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1	Dose-response of sodium bicarbonate ingestion highlights individuality in time course of blood
2	analyte responses.
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18 **Running title:** Dose-response to sodium bicarbonate ingestion

### 19 ABSTRACT

20 To defend against hydrogen cation accumulation and muscle fatigue during exercise, sodium bicarbonate (NaHCO<sub>3</sub>) ingestion is commonplace. The individualised dose-response relationship 21 22 between NaHCO<sub>3</sub> ingestion and blood biochemistry is unclear. The present study investigated the 23 bicarbonate, pH, base excess and sodium responses to NaHCO<sub>3</sub> ingestion. Sixteen healthy males  $(23\pm2)$ 24 years; 78.6±15.1 kg) attended three randomised order-balanced, non-blinded sessions, ingesting a single dose of either 0.1, 0.2 or 0.3 gkg<sup>-1</sup>BM of NaHCO<sub>3</sub> (Intralabs, UK). Fingertip capillary blood was 25 26 obtained at baseline and every 10 min for 1 h, then every 15 min for a further 2 h. There was a significant 27 main effect of both time and condition for all assessed blood analytes ( $P \le 0.001$ ). Blood analyte responses were significantly lower following 0.1 g kg<sup>-1</sup>BM compared with 0.2 g kg<sup>-1</sup>BM; bicarbonate 28 concentrations and base excess were highest following ingestion of 0.3 g·kg<sup>-1</sup>BM (P≤0.01). Bicarbonate 29 concentrations and pH significantly increased from baseline following all doses; the higher the dose the 30 31 greater the increase. Large inter-individual variability was shown in the magnitude of the increase in 32 bicarbonate concentrations following each dose  $(+2.0-5; +5.1-8.1; and +6.0-12.3 mmol \cdot L^{-1}$  for 0.1, 0.2 and 0.3 g kg<sup>-1</sup>BM) and in the range of time to peak concentrations (30-150; 40-165; and 75-180 min for 33 0.1, 0.2 and 0.3 gkg<sup>-1</sup>BM). The variability in bicarbonate responses was not affected by normalisation 34 35 to body mass. These results challenge current practices relating to NaHCO<sub>3</sub> supplementation and clearly 36 show the need for athletes to individualise their ingestion protocol and trial varying dosages prior to competition. 37

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### **39 Key words:** Extracellular buffering, pH, fatigue

#### 40 INTRODUCTION

High-intensity exercise increases hydrogen cation (H<sup>+</sup>) production in the working muscle (Hill and 41 Lupton, 1923). The majority of these H<sup>+</sup> are buffered, with only a small fraction being free in the 42 cytosol to cause a decline in intracellular pH (Sahlin, 2014). It has been proposed that the decreased 43 44 intracellular pH is a critical factor in the development of fatigue during high-intensity exercise, either via a direct effect on the muscle contractile machinery or by disruption to muscle energetics (Fitts, 45 1996). The ability to deal with this proton production is an important determinant of exercise 46 47 performance and capacity. Two defence mechanisms against intramuscular acidosis are evident, 48 namely, intramuscular physicochemical buffers and dynamic buffering (*i.e.*, the ability to transport H<sup>+</sup> out of the muscle and into the blood). Whilst the first line of defence is intramuscular physicochemical 49 buffering, the main controller of pH during high-intensity exercise is dynamic buffering, this process 50 allows the bicarbonate buffering system to minimise disruption to intramuscular pH (McNaughton et 51 52 al. 2008; Carr et al. 2011)

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54 Supplementation with sodium bicarbonate (NaHCO<sub>3</sub>) increases the efflux of H<sup>+</sup> out of the muscle and the extracellular buffering capacity, thus delaying the onset of muscle fatigue, and maintaining exercise 55 performance (Cairns, 2006; Thomas et al. 2005). It is unsurprising that NaHCO<sub>3</sub> ingestion has been a 56 57 focus of researchers and athletes for over 30 years (Matson & Tran, 1993; Linderman & Gaosselink, 1994), with mixed findings in regards to the ergogenic efficacy of NaHCO<sub>3</sub> (for review see Peart et al. 58 2012). Some of these differences might be explained by differences dosing strategies. Several dosing 59 strategies are employed within with the sporting field, with NaHCO<sub>3</sub> doses of 0.2-0.5 gkg<sup>-1</sup>BM being 60 consumed to enhance exercise performance (McNaughton et al. 1991). Nonetheless, ingestion of 0.3 61 g·kg<sup>-1</sup>BM is most commonplace, consumed 60-90 min prior to exercise (Renfree, 2007; Price & Singh, 62 2008; Siegler et al. 2010) in flavoured water or capsules (Peart et al. 2012). Consumption of 0.3 gkg<sup>-</sup> 63 64 <sup>1</sup>BM typically increases blood bicarbonate concentrations by  $\sim$ 5–6 mmol<sup>-L-1</sup> from baseline (Matson & Tran, 1993; Price et al. 2003; Robergs et al. 2005; Saunders et al. 2014; Miller et al. in press), which 65

has been suggested to enhance the buffering process sufficiently to result in an ergogenic benefit (Carret al. 2011).

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Ingestion strategies can result in significant alterations in blood parameters, with peak acid-base 69 70 disturbances occurring between 60 and 90 min post ingestion of 0.3 gkg<sup>-1</sup>BM of NaHCO<sub>3</sub> (Renfree, 71 2007; Price & Singh, 2008; Siegler et al. 2010). Although there remains uncertainty as to how different doses affect the inter-individual variability in blood acid-base responses. Siegler et al. (2010) showed 72 73 that blood bicarbonate peaked 65 min post ingestion of 0.3 g kg<sup>-1</sup>BM, although due to blood samples 74 assessed at 20 min intervals some important aspects of the temporal pattern in acid-base responses might have been overlooked. It has been proposed that the blood buffering responses to NaHCO<sub>3</sub> ingestion 75 are highly individual (Peart et al. 2012; Saunders et al. 2014b) and in order to optimise ergogenic 76 potential, individualising the timing of exercise based on acid-base responses to NaHCO<sub>3</sub> ingestion 77 78 should be undertaken (Miller et al. in press). This highlights the need to examine how individuals 79 respond to varying NaHCO<sub>3</sub> doses.

