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Habitual physical activity is associated with the maintenance of neutrophil migratory dynamics in healthy older adults

David B. Bartlett¹, Oliver Fox¹, Clare L. McNulty², Hannah L. Greenwood¹, Laura Murphy¹, Elizabeth Sapey¹, Martin Goodman³, Nicola Crabtree⁴, Cecilie Thøgersen-Ntoumani², James P. Fisher², Anton J.M. Wagenmakers⁵ and Janet M. Lord¹

¹MRC-ARUK Centre for Musculoskeletal Ageing Research, Institute of Inflammation and Ageing, University of Birmingham, Birmingham B15 2TT.

²School of Sport, Exercise and Rehabilitation Sciences, University of Birmingham, Birmingham B15 2TT.

³Geriatric Medicine, University Hospitals Birmingham NHS Foundation Trust, Birmingham, B15 2WB.

⁴NIHR/Wellcome Trust Clinical Research Facility, Queen Elizabeth Hospital, Birmingham, B15 2WB.

⁵Research Institute for Sport & Exercise Sciences, Liverpool John Moores University, Liverpool, L3 3AF.

*Corresponding Author:

David B. Bartlett, PhD

MRC-ARUK Centre for Musculoskeletal Ageing Research

College of Medical and Dental Sciences

University of Birmingham

Birmingham

B15 2WB

Email: d.b.bartlett@bham.ac.uk

Abstract

Background: Dysfunctional neutrophils with advanced age are a hallmark of immunosenescence. Reduced migration and bactericidal activity increase the risk of infection. It remains unclear why neutrophil dysfunction occurs with age. Physical activity and structured exercise have been suggested to improve immune function in the elderly. The aim of this study was to assess a comprehensive range of neutrophil functions and determine their association with habitual physical activity.

Method: Physical activity levels were determined in 211 elderly (67 ± 5 years) individuals by 7-days of accelerometry wear. Twenty of the most physically active men and women were matched for age and gender to twenty of the least physically active individuals. Groups were compared for neutrophil migration, phagocytosis, oxidative burst, cell surface receptor expression, metabolic health parameters and systemic inflammation. Groups were also compared against ten young participants (23 ± 4 years).

Results: The most active group completed over twice as many steps/day as the least active group ($p < 0.001$), had lower BMI's ($p = 0.007$) and body fat percentages ($p = 0.029$). Neutrophils migrated towards IL-8 better in the most active group compared to the least active ($p < 0.05$) and was comparable to that of the young ($p > 0.05$). These differences remained after adjusting for BMI, body fat and plasma metabolic markers which were different between groups. Correlations revealed that steps/day, higher adiponectin and lower insulin were positively associated with migratory ability ($p < 0.05$). There was no difference in expression of the chemokine receptors CXCR1 or CXCR2 ($p > 0.05$ for both). CD11b was higher in the most active group compared to the least active ($p = 0.048$). No differences between activity groups or young controls were observed for neutrophil phagocytosis or oxidative

burst in response to *E.coli* ($p>0.05$). The young group had lower concentrations of IL-6, IL-8, MCP-1, CRP, IL-10 and IL-13 ($p<0.05$ for all) with no differences between the two older groups.

Conclusion: These data suggest that impaired neutrophil migration, but not bactericidal function, in older adults may be, in part, the result of reduced physical activity. A 2-fold difference in physical activity is associated with better preserved neutrophil migratory dynamics in healthy older people. As a consequence increasing habitual physical activity may be beneficial for neutrophil mediated immunity.

1. Introduction

Innate immunity is the first line of defence against invading microorganisms. With advancing age there is a chronic deterioration of immune function, termed immunosenescence, leading to increased susceptibility to infection [1, 2]. Neutrophils are the most abundant leukocyte in blood and the first immune cell to arrive at sites of bacterial infection, making them critical intermediaries in the effective resolution of infection. Neutrophil dysfunction is evident in healthy older adults and is characterised by reduced directional migration, phagocytosis, oxidative burst and neutrophil extracellular trap (NET) production [3-6]. Reduced chemotaxis may be particularly detrimental as it will delay recruitment of neutrophils to sites of infection, but it may also contribute to heightened inflammation during infection and pro-inflammatory insults in older adults. This is because neutrophils release proteases such as neutrophil elastase to aid migration through tissue [7], thus damaging healthy tissue and inducing inflammation [4]. Inefficient chemotaxis will increase this unintentional damage and inflammation. It remains unclear why neutrophil dysfunction occurs with ageing but it is likely that defects are intrinsic to the cellular maturation process as neutrophils exit the bone marrow as mature non-proliferating cells [6, 8, 9]. Our own recent work has shown that signalling pathways central to chemotaxis, namely phosphoinositide 3 kinase (PI3K), is dysregulated in neutrophils from older adults and reduced chemotaxis can be corrected by selective inhibition of this pathway [4]. Identifying ways to prevent or reverse neutrophil immunosenescence is critical to improving immunity in older adults.

One of the features of ageing in humans is a reduced level of physical activity. Less than half of over 65 year olds in the UK meet national guidelines for habitual physical activity [10]. Importantly, a growing body of evidence suggests that regular

participation in physical exercise can improve or maintain immune function in old age, potentially slowing the progression of immunosenescence [11, 12]. The question thus arises of whether a significant part of immunosenescence is the consequence of an inactive lifestyle, rather than being due to the intrinsic ageing process. Despite the extent of neutrophil dysfunction with age and its potential contribution to ill-health, there are relatively few studies assessing the effects of exercise on neutrophil function in the elderly. Yan and colleagues showed that phagocytosis was better maintained in physically active compared to sedentary older men [13]. In overweight older women, 6-weeks of aerobic exercise training reduced circulating neutrophil numbers – a potentially beneficial anti-inflammatory effect [14]. In young participants, neutrophil chemotaxis and phagocytosis was improved following 2 months of moderate aerobic exercise training [15]. These studies highlight the potential for regular exercise to positively modify neutrophil function and potentially lower the risk of infectious diseases in later life.

Understanding the effects of habitual physical activity on immune function in the elderly could lead to new models of exercise prescription for older adults. To date, no studies have assessed broad neutrophil functions in healthy older individuals with a range of physical activity levels. The aim of this study was thus to assess a comprehensive range of neutrophil functions known to be reduced with age and determine the association with habitual physical activity. We hypothesized that regular physical activity in later life would be associated with maintenance of neutrophil function. Specifically we assessed directional migration, cell receptor expression, phagocytosis of *E.coli* and the oxidative burst response to *E.coli*.

2. Materials and Methods

2.1.1. Participants and assessment of habitual physical activity

An assessment of physical activity and a broad range of physiological function were completed in 211 healthy older participants, 100 men (67 ± 5 years) and 111 women (66 ± 5 years), at the Wellcome Trust Clinical Research Facility (WTCRF), Birmingham, UK. For assessment of habitual activity levels participants wore a GT3X accelerometer (Actigraph, FL, USA) around the waist above the left leg for 7 days, except for during water based activities. Total steps per day were calculated using ActiLife Software v5 (Actigraph, FL, USA). Following 7 continuous days wearing an accelerometer the 20 most active (MA) and 20 least active (LA) participants were identified and asked to return to the WTCRF for an additional blood sampling. All participants were grouped (quintiles) by physical activity and then stratified for age and gender. The top ten MA males were then matched for age (± 2 years) with ten LA males and the same was done for women. Upon recruitment to the study, participants again wore the accelerometer to ensure physical activity levels were similar to their first visit and completed the International Physical Activity Questionnaire (IPAQ). Details of participant age, physical activity levels, clinical and physiological characteristics are described in Table 1. Ten healthy young individuals (23 ± 4 years) were also recruited in order to compare neutrophil functions and serological status with age.

