1 A maritime Antarctic study on the impact of marine vertebrates on terrestrial micro-

2 arthropods via nutrient content of vegetation

- 3 Trophic cascades in the Antarctic
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17 Abstract

Traits of primary producers associated with tissue quality are commonly assumed to play a strong control on higher trophic levels. However, this view is largely based on studies of vascular plants, and cryptogamic vegetation has received far less attention. In this study we utilised natural gradients in nutrient concentrations in cryptogams associated with the proximity of penguin colonies on a maritime Antarctic island, to quantify the impact of nitrogen content for micro-arthropod communities.

Proximity to penguin colonies increased the nitrogen concentration of cryptogams and the penguin source was confirmed by decreasing $\delta^{15}N$ values at greater distance from colonies. Micro-arthropod abundance, diversity (H') and richness declined with distance from the penguin colonies, and was positively correlated with the nitrogen concentrations of cryptogams. $\Delta^{15}N$ of micro-arthropods was positively correlated (r² = 0.865, *P* < 0.01) with $\delta^{15}N$ of the moss *Andreaea depressinervis*, indicating that penguin-derived nitrogen moves through Antarctic food webs across multiple trophic levels.

Nitrogen content of cryptogams was correlated with associated micro-arthropods indicating that biotic interactions affect community development in Antarctic terrestrial ecosystems. The spatial patterns of Antarctic biodiversity can therefore be affected by local factors, such as marine vertebrates, beyond existing latitudinal patterns of temperature and water availability.

35 Keyv

Keywords: Acari; Collembola; Isotope; Lichen; Moss; Nitrogen

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38 Introduction

Functional traits of vascular plant primary producers can be major drivers of the 39 community composition of herbivores and decomposers (Grime et al. 1996; Agrawal and 40 Fishbein 2006; Loranger et al. 2012). Nutrient content of plant tissues, in particular, plays an 41 important role in supporting the diversity and abundance of consumers (Loranger et al. 2012). 42 However, patterns described to date are primarily based on studies of vascular plants (Mattson 43 1980; Grime 2001). Cryptogams (bryophytes, lichens and algae), in contrast have received 44 less attention despite the large and even dominant role these primary producers play in many 45 ecosystems, especially at higher latitudes and altitudes, and in the global carbon cycle (Davis 46 1981; Convey 2013; Convey et al. 2014). The few studies examining cryptogam traits in 47 relation to consumption have mainly focused on the inhibitory function of secondary 48 compounds in preventing herbivory (Gauslaa 2005; Asplund and Wardle 2013). The most 49 extensive study to date on the impacts of physiological traits of lichens for invertebrate 50 51 communities found variable responses from different invertebrate groups to lichen nutrient content which was, in part, due to the variety in lichen growth forms across the different 52 growth locations that were studied (Bokhorst et al. 2015). To overcome the variability caused 53 by differences in growth form and growth location, natural gradients in intra-specific lichen 54 nutrient concentrations are required. 55

Mosses,lichens and algae are the major macroscopic primary producer components of food webs in many polar terrestrial ecosystems (Convey 2013) and can often be found in the vicinity of penguin colonies. These penguin colonies and other vertebrate aggregations along the coast of the Antarctic Peninsula and many offshore islands provide large quantities of nitrogen that encourage plant growth (Lindeboom 1984; Erskine et al. 1998). Bird-derived nitrogen sources are well known to support vigorous growth of primary producers globally

(Anderson and Polis 1999; Sanchez-Pinero and Polis 2000; Ellis 2005) and to increase 62 63 ecosystem process rates (Bokhorst et al. 2007a). The impacts generally diminish with increasing distance from the source, creating a nutrient gradient within the vegetation which 64 also includes declines in δ^{15} N (Erskine et al. 1998; Zmudczyńska et al. 2012; Crittenden et al. 65 2015). Such gradients in δ^{15} N are useful to determine whether higher trophic levels feed on 66 the penguin-affected vegetation, as $\delta^{15}N$ levels typically increase by 2-3 units per step in 67 trophic level (Peterson and Fry 1987). To date, the existence or magnitude of any impact of 68 increased nutrient content of primary producers for higher trophic levels in Antarctic 69 terrestrial ecosystems is poorly understood (Davis 1981). 70