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NaHCO<sub>3</sub> ingestion can result in gastrointestinal (GI) distress (Carr et al. 2011; Siegler et al. 2012; Peart 81 82 et al. 2012), with 10% of participants not tolerating the doses needed to gain a beneficial performance 83 effect (McNaughton et al. 2008). As dose increases, GI discomfort is more commonplace, often without additional performance improvements (McNaughton, 1991; Kahle et al. 2013). To combat GI 84 symptoms, stacking dose strategies has been implemented (Sale et al. 2011; Saunders et al. 2014a); 85 splitting larger doses (0.3 gkg<sup>-1</sup>BM) into smaller separate doses across a longer timeframe (0.2 gkg<sup>-1</sup>BM) 86 87 <sup>1</sup>BM followed by 0.1 g kg<sup>-1</sup>BM). How blood bicarbonate concentrations are altered following different 88 dosages of NaHCO<sub>3</sub> requires further investigation.

- 90 Therefore, the present study investigated bicarbonate, pH, base excess and sodium (Na<sup>+</sup>) responses to
- 91 three different doses of NaHCO<sub>3</sub> to determine the time course of changes and the inter-individual
- 92 variability in responses.

#### 93 METHODS

#### 94 **Participants**

Eighteen participants volunteered to participate in this non-blinded, order-balanced, crossover study.
Two participants withdrew due to GI distress, meaning that sixteen healthy males (age, 23±2 years;
height, 1.80±0.07 m; body mass, 78.6±15.1 kg) completed all aspects of the study. Participants provided
written informed consent and completed a health screen questionnaire prior to taking part in the study,
which was first approved by the Nottingham Trent University Ethical Advisory Committee. Participants
had not ingested any nutritional supplement or suffered from any GI problems in the previous 6 months.

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## **102 Protocol and measurements**

Participants attended three supplementation sessions at the same time of day, in at least a 4 h postprandial state and having replicated 24 h dietary intake. Participants were instructed to abstain from alcohol and strenuous/unaccustomed exercise for 24 h prior to each assessment, with caffeine prohibited on test days. Compliance with these requests was verbally confirmed prior to each session. Participants ingested a single dose of either 0.1, 0.2 or 0.3 g·kg<sup>-1</sup>BM of NaHCO<sub>3</sub> (Intralabs, UK) in clear gelatine capsules. Supplements were independently tested by HFL Sports Science, UK, ensuring no contamination with steroids or stimulants according to ISO 17025 accredited tests.

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Fingertip capillary blood was obtained before participants ingested NaHCO<sub>3</sub> with 500 ml of water. Following ingestion, blood was obtained every 10 min for 1 h, and then every 15 min for a further 2 h, during which time participants rested in a seated position. 80  $\mu$ L of whole blood was collected in a heparin-coated clinitube (Radiometer Ltd, UK), and immediately analysed for pH, bicarbonate and Na<sup>+</sup> concentrations with base excess being calculated (Radiometer ABL 900, UK).

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## 117 Statistical Analysis

118 Based on an *a priori* power calculation (using Ducker et al. 2013); a minimum of 12 participants were 119 required to achieve 95% power at P<0.01, with 18 participants recruited to allow for dropouts. Statistical analyses were completed using SPSS version 22 (SPSS Inc., Chicago, IL, USA) and Microsoft Excel 120 121 (Microsoft Inc., USA). Data were analysed using two-way (condition X time) repeated measures 122 ANOVA. Assessed variables were tested for normality using the Shapiro-Wilks test, and for homogeneity using the Levene test. A Greenhouse-Geisser correction was applied when Mauchly's test 123 indicated that sphericity assumptions were violated. Blood analytes at each time-point were compared 124 using a one-way ANOVA, with significance based on Bonferroni-corrected p-values. Net area under 125 the curve (AUC) was calculated (as per Gannon et al. 1989), and compared using a one-way ANOVA 126 with Bonferroni-corrected post hoc analysis. Linear regression analyses were performed to investigate 127 relationships between baseline and absolute changes in bicarbonate concentrations. Statistical 128 129 significance was accepted at P $\leq$ 0.05, with data presented as mean  $\pm$  1 standard deviation (SD).

## 131 **RESULTS**

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**132** The AUC was significantly greater for bicarbonate (0.1 g·kg<sup>-1</sup>BM:  $314.4\pm96.1$  mmol·L<sup>-1</sup>.180min<sup>-1</sup>; 0.2

- g'kg<sup>-1</sup>BM: 697.7±122.8 mmol·L<sup>-1</sup>.180min<sup>-1</sup>, 0.3 g'kg<sup>-1</sup>BM: 915.7±182.2 mmol·L<sup>-1</sup>.180min<sup>-1</sup>), pH (0.1
  g'kg<sup>-1</sup>BM: 5.05±2.60 pH units 180min<sup>-1</sup>; 0.2 g'kg<sup>-1</sup>BM: 9.03±2.93 pH units 180min<sup>-1</sup>; 0.3 g'kg<sup>-1</sup>BM:
- 135  $10.35\pm3.97$  pH units 180min<sup>-1</sup>), base excess (0.1 gkg<sup>-1</sup>BM: 379.6±122.0 mEq·L<sup>-1</sup>.180min<sup>-1</sup>; 0.2 gkg<sup>-1</sup>
- $136 \qquad ^{1}BM: 824.4 \pm 156.7 \ \text{mEq}\cdot\text{L}^{\text{-1}}180 \text{min}^{\text{-1}}; \ 0.3 \ \text{g}\cdot\text{kg}^{\text{-1}}BM: 1078.6 \pm 210.3 \ \text{mEq}\cdot\text{L}^{\text{-1}}180 \text{min}^{\text{-1}}) \ \text{and} \ \text{Na}^{\text{+}} \ (0.1 \ \text{g}\cdot\text{kg}^{\text{-1}}) = 0.3 \ \text{mEq}\cdot\text{L}^{\text{-1}} = 0.3 \ \text{mEq}\cdot\text$
- 137 <sup>1</sup>BM: -48.7 $\pm$ 195.7 mmol·L<sup>-1</sup>.180min<sup>-1</sup>; 0.2 g·kg<sup>-1</sup>BM: 111.4 $\pm$ 223.5 mmol·L<sup>-1</sup>.180min<sup>-1</sup>; 0.3 g·kg<sup>-1</sup>BM:

358.6±292.5 mmol·L<sup>-1</sup>·180min<sup>-1</sup>) following 0.3 g·kg<sup>-1</sup>BM compared to 0.2 g·kg<sup>-1</sup>BM (with the exception