All participants were asked to abstain from physical exercise for 48 hours before arriving at the clinic. Exclusion criteria was specified to ensure relatively good health and included: no general medical history in the previous 3 years: dementia, Parkinson's, stroke, transient ischaemic attacks, liver disease, cancer, myocardial infarction, cardiomyopathy, valvular disease, angina pectoris, sudden chest pains,

blood pressure > 190/120 or tendencies to faint, asthma, chronic obstructive pulmonary disease, rheumatoid arthritis or osteoarthritis, use of corticosteroids, type 1 or 2 diabetes, leg pain during exertion, and consultation with GP for feeling unwell in the previous 10-days. Ethical approval was granted by the Black Country Research Ethics committee (10/H1202/77) and all participants gave written informed consent.

2.1.2. Clinical Assessment

Clinical assessments were taken by trained research nurses in the WTCRF. Briefly height and weight were recorded and body composition was measured using dual-energy x-ray absorptiometry (GE Lunar iDXA, GE Healthcare, Madison, WI) using Encore Software v13.6, as previously described [16]. Blood pressure (BP) and heart rate were taken after 15 minutes sitting, using an Omron M5-I automated BP machine (Omron Healthcare, Lake Forest, USA). An average of 3 BP readings taken over 5 minutes was taken as clinical BP [17].

2.1.3 Functional Testing

Each participant completed a short battery of physical functioning tests which are indicative of health and progression towards frailty in older people [18-21]: Hand grip strength was determined by measuring both left and right hands using a hand held dynamometer (Takei TKK-5401, Takei Scientific Instruments, Japan) and the best scores used. Participants also completed the 6-metre timed-up and go (TUG) test, briefly, participants were timed to stand from sitting, walk 3-metres at their regular walking speed, turn around, walk back and sit down. The Berg Balance Scale (BBS), a measure of balance and postural stability, was also carried out in which participants were asked to complete 14 balance tasks and scores given. Scoring gives a risk of falling, with 41 – 56 a 'low risk of falling'.

2.1.4. Blood Sampling

Participants arrived at the clinic at 8am having fasted overnight and peripheral venous blood samples were taken into vacutainers containing either heparin, EDTA or a clotting agent for serum. Samples were then processed immediately for serum, plasma and immune cell isolation. Serum and plasma were separated from blood by centrifugation (3000xg for 10 minutes), snap frozen in liquid nitrogen and stored at -80 °C until analysis.

2.1.5. Neutrophil Isolation

Neutrophils were isolated from heparinised blood following dextran induced sedimentation and separation on a discontinuous Percoll gradient as previously described [3]. Neutrophil purity and viability was determined by Giemsa staining (Diff-Quik; Gentaur Europe, Brussels, Belgium) and trypan blue (Sigma Aldrich, Dorset, UK) exclusion, respectively. Neutrophil purity and viability were consistently $\geq 96\%$. Neutrophils were then resuspended at $1 \times 10^6 \text{ ml}^{-1}$ in RPMI 1640 medium (Sigma-Aldrich) containing 0.15% bovine serum albumin (Sigma-Aldrich).

2.1.6. Neutrophil Migration

Neutrophil migratory dynamics were assessed using an Insall chamber (Weber Scientific International Ltd., Teddington, UK) as previously described [4, 22]. Briefly, coverslips were coated with 7.5% culture tested bovine serum albumin (Sigma-Aldrich) and neutrophils adhered to this surface for 20 minutes at room temperature. The use of this bovine serum albumin has previously been shown to mimic the ligand for CD11b and CD18, ICAM-1 [23]. The coverslip was then inverted on the Insall chamber before addition of buffer (RPMI-1640) alone as a control or buffer containing 100 nM of interleukin (IL)-8 (R&D Systems; Abingdon, UK).

Neutrophil migration was monitored using a Zeiss Axiovert 100 inverted microscope fitted with a Fast Mono 12-bit QICAM digital camera. Time-lapse recordings and calculation of neutrophil migratory dynamics were performed as previously described [4]. Briefly, the Insall chamber allows the formation of stable chemoattractant gradients, with defined, consistent direction in the y direction for each experiment [22]. Only distance travelled in the y direction over time was included in calculations of chemotaxis. Migration was assessed using 3 parameters: Average cell speed ($\mu\text{m}/\text{min}$) of movement towards a chemokine (e.g. IL-8, termed chemokinesis), average velocity ($\mu\text{m}/\text{min}$) of cells (termed chemotaxis) and accuracy of movement (termed chemotactic index). Chemotactic index is expressed in a comparative scale (CS) ranging from -1 to +1. Movement directly towards the chemoattractant is +1 whilst directly away is -1.

Recordings lasted 20 minutes per experiment, with 20 slides captured using Improvision OpenLab software. The Java software ImageJ (Wayne Rasband, NIH, Bethesda, MD) was used to analyse cell tracks. All analysis was carried out by a single analyst, blinded to subject group and cell conditions.

2.1.7. Neutrophil Phagocytosis and Oxidative Burst

Phagocytosis and oxidative burst were assessed in whole blood using commercially available kits and manufacturers' guidelines (BD Biosciences, Oxford, UK). Briefly, phagocytosis was assessed in heparin treated whole blood and incubated at 4°C (control) or 37°C (test) with opsonised FITC-labelled *E.coli*. Phagocytosis was halted by the addition of cold phosphate buffered saline (PBS) whilst cell surface bound FITC was quenched by addition of Trypan Blue solution. Unbound free bacteria were removed by washing in PBS and erythrocytes lysed and leukocytes fixed using 1%

fix/lyse solution provided in the kit. Cell DNA was counterstained by addition of propidium iodide (PI) before flow cytometry analysis was performed.

Oxidative burst was assessed in heparin treated whole blood which was incubated at 37°C with opsonised *E.coli* (test) or PBS (control). Solution containing dihydrorhodamine-123, which is converted to fluorescent rhodamine-123 in the presence of reactive oxidants, was included for 30 minutes at 37°C. Oxidative burst was halted by the addition of erythrocyte lysis/leukocyte fixation buffer before leukocyte DNA was stained and flow cytometry analysis performed.

Phagocytosis and oxidative burst quantitation was performed on a CyAn_{ADP}TM 430 flow cytometer (Beckman-Coulter, High Wycombe, UK) equipped with 3 solid-state lasers. FITC and R-123 were detected in FL1 whilst PI was detected in FL2 using the Argon (405nm) laser. 10,000 neutrophils were acquired for analysis. Following compensation of FL1 v FL2 phagocytic and oxidative burst were determined by the relative increase in percentage and median fluorescence intensity (MFI) in FL1 compared to negative controls. Data were analysed using Summit v4.3 (DAKO, Cambridgeshire, UK).

2.1.8. Measurement of Surface Receptor Expression

Freshly isolated neutrophils ($1 \times 10^5 \text{ ml}^{-1}$) were stained with anti-CXCR1-FITC (eBioscience, Hatfield, UK, clone 8F1-1-4), anti-CXCR2-PE (eBioscience, clone 5E8-C7-F10), anti-CD16-APC (eBioscience, clone CB16), anti-CD16-FITC (BD Bioscience, clone 3G8), anti-CD11b-APC (BD Bioscience, clone ICRF44), anti-CD18-PE (BD Bioscience, clone 6.7) or their relevant concentration-matched isotype control for 30 minutes on ice in the dark. Following incubation, cells were washed in PBS/1% BSA and resuspended in PBS/1% BSA for analysis by flow cytometry.