To address these issues we tested whether proximity to penguin colonies was 71 associated with increased nitrogen concentrations and $\delta^{15}N$ values in cryptogams on Signy 72 Island (maritime Antarctic) and whether this, in turn, correlated with the community 73 composition of the most prominent elements of the terrestrial fauna, springtails and mites 74 (Convey 2013). We hypothesised that: 1) the nitrogen concentrations and $\delta^{15}N$ values of 75 cryptogams would decrease with increasing distance from penguin colonies; and 2) that, as a 76 consequence, this would be associated with decreased abundance, diversity (H') and species 77 richness of micro-arthropods living among these cryptogams. Confirmation of these 78 hypotheses would provide the first direct evidence for trophic cascades in the Antarctic 79 terrestrial environment, and add support to recent studies (Caruso et al. 2013) challenging the 80 long-held view that abiotic factors are the driving forces underlying spatial patterns and 81 processes in the Antarctic terrestrial biota (Convey 1996; Hogg et al. 2006). 82

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84 Materials and Methods

85 *Study sites*

This study was performed on Signy Island (60°17'S 45°59'W) in the maritime 86 Antarctic region north-east of the Antarctic Peninsula. The island is part of the South Orkney 87 Islands archipelago, has an annual temperature of around -2°C and receives about 400 mm yr⁻ 88 ¹ of precipitation of which most falls as snow (Walton 1982; Royles et al. 2013). Up to 50% 89 of the island's surface is free of snow and ice during summer (December-February), exposing 90 some of the best-developed and most diverse cryptogam communities in the Antarctic. There 91 are four areas hosting large penguin colonies along the coastline of Signy Island (Fig. 1). The 92 largest of these is at North Point, which is inhabited by Adélie (Pvgoscelis adeliae), chinstrap 93 (P. antarctica) and gentoo penguins (P. papua) (total of 13000 breeding pairs). The colony at 94 Gourlay Peninsula is dominated by Adélie and chinstrap penguins (12900 breeding pairs). 95 Smaller colonies are situated along the west coast of the island, with the Cummings Cove area 96 hosting 7000 chinstrap penguin pairs, and Spindrift Rocks 2000 pairs of Adélie penguins. 97

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99 *Sampling regime*

100 During December 2013 we sampled mosses, algae and lichens as close as possible to the four penguin colonies and at four additional sites at increasing distances away from each 101 colony along each of three replicate transects, with sampling points separated by at least 10 m 102 from each other. Due to the differences in vegetation development and topography between 103 the four colonies we were unable to collect all cryptogam species from each transect and 104 sampling distances from the penguin colonies differed (for details see Table 1, Fig. 1). To 105 determine the impact of cryptogam nitrogen concentrations on micro-arthropod communities 106 across larger geographical scales we also sampled lichens along the north-south axis of Signy 107 Island, thereby extending the transect's starting from the North Point penguin colony. In 108 addition, as some of the transects increased in altitude (up to 200 m asl) at greater distance 109 from the penguin colonies, we also included three transects along a hillside remote from any 110

penguin colonies in order to control for any potential effect of altitude on micro-arthropodcommunity composition (See Fig. 1).

We focussed on the dominant lichen species, *Usnea antarctica* (DuRietz), and *Umbilicaria antarctica* (Frey & I.M. Lamb), the moss *Andreaea depressinervis* (Card.) and the foliose alga *Prasiola crispa* (Lightfoot). Henceforth these species will be referred to by their genus name alone. Sampling from extensive moss carpets or turves was avoided in order to avoid artefacts caused by autocorrelation with the presence or extent of moss, which is known to be a preferred micro-habitat for many micro-arthropods (Davis 1981; Usher and Booth 1984; Bokhorst et al. 2014).

120 At each sampling site, *Prasiola* was collected with a PVC corer (7 cm diameter) including the underlying soil (at most 1 cm if any present) and stored in a plastic container. 121 Moss and lichen clumps were collected by hand and also stored in individual plastic 122 containers. Mean cryptogam dry mass was: 5.8 g (sd = 2.8) for Andreaea, 3.0 g (1.7) for 123 Umbilicaria, 3.7 g (1.6) for Usnea and 1.8 g (0.9) for Prasiola. All samples were kept in the 124 125 shade at ambient temperatures while in the field and at approximately 5°C in the dark when stored at the station until extraction (within 24 h) in a modified Tullgren extractor for 24 h. 126 Extracted Collembola and Acari were preserved in 70% ethanol and identified to species 127 level, except for the smallest Prostigmata which were grouped together. Micro-arthropod 128 abundance was expressed as the number of individuals per gram moss, algae or lichen. To 129 obtain dry vegetation mass of each sample all soil and rock (if any) were removed and the 130 remaining vegetation weighed when dry after completion of the micro-arthropod extractions. 131 Moisture content of the sample was quantified by weighing each sample before and after 132 placement in the Tullgren funnels. 133

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135 Analyses of nitrogen and $\delta^{15}N$.

