGCCCGC
<i>Euaethiopina loveridgei</i>	ACCCCGGGGC ATTCTTTCTT TCTTTCT--- -----AGAA GA GTCTGCCCGC
<i>Achatina achatina</i>	ACCCCGGGGC ATTCTTTCTG TTCTTTC--- -----AGCA GTCTGCCCGC
<i>Archachatina marginata</i>	ACCCCGGGGC ATTCTTTGTC AGCAGT--- -----CAA GTCTGCCCGC
<i>Achatina bisculpta</i>	ACCCCGGGGC ATTCTTTATT A---TTCGTT ATTTCG--- -----AGA GTCTGCCCGC
<i>Achatina damarensis</i>	ACCCCGGGGC ATTCTTTATT ACTCTTCGTT ATT-----AGG GTCTGCCCGC
<i>Achatina stuhlmanni</i>	ACCCCGGGGC ATTCTTTATA A---TTCGTT ATT-----AGG GTCTGCCCGC
<i>Atopocochlis exarata</i>	ACCCCGGGGC ATTCTTTATT A---ATCGTT TAA-----AGG GTCTGCCCGC
<i>Limicolaria kambeul</i>	ACCCCGGGGC ATTCTTTATT A---TTCGTT -----AGG GTCTGCCCGC
<i>Limicolaria martenssii</i>	ACCCCGGGGC ATTCTTTATT A---TTCGTT -----AGG GTCTGCCCGC
<i>Limicolariaopsis sp.</i>	ACCCCGGGGC ATTCTTTATT ATA-TTCGTT TT-----AGG GTCTGCCCGC
<i>Limicolariaopsis ruwenzori</i>	ACCCCGGGGC ATTCTTTATT A---TTCGTT -----AGG GTCTGCCCGC
<i>Cochlitoma varicosa</i>	ACCCCGGGGC ATTCTTTCTT T--CTTT--- -----CGTGATTAAG GTCTGCCCGC
<i>Cochlitoma dimidiata</i>	ACCCCGGGGC ATTCTTTCTT T--CTTT--- -----A GTCTGCCCGC
<i>Cochlitoma sp. cf. vestita</i>	ACCCCGGGGC ATTCTTTCTG G-----ATTAAG GTCTGCCCGC
<i>Cochlitoma marinae</i>	ACCCCGGGGC ATTCTTTCTG G-----ATTAAG GTCTGCCCGC
<i>Cochlitoma churchilliana</i>	ACCCCGGGGC ATTCTTTCTT T--CTTTCT T-----GATTAA GTCTGCCCGC
<i>Cochlitoma granulata</i>	ACCCCGGGGC ATTCTTTCTT TTTCTT--- -----G ATTAATTAAG GTCTGCCCGC
<i>Cochlitoma simplex</i>	ACCCCGGGGC ATTCTTTCTT T--CTTT--- -----A GTCTGCCCGC
<i>Cochlitoma kilburni</i>	ACCCCGGGGC ATTCTTTCTT T--CTTT--- -----ATTA ATTAATTAAG GTCTGCCCGC
<i>Cochlitoma montistempli</i>	ACCCCGGGGC ATTCTTTCTT TTTCTT--- -----G ATTAATTAAG GTCTGCCCGC
<i>Cochlitoma omissa</i>	ACCCCGGGGC ATTCTTTCTT TTTCTT--- -----G ATTAATTAAG GTCTGCCCGC
<i>Cochlitoma semidecussata</i>	ACCCCGGGGC ATTCTTTCTT TTTCTT--- -----G ATTAATTAAG GTCTGCCCGC
<i>Cochlitoma ustulata</i>	ACCCCGGGGC ATTCTTTCTG G-----ATTAAG GTCTGCCCGC
<i>Cochlitoma zebra</i>	ACCCCGGGGC ATTCTTTCTG G-----ATTAAG GTCTGCCCGC
<i>Metachatina kraussi</i>	ACCCCGGGGC ATTCTTTCTT T--CTT--- -----AAA GTCTGCCCGC
<i>Rumina decollata</i>	ACCCCGGGGC AATCTTTCTC GAGCTA--- -----AGG AGAAAAAGAA GTCTGCCCGC
NUCLEOTIDES INCLUDED	mmmmmmmmmm -----m mmmmmmmmmmm

	2530 2540 2550 2560 2570 2580
<i>Achatina fulica</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Achatina immaculata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Achatina reticulata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Achatina zanzibarica</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Euaethiopina loveridgei</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Achatina achatina</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Archachatina marginata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Achatina bisculpta</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Achatina damarensis</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Achatina stuhlmanni</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Atopocochlis exarata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Limicolaria kambeul</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Limicolaria martenssii</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Limicolariopsis sp.</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Limicolariopsis ruwenzori</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma varicosa</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma dimidiata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma sp. cf. vestita</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma marinae</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma churchilliana</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma granulata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma simplex</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma kilburni</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma montistempli</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma omissa</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma semidecussata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma ustulata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Cochlitoma zebra</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Metachatina kraussi</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
<i>Rumina decollata</i>	AGCCTATCGA AAGGGAATCG GGTTAATATT CCCGAACCTG GACACGGAGA TTGGTCCTCA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2590 2600 2610 2620 2630 2640
<i>Achatina fulica</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Achatina immaculata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Achatina reticulata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Achatina zanzibarica</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Euaethiopina loveridgei</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Achatina achatina</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Archachatina marginata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Achatina bisculpta</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Achatina damarensis</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Achatina stuhlmanni</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Atopocochlis exarata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Limicolaria kambeul</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Limicolaria martenssii</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Limicolariopsis sp.</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Limicolariopsis ruwenzori</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma varicosa</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma dimidiata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma sp. cf. vestita</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma marinae</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma churchilliana</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma granulata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma simplex</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma kilburni</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma montistempli</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma omissa</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma semidecussata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma ustulata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Cochlitoma zebra</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Metachatina kraussi</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
<i>Rumina decollata</i>	GGGGCCACGT GCGGCAACGC AAACGAAGTG GGGGACGTCG GCGGGAGCCC CGGGAAGAGT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2650 2660 2670 2680 2690 2700
<i>Achatina fulica</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Achatina immaculata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Achatina reticulata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Achatina zanzibarica</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Euaethiopina loveridgei</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Achatina achatina</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Archachatina marginata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Achatina bisculpta</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Achatina damarensis</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Achatina stuhlmanni</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Atopocochlis exarata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Limicolaria kambeul</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Limicolaria martenssii</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Limicolariopsis sp.</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Limicolariopsis ruwenzori</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma varicosa</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma dimidiata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma sp. cf. vestita</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma marinae</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma churchilliana</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma granulata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma simplex</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma kilburni</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma montistempli</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma omissa</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma semidecussata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma ustulata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Cochlitoma zebra</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Metachatina kraussi</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
<i>Rumina decollata</i>	TCTCTTTTCT TTGTAAGGAG CCACATCCCT GGAATCGGCT TGCCCGGAGA TAGGGACCGC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2710 2720 2730 2740 2750 2760
<i>Achatina fulica</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Achatina immaculata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Achatina reticulata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Achatina zanzibarica</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Euaethiopina loveridgei</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Achatina achatina</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Archachatina marginata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Achatina bisculpta</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Achatina damarensis</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Achatina stuhlmanni</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Atopocochlis exarata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Limicolaria kambeul</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Limicolaria martenssii</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Limicolariopsis sp.</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Limicolariopsis ruwenzori</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma varicosa</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma dimidiata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma sp. cf. vestita</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma marinae</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma churchilliana</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma granulata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma simplex</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma kilburni</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma montistempli</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma omissa</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma semidecussata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma ustulata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Cochlitoma zebra</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Metachatina kraussi</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
<i>Rumina decollata</i>	GGCCCCGTAA AGCACC GCGG CTCTTGCGGT GTCCGGTGCG CTCCCGTCGG CCCTTGAAAA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	2770	2780	2790	2800	2810	2820	
<i>Achatina fulica</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Achatina immaculata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Achatina reticulata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Achatina zanzibarica</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Euaethiopina loveridgei</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Achatina achatina</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Archachatina marginata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Achatina bisculpta</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Achatina damarensis</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Achatina stuhlmanni</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Atopocochlis exarata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Limicolaria kambeul</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Limicolaria martenssii</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Limicolariopsis sp.</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Limicolariopsis ruwenzori</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma varicosa</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma dimidiata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma sp. cf. vestita</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma marinae</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma churchilliana</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma granulata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma simplex</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma kilburni</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma montistempli</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma omissa</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma semidecussata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma ustulata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Cochlitoma zebra</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Metachatina kraussi</i>	CCCCACGNAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
<i>Rumina decollata</i>	CCCCACGGAG	ACGGTGTGAA	TTTCGTGCCA	GGCCGTACCC	ATATCCGCAG	CAGGTCTCCG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2830	2840	2850	2860	2870	2880	
<i>Achatina fulica</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Achatina immaculata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Achatina reticulata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Achatina zanzibarica</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Euaethiopina loveridgei</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Achatina achatina</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Archachatina marginata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Achatina bisculpta</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Achatina damarensis</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Achatina stuhlmanni</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Atopocochlis exarata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Limicolaria kambeul</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Limicolaria martenssii</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Limicolariopsis sp.</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Limicolariopsis ruwenzori</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma varicosa</i>	AGGTGCACAG	TCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma dimidiata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma sp. cf. vestita</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma marinae</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma churchilliana</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma granulata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma simplex</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma kilburni</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma montistempli</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma omissa</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma semidecussata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma ustulata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Cochlitoma zebra</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Metachatina kraussi</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
<i>Rumina decollata</i>	AGGTGCACAG	CCTCTAGTCG	ATAGAACAAT	GTAGGTAAGG	GAAGTCGGCA	AATTGGATCC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	2890	2900	2910	2920	2930	2940	
<i>Achatina fulica</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Achatina immaculata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Achatina reticulata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Achatina zanzibarica</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Euaethiopina loveridgei</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Achatina achatina</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Archachatina marginata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Achatina bisculpta</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Achatina damarensis</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Achatina stuhlmanni</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Atopocochlis exarata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Limicolaria kambeul</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Limicolaria martenssii</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Limicolariopsis sp.</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Limicolariopsis ruwenzori</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma varicosa</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma dimidiata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma sp. cf. vestita</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma marinae</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma churchilliana</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma granulata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma simplex</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma kilburni</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma montistempli</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma omissa</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma semidecussata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma ustulata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Cochlitoma zebra</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Metachatina kraussi</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
<i>Rumina decollata</i>	GTAAC	TTCGG	GAAAAGGATT	GGCTCTGAGG	GCTGGGTCAG	TCGGGCCGGG	GTACGAAGCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	2950	2960	2970	2980	2990	3000	
<i>Achatina fulica</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Achatina immaculata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Achatina reticulata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Achatina zanzibarica</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Euaethiopina loveridgei</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Achatina achatina</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Archachatina marginata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Achatina bisculpta</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Achatina damarensis</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Achatina stuhlmanni</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Atopocochlis exarata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Limicolaria kambeul</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Limicolaria martenssii</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Limicolariopsis sp.</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Limicolariopsis ruwenzori</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma varicosa</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma dimidiata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma sp. cf. vestita</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma marinae</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma churchilliana</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma granulata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma simplex</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma kilburni</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma montistempli</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma omissa</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma semidecussata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma ustulata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Cochlitoma zebra</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Metachatina kraussi</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
<i>Rumina decollata</i>	GGACTGGGAT	GGGCCCGGGC	TGGGCGAGGC	CGCCGCCGCT	AGCCGGCCGG	CCGGCCGAGC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3010	3020	3030	3040	3050	3060	
Achatina fulica	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Achatina immaculata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Achatina reticulata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Achatina zanzibarica	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Euaethiopina loveridgei	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Achatina achatina	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Archachatina marginata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Achatina bisculpta	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Achatina damarensis	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Achatina stuhlmanni	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Atopocochlis exarata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Limicolaria kambeul	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Limicolaria martenssii	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Limicolariopsis sp.	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Limicolariopsis ruwenzori	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma varicosa	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma dimidiata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma sp. cf. vestita	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma marinae	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma churchilliana	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma granulata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma simplex	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma kilburni	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma montistempli	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma omissa	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma semidecussata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma ustulata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Cochlitoma zebra	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Metachatina kraussi	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCTCCCCGG	
Rumina decollata	TCGGAACGCG	GCTGCAACCT	TCCCCTGGAC	CGCCCCAGCT	ATGCGGCGGC	GCCT-CCCCG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3070	3080	3090	3100	3110	3120	
Achatina fulica	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina immaculata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina reticulata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina zanzibarica	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Euaethiopina loveridgei	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina achatina	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Archachatina marginata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina bisculpta	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina damarensis	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Achatina stuhlmanni	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Atopocochlis exarata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Limicolaria kambeul	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Limicolaria martenssii	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Limicolariopsis sp.	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Limicolariopsis ruwenzori	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma varicosa	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma dimidiata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma sp. cf. vestita	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma marinae	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma churchilliana	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma granulata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma simplex	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma kilburni	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma montistempli	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma omissa	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma semidecussata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma ustulata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Cochlitoma zebra	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Metachatina kraussi	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
Rumina decollata	CGTCGTTCCG	GTCGGCTGGC	ATTCAACAGC	CAACTCAGAA	CTGGTACGGA	CCAGGGGAAT	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3130	3140	3150	3160	3170	3180	
<i>Achatina fulica</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Achatina immaculata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Achatina reticulata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Achatina zanzibarica</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Euaethiopina loveridgei</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Achatina achatina</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Archachatina marginata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Achatina bisculpta</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Achatina damarensis</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Achatina stuhlmanni</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Atopocochlis exarata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Limicolaria kambeul</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Limicolaria martenssii</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Limicolaria ruwenzori</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma varicosa</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma dimidiata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma sp. cf. vestita</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma marinae</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma churchilliana</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma granulata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma simplex</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma kilburni</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma montistempli</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma omissa</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma semidecussata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma ustulata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Cochlitoma zebra</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Metachatina kraussi</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
<i>Rumina decollata</i>	CCGACTGTCT	AATTA AAAACA	AAGCATTGCG	ACGGCCGTCA	CCCGGTGTTG	ACGCAATGTG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3190	3200	3210	3220	3230	3240	
<i>Achatina fulica</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Achatina immaculata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Achatina reticulata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Achatina zanzibarica</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Euaethiopina loveridgei</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Achatina achatina</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Archachatina marginata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Achatina bisculpta</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Achatina damarensis</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Achatina stuhlmanni</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Atopocochlis exarata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Limicolaria kambeul</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Limicolaria martenssii</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Limicolaria ruwenzori</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma varicosa</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma dimidiata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma sp. cf. vestita</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma marinae</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma churchilliana</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma granulata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma simplex</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma kilburni</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma montistempli</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma omissa</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma semidecussata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma ustulata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Cochlitoma zebra</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Metachatina kraussi</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
<i>Rumina decollata</i>	ATTTCTGCCC	AGTGCTCTGA	ATGTCAAAGT	GAAGAAATTC	AACCAAGCGC	GGGTAACCGG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3250 3260 3270 3280 3290 3300
<i>Achatina fulica</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Achatina immaculata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Achatina reticulata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Achatina zanzibarica</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Euaethiopina loveridgei</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Achatina achatina</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Archachatina marginata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Achatina bisculpta</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Achatina damarensis</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Achatina stuhlmanni</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Atopocochlis exarata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Limicolaria kambeul</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Limicolaria martenssii</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Limicolariopsis sp.</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Limicolariopsis ruwenzori</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma varicosa</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma dimidiata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma sp. cf. vestita</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma marinae</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma churchilliana</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma granulata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma simplex</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma kilburni</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma montistempli</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma omissa</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma semidecussata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma ustulata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Cochlitoma zebra</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Metachatina kraussi</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
<i>Rumina decollata</i>	CGGGAGTAAC TATGACTCTC TTAAGGTAGC CAAATGCCTC GTCATCTAAT TAGTGACGCG
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3310 3320 3330 3340 3350 3360
<i>Achatina fulica</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Achatina immaculata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Achatina reticulata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Achatina zanzibarica</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Euaethiopina loveridgei</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Achatina achatina</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Archachatina marginata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Achatina bisculpta</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Achatina damarensis</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Achatina stuhlmanni</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Atopocochlis exarata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Limicolaria kambeul</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Limicolaria martenssii</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Limicolariopsis sp.</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Limicolariopsis ruwenzori</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma varicosa</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma dimidiata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma sp. cf. vestita</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma marinae</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma churchilliana</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma granulata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma simplex</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma kilburni</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma montistempli</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma omissa</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma semidecussata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma ustulata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Cochlitoma zebra</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Metachatina kraussi</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
<i>Rumina decollata</i>	CATGAATGGA TTAACGAGAT TCCCCTGTG CCTATCTACT ATCTAGCGAA ACCACAGCCA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3370 3380 3390 3400 3410 3420
<i>Achatina fulica</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Achatina immaculata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Achatina reticulata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Achatina zanzibarica</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Euaethiopina loveridgei</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Achatina achatina</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Archachatina marginata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Achatina bisculpta</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Achatina damarensis</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Achatina stuhlmanni</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Atopocochlis exarata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Limicolaria kambeul</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Limicolaria martenssii</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Limicolariopsis sp.</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Limicolariopsis ruwenzori</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma varicosa</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma dimidiata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma sp. cf. vestita</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma marinae</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma churchilliana</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma granulata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma simplex</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma kilburni</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma montistempli</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma omissa</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma semidecussata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma ustulata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Cochlitoma zebra</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Metachatina kraussi</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
<i>Rumina decollata</i>	AGGGAACGGG CTTGGTAGAA TCAGCGGGGA AAGAAGACCC TGTTGAGCTT GACTCTAGTC
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3430 3440 3450 3460 3470 3480
<i>Achatina fulica</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Achatina immaculata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Achatina reticulata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Achatina zanzibarica</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Euaethiopina loveridgei</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Achatina achatina</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Archachatina marginata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Achatina bisculpta</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Achatina damarensis</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Achatina stuhlmanni</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Atopocochlis exarata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Limicolaria kambeul</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Limicolaria martenssii</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Limicolariopsis sp.</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Limicolariopsis ruwenzori</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma varicosa</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma dimidiata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma sp. cf. vestita</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma marinae</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma churchilliana</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma granulata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma simplex</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma kilburni</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma montistempli</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma omissa</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma semidecussata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma ustulata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Cochlitoma zebra</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Metachatina kraussi</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
<i>Rumina decollata</i>	CGACTTTGTG AAGACACATG AAGGGTGTAG CATAGGTGGG AGCGCAAGCG CAATTGAAAT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3490	3500	3510	3520	3530	3540	
<i>Achatina fulica</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Achatina immaculata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Achatina reticulata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Achatina zanzibarica</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Euaethiopina loveridgei</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Achatina achatina</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Archachatina marginata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Achatina bisculpta</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Achatina damarensis</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Achatina stuhlmanni</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Atopocochlis exarata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Limicolaria kambeul</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Limicolaria martenssii</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Limicolariopsis sp.</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Limicolariopsis ruwenzori</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma varicosa</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma dimidiata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma sp. cf. vestita</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma marinae</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma churchilliana</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma granulata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma simplex</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma kilburni</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma montistempli</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma omisa</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma semidecussata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma ustulata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Cochlitoma zebra</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Metachatina kraussi</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
<i>Rumina decollata</i>	ACCACTACTT	TTATCGTTTC	TTTACTTATT	CAGTCAAGCG	GAGAGCGGGG	CGCAAGCCCC	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3550	3560	3570	3580	3590	3600	
<i>Achatina fulica</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Achatina immaculata</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Achatina reticulata</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Achatina zanzibarica</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Euaethiopina loveridgei</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Achatina achatina</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Archachatina marginata</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Achatina bisculpta</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Achatina damarensis</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Achatina stuhlmanni</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Atopocochlis exarata</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Limicolaria kambeul</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Limicolaria martenssii</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Limicolariopsis sp.</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Limicolariopsis ruwenzori</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma varicosa</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma dimidiata</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma sp. cf. vestita</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma marinae</i>	TCGCTTCTGG	AATTAAGCGG	CAACAGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma churchilliana</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma granulata</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma simplex</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-TCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma kilburni</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma montistempli</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma omisa</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma semidecussata</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma ustulata</i>	TCGCTTCTGG	AGTTAAGCGG	CAATCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Cochlitoma zebra</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Metachatina kraussi</i>	TCGCTTCTGG	AGTTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
<i>Rumina decollata</i>	TCGCTTCTGG	AATTAAGCGG	CAACCGGCGC	TCGTCGTCG-	-GCCGCGATC	CGCTCTGAAG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3610 3620 3630 3640 3650 3660
<i>Achatina fulica</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Achatina immaculata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Achatina reticulata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Achatina zanzibarica</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Euaethiopina loveridgei</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Achatina achatina</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Archachatina marginata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Achatina bisculpta</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Achatina damarensis</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Achatina stuhlmanni</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Atopocochlis exarata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Limicolaria kambeul</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Limicolaria martenssii</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Limicolariopsis sp.</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Limicolariopsis ruwenzori</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma varicosa</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma dimidiata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma sp. cf. vestita</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma marinae</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma churchilliana</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma granulata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma simplex</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma kilburni</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma montistempli</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma omissa</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma semidecussata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma ustulata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Cochlitoma zebra</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Metachatina kraussi</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
<i>Rumina decollata</i>	ACAGTGTCCAG GCGGGGAGTT TGACTGGGGC GGTACATCTG TCAAAAAGGTA ACGCAGGTGT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3670 3680 3690 3700 3710 3720
<i>Achatina fulica</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Achatina immaculata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Achatina reticulata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Achatina zanzibarica</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Euaethiopina loveridgei</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Achatina achatina</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Archachatina marginata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Achatina bisculpta</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Achatina damarensis</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Achatina stuhlmanni</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Atopocochlis exarata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Limicolaria kambeul</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Limicolaria martenssii</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Limicolariopsis sp.</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Limicolariopsis ruwenzori</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma varicosa</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma dimidiata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma sp. cf. vestita</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma marinae</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma churchilliana</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma granulata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma simplex</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma kilburni</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma montistempli</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma omissa</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma semidecussata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma ustulata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Cochlitoma zebra</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Metachatina kraussi</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
<i>Rumina decollata</i>	CCTAAGGCGA GCTCAGCGAG GACGGAAACC TCGCGTAGAG CAAAAGGGCA AAAGCTCGCT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	3730	3740	3750	3760	3770	3780	
<i>Achatina fulica</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Achatina immaculata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Achatina reticulata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Achatina zanzibarica</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Euaethiopina loveridgei</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Achatina achatina</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Archachatina marginata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Achatina bisculpta</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Achatina damarensis</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Achatina stuhlmanni</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Atopocochlis exarata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Limicolaria kambeul</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Limicolaria martenssii</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Limicolariopsis sp.</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Limicolariopsis ruwenzori</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma varicosa</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma dimidiata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma sp. cf. vestita</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma marinae</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma churchilliana</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma granulata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma simplex</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma kilburni</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma montistempli</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma omissa</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma semidecussata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma ustulata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Cochlitoma zebra</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Metachatina kraussi</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
<i>Rumina decollata</i>	TGATTTTGAT	TTTCAGTACG	AATACAGACC	GTGAAAGCGT	GGCCTATCGA	TCCTTTTGAC	
NUCLEOTIDES INCLUDED	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	

	3790	3800	3810	3820	3830	3840	
<i>Achatina fulica</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Achatina immaculata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Achatina reticulata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Achatina zanzibarica</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Euaethiopina loveridgei</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Achatina achatina</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Archachatina marginata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Achatina bisculpta</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Achatina damarensis</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Achatina stuhlmanni</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Atopocochlis exarata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Limicolaria kambeul</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Limicolaria martenssii</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Limicolariopsis sp.</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Limicolariopsis ruwenzori</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma varicosa</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma dimidiata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma sp. cf. vestita</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma marinae</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma churchilliana</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma granulata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma simplex</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma kilburni</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma montistempli</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma omissa</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma semidecussata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma ustulata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Cochlitoma zebra</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Metachatina kraussi</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
<i>Rumina decollata</i>	TTTAAGAGTT	TTAAGCAAGA	GGTGTCAGAA	AAGTTACCAC	AGGGATAACT	GGCTTGTGGC	
NUCLEOTIDES INCLUDED	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	

	3850	3860	3870	3880	3890	3900
<i>Achatina fulica</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Achatina immaculata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Achatina reticulata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Achatina zanzibarica</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Euaethiopina loveridgei</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Achatina achatina</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Archachatina marginata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Achatina bisculpta</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Achatina damarensis</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Achatina stuhlmanni</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Atopocochlis exarata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Limicolaria kambeul</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Limicolaria martenssii</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Limicolariopsis sp.</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Limicolariopsis ruwenzori</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma varicosa</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma dimidiata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma sp. cf. vestita</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma marinae</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma churchilliana</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma granulata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma simplex</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma kilburni</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma montistempli</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma omissa</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma semidecussata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma ustulata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Cochlitoma zebra</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Metachatina kraussi</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
<i>Rumina decollata</i>	AGCCAAGCGT	TCATAGCGAC	GTTGCTTTT	GATCCTTCGA	TGTCGGCTCT	TCCTATCATT	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3910	3920	3930	3940	3950	3960
<i>Achatina fulica</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Achatina immaculata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Achatina reticulata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Achatina zanzibarica</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Euaethiopina loveridgei</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Achatina achatina</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Archachatina marginata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Achatina bisculpta</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Achatina damarensis</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Achatina stuhlmanni</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Atopocochlis exarata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Limicolaria kambeul</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Limicolaria martenssii</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Limicolariopsis sp.</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Limicolariopsis ruwenzori</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma varicosa</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma dimidiata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma sp. cf. vestita</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma marinae</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma churchilliana</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma granulata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma simplex</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma kilburni</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma montistempli</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma omissa</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma semidecussata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma ustulata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Cochlitoma zebra</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Metachatina kraussi</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
<i>Rumina decollata</i>	GCGAAGCAGA	ATTTCGCCAAG	CGTTGGATTG	TTCACCCACT	AATAGGGAAC	GTGAGCTGGG	
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	3970 3980 3990 4000 4010 4020
<i>Achatina fulica</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Achatina immaculata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Achatina reticulata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Achatina zanzibarica</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Euaethiopina loveridgei</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Achatina achatina</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Archachatina marginata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Achatina bisculpta</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Achatina damarensis</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Achatina stuhlmanni</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Atopocochlis exarata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Limicolaria kambeul</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Limicolaria martenssii</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Limicolariopsis sp.</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Limicolariopsis ruwenzori</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma varicosa</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma dimidiata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma sp. cf. vestita</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma marinae</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma churchilliana</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma granulata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma simplex</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma kilburni</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma montistempli</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma omissa</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma semidecussata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma ustulata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Cochlitoma zebra</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Metachatina kraussi</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
<i>Rumina decollata</i>	TTTAGACCGT CGTGAGACAG GTTAGTTTTA CCCTACTGAT GACAGGTCGT TGCTACGGTA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

	4030 4040 4050 4060 4070 4080
<i>Achatina fulica</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Achatina immaculata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Achatina reticulata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Achatina zanzibarica</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Euaethiopina loveridgei</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Achatina achatina</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Archachatina marginata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Achatina bisculpta</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Achatina damarensis</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Achatina stuhlmanni</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Atopocochlis exarata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Limicolaria kambeul</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Limicolaria martenssii</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Limicolariopsis sp.</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Limicolariopsis ruwenzori</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma varicosa</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma dimidiata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma sp. cf. vestita</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma marinae</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma churchilliana</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma granulata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma simplex</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma kilburni</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma montistempli</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma omissa</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma semidecussata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma ustulata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Cochlitoma zebra</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Metachatina kraussi</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
<i>Rumina decollata</i>	ATCCTGCTCA GTACGAGAGG AACCGCAGGT TCAGACATTT GGTTCACGTG CTTGGCTGAT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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.....|.....| .....|.....| .....|.....| .....|...
          4090          4100          4110
Achatina fulica      AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Achatina immaculata AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Achatina reticulata AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Achatina zanzibarica AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Euaethiopina loveridgei AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Achatina achatina    AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Archachatina marginata AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Achatina bisculpta   AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Achatina damarensis  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Achatina stuhlmanni  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Atopocochlis exarata AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Limicolaria kambeul  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Limicolaria martenssii AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Limicolariopsis sp. AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Limicolariopsis ruwenzori AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma varicosa  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma dimidiata AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma sp. cf. vestita AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma marinae  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma churchilliana AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma granulata AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma simplex  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma kilburni  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma montistempli AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma omissa    AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma semidecussata AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma ustulata  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Cochlitoma zebra     AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Metachatina kraussi  AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
Rumina decollata     AAGCCAATGG TCGGAGGCTA CCATCTGAGG GATTATGG
NUCLEOTIDES INCLUDED mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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Appendix 4.2: Alignment of the actin gene for the Achatinidae and the subulinid outgroup *Rumina decollata*. Note that sequencing was unsuccessful for *Limicolariopsis ruwenzoriensis*.

	10	20	30	40	50	60
Achatina fulica	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Achatina immaculata	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Achatina reticulata	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Achatina zanzibarica	TCCAAGAGGG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Euaethiopina loveridgei	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Achatina achatina	TCCAAGAGAG	GTATYCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Archachatina marginata	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Achatina bisculpta	TCYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Achatina damarensis	TYYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Achatina stuhlmanni	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Atopocochlis exarata	TCYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Limicolaria kambeul	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Limicolaria martenssii	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Limicolariopsis sp.	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma varicosa	TCCAAGAGAG	GYATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma dimidiata	TCCAAGAGAG	GYATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma sp. cf. vestita	TCCAAGAGAG	GYATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma marinae	TCTAAGAGGG	GTATTCTCAC	ACTCAAGTAC	CCCATTGAGC	ATGGTATTGT	CACAACTGG
Cochlitoma churchilliana	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma granulata	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma simplex	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cpchlilitoma kilburni	TCCAAGRGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma montistempli	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma omissa	TCCAAGAGAG	GYATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma semidecussata	TCCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma ustulata	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Cochlitoma zebra	TCCAAGAGAG	GCATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Metachatina kraussi	TYCAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG
Rumina decollata	TCYAAGAGAG	GTATCCTCAC	TCTCAAGTAT	CCCATTGAGC	ATGGTATTGT	CACCAACTGG

	70	80	90	100	110	120
Achatina fulica	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTSAGAGT	TGCACCAGAA
Achatina immaculata	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCWCCAGAA
Achatina reticulata	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCRGAA
Achatina zanzibarica	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Euaethiopina loveridgei	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Achatina achatina	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGTTGAGAGT	TGCACCAGAA
Archachatina marginata	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGYTGAGAGT	TGCACCAGAA
Achatina bisculpta	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Achatina damarensis	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCWCCAGAA
Achatina stuhlmanni	GATGATATGG	AGAAAATATG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Atopocochlis exarata	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTAYAATG	ARCTGAGAGT	TGCACCAGAA
Limicolaria kambeul	GATGAYATGG	AGAAGATCTG	GCATCACACC	TTCTAYAATG	AGCTGAGAGT	TGCACCAGAA
Limicolaria martenssii	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTATAATG	AGCTGAGAGT	TGCACCAGAA
Limicolariopsis sp.	GATGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma varicosa	GATGATATGG	AGAAGATCTG	GCAYCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma dimidiata	GATGATATGG	AGAAGATCTG	GCAYCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma sp. cf. vestita	GATGATATGG	AGAAGATCTG	GCAYCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma marinae	GACGATATGG	AGAAGATCTG	GCATCACACC	TTCTACARCG	AGCTTAGAGT	TGCCCCCTGAA
Cochlitoma churchilliana	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma granulata	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma simplex	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cpchlilitoma kilburni	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma montistempli	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma omissa	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma semidecussata	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma ustulata	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Cochlitoma zebra	GATGATATGG	AGAAGATCTG	GCACCACACC	TTCTACAATG	AGCTGAGAGT	TGCACCAGAA
Metachatina kraussi	GATGATATGG	AGAARATCTG	GCAYCACACC	TTCTACAATG	AGCTGAGAGT	TGCWCCAGAA
Rumina decollata	GAYGATATGG	AGAAGATCTG	GCATCACACC	TTCTACAATG	AGCTGAGAGT	TGCWCCAGAA

	130 140 150 160 170 180
<i>Achatina fulica</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Achatina immaculata</i>	GAGCATCCAG TTCTGCTTAC AGAAGCTCCA CTCAACCCCA AGGCCAAYAG AGAGAAGATG
<i>Achatina reticulata</i>	GARCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCYA AGGCCAACAG AGAGAAGATG
<i>Achatina zanzibarica</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Euaethiopina loveridgei</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Achatina achatina</i>	GAGCATCCAG TTCTRCKTAC AGAGGCTCCA CTCAATCCCA AGGCCAACAG AGAGAAGATG
<i>Archachatina marginata</i>	GAGCATCCAG TTCTRCKTAC AGAGGCTCCA CTCAATCCCA AGGCCAACAG AGAGAAGATG
<i>Achatina bisculpta</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Achatina damarensis</i>	GARCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Achatina stuhlmanni</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Atopocochlis exarata</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Limicolaria kambeul</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAAYCCCA AGGCCAACAG AGAGAAGATG
<i>Limicolaria martenssii</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Limicolariopsis sp.</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAAYCCCA AGGCCAACAG AGAGAAGATG
<i>Cochlitoma varicosa</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCD CTCAACCCCA AGGCCAAYAG AGAGAAGATG
<i>Cochlitoma dimidiata</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Cochlitoma sp. cf. vestita</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAAYAG AGAGAAGATG
<i>Cochlitoma marinae</i>	GAACATCCCG TTTTGCTGAC AGAGGCTCCT CTCAACCCAA AAGCCAACAG AGAGAAGATG
<i>Cochlitoma churchilliana</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Cochlitoma granulata</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Cochlitoma simplex</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Cpchlitoloma kilburni</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAATAG AGAGAAGATG
<i>Cochlitoma montistempli</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Cochlitoma omissa</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Cochlitoma semidecussata</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Cochlitoma ustulata</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Cochlitoma zebra</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCA CTCAACCCCA AGGCCAATAG AGAGAAGATG
<i>Metachatina kraussi</i>	GAGCATCCAG TTCTGCTTAC AGAGGCTCCM CTCAACCCCA AGGCCAACAG AGAGAAGATG
<i>Rumina decollata</i>	GAGCACCAG TTYTSCTKAC AGAGGCTCCA CTCAACCCCA AGGCCAACAG AGAGAAGATG

	190 200 210 220 230 240
<i>Achatina fulica</i>	ACCCAGATCA TGTTTGA AAC YTTCAAYTCT CCAGCCATGT ATGTCGCCAT YCAAGCYGTG
<i>Achatina immaculata</i>	ACCCAGATCA TGTTTGA AAC TTTCAATTCT CCAGCCATGT ATGTYGCCAT TCAAGCYGTG
<i>Achatina reticulata</i>	ACCCAGATCA TGTTTGA AAC YTTCAAYTCT CCAGCCATGT AYGTYGCCAT TCAAGCTGTG
<i>Achatina zanzibarica</i>	ACCCAGATCA TGTTTGA AAC TTTCAATTCT CCAGCCATGT ATGTCGCCAT CCAAGCCGTG
<i>Euaethiopina loveridgei</i>	ACCCAGATCA TGTTTGA AAC TTTCAACTCT CCAGCCATGT ATGTCGCCAT CCAAGCCGTG
<i>Achatina achatina</i>	ACCCAGATCA TGTTTGA AAC YTTCAAYWSY CCAGCYATGT ATGTYGCIAT TCAAGCHGTG
<i>Archachatina marginata</i>	ACCCAGATCA TGTTTGA AAC CTTCAAYACC CCAGCYATGT ATGTTGCCAT CCAAGCYGTG
<i>Achatina bisculpta</i>	ACCCAGATCA TGTTTGA AAC YTTCAAYWCC CCAGCYATGT ATGTTGCCAT YCAAGCCGTG
<i>Achatina damarensis</i>	ACCCAGATCA TGTTTGA AAC YTTCAACWCY CCAGCYATGT ATGTTGCCAT TCAAGCYGTG
<i>Achatina stuhlmanni</i>	ACCCAGATCA TGTTTGA AAC TTTCAACTCT CCAGCCATGT ATGTTGCCAT TCAAGCAGTC
<i>Atopocochlis exarata</i>	ACCCAGATCA TGTTTGA AAC YTTCAACTCY CCAGCCATGT AYGTTGCIAT TCAAGCTGTG
<i>Limicolaria kambeul</i>	ACCCAGATCA TGTTTGA AAC YTTCAAYWCY CCAGCYATGT AYGTTGCCAT YCAAGCYGTG
<i>Limicolaria martenssii</i>	ACCCAGATCA TGTTTGA AAC CTTCAATACC CCAGCYATGT AYGTTGCCAT YCAAGCYGTG
<i>Limicolariopsis sp.</i>	ACCCAGATCA TGTTTGA AAC YTTCAAYWCY CCAGCCATGT AYGTYGCCAT TCAAGCYGTG
<i>Cochlitoma varicosa</i>	ACCCAGATCA TGTTTGA AAC CTTCAAYWCY CCRGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Cochlitoma dimidiata</i>	ACCCAGATCA TGTTTGA AAC CTTCAAYWCY CCAGCCATGT AYGTYGCCAT TCAAGCTGTG
<i>Cochlitoma sp. cf. vestita</i>	ACCCAGATCA TGTTTGA AAC CTTCAAYWCY CCRGCCATGT AYGTYGCIAT TCAAGCTGTG
<i>Cochlitoma marinae</i>	ACCCAGATCA TGTTTGA AAC CTTCAACACY CCAGCTATGT ATGTTGCTAT CCAGGCTGTG
<i>Cochlitoma churchilliana</i>	ACCCAGATCA TGTTTGA AAC CTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Cochlitoma granulata</i>	ACCCAGATCA TGTTTGA AAC TTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Cochlitoma simplex</i>	ACCCAGATCA TGTTTGA AAC CTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Cpchlitoloma kilburni</i>	ACCCAGATCA TGTTTGA AAC CTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Cochlitoma montistempli</i>	ACCCAGATCA TGTTTGA AAC TTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Cochlitoma omissa</i>	ACCCAGATCA TGTTTGA AAC TTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Cochlitoma semidecussata</i>	ACCCAGATCA TGTTTGA AAC TTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Cochlitoma ustulata</i>	ACCCAGATCA TGTTTGA AAC CTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCCGTG
<i>Cochlitoma zebra</i>	ACCCAGATCA TGTTTGA AAC CTTCAACTCT CCAGCCATGT ATGTCGCCAT TCAAGCTGTG
<i>Metachatina kraussi</i>	ACCCAGATCA TGTTTGA AAC CTTCAAYWCY CCAGCYATGT AYGTYGCIAT YCAAGCYGTG
<i>Rumina decollata</i>	ACACAGATCA TGTTTGA AAC CTTCAAYACC CCAGCHATGT AYGTGCIAT YCARGCCGTG

	250	260	270	280	290	300		
<i>Achatina fulica</i>	CTTTCTTTTRT	ATGCWTCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Achatina immaculata</i>	CTTTCTTTTRT	AYGCATCAGG	TCGTACAACCT	GGTATTGTGY	TGGATTCTGG	TGATGGTGTGTA		
<i>Achatina reticulata</i>	CTTTCCYYTRT	AYGCWTCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Achatina zanzibarica</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTAC	TGGATTCTGG	TGATGGTGTGTA		
<i>Euaethiopina loveridgei</i>	CTTTCTTTTAT	ACGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Achatina achatina</i>	CTTTCTTTTRT	ATGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Archachatina marginata</i>	CTTTCTTTTRT	ATGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Achatina bisculpta</i>	CTTTCTTTTRT	AYGCWTCAGG	TCGTACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTR		
<i>Achatina damarensis</i>	CTTTCTTTTRT	ATGCWTCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTR		
<i>Achatina stuhlmanni</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Atopocochlis exarata</i>	CTTTCTTTTRT	AYGCWTCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Limicolaria kambeul</i>	CTTTCCYYTRT	ATGCWTCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Limicolaria martenssii</i>	YTTTCYTTTRT	AYGCWTCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTR		
<i>Limicolariopsis sp.</i>	CTTTCTTTTRT	AYGCWTCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma varicosa</i>	CTTTCTTTTRT	ATGCATCAGG	TCGTACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma dimidiata</i>	CTTTCTTTTRT	ATGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma sp. cf. vestita</i>	CTTTCTTTTRT	ATGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGAYTCTGG	TGATGGTGTGTM		
<i>Cochlitoma marinae</i>	CTGTCACTGT	ATGCATCAGG	TCGTACAACCT	GGTATTGTTC	TTGACTCAGG	AGATGGTGTGTC		
<i>Cochlitoma churchilliana</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma granulata</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma simplex</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cpchlitoloma kilburni</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma montistempli</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma omissa</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma semidecussata</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma ustulata</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTA		
<i>Cochlitoma zebra</i>	CTTTCTTTTAT	ATGCATCAGG	TCGTACAACCT	GGTATTGTGC	TGGATTCTGG	TGATGGTGTGTA		
<i>Metachatina kraussi</i>	CTTTCTTTTRT	ATGCWTCAGG	TCGYACAACCT	GGTATTGTTC	TGGATTCTGG	TGATGGTGTGTV		
<i>Rumina decollata</i>	CTKTCYYTGT	ATGCCTCTGG	TCGTACAACCT	GGTATTGTGC	TAGATTCTGG	TGATGGTGTGTT		

	310	320	330	340	350	360	
<i>Achatina fulica</i>	ACCCACACTG	TCCCAATTTA	TGAAGGTTAT	GCTCTACCTC	ATGCCATCAT	GAGACTGGAC	
<i>Achatina immaculata</i>	ACCCACACTG	TCCCAATYTA	TGAAGGTTAT	GCTCTACCTC	ATGCCATCAT	GAGACTGGAC	
<i>Achatina reticulata</i>	ACCCACACTG	TCCCAATYTA	TGAAGGTTAT	GCTCTTCCCC	AYGCCATCAT	GAGACTGGAC	
<i>Achatina zanzibarica</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Euaethiopina loveridgei</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Achatina achatina</i>	ACYCAYACTG	THCCCAATMTA	TGARGGTTAT	GCTCTTCCYC	AYGCCATCAT	GAGACTGGAY	
<i>Archachatina marginata</i>	ACYCAYACTG	TCCCAATCTA	YGARGGTTAT	GCTCTTCCYC	ATGCCATYMT	GAGACTGGAC	
<i>Achatina bisculpta</i>	ACCCACACTG	TYCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTRGAY	
<i>Achatina damarensis</i>	ACCCACACTG	TYCCAATCTA	TGAAGGTTAT	GCTCTTCCYC	ATGCCATCAT	GAGACTGGAY	
<i>Achatina stuhlmanni</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Atopocochlis exarata</i>	ACYCAYACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCYC	ATGCCATCAT	GAGACTGGAC	
<i>Limicolaria kambeul</i>	ACYCAYACTG	TYCCAATCTA	TGAAGGTTAT	GCTCTTCCYC	ATGCCATCAT	GAGACTGGAC	
<i>Limicolaria martenssii</i>	ACCCACACTG	TYCCAATCTA	YGAAGGTTAT	GCTCTTCCYC	ATGCCATCAT	GAGACTKGAC	
<i>Limicolariopsis sp.</i>	ACCCACACTG	TYCCAATCTA	YGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma varicosa</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma dimidiata</i>	ACYCAYACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAY	
<i>Cochlitoma sp. cf. vestita</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAY	
<i>Cochlitoma marinae</i>	ACTCACACTG	TGCCCAATCTA	TGAAGGTTAT	GCCCTTCCCC	ATGCCATCAT	GAGACTGGAT	
<i>Cochlitoma churchilliana</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma granulata</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma simplex</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cpchlitoloma kilburni</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma montistempli</i>	ACCCATACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma omissa</i>	ACCCAYACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma semidecussata</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma ustulata</i>	ACTCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Cochlitoma zebra</i>	ACCCACACTG	TCCCAATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCAT	GAGACTGGAC	
<i>Metachatina kraussi</i>	ACCCACACTG	TCCCMATCTA	TGAAGGTTAT	GCTCTTCCCC	ATGCCATCMW	GAGACTGGAC	
<i>Rumina decollata</i>	ACCCAYACTG	TCCCAATCTA	TGAAGGTTAT	GCCCTTCCCT	ATGCCATCAT	GAGAYTGGAC	

	370	380	390	400	410	420	
<i>Achatina fulica</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Achatina immaculata</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Achatina reticulata</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Achatina zanzibarica</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Euaethiopina loveridgei</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAAATCC	TTACAGAGAG	AGGCTACAGC	
<i>Achatina achatina</i>	TTGGCTGGTC	GTGACCTCAC	AGATTATCTC	ATGAARATCC	TCACAGAGAG	AGGTTACAGC	
<i>Archachatina marginata</i>	TTGGCTGGWC	GTGACCTCAC	AGATTAYCTC	ATGAARATYC	TCACAGAGAG	AGGYTACAGY	
<i>Achatina bisculpta</i>	TTGGCTGGTC	GTGACCTYAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Achatina damarensis</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Achatina stuhlmanni</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Atopocochlis exarata</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Limicolaria kambeul</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGY	
<i>Limicolaria martenssii</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCY	TCACAGAGAG	AGGCTACAGY	
<i>Limicolariopsis sp.</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma varicoso</i>	TTGGCTGGTC	GTGACCTCAC	AGATTATCTS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma dimidiata</i>	TTGGCTGGTC	GTGACCTCAC	AGATTATCTS	ATGAAGATCC	TYACAGAGAG	AGGCTACAGC	
<i>Cochlitoma sp. cf. vestita</i>	TTGGCTGGTC	GTGACCTCAC	AGATTATCTS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma marinae</i>	CTGGCTGGTC	GTGACCTTAC	TGATTATTG	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma churchilliana</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma granulata</i>	TTGGCTGGTC	GTGACCTCAC	AGATTAYCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma simplex</i>	TTGGCTGGTC	GTGACCTCAC	AGATTATCTS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cpchlitoma kilburni</i>	TTGGCTGGTC	GTGACCTCAC	AGATTATCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma montistempli</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma omissa</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma semidecussata</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma ustulata</i>	TTGGCTGGTC	GTGACCTCAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Cochlitoma zebra</i>	TTGGCTGGTC	GTGACCTCAC	AGATTATCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Metachatina kraussi</i>	TTGGCTGGTC	GTGACCTYAC	AGATTAYCWS	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	
<i>Rumina decollata</i>	TTGGCTGGTC	GTGACCTYAC	AGATTACCTC	ATGAAGATCC	TCACAGAGAG	AGGCTACAGC	

	430	440	450	460	470	480	
<i>Achatina fulica</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Achatina immaculata</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Achatina reticulata</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAARCT	TTGCTATGTG	
<i>Achatina zanzibarica</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Euaethiopina loveridgei</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Achatina achatina</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGAYATCA	ARGAGAAGCT	GTGCTATGTG	
<i>Archachatina marginata</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AGGAGAAGCT	GTGCTATGTG	
<i>Achatina bisculpta</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG	
<i>Achatina damarensis</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG	
<i>Achatina stuhlmanni</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG	
<i>Atopocochlis exarata</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG	
<i>Limicolaria kambeul</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTATGTG	
<i>Limicolaria martenssii</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTAYGTG	
<i>Limicolariopsis sp.</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAACT	TTGCTAYGTG	
<i>Cochlitoma varicoso</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma dimidiata</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma sp. cf. vestita</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma marinae</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma churchilliana</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma granulata</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma simplex</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cpchlitoma kilburni</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma montistempli</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma omissa</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma semidecussata</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma ustulata</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Cochlitoma zebra</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGACATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Metachatina kraussi</i>	TTCACCACCA	CAGCTGAGAG	AGAAATTGTT	CGAGAYATCA	AAGAGAAGCT	TTGCTATGTG	
<i>Rumina decollata</i>	TTCACCACCA	CTGCTGAGAG	AGAAATTGTT	CGAGACATCA	AGGAGAAGCT	GTSTATGTG	

	490	500	510	520	530	540	
<i>Achatina fulica</i>	GCCCTTGACT	TTGAGCAAGA	RATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Achatina immaculata</i>	GCCCTTGACT	TTGAGCARGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Achatina reticulata</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCYACAT	CATCTTCATT	AGAGAAGAGT	
<i>Achatina zanzibarica</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTGCTACAT	CATCTTCATT	AGAGAAGAGT	
<i>Euaethiopina loveridgei</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTGCTACAT	CATCTTCATT	AGAGAAGAGT	
<i>Achatina achatina</i>	GCYCTTGACT	TTGAGCAAGA	GATGTCRACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGY	
<i>Archachatina marginata</i>	GCYCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Achatina bisculpta</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Achatina damarensis</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Achatina stuhlmanni</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTGCCACAT	CATCTTYATT	AGAGAAGAGT	
<i>Atopocochlis exarata</i>	GCYCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Limicolaria kambeul</i>	GCCCTTGAYT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGARAAGAGT	
<i>Limicolaria martenssii</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Limicolariopsis sp.</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTKCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma varicosa</i>	GCCCTTGACT	TTGARCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma dimidiata</i>	GCCCTTGACT	TTGARCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma sp. cf. vestita</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma marinae</i>	GCACTTGACT	TTGAGCAAGA	AATGGGTACA	GCAGCAACAT	CATCATCTTT	GGAGAAGAGC	
<i>Cochlitoma churchilliana</i>	GCCCTTGACT	TTGAACAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma granulata</i>	GCCCTTGACT	TTGAGCARGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma simplex</i>	GCCCTTGACT	TTGAACAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cpchlitoloma kilburni</i>	GCCCTTGACT	TTGAACAAGA	GATGGCAACA	GCTTCAACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma montistempli</i>	GCCCTTGACT	TTGAGCAGGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma omissa</i>	GCCCTTGACT	TTGAGCARGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma semidecussata</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCCACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma ustulata</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Cochlitoma zebra</i>	GCCCTTGACT	TTGAACAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Metachatina kraussi</i>	GCCCTTGACT	TTGAGCAAGA	GATGGCAACA	GCTTCCACAT	CATCTTCATT	AGAGAAGAGT	
<i>Rumina decollata</i>	GCTCTTGACT	TTGAACAAGA	AATGGCAACA	GCTGCCACTT	CATCTTCWCT	GGAGAAAAGT	

	550	560	570	580	590	600
<i>Achatina fulica</i>	TATGAATTTC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	ATGTCCAGAA
<i>Achatina immaculata</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAATG	AGCGTTTCAG	ATGTCCAGAA
<i>Achatina reticulata</i>	TATGAAYTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	RTGYCCAGAA
<i>Achatina zanzibarica</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGCAACG	AGCGTTTCAG	GTGCCCAGAA
<i>Euaethiopina loveridgei</i>	TATGAACTGC	CTGATGGACA	GGTCATCACC	ATTGGTAATG	AGCGTTTCAG	GTGCCCAGAA
<i>Achatina achatina</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA
<i>Archachatina marginata</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	RTGYCCAGAA
<i>Achatina bisculpta</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	ATGTCCAGAA
<i>Achatina damarensis</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	ATGTCCAGAA
<i>Achatina stuhlmanni</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGATTTCAG	GTGCCCAGAA
<i>Atopocochlis exarata</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	RTGTCCAGAA
<i>Limicolaria kambeul</i>	TATGAATTTC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	RTGYCCAGAA
<i>Limicolaria martenssii</i>	TATGAATTTC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	ATGTCCAGAA
<i>Limicolariopsis sp.</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	ATGYCCAGAA
<i>Cochlitoma varicosa</i>	TATGAATTTC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA
<i>Cochlitoma dimidiata</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA
<i>Cochlitoma sp. cf. vestita</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA
<i>Cochlitoma marinae</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGYTTTCAG	RTGYCCAGAA
<i>Cochlitoma churchilliana</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	GTGCCCAGAA
<i>Cochlitoma granulata</i>	TATGAATTTC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	GTGCCCAGAA
<i>Cochlitoma simplex</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	GTGCCCAGAA
<i>Cpchlitoloma kilburni</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	GTGCCCAGAA
<i>Cochlitoma montistempli</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	GTGCCCAGAA
<i>Cochlitoma omissa</i>	TATGAATTTC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	GTGCCCAGAA
<i>Cochlitoma semidecussata</i>	TATGAATTAC	CTGATGGACA	GGTCATCACC	ATTGGTAACG	AGCGTTTCAG	GTGCCCAGAA
<i>Cochlitoma ustulata</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAATG	AGCGTTTCAG	GTGCCCAGAA
<i>Cochlitoma zebra</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAATG	AGCGTTTCAG	GTGCCCAGAA
<i>Metachatina kraussi</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATTGGTAAYG	AGCGTTTCAG	ATGTCCAGAA
<i>Rumina decollata</i>	TATGAATTGC	CTGATGGACA	GGTCATCACC	ATCGGCAAYG	AGCGTTTCAG	ATGTCCAGAA

	610 620 630 640 650 660
<i>Achatina fulica</i>	GCCATGTTCC AGCCWCTTTT CCTTGGTATG GARWCCGCAG GTATTCATGA RACCACYTAC
<i>Achatina immaculata</i>	GCCATGTTCC AGCCATCTTT CCTTGGTATG GAAWCCGCAG GTATTCATGA AACCACCTAC
<i>Achatina reticulata</i>	GCMRWRTTTC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GYATTCATGA AACCACCTAC
<i>Achatina zanzibarica</i>	GCAGAATTTC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Euaethiopina loveridgei</i>	GCAGAATTTC AGCCCTCTTT CCTTGGTATG GAGTCTGCAG GCATTCATGA AACCACCTAC
<i>Achatina achatina</i>	GCCATGTTCC AGCCATCTTT CCTTGGTATG GARWCTGCAG GTATTCATGA AACCACCTAC
<i>Archachatina marginata</i>	GCCATGTTCC AGCCATCTTT CCTTGGTATG GARTCTGCAG GTATTCATGA GACCACCTAC
<i>Achatina bisculpta</i>	GCCATGTTCC AGCCATCTTT CCTTGGYATG GAGTCTGCAG GTATTCATGA GACCACSTAC
<i>Achatina damarensis</i>	GCCATGTTCC AGCCATCTTT YCTTGGTATG GARWCYGCAG GTATTCATGA GACCACCTAC
<i>Achatina stuhlmanni</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCACGA GACCACCTAC
<i>Atopocochlis exarata</i>	GCMATGTTCC AGCCATCTTT CCTTGGTATG GAGTCYGCAG GTATTCATGA GACCACCTAC
<i>Limicolaria kambeul</i>	GCCATGTTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCAYGA GACCACCTAY
<i>Limicolaria martenssii</i>	GCCATGTTCC AGCCMTCTTT YCTTGGTATG GAGTCHGCAG GTATTCAYGA GACCACCTAC
<i>Limicolariopsis sp.</i>	GCCATGTTCC AGCCATCTTT YCTTGGTATG GARWCWGCAG GTATTCAYGA GACCACCTAC
<i>Cochlitoma varicoso</i>	GCMRWRTTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma dimidiata</i>	GCMRWRTTCC AGCCWCTTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma sp. cf. vestita</i>	GCMRWRTTCC AGCCATCTTT CCTTGGYATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma marinae</i>	GCAATGTTCC AGCCATCTTT CCTTGGGATG GAGTCTGCTG GTAYTCATGA AACAAACATAC
<i>Cochlitoma churchilliana</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma granulata</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma simplex</i>	GCAGAATTCC AGCCTTCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cpchlitoma kilburni</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma montistempli</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma omisa</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma semidecussata</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma ustulata</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Cochlitoma zebra</i>	GCAGAATTCC AGCCATCTTT CCTTGGTATG GAGTCTGCAG GTATTCATGA GACCACCTAC
<i>Metachatina kraussi</i>	GCCATGTTCC AGCCATCTTT YCTTGGTATG GARWCYGCWAG GTATTCATGA RACCACCTAC
<i>Rumina decollata</i>	GCAATGTTCC AGCCATCTTT CCTTGGTATG GAGTCTGCTG GTATTCATGA GACCACCTAC

	670 680 690 700 710 720
<i>Achatina fulica</i>	AACTCCATCA TGAAGTGTGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Achatina immaculata</i>	AACTCCATCA TGAAGTYGGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Achatina reticulata</i>	AACTCCATCA TGAAGTYGGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Achatina zanzibarica</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Euaethiopina loveridgei</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Achatina achatina</i>	AACTCCATCA TGAAGTYGGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTA
<i>Archachatina marginata</i>	AACTCCATCA TGAAGTGTGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTA
<i>Achatina bisculpta</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTACTG
<i>Achatina damarensis</i>	AACTCCATCA TGAAGTYGGA YGTYGACATC CGTAAAGACT TGTATGCCAA CACTGTAYTG
<i>Achatina stuhlmanni</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTACTT
<i>Atopocochlis exarata</i>	AACTCCATCA TGAAGTYGGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTACTG
<i>Limicolaria kambeul</i>	AACTCCATCA TGAARTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Limicolaria martenssii</i>	AACTCCATCA TGAARTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTR
<i>Limicolariopsis sp.</i>	AACTCCATCA TGAAGTYGGA CGTCGACATC CGTAAAGACT TRTATKCCAA CACTGTMTTG
<i>Cochlitoma varicoso</i>	AACTCCATCA TGAAGTYGGA CGTYGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma dimidiata</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma sp. cf. vestita</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma marinae</i>	AACTCCATCA TGAAGTGCGA TGTTGATATC CGTAAAGACT TGTATGCCAA CACTGTCTTA
<i>Cochlitoma churchilliana</i>	AACTCCATCA TGAAGTYGGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma granulata</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA YACTGTATTG
<i>Cochlitoma simplex</i>	AACTCCATCA TGAAGTGCGA TGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cpchlitoma kilburni</i>	AACTCCATCA TGAAGTGCGA CGTTGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma montistempli</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma omisa</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma semidecussata</i>	AACTCCATCA TGAAGTGCGA CGTTGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma ustulata</i>	AACTCCATCA TGAAGTGCGA CGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Cochlitoma zebra</i>	AACTCCATCA TGAAGTGCGA TGTCGACATC CGTAAAGACT TGTATGCCAA CACTGTATTG
<i>Metachatina kraussi</i>	AACTCCATCA TGAARTGCGA YGTCGACATC CGTAAAGACT TGTATGCCAA CWCTGTATTG
<i>Rumina decollata</i>	AATTCCATCA TGAARTGTGA TGTYGACATC CGTAAAGACT TGTATGCCAA CACTGTCTTG

	730	740	750	760	770	780		
<i>Achatina fulica</i>	TCTGGTGGAW	CTACCATGTA	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATYGYCAAC		
<i>Achatina immaculata</i>	TCTGGTGGAA	CTACCATGTA	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATYGYCAAC		
<i>Achatina reticulata</i>	TCTGGTGGAW	CYACMATGTW	YCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Achatina zanzibarica</i>	TCTGGTGGAT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Euaethiopina loveridgei</i>	TCTGGTGGAT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Achatina achatina</i>	TCTGGTGGAW	CTACCATGTA	TCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACHAAY		
<i>Archachatina marginata</i>	TCTGGTGGAW	CYACCATGTA	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACYAAY		
<i>Achatina bisculpta</i>	TCTGGTGGMT	CYACMATGTW	YCCCGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAC		
<i>Achatina damarensis</i>	TCTGGTGGAT	CYACCATGTW	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACYAAY		
<i>Achatina stuhlmanni</i>	TCTGGTGGCT	CCACAATGTT	TCCAGGCATC	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Atopocochlis exarata</i>	TCTGGTGGAT	CCACAATGTT	CCMYGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACYAAY		
<i>Limicolaria kambeul</i>	TCTGGTGGAT	CYACCATGTA	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAC		
<i>Limicolaria martenssii</i>	TCTGGTGGAT	CTACYATGTA	TCCAGGCMTT	GCTGACCGCA	TGCAGAAGGA	AATCACCAC		
<i>Limicolariopsis sp.</i>	TCTGGTGGAT	CTACCATGTW	TCCAGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCRYAAY		
<i>Cochlitoma varicosa</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGY		
<i>Cochlitoma dimidiata</i>	TCTGGTGGMT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGY		
<i>Cochlitoma sp. cf. vestita</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGY		
<i>Cochlitoma marinae</i>	TCTGGTGGTT	CCACAATGTT	CCAGGCATC	GCTGACCGCA	TGCAGAAGGA	AATCACTGCT		
<i>Cochlitoma churchilliana</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Cochlitoma granulata</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Cochlitoma simplex</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Cpchlitoloma kilburni</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Cochlitoma montistempli</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Cochlitoma omissa</i>	TCTGGTGGCT	CCACAATGTT	CCCKGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Cochlitoma semidecussata</i>	TCTGGTGGCT	CCACAATGTT	CCCGGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Cochlitoma ustulata</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Cochlitoma zebra</i>	TCTGGTGGCT	CCACAATGTT	CCCTGGCATT	GCTGACCGCA	TGCAGAAGGA	AATCACCAGC		
<i>Metachatina kraussi</i>	TCTGGTGGAW	YACCATGTT	CCCTGGCATT	GCTGACCGYA	TGCAGAAGGA	AATYRYCTSY		
<i>Rumina decollata</i>	TCTGGTGGHT	CAACAATGTT	CCAGGCATC	GCTGACCGCA	TGCAGAAGGA	AATCACCAMT		

	790	800	810	820	830	840		
<i>Achatina fulica</i>	CTGGCTCCTT	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Achatina immaculata</i>	CTGGCTCCAT	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	GTACTCTGTC		
<i>Achatina reticulata</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Achatina zanzibarica</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Euaethiopina loveridgei</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Achatina achatina</i>	CTGGCTCCAS	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CAGAACGTAA	ATACTCTGTC		
<i>Archachatina marginata</i>	CTGGCTCCAC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CAGAACGTAA	ATACTCTGTC		
<i>Achatina bisculpta</i>	YTGCTCCWS	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC		
<i>Achatina damarensis</i>	CTGGCTCCTC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATWCTCTGTC		
<i>Achatina stuhlmanni</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC		
<i>Atopocochlis exarata</i>	CTGGCTCCTC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC		
<i>Limicolaria kambeul</i>	CTRGCTCCAC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC		
<i>Limicolaria martenssii</i>	CTRGCTCCAC	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC		
<i>Limicolariopsis sp.</i>	CTRGCTCCAC	CCACAATGAA	GATCAAGATH	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC		
<i>Cochlitoma varicosa</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTS		
<i>Cochlitoma dimidiata</i>	CTGGCTCCTS	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTG		
<i>Cochlitoma sp. cf. vestita</i>	CTGGCTCCTS	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTG		
<i>Cochlitoma marinae</i>	CTTGCTCCAC	CCACAATGAA	AATCAAGATT	ATTGCTCCAC	CAGAAAGAAA	ATACTCTGTT		
<i>Cochlitoma churchilliana</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTG		
<i>Cochlitoma granulata</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Cochlitoma simplex</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Cpchlitoloma kilburni</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Cochlitoma montistempli</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATT	ATTGCTCCAC	CTGAACGTAA	ATACTCTGTC		
<i>Cochlitoma omissa</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATT	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Cochlitoma semidecussata</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Cochlitoma ustulata</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Cochlitoma zebra</i>	CTGGCTCCTG	CCACAATGAA	GATCAAGATC	ATTGCTCCAC	CTGAGCGTAA	ATACTCTGTC		
<i>Metachatina kraussi</i>	CTGGCTCCAC	CCACAATGAA	GATCAAGATC	ATTGCTCCWC	CTGAGMGTAA	ATACTCTGTG		
<i>Rumina decollata</i>	CTGGCTCCAC	CCACAATGAA	GATCAAAATC	ATTGCTCCTC	CTGAGAGAAA	ATACTCTGTC		

	850	860	870		
<i>Achatina fulica</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Achatina immaculata</i>	TGGATYGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Achatina reticulata</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Achatina zanzibarica</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Euaethiopina loveridgei</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Achatina achatina</i>	TGGATTGGAG	GTTCCATCT	GGCTTCTCTG	TCC	
<i>Archachatina marginata</i>	TGGATTGGAG	GTTCCATCT	GGCTTCTCTG	TCC	
<i>Achatina bisculpta</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Achatina damarensis</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Achatina stuhlmanni</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Atopocochlis exarata</i>	TGGATTGGAG	GATCCATCCT	WGCCTCTCTG	TCC	
<i>Limicolaria kambeul</i>	TGGATTGGAG	GATCCATCCT	GGCTTCTCTG	TCC	
<i>Limicolaria martenssii</i>	TGGATTGGAG	GATCCATCCT	GGCTTCTCTG	TCC	
<i>Limicolariopsis sp.</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma varicosa</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma dimidiata</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma sp. cf. vestita</i>	TGGATTGGAG	GWTCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma marinae</i>	TGGATTGGAG	GCTCCATTT	GGCTTCTCTA	TCC	
<i>Cochlitoma churchilliana</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma granulata</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma simplex</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cpchlitoma kilburni</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma montistempli</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma omissa</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma semidecussata</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Cochlitoma ustulata</i>	TGGATTGGAG	GATCCATCCT	GGCTTCTCTG	TCC	
<i>Cochlitoma zebra</i>	TGGATTGGAG	GATCCATCCT	GGCCTCTCTG	TCC	
<i>Metachatina kraussi</i>	TGGATTGGAG	GATCCATCCT	GGCTYCTCTG	TCC	
<i>Rumina decollata</i>	TGGATTGGAG	GTTCCATCCT	GGCCTCTCTG	TCC	

Appendix 4.3: Alignment of the histone 3 gene for the Achatinidae and the subulinid outgroup *Rumina decollata*. Note that sequencing was unsuccessful for *Euaethiopina loveridgei* and *Limicolariopsis ruwenzoriensis*.

	10 20 30 40 50 60
<i>Achatina fulica</i>	TCGTAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Achatina immaculata</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Achatina reticulata</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CKGCGYAGAAA
<i>Achatina zanzibarica</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Achatina achatina</i>	TCGTAATCC ACCGGAGGCA AGGCTCCCCG CAAGCARCTG GCCACCAAGG CKGCCAGRAA
<i>Archachatina marginata</i>	TMGYAAATCN ACCGGAGGCA AGGCTCCCCG YMAGCAGCTG GCCCCCAAGG CAGCCAGRAA
<i>Achatina bisculpta</i>	TCGCAATCC ACCGGAGGTA AGGCTCCYCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Achatina damarensis</i>	TCGCAATCC ACCGGAGGTA AGGCTCCTCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Achatina stuhlmanni</i>	TCGCAATCC ACCGGAGGTA AGGCTCCTCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Atopocochlis exarata</i>	TCGCAATCC ACCGGAGGTA AGGCTCCTCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Limicolaria kambeul</i>	TCGCAATCC ACCGGAGGCA AGGCTCCTCG CAAACAGCTT GCCACTAAGG CGGCTAGAAA
<i>Limicolaria martenssii</i>	TCGCAATCC ACCGGAGGCA AGGCTCCTCG CAAACAGCTT GCCACYAAGG CGGCTAGAAA
<i>Limicolariopsis sp.</i>	TCGCAATCC ACCGGAGGTA AGGCTCCTCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Cochlitoma varicosa</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACMAAGG CGGCTAGGAA
<i>Cochlitoma dimidiata</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAARCARCTK GCCACCAAGG CRGCGYAGGAA
<i>Cochlitoma sp. cf. vestita</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CMAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma marinae</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma churchilliana</i>	TYGCAATCC MCCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma granulata</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGAAA
<i>Cochlitoma simplex</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma kilburni</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma montistempli</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma omissa</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma semidecussata</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma ustulata</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Cochlitoma zebra</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAACAGCTT GCCACCAAGG CGGCTAGGAA
<i>Metachatina kraussi</i>	TCGCAATCC ACCGGAGGTA AGGCTCCCCG CAAGCAGCTT GCCACGAAGG CGGCTAGAAA
<i>Rumina decollata</i>	TCGCAATCC ACAGGAGGAA AAGCTCCCCG CAAACAGCTT GCCACTAAGG CGGCTAGAAA

	70 80 90 100 110 120
<i>Achatina fulica</i>	GTCGGCCCCG GCCACAGGAG GTGTCAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Achatina immaculata</i>	GTCGGCCCCA GCCACAGGAG GTGTCAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Achatina reticulata</i>	GTCGGCCCCA GCCACAGGRG GTGTCAAGAA ACCCCAYMGA TACAGGCCCG GCACAGTCCG
<i>Achatina zanzibarica</i>	GTCGGCCCCA GCCACAGGGG GTGTCAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Achatina achatina</i>	RTCGGCCCCR GCCACMGGRG GTGTSAAGAA RCCCCATCGW TACAGGCCCG GCACAGTCCG
<i>Archachatina marginata</i>	RTCKGICYCCR SCCCCGGWGT GTGTCAAGAA RCCCCAYSgy TACAGGCCSG SSACCGTGGC
<i>Achatina bisculpta</i>	GTCGGCCCCA GCCACMGGAG GTGTGAAGAA GCCCCAYMGA TACAGGCCCG GCACAGTCCG
<i>Achatina damarensis</i>	GTCGGCCCCA GCCACAGGAG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Achatina stuhlmanni</i>	GTCGGCCCCA GCCACAGGAG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Atopocochlis exarata</i>	GTCTGCCCCA GCCACAGGAG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Limicolaria kambeul</i>	GTCGGCCCCA GCCACAGGAG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Limicolaria martenssii</i>	GTCGGCCCCA GCCACAGGAG GTGTGAARAA GCCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Limicolariopsis sp.</i>	ATCGGCCCCA GCTACAGGAG GTGTGAAGAA GCCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma varicosa</i>	GTCGGCCCCA GCCACMGGAG GWGTGAAGAA ACCCCACMGA TACMGGCCCG GMACMGTCCG
<i>Cochlitoma dimidiata</i>	RTCGGCCCCA GCCACMGGWGT GTGTGAAGAA ACCCCAYMGA TACAGGCCCG GCACMGTCCG
<i>Cochlitoma sp. cf. vestita</i>	GTCGGCCCCA GCCACAGGAG GKGKGAAGAA ACCCCACAGA TMCAGACCCG GCMCAGTCCG
<i>Cochlitoma marinae</i>	GTCGGCCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma churchilliana</i>	GTCGGCCCCA GCCACAGGAG GTGTGAARAA ACCCCACARA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma granulata</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma simplex</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma kilburni</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGACCCG GCACAGTCCG
<i>Cochlitoma montistempli</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma omissa</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma semidecussata</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma ustulata</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Cochlitoma zebra</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Metachatina kraussi</i>	GTCGGGCCCA GCCACAGGAG GTGTGAAGAA ACCCCACAGA TACAGGCCCG GCACAGTCCG
<i>Rumina decollata</i>	GTCGGGCCCG GCCACAGGAG GCGTGAAGAA ACCCCAYCGA TACAGGCCCG GCACGGTCCG

	130 140 150 160 170 180
<i>Achatina fulica</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACTGAGCTT CTCATCCGCA AACTGCCCTT
<i>Achatina immaculata</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Achatina reticulata</i>	WCTKMGAGAG ATCCGTCGTT ACCAGAAGAG CACKGAGCTK CTCATYCGCA ARCTGCCCTT
<i>Achatina zanzibarica</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Achatina achatina</i>	TCTKCGAGAR ATCCGTCGTT ACCAGAAGAG CACKGAGCTC CTCATCCGCA AGCTGCCCTT
<i>Archachatina marginata</i>	CCTSMGAGAR ATCCGTCGTT ACCAGAAGAG CASGGAGCTG CTCATYCGCA AGYTGCCCTT
<i>Achatina bisculpta</i>	WCTTCGAGAG ATCCGTCGTT ACCAGAAGAG YACGGAGCTT CTCATCCGCA ARCTGCCATT
<i>Achatina damarensis</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG TACGGAGCTT CTCATCCGCA AACTGCCATT
<i>Achatina stuhlmanni</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCATT
<i>Atopocochlis exarata</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCATT
<i>Limicolaria kambeul</i>	ACTTCGAGAG ATCCGTCGTT ACCAAAAAAG CACGGAGCTT CTCATCCGCA AACTGCCATT
<i>Limicolaria martenssii</i>	ACTTCGAGAG ATCCGTCGTT ACCARAARAG CACGGAGCTT CTCATCCGCA AACTGCCATT
<i>Limicolariopsis sp.</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma varicosa</i>	WCTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA ARCTGCCCTT
<i>Cochlitoma dimidiata</i>	WCTKCGAGAR ATCCGTCGTT ACCAGAAGAG CACWGAGCTT CTCATCCGCA ARCTGCCCTT
<i>Cochlitoma sp. cf. vestita</i>	ACTTCGAGAG ATCCGTCGTT ACCARAARAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma marinae</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAAAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma churchilliana</i>	ACTTCGAGAG ATCCGTCGTT ACCARAARAG CACGGAGCTT YTCATCCGCA AACTGCCCTT
<i>Cochlitoma granulata</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma simplex</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma kilburni</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma montistempli</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma omissa</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma semidecussata</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma ustulata</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Cochlitoma zebra</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Metachatina kraussi</i>	ACTTCGAGAG ATCCGTCGTT ACCAGAAGAG CACGGAGCTT CTCATCCGCA AACTGCCCTT
<i>Rumina decollata</i>	GCTTCGAGAG ATCCGACGTT ACCAGAAGAG CACAGAGCTT CTCATCCGCA AGCTGCCCTT

	190 200 210 220 230 240
<i>Achatina fulica</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTTAAGACC GATCTGCGTT TCCAGAGCTC
<i>Achatina immaculata</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTTAAGACC GATCTGCGTT TCCAGAGCTC
<i>Achatina reticulata</i>	CCAGCGGCTG GTSAGAGAGA TYGCCAGGA CTTYAAGACC GATCTGCGYT TCCAGAGCTC
<i>Achatina zanzibarica</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTTAAGACC GATCTGCGTT TCCAGAGCTC
<i>Achatina achatina</i>	CCAGCGCCTG GTCAGAGAAA TCGCCAGGA CTTCAAGACY GACCTGCGCT TCCAGAGCTC
<i>Archachatina marginata</i>	CCAGCGCYTG GTGAGAGARA TCGCCARGA CTTCAAGACH GAYCTGCGYT TCCAGAGCTC
<i>Achatina bisculpta</i>	CCAGCGGCTG GTCAGAGAGA TCGCMCAGGA CTTCAAGACY GATCTGCGYT TCCAGAGCTC
<i>Achatina damarensis</i>	CCAGCGGCTG GTCAGAGAGA TCGCACAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Achatina stuhlmanni</i>	CCAGCGGCTG GTCAGAGAGA TCGCACAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Atopocochlis exarata</i>	CCAGCGGCTG GTCAGAGAGA TCGCACAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Limicolaria kambeul</i>	CCAGCGGCTG GTCAGAGAGA TCGCACAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Limicolaria martenssii</i>	CCAGCGGCTG GTCAGAGAGA TCGCACAGGA CTTCAAGACC GATCTGCGTT TYCAGAGCTC
<i>Limicolariopsis sp.</i>	CCAGCGGCTG GTCAGAGAGA TCGCACAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma varicosa</i>	CCAGCGGYTG GTCAGAGAGA TCGCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma dimidiata</i>	CCAGCGGCTG GTSMGAGARA TCGCYCAGGA CTTCAAGACC GATCTGCGYT TCCAGAGCTC
<i>Cochlitoma sp. cf. vestita</i>	CCAGCGGCTG GTCARARARA TCGCCAGGA CTTCAAGACT GATYTGCGTT TCCAGAGCTC
<i>Cochlitoma marinae</i>	CCAGCGGCTT GTCAGAGAGA TCGCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma churchilliana</i>	CCAGCGGCTG GTCARAGAGA TCGCCAGGA CTTCAARACC GATYTGCGTT TCCARAGCTC
<i>Cochlitoma granulata</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma simplex</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma kilburni</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTCAAGACT GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma montistempli</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma omissa</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma semidecussata</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma ustulata</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Cochlitoma zebra</i>	CCAGCGGCTG GTCAGAGAGA TCGCCAGGA CTTCAAGACT GATCTGCGTT TCCAGAGCTC
<i>Metachatina kraussi</i>	CCAACGGCTG GTCAGAGAGA TCGCCAAGA CTTCAAGACC GATCTGCGTT TCCAGAGCTC
<i>Rumina decollata</i>	CCAGCGCCTT GTCAGAGAGA TCGCCAGGA TTTCAAGACT GACCTGCGTT TCCAGAGCTC

	250	260	270	280	290	300	
<i>Achatina fulica</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Achatina immaculata</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Achatina reticulata</i>	VGCTGTCATG	GCWCTGCAAG	AGGCYAGCGA	RGCMTACCTG	GTGGGTCTGT	TYGAGGACAC	
<i>Achatina zanzibarica</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Achatina achatina</i>	GGCYGTCATG	GCCCTGCAAG	AGGCYAGCGA	RGCTACCTG	GTGGGTCTGT	TYGAGGACAC	
<i>Archachatina marginata</i>	GGCTGTCATG	GCYCTGCARG	AGGCYAGCGA	GGCMTACCTG	GTGGGCCTGT	TYGAGGACAC	
<i>Achatina bisculpta</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Achatina damarensis</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Achatina stuhlmanni</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Atopocochlis exarata</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Limicolaria kambeul</i>	AGCTGTCATG	GCACTGCAAG	AAGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Limicolaria martenssii</i>	AGCTGTCATG	GCACTGCAAG	AAGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGARGACAC	
<i>Limicolariopsis sp.</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma varicosa</i>	GGCTGTCATG	GCACTGCAAG	AGGCYAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma dimidiata</i>	GGCTGTCATG	GCACTGCAAG	AGGCYAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma sp. cf. vestita</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma marinae</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma churchilliana</i>	GGCTGTCATG	GCACTGCAAR	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma granulata</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma simplex</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma kilburni</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma montistempli</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma omissa</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma semidecussata</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma ustulata</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Cochlitoma zebra</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTG	GTGGGTCTGT	TTGAGGACAC	
<i>Metachatina kraussi</i>	GGCTGTCATG	GCACTGCAAG	AGGCTAGCGA	GGCCTACCTA	GTGGGTCTGT	TTGAGGACAC	
<i>Rumina decollata</i>	RGCYGTTCATG	GCACTGCAGG	AGGCTAGCGA	GGCMTACYTR	GTGGGCCTGT	TTGAGGACAC	

	310	320	
<i>Achatina fulica</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Achatina immaculata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Achatina reticulata</i>	CAACCTGTGC	GCCATCCAYG	CCAARMGT
<i>Achatina zanzibarica</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Achatina achatina</i>	YAACYTGTGC	GCYATCCACG	CCAARCGT
<i>Archachatina marginata</i>	CMACCTKTGC	GCCATCCAYG	CCARVCGT
<i>Achatina bisculpta</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Achatina damarensis</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Achatina stuhlmanni</i>	CAACCTGTGC	GCCATYCACG	CCAAACGT
<i>Atopocochlis exarata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Limicolaria kambeul</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Limicolaria martenssii</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Limicolariopsis sp.</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma varicosa</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma dimidiata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma sp. cf. vestita</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma marinae</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma churchilliana</i>	CAACTTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma granulata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma simplex</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma kilburni</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma montistempli</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma omissa</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma semidecussata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma ustulata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Cochlitoma zebra</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Metachatina kraussi</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT
<i>Rumina decollata</i>	CAACCTGTGC	GCCATCCACG	CCAAACGT

Appendix 4.4: Alignment of the CO1 gene for the Achatinidae and the subulinid outgroup *Rumina decollata*. Note that sequencing was unsuccessful for *Euaethiopina loveridgei* and *Limicolariopsis ruwenzoriensis*.