10,000 CD16⁺ neutrophils were acquired for analysis on a CyAn_{ADP}TM 430 flow cytometer and data analysed using Summit v4.3 software.

2.2.0. Serological and Metabolic Analysis

All measurements were made in duplicate using commercially available kits and manufacturers guidelines. Serum IL-1 β , IL-4, IL-6, IL-8, IL-10, IL-13, IL-17, GMC-SF, TNF α and MCP-1 were simultaneously measured by multiplex luminometry (Bio-Rad, Hemel Hempstead, UK). Samples were analysed using a Bio-Plex Lumines²⁰⁰ platform equipped with a 635nm red and 532nm green laser using Bio-Plex ManagerTM software. Detection of C-reactive protein (CRP) was by high sensitivity ELISA using a commercial kit (IBL International, Hamburg, Germany). Plasma adiponectin and leptin were assessed separately by solid phase sandwich ELISA (R&D Systems, Abingdon, UK). Plasma insulin levels were determined by sandwich ELISA (Life Technologies, Paisley, UK). Plasma glucose, cholesterol and triglycerides (Instrumentation Laboratories, Warrington, UK) were determined photometrically on an automated chemistry analyser (iLab 650, Instrumentation Laboratories, Warrington, UK).

2.2.1. Statistical Analysis

All analyses were conducted using PASW version 18.0 (Chicago, IL, USA) and all data presented as mean \pm standard deviation (SD) unless otherwise stated. Normality was assessed using Kolmogorov-Smirnov analysis; natural log transformation of distributed variables violating normality was completed. Analysis of covariance (ANCOVA) with Bonferroni correction and independent sample T-tests were used to determine differences between the groups. Covariates were included in the analyses if they were different between the most and least active elderly groups. Pearson correlation analysis was conducted to determine relationships between

migratory dynamics and steps/day as well as metabolic variables. The homeostasis model assessment of insulin resistance (HOMA-IR) was calculated as described previously using the HOMA2-IR online calculator [24, 25]. As previously described, all experiments were preceded by studies of intra-subject and inter-subject variance to allow appropriate power calculations to be performed [4]. Statistical significance was accepted as $p < 0.05$.

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3. Results

3.1. Participant characteristics

The physical and serological characteristics of the participants are shown in Table 1. Physical activity levels ranged from 1518 – 19,161 steps/day and as is typical with older people a low percentage, only 13%, completed more than 10,000 steps/day, causing a left-shift in distribution. There were no differences between the first and second wearing of the accelerometers and both were highly correlated, ($r(38) > .784$; $p < 0.001$ for all), therefore data presented here are from the second wear time only. The most active (MA) group was over twice as active for steps/day as the least active (LA) group ($p < 0.001$). These data highlighted that not only were the MA group walking more, but were less sedentary. Results from self-perceived activity reports (IPAQ) were similar to accelerometer data with approximately a two-fold difference between the two groups ($p = 0.026$).

Although attempts were made to match the groups for as many potential confounding variables as possible, inevitably a number of differences were apparent. In particular, body mass index (BMI; $p = 0.007$) and body fat percentage ($p = 0.029$) were higher in the LA group which likely accounted for a number of metabolic differences. Fasting glucose ($p = 0.010$), insulin ($p = 0.004$) and leptin ($p = 0.036$) were all higher whilst adiponectin ($p = 0.047$) was lower, in the LA group. Importantly, none of these measures were clinically abnormal and other measures of health including blood pressure, resting heart rate, triglycerides, total cholesterol, grip strength, balance, walk speed, statin medication, and smoking were similar between groups ($p > 0.05$ for all). There were also no differences in total immune cell counts for neutrophils, monocytes or lymphocytes between the two groups (Table 1). Taken together, these data show that our two populations were relatively healthy and we

can be confident that any group differences are likely driven by differences in physical activity levels rather than morbidity.

3.2. Neutrophil migration and physical activity

Efficient neutrophil migration is a critical step in the resolution of infection and has recently been shown to be impaired with advancing age [4]. In agreement with published data, isolated neutrophils from young participants migrated towards the chemokine IL-8 with a greater directional speed [chemotaxis ($p=0.038$)] and an overall greater chemotactic accuracy [chemotactic index ($p=0.020$)], but not overall speed [chemokinesis ($p=0.098$)], than older participants (Table 2). When split into their respective activity groups, the LA group had significantly reduced neutrophil chemotaxis (Figure 1A) compared to both the MA ($p=0.044$) and young participants ($p=0.021$) and there were still no differences between groups for chemokinesis (Figure 1B). Chemotactic index for the LA group (Figure 1C) was also reduced compared to the MA group ($p=0.002$) and young participants ($p=0.001$).

Correlation analysis revealed that increased steps/day were associated with increased neutrophil chemotactic index ($r(38)=-.689$, $p=0.001$) but not chemotaxis or chemokinesis (both $p>0.05$). These data suggest being physically active in later life could maintain neutrophil migratory dynamics, specifically accuracy and velocity, and that the amount of physical activity is related to the degree of migratory benefit.

3.3. Neutrophil migration and a relationship with metabolic phenotype

In order to determine if the relationship between neutrophil migration and physical activity might be mediated by differences in metabolic parameters between the LA and MA groups, data was analysed to include BMI, body fat percentage, adiponectin, leptin, glucose and insulin concentrations as covariates. Inclusion of these variables

did not statistically alter our findings for any individual migratory parameters, chemokinesis, chemotaxis or chemotactic index (Supplementary Table 1). Correlation analysis did however reveal that certain metabolic markers were associated with the overall chemotactic index. Figure 2A shows that adiponectin ($r(38) = .515, p=0.05$) and insulin (Figure 2D: $r(38) = -.617, p=0.01$) were associated with chemotactic index, whilst leptin (Figure 2B: $r(38) = -.359, p=0.22$) and glucose (Figure 2C: $r(38) = -.327, p=0.263$) were not. These data suggest that exposure to raised adiponectin and reduced insulin may play a role in mediating the effects of exercise on neutrophil migration.

3.4. Phagocytosis and oxidative burst and relation to physical activity

A primary mechanism of bacterial clearance by neutrophils is ingestion through phagocytosis and subsequent killing in the phagolysosome through exposure to reactive oxygen species such as superoxide anions. All neutrophils ingested bacteria and produced an oxidative burst. Phagocytosis of opsonised FITC-labelled *E.coli* showed no differences between young, LA or MA groups (Figure 3A) and the oxidative burst also did not differ between groups (Figure 3B).

3.5. Neutrophil receptor expression

To understand whether the differences in migration were due to the detection of the chemokine IL-8, or the adhesion to the coverslip chemokine and adhesion receptor expression was assessed on isolated neutrophils (Table 3). All neutrophils expressed the receptors measured. No differences were observed between groups for the IL-8 receptors CXCR1 and CXCR2, or the adhesion molecule CD18. There was a small group effect for the adhesion molecule CD11b [$F_{(2, 39)}=3.64, p=0.036, \eta^2=.168$] which was due to lower expression on the LA neutrophils compared to the MA ($p=0.048$) and young ($p=0.041$) groups. CD11b was included as a covariate in

the analysis of Chemotactic Index but did not alter our findings. Our data suggest that differences in neutrophil migration are not the result of chemokine receptor expression.