Dried cryptogam samples were transported to Europe for nitrogen and stable isotope $\delta^{15}N$ 136 analyses. Nitrogen and $\delta^{15}N$ were analysed using a Fisons NA 1500 elemental analyser 137 coupled to a Finnigan conflo II interface, and a Finnigan MAT Delta S isotope ratio mass 138 spectrometer. In addition, we measured $\delta^{15}N$ values of adult individuals of the springtail 139 Cryptogpygus antarcticus (c. 20 individuals per sample; 210 µg) and the mites Alaskozetes 140 antarcticus and Halozetes belgicae (both c. 6 individuals per sample; 280 ug) to determine if 141 penguin-derived nitrogen flows from the primary producers to these grazers. The $\delta^{15}N$ 142 analyses of micro-arthropods were carried out on samples extracted from Andreaea as the 143 moss contained the highest micro-arthropod abundance. To remove any ethanol before 144 analyses the micro-arthropods were placed in pre-weighed tin cups and oven dried (60°C) for 145 24 h. To quantify the biomass of the springtails and mites in each sample the cups were 146 weighed again after drying before analysis of δ^{15} N (mass ranged from 89-612 µg/sample). 147

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149 *Calculations and statistical analyses.*

To determine differences in nitrogen concentrations, $\delta^{15}N$, water content, abundance 150 of total micro-arthropods, diversity (H') and species richness along the penguin transects we 151 used a mixed effect model with distance to the penguin colony as fixed factor and site as a 152 random factor for each of the cryptogam species separately. To compare the rate of change of 153 cryptogam traits (%N, δ^{15} N and water content) and the micro-arthropod community along 154 transects we compared the slopes of linear regression lines through the transect data points 155 using cryptogam species as fixed factor and site as a random factor. To determine the impact 156 of penguin colonies on the micro-arthropod variables along greater spatial distances we used 157 the same mixed effect model as described above but limited to Usnea using data from the 158 North Point and the N-S transects. To account for potential impacts of altitude (as a proxy for 159 temperature) differences along the transects we compared the differences in Usnea N 160

concentrations, $\delta^{15}N$, water content and micro-arthropod variables with altitude along 161 penguin-influenced transects (North Point, Cummings, and Spindrift Rocks) and non-affected 162 transects (Moraine Valley) with altitude as a fixed factor and site as random factor. 163 Correlation coefficients were calculated between the nitrogen, $\delta^{15}N$ and water content of 164 cryptogams and the micro-arthropod variables to explore which if any of these was a stronger 165 driver behind observed patterns in community composition. Abundance data were log 166 transformed to achieve homogeneity of variance. All statistical analyses were carried out 167 using SPSS 22.0 (IBM SPSS Statistics version 22.0. Armonk NY, USA). 168

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171 **Results**

172 *Cryptogam nitrogen and water content*

Nitrogen concentrations of cryptogams declined by nearly two-thirds with increasing 173 distance from penguin colonies (Fig. 2 a-d, Table 2). $\Delta^{15}N$ declined by between 5 to 15 units 174 as the distance to penguin colonies increased, although the variation between species and 175 different transects was large (Fig. 2e-h). For example, Prasiola showed no change in %N or 176 δ^{15} N along the Gourlay transect but it did along the North Point transects (Fig. 2b) while 177 Andreaea showed declines in N along transects of both sites (Fig. 2a). The horizontal extent 178 of the impact of the penguin colonies on cryptogam N concentrations and δ^{15} N was c. 700-179 1000 m for Andreaea, 500-750 m for Prasiola, 300-500 m for Usnea and 700-1500 m for 180 Umbilicaria. Mean nitrogen concentrations and $\delta^{15}N$ values were highest (F_{3,133} = 64.1 and 181 104.7 respectively; P < 0.001) for Prasiola (4.1%, 10.4%), followed by Andreaea (2.1%, 182 2.7‰), Usnea (1.1%, -2.4‰) and Umbilicaria (1.1%, -5.9‰). There were some differences in 183 water content for individual cryptogams between sites and along transects (Table 2) but not in 184 a consistent pattern along transects (Fig. S1). Mean water content was highest (ANOVA F_{3,133} 185 = 30.4 P <0.001) for Prasiola (67%), followed by Umbilicaria (60%), Andreaea (52%) and 186 187 Usnea (25%) (Table 2).