	10	20	30	40	50	60	
Achatina fulica	TTTTTGGTGT	ATGATGTGGG	TTAGTTGGCA	CAGGCTTGTC	ACTCTTAATT	CGGTTAGAGC	
Achatina immaculata	TTTTTGGGRT	TTGATGTGGC	TTGGTAGGAA	CCGGCCTTTC	ATTATTAATT	CGATTGGAGC	
Achatina reticulata	TCCTCGGGGT	ATGATGTGGT	TTGGTTGGTA	CTGGTTTATC	ATTATTAATT	CGTCTAGAGC	
Achatina zanzibarica	TATTTGGTGT	TTGGTGCGGC	TTGGTGGGGA	CAGGCTTATC	ATTACTAATT	CGACTAGAGT	
Achatina achatina	TTTTTGGGGT	TTGATGTGGC	CTAGTAGGTA	CAGGCCTATC	ATTGTTAATT	CGATTAGAAT	
Archachatina marginata	TTTTTGGGGT	TTGATGTGGC	CTTGTGGGAA	CAGGCTTATC	TTTTTAATT	CGGTTAGAAT	
Achatina bisculpta	TTTTTGGTGT	TTGATGTGGT	TTGGTAGGAA	CTGGGCTATC	TTTACTAATT	CGGTTAGAGT	
Achatina damarensis	TTTTTGGTGT	GTGGTGTGGG	TTAGTCCGGA	CTGGGTTATC	ATTACTAATT	CGGTTAGAGT	
Achatina stuhlmanni	TTTTTGGGGT	CTGGTGTGGA	TTAGTTGGTA	CCGGCTTGTC	TTTACTTATC	CGGCTAGAAC	
Atopocochlis exarata	TCTTTGGGGT	TTGATGTGGG	CTAGTGGGTA	CTGGCTTATC	CTGGCTTATC	CGATTAGAAC	
Limicolaria kambeul	TTTTTGGAAAT	TTGATGTGGG	TTGGTTGGGA	CAGGCCTATC	GTTTTTAATT	CGTTTAGAAC	
Limicolaria martenssii	TCTTTGGAGT	ATGATGTGGT	TTAGTGGGGA	CAGGTTTATC	ATTACTGATT	CGTTTAGAGT	
Limicolariopsis sp.	TCTTAGGGGT	ATGATGTGGG	TTAGTGGGAA	CAGGCCTATC	ATTATTAATT	CGACTAGAAC	
Cochlitoma varicosa	TTTTCGGGGT	GTGATGTGGT	TTAGTTGGGA	CTGGGCTATC	TTTACTTATT	CGATTAGAGT	
Cochlitoma dimidiata	TTTTTGGTGT	TTGATGTGGT	TTGGTTGGGA	CCGGGCTATC	TCTACTAATC	CGTTTAGAGC	
Cochlitoma sp. cf. vestita	TTTTTGGTGT	GTGATGTGGT	TTGGTTGGGA	CTGGGCTATC	CTTACTAATC	CGTCTAGAGT	
Cochlitoma marinae	TCTTCGGGAC	CTGGTGTGGC	TTGGTGGGCA	CTGGGCTGTC	TTTACTAATC	CGTTTAGAGT	
Cochlitoma churchilliana	TTTTCGGGGT	CTGGTGTGGG	TTAGTGGGGA	CTGGTTTATC	CCTATTAATT	CGATTAGAGC	
Cochlitoma granulata	TCTTTGGATT	CTGATGCGGG	TTAGTTGGTA	CTGGGTTATC	GTTATTGATC	CGGTTAGAAC	
Cochlitoma simplex	TTTTTGGAGT	TTGATGTGGA	CTAGTGGGAA	CTGGGTTATC	CTTACTAATC	CGACTAGAAC	
Cochlitoma kilburni	TTTTTGGAGT	GTGATGTGGC	TTGGTTGGGA	CAGGACTATC	ACTGCTTATC	CGTTTAGAGT	
Cochlitoma montistempli	TTTTTGGGTT	TTGGTGCGGG	TTGGTTGGAA	CAGGGCTATC	ATTATTAATT	CGATTAGAGC	
Cochlitoma omissa	TTTTCGGGTT	CTGATGCGGG	TTAGTTGGTA	CTGGGTTATC	ATTATTGATC	CGGTTAGAGC	
Cochlitoma semidecussata	TTTTCGGGTT	TTGGTGCGGG	TTGGTTGGTA	CAGGGCTCTC	GTTATTAATT	CGATTAGAGC	
Cochlitoma ustulata	TTTTTGGGGT	ATGGTGTGGT	CTAGTGGGCA	CAGGCTTATC	TTTATTGATC	CGACTAGAGC	
Cochlitoma zebra	TTTTCGGAGT	GTGATGTGGA	CTGGTGGGGA	CWGRTTTATC	TTTAYTKATY	CGGYTAGAGT	
Metachatina kraussi	TTTTTGGGGT	ATGATGTGGC	TTAGTTGGCA	CAGGCCTATC	TCTTTTAATT	CGATTAGAAC	
Rumina decollata	TTTTTGGGGT	CTGATGCGGT	TTAGTTGGTA	CTGGTTTATC	TCTTCTTATT	CGTTTAGAAC	

	70	80	90	100	110	120	
Achatina fulica	TTGGAACAGT	GGGAACCTTA	ACTGATGATC	ACTTTTTTCAA	CGTGGTTGTA	ACTGCGCATG	
Achatina immaculata	TAGGGTCAGT	TGGGACTTTA	ACAGATGATC	ATTTTTTTTAA	TGTTGTAGTA	ACTGCGCATG	
Achatina reticulata	TCGGTTCGGT	GGGACGTTA	ACTGACGATC	ATTTTTTTTAA	TGTTGTAGTA	ACCGCACATG	
Achatina zanzibarica	TAGGGACTAC	TGGTACCCCT	ACAGATGACC	ATTTCTTTTAA	TGTTGTAGTT	ACTGCACATG	
Achatina achatina	TAGGGACAGC	TGGGTTTTTA	ACCGATGATC	ATTTCTTTTAA	TGTTGTAGTT	ACAGCCATG	
Archachatina marginata	TAGGGACAGC	AGGGGTATTA	ACTGATGATC	ACTTTTTTTAA	TGTGGTAGTA	ACTGCTCATG	
Achatina bisculpta	TAGGGAATGT	TGTTACTTTG	ATTGATGATC	ACTTTTTTCAA	CGTGGTGGTC	ACTGCTCATG	
Achatina damarensis	TAGGGAAGTAC	TGGTACCTTA	ACTGATGATC	ATTTTTTATA	TGTAGTAGTT	ACTGCTCATG	
Achatina stuhlmanni	TAGGGAGTAT	TGGAACACTG	ACTGACGATC	ATTTTTTTTAA	TGTTGTAGTT	ACTGCTCATG	
Atopocochlis exarata	TAGGAAATGT	AGGGACATTA	ACTGATGACC	ACTTTTACAA	TGTAATTGTG	ACTGCTCATG	
Limicolaria kambeul	TAGGGAACAT	TGGAACCTTA	ATTGATGATC	ATTTGTATAA	TGTAGTAGTT	ACCGCCACG	
Limicolaria martenssii	TAGGTAATGT	GGGTACCTTG	ATAGATGATC	ATTTTTTATA	TGTTATTGTA	ACAGCTCAGC	
Limicolariopsis sp.	TTGGGAACAC	AGGAACCTTA	ACTGATGATC	ATTTTTTTTAA	TGTAGTAGTA	ACTGCTCATG	
Cochlitoma varicosa	TAGGTACTAC	CTCTACCTTA	ACAGACGATC	ATTTCTTTTAA	TGTTATTGTA	ACAGCACATG	
Cochlitoma dimidiata	TCGGGACAAC	CTCTACATTA	ACAGATGACC	ATTTCTACAA	TGTAATTGTA	ACAGCACATG	
Cochlitoma sp. cf. vestita	TAGGTACGAC	CTCTACACTA	ACTGATGATC	ATTTTTTCAA	TGTAATTGTG	ACGGCGCACG	
Cochlitoma marinae	TAGGTACCAC	ATCCACACTT	ACCGATGATC	ATTTCTACAA	TGTAATTGTG	ACAGCACATG	
Cochlitoma churchilliana	TAGGGACTGT	GTCTACTTTG	ACAGATGACC	ATTTTTTTTAA	TGTCATTGTG	ACTGCACATG	
Cochlitoma granulata	TGGGGACTAC	TTCCGACTTTA	ACAGATGATC	ATTTTTTTTAA	TGTTATTGTT	ACAGCCATG	
Cochlitoma simplex	TGGGGACTAC	CACTACTCTA	ACCGATGATC	ATTTTTTACAA	CGTGATCGTT	ACTGCACATG	
Cochlitoma kilburni	TAGGTACGAC	CTCTACACTA	ACTGATGATC	ATTTCTTCAA	TGTTATTGTA	ACGGCTCAGC	
Cochlitoma montistempli	TAGGGACAAC	AACACTCTTA	ACAGATGATC	ATTTTTTTTAA	TGTTATTGTA	ACGGCACAGC	
Cochlitoma omissa	TGGGGACTAC	CTCGACTCTA	ACAGATGATC	ATTTTTTTTAA	TGTTATTGTT	ACAGCTCATG	
Cochlitoma semidecussata	TAGGGACAAC	GTCTACACTA	ACGGATGATC	ATTTTTTTTAA	CGTTATTGTA	ACGGCACAGC	
Cochlitoma ustulata	TAGGAACGAC	CTCTACTTTA	ACTGATGATC	ATTTCTTTTAA	TGTTATCGTG	ACAGCCACG	
Cochlitoma zebra	TAGGWACTAC	CTCYACTTTA	ACKGATGATC	ATTTCTTTTAA	TGTTATTGTR	ACAGCTCAGC	
Metachatina kraussi	TAGGGATAAC	CTCTACTTTA	ACAGATGATC	ATTTTTTTTAA	TGTTGTAGTT	ACTGCTCAGC	
Rumina decollata	TAGGTACAGC	AGGAGTGTTA	ACAGATGACC	ATTTTTTTTAA	TGTGGTAGTT	ACTGCCCATG	

	130	140	150	160	170	180	
<i>Achatina fulica</i>	CTTTTGTGTCAT	AATTTTTTTTT	ATGGTTATAC	CAATTATAAT	TGGCGGATTT	GGAAACTGGA	
<i>Achatina immaculata</i>	CGTTTGTGTTAT	AATTTTCTTT	ATGGTTATGC	CAATTATGAT	TGGGGGATTT	GGTAATTGAA	
<i>Achatina reticulata</i>	CTTTTGTGTTAT	AATTTTTTTTT	ATAGTTATAC	CAATTATAAT	CGGAGGATTT	GGGAACTGAA	
<i>Achatina zanzibarica</i>	CATTTGTGTTAT	AATTTTTTTTT	ATAGTGATAC	CAATTATGAT	TGGGGGGTTT	GGGAACTGAA	
<i>Achatina achatina</i>	CTTTTGTGTTAT	AATTTTTTTTT	ATAGTCATAC	CAATTATGAT	TGGGGGGTTT	GGTAATTGAA	
<i>Archachatina marginata</i>	CATTTGTGTAAT	AATTTTTTTTT	ATGGTTATGC	CAATTATAAT	TGGGGGGTTT	GGAAACTGAA	
<i>Achatina bisculpta</i>	CTTTTGTGTAAT	GATTTTCTTT	ATAGTAATGC	CTATTATAAT	TGGGGGGTTT	GGTAATTGAA	
<i>Achatina damarensis</i>	CTTTTGTGTAAT	AATCTTCTTC	ATAGTTATAC	CTATTATAAT	TGGAGGATTT	GGAAACTGAA	
<i>Achatina stuhlmanni</i>	CTTTTGTGAT	AATTTTTTTTT	ATAGTGATAC	CAATTATAAT	TGGTGGGTTT	GGAAACTGGA	
<i>Atopocochlis exarata</i>	CTTTCGTAAT	AATTTTCTTT	ATAGTTATAC	CAATCATAAT	TGGCGGGTTT	GGGAACTGAA	
<i>Limicolaria kambeul</i>	CTTTTGTGTAAT	AATCTTCTTC	ATAGTTATAC	CTATTATAAT	TGGAGGTTTC	GGAAATTGGA	
<i>Limicolaria martenssii</i>	CTTTTGTGTTAT	AATTTTCTTT	ATAGTTATAC	CCATTATAAT	TGGAGGATTT	GGAAATTGAA	
<i>Limicolariopsis sp.</i>	CCTTCGTAAT	AATTTTTTTTT	ATAGTTATAC	CTATTATAAT	TGGTGGATTT	GGTAATTGAA	
<i>Cochlitoma varicosa</i>	CATTTGTGTAAT	AATTTTTTTTT	ATAGTGATAC	CCATTATAAT	TGGTGGTTTT	GGGAACTGGA	
<i>Cochlitoma dimidiata</i>	CTTTCGTTAT	AATTTTCTTC	ATAGTAATAC	CCATTATAAT	TGGGGGGTTT	GGGAACTGAA	
<i>Cochlitoma sp. cf. vestita</i>	CATTTCGTTAT	AATTTTTTTTT	ATGGTAATAC	CAATTATAAT	TGGGGGGTTT	GGTAATTGAA	
<i>Cochlitoma marinae</i>	CATTTGTGAT	GATTTTTTTC	ATAGTAATAC	CAATTATGAT	TGGGGGGTTT	GGGAACTGGA	
<i>Cochlitoma churchilliana</i>	CGTTTGTGTCAT	GATCTTCTTC	ATGGTAATGC	CTATTATAAT	TGGTGGTTTT	GGAAATTGAA	
<i>Cochlitoma granulata</i>	CATTTCGTTAT	AATTTTTTTC	ATAGTAATAC	CAATTATAAT	TGGGGGGTTT	GGTAACTGAA	
<i>Cochlitoma simplex</i>	CTTTTGTGTTAT	GATTTTTTTC	ATGGTGATGC	CCATTATAAT	CGGGGGCTTT	GGAAATTGGA	
<i>Cochlitoma kilburni</i>	CATTTCGTTAT	AATTTTCTTC	ATAGTAATAC	CTATTATAAT	TGGGGGGTTT	GGCAACTGGA	
<i>Cochlitoma montistempli</i>	CGTTTGTGTTAT	AATTTTCTTT	ATAGTAATAC	CTATTATAAT	TGGGGGGTTT	GGGAACTGGA	
<i>Cochlitoma omissa</i>	CATTTCGTTAT	AATTTTTTTTT	ATAGTAATAC	CTATTATAAT	CGGGGGTTT	GGTAACTGAA	
<i>Cochlitoma semidecussata</i>	CATTTGTGTTAT	AATTTTTTTTT	ATAGTAATAC	CTATTATAAT	TGGGGGGTTT	GGGAATTGAA	
<i>Cochlitoma ustulata</i>	CTTTTGTGTAAT	AATTTTCTTC	ATAGTAATAC	CCATTATAAT	TGGGGGGTTT	GGTAATTGAA	
<i>Cochlitoma zebra</i>	CTTTTGTGTTAT	AATCTTCTTC	ATGGTAATAC	CCATTATAAT	TGGGGGATTC	GGAAATTGRA	
<i>Metachatina kraussi</i>	CATTTGTGTTAT	GATTTTTTTTT	ATAGTTATGC	CTATTATAAT	TGGTGGATTT	GGAAATTGGA	
<i>Rumina decollata</i>	CATTTGTGAT	AATTTTTTTTT	ATAGTAATGC	CTATTATAAT	CGGTGGTTTT	GGAAATTGAA	

	190	200	210	220	230	240	
<i>Achatina fulica</i>	TGGTCCCAAT	ACTTATTGGT	GTCCTGATA	TAAGATTTC	ACGAATAAAT	AATATAAGGT	
<i>Achatina immaculata</i>	TGGTTCCCT	TTTAATTGGG	GTCCTGATA	TAAGTTTTC	CCGGATGAAT	AATATGAGGT	
<i>Achatina reticulata</i>	TGGTTCCCAT	ACTTATTGGG	GTCCTGATA	TAAGTTTTC	ACGTATAAAC	AATATAAGAT	
<i>Achatina zanzibarica</i>	TGGTACCCT	ATTAATTGGA	GCACCTGACA	TAAGATTCC	ACGCATAAAT	AATATAAGAT	
<i>Achatina achatina</i>	TAGTCCCACT	ATTAATTGGG	GTCCTGATA	TAAGATTCC	TCCGATGAAT	AATATAAGGT	
<i>Archachatina marginata</i>	TAGTCCCTCT	ATTAATTGGG	GTCCTGATA	TAAGTTTTC	TCCGATAAAT	AATATAAGGT	
<i>Achatina bisculpta</i>	TGGTCCCACT	TTTAATTGGT	GCACCAGATA	TAAGATTTC	TCCAATAAAT	AATATAAGAT	
<i>Achatina damarensis</i>	TGGTGCCTCT	TTTGATTGGT	GCACCAGACA	TAAGATTTC	TCCGATAAAC	AACATAAGGT	
<i>Achatina stuhlmanni</i>	TGGTCCCAT	ACTGATTGGT	GTCCTGATA	TAAGTTTTC	GCCGATAAAT	AACATAAGAT	
<i>Atopocochlis exarata</i>	TAGTCCCAT	ATTAATTGGG	GCCCCGACA	TAAGTTTTC	TCCAATAAAT	AACATAAGAT	
<i>Limicolaria kambeul</i>	TAGTCCCTT	ATTAATTGGT	GTCCTGATA	TAAGATTCC	CCGAATGAAT	AATATAAGAT	
<i>Limicolaria martenssii</i>	TAGTACCATT	ATTGATTGGT	GCACCAGATA	TAAGCTTTC	TCCGATGAAT	AATATAAGAT	
<i>Limicolariopsis sp.</i>	TAGTACCCT	GTTAATTGGG	GCACCTGACA	TAAGATTTC	TCCGATAAAC	AATATAAGGT	
<i>Cochlitoma varicosa</i>	TAGTCCCTT	GCTAATTGGT	GCCCCGATA	TAAGCTTTC	CCGAATGAAT	AATATAAGTT	
<i>Cochlitoma dimidiata</i>	TAGTCCCTT	ATTGATTGGG	GCACCAGATA	TGAGATTTC	TCCGATAAAC	AATATAAGCT	
<i>Cochlitoma sp. cf. vestita</i>	TGGTCCCTT	ACTGATCGGG	GCCCCGATA	TAAGTTTTC	CCGAATGAAT	AATATAAGTT	
<i>Cochlitoma marinae</i>	TAGTACCCTT	ATTGTTGGT	GCCCCGATA	TAAGATTTC	TCCAATAAAT	AATATAAGTT	
<i>Cochlitoma churchilliana</i>	TGATTCCCT	ACTAATTGGG	GTCCTGATA	TAAGTTTTC	GCGAATAAAC	AACATAAGGT	
<i>Cochlitoma granulata</i>	TAGTACCAT	GTTAATCGGG	GCCCCGATA	TAAGTTTTC	TCCGATAAAT	AATATAAGCT	
<i>Cochlitoma simplex</i>	TAGTCCCTT	GCTAATTGGG	GCACCTGACA	TGAGATTTC	TCCGATAAAC	AATATAAGCT	
<i>Cochlitoma kilburni</i>	TGGTCCCTT	GTTGATTGGG	GTCCTGATA	TAAGTTTTC	CCGAATGAAT	AATATAAGCT	
<i>Cochlitoma montistempli</i>	TAGTACCATT	GTTAATCGGG	GCCCCGATA	TAAGTTTTC	TCCGATAAAT	AATATAAGCT	
<i>Cochlitoma omissa</i>	TAGTCCCAT	GTTAATCGGG	GCCCCGATA	TAAGATTTC	TCCGATAAAT	AATATAAGCT	
<i>Cochlitoma semidecussata</i>	TAGTCCCAT	GTTAATCGGG	GCCCCGATA	TAAGATTTC	TCCGATAAAT	AATATAAGCT	
<i>Cochlitoma ustulata</i>	TAACTCCCT	TTTAATCGGG	GTCCTGATA	TAAGCTTTC	TCCGATAAAT	AACATGAGCT	
<i>Cochlitoma zebra</i>	TAATYCCCT	ACTTATTGGG	GTCCTGATA	TAAGCTTTC	CCGGATAAAT	AACATRAGYT	
<i>Metachatina kraussi</i>	TAGTCCCT	ATTAATTGGG	GTCCTGATA	TAAGCTTTC	ACGAATAAAT	AATATAAGCT	
<i>Rumina decollata</i>	TAGTCCCT	ATTAATTGGG	GCACCAGATA	TAAGTTTTC	TCCAATAAAT	AATATGAGAT	

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250 260 270 280 290 300
Achatina fulica TTTGACTTTT ACCACCTTCA TTTACTTTGT TAATCTGTTT AAGTATAGTG GAAGGAGGGG
Achatina immaculata TTTGGCTTCT CCCACCATCG TTTACTTTAT TAATTAGGTC AAGCCTGGTT GAAGGTGGTG
Achatina reticulata TTTGACTTTT ACCCCCTGCG TTTACATTAT TAATCTGCTC AAGTATAGTC GAAGGTGGGG
Achatina zanzibarica TTTGACTTCT ACCACCTTCT TTTTGTCTTT TAATTAGATC GAGAATGGTA GAGGGGGGGG
Achatina achatina TCTGATTATT ACCACCTTCA TTTACCTTAC TTATTCTTTC TAGTATAGTT GAGGGAGGAG
Archachatina marginata TTTGATTGCT TCCACCTTCA TTTGTCTGTC TAATCTCTC GAGGATGGTC GAGGGTGGAG
Achatina bisculpta TCTGACTTTT ACCCCCATCT TTTACTCTTT TAATCCTATC GAGTATAGTT GAAGGTGGAG
Achatina damarensis TTTGACTTTT ACCACCTTCT TTTACTTTAC TAATTTTATC AAGTATGGTT GAAGGTGGAG
Achatina stuhlmanni TTTGACTACT TCCCCATCA TTTTCTTTC TAATCTCTC AAGTATAGTA GAAGGTGGGG
Atopocochlis exarata TTTGGTTACT TCCACCATCA TTTACCCTTT TAATTTTATC TAGAATAGTA GAAGGAGGGG
Limicolaria kambeul TTTGATTACT ACCACCTTCT TTTACTTTAC TAATCTTATC AAGCATAGTA GAGGGTGGGG
Limicolaria martenssii TCTGGTTACT ACCCCCAGCT TTTACTTTAT TAATCTCTC TAGGATAGTA GAAGGAGGCG
Limicolariopsis sp. TCTGATTACT TCCTCCTGCA TTTACTTTAC TAATTTTATC AAGGATAGTA GAAGGAGGCG
Cochlitoma varicosa TCTGACTATT ACCACCTTCA TTTCTCCCTGT TAATCTATC AAGGATAGTT GAGGGGGGTG
Cochlitoma dimidiata TTTGATTGCT CCCACCTTCA TTTTCTCTTT TGATTTTATC AAGAATAGTG GAAGGGGGAG
Cochlitoma sp. cf. vestita TTTGATTACT TCCACCCTCG TTTTCTCTAT TAATTTTATC AAGAATGGTA GAAGGTGGTG
Cochlitoma marinae TCTGATTATT ACCACCCTCA TTCTCACTAC TAATCTTATC TAGTATAGTA GAAGGGGGAG
Cochlitoma churchilliana TCTGATTACT TCCCCCTCT TTTTCTGTTAC TGATTTTATC TAGTATAGTG GAAAGTGGTG
Cochlitoma granulata TTTGACTCCT ACCCCCCTCA TTCTCTCTC TAATCTTATC AAGAATGGTT GAGGGTGGTG
Cochlitoma simplex TCTGACTGCT TCCTCCTTCA TTTTCTCTTC TAATCTTATC AAGACTAGTG GAGGGTGGGG
Cochlitoma kilburni TTTGATTACT CCCACCTTCA TTTTCACTTC TAATCTTATC AAGAATGGTA GAGGGTGGTG
Cochlitoma montistempli TCTGGCTCCT ACCCCCCTCT TTTTCCCTCT TAATTTTATC AAGAATAGTA GAGGGCGGAG
Cochlitoma omissa TTTGACTCCT GCCCCCCTCG TTCTCTCTTT TAATCTTATC TAGAATGGTT GAGGGTGGTG
Cochlitoma semidecussata TCTGGCTCCT ACCCCCCTCG TTTTCCCTCT TAATTTTATC AAGAATAGTA GAGGGTGGGG
Cochlitoma ustulata TTTGATTGTT ACCCCCTTCC TTTTCACTTC TTATCTTGTG AAGAATAGTG GAGGGGGGTG
Cochlitoma zebra TTTGATTACT CCCMCCTTCA TTCTCTCTAC TAATTTTATC AAGRATAGTA GAGGGAGGTG
Metachatina kraussi TTTGGTTATT ACCGCCGTCT TTTACATTAT TGATTTTGTG AAGAATAGTA GAAGGAGGTG
Rumina decollata TTTGACTACT TCCTCCTTCT TTTATTTTAT TAATTATATC TAGGATAGTA GAAGGTGGGG

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310 320 330 340 350 360
Achatina fulica CTGGAAGTGG GTGGACTGTG TACCCGCCCT TAAGTCTCTG CTTAGGACAC AGAGGGGCTT
Achatina immaculata CTGGGACTGG TTGAAGTGTG TACCCGCCGT TAAGTCTCTA TATGGGTCAC AGGGGGGCCCT
Achatina reticulata CTGGCACAGG TTGAAGTGTG TACCCACCTT TAAGCTCTTA TTTAGGGCAT AGTGGGGCAT
Achatina zanzibarica CAGGTACAGG TTGGACTGTG TACCCACCAT TGAGCTCAAT TATTGGGCAT AGGGGCCCAT
Achatina achatina CTGGTACTGG GTGAACAGTT TATCCACCTT TAAGATCTAC TTTAGGTCAT AGAGGTCCCT
Archachatina marginata CAGGTACTGG GTGGACAGTT TACCCCCCTT TTAGGTCAGT CTTAGGACAT AGTGGAGCTT
Achatina bisculpta CGGGAAGTGG CTGAACAGTT TATCCACCAC TTAGTTCATT AATTGGACAT AGTGGGACAT
Achatina damarensis CAGGAAGTGG TTGACAGTT TACCCACCTC TTAGTCTCTT GATTGGTCAT AGGGGAGCAT
Achatina stuhlmanni CTGGGACAGG ATGGACAGTT TACCCTCCTC TAAGTTCATT AATTGGCCAT AGTGGAGCAT
Atopocochlis exarata CAGGGACTGG TTGAACGGTG TACCCCCCTC TTAGCTCTAT GCTTGGCCAC AGGGGTGCCCT
Limicolaria kambeul CTGGAAGTGG TTGAACAGTT TACCCTCCTC TTAGAGCTAT TACAGGCCAT AGAGGTGCTT
Limicolaria martenssii CGGGAACAGG GTGAAGTGTG TATCCTCCTT TAAGATCCCT AGTGGGTCAT AGAGGGGCCA
Limicolariopsis sp. CAGGAAGTGG TTGAAGTGTG TACCCCCCTC TAAGTCTAT AATTGGACAT AGAGGAGCCT
Cochlitoma varicosa CAGGGACTGG TTGAACAGTT TACCCCCCTC TGAGTCTAT AATAGGACAT AGAGGAGCGT
Cochlitoma dimidiata CAGGGACAGG CTGGACGGTA TACCCGCCCT TAAGTTCGAT TATAGGTCAC AGAGGGGCTT
Cochlitoma sp. cf. vestita CAGGTACTGG GTGAAGTGTG TACCCACCTC TCAGGCTAT GTTAGGGCAT AGAGGTGCGT
Cochlitoma marinae CAGGCACCGG TTGGACTGTG TACCCGCCCT TAAGCTCTAT TATTGGTCAT AGGGGCGCTT
Cochlitoma churchilliana CTGGGACAGG GTGAACAGTA TACCCTCCCC TGAGGTCAAC TATTGGTCAT AGGGGCGCAT
Cochlitoma granulata CAGGTACAGG TTGAAGTGTG TACCCTCCTT TGAGTCTGT AATAGGCCAT AGAGGTGCAT
Cochlitoma simplex CAGGAACAGG CTGAACAGTA TACCCTCCCC TTAGTCTAT TATTGGTCAC AGAGGAGCTT
Cochlitoma kilburni CAGGTACTGG TTGAAGTGTG TACCCACCTC TTAGATCTAT ATTAGGGCAT AGGGGTGCAT
Cochlitoma montistempli CAGGTACTGG TTGAACAGTT TACCCCCCTT TAAGTCAAT TATTGGCCAT AGAGGTGCAT
Cochlitoma omissa CAGGTACTGG TTGAACAGTT TACCCCCCTT TGAGCTCTAT AATAGGTCAT AGAGGTGCAT
Cochlitoma semidecussata CAGGTACTGG TTGAACGGTT TACCCTCCTT TAAGTCAAT TATTGGCCAT AGAGGAGCAT
Cochlitoma ustulata CAGGTACAGG TTGAAGTGTG TATCCACCTT TAAGTCTAT CATAGGTCAT AGGGGAGCAT
Cochlitoma zebra CWGGTACWGG CTGRACAGTG TATCCCCCTT TAAGTTCAT GCTWGGACAT AGRGGWGCAT
Metachatina kraussi CCGGGACTGG TTGAAGTGTG TACCCACCGT TAAGTCTAT TCTTGGGCAT AGGGGAGCAT
Rumina decollata CAGGAAGTGG TTGAACAGTG TATCCACCTT TAAGCAGAAT CATAGGACAT AGTGGGGCAT

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	370	380	390	400	410	420	
<i>Achatina fulica</i>	CAGTTGATTT	AGCTATTTTT	TCTTTACATT	TGGCTGGTGT	ATCCTCAATT	TTAGGGGCAA	
<i>Achatina immaculata</i>	CAGTTGATTT	AGCAATTTTT	TCCCTTCATC	TTGCTGGTGT	CTCATCGATT	TTAGGGGCAA	
<i>Achatina reticulata</i>	CAGTAGATTT	AGCTATCTTC	TCTCTCCACT	TAGCTGGGGT	ATCCTCAATC	TTAGGGGGCTA	
<i>Achatina zanzibarica</i>	CTGTTGATTT	AGCGATTTTT	TCACTTCATT	TGGCAGGAGT	TTCGTCAATT	TTAGGGGCAA	
<i>Achatina achatina</i>	CGGTAGATTT	AGCGATTTTT	TCATTACATT	TGGCTGGTAT	ATCATCTATC	TTAGGGGGCTA	
<i>Archachatina marginata</i>	CTGTGGACCT	GGCTATTTTT	TCTTTGCACT	TAGCCGGTAT	ATCCTCTAAT	CTTGGTGCAA	
<i>Achatina bisculpta</i>	CAGTGGATTT	AGCAATTTTT	TCTTTACACT	TAGCTGGTAT	ATCCTCAATC	CTTGGAGCCA	
<i>Achatina damarensis</i>	CTGTAGATTT	AGCAATTTTT	TCTTTACATC	TAGCTGGGAT	ATCCTCAATC	CTTGGTGCTA	
<i>Achatina stuhlmanni</i>	CAGTGGACTT	GGCAATTTTT	TCTCTACACT	TAGCCGGAAT	ATCCTCTAAT	CTCGGAGCTA	
<i>Atopocochlis exarata</i>	CGGTTGACTT	AGCAATTTTT	TCACTTCATC	TAGCAGGTAT	ATCCTCGATT	CTTGGGGCTA	
<i>Limicolaria kambeul</i>	CAGTAGACTT	AGCAATCTTC	TCTCTTCACC	TTGCAGGAAT	GTCTTCAATC	CTAGGAGCAA	
<i>Limicolaria martenssii</i>	GGGTTGACTT	AGCTATTTTT	TCCTTACACT	TAGCTGGAAT	ATCTTCCATT	CTAGGGGCTA	
<i>Limicolariopsis sp.</i>	CAGTAGATTT	AGCTATTTTT	TCATTACATC	TAGCAGGTAT	GTCTTCAATC	TTAGGGGGCTA	
<i>Cochlitoma varicosa</i>	CTGTAGATCT	GGCTATCTTC	TCTCTTCATC	TTGCTGGGAT	GTCTTCAATC	TTAGGGGGCTA	
<i>Cochlitoma dimidiata</i>	CTGTTGATTT	GGCCATTTTT	TCCCTTACATC	TGGCAGGAAT	ATCGTCAATT	TTAGGTGCTA	
<i>Cochlitoma sp. cf. vestita</i>	CTGTGATTT	AGCTATCTTC	TCACTACATC	TTGCAGGGAT	ATCCTCGATT	TTAGGAGCCA	
<i>Cochlitoma marinae</i>	CAGTGGATCT	AGCTATTTTT	TCTCTTCACC	TAGCTGGGAT	ATCCTCTAAT	TTAGGTGCTA	
<i>Cochlitoma churchilliana</i>	CAGTGGATTT	AGCAATCTTC	TCCCTTCACT	TGGCGGGGAT	ATCCTCAATC	CTAGGTGCTA	
<i>Cochlitoma granulata</i>	CAGTAGATTT	AGCTATTTTT	TCCCTTCACC	TCGCAGGTCT	GTCTTCAATC	TTGGGTGCTA	
<i>Cochlitoma simplex</i>	CTGTAGATCT	GGCTATTTTT	TCCCTTACATC	TAGCAGGAAT	GTCTTCAATC	TTAGGGGGCTA	
<i>Cochlitoma kilburni</i>	CTGTGATTT	AGCAATCTTC	TCACTACATC	TTGCAGGGAT	ATCCTCAATT	CTAGGTGCTA	
<i>Cochlitoma montistempli</i>	CTGTAGATCT	GGCTATTTTT	TCGCTACATC	TTGCGGGTAT	ATCCTCAATT	CTAGGTGCTA	
<i>Cochlitoma omissa</i>	CTGTGGATTT	AGCTATTTTT	TCTCTTCACC	TTGCGGGTCT	GTCTTCAATC	CTAGGTGCTA	
<i>Cochlitoma semidecussata</i>	CGGTTGACCT	TGCAATCTTC	TCACTTCATC	TTGCGGGTAT	ATCCTCAATT	CTTGGTGCTA	
<i>Cochlitoma ustulata</i>	CAGTGGATTT	GGCTATCTTC	TCTCTACACT	TAGCTGGTAT	ATCCTCTAAT	CTCGGGGGCTA	
<i>Cochlitoma zebra</i>	CCGTSGATCT	RGCTATCTTC	TCTCTACATY	TMGCTGGCAT	ATCCTCTAAT	TTAGGKGCCA	
<i>Metachatina kraussi</i>	CAGTTGACTT	AGCTATTTTT	TCTTTACATT	TAGCTGGGAT	ATCATCTAAT	TTAGGAGCTA	
<i>Rumina decollata</i>	CTGTTGACTT	AGCTATTTTT	TCTCTTCATT	TAGCAGGGAT	ATCCTCTAAT	TTAGGTGCAA	

	430	440	450	460	470	480	
<i>Achatina fulica</i>	TCAATTTTAT	TACGACGGTG	TATAATATAC	GAGCCTCTGG	GCTAACTATA	GAACGGGTAA	
<i>Achatina immaculata</i>	TTAATTTTAT	TACAACAGTA	TATAATATAC	GTTCTTCAGG	GTTAACCATA	GAACGAGTAA	
<i>Achatina reticulata</i>	TTAATTTTAT	TACTACAGTT	TACAATATAC	GTGCTTCTGG	GTTAACTATA	GAACGAGTAA	
<i>Achatina zanzibarica</i>	TTAATTTTAT	TACCACAGTC	TATAATATGC	GGGCTTCGGG	CTTAGAGATA	GAACGGATGA	
<i>Achatina achatina</i>	TTAATTTTAT	TACTACAATT	TTAATATAC	GAGCATCAGG	GATATCTATA	GAACGGGTAA	
<i>Archachatina marginata</i>	TTAATTTTAT	CACAACATTT	TTCAACATGC	GAGCTCCCGG	TCTTACAATG	GAGCGGGTAA	
<i>Achatina bisculpta</i>	TTAATTTTAT	TACTACTATT	TTAATATAC	GGGTGTCTGG	TTTGTCTATG	GAACGGGTAA	
<i>Achatina damarensis</i>	TTAATTTTAT	TACTACTATC	TTCAATATGC	GGGTGTCTGG	GCTAACTATA	GAACGGGTAA	
<i>Achatina stuhlmanni</i>	TCAATTTTAT	TACTACAATT	TTAACATAC	GAGTATCTGG	ACTATCTATG	GAACGGGTAA	
<i>Atopocochlis exarata</i>	TCAATTTTAT	TACCACAATT	TATAATATAC	GGGTAGGGG	ATTGACAATA	GAACGAGTAA	
<i>Limicolaria kambeul</i>	TTAATTTTAT	TACTACAATT	TTAATATAC	GAGCATCAGG	GTTATCTTTA	GAGCGTATAA	
<i>Limicolaria martenssii</i>	TTAATTTTAT	TACTACTATT	TTAATATAC	GAGTAAAAGG	CTTATCTATA	GAACGCTATA	
<i>Limicolariopsis sp.</i>	TTAATTTTAT	TACAACAGTA	TTAATATGC	GAGCATCAGG	GTTAACTATA	GAACGAGTAA	
<i>Cochlitoma varicosa</i>	TTAATTTTAT	TACCACAGTA	TTAATATAC	GAGCGCAGGG	GTTATCTATG	GAGCGAGTCA	
<i>Cochlitoma dimidiata</i>	TTAATTTTAT	TACAACAGTA	TTAATATAC	GGGTGCAAGG	ACTCTCAATG	GAACGGGTAA	
<i>Cochlitoma sp. cf. vestita</i>	TTAATTTTAT	TACTACAGTA	TTAATATAC	GAGCACAAGG	CTTATCAATA	GAACGTGTCA	
<i>Cochlitoma marinae</i>	TTAATTTTAT	CACCACATTT	TTAATATAC	GGGCGCAAGG	TTTAACTATA	GAACGAGTGA	
<i>Cochlitoma churchilliana</i>	TTAATTTTAT	TACCACAGTC	TTAATATAC	GTACTCCTGG	TTTAACTATA	GAACGGGTAA	
<i>Cochlitoma granulata</i>	TTAATTTTAT	TACTACAGTG	TTAACATAC	GGGCACAAGG	ATTATCTATG	GAACGGGTCA	
<i>Cochlitoma simplex</i>	TTAATTTTAT	TACCACAGTG	TTCAATATGC	GGGTGAAAAG	CCTGTCAATA	GAACGAGTGA	
<i>Cochlitoma kilburni</i>	TTAATTTTAT	TACTACAGTA	TTCAATATAC	GAGCACAAGG	ACTATCAATA	GAACGAGTCA	
<i>Cochlitoma montistempli</i>	TTAATTTTAT	TACTACAGTG	TTAATATAC	GGGCCAGGG	ATTATCAATG	GAACGAGTAA	
<i>Cochlitoma omissa</i>	TTAATTTTAT	TACAACAGTG	TTAATATAC	GGGCACAAGG	GTTAACTATG	GAACGAGTCA	
<i>Cochlitoma semidecussata</i>	TCAACTTTAT	TACTACAGTG	TTAATATAC	GGGCCAGGG	AATATCTATA	GAACGAGTAA	
<i>Cochlitoma ustulata</i>	TTAATTTTAT	CACAACATTC	TTAATATAC	GAGCTCAAGG	CTTATCTATG	GAGCGTATAA	
<i>Cochlitoma zebra</i>	TTAATTTTAT	YACTACTGTA	TTAATATAC	GAGCACAGGG	GTTGTCAATG	GAACGAGTCA	
<i>Metachatina kraussi</i>	TTAATTTTAT	TACTACCGTC	TTAATATAC	GAACCCGGG	GTTATCTATA	GAGCGGGTAA	
<i>Rumina decollata</i>	TTAATTTTAT	TACTACTATT	TATAATATAC	GGTCCCCAGG	TCTTTCAATA	GAACGGGTAA	

	490	500	510	520	530	540	
<i>Achatina fulica</i>	GCTTATTG	GTGGTCAATT	TTAGTTACAG	TGTTTCTACT	ATTGTTATCA	TTGCCAGTTC	
<i>Achatina immaculata</i>	GGCTATTG	CTGGTCTATT	TTAGTAACCG	TTTTTTTATT	ACTGCTATCC	CTACCAGTTC	
<i>Achatina reticulata</i>	GATTATTG	GTGATCAATT	TTGGTTACCG	TGTTTCTTTT	ACTACTATCA	TTACCTGTAC	
<i>Achatina zanzibarica</i>	GTTTATTG	TTGATCAATT	TTAGTCACCG	TATTTTACT	ACTACTCTCA	TTACCTGTAT	
<i>Achatina achatina</i>	GACTGTTG	ATGATCTATT	TTAGTTACAG	TATTTTACT	GCTTTTATCT	CTCCCAGTAT	
<i>Archachatina marginata</i>	GTCTTTTG	GTGGTCTATT	TTGGTGACAG	TTTTTCTGTT	GTTGTTGTCT	TTACCAGTGT	
<i>Achatina bisculpta</i>	GTTTATTG	TTGATCTATT	CTTGTAAC	TATTTCTTTT	ACTTCTCTCA	TTGCCAGTCC	
<i>Achatina damarensis</i>	GTTTATTG	TTGGTCAATT	CTTGTTACAG	TGTTTCTTTT	ACTTCTTTCT	TTGCCCTGTCT	
<i>Achatina stuhlmanni</i>	GGTTATTG	ATGATCAATT	CTTGTTACTG	TTTTCTTTT	ACTTTTATCT	CTACCAGTCT	
<i>Atopocochlis exarata</i>	GGTTATTG	GTGGTCAATT	TTGGTTACAG	TATTCCTTTT	ACTCTTATCA	TTACCAGTAT	
<i>Limicolaria kambeul</i>	GCTTATTG	TTGGTCTATT	CTTGTTACAG	TATTTTACT	ACTTCTCTCT	CTCCCTGTTT	
<i>Limicolaria martenssii</i>	GTCTGTTG	ATGATCTATC	TTAGTAACCG	TCTTTTATT	ACTCCTATCT	CTTCCAGTAC	
<i>Limicolariopsis sp.</i>	GGTTATTG	ATGATCTATT	TTAATTACTG	TATTTCTTTT	ACTCTTATCT	CTACCAGTCC	
<i>Cochlitoma varicosa</i>	GCTTATTG	GTGGTCAATC	CTTGTCAC	TATTTTACT	TCTTTTATCG	CTACCTGTTT	
<i>Cochlitoma dimidiata</i>	GCCTATTG	GTGGTCTATC	TTAGTTACCG	TGTTTACT	ACTTCTCTCC	CTTCCCTGCT	
<i>Cochlitoma sp. cf. vestita</i>	GGTTATTG	GTGGTCAATT	TTAGTAAC	TATTTTACT	ACTTCTCTCT	CTCCCAGTTC	
<i>Cochlitoma marinae</i>	GGTTGTTG	GTGATCAATC	CTAGTAACCG	TGTTTACT	ATTATATATCC	TTACCAGTTC	
<i>Cochlitoma churchilliana</i>	GTTTGTG	ATGGTCTGTT	CTAGTGACAG	TATTTACT	ACTTTTGTCT	TTACCTGTAT	
<i>Cochlitoma granulata</i>	GTTTATCG	TTGGTCTATT	TTGATTACTG	TTTTTTACT	CCTTCTCTCT	CTTCCCTGTT	
<i>Cochlitoma simplex</i>	GCCTCTTG	GTGGTCAATC	TTAGTCACCG	TGTTTACT	ACTTCTCTCC	CTCCCTGTCT	
<i>Cochlitoma kilburni</i>	GGTTATTG	ATGGTCAATT	TTAGTAAC	TATTTTACT	TATTTTATCT	CTTCCAGTTC	
<i>Cochlitoma montistempli</i>	GCTTGTG	ATGGTCAATC	TTGGTAACAG	TATTTTACT	TCTTCTCTCC	CTTCCGGTTT	
<i>Cochlitoma omissa</i>	ATTTATCG	ATGGTCTATT	TTAGTTACTG	TATTTTACT	CCTTCTCTCT	CTTCCCTGTT	
<i>Cochlitoma semidecussata</i>	GCTTATTG	GTGGTCCATC	TTGGTAACAG	TTTTTTACT	TCTTCTCTCC	CTTCCCTGTT	
<i>Cochlitoma ustulata</i>	GATTATTG	ATGGTCTATT	TTAGTTACTG	TTTTCTACT	GCTCTTATCT	CTACCAGTAC	
<i>Cochlitoma zebra</i>	GMCTATCG	MTGRTCCATT	YTAGTAACAG	TTTTCTACT	SCTCTTATCA	TTACCCGTCT	
<i>Metachatina kraussi</i>	GATTATTG	TTGATCTATT	TTAGTGACCG	TGTTCTTATT	ACTTCTTTCT	TTACCAGTTC	
<i>Rumina decollata</i>	GCCTATTG	GTGATCTATT	TTAGTAACCG	TTTTTCTATT	GCTTTTATCC	CTACCAGTAC	

	550	560	570	580	590	600	
<i>Achatina fulica</i>	TAGCGGGGC	AATTACTATA	CTATTAACCG	ATCGTAAC	TAACACATCT	TTTTTTGACC	
<i>Achatina immaculata</i>	TTGCCGGAGC	TATTACTATA	CTGCTAACAG	ACCGTAATTT	TAATACCTCA	TTTTTTGACC	
<i>Achatina reticulata</i>	TGGCTGGGGC	TATTACTATA	TTATTAACAG	ACCGTAATTT	TAATACATCT	TTTTTTGATC	
<i>Achatina zanzibarica</i>	TAGCAGGGGC	AATTACTATA	TTATTAACAG	ACCGAAACTT	TAACACATCA	TTTTTTGACC	
<i>Achatina achatina</i>	TAGCGGGTGC	AATTACTATA	CCTTCTGACCG	ATCGTAAC	TAATACATCT	TTTTTTGATC	
<i>Archachatina marginata</i>	TAGCTGGTGC	TATTACTATA	CTTTTAACTG	ATCGAAATTT	TAATACATCT	TTTTTTGACC	
<i>Achatina bisculpta</i>	TAGCTGGGGC	AATTACAATA	TTATTAACAG	ATCGAAATTT	TAATACATCA	TTTTTTGATC	
<i>Achatina damarensis</i>	TAGCTGGAGC	AATTACAATG	TTATTAAC	ATCGAAATTT	TAATACATCG	TTTTTTGACC	
<i>Achatina stuhlmanni</i>	TAGCTGGGGC	GATTACTATA	CTATTAAC	ATCGAAATTT	TAATACATCT	TTTTTTGATC	
<i>Atopocochlis exarata</i>	TAGCAGGGGC	AATTACTATA	CTATTAACAG	ATCGAAATTT	TAATACTTCA	TTTTTTGACC	
<i>Limicolaria kambeul</i>	TAGCTGGAGC	TATTACTATA	TTATTAAC	ATCGTAATTT	TAATACATCT	TTTTTTGACC	
<i>Limicolaria martenssii</i>	TGGCAGGGGC	TATTACTATA	TTATTAACAG	ATCGTAATTT	TAATACTCT	TTTTTTGACC	
<i>Limicolariopsis sp.</i>	TTGCAGGTGC	AATCACTATA	CTATTAAC	ATCGTAAC	TAATACGTCA	TTTTTTGATC	
<i>Cochlitoma varicosa</i>	TAGCAGGGGC	TATCACTATA	CTATTAACAG	ATCGAAATTT	TAACACTTCT	TTTTTTGATC	
<i>Cochlitoma dimidiata</i>	TAGCGGGGGC	TATCACTATA	CTACTTACCG	ACCGAAATTT	TAACACTTCA	TTTTTTGACC	
<i>Cochlitoma sp. cf. vestita</i>	TAGCAGGGGC	TATTACTATG	CTTTTAACTG	ATCGTAATTT	TAATACTTCC	TTTTTTGACC	
<i>Cochlitoma marinae</i>	TAGCAGGTGC	AATCACAAATG	CTCTTGACTG	ATCGGAATTT	TAATACTTCC	TTTTTTGATC	
<i>Cochlitoma churchilliana</i>	TGGCAGGGGC	GATTACTATA	CTGCTTACTG	ACCGGAATTT	TAATACTCT	TTTTTCGACC	
<i>Cochlitoma granulata</i>	TAGCTGGGGC	TATTACTATA	CTTTTAACTG	ATCGCAATTT	TAACACTTCC	TTCTTTGACC	
<i>Cochlitoma simplex</i>	TGGCCGGGGC	TATTACTATG	CTTCTAACAG	ACCGAAATTT	TAATACTTCA	TTCTTTGACC	
<i>Cochlitoma kilburni</i>	TAGCAGGGGC	TATTACCATA	CCTTCTGACAG	ACCGTAATTT	TAATACTTCT	TTTTTTGATC	
<i>Cochlitoma montistempli</i>	TGGCAGGAGC	TATTACTATG	CTTTTAACTG	ATCGAAATTT	TAACACTTCC	TTCTTTGATC	
<i>Cochlitoma omissa</i>	TAGCAGGGGC	TATTACTATA	CTTTTAACTG	ACCGAAATTT	TAACACTTCC	TTCTTCGACC	
<i>Cochlitoma semidecussata</i>	TAGCAGGGGC	TATTACTATA	CTTTTAACTG	ATCGGAATTT	CAACACTTCA	TTTTTTGATC	
<i>Cochlitoma ustulata</i>	TAGCAGGGGC	TATTACAATA	CTCCTTACTG	ATCGTAATTT	TAACACATCC	TTTTTCGACC	
<i>Cochlitoma zebra</i>	TAGCGGGGGC	TATTACCATA	TTATTAACAG	ATCGAAATTT	TAATACATCT	TTTTTTGATC	
<i>Metachatina kraussi</i>	TAGCTGGAGC	TATTACTATA	CCTTCTACTG	ATCGAAATTT	TAATACTTCT	TTTTTTGACC	
<i>Rumina decollata</i>	TAGCAGGAGC	TATTACAATA	CTACTAAC	ACCGTAAC	TAACACTTCA	TTCTTTGATC	

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      ....|....| ....|....| ....|....| ....|....| .
            610      620      630      640
Achatina fulica      CAGCTGGAGG GGGGGACCCA GTCCTGTACC AGCACCTATT T
Achatina immaculata CAGCAGGGGG TGGTGATCCA ATTTTATACC AACATTTATT T
Achatina reticulata CTGCTGGAGG TGGGGATCCG GTCTTATATC AACACTTGTT T
Achatina zanzibarica CAGCGGGAGG GGGAGACCCA ATTTTGTATC AGCACTTGTT T
Achatina achatina    CAGCTGGAGG TGGTGATCCA GTGCTTTACC AGCACCTATT C
Archachatina marginata CGGCAGGGAGG TGGGGACCC TATTCTTATC AGCATTTGTT T
Achatina bisculpta   CTGCTGGAGG GGGTGATCCT GTCTTGTATC AACACTTGTT T
Achatina damarensis CTGCCGGTGG TGGTGACCCG ATTTTATATC AACATTTATT T
Achatina stuhlmanni  CGGCAGGGGG TGGTGATCCA ATCCTATACC AGCAYCTATT C
Atopocochlis exarata CGGCAGGGGG TGGTGACCC TGTCTATATC AACATTTGTT T
Limicolaria kambeul  CTGCAGGCGG CGGGGATCCT ATTTTGTATC AGCACTTATT C
Limicolaria martenssii CAGCTGGAGG GGGGGATCCT ATTCTTTATC AGCATTTATT C
Limicolariopsis sp. CTGCAGGAGG AGGGGATCCA GTCTTATATC AACATTTATT T
Cochlitoma varicosa  CGGCCGGGGG GGGTGACCCT ATTCTATATC AGCATCTATT C
Cochlitoma dimidiata CAGCTGGAGG TGGGGACCC TATCTTGTACC AGCACCTCTT C
Cochlitoma sp. cf. vestita CGGCTGGGGG AGGCGATCCT ATTTTGTATC AACATCTCTT T
Cochlitoma marinae   CTGCAGGAGG TGGGGACCC TATTTATACC AGCATTTATT T
Cochlitoma churchilliana CTGCAGGAGG GGGAGACCCG ATTTTGTACC AACACCTTTT C
Cochlitoma granulata CAGCTGGTGG TGGGGACCC TATTTATATC AGCACCTTTT T
Cochlitoma simplex   CAGCTGGAGG TGGAGACCCT ATTTCTATACC AGCACCTCTT T
Cochlitoma kilburni  CAGCTGGTGG TGGTGACCCT ATTTTATACC AACATCTCTT T
Cochlitoma montistempli CGGCTGGTGG AGGTGATCCC ATTTTATATC AACATCTTTT T
Cochlitoma omissa    CAGCTGGTGG TGGGGACCC TATTTATATC AGCACCTTTT T
Cochlitoma semidecussata CTGCTGGGGG TGGTGATCCT ATTTTATATC AACACCTTTT C
Cochlitoma ustulata  CGGCAGGGGG AGGAGATCCT ATCTTGTATC AGCATCTCTT C
Cochlitoma zebra     CGGCTGGAGG GGGWGAYCCA ATCTTATACC AACATCTATT T
Metachatina kraussi  CTGCAGGAGG CGGAGATCCT ATTTTATATC AGCATCTTTT T
Rumina decollata    CTGCAGGGGG AGGTGATCCA ATTCTATATC AGCATTTATT T

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Appendix 4.5: Alignment of the 16S rRNA gene for the Achatinidae and the subulinid outgroup *Rumina decollata*. Positions included for the phylogeny analyses were marked by “m” within “NUCEOTIDES INCLUDED”. Note that sequencing was unsuccessful for *Limicolariopsis ruwenzoriensis*.

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      ....|....|....|....|....|....|....|....|....|....|
      10      20      30      40      50      60
Achatina fulica      TAGCTTAAAG A-GTGCCTAA CACAGATTTT AAGTAACCCC TGCCCGGTGA GTTATAGA--
Achatina immaculata TAGTTTAAAG AA----ATTA AAAAGATTTT AAGTGATCCC TGCCCGGTGA GTAAAA----
Achatina reticulata TAGCTTAAAG CG----TWCC TAAAGATTTT AAGTAATCTC TGCCCGKGA GTTAC-----
Achatina zanzibarica TAGCTTAAAG GA-----CC AAAMCATTAT AAGTAACCCC TGCCCGGTGG GCTGT-----
Euaethiopia loveridgei CAGCTTGAAG TT-----TAA AAAAGATTTT AAGTGATCCC TGCCCGGTGA GATTTGTT--
Achatina achatina    TAGCTTAGG  A----- TTTAAATCTT TAGTACCTCC TGCCAGTGA TAGGTTAC--
Archachatina marginata TAGCTCTAGG A----- CTTAAATCTT GAGTACACCC TGCCAGTGA CAATTT----
Achatina bisculpta   TRGKTARTAG T----- TTATGATTAT TAGTGGATCC TGCCAGTGG AATTTTAA--
Achatina damarensis TAGCCAGTAG C-----TAG CTAGGATTGC TGGTGGATCC TGCCAGTGG AAAATTCTC-
Achatina stuhlmanni TAGCTAATAG G---CTCTTT CCATAATTAT TAGTGGACCC TGCCCGGTGA AGTTAA----
Atopocochlis exarata TAGCTAAAAG T----- -AGAAATTTT TAGTGTGTCC TGCCACTGG GTTCC-----
Limicolaria kambeul TAGCTAATAG A----- -ATAGATTAT TAGTATTCCC TGCCAGTGA AAATATTTCA
Limicolaria martenssii TAGCTAATAG C-----ATTT TATATATTGT TAGTGTGTCC TGCCAGTGG AGAACATA-
Limicolariopsis sp. TAGYTGAAC  G-----TT AGARATTTT CAGTATATTC TGCCAGTGA GATAAT----
Cochlitoma varicosa TAGCTTAAAG T----- AATTTATCTT AAGTGGTCCC TGCCCGTGA AAGTT-----
Cochlitoma dimidiata TAGCTKAAG  A-----A GTTTGATTTT AAGTACCCC TGCCAGTGG TAAATC---
Cochlitoma sp. cf. vestit TAGCTTGAAG T----- TTTTCTTTT AAGTGGTCCC TGCCCGTGG ATTTA-----
Cochlitoma marinae TAGCTTAAGG G-----T ATTAGTYTT AAGTAGTCCC TGCCCGTGG TAATAA----
Cochlitoma churchilliana TAGCTTAAAG T----TCTA AGTCTATTTT AAGTAGTCCC TGCCCGTGA GTAGAGAAA-
Cochlitoma granulata TAGCTTGAGA G-----T ACTTTATTTT AAGTGTCCC TGCCAGTGA ACAATAA---
Cochlitoma simplex TAGCTTGAAG C-----AC ATTTGATTTT AAGTACCCC TGCCCGTGG CGCAGTAGCT
Cochlitoma kilburni TAGCTTGAAG T----- TTTTCTTTT AAGTGTCCC TGCCCGTGG TGTAA-----
Cochlitoma montistempli TAGCTTGAGA G-----T ATTTTATTTT AAGTGTCCC TGCCAGTGA TAATTA----
Cochlitoma omissa TAGCTTGRGA G-----T ACTTCATTTT AAGTGTCCC TGCCAGTGA TCAATAA---
Cochlitoma semidecussata TAGCCTAAGA G-----T GCTTTATTTT AGGTGTCCC TGCCAGTGA TAATTA----
Cochlitoma ustulata TAGCCTGCTG T-----A GATTTATTAC AGGTGTCCC TGCCCGTGG GGCCTATT--
Cochlitoma zebra TAGCTTTAAG T----- AATTCATTTT AAGTGGTCCC TGCCCGTGA GAATC-----
Metachatina kraussi TTGCTTGAAG T-----AGT ATTTTATTTT AAGTAATCCC TGCCCGTGG GATAATA---
Rumina decollata    -----TTT AGGTAACCCC TGCCAGCGA TATAT-----
NUCLEOTIDES INCLUDED -----mmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm m-----

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      ....|....|....|....|....|....|....|....|....|....|
      70      80      90      100     110     120
Achatina fulica      ---AACTTTA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTCT
Achatina immaculata ---TATTTTA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Achatina reticulata ---TACTTTA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Achatina zanzibarica ---GGCTTAA ACGGCTGCAG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTAT
Euaethiopia loveridgei ---TTCTTTA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Achatina achatina    ---CTATTTA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Archachatina marginata ---TGACTTA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Achatina bisculpta   ---ATTCTTA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Achatina damarensis ---ATTCTTA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Achatina stuhlmanni ---CCTTTTA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Atopocochlis exarata ---CTCCTTA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Limicolaria kambeul TAGATTTTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Limicolaria martenssii ---TACCTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATCA TTTGTCCTTT
Limicolariopsis sp. ---ATCTTTA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma varicosa ---AGTTTTA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma dimidiata ---ACCCTAA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma sp. cf. vestit ---AACCTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma marinae ---ACTCTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma churchilliana ---TACTTCA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma granulata ---ACCTTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma simplex ---GCCCTAA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma kilburni ---AACCTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma montistempli ---ACCTTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma omissa ---ACCTTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma semidecussata ---ACCTTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Cochlitoma ustulata ---ACCCTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATCA TTTGTCCTTT
Cochlitoma zebra ---AGTTTTA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Metachatina kraussi ---AACCTAA ACGGCCGCG TACCTTGACT GTGCAAAGGT AGCATAATAA TTTGTCCTTT
Rumina decollata    ---AAGTTTA ACGGCCGCG TACCTTGACT GTGCTAAGGT AGCATAATAA TTTGTCCTTT
NUCLEOTIDES INCLUDED -----mmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm mmmmmmmmm

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	130	140	150	160	170	180
Achatina fulica	AATTAAGGTC	TGGAATGAAG	GGGGACA--C	AGGGGAG-AG	CTGTCTCCAA	TAAGGTTAAT
Achatina immaculata	AATTGAGGTC	TAGTATGAAT	GGGTTCA--C	AGGGGAA-AG	CTGTCTCAAG	GGAGATTAAT
Achatina reticulata	AATTGAGGTC	TGGAATGAAT	GAGAAACA--C	AGGGGAG-AG	CTGTCTCTAG	WARGGTTAAC
Achatina zanzibarica	AATTGAGGTC	TGATATGAAT	GGGTTCA--C	GGGGGAA-AC	CTGTCTTTAT	TAAGGTAACC
Euaethiopina loveridgei	AATTGAGGTC	TGATATGAAT	GGGGGTA--C	GGGCGGG-TG	CTGTCTTTAC	TAAAGGTTAC
Achatina achatina	AATTAGGGTC	TGATATGAAA	GGATTTT--C	GGGAGTT-TA	CTGTCTTAGT	GAGGTTAATA
Archachatina marginata	AATTGAGGTC	TGGAATGAAT	GGGCTAC--C	GGGGGTT-GG	CTGTCTGCTA	-AAGGATAAT
Achatina bisculpta	AATTAAGGTC	TTGTATGAAA	GGGGGAA--C	GGGGGTT-AA	CTGTCTCTAA	GGGGTTTCAG
Achatina damarensis	AATTAAGGTC	TTGTATGAAA	GGGAGAA--C	GGGGGAT-AA	CTGTCTCTTA	TGGGGTTTAA
Achatina stuhlmanni	AATTAAGGTC	TTGCATGAAT	GGGTGTT--C	GAGGAGT-CG	CTGTCTTTAT	TAGGGTTTAA
Atopocochlis exarata	GATTGGGGTC	TAGAATGAAT	GGATGAT--T	GGGGGAA-AA	CTGTCTTCTT	CATGTTAATA
Limicolaria kambeul	AAATGAGGTC	TTGTATGAAT	GGGATAA--C	GGGGGTT-AG	CTGTCTTTAT	ATAGTTTCTT
Limicolaria martenssii	AAATGAGGTC	TTGTATGAAA	GGATTTA--C	GGGGGAG-AG	CTGTCTTATC	TATAAAAAAC
Limicolariopsis sp.	AATTAGGGTC	TAGAATGAAA	GAACGCA--C	GATATAA-AG	CTGTCTCAAA	ATGGGTATT
Cochlitoma varicosa	AATTGGGGTC	TAGTATGAAT	GGGTTAA--C	GGGTATT-AG	CTGTCTCAGG	GGGGTCTTAT
Cochlitoma dimidiata	AATTGAGGTC	TGGAATGAAC	GGGTTAGA-C	GAATGTT-AG	CTGTCTTATG	TTAGGTCCTA
Cochlitoma sp. cf. vestit	AATTGAGGTC	TGGAATGAAT	GGGTTAAA-C	GAGTATT-AG	CTGTCTTATG	TTGGTTAATG
Cochlitoma marinae	AATTGAGGTC	TAGAATGAAT	GGGAAAAG-C	GGGTGAT-AG	CTGTCTTAA	TCAGTTTTTA
Cochlitoma churchilliana	AATTAAGGTC	TGATATGAAT	GGGTTAA--C	GGGTGTT-GA	CTGTCTCAGG	GATTGATATT
Cochlitoma granulata	AATTGAGGTC	TGATATGAAT	GGGTTAGA-C	GGGTATA-AG	CTGTCTTAA	TTGGTTATCT
Cochlitoma simplex	AATTGAGGTC	TAGAATGAAT	GGGTTAAAAC	GGGTGTT-AG	CTGTCTCAGG	TTGGGTTTTT
Cochlitoma kilburni	AATTGAGGTC	TGGAATGAAT	GGGTTTAA-C	GGGTGTT-AG	CTGTCTCATG	CTGGTTAATG
Cochlitoma montistempli	AATTGAGGTC	TGGAATGAAT	GGGTTAGA-C	GGGTATA-AG	CTGTCTTAA	TTGGTTATCT
Cochlitoma omissa	AATTGAGGTC	TGGAATGAAT	GGGTTAGA-C	GGGTATG-AG	CTGTCTTAA	TTGGTTATTT
Cochlitoma semidecussata	AATTGAGGTC	TGGAATGAAT	GGGTTAGA-C	GGGTATA-AG	CTGTCTTAA	TTGGTTATCC
Cochlitoma ustulata	AATTGAGGTC	TGGAATGAAA	GGGAAAGAAC	GGGTATG-AG	CTGTCTCTAG	TTAGGTCCTT
Cochlitoma zebra	AATTGAGGTC	TGGAATGAAT	GGGTTTAA-C	GGGTAAA-AG	CTGTCTTAGG	AAGGTCCTAT
Metachatina kraussi	AATTGAGGTC	TGGAATGAAT	GGGAAA--C	GGGTATA-AG	CTGTCTCAAG	CTATTTTTGT
Rumina decollata	AATTGAGGTC	TTGAATGAAC	GGGTTTAA--C	GAAAAATGTT	CTGTCTCCTG	TACAAATTTT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmm-----m	mmmm-----m	mmmmmmmmmm--	-----

	190	200	210	220	230	240
Achatina fulica	T--TAACCTT	C--TTATCAGG	TGAAAATTC	TGAGCCT--CC	GATGAAAGAC	GAGAAGACCC
Achatina immaculata	C--TAATTTA	C--TTATCAGG	TGAAAATTC	TGAGGAT-AA	AAGGAAAGAC	GAGAAGACCC
Achatina reticulata	T--TAACCTC	C--TTATCAGG	TGAAAATTC	TGAGCTT-CT	TTGGAAAGAC	GAGAAGACCC
Achatina zanzibarica	T--AAATTTG	C--TTATCAGG	TGAAAATTC	TGAATTTATA	AAGGAAAGAC	GAGAAGACCC
Euaethiopina loveridgei	CT-AAACTTA	C--TTATCAGG	TGAAAATTC	TGAACTT-TT	AAGGAAAGAC	GAGAAGACCC
Achatina achatina	T--GAATTTA	C--TCAATAAG	GTGAAAATTC	CTTAAAATTA	GATAAAAGAC	GAGAAGACCC
Archachatina marginata	TT-GAATTTG	C--TTATAAAG	TGAAAATTC	TTTAAATTTA	GATAAAAGAC	GAGAAGACCC
Achatina bisculpta	T--GAATTTT	C--TCAGGAGG	TGAAAATTC	TCTGATATTT	GACAAAAGAC	GAGAAGACCC
Achatina damarensis	T--AAATTTA	C--TTAAGAGG	TGAAAATTC	TCTAATTT-GT	GACAAAAGAC	GAGAAGACCC
Achatina stuhlmanni	TT-AAATTTA	C--TCAAGAGG	TGAAAATTC	TCTAATTTAA	GATAAAAGAC	GAGAAGACCC
Atopocochlis exarata	T--TAACCTA	C--TTAAAAGG	TGAAAATTC	TTTAGTT-AA	GAAAAAAGAC	GAGAAGACCC
Limicolaria kambeul	T--GAACCTA	CTAAATTAGG	TGAAAATTC	TACACTTATA	TACAAAAGAC	GAGAAGACCC
Limicolaria martenssii	---GAAATTA	CTAATTAGG	TGAAAATTC	TATAAAT-AA	GAAAAAAGAC	GAGAAGACCC
Limicolariopsis sp.	TT-TAATTTG	C--TAAAGAGG	TGAAAATTC	TTTAAATTTT	TATAAAAGAC	GAGAAGACCC
Cochlitoma varicosa	T--TAATTTA	T--TTAGAAGG	TGAAAATTC	TTTAAATTTA	TATAGTAGAC	GAGAAGACCC
Cochlitoma dimidiata	T--TAACCTA	A--TTAGAAGG	TGAAAATTC	TTTATGTATT	ATTAGAAGAC	GAGAAGACCC
Cochlitoma sp. cf. vestit	C--TAACCTA	A--TTAGAAGG	TGAAAATTC	TTTATGTATT	ATTAGAAGAC	GAGAAGACCC
Cochlitoma marinae	T--TAATTTA	A--TTAGGGGG	KGAAAATTC	CCTATTCCTA	AACAGTAGAC	GAGAAGACCC
Cochlitoma churchilliana	ATTGAATTTA	A--TCATAAGG	TGAAAATTC	TTTACTTATT	AACAGAAGAC	GAGAAGACCC
Cochlitoma granulata	T--AAATTTA	A--TTAAAAGG	TGAAAATTC	TTTAAAATAA	GATAGAAGAC	GAGAAGACCC
Cochlitoma simplex	T--TAACCTA	ATCTAGAAGG	TGAAAATTC	TTCAAGAATA	GACAGAAGAC	GAGAAGACCC
Cochlitoma kilburni	T--CAACTTA	A--TTAGAAGG	TGAAAATTC	TTTATGTATT	AATAGAAGAC	GAGAAGACCC
Cochlitoma montistempli	T--AACTTTA	A--TTAGAAGG	TGAAAATTC	TTTAAATATAT	TACAGAAGAC	GAGAAGACCC
Cochlitoma omissa	T--AAATTTA	A--TTAAAAGG	TGAAAATTC	TTTAAAGTGA	AATAGAAGAC	GAGAAGACCC
Cochlitoma semidecussata	T--GAACCTA	A--TGAAAAGG	TGAAAATTC	TTTAAAGT-GT	TACAGAAGAC	GAGAAGACCC
Cochlitoma ustulata	T--AAATTTA	A--TTGTTAGG	TGAAAATTC	TATACCT-TT	AACAGAAGAC	GAGAAGACCC
Cochlitoma zebra	T--TAACCTA	A--TTAGAAGG	TGAAAATTC	TTTAAAGTATA	AAAAGTAGAC	GAGAAGACCC
Metachatina kraussi	T--GAACCTA	CTTTAGAAGG	TGAAAATTC	TTTGTTT-TA	TTTAGAAGAC	GAGAAGACCC
Rumina decollata	ACTGAACCTA	C--TTAAGAGG	TGAAAATTC	TTTAGTA-AT	GAAAAAAGAC	GAGAAGACCC
NUCLEOTIDES INCLUDED	---mmmmmmmm	m--mmmmmmmm	mmmmmmmmmm	mmmm-----	mmmmmmmmmm	mmmmmmmmmm

	250 260 270 280 290 300
<i>Achatina fulica</i>	TTAGAGTTTT TATTATAACT TGATA----- TATTAAGATT CTTGTTCTGT TTTTGTGGG
<i>Achatina immaculata</i>	TTAGAGCTTT TATGAAAATT TGAGA----- AGTTACTAAT CTAATATCGT TTTTGTGGG
<i>Achatina reticulata</i>	TTAGAGTTTT AATAAATATT TAGAT----- AAATAAAATT CTAATATCGT TTTTGTGGG
<i>Achatina zanzibarica</i>	TTAGAGTTTT CATTAAAGCC TAGGA----- --GTAATAGC ATGGTTATGT TTTTGTGGG
<i>Euaethiopina loveridgei</i>	TTAGAGTTTT TACTACAAC T AGAAA----- -TGTCATATT CTAGTATTGT TTTTGTGGG
<i>Achatina achatina</i>	TTGGAGTTTT TATTATATAG ATAGA----- ----- TTCTATCTT TTTTCGTTGGG
<i>Archachatina marginata</i>	TTGGAATTTT AAGGAATTTG TAAGC----- -----T TTACATAAAT TTTTCGTTGGG
<i>Achatina bisculpta</i>	TTAGAGCTTT AATAAAAAGT AAATC----- ----ACTATT TTACTTAAGT TTTTGTGGG
<i>Achatina damarensis</i>	TTAGAGCTTT TACAAAAAGT AGACAAGTAG TGTACTATTT CTACTTAAGT TTTTGTGGG
<i>Achatina stuhlmanni</i>	TTAGAGTTTT AATGAAAAGT AAAC----- -----T TTACTTAAGT TTTTGTGGG
<i>Atopocochlis exarata</i>	TTAGAAAATTT GTAGAAAATG CAAAT----- ----TCAAT TTGCTTACTT TTGTGTTGGG
<i>Limicolaria kambeul</i>	TATGAGTTTT TATGAATAAA TGAAA----T CATATTTTAT CATACTAAGT TTTTGTGGG
<i>Limicolaria martenssii</i>	TAAGAGTTTT AATGAGAAATA CTATT----- AGGTAATTTAA ATAGAGAAGT TTTTGTGGG
<i>Limicolariopsis sp.</i>	TAAGAGCTTT TAATAACTTA AATAT----- -AAAAATTTA TATTTAAAGT TTTTGTGGG
<i>Cochlitoma varicoso</i>	TTAGAGTTTC CATGAAAAGT TATAA----- --TCTTTAT ATATCTAAGT TTTTCGTTGGG
<i>Cochlitoma dimidiata</i>	TTAGAGTTTT AATGAAAACAA TATAT----- TTTACAATTG TATTTTATGT TTTTCGTTGGG
<i>Cochlitoma sp. cf. vestit</i>	TTAGAGTTTA AAATGAATAA ATATT----A ATCTTACGAT TATTTTATGT TTTTCGTTGGG
<i>Cochlitoma marinae</i>	TTAGAGTTTT TATGAAAAGT GTCTA--AGG TTATCTAGTA TACTACTAGT TTTTCGTTGGG
<i>Cochlitoma churchilliana</i>	TTAGAGTTTT TATGAAGAAG TTATA--TAT ATATCAAATA AGCTTTAAGT TTTTCGTTGGG
<i>Cochlitoma granulata</i>	TTAGAGTTTT TATAAATAAA TATAG---AA CAGATGAGTT ATATTTTATGT TTTTGTGGG
<i>Cochlitoma simplex</i>	TTAGAGTTTT AATGAAAATAA TATAG----- TTTAAAGTAG GTTTTTTATGT TTTTCGTTGGG
<i>Cochlitoma kilburni</i>	TTAGAGTTTT AAATGAATAA ATACT----A ATATTATAAT TATTTTATGT TTTTCGTTGGG
<i>Cochlitoma montistempli</i>	TTAGAGTTTT TATAAATAAA TATAG---A ACAATAAGTT GTATTTTATGT TTTTGTGGG
<i>Cochlitoma omissa</i>	TTAGAGTTTT TATAAACAAA TGTAG---A ACAATGAGTT ATATTTTATGT TTTTGTGGG
<i>Cochlitoma semidecussata</i>	TTAGAGTTTT TATAAATAAA TGTAG---A ACAATGCGTT GTATTTTATGT TTTTCGTTGGG
<i>Cochlitoma ustulata</i>	TTAGAGTTTT TATGATAAGC ATTCT----- ----ACATA TGCTGTCAGT TTTTCGTTGGG
<i>Cochlitoma zebra</i>	TTAGAGTTTT TATAAGAATA TATAA----- --CTTTTTT ATATTTTATGT TTTTCGTTGGG
<i>Metachatina kraussi</i>	TTAGAGTTTT TATAAATAAA TATGG---TA ATTATTTAAA TATATTGAGT TTTTCGTTGGG
<i>Rumina decollata</i>	TTAGAATTTT AATAAAAAC TGCAT----- ----TGCGTG ATTGTTTATGT TTTTCGTTGGG
NUCLEOTIDES INCLUDED	mmmmmmmmmm m----- ----- ----- mmmmmmmmmmm

	310 320 330 340 350 360
<i>Achatina fulica</i>	GCGACAGGGT TACA----- -----A TAGATAACTT ACCCTACCAT A--TTTTTAT
<i>Achatina immaculata</i>	GCGACAGGGT AGCA----- ----- TATTTAAACCT ACCTGATCTT T--AAAGTAT
<i>Achatina reticulata</i>	GCGACAGGGT AACA----- -----AT AGGTAAACCT ACCCATTCAT ATATTTTTAT
<i>Achatina zanzibarica</i>	GCGACGGGGT TACA----- ----- GTTTGAACCT ACCTAGAATA C-TTCTGTT
<i>Euaethiopina loveridgei</i>	GCGACAGGGT GACA----- -----A TAATTAACCT ACCAAGACAA TATTATATAT
<i>Achatina achatina</i>	GCAGCGAGAT TACA----- ----- -TCTTTA CTTTTAATTA ATCTTTACTT
<i>Archachatina marginata</i>	GCGGCGAGAC TACA----- ----AGGTT TCATTAACCTA GTCTTACATT ATAATTTGGT
<i>Achatina bisculpta</i>	GCAACAAGAT GGCA----- -AATGAAAAT AAACCCATCT AATATTCCTA TTTTATTTGA
<i>Achatina damarensis</i>	GCAACAAGAT GGCA----- ----AAGTT TAACCCATCC A-TATCTAT GTTAATTTGA
<i>Achatina stuhlmanni</i>	GCAACAAGAT GGCAAAAATT AGTAATACTT AAACCCATCT AATTTTCTAA AGTGAATCAG
<i>Atopocochlis exarata</i>	GCAACAGAAT GGCA----- ----AATATA TAACCCATT AAACCTTAAG CTTAAATCTA
<i>Limicolaria kambeul</i>	GCAACAAGAT ATCA----- ----- AGATTAATAT ATCTTTATTA TAGTAAATAA
<i>Limicolaria martenssii</i>	GCAACAGAAT TCCA----- ----- GATTAATGA ATTTATCTAT ATTATATAT
<i>Limicolariopsis sp.</i>	GCAACAAGAT ATCA----- ----AA AAGTTAACAT ATCTGTTTTT CTGGAATACA
<i>Cochlitoma varicoso</i>	GCAACGGGGT CGTA----- ----- AAGTTAACCG ACCTGTATAG ATGGTAAATG
<i>Cochlitoma dimidiata</i>	GCAACGGAAT CGCA----- ----G TGTTTAACCG GTTTAA--AA GTAATACACA
<i>Cochlitoma sp. cf. vestit</i>	GCAACGGAGT CGCA----- -----A TATTTAACCG GCTAACGAAA TAAACTAAGA
<i>Cochlitoma marinae</i>	GCAACGGATC TGCA----- ----AT TAGTTAAACA GATTAATTTG TTTAGGGTTT
<i>Cochlitoma churchilliana</i>	GCAACGGGGT AGCA----- ----- ATATTAACCT ACTTGTTAGG GAAGTAATAA
<i>Cochlitoma granulata</i>	GCAACAGGTT CGCA----- -----A GCATAAACCG AATTATTTAG TAACTAACAT
<i>Cochlitoma simplex</i>	GCAACGGAAC CGCA----- ----GA GCTTGAACCG GTTTAATAAA TAATGCATAT
<i>Cochlitoma kilburni</i>	GCAACGGAAT CGCA----- -----A TACTTAACCT GTTAACGAAA TAAACTAATA
<i>Cochlitoma montistempli</i>	GCAACGGAAT CGCA----- -----A ATACTAACCG ATTTATCAAA TAATTAATGA
<i>Cochlitoma omissa</i>	GCAACAGTTT CGCA----- -----A GCATAAACCG AATTATTTAG TAACTAACAT
<i>Cochlitoma semidecussata</i>	GCAACAGAAT CGCA----- ----A ATATTAACCG ATCTAT-TAA TACCTAGTGG
<i>Cochlitoma ustulata</i>	GCAACGAGAT TGCA----- ----TG AAATAAACCA ATCAT--ACG TTAATAGTAG
<i>Cochlitoma zebra</i>	GCAACGGGGT CGAA----- ----- AAATTAACCG ATCAAAAAAT TTAATTAATAA
<i>Metachatina kraussi</i>	GCAACGAGAT TTCA---TTA TTTTAATATA TCTGTATAAA ATTTTATC--
<i>Rumina decollata</i>	GCGACAGGGT AGCA----- ----AT CAAATAACCT ACCCTAATAT ATTTAATACA
NUCLEOTIDES INCLUDED	mmmmmmmmmm mmmmm----- ----- ----- mmmmmmmmmmm

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      370      380      390      400      410      420
Achatina fulica      ATGGCGATCA TTATACTTT- ----- ---TAATTAA GCTACCTAAG GGATAACAGC
Achatina immaculata ATACCGATGG TAATATTAA- ----- ---CACGAAA GCTACCTAAG GGATAACAGC
Achatina reticulata CCGTCGATAG TTATACTAA- ----- ---TAATTAA ACTACCTAAG GGATAACAGC
Achatina zanzibarica TTGACGACTT TCTTACCTA- ----- ---TAATAAA ACTACCTAAG GGATAACAGC
Euaethiopina loveridgei CCAGCGATTA TTATACTAG- ----- ---TAAATAA GCTACCTAAG GGATAACAGC
Achatina achatina    ATCTAAACAT TTGCCAAGTA TTTATCTTAA GAAAAACTAA ACTACCCAAG GGATAACAGC
Archachatina marginata CAAGTTTATT AAGAA----- ----- ---GAATTAA ATTACCCAAG GGATAACAGC
Achatina bisculpta   AGAGG-TTTC TAAGAG---- ----- ---TAGTTAA GCTACCTAAG GGATAACAGC
Achatina damarensis AGAGTTGTTT TAAGAG---- ----- ---TAGATAA GCTACCTAAG GGATAACAGC
Achatina stuhlmanni TCAATTATTA TAAGAA----- ----- ---CAGAAAA ACTACCTAAG GGATAACAGC
Atopocochlis exarata CTAGTTAATT TTAATAAGAA ----- ---TGAAGAA TTTACCTAAG GGATAACAGC
Limicolaria kambeul  ATCGATTATG ATAATGG--- ----- ---AAACTAA ACTACCTTAG GGATAACAGC
Limicolaria martenssii AAGAAGAAAG GTAATAAGAA ----- ---AAAATAA GCTACCTTAG GGATAACAGC
Limicolariopsis sp. TACCGACTAG TTAGTATAAA ----- ---TGATCAA GCTACCTTAG GGATAACAGC
Cochlitoma varicosa  VATATGCCAA TTTTATAAAG AA----- ---TAATTAA ACTACCTAAG GGATAACAGC
Cochlitoma dimidiata TATATGCCAA TTTTAAAAG AA----- ---TAAGTAA ACTACCTAAG GGATAACAGC
Cochlitoma sp. cf. vestit GATTTGCCAA TATTAATAAG AA----- ---TAAGAAA ACTACCTAAG GGATAACAGC
Cochlitoma marinae  TATCTGCCAA TTTATTATAA GGG----- ---TAAATAA ACTACCTAAG GGATAACAGC
Cochlitoma churchilliana ACA-TGCGTC AATATTTTAT TCGTT----- ---TAACAAA ACTACCTAAG GGATAACAGC
Cochlitoma granulata ATA-TGACAA TTCCTATAAG AA----- ---TAAGTAA ACTACCTAAG GGATAACAGC
Cochlitoma simplex  ATT-TGTCCA CTTTCAAAG AA----- ---TAAGTAA ACTACCTAAG GGATAACAGC
Cochlitoma kilburni CATTTGCCAA TATTAATAAG AA----- ---TAAGAAA ACTACCTAAG GGATAACAGC
Cochlitoma montistempli ATA-TGACAA TTCGTATGAG AG----- ---TAAATAA ACTACCTAAG GGATAACAGC
Cochlitoma omissa   ATA-TGACAA TTTATATAAG TA----- ---TAAGTAA ACTACCTAAG GGATAACAGC
Cochlitoma semidecussata ATA-AGACAA TTCGAATAAG AA----- ---TAAGCAA ACTACCTAAG GGATAACAGC
Cochlitoma ustulata ATA-AGTCAA TCTTAATAGG TA----- ---TAATTAA ACTACCTAAG GGATAACAGC
Cochlitoma zebra    AATTTGATAA TCTTTATAAG AA----- ---TAATTAA ACTACCTAAG GGATAACAGC
Metachatina kraussi ----TGCCAA TTATTCTATA AA----- ---CAAGTAA GCTACCTAAG GGATAACAGC
Rumina decollata    CCGAAATCTA TTAACA----- ---TAATTAA ATTACCTAAG GGATAACAGC
NUCLEOTIDES INCLUDED -----

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      430      440      450      460      470      480
Achatina fulica      GTAATCTTTT T--TTTGGT TTGCGACCTC GATGTTGGAC TAGGGCCCTA CTGGTTAGTT
Achatina immaculata GTAATCTTTG ---TTATGGT TTGCGACCTC GATGTTGGAC TAGGAACCTT TTGGTTAGTT
Achatina reticulata GTAATCTTTT T--TTTGGT TTGCGACCTC GATGTTGGAC TAGGGCCCTA CTGGTTAGCA
Achatina zanzibarica GTAATCTAAA ---AGTAGT TTGCGACCTC GATGTTGGAC TAGGGTCCAC TTGGTTAGCC
Euaethiopina loveridgei GTAATCTTTT ---TTAAGGC TTGCGACCTC GATGTTGGAC TAGGGACCTG TTGGTTAGTT
Achatina achatina    ATAATTCCTT ---AGGGAGA TTGTGACCTC GATGTTGGAC TAGGGCTCTA TTGGTTAACT
Archachatina marginata ATAATTCCTA ---ATGGAGT TTGTGACCTC GATGTTGGAC TAGGGCCCTT ATGATTAECT
Achatina bisculpta   GTAATCTAAA ATGTTTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCAA ATGATTAGAT
Achatina damarensis GTAATCTAAT AG-TTTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCAA ATGATTAGAT
Achatina stuhlmanni GTAATCTAAT ---ATTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA ATGATGAGAT
Atopocochlis exarata GTAATCTAGC ---ATTAGGA TTACGACCTC GATGTTGGAC TAGGGACCAA ATGATAAGAT
Limicolaria kambeul  GTAATCTAAA ---TTTAGGT TTGCGACCTC GATGTTGGAC TAGGGCCCTA ATGATTAGTC
Limicolaria martenssii GTAATCTATT A--GTTAGGT TTGCGACCTC GATGTTGGAC TAGGGACCGA ATGATTAGC
Limicolariopsis sp. GTAATCTATT ---AATAGGA TTGCGACCTC KATGTTGGAC TAGGGWCCAA ATGATTWGT
Cochlitoma varicosa  GTAATCTTCT ---TTTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA TTGGTTAKTA
Cochlitoma dimidiata GTAATCTAYT T--TTTAGGY TTGCGACCTC GATGTTGGAC TAGGGACCTA TTGGTTAGCA
Cochlitoma sp. cf. vestit GTAATTTATT ---TTAAGA TTGCGACCTC GATGTTGGAC TAGGGACCTT TTGGTTAGTA
Cochlitoma marinae  GTAATCTTAC T--ATTGGGA TTGCGACCTC GATGTTGGAC TAGGGACCCA TTGGTTAGTA
Cochlitoma churchilliana GTAATCTCTC ---TTTTGGA TTGCGACCTC GATGTTGGAC TAGGGACTGA ATGGTTAACA
Cochlitoma granulata GTTATCTACT ---ATTAGGA TTGCGACCTC GATGTTGGAC TAGGGACTTA TCGGTAATCA
Cochlitoma simplex  GTGATCTATT ---CATAGT TTGCGACCTC GATGTTGGAC TAGGGACCCA TTGGTTAACA
Cochlitoma kilburni GTAATTTATT ---TTAAGA TTGCGACCTC GATGTTGGAC TAGGGACCTT TTGGTTAGTA
Cochlitoma montistempli GTAATCTACT A--TTTGGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA TTGGTTAGCA
Cochlitoma omissa   GTTATCTACT ---ATTAGGA TTGCGACCTC GATGTTGGAC TAGGGACTTA TTGGTTAACA
Cochlitoma semidecussata GTAATCTACT ---ATTGGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA TTGGTTAGTA
Cochlitoma ustulata GTAATCTTAT ---TTTAGGA TTGCGACCTC GATGTTGGAC TAGGGACCCA GTGGTGAGAA
Cochlitoma zebra    GTAATCTTCT ---TTTTGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA ATGGTTAGTA
Metachatina kraussi GTAATCTTT- ---TGAAGGA TTGCGACCTC GATGTTGGAC TAGGGACCTA TCGGTTAGCA
Rumina decollata    ATAATACTAA ---TTAGTGT TTGTGACCTC GATGTTGGAC TAGGGACTAA TTGGTTAACC
NUCLEOTIDES INCLUDED -----

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Appendix 4.6: Log likelihood scores of the different models used for the Achatinidae using (A) 3648 unambiguously aligned nucleotides from the rRNA gene cluster; (B) 873 unambiguously aligned nucleotides of actin; (B.1) 582 unambiguously aligned nucleotides of the 1st and 2nd codon positions of actin; (B.2) 291 unambiguously aligned nucleotides of the 3rd codon position of actin; (C) 328 unambiguously aligned nucleotides of histone 3; (C.1) 218 unambiguously aligned nucleotides of the 1st and 2nd codon positions of histone 3; (C.2) 110 unambiguously aligned nucleotides of the 3rd codon position of histone 3; (D) 641 unambiguously aligned nucleotides of CO1; (D.1) 427 unambiguously aligned nucleotides of the 1st and 2nd codon positions of CO1; (D.2) 214 unambiguously aligned nucleotides of the 3rd codon position of CO1; and (E) 310 unambiguously aligned nucleotides of the 16S rRNA gene. Scores in **bold** belong to optimal models as determined by Likelihood Ratio Test. Note that the K2P and F81 models are non-nested and therefore cannot be compared for significant difference using the LRT.