- of pH responses; P $\leq$ 0.05) and 0.1 g·kg<sup>-1</sup>BM doses (P $\leq$ 0.05). Overall responses to 0.2 g·kg<sup>-1</sup>BM were
- 140 significantly greater than 0.1 g kg<sup>-1</sup>BM (P $\leq$ 0.05).
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Baseline bicarbonate ( $F_{(2,30)}=2.0$ ; P=0.20), pH ( $F_{(2,30)}=0.7$ ; P $\leq 0.51$ ), base excess ( $F_{(2,30)}=1.7$ ; P $\leq 0.20$ ) and Na<sup>+</sup> ( $F_{(2,30)}=0.3$ ; P $\leq 0.78$ ) levels (Table 1) were not significantly different between doses. There was a significant main effect of time for bicarbonate ( $F_{(14,210)}=72.6$ ; P $\leq 0.001$ ), pH ( $F_{(14,210)}=39.8$ ; P $\leq 0.001$ ), base excess ( $F_{(14,210)}=70.5$ ; P $\leq 0.001$ ) and Na<sup>+</sup> ( $F_{(14,210)}=11.3$ ; P $\leq 0.001$ ) levels, with increases following NaHCO<sub>3</sub> ingestion under all supplemental conditions (Table 1).

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There was a significant main effect of NaHCO<sub>3</sub> dose on bicarbonate ( $F_{(2,30)}$ =53.0; P $\leq$ 0.001), pH 148  $(F_{(2,30)}=18.4; P \le 0.001)$ , base excess  $(F_{(2,30)}=56.2; P \le 0.001)$  and Na<sup>+</sup>  $(F_{(2,30)}=27.0; P \le 0.001)$  levels. Post 149 150 *hoc* analysis showed that the responses of all blood analytes were significantly lower following 0.1 g kg<sup>-</sup> 151 <sup>1</sup>BM than following 0.2 g·kg<sup>-1</sup>BM (all P≤0.001 with the exception of pH [P>0.05]) and 0.3 g·kg<sup>-1</sup>BM doses (P≤0.003; Table 1). Bicarbonate concentrations (P≤0.01) and base excess (P≤0.001) were 152 significantly higher following 0.3 g kg<sup>-1</sup>BM compared to 0.2 g kg<sup>-1</sup>BM, although there were no 153 significant differences in pH and Na<sup>+</sup> concentrations between these doses. There were significant dose 154 by time interactions for bicarbonate (F<sub>(28,420)</sub>=17.2; P≤0.001), pH (F<sub>(28,420)</sub>=5.4; P≤0.001), base excess 155

156  $(F_{(28,420)}=18.4; P \le 0.001)$ , and Na<sup>+</sup>  $(F_{(28,420)}=5.0; P \le 0.001)$  responses; time point comparisons for blood 157 analytes are displayed in Table 1.

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Across each time interval there was large variability in the responses of blood analytes following each NaHCO<sub>3</sub> dose (Table 1). From this point the results will focus solely on blood bicarbonate concentrations in the interests of brevity and given that this is the primary outcome measure of interest. With respect to bicarbonate concentrations, the greatest variability in responses occurred between ~20 and 75 min after ingestion (Table 1). Variability was not reduced when data were normalised for body mass (data not shown). Individual blood bicarbonate responses are displayed in Figure 1.

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166 The absolute increases in bicarbonate concentrations from baseline to peak values (Figure 2) were 167 significantly greater following the ingestion of 0.3 g kg<sup>-1</sup>BM ( $8.2\pm1.4$  mmol·L<sup>-1</sup>) than ingestion of 0.1  $gkg^{-1}BM$  (3.6±0.8 mmol·L<sup>-1</sup>; P≤0.001) or 0.2  $gkg^{-1}BM$  (6.1±0.9 mmol·L<sup>-1</sup>; P≤0.001). The magnitude 168 of responses ranged from 2.0-5.0 mmol·L<sup>-1</sup> for 0.1 g·kg<sup>-1</sup>BM, 5.1-8.1 mmol·L<sup>-1</sup> for 0.2 g·kg<sup>-1</sup>BM and 169 170 6.0-12.3 mmol·L<sup>-1</sup> for 0.3 g·kg<sup>-1</sup>BM doses (Figure 2). One participant achieved an increase of 5 mmol·L<sup>-</sup> <sup>1</sup> from baseline, with none achieving an increase of 6 mmol· $L^{-1}$  from baseline following 0.1 gkg<sup>-1</sup>BM 171 (Table 2). With the ingestion of 0.2 gkg<sup>-1</sup>BM, all 16 participants achieved an increase of 5 mmol· $L^{-1}$ 172 from baseline and 9 participants achieved an increase of 6 mmol·L<sup>-1</sup> from baseline (Table 2). All 173 participants achieved an increase of 6 mmol  $\cdot$ L<sup>-1</sup> from baseline following the ingestion of 0.3 g kg<sup>-1</sup>BM 174 175 (Table 2).

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Individual magnitudes of responses between baseline and peak values were ranked, with only three participants (1, 7 and 10) consistently in the greatest 8 responders and three participants (4, 6 and 15) consistently in the least 8 responders on each dose (Table 2). The magnitudes of the responses were more consistent within participants when comparing 0.2 and 0.3 gkg<sup>-1</sup>BM doses (Table 2); 6 participants (1, 7, 9, 10, 12, 16) were in the greatest 8 responders and 6 participants (4, 6, 8, 13, 14, 15)

were in the least 8 responders. There was a significant difference in time-to-peak blood bicarbonate concentrations ( $F_{(2,30)}=15.7$ , P $\leq 0.001$ ) between doses (Table 2). The time between ingestion and peak responses of blood bicarbonate demonstrated high inter-individual variability, with times ranging from 30-150 min (mean: 78 min; CoV: 44%) following 0.1 g kg<sup>-1</sup>BM, 40-165 min (mean: 98; CoV: 32%) following 0.2 g kg<sup>-1</sup>BM and 75-180 min (mean: 123 min; CoV: 29%) following 0.3 g kg<sup>-1</sup>BM. No relationship between baseline bicarbonate concentrations and the subsequent increase in response to NaHCO<sub>3</sub> supplementation was shown for any dose (0.1 g kg<sup>-1</sup>BM: R<sup>2</sup>=0.01; 0.2 g kg<sup>-1</sup>BM: R<sup>2</sup>=0.19; 0.3

189  $g k g^{-1} BM: R^2 = 0.01$ ).