188 Micro-arthropod communities in relation to distance from penguin colonies, cryptogam
189 nitrogen and water content

There were consistent declines in Acari abundance (3.9 ind./100 m), species richness (0.2 species/100m) and diversity (H') (0.07/100 m) across cryptogam species and sites (Fig. 3, Table 2) resulting in lower (Tukey HSD P < 0.01) values at the furthest transect points compared to points closest to the penguin colonies. Change in Collembola abundance was variable between species and sites with larger changes along Gourlay compared to North
Point transects among *Prasiola* and *Umbilicaria* resulting in no overall change in Collembola
abundance along transects within the *Prasiola* samples (Fig. 3e). These variable Collembola
patterns were also responsible for the significant species-site interactions for changes in total
micro-arthropods (Table 2).

Micro-arthropod communities were positively correlated with nitrogen concentrations 199 of the cryptogam species but not with that of Prasiola (Fig. 4). Acari abundance was 200 significantly correlated with the nitrogen concentrations of all cryptogam species while 201 Collembola abundance was significantly correlated with the nitrogen concentrations of 202 Andreaea only (Fig 4a). There were positive correlations between micro-arthropod diversity 203 204 (H') and the nitrogen concentrations of the cryptogams (but not that of *Prasiola*), and of species richness and nitrogen concentrations of the two lichens (Fig. 4). $\Delta^{15}N$ for the 205 collembolan *Cryptopygus antarcticus* was positively correlated with the $\delta^{15}N$ values of 206 Andreaea (Fig. 5a), and on average 3.6% (\pm 0.47) higher than the δ^{15} N of the moss they were 207 extracted from. Positive correlations were also found for $\delta^{15}N$ of the mites *Alaskozetes* 208 antarcticus and Halozetes belgicae with that of Andreaea (Fig. 5b), and $\delta^{15}N$ was 5.8% (± 209 0.3) higher compared to the moss they were extracted from. There was no correlation between 210 micro-arthropods (total abundance, Acari and Collembola) and tissue water content within 211 cryptogam species. However, when compared across all samples (n = 166), total micro-212 arthropod and Collembola abundance were positively correlated ($r^2 = 0.15$ and 0.24 213 respectively P < 0.05) with cryptogam water content, reflecting the potential impact of the 214 215 different cryptogam growth forms and water holding capacity.

216 Micro-arthropod communities across Signy Island and in relation to altitude

There were no differences in micro-arthropod abundance, diversity (H') or species 217 richness along the N-S transect across the island (Fig. 6). Nitrogen concentrations and $\delta^{15}N$ 218 did not differ along the N-S transect and the penguin impact on Usnea N concentrations was 219 220 limited to c. 300-500 m distance (Fig. 6c). There were no correlations between Usnea N concentrations and the micro-arthropod community along the N-S transect (data not shown). 221 The penguin colony impact on micro-arthropod abundance, diversity (H') and species 222 richness in Usnea appears limited to c. 300-500 m distance as all more distant sampling sites 223 showed no further differences (Fig. 6a,b). There was no difference in the micro-arthropod 224 abundance or species richness across transects from Moraine Valley to Green Gable (Fig. 225 226 6d,e) indicating that there was no effect of altitude on the micro-arthropod community along the study transects. $\delta^{15}N$ of Usnea did not change with altitude along the Moraine Valley 227 transects (Fig. 6f) and there was no correlation with the micro-arthropod abundance or species 228 229 richness. In comparison with the penguin colony-associated transects the micro-arthropod abundance, diversity (H') and richness were consistently lower (Tukey HSD P < 0.05) along 230 the Moraine Valley transect (Table 2, Fig. 6). 231

232 *Micro-arthropod communities across cryptogam species*

Species richness was highest (Tukey HSD P < 0.01) in *Andreaea* (4 species on average) compared to the other cryptogams (2 to 3 species) (Table 2). The lowest microarthropod diversity (H') was found among *Prasiola* (mean H' of 0.3 across all samples) while the other cryptogam species had higher (Tukey HSD P < 0.05) mean values ranging from 0.5 to 0.7. In contrast, total micro-arthropod abundance was highest (Tukey HSD P < 0.05) in *Prasiola* followed by *Andreaea, Umbilicaria* and *Usnea* (data not shown). Acari abundance did not differ between cryptogam species. Collembola abundance was consistently higher 240 (Tukey HSD *P* < 0.001) in *Prasiola* compared to the other cryptogam species, and *Andreaea*241 supported more Collembola than the two lichen species.