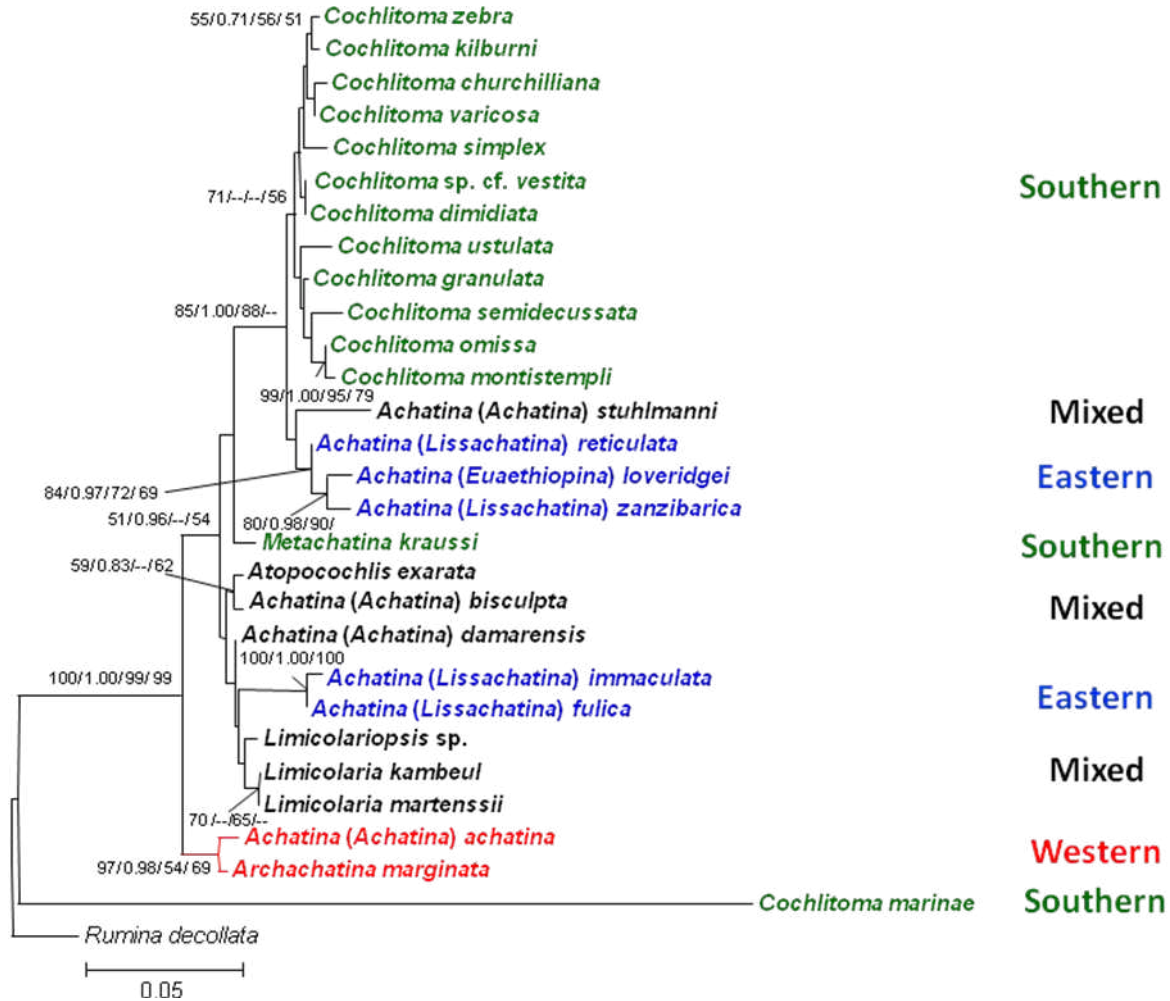
	(A)	(B)	(B.1)	(B.2)	(C)	(C.1)	(C.2)	(D)	(D.1)	(D.2)	(E)
Model	rRNA gene cluster	Actin-combined	Actin-1 st +2 nd codons	Actin 3 rd codons	Histone 3	Histone 3 1 st +2 nd codons	Histone 3 3 rd codons	CO1	CO1 1 st +2 nd codons	CO1 3 rd codons	16S rRNA
JC69	6248.60194	3079.30195	1033.47821	1770.38462	961.35579	330.69952	547.08597	10577.13076	2639.75972	6467.22043	3410.68577
JC69+ Γ	6274.56535	2941.42677	1019.80247	1748.03013	944.88406	330.69952	545.05987	9044.70208	2207.91549	6415.07916	3057.86503
F81	6187.62669	3067.34670	1028.44871	1747.01512	951.97785	325.87231	537.40127	10519.45591	2597.94022	6379.86740	3392.31335
F81+ Γ	6213.92744	2928.60315	1015.11409	1722.19734	938.63868	325.87231	535.75438	8289.79099	2137.55284	6334.75953	3022.41771
K2P	6217.14469	2999.90791	1029.51509	1690.58148	942.44107	330.63592	520.04898	10305.12794	2494.32907	6291.12771	3332.43652
K2P+ Γ	6233.51676	2856.89829	1018.80719	1657.31863	927.23046	330.63592	525.37382	8654.44378	2053.68656	6219.17666	2961.52886
HKY85	6155.43566	2987.60266	1024.45322	1677.30224	932.61470	325.81310	514.06374	10214.37634	2447.65021	6026.06510	3311.76468
HKY85+ Γ	6173.49112	2847.31250	1013.84964	1646.32805	909.37626	325.81310	513.02241	8294.44610	2020.38852	5887.87774	2902.18513
TrN93	6146.48364	2976.74300	1019.97904	1677.26346	932.53700	324.88259	513.02953	10210.89277	2422.04372	6023.81427	3308.08745
TrN93+ Γ	6164.98831	2837.76616	1013.43024	1644.81901	909.25171	324.88259	512.04887	8289.71414	2015.28913	5879.85256	2900.05462
GTR	6143.87015	2967.76023	1018.79267	1663.08220	931.83493	321.54407	512.77514	9954.78203	2405.51236	6018.45857	3243.11777
GTR+ Γ	6157.24483	2821.80122	1011.67014	1634.16756	908.90524	321.54420	511.78485	8289.46582	1995.68125	5878.05759	2876.32253

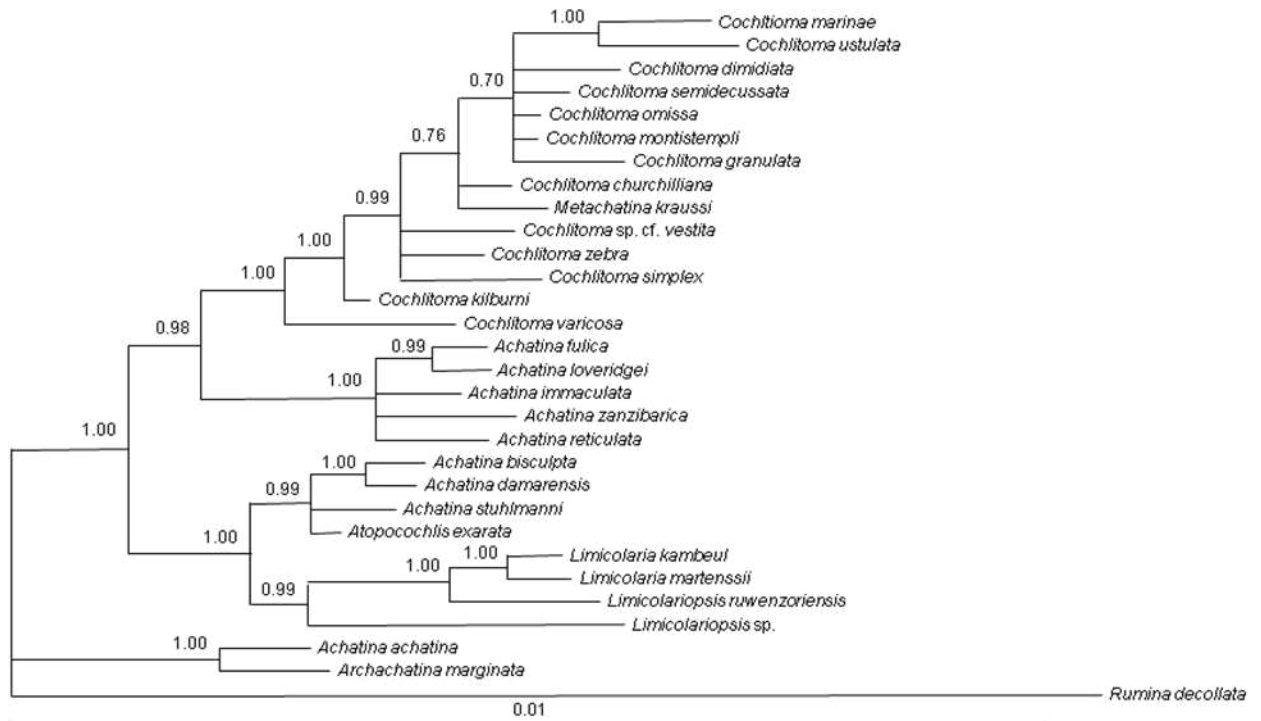
Appendix 4.7: Summary of ambiguous sites for the actin gene for the Achatinidae

Species	Total ambiguous sites	% ambiguous sites (out of 873 total)	Total ambiguous 3 rd codon positions	% ambiguous 3 rd codon positions relative to all ambiguous sites	Number of RY sites at ambiguous 3 rd codon positions	% RY sites relative to all ambiguous 3 rd codon positions	Number of ambiguous amino acids	% ambiguous amino acids (out of 291 total)
WEST AFRICAN								
acha	45	5.15	40	88.89	31	77.50	4	1.37
arcm	27	3.09	23	85.19	21	91.30	3	1.03
EAST AFRICAN								
achf	18	2.06	16	88.89	12	75.00	2	0.69
achi	15	1.72	12	80.00	11	91.67	1	0.34
achr	33	3.78	26	78.79	23	88.46	4	1.37
achz	1	0.11	0	0	0	0	1	0.34
euel	0	0	0	0	0	0	0	0
OTHERS								
achb	32	3.67	26	81.25	17	65.38	4	1.37
achd	34	3.89	27	79.41	25	92.59	6	2.06
achs	2	0.23	1	50.00	1	100.00	1	0.34
atcx2	29	3.32	25	86.21	21	84.00	3	1.03
lim	30	3.44	27	90.00	25	92.59	2	0.69
limm	33	3.78	27	81.82	23	85.19	7	2.41
lip	40	4.58	32	80.00	27	84.38	7	2.41
SOUTHERN AFRICAN								
achv	27	3.09	24	88.89	18	75.00	2	0.69
arcd	31	3.55	27	87.10	20	74.07	3	1.03
arcv	30	3.32	26	86.67	19	73.08	3	1.03
coia	4	0.46	2	50.00	1	50.00	2	0.69
coic	0	0	0	0	0	0	0	0
coik	1	0.11	0	0	0	0	0	0
coig	9	1.03	9	100.00	8	88.89	0	0
coii	2	0.23	2	100.00	0	0	0	0
coim	0	0	0	0	0	0	0	0
coio	6	0.69	6	100.00	5	83.33	0	0
cois	0	0	0	0	0	0	0	0
coiu	0	0	0	0	0	0	0	0
coiz	0	0	0	0	0	0	0	0
metk	52	5.96	38	73.08	29	76.32	9	3.09

NOTE: primers used: ActF1 and ActR

Appendix 4.8. Maximum likelihood phylogenetic tree of the Achatinidae based on 873 unambiguously aligned nucleotide sites of the actin gene. The phylogenies were rooted on the subulinid *Rumina decollata*. Values on the nodes represent bootstrap support (1000 replicates) for ML, posterior probabilities (based on the last 1000 trees) for BI, and bootstrap supports for NJ (1000 replicates) and MP (69 replicates only after the PAUP* version 4.0b10 was terminated after running for 135 hours). The scale bar represents 5 substitutional changes per 100 nucleotide positions.

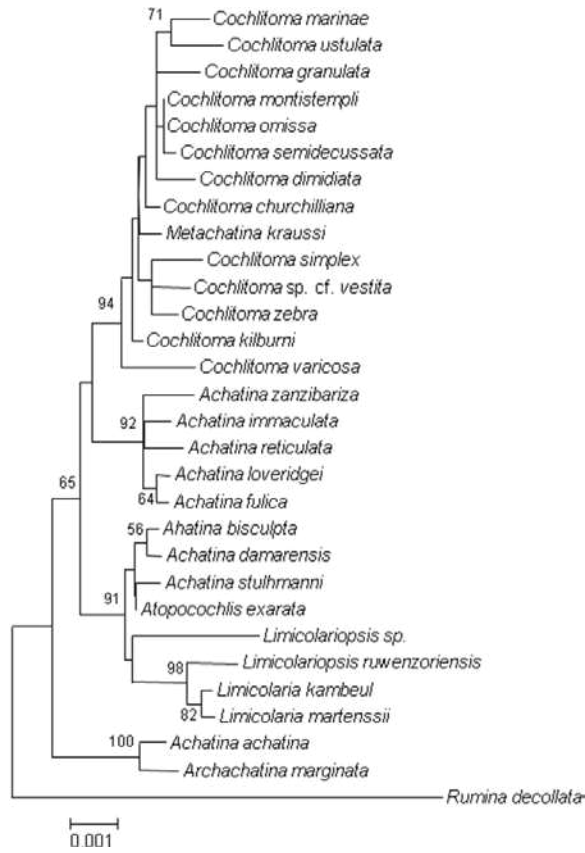




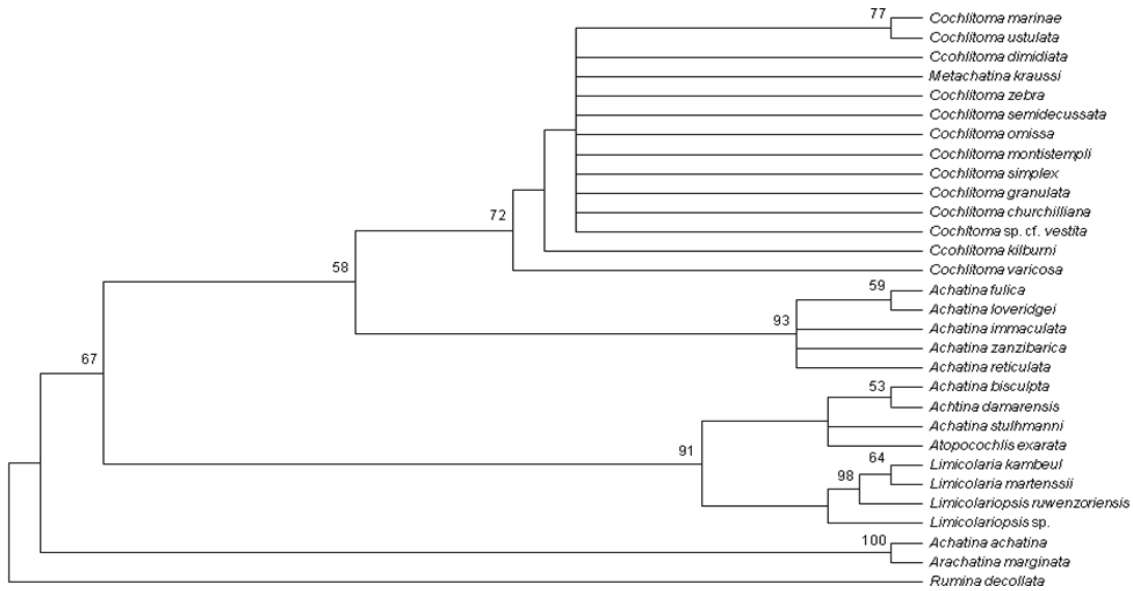
(A) BI (LSU rRNA)

Appendix 4.9: Phylogenetic trees of the Achatinidae based on the large subunit rRNA gene using (A) Bayesian analysis, (B) neighbor-joining (p. 512) and (C) maximum parsimony based on 25 equally parsimonious trees (p. 512). The phylogenies were constructed from 3648 unambiguously aligned nucleotide sites and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 100 nucleotides and that of (B) represents 1 substitutional change per 1000 nucleotides.

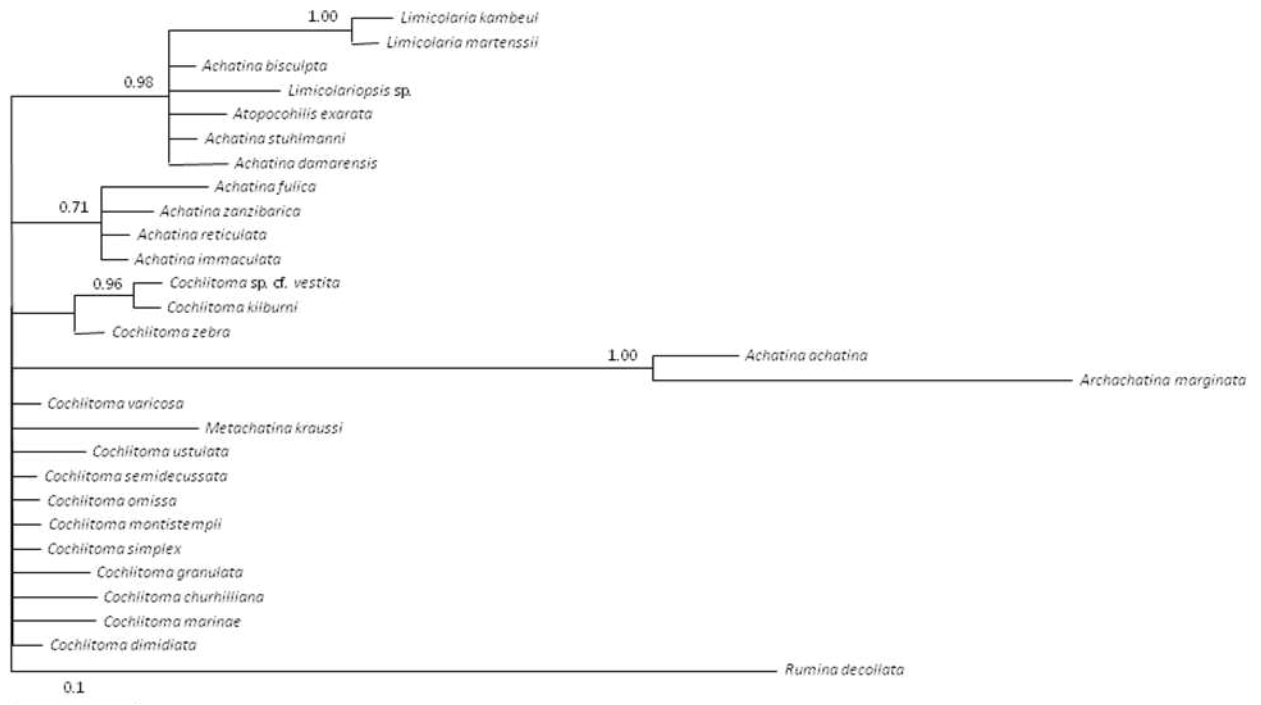
Appendix 4.9 (contd.)



(B) NJ (LSU rRNA)



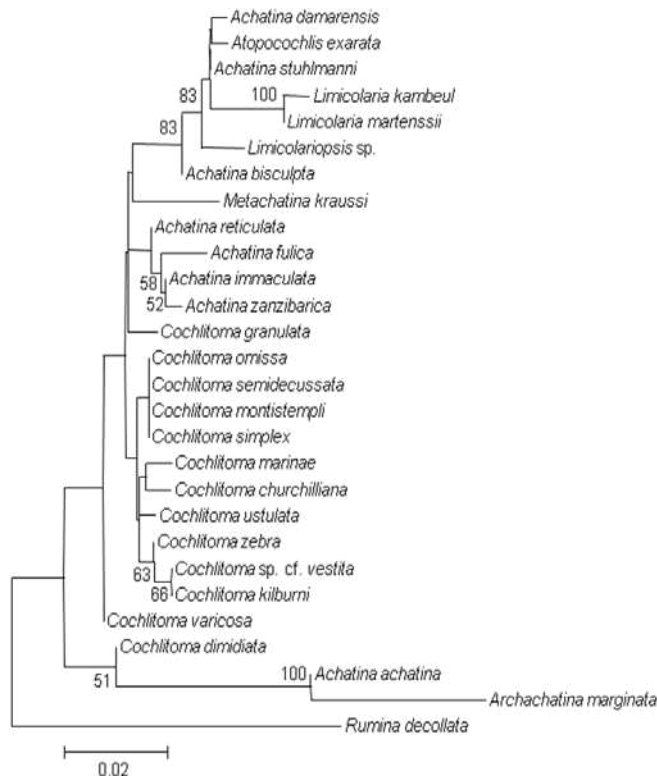
(C) MP (LSU rRNA)



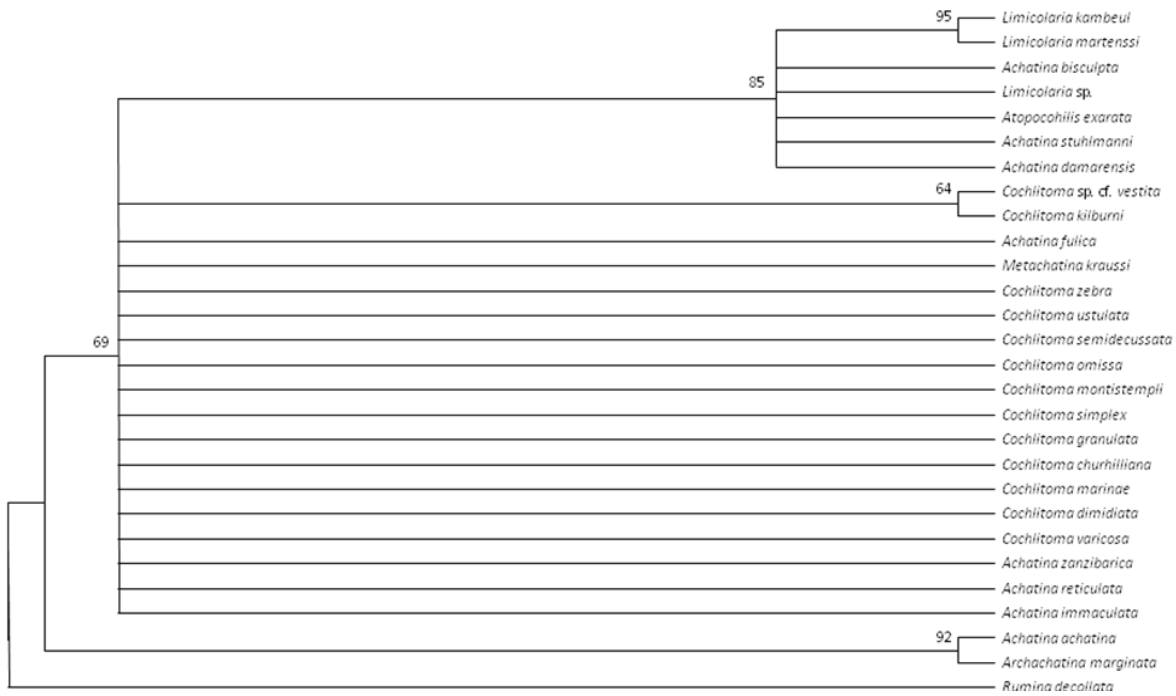
(A) BI (histone 3)

Appendix 4.10: Phylogenetic trees of the Achatinoidea based on the histone 3 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 514) and (C) maximum parsimony based on 8404 equally parsimonious trees (p. 514). The phylogenies were constructed from 328 unambiguously aligned nucleotide sites and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 10 nucleotides while that of (B) represents 2 substitutional changes per 100 nucleotides. Note that *Achatina loveridgei* and *Limicolariopsis ruwenzoriensis* were not included in the phylogenies as they were not successfully sequenced.

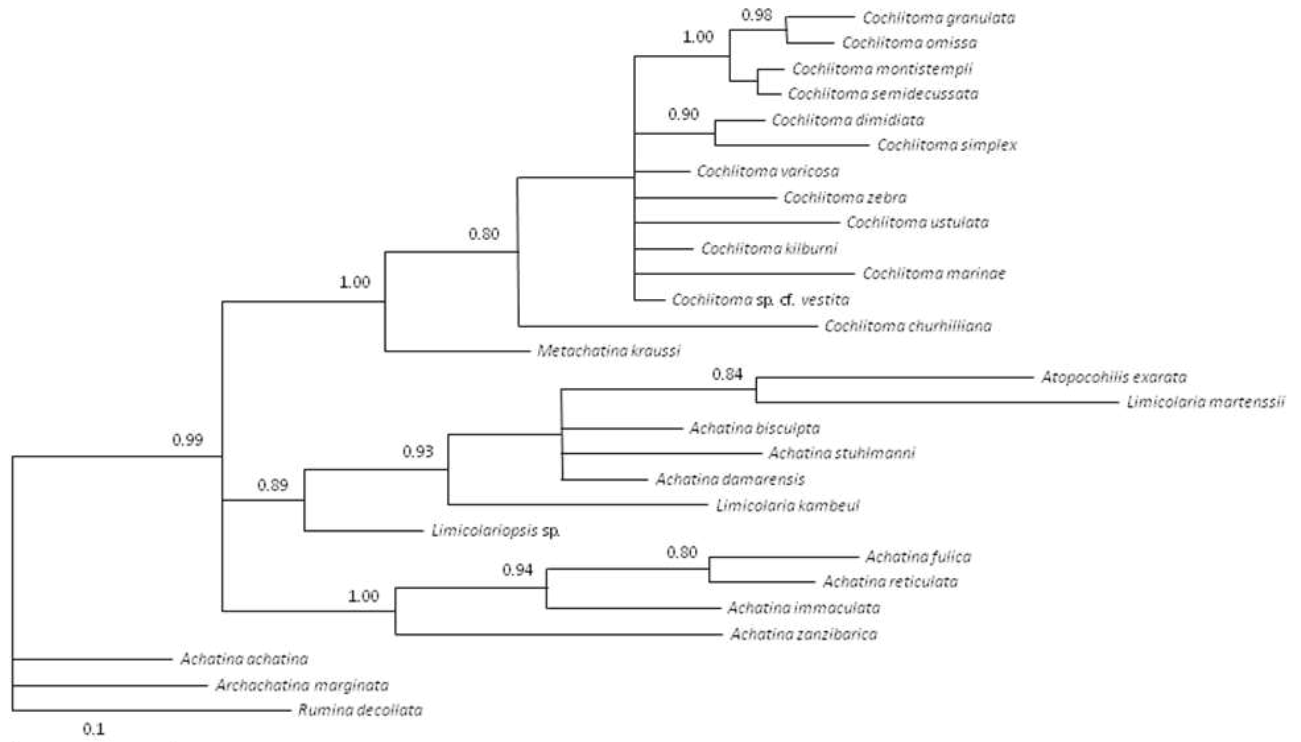
Appendix 4.10 (contd.)



(B) NJ (histone 3)



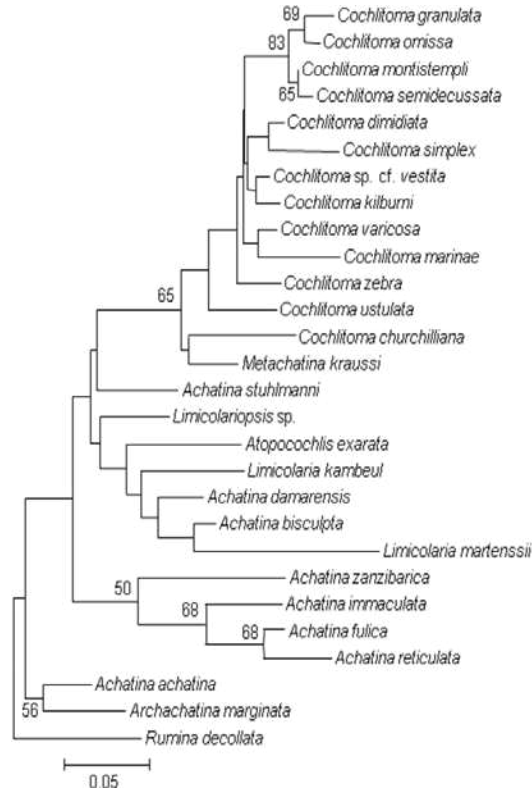
(C) MP (histone 3)



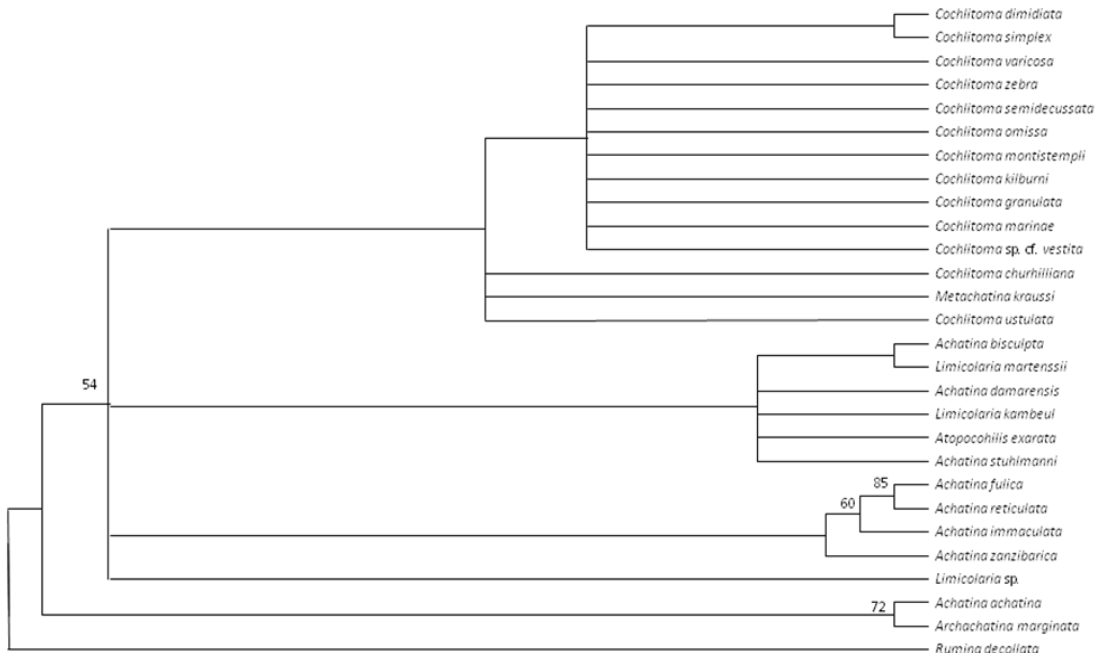
(A) BI (1st and 2nd codon positions of CO1 gene)

Appendix 4.11: Phylogenetic trees of the Achatinidae based on the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 516) and (C) maximum parsimony based on 127 equally parsimonious trees (p. 516). The phylogenies were constructed from 427 unambiguously aligned nucleotide sites and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) represents 1 substitutional change per 10 nucleotides and that of (B) represents 5 substitutional changes per 100 nucleotides. Note that *Achatina loveridgei* and *Limicolariopsis ruwenzoriensis* were not included in the phylogenies as they were not successfully sequenced.

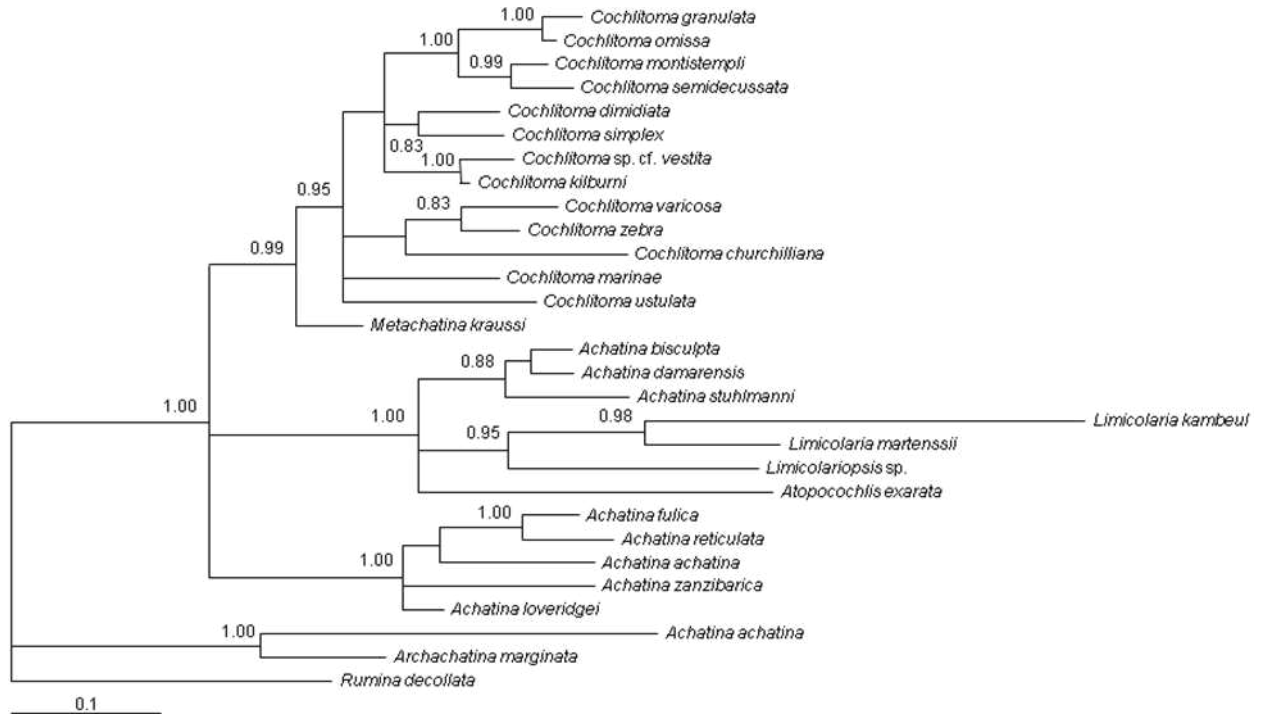
Appendix 4.11 (contd.)



(B) NJ (1st and 2nd codon positions of the CO1 gene)



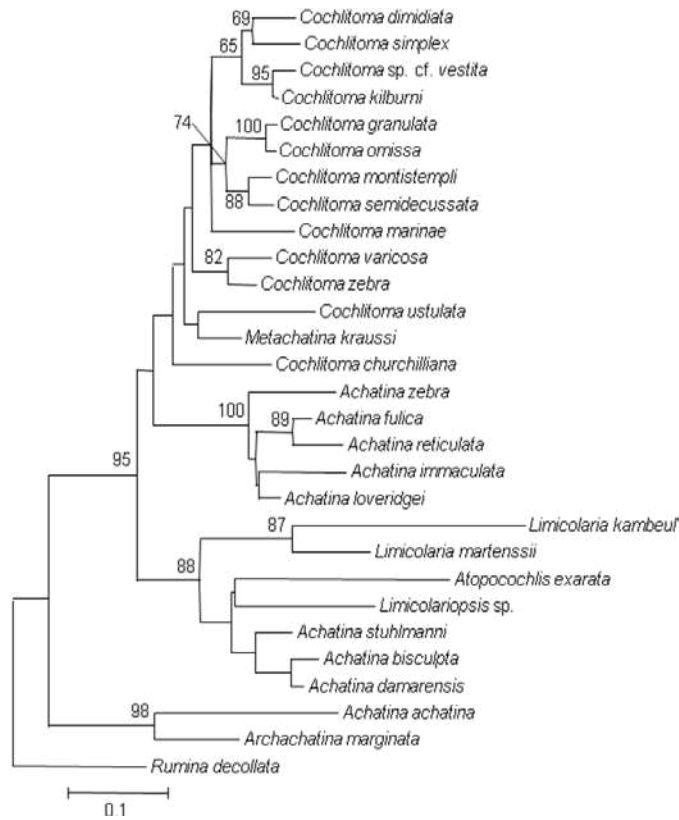
(C) MP (1st and 2nd codon positions of the CO1 gene)



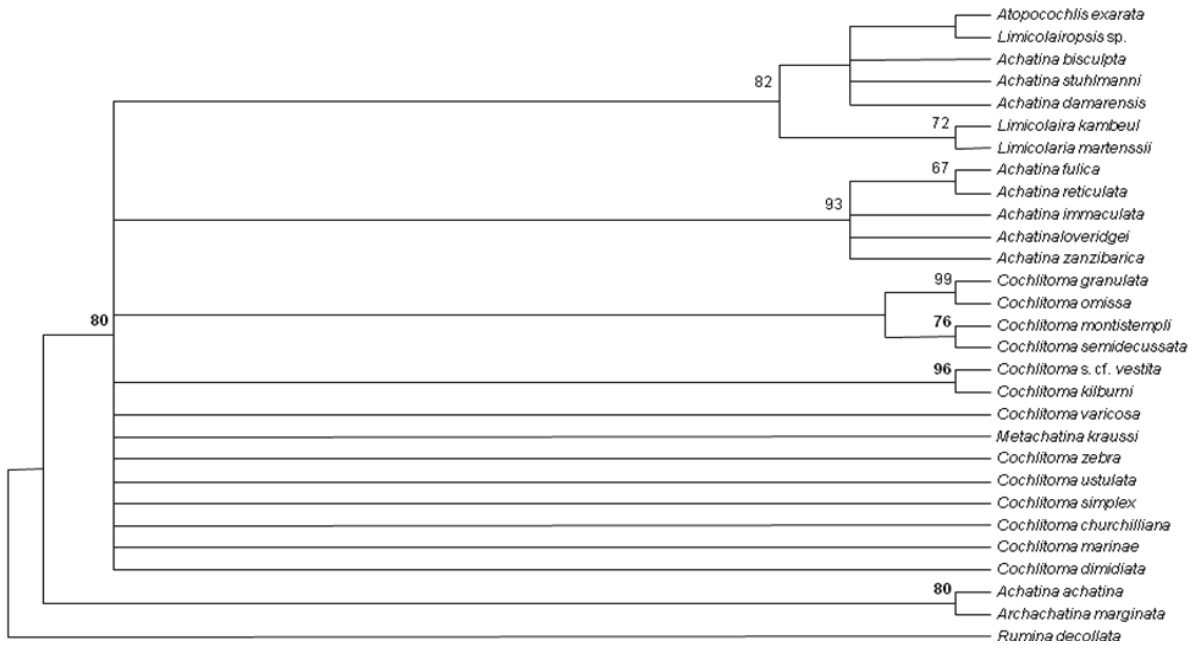
(A) BI (16S rRNA gene)

Appendix 4.12: Phylogenetic trees of the Achatinidae based on the 16S rRNA gene using (A) Bayesian analysis, (B) neighbor-joining (p. 518) and (C) maximum parsimony based on 11 equally parsimonious trees (p. 518). The phylogenies were constructed from 310 unambiguously aligned nucleotide sites and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bar for (A) and (B) represents 1 substitutional change per 10 nucleotides. Note that *Limicolariopsis ruwenzoriensis* was excluded in the analyses as it was not successfully sequenced.

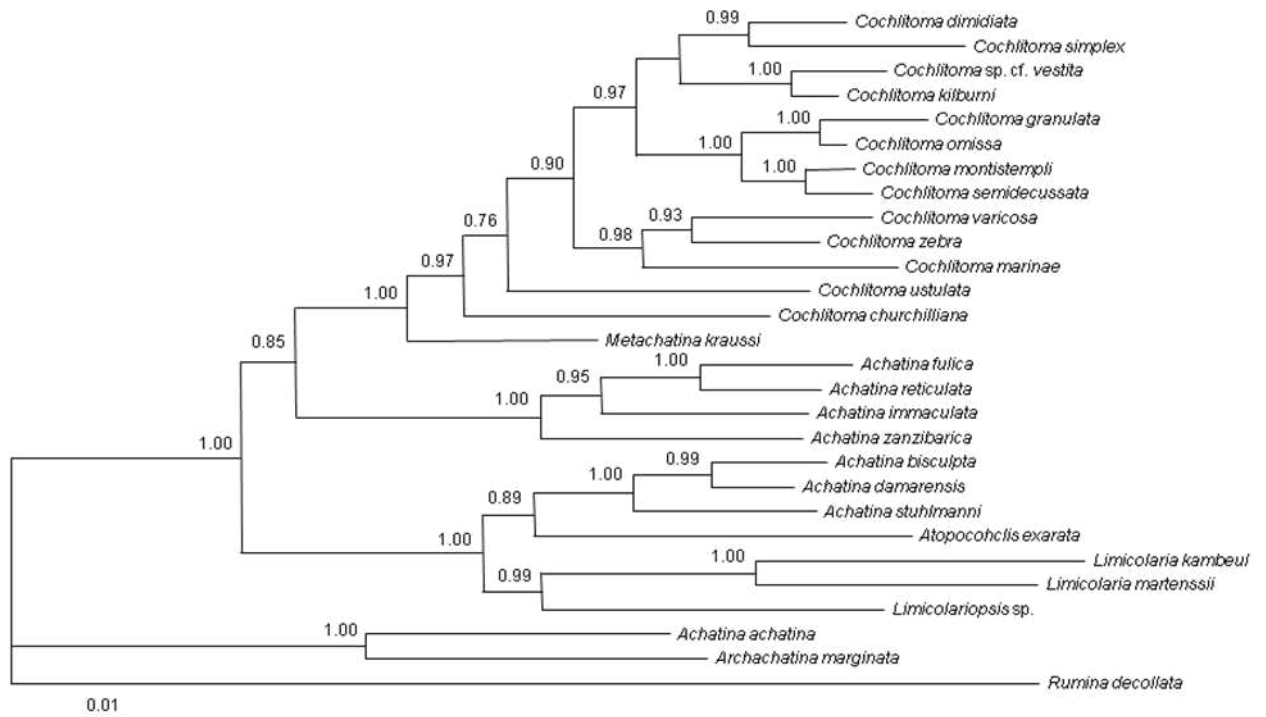
Appendix 4.12 (contd.)



(B) NJ (16S rRNA gene)



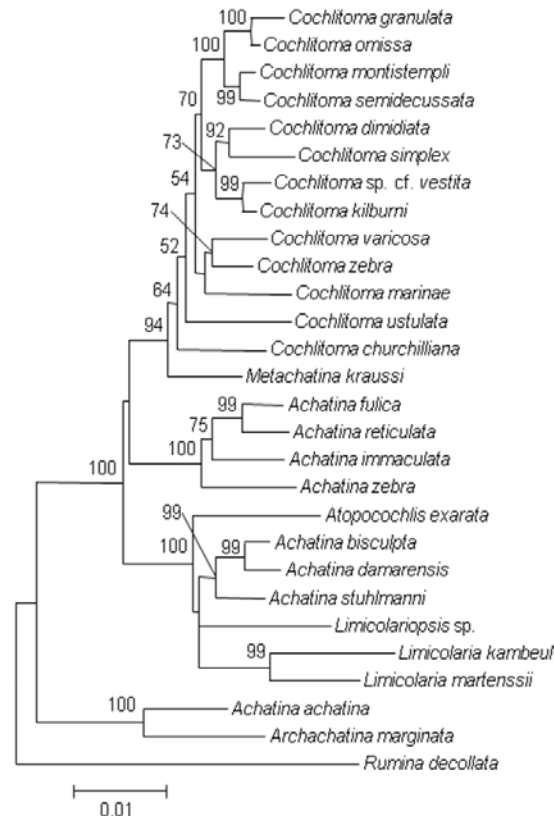
(C) MP (16S rRNA gene)



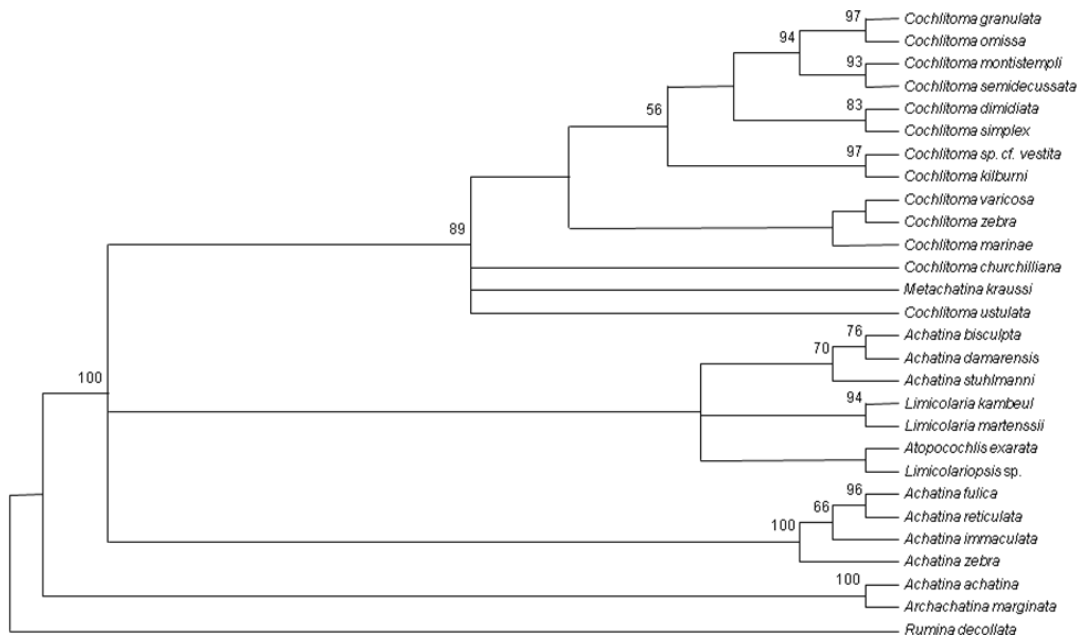
(A) BI (concatenated sequence – taxa with complete datasets only)

Appendix 4.13: Phylogenetic trees of the Achatinidae (taxa with complete datasets only) based on the combined dataset of the LSU rRNA, histone 3, the 1st and 2nd codon positions of the CO1 gene and the 16S rRNA using (A) Bayesian analysis, (B) neighbor-joining (p. 520) and (C) maximum parsimony based on four equally parsimonious trees (p. 520). The phylogenies were constructed from a concatenated sequence of 4713 nucleotides and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bars for (A) and (B) represent 1 substitutional change per 100 nucleotides. Note that *Achatina loveridgei* and *Limicolariopsis ruwenzoriensis* were not included in the phylogenies as they lack at least one of the genes used for the combined analyses.

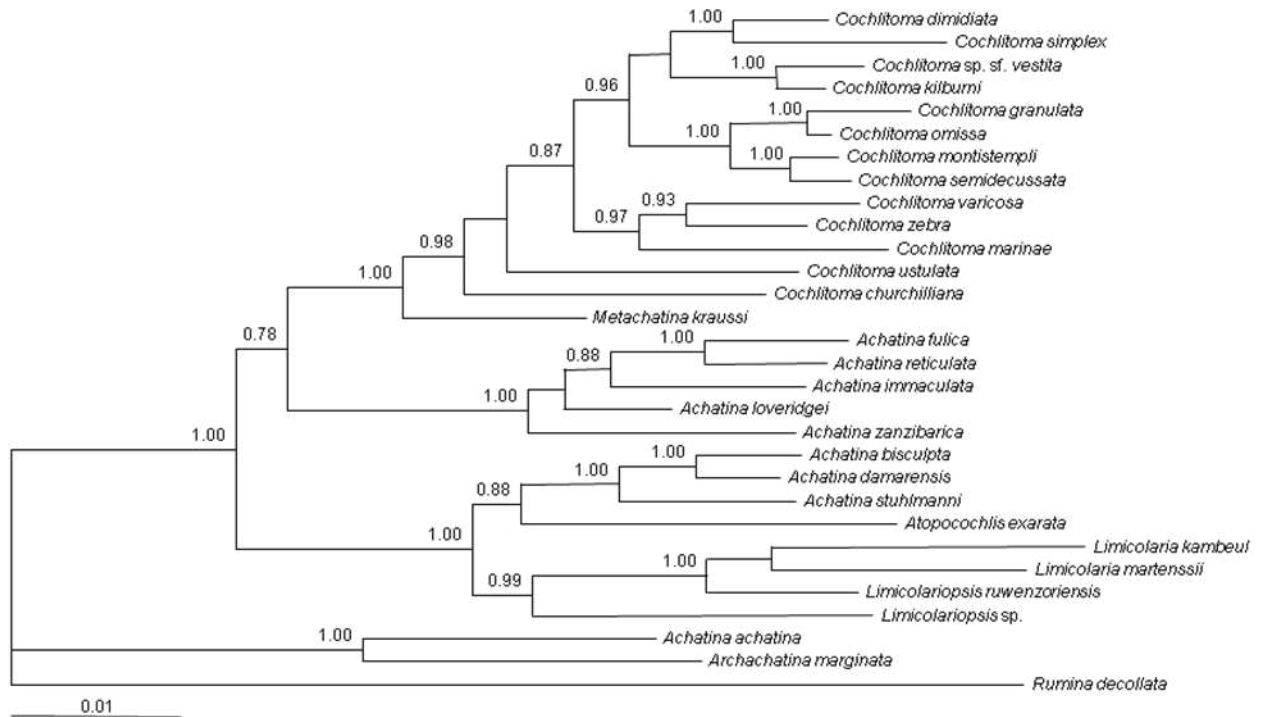
Appendix 4.13 (contd.)



(B) NJ (concatenated sequence – taxa with complete datasets only)



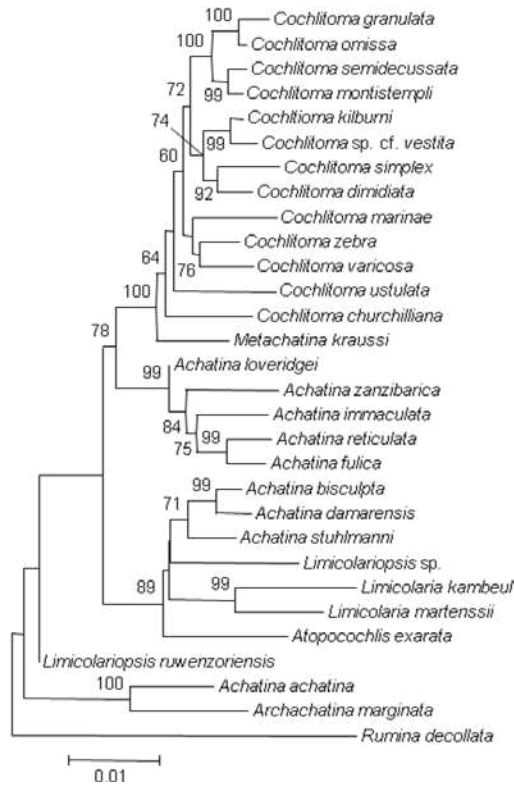
(C) MP (concatenated sequence – taxa with complete datasets only)



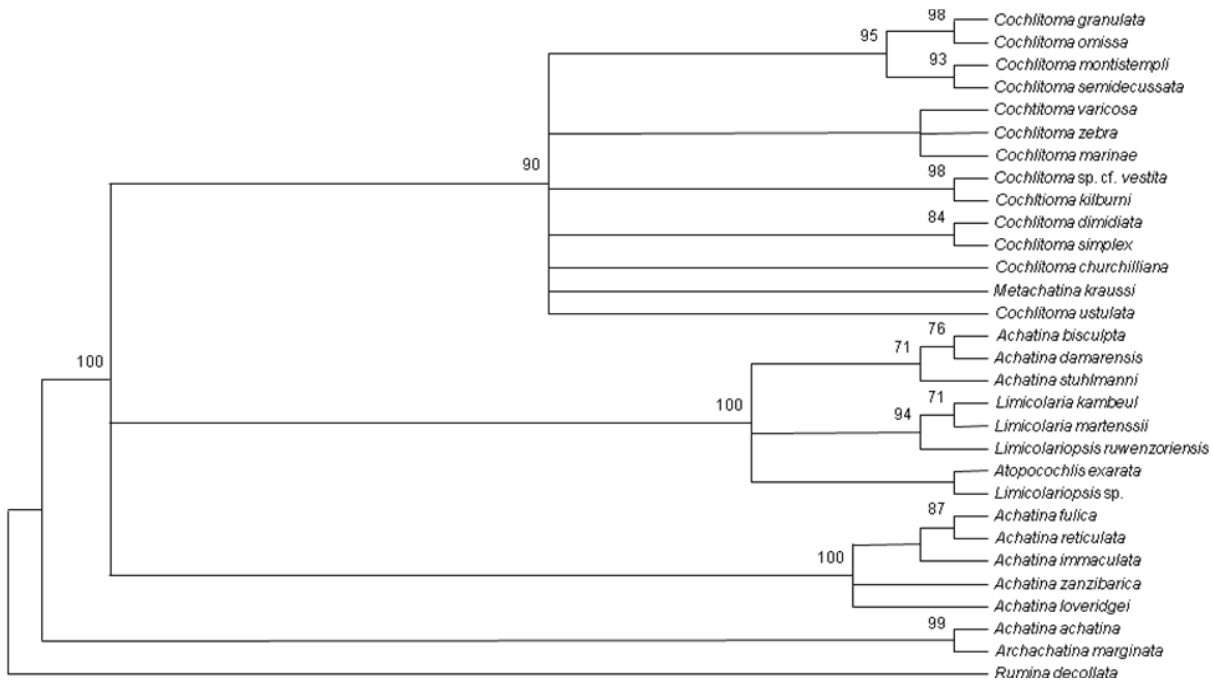
(A) BI (concatenated sequence – all taxa)

Appendix 4.14: Phylogenetic trees of the Achatinidae (all taxa) based on the combined dataset of the LSU rRNA, actin and histone 3 genes and the 1st and 2nd codon positions of the CO1 gene using (A) Bayesian analysis, (B) neighbor-joining (p. 522) and (C) maximum parsimony based on eight equally parsimonious trees (p. 522). The phylogenies were constructed from a concatenated sequence of 4713 nucleotides and were rooted on the subulinid *Rumina decollata*. Bayesian posterior probabilities in (A) indicate support for individual branches in the Bayesian analysis based on the last 1000 trees. Posterior probabilities lower than 0.7 are not shown. Bootstrap values in (B) and (C) indicate the percentage support for individual branches based on 1000 replicates. Bootstrap values lower than 50% are not shown. The scale bars for (A) and (B) represent 1 substitutional change per 100 nucleotides.

Appendix 4.14 (contd.)



(B) NJ (concatenated sequence – all taxa)



(C) MP (concatenated sequence – all taxa)

Appendix 5.1: Pair-wise uncorrected distances of the East African *Achatina* species using the SSCP fragment of the 16S rRNA gene (293 nucleotides for *A. fulica*).

	<i>A. fulica</i>	<i>A. zanzibarica</i>	<i>A. immaculata</i>
<i>A. fulica</i>	--		
<i>A. zanzibarica</i>	0.24518		
<i>A. immaculata</i>	0.22864	0.26893	
<i>A. reticulata</i>	0.14414	0.24952	0.22459

Appendix 5.2: Alignment of the 15 haplotypes for the global *Achatina fulica* populations based on 293 nucleotides of the 16S rRNA gene. Sequence for *Achatina reticulata* was used as outgroup.

	10	20	30	40	50	60
Haplotype A	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype B	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype C	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype D	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype E	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype F	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype G	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype H	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype I	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype J	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype K	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype L	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype M	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype N	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
Haplotype O	TAATTTGTCC	TCTAATTAAG	GTCTGGAATG	AAGGGGGACA	CAGGGGAGAG	CTGTCTCCAA
<i>A. reticulata</i>	TAATTTGTCC	TTTAATTTAG	GTCTGGAATG	AATGAGAACA	CAGGGGAGAG	CTGTCTCTAG

	70	80	90	100	110	120
Haplotype A	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype B	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype C	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype D	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype E	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype F	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype G	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCTGATG	AAAGACGAGA
Haplotype H	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype I	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype J	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GTCTCCGATG	AAAGACGAGA
Haplotype K	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
Haplotype L	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GGCTCCGATG	AAAGACGAGA
Haplotype M	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GTCTCCGATG	AAAGACGAGA
Haplotype N	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GTCTCCGATG	AAAGACGAGA
Haplotype O	TAAGGTTAAT	TTAACTTTCT	TATCAGGTGA	AAATTCCTGA	GCCTCCGATG	AAAGACGAGA
<i>A. reticulata</i>	TAAGGTTAAC	TTAACTTCT	TATCAGGTGA	AAATTCCTGA	GCTTCTTTGG	AAAGACGAGA

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130      140      150      160      170      180
Haplotype A  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATTA AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype B  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATTA AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype C  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATTA AAGAT TCTTGTTCTG TTTTGTGTTGG
Haplotype D  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATTA AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype E  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATTA AAGAT TCTTGTTCTG TTTTGTGTTGG
Haplotype F  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATCA AAGAT TCTTGTTCTG TTTTGTGTTGG
Haplotype G  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATTA AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype H  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATTA AAGAT TCTTGTTCTG TTTTGTGTTGG
Haplotype I  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT GTATTT AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype J  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT GTATTT AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype K  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT ATATTA AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype L  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT GTATTT AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype M  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT GTATTT AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype N  AGACCCTTAG AGTTTTTTATT ATAACCTTGAT GTATTT AAAAT TCTTGTTCTG TTTTGTGTTGG
Haplotype O  AGACCCTTAG AGTTTTTTATT ATAATTTGAT ATATTA AAAAT TCTTGTTCTG TTTTGTGTTGG
A. reticulata AGACCCTTAG AGTTTTTAATA ATTATTTAGA TAAATA AAAAT TCTACTATG TTTTGTGTTGG

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190      200      210      220      230      240
Haplotype A  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype B  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype C  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype D  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype E  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype F  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype G  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype H  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype I  GGCGACAGGG TTACAACAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype J  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype K  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype L  GGCGACAGGG TTACAACAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype M  GGCGACAGGG TTACAACAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype N  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
Haplotype O  GGCGACAGGG TTACAATAGA TAA-C TTACC CTACCATA-- TTTTATATG GCGATCATT
A. reticulata GGCGACAGGG TAACAATAGG TAAACTTACC CATTCATATA TTTTATGCG TCGATAGTTA

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.....|.....| .....|.....| .....|.....| .....|.....| .....|..
250      260      270      280      290
Haplotype A  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTTG GTTTGC
Haplotype B  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTTG GTTTGC
Haplotype C  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTT-G GTTTGC
Haplotype D  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTT-G GTTTGC
Haplotype E  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTT-G GTTTGC
Haplotype F  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTT-G GTTTGC
Haplotype G  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTT-G GTTTGC
Haplotype H  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTTG GTTTGC
Haplotype I  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTTG GTTTGC
Haplotype J  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTATTT-G GTTTGC
Haplotype K  TACTTTTAAT TAAGCTACCT GAGGGATAAC AGCGTAATCT TTTTTTTT--G GTTTGC
Haplotype L  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTT--G GTTTGC
Haplotype M  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTT-G GTTTGC
Haplotype N  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTT-G GTTTGC
Haplotype O  TACTTTTAAT TAAGCTACCT AAGGGATAAC AGCGTAATCT TTTTATTTT-G GTTTGC
A. reticulata TACTAATAAT TAAACTACCT AAGGGATAAC AGCGTAATCT TTTTTTTTT-G GTTTGC

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Appendix 5.3: Log likelihood scores of the different models used for comparison for the 16S haplotypes of the global *A. fulica* populations using 293 nucleotides of the 16S rRNA gene. Likelihood Ratio Test revealed that GTR and TrN93 models were not significantly better than HKY85; the latter was therefore used for the phylogenetic analysis. Note that the K2P and F81 models are non-nested and therefore cannot be compared using LRT.

Model	Log-Likelihood Score
JC69	697.00855
JC69+ Γ	696.73625
F81	676.85077
F81+ Γ	676.50788
K2P	692.34383
K2P+ Γ	691.96730
HKY85	670.85793
HKY85+ Γ	670.25753
TN93	670.82611
TN93+ Γ	670.23779
GTR	666.27323
GTR+ Γ	666.03208

Appendix 6.1: Nematode species used for the *Angiostrongylus cantonensis* survey as found in GenBank Release No. 157 (22 December 2006) or GenBank Release No. 158 (15 February 2007) as indicated by (*).

Species	Accession No.	Taxonomy
<i>Gordius aquaticus</i>	X87985	Nematomorpha; Gordioida
<i>Chordodes morgani</i>	AF036639	Nematomorpha; Gordioida
<i>Priapulius caudatus</i>	Z38009	Priapulida; Priapulidae
<i>Brachionus plicatilis</i>	U49911	Rotifera; Monogononta
<i>Axonolaimus helgolandicus</i>	AY854232	Nematoda; Chromadorea; Araeolaimida; Axonolaimoidea; Axonolaimidae
<i>Ascolaimus elongatus</i>	AY854231	Nematoda; Chromadorea; Araeolaimida; Axonolaimoidea; Axonolaimidae
<i>Odontophora rectangula</i>	AY854233	Nematoda; Chromadorea; Araeolaimida; Axonolaimoidea; Axonolaimidae
<i>Cylindrolaimus</i> sp. 202149	AF202149	Nematoda; Chromadorea; Araeolaimida; Cylindrolaimidae
<i>Tylocephalus auriculatus</i>	AF202155	Nematoda; Chromadorea; Araeolaimida; Plectoidea; Plectidae
<i>Plectus acuminatus</i> strain BS9	AF037628	Nematoda; Chromadorea; Araeolaimida; Plectoidea; Plectidae
<i>Plectus aquatilis</i>	AF036602	Nematoda; Chromadorea; Araeolaimida; Plectoidea; Plectidae
<i>Anaplectus</i> sp. PDL-2005	AJ966473	Nematoda; Chromadorea; Araeolaimida; Plectoidea; Plectidae
<i>Anisakis</i> sp. WKT	U81575	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Anisakis</i> sp. Nadler	U94365	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Contraecum multipapillatum</i>	U94370	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Pseudoterranova decipiens</i>	U94380	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Raphidascaris acus</i> *	DQ503460	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Terranova caballeroi</i>	U94382	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Anisakidae
<i>Ascaris suum</i>	U94367	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Ascaris lumbricoides</i>	U94366	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Baylisascaris procyonis</i>	U94368	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Baylisascaris transfuga</i>	U94369	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Parascaris equorum</i>	U94378	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Porrocaecum depressum</i>	U94379	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Toxascaris leonine</i>	U94383	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Ascarididae
<i>Heteroncheilus tunicatus</i>	U94373	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Heterocheilidae
<i>Goezia pelagia</i>	U94372	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Hysterothylacium fortalezae</i>	U94374	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Hysterothylacium pelagicum</i>	U94375	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Hysterothylacium reliquens</i>	U94376	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Iheringascaris inquires</i>	U94377	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Raphidascarididae
<i>Toxocara canis</i>	U94382	Nematoda; Chromadorea; Ascaridida; Ascaridoidea; Toxocaridae
<i>Nemhelix bakeri</i>	DQ118537	Nematoda; Chromadorea; Ascaridida; Cosmocercidae; Cosmocercidae
<i>Raillietnema</i> sp. V3060*	DQ503461	Nematoda; Chromadorea; Ascaridida; Cosmocercidae; Cosmocercidae
<i>Cruzia Americana</i>	U94371	Nematoda; Chromadorea; Ascaridida; Cosmocercidae; Kathlaniidae
<i>Paraspidodera</i> sp. 21303	AF083005	Nematoda; Chromadorea; Ascaridida; Heterakoidea; Aspidoderidae

Species	Accession No.	Taxonomy
<i>Heterakis gallinarum*</i>	DQ503462	Nematoda; Chromadorea; Ascaridida; Heterakoidea; Heterakidae
<i>Heterakis sp.14690</i>	AF083003	Nematoda; Chromadorea; Ascaridida; Heterakoidea; Heterakidae
<i>Chromadoropsis vivipara</i>	AF047891	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Chromadora nudicapitata</i>	AY854205	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Chromadora sp. BHMM_2005</i>	AY854206	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Atrochromadora microlaima</i>	AY854204	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Chromadorita tentabundum</i>	AY854208	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Chromadorina germanica</i>	AY854207	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Dichromadora sp. BHMM_2005</i>	AY854209	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Neochromadora BHMM_2005</i>	AY854210	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Spilophorella paradoxa</i>	AY854211	Nematoda; Chromadorea; Chromadorida; Chromadoridae
<i>Paracanthonchus caecus</i>	AF047888	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Paracyatholaimus intermedius</i>	AJ966495	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Praeacanthonchus punctatus</i>	AY854214	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Praeacanthonchus sp.</i>	AF036612	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Cyatholaimus sp. BHMM_2005</i>	AY854213	Nematoda; Chromadorea; Chromadorida; Cyatholaimidae
<i>Spirinia parasitifera</i>	AY854217	Nematoda; Chromadorea; Desmodorida; Desmodoridae
<i>Acanthopharynx micans</i>	Y16911	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Desmodorinae
<i>Desmodora ovigera</i>	Y16913	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Desmodorinae
<i>Desmodora communis</i>	AY854215	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Desmodorinae
<i>Xyzzors sp.</i>	Y16923	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Desmodorinae
<i>Metachromadora sp.</i>	AF036595	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Spiriniinae
<i>Metachromadora remanei</i>	AY854216	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Spiriniinae
<i>Catanema sp.</i>	Y16912	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Eubostrichus diana</i>	Y16915	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Eubostrichus topiarus</i>	Y16917	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Eubostrichus parasitiferus</i>	Y16916	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Laxus oneistus</i>	Y16919	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Laxus cosmopolites</i>	Y16918	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Leptonemella sp.</i>	Y16920	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Robbea hypermnestra</i>	Y16921	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Stilbonema majum</i>	Y16922	Nematoda; Chromadorea; Desmodorida; Desmodoridae; Stilbonematinae
<i>Monoposthia costata</i>	AY854221	Nematoda; Chromadorea; Desmodorida; Monoposthiidae
<i>Nudora bipapillata</i>	AY854222	Nematoda; Chromadorea; Desmodorida; Monoposthiidae
<i>Calomicrolaimus parahonestus</i>	AY854218	Nematoda; Chromadorea; Desmodorida; Richtersioidea; Microlaimidae
<i>Calomicrolaimus sp. BHMM_2005</i>	AY854219	Nematoda; Chromadorea; Desmodorida; Richtersioidea; Microlaimidae
<i>Molgolaimus demani</i>	AY854220	Nematoda; Chromadorea; Desmodorida; Richtersioidea; Microlaimidae
<i>Diplogaster lethieri</i>	AF036643	Nematoda; Chromadorea; Diplogasterida; Diplogasteridae

Species	Accession No.	Taxonomy
<i>Aduroscopiculum halicti</i>	U61759	Nematoda; Chromadorea; Diplogasterida; Diplogasteridae
<i>Pristionchus lheritieri</i>	AF036640	Nematoda; Chromadorea; Diplogasterida; Neodiplogasteridae
<i>Pristionchus pacificus</i>	U81584	Nematoda; Chromadorea; Diplogasterida; Neodiplogasteridae
<i>Pristionchus pacificus</i> PS312	AF083010	Nematoda; Chromadorea; Diplogasterida; Neodiplogasteridae
<i>Sabatieria punctata</i> Strain 343	AY854237	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria sp.</i> 355 BHMM_2005	AY854239	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria celtica</i>	AY854234	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria punctata</i> Strain 200	AY854235	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria punctata</i> Strain 223	AY854236	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Sabatieria sp.</i> 210 BHM_2005	AY854238	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Setosabatieria hilarula</i>	AY854240	Nematoda; Chromadorea; Monhysterida; Comesomatidae
<i>Desmolaimus zeelandicus</i>	AY854229	Nematoda; Chromadorea; Monhysterida; Linhomoeidae
<i>Terschellingia longicaudata</i>	AY854230	Nematoda; Chromadorea; Monhysterida; Linhomoeidae
<i>Cyartonema elegans</i>	AY854203	Nematoda; Chromadorea; Monhysterida; Monhysterida incertae sedis
<i>Tridentulus sp.</i> PDL-2005	AJ966507	Nematoda; Chromadorea; Monhysterida; Monhysteridae
<i>Diplolaimelloides meylli</i>	AJ966507	Nematoda; Chromadorea; Monhysterida; Monhysteridae
<i>Diplolaimella dievengatensis</i>	AJ966482	Nematoda; Chromadorea; Monhysterida; Monhysteridae
<i>Geomonhystera disjuncta</i>	AJ966485	Nematoda; Chromadorea; Monhysterida; Monhysteridae
<i>Sphaerolaimus hirsutus</i>	AY854228	Nematoda; Chromadorea; Monhysterida; Sphaerolaimidae
<i>Theristus acer</i>	AJ966505	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema procerus</i>	AF047889	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema hirsutum</i>	AY854223	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema normandicum</i>	AY854224	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema oxycerca</i>	AY854225	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Daptonema setosum</i>	AY854226	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Metadesmolaimus sp.</i> PDL-2005	AJ966491	Nematoda; Chromadorea; Monhysterida; Xyalidae
<i>Dentostomella sp.</i>	AF036590	Nematoda; Chromadorea; Oxyurida; Oxyuroidea; Heteroxyematidae
<i>Bunonema franzi</i>	AJ966477	Nematoda; Chromadorea; Rhabditida; Bunonematoidea; Bunonematidae
<i>Bunonema sp.</i>	U81582	Nematoda; Chromadorea; Rhabditida; Bunonematoidea; Bunonematidae
<i>Seleborca complexa</i>	U81577	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobida
<i>Cephaloboides sp.</i> SB227	AF083027	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobelloides nanus</i>	DQ102707	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobelloides bodenheimeri</i> PS1158	AF202159	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobelloides sp.</i> PS1146	AF034391	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobeles ciliates</i>	AF202148	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobeles sp.</i> PS1156	U81576	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Acrobeles complexus</i> WCUG2	U81577	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Cephalobus cubaensis</i>	AF202161	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae
<i>Cephalobus sp.</i> PS1143	AF202158	Nematoda; Chromadorea; Rhabditida; Cephaloboidea; Cephalobidae

Species	Accession No.	Taxonomy
<i>Cephalobus</i> sp. PS1196	AF202160	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Cephalobus oryzae</i> PS1165	AF034390	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Cervidellus alutus</i>	AF202152	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Pseudacrobeles variabilis</i>	AF202150	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Triligulla alluta</i>	AF331911	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Zeldia punctata</i>	U61760	Nematoda; Chromadorea; Rhabditida; Cephaloidea; Cephalobidae
<i>Myolaimus</i> sp U81585	U81585	Nematoda; Chromadorea; Rhabditida; Myolaimoidea; Myolaimidae
<i>Rhabditophanes</i> sp. KR3021	AF202151	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Alloionematidae
<i>Brevibucca</i> sp. SB261	AF202163	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Brevibuccidae
<i>Halicephalobus gingivalis</i>	AF202156	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrobelus stammeri</i>	AF202153	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Plectonchus</i> sp. PDL0025	AF202154	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Turbatrix aceti</i>	AF202165	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrellus redivivus</i>	AF036599	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrellus redivivus</i> PS1163	AF083007	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus subelongatus</i>	AY284681	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus davidi</i>	AJ567385	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus cf rigidus</i> AF40	DQ285636	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm7	AF430487	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm5	AF430485	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm6	AF430486	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm4	AF430484	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Panagrolaimus</i> sp. Sourhope farm3	AF430483	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Panagrolaimidae
<i>Steinernema carpocapsae</i>	AF036604	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Steinernematidae
<i>Strongyloides stercoralis</i>	M84229	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Strongyloididae
<i>Strongyloides ratti</i>	U81581	Nematoda; Chromadorea; Rhabditida; Panagrolaimoidea; Strongyloididae
<i>Diploscapter</i> sp PS1897	AF083009	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Diploscapteridae
<i>Diploscapter</i> sp PS2017	U81586	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Diploscapteridae
<i>Heterorhabditis bacteriophora</i>	AF036593	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Heterorhabditidae
<i>Heterorhabditis hepialus</i>	AF083004	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Heterorhabditidae
<i>Heterorhabditis zelandica</i>	AJ920368	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Heterorhabditidae
<i>Phasmarhabditis hermaphrodita</i> *	DQ639981	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Phasmarhabditis neopapillosa</i> *	DQ639982	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Parasitorhabditis</i> sp SB281	AF083028	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Rhabditoides inermiformis</i>	AF083017	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae

Species	Accession No.	Taxonomy
<i>Rhabditoides inermis</i> DF5001	AF082996	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Rhabditoides regina</i>	AF082997	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Poikilolaimus oxycerca</i> SB200	AF083023	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Poikilolaimus regenfussi</i> SB199	AF083022	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Choriorhabditis dudichi</i>	AF083012	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Distolabrellus veechi</i>	AF083011	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Distolabrellus veechi</i> DF5024	AF082999	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae
<i>Protorhabditis</i> sp.	AF083024	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Protorhabditis</i> sp. DF5055	AF083001	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Cruzinema tripartitum</i> DF5015	U73449	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Mesorhabditis</i> sp. PS1179	U73452	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Mesorhabditis spiculigera</i> SB157	AF083016	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Mesorhabditis anisomorpha</i> SB123	AF083013	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Teratorhabditis palmarum</i> DF5019	U13937	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Teratorhabditis synpapillata</i> SB131	AF083015	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Mesorhabditinae
<i>Caenorhabditis briggsae</i> PB102	U13929	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis elegans</i> N2	X03680	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. CB5161	U13930	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. PS1010	AF083006	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis japonica</i>	AY602182	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. DF5170	AY602181	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. SB341	AY602180	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis plicata</i>	AY602178	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis</i> sp. CB5161	U13930	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis vulgaris</i>	U13931	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis sonorae</i>	AF083026	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Caenorhabditis drosophilae</i>	AF083025	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Pellioditis marina</i>	AF083021	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Pellioditis mediterranea</i>	AF083020	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Peloderinae
<i>Prodontorhabditis wirthi</i>	AY602179	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Protorhabditinae
<i>Crustorhabditis scanica</i> SB125	AF083014	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Dolichorhabditis</i> sp. CEW1 (=Oscheius)	AF036591	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Oscheius</i> sp. DF5000	AF082995	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Oscheius</i> sp. BW282	AF082994	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Oscheius myriophila</i> EM435	U81588	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae

Species	Accession No.	Taxonomy
<i>Oscheius insectivora</i>	AF083019	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Oscheius dolichuroides</i>	AF082998	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis blumi</i> DF5010	U13935	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis sp</i> PS1191	AF083008	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis sp</i> PS1010	AF083006	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis myriophila</i> EM435	U13936	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditis colombiana*</i>	AY751546	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditella axei</i> DF5006	U13934	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Rhabditella sp.</i> DF5044	AF083000	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Cuticularia sp.</i> PS2083	U81583	Nematoda; Chromadorea; Rhabditida; Rhabditoidea; Rhabditidae; Rhabditinae
<i>Necator americanus</i>	AY295811	Nematoda; Chromadorea; Rhabditida; Strongylida; Ancylostomatoidea; Ancylostomatidae; Bunostominae
<i>Kalicephalus cristatus</i>	AJ920349	Nematoda; Chromadorea; Rhabditida; Strongylida; Diaphanocephaloidea; Diaphanocephalidae
<i>Ancylostoma caninum</i>	AJ920347	Nematoda; Chromadorea; Rhabditida; Strongylida; Ancylostomatoidea; Ancylostomatidae; Ancylostomatinae
<i>Angiostrongylus cantonensis</i>	AY295804	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Angiostrongylus costaricensis</i>	This study	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Angiostrongylus malaysiensis</i>	This study	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Angiostrongylus dujardini</i>	This study	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Angiostrongylus vasorum</i>	This study	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Aelurostrongylus abstrusus</i>	AJ920366	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Didelphostrongylus hayesi</i>	AY295806	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Angiostrongylidae
<i>Crenosoma mephitidis</i>	AY295805	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Crenosomatidae
<i>Crenosoma vulpis</i>	AJ920367	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Crenosomatidae
<i>Otostrongylus circumlitus</i>	AY295813	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Crenosomatidae
<i>Troglostrongylus wilsoni</i>	AY295820	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Crenosomatidae
<i>Filaroides martis</i>	AY295807	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Filaroididae
<i>Oslerus osleri</i>	AY295812	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Filaroididae
<i>Parafilaroides decorus</i>	AY295814	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Filaroididae
<i>Metastrongylus salmi</i>	AY295809	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Metastrongylidae
<i>Metastrongylus elongatus</i>	AJ920363	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Metastrongylidae
<i>Skrjabingylus chitwoodrum</i>	AY295819	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Metastrongylidae
<i>Parelaphostrongylus odocoilei</i>	AY295815	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Protostrongylidae; Elaphostrongylinae
<i>Muellerius capillaries</i>	AY295810	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Protostrongylidae; Elaphostrongylinae
<i>Protostrongylus rufescens</i>	AJ920364	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Protostrongylidae; Protostrongylinae
<i>Halocercus invaginatus</i>	AY295808	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Pseudaliidae

Species	Accession No.	Taxonomy
<i>Pseudalius inflexus</i>	AY295816	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Pseudaliidae
<i>Stenurus minor</i>	AY295817	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Pseudaliidae
<i>Syngamus trachaea</i>	AJ920344	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Syngamidae
<i>Torynurus convolutus</i>	AY295818	Nematoda; Chromadorea; Rhabditida; Strongylida; Metastrongyloidea; Unclassified Metastrongyloidea
<i>Nematodirus battus</i>	U01230	Nematoda; Chromadorea; Rhabditida; Strongylida; Molineoidea; Molineidae; Nematodirinae
<i>Petrovinema poculatum</i>	AJ920343	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Strongylidae
<i>Strongylus equines</i>	DQ094176	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Strongylidae
<i>Cylicocyclus insignis</i>	AJ920342	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Strongylidae
<i>Zoniolaimus mawsonae</i>	AJ920338	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Cloacinidae
<i>Labiostrongylus bipapillosus</i>	AJ920337	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Cloacinidae
<i>Hypodontus macropi</i>	AJ920339	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Cloacinidae
<i>Chabertia ovina</i>	AJ920341	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Chabertiidae
<i>Cyclodontostomum purvisi</i>	AJ920340	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Chabertiidae
<i>Stephanurus dentatus</i>	AJ920345	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Stephanuridae
<i>Deletrocephalus dimidiatus</i>	AJ920346	Nematoda; Chromadorea; Rhabditida; Strongylida; Strongyloidea; Deletrocephalidae
<i>Dictyocaulus eckerti</i> P7B8	AY168857	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus capreolus</i> P3B2	AY168859	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus sp.</i> P6A1	AY168860	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus filarial</i>	AY168861	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus capreolus</i> P2C10	AY168862	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus viviparus</i>	AY168856	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus filarial</i>	AJ920362	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Dictyocaulus viviparus</i>	AJ920361	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dictyocaulidae; Dictyocaulinae
<i>Haemonchus sp.</i> V3091*	DQ503465	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Haemonchinae
<i>Haemonchus contortus</i>	L04153	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Haemonchinae
<i>Haemonchus placei</i>	L04154	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Haemonchinae
<i>Haemonchus similis</i>	L04152	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Haemonchinae
<i>Ostertagia ostertagi</i>	AJ920352	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Ostertagiinae
<i>Ostertagia leptospicularis</i>	AJ920351	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Haemonchidae; Ostertagiinae
<i>Nippostrongylus brasiliensis</i>	AJ920356	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Heligmonellidae; Nippostrongylinae
<i>Heligmosomoides polygyrus</i>	AJ920355	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Heligmosomatidae
<i>Trichostrongylus colubriformis</i>	AJ920350	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Trichostrongylidae
<i>Tetrabothriostongylus mackerrasae</i>	AJ920359	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Mackerrastrongylidae
<i>Herpetostrongylus pythonis</i>	AJ920358	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Herpetostrongylidae

Species	Accession No.	Taxonomy
<i>Nicollina cameroni</i>	AJ920357	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Nicollinidae
<i>Filarinema flagrifer</i>	AJ920354	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Dromaeostrongylidae
<i>Amidostomum cygni</i>	AJ920353	Nematoda; Chromadorea; Rhabditida; Strongylida; Trichostrongyloidea; Amidostomatidae
<i>Teratocephalus lirellus</i>	AF036607	Nematoda; Chromadorea; Rhabditida; Teratocephaloidea; Teratocephalidae; Teratocephalinae
<i>Brumptaemilius justini</i>	AF036589	Nematoda; Chromadorea; Rhigonematida
<i>Anguillicola crassus</i>	DQ118535	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Anguillicolidae
<i>Dracunculus medinensis</i>	AY852268	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Dracunculidae
<i>Dracunculus oesophageus</i>	AY852269	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Dracunculidae
<i>Dracunculus sp. V3104*</i>	DQ503457	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Dracunculidae
<i>Philonema sp. A</i>	U81574	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Philometridae
<i>Philometra obturans</i>	AY852267	Nematoda; Chromadorea; Spirurida; Dracunculoidea; Philometridae
<i>Camallanus oxycephalus*</i>	DQ503463	Nematoda; Chromadorea; Spirurida; Camallanida; Camallanidae
<i>Acanthocheilonema viteae</i>	DQ094171	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Loa loa</i>	DQ094173	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Onchocerca cervicalis</i>	DQ094174	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Dirofilaria immitis</i>	AF036638	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Brugia malayi</i>	AF036588	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Wuchereria bancrofti</i>	AF227234	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Litomosoides sigmodontis</i>	AF227233	Nematoda; Chromadorea; Spirurida; Filarioidea; Onchocercidae
<i>Setaria digitata</i>	DQ094175	Nematoda; Chromadorea; Spirurida; Filarioidea; Setariidae
<i>Gnathostoma turgidum</i>	Z96948	Nematoda; Chromadorea; Spirurida; Gnathostomatoidea; Gnathostomatidae
<i>Gnathostoma neoprocyonis</i>	Z96947	Nematoda; Chromadorea; Spirurida; Gnathostomatoidea; Gnathostomatidae
<i>Gnathostoma binuncleatum</i>	Z96946	Nematoda; Chromadorea; Spirurida; Gnathostomatoidea; Gnathostomatidae
<i>Physaloptera alata</i>	AY702703	Nematoda; Chromadorea; Spirurida; Physalopteroidea; Physalopteridae
<i>Physaloptera turgid*</i>	DQ503459	Nematoda; Chromadorea; Spirurida; Physalopteroidea; Physalopteridae
<i>Ascarophis arctica</i>	DQ094172	Nematoda; Chromadorea; Spirurida; Spiruroidea; Cystidicolidae
<i>Spinitectus carolini*</i>	DQ503565	Nematoda; Chromadorea; Spirurida; Spiruroidea; Cystidicolidae
<i>Thelazia lacrymalis*</i>	DQ503458	Nematoda; Chromadorea; Spirurida; Thelazioidea; Thelaziidae
<i>Aphelenchus avenae</i>	AF036586	Nematoda; Chromadorea; Tylenchida; Aphelenchoidea; Aphelenchidae
<i>Aphelenchoides fragariae</i>	AJ966475	Nematoda; Chromadorea; Tylenchida; Aphelenchoidea; Aphelenchidae
<i>Bursaphelenchus sp.</i>	AF037369	Nematoda; Chromadorea; Tylenchida; Aphelenchoideoidea; Aphelenchoididae
<i>Deladenus sp.</i>	AJ966481	Nematoda; Chromadorea; Tylenchida; Sphaerulariidae
<i>Criconema sp.</i>	AJ966480	Nematoda; Chromadorea; Tylenchida; Tylenchina; Criconematoidea; Criconematidae; Criconematinae
<i>Hemicycliophora conida</i>	AJ966471	Nematoda; Chromadorea; Tylenchida; Tylenchina; Criconematoidea; Criconematidae; Hemicycliophorinae
<i>Pratylenchus thornei</i>	AJ966499	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Pratylenchinae
<i>Paratylenchus dianthus</i>	AJ966496	Nematoda; Chromadorea; Tylenchida; Tylenchina; Criconematoidea; Tylenchulidae; Paratylenchidae
<i>Tylenchulus semipetrans</i>	AJ966511	Nematoda; Chromadorea; Tylenchida; Tylenchina; Criconematoidea; Tylenchulidae; Tylenchulinae

Species	Accession No.	Taxonomy
<i>Ditylenchus angustus</i>	AJ966483	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Anguinidae
<i>Subanguina radicola</i>	AF202164	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Anguinidae; Anguininae
<i>Tylenchorhynchus maximus</i>	AY993979	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Belonolaimidae
<i>Geocenamus quadrifer</i>	AY993977	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Belonolaimidae
<i>Globodera pallid</i>	AFO36592	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Heteroderidae; Heteroderinae
<i>Scutellonema bradys</i>	AJ966504	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Hoplolaimidae; Hoplolaiminae
<i>Helicotylenchus dihystra</i>	AJ966486	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Hoplolaimidae; Hoplolaiminae
<i>Rotylenchus robustus</i>	AJ966503	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Hoplolaimidae; Rotylenchulinae
<i>Meloidogyne arenaria</i>	U42342	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne incognita</i>	U81578	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne javanica</i>	AF442193	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne artiellia</i>	AF442192	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne duytsi</i>	AF442197	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne exigua</i>	AF442200	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne hapla</i>	AF442194	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne ichinohei</i>	AF442191	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne maritima</i>	AF442199	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Meloidogyne microtyla</i>	AF442198	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Meloidogynidae; Meloidogyninae
<i>Nacobbus aberrans</i>	AJ966494	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Nacobbininae
<i>Pratylenchoides ritteri</i>	AJ966497	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Pratylenchinae
<i>Pratylenchoides magnicauda</i>	AF202157	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Pratylenchinae
<i>Hirschmanniella</i> sp. JH_2003	AY284616	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Pratylenchinae
<i>Radopholus similis</i>	AJ966502	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Radopholinae
<i>Pratylenchus goodyeri</i>	AJ966498	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Pratylenchidae; Radopholinae
<i>Boleodorus thylactus clone2</i>	AY993976	Nematoda; Chromadorea; Tylenchida; Tylenchina; Tylenchoidea; Tylenchidae; Boleodorinae
<i>Paractinolaimus macrolaimus</i>	AY993978	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Actinolaimoidea; Paractinolaiminae
<i>Aporcelaimellus obtusicaudatus</i>	DQ141212	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Aporcelaimidae; Aporcelaiminae
<i>Wilsonema schuurmansstekhoveni</i>	AJ966513	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Dorylaimidae
<i>Mesodorylaimus</i> cf. <i>nigritulus</i> AV-2005	AJ966490	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Dorylaimidae; Laimydorinae
<i>Mesodorylaimus japonicus</i>	AJ966489	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Dorylaimidae; Laimydorinae
<i>Mesodorylaimus bastiani</i>	AJ966488	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Dorylaimidae; Laimydorinae
<i>Pungentus</i> sp. PDL-2005	AJ966501	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Nordiidae; Pungentinae
<i>Allodorylaimus</i> sp. PDL-2005	AJ966472	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Qudsianematidae
<i>Eudorylaimus carteri</i>	AJ966484	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Qudsianematidae; Qudsianematinae

Species	Accession No.	Taxonomy
<i>Microdorylaimus</i> sp. PDL-2005	AJ966492	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Dorylaimoidea; Qudsianematidae; Qudsianematinae
<i>Longidorus elongates</i>	AF036594	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Longidoroidea; Longidoridae
<i>Xiphinema rivesi</i>	AF036610	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Longidoroidea; Longidoridae
<i>Tylencholaimus</i> sp. PDL-2005	AJ966510	Nematoda; Enoplea; Dorylaimida; Dorylaimina; Tylencholaimoidea; Tylencholaimidae; Tylencholaiminae
<i>Anoplostoma</i> sp. BHMM_2005	AY854194	Nematoda; Enoplea; Enoplida; Enoploidea; Anoplostomatidae
<i>Adoncholaimus fuscus</i>	AY854195	Nematoda; Enoplea; Enoplida; Enoploidea; Enopliidae
<i>Enoplus meridionalis</i>	Y16914	Nematoda; Enoplea; Enoplida; Enoploidea; Enopliidae
<i>Enoplus brevis</i>	U88336	Nematoda; Enoplea; Enoplida; Enoploidea; Enopliidae
<i>Enoplus communis</i>	AY854192	Nematoda; Enoplea; Enoplida; Enoploidea; Enopliidae
<i>Enoploides brunettii</i>	AY854193	Nematoda; Enoplea; Enoplida; Enoploidea; Thoracostomopsidae
<i>Syringolaimus striatocaudatus</i>	AY854200	Nematoda; Enoplea; Enoplida; Ironoidea; Ironidae
<i>Ironus dentifurcatus</i>	AJ966487	Nematoda; Enoplea; Enoplida; Ironoidea; Ironidae
<i>Calyptronema maxweberi</i>	AY854199	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Enchelidiidae
<i>Viscosia viscosa</i>	AY854198	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Oncholaimidae
<i>Viscosia</i> sp. BHMM_2005	AY854197	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Oncholaimidae
<i>Pontonema</i> sp.	AF047890	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Oncholaimidae
<i>Oncholaimus</i> sp. BHMM_2005	AY854196	Nematoda; Enoplea; Enoplida; Oncholaimoidea; Oncholaimidae
<i>Alaimus</i> sp. PDL-2005	AJ966514	Nematoda; Enoplea; Enoplida; Oxystominoidea; Alaimidae
<i>Prismatolaimus intermedius</i>	AF036603	Nematoda; Enoplea; Enoplida; Tripyloidea; Prismatolaimidae
<i>Tobrilus gracilis</i>	AJ966506	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripylidae
<i>Tripyla</i> cf. <i>filicaudata</i> JH-2004 isolate TripFil2Z	AY284731	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripylidae
<i>Bathylaimus</i> sp. PB-2005	AM234619	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Bathylaimus assimilis</i>	AJ966476	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Bathylaimus</i> sp. BHMM_2005	AY854201	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Tripyloides</i> sp. BHMM_2005	AY854202	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Trischistoma monohystera</i>	AJ966509	Nematoda; Enoplea; Enoplida; Tripyloidea; Tripyloididae
<i>Mermis nigrescens</i>	AF036641	Nematoda; Enoplea; Mermithida; Mermithoidea; Mermithidae
<i>Mylonchulus arenicolus</i>	AF036596	Nematoda; Enoplea; Mononchida; Mononchina; Mononchoidea; Mylonchulidae; Mylonchulinae
<i>Anatonchus tridentatus</i>	AJ966474	Nematoda; Enoplea; Mononchida; Mononchina; Anatonchoidea; Anatonchidae
<i>Mononchus truncates</i>	AJ966493	Nematoda; Enoplea; Mononchida; Mononchina; Mononchoidea; Mononchidae; Mononchinae
<i>Prionchulus muscorum</i>	AJ966500	Nematoda; Enoplea; Mononchida; Mononchina; Mononchoidea; Mononchidae; Prionchulinae
<i>Clarkus</i> sp. PDL-2005	AJ966479	Nematoda; Enoplea; Mononchida; Mononchina; Mononchoidea; Mononchidae; Prionchulinae
<i>Trichinella spiralis</i>	TSU60231	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella papuae</i>	AY851263	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella britovi</i>	AY851257	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella murelli</i>	AY851259	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella pseudospiralis</i>	AY851258	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella native</i>	AY851256	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella zimbabwensis</i>	AY851264	Nematoda; Enoplea; Trichocephalida; Trichinellidae
<i>Trichinella nelson</i>	AY851261	Nematoda; Enoplea; Trichocephalida; Trichinellidae

Species	Accession No.	Taxonomy
<i>Trichuris suis</i>	AY851265	Nematoda; Enoplea; Trichocephalida; Trichiuridae
<i>Trichuris trichiura</i>	DQ118536	Nematoda; Enoplea; Trichocephalida; Trichiuridae
<i>Trichuris muris</i>	AF036637	Nematoda; Enoplea; Trichocephalida; Trichiuridae
<i>Tyolaimophorus minor</i>	AJ966512	Nematoda; Enoplea; Triplonchida; Diptherophorina; Diptherophoroidea; Diptherophoridae
<i>Paratrichodorus pachydermus</i>	AF036601	Nematoda; Enoplea; Triplonchida; Diptherophorina; Trichodoroidea; Trichodoridae
<i>Paratrichodorus anemones</i>	AF036600	Nematoda; Enoplea; Triplonchida; Diptherophorina; Trichodoroidea; Trichodoridae
<i>Trichodorus primitivus</i>	AF036609	Nematoda; Enoplea; Triplonchida; Diptherophorina; Trichodoroidea; Trichodoridae

Appendix 6.2: Alignment of the 5' end of the SSU rRNA gene for the Nematoda. Positions included for the phylogeny analyses were marked by “m” within “NUCEOTIDES INCLUDED”. This alignment included new nematode sequences from Chapter 6 *and* Chapter 7.