3.6. Systemic inflammation and neutrophil migratory dynamics

Exposure to inflammatory mediators is known to influence the function of neutrophils including migration. In order to determine whether systemic inflammation could have contributed to migratory differences levels of pro-inflammatory cytokines, acute phase proteins and anti-inflammatory cytokines were assessed. Table 4 reveals the young subjects had lower levels of circulating IL-6 ($p=0.011$), IL-8 ($p=0.049$), MCP-1 ($p=0.002$), CRP ($p=0.039$), IL-10 ($p=0.004$) and IL-13 ($p=0.033$) than the combined older participants. No differences between activity groups for the LA and MA groups were evident.

4. Discussion

With a growing body of evidence to suggest that physical activity in later life reduces the risk of age related diseases it is imperative to understand which specific aspects of physiology benefit from a healthy lifestyle [26, 27]. The immune system characteristically becomes dysregulated with age leading to a severe functional loss and exposing older individuals to increased risk of infection and disease [28, 29]. The beneficial effects and impact of physical activity on the immune system, especially adaptive immunity, have been extensively studied in young healthy individuals [30-32]. However few studies have specifically focused on the neutrophil in older adults and the effects of habitual physical activity in later life. This study is the first to compare neutrophil phagocytosis, superoxide production and detailed migratory dynamics towards the chemokine IL-8 in habitually physically active and less active healthy older individuals. We show for the first time that a 2-fold difference in physical activity is associated with better preserved neutrophil migratory dynamics in healthy older people.

Neutrophil migration is a key component for the resolution of infection and control of inflammation. Along with age-associated reductions in bacterial clearance, migratory dysfunction can lead to increased morbidity and mortality from bacterial infections [33]. Defective migration results in poor resolution of inflammation and wound repair in aged mice [34]. Additionally, if the neutrophil does not migrate accurately between tissues, collateral damage is augmented by elevated proteinase release from the cell, enhancing the inflammatory milieu and potentially contributing to the increased infection associated frailty [35-37]. Recent work from our group showed that neutrophils from older adults migrated with a similar speed but reduced velocity and accuracy when compared to younger participants [4]. Our findings here show that

velocity and accuracy were similarly reduced in our older cohort compared to young subjects, but we now add that the reduced habitual activity of our elders is associated with this loss of migratory accuracy.

A number of studies have highlighted the beneficial effects of physical exercise on immune function in the young and to a lesser extent the old. However, few studies have assessed whether neutrophil migration is associated with physical activity. Neutrophil migration was improved following six months of moderate exercise training in hypertensive older women (65 ± 5 years) to a similar level of migration as that in age-matched healthy women [38]. Furthermore, migration was also improved 2-fold in young sedentary individuals following a 2-month moderate intensity cycling exercise program [15]. Interestingly, in both these studies following a period of no-training or de-training neutrophil migration reverted back to levels prior to commencing training. These data suggest that efficient neutrophil migration is not only improved by physical exercise but is highly plastic and fundamentally dependent on it. In support of a physical activity effect on neutrophil function, we show that just a 2-fold difference in habitual physical activity levels was sufficient to be associated with increased neutrophil chemotaxis and chemotactic index. Moreover, none of our participants were actively engaged in structured exercise training as stated in their IPAQ's, thus higher habitual physical activity levels resulted in significantly better migration.

To investigate the potential mechanisms we assessed whether cell surface receptor expression related to migratory dynamics was altered with physical activity levels. The two main neutrophil receptors for IL-8 are the G-coupled protein receptors CXCR1 and CXCR2, which signal through a range of pathways, with the PI3K/AKT and Rac GTPase pathways central to chemotaxis and migratory accuracy [42].

CXCR2 has been shown to increase in muscle fibres and vascular endothelium following acute exercise, likely stimulating angiogenesis in the working tissue [39]. However we found that physical activity levels did not correlate with CXCR1 or CXCR2 expression on neutrophils. Our group recently showed that neither CXCR1 nor CXCR2 were altered with age and instead migratory dysfunction was associated with constitutive activation of PI3K, which if inhibited pharmacologically led to improved chemotaxis [4]. Whether higher habitual physical activity can reduce the constitutive PI3K activity seen in neutrophils with age was not determined here, but could offer an explanation for the association observed. Thus migratory defects due to physical inactivity are likely intrinsic to downstream signalling differences and possibly through modification of PI3K activity.

Of the adhesion receptors measured only CD11b was different between active and inactive elders, with around a 2-fold reduced expression in less active elders. CD11b is an α -integrin which forms a complex with the β 2-integrin CD18 to form the adhesion complex MAC-1. Together this complex is important for firm adhesion to vascular endothelial walls where it binds to ICAM-1 before transmigrating into the tissue. In our model, the albumin used to coat the coverslips mimics ICAM-1 [23]. Basal neutrophil expression of CD11b has been shown to be unaltered with age [3, 40]. Priming neutrophils with TNF α up-regulates expression of MAC-1 and promotes migratory function [41]. Interestingly, TNF α mediated MAC-1 expression is dependent on PI3K/Akt and p38 MAPK activity [41]. Therefore it may be that the increased expression of CD11b on neutrophils from active elders confers an efficient migratory phenotype through a functional PI3K/Akt signalling pathway similar to younger individuals. It remains unclear whether an increased basal expression of CD11b results in improved neutrophil migration in elders who are physically active.

CD11b but not CD18 neutrophil expression has been shown to be upregulated following acute bouts of exercise and exercise training [43]. Therefore it may be that regular physical activity promotes surface expression of CD11b as a surveillance mechanism readying neutrophils to migrate to sites of infection.

One unexpected result was the lack of a difference in phagocytosis or ROS production between any of the groups. We have previously shown age-associated reductions in phagocytosis and others have reported reductions in ROS production in response to *Staphylococcus Aureus* but not to *E.coli*, findings for these parameters are thus variable in the literature [3, 44]. The data for habitual physical activity do suggest however that this does not modify these aspects of neutrophil function. One difference between these bactericidal assays and neutrophil migration is the stimulus for migration was a single chemokine (IL-8), whereas ingestion of the bacteria and ROS generation is effected by a range of signalling pathways and receptors which may not be sensitive to physical activity.

Neutrophils exposed to chronic inflammatory stimulants like TNF α and IL-1 β , such as occurs in rheumatoid arthritis, have altered function [45]. This includes extended lifespan and increased inflammatory cytokine and chemokine production augmenting the inflammatory milieu [46-48]. Migratory activity is also altered in these conditions with cells becoming more adherent [49-52]. Increased systemic inflammation with age, inflammaging, has been well documented and we observed higher systemic inflammation in the older group as a whole compared to the young. However no differences were observed between high and low physical activity groups. This is in contrast to a number of studies suggesting that exercise can reduce systemic inflammation and that physical inactivity is associated with elevated inflammation [53-55]. This might be reflective of our recruitment criteria and the health of our

participants. In addition it is possible that our sample size lacked power to detect a difference in the older group with activity level. Thus, the migratory differences observed between activity groups and the young were likely not a consequence of chronic exposure to systemic inflammation.