#### 190 **DISCUSSION**

191 This is the first study to report blood analyte responses from 15 time points over 3 hrs, with a high temporal frequency of sampling, following NaHCO<sub>3</sub> ingestion at three differing doses. Despite 192 193 individualising NaHCO<sub>3</sub> dosing (based on individual body mass) a high degree of inter-individual 194 variability existed with regards to the magnitude of change in blood analyte levels and the time to peak. 195 The magnitude of the increase in blood analytes was dose-dependent, with greater increases achieved with larger doses of NaHCO<sub>3</sub>, although the range in responses was also greater at these highest dose. 196 197 These data challenge the most commonly suggested supplementation protocol of 0.3 gkg<sup>-1</sup>BM 198 administered ~60 min prior to performance (McNaughton, 1991; Siegler et al. 2012; Duncan et al. 2014), which is unlikely to result in optimal blood biochemistry for all individuals. It is difficult to 199 200 compare the time-course relationship following ingestion due to existing data being focused on either pre- to post-exercise comparisons, or due to infrequent sample collection (Renfree et al. 2007; Siegler 201 202 et al. 2010; Carr et al. 2011; Miller et al. in press). Here we extend previous work examining the effect 203 of NaHCO<sub>3</sub> ingestion on acid-base responses (Renfree et al. 2007; Siegler et al. 2010; Carr et al. 2011; Miller et al. *in press*), by employing a much greater temporal resolution (every 10 min) in sampling. 204 The mean time-to-peak for bicarbonate and pH responses following ingestion of 0.3 gkg<sup>-1</sup>BM was 205 206 greater than the 60-90 min previously documented (Renfree, 2007; Price & Singh, 2008; Siegler et al. 207 2010); even when ingesting smaller doses (>60 min). Time-to-peak for all variables increased in a stepwise manner relative to dose; blood pH peaked at 75 (0.1 gkg<sup>-1</sup>BM), 105 (0.2 gkg<sup>-1</sup>BM) and 120 min 208 (0.3 g·kg<sup>-1</sup>BM) post-ingestion. Our data suggest that the time intervals used in previous studies might 209 210 lead to some misinterpretation of findings relating to optimal blood analyte responses. It remains unclear as to why high variability exists in time-to-peak when ingestion of NaHCO<sub>3</sub> was conducted 211 212 within a small and structured time period (10 min). Numerous factors which could explain this 213 variability, thus providing an avenue for future investigation.

If we use the 6 mmol·L<sup>-1</sup> above baseline cut-off for blood bicarbonate responses, as suggested by Carr et al. (2011) to provide an ergogenic effect, it is clear that a dose of 0.3 g·kg<sup>-1</sup>BM remains the most

217 relevant to ensure that all individuals reach this zone (Figure 2). Following 0.3 gkg<sup>-1</sup>BM, absolute changes in blood bicarbonate ranged between 6.0 and 12.3 mmol<sup>-1</sup>, with time-to-peak varying 218 between 75 and 180 min. This demonstrates that the time taken for individuals to achieve peak 219 220 concentrations or even performance relevant blood bicarbonate changes (Carr et al. 2011) is highly 221 variable, suggesting a need to consider individual responses to NaHCO<sub>3</sub> supplementation (Figure 1). Practically, an *a priori* knowledge of an individual's blood responses following ingestion is required to 222 223 optimise outcomes. What is not yet clear is whether or not individuals respond consistently to the same 224 dose of  $NaHCO_3$  or what factors influence bicarbonate release (e.g., nutritional impact of gastric 225 emptying), providing an avenue for further work.

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The current investigation might also help to explain discrepancies previously shown in relation to the 227 ergogenic effect of NaHCO<sub>3</sub> ingestion (for review see Carr et al. 2011), where numerous 228 229 methodological differences relating dosing strategy were employed. In the current study, to provide consistency, participants were instructed to consume all capsules within 10 min, as per Siegler et al. 230 (2010). The time taken to ingest NaHCO<sub>3</sub> is often unreported or is >30 min (Carr et al. 2011), which 231 232 would theoretically cause more variability in individual peak responses that those reported in the current study, as such comparisons to previous blood analyte responses are confounded. Gastric emptying has 233 234 shown considerable inter-individual variation (Paintaud et al. 1998; Barbosa et al. 2005), although there 235 is some consistency in intra-individual responses (Paintaud et al. 1998; Barbosa et al. 2005). These findings suggest that it might be important to replicate dietary intake prior to ingestion in order to 236 237 develop a more consistent response to NaHCO<sub>3</sub> ingestion. Participants in the current investigation replicated their 24 h dietary intake and remained fasted for 4 hrs prior to supplementation, where 90% 238 of food would be emptied from the stomach (Tougas et al. 2000). Meal volume, composition and texture 239 would, however, influence gastric emptying rates (Donohoe et al. 2009). An overnight fast would not 240 241 be representative of athlete behaviour and so we decided to use a 4 h fast to provide a balance between 242 experimental control and ecological validity. It should, however, be noted that the results of future studies might differ with alternative dietary intake patterns. During the current investigation non-243

244 arterialised fingertip capillary blood samples have been used to assess blood analyte responses. The PO<sub>2</sub> values for the current investigation were  $75.38 \pm 2.14$  for the 0.1 g·kg<sup>-1</sup>BM condition,  $74.14 \pm 2.61$  for 245  $0.2 \text{ gkg}^{-1}BM$  and  $73.18 \pm 2.63$  for the 0.3 gkg $^{-1}BM$  condition as an average across all time points. Non-246 warmed capillary blood samples are a useful and practical tool, reporting a strong correlation with 247 248 arterial samples for pH, HCO<sub>3</sub> and base excess variables (Yildizdas et al. 2004). This method is also inline with a number of previous investigations (Price & Simons, 2010; Bellinger et al. 2012; Siegler et 249 al. 2013; Saunders et al. 2014). In a small independent study, we confirmed that blood arterialisation 250 via warming the hand in a water bath (42°C) for 10 minutes did not alter blood gas parameters. 251 252 Nonetheless, it is important to suggest caution when comparing non-arterialised with arterialised 253 samples.