	10	20	30	40	50	60
Gordius_aquaticus	---CTGGTTG	ATYCTGCCAG	TAGTCATATG	CTT-----	-----	-----
Priapulul_caudatus	AACCTGGTTG	ATCCTGCCAG	TAGTCATATG	CTT-----	-----	-----
Brachionus_plicatilis	-ACCTGGTTG	ATCCTGCCAG	TAGTCATATG	CTT-----	-----	-----
Chordodes_morgani	-----	-----	---AAGCTTG	CTT-----	-----	-----
Paractinolaimus_macrolaimus	-----	-----	-----	-----	-----	-----
Aporcelaimellus_obtusicaudatus	-----	-----	-----	-----	-----	-----
Wilsonema_schuermansstekhoveni	-----	-----	-----	-----	-----	-----
Mesodorylaimus_sp_cf_nigritul	-----	-----	-----	-----	-----	-----
Mesodorylaimus_bastiani	-----	-----	-----	-----	-----	-----
Mesodorylaimus_japonicus	-----	-----	-----	-----	-----	-----
Pungentus_sp._PDL_2005	-----	-----	-----	-----	-----	-----
Allodorylaimus_sp	-----	-----	-----	-----	-----	-----
Eudorylaimus_carteri	-----	-----	-----	-----	-----	-----
Microdorylaimus_sp	-----	-----	-----	-----	-----	-----
Longidorus_elongatus	-----	-----	-----	-----	-----	-----
Xiphinema_rivesi	-----	-----	-----	-----	-----	-----
Tylencholaimus_sp	-----	-----	-----	-----	-----	-----
Anoplostoma_sp._BHMM_2005	-----	-----	-----	-----	-----	-----
Adoncholaimus_fuscus	-----	-----	-----	-----	-----	-----
Enoplus_meridionalis	-----	-----	GAATTCGTCG	ACAACCTGGT	TGATCCTGCC	AGTAGTCATA
Enoplus_brevis_U88336	-----	-----	-----	--AGCCTGGT	TGATCCTGCC	AGTAGTCATA
Enoplus_communis	-----	-----	-----	-----	-----	-----
Enoploides_brunettii	-----	-----	-----	-----	-----	-----
Syringolaimus_striatocaudatus	-----	-----	-----	-----	-----	-----
Ironus_dentifurcatus	-----	-----	-----	-----	-----	-----
Calyptronema_maxweberi	-----	-----	-----	-----	-----	-----
Viscosia_sp._BHMM_2005	-----	-----	-----	-----	-----	-----
Viscosia_viscosa	-----	-----	-----	-----	-----	-----
Pontonema_vulgare	-----	-----	-----	--AACCTGGT	TGATCCTGCC	AGTAGTCATA
Oncholaimus_sp._BHMM_2005	-----	-----	-----	-----	-----	-----
Alaimus_sp._PDL_2005	-----	-----	-----	-----	-----	-----
Prismatolaimus_intermedius	-----	-----	-----	-----	-----	-----
Tobrilus_gracilis	-----	-----	-----	-----	-----	-----
Tripyla_cf_filicaudata_JH_2004	-----	-----	-----	-----	-----	-----
Bathylaimus_sp	-----	-----	-----	-----	-----	-----
Bathylaimus_assimilis	-----	-----	-----	-----	-----	-----
Bathylaimus_sp._BHMM_2005	-----	-----	-----	-----	-----	-----
Tripylodes_sp._BHMM_2005	-----	-----	-----	-----	-----	-----
Trischistoma_monohystera	-----	-----	-----	-----	-----	-----
Mermis_nigrescens	-----	-----	-----	-----	-----	-----
Mylonchulus_arenicolus	-----	-----	-----	-----	-----	-----
Anatonchus_tridentacus	-----	-----	-----	-----	-----	-----
Mononchus_truncatus	-----	-----	-----	-----	-----	-----
Prionchulus_muscorum	-----	-----	-----	-----	-----	-----
Clarkus_sp	-----	-----	-----	-----	-----	-----
Trichinella_spiralis	-----	-----	-----	-----	-----	-----AAGCT
Trichinella_papuae	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA
Trichinella_britovi	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA
Trichinella_murrelli	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA
Trichinella_pseudospiralis	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA
Trichinella_nativa	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA
Trichinella_zimbabwensis	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA
Trichinella_nelsoni	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA
Trichuris_suis	-----	-----	-----	--TACCTGGT	TGATCCTGCC	AGTAGTCATA
Trichuris_trichiura	-----	-----	-----	-----	-----	-----
Trichuris_muris	GCTTGCATGC	CTGCAGGTCG	ACTCTAGAGG	ATCCCCGGGT	ACCGAGCTCG	AATTCAAGCT
Tylolaimophorus_minor	-----	-----	-----	-----	-----	-----
Paratrichodorus_pachydermus	-----	-----	-----	-----	-----	-----
Paratrichodorus_anemones	-----	-----	-----	-----	-----	-----
Trichodorus_primitivus	-----	-----	-----	-----	-----	-----
Axonolaimus_helgolandicus	-----	-----	-----	-----	-----	-----
Ascolaimus_elongatus	-----	-----	-----	-----	-----	-----
Odontophora_rectangula	-----	-----	-----	-----	-----	-----
Cylindrolaimus_sp._202149	-----	-----	-----	-----	-----	-----
Tylocephalus_auriculatus	-----	-----	-----	-----	-----	-----
Plectus_acuminatus_BS9	-----	-----	-----	-----	-----	-----
Plectus_aquatilis	-----	-----	-----	-----	-----	-----
Anaplectus_sp	-----	-----	-----	-----	-----	-----
Anisakis_sp_WKT	-----	-----	-----	-----	-----	-----
Anisakis_sp_Nadler	-----	-----	-----	-----	-----	---GGTTATA
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	10	20	30	40	50	60
Contraecaeum_multipapillatum	-----	-----	-----	-----	-----	---	GGTTATA
Pseudoterranova_decipiens	-----	-----	-----	-----	-----	---	GGTTATA
Raphidascaris_acus	-----	-----	-----	-----	-----	---	ATA
Terranova_caballeroi	-----	-----	-----	-----	-----	---	GGTTATA
Ascaris_suum	-----	-----	-----	-----	-----	---	GGTTATA
Ascaris_lumbricoides	-----	-----	-----	-----	-----	---	GGTTATA
Baylisascaris_procyonis	-----	-----	-----	-----	-----	---	GGTTATA
Baylisascaris_transfuga	-----	-----	-----	-----	-----	---	GGTTATA
Parascaris_equorum	-----	-----	-----	-----	-----	---	GGTTATA
Porrocaecum_depressum	-----	-----	-----	-----	-----	---	GGTTATA
Toxascaris_leonina	-----	-----	-----	-----	-----	---	GGTTATA
Heterocheilus_tunicatus	-----	-----	-----	-----	-----	---	GGTTATA
Goezia_pelagia	-----	-----	-----	-----	-----	---	GGTTATA
Hysterothylacium_fortalezae	-----	-----	-----	-----	-----	---	GGTTATA
Hysterothylacium_pelagicum	-----	-----	-----	-----	-----	---	GGTTATA
Hysterothylacium_reliquens	-----	-----	-----	-----	-----	---	GGTTATA
Iheringascaris_inquies	-----	-----	-----	-----	-----	---	GGTTATA
Toxocara_canis	-----	-----	-----	-----	-----	---	GGTTATA
Nemhelix_bakeri	-----	-----	-----	-----	-----	---	CA
Raillietnema_sp._V3060	-----	-----	-----	-----	-----	---	
Cruzia_americana	-----	-----	-----	-----	-----	---	GGTTATA
Heterakis_sp_14690	-----	-----	-----	-----	-----	---	CGGTTATA
Heterakis_gallinarum	-----	-----	-----	-----	-----	---	ATA
Paraspidodera_sp_21303	-----	-----	-----	-----	-----	---	CGGTCATA
Chromodora_nudicapitata	-----	-----	-----	-----	-----	---	
Chromadora_sp._BHMM_2005	-----	-----	-----	-----	-----	---	
Atrochromadora_microlaima	-----	-----	-----	-----	-----	---	
Chromadorina_germanica	-----	-----	-----	-----	-----	---	
Chromadorita_tentabundum	-----	-----	-----	-----	-----	---	
Dichromadora_sp._BHMM_2005	-----	-----	-----	-----	-----	---	
Neochromadora_BHMM_2005	-----	-----	-----	-----	-----	---	
Spilophorella_paradoxa	-----	-----	-----	-----	-----	---	
Paracanthonchus_caecus	-----	-----	-----	-----	-----	---	CGGT TGATCCTGCC AGTAGTCATA
Paracyatholaimus_intermedius	-----	-----	-----	-----	-----	---	
Praeacanthonchus_punctatus	-----	-----	-----	-----	-----	---	
Praeacanthonchus_sp	-----	-----	-----	-----	-----	---	
Cyatholaimus_sp._BHMM_2005	-----	-----	-----	-----	-----	---	
Spirinia_parasitifera	-----	-----	-----	-----	-----	---	
Acanthopharynx_micans	-----	-----	-----	-----	-----	---	GA ATTCGGCTTG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Xyzzors_sp	-----	-----	-----	-----	-----	---	TG GAATTCGGCT TAATTCGTCG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Metachromadora_sp	-----	-----	-----	-----	-----	---	
Metachromadora_remanei	-----	-----	-----	-----	-----	---	
Catanema_sp	CCAGTGTGCT	GGAATTCGGC	TTATTGCTCG	ACAACCTGGT	TGATCCTGCC	AGTGGTCATA	
Eubostriechus_dianae	-----	-----	-----	-----	-----	---	GA ATTCG--TCG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Eubostriechus_topiarus	-----	-----	-----	-----	-----	---	GA ATTCG--TCG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Eubostriechus_parasitiferus	-----	-----	-----	-----	-----	---	GAATT CGGCTTCCGA ATTCGT-CGA CCAACCTGGT TGATCCTGCC AGTGGTCATA
Laxus_oneistus	-----	-----	-----	-----	-----	---	GAATTCG--TCG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Laxus_cosmopolitus	-----	-----	-----	-----	-----	---	TTCG--TCG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Leptonemella_sp	-----	-----	-----	-----	-----	---	TGAATTCG--TCGA CAACCTGGT TGATCCTGCC AGTGGTCATA
Robbea_hypermnestra	-----	-----	-----	-----	-----	---	GAATTCG--TCG -CAACCTGGT TGAACCTGCC AGTGGTCATA
Stilbonema_majum	-----	-----	-----	-----	-----	---	GAATTCG--TCG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Monoposthia_costata	-----	-----	-----	-----	-----	---	
Nudora_bipapillata	-----	-----	-----	-----	-----	---	
Calomicrolaimus_parahonestus	-----	-----	-----	-----	-----	---	
Calomicrolaimus_sp._BHMM_2005	-----	-----	-----	-----	-----	---	
Molgolaimus_demani	-----	-----	-----	-----	-----	---	
Diplogaster_lethieri	-----	-----	-----	-----	-----	---	AAGCT
Aduncospiculum_halicti	-----	-----	-----	-----	-----	---	
Pristionchus_lheritieri	-----	-----	-----	-----	-----	---	AAGCT
Pristionchus_pacificus	-----	-----	-----	-----	-----	---	
Pristionchus_pacificus	-----	-----	-----	-----	-----	---	
Sabatieria_punctata_STRAIN_343	-----	-----	-----	-----	-----	---	
Sabatieria_sp._355_BHMM_2005	-----	-----	-----	-----	-----	---	
Sabatieria_celtica	-----	-----	-----	-----	-----	---	
Sabatieria_punctata_STRAIN_200	-----	-----	-----	-----	-----	---	
Sabatieria_punctata_STRAIN_223	-----	-----	-----	-----	-----	---	
Sabatieria_sp._210_BHM_2005	-----	-----	-----	-----	-----	---	
Setosabatieria_hilarula	-----	-----	-----	-----	-----	---	
Desmolaimus_zeelandicus	-----	-----	-----	-----	-----	---	
Terschellingia_longicaudata	-----	-----	-----	-----	-----	---	
Cyartonema_elegans	-----	-----	-----	-----	-----	---	
Tridentulus_sp	-----	-----	-----	-----	-----	---	
Diplolaimelloides_meyli	-----	-----	-----	-----	-----	---	
Diplolaimella_diavengatensis	-----	-----	-----	-----	-----	---	
Geomonhystera_disjuncta	-----	-----	-----	-----	-----	---	
Sphaerolaimus_hirsutus	-----	-----	-----	-----	-----	---	
Theristus_acer	-----	-----	-----	-----	-----	---	
Daptonema_procerus	-----	-----	-----	-----	-----	---	GGTCATA
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	---	

	10 20 30 40 50
Daptonema_hirsutum	-----
Daptonema_normandicum	-----
Daptonema_oxycerca	-----
Daptonema_setosum	-----
Desmodora_communis	-----
Desmodora_ovigera	-----GA ATTCG--TCG ACAACCTGGT TGATCCTGCC AGTGGTCATA
Metadesmolaimus_sp	-----
Dentostomella_sp	-----
Bunonema_franzi	-----
Bunonema_sp	-----T
Seleborca_complexa	-----
Acrobelloides_nanus	-----
Acrobelloides_bodenheimeri	-----
Acrobelloides_sp_PS1146	-----
Acrobeles_ciliatus	-----
Acrobeles_sp_PS1156	-----
Acrobeles_complexus_WCUG2	-----
Cephaloboides_sp_SB227	-----CTGA TTGATTCTGT CAGCGATGTA
Cephalobus_cubaensis	-----
Cephalobus_sp._PS1143	-----
Cephalobus_sp._PS1196	-----
Cephalobus_oryzae_PS1165	-----
Cervidellus_alutus	-----
Pseudacrobeles_variabilis	-----
Triligulla_aluta	-----
Zeldia_punctada	-----
Myolaimus_sp_U81585	-----
Rhabditophanes_sp._KR3021	-----
Brevibucca_sp._SB261	-----
Halicephalobus_gingivalis	-----
Panagrobelus_stammeri	-----
Plectonchus_sp._PDL0025	-----
Turbatrix_aceti	-----
Panagrellus_redivivus	-----
Panagrellus_redivivus_PS1163	-----
Panagrolaimus_subelongatus	-----
Panagrolaimus_davidi	-----
Panagrolaimus_cf_rigidus_AF40	-----T
Panagrolaimus_sp._Sourhope_ED2	-----
Panagrolaimus_sp._Sourhope_ED2	-----
Panagrolaimus_sp._Sourhope_ED2	-----
Panagrolaimus_sp._Sourhope_ED2	-----
Panagrolaimus_sp._Sourhope_ED2	-----
Steinernema_carpocapsae	-----
Strongyloides_stercoralis	-----ACCTGGT TGATCCTGCC AGTAGTCATA
Strongyloides_ratti	-----
Diploscapter_sp_PS1897	-----
Diploscapter_sp_PS2017	-----CGGTATA
Heterorhabditis_bacteriophora	-----
Heterorhabditis_hepialus	-----CGCTATA
Heterorhabditis_zelandica	-----CGCTATA
Parasitorhabditis_sp_SB281	-----GACGATA
Rhabditoides_inermiformis	-----
Rhabditoides_inermis_DF5001	-----GATCATA
Rhabditoides_regina_DF5012	-----GGTGATA
Poikilolaimus_oxycerca_SB200	-----
Poikilolaimus_regenfussi_SB199	-----
Distolabrellus_veechi_DWF1604	-----GGTCTGTC AGTGGCGATA
Distolabrellus_veechi_DF5024	-----GATTCTGTC AGTGGCGATA
Choriorhabditis_dudichi	-----
Protorhabditis_sp	-----CGGTATA
Protorhabditis_sp_DF5055	-----
Cruzema_tripartitum_DF5015	-----
Mesorhabditis_sp_PS1179	-----
Mesorhabditis_spiculigera_SB15	-----
Mesorhabditis_anisomorpha_SB12	-----
Teratorhabditis_palmarum_DF501	-----
Teratorhabditis_synpapillata_S	-----GCC GCTACCTGAT TGATTCTGTC AGTGGCGATA
Caenorhabditis_briggsae_PB102	-----
Caenorhabditis_elegans_N2	-----ATACCTGA TTGATTCTGT CAGCGCGATA
Caenorhabditis_sp_CB5161	-----
Caenorhabditis_sp_PS1010	-----CGCGATA
Caenorhabditis_japonica	-----CGCGATA
Caenorhabditis_sp_DF5170	-----GCGCGATA
Caenorhabditis_sp_SB341	-----CGCTATA
Caenorhabditis_plicata	-----CGCGATA
Caenorhabditis_vulgaris	-----
Caenorhabditis_sonorae	-----CGCTATA
NUCLEOTIDES INCLUDED	-----

	10 20 30 40 50
Caenorhabditis_drosophilae	-----CGCGATA
Phasmarhabditis_hermaphrodita	-----
Phasmarhabditis_neopapillosa	-----
Phasmarhabditis_neopapillosa	-----CGCTGTA
Pellioditis_mediterranea_SB173	-----CGCTGTA
Pellioditis_marina	-----CGCTGTA
Prodontorhabditis_wirthi	-----T
Crustorhabditis_scanica	-----
Dolichorhabditis_sp_CEW1	-----
Oscheius_sp_DF5000	-----GA TTGATTCTGT CAGCGCTGTA
Oscheius_sp_BW282	-----CGCTGTA
Oscheius_myriophila_EM435	-----
Oscheius_insectivora	-----CGCTGTA
Oscheius_dolichuroides	-----CGCTGTA
Rhabditis_blumi_DF5010	-----
Rhabditis_sp_PS1191	-----CGCTGTA
Rhabditis_sp_PS1010	-----
Rhabditis_myriophila_EM435	-----
Rhabditis_colombiana	-----
Rhabditella_axei_DF5006	-----
Rhabditella_sp_DF5044	-----CGATGTA
Cuticularia_sp_PS2083	-----
Necator_americanus	-----
Kalicephalus_cristatus	-----GTACCTGA TTGATTCTGT CAGCGCTATA
Ancylostoma_caninum	-----CGCTATA
Angiostrongylus_cantonensis	-----
Angiostrongylus_costaricensis	-----
Angiostrongylus_malaysiensis	-----
Angiostrongylus_dujardini	-----
Angiostrongylus_vasorum	-----
Aulurostrongylus_abstrusus	-----CGATATA
Didelphostrongylus_hayesi	-----TATAT
Crenosoma_mephitidis	-----TATAT
Crenosoma_vulpis	-----CGCTATA
Otostrongylus_circumlitus	-----
Troglostrongylus_wilsoni	-----
Filaroides_martis	-----
Oslerus_osleri	-----
Parafilaroides_decorus	-----
Metastrongylus_salmi	-----
Metastrongylus_elongatus	-----CGCTATA
Skrjabinstrongylus_chitwoodrum	-----TATA
Parelaphostrongylus_odocoilei	-----
Muelerius_capillaris	-----
Protostrongylus_rufescens	-----CGCTATA
Halocercus_invaginatus	-----
Pseudalius_inflexus	-----
Stenurus_minor	-----
Torynurus_convolutus	-----
Syngamus_trachea	-----TACCTGA TTGATTCTGT CAGCGCTATA
Stephanurus_dentatus	-----CGCTATA
Nematodirus_battus	-----CGCTATA
Strongylus_equinus	-----TAAGCC
Labiostrongylus_bipapillosus	-----TACCTGA TTGATTCTGT CAGCGCTATA
Petrovinema_poculatum	-----CGCTATA
Cylicocyclus_insignis	-----CGCTATA
Chabartia_ovina	-----CGCTATA
Cyclodontostomum_purvisi	-----CGCTATA
Zoniolaimus_mawsonae	-----CGCTATA
Hypodontus_macropi	-----TACCTGA TTGATTCTGT CAGCGCTATA
Deletrocephalus_dimidiatus	-----CGCTATA
Dictyocaulus_eckerti_P7B8	-----
Dictyocaulus_capreolus_P3B2	-----
Dictyocaulus_sp_P6A1	-----
Dictyocaulus_filaria	-----
Dictyocaulus_capreolus_P2C10	-----
Dictyocaulus_viviparus	-----
Haemonchus_sp_V3091	-----
Haemonchus_contortus	-----TACCTGA TTGATTCTGT CAGCGCTATA
Haemonchus_placei	-----TACCTGA TTGATTCTGT CAGCGCTATA
Haemonchus_similis	-----TACCTGA TTGATTCTGT CAGCGCTATA
Ostertagia_ostertagi	-----CGCTATA
Ostertagia_leptospicularis	-----CGCTATA
Nippostrongylus_brasileinsis	-----CGCTATA
Heligmosomoides_polygyrus	-----CGCTATA
Trichostrongylus_colubriformis	-----CGCTATA
Tetrabothriostrongylus_mackerr	-----CGCTATA
Herpestrostrongylus_pythonis	-----CGCTATA
NUCLEOTIDES INCLUDED	-----

	10	20	30	40	50	
Nicollina cameroni	-----	-----	-----	-----	-----	---CGCTATA
Filarimena flagrifer	-----	-----	-----	-----	-----	---CGCTATA
Amidostomum cygni	-----	-----	-----	-----	-----	---CGCTATA
Teratocephalus lirellus	-----	-----	-----	-----	-----	-----
Brumptaemilius justini	-----	-----	-----	-----	-----	-----
Anguillicola crassus	-----	-----	-----	-----	-----	-----
Dracunculus medinensis	-----	-----	-----	-----	-----	-----
Dracunculus oesophageus	-----	-----	-----	-----	-----	-----
Dracunculus sp. V3104	-----	-----	-----	-----	-----	-----
Philonema sp A	-----	-----	-----	-----	-----	-----
Philometra obturans	-----	-----	-----	-----	-----	-----
Camallanus oxycephalus	-----	-----	-----	-----	-----	-----
Acanthocheilonema viteae	-----	-----	-----	-----	-----	-----
Loa loa	-----	-----	-----	-----	-----	-----
Onchocerca cervicalis	-----	-----	-----	-----	-----	-----
Dirofilaria immitis	-----	-----	-----	-----	-----	-----
Brugia malayi	-----	-----	-----	-----	-----	-----
Wuchereria bancrofti	-----	-----	-----	-----	-----	-----
Litomosoides sigmodontis	-----	-----	-----	-----	-----	-----
Setaria digitata	-----	-----	-----	-----	-----	-----
Gnathostoma turgidum	-----	-----	---	TCTCCGAT	TGATTCTGTC	GGCGATTATA
Gnathostoma neoprocyonis	-----	-----	---	TCTCCGAT	TGATTCTGTC	GGCGATTATA
Gnathostoma binucleatum	-----	-----	---	TCTCCGAT	TGATTCTGTC	GGCGATTATA
Physaloptera alata	-----	-----	-----	-----	-----	-----
Physaloptera turgida	-----	-----	-----	-----	-----	-----
Ascarophis arctica	-----	-----	-----	-----	-----	-----
Spinitectus carolini	-----	-----	-----	-----	-----	-----
Thelazia lacrymalis	-----	-----	-----	-----	-----	-----
Aphelenchus avenae	-----	-----	-----	-----	-----	-----
Aphelenchoides fragariae	-----	-----	-----	-----	-----	-----
Bursaphelenchus sp	-----	-----	-----	-----	-----	-----
Deladenus sp	-----	-----	-----	-----	-----	-----
Criconema sp	-----	-----	-----	-----	-----	-----
Hemicycliophora conida	-----	-----	-----	-----	-----	-----
Paratylenchus dianthus	-----	-----	-----	-----	-----	-----
Pratylenchus thornei	-----	-----	-----	-----	-----	-----
Tylenchulus semipentrans	-----	-----	-----	-----	-----	-----
Ditylenchus angustus	-----	-----	-----	-----	-----	-----
Subanguina radicola	-----	-----	-----	-----	-----	-----
Tylenchorhynchus maximus	-----	-----	-----	-----	-----	-----
Geocenamus quadrifer	-----	-----	-----	-----	-----	-----
Globodera pallida	-----	-----	-----	-----	-----	-----
Scutellonema bradys	-----	-----	-----	-----	-----	-----
Helicotylenchus dihystra	-----	-----	-----	-----	-----	-----
Rotylenchus robustus	-----	-----	-----	-----	-----	-----CA
Meloidogyne arenaria	-----	-----	---	ATATCTGGT	TGATCCTGCC	TGAACTGATG
Meloidogyne incognita KT	-----	-----	-----	-----	-----	-----
Meloidogyne javanica	-----	-----	-----	-----	-----	-----
Meloidogyne artiellia	-----	-----	-----	-----	-----	-----
Meloidogyne duytsi	-----	-----	-----	-----	-----	-----
Meloidogyne exigua	-----	-----	-----	-----	-----	-----
Meloidogyne hapla	-----	-----	-----	-----	-----	-----
Meloidogyne ichinohei	-----	-----	-----	-----	-----	-----G
Meloidogyne maritima	-----	-----	-----	-----	-----	-----
Meloidogyne microtyla	-----	-----	-----	-----	-----	-----
Nacobbus aberrans	-----	-----	-----	-----	-----	-----
Pratylenchoides ritteri	-----	-----	-----	-----	-----	-----
Pratylenchoides magnicauda	-----	-----	-----	-----	-----	-----
Hirschmanniella sp. JH_2003	-----	-----	-----	-----	-----	-----
Radopholus similis	-----	-----	-----	-----	-----	-----
Pratylenchus goodeyi_VF	-----	-----	-----	-----	-----	-----
Boleodorus thylactus_clone2	-----	-----	-----	-----	-----	-----T
Philippine Sequence 1	-----	-----	-----	-----	-----	-----
Philippine Sequence 2	-----	-----	-----	-----	-----	-----
Philippine Sequence 3	-----	-----	-----	-----	-----	-----
Philippine Sequence 4	-----	-----	-----	-----	-----	-----
Philippine Sequence 5	-----	-----	-----	-----	-----	-----
Philippine Sequence 6	-----	-----	-----	-----	-----	-----
Philippine Sequence 7	-----	-----	-----	-----	-----	-----
Tahiti Sequence 1	-----	-----	-----	-----	-----	-----
Tahiti Sequence 2	-----	-----	-----	-----	-----	-----
Thailand Sequence 1	-----	-----	-----	-----	-----	-----
Ogasawara Sequence 1	-----	-----	-----	-----	-----	-----
Ogasawara Sequence 2	-----	-----	-----	-----	-----	-----
Ivory Coast Sequence 1	-----	-----	-----	-----	-----	-----
Ivory Coast Sequence 2	-----	-----	-----	-----	-----	-----
Ivory Coast Sequence 3	-----	-----	-----	-----	-----	-----
Ivory Coast Sequence 4	-----	-----	-----	-----	-----	-----
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	10 20 30 40 50
Ivory Coast Sequence 5	-----
Ivory Coast Sequence 6	-----
Ivory Coast Sequence 7	-----
Singapore Sequence 1	-----
Singapore Sequence 2	-----
Singapore Sequence 3	-----
Singapore Sequence 4	-----
Nigerian Sequence 1	-----
Nigerian Sequence 2	-----
Nigerian Sequence 3	-----
NUCLEOTIDES INCLUDED	-----

	70 80 90 100 110 120
Gordius aquaticus	----GTCTC AAAGATTAAG CCATGCATGT ATAA-GT-AA T-AAAC-TA- --ACTTA-AA
Priapulus caudatus	----GTCTC AAAGATTAAG CCATGCATGT CTAA-GT-AA C-AGAC-GG- --ATACA-AA
Brachionus plicatilis	----GTCTC AAAGATTAAG CCATGCATGT CTAA-GT-AA C-ATAC-CT- --TAGCA-CG
Chordodes morgani	----GTCTC AAAGATTAAG CCATGCATGT CTAA-GT-AA T-AAAC-TT- --ACTAC-Aa
Paractinolaimus macrolaimus	TGCTTGCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Aporcelaimellus obtusicaudatus	-----GAT-AAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Wilsonema schuurmansstekhoveni	-----CTC AAAGATTAAG CCATGCATGT CTAT-GC-AA C-AAGC-CG- --ATTAA-TG
Mesodorylaimus_sp_cf_nigriritul	-----GATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Mesodorylaimus bastiani	----TCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TTATA-AG
Mesodorylaimus japonicus	-----CTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CA- --TTTAA-TG
Puangentus_sp._PDL_2005	-----TTT AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Allodorylaimus_sp	-----CTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Eudorylaimus carteri	-----TCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Microdorylaimus_sp	-----TC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Longidorus elongatus	--GCTGTCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Xiphinema rivesi	-----TAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Tylencholaimus_sp	-----CC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CT- --TAATA-CG
Anoplostoma_sp._BHMM_2005	----TGCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-CG- --ATTAA-TG
Adoncholaimus fuscus	----TGCTC AAAGATTAAG CCATGCATGT CTAG-GC-AA T-AAAT-GA- --ATCAA-TT
Enoplus meridionalis	TGCTTGCTC AAAGATTAAG CCATGCATGT CTAA-GT-AA C-ATAC-TG- --ATTAA-TA
Enoplus brevis_U88336	T-CTTGCTC AAAGATTAAG CCATGCATGT CTAA-GT-AA C-ACAC-TG- --ATTAA-TA
Enoplus communis	----TGCTC AAAGATTAAG CCATGCATGT CTAA-GT-AA C-ACAC-TG- --ATTAA-TA
Enoploides brunettii	----TGCTC AAAGATTAAG CCATGCATGT CTCA-GT-AA C-ATAC-TG- --ATTAA-TA
Syringolaimus striatocaudatus	----TGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA T-GAGC-CT- --CAAAA-TG
Ironus dentifurcatus	-----CTC AAAGATTAAG CCATGCAAGT CTAA-GC-AA C-ATGC-CG- --ATTAA-AG
Calyptronema_maxweberi	----TGCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-AAGC-TA- --TTTAA---
Viscosia_sp._BHMM_2005	----TGCTC AAAGATTAAG CCATGCATGT CTCA-GC-AA T-AAAT-AA- --ATTCCATT
Viscosia viscosa	----TGCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA T-AAAT-GA- --ATTCA-TA
Pontonema vulgare	TGCTTGCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-AAAC-TA- --TT-AA-TT
Oncholaimus_sp._BHMM_2005	----TGCTC AAAGATTAAG CCATGCATGT CTAA-GT-AA T-GAAC-TA- --TCTAG---
Alaimus_sp_PDL_2005	-----AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ACGC-CG- --ATAAA-TG
Prismatolaimus intermedius	-----AAAGATTAAG CCATGCATGT CTAA-GC-AA C-AAGC-CG- --AAGAA-TG
Tobrilus gracilis	-----CTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-AAGC-CC- --TTGAA-TG
Tripyla_cf_filicaudata_JH_2004	-----CTC AAAGATTAAG CCATGCATGT CTAA-GC- --AGAGC-CC- --TTGAA-TG
Bathylaimus_sp	----TCTC AAAGATTAAG CCATGCAAGT CTTA-GC-AA C-AAGC-CG- --TTAGTATG
Bathylaimus assimilis	----TCTC AAAGATTAAG CCATGCAAGT CTTA-GC-AA C-AAGC-CG- --TTAGTATG
Bathylaimus_sp._BHMM_2005	----TGCTC AAAGATTAAG CCATGCAAGT CTAA-GC-AA C-AAGC-CG- --TCAGTATG
Tripyloides_sp._BHMM_2005	----TGCTC AAAGATTAAG CCATGCAAGT CTAA-GC-AA C-AAGC-CG- --TTAAA-TG
Trischistoma monohystera	----TCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-AAGC-CG- --AA-CAATG
Mermis nigrescens	-GCTTGCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-C- --TTAAAACG
Mylonchulus arenicolus	-----AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ATGC-C- --TTAAAACG
Anatonchus tridentacus	-----AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ATGC-C- --TTAAAATG
Mononchus truncatus	----TCTC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ATGC-C- --TTAAAATG
Prionchulus muscorum	-----AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ATGC-C- --TTAAAATG
Clarkus_sp	----TCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ATGC-C- --TTAAAACG
Trichinella spiralis	TGCTTGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ACAC-TGT T-TTAAAA--
Trichinella papuae	TGCTTGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ACAC-TGT T-TTAAAA--
Trichinella britovi	TGCTTGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ACAC-TGT T-TTAAAA--
Trichinella murrelli	TGCTTGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ACAC-TGT T-TTAAAA--
Trichinella pseudospiralis	TGCTTGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ACAC-TGT T-TTAAAA--
Trichinella nativa	TGCTTGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ACAC-TGT T-TTAAAA--
Trichinella zimbabwensis	TGCTTGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ACAC-TGT T-TTAAAA--
Trichinella nelsoni	TGCTTGCTC AAAGATTAAG CCATGCATGT GTAA-GC-AA C-ACAC-TGT G-TTAAAA--
Trichuris suis	TGCTTGCTC AAAGATTAAG CCATGCAAGT GCAA-GC-G C-ACAC-CG- --CTGAA-CG
Trichuris trichiura	-----AAAGATTAAG CCATGCAAGT GCAA-GC-G C-ATAT-CG- --GTCTA-CG
Trichuris muris	TGCTTGCTC AAAGATTAAG CCATGCAAGT GCAA-GC-G C-ACAC-CG- --CTGAA-CG
Tyololaimophorus minor	-----C AAAGATTAAG CCATGCATGT GCCA-GC-AA C-ACGC-CC- --TTGTA-AG
Paratrichodorus pachydermus	-----C AAAGATTAAG CCATGCATGT CTAA-GC-AA C-AAGC-CC- --TTGAA-TG
Paratrichodorus anemones	-----TC AAAGATTAAG CCATGCATGT CTAA-GC-AA C-ACGC-CC- --TTGAA-TG
Trichodorus primitivus	-----AAAGATTAAG CCATGCATGT CTAT-GC-AA C-ACGC-CC- --TTGAA-TG
Axonolaimus helgolandicus	----TGCTC AAAGATTAAG CCATGCATGT CTAT-GC-AA C-AAGC-CT- --ACTA-AG
Ascolaimus elongatus	----TGCTC AAAGATTAAG CCATGCATGT TTAT-GC-AA C-AAGC-CT- --ACTA-AG
NUCLEOTIDES INCLUDED	-----

	70	80	90	100	110	120
Odontophora_rectangula	-----	-----	-----	-----	---aacg-ct-	---aata--ag
Cylindrolaimus_sp_202149	TGGCTGTCTC	AAAGATTAAG	CCATGCATGT	CTAT-GC--A	C-AAGC-CA-	---AATTA-AG
Tylocephalus_auriculatus	-----	-----	-----CATGT	CTAA-GC--A	C-AAGC-CG-	---ATTAA-TG
Plectus_acuminatus_BS9	-----	-----	-----	-----	---AAGC-CG-	---TTTAA-TG
Plectus_aquatilis	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAT-GC--A	C-AAGC-CG-	---AATA--TG
Anaplectus_sp	-----CTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-CG-	---AATA--CG
Anisakis_sp_WKT	-----	-----	-----TGT	CTAA-GTTCA	AATGGC-CT-	---ATAGA-AG
Anisakis_sp_Nadler	TGCTTATCTC	GAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAGA-AG
Contracaecum_multipapillatum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAT-GTTCA	AATGGC-CT-	---ATATA--AG
Pseudoterranova_decipiens	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAGA-AG
Raphidascaris_acus	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	---ATAAA--G
Terranova_caballeri	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAAA-AG
Ascaris_suum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAA--AG
Ascaris_lumbricoides	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---AAAA--AG
Baylisascaris_procyonis	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAA--AG
Baylisascaris_transfuga	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAA--AG
Parascaris_equorum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAA--AG
Porrocaecum_depressum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAA--AG
Toxascaris_leonina	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---ATAA--AG
Heterocheilus_tunicatus	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTCA-GTTCA	AATGGC-CT-	---TAA---AG
Goezia_pelagia	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---TTAA--AG
Hysterothylacium_fortalezae	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	---AAA---AG
Hysterothylacium_pelagicum	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	---ATAA--AG
Hysterothylacium_reliquens	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	---ATAA--AG
Iheringascaris_inquies	TGCTTATCTG	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAGC-CT-	---ATAA--AG
Toxocara_canis	TGCTTATCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATGGC-CT-	---TTAA--AG
Nemhelix_bakeri	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GTTCA	AATAAC-CT-	---ATAA--TG
Raillietnema_sp_V3060	-----	---AGGCTAAG	CCATGCATGT	CTAA-GTTCA	TATGAC-CT-	---TTAA--AG
Cruzia_americana	TGCTTGTCTC	AAAGGCTAAG	CCATGCATGT	CTAA-GTTCA	AATAAC-CT-	---ATAA--TG
Heterakis_sp_14690	TGCTTGTCTC	AAAGGCTAAG	CCATGCATGT	GTAT-GTTCA	AATAAC-CT-	---ATAA--AG
Heterakis_gallarum	TGCTTGTCTC	AAAGGCTAAG	CCATGCATGT	GTAT-GTTCA	AATAAC-CT-	---ATAA--AG
Paraspidodera_sp_21303	TGCT--GTCTC	AAAGGCTAAG	CC--TGCATGT	CTAA-GTTCA	AATAAC-CT-	---TTAA--TG
Chromadora_nudicapitata	----TGTCTC	AAAGATTAAG	CCATGCATGT	GTAA-GA--A	T-AAAC-CG-	---AATA--TG
Chromadora_sp_BHMM_2005	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	T-AAAC-CG-	---AATA--TG
Atrochromadora_microloaima	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	T-AAAC-CG-	---AATA--TG
Chromadorina_germanica	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	T-AAAC-CG-	---AATA--TG
Chromadorita_tentabundum	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---AATA--TG
Dichromadora_sp_BHMM_2005	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	T-AAAC-CG-	---AATA--TG
Neochromadora_BHMM_2005	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAAC-CG-	---AATA--TG
Spilophorella_paradoxa	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---AATA--TG
Paracanthonchus_caecus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---ATTAA-TG
Paracyatholaimus_intermedius	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-CA-	---TAGAA-TG
Praeacanthonchus_punctatus	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---ATTAA-TG
Praeacanthonchus_sp	-----YMTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---ATAAA-TG
Cyatholaimus_sp_BHMM_2005	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---ATTAA-TG
Spirinia_parasitifera	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	---AATAA-TG
Acanthopharynx_micans	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CA-	---AATAA-TG
Xyzzors_sp	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	---AATAA-TG
Metachromadora_sp	----CTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	---AATTA-TG
Metachromadora_remanei	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	---AATTA-TG
Catanema_sp	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	---AATTA-TG
Eubostrichus_dianae	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAAAGC--A	T-GAGC-CG-	---ATTTA-TG
Eubostrichus_topiarus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---ATAAA-TG
Eubostrichus_parasitiferus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---ATAAA-TG
Laxus_oneistus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	---AATTA-TG
Laxus_cosmopolitus	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	---AATTA-TG
Leptonemella_sp	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AGCC--G-	---AATTA-TG
Robbea_hypermnestra	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CG-	---AATTA-TG
Stilbonema_majum	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-AAGC-CG-	---AATTA-TG
Monoposthia_costata	----TGTCTC	AAAGATTAAG	CCATGCATGT	GCAA-GT--A	-TGAAC-CA-	---ATTGAACT
Nudora_bipapillata	----TGTCTC	AAAGATTAAG	CCATGCATGT	TTAA-GC--A	C-AAGC-CG-	---ATTAA-TG
Calomicrolaimus_parahonestus	----TGTCTC	AAAGATTAAG	CCATGCACGT	TTAA-AT--A	C-AAGC-CT-	---CAAAA-TG
Calomicrolaimus_sp_BHMM_2005	----TGTCTC	AAAGATTAAG	CCATGCACGT	TTAA-AT--A	C-AAGC-CT-	---CAAAA-TG
Molgolaimus_demani	----TGTCTC	AAAGATTAAG	CCATGCACGT	TTAT-AT--A	C-AAAC-CT-	---TCAAA-TG
Diplogaster_lethieri	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	C-ATAT-GT-	---GTAAA--C
Aduncospiculum_halicti	-----	-----	-----	-----	G-ATAG-TT-	---CTTGA--A
Pristionchus_lheritieri	TGCTTGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	C-ATAT-GT-	---GTAAA--C
Pristionchus_pacificus	-----	-----	-----TGT	CTAA-GA--A	C-ATAT-GT-	---GTAAA--C
Pristionchus_pacificus	-----A	AAAGATTAAG	CCATGCATGT	CTAA-GA--A	C-ATAT-GT-	---GTAAA--C
Sabatieria_punctata_STRAIN_343	----TGTCTC	AAAGATTAAG	CCATGCATGT	TTAA-GC--A	G-AAGC-CG-	---CACAA-TG
Sabatieria_sp_355_BHMM_2005	----TGTCTC	AAAGATTAAG	CCATGCATGT	TTAA-GC--A	G-AAGC-CG-	---CACAA-TG
Sabatieria_celtica	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	---TACAC--G
Sabatieria_punctata_STRAIN_200	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	---AACAA-CG
Sabatieria_punctata_STRAIN_223	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	---AACAA-TG
Sabatieria_sp_210_BHM_2005	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	---AACAA-CG
Setosabatieria_hilarula	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	G-AAGC-CG-	---TACTA-CG
Desmolaimus_zeelandicus	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	C-AAGC-TG-	---AATA--CG
Terschellingia_longicaudata	----TGTCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AAGC-TA-	---AATTA-AA
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	70	80	90	100	110	120
Caenorhabditis_sp_CB5161	-----	---GATTAAG	CCATGCATG-	CTTT-G---A	T-TCAT-CA-	--A-----
Caenorhabditis_sp_PS1010	TGCTCAAGTA	AA-GATTAAG	CCATGCATG-	CTT--G---A	T-TCAT-CA-	--A-----
Caenorhabditis_japonica	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	CTTT-G---A	T-TCAT-AA-	--A-----
Caenorhabditis_sp_DF5170	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	CTA--G---A	T-TCAT-CA-	--A-----
Caenorhabditis_sp_SB341	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	TTG--G---T	T--CAT-CG-	--TAAG----
Caenorhabditis_plicata	TGCTTAAGTA	AAAGATTAAG	CCATGCATG-	CTTT-G---A	T-TCAT-AA-	--A-----
Caenorhabditis_vulgaris	-----	---GATTAAG	CCATGCATG-	CTTT-G---A	T-TCAT-CA-	--A-----
Caenorhabditis_sonorae	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	TTG--G---T	T-CATC-GT-	--AA-----
Caenorhabditis_drosophilae	TGCTCAAGTA	AAAGATTAAG	CCATGCATG-	CTG--G---A	T-TCAT-CA-	--A-----
Phasmarhabditis_hermaphrodita	-----GCT	TGCTCTCAAAG	ATTAAGCCAT	GCA--TG---	--TCGAGTT	TATATATAAA
Phasmarhabditis_neopapillosa	-----GCT	TGCTCTCAAAG	ATTAAGCCAT	GCA--TG---	--TCGAGTT	CATTTT---A
Phasmarhabditis_neopapillosa	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--T	C-AGCT--T	-----T
Pellioiditis_mediterranea_SB173	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--T	C-AACT--T	-----A
Pellioiditis_marina	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--T	C-AACT--T	-----T
Prodonatorhabditis_wirthi	TGCTCAAGTA	AAAGATTAAG	CCATGCATGC	A--CAGT--C	C-AATC-GA-	--CTTTTCGA
Crustorhabditis_scanica	-----	---GATTAAG	CCATGCATGA	TGTCAGT--G	C-GACC-AC-	--ACGTATTG
Dolichorhabditis_sp_CEW1	-----	---CGCATGT	AAAAAGT--T	C-AACT--T	--AT-----	-----A
Oscheius_sp_DF5000	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--T	C-AACC--T	--AT-----G
Oscheius_sp_BW282	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	AAAAAGT--T	C-AACT--T	--TT-----
Oscheius_myriophila_EM435	-----	---GATTAAG	CCATGCATGT	AAAA--GT--	C-AACT--T	--T-----
Oscheius_insectivora	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	ATAAAGT--A	C-AGCT--T	--TA-----
Oscheius_dolichuroides	TGCTCAAGTA	AAAGATTAAG	CCATGCATGT	GAAAAGT--T	C-AACC--T	--AT-----G
Rhabditis_blumi_DF5010	-----	---GACTAAG	CCATGCATGT	CTCA--G---A	T-CCCC-AT-	--TT-----CA
Rhabditis_sp_PS1191	TGCTCACGCT	AAAGATTAAG	CCATGCATGC	TTGAGAT--	C-ACCC--T	--TT-----
Rhabditis_sp_PS1010	-----	---TG---	CTT--G---A	T-TCAT-CA-	--A-----	-----
Rhabditis_myriophila_EM435	-----	---GATTAAG	CCATGCATGT	A-AAAGT--T	C-AACT--T	--T-----
Rhabditis_colombiana	-----	---AAAGT--	A-GCTT--T	A-GCTT--T	--A-----	-----AA
Rhabditella_axei_DF5006	-----	---GATTAAG	CCATGCATGT	AT--AGT--T	C-ATTT--T	--T-----
Rhabditella_sp_DF5044	TGCTCACGTA	AAAGATTAAG	CCATGCATGT	AT--AGT--T	C-ATTT--T	--T-----
Cuticularia_sp_PS2083	-----	---CGT	GT-AAGT--T	C-ACAT-CT-	-----TTTA--	---TG
Necator_americanus	-----	---ATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A	-----AA
Kalicephalus_cristatus	TGCTCAGTTT	AAAGACTAAG	CCATGCATGT	CGA--GT--T	C-ATCTTT--	-----GA
Ancylostoma_caninum	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A	-----AA
Angiostrongylus_cantonensis	-----	---ATTAAG	CCATGCATGA	GGA--GT--T	C-AGCT--T	-----AA
Angiostrongylus_costaricensis	-----	-----A	GGA--GT--T	C-AGCT--TC	-----AA	-----AA
Angiostrongylus_malaysiensis	-----	-----A	GGA--GT--T	C-AGCT--TC	-----AA	-----AA
Angiostrongylus_dujardini	-----	-----A	GGA--GT--T	C-AGCT--TC	-----AA	-----AA
Angiostrongylus_vasorum	-----	-----A	GGA--GT--T	C-AGCT--TC	-----AA	-----AA
Aulurostrongylus_abstrusus	TGTTCAAGTA	AAAGGTTAAG	CCATGCACGA	GGA--GT--T	C-AACT--TC	-----AA
Didelphostrongylus_haysi	GCCTCAGTTAA	AAGATT--AAG	CCATGCATGT	GGA--GT--T	C-AGCT--TT	--A-----AA
Crenosoma_mephitidis	GCCTCAGTTAA	AAGATTGAAG	CCATGCATGT	GGA--GT--T	C-AGCT--TC	-----AA
Crenosoma_vulpis	TGCTCGGTTA	AAAGATTACA	AGCCATGCAT	GT--GT--A	G-TTCAGTA-	-----AA
Otostongylus_circumlitus	-----	---GATTAAG	CCATGCATGT	CGA--GT--T	C-AGCT--TC	-----AA
Troglostongylus_wilsoni	-----	---GATTAAG	CCATGCATGT	TGA--GT--T	C-AGCT--TC	-----AA
Filaroides_martis	-----	---GATTAAG	CCATGCATGT	CGA--GT--T	C-AGCT--TC	-----AA
Oslerus_osleri	-----	---GATTAAG	CCATGCATGT	CGA--GT--T	C-AACT--TC	-----AA
Parafilaroides_decorus	-----	---ATTAAG	CCATGCATGT	CGA--GT--T	C-AGCT--TC	-----AA
Metastrongylus_salmi	-----	---TTAAG	CCATGCATGT	CGA--GT--T	C-AACT--TC	-----AA
Metastrongylus_elongatus	TGCTCAGTTA	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-AACT--TC	-----AA
Skrjabinstrongylus_chitwoodrum	TGCTCAGTTA	AAAGATTAAG	CCATGCATGT	TGGA--GT--T	C-AGCT--TC	-----AA
Parelaphostrongylus_odocoilei	-----	---ATTAAG	CCATGCATGT	GGA--GT--T	C-AACT--TC	--A-----AA
Muelerius_capillaris	-----	---TGT	GGA--GT--T	C-AATT--TC	--T-----AT	-----
Protostrongylus_rufescens	TGCTCAGTTA	AAAGATTAAG	CCATGCATGT	GGA--GT--T	C-AACT--TC	-----AA
Halocercus_invasivatus	-----	---TGT	CGA--GT--T	C-AGCT--TT	--G-----AA	-----
Pseudalius_inflexus	-----	---CATGCATGT	CGA--GT--T	C-AACT--TC	-----AA	-----
Stenurus_minor	-----	---ATTAAG	CCATGCATGT	CGA--GT--T	C-AGCT--TC	-----AA
Torynurus_convolutus	-----	---ATTAAG	CCATGCATGT	CGA--GT--T	C-AGCT--TC	-----AA
Syngamus_trachea	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCTT--	-----GA
Stephanurus_dentatus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A	-----TG
Nematodirus_battus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--TT	-----GA
Strongylus_equinus	ATGCTTGTC	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--T	-----AA
Labiostongylus_bipapillosus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCTA--	-----AA
Petrovina_poculatum	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCA--C	-----AA
Cylicocyclus_insignis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--C	-----AA
Chabartia_ovina	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A	-----AA
Cyclodontostomum_purvisi	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A	-----AA
Zoniolaimus_mawsonae	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A	-----AA
Hypodontus_macropi	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCTA--	-----AA
Deletrocephalus_dimidiatus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--A	-----AA
Dictyocaulus_eckerti_P7B8	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--T	C-AACT--TG	--TA-----TA
Dictyocaulus_capreolus_P3B2	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--T	C-AACT--TG	--TG-----AA
Dictyocaulus_sp_P6A1	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--T	C-AACT--TT	--AA-----A
Dictyocaulus_filaria	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--T	C-AACT--AA	--AA-----AA
Dictyocaulus_capreolus_P2C10	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--T	C-AACT--TG	--TG-----AA
Dictyocaulus_viviparus	-----	---AAGATTAAG	CCATGCATGT	CGA--GT--T	C-AACT--TT	--TC-----AA
Haemonchus_sp_V3091	-----	---CATGCATGT	CGA--GT--T	C-ATCT--TT	--G-----AA	-----
Haemonchus_contortus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--TT	--G-----AA
Haemonchus_placei	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--TT	--G-----AA
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	70	80	90	100	110	120
Haemonchus similis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-TT	--G-----AA
Ostertagia ostertagi	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-AA	-----AA
Ostertagia leptospicularis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-AA	-----AA
Nippostrongylus brasileinsis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-AA	-----TA
Heligmosomoides polygyrus	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-AA	-----AA
Trichostrongylus colubriformis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	GGA--GT--T	C-ATCT-AA	-----AA
Tetrabothriostromylus mackerr	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT-TT	-----GA
Herpestrostrongylus pythonis	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--C	-----AA
Nicollina cameroni	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCT--C	-----AA
Filarimena flagrifer	TGCTCAGTTT	AAAGATTAAG	CCATGCATGT	CGA--GT--T	C-ATCC-CT	-----GA
Amidostomum cygni	TGCTCAGTTT	AAAGACTAAG	CCATGCATGT	CGA--GT--T	C-ATCT---	-----AA
Teratocephalus lirellus	-----TC	AAAGATTAAG	CCATGCATGT	CTAA-GC--A	T-GAGC-CA	--ATTAA-TG
Brumptaemilius justini	-----	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	C-AAAT-AA	--CCTAA-AG
Anguillicola crassus	-----C	AAAGATTAAG	CCATGCATGT	CTAT-GT--T	T-AAAT-AT	-CCTATAAAG
Dracunculus medinensis	-----	-----	-----	-----	--ATTG-CC	--TATAA-TG
Dracunculus oesophageus	-----	-----	-----	-----	--TTG-CC	--TATAA-TG
Dracunculus sp. V3104	-----	-----	-----	--A-GT--T	C-ATAC-TG	-CCTTAAATG
Philonema sp A	-----	-----	-----TGT	CCAA-GT--T	C-AAAT-TG	-CCTATAATG
Philometra obturans	-----	-----	-----	-----	--ATTG-CC	--TATAA-TG
Camallanus oxycephalus	-----	-----	-----TG	TCAAAGT--T	C-AAATGTC	--TTATAATG
Acanthocheilonema viteae	-----	-----TTAAG	CCATGCATGT	GTAA-GT--T	C-AAATAAA	-CCTATAAAG
Loa loa	-GCTTGTCCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	C-AAATAAA	-CCTATAATG
Onchocerca cervicalis	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	C-AAATAAA	-CCTATAATG
Dirofilaria immitis	-----	-----	-----GCCATGCA	TGTCTAATTT	C-AAATAAA	-CCTATAATG
Brugia malayi	-----	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	C-AAATAAA	-CCTATAATG
Wuchereria bancrofti	-----	-----AG	CCATGCATGT	CTAA-GT--T	C-AAATAAA	-CCTATAATG
Litomosoides sigmodontis	-----	-----	-----	-----GT-CT	C-AAATAAA	-CCTATAAAG
Setaria digitata	-----TC	AAAGATTAAG	CCATGCATGT	GTAA-GT--T	C-AAATAAA	-CCTAAAATG
Gnathostoma turgidum	TGCTTGTCCTC	AAAGGTTAAG	CCATGCATGT	CTGC-GT--G	C-CAAC-TC	--TCGAA-AA
Gnathostoma neoprocyonis	TGCTTGTCCTC	AAAGGTTAAG	CCATGCATGT	CTGC-GT--G	C-CAAC-TC	--TCGAA-AA
Gnathostoma binucleatum	TGCTTGTCCTC	AAAGGTTAAG	CCATGCATGT	CTGC-GT--G	C-CTAC-TC	--TTGAA-AA
Physaloptera alata	-----	-----	-----ATGCATGT	CTAA-GT--T	C-AAAT-AA	-CCTATAATG
Physaloptera turgida	-----	-----G	CCATGCATGT	CTAT-GT--T	C-AAAT-AA	-CCTATAATG
Ascarophis arctica	-----	-----TTAAG	CCATGCATGT	CTAA-GT--T	C-AAAT-AA	-CCTATAATG
Spinitectus carolini	-----	-----	-----	CTAA-GT--T	C-AAAC-AA	-CCTATAATG
Thelazia lacrymalis	-----	-----AG	CCATGCATGT	CTAA-GT--T	C-AAAT-AA	-CCTAAAATG
Aphelenchus avenae	-----	-----	-----	-----	-----	-----a
Aphelenchoides fragariae	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	GTAA-GT--A	G-AGTT-GA	--TATAA--A
Bursaphelenchus sp	-----	-----	-----	-----	--GGAG-TA	-----TTATA-CA
Deladenus sp	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AANC-GA	--TTTAA-TC
Criconema sp	-----	--AGATTAAG	CCATGCATGT	ATAA-GT--A	C-ACGC-CT	--TGAAA-AG
Hemicycliophora conida	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATTA-GT--A	C-AAGC-CT	--TAACA-AG
Paratylenchus dianthus	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	C-AAAC-CT	--TCATA-AG
Pratylenchus thornei	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GC	--TTTAA-GC
Tylenchulus semipetrans	-----CTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AATC-GT	--CTTGA-AC
Ditylenchus angustus	-----C	AAAGATTAAG	CCATGCATGT	CTAA-GT--T	T-AAAC-GA	--TTAAT--C
Subanguina radicola	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA	--TTTAA-TC
Tylenchorhynchus maximus	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA	--CTTGA-TC
Geocenamus quadrifer	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA	--TTTAA-TC
Globodera pallida	-----	AAAGATTAAG	CCATGCATGT	ATAA-GT--G	T-AACC-TG	--CCAGACAG
Scutellonema bradys	-GCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AACC-TG	--CCAGA-CA
Helicotylenchus dihystrera	-----TCTC	AAAGATTAAG	CCATGCATGT	CTAA-GT--A	T-AACC-TG	--CCAGA-CA
Rotylenchus robustus	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AACC-TG	--CCAGA-CA
Meloidogyne arenaria	TGTTTATTTT	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT	--TTA---TC
Meloidogyne incognita_KT	-----	-----	-----TGT	ATAA-GT--T	T-AATC-GT	--TTA---TC
Meloidogyne javanica	-----	-----C	CCAGGCMSSM	AWAA-GT--T	T-ATTC-GT	--TTA---TC
Meloidogyne artiellia	---TTGTTTT	AAAGATTAAG	CCATGCATGT	ACAA-GT--T	T-AATC-GC	--ATTA--GC
Meloidogyne duytsi	-GCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT	--TTTC--AC
Meloidogyne exigua	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GA	--TTTT--TC
Meloidogyne hapla	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT	--ATTT--AC
Meloidogyne ichinohei	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATCA-GT--T	T-AATC-GC	--ATCA--GC
Meloidogyne maritima	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT	--TTTT--AC
Meloidogyne microtyla	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--T	T-AATC-GT	--TTTT--AC
Nacobbus aberrans	TGCTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA	--CTTGA-TC
Pratylenchoides ritteri	-----TTT	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA	--TTTTA-TC
Pratylenchoides magnicauda	-----	-----	-----	-----	--aaac-ga	--tttta-tc
Hirschmanniella sp. JH_2003	-----CTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GA	-CTTTGA-TC
Radopholus similis	-----	AAAGATTAAG	CCATGCATGT	C-AA-GT--A	TAAATC-GA	--AA---CG
Pratylenchus goodeyi_VF	-----C	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	T-AAAC-GC	--ATA---GC
Boleodorus thylactus_clone2	GCTTTGTCCTC	AAAGATTAAG	CCATGCATGT	ATAA-GT--A	C-AAAC-GA	--CTCGA-TC
Philippine Sequence 1	-----	-----	-----	-----	-----	-----
Philippine Sequence 2	-----	AAGGATTAAG	CCATGCATGA	GGA--GT--T	C-AGCT-TT	-----AA
Philippine Sequence 3	-----	-----	-----T	ATAA-AT--T	C-AACG-T	-----AA
Philippine Sequence 4	-----	-----	-----	CTAA-GT--A	C--ATG-TG	-----TTTAAAC
Philippine Sequence 5	-----	-----	-----T	CGA--GT--T	C-ATCT--A	-----AA
Philippine Sequence 6	-----	-----	-----T	ATAAAGT--T	C-ATCA---	-----TTTT
Philippine Sequence 7	-----	-----	-----	CTTT-G--A	T-TCAT-CA	--A-----
Tahiti Sequence 1	-----	-----	-----	-----	-----	-----
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	130	140	150	160	170	180
Trichuris muris	GTG-AAG-CC	GCGAATGGCT	CATTAT-AAC	AGCCA-TTGT	-TCGCAAGAA	CTG-AT-AT-
Tylolaimophorus minor	GCG-AAG-CC	GCGAATGGCT	CATTAT-AAC	AGCCG-TTGT	-TTACTAGAT	GAT-CC-AT-
Paratrichodorus pachydermus	GTG-AAG-CC	GCGAAAAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-TA-CC-
Paratrichodorus anemones	GCG-AAG-CC	GCGAAAAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-TC-CT-
Trichodorus primitivus	GTG-AAG-CC	GCGAAAAGCT	CATTAC-AAC	ASCCR-TAGT	-TTATTGGAT	GTT-TA-CG-
Axonolaimus helgolandicus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TTGT	-TTACTGGAT	CTT-AA-TA-
Ascolaimus elongatus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TTGT	-TTACTGGAT	CTT-AA-TA-
Odontophora rectangula	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TTGT	-TTACTGGAT	CTT-AA-AA-
Cylindrolaimus_sp._202149	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTACTGGAT	CTT-AA-AA-
Tylocephalus auriculatus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTGGAT	CTT-GA-CT-
Plectus acuminatus_BS9	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTGGAT	CTT-GA-CT-
Plectus aquatilis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTGGAT	CTT-GA-TT-
Anaplectus_sp	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-CTGT	-TTACTGGAT	CTT-GA-TT-
Anisakis_sp_WKT	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	GTT-GA-TA-
Anisakis_sp_Nadler	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	GTT-GA-TA-
Contracecum multipapillatum	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-CG-
Pseudoterranova decipiens	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	GTT-GA-TA-
Raphidascaris acus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	GTT-GA-TA-
Terranova caballeroi	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	GTT-GA-TA-
Ascaris suum	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Ascaris lumbricoides	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Baylisascaris procyonis	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Baylisascaris transfuga	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Parascaris equorum	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Porrocaecum depressum	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Toxascaris leonina	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TC-
Heterocheilus tunicatus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TAAT	-ATACTGGAT	CTT-GA-TA-
Goezia pelagia	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Hysterothylacium fortalezae	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Hysterothylacium pelagicum	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Hysterothylacium reliquens	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Iheringascaris iniquus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Toxocara canis	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TG-
Nemhelix bakeri	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TTAT	-TTACTGGAT	CTT-GA-TA-
Raillietnema_sp._V3060	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TTAT	-TTACTGGAT	CTT-GA-TA-
Cruzia americana	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Heterakis_sp_14690	GTG-AAA-CC	GCGAACGGCT	CATTAT-ACC	AGCTA-TAAT	-ATACTGGAT	CTT-GACTA-
Heterakis gallinarum	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TAAT	-ATACTGGAT	GTT-GATTA-
Paraspidodera_sp_21303	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TTAT	-ATACTGGAT	CTT-GA-TA-
Chromodora nudicapitata	GTA-AAT-CC	GCGAATGGCT	CATTAT--TC	AGCCA-CAAA	-TCATTGGAT	CTA-AT-CA-
Chromadora_sp._BHMM_2005	GTA-AAT-CC	GCGAATGGCT	CATTAC-AAC	AGCTT-TAGT	-TTGTTGGAT	CTA-AT-CC-
Atrochromadora microlaima	GTA-AAT-CC	GCGAATGGCT	CATTAC-AAC	AGCCT-TAGT	-TTGTTGGAT	CTA-AA-TC-
Chromadorina germanica	GTA-AAT-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAAT	-TTATTGGAT	CTA-AC-AT-
Chromadorita tentabundum	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-AG-AG-
Dichromadora_sp._BHMM_2005	GTA-AAT-CC	GCGAATGGCT	CATTAC-AAC	AGCCT-TAGT	-TTGTTGGAT	CTA-AA-TC-
Neochromadora_BHMM_2005	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-AG-AG-
Spilophorella paradoxa	GTT-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCA-TAGT	-TTGTTGGAT	CTA-AA-AT-
Paracanthonchus caecus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Paracyatholaimus intermedius	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Praeacanthonchus punctatus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Praeacanthonchus_sp	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Cyatholaimus_sp._BHMM_2005	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	-TTATTGGAT	CTT-CT-CT-
Spirinia parasitifera	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GA-TT-
Acanthopharynx micans	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GT-TT-
Xyzzors_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCT-TTGT	-TTCTTGGAT	CTT-GA-AC-
Metachromadora_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GA-TT-
Metachromadora_remanei	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-GA-TT-
Catanema_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Eubostrichus dianae	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AG-TT-
Eubostrichus topiarus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Eubostrichus parasitiferus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Laxus oneistus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Laxus cosmopolitus	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Leptonemella_sp	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCTG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Robbea hypermnestra	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Stilbonema majum	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTT-AC-TT-
Monoposthia_costata	GTG-AAA-CT	STSAATGGCT	CATTAA-ATC	AGTTA-TAGT	-TTGTTGGAT	GGT-AC-GT-
Nudora bipapillata	GTG-AAG-CC	GCGGACAGCT	CATTAA-TAC	ACTCC-TAAT	-CCTAGCAAT	GCT-TA-CC-
Calomicrolaimus parahonestus	GTG-AAA-TC	GCGAATAGCT	CATTAC-AAC	AGCCA-TTGT	-TTCTTGGAT	CTT-AC-TT-
Calomicrolaimus_sp._BHMM_2005	GTG-AAA-TC	GCGAATAGCT	CATTAC-AAC	AGCCA-TTGT	-TTCTTGGAT	CTT-AC-TT-
Molgolaimus demani	GTG-AAA-TC	GCGAATAGCT	CATTAC-AAC	AGCCA-TTGT	-TTCTTGGAT	CTT-AC-TT-
Diplogaster lethieri	ATG-AAT-CT	GCGAACGGCT	CATTATTAAC	ACCCG-TAAT	-CTACCCAGT	TTT-CG-TA-
Aduncospiculum halicti	CTG-AAT-CT	GCGAACGGCT	CATTAA-AAC	GGCTA-TAAT	-TTACTCGAA	TTA-TC-A--
Pristionchus lherieri	ATG-AAT-CT	GCGAACGGCT	CATTATTAAC	ACCCG-TAAT	-CTACCCAGT	TTT-CG-TA-
Pristionchus pacificus	ATG-AAT-CT	GCGAACGGCT	CATTATTAAC	ACCCA-TAAT	-CTACCCAGT	TTT-CG-TA-
Pristionchus pacificus	ATG-AAT-CT	GCGAACGGCT	CATTATTAAC	ACCCA-TAAT	-CTACCCAGT	TTT-CG-TA-
Sabatieria punctata_STRAIN_343	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-TA-AT-
Sabatieria_sp._355_BHMM_2005	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTTGGAT	CTC-TA-AT-
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmmmm	mmmmmmmm-mm	mmmmmm-mmmm	mmmmmmmmmm	mmmm-----

	130	140	150	160	170	180
Sabatieria_celtica	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTGGAT	CTC-TC-GA-
Sabatieria_punctata_STRAIN_200	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTGGAT	CTC-TT-AT-
Sabatieria_punctata_STRAIN_223	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTGGAT	CTC-TC-AT-
Sabatieria_sp._210_BHM_2005	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTGGAT	CTC-TT-AT-
Setosabatieria_hilarula	GTA-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTGGAT	CTC-TC-GA-
Desmolaimus_zeelandicus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TAGT	CTACTTGGT	CTT-GA-AA-
Terschellingia_longicaudata	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	TCCCTTGAGA	CTA-TG-AA-
Cyartonema_elegans	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	TCCCTAGAGA	CTA-CG-A--
Tridentulus_sp	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-TT-CT-
Diplolaimelloides_meyli	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-TT-CT-
Diplolaimella_diavengatensis	ATG-AAG-CC	GCGAATAGCT	CATTAT-AAC	AGCCG-TTGT	-TTCTGGAT	CTT-CT-CT-
Gecmonhystrera_disjuncta	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAT	AGCCG-TTGT	-TTCTGGAT	CCT-GA-AT-
Sphaerolaimus_hirsutus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-AT-CC-
Theristus_acer	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TCGT	-TTCTTAGAT	CTC-CG-AT-
Daptonema_procerus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-CG-TT-
Daptonema_hirsutum	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-CG-CT-
Daptonema_normandicum	GTA-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-CG-CA-
Daptonema_oxycerca	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-CG-TT-
Daptonema_setosum	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-CG-CT-
Desmodora_communis	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTT-GT-TT-
Desmodora_ovigera	GTG-AAG-CC	GCGAATGGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTT-GT-TT-
Metadesmolaimus_sp	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCG-TTGT	-TTCTGGAT	CTC-CG-CT-
Dentostomella_sp	GTG-AAA-CC	GCGAAGGCT	CATTAT-AAC	AGCTA-TTAT	-TTACTTGT	ATT-GA-TA-
Bunonema_franzi	GCG-AAT-TC	GCGGACGGCT	CATTAC-AAC	AGAGA-TAAT	-TTGCTTGT	TTA-GA-TT-
Bunonema_sp	GCA-AAT-CC	GCGGACGGCT	CATTAC-AAC	AGAGC-TAAT	-TTTCTTGT	TTT-GA-CG-
Seleborca_complexa	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTT-TGA-	-TTCTTGA	CGT-AG-AC-
Acrobeloides_nanus	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	GTT-GA-CA-
Acrobeloides_bodenheimeri	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CA-
Acrobeloides_sp_PS1146	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CA-
Acrobeles_ciliatus	GTG-AAA-CC	GCGAATGGCT	CATTAA-TAC	AGTAA-AAAC	-CTACAAGAT	GTT-GA-CA-
Acrobeles_sp_PS1156	GTG-AAA-CC	GCGAATGGCT	CATTAA-TAC	AGTAA-AAAC	-CTACAAGAT	GTT-GA-CA-
Acrobeles_complexus_WCUG2	GTG-AAA-CC	GCGAATGGCT	CATTAA-TAC	AGTAA-AAAC	-CTACAAGAT	GTT-GA-CA-
Cephaloboides_sp_SB227	TTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTT-TTGCT	G-----
Cephalobus_cubaensis	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CT-
Cephalobus_sp_PS1143	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CA-
Cephalobus_sp_PS1196	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CA-
Cephalobus_oryzae_PS1165	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CA-
Cervidellus_alutus	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CA-
Pseudacrobelus_variabilis	TTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	GTT-GA-CT-
Triligulla_aluta	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CA-
Zeldia_punctada	GTG-AAA-CC	GCGAATGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	CTT-GA-CA-
Myolaimus_sp_U81585	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGT	TTT-GA-TA-
Rhabditophanes_sp_KR3021	GTG-AAA-CC	GCGGAAAGCT	CATTAT-AAC	AGCTA-TAAT	-TTACACGGC	AAA-TT-CC-
Brevibucca_sp_SB261	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TTAT	-TTACTTGAC	GAG-AC-AA-
Halicephalobus_gingivalis	ACG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-AAAT	-GTATACGGT	GTT-GA-CA-
Panagrobelus_stammeri	ACG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTT-TAAC	-CTACTTGT	TTA-GA-CA-
Plectonchus_sp_PDL0025	ACG-AAA-CC	GCGAACGGCT	CATTAC-AAC	GGCTT-TACT	-TTACTTGT	TTT-GA-CT-
Turbatrix_aceti	GCG-AAA-CC	GCGCATAGCT	CATTAT-AAC	AGTCC-TAAT	-TTACAAGTA	TTT-GA-TT-
Panagrellus_redivivus	ACG-AAA-CC	GCGTATGGCT	CATTAC-AAC	AGCTA-TGAT	-TCTATTGAT	CAT-GA-CA-
Panagrellus_redivivus_PS1163	ACG-AAA-CC	GCGTATGGCT	CATTAC-AAC	AGCTA-TGAT	-TCTATTGAT	CAT-GA-CA-
Panagrolaimus_subelongatus	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGT	TTT-GA-CT-
Panagrolaimus_davidi	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGT	TTT-GA-CT-
Panagrolaimus_cf_rigidus_AF40	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGT	TTT-GA-CT-
Panagrolaimus_sp_Sourhope_ED2	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGT	TTT-GA-CT-
Panagrolaimus_sp_Sourhope_ED2	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGT	TTT-GA-CT-
Panagrolaimus_sp_Sourhope_ED2	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGT	TTT-GA-CT-
Panagrolaimus_sp_Sourhope_ED2	ACG-AAA-CC	GCGTATGGCT	CATTAT-AAC	AGCTA-AAAT	-TTACTTGT	TTT-GA-CT-
Steinernema_carpocapsae	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCGA-TAAT	-TGACTAAAC	GAT-GT-TA-
Strongyloides_stercoralis	GTG-AAA-CT	GCGAATGGCT	CATTAA-ATC	AGTTA-TAGT	-TTATTTGAT	GGT-TT-CT-
Strongyloides_ratti	ATG-AAA-CC	GCGGAAAGCT	CATTAT-AAC	AGCTA-TAGA	-CTACACGGT	AAA-TA-TT-
Diploscapter_sp_PS1897	GTG-AAG-CT	GCGCACGGCT	CATTAG-CAC	GGTTT-CGAC	CTTTTTCGGA	TAT-CT----
Diploscapter_sp_PS2017	GTG-AAG-CT	GCGCACGGCT	CATTAG-CAC	GGTTT-CGAC	CTTTTTCGGA	TAT-CT----
Heterorhabditis_bacteriophora	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAT	-TTATTCGGT	AAT-A-----
Heterorhabditis_hepialus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAT	-TTATTCGGT	AAT-A-----
Heterorhabditis_zelandica	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAT	-TTATTCGGT	AAT-A-----
Parasitorhabditis_sp_SB281	GCA-GTA-CT	GTGAACGGCT	CATTAT-AAC	AGTTA-AAAT	-TTGCAAGAA	GCT-TT-TC-
Rhabditoides_inermiformis	GTG-AAA-CT	GCGAACGGCT	AATTAT-ACC	AGCTA-TAAT	-TTACTAGAT	TTT-GA-CA-
Rhabditoides_inermis_DF5001	GTG-AAA-CT	GCGGGCGGCT	CATTAT-ATC	AGTTA-TGAT	-TTACTTGT	TTT-GA-TT-
Rhabditoides_regina_DF5012	GTT-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGCCA-AAAT	-CAGCATGAG	GTT-GA-C--
Poikilolaimus_oxycerca_SB200	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TAAT	-TTACTTGT	GTT-GA-CT-
Poikilolaimus_regenfussi_SB199	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TAAT	-TTACTTGT	GTT-GA-CA-
Distolabrellus_veechi_DWF1604	GTG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGTTA-AAAT	-CAGCGTGAA	GCT-TA-C--
Distolabrellus_veechi_DF5024	GTG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGTTA-AAAT	-CAGCGTGAA	GCT-TA-C--
Choriorhabditis_dudichi	GAG-AAA-CT	GCGTACGGCT	CATTAG-AGC	AGATA-TGCA	-CTTGTCGAG	GTT-Y-----
Protorhabditis_sp	GTG-AAA-CT	GCGCACGGCT	CATTAG-AAC	GGTTT-ATA-	-CCTATTTCCG	GAT-GT-----
Protorhabditis_sp_DF5055	GCG-AAA-CT	GCGAATGGCT	CATTAG-AGC	AGTTA-ATAG	-CCTTTCCGT	TGA-TC-----
Cruzanema_tripartitum_DF5015	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TATT	-TTATTTCCG	TGA-TC-C--
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmmmm	mmmmmm-mm	mmmmmm-mm	mmmmmmmmmm	mmmm-----

	130	140	150	160	170	180
Mesorhabditis_sp_PS1179	GGT-AGA-CT	GTGAACGGCT	CATTAT-AAC	AGTTA-AAAT	-CCTCGCGAA	GTT-AG-C--
Mesorhabditis_spiculigera_SB15	GTG-GTA-CT	GTGAACGGCT	CATTAC-AAC	AGTTG-AAAT	-TCGCGTGAA	ATC-GT-TC-
Mesorhabditis_anisomorpha_SB12	GTT-AGA-CT	GTGAACGGCT	CATTAC-AAC	GGTCA-AAAT	-CGGCACGAA	GTA-GT-C--
Teratorhabditis_palmarum_DF501	GCC-CAT-CT	GCGAACGGCT	CATTAC-AAC	AGTTA-AAAT	-TCGCATGAC	GAT-TG-T--
Teratorhabditis_synpapillata_S	ACC-TAT-CT	GCGAACGGCT	CATTAC-AAC	AGTTA-AAAT	-CCGCATGAC	GAT-TG-T--
Caenorhabditis_briggsae_PB102	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTGTTCCG	AAT-CC-T--
Caenorhabditis_elegans_N2	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTATCCGG	GAT-CC-G--
Caenorhabditis_sp_CB5161	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTATCCGG	AAT-CC----
Caenorhabditis_sp_PS1010	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAT	-CTCGTTCGG	AAT-TC----
Caenorhabditis_japonica	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTCTCCGG	AAT-CC----
Caenorhabditis_sp_DF5170	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAT	-TTTACTCCG	AAT-TC----
Caenorhabditis_sp_SB341	-AG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAC	-ATCCTTCGG	CAT-TC----
Caenorhabditis_plicata	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATG-TATT	-CTCATTCCG	CGG-TC----
Caenorhabditis_vulgaris	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAC	-CTTGTTCGG	AAT-CC----
Caenorhabditis_sonorae	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAC	-TTCCCTCCG	CGA-TC----
Caenorhabditis_drosophilae	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAT	-TTTACTCCG	AAT-TC----
Phasmarhabditis_hermaphrodita	TAG-AAA-CT	GCGTACGGCT	CATTAG-AGC	AGATTTAAAT	-GATTCGGG	TAT-C----
Phasmarhabditis_neopapillosa	AAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATTTAAAT	-GATTCAGG	TAT-C----
Phasmarhabditis_neopapillosa	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTCCG	CTA-TC----
Pellioditis_mediterranea_SB173	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTCCG	GTA-TC----
Pellioditis_marina	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTCCG	CTA-TC----
Prodontorhabditis_wirthi	ATG-AAA-CT	GCGGACGGCT	CATTAT-AGC	ATTGA-AACG	-TTATTCTCG	CAC-CA-T--
Crustorhabditis_scanica	GTT-CGA-CT	GTGAACGGCT	CATTAT-AAC	AGTTA-AAAT	-CAGCATGAA	GTA-TA-T--
Dolichorhabditis_sp_CEW1	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TAAT	-ATACCCGAG	ATA-TC-T--
Oscheius_sp_DF5000	GTG-AAA-CT	GCGAACGGCT	CATTAT-AGC	AGATA-TAAT	-GTACCCGAG	AAT-AT-C--
Oscheius_sp_BW282	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTCCG	ATA-TC-C--
Oscheius_myriophila_EM435	CTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTCCG	ATA-TC-C--
Oscheius_insectivora	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTCCG	ATA-TC-C--
Oscheius_dolichuroides	GTG-AAA-CT	GCGAACGGCT	CATTAT-AGC	AGATA-TAAT	-GTACCCGAG	ATA-TT-C--
Rhabditis_blumi_DF5010	CGG-AAT-CG	GCGAACGGCT	CAATAT-TAC	AGCTA-TCAC	-TTATT-CGG	TGA-T----
Rhabditis_sp_PS1191	GGG-AAATCA	GCGGCTAGCT	CATTAC-AAC	AGATC-TTAC	-TTATTTGGA	AGA-GA-T--
Rhabditis_sp_PS1010	-TG-AAA-TT	GCGTACGGCT	CATTAG-AGC	AGATA-TCAT	-CTCGTTCGG	AAT-TC-T--
Rhabditis_myriophila_EM435	CTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTCCG	ATA-TC-C--
Rhabditis_colombiana	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGCTA-TCAT	-TCTCTCCG	ATA-TC-C--
Rhabditella_axei_DF5006	CTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATT-TTAC	-CTATTCCGA	AAT-CT----
Rhabditella_sp_DF5044	CTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATT-TTAC	-CTATTCCGA	AAT-CT----
Cuticularia_sp_PS2083	GTG-AAA-CC	GCGAATGGCT	CATTAC-AAC	AGCTA-TAAT	-TTACTGGAT	GTT-GA-CT-
Necator_americanus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Kalicephalus_cristatus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	ATG-TC----
Ancylostoma_caninum	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Angiostrongylus_cantonensis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTCCGA	AAA-TC----
Angiostrongylus_costaricensis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTCCGA	AAA-TC----
Angiostrongylus_malaysiensis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTCCGA	AAA-TC----
Angiostrongylus_dujardini	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Angiostrongylus_vasorum	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Aulurostrongylus_abstrusus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAG-TT----
Didelphostrongylus_hayesi	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TATT	-TTAATCCGT	AAT-TT----
Crenosoma_mephitidis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Crenosoma_vulpis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTGGA	AAA-TC----
Otostrongylus_circumlitus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Troglostrongylus_wilsoni	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Filaroides_martis	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Oslerus_osleri	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAG-AC----
Parafilaroides_decorus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Metastrongylus_salmi	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Metastrongylus_elongatus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Skrjabinogylus_chitwoodrum	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATA-TCAT	-TTATTCCGA	AAT-TC----
Parelaphostrongylus_odoccoilei	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC-C--
Muelerius_capillaris	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TG----
Protostrongylus_rufescens	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TT----
Halocercus_invaginatus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Pseudalius_inflexus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC-T--
Stenurus_minor	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Torynurus_convolutus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC----
Syngamus_trachea	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	ACG-TG----
Stephanurus_dentatus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAG-TC----
Nematodirus_battus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTCCGA	AAA-TC-C--
Strongylus_equinus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Labiostongylus_bipapillosus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAA-TC----
Petrovinema_poculatum	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Cylicocyclus_insignis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Chabartia_ovina	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Cyclodontostomum_purvisi	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Zoniolaimus_mawsonae	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Hypodontus_macropi	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAG-TC----
Deletocephalus_dimidiatus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAC	-TTATTCCGA	AAA-TC----
Dictyocaulus_eckerti_P7B8	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTCCGA	ATT-AT-T--
Dictyocaulus_capreolus_P3B2	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTCCGA	ATT-AT-T--
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmmmm	mmmmmm-mmm	mmmmmm-mmmmm	mmmmmmmmmm	mmmm------

	130	140	150	160	170	180
Dictyocaulus_sp_P6A1	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTTCGGA	ATT-AT-T--
Dictyocaulus_filaria	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTCATTCGGA	AAA-TC-C--
Dictyocaulus_capreolus_P2C10	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTTCGGA	ATT-AT-t--
Dictyocaulus_viviparus	GTG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TAAT	-TTATTTCGGA	ATT-AT-T--
Haemonchus_sp_V3091	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Haemonchus_contortus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Haemonchus_placei	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Haemonchus_similis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Ostertagia_ostertagi	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----
Ostertagia_leptospicularis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACA-TC----
Nippostrongylus_brasileinsis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Heligmosomoides_polygyrus	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Trichostrongylus_colubriformis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Tetrabothriostromylus_mackerr	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Herpestrostrongylus_pythionis	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Nicollina_cameroni	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAG-TC----
Filarimena_flagrifer	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	ACG-TC----
Amidostomum_cygni	GAG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TCAT	-TTATTTCGGA	AAA-TC----
Teratocephalus_lirellus	GTG-AAG-CC	GCGAATAGCT	CATTAC-AAC	AGCCA-TTGT	-TTACTTGAT	CTT-GA-TA-
Brumptaemilius_justini	GTC-AAA-CC	GCGTACGGCT	CATTAT-AAC	AGCCA-TAAT	-ATACTTGAC	GTT-GA-CT-
Anguillicola_crassus	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGGT	CTT-GA-TA-
Dracunculus_medinensis	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	CTT-GG-AT-
Dracunculus_oesophageus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	CTT-GA-TT-
Dracunculus_sp_V3104	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	CTT-GA-TT-
Philonema_sp_A	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-ATACTTGAT	TCT-GA-TT-
Philometra_obturanus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TTAT	-TTACTTGAT	TTT-GA-TT-
Camallanus_oxycepahalus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCCA-TAAT	-ATACTTGAT	GTT-GA-TT-
Acanthocheilonema_viteae	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-
Loa_loa	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-
Onchocerca_cervicalis	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-
Dirofilaria_immitis	GTG-AAA-CC	GCGAACGGCT	CAT--T-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-
Brugia_malay	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-
Wuchereria_bancrofti	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-
Litomosoides_sigmodontis	GTG-AAA-CC	GCGAACGGCT	CATNAT-NAC	AGCCTCTAAT	-GTACTTGAT	GCT-GA-CT-
Setaria_digitata	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-
Gnathostoma_turgidum	GTG-AAA-CC	GCGAATGGCT	CATTAC-GAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-TA-
Gnathostoma_neoprocyonis	GTG-AAA-CC	GCGAATGGCT	CATTAC-GAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-TA-
Gnathostoma_binucleatum	GTG-AAA-CC	GCGAATGGCT	CATTAC-GAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-TA-
Physaloptera_alata	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	CTT-GA-TC-
Physaloptera_turgida	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	CTT-GA-TT-
Ascarophis_arctica	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	CTT-GA-TA-
Spinitectus_carolini	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	CTT-GA-TA-
Thelazia_lacrymalis	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-GTACTTGAT	GTT-GA-TT-
Aphelenchus_avenae	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGGTA-TAAT	-TTACTTGAT	CTT-GA-AA-
Aphelenchoides_fragariae	GCG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGATA-TAAT	-TTACTTGT	CTA-TT-CC-
Bursaphelenchus_sp	GCG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGATA-TAAT	-TTACTAGTT	CTT-GA-TC-
Deladenus_sp	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	TTT-CA-CT-
Criconema_sp	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TAAT	-GTACTTGAT	CTT-GA-AC-
Hemicycliophora_conida	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCCA-TAAT	-TTACTCGAT	CTT-GA-AC-
Paratylenchus_dianthus	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTAGAC	CTT-GA-GC-
Pratylenchus_thornei	GTG-AAA-CT	GCGTACGGCT	CATTAC-AAC	AGCAA-TAAT	-TTACTTGAT	CTT-GA-CA-
Tylenchulus_semipentrans	GAG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-AC-
Ditylenchus_angustus	GTG-AAA-CC	GCGAACGGCT	CATTAC-AAC	AGCTA-TGAT	-TTACTTGAT	CTT-GA-CC-
Subanguina_radicola	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-
Tylenchorhynchus_maximus	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-
Geocenamus_quadriker	GTG-AAA-CC	GCGAACAGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-
Globodera_pallida	GGG-AAA-CT	GCGTACGGCT	CATTAC-ACC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-
Scutellonema_bradys	GGG-AAA-CT	GCGTACGGCT	CATTAC-ACC	AGCTG-TAAT	-TTACTTGAT	CTT-GA-CA-
Helicotylenchus_dihystera	GGG-AAA-CT	GCGTACGGCT	CATTAC-ACC	AGCCT-TAAT	-TTACTTGAT	CTT-GA-CA-
Rotylenchus_robustus	GGG-AAA-CT	GCGTACGGCT	CATTAC-ACC	AGCCT-TAAT	-TTACTTGAT	CTT-GA-CA-
Meloidogyne_arenaria	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TTAT	-TTACTTGAT	CTT-GA-TT-
Meloidogyne_incognita_KT	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TTAT	-TTACTTGAT	CTT-GA-TT-
Meloidogyne_javanica	GAG-AAA-CC	GCGAACGGCT	MAWAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TT-
Meloidogyne_artiellia	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TTAT	-TTACTTGAT	CTT-GA-CA-
Meloidogyne_duytsi	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TA-
Meloidogyne_exigua	GAG-AAA-CC	GCGTACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TT-
Meloidogyne_hapla	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TA-
Meloidogyne_ichinohei	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TTAT	-TTACTTGAT	CTT-GA-TCA
Meloidogyne_maritima	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TTA
Meloidogyne_microtyla	GAG-AAA-CC	GCGAACGGCT	CATTAC-AAT	GGCCA-TGAT	-TTACTTGAT	CTT-GA-TA-
Nacobbus_aberrans	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-
Pratylenchoides_ritteri	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-
Pratylenchoides_magnicauda	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CT-
Hirschmanniella_sp_JH_2003	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-
Radopholus_similis	ATG-AAA-CC	GCAAACGGCT	CATTAC-ACC	AGCAG-TGAT	-CTATTGAC	CTT-GA-TA-
Pratylenchus_goodeyi_VF	GTG-AAA-CT	GCGAACGGCT	CATTAC-AAC	AGCTG-TAAT	-TTACTTGAT	CTT-GA-AA-
Boleodorus_thylactus_clone2	GTG-AAA-CC	GCGAACGGCT	CATTAT-AAC	AGCTA-TAAT	-TTACTTGAT	CTT-GA-CA-
Philippine Sequence 1	-TG-AAA-CT	GCGAACGGCT	CATTAG-AGC	AGATG-TGAT	-TTATTTCGGA	AAA-TC----
NUCLEOTIDES INCLUDED	-mm-mmm-mm	mmmmmmmmmm	mmmmmmmm-mm	mmmmmm-mmmm	mmmmmmmmmm	mmmm-----