Regular physical activity is known to positively alter the metabolism of individuals and reduce the risk for type-2 diabetes and associated cardiovascular disorders [56]. Indeed, we observed a significantly lower measure of insulin resistance and diabetes risk (HOMA-IR) in the MA group ($p=0.004$). Lower circulating insulin and fasting glucose levels coupled with reduced body fat and adipocyte related inflammation are a common feature of physically active individuals. Additionally, insulin signals through the PI3K/AKT pathway also and may account for altered PI3K activation in the elderly with increased circulating insulin concentrations [57]. Recently neutrophils were shown to facilitate insulin resistance in obese mice, highlighting a specific connection between innate immune function and metabolism [58]. Adipokines such as adiponectin and leptin are important regulators of energy metabolism and their ratio is indicative of insulin sensitivity [59, 60]. Additionally both have been suggested to modify neutrophil function [61-63]. In this study we found no association between serum leptin and chemotactic index. Adiponectin on the other hand plays an important role in the regulation of energy metabolism via AMPK signalling and has inherent anti-inflammatory effects. Here adiponectin levels were positively associated with chemotactic index. Our group showed adiponectin delayed spontaneous neutrophil apoptosis whilst others have shown reduced IL-8 production in obese individuals associated with reduced serum adiponectin [64, 65]. Recently, adiponectin was associated with improved neutrophil migration towards acute kidney injury in mice [66]. A recent study suggested that the addition of 20-weeks aerobic

exercise to dieting in overweight postmenopausal women resulted in increased adiponectin concentrations [67]. Together these findings suggest a positive role for adiponectin on neutrophil function and migration. Further work is required to understand metabolism in neutrophils and the role exercise plays.

Our study is not without limitations. We did not measure body composition, physical activity levels or physical functioning in our young cohort. Our data suggest that neutrophil dysfunction, specifically chemotactic index, may be a consequence of physical inactivity. This would have been further supported had the activity levels of the young cohort matched the most active elderly. Furthermore we did not consider dietary differences between groups. It is clear that different compositions (carbohydrate, protein, fats) of diet can have effects on metabolism and therefore potentially cell function. However, we measured a number of inflammatory and endocrine markers which are associated with poor diet and increased adiposity and determined they did not have a direct effect on neutrophil migration. Future studies assessing immunological differences between young and old individuals should consider physical activity and dietary differences. Finally we have previously observed that PI3K is constitutively activated in elderly individuals and blocking PI3K in these cells restores migratory dynamics. Future studies should assess PI3K expression associated with physical activity and determine where the signalling defect occurs.

In conclusion our study suggests impaired neutrophil migration, but not bactericidal function, in older adults may be in part the result of physical inactivity. It is unlikely that these effects are driven by systemic inflammation or reduced surface chemokine receptor expression but a positive correlation with adiponectin levels suggests an

influence of this metabolic hormone. Increasing habitual physical activity levels in elderly individuals may therefore be beneficial for neutrophil mediated immunity.

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Conflicts of Interest

All authors declare they have no conflicts of interest.

References

1. Gavazzi, G. and K.H. Krause, *Ageing and infection*. The Lancet Infectious Diseases, 2002. **2**: p. 659 - 66.
2. Butcher, S.K., et al., *Effect of age on susceptibility to post-traumatic infection in the elderly*. Biochemical Society Transactions, 2003. **31**(2): p. 449-51.
3. Butcher, S.K., et al., *Senescence in innate immune responses: reduced neutrophil phagocytic capacity and CD16 expression in elderly humans*. Journal of Leukocyte Biology, 2001. **70**(6): p. 881-6.
4. Sapey, E., et al., *Phosphoinositide 3 kinase inhibition restores neutrophil accuracy in the elderly: towards targeted treatments for immunosenescence*. Blood, 2013.
5. Hazeldine, J., et al., *Impaired neutrophil extracellular trap formation: a novel defect in the innate immune system of aged individuals*. Aging Cell, 2014. **13**(4): p. 690-698.
6. Shaw, A., et al., *Aging of the innate immune system*. Current Opinion in Immunology, 2010. **22**(4): p. 507 - 513.
7. Cepinskas, G., M. Sandig, and P.R. Kvietys, *PAF-induced elastase-dependent neutrophil transendothelial migration is associated with the mobilization of elastase to the neutrophil surface and localization to the migrating front*. Journal of Cell Science, 1999. **112**(12): p. 1937-1945.
8. Ward, J.R., et al., *Regulation of Neutrophil Senescence by MicroRNAs*. PLoS ONE, 2011. **6**(1): p. e15810.
9. Fortin, C.F., et al., *Aging and neutrophils: there is still much to do*. Rejuvenation Research, 2008. **11**(5): p. 873-82.
10. House of Lords Select Committee on Public Service and Demographic Change. *First Report: Ready for Ageing*. 2013 [cited 2014 June]; Available from: <http://www.publications.parliament.uk/pa/ld201213/ldselect/ldpublic/140/14002.htm>.
11. Walsh, N.P., et al., *Position statement. Part one: Immune function and exercise*. Exercise Immunology Review, 2011. **17**: p. 6-63.
12. Simpson, R.J., et al., *Exercise and the aging immune system*. Ageing Research Reviews, 2012(0).
13. Yan, H., et al., *Effect of moderate exercise on immune senescence in men*. European Journal of Applied Physiology, 2001. **86**(2): p. 105-11.
14. Michishita, R., et al., *Effect of exercise therapy on monocyte and neutrophil counts in overweight women*. The American Journal of the Medical Sciences, 2010. **339**(2): p. 152-6.
15. Syu, G.D., H.I. Chen, and C.J. Jen, *Differential effects of acute and chronic exercise on human neutrophil functions*. Medicine & Science in Sports & Exercise, 2012. **44**(6): p. 1021-7.
16. Schousboe, J.T., et al., *Executive summary of the 2013 international society for clinical densitometry position development conference on bone densitometry*. Journal of Clinical Densitometry, 2013. **16**(4): p. 455-466.
17. Omboni, S., et al., *Validation of the Omron M5-I, R5-I and HEM-907 automated blood pressure monitors in elderly individuals according to the International Protocol of the European Society of Hypertension*. Blood Pressure Monitoring, 2007. **12**(4): p. 233-242.