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255 Blood bicarbonate concentrations were similar over the first 30 min following ingestion of all NaHCO<sub>3</sub> 256 doses; blood pH also followed a similar pattern for the first 60 min post-ingestion. These findings questions the use of doses above 0.1 gkg<sup>-1</sup>BM when the time between ingestion and performance is 257 relatively short (*i.e.*, following a high-intensity warm-up or when an athlete has multiple events over a 258 259 short period of time), especially when the same level of bicarbonate manipulation is achievable. In these situations it would also be advisable to consume 0.1 g·kg<sup>-1</sup>BM of NaHCO<sub>3</sub>, given that lower doses 260 261 reduce the intensity and/or frequency of negative GI symptoms (McNaughton, 1992; Kahle et al. 2013), which would benefit athletes in the competitive setting. Some athletes require co-ingestion of NaHCO<sub>3</sub> 262 with food and fluid in order to reduce GI symptoms, therefore lowering the dose could lead to a 263 264 reduction in the amount of food/fluid ingested, vital for athletes competing numerous times within a short period. 265

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Following large quantities of NaHCO<sub>3</sub>, carbonic acid formation occurs in the stomach and Na<sup>+</sup> absorption and Na<sup>+</sup> plasma concentration both increase (Heigenhauser, 1991). As the physiochemical equilibrium shifts, water and CO<sub>2</sub> increase in the blood, thereby increasing CO<sub>2</sub> partial pressure (as described by the Henderson-Hasselbalch equation). This mechanism alters the already acidic

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271 environment of the stomach, which can result in GI distress, including stomach bloating, nausea, and 272 diarrhoea (McNaughton, 1992; Siegler et al. 2012). In the present study following NaHCO<sub>3</sub> ingestion we have shown increased plasma Na<sup>+</sup> concentrations, with the mean change being two times greater 273 following 0.3 gkg<sup>-1</sup>BM (4 mmol<sup>-1</sup>) compared to 0.1 gkg<sup>-1</sup>BM (2 mmol<sup>-1</sup>). The peak change in Na<sup>+</sup> 274 275 concentrations following 0.2 and 0.3 gkg<sup>-1</sup>BM occurred ~105 minutes post-ingestion, which broadly corresponds to the timeframe of the greatest incidence of GI distress (~90 min following ingestion; Carr 276 et al. 2011). The inter-individual variability in the magnitude of change in Na<sup>+</sup> concentrations might 277 explain why some individuals report GI distress, whilst others do not, even at the same NaHCO<sub>3</sub> dose. 278

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In conclusion, the present data challenges the most commonly implemented NaHCO<sub>3</sub> supplementation protocol and its efficacy to enhance buffering capacity and exercise performance for all individuals. Due to the large inter-individual responses shown, individual and mean responses should be included in future research and knowledge of the individual responses to NaHCO<sub>3</sub> supplementation is essential in the applied setting. For individuals needing to ingest NaHCO<sub>3</sub>  $\leq$  30 min prior to the onset of exercise, smaller doses can be ingested with no negative consequences for the additional extracellular buffering potential.

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 TS, GGA, BS and CS; data were collected and analysed by RLS; data interpretation and manuscript

- 296 preparation were undertaken by RLS, TS, SC, GGA, BS and CS. All authors approved the final version
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#### 298 **REFERENCES**

- Afman, G., Garside, R.M., Dinan. N., Gant. N., Betts, J.A., & Williams, C. (2014). Effect of
  carbohydrate or sodium bicarbonate ingestion on performance during a validated basketball simulation
  test. *International Journal of Sports Nutrition Exercise Metabolism*, 24, 6, 632-644.
- Barbosa, L., Vera, H., Moran, S., Del Prado, M., & Lopez Alarcon, M. (2005). Reproducibility and
  reliability of the 13C-acetate breath test to measure gastric emptying of liquid meal in
  infants. *Nutrition*, 21, 289–294.
- Bruton, J.D., Lannergren, J., & Westerblad, H. (1998). Effects of CO<sub>2</sub>-induced acidification on the
- fatigue resistance of single mouse muscle fibers at 28°C. *Journal of Applied Physiology*, 85, 478–483.
- Burke, L.M., & Pyne, D.B. (2007). Bicarbonate loading to enhance training and competitive
  performance. *International Journal of Sports Physiology and Performance*, 2, 93-97.
- Cairns, S.P. (2006). Lactic acid and exercise performance: culprit or friend? *Sports Medicine*, 36, 279–
  291.
- 311 Carr, A.J., Hopkins, W.G., & Gore, C.J. (2011). Effects of acute alkalosis and acidosis on performance:
- 312 A meta-analysis. *Sports Medicine*, 41, 10, 801-814.
- Carr, A.J., Slater, G.J., Gore, C.J., Dawson, B. & Burke, L.M. (2011). Effect of sodium bicarbonate on
  [HCO<sub>3</sub><sup>-</sup>], pH, and gastrointestinal symptoms. *International Journal of Sport Nutrition and Exercise Metabolism*, 21, 189-194.
- 316 Duncan, M.J., Weldon, A., & Price, M.J. (2014). The effect of sodium bicarbonate ingestion on back
- squat and bench press exercise to failure. *Journal of Strength and Conditioning Research*, 1358–1366.
- 318 Flinn, S., Herbert, K., Graham, K., & Siegler, J.C. (2014). Differential effect of metabolic alkalosis and
- 319 hypoxia on high-intensity cycling performance. Journal of Strength and Conditioning Research, 28,
- **320** 2852–2858.

- Forbes, S.C., Raymer, G.H., Kowalchuk, J.M., & Marsh, G.D. (2005). NaHCO<sub>3</sub>-induced alkalosis
  reduces the phosphocreatine slow component during heavy intensity forearm exercise. *Journal of Applied Physiology*, 99, 1668–1675.
- 324 Gannon, M.C., Nuttall, F.Q., Westphal, S.A., Neil, B.J., & Seaquist, E.R. (1989). Effects of dose of
- ingested glucose on plasma metabolite and hormone responses in type II diabetic subjects. *Diabetes Care*, 12, 544–552.
- Heigenhauser, G.J.F. (1991). Ergogenic enhancement of performance in exercise and sport: Bicarbonate
- 328 loading. In: D. R. Lamb and M. H. Williams (Ed.), *Prospective in Exercise Science and Sports Medicine*
- 329 (Vol. 4; pp. 183–212). Carmel, In: Cooper Publishing Group LLC.
- Hill, A.V. & Lupton, H. (1923). Muscular exercise, lactic acid, and the supply and utilization of oxygen. *QJM*, 16, 135-171.
- Joyce, S., Minahan, C., Anderson, M., & Osborne, M. (2011). Acute and chronic loading of sodium
  bicarbonate in highly trained swimmers. *European Journal of Applied Physiology*, 112, 1–9.
- Kahle, L., Kelly, P., Eliot, K., & Weiss, E. (2013). Acute sodium bicarbonate loading has negligible
  effects on resting and exercise blood pressure but causes gastrointestinal distress. *Nutrition Research*,
  33, 479–86.
- 337 Kozac-Collins, K., Burke, E.R., & Schoene, R. (1994). Sodium bicarbonate ingestion does not improve
- performance in women cyclists. *Medicine and Science in Sports and Exercise*, 26, 12, 1510-5.
- Kupcis, P.D., Slater, G.J., Pruscino, C.L., & Kemp, J.G. (2012). Influence of sodium bicarbonate on
  performance and hydration in lightweight rowing. *International Journal of Sport Physiological Performance*, 7, 11–18.
- Linderman J.K., & Gosselink K.L. (1994). The effects of sodium bicarbonate ingestion on exercise
  performance. *Sports Medicine*, 18, 75–80.
- 344 Matson, L.G., & Tran, Z.V. (1993). Effects of sodium bicarbonate ingestion on anaerobic performance:
- A meta-analytic review. *International Journal of Sport Nutrition*, 3, 1, 2–28.