	130 140 150 160 170 180
Philippine Sequence 2	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TGAT -TTATTCCGGA AAA-TC----
Philippine Sequence 3	GTG-AAA-TT GCGAACGGCT CATTAG-AGC AGCTA-TTAT TCTCTTCGGT ATCAT-----
Philippine Sequence 4	ACG-AAA-CC GCGTATGGCT CATTAT-AAC AGCTA-TAAT -TTACTTGAT TTT-GA-CA-
Philippine Sequence 5	GAG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAC -TTATTCCGGA AAG-TC----
Philippine Sequence 6	GAG-AAA-CT GCGAACGGCT CATTAG-AGC AGATA-TTAT -TTCTTCGGA ATA-T-----
Philippine Sequence 7	-TG-AAA-TT GCGTACGGCT CATTAG-AGC AGATA-TCAC -CTTGTTCCGG AAT-AT----
Tahiti Sequence 1	-TG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TGAT -TTATTCCGGA AAA-TC----
Tahiti Sequence 2	GTG-AAA-CT GCGAACGGCT TATTAG-AGC AGATG-TGAT -TTATTCCGGA AAA-TC----
Thailand Sequence 1	GTG-AAA-CC GCGAACGGCT CATTAC-AAC AGCTA-TAAT -ATACTTGAT CTT-GACTA-
Ogasawara Sequence 1	ACG-AAA-CC GCGTATGGCT CATTAT-AAC AGCTA-TAAT -TTACTTGAT TTT-GA-CA-
Ogasawara Sequence 2	ATG-AAA-CT GCGAACGGCT CATTAG-AGC AGATATTAAT GATTGCAGAG -TA-TC----
Ivory Coast Sequence 1	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAT -TTATTCCGGA AAA-TC----
Ivory Coast Sequence 2	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAT -TTATTCCGGA AAG-AC----
Ivory Coast Sequence 3	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAT -TTATTCCGGA AAA-AT----
Ivory Coast Sequence 4	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAT -TTACTCGGT AAAGAC----
Ivory Coast Sequence 5	GTG-AAA-CC GCGAACGGCT CATTAC-AAC AGCTA-TAAT -ATACTTGAT CTT-GACTA-
Ivory Coast Sequence 6	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAT -TTACTCGGT AAA-GAC----
Ivory Coast Sequence 7	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAT -TTATTCCGGA AA--GAC----
Singapore Sequence 1	GTG-AAA-TT GCGAACGGCT CATTAG-AGC AGCTA-TTAT -TCTCTTCGG -TA-TC----
Singapore Sequence 2	GTG-AAR-CC GCGAACGGCT CATTAC-AAC AGCTA-TAAT -ATACTTGAT CTT-GACTA-
Singapore Sequence 3	-TG-AAA-TT GCGTACGGCT CATTAG-AGC AGATA-TCAC -CTTGTTCCGG AAT-CC-T--
Singapore Sequence 4	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAT -TTATTCCGGA AAG-AC----
Nigerian Sequence 1	-TG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TTAT -TTATTCCGGA AAA-TA----
Nigerian Sequence 2	GAG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCWC -TTATTCCGGA AAG-TC----
Nigerian Sequence 3	GTG-AAA-CT GCGAACGGCT CATTAG-AGC AGATG-TCAT -TTATTCCGGA AAG-ATGTA-
NUCLEOTIDES INCLUDED	-mm-mmm-mm mmmmmmmmmmm mmmmmmm-mmmmm mmmmm-mmmmm -mmmmmmmmmm mmm-----

	190 200 210 220 230 240
Gordius aquaticus	-----CC-- -----CACA TGGATAAC -- TGTGGTAATT CTAGAGCTAA TACATGCT-A
Priapulius caudatus	-----TCC- -----TACT TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCA-A
Brachionus plicatilis	-----AC-- -----TACA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCA--
Chordodes morgani	-----AC-- -----CACA TGGATAAC -- TGTGGTAATT CTAGAGCTAA TACATGCT-G
Paractinolaimus macrolaimus	-----ATCC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Aporcelaimellus obtusicaudatus	-----ATCC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Wilsonema schuurmansstekhoveni	-----ATCC- -----TACT TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCA-A
Mesodorylaimus_sp_cf_nigritul	-----ATCT- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Mesodorylaimus bastiani	-----ATCT- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Mesodorylaimus japonicus	-----TTCC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Pungentus_sp_PDL_2005	-----ATCC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Allodorylaimus_sp	-----ATCT- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Eudorylaimus carteri	-----ATCC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Microdorylaimus_sp	-----ATCT- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCG-A
Longidorus elongatus	-----ATCC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Xiphinema rivesi	-----ATCC- -----TACT CGGATAAC-- TGTGGCAATT CTAGAGCTAA TACGTCG-A
Tylencholaimus_sp	-----TTCC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Anoplostoma_sp_BHMM_2005	-----T--- -----TGAT AGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCA-C
Adoncholaimus fuscus	-----T--- -----TACA TGGATAAC-- TGTGGTAATT CTACAGCTAA TACACGCA-T
Enoplus meridionalis	-----GT--- -----TACA TGGATAAC-- TGTGGTAACC TAAGAGCTAA TACGCGCA-A
Enoplus brevis_U88336	-----GT--- -----TACA CGGATAACC TGTGGTAACC TAAGAGCTAA TACACGCA-A
Enoplus communis	-----GT--- -----TACA CGGATAACC TGTGGTAACC TAAGAGCTAA TACGCGCA-A
Enoploides brunettii	-----C--- -----TACT TGGATAAC-- TGTGGTAACC TAAGAGCTAA TACATGCA-A
Syringolaimus striatocaudatus	-----TTT- -----TACA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACACGCA-T
Ironus dentifurcatus	-----TCC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-A
Calyptronema maxweberi	-----TAC- -----TACA TGGATAAC-- TGTGGTAATT CTACAGCTAA TACACGCT-C
Viscosia_sp_BHMM_2005	-----TCC- -----TACA TGGATAAC-- CGTGGTAATT CTACGCTAA TACACGCT-T
Viscosia viscosa	-----TCC- -----TACA TGGATAAC-- TGTGGTAATT CTACAGCTAA TACACGCA-T
Pontonema vulgare	-----AAT- -----TACA TGGATAAC-- TGTGGTAATT CTACAGCTAA TACACGCA-T
Oncholaimus_sp_BHMM_2005	-----AT--- -----TACA TGGATAAC-- TGTGGTAATT CTACAGCTAA TACACGCA-T
Alaimus_sp_PDL_2005	-----TCC- -----TACA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCC-T
Prismatolaimus intermedius	-----TCC- -----TACT TGGATAAC-- TGTGGTAATT CTAGAGCTA- TACACGCA-T
Tobrilus gracilis	-----TCC- -----TACT TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACACGCA-G
Tripyla_cf_filicaudata_JH_2004	-----TCC- -----TACT TGGATAAC-- TGAGCTAATT GTTGAGCTAA TACATGCA-C
Bathylaimus_sp	-----TAC- -----TACA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCA-A
Bathylaimus assimilis	-----TAC- -----TACA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCA-A
Bathylaimus_sp_BHMM_2005	-----ACC- -----TACA TGGATAAC-- TGAGGTAATT CTTGAGCTAA TACATGCT-A
Tripylloides_sp_BHMM_2005	-----TAC- -----TACA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCT-A
Trischiostoma monohystera	-----TCC- -----TACT TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACATGCC-T
Mermis nigrescens	-----ATC- -----TACT TGGATAAC-- TGCGGTAATT CTAGAGCTAA TACATGCA-A
Mylonchulus arenicolus	-----TCT- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-C
Anatonchus tridentacus	-----ATC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-C
Mononchulus truncatus	-----ATC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-C
Prionchulus muscorum	-----ATC- -----TACT CGGATAAC-- TGTGGCAATT CTAGAGCTAA TACGTCGCA-C
Clarkus_sp	-----ATC- -----TACT TGGATAAC-- TGTGGCAATT CTAGAGCTAA TACATGCA-C
Trichinella spiralis	-----ATCA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACACGCA-C
Trichinella papuae	-----TACA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACACGCA-C
Trichinella britovi	-----ATCA TGGATAAC-- TGTGGTAATT CTAGAGCTAA TACACGCA-C
NUCLEOTIDES INCLUDED	-----mmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm--

	190	200	210	220	230	240	
<i>Trichinella murrelli</i>	-----	-ATCA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C	
<i>Trichinella pseudospiralis</i>	-----	-TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C	
<i>Trichinella nativa</i>	-----	-ATCA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C	
<i>Trichinella zimbabwensis</i>	-----	-TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C	
<i>Trichinella nelsoni</i>	-----	-CTCA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C	
<i>Trichuris suis</i>	-----	-CACT	TGGATAAC--	TGTGGAAATT	CTAGAGCTAA	TAC-TGCC-T	
<i>Trichuris trichiura</i>	-----	-C----	-CACT	TGGATAAC--	TATGGAAATG	CTAGAGCTAA	TACATGCC-T
<i>Trichuris muris</i>	-----	-C----	-CACT	TGGATAAC--	TGTGGAAATT	CTAGAGCTAA	TACATGCC-T
<i>Tyolaimophorus minor</i>	-----	-TACA	TGGATAAC--	TGTGGTAATT	CAAGAGCTAA	TACATGCC--	
<i>Paratrichodorus pachydermus</i>	-----	-TTCT-	-TACA	CGGATAAC--	TGCGGTAATT	CTGGAGCTAC	TACGTGCA-T
<i>Paratrichodorus anemones</i>	-----	-TCT--	-TACA	CGGATAAC--	TGCGGTAATT	CTGGAGCTAC	TACGTGCA-T
<i>Trichodorus primitivus</i>	-----	-TTCC-	-TACA	AGGACACC--	TGCGGTAATT	CTGGAGCTAA	TACTTGCC--
<i>Axonolaimus helgolandicus</i>	-----	-TCC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA--
<i>Ascolaimus elongatus</i>	-----	-TCC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA--
<i>Odontophora rectangula</i>	-----	-TCC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Cylindrolaimus sp. 202149</i>	-----	-TCC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Tylocephalus auriculatus</i>	-----	-ATCC-	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Plectus acuminatus_BS9</i>	-----	-ATCC-	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Plectus aquatilis</i>	-----	-ATCC-	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Anaplectus sp</i>	-----	-ATCC-	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Anisakis sp_WKT</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Anisakis sp_Nadler</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Contracecum multipapillatum</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Pseudoterranova decipiens</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Raphidascaris acus</i>	-----	-tcc--	-tacg	tggataac--	tgtggttaatt	ctagagctaa	tacatgca-c
<i>Terranova caballeroi</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Ascaris suum</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Ascaris lumbricoides</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Baylisascaris procyonis</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Baylisascaris transfuga</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Parascaris equorum</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Porrocaecum depressum</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Toxascaris leonina</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Heterocheilus tunicatus</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Goezia pelagia</i>	-----	-TCCC-	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Hysterothylacium fortalezae</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Hysterothylacium pelagicum</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Hysterothylacium reliquens</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Iheringascaris iniquies</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Toxocara canis</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Nemhelix bakeri</i>	-----	-TCC--	-TACT	TGGATAAC--	TGAGGTAATT	CTAGAGCTAA	TACAATGCAG
<i>Raillietnema sp. V3060</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Cruzia americana</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Heterakis sp_14690</i>	-----	-TCC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Heterakis gallinarum</i>	-----	-TCC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Paraspidodera sp_21303</i>	-----	-TCC--	-TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
<i>Chromadora nudicapitata</i>	-----	-GT--	-TACT	TGGATAAC--	TGTTCAAAG	GAAGAGCTAA	GACATGCC-T
<i>Chromadora sp. BHMM_2005</i>	-----	-----	-CACG	GCAATAAC--	TGTGGAAAAG	CCAGAGCTAA	CTC-TGCA-T
<i>Atrochromadora microlaima</i>	-----	-----	-CACG	GCAATAAC--	TGTGGAAAAG	CCAGAGCTAA	CTC-TGCA-T
<i>Chromadorina germanica</i>	-----	-CC--	-TACT	TGGATAAC--	TGTGGAAAAT	CCAGAGCTAA	TACATGCACC
<i>Chromadorita tentabundum</i>	-----	-TCC--	-TACT	TGGATAAC--	TGTGGAAAAT	CTAGAGCTAA	TACACGCA-A
<i>Dichromadora sp. BHMM_2005</i>	-----	-----	-CACG	GCAATAAC--	TGTGGAAAAG	CCAGAGCTAA	CTCTGCAT--
<i>Neochromadora BHMM_2005</i>	-----	-TCC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Spilophorella paradoxa</i>	-----	-CC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Paracanthochus caecus</i>	-----	-CA--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Paracyatholaimus intermedius</i>	-----	-CA--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Praeacanthochus punctatus</i>	-----	-C----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Praeacanthochus sp</i>	-----	-C----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Cyatholaimus sp. BHMM_2005</i>	-----	-CA--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-A
<i>Spirinia parasitifera</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGTG-A
<i>Acanthopharynx micans</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Xyzzors sp</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Metachromadora sp</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Metachromadora remanei</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Catanema sp</i>	-----	-----	-TACT	CGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACGTGCC-A
<i>Eubostrichus diana</i>	-----	-T--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Eubostrichus topiarus</i>	-----	-T--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Eubostrichus parasitiferus</i>	-----	-T--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Laxus oneistus</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACGTGCC-A
<i>Laxus cosmopolitus</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Leptonemella sp</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Robbea hypermnestra</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGTA-T
<i>Stilbonema majum</i>	-----	-----	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
<i>Monoposthia costata</i>	-----	-GC--	-TACT	CGGATAAC--	CGTAGTAATT	CTAGAGCTAA	TACGTGCC-A
<i>Nudora bipapillata</i>	-----	-----	-AATA	TGGATATC--	TGTGCTAATT	GTAGAGCTAA	TACACGCA-G
<i>Calomicrolaimus parahonestus</i>	-----	-TC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Calomicrolaimus sp. BHMM_2005</i>	-----	-TC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
<i>Molgolaimus demani</i>	-----	-TC--	-TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA--
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	-----

	190	200	210	220	230	240
Diplogaster_lethieri	-----TCC-----	AAAA	CGGATATC--	TGCGTTAATT	TTGGAGCTAA	TACGTGCA-C
Aduncopticulum_halicti	-----TCC-----	TAAA	CGGATATC--	TGCGATAATT	TTGGAGCTAA	TACGTGCC-C
Pristionchus_lheritieri	-----TCC-----	AAAA	CGGATATC--	TGCGTTAATT	TTGGAGCTAA	TACGTGCA-C
Pristionchus_pacificus	-----TCC-----	AAAA	CGGATATC--	TGCGTTAATT	TTGGAGCTAA	TACGTGCA-C
Pristionchus_pacificus	-----TCC-----	AAAA	CGGATATC--	TGCGTTAATT	TTGGAGCTAA	TACGTGCA-C
Sabatieria_punctata_STRAIN_343	-----TT--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Sabatieria_sp._355_BHMM_2005	-----TT--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Sabatieria_celtica	-----TC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCC-T
Sabatieria_punctata_STRAIN_200	-----T--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Sabatieria_punctata_STRAIN_223	-----TT--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Sabatieria_sp._210_BHM_2005	-----TT--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Setosabatieria_hilarula	-----TC--	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCC-T
Desmolaimus_zeelandicus	-----TCC-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA--
Terschellingia_longicaudata	-----TCC-----	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACACGCA-C
Cyartonema_elegans	-----TCC-----	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACACGCC-C
Tridentulus_sp	-----C---	-----TACT	TGGATAATC--	TCTGGTAATT	CTAGAGCTAA	TACATGCA--
Diplolaimelloides_meyli	-----C---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCAA-
Diplolaimella_diavengatensis	-----C---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCCT-T
Geomonhyстера_disjuncta	-----T---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCCA-
Sphaerolaimus_hirsutus	-----T---	-----TACT	TGGATAAC--	TGTCGAAATG	CGAGAGCTAA	TACATGCAA-
Theristus_acer	-----T---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACACGCAT-T
Daptonema_procerus	-----T---	-----TACT	TGGATAATG--	TGCTGCAATT	CAGGACTTAA	TACATGCAA-
Daptonema_hirsutum	-----TT--	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCAA-
Daptonema_normandicum	-----A---	-----TACT	TGGATAAC--	TGAGGTAATT	CTTGAGCTAA	TACACGCCA-
Daptonema_oxycerca	-----T---	-----TACT	TGGATAATG--	TGCTGCAATT	CAGGACTTAA	TACATGCAA-
Daptonema_setosum	-----TT--	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCAA-
Desmodora_communis	-----T---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCAA-
Desmodora_ovigera	-----T---	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-A
Metadesmolaimus_sp	-----TT--	-----TACT	TGGATAAC--	TGTGGCAATT	CTAGAGCTAA	TACATGCAA-
Dentostomella_sp	-----TCC-----	-----TACG	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Bunonema_franzi	-----CC--	-----CAAG	TGGATAAC--	TTTGGTAATT	CTGGAGCTAA	TACATGCCT-T
Bunonema_sp	-----TCC-----	-----CAAT	TGGATAAC--	TTTGGTAATT	CTGGAGCTAA	TACATGCCT-T
Seleborca_complexa	-----ACAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Acrobeloides_nanus	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Acrobeloides_bodenheimeri	-----CTAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Acrobeloides_sp_PS1146	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Acrobeles_ciliatus	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Acrobeles_sp_PS1156	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Acrobeles_complexus_WCUG2	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Cephaloboides_sp_SB227	-----TCCT	C---TATG	AGGATAAC--	TGCGATAAAT	TTGGAGCTAA	TACATATT-T
Cephalobus_cubaensis	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Cephalobus_sp._PS1143	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Cephalobus_sp._PS1196	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Cephalobus_oryzae_PS1165	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Cervidellus_alutus	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Pseudacrobeles_variabilis	-----TCAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Triligulla_aluta	-----CAAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Zeldia_punctada	-----CTAT	CC---TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Myolaimus_sp_U81585	-----TCC-----	-----TACT	TGGACATATC	TGAGGAAAT	CTTGAGCTAA	TACATGCA-C
Rhabditophanes_sp._KR3021	-----C---	-----TAGT	TGGATAAC--	TGAGGTAATT	CTTGAGCTAA	TACACGCAAT
Brevibucca_sp._SB261	-----TCC-----	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Halicephalobus_gingivalis	-----CAT-	-----TAAA	TGGATAATC--	TGCGGTAAT	CTGGAGCTAA	TACATGCA-A
Panagrobelus_stammeri	-----TNTA	TCT---TACA	TGGATAAC--	NGTGGTAATT	CTGGAGCTAA	TACATGCAT-T
Plectonchus_sp._PDL0025	-----TTAT	CC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-C
Turbatrix_aceti	-----TATC	-----TACA	CGGATAAC--	TTTGGCAATT	CTGGAGCTAA	TACGTGCA-T
Panagrellus_redivivus	-----CTTT	CC---TATA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrellus_redivivus_PS1163	-----CTTT	CC---TATA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus_subelongatus	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus_davidi	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus_cf_rigidus_AF40	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus_sp._Sourhope_ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus_sp._Sourhope_ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus_sp._Sourhope_ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Panagrolaimus_sp._Sourhope_ED2	-----TTTA	TCC---TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-T
Steinernema_carpocapsae	-----ATCC	TT---TAAA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGGT-T
Strongyloides_stercoralis	-----TGC-----	-----TACA	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCTKA
Strongyloides_ratti	-----T---	-----TAGT	TGGATAAC--	TGAGGTAATT	CTTGAGCTAA	TACACGCC-T
Diploscapter_sp_PS1897	-----T---	-----CAAT	GGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACCTGCA-C
Diploscapter_sp_PS2017	-----T---	-----CAAT	GGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACCTGCA-C
Heterorhabditis_bacteriophora	-----T---	-----TTTT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Heterorhabditis_hepialus	-----T---	-----TTTT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Heterorhabditis_zelandica	-----T---	-----TTTT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Parasitorhabditis_sp_SB281	-----C---	-----CGTA	TGGATAAC--	TCTAGTAATT	CTGGAGCTAA	TACATFACT-A
Rhabditoides_inermiformis	-----ATCC	-----TACA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-C
Rhabditoides_inermis_DF5001	-----A---	-----TCTA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	AACATGCA-T
Rhabditoides_regina_DF5012	-----T---	-----CTTA	TGGATAAC--	TGACGTAATT	CTGGAGCTAA	TACACGTG-C
Poikilolaimus_oxycerca_SB200	-----ATCC	-----TACT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	190	200	210	220	230	240
Poikilolaimus regenfussi_SB199	-----ATCC	-----TACT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Distolabrellus veechi_DWF1604	-----	-----TGAG	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TACATGTT-C
Distolabrellus veechi_DF5024	-----	-----TGAG	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TACATGTT-C
Choriorhabditis dudichi	-----	-----CCCM	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Protorhabditis_sp	-----ATCC	-----TTTT	GGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACCTGCA-C
Protorhabditis_sp_DF5055	-----	-----CCTT	GGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACCATTA-A
Cruzema tripartitum_DF5015	-----	-----AAGA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Mesorhabditis_sp_PS1179	-----	-----ATTT	TGGATACT--	TCTAGTAATT	CTGGAGCTAA	TACACGTG-T
Mesorhabditis spiculigera_SB15	-----	-----TCAA	TGGATACT--	TCTAGTAATT	CTGGAGCTAA	TACATGCA-A
Mesorhabditis anisomorpha_SB12	-----	-----TCCC	CGGATACT--	TCTAGTAATT	CTGGAGCTAA	TACGGCGC-A
Teratorhabditis palmarum_DF501	-----	-----TTCT	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TAC--GTA-A
Teratorhabditis synpapillata_S	-----	-----TTTC	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TAC--GTA-C
Caenorhabditis briggsae_PB102	-----	-----AATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis elegans_N2	-----GATCC	-----TCATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis_sp_CB5161	-----	-----TATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis_sp_PS1010	-----	-----TAATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis japonica	-----	-----TGAAA	AGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCG-A
Caenorhabditis_sp_DF5170	-----	-----TATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-T
Caenorhabditis_sp_SB341	-----	-----CAAA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis plicata	-----	-----CTTAAA	TGGATACT--	TGCGGAAATT	CTGGAGCTAA	TACATGCC-G
Caenorhabditis vulgaris	-----	-----TATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis sonoreae	-----	-----TCTTA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Caenorhabditis drosophilae	-----	-----CATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-T
Phasmarhabditis hermaphrodita	-----	-----CTAT	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-A
Phasmarhabditis neopapillosa	-----	-----CTTT	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-A
Phasmarhabditis neopapillosa	-----	-----CTGA	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Pellioiditis mediterranea_SB173	-----	-----CTAG	TGGATAAC--	TTGCGTAATT	CTGGAGCTAA	TACATGCA-A
Pellioiditis marina	-----	-----CTGA	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Prodontorhabditis wirthi	-----	-----AACG	AGGATACT--	TGCGGAAATA	CTGGAGCTAA	TACTTGCTA
Crustorhabditis scanica	-----	-----CTTT	TGGATACT--	TTCAGTAATT	CTGGAGCTAA	TACATGCC-C
Dolichorhabditis_sp_CEW1	-----	-----TTAG	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGTC-A
Oscheius_sp_DF5000	-----	-----TTAA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Oscheius_sp_BW282	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Oscheius myriophila_EM435	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Oscheius insectivora	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Oscheius dolichuroides	-----	-----TTAA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Rhabditis blumi_DF5010	-----	-----CCTT	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Rhabditis_sp_PS1191	-----	-----CATT	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCA-A
Rhabditis_sp_PS1010	-----	-----AATA	TGGATAAC--	TGCGGAAATA	CTGGAGCTAA	TACATGCC-A
Rhabditis myriophila_EM435	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Rhabditis colombiana	-----	-----TTTA	TGGATAAC--	TGCGGAAATT	CTGGAGCTAA	TACATGCA-A
Rhabditella_axei_DF5006	-----	-----TATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Rhabditella_sp_DF5044	-----	-----TATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Cuticularia_sp_PS2083	-----	-----TACT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-T
Necator americanus	-----ATCC	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Kalichephalus cristatus	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Ancylostoma caninum	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Angiostrongylus cantonensis	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-T
Angiostrongylus costaricensis	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-T
Angiostrongylus malaysiensis	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-T
Angiostrongylus dujardini	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-G
Angiostrongylus vasorum	-----	-----TTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Aulurostrongylus abstrusus	-----	-----CACC	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-G
Didelphostrongylus hayesi	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-G
Crenosoma mephitidis	-----	-----CATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Crenosoma vulpis	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Otostrongylus circumlitus	-----	-----CATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Troglostrongylus wilsoni	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Filaroides martis	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Oslerus osleri	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Parafilaroides decorus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Metastrongylus salmi	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Metastrongylus elongatus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Skrjabinigylus chitwoodrum	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Parelaphostrongylus odocoilei	-----	-----TTAA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATG-C
Muellerius capillaris	-----	-----ACAA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Protostrongylus rufescens	-----	-----AACT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Halocercus invaginatus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-a
Pseudalius inflexus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATTCT-A
Stenurus minor	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-A
Torynurus convolutus	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-A
Syngamus trachea	-----	-----GTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Stephanurus dentatus	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Nematodirus battus	-----	-----TTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-T
Strongylus equinus	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Labiostongylus bipapillosus	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Petrovinaema poculatum	-----	-----CTTT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
Cylicocyclus insignis	-----	-----CTAT	TGGATAAC--	TGCGGCAATT	CTGGAGCTAA	TACATGCC-A
NUCLEOTIDES INCLUDED	-----	-----nnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn	nnnnnnnnnnnn

	190	200	210	220	230	240
Chabartia_ovina	-----	CTTT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCG-A
Cyclodontostomum_purvisi	-----	CTTT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCG-A
Zoniolaimus_mawsonae	-----	CTAT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCG-A
Hypodontus_macropi	-----	CTAT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCG-A
Deletrocephalus_dimidiatus	-----	CTAT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Dictyocaulus_eckerti_P7B8	-----	CGAA	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Dictyocaulus_capreolus_P3B2	-----	CGAA	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Dictyocaulus_sp._P6A1	-----	CGAA	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Dictyocaulus_filaria	-----	ACTA	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Dictyocaulus_capreolus_P2C10	-----	CGAA	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Dictyocaulus_viviparus	-----	CGAA	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Haemonchus_sp._V3091	-----	CTTT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Haemonchus_contortus	-----	CTTT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Haemonchus_placei	-----	CTTT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Haemonchus_similis	-----	CTTT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Ostertagia_ostertagi	-----	CTAT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ostertagia_leptospicularis	-----	CTAT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Nippostrongylus_brasileinsis	-----	CTAT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Heligmosomoides_polygyrus	-----	CTAT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Trichostrongylus_colubriformis	-----	CTAT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGTA-A
Tetrahelminthostrongylus_mackerr	-----	TTTT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCG-A
Herpestrongylus_pythonis	-----	CTAT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Nicollina_cameroni	-----	CTAT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Filarimena_flagrifer	-----	GTTT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Amidostomum_cygni	-----	CTTT	TGGATAAC	--	TGCGGCAATT	CTGGAGCTAA	TACATGCA-A
Teratocephalus_lirellus	-----	TCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACACGCA-C
Brumptaemilius_justini	-----	TCC	TGGATAAC	--	TGTGRTAATT	CTAGAGCTAA	TACATGCA-C
Anguillicola_crassus	-----	TAC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Dracunculus_medinensis	-----	TCCA	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Dracunculus_oesophageus	-----	TCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Dracunculus_sp._V3104	-----	TCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Philonema_sp_A	-----	TCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Philometra_obturus	-----	CCA	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Camallanus_oxyccephalus	-----	TCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Acanthocheilonema_viteae	-----	ATCC	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Loa_loa	-----	ATCC	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Onchocerca_cervicalis	-----	ATCC	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Dirofilaria_immitis	-----	ATCC	TGGATAAC	--	TGTGGCAATT	CTAGTGTGA	TACATGCA-C
Brugia_malay	-----	ATCT	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Wuchereria_bancrofti	-----	ATCC	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Litomosoides_sigmodontis	-----	GGCC	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Setaria_digitata	-----	ATCC	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Gnathostoma_turgidum	-----	CTC	TGGATATC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Gnathostoma_neoprocyonis	-----	TTC	TGGATATC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Gnathostoma_binucleatum	-----	TTC	TGGATATC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Physaloptera_alata	-----	ATCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Physaloptera_turgida	-----	ATCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Ascarophis_arctica	-----	ATCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Spinitectus_carolini	-----	ATCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Thelazia_lacrymalis	-----	TATCC	TGGATAAC	--	TGTGGCAATT	CTAGAGCTAA	TACATGCA-C
Aphelenchus_avenae	-----	GAAT	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCC-C
Aphelenchoides_fragariae	-----	TT	TGGATAAC	--	TGCGGTAATT	CTGGAGCTAA	TACATGCC-C
Bursaphelenchus_sp	-----	C	TGGATAAC	--	TGCGGTAATT	TGGAGCTAA	TACATGCA-T
Deladenus_sp	-----	ATAC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-T
Criconema_sp	-----	AGT	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Hemicyclophora_conida	-----	CCAC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACTTGCA-C
Paratylenchus_dianthus	-----	AAC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Pratylenchus_thornei	-----	ACC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Tylenchulus_semipentrans	-----	AGT	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Ditylenchus_angustus	-----	TATA	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Subanguina_radicola	-----	ATCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Tylenchorhynchus_maximus	-----	ACC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Geocenamus_quadriifer	-----	TCC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Globodera_pallida	-----	TCT	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Scutellonema_bradys	-----	ACC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Helicotylenchus_dihystera	-----	ACC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Rotylenchus_robustus	-----	ACC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Meloidogyne_arenaria	-----	GTC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_incognita_KT	-----	GTC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_javanica	-----	GTC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_artiellia	-----	CTCC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_duytsi	-----	ATCC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_exigua	-----	TATC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_hapla	-----	ATCC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_ichinohei	-----	ATCC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_maritima	-----	ATCC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Meloidogyne_microtyla	-----	ATCC	TGGATAAC	--	TGTGGAAAAG	CTAGAGCTAA	TACATGCA-C
Nacobbus_aberrans	-----	ACC	TGGATAAC	--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
NUCLEOTIDES INCLUDED	-----	mmmm	mmmmmmmm	--	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	190	200	210	220	230	240
Pratylenchoides ritteri	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Pratylenchoides magnicauda	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Hirschmanniella_sp._JH_2003	-----CACC	-----TACM	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Radopholus similis	-----ACC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Pratylenchus goodeyi_VF	-----CCCA	-----CACA	CGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Boleodorus thylactus_clone2	-----ATCC	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Philippine Sequence 1	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Philippine Sequence 2	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Philippine Sequence 3	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Philippine Sequence 4	-----TAAA	TCC--TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-C
Philippine Sequence 5	-----	-----CTTT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Philippine Sequence 6	-----	-----TTTC	AGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Philippine Sequence 7	-----	-----TATA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGTC-A
Tahiti Sequence 1	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Tahiti Sequence 2	-----	-----CTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-T
Thailand Sequence 1	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Ogasawara Sequence 1	-----TAAA	TCC--TACA	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-C
Ogasawara Sequence 2	-----	-----TATTT	TGGATAAC--	TGTGGTAATT	CTGGAGCTAA	TACATGCA-C
Ivory Coast Sequence 1	-----	-----CAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 2	-----	-----AAAC	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 3	-----	-----TAATT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 4	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 5	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Ivory Coast Sequence 6	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Ivory Coast Sequence 7	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATTCT-A
Singapore Sequence 1	-----	-----ATCTAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACACGTA-A
Singapore Sequence 2	-----TCC-	-----TACT	TGGATAAC--	TGTGGTAATT	CTAGAGCTAA	TACATGCA-C
Singapore Sequence 3	-----	-----AATA	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCA-A
Singapore Sequence 4	-----	-----GAAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Nigerian Sequence 1	-----	-----TAAC	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Nigerian Sequence 2	-----	-----CTTT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCG-A
Nigerian Sequence 3	-----	-----ATAT	TGGATAAC--	TGCGGTAATT	CTGGAGCTAA	TACATGCT-A
NUCLEOTIDES INCLUDED	-----	-----m	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	250	260	270	280	290	300
Gordius aquaticus	AAAAG-TCGA	ATC-----	-----	-----	GCGAGGTCGG	ACCGCTTTT-
Priapulid caudatus	TCAAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCTTTT-
Brachionus plicatilis	AAAAGCTCCG	ACC-----	-----	-----	GTATGGGAAG	AGCGCTTTT-
Chordodes morgani	AAACG-TCGA	ATC-----	-----	-----	GCGAGGTTTCG	ACTGCTTTT-
Paractinolaimus macrolaimus	AAAAGCTCTG	CCC-----	-----	-----	GAAAGGAACG	AGCGCATTT-
Aporcelaimellus obtusicaudatus	AAAAGCTCCG	CCC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Wilsonema schuurmansstekhoveni	ACAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCATTT-
Mesodorylaimus_sp._cf._nigritul	AAAAGCTCCG	ACC-----	-----	-----	GAAAGGAACG	AGCGCATTT-
Mesodorylaimus bastiani	AAAAGCTCTG	ACC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Mesodorylaimus japonicus	AAAAGCTCTG	CCCT-----	-----	-----	TCACGGAAAG	AGCGCATTT-
Pungentus_sp._PDL_2005	TAAAGCTCTG	CCC-----	-----	-----	GTAAGGAACG	AGCGCATTT-
Alloorylaimus_sp	TAAAGCTCCG	CCC-----	-----	-----	GAAAGGAACG	AGCGCATTT-
Eudorylaimus carteri	TAAAGCTCTG	CCC-----	-----	-----	GCAAGGAACG	AGCGCATTT-
Microdorylaimus_sp	TAAAGCTCTG	ACC-----	-----	-----	GAAAGGAACG	AGCGCATTT-
Longidorus elongatus	AAAAGCTCAG	ACT-----	-----	-----	GAAAGGAATG	AGCGCATTT-
Xiphinema rivesi	AAAAGCTCGT	TTC-----	-----	-----	GCAAGATTCG	AGCGCATTT-
Tylencholaimus_sp	AAAAGCTCCG	CCC-----	-----	-----	GAAAGGAACG	AGCGCATTT-
Anoplostoma_sp._BHMM_2005	TTATATCCAG	ACC-----	-----	-----	TTACGGAATG	GATGCAGTT-
Adoncholaimus fuscus	CAAAACCCCG	AC-----	-----	-----	-TAACGAAGG	GGTGCGTTT-
Enoplus meridionalis	TTATGCCCCTG	ACT-----	-----	-----	TCACAGGAAG	GGCGCGGTT-
Enoplus brevis_U88336	TTAAGTCCAG	ACC-----	-----	-----	TCACGGAACG	GACGCGGTT-
Enoplus communis	TTAAGTCCAG	ACCT-----	-----	-----	-CACGGAACG	GACGCGGTT-
Enoploides brunettii	CAAAAGCCCTA	TTGC-----	-----	-----	-----AG	GGCGCATTT-
Syringolaimus striatocaudatus	TTATGCTCAG	ACC-----	-----	-----	TTACGGAATG	AGCGCATTT-
Ironus dentifurcatus	CAAAAGCCCTG	ACC-----	-----	-----	GTAAGGAAGG	GTTGCACCTT-
Calyptonema_maxweberi	CAAAACCCCA	ACT-----	-----	-----	-TTACGGAGG	GGTGCGTTT-
Viscosia_sp._BHMM_2005	CAAAACCCAA	CCT-----	-----	-----	-TACGGTGG	GGTGCGTTT-
Viscosia viscosa	CAAAACCCAA	CCT-----	-----	-----	-TACGGTGG	GGTGCGTTT-
Pontonema_vulgare	CAAAACACCC	GACT-----	-----	-----	TCGATGGAGG	GGTGCGATTT
Oncholaimus_sp._BHMM_2005	CAAAAGCCCTG	ACT-----	-----	-----	-TCGGAAGG	GGTGCGTTT-
Alaimus_sp._PDL_2005	TGAAGCCGAA	ACC-----	-----	-----	TTACGGAATG	GTTGCACCTT-
Prismatolaimus intermedius	GAAAGCTCTG	ACC-----	-----	-----	GTTTCGGGAGG	AGCGCATTT-
Tobrilus gracilis	CAAAAGCTCGG	ACC-----	-----	-----	TCACG-AAAC	AGCGCATTT-
Tripyla_cf._filicaudata_JH_2004	CAAAAGTTCG	ACC-----	-----	-----	TTACGGAAGG	AGCGCATTT-
Bathylaimus_sp	CAAA-CGGCA	CCC-----	-----	-----	TTATGGGAGC	CGCGCTTTT-
Bathylaimus assimilis	CAAA-CGGCA	CCC-----	-----	-----	TTATGGGAGC	CGCGCTTTT-
Bathylaimus_sp._BHMM_2005	CAAA-CGGCG	ACC-----	-----	-----TTTC	ACGAGGGAGC	CGCGCTTTT-
Tripylodes_sp._BHMM_2005	CAAA-CTGTG	ACTTC-----	-----	-----GC	AAGAAGAAGC	AGTGCTTTT-
Trischistoma monohystera	TCAA-CTCTG	ACC-----	-----	-----	TTACGGAAGG	AGCGCGTTT-
Mermis nigrescens	GAAAGTCCG	ACC-----	-----	-----	TTCCGGAAAG	AGCGCATTT-
Mylonchulus arenicolus	GAAAGTCCG	ACCC-----	-----	-----	TTGAGGAAGG	AGCGCATTT-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	250	260	270	280	290	300
Anatonchus tridentacus	GTAAGCTCGG	ACC-----	-----	-----	TTAGGAAAAG	AGCGCATTT-
Mononchus truncatus	GAAAGCTCGG	ACC-----	-----	-----	TTT-GGAAAG	AGCGCATTT-
Prionchulus muscorum	GAAGGCTCGG	ACC-----	-----	-----	TTTTGGGAAG	AGCGCATTT-
Clarkus_sp	GCAGGCTCGG	ACC-----	-----	-----	TTTTGGGAAG	AGCGCATTT-
Trichinella spiralis	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCCTTT-
Trichinella papuae	AAAAACTTCA	ACTG-----	-----	-----	AAATGGTTGA	AGTGCCTTT-
Trichinella britovi	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCCTTT-
Trichinella murrelli	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCCTTT-
Trichinella pseudospiralis	AAAAACTTCA	ACTA-----	-----	-----	TA CAATGGTTGA	AGTGCCTTT-
Trichinella nativa	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCCTTT-
Trichinella zimbabwensis	AAAAACTTCA	ACTG-----	-----	-----	AAATGGTTGA	AGTGCCTTT-
Trichinella nelsoni	AAAAACTTCA	ACTG-----	-----	-----	AATTGGTTGA	AGTGCCTTT-
Trichuris suis	CGAAGCTCGG	TCGCG-----	-----	-----	CG CAACGCTCGG	AGCGCATTT-
Trichuris trichiura	CGAAGCTCAG	TCGC-----	-----	-----	-GCGC TCGCGCTCGG	AGCGCGTTT-
Trichuris muris	CAAAGCTTCG	GCCG-----	-----	-----	-GC AATGCGCGGA	GACGCATTT-
Tylolaimophorus minor	AAAAACTCAG	ACC-----	-----	-----	TTCGGGAATG	AGTGCCTTT-
Paratrachodorus pachydermus	TAAA-CGACG	ACC-----	-----	-----	TTACGGAAAGT	CGTGCATTT-
Paratrachodorus anemones	TAAA-CGACA	ACC-----	-----	-----	TCACGGAAAGT	CGTGCATTT-
Trichodorus primitivus	AAATGCGACA	ACC-----	-----	-----	TCACGGGAGT	CGTGCATTT-
Axonolaimus helgolandicus	AACAGCTCTG	ACC-----	-----	-----	GCAAGGGATG	AGCGCATTT-
Ascolaimus elongatus	AACAGCTCTG	ACC-----	-----	-----	GCAAGGGATG	AGCGCATTT-
Odontophora rectangula	C-AAGCTCTG	ACC-----	-----	-----	GCAAGGGATG	AGCGCATTT-
Cylindrolaimus_sp._202149	CAAAGCTCTG	ACC-----	-----	-----	GTAAGGGAGG	AGTGCATTT-
Tylocephalus auriculatus	AAAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCATTT-
Plectus acuminatus_BS9	TAAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCATTT-
Plectus aquatilis	TAAAGCTCCG	ACC-----	-----	-----	TTACGGGACG	AGCGCATTT-
Anaplectus_sp	CCAAGCTCCG	ACC-----	-----	-----	GTAAGGGAGG	AGCGCATTT-
Anisakis_sp_WKT	CAAAGCTCCG	AT-----	-----	-----	TA-TTTGACG	AGCGCATCT-
Anisakis_sp_Nadler	CAAAGCTCCG	AT-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Contracaecum multipapillatum	CAAAGCTCCG	AT-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Pseudoterranova decipiens	CAAAGCTCCG	AT-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Raphidascaris acus	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Terranova caballeroi	CAAAGCTCCG	AA-----	-----	-----	TC-ATTGACG	AGCGCATCT-
Ascaris suum	CAAAGCTCCG	AT-----	-----	-----	TT-TCTGACG	AGCGCATCT-
Ascaris lumbricoides	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Baylisascaris procyonis	CAAAGCTCCG	AT-----	-----	-----	TT-TCTGACG	AGCGCATCT-
Baylisascaris transfuga	CAAAGCTCCG	AT-----	-----	-----	TT-TCTGACG	AGCGCATCT-
Parascaris equorum	CAAAGCTCCG	AT-----	-----	-----	TT-TCTGACG	AGCGCATCT-
Porrocaecum depressum	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Toxascaris leonina	CAAAGCTCAG	AT-----	-----	-----	TT-TCTGACG	AGCGCATTT-
Heterocheilus tunicatus	CAAAGCTCCG	AC-----	-----	-----	TC-GTCGACG	AGCGCATCT-
Goezia pelagia	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Hysterothylacium fortalezae	CAAAGCTCCG	AA-----	-----	-----	TC-TTTGACG	AGCGCATCT-
Hysterothylacium pelagicum	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Hysterothylacium reliquens	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Iheringascaris iniques	CAAAGCTCCG	AA-----	-----	-----	TT-TTTGACG	AGCGCATCT-
Toxocara canis	CAAAGCTCCG	AT-----	-----	-----	TT-TGTGACG	AGCGCATCT-
Nemhelix bakeri	CAAAGCTCCG	AT-----	-----	-----	TT-T-TGACG	AGCGCATCT-
Raillietnema_sp._V3060	CAAAGCCCGG	AT-----	-----	-----	TT-T-TGACG	GGCGCATCT-
Cruzia americana	CAAAGCTCCG	AC-----	-----	-----	TC-GTTGACG	AGCGCATCT-
Heterakis_sp_14690	CAAAGCTCCG	AT-----	-----	-----	---AATGACG	AGCGCATCT-
Heterakis gallinarum	CAAAGCTCCG	AT-----	-----	-----	---TTTGACG	AGCGCATCT-
Paraspidodera_sp_21303	CGAAGCTCTG	AT-----	-----	-----	-T-TTTGACG	AGCGCATCT-
Chromadora nudicapitata	CGAAGGTCAA	GC-----	-----	-----	GC-AAGCTTG	ATCGCATTT-
Chromadora_sp._BHMM_2005	TCAAGTCCCG	AC-----	-----	-----	TT-CGGAAGG	GACGCATTT-
Atrochromadora microlaima	TCAAGTCCCG	AC-----	-----	-----	TT-CGGAAGG	GACGCATTT-
Chromadora_germanica	AACAG--CCA	AC-----	-----	-----	TT-CGGAAGG	CTGGCAATTT-
Chromadorita tentabundum	TCAAGCCCTA	ACCT-----	-----	-----	TT-TGAAAGG	GGCGCATTT-
Dichromadora_sp._BHMM_2005	TCAAGTCCCG	AC-----	-----	-----	TT-CGGAAGG	GACGCATTT-
Neochromadora_BHMM_2005	TCAAGCCCTA	AC-----	-----	-----	TGACGGGCGG	GGCGCATTT-
Spilophorella paradoxa	TTAAACCCCA	AAC-----	-----	-----	TTACGTGCGG	GGTGCATTT-
Paracanthonus caecus	CAAAACCCTG	AC-----	-----	-----	TT-CGGAAGG	GGTGCATTT-
Paracyatholaimus intermedius	CAAAACCCTG	AC-----	-----	-----	TT-CGGAAGG	GGTGCATTT-
Praeacanthonus punctatus	GAAAGCCCGG	AC-----	-----	-----	TT-CGGGAGG	GGCGCATTT-
Praeacanthonus_sp	GAAAGCCC-G	AC-----	-----	-----	TT-CGGGAGG	GGCGCATTT-
Cyatholaimus_sp._BHMM_2005	CAAAACCCTG	AC-----	-----	-----	TT-CGGAAGG	GGTGCATTT-
Spirinia parasitifera	AAAAGCTCCA	ACC-----	-----	-----	TTACGGAAGG	AGTGCATTT-
Acanthopharynx micans	CCAAGCTCTG	ACC-----	-----	-----	TTACGGAAGG	AGCGCATTT-
Xyzzors_sp	CCAAGCTCTG	ACC-----	-----	-----	TTGGGGAAGG	AGTGCATTT-
Metachromadora_sp	CCAAGCTCCG	ACC-----	-----	-----	TTGCGGAAGG	AGCGCATTT-
Metachromadora remanei	CCAAGCTCCG	ACC-----	-----	-----	TTGCGGAAGG	AGCGCATTT-
Catanema_sp	ACAAGCTCCG	ACT-----	-----	-----	TCGCGGAAGG	AGCGCATTT-
Eubostriechus diana	TCAGGCTGCG	ACCT-----	-----	-----	TTTTGGAAGC	AGTGCATTT-
Eubostriechus topiarus	CCAAGCTCCA	ACC-----	-----	-----	TTACGGAAGG	AGCGCATTT-
Eubostriechus parasitiferus	CCAAGCTCCA	ACC-----	-----	-----	TTACGGAAGG	AGCGCATTT-
Laxus oneistus	TCAAGCTCCG	ACC-----	-----	-----	TTACGGAAGG	AGTGCATTT-
Laxus cosmopolitus	TCAAGCTCCG	ACC-----	-----	-----	TTGCGGAAGG	AGCGCATTT-
Leptonemella_sp	CTCAGCTTCG	ACC-----	-----	-----	TTACGGAAGG	AGCGCATTT-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	250	260	270	280	290	300
Robbea hypermnestra	TCAAGCTCCG	CCC	-----	-----	TCACGGAAGG	AGTGCATTT-
Stilbonema majum	CAAAGCTCCG	ACC	-----	-----	TTACGGAAAG	AGCGCATTT-
Monoposthia costata	ACAACCCCG	ACT	-----	-----	TCT-GGGAGG	GGCGCATTT-
Nudora bipapillata	AAAAGCCCAA	GTT	-----	-----	TCT-----TG	GGTGCATTT-
Calomicrolaimus parahonestus	ACAAGTTCCG	ACG	-----	-----	--TAAGGAGG	AACGCATTT-
Calomicrolaimus sp. BHMM_2005	ACAAGTTCCG	ACG	-----	-----	--TAAGGAGG	AACGCATTT-
Molgolaimus demani	TCAAGTTCCA	ACG	-----	-----	--CAAGAAGG	AACGCATTT-
Diplogaster lethieri	CAAAGCACCG	CCA	-----	-----	GCAATGGTAG	TGCGCACAT-
Aduncospiculum halicti	CGAGTCAATG	GCC	-----	-----	TC-ACGGCGT	TGGCATT-
Pristionchus lheritieri	CAAAGCACCG	CCA	-----	-----	GCAATGGTAG	TGCGCACAT-
Pristionchus pacificus	CAACGTGCTG	CTA	-----	-----	GCAATAGTGG	CACGCATT-
Pristionchus pacificus	CAACGTGCTG	CTA	-----	-----	GCAATAGTGG	CACGCATT-
Sabatieria punctata STRAIN_343	TAAAGCTCTG	ACC	-----	-----	TTACGGGACG	AGCGCATTT-
Sabatieria sp. 355 BHMM_2005	TAAAGCTCTG	ACC	-----	-----	TTACGGGACG	AGCGCATTT-
Sabatieria celtica	TGAAGTCTG	ACC	-----	-----	TTCGGGGACG	AGCGCATTT-
Sabatieria punctata STRAIN_200	TAAAGCTCCG	ACC	-----	-----	TTACGGAACG	AGCGCATTT-
Sabatieria punctata STRAIN_223	CAAAGCTCCG	ACC	-----	-----	TTACGGGACG	AGCGCATTT-
Sabatieria sp. 210 BHM_2005	TAAAGTCCG	ACC	-----	-----	TTACGGAACG	AGCGCATTT-
Setosabatieria hilarula	TGAAGTCTG	ACC	-----	-----	TTCGGGGACG	AGCGCATTT-
Desmolaimus zeelandicus	AAGAACTCTG	CCC	-----	-----	TTCGGGAACG	AGTGCCTTT-
Terschellingia longicaudata	TGAAGCCCTG	ACC	-----	-----	TTCGGGGACG	GGTGCATTT-
Cyartonema elegans	TAAAGCCCG	ACC	-----	-----	TTCGGGAAGG	GGTGCATTT-
Tridentulus sp	TCAAGACCCG	ATC	-----	-----	--CAAGGACG	GTGCAATT-
Diplolaimelloides meylli	CTAATCCCG	ATC	-----	-----	GCAAGTGACG	GGAGCATT-
Diplolaimella diavengatensis	TGTAGTCTG	ATC	-----	-----	GTAAGTGATG	AGCGCATTT-
Geomonhystra disjuncta	CTAAGGGCCG	AAC	-----	-----	TCACGTGACG	CTTGCAATT-
Sphaerolaimus hirsutus	-GAAGCCCTG	ACC	-----	-----	TTTTGGGATG	GGTGCATTT-
Theristus acer	TCGAGTCCG	CCC	-----	-----	GCAAGGAACG	AGCGCATTT-
Daptonema procerus	TCGAGCCCCG	ACT	-----	-----	TCACAGGACG	GGCGCATTT-
Daptonema hirsutum	TCGAGTCCG	ACT	-----	-----	TAACAGGACG	AGCGCATTT-
Daptonema normandicum	TCGAGTCTG	ACC	-----	-----	-TTTTGGGACG	AGTGCATTT-
Daptonema oxycerca	TCGAGCCCCG	ACT	-----	-----	TCACAGGACG	GGCGCATTT-
Daptonema setosum	TCGAGTCCG	ACT	-----	-----	TAACAGGACG	AGCGCATTT-
Desmodora communis	ACAAGTCCG	ACC	-----	-----	TTACGGAAGG	AGCGCATTT-
Desmodora ovigera	CCAAGTCCG	ACC	-----	-----	TT-TGGAAGG	AGTGCCTTT-
Metadesmolaimus sp	TCGAGTCCG	ACTT	-----	-----	AACAGGACG-	AGCGCATTT-
Dentostomella sp	CAAAGTCTG	AT	-----	-----	TTATTGTGG	AGCGCATTT-
Bunonema franzi	CGATGCCCTG	ACC	-----	-----	GTGAGGAAAG	GGTGCAGTT-
Bunonema sp	CGATGCCCTG	ACC	-----	-----	GCAAGGAAAG	GGTGCAGTT-
Seleborca complexa	TAAAGTCCG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Acrobeloides nanus	TAAAGTCTG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Acrobeloides bodenheimeri	TAAAGTCTG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Acrobeloides sp PS1146	TAAAGTCCG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Acrobeles ciliatus	CTAAGTCTG	CCT	-----	-----	TCACGGGTAG	AGTGCATTT-
Acrobeles sp PS1156	CTAAGTCTG	CCT	-----	-----	TCACGGGTAG	AGTGCATTT-
Acrobeles complexus WCUG2	CTAAGTCTG	CCT	-----	-----	TCACGGGTAG	AGTGCATTT-
Cephaloboides sp SB227	AAAAG-CATG	CTC	-----	-----	TCACGAGCAT	-GTACCATT-
Cephalobus cubaensis	TAAAGTCTG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Cephalobus sp. PS1143	TAAAGTCTG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Cephalobus sp. PS1196	TAAAGTCTG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Cephalobus oryzae PS1165	TAAAGTCTG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Cervidellus alutus	TAAAGTCTG	ACC	-----	-----	TAACGGGAAAG	AGTGCATTT-
Pseudacrobeles variabilis	TAAAGTCTG	ACC	-----	-----	TTCCGGGAAAG	AGTGCATTT-
Triligulla aluta	TAAAGTCTG	ACC	-----	-----	TAACGGGAAAG	AGTGCATTT-
Zeldia punctata	TAAAGTCTG	ACC	-----	-----	TCACGGGAAAG	AGTGCATTT-
Myolaimus sp U81585	CAAAGTCCG	A	-----	-----	-TTCATGACG	AGCGCATTT-
Rhabditophanes sp. KR3021	CTAAGCCACA	TCC	-----	-----	-----GT	GGTGCCTTT-
Brevibucca sp. SB261	CAAATGCTCC	GAC	-----	-----	-TTCGGGACG	AGCGCACAT-
Halicephalobus gingivalis	TATATGTGT	ATA	-----	-----GT	TATCTATATG	ACGGCAGTT-
Panagrobelus stammeri	AAAAGCCCGC	CAG	-----	-----	-TAATGGTTG	GTTGCATTT-
Plectonchus sp. PDL0025	TAAAGCCCGG	CTA	-----	-----	GCGATAGTGG	GGTGCATTT-
Turbatrix aceti	TGATGCCTAA	CCT	-----	-----	----TTGTTG	GGTGCAGTT-
Panagrellus redivivus	AAAAGCGGGG	TCA	-----	-----C	TCGTTGGCAC	CGTGCATTT-
Panagrellus redivivus PS1163	AAAAGCGGGG	TCA	-----	-----C	TCGTTGGCAC	CGTGCATTT-
Panagrolaimus subelongatus	AAAAGATAWA	ACA	-----	-----TT	TTTWTGTTWT	ATTGCGTAT-
Panagrolaimus davidi	AAAAGATGTT	TTT	-----	-----	GTAATAA AAC	ATTGCATAT-
Panagrolaimus cf rigidus AF40	AAAAGATTAT	ACA	-----	-----TT	TTTATGTATA	ATTGCGTAT-
Panagrolaimus sp. Sourhope ED2	AAAAGACAAA	TTT	-----	-----CT	TATGGGATTT	GTTGCATAT-
Panagrolaimus sp. Sourhope ED2	AAAAGACAAA	TTT	-----	-----CT	TATGGGATTT	GTTGCATAT-
Panagrolaimus sp. Sourhope ED2	AAAAGATGTT	ATA	-----	-----CT	TATGGGATTT	GTTGCATAT-
Panagrolaimus sp. Sourhope ED2	AAAAGATGTT	ATA	-----	-----TT	TGATTGTAAC	ATTGCGTAT-
Steinernema carpocapsae	TGTAACAAA	CGT	-----	-----	-TTATCGGTT	TGTTCAATT-
Strongyloides stercoralis	AAAGCCCGA	CTTCT	-----	-----	-----GGAAGG	GGTGCATTT-
Strongyloides ratti	TTTATACCAC	ATT	-----	-----	-----AGT	GGTGCCTTT-
Diploscapter sp PS1897	TTAAAACCC	GAC	-----	-----	-GTAAGTAGG	GTTGCAATT-
Diploscapter sp PS2017	TTAAAACCC	GAC	-----	-----	-GTAAGTAGG	GTTGCAATT-
Heterorhabditis bacteriophora	CAATGCCCA	ACT	-----	-----	--TCGGAAGG	GGTGCATTT-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	250	260	270	280	290	300
Heterorhabditis hepialus	CTATGCCCCA	ACT	-----	-----	--TCGGAAGG	GGTGCAATT-
Heterorhabditis zelandica	CTATGCCCCA	ACT	-----	-----	TC--GGAAGG	GGTGCAATT-
Parasitorhabditis_sp_SB281	GCGCTTC	---	-----	-----	-----G	GCGTCATTT-
Rhabditoides inermiformis	CAAAGCTCTG	ACC	-----	-----	GTAAGGAACG	AGCGCATTT-
Rhabditoides inermis_DF5001	CAAACCCAG	ATC	-----	-----	GTAAGAGGCC	GGTGCAATT-
Rhabditoides regina_DF5012	GACCTT	---	-----	-----	-----CG	GGTTCATTT-
Poikilolaimus oxycerca_SB200	CAAAGCTCTG	ACA	-----	-----	---CCCTCGG	AGCGCATTT-
Poikilolaimus regenfussi_SB199	CAAAGCTCTG	ACA	-----	-----	---CCCTCGG	AGCGCATTT-
Distolabrellus veechi_DWF1604	GCACGGGCAA	CCG	-----	-----	-----	-TGTCATTT-
Distolabrellus veechi_DF5024	GCACGGGCAA	CCG	-----	-----	-----	-TGTCATTT-
Choriorhabditis dudichi	TATGCCCCGA	CTC	-----	-----	--ACGGAGG	GGTGCAATT-
Protorhabditis_sp	ACATAACCGG	ACG	-----	-----	--CAAGTACG	GTTGCAATT-
Protorhabditis_sp_DF5055	CAAAGCTTGG	AAG	-----	-----	--CAATTCCA	AGTGCAGTT-
Cruz nema tripartitum_DF5015	CGAAACCCCG	AC	-----	-----	-TTCCGGAAGG	GGTGCAATT-
Mesorhabditis_sp_PS1179	GCACTTC	---	-----	-----	-----G	GTGTCATTT-
Mesorhabditis spiculigera_SB15	GTGCTTC	---	-----	-----	-----G	GCACCATTT-
Mesorhabditis anisomorpha_SB12	CTTCGGY	---	-----	-----	-----	--GTACATC-
Teratorhabditis palmarum_DF501	GCGTGCTTCG	G	-----	-----	-----	-CACCACTT-
Teratorhabditis synpapillata_S	TCGTGCTTCG	G	-----	-----	-----	-CACCACTT-
Caenorhabditis briggsae_PB102	CTAAACCCCA	ACG	-----	-----	--TAAGGCGG	GGTGCAATT-
Caenorhabditis elegans_N2	CTATACCCCA	ACG	-----	-----	--CAAGGCGG	GGTGCAATT-
Caenorhabditis_sp_CB5161	CTATACCCCA	ACG	-----	-----	--CAAGGCGG	GGTGCAATT-
Caenorhabditis_sp_PS1010	TTAAACCTCG	ACG	-----	-----	--CAAGGACG	GGTGCAATT-
Caenorhabditis japonica	CTATACTCCA	ACG	-----	-----	--CAAGGCGG	GGTGCAATT-
Caenorhabditis_sp_DF5170	TTAAACCTCG	ACG	-----	-----	--TAAGGAGA	GGTGCAATT-
Caenorhabditis_sp_SB341	CTAAGCCCTG	ACG	-----	-----	--TAAGGAGG	GGTGCAATT-
Caenorhabditis plicata	AAATTCCTCG	ACG	-----	-----	--CAAGGAGG	GGGGCAATA-
Caenorhabditis vulgaris	CTAAGCCCA	ACG	-----	-----	--TAAGGCGG	GGTGCAATT-
Caenorhabditis sonoreae	CTAAGCCCTG	ACG	-----	-----	--CAAGGAGG	GGTGCAATT-
Caenorhabditis drosophilae	TTAAACCTCA	ACG	-----	-----	--TAAGGCGA	GGTGCAATT-
Phasmarhabditis hermaphrodita	CAATGCTCAG	TTT	-----	-----	---CGGCTG	AGTGCAGTT-
Phasmarhabditis neopapillosa	CAATGCTCAG	TTT	-----	-----	---CGGCTG	AGTGCAGTT-
Phasmarhabditis neopapillosa	TAAAGCCTCG	ACT	-----	-----	TC--GGAAGA	GGTGCAGAT-
Pellioiditis mediterranea_SB173	TAAAGCCTCG	ACT	-----	-----	TC--GGAAGA	GGTGCAGAT-
Pellioiditis marina	TAAAGCCTCG	ACT	-----	-----	TC--GGAAGA	GGTGCAGAT-
Prodontorhabditis wirthi	TGATGTCTCG	CCCGAGC	---	-----	--TAAAGTCA	GCTGCGGTT-
Crustorhabditis scanica	--ACGTCA	---	-----	-----	---YTCTGA	CGT-CATTT-
Dolichorhabditis_sp_CEW1	CAAAGCTAG	GCTC	-G	-----	-----CTTA	GCTGCAGGT-
Oscheius_sp_DF5000	TTAAAGCTAG	G-TTC	---	-----	-----GTCTA	GCTGCAGGT-
Oscheius_sp_BW282	CAAACCTCG	ACTTT	---	-----	-----GAAGA	GGTGCAGAT-
Oscheius myriophila_EM435	CAAACCTCG	ACTTT	---	-----	-----GAAGA	GGTGCAGAT-
Oscheius insectivora	TAAACCCCG	ACTTT	---	-----	-----GAAGG	GGTGCAGAT-
Oscheius dolichuroides	AAAAGCTAG	ATTC	---	-----	-----GTCTA	GCTGCAGGT-
Rhabditis blumi_DF5010	C-ATGCCGTG	TC	TTT-G	-----	-----ACAC	GGTGCATTT-
Rhabditis_sp_PS1191	CGAAACCCCG	ACTTT	---	-----	-----GAAGG	GGTGCAGAT-
Rhabditis_sp_PS1010	TTAAACCTCG	ACGCAAG	---	-----	-----GACG	GGTGCATTT-
Rhabditis myriophila_EM435	CAAACCTCG	ACTTT	---	-----	-----GAAGA	GGTGCAGAT-
Rhabditis colombiana	TTAAAGCCCG	AC	---	-----	TC--ACGAAG	GGTGCAGAT-
Rhabditella axei_DF5006	TAAAGCCCAT	CCTTACG	---	-----	-----GGTG	GGTGCAGAT-
Rhabditella_sp_DF5044	TAAAGCCCAT	CCTTACG	---	-----	-----GGTG	GGTGCAGAT-
Cuticularia_sp_PS2083	CAAAGCTCTG	ACACCTT	---	-----	-----CGG-	AGCGCATTT-
Necator americanus	ATAAACCTCG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Kalicephalus cristatus	TTATACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Ancylostoma caninum	ATAAACCTCG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus cantonensis	ATAAACCTCG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus costaricensis	ATAAACCTCG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus malaysiensis	ATAAACCTCG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus dujardini	TTAAGCCCTG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Angiostrongylus vasorum	TTAAACCTCG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Aulurostrongylus abstrusus	TAGGACCTCG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Didelphostrongylus hayesi	ATAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Crenosoma mephitidis	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Crenosoma vulpis	TTAGACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Otostrongylus circumlitus	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Troglostrongylus wilsoni	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Filaroides martis	TTAAACCTCG	ACT	---	-----	TT--CGAAAG	GGTGCATTT-
Oslerus osleri	TAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Parafilaroides decorus	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Metastrongylus salmi	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Metastrongylus elongatus	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Skrjabingylus chitwoodrum	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Parelaphostrongylus odocoilei	ATAAACCTCG	ACT	---	-----	CT--GTGAAAG	GGTGCATTT-
Muellerius capillaris	TAAATCCTG	ACT	---	-----	TT--TGAAAG	GATGCAATT-
Protostrongylus rufescens	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Halocercus invaginat	TTAGACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Pseudalius inflexus	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Stenurus minor	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
Torynurus convolutus	TTAAACCTCG	ACT	---	-----	TT--TGAAAG	GGTGCATTT-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	250	260	270	280	290	300
Syngamus trachea	GTA AACCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Stephanurus dentatus	ATAAGCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Nematodirus battus	TTAGGCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Strongylus equinus	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Labiostrongylus bipapillosus	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Petrovina poculatum	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Cylicocyclus insignis	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Chabertia ovina	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Cyclodontostomum purvisi	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Zoniolaimus mawsonae	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Hypodontus macropi	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Deletocephalus dimidiatus	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Dictyocaulus eckerti P7B8	TAAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Dictyocaulus capreolus P3B2	TAAAACCCCTA	ACG	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Dictyocaulus sp. P6A1	TAAAACCCCG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Dictyocaulus filaria	AATAACCCCTG	ACG	-----	-----	TT--TTGAAAG	GGTGCAAGTT-
Dictyocaulus capreolus P2C10	TAAAACCCCTA	ACG	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Dictyocaulus viviparus	TAGAACCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Haemonchus sp. V3091	ATAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Haemonchus contortus	ATAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Haemonchus placei	ATAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Haemonchus similis	ATAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Ostertagia ostertagi	TTAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Ostertagia leptospicularis	TTAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Nippostrongylus brasiliensis	TAAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Heligmosomoides polygyrus	GAAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Trichostrongylus colubriformis	ATAAACCCCTG	ACT	-----	-----	TT--CGAAAG	GGTGCAAGTT-
Tetrastrongylus mackerr	TTAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Herpestrongylus pythons	AGAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Nicollina cameroni	AGAAACCCCTG	ACT	-----	-----	TT--TGAAAG	GGTGCAAGTT-
Filarimena flagrifer	TGAAACCCCTG	ACT	-----	-----	CT--TGAAAG	GGTGCAAGTT-
Amidostomum cygni	ATATAACCCCTG	ACT	-----	-----	AT--TGAAAG	GGTGCAAGTT-
Teratocephalus lirellus	CAAAGGTCAG	ACC	-----	-----	TTTCGGGACG	AGCGCATTT-
Brumptaemilius justini	CAAAGTCCG	ATT	-----	-----	TT--TGAAAG	AGCGCATCT-
Anguillicola crassus	CAACGCTCCG	ATT	-----	-----	TAC--TGACG	AGCGCATCT-
Dracunculus medinensis	CAAAGTCCG	GAT	-----	-----	TTTATTGACG	AGCGCATCT-
Dracunculus oesophageus	CAAAGTCTG	AGA	-----	-----	TATATCGAGG	AGCGCATCT-
Dracunculus sp. V3104	CAAAGTCCG	ATT	-----	-----	TTA--TTGACG	AGCGCATCT-
Philonema sp A	CAAAGTCCA	AAT	-----	-----	TTC--TGACG	AGCGCATCT-
Philometra obturans	CAAAGTCCG	ACTAC	-----	-----	AAA--TTT--AGACG	AGCGCATCT-
Camallanus oxycephalus	CAAAGTCTG	AT	-----	-----	TTTT--TGACG	AGCGCATCT-
Acanthocheilonema viteae	CAAAGTCCG	AC	-----	-----	TTAT--GGACG	AGCGCATCT-
Loa loa	CAAAGTCCG	AC	-----	-----	TTTT--GAACG	AGCGCATCT-
Onchocerca cervicalis	CAAAGTCCG	AC	-----	-----	TTTT--GGACG	AGCGCATCT-
Dirofilaria immitis	CAAAGTCTG	AC	-----	-----	TTTTTTGAACG	AGCGCATCT-
Brugia malayi	CAAAGTCCG	AA	-----	-----	TTTT--AAACG	AGCGCATCT-
Wuchereria bancrofti	CAAAGTCCG	AT	-----	-----	TTTT--AGACG	AGCGCATCT-
Litomosoides sigmodontis	CAAAGTCCG	AC	-----	-----	TGTTTGAACG	AGCGCATCT-
Setaria digitata	CAAAGTCCG	AC	-----	-----	TTTT--GGACG	AGCGCATCT-
Gnathostoma turgidum	CAAAGTCCG	AC	-----	-----	TCTG--TGACG	AGCGCATTT-
Gnathostoma neoprocyonis	CAAAGTCCG	AC	-----	-----	TTTG--TGACG	AGCGCATTT-
Gnathostoma binucleatum	CAAAGTCCG	AC	-----	-----	TCTG--TGACG	AGCGCATTT-
Physaloptera alata	CAAAGTCTG	AC	-----	-----	TTC--GGATG	AGCGCATCT-
Physaloptera turgida	CAAAGTCTG	AC	-----	-----	TTT--GGACG	AGCGCATCT-
Ascarophis arctica	CAAAGTCCG	AC	-----	-----	TACTTGACG	AGCGCATCT-
Spinitectus carolini	CAAAGTCCG	AC	-----	-----	TTAGTGGACG	AGCGCATCT-
Thelazia lacrymalis	CAAAGTCAA	AC	-----	-----	TAC--GGATG	AGCGCATCT-
Aphelenchus avenae	TAAAGTCTG	AC	-----	-----	C--TCGGGGGAAAG	AGTGCAATTT-
Aphelenchoides fragariae	ATAAGTACTT	TTCA	-----	-----	G--CAATGAGAAG	TATGCACTTT-
Bursaphelenchus sp	CGAGCTCCG	CC	-----	-----	T--TCGGTTCCG	GGTGCAATTT-
Deladenus sp	ACATGCTCCG	ACC	-----	-----	TTGCGGAAAG	AGTGCAATTT-
Criconema sp	CGATGCTGTC	CC	-----	-----	TCGGGGGGC	AGTGCAATTT-
Hemicycliophora conida	CGAAG--CTG	TC	-----	-----	CCCCAAGGGC	AGTGCAATTT-
Paratylenchus dianthus	CGAAGTTGTT	CC	-----	-----	TCACGGAGC	AATGCAATTT-
Pratylenchus thornei	CAAAGTCCG	AC	-----	-----	CC--GCGAGGGGAG	AGCGCATTT-
Tylenchulus semipentrans	CGAAGTACC	TC	-----	-----	TTGC--GGGGT	AGTGCAATTT-
Ditylenchus angustus	CTAAGCTT	ACC	-----	-----	TTACGGAAAG	AGTGCAATTT-
Subanguina radicola	CAAAGTCTG	ACC	-----	-----	TTACGGAAAG	AGTGCAATTT-
Tylenchorhynchus maximus	CAAAGTCCG	ACC	-----	-----	GTAAGGGGAAAG	AGCGCATTT-
Geocnamus quadriifer	CAAAGTCTG	ACC	-----	-----	TTACGGGAAAG	AGCGCATTT-
Globodera pallida	CAAAGTCCA	ATCCT	-----	-----	CCAGAGCGG	AGCGCATTT-
Scutellonema bradys	CAAAGTCTG	ATCCCTT	-----	-----	GA--GGAGAGG	AGTGCAATTT-
Helicotylenchus dihystra	CAAAGTCCG	ATCCCT	-----	-----	GC--GGAGAGG	AGCGCATTT-
Rotylenchus robustus	CAAAGTCCG	ATCCCTT	-----	-----	GCTGGAGAGG	AGCGCATTT-
Meloidogyne arenaria	TAAAGCTTTG	TCC	-----	-----	TTACGG--AAA	AGCGCATTT-
Meloidogyne incognita KT	TAAAGCTTTG	TCC	-----	-----	TTACGGAAAA	--GCGCATTT-
Meloidogyne javanica	CAAAGCTTTG	TCC	-----	-----	TTACGGWAAA	--GCGCATTT-
Meloidogyne artiellia	CAAAGCTTTG	ACCCTTT	-----	-----	ACGGGGAAAA	AGCGCATTT-
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	250	260	270	280	290	300
Meloidogyne_duytsi	CAAAGCTTTG	TCC	-----	-----	-----	TTCCGGAAA-	AGCGCATTT-
Meloidogyne_exigua	TAAAGCTTTG	TCC	-----	-----	-----	TTACGGAAA-	AGCGCATTT-
Meloidogyne_hapla	TAAAGCTTTG	TCC	-----	-----	-----	TTACGGAAA-	AGCGCATTT-
Meloidogyne_ichinohei	AAAAGCTCCG	ACCC	-----	-----	-----	CTCGGGGAG	AGCGTATTT-
Meloidogyne_maritima	TAAAGCTTTG	TCC	-----	-----	-----	TTACGGAAA-	AGCGCATTT-
Meloidogyne_microtyla	TAAAGCTTTG	TCC	-----	-----	-----	TTACGGAAA-	AGCGCATTT-
Nacobbus_aberrans	AAAAGCTCCG	ACC	-----	-----	-----	TAAAGGAAA	AGCGCATTT-
Pratylenchoides_ritteri	CAAAGCTCCG	ACC	-----	-----	-----	TTACGGGAA	AGCGCATTT-
Pratylenchoides_magnicauda	CAAAGCTCCG	ACC	-----	-----	-----	TTACGGGAA	AGCGCATTT-
Hirschmanniella_sp._JH_2003	CAAAGCTCCG	ACC	-----	-----	-----	GCGAGGGAA	GGCGCATTT-
Radopholus_similis	AAAACCTCCC	AAC	-----	-----	-----	GCAAGGAGG	AGCGCATTT-
Pratylenchus_goodeyi_VF	CAAAGCTCTG	ATCCGCCTT	-----	-----	-----	GCGTGGAAG	AGCGCATTT-
Boleodorus_thylactus_clone2	CAAAGCTCCG	ACC	-----	-----	-----	TCGCGGGAG	AGCGCATTT-
Philippine Sequence 1	ATAAACCTCG	ACT	-----	-----	-----	TT--CGAAG	GGTGCAATT-
Philippine Sequence 2	ATAAACCTCG	ACT	-----	-----	-----	TT--CGAAG	GGTGCAATT-
Philippine Sequence 3	TTTAGCCCCG	ACT	-----	-----	-----	CA--CGAAG	GGTACGGTT-
Philippine Sequence 4	AAAAGTTGAA	TCT	-----	-----	-----	TA-TGGATC	AATGCGTAT-
Philippine Sequence 5	ATAAACCTCG	ACT	-----	-----	-----	TT--CGAAG	GGTGCAATT-
Philippine Sequence 6	CAAAGCCCTG	ACT	-----	-----	-----	TC--GGAAG	GGTGCAATT-
Philippine Sequence 7	TACATCCCCA	ACG	-----	-----	-----	--TAAGCGG	GGTGCAATT-
Tahiti Sequence 1	ATAAACCTCG	ACT	-----	-----	-----	TT--CGAAG	GGTGCAATT-
Tahiti Sequence 2	ATAAACCTCG	ACT	-----	-----	-----	TT--CGAAG	GGTGCAATT-
Thailand Sequence 1	CAAAGCTCCG	AT	-----	-----	-----	-----TTGAG	AGCGCATCT-
Ogasawara Sequence 1	AAAAGTTGAA	ACT	-----	-----	-----	-TATGGTTTC	AATGCGTAT-
Ogasawara Sequence 2	CAATGCTCAG	TTT	-----	-----	-----	----CGGCTG	AGTGCAGTT-
Ivory Coast Sequence 1	TTAAACCTCG	ACT	-----	-----	-----	TT--TGAAG	GGTGCAATT-
Ivory Coast Sequence 2	TTAAACCTCG	ACT	-----	-----	-----	TT--TGAAG	GGTGCAATT-
Ivory Coast Sequence 3	YTAACCTCG	ACT	-----	-----	-----	TT--TGAAG	GGTGCAATT-
Ivory Coast Sequence 4	TTAAACCTCG	ACT	-----	-----	-----	TT--TGAAG	GGTGCAATT-
Ivory Coast Sequence 5	CAAAGCTCCG	AT	-----	-----	-----	----TTGAC	AGCGCATCT-
Ivory Coast Sequence 6	TTAAACCTCG	ACT	-----	-----	-----	TT--TGAAG	GGTGCAATT-
Ivory Coast Sequence 7	TAAACCTCG	ACT	-----	-----	-----	TT--TGAAG	GGTGCAATT-
Singapore Sequence 1	TTTAGCCCCG	ACT	-----	-----	-----	CA--CGAAG	GGTACGGTT-
Singapore Sequence 2	CAAAGCTCCG	AT	-----	-----	-----	----TTGAG	AGCGCATCT-
Singapore Sequence 3	CTAAACCTCG	ACG	-----	-----	-----	--TAAGCGG	GGTGCAATT-
Singapore Sequence 4	TAAACCTCG	ACT	-----	-----	-----	TT--TGAAG	GGTGCAATT-
Nigerian Sequence 1	CTAAACCTCG	ACT	-----	-----	-----	TT--TGAAG	GGTGCAATT-
Nigerian Sequence 2	ATAACCTCG	ACT	-----	-----	-----	TT--CGAAG	GGTGCAATT-
Nigerian Sequence 3	GTAACCTCG	ACT	-----	-----	-----	TA--TGAAG	GGTGCAATT-
NUCLEOTIDES INCLUDED	GTAACCTCG	ACT	-----	-----	-----	TA--TGAAG	GGTGCAATT-

	310	320	330	340	350	360
Gordius_aquaticus	A-TTAGA---	ACAAAAACTA	ACG	-----	-----	-----	-----
Priapulius_caudatus	A-TTAGA---	CC-AAACCAA	TCG	-----	-----	-----GG	TTT-----CG
Brachionus_plicatilis	A-TTAGA---	CCAAAACCAA	TGG	-----	-----	-----GG	T-----CG
Chordodes_morgani	A-TTAGA---	ACAAAAACTA	ACA	-----	-----	-----	-----
Paractinolaimus_macrolaimus	A-TTAGAA--	TAAAAACCAA	TCA	-----	-----	-----GG	TTT-----CG
Aporcelaimellus_obtusicaudatus	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----GG	TTT-----CG
Wilsonema_schuurmansstekhoveni	A-TTAGAA--	CCAAAACCAA	TCG	-----	-----	-----GG	CTT-----CG
Mesodorylaimus_sp_cf_nigritul	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----GG	CCT-----CG
Mesodorylaimus_bastiani	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----GG	TTT-----CG
Mesodorylaimus_japonicus	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----GG	CTT-----CG
Pungentus_sp._PDL_2005	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----GG	TTT-----CG
Allodorylaimus_sp	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----AG	CTT-----CG
Eudorylaimus_carteri	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----AG	CTT-----CG
Microdorylaimus_sp	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----AG	CTT-----CG
Longidorus_elongatus	A-TTAKAA--	TAAAAACCAA	TCG	-----	-----	-----GG	TCT-A---AAA
Xiphinema_rivesi	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----GG	CCCAT---AAA
Tylencholaimus_sp	A-TTAGAA--	TAAAAACCAA	TCG	-----	-----	-----GG	CCT---CGT
Anoplostoma_sp._BHMM_2005	A-TTAGAA--	TAAA-ACCAA	GCA	-----	-----	-----GG	CTT---GCC
Adoncholaimus_fuscus	G-TTACT---	TCAA-ACCAA	TCG	-----	-----	-----GG	CTT---CG
Enoplus_meridionalis	A-TTAGAA--	CCAAAACCAA	TCG	-----	-----	-----GG	CTT---GT
Enoplus_brevis_U88336	A-TTAGAA--	CCAAAACCAA	TCG	-----	-----	-----GG	CTT---G
Enoplus_communis	A-TTAGAA--	CCAAAACCAA	TCG	-----	-----	-----GG	CTT---GT
Enoploides_brunettii	A-TTAGG---	ACAAAACCAA	TCG	-----	-----	-----GG	CTT---GCC
Syngolaimus_striatocaudatus	A-TTAGAA--	ACAAAACCAA	TCG	-----	-----	-----GA	CTT---CGG
Ironus_dentifurcatus	A-TTAGAA--	ACAAAACCAA	TCG	-----	-----	-----AG	CCT---CTG
Calyptonema_maxweberi	G-TTACT---	TCAA-ATCAA	TCG	-----	-----	-----GA	CTT---CG
Viscosia_sp._BHMM_2005	G-TTACA---	ACAA-ACCGA	TCA	-----	-----	-----GA	CCT---CG
Viscosia_viscosa	G-TTACT---	TCAA-ACCAA	TCG	-----	-----	-----GA	CTT---CG
Pontonema_vulgare	GATTACT---	TCAA-ATCAA	TCG	-----	-----	-----GA	CTT---CG
Oncholaimus_sp._BHMM_2005	G-TTACT---	TCAA-ACTGA	TCG	-----	-----	-----GT	CTT---CG
Alaimus_sp._PDL_2005	A-TTAGAA--	TCAAACCAA	TCG	-----	-----	-----GA	CTT---CG
Prismatolaimus_intermedius	A-TTAGAA--	ACAAAACCAA	TCG	-----	-----	-----GG	CCTT---CG
Tobrilus_gracilis	A-TTAGAA--	GCAAAAACCAA	TCG	-----	-----	-----GG	CTT---CG
Tripyla_cf_filicaudata_JH_2004	A-TTAGAA--	CTAAAGCCAA	TCG	-----	-----	-----GA	CCT---TCGG
NUCLEOTIDES INCLUDED	m-mmmmm---	mmmm-mmmmm	mmmm	-----	-----	-----	-----