18. Thrane, G., R.M. Joakimsen, and E. Thornquist, *The association between timed up and go test and history of falls: the Tromso study*. BMC Geriatrics, 2007. **7**: p. 1.
19. Nordin, E., et al., *Prognostic validity of the Timed Up-and-Go test, a modified Get-Up-and-Go test, staff's global judgement and fall history in evaluating fall risk in residential care facilities*. Age & Ageing, 2008. **37**(4): p. 442-8.
20. Muir, S.W., et al., *Use of the Berg Balance Scale for predicting multiple falls in community-dwelling elderly people: a prospective study*. Physical Therapy, 2008. **88**(4): p. 449-59.
21. Muir, S.W., et al., *Balance impairment as a risk factor for falls in community-dwelling older adults who are high functioning: a prospective study*. Physical Therapy, 2010. **90**(3): p. 338-47.
22. Muinonen-Martin, A.J., et al., *An Improved Chamber for Direct Visualisation of Chemotaxis*. PLoS ONE, 2010. **5**(12): p. e15309.
23. Zhu, X., et al., *A surrogate method for assessment of beta(2)-integrin-dependent adhesion of human eosinophils to ICAM-1*. Journal of Immunological Methods, 2000. **240**(1-2): p. 157-64.
24. The Oxford Center for Diabetes. Endocrinology & Metabolism Diabetes Trial Unit. *HOMA Calculator*. [cited 2014 June]; Available from: <http://www.dtu.ox.ac.uk/>.
25. Wallace, T.M., J.C. Levy, and D.R. Matthews, *Use and Abuse of HOMA Modeling*. Diabetes Care, 2004. **27**(6): p. 1487-1495.
26. Brandt, C. and B.K. Pedersen, *The role of exercise-induced myokines in muscle homeostasis and the defense against chronic diseases*. Journal of Biomedicine and Biotechnology, 2010. **2010**: p. 520258.
27. Pedersen, B.K., *Exercise-induced myokines and their role in chronic diseases*. Brain, Behavior, and Immunity, 2011. **25**(5): p. 811-6.
28. Pawelec, G., et al., *Human immunosenescence: is it infectious?* Immunological Reviews, 2005. **205**: p. 257-68.
29. Panda, A., et al., *Human innate immunosenescence: causes and consequences for immunity in old age*. Trends in Immunology, 2009. **30**(7): p. 325-33.
30. LaVoy, E.C., et al., *Acute aerobic exercise in humans increases cytokine expression in CD27(-) but not CD27(+) CD8(+) T-cells*. Brain, Behavior, and Immunity, 2013. **27**(1): p. 54-62.
31. Simpson, R.J., et al., *Senescent phenotypes and telomere lengths of peripheral blood T-cells mobilized by acute exercise in humans*. Exercise Immunology Review, 2010. **16**: p. 40-55.
32. Spielmann, G., et al., *Aerobic fitness is associated with lower proportions of senescent blood T-cells in man*. Brain, Behavior, and Immunity, 2011. **25**(8): p. 1521 - 1529.
33. Liang, S.Y. and P.A. Mackowiak, *Infections in the elderly*. Clinics in Geriatric Medicine, 2007. **23**(2): p. 441-456.
34. Brubaker, A.L., et al., *Reduced neutrophil chemotaxis and infiltration contributes to delayed resolution of cutaneous wound infection with advanced age*. The Journal of Immunology, 2013. **190**(4): p. 1746-1757.
35. Liou, T.G. and E.J. Campbell, *Quantum proteolysis resulting from release of single granules by human neutrophils: a novel, nonoxidative mechanism of extracellular proteolytic activity*. The Journal of Immunology, 1996. **157**(6): p. 2624-2631.

36. [Pham, C.T., *Neutrophil serine proteases: specific regulators of inflammation*. Nature Reviews Immunology, 2006. 6\(7\): p. 541-550.](#)
37. [Fleming, D.M. and A.J. Elliot, *The impact of influenza on the health and health care utilisation of elderly people*. Vaccine, 2005. 23, Supplement 1: p. S1-S9.](#)
38. [Fuente, M.D.L., A. Hernanz, and M. Vallejo, *The immune system in the oxidative stress conditions of aging and hypertension: favorable effects of antioxidants and physical exercise*. Antioxidants & Redox Signaling, 2005. 7\(9-10\): p. 1356-1366.](#)
39. [Frydelund-Larsen, L., et al., *Exercise induces interleukin-8 receptor \(CXCR2\) expression in human skeletal muscle*. Experimental Physiology, 2007. 92\(1\): p. 233-240.](#)
40. [Murciano, C., et al., *Influence of aging on murine neutrophil and macrophage function against Candida albicans*. FEMS Immunology & Medical Microbiology, 2008. 53\(2\): p. 214-221.](#)
41. [Montecucco, F., et al., *Tumor necrosis factor-alpha \(TNF-alpha\) induces integrin CD11b/CD18 \(Mac-1\) up-regulation and migration to the CC chemokine CCL3 \(MIP-1alpha\) on human neutrophils through defined signalling pathways*. Cellular Signalling, 2008. 20\(3\): p. 557-568](#)
42. [King, J.S. and R.H. Insall, *Chemotaxis: finding the way forward with dityostelium*. Trends in Cell Biology, 2009. 19:523-530.](#)
43. [Jordan, J., et al., *Regulation of MAC-1 \(CD11b/CD18\) expression on circulating granulocytes in endurance runners*. Medicine & Science in Sports & Exercise, 1999. 31\(3\): p. 362-7.](#)
44. [Wenisch, C., et al. *Effect of age on human neutrophil function*. Journal of Leukocyte Biology, 2000. 67 \(1\), p. 40-45.](#)
45. [Wright, H.L., et al., *Neutrophil function in inflammation and inflammatory diseases*. Rheumatology, 2010. 49\(9\): p. 1618-1631.](#)
46. [Raza, K., et al., *Synovial fluid leukocyte apoptosis is inhibited in patients with very early rheumatoid arthritis*. Arthritis Research & Therapy, 2006. 8\(4\): p. R120.](#)
47. [Cross, A., et al., *Secretion of oncostatin M by neutrophils in rheumatoid arthritis*. Arthritis & Rheumatism, 2004. 50\(5\): p. 1430-1436.](#)
48. [Parsonage, G., et al., *Prolonged, granulocyte-macrophage colony-stimulating factor-dependent, neutrophil survival following rheumatoid synovial fibroblast activation by IL-17 and TNFalpha*. Arthritis Research & Therapy, 2008. 10\(2\): p. R47.](#)
49. [Álvarez-Rodríguez, L., et al., *Phagocyte dysfunction in polymyalgia rheumatica and other age-related, chronic, inflammatory conditions*. Journal of Leukocyte Biology, 2013. 94\(5\): p. 1071-1078.](#)
50. [Aglas, F., J. Hermann, and G. Egger, *Abnormal directed migration of blood polymorphonuclear leukocytes in rheumatoid arthritis. Potential role in increased susceptibility to bacterial infections*. Mediators of Inflammation, 1998. 7\(1\): p. 19-23.](#)
51. [Paino, I.M.M., et al., *Phagocytosis and nitric oxide levels in rheumatic inflammatory states in elderly women*. Journal of Clinical Laboratory Analysis, 2011. 25\(1\): p. 47-51.](#)
52. [Stockley, J.A., et al., *Aberrant neutrophil functions in stable chronic obstructive pulmonary disease: The neutrophil as an immunotherapeutic target*. International Immunopharmacology, 2013. 17\(4\): p. 1211-7.](#)