- Maughan, R.J., King, D.S., & Lea, T. (2004). Dietary supplements. *Journal of Sports Sciences*, 22, 95–
  113.
- 348 McNaughton, L.R. (1992). Bicarbonate ingestion: Effects of dosage on 60s cycle ergometry. *Journal*349 *of Sports Sciences*, 10, 415-423.
- McNaughton, L.R., Siegler, J.C., & Midgley, A. (2008). The ergogenic effect of sodium bicarbonate.
   *Current Sports Medicine Reports*, 7, 230–236.
- Miller, P., Robinson, A., Sparks, A., Bridge, C.A., Bentley, D., & McNaughton, L.R. (*in press*). The
  effects of novel ingestion of sodium bicarbonate on repeated sprint ability. *Journal of Strength and Conditioning Research*.
- 355 Paintaud, G., Thibault, P., Queneau, P.E., Magnette, J., Berard, M., Rumbach, L., Bechtel, P.R., &
- 356 Carayon, P. (1998). Intraindividual variability of paracetamol absorption kinetics after a semisolid meal
- in healthy subjects. European Journal of Clinical Pharmacology, 53, 355–359.
- 358 Peart, D., Siegler, J., & Vince, R. (2012). Practical recommendations for coaches and athletes: a meta-
- analysis of sodium bicarbonate use for athletic performance. *The Journal of Strength & Conditioning Research*, 26, 1975–1983.
- 361 Pilegaard, H., Domino, K., Noland, T., Juel C., Hellsten Y. & Halestrap A.P. (1999). Effect of high-
- 362 intensity exercise training on lactate/H transport capacity in human skeletal muscle. American Journal
- 363 *of Physiology Endocrinology and Metabolism*, 276, 2, 255–261.
- Price, M.J., & Singh, M. (2008). Time course of blood bicarbonate and pH three hours after sodium
- 365 bicarbonate ingestion. *International Journal of Sports Physiology and Performance*, 3, 240–242.
- Renfree, A. (2007). The time course for changes in plasma [H<sup>+</sup>] after sodium bicarbonate ingestion.
- 367 International Journal of Sports Physiology and Performance, 2, 323–326.
- 368 Robergs, R.A. (2002). Blood acid-base buffering: explanation of the effectiveness of bicarbonate and
- 369 citrate ingestion. *Journal of Exercise Physiology*, 5, 3, 1-5.

- Sale, C., Hill, C.A., Ponte, J., & Harris, R.C. (2012). β-alanine Supplementation Improves Isometric
  Endurance of the Knee Extensor Muscles. *Journal of the International Society of Sports Nutrition*, 9, 1,
  26.
- Sale, C., Saunders, B., Hudson, S., Wise, J.A., Harris, R.C., & Sunderland, C.D. (2011). Effect of β-
- alanine plus sodium bicarbonate on high-intensity cycling capacity. *Medicine and Science in Sports and*
- 375 *Exercise*, 43 (10), 1972–1978.
- 376 Saunders, B., Sale, C., Harris, R.C., & Sunderland, C. (2014a). Effect of sodium bicarbonate and Beta-
- alanine on repeated sprints during intermittent exercise performed in hypoxia. *International Journal of*
- 378 Sport Nutrition and Exercise Metabolism, 24, 196–205.
- 379 Saunders, B., Sale, C., Harris, R.C., & Sunderland, C. (2014b). Sodium Bicarbonate and High-Intensity-
- 380 Cycling Capacity: Variability in Responses. International Journal of Sports Physiology and
  381 Performance, 9, 627-632.
- 382 Siegler, J.C., Marshall, P.W.M., Bray, J., & Towlson, C. (2012). Sodium bicarbonate supplementation
- and ingestion timing. *Journal of Strength and Conditioning Research*, 26, 1953–1958.
- 384 Siegler, J.C., Midgley, A.W., Polman, R.C., & Lever, R. (2010). Effects of various sodium bicarbonate
- 385 loading protocols on the time-dependent extracellular buffering profile. Journal of Strength and
- **386** *Conditioning Research,* 24, 2551–2557.
- 387 Stellingwerff, T., Boit, M.K., & Res, P.T. (2007). Nutritional strategies to optimize training and racing
- in middle distance athletes. *Journal of Sports Sciences*, 25, S1, S17-S28.
- 389 Thomas, C., Perrey, S., Lambert, K., Hugon, G., Mornet, D., & Mercier, J. (2005). Monocarboxylate
- transporters, blood lactate removal after supramaximal exercise, and fatigue indexes in humans. *Journal*
- *of Applied Physiology*, 98, 804-809.
- Vanhatalo, A., McNaughton, L.R., Siegler, J., & Jones, A.M. (2010). Effect of induced alkalosis on the
- 393 power-duration relationship for "All-out" Exercise. *Medicine and Science in Sports and Exercise*, 42,
- **394** 563–570.