	310	320	330	340	350	360
Catanema_sp	A-TTAGG---	CCAAAACCAA	TCG-----	-----	GGC CT----	CGG
Eubostrichus_dianae	A-TTAAA---	CCAAAACCAA	TCA-----	-----	GAC TT----	CGG
Eubostrichus_topiarus	A-TTAGA---	CCAAAACCAA	TCA-----	-----	GGC TC----	CGG
Eubostrichus_parasitiferus	A-TTAGA---	CCAAAACCAA	TCA-----	-----	GGC TT----	CGG
Laxus_oneistus	A-TTAGA---	CCAAAATCAA	TCA-----	-----	GGC TT----	TGC
Laxus_cosmopolitus	A-TTAGA---	CCAAAATCAA	TCG-----	-----	GGC TT----	TGC
Leptonemella_sp	A-TTAGA---	CCAAAACCAA	TCG-----	-----	GGC TC----	GTC
Robbea_hypermnestra	A-TTAGA---	CCAAAGACCAA	TCA-----	-----	GGC TC----	TGC
Stilbonema_majum	A-TTAGA---	CCAAAACCAA	TCA-----	-----	GGC TT----	CGG
Monoposthia_costata	A-TTAGA---	TAAAAGGCTG	ACGCG--	-----	GGC TC----	TGC
Nudora_bipapillata	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GGC TT----	GT
Calomicrolaimus_parahonestus	A-TTAGA---	ACTAAAACCAA	TCG-----	-----	GGC TT----	GC
Calomicrolaimus_sp._BHMM_2005	A-TTAGA---	ACTAAAACCAA	TCG-----	-----	GGC TT----	GC
Molgolaimus_demani	A-TTAGA---	ACTAAAACCAA	TCG-----	-----	GGC TT----	GC
Diplogaster_ lethieri	A-TTAGA---	TCAAGACCTA	TTG-----	-----	GGG CA----	AC
Aduncospiculum_halicti	A-TTAGA---	ACAAGACCAA	TCG-----	-----	TGC TT----	GC
Pristionchus_lheritieri	A-TTAGA---	TCAAGACCTA	TTG-----	-----	GGG CA----	AC
Pristionchus_pacificus	A-TTAAA---	TCAAAGCCGA	TTG-----	-----	GGG CA----	AC
Pristionchus_pacificus	A-TTAGA---	TCAAAGCCGA	TTG-----	-----	GGG CA----	AC
Sabatieria_punctata_STRAIN_343	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GGC TT----	TGG
Sabatieria_sp._355_BHMM_2005	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GGC TT----	TGG
Sabatieria_celtica	A-TTAGA---	ACAAAACCAA	TGG-----	-----	GGT T-----	CG
Sabatieria_punctata_STRAIN_200	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GGT TT----	CGG
Sabatieria_punctata_STRAIN_223	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GGC TT----	CGG
Sabatieria_sp._210_BHM_2005	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GGT TT----	CGG
Setosabatieria_hilarula	A-TTAGA---	ACAAAACCAA	TGG-----	-----	GGT T-----	CGC
Desmolaimus_zeelandicus	A-TTAGA---	CCAAAACCAA	TCG-----	-----	GTC TT----	CGG
Terschellingia_longicaudata	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GGG CA----	ACC
Cyartonema_elegans	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GGG CA----	ACC
Tridentulus_sp	A-TTAGA---	ACAAGACCAT	CCG-----	-----	GTC CT----	CGG
Diplolaimelloides_meyli	A-TTAGA---	CCAAAGCCAT	CCG-----	-----	G C TT----	CGG
Diplolaimella_diavengatensis	A-TTAGA---	TCAAGACCA-	TCC-----	-----	GGC TT----	CGG
Geomonhystera_disjuncta	A-TTAGA---	TCAAAGACTA	CCG-----	-----	GCT T-----	CGG
Sphaerolaimus_hirsutus	A-TTAGA---	GCAAAAACCAA	TCA-----	-----	GCC T-----	CGG
Theristus_acer	A-TTAGA---	CTAAAACCTA	TCT-----	-----	GCA CT----	TGT
Daptonema_procerus	A-TTAGC---	TCAAAAACCAA	TCA-----	-----	GCT T-----	CGG
Daptonema_hirsutum	A-TTAGA---	CCAAAACCAA	TCG-----	-----	GCG CA----	AGC
Daptonema_normandicum	A-TTAGA---	ACAAAACCAA	TCG-----	-----	GTG CT----	TGC
Daptonema_oxycerca	A-TTAGC---	TCAAAAACCAA	TCA-----	-----	GCT T-----	CGG
Daptonema_setosum	A-TTAGA---	CCAAAACCAA	TCG-----	-----	GCG C-----	AAG
Desmodora_communis	A-TTAGA---	CCAAGACCAA	TCA-----	-----	GTC TC----	TGA
Desmodora_ovigera	A-TTAGA---	CCAAGACCAA	TCA-----	-----	GAC AT----	TCG
Metadesmolaimus_sp	A-TTAGA---	CCAAAACCAA	TCG-----	-----	GCG CA----	AGC
Dentostomella_sp	A-TTACA---	CTACAACCAA	TCG-----	-----	GGC GT----	CAG
Bunonema_franzi	A-TTAGA---	TCAAAAACCAA	ACG-----	-----	GGC TT----	GCC
Bunonema_sp	A-TTAGA---	ACAAAACCAA	ACG-----	-----	GGC TC----	GCC
Seleborca_complexa	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Acrobeloides_nanus	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Acrobeloides_bodenheimeri	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Acrobeloides_sp_PS1146	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Acrobelus_ciliatus	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Acrobelus_sp_PS1156	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Acrobelus_complexus_WCUG2	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Cephaloboides_sp_SB227	A-TTAGT---	ACAA-ACCAT	GCA-----	-----	G-----	---
Cephalobus_cubaensis	A-TTAGG---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Cephalobus_sp._PS1143	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Cephalobus_sp._PS1196	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Cephalobus_oryzae_PS1165	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Cervidellus_alutus	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Pseudacrobelus_variabilis	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Triligulla_aluta	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Zeldia_punctata	A-TTAGG---	ACAAAACCAA	GCG-----	-----	GCT T-----	CGG
Myolaimus_sp_U81585	A-TTAGA---	CCAAAACCAA	TCG-----	-----	GGC T-----	TGC
Rhabditophanes_sp._KR3021	A-TTGA---	TTAA-ACTA-	-----	-----	-----	---
Brevibucca_sp._SB261	A-TTAGA---	CCGAAAACCAA	CCG-----	-----	GG CTT----	GCC
Halicephalobus_gingivalis	A-TTAGA---	ACAAAACCAA	TGG-----	-----	GC TTT----	GCC
Panagrobelus_stammeri	A-TTAGA---	ACAAAACCAA	GCG-----	-----	GC TTT----	GCC
Plectonchus_sp._PDL0025	A-TTAGT---	ACAAAACCAA	GCG-----	-----	GC TTT----	GCC
Turbatrix_aceti	A-TTAGT---	ACAAAACCAT	CCG-----	-----	AC TT----	CGG
Panagrellus_redivivus	A-TTAGA---	ACAAAACCAA	C-----	-----	TT-----	GTT
Panagrellus_redivivus_PS1163	A-TTAGA---	ACAAAACCAA	C-----	-----	TT-----	GTT
Panagrolaimus_subelongatus	A-TTAGTA--	TTAAAACCAA	GCA-----	-----	TT CTT----	YGG
Panagrolaimus_davidi	A-TTAGTA--	TTAAAACCAA	GCA-----	-----	TT CTT----	CGG
Panagrolaimus_cf_rigidus_AF40	A-TTAGTA--	TTAAAACCAA	GCA-----	-----	TT CTT----	CGG
Panagrolaimus_sp._Sourhope_ED2	A-TTAGTA--	TTAAAACCAA	GCA-----	-----	GT CCT----	TTGG
Panagrolaimus_sp._Sourhope_ED2	A-TTAGTA--	TTAAAACCAA	GCA-----	-----	GT CCT----	TTGG
Panagrolaimus_sp._Sourhope_ED2	A-TTAGTA--	TTAAAACCAA	GCA-----	-----	GT CCT----	TTGG
Panagrolaimus_sp._Sourhope_ED2	A-TTAGTA--	TTAAAACCAA	GCA-----	-----	TT CTT----	CG
NUCLEOTIDES INCLUDED	m-mmmmm--	mmmm-mmmmm	mmmm-----	-----	-----	---

	310	320	330	340	350	360
Panagrolaimus sp. Sourhope ED2	A-TTAGTA--	TTAAAACCAA	GCA-----	-----	-----TT	CTT----CG
Steinernema carpocapsae	A-TTAGA--	TCAAAACCAA	GCG-----	-----	-----AC	TC----CGG
Strongyloides stercoralis	A-TTAGAT--	AAAAAACCAA	TGA-----	-----	-----C	TT----CGG
Strongyloides ratti	A-TTTGA--	TTAA-ACCAT	TT-----	-----	-----T	TT----CGG
Diploscapter sp PS1897	A-TTAGG--	CCAA-ACCAA	TCG-----	-----	-----TC	CT----CGT
Diploscapter sp PS2017	A-TTAGG--	CCAA-ACCAA	TCG-----	-----	-----TC	CT----CGT
Heterorhabditis bacteriophora	A-TTAGA--	ACAG-ACCAA	TCG-----	-----	-----TC	CT----TGG
Heterorhabditis hepialus	A-TTAGA--	ACAG-ACCAA	TCG-----	-----	-----TC	CT----TGG
Heterorhabditis zelandica	A-TTAGA--	ACAG-ACCAA	TCG-----	-----	-----TC	CT----CGG
Parasitorhabditis sp SB281	A-TTAGA--	TTC--ACCAA	CCG-----	-----	-----C	TT----CGG
Rhabditoides inermiformis	A-TTAGA--	ACAAAACCAA	TCG-----	-----	-----GC	TT----CGG
Rhabditoides regina DF5001	A-TTAGA--	CTTATACCAG	CCG-----	-----	-----GG	CA----GCC
Rhabditoides regina DF5012	T-TTAGA--	TTC--ACTAA	CCG-----	-----	-----GG	CAA---CCG
Poikilolaimus oxycerca SB200	A-TTAGA--	ACAAAACCAA	TCG-----	-----	-----GC	TT----TGG
Poikilolaimus regenfussi SB199	A-TTAGA--	ACAAAACCAA	TCG-----	-----	-----GC	TT----CGG
Distolabrellus veechi DWF1604	A-TTAGA--	ATC--ACCAA	CCG-----	-----	-----CT	TC----GG
Distolabrellus veechi DF5024	A-TTAGA--	ATC--ACCAA	CCG-----	-----	-----CT	TC----GG
Choriorhabditis dudichi	A-TTAGT--	ACAG-ACCAA	TCC-----	-----	-----T	TC----AAAA
Protorhabditis sp	A-TTAGG--	CCAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Protorhabditis sp DF5055	A-TTAGC--	GAAG-ACCAA	TCC-----	-----	-----TC	CT----CGG
Cruznema tripartitum DF5015	A-TTAGT--	GCAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Mesorhabditis sp PS1179	A-TTAGA--	TTC--ACCAA	CCG-----	-----	-----C	TT----CGG
Mesorhabditis spiculigera SB15	A-TTAGA--	TTC--ACCAA	CCA-----	-----	-----C	-----CGG
Mesorhabditis anisomorpha SB12	A-TCAGA--	GCAG-ACCAA	CCG-----	-----	-----C	CT----CGG
Teratorhabditis palmarum DF501	A-TTAGA--	-TCG-ACCAA	CCG-----	-----	-----C	TT----AGG
Teratorhabditis synpapillata S	A-TTAGA--	-TCG-ACCAA	CCG-----	-----	-----C	TT----CGG
Caenorhabditis briggsae PB102	A-TTAGA--	ACAG-ACCAA	ACG-----	-----	-----TT	TT----CGG
Caenorhabditis elegans N2	A-TTAGA--	ACAG-ACCAA	ACG-----	-----	-----TT	TT----CGG
Caenorhabditis sp CB5161	A-TTAGA--	ACAG-ACCAA	ACG-----	-----	-----TT	CT----CGG
Caenorhabditis sp PS1010	A-TTAGA--	ACAA-GCCAG	ACG-----	-----	-----TT	CT----CGG
Caenorhabditis japonica	A-TTAGA--	ACAG-ACCAA	ACG-----	-----	-----TT	TT----CGG
Caenorhabditis sp DF5170	A-TTAGA--	ACAG-ATCAA	ACG-----	-----	-----TT	TT----CGG
Caenorhabditis sp SB341	A-TTAGA--	ACAG-ACCAA	CCG-----	-----	-----TC	CT--AACGG
Caenorhabditis plicata	A-TTAGA--	TTAC-ACCAA	CCG-----	-----	-----TC	TT----CGG
Caenorhabditis vulgaris	A-TTAGA--	ACAG-ACCAA	ACG-----	-----	-----TT	TT----CGG
Caenorhabditis sonoreae	A-TTAGA--	ACAG-ACCAA	CCG-----	-----	-----TC	CT----CGG
Caenorhabditis drosophilae	A-TTAGA--	ACAA-ATCAA	ACG-----	-----	-----TT	TT----CGG
Phasmarhabditis hermaphrodita	A-TTAGT--	ACGA-ACCAA	TCG-----	-----	-----CT	TT----CGG
Phasmarhabditis neopapillosa	A-TTAGT--	ACGA-ACCAA	TCG-----	-----	-----CT	CT----CGG
Phasmarhabditis neopapillosa	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TT	TT----CGG
Pellioiditis mediterranea SB173	A-TTAGT--	TCAA-ACCAA	TCG-----	-----	-----TC	C----TGG
Pellioiditis marina	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TT	TT----CGG
Prodontorhabditis wirthi	A-TTAGT--	GAAG-GCCAA	CCG-----	-----	-----TC	TC----TGA
Crustorhabditis scanica	A-TTAGA--	TTC--ACCAA	CCG-----	-----	-----CT	TC----GGC
Dolichorhabditis sp CEW1	A-TTAGT--	GCAA-ACCAA	TCG-----	-----	-----TT	TT----CGG
Oscheius sp DF5000	A-TTATT--	TCAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Oscheius sp BW282	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Oscheius myriophila EM435	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Oscheius insectivora	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Oscheius dolichuroides	A-TTATT--	TCAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Rhabditis blumi DF5010	A-TTAGT--	ACAA-ATCAA	TCG-----	-----	-----CT	TT----CGG
Rhabditis sp PS1191	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Rhabditis sp PS1010	A-TTAGA--	ACAA-GCCAG	ACG-----	-----	-----TT	CT----CGG
Rhabditis myriophila EM435	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Rhabditis colombiana	TATTAGT--	ACAA-ACCAA	TCG-----	-----	-----TC	TT----CGG
Rhabditella axei DF5006	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TC	T----CTG
Rhabditella sp DF5044	A-TTAGT--	ACAA-ACCAA	TCG-----	-----	-----TC	T----CTG
Cuticularia sp PS2083	A-TTAGA--	ACAAAACCAA	TCG-----	-----	-----GC	TT----TGG
Necator americanus	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TC	TT----CGG
Kalicephalus cristatus	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TC	TT----CGG
Ancylostoma caninum	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----CC	TT----CGG
Angiostrongylus cantonensis	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Angiostrongylus costaricensis	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Angiostrongylus malaysiensis	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Angiostrongylus dujardini	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Angiostrongylus vasorum	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Aulurostrongylus abstrusus	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Didelphostrongylus hayesi	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TC	TT----CGG
Crenosoma mephitidis	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Crenosoma vulpis	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Otostrongylus circumlitus	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Troglostrongylus wilsoni	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Filaroides martis	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Oslerus osleri	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TC	TA----CGG
Parafilaroides decorus	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
Metastrongylus salmi	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TA----CGG
Metastrongylus elongatus	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TA----CGG
Skrjabinstrongylus chitwoodrum	A-TTAGA--	GCAA-ATCAA	TCA-----	-----	-----TT	TT----CGG
NUCLEOTIDES INCLUDED	m-mmmmmmm	mmmmmm-mmmmmmm	mmmm-----	-----	-----	-----

	310	320	330	340	350	360
Parelaphostrongylus_odocoilei	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT----CGG
Muellerius_capillaris	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Protostrongylus_rufescens	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Halocercus_invaginatus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT TT----CGG
Pseudalius_inflexus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT TT----CGG
Stenurus_minor	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT TT----CGG
Torynurus_convolutus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TT TT----CGG
Syngamus_trachea	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Stephanurus_dentatus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Nematodirus_battus	A-TTAGA---	GCAA-ATCAA	TCT-----	-----	-----	CC TT----CGG
Strongylus_equinus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Labiostongylus_bipapillosus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Petrovinema_poculatum	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Cylicocyclus_insignis	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Chabartia_ovina	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Cyclodontostomum_purvisi	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Zoniolaimus_mawsonae	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Hypodontus_macropi	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Deletrocephalus_dimidiatus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Dictyocaulus_eckerti_P7B8	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----CGG
Dictyocaulus_capreolus_P3B2	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----CGG
Dictyocaulus_sp._P6A1	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----CGG
Dictyocaulus_filaria	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TCCT TA----CGG
Dictyocaulus_capreolus_P2C10	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----CGG
Dictyocaulus_viviparus	A-TTAGA---	GCAA-ATCAG	TGA-----	-----	-----	TC TT----CGG
Haemonchus_sp._V3091	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CT TT----CGG
Haemonchus_contortus	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CT TT----CGG
Haemonchus_placei	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CT TT----CGG
Haemonchus_similis	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CT TT----CGG
Ostertagia_ostertagi	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Ostertagia_leptospicularis	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Nippostrongylus_brasileinsis	A-TTAGT---	GCAA-ACCAA	TCA-----	-----	-----	TC TT----CGG
Heligmosomoides_polygyrus	A-TTAGA---	GCAA-ACCAA	TCA-----	-----	-----	CC TT----CGG
Trichostrongylus_colubriformis	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CC TT----CGG
Tetraothriostongylus_mackerr	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	TC TT----CGG
Herpestongylus_pythonis	A-TTAGA---	GCAA-ACCAA	TCA-----	-----	-----	CC TT----CGG
Nicollina_cameroni	A-TTAGA---	GCAA-ACCAA	TCA-----	-----	-----	CC TT----CGG
Filarimena_flagrifer	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CC CT----CGG
Amidostomum_cygni	A-TTAGA---	GCAA-ATCAA	TCA-----	-----	-----	CC TT----CGG
Teratocephalus_lirellus	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	GGC TT----CGG
Brumptaemilius_justini	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GGT CT----CGG
Anguillicola_crassus	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	AGC TG----CGG
Dracunculus_medinensis	A-TAAGGA--	TAAAAACCAA	GTC-----	-----	-----	GTAGTCT TAAAA--CGG
Dracunculus_oesophageus	A-TTAGA---	CTAAAACCAA	TCG-----	-----	-----	AGCGGT TTTAAAAC TG
Dracunculus_sp._V3104	A-TTAGA---	CTAAAACCAA	TCG-----	-----	-----	AGCCAA TTTAAA--CGG
Philonema_sp_A	A-TTAGA---	TAAAAACCAA	TCGGGCATAT	AACTTCATT	GTGGAGTGAT	A-----GG
Philometra_obturans	A-TTAGAC--	TAAAAACCAA	TCGAGATGGT	GTCTCTCCGC	GCCATGCGGG	-TGGCATAAC
Camallanus_oxyccephalus	A-TTAGAC--	CAAAAACCAA	TCG-----	-----	-----	AGA TTATTCTGC
Acanthocheilonema_viteae	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GGT TATTGGCTTA
Loa_loa	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GGT TATT----AG
Onchocerca_cervicalis	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GGT TATT----GG
Dirofilaria_immitis	A-TTAGA---	TAAAAACCAA	TCGGAACAAA	-----	-----	TTT GATCT--GA
Brugia_malayi	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GATT ATTAATTTTA
Wuchereria_bancrofti	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GATT ATTAATTAAC
Litomosoides_sigmodontis	A-TTAGA---	TAAAAACCAA	TGG-----	-----	-----	GGTGTC ATCAGCTGCT GCTGCTGGCC
Setaria_digitata	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GGT TATTT--GA
Gnathostoma_turgidum	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	AGC TTC----GG
Gnathostoma_neoprocyonis	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	AGC TTC----GG
Gnathostoma_binucleatum	A-TTAGA---	ACAAAACCAA	TCG-----	-----	-----	AGC TTC----GG
Physaloptera_alata	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GGC TTC----CG
Physaloptera_turgida	A-TTAGA---	TAAAAACCAA	TCG-----	-----	-----	GGC TTC----AA
Ascarophis_arctica	A-TTAGG---	TAAAAACCAA	TCG-----	-----	-----	GGC TT----TG
Spinitectus_carolini	A-TTAGA---	TTAAATCCAA	TCG-----	-----	-----	GTG AAC----TT
Thelazia_lacrymalis	A-TTAGAATG	AAAAAACCAA	TGG-----	-----	-----	GGT TT----ATG
Aphelenchus_avenae	A-TTAGA---	TCAAAACCAA	GCG-----	-----	-----	G-C TT----CG
Aphelenchoides_fragariae	A-TCGGA---	GCAAAACTAA	TCT-----	-----	-----	-----T----
Bursaphelenchus_sp	A-TTGGA---	GCAAAAACCG	TTT-----	-----	-----	G- TA-----
Deladenus_sp	A-TTAGA---	ACAAAACCG	GCG-----	-----	-----	G-C TT----CGG
Criconema_sp	G-TTGGA---	GCAAAAACCG	GCG-----	-----	-----	G-C TT----CGG
Hemicyclophora_conida	A-TTGGA---	GCAAAAACCG	GCG-----	-----	-----	AC TT----CGG
Paratylenchus_dianthus	A-TTTGA---	GCAAAAACCG	GCG-----	-----	-----	G-C TT----CGG
Pratylenchus_thornei	A-TTAGA---	ACAAAACCAA	GCG-----	-----	-----	G-C TT----CGG
Tylenchulus_semipentrans	A-TTGGA---	GCAAAAACCAA	GCG-----	-----	-----	G-C TT----CGG
Ditylenchus_angustus	A-TTAGA---	ACAAAACCG	GCG-----	-----	-----	G-C TT----G
Subanguina_radicola	A-TTAGA---	ACAAAACCAA	GCG-----	-----	-----	AC TT----CGG
Tylenchorhynchus_maximus	A-TTAGA---	ACAAAACCAA	GCG-----	-----	-----	G-C TT----CGG
Geocenamus_quadriker	A-TTCGA---	ACAAAACCAA	GCG-----	-----	-----	G-C TT----CGG
Globodera_pallida	G-TTCGCC--	ACAAAACCAA	GCG-----	-----	-----	C-C TT----CGG
NUCLEOTIDES INCLUDED	m-mmmmm--	mmmm-mmmmm	mmmm-----	-----	-----	-----

	370	380	390	400	410	420
Viscosia viscosa	G--TCTG---	-----	-----	AAATCAAGTG	A--TATTGAA	CAATTTA---
Pontonema vulgare	G--TCTG---	-----	-----	ATGTCAAATG	A--TATTGAA	CAATTTA---
Oncholaimus_sp._BHMM_2005	G--ACTG---	-----	-----	TAATCAAGTG	A--GATTGAA	CTATTTA---
Alaimus_sp._PDL_2005	G--TCCG---	-----	-----	CTCTTTGGTG	A--ATCTGAA	TAACTCA---
Prismatolaimus_intermedius	G--GCCCG---	-----	-----	CCAGTTGGTG	A--ATCTGAA	TAACTTT---
Tobrilus_gracilis	G--CCCG---	-----	-----	TTCTTTGGTG	A--ATCTGAA	TAACTCT---
Tripyla_cf_filicaudata_JH_2004	G--TCCG---	-----	-----	TCCATTGGTG	A--TTCTAAA	TAACTCT---
Bathylaimus_sp	G--CATTGTT	GCG-----	-----	TAATTTGGTG	A--GTCTGAA	TAACTTG---
Bathylaimus_assimilis	G--CATTGTT	GCG-----	-----	TAATTTGGTG	A--GTCTGAA	TAACTTG---
Bathylaimus_sp._BHMM_2005	G--CTGCAGT	GCG-----	-----	TCATTTGGTG	A--GTCTGAA	TAACTTG---
Tripylodes_sp._BHMM_2005	G--TGGTGCG	-----	-----	TTATTTGGTG	A--ATCTGAA	TAACTTG---
Trischiostoma_monohystera	G--CCCG---	-----	-----	TCATTTGGTG	A--ATCTGAA	TAACTTT---
Mermis_nigrescens	G--CTCG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACTCA---
Mylonchulus_arenicolus	G--TGCTTG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACTTA---
Anatonchus_tridentacus	G--CTTG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACTAA---
Mononchus_truncatus	G--CTTG---	-----	-----	ATTGTTGGTG	A--CTCTGAA	TAACTAA---
Prionchulus_muscorum	G--CTTG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACTAA---
Clarkus_sp	G--CTTG---	-----	-----	TGTTTTGGTG	A--CTCTGAA	TAACTAA---
Trichinella_spiralis	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACTATT---
Trichinella_papuae	C--TGATGCT	GAACGTCGAA	A-----	CTCTTTGGTG	A--ATCTGAA	TAACTATT---
Trichinella_britovi	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACTATT---
Trichinella_murrelli	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACTATT---
Trichinella_pseudospiralis	C--TTGTGTT	GAACGTCGAA	A-----	CTCTTTGGTG	A--ATCTGAA	TAACTATT---
Trichinella_nativa	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACTATT---
Trichinella_zimbabwensis	C--TGATGCT	GAACGTCGAA	A-----	CTCTTTGGTG	A--ATCTGAA	TAACTATT---
Trichinella_nelsoni	C--GAATGTT	GAACGTCGAA	A-----	CTGTTTGGTG	A--ATCTGAA	TAACTATT---
Trichuris_suis	G--CTATCTA	TTGGTCTGGG	TCCGCGA---	AAGGTTGGTG	A--ATCGGAA	TAACTAT---
Trichuris_trichiura	G--CTAGCTA	TTGGTCTGAG	TCCGCGA---	AATGTTGGTG	A--ATCGGAA	TAACTAT---
Trichuris_muris	C--TCTTTGC	GTCCTGCCGT	CA-----	ACGTTTGGTG	A--ATCGGAA	TAACTAT---
Tyololaimophorus_minor	G--TCCG---	-----	-----	CTCTTTGGTG	A--ATCTGAA	TAACTTA---
Paratrachodorus_pachydermus	G--ACCG---	-----	-----	CCATTTGGTG	A--ACTCTGAA	TAACTTT---
Paratrachodorus_anemones	G--ACCG---	-----	T-----	CCATTTGGTG	A--CTCTGAA	TAATTTCC---
Trichodorus_primitivus	G--ACCG---	-----	-----	CCATTTGGTG	A--ACTCTGAA	TAACTTA---
Axonolaimus_helgolandicus	---T-----	-----	-----	GTCTTTGGTG	G--ATCTGAA	TAACTCA---
Ascolaimus_elongatus	---T-----	-----	-----	GTCTTTGGTG	G--ATCTGAA	TAACTCA---
Odontophora_rectangula	---CCC-----	-----	-----	GTCTTTGGTG	G--ATCTGAA	TAACTCA---
Cylindrolaimus_sp._202149	---CCT-----	-----	-----	GTGTTTGGTG	G--ATCTGAA	TAACTCA---
Tylocephalus_auriculatus	---CCTG---	-----	A-----	AACGTTGGTG	A--CTCTGAA	TAACTGA---
Plectus_acuminatus_BS9	---CCTG---	-----	-----	AACGGTGGTG	A--CTCTGAA	TAACTGA---
Plectus_aquatilis	---CCTG---	-----	-----	AACGGTGGTG	A--CTCTGAA	TAACTGA---
Anaplectus_sp	---CCTG---	-----	-----	AGGTTTGGTG	A--ATCTGAA	TAACTGA---
Anisakis_sp_WKT	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATA---
Anisakis_sp_Nadler	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATG---
Contraecaeum_multipapillatum	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATA---
Pseudoterranova_decipiens	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATA---
Raphidascaris_acus	---CCCG---	-----	-----	TTTGTGGTG	A--CTCTGAA	TAACTATA---
Terranova_caballeri	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTAAG---
Ascaris_suum	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATA---
Ascaris_lumbricoides	---CCCG---	-----	-----	TAAATTGGTG	A--CTCTGAA	TAACTATA---
Baylisascaris_procyonis	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATA---
Baylisascaris_transfuga	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATA---
Parascaris_equorum	---CCCG---	-----	-----	TAAATTGGTG	A--CTCTGAA	TAACTATA---
Porrocaecum_depressum	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATA---
Toxascaris_leonina	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATT---
Heterocheilus_tunicatus	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATG---
Goezia_pelagia	---CCCG---	-----	-----	TTTGTGGTG	A--CTCTGAA	TAACTATG---
Hysterothylacium_fortalezae	---CCCG---	-----	-----	TTCGTTGGTG	A--CTCTGAA	TAACTGTA---
Hysterothylacium_pelagicum	---CCCG---	-----	-----	TTTGTGGTG	A--CTCTGAA	TAACTATA---
Hysterothylacium_reliquens	---CCCG---	-----	-----	TTTGTGGTG	A--CTCTGAA	TAACTATG---
Iheringascaris_inquies	---CCCG---	-----	-----	TTTGTGGTG	A--CTCTGAA	TAACTATG---
Toxocara_canis	---CCCG---	-----	-----	TAAATTGGTG	A--CTCTGAA	TAACTGTA---
Nemhelix_bakeri	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAGCTATG---
Raillietnema_sp._V3060	---TCCG---	-----	-----	TCATTTGGTG	A--CTCTGAA	TAGCTATC---
Cruzia_americana	---CCCG---	-----	-----	TCAATTGGTG	A--CTCTGAA	TAACTATG---
Heterakis_sp_14690	---CCTG---	-----	-----	TTCATTGGTG	A--CTCTGAA	TAGCTCA---
Heterakis_gallarum	---CCTG---	-----	-----	TTTATTGGTG	A--CTCTGAA	TAGCTTA---
Paraspidodera_sp_21303	---CCCG---	-----	-----	TCAACTGGTG	A--CTCTGAA	TAGCTATA---
Chromadora_nudicapitata	C---CA---	-----	-----	TCCATTGGTG	A--ATCTCC	GAAGAAA---
Chromadora_sp._BHMM_2005	C---CA---	-----	-----	TCTATTGGTG	A--ATCTGAG	CAACTTA---
Atrochromadora_microlaima	C---CA---	-----	-----	TCTATTGGTG	A--ATCTGAG	CAACTTA---
Chromadorina_germanica	C---CA---	-----	-----	TCTATTGGTG	A--ATCTGAA	TAACTAC---
Chromadorita_tentabundum	C---CA---	-----	-----	TGAATTGGTG	A--ATCTGAA	TAACTAC---
Dichromadora_sp._BHMM_2005	C---CA---	-----	-----	TCTATTGGTG	A--ATCTGAG	CAACTTA---
Neochromadora_BHMM_2005	C---CA---	-----	-----	TAAATTGGTG	A--ATCTGAA	TAACTAC---
Spilophorella_paradoxa	C---CA---	-----	-----	TTCTTTGGTG	A--ATCTGAA	TAACTAC---
Paracanthonchus_caecus	T---CCG---	-----	-----	TAGTTTGGTG	A--ATCTGAA	TAACTCA---
Paracyatholaimus_intermedius	C---CCG---	-----	-----	TCGTTTGGTG	A--ATCTGAA	TAACTCA---
Praeacanthonchus_punctatus	C---CCG---	-----	-----	TCGTTTGGTG	A--ATCTGAA	TAACTCA---
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----

	370	380	390	400	410	420	
Panagrolaimus_subelongatus	--AATG-----	-----	-----	-----	TTTTGTGTTG	A--AKCTGAA	TATATCAT--	
Panagrolaimus_davidi	G-ATG-----	-----	-----	-----	TTTTGTGTTG	A--AACTGAA	TATATCAT--	
Panagrolaimus_cf_rigidus_AF40	G-ATG-----	-----	-----	-----	TTTTGTGTTG	A--ACCTGAA	TATATCT---	
Panagrolaimus_sp_Sourhope_ED2	G-ATTG-----	-----	-----	-----	TTTAGTGTG	A--ATCTGAA	TATATCT---	
Panagrolaimus_sp_Sourhope_ED2	G-ATTG-----	-----	-----	-----	TTTAGTGTG	A--ATCTGAA	TATATCT---	
Panagrolaimus_sp_Sourhope_ED2	G-AATG-----	-----	-----	-----	TTTAGTGTG	A--ACCTGAA	TATATCT---	
Panagrolaimus_sp_Sourhope_ED2	G-AATG-----	-----	-----	-----	TTTAGTGTG	A--ACCTGAA	TATATCT---	
Steinernema_carpocapsae	--TCG-----	-----	-----	TT	TATGTTGGTG	A--CTCTAG	TAACFTT---	
Strongyloides_stercoralis	-----	-----	-----	G	CTCCTGGTG	A--TTCATAA	TAACFTC---	
Strongyloides_ratti	-----	-----	-----	-----	-----	TTG	A--CTCAAAA	TATCCTC---
Diploscapter_sp_PS1897	-GACG-----	-----	-----	-----	TATTGTGGCG	A--CTCCAAA	TAGCCCA---	
Diploscapter_sp_PS2017	-GACG-----	-----	-----	-----	TATTGTGGCG	A--CTCCAAA	TAGCCCA---	
Heterorhabditis_bacteriophora	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Heterorhabditis_hepialus	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Heterorhabditis_zelandica	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Parasitorhabditis_sp_SB281	--CG-----	-----	-----	-----	TAAAGTGATG	A--ATCTGAA	TATCTT---	
Rhabditoides_inermiformis	--TCG-----	-----	-----	T	TTGTTTGTG	A--CTCTGAA	TAACFTA---	
Rhabditoides_inermis_DF5001	--GAT-----	-----	-----	-----	---ACTGGTG	A--CTCTGAA	TAACCCA---	
Rhabditoides_regina_DF5012	-----	-----	-----	-----	TTGAGTGATG	A--CTCTGAT	TATCTTG---	
Poikilolaimus_oxycerca_SB200	--CCG-----	-----	-----	-----	TTTGTGTTG	A--CTCTGAA	TAACFTA---	
Poikilolaimus_regenfussi_SB199	--CCG-----	-----	-----	-----	TTTGTGTTG	A--CTCTGAA	TAACFTA---	
Distolabrellus_veechi_DWF1604	--C-G-----	-----	-----	-----	TGTAGTGATG	A--CTCTGAA	TATCTC---	
Distolabrellus_veechi_DF5024	--C-G-----	-----	-----	-----	TATAGTGATG	A--CTCTGAA	TATCTC---	
Choriorhabditis_dudichi	--GAG-----	-----	-----	G	ACGTTTGTG	A--CTCTGAA	TAACFTA---	
Protorhabditis_sp	--ACG-----	-----	-----	-----	TATTGTGGCG	A--CTCCAAA	TAGCCCA---	
Protorhabditis_sp_DF5055	--AGG-----	-----	-----	-----	TTTTGTGTTG	A--ATCTGAG	TAAGTTT---	
Cruzinema_tripartitum_DF5015	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAG	TAATGCA---	
Mesorhabditis_sp_PS1179	--CG-----	-----	-----	-----	TCAAAGTGATG	A--TTCTGAA	TATCTT---	
Mesorhabditis_spiculigera_SB15	--CAACG----	-----	-----	GTTGT	TTCGGTGATG	A--TTCTGAA	TAACFTA---	
Mesorhabditis_anisomorpha_SB12	--CG-----	-----	-----	-----	TTGAGTGATG	A--TTCTGAA	TGCTTT---	
Teratorhabditis_palmarum_DF501	--CG-----	-----	-----	-----	TTTTGTGTTG	A--CTCTGAA	TATTTT---	
Teratorhabditis_synpallata_S	--CG-----	-----	-----	-----	TTTTGTGTTG	A--CTCTGAA	TATTTT---	
Caenorhabditis_briggsae_PB102	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TAAAGCA---	
Caenorhabditis_elegans_N2	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TAAAGCA---	
Caenorhabditis_sp_CB5161	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TAAAGCA---	
Caenorhabditis_sp_PS1010	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAAAGCA---	
Caenorhabditis_japonica	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TACCCTC---	
Caenorhabditis_sp_DF5170	--ACG-----	-----	-----	-----	TAATTTGTG	A--CTCTGAA	TAAAGCA---	
Caenorhabditis_sp_SB341	--ACG-----	-----	-----	-----	TTGACTGTTG	A--CTCTGAA	TAATGCA---	
Caenorhabditis_plicata	--ACG-----	-----	-----	-----	TTTTGTGTTG	A--CTCTGAA	CAATCCC---	
Caenorhabditis_vulgaris	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TAAAGCA---	
Caenorhabditis_sonorae	--ACG-----	-----	-----	-----	TTGACTGTTG	A--CTCTGAA	TAAAGCA---	
Caenorhabditis_drosophilae	--ACG-----	-----	-----	-----	TAATTTGTG	A--CTCTGAA	TAAAGCA---	
Phasmarhabditis_hermaphrodita	--GCG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TACCFTAG---	
Phasmarhabditis_neopapillosa	--GCG-----	-----	-----	-----	TAGTTTGTG	A--ATCTAAA	TACCFTAG---	
Phasmarhabditis_neopapillosa	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TACCFTA---	
Pellioditis_mediterranea_SB173	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TACCFTA---	
Pedlioditis_marina	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TACCFTA---	
Prodontorhabditis_wirthi	--CGG-----	-----	-----	-----	ACTGTG-TTG	A--CACTGAA	TAACGCTC---	
Crustorhabditis_scanica	--GTW-----	-----	-----	-----	AAGTGATGAC	T--CTGGATA	TCCTGYA---	
Dolichorhabditis_sp_CEW1	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TACC-CA---	
Oscheius_sp_DF5000	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TACCAG---	
Oscheius_sp_BW282	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TATCGCA---	
Oscheius_myriophila_EM435	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TATCGCA---	
Oscheius_insectivora	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TAACFTA---	
Oscheius_dolichuroides	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TACCCA---	
Rhabditis_blumi_DF5010	--GCG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TAACFTA---	
Rhabditis_sp_PS1191	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACFTA---	
Rhabditis_sp_PS1010	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAAAGCA---	
Rhabditis_myriophila_EM435	--ACG-----	-----	-----	-----	TTGTTTGTG	A--CTCTGAA	TATCGCA---	
Rhabditis_colombiana	--ACG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCT---	
Rhabditella_axei_DF5006	--ACG-----	-----	-----	-----	AAGTTTGTG	A--CTCTGAA	TATCGTA---	
Rhabditella_sp_DF5044	--ACG-----	-----	-----	-----	AAGTTTGTG	A--CTCTGAA	TATCGTA---	
Cuticularia_sp_PS2083	--CCG-----	-----	-----	-----	TTTGTGTTG	A--CTCTGAA	TAACFTA---	
Necator_americanus	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTAAA	TAACGCT---	
Kalicephalus_cristatus	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCT---	
Ancylostoma_caninum	--GTG-----	-----	-----	-----	TAGTTTGTG	A--CTCTAAA	TAACGCT---	
Angiostrongylus_cantonensis	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCT---	
Angiostrongylus_costaricensis	--ATG-----	-----	-----	-----	CAGTTTGTG	A--CTCTGAA	TAACGCA---	
Angiostrongylus_malaysiensis	--ATG-----	-----	-----	-----	CAGTTTGTG	A--CTCTGAA	TAACGCA---	
Angiostrongylus_dujardini	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Angiostrongylus_vasorum	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Aulurostrongylus_abstrusus	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Didelphostrongylus_hayesi	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Crenosoma_mephitidis	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Crenosoma_vulpis	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
Otostrongylus_circumlitus	--ATG-----	-----	-----	-----	TAGTTTGTG	A--CTCTGAA	TAACGCA---	
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	-----	-----	-----	

	370	380	390	400	410	420
Troglostrongylus_wilsoni	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Filaroides_martis	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Oslerus_osleri	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Parafilaroides_decorus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Metastrongylus_salmi	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Metastrongylus_elongatus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Skrjabinstrongylus_chitwoodrum	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Parelaphostrongylus_odocoilei	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Muellerius_capillaris	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Protostrongylus_rufescens	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Halocercus_invaginatus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Pseudalius_inflexus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Stenurus_minor	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Torynurus_convolutus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Syngamus_trachea	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTGAA	TAACGCC---
Stephanurus_dentatus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Nematodirus_battus	--GTG-----	-----	-----	CTGTTTGCTG	A--CTCTGAA	TAATGCA---
Strongylus_equinus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Labiostongylus_bipapillosus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Petrovinema_poculatum	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Cylicocycclus_insignis	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Chabartia_ovina	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Cyclodontostomum_purvisi	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Zoniolaimus_mawsonae	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Hypodontus_macropi	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Deletrocephalus_dimidiatus	--ATG-----	-----	-----	TAGTTTGCTG	A--CTCTAAA	TAACGCT---
Dictyocaulus_eckerti_P7B8	--ATG-----	-----	-----TA	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Dictyocaulus_capreolus_P3B2	--ATG-----	-----	-----TA	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Dictyocaulus_sp._P6A1	--ATG-----	-----	-----TA	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Dictyocaulus_filaria	--ATG-----	-----	-----CAA	AAGTTTGCTG	A--CTCTGAA	TAACGCA---
Dictyocaulus_capreolus_P2C10	--ATG-----	-----	-----TA	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Dictyocaulus_viviparus	--ATG-----	-----	-----TA	TAGTTTGCTG	A--CTCTGAA	TAACGCA---
Haemonchus_sp._V3091	--GTG-----	-----	-----	CAGTTTGCTG	A--CTCTGAA	TAACGCA---
Haemonchus_contortus	--GTG-----	-----	-----	CAGTTTGCTG	A--CTCTGAA	TAACGCA---
Haemonchus_placei	--GTG-----	-----	-----	CAGTTTGCTG	A--CTCTGAA	TAACGCA---
Haemonchus_similis	--GTG-----	-----	-----	CAGTTTGCTG	A--CTCTGAA	TAACGCA---
Ostertagia_ostertagi	--ATG-----	-----	-----	AAGTTTGCTG	A--CTCTGAA	TAACGCA---
Ostertagia_leptospicularis	--ATG-----	-----	-----	AAGTTTGCTG	A--CTCTGAA	TAACGCA---
Nippostrongylus_brasileinsis	--ATG-----	-----	-----	TTGTTTGCTG	A--CTCTGAA	TAACGCA---
Heligmosomoides_polygyrus	--ATG-----	-----	-----	CTGTTTGCTG	A--CTCTGAA	TAACGCA---
Trichostrongylus_colubriformis	--GTG-----	-----	-----	CTGTTTGCTG	A--CTCTGAA	TAACGCA---
Tetraothriostongylus_mackerr	--GTG-----	-----	-----	TTGTTTGCTG	A--CTCTGAA	TAACGCA---
Herpestrongylus_pythonis	--GTG-----	-----	-----	CTGTTTGCTG	A--CTCTGAA	TAACGCA---
Nicollina_cameroni	--GTG-----	-----	-----	CTGTTTGCTG	A--CTCTGAA	TAACGCA---
Filarimena_flagrifer	--GTG-----	-----	-----	CTGTTTGCTG	A--CTCTGAA	TAACGCA---
Amidostomum_cygni	--GTG-----	-----	-----	CTGTTTGCTG	A--CTCTGAA	TAACGCA---
Teratocephalus_lirellus	--CTC-----	-----	-----	GCCGTTGGTG	A--CTCTGAA	TAACCTCA---
Brumptaemilius_justini	C-CCG-----	-----	-----	TCTCTGGTG	A--CTCTGAA	TAGCTTCA---
Anguillicolu_crassus	C-TCG-----	-----	-----	TAAGTTGGTG	A--CTCTGAA	TAACCTAT---
Dracunculus_medinensis	C-TCGT-----	-----	-----	CTAATTGGTG	A--CTCTGAA	TAGCTTA---
Dracunculus_oesophageus	T-TCG-----	-----	-----	TTAATTGGTG	A--CTCTGAA	TAGCTTA---
Dracunculus_sp._V3104	C-TCG-----	-----	-----	TTAATTGGTG	A--CTCTGAA	TAGCTTA---
Philonema_sp_A	C-TCG-----	-----	-----	TCAATTGGTG	A--CTCTGAA	TAGCTCA---
Philonema_obturans	TTCTCG-----	-----	-----T	CAATTGGTG	A--CTCTGAA	TAGCTTA---
Camallanus_oxyccephalus	ATTCTGTGCTC	GAAC-----	---TTCTCGT	AACCTTGGTG	A--CTCTGAA	TAGCTTA---
Acanthocheilonema_viteae	ACCCG-----	-----	-----	TAAATTGGTG	A--CTCTGAA	TAGCTATG---
Loa_loa	C-CCG-----	-----	-----	TTAATTGGTG	A--CTCTGAA	TAGCTATG---
Onchocerca_cervicalis	C-CCG-----	-----	-----	TTAATTGGTG	A--CTCTGAA	TAGCTATG---
Dirofilaria_immitis	T-CCG-----	-----	-----T	TAATTGGTG	A--CTCTAAA	TAGCTATA---
Brugia_malayi	ATCCG-----	-----	-----	TTAATTGGTG	A--CTCTGAA	TAGCTATG---
Wuchereria_bancrofti	--CCG-----	-----	-----	TTAATTGGTG	A--CTCTGAA	TAGCTATG---
Litomosoides_sigmodontis	ACCCG-----	-----	-----TT	CAGATTGGTG	A--CTCTGAA	TAGCTATT---
Setaria_digitata	C-CCG-----	-----	-----	TTAATTGGTG	A--CTCTGAA	TAGCTATG---
Gnathostoma_turgidum	C-TCG-----	-----	-----	GTTGTTGGTG	A--CTCTGAA	TAACCTACG---
Gnathostoma_neoprocyonis	C-TCG-----	-----	-----	GTTGTTGGTG	A--CTCTGAA	TAACCTACG---
Gnathostoma_binucleatum	C-TCG-----	-----	-----	GTTGTTGGTG	A--CTCTGAA	TAACCTACG---
Physaloptera_alata	C-CCG-----	-----	-----	TCACTTGGTG	A--CTCTGAA	TAGCTATG---
Physaloptera_turgida	C-CCG-----	-----	-----	TCATTGGTG	A--CTCTGAA	TAGCTATG---
Ascarophis_arctica	C-TCG-----	-----	-----	TCAATTGGTG	A--CTCTGAA	TAGCTATG---
Spinitectus_carolini	AACGGTTTGC	CG-----	-----	TTAATTGGTG	A--CTCTGAA	TAGCTCA---
Thelazia_lacrymalis	CAAATAAGCT	CGCAAAAAA-	-----	AAAATTGGTG	A--ACTGAA	TAGCTATA---
Aphelenchus_avenae	G-CCG-----	-----	-----	TCTACCCTG	A--CTTAAAG	TAACCTAA---
Aphelenchoides_fragariae	-----	-----	-----	GTTTATGCTA	A--CTCTGGG	TAACCT---
Bursaphelenchus_sp	-----	-----	-----	CTGTTTGCTA	A--CT-TGGA	TAATTT---
Deladenus_sp	--TCG-----	-----	-----	TTTTTTGTTG	A--CTCAGAA	TAACCTTA---
Criconema_sp	--CCG-----	-----	-----	TTTTTTGCTG	A--CTCAGAA	CAACCTTA---
Hemicycliophora_conida	--TCG-----	-----	-----	TTCTTGCTG	A--CTCAGAA	TAACCTAA---
Paratylenchus_dianthus	--CCG-----	-----	-----	TTTTCTGTTG	A--TTCAGAA	TAACCTAA---
NUCLEOTIDES INCLUDED	-----	-----	-----	-----	m-----	-----

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370      380      390      400      410      420
Pratylenchus_thornei      --CCG-----TCCATTGTTG A--CTCAGAA TAACTAA---
Tylenchulus_semipentrans --CCG----- -TTTTTGCTG A--CTCAGAA TAACTAA---
Ditylenchus_angustus     --CCG----- TGTTTTGTG A--CTCAAA TAACTCA---
Subanguina_radicola      --TCG----- TTTTTTGTG A--CTCAGAA TAACTAA---
Tylenchorhynchus_maximus --CCG----- TCCAATGTTG A--CTCAGAA TAACTAA---
Geocenamus_quadrifer     --CCG----- TCCGCAGTTG A--CTCAGAA TAACTAA---
Globodera_pallida        --GCG----- TCCAGTGTG A--CTCAGAA CAACTAA---
Scutellonema_bradys      --TCG----- TTCAGTGTG A--ACCAGAA CAACTAA---
Helicotylenchus_dihystera --GCG----- TCCATTGCTG A--ACCAGAA CAACTCA---
Rotylenchus_robustus     --GCG----- TTCAGTGTG A--CTCAGAA TAACTAA---
Meloidogyne_arenaria     --CTG----- CTTCTTGTG A--CTCAGAA TAACTTA---
Meloidogyne_incognita_KT --CTG----- CTTCTTGTG A--CTCAGAA TAACTTA---
Meloidogyne_javanica     --CTG----- CTTYTTGTG A--CTCAGAA TAACTWA---
Meloidogyne_artiellia    --CTGTCAA--- ACGTTTGTG A--CTCAGAA TAACTAA---
Meloidogyne_duytsi       --CTG----- CTTCTTGTG A--CTCAGAA TAACTAA---
Meloidogyne_exigua       --CTG----- CCTTTTGTG A--CTCAGAA TAACTAA---
Meloidogyne_hapla        --CTG----- CTTCTTGTG A--CTCAGAA TAACTAA---
Meloidogyne_ichinohei    --CTG----- TTCATTGTTG A--CTCAGAA TAACTCA---
Meloidogyne_maritima     --CTG----- CTTCTTGTG A--CTCAGAA TAACTAA---
Meloidogyne_microtyla    --CTG----- CTTCTTGTG A--CTCAGAA TAACTAA---
Nacobbus_aberrans        --CCG----- TCCACTGTTG A--ACCAGAA TAACTAA---
Pratylenchoides_ritteri  --CCG----- TTCGCAGTTG A--CTCAGAA TAACTAA---
Pratylenchoides_magnicauda --CCG----- TTCGCAGTTG A--CTCAGAA TAACTAA---
Hirschmanniella_sp._JH_2003 --CCG----- TCCATGAGTTG A--CTCAGAA TAACTAA---
Radopholus_similis       --GGG----- TTGAGTGTG A--ATCAGAG CAACTAA---
Pratylenchus_goodeyi_VF  --CCG----- TCCATTGTTG A--CTCAGAA TAACTAA---
Boleodorus_thylactus_clone2 A-CGG-TGCT CG----- TGATTGTG A--CTCAGAA TAACTAA---
Philippine Sequence 1    --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Philippine Sequence 2    --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Philippine Sequence 3    --ACG----- -CGGCTGTTG A--CTCTGAA TAACGCT---
Philippine Sequence 4    --ATG----- TTTTGTGTTG A--AACTGAA TATATCT---
Philippine Sequence 5    --GTG----- TAGTTGCTG A--CTCTAAA TAACGCT---
Philippine Sequence 6    --ACG----- TTGTTTGTG A--CTCTGAA TATCGTA---
Philippine Sequence 7    --ACG----- TTGTTTGTG A--CTCTGAA TAAAGCA---
Tahiti Sequence 1        --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Tahiti Sequence 2        --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Thailand Sequence 1      ---CCTG--- TTCATTGGT A--CTCTGAA TAGCTTA---
Ogasawara Sequence 1    --ATG----- TTTAGTGTG A--AACTGAA TATATCT---
Ogasawara Sequence 2    --GCG----- TTGTTTGTG A--CTCTGAA TACCAG---
Ivory Coast Sequence 1  --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Ivory Coast Sequence 2  --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Ivory Coast Sequence 3  --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Ivory Coast Sequence 4  --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Ivory Coast Sequence 5  ---CCTG--- TTCATTGGT A--CTCTGAA TAGCTTA---
Ivory Coast Sequence 6  --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Ivory Coast Sequence 7  --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Singapore Sequence 1    --ACG----- -CGGCTGTTG A--CTCTGAA TAACGCT---
Singapore Sequence 2    ---CCTG--- TTCATTGGT A--CTCTGAA TAGCTTA---
Singapore Sequence 3    --ACG----- TTGTTTGTG A--CTCTGAA TAAAGCA---
Singapore Sequence 4    --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Singapore Sequence 1    --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
Nigerian Sequence 2     --GTG----- TAGTTGCTG A--CTCTAAA TAACGCT---
Nigerian Sequence 3     --ATG----- TAGTTGCTG A--CTCTGAA TAACGCA---
NUCLEOTIDES INCLUDED

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430      440      450      460      470      480
Gordius_aquaticus        GCCGATCGCA --CGGTCTC -GT-ACCGGC GACGTATCT- TTC--AAATG TCTGC-CTTA
Priapulus_caudatus      GCTGATCGCA --CGGTCTC -GC-ACCGGC GACGTATCT- TTC--AAATG TCTGC-CTTA
Brachionus_plicatilis   GCTGATCGCA --TGGCCTA -GA-GCCGGC GACGTATCT- TTC--AAGTG TCTGC-CCTA
Chordodes_morgani       GCCGATCGCA --TGGTCTC -GT-ACCGGC GACGTATCT- TTC--AAATG TCTGC-CTTA
Paractinolaimus_macrolaimus GCCAATCGCA --CAGTCCTA -GT-ACTGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Aporcelaimellus_obtusicaudatus GCCAATCGCA --CAGTCCTA -GT-ACTGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Wilsonema_schuurmansstekhoveni GCTGATCGCA --TGGTCTTT -GT-ACCGGC GACGTATCT- TTC--AAGTG TCTGC-CTTA
Mesodorylaimus_sp_cf_nigritul GCCAATCGCA --CAGTCCTA -GC-ACTGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Mesodorylaimus_bastiani GCCAATCGCA --CAGTCCTA -GT-ACTGGC GATGTATCT- TTC--AAATG TCTGC-CTTA
Mesodorylaimus_japonicus GCCAATCGCA --CAGTCCTA -GT-ACTGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Pungentus_sp._PDL_2005  GCCAATCGCA --CAGTCCTA -GT-ACTGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Allodorylaimus_sp       GCCAATCGCA --CAGTCCTA -GT-ACTGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Eudorylaimus_carteri    GCCAATCGCA --CAGTCCTA -GT-ACTGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Microdorylaimus_sp      GCCAATCGCA --CAGTCCTA -GT-ACTGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Longidorus_elongatus    GCTGATCGCA --CGGTCTA -GT-ACCGGC GAMGTATCT- TTC--AAGTG TCTGC-CTTA
Xiphinema_rivesi       GCCGATCGCA --CGGTCTA -GT-ACCGGC GACGTATCT- TTC--AAGTG TCTGC-CTTA
Tylencholaimus_sp      GCCAATCGCA --CGGTCTA -GT-ACCGGC GATGTATCT- TTC--AAGTG TCTGC-CTTA
Anoplostoma_sp._BHMM_2005 G-TGATCGCA --CGGTCTT -GT-ACCGGC GACATATCC- TTC--AAATG TCTGC-CTTA
Adoncholaimus_fuscus   GCTGATCGCA --CGGTCTAT -GC-ACCGGC GACATATCT- TTC--AAACG TCTGC-CTTA
Enoplus_meridionalis   GTAGATCGCA --CGGTCTC -GA-ACCGGC GACTCGTCA- TTC--AAATG TCTGC-CTTA
NUCLEOTIDES INCLUDED

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	430	440	450	460	470	480
<i>Enoplus brevis</i> U88336	GCAGATCGCA	--CGGTCTC	-GC-ACCGGC	GACATGTCA	TTC--AAATG	TCTGC-CTTA
<i>Enoplus communis</i>	GCAGATCGCA	--CGGTCTC	-GC-ACCGGC	GACATGTCA	TTC--AAATG	TCTGC-CTTA
<i>Enoploides brunettii</i>	GCTGATCGCA	--CAGTCTC	-GT-ACCTGG	GATGTATCT	TTC--AAATA	TCTGC-CTTA
<i>Syringolaimus striatocaudatus</i>	GCAGATCGCA	--TGGTCTT	-GC-ACCGGC	GACGTATCT	TTC--AAATG	TCTGC-CTTA
<i>Ironus dentifurcatus</i>	GCCGATCGAA	--CGGTCTT	-GA-ACCGTC	GACGTATCT	TTC--AAATG	TCTGC-CTTA
<i>Calyptronema maxweberi</i>	GCTGAACGAA	--CGGTCTAA	-GA-ACCGAC	GTCATATCC	TTC--AAACG	TCTGC-CTTA
<i>Viscosia sp.</i> BHMM_2005	GCTGATCGCA	--CGGTCCA	-GA-ACCGGC	GACATATCC	TTC--AAATG	TCTGC-CTTA
<i>Viscosia viscosa</i>	GCTGATCGCA	--CGGTCTGA	-GA-ACCGGC	GACATATCC	TTC--AAATG	TCTGC-CTTA
<i>Pontonema vulgare</i>	NNNNAACGCA	--CGGTCCGA	-GT-ATTGGC	GACATATCC	TTC--AAATG	TCTGC-CTTA
<i>Oncholaimus sp.</i> BHMM_2005	GCTGATCGCA	--CGGTCTAA	-GC-ACCGGC	GACATATCC	TTC--AAATG	TCTGC-CTTA
<i>Alaimus sp.</i> PDL_2005	GCCGATCGCA	--GGTCTC	-GA-ACCGGC	GACGTATCT	TTC--AAATG	TCTGC-CTTA
<i>Prismatolaimus intermedius</i>	GCAGATCGCA	--CGGTCTC	-GT-ACCGGC	GACGTATCT	TTC--AAATG	TCTGC-CTTA
<i>Tobrilus gracilis</i>	GCAGATCGCA	--CGGTCTC	-GC-ACCGGC	GACGCATCT	TTC--AAATG	TCTGC-CTTA
<i>Tripyla cf. filicaudata</i> JH_2004	GCCGATCGCA	--TGGTCTC	-GC-ACCGGC	GACGCACCT	TTC--AAATG	TCTGT-CTCA
<i>Bathylaimus sp.</i>	GCAGATCGCA	--CGGTCTTA	-GC-ACCGGC	GACGAATCT	TTC--AAATG	TCTGC-CTTA
<i>Bathylaimus assimilis</i>	GCAGATCGCA	--CGGTCTA	-GC-ACCGGC	GACGAATCT	TTC--AAATG	TCTGC-CTTA
<i>Bathylaimus sp.</i> BHMM_2005	GCAGATCGCA	--CGGTCTA	-GT-ACCGGC	GACGCATCT	TTC--AAATG	TCTGC-CTTA
<i>Tripylodes sp.</i> BHMM_2005	GCAGATCGCA	--TGGTCTA	-GT-ACCGGC	GACGCATCT	TTC--AAATG	TCTGC-CTTA
<i>Trischistoma monohystera</i>	GCCGATCGCA	--TGGTCTAA	-GT-ACCGGC	GACGCATCT	TTC--AAATG	TCTGC-CTTA
<i>Mermis nigrescens</i>	GTTGATCGCA	--CAGTCTC	-GC-ACCTGG	GACGTATCT	TTC--AAATG	TCTGC-CTTA
<i>Mylonchulus arenicolus</i>	GTTGATCGCA	--CAGTCTTA	-GT-ACCTGG	GACGTATCT	TTC--AAATG	TCTGC-CTTA
<i>Anatonchus tridentacus</i>	GTTGATCGCA	--CAGTCTTA	-GT-ACCTGG	GACGTATCT	TTC--AAATG	TCTGC-CTTA
<i>Mononchus truncatus</i>	GTTGATCGCA	--CAGTCTT	-GT-ACCTGG	GACGTATCT	TTC--AAATG	TCTGC-CTTA
<i>Prionchulus muscorum</i>	GTCGATCGCA	--CAGTCTTA	-GT-ACCTGG	GACGTATTT	TTC--AAATG	TCTGC-CTTA
<i>Clarkus sp.</i>	GTCGATCGCA	--CAGTCTTA	-GT-ACCTGG	GACGTATTT	TTC--AAATG	TCTGC-CTTA
<i>Trichinella spiralis</i>	TCTGATCGCA	--TGGTCTA	-GC-ACCGGC	GACATCACT	TTC--AAGTG	TCTGC-CTTA
<i>Trichinella papuae</i>	TCTGATCGCA	--TGGTCTA	-GC-ACCGGC	GACATCACT	TTC--AAGTG	TCTGC-CTTA
<i>Trichinella britovi</i>	TCTGATCGCA	--TGGTCTA	-GC-ACCGGC	GACATCACT	TTC--AAGTG	TCTGC-CTTA
<i>Trichinella murrelli</i>	TCTGATCGCA	--TGGTCTA	-GC-ACCGGC	GACATCACT	TTC--AAGTG	TCTGC-CTTA
<i>Trichinella pseudospiralis</i>	TCTGATCGCA	--TGGTCTA	-GC-ACCGGC	GACATCACT	TTC--AAGTG	TCTGC-CTTA
<i>Trichinella nativa</i>	TCTGATCGCA	--TGGTCTA	-GC-ACCGGC	GACATCACT	TTC--AAGTG	TCTGC-CTTA
<i>Trichinella zimbabwensis</i>	TCTGATCGCA	--TGGTCTA	-GC-ACCGGC	GACATCACT	TTC--AAGTG	TCTGC-CTTA
<i>Trichinella nelsoni</i>	TCTGATCGCA	--TGGTCTA	-GC-ACCGGC	GACATCACT	TTC--AAGTG	TCTGC-CTTA
<i>Trichuris suis</i>	GCTGATCGCA	--CGGTCCA	-GC-ACCGGC	GACGAATCT	TTC--AAATG	ACTTG-CTCA
<i>Trichuris trichiura</i>	GCTGATCGCA	--CGGTCCA	-GT-ACCGGC	GACGATGCT	TTC--AAACG	ACTTG-CTCA
<i>Trichuris muris</i>	GCTGATCGCA	--CGGTCCA	-GC-ACCGGC	GACGAATCT	TTC--AAATG	ACTTG-CTCA
<i>Tylolaimophorus minor</i>	GCTGATCGCA	--CGGTCTC	-GT-ACCGGC	GACAAATCT	TTC--AAATG	TCTGT-TTTA
<i>Paratrichodorus pachydermus</i>	GCTGATCGCA	--CAGTCTC	-GT-ACCTGG	GACAAATCT	TTC--AAGTG	TCTGC-CTTA
<i>Paratrichodorus anemonis</i>	GCTGATCGCA	--CAGTCTC	-GT-ACCTGG	GACAAATCT	TTC--AAGTG	TCTGC-CTTA
<i>Trichodorus primitivus</i>	GCTGATCGCA	--CAGTCTC	-GT-ACCTGG	GACAAATCT	TTC--AAGTG	TCTGC-CTTA
<i>Axonolaimus helgolandicus</i>	GTTGATCGCA	--TGGTCTC	-GC-ACCGGC	GACGTATCT	TTC--AAGTG	TCTGC-CTTA
<i>Ascolaimus elongatus</i>	GTTGATCGCA	--TGGTCTC	-GC-ACCGGC	GACGTATCT	TTC--AAGTG	TCTGC-CTTA
<i>Odontophora rectangula</i>	GCTGATCGCA	--TGGTCTC	-GC-ACCGGC	GACGTATCT	TTC--AAGTG	TCTGC-CTTA
<i>Cylindrolaimus sp.</i> 202149	GCTGATCGCA	--TGGTCTC	-GT-ACCGGC	GACGTATCT	TTC--AAGTG	TCTGC-CTTA
<i>Tylocephalus papiculatus</i>	GCTGATCGCA	--TGGTCTC	-GT-ACCGGC	GACGTATCT	TTC--AAGTG	TCTGC-CTTA
<i>Plectus acuminatus</i> BS9	GCTGATCGCA	--TGGTCTT	-GT-ACCGGC	GACGCATCT	TTC--AAGTG	TCTGC-CTTA
<i>Plectus aquatilis</i>	GCTGATCGCA	--TGGTCTT	-GT-ACCGGC	GACGCATCT	TTC--AAGTG	TCTGC-CTTA
<i>Anaplectus sp.</i>	GCTGATCGCA	--TGGTCTT	-GT-ACCGGC	GACGCATCT	TTC--AAGTG	TCTGC-CTTA
<i>Anisakis sp.</i> WKT	GCTGATCGCA	--TGGTCTT	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Anisakis sp.</i> Nadler	GCTGATCGCA	--TGGTCTT	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Contracaecum multipapillatum</i>	GCTGATCGCA	--TGGTCTT	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Pseudoterranova decipiens</i>	GCTGATCGCA	--TGGTCTT	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Raphidascaris acus</i>	GCTGATCGCA	--TGGTCTA	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Terranova caballeri</i>	GCTGATCGCA	--TGGTCTT	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Ascaris suum</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Ascaris lumbricoides</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Baylisascaris procyonis</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Baylisascaris transfuga</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Parascaris equorum</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Porrocaecum depressum</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Toxascaris leonina</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Heterocheilus tunicatus</i>	GCTGATCGCA	--TGGTCTC	-GC-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Goezia pelagia</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Hysterothylacium fortalezae</i>	GCTGATCGCA	--TGGTCTC	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Hysterothylacium pelagicum</i>	GCTGATCGCA	--TGGTCTA	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Hysterothylacium reliquens</i>	GCTGATCGCA	--TGGTCTA	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Iheringascaris iniquus</i>	GCTGATCGCA	--TGGTCTA	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Toxocara canis</i>	GCTGATCGCA	--TGGTCTA	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Nemhelix bakeri</i>	GCTGATCGCA	--TGGTCTA	-GT-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Raillietnema sp.</i> V3060	GCTGATCGCA	--TGGTCTT	-GT-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Cruzia americana</i>	GCTGATCGCA	--TGGTCTC	-GC-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Heterakis sp.</i> 14690	GCTGATCGCA	--CGGTCTT	-GC-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Heterakis gallinarum</i>	GCTGATCGCA	--TGGTCTT	-GA-ACCGGC	GACGTGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Paraspidodera sp.</i> 21303	GCTGATCGCA	--TGGTCTC	-GC-ACCGGC	GACGYGTCT	ATC--AAGTG	TCTGC-CTTA
<i>Chromodora nudicapitata</i>	GCAGATCGCA	--CGGTCTA	-GT-ACCGGC	GACATATCC	TTC--ATGTG	TCTGC-CTTA
<i>Chromadora sp.</i> BHMM_2005	GCAGATCGCA	--TGGGTCTC	-GT-ACCGGC	GACATATCC	TTC--AAGTG	TCTGC-CTTA
<i>Atrochromadora microlaima</i>	GCAGATCGCA	--TGGGTCTC	-GT-ACCGGC	GACATATCC	TTC--AAGTG	TCTGC-CTTA
<i>Chromadorina germanica</i>	GCTGATCGCA	--CGAGCTC	-GT-CTCGGC	GACATATCC	TTC--AAGTG	TCTGC-CTTA

	430	440	450	460	470	480
Chromadorita_tentabundum	GCTGATCGCA	--CGGTCTC-	-GT-ACCGGC	GACGTATCC-	TTC--AAGTG	TCTGC-CTTA
Dichromadora_sp._BHMM_2005	GCAGATCGCA	--TGGGCTC-	-GT-CCCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA
Neochromadora_BHMM_2005	GCTGATCGCA	--CGGTCTC-	-GT-ACCGGC	GACGTATCC-	TTC--AAGTG	TCTGC-CTTA
Spilophorella_paradoxa	GCTGATCGCA	--TGGGCTC-	-GT-CCCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA
Paracanthonchus_caecus	GTTGATCGCA	--CAGTCTC-	-GC-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Paracyatholaimus_intermedius	GTGGATCGCA	--CAGTCTC-	-GT-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Praeacanthonchus_punctatus	GTTGATCGCA	--CAGTCTC-	-GA-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Praeacanthonchus_sp	GTTGATCGCA	--CAGTCTC-	-GA-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Cyatholaimus_sp._BHMM_2005	GTTGATCGCA	--CAGTCTC-	-GC-ACTGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Spirinia_parasitifera	GCTGATCACA	--CGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Acanthopharynx_micans	GCTGATCACA	--CGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Xyzors_sp	GCTGATCACA	--CGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Metachromadora_sp	GCTGATCACA	--CGGTCTA-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Metachromadora_remanei	GCTGATCACA	--CGGTCTA-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Catanema_sp	GCAGATCACA	--CGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Eubostrichus_dianae	GCTGATCACA	--TGGTCTC-	-GT-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Eubostrichus_topiarus	GCTGATCACA	--TGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Eubostrichus_parasitiferus	GCTGATCACA	--CGGTCTC-	-GT-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Laxus_oneistus	GCTGATCACA	--TGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Laxus_cosmopolitus	GCTGATCACA	--CGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Leptonemella_sp	GCTGATCACA	--CAGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Robbea_hypermnestra	GCTGATCGCA	--TGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Stilbonema_majum	GCTGATCACA	--CGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CCTA
Monoposthia_costata	ACGGATCGCA	--CGGTCTC-	-GT-GCCGGC	GACGATCT-	TTC--AAATT	TCTGC-CCTA
Nudora_bipapillata	GCAGATCGCA	--TGGTCTC-	-GT-ACCGGC	GACACATTC-	CAG--GAGGG	AGTGC-CTTA
Calomicrolaimus_parahonestus	GCAGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATATCT-	TTC--AAGTG	TCTGC-CTTA
Calomicrolaimus_sp._BHMM_2005	GCAGATCGCA	--TGGTCTA-	-GC-ACCGGC	GACATATCT-	TTC--AAGTG	TCTGC-CTTA
Molgolaimus_demani	GCAGATCGCA	--TGGTCTA-	-GT-ACCGGC	GACATATCT-	TTC--AAGTG	TCTGC-CTTA
Diplogaster_lethieri	GCGGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGATCTG-	GTC--GAGCG	GGTGC-CCTA
Aduncospiculum_halicti	GCTGATCGCA	--CGGTCTT-	-GCAACCGGC	GACGTATCT-	TTC--GAGTG	TCTGC-CTTA
Pristionchus_lheritieri	GCGGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGATCTG-	GTC--GAGCG	GGTGC-CCTA
Pristionchus_pacificus	GCGGATCGTA	--TGGTCTT-	-GT-ACCGGC	GACGATCTG-	GTC--GAGTG	GGTGC-CCTA
Pristionchus_pacificus	GCGGATCGTA	--TGGTCTT-	-GT-ACCGGC	GACGATCTG-	GTC--GAGTG	GGTGC-CCTA
Sabatieria_punctata_STRAIN_343	GCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Sabatieria_sp._355_BHMM_2005	GCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Sabatieria_celtica	TCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGATCT-	TTC--AAGTG	TCTGC-CTTA
Sabatieria_punctata_STRAIN_200	GCCGATCGCA	--CGGTCTC-	-GT-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Sabatieria_punctata_STRAIN_223	GCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGATCT-	TTC--AAGTG	TCTGC-CTTA
Sabatieria_sp._210_BHM_2005	GCCGATCGCA	--CGGTCTC-	-GT-ACCGGC	GACGTATCT-	TTC--AAGTG	TCTGC-CTTA
Setosabatieria_hilarula	TCCGATCGCA	--CGGTCTC-	-GC-ACCGGC	GACGATCT-	TTC--AAGTG	TCTGC-CTTA
Desmolaimus_zeelandicus	GCCGATCGCA	--CAGTCTT-	-GT-ACTGGC	GACGGACCA-	TTC--GAGTG	TCTGC-CTTA
Terschellingia_longicaudata	GCTGATCGCA	--CGGTCTT-	-GC-ACCGGC	GACGATCT-	TTC--AAGTG	TCTGC-CTTA
Cyartonea_elegans	GCCGATCGCA	--CGGTCTT-	-GC-ACCGGC	GACGATCT-	TTC--AAGTG	TCTGC-CTTA
Tridentulus_sp	GCAAAATCACA	--TGGTCTC-	-GT-ACCGGT	GATGTATCT-	TTC--AAGTG	TCTGC-CTTA
Diplolaimelloides_meyli	GCAGATCGCA	--TGGGCTC-	-GT-CCCGGC	GACAAATCT-	TTC--AAGTG	TCTGC-CTTA
Diplolaimella_diavengatensis	GCAGATCGCA	--TGGGCTC-	-GT-CCCGGT	GCAAAATCT-	TTC--AAGTG	TCTGC-CTTA
Geomonhystera_disjuncta	GCTGATCGCA	--CGGTCTA-	-GT-GCCGGC	GACATATCT-	TTC--AAGTG	TCTGC-CTTA
Sphaerolaimus_hirsutus	GCGGATCGCA	--CGGTCTT-	-GT-ACCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA
Theristus_acer	GCTGATCGCT	T-CGGTCTC-	-GT-ACCGGC	GACATATCT-	TTC--AAGTG	TCTGC-CTTA
Daptonema_procerus	GCAGATCGCT	--TGGTTTA-	-AT-ACCGGC	GACGTATCC-	TTC--AAGTG	TCTGC-CTTA
Daptonema_hirsutum	GCAGATCGCT	T-CGGTTTC-	-AC-ACCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA
Daptonema_normandicum	GCAGATCGCT	T-CGGTCTC-	-GT-ACCGGC	GATGTATCC-	TTC--AAGTG	TCTGC-CTTA
Daptonema_oxycerca	GCAGATCGCT	T-CGGTTTA-	-AT-ACCGGC	GACGTATCC-	TTC--AAGTG	TCTGC-CTTA
Daptonema_setosum	GCAGATCGCT	T-CGGTTTC-	-AC-ACCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA
Desmodora_communis	GCTGATCACA	--CGGTCTC-	-GC-ACCGGT	GACATATCT-	TTC--AAGTG	TCTGC-CTTA
Desmodora_ovigera	GCTGATCACA	--CAGTCTA-	-GC-ACTGGT	GACATATCT-	TTC--AAGTG	TCTGC-CTTA
Metadesmolaimus_sp	GCAGATCGCT	T-CGGTTTC-	-AC-ACCGGC	GACATATCC-	TTC--AAGTG	TCTGC-CTTA
Dentostomella_sp	GCTGATCCCA	--CGGTAA-	-GC-ACCGGC	GCGTCTTCT-	ATC--AAGTA	TCTGC-CTTA
Bunonema_franzi	TCCGATCGCA	--TGGTCTC-	-GT-ACCGGC	GACGTATCAT	TTC--AAGAC	TCTGC-CTTA
Bunonema_sp	GCCGATCGCA	--TGGCTCT-	-GG-GCCGGC	GACGATCAT	ATC--AAGTC	TCTGC-CTTA
Seleborca_complexa	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Acrobeloides_nanus	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Acrobeloides_bodenheimeri	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Acrobeloides_sp_PS1146	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Acrobeles_ciliatus	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--GAGTT	TCTGC-CGTA
Acrobeles_sp_PS1156	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--GAGTT	TCTGC-CGTA
Acrobeles_complexus_WCUG2	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--GAGTT	TCTGC-CGTA
Cephaloboides_sp_SB227	GATAATCGAA	---GGTT---	-AA-TCCATC	GATTTAAGC-	ATT--AGGTG	TCTGC-CCTA
Cephalobus_cubaensis	GCTGATCGCA	--CGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Cephalobus_sp_PS1143	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Cephalobus_sp_PS1196	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Cephalobus_oryzae_PS1165	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Cervidellus_alutus	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Pseudacrobelus_variabilis	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Triligulla_aluta	GCTGATCGCA	--TGGTCTT-	-GC-ACCGGC	GACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Zeldia_punctata	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	-ACGTGTCT-	TTC--AAGTA	TCTGC-CGTA
Myolaimus_sp_U81585	GCTGATCGCA	--TGGTCTT-	-GT-ACCGGC	GACAGTCT-	TTC--AAGTT	TCTGC-CTTA
Rhabditophanes_sp_KR3021	GCTGAAC---	-----TA-	-CT-CACGTA	GTCATGCCG-	TAT--AAGTA	TCTGC-TTTA
NUCLEOTIDES INCLUDED	mmmmmmmmmm--	-----	-----	-----	-----	mmmmmmmmmm

	430	440	450	460	470	480
Angiostrongylus_dujardini	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Angiostrongylus_vasorum	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Aulurostrongylus_abstrusus	GCATATCG-G	--CGGCTT--	-GA--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Didelphostrongylus_hayesi	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Crenosoma_mephitidis	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Crenosoma_vulpis	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Otostrongylus_circumlitus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Troglostrongylus_wilsoni	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Filaroides_martis	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Oslerus_osleri	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Parafilaroides_decorus	GCATATCG-A	--CGGCTT--	-GT--TCGTC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Metastrongylus_salmi	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Metastrongylus_elongatus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Skrjabinstrongylus_chitwoodrum	GCATATCG-G	--CGGCTT--	-GA--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Parelaphostrongylus_odocoilei	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Muelerius_capillaris	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Protostrongylus_rufescens	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Halocercus_invinatus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AGA--AAGTG	TCTGC-CCTA
Pseudalius_inflexus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Stenurus_minor	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Torynurus_convolutus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Syngamus_trachea	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AGGTG	TCTGC-CCTA
Stephanurus_dentatus	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Nematodirus_battus	GCATATCG-G	--CGGCTT--	-GA--CCGCT	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Strongylus_equinus	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Labiostromylus_bipapillosus	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Petrovina_poculatum	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Cylicocyclus_insignis	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Chabartia_ovina	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Cyclodontostomum_purvisi	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Zoniolaimus_mawsonae	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Hypodontus_macropi	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Deletrocephalus_dimidiatus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_eckerti_P7B8	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_capreolus_P3B2	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_sp._P6A1	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_filaria	GCATATCG-G	--CGGCTTG-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_capreolus_P2C10	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Dictyocaulus_viviparus	GCATATCG-G	--CGGCATA-	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Haemonchus_sp._V3091	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Haemonchus_contortus	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Haemonchus_placei	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Haemonchus_similis	GCATATCG-G	--CGGCTT--	-GT--TCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Ostertagia_ostertagi	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Ostertagia_leptospicularis	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Nippostrongylus_brasileinsis	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Heligmosomoides_polygyrus	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Trichostrongylus_colubriformis	GCATATCG-G	--CGGCTT--	-GT--YCGCC	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Tetraothostrongylus_mackerr	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Herpestrostrongylus_pythonis	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Nicollina_cameroni	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Filarimena_flagrifer	GCATATCG-G	--CGGCTT--	-GT--CCGCC	GATAATCCG-	AAA--AAGTG	TCTGC-CCTA
Amidostomum_cygni	GCATATCG-G	--CGGCTT--	-GT--CCGCT	GATATTCCG-	AAA--AAGTG	TCTGC-CCTA
Teratocephalus_lirellus	GCTGATCGCA	--TGGTCTT--	-GC-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Brumptaemilius_justini	GCTGATCGCA	--TGGTCTT--	-GA-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Anguillicola_crassus	GCTGATCGCA	--TGGTCTT--	-GA-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Dracunculus_medinensis	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Dracunculus_oesophageus	GCCGATCGTA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Dracunculus_sp._V3104	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Philonema_sp_A	GCTGATCGCA	--TGGTCTT--	-GC-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Philometra_obturans	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Camallanus_oxycapahalus	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Acanthocheilonema_viteae	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Loa_loa	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Onchocerca_cervicalis	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Dirofilaria_immitis	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Brugia_malay	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Wuchereria_bancrofti	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Litomosoides_sigmodontis	GCTGATCGCA	--TGGTCTT--	-GC-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Setaria_digitata	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Gnathostoma_turgidum	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Gnathostoma_neoprocyonis	GCTGATCGCA	--TGGTCTT--	-GC-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Gnathostoma_binucleatum	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Physaloptera_turgida	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Physaloptera_turgida	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Ascarophis_arctica	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Spinitectus_carolini	GCTGATCGCA	--TGGTCTT--	-GC-ACCGGC	GACATATCT-	ATC--AAGTG	TCTGC-CCTA
Thelazia_lacrymalis	GCTGATCGCA	--TGGTCTT--	-GT-ACCGGC	GACATATCT-	CAC--AAGTG	TCTGC-CCTA
NUCLEOTIDES INCLUDED	mmmmmmmmmm--	-----	-----	-----	-----	mmmmmmmmmm

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      430      440      450      460      470      480
Aphelenchus_avenae      GCTGATCGTA --TGGTCTT- -GT-ACCGAC GACGTATCT- TTC--AAGTA TCTGC-CTTA
Aphelenchoides_fragariae GCTTATCGCA --CGACTTT- -AC-GTCGGC GAAATTACA- TTC--AAGTA TCTGC-CTTA
Bursaphelenchus_sp      GCTTATCGCA --TGGTCTC- -GT-ACCGGC GAAGTTTCT- TTC--AAGTA T-TGC-TTTA
Deladenus_sp            GCTGATCGCA --TGGTCTT- -GT-ACCGGC GACGTGTCT- TTC--AAGTA TCTGC-CTTA
Criconema_sp            GCTGATCGCA --TGGGCTT- -GT-CCCGGC GACGTACCG- TCC--GAGTG TCTGC-CTTA
Hemicycliophora_conida  GCTGACCGCA --TGGGCTT- -GT-CCCGGC GCGGTGTCT- TTC--AAGTG TCTGC-CTTA
Paratylenchus_dianthus  GCTGATCGCA --TGGGCTT- -GT-CCCGGC GACGTGTCT- TTC--AAGTG TCTGC-CTTA
Pratylenchus_thornei    GCTGATCGTA --CGGTCTT- -GT-ACCGAC GACGTGTCT- TTC--AAGTG TCTGC-CTTA
Tylenchulus_sempentrans GCTGATCGTA --TGGGCTT- -GT-CCCGAC GACGTGTCT- TTC--AAGTG TCTGC-CTTA
Ditylenchus_angustus   GCTGATCGCA --TGGTCTT- -GT-ACCGGC GATGTGTCA- TTC--AAGTA TCTGC-CTTA
Subanguina_radicola    GCTGATCGCA --TGGTCTT- -GC-ACCGGC GACGTGTCT- TTC--AAGTA TCTGC-CTTA
Tylenchorhynchus_maximus GCTGATCGCA --TGGTCTT- -GT-ACCGGC GACGTGTCT- TTC--AAGTA TCTGC-CTTA
Geocenamus_quadrifer    GCTGATCGCA --TGGTCTT- -GC-ACCGGC GACGTGTCT- TTC--AAGTA TCTGC-CTTA
Globodera_pallida      GCTGATCGCA --CGGTCTT- -GT-ACCGGC GACGTGTCT- TTC--AAGTG TCTGC-CTTA
Scutellonema_bradys     GCTGATCGCA --CGGTCTT- -GGT-ACCGGC GACGTGTCT- TTC--AAGTG TCTGC-CTTA
Helicotylenchus_dihystera GCTGATCGCA --CGGTCTT- -GC-ACCGGC GACGTGTCT- TTC--AAGTG TCTGC-CTTA
Rotylenchus_robustus   GCTGATCGCA --CGGTCTT- -GC-ACCGGC GACGTGTCT- TTC--AAGTG TCTGC-CTTA
Meloidogyne_arenaria   GCTGACCGCA --TGGGCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Meloidogyne_incognita_KT GCTGACCGCA --TGGGCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Meloidogyne_javanica   GCTGACCGCA --TGGGCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Meloidogyne_articollis GCTGACCGCA --CGGTCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Meloidogyne_duytsi     GCTGACCGCA --TGGGCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Meloidogyne_exigua     GCTGACCGCA --TGGGCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Meloidogyne_hapla      GCTGACCGCA --TGGGCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Meloidogyne_ichinohei  ACTGACCGCA --CGGCCCC- -GA-GCCGGC GGCGTGTCA- TTC--AAGTG TCTAC-TTTA
Meloidogyne_maritima   GCTGACCGCA --TGGGCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Meloidogyne_microtyla  GCTGACCGCA --TGGGCTT- -GT-GCCGGC GGCGTGTCT- TTC--AAGCG TCCAC-TTTA
Nacobbus_aberrans      GCTGACCGCA --TGGTCTT- -GT-ACCGGC GGCATGTCT- TTC--AAGTA TCTGC-CTTA
Pratylenchoides_ritteri GCTGATCGCA --TGGTCTT- -GA-ACCGGC GACGTGTCT- TTC--AAGTA TCTGC-CTTA
Pratylenchoides_magnicauda GCTGATCGCA --TGGTCTT- -GA-ACCGGC GACGTGTCT- TTC--AAGTA TCTGC-CTTA
Hirschmanniella_sp._JH_2003 GCTGATCGCA --TGGGCTT- -GC-GCCGGC GACGTGTCT- TTC--AAGTA TCTGC-CTTA
Radopholus_similis     GCTGACCGCA --CGGTCTT- -GC-ACCGGC GCCGTATCA- TTC--GAGCT TCTGC-CCTA
Pratylenchus_goodeyi_VF GCTGACCGCA --TGGTCTT- -GA-ACCGGC GGCGTGTCT- TTC--AAGCA TCTGC-ATTA
Boleodorus_thylactus_clone2 GCTGATCGCA --TGGTCTT- -GC-ACCGGC GACGTGTCT- TTC--AAGTA TCTGC-CTTA
Philippine Sequence 1  GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Philippine Sequence 2  GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Philippine Sequence 3  GTTTATCG-T --CGGTTT-- -AT--CCGAC GAAAACCGA- AAG--AAGTG TCTGC-CCTA
Philippine Sequence 4  GCTGATCGTA --TGGTCTT- -GT-ACCGAC GACATGTCA- TTC--AAGTG TCTGA-TCTA
Philippine Sequence 5  GCATATCG-G --CGGCTT-- -GT--CCGCC GATATTCCG- AAA--AAGTG TCTGC-CCTA
Philippine Sequence 6  GCTTATCG-A --CGGTCT-- -AT--CCGTC GATGATCGA- AAA--ATGTG TCTGC-CCTA
Philippine Sequence 7  GTTTACTG-T --CAGTTTC- -GA--CTGAC TCTATCCGA- AAA--GGGTG TCTGC-CCTT
Tahiti Sequence 1      GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Tahiti Sequence 2      GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Thailand Sequence 1     GCTGATCGCA --TGGTCTT- -GT-ACCGGC GACGTGTCT- ATC--AAGTG TCTGC-CTTA
Ogasawara Sequence 2   GCTGATCGTA --TGGTCTT- -GT-ACCGAC GACATGTCA- TTC--AAGTG TCTGA-TCTA
Ogasawara Sequence 1   TCTATCGG-- --CGGTTT-- -AT--CCGCT GAATCTGC- AAT-TTTGTG TCTGC-CCTA
Ivory Coast Sequence 1 GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Ivory Coast Sequence 2 GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Ivory Coast Sequence 3 GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Ivory Coast Sequence 4 GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Ivory Coast Sequence 5 GCTGATCGCA --TGGTCTT- -GC-ACCGGC GACGTGTCT- ATC--AAGTG TCTGC-CTTA
Ivory Coast Sequence 6 GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Ivory Coast Sequence 7 GCATATCG-G --CGGCAT-- -GA--CCGCT GATAATCCG- AAA--AAGTG TCTGC-CCTA
Singapore Sequence 1   GTTTATCG-T --CGGTTT-- -AT--CCGAC GAAAACCGA- AAG--AAGTG TCTGC-CCTA
Singapore Sequence 2   RCTGATCGCA --TGGTCTT- -GT-ACCGGC GACGTGTCT- ATC--AAGTG TCTGC-CTTA
Singapore Sequence 3   GTTTACTG-T --CAGTTTC- -GA--CTGAC TCTATCCGA- AAA--GGGTG TCTGC-CCTT
Singapore Sequence 4   GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Nigerian Sequence 1    GCATATCG-G --CGACTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
Nigerian Sequence 2    GCATATCG-G --CGGCTT-- -GT--CCGCT GATAATCCG- AAA--AAGTG TCTGC-CCTA
Nigerian Sequence 3    GCATATCG-G --CGGCTT-- -GT--TCGCC GATAATCCG- AAA--AAGTG TCTGC-CCTA
NUCLEOTIDES INCLUDED  mmmmmmmmm-- -----

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      490      500      510      520      530      540
Gordius_aquaticus      TCA-ACTGTC GATGGTAGGT TATGCGCCT- ACCATGGTGC TAACGGG-TA ACGGAGAATC
Priapulus_caudatus     TCA-ACTTTC GATGGTAGAT TATGTGCCT- ACCATGGCTG TAACGGG-TA ACGGGGAATC
Brachionus_plicatilis  TCA-ACTTTC GATGGTAAGC GATTGCTT- ACCATGGTTG TAACGGG-TA ACGGGGAATC
Chordodes_morgani      TCA-ACTGTC GATGGTAGGT TATATGCCT- ACCATGGTTG TAACGGG-TA ACGGAGAATC
Paractinolaimus_macrolaimus TCA-ACTTTC GATGGTAGGT TATACGCCT- ACCATGGTAG TAACGGG-TA ACGGAGAATA
Aporcelaimellus_obtusicaudatus TCA-ACTTTC GATGGTAGGT TATACGCCT- ACCATGGTAG TAACGGG-TA ACGGAGAATA
Wilsonema_schuurmansstekhoveni TCA-ACTTTC GATGGTAGGT TATGTGCCT- ACCATGGTTG TTACGGG-TA ACGGAGAATA
Mesodorylaimus_sp_cf_nigritul TCA-ACTTTC GATGGTAGGT TATACGCCT- ACCATGGTAG TAACGGG-TA ACGGAGAATA
Mesodorylaimus_bastiani TCA-ACTTTC GATGGTAGGT TATACGCCT- ACCATGGTAG TAACGGG-TA ACGGAGAATA
Mesodorylaimus_japonicus TCA-ACTTTC GATGGTAGGT TCTACGCCT- ACCATGGTAG TAACGGG-TA ACGGAGAATA
Pungentus_sp._PDL_2005 TCA-ACTTTC GATGGTAGGT TATACGCCT- ACCATGGTAG TAACGGG-TA ACGGAGAATA
Allo-dorylaimus_sp     TCA-ACTTTC GATGGTAGGT TATACGCCT- ACCATGGTAG TAACGGG-TA ACGGAGAATA
Eudorylaimus_carteri   TCA-ACTTTC GATGGTAGGT TATACGCCT- ACCATGGTAG TAACGGG-TA ACGGAGAATA
NUCLEOTIDES INCLUDED  mmm-mmm--m mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmm-m mmmmmmmmmmmmm