53. Pischon, T., et al., *Leisure-time physical activity and reduced plasma levels of obesity-related inflammatory markers*. *Obesity Research*, 2003. **11**(9): p. 1055-64.
54. Colbert, L.H., et al., *Physical activity, exercise, and inflammatory markers in older adults: findings from the Health, Aging and Body Composition Study*. *Journal of the American Geriatrics Society*, 2004. **52**(7): p. 1098-104.
55. Nicklas, B.J., et al., *Exercise training and plasma C-reactive protein and interleukin-6 in elderly people*. *Journal of the American Geriatrics Society*, 2008. **56**(11): p. 2045-52.
56. Gleeson, M., et al., *The anti-inflammatory effects of exercise: mechanisms and implications for the prevention and treatment of disease*. *Nature Reviews Immunology*, 2011. **11**(9): p. 607-615.
57. Schulman, I.H., et al., *Dissociation between metabolic and vascular insulin resistance in aging*. *American Journal of Physiology - Heart and Circulatory Physiology*, 2007. **293**(1): p. H853-H859.
58. Talukdar, S., et al., *Neutrophils mediate insulin resistance in mice fed a high-fat diet through secreted elastase*. *Nature Medicine*, 2012. **18**(9): p. 1407-1412.
59. Inoue, M., et al., *Correlation between the adiponectin-leptin ratio and parameters of insulin resistance in patients with type 2 diabetes*. *Metabolism*, 2005. **54**(3): p. 281-286.
60. Inoue, M., et al., *Relationship between the adiponectin-leptin ratio and parameters of insulin resistance in subjects without hyperglycemia*. *Metabolism*, 2006. **55**(9): p. 1248-1254.
61. Caldefie-Chezet, F., et al., *Leptin: a potential regulator of polymorphonuclear neutrophil bactericidal action?* *Journal of Leukocyte Biology*, 2001. **69**(3): p. 414-418.
62. Caldefie-Chezet, F., A. Poulin, and M.-P. Vasson, *Leptin regulates functional capacities of polymorphonuclear neutrophils*. *Free Radical Research*, 2003. **37**(8): p. 809-814.
63. Lang, K. and J. Ratke, *Leptin and Adiponectin: new players in the field of tumor cell and leukocyte migration*. *Cell Communication and Signaling*, 2009. **7**(1): p. 27.
64. Rossi, A. and J. Lord, *Adiponectin inhibits neutrophil apoptosis via activation of AMP kinase, PKB and ERK 1/2 MAP kinase*. *Apoptosis*, 2013. **18**(12): p. 1469-1480.
65. Trellakis, S., et al., *Low Adiponectin, High Levels of Apoptosis and Increased Peripheral Blood Neutrophil Activity in Healthy Obese Subjects*. *Obesity Facts*, 2012. **5**(3): p. 305-318.
66. Jin, X., et al., *Genetic deficiency of adiponectin protects against acute kidney injury*. *Kidney International*, 2013. **83**(4): p. 604-614.
67. Wang, X., et al., *Addition of Exercise Increases Plasma Adiponectin and Release from Adipose Tissue*. *Medicine & Science in Sports & Exercise*, 2015.

Table 1: Characteristics of the forty older participants assessed.

	Elderly Activity Group	
	Most Active (MA)	Least Active (LA)
Gender (M/F)	10 / 10	10 / 10
Age (years)	67.4 ± 5.0	66.6 ± 4.9
<u>Physical Activity</u>		
Steps (per day)	10370 ± 3221	4746 ± 1351***
IPAQ (AU)	29.4 ± 16.9	13.4 ± 11.3*
<u>Physical Functioning</u>		
Grip Strength (kg)	30 ± 9	32 ± 9
Berg Balance Score (AU)	55.2 ± 1.4	55.3 ± 1.1
Timed-up-and-go (seconds)	8.1 ± 1.5	8.4 ± 1.4
<u>Clinical</u>		
BMI (kg/m ²)	23.5 ± 3.1	26.7 ± 3.9**
Body Fat (%)	26.2 ± 7.9	31.5 ± 6.7*
Blood Pressure		
Systolic (mmHg)	128 ± 16	127 ± 13
Diastolic (mmHg)	77 ± 6	77 ± 7
Resting Heart Rate (bpm)	65 ± 9	64 ± 8
Current Smoker (Y/N)	0 / 20	2 / 18
Use of Statins (Y/N)	1 / 19	5 / 15
<u>Physiological</u>		
White Blood Cells		
Neutrophils (10 ⁹ /L)	3.08 ± 0.8	3.34 ± 0.8
Monocytes (10 ⁹ /L)	0.27 ± 0.08	0.32 ± 0.12
Lymphocytes (10 ⁹ /L)	1.28 ± 0.28	2.00 ± 1.64
Fasting Glucose (mmol/L)	5.0 ± 0.4	5.4 ± 0.5*

Fasting Insulin (μ IU/ml)	12.3 \pm 4.1	20.9 \pm 11.5**
HOMA-IR (AU)	2.8 \pm 1.1	5.1 \pm 3.2**
Triglycerides (mmol/L)	1.14 \pm 0.56	1.16 \pm 0.62
Cholesterol [statin ⁺ (mmol/L)]	5.80	4.15 \pm 0.47
Cholesterol [statin ⁻ (mmol/L)]	5.75 \pm 0.97	5.04 \pm 1.34
Leptin (ng/ml)	9.9 \pm 7.8	15.7 \pm 8.7*
Adiponectin (μ g/ml)	4.2 \pm 2.0	3.1 \pm 1.1*
Leptin: Adiponectin Ratio	2.5 \pm 1.9	5.5 \pm 3.7**

IPAQ (International Physical Activity Questionnaire); AU (Arbitrary Units); BMI (Body Mass Index); HOMA-IR (Homeostasis Model Assessment for Insulin Resistance); Statin⁺ (Prescribed statins); Statin⁻ (Not prescribed statins). Data are mean \pm SD unless indicated otherwise. * p <0.05, ** p <0.01, *** p <0.001 compared to MA.

Table 2: Surface receptor expression (MFI) of key mediators of neutrophil adhesion and migration between young (n=10) and older participants (n=40). Data are mean \pm SEM.

	Young		Old					
			All	MA	LA			
<u>Chemokine</u>								
<u>Recognition</u>								
CXCR1	55.	\pm 2.8	58.	1.	57.	2.	59.	\pm 1.6
	1		3	5	3	5	3	
CXCR2	61.	\pm 12.	47.	4.	53.	6.	41.	\pm 5.4
	9	2	4	2	5	3	6	
<u>Adhesion Molecules</u>								
CD11b	26.	\pm 4.5	19.	2.	25.	4.	14.	1.6*
	6		8	3	2	2	6	#
CD18	39.	\pm 5.3	40.	3.	45.	5.	35.	\pm 4.3
	3		2	7	6	9	1	

CXCR (CXC Chemokine Receptor); CD (Cluster of Differentiation);. * $p < 0.05$ and # $p < 0.05$ compared to MA and young respectively

Table 3: Concentrations of pro- and anti-inflammatory mediators between the young (n=10) and older (n=40) participants. Values are pg/ml unless stated otherwise and data are mean \pm SEM.

	Young		Old					
			All		MA (n=20)		LA (n=20)	
<u>Pro-</u>								
<u>Inflammatory</u>								
IL-1 β (pg/ml) ^a	0.21	\pm 0.05	0.33	\pm 0.04	0.31	\pm 0.05	0.35	\pm 0.07
IL-6 (pg/ml) ^a	1.36	\pm 0.92	5.11	\pm 1.49 [#]	5.71	\pm 2.27	4.46	\pm 1.99
IL-8 (pg/ml)	5.63	\pm 0.58	6.79	\pm 0.30 [#]	6.49	\pm 0.32	7.11	\pm 0.51
IL-17 (pg/ml) ^a	10.0	\pm 10.0	16.0	\pm 6.55	17.4	\pm 11.3	14.4	\pm 6.13
	9	9	1		0	3	8	
TNF α (pg/ml) ^a	0.23	\pm 0.11	3.99	\pm 2.47	5.36	\pm 4.40	2.48	\pm 1.96
CRP (mg/L) ^a	1.30	\pm 0.81	2.13	\pm 0.45 [#]	2.46	\pm 0.81	1.77	\pm 0.29
GM-CSF (pg/ml) ^a	N		2.13	\pm 1.52	2.44	\pm 2.44	1.79	\pm 1.79
	D							
MCP-1 (pg/ml)	27.6	\pm 2.44	43.8	\pm 2.57 [#]	43.3	\pm 3.84	44.4	\pm 3.44
	3		4	#	1	#	2	#
<u>Anti-</u>								

Inflammatory

IL-4	N							
(pg/ml) ^a	D			0.01 ± 0.01	0.02 ± 0.01	0.01 ± 0.01		
IL-10				19.3	3.45 [#]	17.3	21.5	5.19
(pg/ml) ^a		7.12 ± 3.12		± 4	± #	± 4.67	± 6	± #
IL-13								
(pg/ml)		4.13 ± 1.20		7.15 ± 0.71 [#]	6.29 ± 0.87	8.10 ± 1.14		

ND (not-detectable); IL (Interleukin); TNF α (tumour necrosis factor- α); CRP (C-Reactive Protein); GM-CSF (Granulocyte/Monocyte Colony Stimulating Factor); MCP-1 (Monocyte Chemoattractant Protein-1). ^aNot normally distributed and log-transformed. [#] $p < 0.05$ and ^{##} $p < 0.01$ compared to young group.