The predatory mite *Gamasellus racovitzai* was found across all cryptogams but abundances were higher in *Andreaea* and *Prasiola* (data not shown). Abundance of the predatory mite *G. racovitzai* was positively correlated with Collembola (prey) abundance across all (n = 126) cryptogam samples along transects ($r^2 = 0.113$, P < 0.05) but this correlation improved ($r^2 = 0.585$, P < 0.01) when only the *Usnea* data (n = 42) were used, as no significant correlations were present among the other cryptogams individually (data not shown).

249

251 **Discussion**

This is one of the first studies to examine the role of cryptogam nutrient traits in 252 relation to micro-arthropod communities, and the first to do this in the Antarctic. Our data 253 provide clear support for the role of primary producers, as distinct from abiotic interactions, 254 as a factor contributing to ecosystem development in terrestrial ecosystems of the maritime 255 256 Antarctic region (Hogg et al. 2006; Caruso et al. 2013). Although the spatial distribution of Antarctic terrestrial biodiversity is heavily influenced by temperature and water availability 257 (Convey et al. 2014), marine vertebrates clearly have a strong impact at local scales (Erskine 258 et al. 1998; Ellis 2005). Our findings are consistent with general patterns observed for 259 vascular plants in more temperate regions (Loranger et al. 2012) and therefore extend general 260 understanding of the drivers of invertebrate abundance and diversity through traits of other 261 important primary producers such as cryptogams (Bokhorst et al. 2015). 262

Hypothesis 1 was supported by the decrease in nitrogen concentrations and $\delta^{15}N$ of 263 cryptogams with increasing distance from penguin colonies. These findings are consistent 264 with other studies from polar regions, where plant productivity increased closer to penguin 265 colonies (Lindeboom 1984) and the stable isotope nitrogen signature of the primary producers 266 267 was directly linked to the trophic level of the animals producing the faecal matter (Erskine et al. 1998; Crittenden et al. 2015). Like many plants in Arctic and temperate regions, Antarctic 268 primary producers also tend to be nitrogen-limited in their growth (Davey and Rothery 1992; 269 Wasley et al. 2006) and penguin colonies greatly increase the availability of nitrogen for 270 cryptogams. The direct impact of penguin colonies here on the nitrogen content (% N and 271 δ^{15} N) of cryptogams varied greatly between species and sites, ranging from 300-1500 m 272 273 distance, which is within the range of observations from Marion Island (Erskine et al. 1998),

although Crittenden et al. (2015) reported detectable impacts of penguin-derived ammonia on
lichen physiological activity at distances of up to 10 km.

The variation in N concentrations between species and sites with distance to penguin 276 colonies was most likely caused by site specific factors. For instance, N concentrations of 277 Andreaea and Prasiola both declined along North Point transects but along the Gourlay 278 279 transects this pattern was only found for Andreaea. This lack of response by Prasiola to the obvious N gradients that exist along the Gourlay transects, as shown by N concentration in 280 Andreaea and Umbilicaria, may be caused by the decaying moss bank that underlies these 281 transects (Smith 1988) from which Prasiola may receive additional N, while Andreaea and 282 Umbilicaria were sampled from rock and therefore not so open to influence by N sources 283 from below. However, there is no good explanation available for the differences in N content 284 changes between Usnea and Umbilicaria along the Cummings Cove transects. Despite Signy 285 Island being a relatively small island and nutrient inputs from sea spray and marine 286 vertebrates generally being assumed to reach across the island (Allen et al. 1967) there were 287 clear changes and limits (c. 300-1500 m horizontal distance) detected here to the impact of 288 penguin colonies on the nitrogen concentrations or $\delta^{15}N$ of cryptogams. Additional factors 289 that may have affected cryptogam δ^{15} N values include the penguin population size, dominant 290 penguin species within the colony and overall wind direction that carries the nitrogen towards 291 the vegetation. A larger comparative study than that presented here would be required to take 292 these factors into account. 293