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	490	500	510	520	530	540
Microdorylaimus_sp	TCA-AC	TTCGAT	GGTAC	GCCTAT	ACCGG	-TAA
Longidorus_elongatus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Xiphinema_rivesi	TCC-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Tylencholaimus_sp	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Anoplostoma_sp._BHMM_2005	TCA-AC	CGTCCG	GATGG	TAGTAT	GCCTAT	ACCGG
Adoncholaimus_fuscus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Enoplus_meridionalis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Enoplus_brevis_U88336	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Enoplus_communis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Enoploides_brunettii	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Syringolaimus_striatocaudatus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Ironus_dentifurcatus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Calyptronema_maxweberi	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Viscosia_sp._BHMM_2005	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Viscosia_viscosa	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Pontonema_vulgare	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Oncholaimus_sp._BHMM_2005	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Alaimus_sp_PDL_2005	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Prismatolaimus_intermedius	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Tobrillus_gracilis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Tripyla_cf_filicaudata_JH_2004	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Bathylaimus_sp	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Bathylaimus_assimilis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Bathylaimus_sp._BHMM_2005	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Tripyloides_sp._BHMM_2005	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trischiostoma_monohystera	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Mermis_nigrescens	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Mylonchulus_arenicolus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Anatonchus_tridentatus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Mononchus_truncatus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Prionchulus_muscorum	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Clarkus_sp	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichinella_spiralis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichinella_papuae	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichinella_britovi	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichinella_murrelli	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichinella_pseudospiralis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichinella_nativa	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichinella_zimbabwensis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichinella_nelsoni	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichuris_suis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichuris_trichiura	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichuris_muris	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Tylolaimophorus_minor	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Paratrichodorus_pachydermus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Paratrichodorus_anemones	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Trichodorus_primitivus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Axonolaimus_helgolandicus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Ascolaimus_elongatus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Odontophora_rectangula	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Cylindrolaimus_sp._202149	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Tylocephalus_auriculatus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Plectus_acuminatus_BS9	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Plectus_aquatilis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Anaplectus_sp	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Anisakis_sp_WKT	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Anisakis_sp_Nadler	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Contraecaeum_multipapillatum	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Pseudoterranova_decipiens	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Raphidascaris_acus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Terranova_caballeroi	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Ascaris_suum	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Ascaris_lumbricoides	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Baylisascaris_procyonis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Baylisascaris_transfuga	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Parascaris_equorum	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Porrocaecum_depressum	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Toxascaris_leonina	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Heterocheilus_tunicatus	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Goezia_pelagia	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Hysterothylacium_fortalezae	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Hysterothylacium_pelagicum	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Hysterothylacium_reliquens	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Iheringascaris_inquies	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Toxocara_canis	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Nemhelix_bakeri	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Raillietnema_sp._V3060	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
Cruzia_americana	TCA-AC	TTCGAT	GGTAG	GCCTAT	ACCGG	-TAA
NUCLEOTIDES INCLUDED	mmmm	mmmm	mmmm	mmmm	mmmm	mmmm

	490	500	510	520	530	540
Heterakis_sp_14690	TCA-AC	TTC	GATGG	TAGTT	TAAATGC	CTT
Heterakis_gallinarum	TCA-AC	TTC	GATGG	TAGTT	TAAC	TGCCT
Paraspidodera_sp_21303	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Chromadora_nudicapitata	TCA-AC	TGTC	GATGG	TAGTT	TATTGG	ACT
Chromadora_sp._BHMM_2005	TCA-AC	TGTC	GATGG	TAGTT	TATTGG	ACT
Atrochromadora_microlaima	TCA-AC	TGTC	GATGG	TAGTT	TATTGG	ACT
Chromadorina_germanica	TCA-AC	TGTC	GATGG	TAGTT	TATTGG	ACT
Chromadorita_tentabundum	TCA-AC	TTC	GATGG	TAGTC	TATAAG	CCT
Dichromadora_sp._BHMM_2005	TCA-AC	TGTC	GATGG	TAGTT	TATTGG	ACT
Neochromadora_BHMM_2005	TCA-AC	TTC	GATGG	TAGTT	TACATG	ACT
Spilophorella_paradoxa	TCA-AC	TTC	GATGG	TAGTT	TACATG	ACT
Paracanthonchus_caecus	TCA-AC	TTC	GATGG	TAGTT	TATATG	ACT
Paracyatholaimus_intermedius	TCA-AC	TGTC	GATGG	TAGTT	TATATG	ACT
Praeacanthonchus_punctatus	TCA-AC	TTC	GATGG	TAGTT	TACATG	ACT
Praeacanthonchus_sp	TCA-AC	TTC	GATGG	TAGTT	TACATG	ACT
Cyatholaimus_sp._BHMM_2005	TCA-AC	TTC	GATGG	TAGTT	TATATG	ACT
Spirinia_parasitifera	TCA-AC	TTC	GATGG	TAGTT	TACGTGC	CTT
Acanthopharynx_micans	TCA-AC	TGTC	GATGG	TAGTT	TATGTGC	CTT
Xyzzors_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Metachromadora_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Metachromadora_remanei	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Catanema_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Eubostrichus_dianae	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Eubostrichus_topiarus	TCA-AC	TTC	GATGG	TAGTT	TATATGC	CTT
Eubostrichus_parasitiferus	TCA-AC	TTC	GATGG	TAGTT	TATATGC	CTT
Laxus_oneistus	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Laxus_cosmopolitus	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Leptonemella_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Robbea_hypermnestra	TCA-AC	TAC	GATGG	TAGTT	TATGTGC	CTT
Stilbonema_majum	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Monoposthia_costata	TCA-AC	TTC	GATGG	TAGTA	TAGGGC	CTT
Nudora_bipapillata	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Calomicrolaimus_parahonestus	TCA-AC	TTC	GATGG	TAGTT	TATGTG	ACT
Calomicrolaimus_sp._BHMM_2005	TCA-AC	TTC	GATGG	TAGTT	TATGTG	ACT
Molgolaimus_demani	TCA-AC	TTC	GATGG	TAGTT	TATATG	ACT
Diplogaster_lethieri	TCA-AC	TATT	GATGG	TAGTC	TATGTG	TCT
Aduncospiculum_halicti	TCA-AC	TTC	GATGG	TAGTC	TATGTG	ACT
Pristionchus_lheritieri	TCA-AC	TATT	GATGG	TAGTC	TATGTG	TCT
Pristionchus_pacificus	TCA-AC	TATT	GATGG	TAGTC	TATGTG	TCT
Pristionchus_pacificus	TCA-AC	TATT	GATGG	TAGTC	TATGTG	TCT
Sabatieria_punctata_STRAIN_343	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_sp._355_BHMM_2005	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_celtica	TCA-AC	TGTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_punctata_STRAIN_200	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_punctata_STRAIN_223	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Sabatieria_sp._210_BHM_2005	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Setosabatieria_hilarula	TCA-AC	TGTC	GATGG	TAGTT	TATGTGC	CTT
Desmolaimus_zeelandicus	TCA-AC	TATT	GATGG	TAGTT	TATGTG	ACT
Terschellingia_longicaudata	TCA-AC	TTC	GATGG	TAGTT	TATGTG	ACT
Cyarttonema_elegans	TCA-AC	TTC	GATGG	TAGTT	TATGTG	ACT
Tridentulus_sp	TCA-AC	TTC	GATGG	TAGTT	TATGTG	ACT
Diplolaimelloides_meyli	TCA-AC	TTC	GATGG	TAGTT	TCTGTG	ACT
Diplolaimella_diavengatensis	TCA-AC	TTC	GATGG	TAGTT	TATGTG	ACT
Geomonhystera_disjuncta	TCA-AC	TGTC	GATGG	TAGTT	TATGTG	ACT
Sphaerolaimus_hirsutus	TCA-AC	TTC	GATGG	TAGTT	TATATGC	CTT
Theristus_acer	TCA-AC	TTC	GATGG	TAGTT	TATGTG	ACT
Daptonema_procerus	TCA-AC	TTC	GATGG	TAGTT	TCTACG	CTT
Daptonema_hirsutum	TCA-AC	TTC	GTTGG	TAGTT	TATACG	CTT
Daptonema_normandicum	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Daptonema_oxycerca	TCA-AC	TTC	GATGG	TAGTT	TCTACG	CTT
Daptonema_setosum	TCA-AC	TTC	GTTGG	TAGTT	TATACG	CTT
Desmodora_communis	TCA-AC	TGTC	GATGG	TAGTT	TATGTGC	CTT
Desmodora_ovigera	TCA-AC	TTC	GATGG	TAGTT	TATGTGC	CTT
Metadesmolaimus_sp	TCA-AC	TTC	GTTGG	TAGTT	TATACG	CTT
Dentostomella_sp	TCA-AC	TTC	GATGG	TGGT	TATGTGC	CTT
Bunonema_franzi	TCA-AC	TCC	GATGG	GAGT	CATGTG	CTT
Bunonema_sp	TCA-AC	TCC	GATGG	GAGT	TATGTGC	CTT
Seleborca_complexa	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
Acrobeloides_nanus	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
Acrobeloides_bodenheimeri	TCA-AC	TTC	GATGG	TGGT	TTTCTG	ACT
Acrobeloides_sp_PS1146	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
Acrobeles_ciliatus	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
Acrobeles_sp_PS1156	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
Acrobeles_complexus_WCUG2	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
Cephaloboides_sp_SB227	TCA-AC	--T	GATGG	TAGTT	TATTAG	ACT
Cephalobus_cubaensis	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
Cephalobus_sp._PS1143	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
Cephalobus_sp._PS1196	TCA-AC	TTC	GATGG	TAGTG	TATCTG	ACT
NUCLEOTIDES INCLUDED	mnm	mnm	--m	mnmnmnmnmnmnm	mnmnmnmnmnmnm	mnmnmnmnmnmnm

	490	500	510	520	530	540
Cephalobus_oryzae_PS1165	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Cervidellus_alutus	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Pseudocroboles_variabilis	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Triligulla_aluta	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Zeldia_punctada	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Myolaimus_sp_U81585	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditophanes_sp_KR3021	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Brevibucca_sp_SB261	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Halicephalobus_gingivalis	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrobelus_stameri	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Plectonchus_sp_PDL0025	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Turbatrix_aceti	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrellus_redivivus	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrellus_redivivus_PS1163	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrolaimus_subelongatus	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrolaimus_davidi	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrolaimus_cf_rigidus_AF40	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrolaimus sp. Sourhope ED2	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrolaimus sp. Sourhope ED2	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrolaimus sp. Sourhope ED2	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrolaimus sp. Sourhope ED2	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Panagrolaimus sp. Sourhope ED2	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Steinernema_carpocapsae	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Strongyloides_stercoralis	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Strongyloides_ratti	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Diploscapter_sp_PS1897	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Diploscapter_sp_PS2017	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Heterorhabditis_bacteriophora	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Heterorhabditis_hepialus	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Heterorhabditis_zelandica	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Parasitorhabditis_sp_SB281	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditoides_inermiformis	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditoides_inermis_DF5001	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditoides_regina_DF5012	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Poikilolaimus_oxycerca_SB200	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Poikilolaimus_regenfussi_SB199	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Distolabrellus_veechi_DWF1604	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Distolabrellus_veechi_DF5024	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Choriorhabditis_dudichi	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Protorhabditis_sp	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Protorhabditis_sp_DF5055	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Cruzema_tripartitum_DF5015	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Mesorhabditis_sp_PS1179	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Mesorhabditis_spiculigera_SB15	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Mesorhabditis_anisomorpha_SB12	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Teratorhabditis_palmarum_DF501	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Teratorhabditis_synpapillata_S	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_briggsae_PB102	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_elegans_N2	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_sp_CB5161	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_sp_PS1010	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_japonica	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_sp_DF5170	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_sp_SB341	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_plicata	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_vulgaris	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_sonorae	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Caenorhabditis_drosophilae	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Phasmarhabditis_hermaphrodita	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Phasmarhabditis_neopapillosa	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Phasmarhabditis_neopapillosa	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Pellioiditis_mediterranea_SB173	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Pellioiditis_marina	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Prodontorhabditis_wirathi	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Crustorhabditis_scanica	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Dolichorhabditis_sp_CEW1	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Oscheius_sp_DF5000	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Oscheius_sp_BW282	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Oscheius_myriophila_EM435	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Oscheius_insectivora	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Oscheius_dolichuroides	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditis_blumi_DF5010	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditis_sp_PS1191	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditis_sp_PS1010	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditis_myriophila_EM435	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditis_colombiana	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditella_axei_DF5006	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
Rhabditella_sp_DF5044	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA
NUCLEOTIDES INCLUDED	TCA-AC	TTC	GATGGT	TAGTGT	TATCTG	ACTGACT- ACCATGGTGA TCACGGG-TA ACGGAGAATA

	490	500	510	520	530	540
Cuticularia_sp_PS2083	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	CAACGGG-TA	ACGGAAAATA
Necator_americanus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Kalichephalus_cristatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Ancylostoma_caninum	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Angiostrongylus_cantonensis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Angiostrongylus_costaricensis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Angiostrongylus_malaysiensis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Angiostrongylus_dujardini	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Angiostrongylus_vasorum	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Aulurostrongylus_abstrusus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Didelphostrongylus_hayesi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Crenosoma_mephitidis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Crenosoma_vulpis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Otostrongylus_circumlitus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Troglostrongylus_wilsoni	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Filaroides_martis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Oslerus_osleri	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Parafilaroides_decorus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Metastrongylus_salmi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Metastrongylus_elongatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Skrjabinogylus_chitwoodrum	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Parelaphostrongylus_odocoilei	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Muellerius_capillaris	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Protostrongylus_rufescens	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Halocercus_invaginatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Pseudalius_inflexus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Stenurus_minor	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Torynurus_convolutus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Syngamus_trachea	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Stephanurus_dentatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Nematodirus_battus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Strongylus_equinus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Labiostongylus_bipapillosus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Petrovinema_poculatum	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Cylicocyclus_insignis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Chabartia_ovina	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Cyclodontostomum_purvisi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Zoniolaimus_mawsonae	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Hypodontus_macropi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Deletrocephalus_dimidiatus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Dictyocaulus_eckerti_P7B8	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Dictyocaulus_capreolus_P3B2	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Dictyocaulus_sp._P6A1	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Dictyocaulus_filaria	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Dictyocaulus_capreolus_P2C10	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Dictyocaulus_viviparus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Haemonchus_sp._V3091	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Haemonchus_contortus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Haemonchus_placei	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Haemonchus_similis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Ostertagia_ostertagi	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Ostertagia_leptospicularis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Nippostrongylus_brasileinsis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Heligmosomoides_polygyrus	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Trichostrongylus_colubriformis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Tetrabothriostongylus_mackerr	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Herpestrongylus_pythonis	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Nicollina_cameroni	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Filarimena_flagrifer	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Amidostomum_cygni	TCA-ACC--T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
Teratocephalus_lirellus	TCA-ACTTTC	GATGGTAGAT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Brumptaemilius_justini	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Anguillicola_crasus	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Dracunculus_medinensis	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Dracunculus_oesophageus	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Dracunculus_sp._V3104	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Philonema_sp_A	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Philonema_sp_B	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Camallanus_obturus	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Camallanus_oxyccephalus	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Acanthocheilonema_viteae	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Loa_loa	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Onchocerca_cervicalis	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Dirofilaria_immitis	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Brugia_malayi	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Wuchereria_bancrofti	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Litomosoides_sigmodontis	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Setaria_digitata	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
Gnathostoma_turgidum	TCA-ACTTTC	GATGGTAGTT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
NUCLEOTIDES INCLUDED	mnmn-mnmn--m	mnmnmnmnmnmnm	mnmnmnmnmnmnm	mnmnmnmnmnmnm	mnmnmnmnmnm	mnmnmnmnmnmnm

	490	500	510	520	530	540
<i>Gnathostoma_neoprocyonis</i>	TCA-AC	TTC	GATGGTAGGT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
<i>Gnathostoma_binucleatum</i>	TCA-AC	TTC	GATGGTAGGT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
<i>Physaloptera_alata</i>	TCA-AC	TTC	GATGGTAGGT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
<i>Physaloptera_turgida</i>	TCA-AC	TTC	GATGGTAGGT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
<i>Ascarophis_arctica</i>	TCA-AC	TTC	GATGGTAGGT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
<i>Spinitectus_carolini</i>	TCA-AC	TTC	GATGGTAGGT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
<i>Thelazia_lacrymalis</i>	TCA-AC	TTC	GATGGTAGGT	TATGTGCCT-	ACCATGGTTG	TAACGGG-TA	ACGGAGAATA
<i>Aphelenchus_avenae</i>	TCA-AC	TTC	GATGGTAGTG	TAGTGGACT-	ACCATGGTTG	TGACGGG-TA	ACGGAGGATA
<i>Aphelenchoides_fragariae</i>	TCA-AC	TTC	GATGGTAGTG	TATTGGACT-	ACCATGGTTG	TGACGGG-TA	ACGGAGAATC
<i>Bursaphelelenchus_sp</i>	TCA-AC	TATC	GTTGGTAGTT	TATTGGACT-	ACCATGGTGT	TGACGGG-TA	ACGGAGAATC
<i>Deladenus_sp</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGGGGATA
<i>Criconea_sp</i>	TCA-AC	TTC	GATGGTAGTG	TACGTGACT-	ACCATGGTTG	TGACGGG-TA	ACGGAGGATA
<i>Hemicycliophora_conida</i>	TCA-AC	TTC	GATGGTAGTG	TACGTGACT-	ACCATGGTTG	TGACGGG-TA	ACGGAGGATA
<i>Paratylenchus_dianthus</i>	TCA-AC	TTC	GATGGTAGTG	TACGTGACT-	ACCATGGTGT	TGACGGG-TA	ACGGAGGATA
<i>Pratylenchus_thornei</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATA
<i>Tylenchulus_semipetrans</i>	TCA-AC	TTC	GATGGTAGTG	TACGTGACT-	ACCATGGTGT	TGACGGG-TA	ACGGAGGATA
<i>Ditylenchus_angustus</i>	TCA-AC	TTC	GATGGTAGTG	TATTGGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATA
<i>Subanguina_radicola</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATA
<i>Tylenchorhynchus_maximus</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGA	TGACGGG-TA	ACGGAGGATA
<i>Geocenamus_quadriifer</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATA
<i>Globodera_pallida</i>	TCA-AC	TTC	GATGGTAGTG	TACCTGACT-	ACCATGGTGA	TGACGGG-TA	ACGGAGGATA
<i>Scutellonema_bradys</i>	TCA-AC	TTC	GATGGTAGTG	TACCTGACT-	ACCATGGTTG	TGACGGG-TA	ACGGAGGATC
<i>Helicotylenchus_dihystera</i>	TCA-AC	TTC	GATGGTAGTG	TACCTGACT-	ACCATGGTTG	TGACGGG-TA	ACGGAGGATA
<i>Rotylenchus_robustus</i>	TCA-AC	TTC	GATGGTAGTG	TACCTGACT-	ACCATGGTGA	TGACGGG-TA	ACGGAGGATA
<i>Meloidogyne_arenaria</i>	TCA-AC	TTC	GACGGGAGCA	TAATCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Meloidogyne_incognita_KT</i>	TCA-AC	TTC	GACGGGAGCA	TAATCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Meloidogyne_javanica</i>	TCA-AM	TTC	GACGGGAGCA	TAATCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Meloidogyne_artiellia</i>	TCA-AC	TTC	GACGGGAGTA	TAATCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGAATC
<i>Meloidogyne_duytsi</i>	TCA-AC	TTC	GACGGGAGCA	TAACCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Meloidogyne_exigua</i>	TCA-AC	TTC	GACGGGAGCA	TAATCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Meloidogyne_hapla</i>	TCA-AC	TTC	GACGGGAGCA	TAATCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Meloidogyne_ichinohei</i>	TCATA	ACTT	GATGTTAGTA	TCAGTGGCT-	AACATGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Meloidogyne_maritima</i>	TCA-AC	TTC	GACGGGAGCA	TAATCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Meloidogyne_microtyla</i>	TCA-AC	TTC	GACGGGAGCA	TAATCGACT-	CCCCTGGTGG	TGACGGA-TA	ACGGAGGATC
<i>Nacobbus_aberrans</i>	TCA-AC	TTC	GATGGTAGCG	TATCTGCCT-	ACCATGGTGA	TGACGGG-TA	ACGGAGGATA
<i>Pratylenchoides_ritteri</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATA
<i>Pratylenchoides_magnicauda</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATA
<i>Hirschmanniella_sp._JH_2003</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATA
<i>Radopholus_similis</i>	TCA-AC	TTC	GATGGTAGTG	TCTCTGGCT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATC
<i>Pratylenchus_goodeyi_VF</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATC
<i>Boleodorus_thylactus_clone2</i>	TCA-AC	TTC	GATGGTAGTG	TATCTGACT-	ACCATGGTGG	TGACGGG-TA	ACGGAGGATA
<i>Philippine Sequence 1</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Philippine Sequence 2</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Philippine Sequence 3</i>	TCA-ACC	-A	GATGGTAGCC	TATTAGACT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATC
<i>Philippine Sequence 4</i>	TCA-AC	TTC	GATGGTAGTG	TATATAAGTA	TATAGTACT-	TATATGGTTT	TGACGGA-TA
<i>Philippine Sequence 5</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Philippine Sequence 6</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Philippine Sequence 7</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTG	TTACGGG-TA	ACGGAGAATA
<i>Tahiti Sequence 1</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Tahiti Sequence 2</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Thailand Sequence 1</i>	TCA-AC	TTC	GATGGTAGTG	TAAATGCCT-	ACCATGGTTG	TTACGGG-TA	ACGGAGAATA
<i>Ogasawara Sequence 1</i>	TCA-AC	TTC	TATATAAGTA	TATAGTACT-	TATATGGTTT	TGACGGA-TA	ACGGAGTATT
<i>Ogasawara Sequence 2</i>	TCA-AC	TTC	GATGTTAGTC	TATTAGTCT-	AACATGGTTG	TAACGGG-TA	ACGGAGAATC
<i>Ivory Coast Sequence 1</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Ivory Coast Sequence 2</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Ivory Coast Sequence 3</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Ivory Coast Sequence 4</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Ivory Coast Sequence 5</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTG	TTACGGG-TA	ACGGAGAATA
<i>Ivory Coast Sequence 6</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Ivory Coast Sequence 7</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Singapore Sequence 1</i>	TCA-ACC	-A	GATGGTAGCC	TATTAGACT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATC
<i>Singapore Sequence 2</i>	TCA-AC	TTC	GATGGTAGTG	TAAATGCCT-	ACCATGGTTG	TTACGGG-TA	ACGGAGAATA
<i>Singapore Sequence 3</i>	TCA-ACT	-A	GATGGTAGTT	TATTGGACT-	ACCATGGTTG	TTACGGG-TA	ACGGAGAATA
<i>Singapore Sequence 4</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Nigerian Sequence 1</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TG	ACGGAGAATA
<i>Nigerian Sequence 2</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
<i>Nigerian Sequence 3</i>	TCA-ACC	-T	GATGGTAGTC	TATTAGTCT-	ACCATGGTTA	TTACGGG-TA	ACGGAGAATA
NUCLEOTIDES INCLUDED	mmmm	-mmm	-mm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	550	560	570	580	590	600
<i>Gordius_aquaticus</i>	AGGGTT	-CG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Priapulus_caudatus</i>	AGGGTT	-CG-	ATTCC--GGA	GAGGGAGCAT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Brachionus_plicatilis</i>	AGGGTT	-CG-	ATTCC--GGA	GAGGGAGCAT	GAGAAACGGC	TACCACATCT	ACGGAAGGCA
<i>Chordodes_morgani</i>	AGGGTT	-CG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Paractinolaimus_macrolaimus</i>	AGGGTT	-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Aporcelaimellus_obtusicaudatus</i>	AGGGTT	-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
NUCLEOTIDES INCLUDED	mmmmmm	-mm-	mmmmmm	-mmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	550	560	570	580	590	600
Wilsonema schuurmansstekhoveni	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Mesodorylaimus_sp_cf_nigritul	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Mesodorylaimus_bastianii	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Mesodorylaimus_japonicus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Pungentus_sp._PDL_2005	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Allodorylaimus_sp	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Eudorylaimus_carteri	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Microdorylaimus_sp	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Longidorus_elongatus	AGGGTT-YG- ACTCC--GGA	GAGGGAGCCT	GWGAAACGGC	TACCACATCC	AAGGAAGGCA	
Xiphinema_rivesi	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Tylencholaimus_sp	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Anoplostoma_sp._BHMM_2005	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Adoncholaimus_fuscus	AGGGTT-TG- ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Enoplus_meridionalis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Enoplus_brevis_U88336	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Enoplus_communis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Enoploides_brunettii	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Syringolaimus_striatocaudatus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ironus_dentifurcatus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Calyptronema_maxweberi	AGGGTT-TG- ATTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Viscosia_sp._BHMM_2005	AGGGTT-TG- ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Viscosia_viscosa	AGGGTT-TG- ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Pontonema_vulgare	AGGGTT-TG- ATTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Oncholaimus_sp._BHMM_2005	AGGGTT-TG- ATTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Alaimus_sp._PDL_2005	AGGGTT-CG- ATACC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Prismatolaimus_intermedius	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Tobrilus_gracilis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Tripyla_cf_filicaudata_JH_2004	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAKCGGC	TACCACATCC	AAGGAAGGCA	
Bathylaimus_sp	AGGGTT-TG- ACTCT--GGA	GAGGCAGCCT	GAGAGACGGC	TACCACATCC	AAGGAAGGCA	
Bathylaimus_assimilis	AGGGTT-TG- ACTCT--GGA	GAGGCAGCCT	GAGAGACGGC	TACCACATCC	AAGGAAGGCA	
Bathylaimus_sp._BHMM_2005	AGGGTT-TG- ACTCT--GGA	GAGGCAGCCT	GAGAGACGGC	TACCACATCC	AAGGAAGGCA	
Tripylodes_sp._BHMM_2005	AGGGTT-TG- ACTCT--GGA	GAGGGAGCCT	GAGAGACGGC	TACCACATCC	AAGGAAGGCA	
Trischistoma_monohystera	AGGGTT-CG- ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Mermis_nigrescens	AGG-TT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Mylonchulus_arenicolus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Anatonchus_tridentacus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Mononchus_truncatus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Prionchulus_muscorum	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Clarkus_sp	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Trichinella_spiralis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichinella_papuae	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichinella_britovi	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichinella_murrelli	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichinella_pseudospiralis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichinella_nativa	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichinella_zimbabwensis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichinella_nelsoni	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichuris_suis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichuris_trichiura	AGGGTT-CG- GCTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichuris_muris	AGGGTTTCG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Tyolaimophorus_minor	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Paratrichodorus_pachydermus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Paratrichodorus_anemones	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Trichodorus_primitivus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Axonolaimus_helgolandicus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Ascolaimus_elongatus	AGGGTT-CG- ACTCC--GGA	RAGGGAGCCT	GAAAATGGC	TACCACATCC	AAGGAAGGCA	
Odontophora_rectangula	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Cylindrolaimus_sp._202149	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Tylocephalus_auriculatus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Plectus_acuminatus_BS9	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Plectus_aquatilis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Anaplectus_sp	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Anisakis_sp_WKT	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Anisakis_sp_Nadler	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Contracaecum_multipapillatum	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Pseudoterranova_decipiens	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Raphidascaris_acus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Terranova_caballeri	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ascaris_suum	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ascaris_lumbricoides	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Baylisascaris_procyonis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Baylisascaris_transfuga	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Parascaris_equorum	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
Porrocaecum_depressum	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Toxascaris_leonina	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Heterocheilus_tunicatus	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Goelia_pelagia	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Hysterothylacium_fortalezae	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
NUCLEOTIDES INCLUDED	mmmmmm-mm-	mmmmmm--mm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	550	560	570	580	590	600
<i>Hysterothylacium pelagicum</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Hysterothylacium reliquens</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Itheringascaris iniquies</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Toxocara canis</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Nemhelix bakeri</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Raillietnema sp. V3060</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Cruzia americana</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Heterakis sp. 14690</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Heterakis gallinarum</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAAAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Paraspidodera sp. 21303</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Chromodora nudicapitata</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGATACGGC	TACCACATCC	AAGGAAGGCA	
<i>Chromadora sp. BHMM_2005</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGATACGGC	TACCACATCC	AAGGAAGGCA	
<i>Atrochromadora microlaima</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGATACGGC	TACCACATCC	AAGGAAGGCA	
<i>Chromadorina germanica</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGATACGGC	TACCACATCC	AAGGAAGGCA	
<i>Chromadorita tentabundum</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Dichromadora sp. BHMM_2005</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGATACGGC	TACCACATCC	AAGGAAGGCA	
<i>Neochromadora BHMM_2005</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Spilophorella paradoxa</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Paracanthonchus caecus</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Paracyatholaimus intermedius</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Praeacanthonchus punctatus</i>	agggtt-cg- actcc--gga	gagggagcct	gagaaacggc	taccacatcc	aaggaaggca	
<i>Praeacanthonchus sp</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Cyatholaimus sp. BHMM_2005</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Spirinia parasitifera</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Acanthopharynx micans</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Xyzzors sp</i>	AGGGTT-TG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Metachromadora sp</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Metachromadora remanei</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Catanema sp</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCG	
<i>Eubostrichus dianae</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Eubostrichus topiarus</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Eubostrichus parasitiferus</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Laxus oneistus</i>	AGGGTT-CG- GCTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Laxus cosmopolitus</i>	AGGGTT-CG- ACTCC--GGA	GAGGGATCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Leptonemella sp</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Robbea hypermestra</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Stilbonema majum</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Monoposthia costata</i>	AGGGTT-CG- ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Nudora bipapillata</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAAAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Calomicrolaimus parahonestus</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Calomicrolaimus sp. BHMM_2005</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Molgolaimus demani</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Diplogaster lethieri</i>	AGGGTT-CG- ACTCC--GGA	GAGCTAGCCT	TAGAAACGGC	TATCACATCC	AAGGAAGGCA	
<i>Aduncospiculum halicti</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGATGGCA	
<i>Pristionchus lheritieri</i>	AGGGTT-CG- ACTCC--GGA	GAGCTAGCCT	TAGAAACGGC	TATCACATCC	AAGGAAGGCA	
<i>Pristionchus pacificus</i>	AGGGTT-CG- ACTCC--GGA	GAGCTAGCCT	TAGAAACGGC	TATCACATCC	AAGGAAGGCA	
<i>Pristionchus pacificus</i>	AGGGTT-CG- ACTCC--GGA	GAGCTAGCCT	TAGAAACGGC	TATCACATCC	AAGGAAGGCA	
<i>Sabatieria punctata STRAIN_343</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Sabatieria sp. 355 BHMM_2005</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Sabatieria celtica</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Sabatieria punctata STRAIN_200</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Sabatieria punctata STRAIN_223</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Sabatieria sp. 210 BHM_2005</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Setosabatieria hilarula</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Desmolaimus zeelandicus</i>	AGGGTT-CG- ACTCC--GGA	GAGGCAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Terschellingia longicaudata</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Cyartonema elegans</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Tridentulus sp</i>	AGGGTT-TG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCT	AAGGAAGGCA	
<i>Diplolaimelloides meyli</i>	AGGGTC-TG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCT	AAGGAAGGCA	
<i>Diplolaimella diavengatensis</i>	AGGGTT-TG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCT	AAGGAAGGCA	
<i>Geomonhystera disjuncta</i>	AGGGTT-TG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCT	AAGGAAGGCA	
<i>Sphaerolaimus hirsutus</i>	AGGGTT-TG- ATTCC--GGA	GAGGGCCTCC	GAGAAATGGC	GACCCTTCC	AAGGAAGGCA	
<i>Theristus acer</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCT	AAGGAAGGCA	
<i>Daptonema procerus</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCT	AAGGAAGGCA	
<i>Daptonema hirsutum</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCC	AAGGAAGGCA	
<i>Daptonema normanicum</i>	AGGGTT-CG- ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCC	AAGGAAGGCA	
<i>Daptonema oxycerca</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCC	AAGGAAGGCA	
<i>Daptonema setosum</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCC	AAGGAAGGCA	
<i>Desmodora communis</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Desmodora ovigera</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAATGGC	TACCACATCC	AAGGAAGGCA	
<i>Metadesmolaimus sp</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCCTTCT	AAGGAAGGCA	
<i>Dentostomella sp</i>	AAGGTT-CG- ACTCC--GGA	GAAGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Bunonema franzi</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Bunonema sp</i>	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
<i>Seleborca complexa</i>	AGGGTT-CG- GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA	
<i>Acrobeloides nanus</i>	AGGGTT-CG- GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA	
<i>Acrobeloides bodenheimeri</i>	AGGGTT-CG- GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA	
<i>Acrobeloides sp. PS1146</i>	AGGGTT-CG- GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA	
NUCLEOTIDES INCLUDED	nnnnnnnn--nnn	nnnnnnnn--nnnn	nnnnnnnnnnnnnnnn	nnnnnnnnnnnnnnnn	nnnnnnnnnnnnnnnn	nnnnnnnnnnnnnnnn

	550	560	570	580	590	600
<i>Acrobeles_ciliatus</i>	AGGGTT-CG-	ACTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Acrobeles_sp_PS1156</i>	AGGGTT-CG-	ACTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Acrobeles_complexus_WCUG2</i>	AGGGTT-CG-	ACTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Cephaloboides_sp_SB227</i>	AGGGTT-CG-	TCTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Cephalobus_cubaensis</i>	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Cephalobus_sp_PS1143</i>	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Cephalobus_sp_PS1196</i>	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Cephalobus_oryzae_PS1165</i>	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Cervidellus_alutus</i>	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Pseudacrobeles_variabilis</i>	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Triligulla_aluta</i>	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Zeldia_punctada</i>	AGGGTT-CG-	GCTCC--GGA	GAGTTCGCCT	GAGAAATCGC	GAACACATCT	AAGGAAGGCA
<i>Myolaimus_sp_U81585</i>	AGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Rhabditophanes_sp_KR3021</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Brevibucca_sp_SB261</i>	AGGGTT-CG-	ACTCC--GGA	GAGGCAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Halicephalobus_gingivalis</i>	AGGGTT-CG-	TCTCC--GGA	GAGAATGCCT	TAAAAACGGC	TTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_stammeri</i>	AGGGTT-CG-	ACTCC--GGA	GAAATCGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Plectonchus_sp_PDL0025</i>	AGGGTT-CG-	ACTCC--GGA	GAAATCGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Turbatrix_aceti</i>	AGGGTT-CG-	ACTCC--GGA	GAAAACGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrellus_redivivus</i>	ATGGTT-CG-	ACTCC--GGA	GAAATAGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrellus_redivivus_PS1163</i>	AGGGTT-CG-	ACTCC--GGA	GAAATAGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_subelongatus</i>	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_davidi</i>	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_cf_rigidus_AF40</i>	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_sp_Sourhope_ED2</i>	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_sp_Sourhope_ED2</i>	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_sp_Sourhope_ED2</i>	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_sp_Sourhope_ED2</i>	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Panagrolaimus_sp_Sourhope_ED2</i>	AGGGTA-CG-	ACTCC--GGA	GAAAATGCCT	GAGAAACGGC	GTTTACATCC	AAGGAAGGCA
<i>Steinernema_carpcapsae</i>	AGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Strongyloides_stercoralis</i>	AGGGTT-CG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Strongyloides_ratti</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Diploscapter_sp_PS1897</i>	GGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Diploscapter_sp_PS2017</i>	GGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Heterorhabditis_bacteriophora</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Heterorhabditis_hepialus</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Heterorhabditis_zelandica</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Parasitorhabditis_sp_SB281</i>	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	TAGAAACGGC	AACCACATCT	AAGGAAGGCA
<i>Rhabditoides_inermiformis</i>	AGGGTT-CG-	ACTCC--GTA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Rhabditoides_inermis_DF5001</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Rhabditoides_regina_DF5012</i>	AGGGTT-TG-	ACTCC--GGA	GAAGATGCCT	TAGAAACGGC	AACTACATCT	AAGGAAGGCA
<i>Poikilolaimus_oxycerca_SB200</i>	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Poikilolaimus_regenfussi_SB199</i>	AGGGTT-TG-	ATTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCT	AAGGAAGGCA
<i>Distolabrellus_veechi_DWF1604</i>	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	TAGAAACGGC	AACCACATCT	AAGGAAGGCA
<i>Distolabrellus_veechi_DF5024</i>	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	TAGAAACGGC	AACCACATCT	AAGGAAGGCA
<i>Choriorhabditis_dudichi</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Protorhabditis_sp</i>	GGGGTT-TG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Protorhabditis_sp_DF5055</i>	GGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Cruzema_tripartitum_DF5015</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Mesorhabditis_sp_PS1179</i>	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	GAGAAACGGC	AACCACATCT	AAGGAAGGCA
<i>Mesorhabditis_spiculigera_SB15</i>	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	GAGAAACGGC	AACCACATCT	AAGGAAGGCA
<i>Mesorhabditis_anisomorpha_SB12</i>	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	GAGACACGGC	AACCACATCT	AAGGAAGGCA
<i>Teratorhabditis_palmarum_DF501</i>	AGGGTT-TG-	ACTCC--GGA	GA-GTATCCT	GAGAGACGGC	AAACACATCT	AAGGAAGGCA
<i>Teratorhabditis_synpapillata_S</i>	AGGGTT-TG-	ACTCC--GGA	GAGTATGCCT	GAGAGACGGC	AAACACATCT	AAGGAAGGCA
<i>Caenorhabditis_briggsae_PB102</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_elegans_N2</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_sp_CB5161</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_sp_PS1010</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_japonica</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_sp_DF5170</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_sp_SB341</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_plicata</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_vulgaris</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_sonorae</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Caenorhabditis_drosophilae</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Phasmorhabditis_hermaphrodita</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Phasmorhabditis_neopapillosa</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Phasmorhabditis_neopapillosa</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Pellioiditis_mediterranea_SB173</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Pellioiditis_marina</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Prodontorhabditis_wirthi</i>	ATG-TT-CG-	TCTCC--GGA	GAGCCGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Crustorhabditis_scanica</i>	AGGGTT-TG-	ACTCC--GGA	GAGGATGCCT	KAGAAACGGC	AGCCACATCT	AAGGAAGGCA
<i>Dolichorhabditis_sp_CEW1</i>	AGGGTT-CG-	GCTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Oscheius_sp_DF5000</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Oscheius_sp_BW282</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Oscheius_myriophila_EM435</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Oscheius_insectivora</i>	AGGGTT-CG-	ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
<i>Oscheius_dolichuroides</i>	AGGGTT-CG-	GCTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
NUCLEOTIDES INCLUDED	mmmmmm-mm-	mmmmmm--mm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	550	560	570	580	590	600		
Rhabditis blumi_DF5010	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditis sp_PS1191	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditis sp_PS1010	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditis myriophila_EM435	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditis colombiana	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditella axei_DF5006	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Rhabditella sp_DF5044	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Cuticularia sp_PS2083	AGGGTT	-TG-	ATTCC	--GGN	GAGGGAGCCT	GAGAAACGGC	TACCACATCT	AAGGAAGGCA
Necator americanus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Kalicephalus cristatus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ancylostoma caninum	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Angiostrongylus cantonensis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Angiostrongylus costaricensis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Angiostrongylus malaysiensis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Angiostrongylus dujardini	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Angiostrongylus vasorum	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Aulurostrongylus abstrusus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Didelphostrongylus hayesi	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Crenosoma mephitidis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Crenosoma vulpis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Otostrongylus circumlitus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Troglostrongylus wilsoni	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Filaroides martis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Oslerus osleri	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Parafilaroides decorus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Metastrongylus salmi	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Metastrongylus elongatus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Skrjabinstrongylus chitwoodrum	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Parelaphostrongylus odocoilei	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Muelerius capillaris	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Protostrongylus rufescens	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Halocercus invaginatus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Pseudalius inflexus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Stenurus minor	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Torynurus convolutus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Syngamus trachea	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Stephanurus dentatus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Nematodirus battus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Strongylus equinus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Labiostongylus bipapillosus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Petrovinema poculatum	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Cylicocyclus insignis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Chabartia ovina	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Cyclodontostomum purvisi	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Zoniolaimus mawsonae	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Hypodontus macropi	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Deletrocephalus dimidiatus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dictyocaulus eckerti_P7B8	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dictyocaulus capreolus_P3B2	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dictyocaulus sp._P6A1	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dictyocaulus filaria	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dictyocaulus capreolus_P2C10	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dictyocaulus viviparus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Haemonchus sp._V3091	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Haemonchus contortus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Haemonchus placei	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Haemonchus similis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ostertagia ostertagi	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Ostertagia leptospicularis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Nippostrongylus brasiliensis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Heligmosomoides polygyrus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Trichostrongylus colubriformis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Tetrastrongylus mackerr	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Herpestrostrongylus pythonis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Nicollina cameroni	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Filarimena flagrifer	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Amidostomum cygni	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA
Teratocephalus lirellus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Brumptaemilius justini	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GRGAAACGGC	TACCACATCC	AAGGAAGGCA
Anguillicola crassus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dracunculus medinensis	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dracunculus oesophageus	AGGGTT	-TG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Dracunculus sp._V3104	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philonema sp_A	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Philometra obturans	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Camallanus oxycephalus	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Acanthocheilonema viteae	AGGGTT	-TG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
Loa loa	AGGGTT	-CG-	ACTCC	--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA
NUCLEOTIDES INCLUDED	nnnnnnnn	-nn-	nnnnnnnn	-nnnn	nnnnnnnnnnnnnnnn	nnnnnnnnnnnnnnnn	nnnnnnnnnnnnnnnn	nnnnnnnnnnnnnnnn

	550	560	570	580	590	600
Onchocerca_cervicalis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Dirofilaria_immitis	AGGGTT-CG- ACNCA--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Brugia_malay	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Wuchereria_bancrofti	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Litosomoides_sigmodontis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Setaria_digitata	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Gnathostoma_turgidum	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Gnathostoma_neoprocyonis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Gnathostoma_binucleatum	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Physaloptera_alata	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Physaloptera_turgida	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ascarophis_arctica	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Spinitectus_carolini	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Thelazia_lacrymalis	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Aphelenchus_avenae	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	GAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Aphelenchoides_fragariae	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCATGTCT	AAGGACAGCA	
Bursaphelenchus_sp	AGGGTT-TG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCATGT-T	AAGGATAGCA	
Deladenus_sp	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Criconema_sp	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Hemicycliophora_conida	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	GACTACGTCT	AAGGATGGCA	
Pratylenchus_dianthus	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Pratylenchus_thornei	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Tylenchulus_semipetrans	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Ditylenchus_angustus	AGGGTT-TG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Subanguina_radicola	AGGGTT-TG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Tylenchorhynchus_maximus	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Geocenamys_quadrifer	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Globodera_pallida	AGGGTT-TG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Scutellonema_bradys	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Helicotylenchus_dihystera	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Rotylenchus_robustus	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_arenaria	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_incognita_KT	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_javanica	AGGGTT-CG- ACTCC--GGT	GAAGGGGCCT	GAGAAATCGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_artiellia	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_duytsi	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_exigua	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_hapla	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_ichinohei	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_maritima	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Meloidogyne_microtyla	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Nacobbus_aberrans	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Pratylenchoides_ritteri	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Pratylenchoides_magnicauda	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Hirschmanniella_sp._JH_2003	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Radopholus_similis	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTATGTCT	AAGGATAGCA	
Pratylenchus_goodeyi_VF	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Boleodorus_thylactus_clone2	AGGGTT-CG- ACTCC--GGA	GAAGGGGCCT	GAGAAATGGC	CACTACGTCT	AAGGATGGCA	
Philippine Sequence 1	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Philippine Sequence 2	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Philippine Sequence 3	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Philippine Sequence 4	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Philippine Sequence 5	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Philippine Sequence 6	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Philippine Sequence 7	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Tahiti Sequence 1	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Tahiti Sequence 2	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Thailand Sequence 1	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ogasawara Sequence 1	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ogasawara Sequence 2	AGGGTT-CG- TCTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ivory Coast Sequence 1	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ivory Coast Sequence 2	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ivory Coast Sequence 3	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ivory Coast Sequence 4	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ivory Coast Sequence 5	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ivory Coast Sequence 6	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Ivory Coast Sequence 7	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Singapore Sequence 1	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Singapore Sequence 2	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Singapore Sequence 3	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Singapore Sequence 4	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Nigerian Sequence 1	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Nigerian Sequence 2	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
Nigerian Sequence 3	AGGGTT-CG- ACTCC--GGA	GAGGGAGCCT	TAGAAACGGC	TACCACATCC	AAGGAAGGCA	
NUCLEOTIDES INCLUDED						

	610	620	630	640	650	660
Gordius_aquaticus	GCAG-GCGCG	CAAA-TTACC	CA---CTCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Priapulius_caudatus	GCAG-GCACG	CAAA-TTACC	CA---TTCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Brachionus_plicatilis	GCAG-GCGCG	CAAA-TTACC	CAC---TCCT	AGA---ACG-	--GGGAGG--	-TAGTG-ACG
Chordodes_morgani	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	GGC---TCG-	--GGGAGG--	-TAGTG-ACG
Paractinolaimus_macrolaimus	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Aporcelaimellus_obtusicaudatus	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Wilsonema_schuurmansstekhoveni	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Mesodorylaimus_sp_cf_nigritul	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Mesodorylaimus_bastiani	GCAG-GCGCG	CAAA-TTACC	CAC---TTTC	AGA---ACG-	--AAGAGG--	-TAGTG-ACG
Mesodorylaimus_japonicus	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Pungentus_sp_PDL_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Allodorylaimus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Eudorylaimus_carteri	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Microdorylaimus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Longidorus_elongatus	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Xiphinema_rivesi	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Tylencholaimus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TTCC	AGA---ACG-	--GAGAGG--	-TAGTG-ACG
Anoplostoma_sp_BHMM_2005	GCAG-GCACG	CAAA-TTACC	CAC---TCTT	GGT---ACG-	--AGGAGG--	-TAGTG-ACG
Adoncholaimus_fuscus	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Enoplus_meridionalis	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	AAC---ACG-	--GGGAGG--	-TAGTG-ACG
Enoplus_brevis_U88336	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	AAT---ACG-	--GGGAGG--	-TAGTG-ACG
Enoplus_communis	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	AAT---ACG-	--GGGAGG--	-TAGTG-ACG
Enoploides_brunettii	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGA---AAG-	--GGGAGG--	-TAGTG-ACG
Syringolaimus_striatocaudatus	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	CGA---ACG-	--GGGAGG--	-TAGTG-ACG
Ironus_dentifurcatus	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---AAG-	--GGGAGG--	-TAGTG-ACG
Calyptronema_maxweberi	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Viscosia_sp_BHMM_2005	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Viscosia_viscosa	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Pontonema_vulgare	GCAG-GCACG	CAAA-TTACC	CAC---TCCG	GCA---CGG-	--GGAGG--	-TAGTG-ACG
Oncholaimus_sp_BHMM_2005	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Alaimus_sp_PDL_2005	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GTGAGG--	-TAGTG-ACG
Prismatolaimus_intermedius	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Tobrilus_gracilis	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Tripyla_cf_filicaudata_JH_2004	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Bathylaimus_sp	GCAG-GCACG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Bathylaimus_assimilis	GCAG-GCACG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Bathylaimus_sp_BHMM_2005	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---TCG-	--GGGAGG--	-TAGTG-ACG
Tripylloides_sp_BHMM_2005	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Trischistoma_monohystera	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	GGC---ACG-	--GGGAGG--	-TAGTG-ACG
Mermis_nigrescens	GCACTGCGCG	CAAA-TTACC	CAC---TCCC	AGT---ACG-	--GGGAGG--	-TAGTG-ACG
Mylonchulus_arenicolus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Anatonchus_tridentatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGA---ACG-	--GGGAGG--	-TAGTG-ACG
Mononchus_truncatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGA---ATG-	--GGGAGG--	-TAGTG-ACG
Prionchulus_muscorum	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGA---ACG-	--GGGAGG--	-TAGTG-ACG
Clarkus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGA---ACG-	--GGGAGG--	-TAGTG-ACG
Trichinella_spiralis	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_papuae	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_britovi	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_murrelli	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_pseudospiralis	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_nativa	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_zimbabwensis	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichinella_nelsoni	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	AGC---TTG-	--GGGAGG--	-TAGTG-ACG
Trichuris_suis	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	AGA---TCG-	--GGGAGG--	-TAGTG-ACG
Trichuris_trichiura	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	AGA---TCG-	--GGGAGG--	-TAGTG-ACG
Trichuris_muris	GCAG-GCACG	CAAA-TTACC	CAC---TCCC	AGA---TCG-	--GGGAGG--	-TAGTG-ACG
Tyololaimophorus_minor	GCAG-GCGCG	CAAA-TTACC	CAC---TCCC	GGT---TCG-	--GGGAGG--	-TAGTG-ACG
Paratrichodorus_pachydermus	GCAG-GCGCG	CAAA-TTACC	CAC---TTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Paratrichodorus_anemones	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Trichodorus_primitivus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--CGGAGG--	-TAGTG-ACG
Axonolaimus_helgolandicus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGA---ACG-	--AGGAGG--	-TAGTG-ACG
Ascolaimus_elongatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGA---ACG-	--AGGAGG--	-TAGTG-ACG
Odontophora_rectangula	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGA---ACG-	--AGGAGG--	-TAGTG-ACG
Cylindrolaimus_sp_202149	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Tylocephalus_auriculatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Plectus_acuminatus_BS9	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Plectus_aquatilis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Anaplectus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Anisakis_sp_WKT	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Anisakis_sp_Nadler	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Contracaecum_multipapillatum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Pseudoterranova_decipiens	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Raphidascaris_acus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Terranova_caballeri	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ascaris_suum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ascaris_lumbricoides	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Baylisascaris_procyonis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Baylisascaris_transfuga	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
NUCLEOTIDES INCLUDED	mmmmmm-mmmmm	mmmmmm-mmmmm	mmmmmm-mmmmm	mmmmmm-mmmmm	mmmmmm-mmmmm	mmmmmm-mmmmm

	610	620	630	640	650	660
Parascaris equorum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Porrocaecum depressum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Toxascaris leonina	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Heterocheilus tunicatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Goezia pelagia	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Hysterothylacium fortalezae	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Hysterothylacium pelagicum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Hysterothylacium reliquens	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Iheringascaris inquires	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Toxocara canis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Nemhelix bakeri	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Raillietnema sp._V3060	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Cruzia americana	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Heterakis sp_14690	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Heterakis gallinarum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ATG-	--AGGAGG--	-TAGTG-ACG
Paraspidodera sp_21303	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Chromodora nudicapitata	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---GCG-	--AGGAGG--	-TAGTG-ACG
Chromodora sp._BHMM_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Atrochromadora microlaima	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Chromadorina germanica	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Chromadorita tentabundum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Dichromadora sp._BHMM_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Neochromadora BHMM_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Spilophorella paradoxa	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Paracanthonchus caecus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Paracatholaimus intermedius	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Praeacanthonchus punctatus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Praeacanthonchus sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Cyatholaimus sp._BHMM_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Spirinia parasitifera	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Acanthopharynx micans	GCTG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Xyzzors sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Metachromadora sp	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	AGT---TTG-	--AGGAGG--	-TAGTG-ACG
Metachromadora remanei	GCAG-GCGCG	CAAA-TTACC	CAA---TCTC	AGT---TTG-	--AGGAGG--	-TAGTG-ACG
Catanema sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---CCG-	--AGGAGG--	-TAGTG-ACG
Eubostrichus dianae	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Eubostrichus topiarus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Eubostrichus parasitiferus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Laxus oneistus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Laxus cosmopolitus	GCAG-GCAGC	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Leptonemella sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---TCG-	--AGGAGG--	-TAGTG-ACG
Robbea hypermestra	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Stilbonema majum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Monoposthia costata	GCAG-GCGCG	CAAA-TTACA	CAA---TGAC	AGCAAATTG-	--TTGATG--	-TAGTG-ACA
Nudora bipapillata	GCAG-GCGCG	CAAA-TTACC	CAA---TGAC	AGCAAATG-	--TTGATG--	-TAGTG-ACA
Calomicrolaimus parahonestus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Calomicrolaimus sp._BHMM_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Molgolaimus demani	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Diplogaster lethieri	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACT
Aduncospiculum halicti	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---GCG-	--AGGAGG--	-TAGTG-ACT
Pristionchus lheritieri	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACT
Pristionchus pacificus	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACT
Pristionchus pacificus	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACT
Sabatieria punctata_STRAIN_343	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Sabatieria sp._355_BHMM_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Sabatieria celtica	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Sabatieria punctata_STRAIN_200	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Sabatieria punctata_STRAIN_223	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Sabatieria sp._210_BHM_2005	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Setosabatieria hilarula	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Desmolaimus zeelandicus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Terschellingia longicaudata	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cyartonema elegans	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Tridentulus sp	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	GGT---ATG-	--AGGAGG--	-TAGTG-ACG
Diplolaimelloides meyli	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Diplolaimella diavengatensis	GCAG-GCGCG	CAAA-TTACC	CAT---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Geomonhystera disjuncta	GCAG-GCGCG	CAAA-TTACA	CAA---TCTC	AGC---ACG-	--AGGATG--	-TAGTG-ACG
Sphaerolaimus hirsutus	GCAG-GCGCG	TAAC-TTACC	CAC---TCTC	AGA---TTG-	--AGGAGG--	-TAGTG-ACG
Theristus acer	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Daptonema procerus	GCAG-GCAGC	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Daptonema hirsutum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Daptonema normandicum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Daptonema oxycerca	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Daptonema setosum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Desmodora communis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TCG-	--AGGAGG--	-TAGTG-ACG
Desmodora ovigera	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAC---TCG-	--AGGAGG--	-TAGTG-ACG
Metadesmolaimus sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Dentostomella sp	GCAG-GCGCG	CAAA-TTACC	CAC---TNTC	GGC---ATG-	--AGGAGT--	-TAGTG-ACG
NUCLEOTIDES INCLUDED	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm

	610	620	630	640	650	660
Bunonema franzi	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-AAT
Bunonema sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-AAT
Seleborca complexa	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeloides nanus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeloides bodenheimeri	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeloides sp_PS1146	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeles ciliatus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeles sp_PS1156	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acrobeles complexus WCUG2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cephaloboides sp_SB227	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TTA-----	--GTGAGA--	-TAGTG-ACT
Cephalobus cubaensis	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cephalobus sp_PS1143	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cephalobus sp_PS1196	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cephalobus oryzae PS1165	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Cervidellus alutus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Pseudacrobeles variabilis	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Triligulla aluta	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Zeldia punctada	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Myolaimus sp_U81585	GCAG-GCGCG	CAAA-TTATC	CAC---TCTC	GGC---ACG-	--AGGAGA--	-TAGTG-ACG
Rhabditophanes sp_KR3021	GCAG-GCGCG	AAAA-TTACC	CAA---TTTT	AGT---TAA-	--AAGAGG--	-TAGTG-ACG
Brevibucca sp_SB261	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Halicephalobus gingivalis	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---GCG-	--AGGAGG--	-TAGTG-ACG
Panagrobelus stammeri	GCAGG-GCGG	AAAA-TTACC	CAC---TCTT	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Plectonchus sp_PDL0025	GCAG-GCGCG	AAAA-TTACC	CAC---TCTT	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Turbatrix aceti	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	GGT---TCG-	--AGGAGG--	-TAGTG-ACG
Panagrellus redivivus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTT	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrellus redivivus PS1163	GCAG-GCGCG	AAAA-TTACC	CAC---TCTT	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus subelongatus	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus davidi	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus cf rigidus AF40	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus sp. Sourhope ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus sp. Sourhope ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus sp. Sourhope ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Panagrolaimus sp. Sourhope ED2	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Steinerinema carpocapsae	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Strongyloides stercoralis	GCAG-GCGCG	AAAA-TTACC	CAA---TTTT	AGT---TAA-	--AAGAGG--	-TAGTG-ACG
Strongyloides ratti	GCAG-GCGCG	AAAA-TTACC	CAA---TTTT	AGT---TAA-	--AAGAGG--	-TAGTG-ACG
Diploscapter sp_PS1897	GCAG-GCGCG	AAAC-TTATC	CAC---TGTC	AAA---A---	--ATGAGA--	-TAGTG-ACT
Diploscapter sp_PS2017	GCAG-GCGCG	AAAC-TTATC	CAC---TGTC	AAA---A---	--ATGAGA--	-TAGTG-ACT
Heterorhabditis bacteriophora	GCAG-GCGCG	TAAC-TTATC	CAC---TC-C	TAA---TCG-	--GTGAGA--	-TAGTG-ACT
Heterorhabditis hepialus	GCAG-GCGCG	TAAC-TTATC	CAC---TC-C	TAA---TCG-	--GTGAGA--	-TAGTG-ACT
Heterorhabditis zelandica	GCAG-GCGCG	TAAC-TTATC	CAC---TC-C	TAA---TCG-	--GTGAGA--	-TAGTG-ACT
Parasitorhabditis sp_SB281	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---KCG-	--AGGAGG--	-TAGTG-AAA
Rhabditoides inermiformis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACA
Rhabditoides inermis DF5001	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---ACG-	--AGGAGG--	-TAGTG-ACT
Rhabditoides regina DF5012	GCAG-GCGCG	TAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACA
Poikilolaimus oxycerca SB200	GCAG-GCGCG	CAAA-TTACC	CAC---TGGA	GGT---GCT-	--CCGAGG--	-TAGTG-ACT
Poikilolaimus regenfussi SB199	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ACG-	--AGGAGG--	-TAGTG-ACT
Distolabrellus veechi DWF1604	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Distolabrellus veechi DF5024	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Choriorhabditis dudichi	GCAG-GCGTG	AAAC-TTATC	CAC---TGCA	TAT---AG-	---TGAGA--	-TAGTG-ACT
Protorhabditis sp	GCAG-GCGCG	AAAC-TTATC	CAC---TGTA	TCA---AA-	---TGAGA--	-TAGTG-ACT
Protorhabditis sp_DF5055	gcag-gcgcg	aaac-ttatac	cac---tggt	gaa---aa-	---tgaga--	-tagtg-act
Cruznema tripartitum DF5015	GCAG-GCGCG	TAAC-TTATC	CAT---TATA	AAT---T---	--ATGAGA--	-TAGTG-ACT
Mesorhabditis sp_PS1179	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Mesorhabditis spiculigera SB15	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---GCG-	--AGGAGG--	-TAGTG-ACA
Mesorhabditis anisomorpha SB12	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---GCG-	--AGGAGG--	-TAGTG-ACA
Teratorhabditis palmarum DF501	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Teratorhabditis synpapillata S	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
Caenorhabditis briggsae PB102	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis elegans N2	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis sp_CB5161	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis sp_PS1010	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAT---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis sp_PS1010	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis japonica	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GTG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis sp_DF5170	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAC---A---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis sp_SB341	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAC---A---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis plicata	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GGG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis vulgaris	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis sonorae	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAC---A---	--ATGAGA--	-TAGTG-ACT
Caenorhabditis drosophilae	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GTG---T---	--ATGAGA--	-TAGTG-ACT
Phasmarhabditis hermaphrodita	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	CTT---T---	--GTGAGA--	-TAGTG-ACT
Phasmarhabditis neopapillosa	ACAG-GCGCG	TAAC-TTATC	CAC---TACT	CTT---T---	--GTGAGA--	-TAGTG-ACT
Phasmarhabditis neopapillosa	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Pellioiditis mediterranea SB173	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	ACA-----	--GTGAGA--	-TAGTG-ACT
Pellioiditis marina	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Prodontorhabditis wirthi	GCAG-GCGCG	AAAC-TTATC	CAC---TATN	ATA---G---	--ATGAAG--	-TAGTG-ACT
Crustorhabditis scanica	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACA
NUCLEOTIDES INCLUDED	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm	mmmm-mmmmm

	610	620	630	640	650	660
Dolichorhabditis_sp_CEW1	GCAG-GCGCG	TAAC-TTATC	CAC---TACC	ACA-----	--GTGAGA--	-TAGTG-ACT
Oscheius_sp_DF5000	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Oscheius_sp_BW282	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Oscheius_myriophila_EM435	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Oscheius_insectivora	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Oscheius_dolichuroides	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Rhabditis_blumi_DF5010	GCAG-GCGCG	TAAC-TTATC	CAC---TACC	AAT---A---	--GTGAGA--	-TAGCG-ACA
Rhabditis_sp_PS1191	GCAG-GCGCG	TAAC-TTATC	CAC---TACC	GAC---A---	--GTGAGA--	-TAGTG-ACA
Rhabditis_sp_PS1010	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAT---T---	--ATGAGA--	-TAGTG-ACT
Rhabditis_myriophila_EM435	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Rhabditis_colombiana	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTG-ACT
Rhabditella_axei_DF5006	GCAG-GCGCG	TAAC-TTATC	CAC---TACC	TTA-----	--GTGAGA--	-TAGTG-ACT
Rhabditella_sp_DF5044	GCAG-GCGCG	TAAC-TTATC	CAC---TACC	TTA-----	--GTGAGA--	-TAGTG-ACT
Cuticularia_sp_PS2083	GCAG-GCGCG	CAAA-TTACC	CAC---TGGA	GGT---GCT-	--CCAAGG--	-TAGTG-ACT
Necator_americanus	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Kalicephalus_cristatus	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Ancylostoma_caninum	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Angiostrongylus_cantonensis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Angiostrongylus_costaricensis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Angiostrongylus_malaysiensis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Angiostrongylus_dujardini	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Angiostrongylus_vasorum	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Aulurostrongylus_abstrusus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Didelphostrongylus_hayesi	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Crenosoma_mephitidis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Crenosoma_vulpis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Otostongylus_circumlitus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Troglostrongylus_wilsoni	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Filaroides_martis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Oslerus_osleri	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Parafilaroides_decorus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Metastrongylus_salmi	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Metastrongylus_elongatus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Skrjabinigylus_CHITWOODRUM	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Parelaphostrongylus_odocoilei	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Muelerius_capillaris	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Protostrongylus_rufescens	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Halocercus_invaginatus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Pseudalius_inflexus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Stenurus_minor	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Torynurus_convolutus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Syngamus_trachea	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Stephanurus_dentatus	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Nematodirus_battus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Strongylus_equinus	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Labiostrongylus_bipapillosus	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Petrovinema_poculatum	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	AAA---GAG-	--ATGAGA--	-TAGTG-ACT
Cylicocyclus_insignis	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	AAA---GAG-	--ATGAGA--	-TAGTG-ACT
Chabartia_ovina	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Cyclodontostomum_purvisi	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Zoniolaimus_mawsonae	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Hypodontus_macropi	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Deletrocephalus_dimidiatus	GCAG-GCGCG	TAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Dictyocaulus_eckerti_P7B8	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Dictyocaulus_capreolus_P3B2	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Dictyocaulus_sp._P6A1	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Dictyocaulus_filaria	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Dictyocaulus_capreolus_P2C10	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Dictyocaulus_viviparus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Haemonchus_sp._V3091	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---CAG-	--ATGAGA--	-TAGTG-ACT
Haemonchus_contortus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---CAG-	--ATGAGA--	-TAGTG-ACT
Haemonchus_placei	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---CAG-	--ATGAGA--	-TAGTG-ACT
Haemonchus_similis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---CAG-	--ATGAGA--	-TAGTG-ACT
Ostertagia_ostertagi	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Ostertagia_leptospicularis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Nippostrongylus_brasileinsis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Heligmosomoides_polygyrus	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Trichostrongylus_colubriformis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Tetrabothriostongylus_mackerr	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Herpestrostrongylus_pythonis	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Nicollina_cameroni	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Filarimena_flagrifer	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Amidostomum_cygni	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Teratocephalus_lirellus	GCAG-GCGCG	TAAA-TTACC	CAA---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Brumptaemilius_justini	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Anguillicola_crassus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Dracunculus_medinensis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---TAAG-	--AGGAGG--	-TAGTGAACG
Dracunculus_oesophageus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---AAG-	--AGGAGG--	-TAGTG-ACG
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	610	620	630	640	650	660
Dracunculus_sp_V3104	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---AAG-	--AGGAGG--	-TAGTG-ACG
Philonema_sp_A	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---GTG-	--AGGANG--	-TAGTG-ACG
Philometra_obturans	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACAG-	--AGGAGG--	-TAGTG-ACG
Camallanus_oxyccephalus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ACG-	--AGGAGG--	-TAGTG-ACG
Acanthocheilonema_viteae	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Loa_loa	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Onchocerca_cervicalis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Dirofilaria_immitis	GCAG-GCGCG	CAAA-TTACC	CA---TCTC	AGA---ATG-	--AGGATG--	-TAGTG-ACG
Brugia_malayi	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Wuchereria_bancrofti	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Litomosoides_sigmodontis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Setaria_digitata	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGA---ATG-	--AGGAGG--	-TAGTG-ACG
Gnathostoma_turgidum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Gnathostoma_neoprocyonis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Gnathostoma_binucleatum	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Physaloptera_alata	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Physaloptera_turgida	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ascarophis_arctica	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Spinitectus_carolini	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Thelazia_lacrymalis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Aphelenchus_avenae	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Aphelenchoides_fragariae	GCAG-GCGCG	CAAC-TTATC	CAC---TGTT	CAT---TGA-	--ACGAGA--	-TAGTG-ACG
Bursaphelenchus_sp	GCAG-GCGCG	AAAA-TTACC	CAA---TATC	AAA---ACG-	--ATGAGG--	-TAGTG-ACG
Deladenus_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGA---ACG-	--AGGAGG--	-TAGTG-AAG
Criconema_sp	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGT---TCG-	--AGGAGG--	-TAGTG-ACG
Hemicycliophora_conida	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGT---TCG-	--AGGAGG--	-TAGTG-ACG
Paratylenchus_dianthus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Pratylenchus_thornei	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---GCG-	--AGGAGG--	-TAGTG-ACG
Tylenchulus_semipetrans	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Ditylenchus_angustus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Subanguina_radicola	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Tylenchorhynchus_maximus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--AGGAGG--	-TAGTG-ACG
Geocnamus_quadriker	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Globodera_pallida	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAC---ATG-	--AGGAGG--	-TAGTG-ACG
Scutellonema_bradys	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAC---ACGC	GGAGGAGG--	-TAGTG-ACG
Helicotylenchus_dihystera	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAC---GCG-	--AGGAGG--	-TAGTG-ACG
Rotylenchus_robustus	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_arenaria	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_incognita_KT	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_javanica	GCAG-GCGCG	CAAA-TTACC	MAC---TTTC	GGC---TCC-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_artiellia	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ACG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_duytsi	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_exigua	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_hapla	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_ichinohei	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---ATG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_maritima	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Meloidogyne_microtyla	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	GGC---TCG-	--AGGAGG--	-TAGTG-ACG
Nacobbus_aberrans	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Pratylenchoides_ritteri	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Pratylenchoides_magnicauda	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ATG-	--AGGAGG--	-TAGTG-ACG
Hirschmanniella_sp._JH_2003	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACG-	--CGGAGGAG	GTAGTG-ACG
Radopholus_similis	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ATG-	--AGGAGG--	-TAGTG-ACG
Pratylenchus_goodeyi_VF	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AAT---TCG-	--AGGAGG--	-TAGTG-ACG
Boleodorus_thylactus_clone2	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGA---ACAC	TGAGGAGG--	-TAGTG-ACG
Philippine Sequence 1	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Philippine Sequence 2	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Philippine Sequence 3	GCAG-GCGCG	TAAC-TTATC	CAC---TACA	TTT-----	--GTGAGA--	-TAGAG-ACG
Philippine Sequence 4	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Philippine Sequence 5	GCAG-GCGCG	TAAC-TTATC	CAC---TCTT	GAA---GAG-	--ATGAGA--	-TAGTG-ACT
Philippine Sequence 6	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	TCA-----	--GTGAGA--	-TAGTA-ACT
Philippine Sequence 7	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Tahiti Sequence 1	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Tahiti Sequence 2	GCAG-GCGCG	AAAC-TTATC	TAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Thailand Sequence 1	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ogasawara Sequence 1	GCAG-GCGCG	AAAA-TTACC	CAC---TCTC	AGT---TCG-	--AGGAGG--	-TAGTG-ACG
Ogasawara Sequence 2	GCAG-GCGCG	TAAC-TTATC	CAC---TACT	CCT---T---	--GTGAGA--	-TAGGG-ACA
Ivory Coast Sequence 1	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 2	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 3	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 4	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 5	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGC---ATG-	--AGGAGG--	-TAGTG-ACG
Ivory Coast Sequence 6	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Ivory Coast Sequence 7	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Singapore Sequence 1	GCAG-GCGCG	TAAC-TTATC	CAC---TACA	TTT-----	--GTGAGA--	-TAGAG-ACG
Singapore Sequence 2	GCAG-GCGCG	CAAA-TTACC	CAC---TCTC	AGT---ATG-	--AGGAGG--	-TAGTG-ACG
Singapore Sequence 3	GCAG-GCGCG	AAAC-TTATC	CAC---TGTT	GAG---T---	--ATGAGA--	-TAGTG-ACT
Singapore Sequence 4	GCAG-GCGCG	AAAC-TTATC	CAA---TCTT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
Nigerian Sequence 1	GCAG-GCGCG	AAAC-TTATC	CAA---TATT	GAA---TAG-	--ATGAGA--	-TAGTG-ACT
NUCLEOTIDES INCLUDED	mmmmmm-mmmmm	mmmmmm-mmmmm	mmmmmm-mmmmm	mmmmmm-mmmmm	mmmmmm-mmmmm	mmmmmm-mmmmm

	670	680	690	700	710	720
<i>Pseudoterranova decipiens</i>	AAAAATAACG	AG-ACCGTTC	TCTCT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Raphidascaris acus</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Terranova caballeroi</i>	AAAAATAACG	AG-ACCGTTC	TCTCT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Ascaris suum</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Ascaris lumbricoides</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Baylisascaris procyonis</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Baylisascaris transfuga</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Parascaris equorum</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Porrocaecum depressum</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Toxascaris leonina</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Heterocheilus tunicatus</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Goezia pelagia</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Hysterothylacium fortalezae</i>	AAAAATAACG	AG-ACCGTTC	TCTTT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Hysterothylacium pelagicum</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Hysterothylacium reliquens</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Itheringascaris inquires</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Toxocara canis</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Nemhelix bakeri</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Raillietnema sp._V3060</i>	AAAAATAACG	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
<i>Cruzia americana</i>	AAAAATAACG	AG-GCCGTTT	TCTAT---GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Heterakis sp_14690</i>	AAAAATAACA	AG-ACCGTTC	TCTTT---AGA	GGCCG-GTTA	TT-GGAATGA	GTA-CAATTT
<i>Heterakis gallinarum</i>	AAAAATAACA	AG-ACCGTTC	TCTAT---GA	GGCCG-GTTA	TT-GGAATGA	GTA-CAATTT
<i>Paraspidodera sp_21303</i>	AAAAATAACG	AG-GCCGTTT	TCTAT---GA	GGCCG-GCCA	TC-GGAATGG	GTA-CAATTT
<i>Chromodora nudicapitata</i>	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACTT
<i>Chromadora sp._BHMM_2005</i>	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Atrochromadora microlaima</i>	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Chromadorina germanica</i>	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Chromadorita tentabundum</i>	AAAAATAACG	AG-ACAGTTC	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACTT
<i>Dichromadora sp._BHMM_2005</i>	AAAAATAACG	AG-ACAGTCT	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Neochromadora BHMM_2005</i>	AAAAATAACG	AG-ACAGTTC	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACTT
<i>Spilophorella paradoxa</i>	AAAAATAACG	AG-ACAGTTC	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACTT
<i>Paracanthonchus caecus</i>	AAAAATAACA	AG-ATGGTTC	TCTAT---GA	GGTCC-ATCA	TT-GGAATGG	GTA-CAATTT
<i>Paracyatholaimus intermedius</i>	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGTCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Praeacanthonchus punctatus</i>	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGTCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Praeacanthonchus sp</i>	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGTCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Cyatholaimus sp._BHMM_2005</i>	AAAAATAACA	AG-ATGGTTC	TCTAT---GA	GGTCC-GTCA	TT-GGAATGG	GTA-CAATTT
<i>Spirinia parasitifera</i>	AAAAATAACG	AG-ATGGTTC	TCTAT---GA	GGCCC-ATCA	TC-GGAATGG	GTA-CAATTT
<i>Acanthopharynx micans</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAACTT
<i>Xyzors sp</i>	AAAAATAACG	AG-ATGGTTC	TCTAT---GA	GGCCC-ATCA	TC-GGAATGG	GTA-CAATTT
<i>Metachromadora sp</i>	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGCCC-GTCA	TC-GGAATGA	GTA-CAATTT
<i>Metachromadora remanei</i>	AAAAATAACG	AG-ACGGTTC	TCTAT---GA	GGCCC-GTCA	TC-GGAATGA	GTA-CAATTT
<i>Catanema sp</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Eubostriechus diana</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Eubostriechus topiarus</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Eubostriechus parasitiferus</i>	AAAAATAACA	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TT-GGAATGG	GTA-CAATTT
<i>Laxus oneistus</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Laxus cosmopolitus</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Leptonemella sp</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Robbea hypermnestra</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATTT
<i>Stilbonema majum</i>	AAAAATAACG	AG-ACGGTTC	TCTTT---GA	GGTCC-GTCA	TC-GGAATGA	GTA-CAATTT
<i>Monoposthia costata</i>	AGAGGTAACG	AG-GCCGTTT	TCTGT---GA	GGTCC-GTCA	TC-GTATCGA	GTA-CAACTT
<i>Nudora bipapillata</i>	AGAAATAACG	AG-ACGGTTC	TCTAT---GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAACTT
<i>Calomicrolaimus parahonestus</i>	AAAAATAACG	AG-ACAGTTC	TCTTT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACTT
<i>Calomicrolaimus sp._BHMM_2005</i>	AAAAATAACG	AG-ACAGTTC	TCTTT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACCC
<i>Molgolaimus demani</i>	AAAAATAACG	AG-ACAGTTC	TCTAT---GA	GGTCT-GTCA	TC-GGAATGG	GTA-CAACCC
<i>Diplogaster lethieri</i>	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGTCT-GTCA	TT-GAAATGA	GCA-CAACTT
<i>Aduncospiculum halicti</i>	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGCCT-GTCA	TC-GTAATGG	GTG-TATATT
<i>Pristionchus lheritieri</i>	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGTCT-GTCA	TT-GAAATGA	GCA-CAACTT
<i>Pristionchus pacificus</i>	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGTCT-GTCA	TT-GCAATGA	GCA-CAACTT
<i>Pristionchus pacificus</i>	ATCAATAACG	AG-ACAGATC	TCTTT---GA	GGTCT-GTCA	TT-GCAATGA	GCA-CAACTT
<i>Sabatieria punctata STRAIN_343</i>	AAAAATAACA	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TT-GGAATGA	GAA-CAATTC
<i>Sabatieria sp._355_BHMM_2005</i>	AAAAATAACA	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TT-GGAATGA	GAA-CAATTC
<i>Sabatieria celtica</i>	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
<i>Sabatieria punctata STRAIN_200</i>	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
<i>Sabatieria punctata STRAIN_223</i>	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
<i>Sabatieria sp._210_BHM_2005</i>	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
<i>Setosabatieria hilarula</i>	AAAAATAACG	AG-ACGGATC	TCTTT---GA	GGCCC-GTTA	TC-GGAATGA	GAA-CAATTC
<i>Desmolaimus zeelandicus</i>	AGAAATAACG	AG-GCGTCTC	TCTTT---GA	GGTCC-GCCA	TC-GGAATGA	GGA-CAATTC
<i>Terschellingia longicaudata</i>	AAAAATAACG	AG-GCGGTTT	TCTAT---GA	GGCCC-GCTA	TC-GGAATGA	GAA-CAATAT
<i>Cyartonema elegans</i>	AAAAATAACG	AG-GCGGTTT	TCTTT---GA	GGCCC-GCTA	TC-GGAATGA	GAA-CAACAC
<i>Tridentulus sp</i>	AAAAATAACA	AG-CCGAGGC	TCATT---GA	GTTTC-GGCA	TT-GGAATGA	GAA-CAATCT
<i>Diplolaimelloides meyli</i>	AAAAATAACA	AG-AGGAGGC	TCATT---GA	GTTTC-GTCA	TT-GGAATGA	GTA-CAATCT
<i>Diplolaimella diavengatensis</i>	AAAAATAACA	AG-GTGAGGC	CCTAT---GG	GTTTC-GCCA	TT-GGAATGA	GAA-CAATAT
<i>Geomonhystera disjuncta</i>	AAACATACCG	AG-GCGATGC	CCTTT---GG	GTGTC-GTCA	TT-GGAATGA	GTC-GAATCT
<i>Sphaerolaimus hirsutus</i>	AAAAATATCG	AG-CTGGTAG	TCTTT---GA	TTATC-AGCA	TC-GGAATGA	GAA-CAATAT
<i>Theristus acer</i>	AAAAATAACG	AG-ATGGGAC	TCTATC---GA	GTACC-ATCA	TC-GGAATGA	GAA-CAATCT
<i>Daptonema procerus</i>	AAAAATAACG	AG-ATAACCC	GCTAT---GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATTT
<i>Daptonema hirsutum</i>	AAAAATAACG	AG-ATAACCC	GCTCTA---GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATCT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mm-mmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	670	680	690	700	710	720
Daptonema_normandicum	AAAAATAACG	AG-ACGATTC	TCTTTT--GA	GTATC-GTCA	TC-GGAATGA	GAA-CAATCC
Daptonema_oxycerca	AAAAATAACG	AG-ATAACCC	GCTATC--GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATTT
Daptonema_setosum	AAAAATAACG	AG-ATAACCC	GCTCTA--GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATCT
Desmodora_communis	AAAAATAACG	AG-ACGGTTC	TCTTT--GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAATCT
Desmodora_ovigera	AAAAATAACG	AG-ACGGTTC	TCTAT--GA	GGCCC-GTCA	TC-GGAATGG	GTA-CAACTT
Metadesmolaimus_sp	AAAAATAACG	AG-ATAACCC	GCTCTA--GC	GAGTT-ATCA	TC-GGAATGA	GTA-CAATCT
Dentostomella_sp	AAAAATAACG	AG-ACCGTTC	TCAAT--GA	GGCCG-GTTA	TC-GGAATGA	ATA-CAATTT
Bunonema_franzi	AGAAATAACA	AA-GTAGATG	TCTTT--GA	TGTCT-ACTA	TT-GGAATGG	GTA-CAATTT
Bunonema_sp	AGAAATAACA	AA-GCAGATG	TCTCT--GA	TGTCT-GCTA	TT-GGAATGG	GTA-CAATTC
Seleborca_complexa	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeloides_nanus	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeloides_bodenheimeri	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeloides_sp_PS1146	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeles_ciliatus	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeles_sp_PS1156	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Acrobeles_complexus_WCUG2	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Cephaloboides_sp_SB227	AAATATAAAG	AG-ACCAATC	CTCT----G	GATTG-GTTA	TT-TCAATGA	GAT-GAGCTT
Cephalobus_cubaensis	AAAAATAACG	AG-ACCATTC	TCTCAT--GA	GGCTG-GTCA	TC-GGAATGG	GTA-CAATTT
Cephalobus_sp_PS1143	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Cephalobus_sp_PS1196	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Cephalobus_oryzae_PS1165	AAAAATAACG	AG-ACCGTTC	TCTTAT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Cervidellus_alutus	AAAAATAACG	AG-ACCGTTC	TCTT-T--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Pseudacrobelus_variabilis	AAAAATAACG	AG-ACCGTTC	TCTTCT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Triligulla_aluta	AAAAATAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Zeldia_punctada	AAAAATAAC-	AG-ACCGTTC	TCTTCT--GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Myolaimus_sp_U81585	AAAAATAACG	AG-GATGTCT	TCTTT--GA	GGCCA-TCTA	TC-GTAATGA	GTA-CAATTT
Rhabditophanes_sp_KR3021	AAAAATGACA	AG-GCTACAA	ATTTT--GT	TTGTA-GTCA	TT-GGAATCC	TTC-GAGTCT
Brevibucca_sp_SB261	AAAAATAACA	AG-GCGTAC	TCTTT--GA	GGCCG-CCTA	TT-GGAATGG	GTA-CAATTT
Halicephalobus_gingivalis	TGAAATGACN	AGATANGTTN	TGTAT--AG	AATNT-ANCA	TT-GGAATGG	TTT-AATTAC
Panagrolaimus_stammeri	AGAAATGACA	AGGCTTACTC	CTTAT--GG	GGTGA-GTCA	TC-GGAATGG	GTA-CAATTT
Plectonchus_sp_PDL0025	TGAAATGACA	AGGCTTATTC	CTTAT--GG	AATGA-GTCA	TT-GGAATGG	GTC-CAATTT
Turbatrix_aceti	TGAAATGGCA	GAACTCTTTT	CTTTA--TGG	AACGG-ATTA	TT-GCAATGA	TGT-GAGTTT
Panagrellus_redivivus	AGAAAGTACA	AGATCTGTCTM	CTTCG--GG	GGCGG-GTCA	TT-GGAATGG	TTT-GAATTT
Panagrellus_redivivus_PS1163	AGAAAGTACA	AGATCTGTCC	CTTCG--GG	GGCGG-GTCA	TT-GGAATGG	TTT-GAATTT
Panagrolaimus_subelongatus	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_davidi	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_cf_rigidus_AF40	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_sp_Sourhope_ED2	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_sp_Sourhope_ED2	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_sp_Sourhope_ED2	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Panagrolaimus_sp_Sourhope_ED2	AGAAATGACA	AGATCTATTC	CTTCG--GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Steinerinema_carpocapsae	AAAAATAACA	AG-ACTGTCT	GCTTT--GC	GAACA-GTTA	TT-GGAATGG	GTA-CAATTT
Strongyloides_stercoralis	AAAAATGACA	ACCAATATTT	ATTAT--TA	ATATT-TGGA	TT-GAAATC	TTC-AAGTTT
Strongyloides_ratti	AAAAATGACA	ACCAATATTT	ATTTT--TA	GTATT-TGGA	TT-GAAATC	TTC-AAGTTT
Diploscapter_sp_PS1897	AAAAATATAA	AA-TCCCTCC	CTCAT--GG	GGAGG-GATA	TT-TAAATGA	GTA-GAGGTT
Diploscapter_sp_PS2017	AAAAATATAA	AA-TCCCTCC	CTCAT--GG	GGAGG-GATA	TT-TAAATGA	GTA-GAGGTT
Heterorhabditis_bacteriophora	AAAAATAAAA	AG-ACCATTC	CT-AT--GG	GAACG-GTTA	TT-TCAATGA	GTA-GATCTT
Heterorhabditis_hepialus	AAAAATAAAA	AG-ACCATTC	CT-AT--GG	GAACG-GTTA	TT-TCAATGA	GTA-GATCTT
Heterorhabditis_zelandica	AAAAATAAAA	AG-ACCATTC	CTAT--GG	GAACG-GTTA	TT-TCAATGA	GTA-GATCTT
Parasitorhabditis_sp_SB281	ACAACTAACG	AG-ACTGTCT	CCTAC--GG	GAACA-GTCA	TC-GGAATGG	GTA-CAATTT
Rhabditoides_inermiformis	ACAACTAACG	AG-ACCGTTC	TCTTT--GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Rhabditoides_inermis_DF5001	AACAATAACG	AG-ACAGTCT	TCTTT--GA	GGTT-GTTA	TC-GGAATGG	GTT-CAATCT
Rhabditoides_regina_DF5012	ACCAATAACG	AG-ACTCAGA	CCTTT--GG	TTTGA-GTCA	TC-GAAATGG	GTC-CAAAGT
Poikilolaimus_oxycerca_SB200	AACAATAACG	AG-ACCGTCT	TCTAT--GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Poikilolaimus_regenfussi_SB199	AACAATAACG	AG-ACCGTCT	TCTTT--GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Distolabrellus_veechi_DWF1604	ACAACTAACG	AG-ACTGCGT	CCTAT--GG	GCGCA-GTCA	TC-GAAATGG	GTA-CAATTT
Distolabrellus_veechi_DF5024	ACAACTAACG	AG-ACTGCGT	CCTAT--GG	GCGCA-GTCA	TC-GAAATGG	GTA-CAATTT
Choriorhabditis_dudichi	AAACATAAAA	AG-ACCATTC	CGTTC--GG	GAGCG-GTTT	TT-TCAATGA	ATA-GAGTTT
Protorhabditis_sp	AAACATATAA	AA-TCCCTCC	CAAAAT--GG	GGAGG-GATA	TT-TAAATGA	GTA-GAGGTT
Protorhabditis_sp_DF5055	AAGAATAAAA	AA-CCCTACC	CTTAG--GG	GGTGG-GGTA	TT-TGAATGA	GTG-AAGTTC
Cruzema_tripartitum_DF5015	ANACATAAAA	AG-ACCATTC	CTAAC--GG	GAACG-GTTA	TT-TCAATGA	TAC-GAGTGT
Mesorhabditis_sp_PS1179	ACAACTAACG	AG-ACTGTCT	CCTAT--GG	GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Mesorhabditis_spiculigera_SB15	ACAACTAACG	AG-ACTGTCT	CCTAT--GG	GAGCA-GTCA	TC-GAAATGG	GTA-AAATTC
Mesorhabditis_anisomorpha_SB12	ACGAATAACG	AG-ACTGTTC	CCTCT--GG	GAACA-GTCA	TC-GAAATGG	GTC-TAATCC
Teratorhabditis_palmarum_DF501	ACAACTAACG	AG-ACTGTCT	CTACG--GG	GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Teratorhabditis_synpapillata_S	ACAACTAACG	AG-ACTGTCT	CCTAT--GG	GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Caenorhabditis_briggsae_PB102	AAAAATATAA	AG-ACTCATC	CTTTT--GG	GATGA-GTTA	TT-TCAATGA	GTT-GAATAC
Caenorhabditis_elegans_N2	AAAAATATAA	AG-ACTCATC	CTTTT--GG	GATGA-GTTA	TT-TCAATGA	GTT-GAATAC
Caenorhabditis_sp_CB5161	AAAAATATAA	AG-ACTCATC	CTTTT--GG	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_sp_PS1010	AAAAATATAA	AG-ACTCATC	CTTTT--GG	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_japonica	AAAAATATAA	AG-ACTCATC	CTTTT--GG	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_sp_DF5170	AAAAATATAA	AG-ACTCATC	CTTCG--GG	GATGA-GTTA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_sp_SB341	AAAAATAAAA	AG-ACCCCTC	CCAAC--GG	GAGGG-GTTA	TT-TCAATGA	GTT-GAATAC
Caenorhabditis_plicata	AAAAATAGAA	AG-AGTCATT	CTTTAT--GG	GATGA-GTCA	TT-TCAATGA	GTT-GAATAT
Caenorhabditis_vulgaris	AAAAATATAA	AG-ACTCATC	CTTTT--GG	GATGA-GTTA	TT-TCAATGA	GTT-GAATAC
Caenorhabditis_sonorae	AAAAATAAAA	AG-ACCCCTC	CTTT--GG	GAGGG-GTTA	TT-TCAATGA	GTT-GAATAC
Caenorhabditis_drosophilae	AAAAATATAA	AG-ACTCATC	CTTC--GG	GGATGAGTTA	TT-TCAATGA	GTT-GAATAT
NUCLEOTIDES INCLUDED	nnnnnnnnnnnn	nnn-nnnnnnnnn	nnnnnn--m	nnnnnnn-nnnnn	nnn-nnnnnnnnn	nnnn-nnnnnnnnn