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Legends to Figures

Figure 1: Differences for neutrophil chemotaxis (A), chemokinesis (B) and chemotactic index (C) for cells isolated from Young (n=10), Most active (MA) and Least active (LA) older participants (n=20 each). Data are mean \pm SEM.

Figure 2: Correlation analyses plus 95% confidence intervals of metabolic variables for all older participants (n=40) and their relationship with Chemotactic Index for Adiponectin (A), Leptin (B), Glucose (C), and Insulin (D).

Figure 3: Differences between the Young, Most (MA) and Least Active (LA) older participants for neutrophil uptake of FITC-labelled *E.coli* (A) and the oxidative burst against ingested *E.coli* (B). Data are mean \pm SEM.

Figure 1

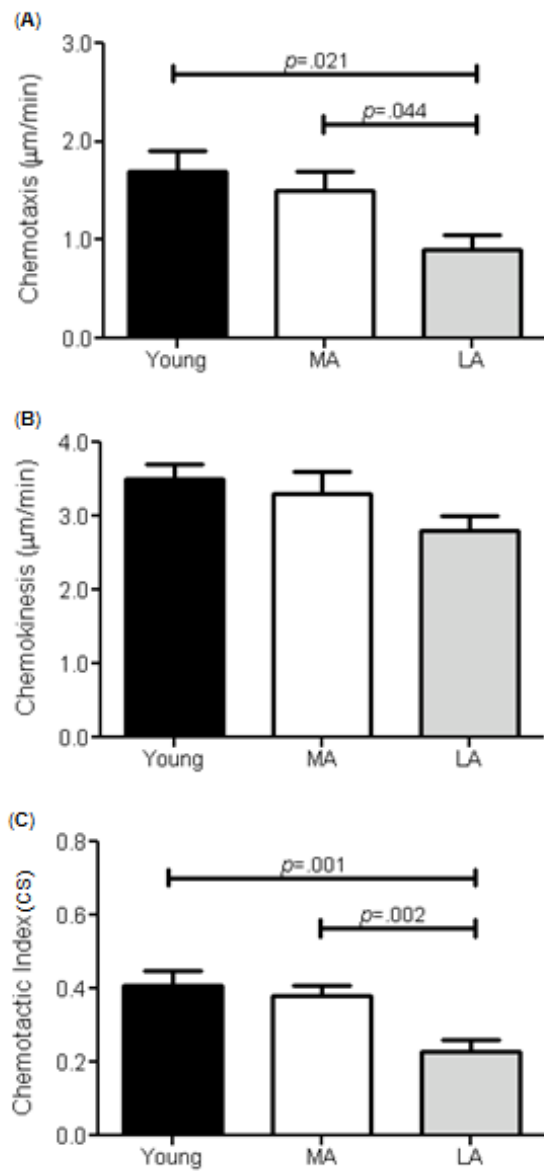


Figure 2

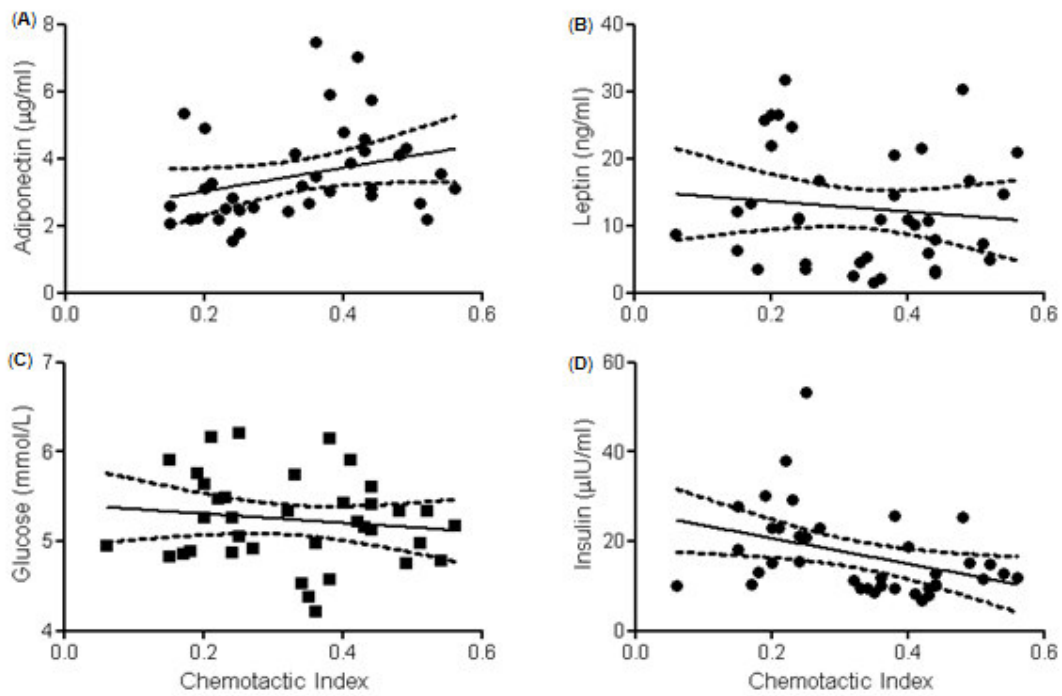
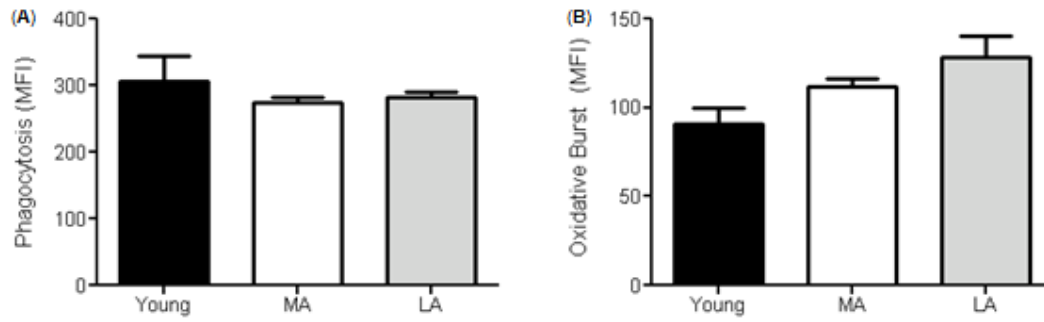


Figure 3



Highlights

- Higher habitual physical activity in over 65 year olds is associated with the maintenance of neutrophil migratory dynamics towards IL-8.
- The effect occurs independently of systemic inflammation or surface chemokine receptor expression.
- Adiponectin levels may positively influence neutrophil migration.

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