The decreases in micro-arthropod abundance, diversity (H') and species richness as distance to the penguin colonies increased provide support for Hypothesis 2. The microarthropod community values also showed strong correlations with cryptogam nitrogen concentrations indicating that this may be a a driving force for community assembly at higher

trophic levels in these Antarctic ecosystems. Water availability and temperature are typically 298 299 considered as strong drivers of biological patterns in the Antarctic (Kennedy 1993; Convey et al. 2014), but we did not observe any effect of intra-specific cryptogam water content or site 300 301 temperature (as altitude proxy) on the micro-arthropod community along the sampling transects. Although site temperature was only studied for Usnea, the Gourlay transects 302 (including other cryptogams) showed strong declines in Acari abundance while only reaching 303 304 a third of the altitude of other transects, indicating that altitude/temperature differences along these transects play a minor role for the studied species. The current data indicate that water 305 and temperature are of less importance than nutrient availability at closer proximity (c. 500 306 307 m) to penguin colonies for Antarctic micro-arthropods at this maritime Antarctic island. This suggests that, at relatively small geographical scales, different drivers of community assembly 308 exist in maritime Antarctic terrestrial ecosystems. The existence of such smaller-scale 309 310 gradients in driving forces behind community development allows for further detailed unravelling of these factors by manipulations of nutrient availability (Wasley et al. 2006) and 311 micro-climate (Bokhorst et al. 2011). Further work should ideally also include other 312 physiological variables such as phosphorus and defence compounds (Bokhorst et al. 2015; 313 Crittenden et al. 2015). 314

The strong correlation between the δ^{15} N of the micro-arthropods and that of the moss *Andreaea* (Fig. 5) indicates that the penguin-derived nitrogen is taken up by the moss and transferred to the micro-arthropods. The mean increase from moss to springtails was 3.6‰ and is consistent with changes generally associated between trophic levels (Peterson and Fry 1987). However, with the current data it is not possible to determine if the springtails feed on the live moss, the epiphytic micro-flora living among the moss shoots, or on decaying moss shoots. The 5.8‰ increase found for the mites compared to the moss (Fig. 5b) is higher than a single trophic level step, suggesting that the mites also feed from other food sources with higher δ^{15} N values such as penguin faecal matter and dead animals (Goddard 1982; Schneider et al. 2004). Despite these uncertainties the δ^{15} N values showed a clear pattern along transects away from penguin colonies in both primary producers and consumers, with consistent declines in nitrogen concentrations and abundance, diversity and species richness of microarthropods. These results provide strong support for the existence of biotic interactions between primary producers and higher trophic levels in Antarctic terrestrial ecosystems.

The abundance of micro-arthropods, notably Collembola, was not always related to 329 the nitrogen concentrations of the primary producers and, particularly, the Prasiola 330 communities showed very poor correlations despite the large range in nitrogen concentrations 331 332 measured (N: 1.6% - 6.2%). This indicates that other characteristics of cryptogams, such as phosphorus, defence compounds against herbivory and growth form affecting the micro-333 habitat conditions, may also have played a role (Asplund and Wardle 2013; Bokhorst et al. 334 335 2015). Overall, the highest micro-arthropod abundance and diversity were found among Andreaea and Prasiola compared to the lichens. These differences most likely reflect the 336 differences in nitrogen concentrations, growth form and water content (there were significant 337 correlations with water content when compared across all sampled cryptogams, as opposed to 338 the low variation within species). The compact growth of moss shoots provides a more stable 339 micro-environment than the open structure of the lichen thallus, and moss has been shown to 340 have a strong positive effect on the abundance of micro-arthropods in other parts of the world 341 (Salmane and Brumelis 2008; Bokhorst et al. 2014). The collembolan Cryptopygus 342 antarcticus is known to have poor control over its body water content (Block and Harrisson 343 1995; Convey et al. 2003) and, therefore, would be less prone to desiccation when within the 344 moss profile. Similarly, the foliose alga Prasiola, whose 3D structure is typified by much 345

folding and many small protected cavities, provides a continuous moist environment optimal 346 347 for these micro-arthropods. The high abundance of micro-arthropods among Prasiola compared to the cryptogams was most likely driven by the preference for algae as a food 348 349 source by at least some of these micro-arthropods (Worland and Lukešová 2000; Bokhorst et al. 2007b) and the availability of optimal moisture conditions. The main driving forces 350 underlying the distribution of consumers within Antarctic terrestrial ecosystems may therefore 351 depend on the specific requirements for growth and avoidance of adverse abiotic conditions 352 of individual species of micro-arthropod. Further work comparing a larger range of 353 cryptogam growth forms than used in this study would be required to determine the role of 354 these for the invertebrate communities in the Antarctic. 355