	670	680	690	700	710	720
Phasmarhabditis hermaphrodita	GAACATAAAA	AG-ACTCATC	TTTCT----	A GATGG-GTTA	TT-TCAATGA	GTT-GAACTT
Phasmarhabditis neopapillosa	GAACATAAAA	AG-ACTCATC	TTTCT----	A GATGG-GTTA	TT-TCAATGA	GTT-GAACTT
Phasmarhabditis neopapillosa	AAAAATAAAA	AG-ACCGATC	CTAT-----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTT
Pellioditis mediterranea_SB173	AAAAATAAAA	AG-ATCAATC	CTAT-----	G GATTG-ATTA	TT-TCAATGA	GTT-GAGCTT
Pellioditis marina	AAAAATAAAA	AG-ACCGATC	CTAT-----	G GATCG-GTTA	TT-TCAATGA	GTT-GAGCTT
Prodontorhabditis wirthi	GGAATAACAA	AA-CCTTTGC	CCTAT---GG	GTAGA-GGTA	TT-TGAATGA	GAA-GATATT
Crustorhabditis scanica	ACAACTAACG	AG-ACCTGTC	TCTAT---GA	GAACA-GTCA	TC-GAAATGG	GTA-CAATTT
Dolichorhabditis_sp_CEW1	AAAAATAAAA	AG-TCCAATC	CTAAC---G	GATCG-GATA	TT-TCAATGA	ATT-GAGCTT
Oscheius_sp_DF5000	AAAAATAAAA	AG-ACCAATC	-TAAC---G	GAACG-GTTA	TT-TCAATGA	ATT-GAGCTT
Oscheius_sp_BW282	AAAAATAAAA	AG-ACCAATC	CTCAC---G	GATCG-GTTA	TT-TCAATGA	GTT-GAGCTT
Oscheius_myriophila_EM435	AAAAATAAAA	AG-ACCAATC	CTCAC---G	GATCG-GTTA	TT-TCAATGA	GTT-GAGCTT
Oscheius_insectivora	AAAAATAAAA	AG-ACCAATC	CTTAC---G	GATCG-GTTA	TT-TCAATGA	GTT-CAGCTT
Oscheius_dolichuroides	AAAAATAAAA	AG-ACCATTC	CTAAC---G	GAACG-GTTA	TT-TCAATGA	ATT-GAGCTT
Rhabditis_blumi_DF5010	GAAAATAAAA	AG-ACCAATC	CTCT-----	GATTG-GTTA	TT-TCAATGA	ATT-GAGCTT
Rhabditis_sp_PS1191	GAAAATAAAA	AG-ACCAATC	CACT-----	GATTG-GTTA	TT-TCAATGA	ATT-GAGCTT
Rhabditis_sp_PS1010	AAAAATATAA	AG-ACTCATC	CTTTT---G	GATGA-GTTA	TT-TCAATGA	GTT-GAATTT
Rhabditis_myriophila_EM435	AAAAATAAAA	AG-ACCAATC	CTCAC---G	GATCG-GTTA	TT-TCAATGA	GTT-GAGCTT
Rhabditis_colombiana	AAAAATAAAA	AG-ACCAATC	CTTAC---G	GATCG-GTTA	TT-TCAATGA	GTT-GAGCTT
Rhabditella_axei_DF5006	AAAAATAAAG	AG-ACCAATC	CTTT-----	GATTG-GTTA	TT-TCAATGA	GTT-GAGCTT
Rhabditella_sp_DF5044	AAAAATAAAG	AG-ACCAATC	CTTT-----	GATTG-GTTA	TT-TCAATGA	GTT-GAGCTT
Cuticularia_sp_PS2083	AACAATAACG	AG-ACCGTCT	TCTAT---GA	AGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Necator_americanus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Kalicephalus_cristatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Ancylostoma_caninum	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_cantonensis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_costaricensis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_malaysiensis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_dujardini	AAAAATAAAA	AA-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Angiostrongylus_vasorum	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Aulurostrongylus_abstrusus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Didelphostrongylus_hayesi	AAAAATAAAA	AA-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Crenosoma_mephitidis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Crenosoma_vulpis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Otostrongylus_circumlitus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Troglstrongylus_wilsoni	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Filaroides_martis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Oslerus_osleri	AAAAATAAAA	AG-ACCGTTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Parafilaroides_decorus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Metastrongylus_salmi	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Metastrongylus_elongatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Skrjabinstrongylus_chitwoodrum	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Parelaphostrongylus_odocoilei	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Muelerius_capillaris	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Protostrongylus_rufescens	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Halocercus_invinatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Pseudalius_inflexus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Stenurus_minor	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Torynurus_convolutus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Syngamus_trachea	AAAAATAAAA	AG-ACCGTTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Stephanurus_dentatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Nematodirus_battus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GAGCTT
Strongylus_equinus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Labiostongylus_bipapillosus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Petrovina_poculatum	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Cylicocyclus_insignis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Chabartia_ovina	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Cyclodontostomum_purvisi	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Zoniolaimus_mawsonae	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Hypodontus_macropi	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Deletrocephalus_dimidiatus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_eckerti_P7B8	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_capreolus_P3B2	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_sp_P6A1	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_filaria	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_capreolus_P2C10	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Dictyocaulus_viviparus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Haemonchus_sp_V3091	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Haemonchus_contortus	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Haemonchus_placei	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Haemonchus_similis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Ostertagia_ostertagi	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Ostertagia_leptospicularis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Nippostrongylus_brasileinsis	AAAAATAAGA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Heligmosomoides_polygyrus	AAAAATAAGA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Trichostrongylus_colubriformis	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Tetrastrongylus_mackerr	AAAAATAAAA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Herpestrongylus_pythonis	AAAAATAAGA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
Nicollina_cameroni	AAAAATAAGA	AG-ACCATTC	CTAT-----	GAAAC-GTCA	TT-TCAATGA	GTT-GATCAT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mm-mmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	670	680	690	700	710	720	
Filarimena flagrifer	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Amidostomum cygni	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Teratocephalus lirellus	AAAAATAACG	AG-ACCGTTC	TCTAT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Brumptoemilius justini	AAAAATAACG	AG-ACCGTTC	TCTWT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Anguillicolia crassus	AAAAATAACG	GG-GCCGTT	TCTTT--	GA	GGCCG-GTTA	CC-GGAATGG	GTA-CAATCT
Dracunculus medinensis	AAAAATAACG	AG-ACCGTTC	TCATT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Dracunculus_oesophageus	AAAAATAACG	GG-ACCGTTC	TCATT--	GA	GGCCG-GTTA	TT-GGAATGG	GTA-CAATTT
Dracunculus_sp_V3104	AAAAATAACG	AG-ACCGTTC	TCATT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Philonema_sp_A	AAAAATAACG	AG-ACCGTTC	TCAAT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Philometra obturans	AAAAATAACG	AG-ACCGTTC	TCGAT--	GA	GGCCG-GTTA	TC-GGAATGG	GCA-CAATCT
Camallanus oxycephalus	AAAAATAACG	AG-ACCGTTC	TCTTC--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Acanthocheilonema viteae	AAAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Loa loa	AAAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Onchocerca cervicalis	AAAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Dirofilaria immitis	AAAAATAACG	AG-ACCGTTC	TCTAT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Brugia malayi	AAAAATAACG	AG-ACCGTTC	TCTTT--	GR	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Wuchereria bancrofti	AAAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Litomosoides sigmodontis	AAAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTTA	TC-GGAATGN	NTA-CAATTT
Setaria digitata	AAAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Gnathostoma turgidum	AAAAATAACG	AA-ACCGATC	TCAAT--	GA	GGCCG-GTTA	TC-GGAATGA	GTC-ACGCTT
Gnathostoma neoprocyonis	AAAAATAACG	AA-ACCGATC	TCAAT--	GA	GGCCG-GTTA	TC-GGAATGA	GTC-ACGCTT
Gnathostoma binucleatum	AAAAATAACG	AA-ACCGATC	TCAAT--	GA	GGCCG-GTTA	TC-GGAATGA	GTC-ACGCTT
Physaloptera alata	AAAAATAACG	AG-ACCGTTC	TCTTT--	GG	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Physaloptera turgida	AAAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Ascarophis arctica	AAAAATAACG	AG-ACCGTTC	TCTTC--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Spinitectus carolini	AAAAATAACG	AG-ACCGTTC	TCTTC--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Thelazia lacrymalis	AAAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTTA	TC-GGAATGG	GTA-CAATTT
Aphelenchus avenae	AAAAATAACG	AG-ACCGTTC	TCTAT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATCT
Aphelenchoides fragariae	AGAAATAACA	AG-TCCGTT	TCTTT--	GA	GGTCC-GACA	TT-GGAATGG	ATG-AAATCT
Bursaphelenchus_sp	AGAAATAACG	AG-ATCGTTC	TCTTT--	GA	GGTCC-GTTA	TT-GGAATGG	GTA-CAATCC
Deladenus_sp	AGAAATAACG	AG-ACCGTTC	TCTAC--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATCC
Criconema_sp	AAAAATAACA	AG-GCTGTT	TCTAC--	GA	GGCCA-GCCA	TT-GGAATGG	GTA-CAACAC
Hemicycliophora conida	AAAAATAACA	AG-GCCGTT	TCTTAT--	GA	GGCCG-GCCA	TT-GGAATGG	GTA-CAACTC
Paratylenchus dianthus	AAAAATAACA	AG-GCCGTT	TCTTT--	GA	GGCCG-GCCA	TT-GGAATGG	GTA-CAACTT
Pratylenchus thornei	AGAAATAACG	AG-GCCGTT	TCTTT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Tylenchulus semipentrans	AAAAATAACG	AG-ACCGTTC	TCTAC--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAACTC
Ditylenchus angustus	AGAAATAACG	AG-ACCGTTC	TTTTT--	AA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Subanguina radicola	AAAAATAACG	AG-ACCGTTC	TCTAT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATCT
Tylenchorhynchus maximus	AGAAATATCG	AG-ACCGTTC	TCTTAA--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Geocenamus quadrifer	AGAAATAACG	AG-ACCGTTC	TCTAC--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Globodera pallida	AGAAATAACG	AG-ACCGATC	TCTTAT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Scutellonema bradys	AGAAATAACG	AG-GCCGTT	TCTTAC--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Helicotylenchus dhystera	AGAAATAACA	AG-ACCGTTC	TCTTAC--	GA	GGCCG-GTTA	TT-GGAATGG	GTA-CAATTT
Rotylenchus robustus	AGAAATAACG	AG-ACCGTTC	TCACAA--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATCT
Meloidogyne arenaria	AGAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Meloidogyne incognita KT	AGAAATAACG	AG-ACTGTT	TCTTT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Meloidogyne javanica	AGAAATAACG	AG-ATTGTT	TCTTT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Meloidogyne artiellia	AGAAATAACA	AG-ACCGTTC	TCTTCA--	GA	GGCCG-GTCA	TT-GGAATGG	GTA-CAACTT
Meloidogyne duytsi	AGAAATAACG	AG-ATCGTTC	TCTTAT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Meloidogyne exigua	AGAAATAACG	AG-ACCGTTC	TCTTT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Meloidogyne hapla	AGAAATAACG	AG-GTCGTT	TCAAAT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Meloidogyne ichinohei	AGAAATAACG	AG-GCCGTT	TCTTCT--	GA	GGCCG-GTCA	TC-GGAATGG	GAA-AAATTT
Meloidogyne maritima	AGAAATAACG	AG-ATCGTTC	TCTTAT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Meloidogyne microtyla	AGAAATAACG	AG-ATCGTTC	TCATAT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Nacobbus aberrans	AGAAATAACG	AG-ACAGTTC	TCTTAT--	GA	GGCCT-GTCA	TC-GGAATGG	GTA-CAATCT
Pratylenchoides ritteri	AGAAATAACG	AG-ACCGTTC	TCTAC--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Pratylenchoides magnicauda	AGAAATAACG	AG-ACCGTTC	TCTAC--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATTT
Hirschmanniella sp_JH_2003	AGAAATAACG	AG-ACTGTT	TCTTCA--	GA	GGCCA-GTCA	TC-GGAATGG	GTA-CAATTT
Radopholus similis	AGAAATAACG	AG-GCCGTT	TCATAC--	GA	GGCCG-GCCA	TT-GGAATGG	AAG-AGGTTT
Pratylenchus goodeyi_VF	AGAAATAACG	AG-ACCGTTC	TCTTCA--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATCC
Boleodorus thylactus_clone2	AGAAATAACG	AG-GCCGTT	TCTAT--	GA	GGCCG-GTCA	TC-GGAATGG	GTA-CAATCT
Philippine Sequence 1	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Philippine Sequence 2	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Philippine Sequence 3	AAAAATAGAA	AG-GGTAAT	CTCT----	G	GATTT-CCTA	TT-TCAATGA	GTT-GAGCCC
Philippine Sequence 4	AGAAATGACA	AG-ATCTATT	CCTTCG--	GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Philippine Sequence 5	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Philippine Sequence 6	AAAAATAAAA	AG-GCCAAT	CTAT----	G	GATCG-GTTA	TT-TCAATGA	GCT-GAGCTT
Philippine Sequence 7	AAAAATATAA	AG-ACTCAT	CTTTT--	G	GATGA-GTTA	TT-TCAATGA	GTT-GAATAC
Tahiti Sequence 1	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTTA	TT-TCAATGA	GTT-GATCAT
Tahiti Sequence 2	AAAAATAAAA	AG-ACCATT	CTAT----	G	GARCG-GTTA	TT-SCAATGA	GTC-GWTCAT
Thailand Sequence 1	AAAAATAACA	AG-ACCGTTC	TCTT--	AGA	GGCCG-GTTA	TT-GGAATGG	GTA-CAATTT
Ogasawara Sequence 1	AGAAATGACA	AGATCTATT	CTTCG--	GG	GATGG-ATCA	TT-GGAATGG	TTT-GATTTT
Ogasawara Sequence 2	GTACATAAAA	AG-ACTCAT	TTTC----	TA	GATGG-GTTA	TT-TCAATGA	GTT-GAATCT
Ivory Coast Sequence 1	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Ivory Coast Sequence 2	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Ivory Coast Sequence 3	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAATCGGTCA	TT-TCAATGA	GTT-GATCAT
Ivory Coast Sequence 4	AAAAATAAAA	AG-ACCATT	CTAT----	G	GAACG-GTCA	TT-TCAATGA	GTT-GATCAT
Ivory Coast Sequence 5	AAAAATAACA	AG-ACCGTTC	TCTAT--	GA	GGCCG-GTTA	TT-GGAATGA	GTA-CAATTT
NUCLEOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	670 680 690 700 710 720
Ivory Coast Sequence 6	AAAAATAAAA AG-ACCATTCTAT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Ivory Coast Sequence 7	AAAAATAAAA AG-ACCATTCTAT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Singapore Sequence 1	AAAAATAGAA AG-GGTAATCTCT-----G GATT-CCTA TT-TCAATGA TTT-GAGCCC
Singapore Sequence 2	AAAAATAACA AG-ACCGTTCCTT---AGA GGCCG-GTTA TT-GGAATGA GTA-CAATAT
Singapore Sequence 3	AAAAATATAA AG-ACCTATCCTTT----G GATGA-GTTA TT-TCAATGA GTT-GAATAC
Singapore Sequence 4	AAAAATAAAA AG-ACCATTCTAT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Nigerian Sequence 1	AAAAATAAAA AG-ACCATTCTAT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Nigerian Sequence 2	AAAAATAAAA AG-ACCATTCTAT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
Nigerian Sequence 3	AAAAATAAAA AG-ACCATTCTAT-----G GAACG-GTCA TT-TCAATGA GTT-GATCAT
NUCLEOTIDES INCLUDED	mmmmmmmmmm mm-mmmmmmm mmmmm-----m mmmmm-mmmmm mm-mmmmmmm mmm-mmmmmmm

	730 740
Gordius_aquaticus	A-AATCCTTT AACGAGGATC TATTGGAG
Priapulus_caudatus	A-AATCCTTT AACGAGGATC TATTGGAG
Brachionus_plicatilis	A-AAACCTTT AACGAGGATC TATTGGAG
Chordodes_morgani	A-AATCCTTT AACGAGGATC TATTGGAG
Paractinolaimus_macrolaimus	A-AATCCTTT AACGAGGATC TATTGGAG
Aporcelaimellus_obtusicaudatus	A-AATCCTTT AACGAGGATC TATTGGAG
Wilsonema_schuurmansstekhoveni	A-AACCCTTT AACGAGGACC TATGAGAG
Mesodorylaimus_sp_cf_nigritul	A-AATCCTTT AACGAGGATC TATTGGAG
Mesodorylaimus_bastiani	A-AATCCTTT AACGAGGATC TATTGGAG
Mesodorylaimus_japonicus	A-AATCCTTT AACGAGGATC TATTGGAG
Pungentus_sp._PDL_2005	A-AATCCTTT AACGAGGATC TATTGGAG
Allodorylaimus_sp	A-AATCCTTT AACGAGGATC TATTGGAG
Eudorylaimus_carteri	A-AATCCTTT AACGAGGATC TATTGGAG
Microdorylaimus_sp	A-AATCCTTT AACGAGGATC TATTGGAG
Longidorus_elongatus	A-AATCCTTT AACGAGGATC TATTGGAG
Xiphinema_rivesi	A-AATCCTTT AACGAGGATC TATTGGAG
Tylencholaimus_sp	A-AATCCTTT AACGAGGATC TATTGGAG
Anoplostoma_sp._BHMM_2005	A-AATCCTTT AACGAGGATC TATTGGAG
Adoncholaimus_fuscus	A-AATCCTTT ATCGAGGATC TATTGGAG
Enoplus_meridionalis	A-AATCCTTT AACGAGGATC TATTGGAG
Enoplus_brevis_U88336	A-AATCCTTT AACGAGGATC TATTGGAG
Enoplus_communis	A-AATCCTTT AACGAGGATC TATTGGAG
Enoploides_brunettii	A-AATCCTTT AACGAGGATC TATTGGAG
Syringolaimus_striatocaudatus	A-AATCCTTT AACGAGGATC CATTGGAG
Ironus_dentifurcatus	A-AATCCTTT AATGAGGATC TATTGGAG
Calyptronema_maxweberi	A-AATCCTTT ATCGAGGATC TATTGGAG
Viscosia_sp._BHMM_2005	A-AATCCTTT ATCGAGGATC TATTGGAG
Viscosia_viscosa	A-AATCCTTT ATCGAGGATC TATTGGAG
Pontonema_vulgare	A-AATCCTTT ATCGAGGATC TATTGGAG
Oncholaimus_sp._BHMM_2005	A-AATCCTTT ATCGAGGATC TATTGGAG
Alaimus_sp_PDL_2005	A-AATCCTTT AACGAGGATC AATTGGAG
Prismatolaimus_intermedius	A-AATCCTTT AACGAGGAT- TATTGGAG
Tobrilus_gracilis	A-AATCCTTT AACGAGGATC TATTGGAG
Tripyla_cf_filicaudata_JH_2004	A-AATCCTTT AACGAGGAAC TATTGGAG
Bathylaimus_sp	A-AATCCTTT AACGAGGATC TATTGGAG
Bathylaimus_assimilis	A-AATCCTTT AACGAGGATC TATTGGAG
Bathylaimus_sp._BHMM_2005	A-AATCCTTT AACGAGGATC TATTGGAG
Tripylloides_sp._BHMM_2005	A-AATCCTTT AACGAGGATC TATTGGAG
Trischiostoma_monohystera	A-AATCCTTT AACGAGGATC TATTGGAG
Mermis_nigrescens	A-AATCCTTT AACGAGGATC TATTAGAG
Mylonchulus_arenicolus	A-AATCCTTT AACGAGGATC TATTAGAG
Anatonchus_tridentatus	A-AAAACCTTT AACGAGGATC TATTAGAG
Mononchus_truncatus	A-AATCCTTT AACGAGGATC TATTAGAG
Prionchulus_muscorum	A-AATCCTTT AACGAGGATC TATTAGAG
Clarkus_sp	A-AATCCTTT AACGAGGATC TATTAGAG
Trichinella_spiralis	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_papuae	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_britovi	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_murrelli	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_pseudospiralis	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_nativa	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_zimbabwensis	A-CACACGCT AACGAGTAGC AATTGGAG
Trichinella_nelsoni	A-CACACGCT AACGAGTAGC AATTGGAG
Trichuris_suis	A-CACAAGCT CGGCTAAATC TATTGGAG
Trichuris_trichiura	A-CATAAGTT CGGCTAAATC TATTGGAG
Trichuris_muris	A-CATAAGCT CGGCTAAATC TATTGGAG
Tylolaimophorus_minor	A-AATCCTTT AACGAGGATC TATTGGAG
Paratrichodorus_pachydermus	A-AATCCTTT AACGAGGATC TATTGGAG
Paratrichodorus_anemones	A-AATCCTTT AATGAGGATC TATTGGAG
Trichodorus_primitivus	A-AATCCTTT AACGAGGATC TATTGGAG
Axonolaimus_helgolandicus	A-AACCTTTT AACGAGGATC TACGAGAG
Ascolaimus_elongatus	A-AACCTTTT AACGAGGATC TACGAGAG
Odontophora_rectangula	A-AACCTTTT AACGAGGATC TACGAGAG
Cylindrolaimus_sp._202149	A-AACCCTTT AACGAGGATC TACGAGAG
NUCLEOTIDES INCLUDED	m-mmmmmmmmm mmmmmmmmmmm mmmmm.....

	730 740
Tylocephalus_auriculatus	A-AACCCCTT AACGAGGACC TATGAGAG
Plectus_acuminatus_BS9	A-AACCCCTT AACGAGGACC TATGAGAG
Plectus_aquatilis	A-AACCCCTT AACGAGGACC TATGAGAG
Anaplectus_sp	A-AACCCCTT AACGAGGACC TATGAGAG
Anisakis_sp_WKT	A-AACCCGTT AACGAGGATC TATGAGAG
Anisakis_sp_Nadler	A-AACCCGTT AACGAGGATC TATGAGAG
Contraecaecum_multipapillatum	A-AACCCGTT AACGAGGATC TATGAGAG
Pseudoterranova_decipiens	A-AACCCGTT AACGAGGATC TATGAGAG
Raphidascaris_acus	A-AACCCGTT AACGAGGATC TATGAGAG
Terranova_caballeroi	A-AACCCGTT AACGAGGATC TATGAGAG
Ascaris_suum	A-AACCCGTT AACGAGGATC TATGAGAG
Ascaris_lumbricoides	A-AACCCGTT AACGAGGATC TATGAGAG
Baylisascaris_procyonis	A-AACCCGTT AACGAGGATC TATGAGAG
Baylisascaris_transfuga	A-AACCCGTT AACGAGGATC TATGAGAG
Parascaris_equorum	A-AACCCGTT AACGAGGATC TATGAGAG
Porrocaecum_depressum	A-AACCCGTT AACGAGGATC TATGAGAG
Toxascaris_leonina	A-AACCCGTT AACGAGGATC TATGAGAG
Heterocheilus_tunicatus	A-AACCCGTT AACGAGGATC TATGAGAG
Goezia_pelagia	A-AACCCGTT AACGAGGATC TATGAGAG
Hysterothylacium_fortalezae	A-AACCCGTT AACGAGGATC TATGAGAG
Hysterothylacium_pelagicum	A-AACCCGTT AACGAGGATC TATGAGAG
Hysterothylacium_reliquens	A-AACCCGTT AACGAGGATC TATGAGAG
Itheringascaris_inquies	A-AACCCGTT AACGAGGATC TATGAGAG
Toxocara_canis	A-AACCCGTT AACGAGGATC TATGAGAG
Nemhelix_bakeri	A-AACCCGTT AACGAGGATC TATGAGAG
Raillietnema_sp._V3060	A-AACCCGTT AACGAGGATC TATAAGAG
Cruzia_americana	A-AACCCGTT AACGAGGATC TATGAGAG
Heterakis_sp_14690	A-AATCCGTT AACGAGGATC TATGAGAG
Heterakis_gallinarum	A-AATCCGTT AACGAGGATC TATGAGAG
Paraspidodera_sp_21303	A-AACCCGTT AACGAGGATC TATGAGAG
Chromodora_nudicapitata	A-AACCCCTT AACGAGGATC TATTGGAG
Chromadora_sp._BHMM_2005	A-AACCCCTT AACGAGGATC TATTGGAG
Atrochromadora_microlaima	A-AACCCCTT AACGAGGATC TATTGGAG
Chromadorina_germanica	A-AACCCCTT AACGAGGATC TATTGGAG
Chromadorita_tentabundum	A-AACCCCTT AACGAGGATC TATTGGAG
Dichromadora_sp._BHMM_2005	A-AACCCCTT AACGAGGATC TATTGGAG
Neochromadora_BHMM_2005	A-AACCCCTT AACGAGGATC TATTGGAG
Spilophorella_paradoxa	A-AACCCCTT AACGAGGATC TATTGGAG
Paracanthonchus_caecus	A-AACCCCTT AACGAGGATC TATTGGAG
Paracyatholaimus_intermedius	A-AACCCCTT AACGAGGATC TATTGGAG
Praeacanthonchus_punctatus	A-AACCCCTT AACGAGGATC TATTGGAG
Praeacanthonchus_sp	A-AACCCCTT AACGAGGATC TATTGGAG
Cyatholaimus_sp._BHMM_2005	A-AACCCCTT AACGAGGATC TATTGGAG
Spirinia_parasitifera	A-AACCCCTT AACGAGGATC TATTGGAG
Acanthopharynx_micans	A-AACCCCTT AACGAGGATC TATTGGAG
Xyzzors_sp	A-AACCCCTT AACGAGGATC TATTGAAG
Metachromadora_sp	A-AACCCCTT AATGAGGATC TATTGGAG
Metachromadora_remanei	A-AACCCCTT AATGAGGATC TATTGGAG
Catanema_sp	A-AACCCCTT AACGAGGATC TATTGGAG
Eubostrichus_dianae	A-AACCCCTT AACGAGGATC TATTGGAG
Eubostrichus_topiarus	A-AACCCCTT AACGAGGATC TATTGGAG
Eubostrichus_parasitiferus	A-AACCCCTT AACGAGGATC TATTGGAG
Laxus_oneistus	A-AACCCCTT AACGAGGATC TATTGGAG
Laxus_cosmopolitus	A-AACCCCTT AACGAGGATC TATTGGAG
Leptonemella_sp	A-AACCCCTT AACGAGGATC TATTGGAG
Robbea_hypermnestra	A-AACCCCTT AACGAGGATC TACTGGAG
Stilbonema_majum	A-AACCCCTT AACGAGGATC TATTGGAG
Monoposthia_costata	A-AAAAATT AACGATA-TC AATTGGAG
Nudora_bipapillata	A-AACCCCTT AACGAGGATC AATTGGAG
Calomicrolaimus_parahonestus	A-AACCCCTT AACGAGGATC TATTGGAG
Calomicrolaimus_sp._BHMM_2005	A-AACCCCTT AACGAGGATC TATTGGAG
Molgolaimus_demani	A-AACCCCTT AACGAGGATC TATTGGAG
Diplogaster_lethieri	A-AAGACTTT AACGAAG-TC TATGGGAG
Aduncospiculum_halicti	A-AACCCCTT AACGAGGATC TATGAGAG
Pristionchus_lheritieri	A-AAGACTTT AACGAAG-TC TATGGGAG
Pristionchus_pacificus	A-AAGACTTT AACGAAG-TC TATGGGAG
Pristionchus_pacificus	A-AAGACTTT AACGAAG-TC TATGGGAG
Sabatieria_punctata_STRAIN_343	A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_sp._355_BHMM_2005	A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_celtica	A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_punctata_STRAIN_200	A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_punctata_STRAIN_223	A-AATCCTTT AACGAGGATC TAGCGGAG
Sabatieria_sp._210_BHM_2005	A-AATCCTTT AACGAGGATC TAGCGGAG
Setosabatieria_hilarula	A-AATCCTTT AACGAGGATC TAGCGGAG
Desmolaimus_zeelandicus	A-AACGACTT ACCGAATT-C TATCAGAG
Terschellingia_longicaudata	A-AATCCTTT AACGAGGATC TAGCGGAG
Cyartonema_elegans	A-AATCCTTT AACGAGGATC TAGCGGAG
Tridentulus_sp	A-AATCCTTT AACGAGGATC TAGTGGAG
NUCLEOTIDES INCLUDED	m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmm


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730          740
Diplolaimelloides_meyli      A-AATCTTTT AACGAGGATC TAGTGGAG
Diplolaimella_diavengatensis A-AATCTTTT AACGAGTAAC AAGTGGAG
Geomonhystera_disjuncta     A-AAAAATTC GGCAAGGATC TAGTGGAG
Sphaerolaimus_hirsutus      A-AATGTGTT ATCGAGTAAC TAGCAGAG
Theristus_acer              A-AACCCTTT AACGAGGACC TAGCGGAG
Daptonema_procerus          A-AATCCTTT AACGAGGACC TAGCGGAG
Daptonema_hirsutum          A-AATCCTTT AACGAGGACC TAGCGGAG
Daptonema_normandicum       A-AATCCTTT AACGAGGACC TAGCGGAG
Daptonema_oxycerca          A-AATCCTTT AACGAGGACC TAGCGGAG
Daptonema_setosum           A-AATCCTTT AACGAGGACC TAGCGGAG
Desmodora_communis          A-AACCCTTT AACGAGGATC TATTGGAG
Desmodora_ovigera           A-AACCCTTT AACGAGGATC TATTGGAG
Metadesmolaimus_sp          A-AATCCTTT AACGAGGACC TAGCGGAG
Dentostomella_sp            A-AACCCTTT AATGAGGATC TATGAGAG
Bunonema_franzi              A-AAACCCAT AACGAGGACC TATGAGAG
Bunonema_sp                  A-AAACCCAT AACGAGGATC TATGAGAG
Seleborca_complexa          A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeloides_nanus          A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeloides_bodenheimeri   A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeloides_sp_PS1146      A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeles_ciliatus          A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeles_sp_PS1156         A-AACCCTTT AACGAGTATC TATGAGAG
Acrobeles_complexus_WCUG2   A-AACCCTTT AACGAGTATC TATGAGAG
Cephaloboides_sp_SB227      A-AATAGCTC TTCGAGGACC CAGTGGAG
Cephalobus_cubaensis        A-AACCCTTT AACGAGTATC TATGAGAG
Cephalobus_sp._PS1143       A-AACCCTTT AACGAGTATC TATGAGAG
Cephalobus_sp._PS1196       A-AACCCTTT AACGAGTATC TATGAGAG
Cephalobus_oryzae_PS1165    A-AACCCTTT AACGAGTATC TATGAGAG
Cervidellus_alutus          A-AACCCTTT AACGAGTATC TATGAGAG
Pseudacrobeles_variabilis   A-AACCCTTT AACGAGTATC TATGAGAG
Triligulla_aluta            A-AACCCTTT AACGAGTATC TATGAGAG
Zeldia_punctada              A-AACCCTTT AACGAGTATC TATGAGAG
Myolaimus_sp_U81585         A-AATCCTTT AACGAGGATC TATGAGAG
Rhabditophanes_sp._KR3021   A-AATAAC-T CGTT-GG-TA AAGGAAAG
Brevibucca_sp._SB261        A-AACCCTTT AACGAGGATC TATGAGAG
Halicephalobus_gingivalis   A-AATTCATT AAAGATTATC AATGAGAG
Panagrobelus_stammeri       A-AACCCTTT AACGATTATC TATGAGAG
Plectonchus_sp._PDL0025     A-AACCCTTG AACGAACG-C AAAAAAGAG
Turbatrix_aceti              ATAACCTTA CAAATTG-C TATGAGAG
Panagrellus_redivivus       A-CAACCTTC CTAGATTATC TATGAGAG
Panagrellus_redivivus_PS1163 A-CAACCTTC CTAGATTATC TATGAGAG
Panagrolaimus_subelongatus   A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus_davidi        A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus_cf_rigidus_AF40 A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus sp. Sourhope ED2 A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus sp. Sourhope ED2 A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus sp. Sourhope ED2 A-AAATCATT AAAGAGTATC AATGAGAG
Panagrolaimus sp. Sourhope ED2 A-AAATCATT AAAGAGTATC AATGAGAG
Steinernema_carpocapsae     A-AACTCTTT AACGAGGACC TATGAGAG
Strongyloides_stercoralis   A-AATMACCT TGTGTAATAA GGAAAGGG
Strongyloides_ratti         A-AATAAC-T TGTGTAATAA GGAAAGGG
Diploscapter_sp_PS1897      A-AAAAGCTC TTCGAGAAAC TAAGAGAG
Diploscapter_sp_PS2017      A-AAAAGCTC TTCGAGAAAC TAAGAGAG
Heterorhabditis_bacteriophora A-AATCTATC TTCGAGTATC TAGTGGAG
Heterorhabditis_hepialus    A-AATCTATC TTCGAGTATC TAGTGGAG
Heterorhabditis_zelandica   A-AATCTATC TTCGAGTACC TAGTGGAG
Parasitorhabditis_sp_SB281  A-AACCCTTT AACGAGGATC TATGAGAG
Rhabditoides_inermiformis   A-AACCCTTT AACGAGGATC TATGAGAG
Rhabditoides_inermis_DF5001 A-AACCCTTT ATCGAGGATC TATGAGAG
Rhabditoides_regina_DF5012  A-AACCCTTT AACGATGATC AATGAGAG
Poikilolaimus_oxycerca_SB200 A-AACCCTTT AACGAGGATC TATGAGAG
Poikilolaimus_regenfussi_SB199 A-AACCCTTT AACGAGGATC TATGAGAG
Distolabrellus_veechi_DWF1604 A-AACCCTTT AACGAGGATC TATGAGAG
Distolabrellus_veechi_DF5024 A-AACCCTTT AACGAGGATC TATGAGAG
Choriorhabditis_dudichi     A-AAACACTT TATGAGGATC AAATGGAG
Protorhabditis_sp           A-AAAACTC TTCGAGAAAC TAAGAGAG
Protorhabditis_sp_DF5055    A-AATAACTT CTCGAGAAAC TAAGAGAG
Cruzanema_tripartitum_DF5015 A-AATATCTC GTAAGGAAC TAGTGGAG
Mesorhabditis_sp_PS1179     A-AACCCTTT AACGAGGATC AATGAGAG
Mesorhabditis_spiculigera_SB15 A-AACCCTTT AACGAGGATC AATGAGAG
Mesorhabditis_anisomorpha_SB12 A-AACCCTTG AACGAGGATC AATGAGAG
Teratorhabditis_palmarum_DF501 A-AACCCTTT AACGAGGATC TATGAGAG
Teratorhabditis_synpapillata_S A-AACCCTTT AACGAGGATC TATGAGAG
Caenorhabditis_briggsae_PB102 A-AATGATTC TTCGAGTATC TAGTGGAG
Caenorhabditis_elegans_N2   A-AATGATTC TTCGAGTAGC AAGGAGAG
Caenorhabditis_sp_CB5161    A-AATGATTC TTCGAGTAGC AAGGAGAG
Caenorhabditis_sp_PS1010    A-AACAATTC TTCGAGTAAC AAGGAGAG
NUCLEOTIDES INCLUDED        m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmm

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Caenorhabditis_japonica	A-AATTATTC	TTCGAGTAGC	AAGGAGAG
Caenorhabditis_sp_DF5170	A-AATCGTTC	TTCAGGTAAC	AAGGAGAG
Caenorhabditis_sp_SB341	A-AATGATTC	TTCGAGGACC	CAGGAGAG
Caenorhabditis_plicata	A-AACTAATC	TTCGAGTAAC	AAGGAGAG
Caenorhabditis_vulgaris	A-AATGATTC	TTCGAGTAGC	AAGGAGAG
Caenorhabditis_sonorae	A-AATGATTC	TTCGAGGACC	CAGGAGAG
Caenorhabditis_drosophilae	A-AATCGTTC	TTCAGGTAAC	AAGGAGAG
Phasmarhabditis_hermaphrodita	A-AATAGTTC	TTCGAGGACC	CAGTGGAG
Phasmarhabditis_neopapillosa	A-AATAGTTC	TTCGAGGACC	CAGTGGAG
Phasmarhabditis_neopapillosa	A-AATAGCTC	TTCGAGGACC	CAGTGGAG
Pellioiditis_mediterranea_SB173	A-AATAGCTC	TTCGAGGACC	CAGTGGAG
Pellioiditis_marina	A-AATAGCTC	TTCGAGGACC	CAGTGGAG
Prodontorhabditis_wirthi	A-AAAATATC	TCCGAGAAAC	TAAGAGAG
Crustorhabditis_scanica	A-AACCCCTT	AACGAGGATC	TATGAGAG
Dolichorhabditis_sp_CEW1	A-AATAGCTC	TATGAGGACC	TAATGGAG
Oscheius_sp_DF5000	A-AATAGCTC	TATGATGATC	TAGTGGAG
Oscheius_sp_BW282	A-AATAGCTC	TTCGAGGATC	TAGTGGAG
Oscheius_myriophila_EM435	A-AATAGCTC	TTCGAGGATC	TAGTGGAG
Oscheius_insectivora	A-AATAGCTG	TTCGAGGACC	TAGTGGAG
Oscheius_dolichuroides	A-AATAGCTC	TATGATGATC	TAGTGGAG
Rhabditis_blumi_DF5010	A-AACAGCTC	TTTGAGTAAC	AAGTGGAG
Rhabditis_sp_PS1191	A-AATAGCTC	TTTGAGTAAC	TAGTGGAG
Rhabditis_sp_PS1010	A-AACAATTC	TTCGAGTAAC	AAGGAGAG
Rhabditis_myriophila_EM435	A-AATAGCTC	TTCGAGGATC	TAGTGGAG
Rhabditis_colombiana	A-AATAGCTC	TTCGAGGACC	TAGTGGAG
Rhabditella_axei_DF5006	A-AATAGCTC	TTCGAGGACC	CAGTGGAG
Rhabditella_sp_DF5044	A-AATAGCTC	TTCGAGGACC	CAGTGGAG
Cuticularia_sp_PS2083	A-AACCCCTT	AACGAGGATC	TATGAGAG
Necator_americanus	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Kalicephalus_cristatus	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Ancylostoma_caninum	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Angiostrongylus_cantonensis	A-AACCTTTT	TTCGAGTATC	CAGTGGAG
Angiostrongylus_costaricensis	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Angiostrongylus_malaysiensis	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Angiostrongylus_dujardini	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Angiostrongylus_vasorum	A-AACCTTTT	TTCGAGTACC	AAGTGGAG
Aulurostrongylus_abstrusus	A-AACTTCTT	TTCGAGTATC	AAGTGGAG
Didelphostongylus_hayesi	A-AACATATC	TTCGAGTATC	AAGTGGAG
Crenosoma_mephitidis	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Crenosoma_vulpis	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Otostrongylus_circumlitus	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Troglostongylus_wilsoni	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Filaroides_martis	a-AACCTTTT	TTCGAGTATC	AAGTGGAG
Oslerus_osleri	A-AACATTTT	TTCAGTATC	AAGTGGAG
Parafilaroides_decorus	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Metastrongylus_salmi	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Metastrongylus_elongatus	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Skrjabinogylus_chitwoodrum	A-AACATTTT	TTCGAGGATC	AAGTGGAG
Parelaphostongylus_odocoilei	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Muellerius_capillaris	A-AACGTTTT	TTCGAGTATC	AAGTGGAG
Protostrongylus_rufescens	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Halocercus_invaginatus	A-AACTATTT	TTCGAGTATC	GAGTGGAG
Pseudalius_inflexus	A-AACTTTTT	TTCGAGTATC	AAGTGGAG
Stenurus_minor	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Torynurus_convolutus	A-AACCTTTT	TTCGAGTATC	AAGTGGAG
Syngamus_trachea	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Stephanurus_dentatus	A-AACTTTTT	TTCGAGGATC	AAGTGGAG
Nematodirus_battus	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Strongylus_equinus	A-AACCTCTT	TTCGAGGATC	AAGTGGAG
Labiostrongylus_bipapillosus	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Petrovina_poculatum	A-AACCTCTT	TTCGAGGATC	AAGTGGAG
Cylicocyclus_insignis	A-AACCTCTT	TTCGAGGATC	AAGTGGAG
Chabertia_ovina	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Cyclodontostomum_purvisi	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Zoniolaimus_mawsonae	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Hypodontus_macropi	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Deletrocephalus_dimidiatus	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Dictyocaulus_eckerti_P7B8	A-AACGTTTT	TTCGAGTATC	GAGTGGAG
Dictyocaulus_capreolus_P3B2	A-AACGTTTT	TTCGAGTATC	GAGTGGAG
Dictyocaulus_sp._P6A1	A-AACGTTTT	TTCGAGTATC	GAGTGGAG
Dictyocaulus_filaria	A-AACCATTT	TTCGAGTATC	AAGTGGAG
Dictyocaulus_capreolus_P2C10	A-AACGTTTT	TTCGAGTATC	GAGTGGAG
Dictyocaulus_viviparus	A-AACGTTTT	TTCGAGTATC	GAGTGGAG
Haemonchus_sp._V3091	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Haemonchus_contortus	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Haemonchus_placei	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Haemonchus_similis	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
Ostertagia_ostertagi	A-AACCTTTT	TTCGAGGATC	AAGTGGAG
NUCLEOTIDES INCLUDED	m-mmmmmmmmm	mmmmmmmmmmmm	mmmmmmmmmm

	730 740
Ostertagia leptospicularis	A-AACCTTTT TTCGAGGATC AAGTGGAG
Nippostrongylus brasiliensis	A-AACCTTTT TTCGAGGAAC AAGTGGAG
Heligmosomoides polygyrus	A-AACCTTTT TTCGAGGATC CAGTGGAG
Trichostrongylus colubriformis	A-AACCTTTT TTCGAGGATC AAGTGGAG
Tetrabothriostrongylus mackerr	A-AACCTTTT TTCGAGGATC AAGTGGAG
Herpestrongylus pythonis	A-AACCTTTT TTCGAGGATC AAGTGGAG
Nicollina cameroni	A-AACCTTTT TTCGAGGATC AAGTGGAG
Filarimena flagrifer	A-AACCTTTT TTCGAGGATC AAGTGGAG
Amidostomum cygni	A-AAACTTTT TTCGAGGATC AAGTGGAG
Teratocephalus lirellus	A-AACCTTTT AACGAGGATC TATGAGAG
Brumptaemilius justini	A-AACCCGTT AACGAGGATC TATGAGAG
Anguillicola crassus	A-AACCTTTT AACGAGGATC TATGAGAG
Dracunculus medinensis	A-AACCTGTT AACGAGGACC TATGAGAG
Dracunculus oesophageus	A-AACCTGTT AACGAGGATC TATGAGAG
Dracunculus sp. V3104	A-AACCTGTT AACGAGGATC TATGAGAG
Philonema sp. A	A-AACCCGTT AACGAGGATC TATGAGAG
Philometra obturans	A-AACCCGTT AACGAGGATC AATGAGAG
Camallanus oxycephalus	A-AAGCCGTT AACAGGATC TATGAGAG
Acanthocheilonema viteae	A-AACCTGTT AACGAGGATC TATGAGAG
Loa loa	A-AACCCGTT AACGAGGATC TATGAGAG
Onchocerca cervicalis	A-AACCCGTT AACGAGGATC TATGAGAG
Dirofilaria immitis	A-AACCTGTT AACGAGGATC TATGAGAG
Brugia malayi	A-AACCTGTT AACGAGGATC TATGAGAG
Wuchereria bancrofti	A-AACCCGTT AACGAGGATC TATGAGAG
Litomosoides sigmodontis	A-AACCTGTT AACGAGGATC TATGAGAG
Setaria digitata	A-AACCCGTT AACGAGGATC TATGAGAG
Gnathostoma turgidum	A-AACCTCTT AACGAGGATC TATGAGAG
Gnathostoma neoprocyonis	A-AACCTCTT AACGAGGATC TATGAGAG
Gnathostoma binucleatum	A-AACCTCTT AACGAGGATC TATGAGAG
Physaloptera alata	A-AACCCGTT AACGAGGATC TATGAGAG
Physaloptera turgida	A-AACCCGTT AACGAGGATC TATGAGAG
Ascarophis arctica	A-AACCCGTT AACGAGGATC TATGAGAG
Spinitectus carolini	A-AACCCGTT AACGAGGATC TATGAGAG
Thelazia lacrymalis	A-AACCTGTT AACGAGGATC TATGAGAG
Aphelenchus avenae	A-AACCTTTT AACGAGTATC TATGAGAG
Aphelenchoides fragariae	A-AACAATTT CATGAGTACC TACGAGAG
Bursaphelenchus sp	A-GACATTTT AACGAGTATC TACGAGAG
Deladenus sp	A-AACCTTTT AACGAGTATC TATGAGAG
Criconema sp	A-AACCTTTT AACGAGTATC AATGAGAG
Hemicycliophora conida	A-AACCTTTT AACGAGTATC AATGAGAG
Paratylenchus dianthus	A-AACCTTTT AACGAGTATC AATGAGAG
Pratylenchus thornei	A-AACCTTTT AACGAGTATC TATGAGAG
Tylenchulus semipetrans	A-AACCTTTT AACGAGTATC AATGAGAG
Ditylenchus angustus	A-AACCTTTT GACGAGTATC TATGAGAG
Subanguina radicola	A-AACCTTTT AACGAGTATC TATGAGAG
Tylenchorhynchus maximus	A-AACCTTTT AACGAGTATC TATGAGAG
Geocenamus quadrifer	A-AACCTTTT AACGAGTATC TATGAGAG
Globodera pallida	A-AACCTTTT AACGAGTATC TATGAGAG
Scutellonema bradys	A-AACCTTTT AACGAGTATC TATGAGAG
Helicotylenchus dihystrera	A-AACCTTTT AACGAGTATC TATGAGAG
Rotylenchus robustus	A-AACCTTTT AACGAGTATC TATGAGAG
Meloidogyne arenaria	A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne incognita_KT	A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne javanica	A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne artiellia	A-AACCTTTT AACGAGTATC TACGAGAG
Meloidogyne duytsi	A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne exigua	A-AACCTTTT AACGAGTATC AAGGAGAG
Meloidogyne hapla	A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne ichinohei	A-AATCCTTT ATCGAGTATC TACGAGAG
Meloidogyne maritima	A-AACCTTTT AACGAGTATC AAGCAGAG
Meloidogyne microtyla	A-AACCTTTT AACGAGTATC AAGCAGAG
Nacobbus aberrans	A-AACCTTTT AACGAGTATC TATGAGAG
Pratylenchoides ritteri	A-AACCTTTT AACGAGTATC TATGAGAG
Pratylenchoides magnicauda	A-AACCTTTT AACGAGTATC TATGAGAG
Hirschmanniella sp. JH_2003	A-AACCTTTT AACGAGTATC TATGAGAG
Radopholus similis	A-AAACCCCTT ATTGAGTATC TACGAGAG
Pratylenchus goodeyi_VF	A-AACCTTTT AACGAGTATC TATGAGAG
Boleodorus thylactus_clone2	A-AACCTTTT AACGAGTATC TATGAGAG
Philippine Sequence 1	A-AACCTTTT TTCGAGTATC CAGTGGAG
Philippine Sequence 2	A-AACCTTTT TTCGAGTATC CAGTGGAG
Philippine Sequence 3	A-AATAGCTC TTGGATGACC CAG....
Philippine Sequence 4	A-AAACCATT AAAGAGTATC AATGAGAG
Philippine Sequence 5	A-AACCTTTT TTCGAGGATC AAG....
Philippine Sequence 6	A-AATAGCTC TACGAGGACC CAGTGGAG
Philippine Sequence 7	A-AATGATTC TTCGAGTATC AAG....
Tahiti Sequence 1	A-AACCTTTT TTCGAGTATC CAGTGGAG
Tahiti Sequence 2	A-AATCTTTT CTTGAGTATC CAG....
Thailand Sequence 1	A-AATCCGTT AACGAGGATC TATGAGAG
NUCLEOTIDES INCLUDED	m-mmmmmmmmm mmmmmmmmmmm mmmmmmmmm

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      ....|....| ....|....| ....|...
      730      740
Ogasawara Sequence 1 A-AAACCATT AAAGAGTATC AATGAGAG
Ogasawara Sequence 2 A-AATAGTTC TTCGAGGACC CAGTGAGG
Ivory Coast Sequence 1 A-AACCTTTT TTCGAGTATC AAGTGGAG
Ivory Coast Sequence 2 A-AACCTTTT TTCGAGTATC AAGTGGAG
Ivory Coast Sequence 3 A-AACCTTTT TTCGAGTATC GAATGGAG
Ivory Coast Sequence 4 A-AACCTTTT TTCGAGTATC AAGTGGAG
Ivory Coast Sequence 5 A-AATCCGTT AACGAGGATC TATGAGAG
Ivory Coast Sequence 6 A-AACCTTTT TTCGAGTATC AAGTGGAG
Ivory Coast Sequence 7 A-AACCTTTT TTCGAGTATC AAGTGGAG
Singapore Sequence 1 A-AATAGCTC TTGAT-GACC CAGTGGAG
Singapore Sequence 2 A-AATCCGTT AACGAGGATC .....
Singapore Sequence 3 A-AATGATTG TTCGAGTAGC AAGGAGAG
Singapore Sequence 4 A-AACCTTTT TTCGAGTATC AAGTGGAG
Nigerian Sequence 1 A-AACCTTTT TTCGAGTATC AAGTGGAG
Nigerian Sequence 2 A-AACCTTTT TTCGAGGATC AAGTGGAG
Nigerian Sequence 3 A-AACCTTTT TTCGAGTATC AAGTGGAG
NUCLEOTIDES INCLUDED m-mmmmmmmmm mmmmmmmmmmm mmm.....

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Appendix 6.3: Log likelihood scores of the different models used for comparison for the (A) the Nematoda and (B) the Rhabditida within Nematoda using the SSU rRNA gene to determine the phylogenetic positions of the nematode sequences from a trial population of *Achatina fulica* and *Laevicaulis alte* in the Philippines. A total of 376 unambiguously aligned nucleotide sites of the 5' end of the SSU rRNA gene were used. Log Likelihood Ratio Tests for both the Nematoda and the Rhabditida revealed GTR+ Γ as the optimal model. Note that the K2P and F81 models are non-nested and therefore cannot be compared using LRT.

(A) Nematoda

(B) Rhabditida

Model	Log-Likelihood Score	Model	Log-Likelihood Score
JC	21909.35435	JC	14354.91018
JC+ Γ	19844.13051	JC+ Γ	13100.12574
F81	21.855.63587	F81	14325.96300
F81+ Γ	19747.80473	F81+ Γ	13031.31768
K2P	21623.78574	K2P	14234.74221
K2P+ Γ	19518.62434	K2P+ Γ	12957.97008
HKY85	21549.39123	HKY	14192.26735
HKY85+ Γ	19361.80971	HKY+ Γ	12844.49245
TN93	21435.08416	TrN93	14129.30390
TN93+ Γ	19353.15979	TN93+ Γ	12846.67437
GTR	21370.61122	GTR	13930.31599
GTR+Γ	19339.54695	GTR+Γ	12725.79552

	130	140	150	160	170	180	
Angiostrongylus cantonensis	ATCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGTATAAA	CCCTGACTTT	
Angiostrongylus costaricensis	ATCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGTATAAA	CCCTGACTTT	
Angiostrongylus malaysiensis	ATCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGTATAAA	CCCTGACTTT	
Angiostrongylus dujardini	ATCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGGTTAAG	CCCTAACTTT	
Angiostrongylus vasorum	ATCT-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT	
Aelurostrongylus abstrusus	GTTT-ACCTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCAGTAGGA	CCCTGACTTT	
Didelphostrongylus hayesi	TTTG-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCAGATAAA	CTCTGACTTT	
Crenosoma mephitiditis	ATCC-ATTTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT	
Otostrongylus circumlitus	ATCC-ATTTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT	
Troglostrongylus wilsoni	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT	
Filaroides martis	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT	
Oslerus osleri	GACG-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATAAAA	CCCTGACTTT	
Parafilaroides decorus	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCTATTAAA	CCCTGACTTT	
Metastrongylus salmi	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCGATTAAA	CCCTGACTTT	
Skrjablingylus chitwoodorum	TTCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGTGATTAAA	CCCTGACTTT	
Parelaphostrongylus odocoilei	ATCCTTAATG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TATGCATAAA	CCCTGACTCT	
Muellerius capillaris	ATGA-CAATG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCAATAAAA	TCCTGACTTT	
Halocercus invaginatus	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCTATTAGA	CCCTGACTTT	
Pseudalius inflexus	ATCTCAATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TTCTATTAAA	CCCTGACTTT	
Stenurus minor	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCTATTAAA	CCCTGACTTT	
Torynurus convolutus	ATCC-AATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCTATTAAA	CCCTGACTTT	
Nematodirus battus	ATCCTTTTTG	GATAACTGCG	GCAATTCTGG	AGCTAATACA	TGCGTTTAGG	CCCTGACTTT	
Nippostrongylus brasiliensis	GTCC-TATTG	GATAACTGCG	GTAATTCTGG	AGCTAATACA	TGCAATAAAA	CCCTGACTTT	
NUCELOTIDES INCLUDED	mmmm-mmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	190	200	210	220	230	240	
Angiostrongylus cantonensis	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Angiostrongylus costaricensis	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGCAGTTT	GCTGACTCTG	
Angiostrongylus malaysiensis	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGCAGTTT	GCTGACTCTG	
Angiostrongylus dujardini	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Angiostrongylus vasorum	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Aelurostrongylus abstrusus	-CGGAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Didelphostrongylus hayesi	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Crenosoma mephitiditis	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Otostrongylus circumlitus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Troglostrongylus wilsoni	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Filaroides martis	-CGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Oslerus osleri	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Parafilaroides decorus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Metastrongylus salmi	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Skrjablingylus chitwoodorum	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Parelaphostrongylus odocoilei	GTGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Muellerius capillaris	-TGAAAGGAT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Halocercus invaginatus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Pseudalius inflexus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Stenurus minor	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Torynurus convolutus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCATTTTCG	GATGTAGTTT	GCTGACTCTG	
Nematodirus battus	-TGAAAGGGT	GCAATTATTA	GAGCAAATCA	ATCTCCTTCG	GGTGCTGTTT	GCTGACTCTG	
Nippostrongylus brasiliensis	-TGAAAGGGT	GCAATTATTA	GTGCAAACCA	ATCATTTTCG	GATGTTGTTT	GCTGACTCTG	
NUCELOTIDES INCLUDED	-mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	250	260	270	280	290	300	
Angiostrongylus cantonensis	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Angiostrongylus costaricensis	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	TGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Angiostrongylus malaysiensis	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	TGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Angiostrongylus dujardini	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Angiostrongylus vasorum	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Aelurostrongylus abstrusus	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Didelphostrongylus hayesi	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Crenosoma mephitiditis	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Otostrongylus circumlitus	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Troglostrongylus wilsoni	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Filaroides martis	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Oslerus osleri	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Parafilaroides decorus	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Metastrongylus salmi	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Skrjablingylus chitwoodorum	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Parelaphostrongylus odocoilei	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Muellerius capillaris	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Halocercus invaginatus	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Pseudalius inflexus	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Stenurus minor	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Torynurus convolutus	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	CGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Nematodirus battus	AATAATGCAG	CATATCGGCG	GCTTGTTCGC	TGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
Nippostrongylus brasiliensis	AATAACGCAG	CATATCGGCG	GCTTGTTCGC	TGATAATCCG	AAAAAGTGTC	TGCCCTATCA	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	310	320	330	340	350	360
Angiostrongylus cantonensis	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Angiostrongylus costaricensis	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Angiostrongylus malaysiensis	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Angiostrongylus dujardini	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Angiostrongylus vasorum	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Aelurostrongylus abstrusus	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Didelphostrongylus hayesi	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Crenosoma mephitiditis	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Otostrongylus circumlitus	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Troglostrongylus wilsoni	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Filaroides martis	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Oslerus osleri	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Parafilaroides decorus	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Metastrongylus salmi	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Skrjabinngylus chitwoodorum	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Parelaphostrongylus odocoilei	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Muellerius capillaris	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Halocercus invaginatus	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Pseudalius inflexus	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Stenurus minor	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Torynurus convolutus	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Nematodirus battus	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
Nippostrongylus brasiliensis	ACCTGATGGT	AGTCTATTAG	TCTACCATGG	TTATTACGGG	TAACGGAGAA	TAAGGGTTCC	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	370	380	390	400	410	420
Angiostrongylus cantonensis	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Angiostrongylus costaricensis	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Angiostrongylus malaysiensis	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Angiostrongylus dujardini	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Angiostrongylus vasorum	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Aelurostrongylus abstrusus	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Didelphostrongylus hayesi	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Crenosoma mephitiditis	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Otostrongylus circumlitus	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Troglostrongylus wilsoni	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Filaroides martis	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Oslerus osleri	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Parafilaroides decorus	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Metastrongylus salmi	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Skrjabinngylus chitwoodorum	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Parelaphostrongylus odocoilei	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Muellerius capillaris	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Halocercus invaginatus	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Pseudalius inflexus	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Stenurus minor	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Torynurus convolutus	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Nematodirus battus	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
Nippostrongylus brasiliensis	ACTCCGGAGA	GGGAGCCTTA	GAAACGGCTA	CCACATCCAA	GGAAGGCAGC	AGGC CGGAAA	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	430	440	450	460	470	480
Angiostrongylus cantonensis	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Angiostrongylus costaricensis	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Angiostrongylus malaysiensis	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Angiostrongylus dujardini	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Angiostrongylus vasorum	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Aelurostrongylus abstrusus	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Didelphostrongylus hayesi	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Crenosoma mephitiditis	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Otostrongylus circumlitus	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Troglostrongylus wilsoni	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Filaroides martis	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Oslerus osleri	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Parafilaroides decorus	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Metastrongylus salmi	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Skrjabinngylus chitwoodorum	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Parelaphostrongylus odocoilei	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Muellerius capillaris	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Halocercus invaginatus	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Pseudalius inflexus	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Stenurus minor	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Torynurus convolutus	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Nematodirus battus	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
Nippostrongylus brasiliensis	CTTATCCAAT	CTTGAATAGA	TGAGATAGTG	ACTAAAAATA	AAAAGACCAT	TCCTATGGAA	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	490	500	510	520	530	540	
Angiostrongylus cantonensis	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Angiostrongylus costaricensis	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Angiostrongylus malaysiensis	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Angiostrongylus dujardini	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Angiostrongylus vasorum	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Aelurostrongylus abstrusus	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Didelphostrongylus hayesi	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Crenosoma mephitiditis	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	ATCCAAGTGA	GGGCAAGTCT	
Otostrongylus circumlitus	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	ATCCAAGTGA	GGGCAAGTCT	
Troglostrongylus wilsoni	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	ATCCAAGTGA	GGGCAAGTCT	
Filaroides martis	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Oslerus osleri	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Parafilaroides decorus	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Metastrongylus salmi	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Skrjablingylus chitwoodorum	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	ATCCAAGTGA	GGGCAAGTCT	
Parelaphostrongylus odocoilei	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Muellerius capillaris	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Halocercus invaginatus	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Pseudalius inflexus	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Stenurus minor	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Torynurus convolutus	CGGTTATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGT	ATCCAAGTGA	GGGCAAGTCT	
Nematodirus battus	CGGTCATTTC	AATGAGTTGA	GCTTAAACCT	TTTTTCGAGG	ATCCAAGTGA	GGGCAAGTCT	
Nippostrongylus brasiliensis	CGGTCATTTC	AATGAGTTGA	TCATAAACCT	TTTTTCGAGG	AACAAGTGA	GGGCAAGTCT	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	550	560	570	580	590	600	
Angiostrongylus cantonensis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Angiostrongylus costaricensis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Angiostrongylus malaysiensis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Angiostrongylus dujardini	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Angiostrongylus vasorum	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Aelurostrongylus abstrusus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Didelphostrongylus hayesi	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Crenosoma mephitiditis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Otostrongylus circumlitus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Troglostrongylus wilsoni	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Filaroides martis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Oslerus osleri	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Parafilaroides decorus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Metastrongylus salmi	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Skrjablingylus chitwoodorum	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Parelaphostrongylus odocoilei	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Muellerius capillaris	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Halocercus invaginatus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Pseudalius inflexus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Stenurus minor	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Torynurus convolutus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Nematodirus battus	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
Nippostrongylus brasiliensis	GGTGCCAGCA	GCCGCGGTAA	TTCCAGCTCC	ACTAGTGTA	ATCGTCATTG	CTGCGGTTAA	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	610	620	630	640	650	660	
Angiostrongylus cantonensis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Angiostrongylus costaricensis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Angiostrongylus malaysiensis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Angiostrongylus dujardini	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Angiostrongylus vasorum	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Aelurostrongylus abstrusus	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Didelphostrongylus hayesi	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	TATTAGCGTT	AATCATTGTT	
Crenosoma mephitiditis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Otostrongylus circumlitus	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Troglostrongylus wilsoni	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Filaroides martis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Oslerus osleri	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Parafilaroides decorus	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Metastrongylus salmi	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Skrjablingylus chitwoodorum	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Parelaphostrongylus odocoilei	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Muellerius capillaris	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Halocercus invaginatus	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Pseudalius inflexus	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATAATTTCG	CTTTGGCGTT	AATCATTGTT	
Stenurus minor	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Torynurus convolutus	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AATGATTTCG	CTTTGGCGTT	AATCATTGTT	
Nematodirus battus	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
Nippostrongylus brasiliensis	AAAGCTCGTA	GTTGGATCTG	AGTTGCATGC	AGTGGTTCGC	CTTTGGCGTT	AATCATTGTT	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	670	680	690	700	710	720
Angiostrongylus cantonensis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Angiostrongylus costaricensis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AACTCTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Angiostrongylus malaysiensis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGA	CTAGCGAGTT	
Angiostrongylus dujardini	GTGACTATTT	----GCTGGT	TTTCTATTGA	AACTCTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Angiostrongylus vasorum	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Aelurostrongylus abstrusus	CGGACTATTT	----GCTGGT	TTTCTGCAGA	GATTCGATT	TCTGTAGTGG	CTAGCGAGTT	
Didelphostrongylus hayesi	GTGACTAGCA	--TTGCTGGT	TTTCTATTGA	GATTCGATT	TCATTAGTGA	CTAGCGAGTT	
Crenosoma mephitiditis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Otostrongylus circumlitus	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Troglostrongylus wilsoni	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Filaroides martis	GTGACTATTT	----GCTGGT	TTTCTATTGA	AACTCTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Oslerus osleri	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGA	CTAGCGAGTT	
Parafilaroides decorus	GTGACTATTT	----GCTGGT	TTTCTATTGA	GATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Metastrongylus salmi	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Skrjabinoglyus chitwoodorum	GTGACTATTT	----GCTGGT	TTTCTATTGA	GATTCGATT	TCAATAGTGG	CTAGCGAGTT	
Parelaphostrongylus odocoilei	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTATAGTGG	CTAGCGAGTT	
Muellerius capillaris	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTATAGTGG	CTAGCGAGTT	
Halocercus invaginatus	GTGACTATCT	RT--GCTGGT	TTTCTATTGA	GATTCGATT	TCAATAGTGG	CTAGCGAGTT	
Pseudalius inflexus	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTATAGTGG	CTAGCGAGTT	
Stenurus minor	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Torynurus convolutus	GTGACTATTT	----GCTGGT	TTTCTATTGA	AATTCGATT	TCTTTAGTGG	CTAGCGAGTT	
Nematodirus battus	GTGACTATTT	----GCTGGT	TTTCTATTGA	GGTTTCGATC	TCTTTAGTGG	CTAGCGAGTT	
Nippostrongylus brasiliensis	GTGACTATTT	----GCTGGT	TTTCTATTAA	AGCTTCGGCT	TTTTTAGTGG	CTAGCGAGTT	
NUCELOTIDES INCLUDED	mmmmmmmmmm	---mmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	730	740	750	760	770	780
Angiostrongylus cantonensis	TACTTTGAAT	AAATTAAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Angiostrongylus costaricensis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Angiostrongylus malaysiensis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Angiostrongylus dujardini	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGA	TCGATCATGG	
Angiostrongylus vasorum	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Aelurostrongylus abstrusus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGA	TCGATCATGG	
Didelphostrongylus hayesi	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Crenosoma mephitiditis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Otostrongylus circumlitus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Troglostrongylus wilsoni	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Filaroides martis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Oslerus osleri	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Parafilaroides decorus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Metastrongylus salmi	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Skrjabinoglyus chitwoodorum	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Parelaphostrongylus odocoilei	TACTTTGAAT	AAATTAAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Muellerius capillaris	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--CA	GCTTGAATGG	TCGATCATGG	
Halocercus invaginatus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTTACGT	GCTTGAATGG	TCGATCATGG	
Pseudalius inflexus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Stenurus minor	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Torynurus convolutus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Nematodirus battus	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
Nippostrongylus brasiliensis	TACTTTGAAT	AAATTAGAGT	GCTCAGAACA	AGCGTT--T	GCTTGAATGG	TCGATCATGG	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	790	800	810	820	830	840
Angiostrongylus cantonensis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Angiostrongylus costaricensis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Angiostrongylus malaysiensis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Angiostrongylus dujardini	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Angiostrongylus vasorum	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Aelurostrongylus abstrusus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Didelphostrongylus hayesi	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Crenosoma mephitiditis	AATAATAGAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Otostrongylus circumlitus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Troglostrongylus wilsoni	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Filaroides martis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Oslerus osleri	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Parafilaroides decorus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Metastrongylus salmi	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Skrjabinoglyus chitwoodorum	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Parelaphostrongylus odocoilei	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Muellerius capillaris	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Halocercus invaginatus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Pseudalius inflexus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Stenurus minor	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Torynurus convolutus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Nematodirus battus	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
Nippostrongylus brasiliensis	AATAATAAAA	GAGGACTTCG	GTTCTATTTA	TTGGTTCAGG	AACGAAGTA	ATGATTAAGA	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	850	860	870	880	890	900
Angiostrongylus cantonensis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Angiostrongylus costaricensis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Angiostrongylus malaysiensis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Angiostrongylus dujardini	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Angiostrongylus vasorum	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Aelurostrongylus abstrusus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Didelphostrongylus hayesi	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Crenosoma mephitiditis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Otostrongylus circumlitus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Troglostrongylus wilsoni	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Filaroides martis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Oslerus osleri	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Parafilaroides decorus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Metastrongylus salmi	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Skrjabinstrongylus chitwoodorum	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Parelaphostrongylus odocoilei	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Muellerius capillaris	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Halocercus invaginatus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Pseudalius inflexus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Stenurus minor	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Torynurus convolutus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Nematodirus battus	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
Nippostrongylus brasiliensis	GGGACAATTC	GGGGGCATTC	GTATCCCTGC	GCGAGAGGTG	AAATTCGTGG	ACCGCAGGGG	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	910	920	930	940	950	960
Angiostrongylus cantonensis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Angiostrongylus costaricensis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Angiostrongylus malaysiensis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Angiostrongylus dujardini	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Angiostrongylus vasorum	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Aelurostrongylus abstrusus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Didelphostrongylus hayesi	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Crenosoma mephitiditis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Otostrongylus circumlitus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Troglostrongylus wilsoni	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Filaroides martis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Oslerus osleri	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Parafilaroides decorus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Metastrongylus salmi	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Skrjabinstrongylus chitwoodorum	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Parelaphostrongylus odocoilei	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Muellerius capillaris	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Halocercus invaginatus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Pseudalius inflexus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Stenurus minor	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Torynurus convolutus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Nematodirus battus	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
Nippostrongylus brasiliensis	GACGCCCTAA	AGCGAAAGCA	TTTGCCAAGA	ATGTCCTCAT	TAATCAAGAA	CGAAAGTCAG	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	970	980	990	1000	1010	1020
Angiostrongylus cantonensis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Angiostrongylus costaricensis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Angiostrongylus malaysiensis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Angiostrongylus dujardini	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Angiostrongylus vasorum	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Aelurostrongylus abstrusus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Didelphostrongylus hayesi	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Crenosoma mephitiditis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Otostrongylus circumlitus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Troglostrongylus wilsoni	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Filaroides martis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Oslerus osleri	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Parafilaroides decorus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Metastrongylus salmi	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Skrjabinstrongylus chitwoodorum	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Parelaphostrongylus odocoilei	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Muellerius capillaris	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Halocercus invaginatus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Pseudalius inflexus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Stenurus minor	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Torynurus convolutus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Nematodirus battus	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
Nippostrongylus brasiliensis	AGGTTTCGAAG	GCGATTAGAT	ACCGCCCTAG	TTCTGACCGT	AAACTATGCC	ATCTAGCGAT	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1030	1040	1050	1060	1070	1080
Angiostrongylus cantonensis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Angiostrongylus costaricensis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Angiostrongylus malaysiensis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Angiostrongylus dujardini	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Angiostrongylus vasorum	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Aelurostrongylus abstrusus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Didelphostrongylus hayesi	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Crenosoma mephitiditis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Otostrongylus circumlitus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Troglostrongylus wilsoni	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Filaroides martis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Oslerus osleri	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Parafilaroides decorus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Metastrongylus salmi	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Skrjabinoglyus chitwoodorum	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Parelaphostrongylus odocoilei	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Muellerius capillaris	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Halocercus invaginatus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Pseudalius inflexus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Stenurus minor	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Torynurus convolutus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Nematodirus battus	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
Nippostrongylus brasiliensis	CCGATGGGGT	ATTGTTGCCT	TGTCGAGGAG	CTTCCCAGAA	ACGAAAGTCT	TTCGGTTCCT
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1090	1100	1110	1120	1130	1140
Angiostrongylus cantonensis	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Angiostrongylus costaricensis	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Angiostrongylus malaysiensis	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Angiostrongylus dujardini	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Angiostrongylus vasorum	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Aelurostrongylus abstrusus	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Didelphostrongylus hayesi	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Crenosoma mephitiditis	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Otostrongylus circumlitus	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Troglostrongylus wilsoni	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Filaroides martis	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Oslerus osleri	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Parafilaroides decorus	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Metastrongylus salmi	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Skrjabinoglyus chitwoodorum	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Parelaphostrongylus odocoilei	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Muellerius capillaris	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Halocercus invaginatus	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Pseudalius inflexus	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Stenurus minor	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Torynurus convolutus	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Nematodirus battus	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
Nippostrongylus brasiliensis	GGGGTAGTAT	GTTTGCAAAG	CTGAAACTTA	AAGAAATGGA	CGGAATGGCA	CCACCAGGAG
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

	1150	1160	1170	1180	1190	1200
Angiostrongylus cantonensis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Angiostrongylus costaricensis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Angiostrongylus malaysiensis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Angiostrongylus dujardini	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Angiostrongylus vasorum	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Aelurostrongylus abstrusus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Didelphostrongylus hayesi	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Crenosoma mephitiditis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Otostrongylus circumlitus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Troglostrongylus wilsoni	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Filaroides martis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Oslerus osleri	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Parafilaroides decorus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Metastrongylus salmi	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Skrjabinoglyus chitwoodorum	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Parelaphostrongylus odocoilei	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Muellerius capillaris	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Halocercus invaginatus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Pseudalius inflexus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Stenurus minor	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Torynurus convolutus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Nematodirus battus	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
Nippostrongylus brasiliensis	TGGAGCCTGC	GGCTTAATTT	GACTCAACAC	GGGAAAACCTC	ACCCGGCCCG	GACACCGTAA
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm

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1210      1220      1230      1240      1250      1260
Angiostrongylus cantonensis  GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Angiostrongylus costaricensis GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Angiostrongylus malaysiensis GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Angiostrongylus dujardini    GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Angiostrongylus vasorum     GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Aelurostrongylus abstrusus  GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Didelphostrongylus hayesi   GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Crenosoma mephitiditis     GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Ostrostrongylus circumlitus GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Troglstrongylus wilsoni    GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Filaroides martis         GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Oslerus osleri           GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Parafilaroides decorus    GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Metastrongylus salmi      GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Skrjablingylus chitwoodorum GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Parelaphostrongylus odocoilei GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Muellerius capillaris     GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Halocercus invaginatus    GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Pseudalius inflexus      GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Stenurus minor           GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Torynurus convolutus     GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Nematodirus battus       G-ATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
Nippostrongylus brasiliensis GGATTGACAG ATTGAAAGCT CTTTCTCGAT TTGGTGGTTG GTGGTGCATG GCCGTTCTTA
NUCELOTIDES INCLUDED     mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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1270      1280      1290      1300      1310      1320
Angiostrongylus cantonensis  GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Angiostrongylus costaricensis GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Angiostrongylus malaysiensis GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Angiostrongylus dujardini    GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Angiostrongylus vasorum     GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Aelurostrongylus abstrusus  GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Didelphostrongylus hayesi   GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Crenosoma mephitiditis     GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Ostrostrongylus circumlitus GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Troglstrongylus wilsoni    GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Filaroides martis         GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Oslerus osleri           GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Parafilaroides decorus    GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Metastrongylus salmi      GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Skrjablingylus chitwoodorum GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Parelaphostrongylus odocoilei GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Muellerius capillaris     GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Halocercus invaginatus    GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Pseudalius inflexus      GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Stenurus minor           GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Torynurus convolutus     GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Nematodirus battus       GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
Nippostrongylus brasiliensis GTTGGTGGAG CGATTTGTCT GGTTTATTCC GATAACGAGC GAGACTCTAG CCTGCTAAAT
NUCELOTIDES INCLUDED     mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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1330      1340      1350      1360      1370      1380
Angiostrongylus cantonensis  AGTGACTAGA TTATTGA--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Angiostrongylus costaricensis AGTGACTAGA TTAGTGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Angiostrongylus malaysiensis AGTGACTAGA TTAGTGA--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Angiostrongylus dujardini    AGTGACTAGA TTAGTGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Angiostrongylus vasorum     AGTGACTAGA TTATTGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Aelurostrongylus abstrusus  AGTGACTAGA TTTTAC--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Didelphostrongylus hayesi   AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Crenosoma mephitiditis     AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Ostrostrongylus circumlitus AGTGACAAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Troglstrongylus wilsoni    AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Filaroides martis         AGTGACTAGA TTTTGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Oslerus osleri           AGTGACTAGA TTTTAC--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Parafilaroides decorus    AGTGACTAGA TTTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Metastrongylus salmi      AGTGACTAGA TTTTAC--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Skrjablingylus chitwoodorum AGTGACTAGA TTAGTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Parelaphostrongylus odocoilei AGTGACTAGA TACGTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Muellerius capillaris     AGTGACTAGA TTCTTAT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Halocercus invaginatus    AGTGACTAGA TTCTTTGTG TGCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Pseudalius inflexus      AGTGACTAGA TTTTGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Stenurus minor           AGTGACTAGA TTTTGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Torynurus convolutus     AGTGACTAGA TTTTGT--- -GTCTAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Nematodirus battus       AGTGCTGGA TTTTAC--- -GTCCAGTCT ACTTCTTAGA GGGATAAGCC GTGTTTA-GC
Nippostrongylus brasiliensis AGTGGCTGAA TTTTCAT--- -GTTTCAGTCT ACTTCTTAGA GGGATAAGCC GTGACTA-GC
NUCELOTIDES INCLUDED     mmmmmmmmmmm mmmmmmmmm--- -mmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm mmmmmmmmmmm

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1390 1400 1410 1420 1430 1440
Angiostrongylus cantonensis CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Angiostrongylus costaricensis CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Angiostrongylus malaysiensis CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Angiostrongylus dujardini CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Angiostrongylus vasorum CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Aelurostrongylus abstrusus CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Didelphostrongylus hayesi CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Crenosoma mephitiditis CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Ostrostrongylus circumlitus CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Troglstrongylus wilsoni CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Filaroides martis CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Oslerus osleri CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Parafilaroides decorus CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Metastrongylus salmi CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Skrjabinngylus chitwoodorum CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Parelaphostrongylus odocoilei CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Muellerius capillaris CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Halocercus invaginatus CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Pseudalius inflexus CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Stenurus minor CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Torynurus convolutus CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Nematodirus battus CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
Nippostrongylus brasiliensis CGCACGAGAT TGAGCGATAA CAGGTCTGTG ATGCCCTTAG ATGTCCGGGG CTGCACGGCG
NUCELOTIDES INCLUDED

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1450 1460 1470 1480 1490 1500
Angiostrongylus cantonensis GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Angiostrongylus costaricensis GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Angiostrongylus malaysiensis GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Angiostrongylus dujardini GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Angiostrongylus vasorum GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Aelurostrongylus abstrusus GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Didelphostrongylus hayesi GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Crenosoma mephitiditis GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Ostrostrongylus circumlitus GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Troglstrongylus wilsoni GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Filaroides martis GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Oslerus osleri GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Parafilaroides decorus GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Metastrongylus salmi GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Skrjabinngylus chitwoodorum GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Parelaphostrongylus odocoilei GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Muellerius capillaris GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Halocercus invaginatus GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Pseudalius inflexus GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Stenurus minor GGTACAATGG AAGATGCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Torynurus convolutus GGTACAATGG AAGATGCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Nematodirus battus GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
Nippostrongylus brasiliensis GGTACAATGG AAGAATCAGC TGGCCTATCC ATTGCCGAAA GGTATTGGTA AACCGTTGAA
NUCELOTIDES INCLUDED

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1510 1520 1530 1540 1550 1560
Angiostrongylus cantonensis ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Angiostrongylus costaricensis ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Angiostrongylus malaysiensis ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Angiostrongylus dujardini ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Angiostrongylus vasorum ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Aelurostrongylus abstrusus ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Didelphostrongylus hayesi ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Crenosoma mephitiditis ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Ostrostrongylus circumlitus ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Troglstrongylus wilsoni ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Filaroides martis ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Oslerus osleri ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Parafilaroides decorus ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Metastrongylus salmi ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Skrjabinngylus chitwoodorum ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Parelaphostrongylus odocoilei ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Muellerius capillaris ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Halocercus invaginatus ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Pseudalius inflexus ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Stenurus minor ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Torynurus convolutus ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Nematodirus battus ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
Nippostrongylus brasiliensis ACTCTTCCGT GACCCGGGATA GGAATTGTA ATTATTTCCC TTGAACGAGG AATTCCTAGT
NUCELOTIDES INCLUDED

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	1570	1580	1590	1600	1610	1620
Angiostrongylus cantonensis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Angiostrongylus costaricensis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Angiostrongylus malaysiensis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Angiostrongylus dujardini	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Angiostrongylus vasorum	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Aelurostrongylus abstrusus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Didelphostrongylus hayesi	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Crenosoma mephitiditis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Otostrongylus circumlitus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Troglostrongylus wilsoni	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Filaroides martis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Oslerus osleri	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Parafilaroides decorus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Metastrongylus salmi	AAGTGTGAGT	CATCAGCTCA	CGTTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Skrjabinstrongylus chitwoodorum	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Parelaphostrongylus odocoilei	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Muellerius capillaris	AAGTGTGAGT	CATTAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Halocercus invaginatus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Pseudalius inflexus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Stenurus minor	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Torynurus convolutus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Nematodirus battus	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
Nippostrongylus brasiliensis	AAGTGTGAGT	CATCAGCTCA	CGCTGATTAC	GTCCCTGCCA	TTTGTACACA	CCGCCCGTCG	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1630	1640	1650	1660	1670	1680
Angiostrongylus cantonensis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Angiostrongylus costaricensis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Angiostrongylus malaysiensis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Angiostrongylus dujardini	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Angiostrongylus vasorum	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCAGGTC	
Aelurostrongylus abstrusus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Didelphostrongylus hayesi	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TATATTGAGG	CCTTCGGGTC	
Crenosoma mephitiditis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATTGAGG	CCTTCGGGTC	
Otostrongylus circumlitus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATTGAGG	CCTTCGGGTC	
Troglostrongylus wilsoni	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATTGAGG	CCTTCGGGTC	
Filaroides martis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Oslerus osleri	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Parafilaroides decorus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Metastrongylus salmi	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Skrjabinstrongylus chitwoodorum	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATTGAGG	CCTTCGGGTC	
Parelaphostrongylus odocoilei	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Muellerius capillaris	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Halocercus invaginatus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Pseudalius inflexus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Stenurus minor	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Torynurus convolutus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTAC	TGTATTGAGG	CCTTCGGGTC	
Nematodirus battus	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATCGAGG	CCTTCGGGTC	
Nippostrongylus brasiliensis	CTGTCCGGGA	CTGAGCTGTC	TCGAGAGGAC	TGCGGACTGC	TGTATCGAGG	CCTTCGGGTC	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	mmmmmmmmmm	

	1690	1700	1710	1720	1730	1740
Angiostrongylus cantonensis	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Angiostrongylus costaricensis	GCGATATAGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Angiostrongylus malaysiensis	GCAATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Angiostrongylus dujardini	GCGATATAGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Angiostrongylus vasorum	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	G.....	
Aelurostrongylus abstrusus	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Didelphostrongylus hayesi	GTGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Crenosoma mephitiditis	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Otostrongylus circumlitus	GTGGTATAGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Troglostrongylus wilsoni	GCGGTATAGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Filaroides martis	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCT-GAA	CCGGG-AAAA	GTCGT-AAACA	
Oslerus osleri	GTGATATGGT	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Parafilaroides decorus	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCG.....	
Metastrongylus salmi	GCGGTATGGT	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Skrjabinstrongylus chitwoodorum	GCGATATAGT	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Parelaphostrongylus odocoilei	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-A...	
Muellerius capillaris	GCGATATGGC	GGGAAA-CAG	TT.....	
Halocercus invaginatus	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	G.....	
Pseudalius inflexus	GTGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Stenurus minor	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Torynurus convolutus	GCGATATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Nematodirus battus	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	GTCGT-AAACA	
Nippostrongylus brasiliensis	GCGGTATGGC	GGGAAA-CAG	TTCAATCGCA	ATGGCTTGAA	CCGGGTAAAA	-TCGT-AAACA	
NUCELOTIDES INCLUDED	mmmmmmmmmm	mmmmmmmmmm	mm.....	

	1750 1760 1770
Angiostrongylus cantonensis	AGGTATCTG.
Angiostrongylus costaricensis	AGGTATCTG.
Angiostrongylus malaysiensis	AGGTATCTG.
Angiostrongylus dujardini	AGGTATCTG.
Angiostrongylus vasorum
Aelurostrongylus ABSTRUSUS	AGGTATCTGT AGGTGAACCT GCAGATGGAT CATCG
Didelphostrongylus hayesi	AGGTATCT.. ..
Crenosoma mephitiditis	AGGTATCT.. ..
Otoststrongylus circumlitus	AGGTATCTGT AGGTGAACCT GG.....
Troglostrongylus wilsoni	AGG.....
Filaroides martis	AG.....
Oslerus osleri	AGGTATCTGT AGGT.....
Parafilaroides decorus
Metastrongylus salmi	AGGTAT... ..
Skrjabingylus chitwoodorum	AGGTATCTGT AGG.....
Parelaphostrongylus odocoilei
Muellerius capillaris
Halocercus invaginatus
Pseudalius inflexus	AG.....
Stenurus minor	AGGTAT... ..
Torynurus convolutus	AGG.....
Nematodirus battus	AGGTATCTGT AGGTGAACCT GCAGATGGAT CATCG
Nippostrongylus brasiliensis	AGGTATCTGT AGGTGAACTG
NUCELOTIDES INCLUDED

Appendix 6.5: Log likelihood scores of the different models used for comparison for the Sub-order Metastrongylina using 1628 unambiguously aligned nucleotide sites from the near full-length SSU rRNA gene. Log Likelihood Ratio Tests revealed GTR+ Γ as the optimal model. Note that the K2P and F81 models are non-nested and therefore cannot be compared using LRT.

Model	Log-Likelihood Score
JC	4572.59040
JC+ Γ	4273.02522
F81	4555.55232
F81+ Γ	4256.28503
K2P	4495.17230
K2P+ Γ	4193.80011
HKY	4477.60518
HKY+ Γ	4172.09473
TN93	4475.64206
TN93+ Γ	4172.09461
GTR	4455.11204
GTR+Γ	4149.04297

Appendix 7.1: Log likelihood scores of the different models used for comparison for the (A) the Nematoda and (B) the Rhabditida within Nematoda using SSU rRNA gene to determine the phylogenetic positions of the nematode type sequences from global populations of *Achatina fulica*, African populations of *Achatina* sp. and Philippine populations of *Laevicaulis alte*. A total of 376 unambiguously aligned nucleotide sites of the 5' end of the SSU rRNA gene were used. Log Likelihood Ratio Tests for both the Nematoda and the Rhabditida reveal GTR+ Γ as the optimal model. Note that the K2P and F81 models are non-nested and therefore cannot be compared using LRT.

(A) Nematoda

(B) Rhabditida

Model	Log-Likelihood Score	Model	Log-Likelihood Score
JC	22626.94349	JC	14988.30034
JC+ Γ	20485.39787	JC+ Γ	13664.52274
F81	22551.59800	F81	14962.62351
F81+ Γ	20366.41757	F81+ Γ	13565.99042
K2P	22346.17796	K2P	14843.88334
K2P+ Γ	20156.78913	K2P+ Γ	13508.78184
HKY85	22261.34758	HKY	14795.57154
HKY85+ Γ	19983.22991	HKY+ Γ	13374.22988
TN93	22163.39719	TN93	14760.22240
TN93+ Γ	19957.42496	TN93+ Γ	13388.72096
GTR	22099.17630	GTR	14645.08708
GTR+Γ	19963.29509	GTR+Γ	13341.76120