- Westerblad, H., Bruton, J.D., & Katz, A. (2010). Skeletal muscle: Energy metabolism, fiber types,
  fatigue and adaptability. *Experimental cell research*, 316, 3093-3099.
- Westerblad, H., Bruton, J.D., & Lannergren, J. (1997). The effect of intracellular pH on contractile
  function of intact, single fibres of mouse muscle declines with increasing temperature. *Journal of Physiology*, 500, 193–204.
- 400 Yildizdas, D., Yapicioglu, H.L., Yilmaz, H.L. & Sertdemir, Y. (2004). Correlation of simultaneously
- 401 obtained capillary, venous and arterial blood gases of pateints in a paediatric intensive care unit.
- 402 Archives of Disease in Childhoof, 89, 176-180.
- 403 Zabala, M., Requena, B., Sanchez-Munoz, C., Gonzalez-Badillo, J.J., Garcia, I., Oopik, V., & Paasuke,
- 404 M. (2008). Effects of sodium bicarbonate ingestion on performance and perceptual responses in a
- 405 laboratory-simulated BMX cycling qualification series. Journal of Strength and Conditioning
- 406 *Research*, 22, 1645–1653.

# 407 **TABLES**

- 408 **Table 1:** Blood bicarbonate, pH, base excess and Na<sup>+</sup> responses across the 3 h duration following
- 409 NaHCO<sub>3</sub> ingestion. Mean time point comparisons are displayed for each blood analyte; <sup>x</sup> denotes a
- 410 significant difference between 0.1 and 0.2 g kg<sup>-1</sup>BM.  $^{\Delta}$  denotes a significant difference between 0.2 and
- 411 0.3 g·kg<sup>-1</sup>BM. All comparisons are based on Bonferroni-corrected p-values of  $\leq 0.003$ .

			Time post ingestion (min)														
			0	10	20	30	40	50	60	75	90	105	120	135	150	165	180
Bicarbonate (mmol·L <sup>-1</sup> )	0.1 g <sup>.</sup> kg <sup>.</sup> <sup>1</sup> BM	Mean	25.7	25.5	26.4	27.1	$27.5^{*}$	$27.9^{*}$	28.0*X	27.9 <sup>*X</sup>	28.0 <sup>*X</sup>	28.1 <sup>*X</sup>	27.9 <sup>*X</sup>	27.4 <sup>*X</sup>	27.4 <sup>*X</sup>	27.2 <sup>*X</sup>	27.2 <sup>*X</sup>
		SD	1.0	1.2	1.7	1.8	1.9	1.9	1.6	1.3	1.0	1.2	1.1	1.1	1.1	0.8	0.7
e (m	0.2 g <sup>-</sup> kg <sup>-</sup> <sup>1</sup> BM	Mean	25.1	25.3	25.9	27.3	28.4	28.9	29.5	30.1	30.5∆	30.5∆	30.1∆	29.9∆	29.6 <sup>∆</sup>	29.3∆	29.1∆
nato		SD	0.87	1.27	1.58	1.74	1.71	1.62	1.19	1.04	0.97	0.85	1.19	1.18	1.24	0.84	0.92
arbo	0.3 g <sup>.</sup> kg <sup>-</sup> <sup>1</sup> BM	Mean	25.5	25.6	26.7	27.9	29.4	30.0	30.6	31.7	32.1	32.3	32.4	32.2	32.2	31.7	31.4
Bic		SD	1.34	1.40	1.65	1.80	1.90	1.96	2.06	1.94	1.98	1.87	2.14	1.89	2.26	1.53	1.40
	0.1 g <sup>.</sup> kg <sup>-</sup> <sup>1</sup> BM	Mean	7.42	7.43	7.44	7.44	7.45	7.46	$7.46^{*}$	$7.46^{*}$	7.46 <sup>*X</sup>	$7.46^{*}$	7.45 <sup>*X</sup>	7.45 <sup>*X</sup>	$7.45^{*}$	$7.45^{*}$	7.44
		SD	0.01	0.02	0.03	0.02	0.03	0.02	0.02	0.02	0.01	0.02	0.02	0.02	0.03	0.02	0.02
μH	0.2 cd.c-	Mean	7.42	7.42	7.43	7.45	7.46	7.47	7.47	7.47	7.48	7.48	7.48	7.48	7.48	7.47	7.46
[d	0.2 g <sup>·</sup> kg <sup>-</sup> <sup>1</sup> BM	SD	0.02	0.02	0.03	0.03	0.03	0.04	0.03	0.02	0.02	0.02	0.02	0.02	0.03	0.03	0.03
	0.3 g·kg <sup>-</sup> <sup>1</sup> BM	Mean	7.42	7.42	7.44	7.45	7.47	7.48	7.48	7.49	7.50	7.50	7.50	7.50	7.49	7.49	7.49
		SD	0.02	0.02	0.03	0.02	0.02	0.02	0.02	0.02	0.02	0.02	0.02	0.03	0.03	0.03	0.02
[-]	0.1 g <sup>.</sup> kg <sup>-</sup> <sup>1</sup> BM	Mean	1.59	1.32	2.51	3.37*	3.84*	4.28 <sup>*X</sup>	4.35 <sup>*X</sup>	4.23 <sup>*X</sup>	4.45 <sup>*X</sup>	4.48 <sup>*X</sup>	4.23*X	3.73 <sup>*X</sup>	3.63 <sup>*X</sup>	3.51 <sup>*X</sup>	3.47 <sup>*X</sup>
EqI		SD	1.29	1.54	2.02	2.23	2.27	2.25	1.93	1.70	1.26	1.41	1.30	1.28	1.28	0.92	0.84
Base Excess (mEqL <sup>-1</sup> )	0.2 g·kg <sup>-</sup> <sup>1</sup> BM	Mean	0.96	1.12	1.71	3.43	4.71	5.28	6.09∆	6.68∆	7.18∆	7.29∆	6.90∆	6.74∆	6.40 <sup>Δ</sup>	6.16 <sup>∆</sup>	5.87∆
xces		SD	1.10	1.55	1.86	2.10	2.11	2.00	1.57	1.30	1.24	1.04	1.32	1.40	1.49	1.08	1.11
se E3	0.3 g <sup>.</sup> kg <sup>.</sup> <sup>1</sup> BM	Mean	1.44	1.53	2.89	4.34	6.01	6.76	7.54	8.66	9.07	9.34	9.58	9.22	9.29	8.75	8.43
Bas		SD	1.66	1.76	2.05	2.25	2.26	2.35	2.41	2.23	2.31	2.12	2.38	2.09	2.43	1.68	1.53
	0.1 g·kg <sup>-</sup> <sup>1</sup> BM	Mean	143	142	142	142	143	143	142*	143*X	142*X	143*	142*	143*x	143*x	142*	142*
$\mathbf{L}^{-1}$		SD	1	1	2	2	1	1	1	1	1	1	2	1	1	1	1
NA <sup>+</sup> (mmol·L <sup>-1</sup> )	0.2 g·kg <sup>-</sup> <sup>1</sup> BM	Mean	142	141	142	142	143	143	143	143	144	144	144	144	143	143	143
		SD	2	1	2	2	2	2	2	1	1	2	1	2	1	1	1
-NA-	0.3 g <sup>.</sup> kg <sup>.</sup> <sup>1</sup> BM	Mean	142	142	142	143	144	144	144	145	145	145	145	145	145	145	145
		SD	2	3	2	2	2	2	2	2	1	1	1	1	2	1	2