356 The current findings indicate a strong link between the nitrogen concentrations of Antarctic primary producers such as mosses and lichens and the community composition of 357 associated micro-arthropods. These data extend the general understanding of the influence of 358 nutrient concentrations in cryptogams on community assembly of invertebrates. Although the 359 data indicate that the impact of marine vertebrates on cryptogam nutrients and micro-360 arthropod community development is important, it also declines at relatively short distances 361 (c. 300-1500 m) from the source. Micro-arthropod biodiversity patterns in the maritime 362 Antarctic are driven by availability of water and temperature when considered across large 363 geographical scales, but biotic interactions can clearly become more important at the local 364 scale. 365

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476 Figures

Figure 1. Map of Signy Island indicating the major penguin colonies. The lines represent the
sampling transects along which samples of cryptogams and micro-arthropods were collected.
The dashed line represents transects without penguin presence from the bottom of Moraine
Valley to the top of Green Gable.

Figure 2. Nitrogen concentrations (%) and $\delta^{15}N$ values of Andreaea depressinervis (a), *Prasiola crispa* (b), Usnea antarctica (c) and Umbilicaria antarctica (d) along transects away from penguin colonies on Signy Island. Note that not all species were present along each transect. Different letters indicate significant differences (Tukey HSD P < 0.05) in N or $\delta^{15}N$ values between sampling sites. For clarity these differences are only presented for one transect but the other transects show similar patterns. Data points are the mean of n = 3 with SE as error bars. •: North Point, \circ : Gourlay, ∇ : Cummings Cove, \blacksquare : Spindrift rocks.

Figure 3. Micro-arthropod abundance, diversity (H') and species richness along transects away from penguin colonies on Signy Island. Regression lines are drawn through the mean of the presented data points. Significant r^2 are indicated by: * P < 0.05, ** P < 0.01 and *** P <0.001. Data points are the mean of n = 3 with error bars as SE. •: North Point, \circ : Gourlay, $\mathbf{\nabla}$: Cummings Cove, $\mathbf{\blacksquare}$: Spindrift rocks.

Figure 4. Regressions between micro-arthropod abundance, diversity (H'), richness with the nitrogen (%) concentration of *Andreaea depressinervis, Prasiola crispa, Usnea antarctica* and *Umbilicaria antarctica*. Data points are individual moss, alga and lichen samples collected along transects away from penguin colonies. Significant r² are indicated by: * P <0.05, ** P < 0.01 and *** P < 0.001. Figure 5. Regressions between $\delta^{15}N$ of *Andreaea depressinervis* and $\delta^{15}N$ of microarthropods. a) $\delta^{15}N$ of *Cryptopgyus antarcticus* from the North Point and Gourlay transects. The upper dashed regression line through the springtail data points was 3.5‰ higher (Y = 1.028 X + 3.54) than that of the moss (Y = X). b) $\delta^{15}N$ of *Alaskozetes antarcticus* and *Halozetes belgicae* from the Gourlay transects. The upper dashed regression line through the mite data points was 5.8‰ higher (Y = 0.974 X + 6.0) than that of the moss (Y = X). Significant r² are indicated by: * P < 0.05, ** P < 0.01 and *** P < 0.001.

Figure 6. Micro-arthropod abundance and richness in Usnea antarctica across Signy Island 505 and in relation to altitude. a) Micro-arthropod abundance, b) species richness, c) Nitrogen 506 (%N) and δ^{15} N of Usnea along the North Point transects away from the penguin colony (grev 507 shaded area) and the north-south transect across Signy Island. Data points with different 508 letters are significantly different (Tukey HSD P < 0.05). Note that, for clarity, in c) only the 509 Tukey differences for $\delta^{15}N$ are shown but the %N follows the same pattern. d) Micro-510 arthropod abundance, e) species richness, f) δ^{15} N of Usnea along the slopes of the penguin-511 affected North Point transects and that of Moraine Valley (without penguins). %N of Usnea 512 shows the same pattern as for $\delta^{15}N$ but is not shown. Different letters indicate significant 513 differences (Tukey HSD P < 0.05) for each site separately. ANOVA statistics are presented in 514 Table 2. Data points are the mean of n = 3 with SE as error bars. 515

Table 1. Details of cryptogam sampling transects in relation to penguin colonies across Signy Island. For the exact location of each penguin colony and the locations of the N-S transect and Moraine Valley see Figure 1. Distances from penguin colonies for the N-S transect were measured from North Point for Robin Peak, Springtail Spur and Jane Col, while Snow Hills and Garnet Hill were measured from the Gourlay Peninsula. –: not present at site.