**Table 2:** Individual blood bicarbonate responses following NaHCO<sub>3</sub> ingestion across supplemental condition. Absolute and percentage change in bicarbonate responses refer to the difference between baseline and peak concentrations; absolute changes of  $\geq$ 5 mmol·L<sup>-1</sup> are highlighted in bold. Position based on response ranks participants on absolute change in descending order, highest response equates to 1, whilst lowest absolute change equates to 16. Significant differences between supplementation conditions for absolute change and time-to-peak are denoted by \* (0.1 and 0.3 g/kg<sup>-1</sup>BM) × (0.1 and 0.2 g/kg<sup>-1</sup>BM) and  $^{\Delta}$  (0.2 and 0.3 g/kg<sup>-1</sup>BM; P≤0.05).

		0.	1 g∙kg⁻¹BM				0.	2 g <sup>.</sup> kg <sup>-1</sup> BM		0.3 g·kg <sup>-1</sup> BM				
Participant number	Baseline (mmol <sup>-</sup> L <sup>-1</sup> )	Absolute Change (mmol·L <sup>-1</sup> )	Percentage change (%)	Time- to-peak (min)	Position based on response	Baseline (mmol·L <sup>-1</sup> )	Absolute Change (mmol·L <sup>-1</sup> )	Percentage change (%)	Time- to-peak (min)	Position based on response	Baseline (mmol·L <sup>-1</sup> )	Absolute Change (mmol <sup>-</sup> L <sup>-1</sup> )	Percentage change (%)	Time- I to-peak b (min) r
1	23.7	3.9	16.5	90	5	23.2	6.9	29.7	90	3	23.7	8.9	37.6	165
2	25.1	4.4	17.5	50	3	25.7	6.0	23.3	90	8	24.8	8.9	35.9	90
3	25.9	2.7	10.4	120	14	25.5	6.0	23.5	120	9	25.5	8.1	31.8	105
4	24.6	2.0	8.1	90	16	23.9	5.5	23.0	105	12	25.4	7.0	27.6	90
5	25.3	3.8	15.0	120	6	24.7	5.6	22.7	105	10	25.1	8.5	33.9	150
6	25.9	3.1	12.0	90	12	25.2	5.3	21.0	120	13	25.2	6.9	27.4	150
7	25.1	3.8	15.1	50	7	24.9	6.4	25.7	90	6	27.1	8.8	32.5	120
8	26.6	4.9	18.4	105	2	25.7	5.1	19.8	105	15	28.5	7.7	27.0	120
9	26.6	2.6	9.8	75	15	25.3	7.1	28.1	120	2	26.7	12.3	46.1	150
10	24.9	4.3	17.3	150	4	24.9	6.6	26.5	135	5	23.1	8.6	37.2	180
11	25.0	3.1	12.4	90	13	24.9	6.4	25.7	90	7	24.2	7.0	28.9	180
12	26.4	3.3	12.5	50	10	24.5	8.1	33.1	90	1	25.9	8.6	33.2	105
13	24.9	5.0	20.1	30	1	26.7	5.5	20.6	40	11	26.7	6.0	22.5	90
14	27.6	3.7	13.4	50	8	26.6	5.2	19.5	165	14	26.1	8.3	31.8	120
15	26.8	3.2	11.9	40	11	25.1	5.1	20.3	60	16	24.9	6.6	26.5	75
16	26.2	3.4	13.0	50	9	25.2	6.7	26.6	50	4	25.4	8.4	33.1	75
Mean	25.7	3.6 *X	14.0	78 *		25.1	6.1 <sup>Δ</sup>	24.3	98 △		25.5	8.2	32.0	123
SD	1.0	0.8	3.3	34		0.9	0.9	3.9	32		1.3	1.4	5.6	36
Min	23.7	2.0	8.1	30		23.2	5.1	19.5	40		23.1	6.0	22.5	75
Max	27.6	5.0	20.1	150		26.7	8.1	33.1	165		28.5	12.3	46.1	180

## FIGURE LEGENDS

Figure 1: Individual blood bicarbonate responses across the 3 hr following NaHCO<sub>3</sub> ingestion at 0.1(A), 0.2 (B) and 0.3 g·kg<sup>-1</sup>BM (C).

Figure 2: Mean absolute change in bicarbonate concentrations across 15 intervals (3 hr) following ingestion of 0.1 (open circles), 0.2 (solid square) and 0.3 g·kg<sup>-1</sup>BM (open triangle) of NaHCO<sub>3</sub>. Zone of ergogenic effect (+6 mmol·L<sup>-1</sup>) is based on concentrations from Carr et al. (2011).

# FIGURES

Figure 1

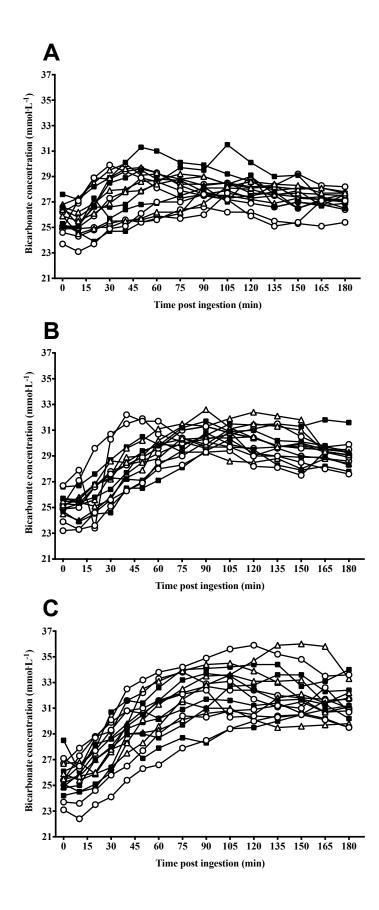


Figure 2