Sampling sites	North Point	Gourlay	Cummings	Spindrift rocks	bindrift rocks N-S transect		Moraine Valley
			altitude (m asl.)				
Penguin species	P. adeliae, P.	P. adeliae	P. antarctica	P. adeliae			
	antarctica, P. papua	P. antarctica					
Breeding pairs (n)	13000	12900	7000	2000			
1	0 (30)	0 (10)	20 (20)	150 (20)	Robin Peak	1000 (220)	40
2	300 (50)	150 (20)	100 (50)	200 (40)	Springtail Spur	1700 (160)	60
3	500 (100)	300 (20)	300 (80)	300 (60)	Jane Col	2700 (160)	90
4	700 (150)	700 (60)	600 (150)	500 (100)	Snow Hills	3600 (265)	120
5	800 (200)	1300 (70)	-	700 (150)	Garnet Hill	2100 (227)	206
Species sampled							
Andreaea	+	+					
Prasiola	+	+					
Usnea	+		+	+	+		+
Umbilicaria	+	+	+				

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Table 2. Mixed effects ANOVA results of cryptogam differences in N content, δ^{15} N, water content and micro-arthropods in relation to distance to penguin colonies. The comparison of change in arthropods along transects compares the slopes of linear regression through the data points (as shown in Fig. 3). The North-South transect statistics show the comparison of the micro-arthropod community among *Usnea* from the North Point penguin colony to the southern tip of the island (see Table 1). The altitude statistics represent a comparison of the micro-arthropod community among *Usnea* along three penguin-associated transects (see Table 1) to that of non-penguin affected transects (Moraine Valley). There were three replicate transects per site and site was used as a random factor in the analyses. Significant differences are indicated by * *P* < 0.05, ** *P* < 0.01, *** *P* < 0.001.

		Cryptogam			Micro-arthropod abundance			Species richness Diversity	
		%N	$\delta^{15}N$	Water content	Total	Acari	Collembola	R	(H')
Andreaea	Distance (8,20)	12.2 ***	5.7 ***	1.9	0.1	5.7 **	2.4	6.2 ***	7.8 ***
	Site (1,20)	14.4 ***	0.3	3.9	0.5	0.4	1.6	2.5	0.0
Prasiola	Distance (7,18)	2.0	3.1 *	2.3	2.2	2.4	2.6	2.8 *	4.3 **
	Site (1,18)	0.2	5.5 *	6.9 *	0.1	0.1	6.5 *	0.3	7.5 *
Usnea	Distance (9,30)	5.5 ***	8.4 ***	7.3 ***	13.1 ***	10.3 ***	10.9 ***	6.5 ***	3.1 **
	Site (2,30)	2.4	6.3 **	12.0 ***	2.6	2.5	1.7	2.0	0.3
Umbilicaria	Distance (10,29)	5.2 ***	1.1	3.9 **	1.5	2.3*	1.3	3.7 **	4.5 **
	Site (2,29)	9.3 **	5.6 **	36.5 ***	0.7	1.6	8.8 **	0.3	1.1
Rate of change in	micro-arthropods along tran	isects							
	Species (3 20)	1.0	1.1	10.8 *	0.7	1.0	0.2	9.0	0.8
	Site (3.20)	2.4	0.7	2.6	0.3	1.5	0.5	1.7	0.3
	Site×Species (3,20)	9.7 ***	8.9 ***	1.4	9.3 ***	1.9	10.0 ***	0.2	1.3
N-S transect acros	s Signy Island								
Usnea	Distance (8,20)	4.9 **	4.5 **	3.4 *	9.6 ***	4.4 **	18.2 ***	2.1	0.7
	Site (1,20)	0.0	1.6	4.8 *	0.1	0.2	0.7	0.1	0.6
Altitude comparis	on								
Usnea	Altitude (12,41)	6.3 ***	7.6 ***	2.7 **	8.6 ***	8.3 ***	7.3 ***	6.0 ***	2.2 *
	Site (2,41)	2.4	11.0 ***	20.1 ***	2.4	4.1 *	2.6	0.1	1.3