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1 **Variation in early-life telomere dynamics in a long-lived**
2 **bird: links to environmental conditions and survival**

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16 **Summary**

17 Conditions experienced during early life can have profound consequences for both short and
18 long-term fitness. Variation in the natal environment has been shown to influence survival
19 and reproductive performance of entire cohorts in wild vertebrate populations. Telomere
20 dynamics potentially provide a link between the early environment and long-term fitness
21 outcomes, yet we know little about how the environment can influence telomere dynamics in
22 early life. We found that environmental conditions during growth have an important
23 influence on early-life telomere length (TL) and attrition in nestlings of a long-lived bird, the
24 European storm petrel *Hydrobates pelagicus*. Nestlings reared under unfavourable
25 environmental conditions experienced significantly greater telomere loss during postnatal
26 development compared with nestlings reared under more favourable natal conditions, which
27 displayed a negligible change in TL. There was, however, no significant difference in pre-
28 fledging TL between cohorts. The results suggest that early-life telomere dynamics could
29 contribute to the marked differences in life-history traits that can arise among cohorts reared
30 under different environmental conditions. Early-life TL was also found to be a significant
31 predictor of survival during the nestling phase, providing further evidence for a link between
32 variation in TL and individual fitness. To what extent the relationship between early-life TL
33 and mortality during the nestling phase is a consequence of genetic, parental and
34 environmental factors is currently unknown, but an interesting area for future research.
35 Accelerated telomere attrition under unfavourable conditions, as observed in this study, might
36 play a role in mediating the effects of the early-life environment on later-life performance.

37 **Introduction**

38 One of the principal aims of evolutionary ecology is to understand the mechanisms
39 underlying individual variation in longevity and fecundity. Telomere dynamics link cellular
40 processes with organismal ageing and thus optimisation of telomere length (TL) and attrition
41 may play a major role in life-history evolution (Monaghan and Haussmann, 2006). Telomeres
42 comprise highly-conserved non-coding DNA sequences that form protective caps at the ends
43 of eukaryotic chromosomes (Blackburn, 2005). By protecting coding sequences from
44 attrition, telomeres play an important role in maintaining genome stability (reviewed by
45 Verdun and Karlseder, 2007). In the absence of the enzyme telomerase, telomeres shorten
46 with each round of somatic cell division; when a critical length is reached, telomeres become
47 dysfunctional and cells enter a state of replicative senescence (Blackburn, 2005; Verdun and
48 Karlseder, 2007). Senescent cells subsequently die or adopt an altered secretory profile,
49 resulting in secretion of inflammatory cytokines, growth factors and degradative enzymes
50 that contribute to age-related declines in tissue and organ function (Campisi, 2005). The
51 accumulation of senescent cells thus appears to be important to the ageing phenotype, thereby
52 influencing lifespan (Campisi, 2005).

53 Longitudinal studies have shown that TL declines progressively with age in many
54 vertebrates (e.g. Zeichner et al., 1999; Brümmendorf and Mak; 2002; Salomons et al., 2009;
55 Aviv et al., 2009; Bize et al., 2009; Heidinger et al., 2012; Barrett et al., 2013). Large within-
56 species variability in TL and the rate of telomere shortening is reported among individuals of
57 the same age (Hall et al., 2004; Aviv et al., 2009; Bize et al., 2009). Inter-individual variation
58 in TL has been found to predict fitness components in natural populations; individuals with
59 the shortest telomeres or the highest loss rate have the poorest survival prospects (Cawthon et
60 al., 2003; Haussmann et al., 2005; Bize et al., 2009; Olsson et al., 2011; Heidinger et al.,
61 2012) and TL was found to be positively correlated with lifetime reproductive success
62 (Pauliny et al., 2006). While early-life TL is partly determined by genetic factors (Njajou et
63 al., 2007; Olsson et al., 2011), much of the inter-individual variation in TL may relate to
64 environmental influences. Exposure to repeated stress (Epel et al., 2004; Kotrschal et al.,
65 2007; Herborn et al.; 2014), large-scale climatic processes (Mizutani et al., 2013), low habitat
66 quality (Angelier and Vleck, 2013) and reproduction (Kotrschal et al., 2007; Heidinger et al.,
67 2012) have all been associated with accelerated telomere attrition, possibly mediated by
68 increased oxidative damage (von Zglinicki, 2002). TL therefore potentially reflects variation
69 in individual state and past experiences.

70 It is well known that early-life conditions can have profound influences on phenotypic
71 development and long-term fitness consequences (Lindström, 1999; Metcalfe and Monaghan,
72 2001). Variation in the pre- and early post-natal environment has been shown to influence
73 survival and reproductive performance in a number of vertebrate species (Albon et al., 1987;
74 Haywood and Perrins, 1992; Sedinger et al., 1995; Rose et al., 1998; Reid et al., 2003).
75 Environmental conditions during early life can affect an entire cohort simultaneously, giving
76 rise to substantial differences in life histories between successive cohorts, which can persist
77 throughout the cohort's lifespan (Albon et al., 1987; Rose et al., 1998; Reid et al., 2003).
78 Several studies on mammalian and avian species have found that telomere loss is greatest
79 during early life, presumably as a result of the rapid growth and cell division that occurs
80 during this period (Zeichner et al., 1999; Baerlocher et al., 2007; Salomons et al., 2009). Poor
81 early nutrition and catch-up growth have been shown to result in accelerated telomere loss in
82 a number of studies, including laboratory rats (*Rattus norvegicus*: Jennings et al., 1999;
83 Tarry-Adkins et al., 2009) and wild birds (European shag *Phalacrocorax aristotelis*: Hall et
84 al., 2004; king penguin *Aptenodytes patagonicus*: Geiger et al., 2012). Developmental stress
85 has also been shown to accelerate telomere shortening through experimental brood
86 enlargement (jackdaw *Corvus monedula*: Boonekamp et al., 2014) and increased nestling
87 exposure to glucocorticoids (European shag: Herborn et al., 2014) in wild birds. Early-life TL
88 was found to be a better predictor of longevity than TL in adulthood in a longitudinal study of
89 captive zebra finches *Taeniopygia guttata* (Heidinger et al., 2012). Early-life telomere
90 dynamics may therefore mechanistically link developmental conditions with later-life
91 senescence (Monaghan, 2010). Despite the potential significance of early-life telomere
92 dynamics in influencing life-history traits, we still know relatively little about how variation
93 in early-life conditions influences telomere loss during development in natural environments.

94 In this study, we examined the effects of inter-annual variation in the natal
95 environment on TL and telomere dynamics during postnatal development in the European
96 storm petrel *Hydrobates pelagicus* (hereafter, storm petrel). The storm petrel is a remarkably
97 long-lived seabird, displaying low annual fecundity (obligate clutch of one) and low adult
98 mortality. Its long lifespan may in part be attributed to superior cellular mechanisms for
99 resistance to oxidative damage (Ogburn et al., 2001) and maintenance of telomere length
100 (Haussmann et al., 2007). The oldest individuals in a population of a related species, the
101 Leach's storm petrel *Oceanodroma leucorhoa*, show little or no accumulation of short
102 telomeres over time (Haussmann and Mauck, 2008). We compared cohorts from two
103 consecutive breeding seasons characterised by different environmental conditions, as

104 indicated by overall reproductive performance of the colony. Productivity was relatively poor
105 in 2011 and significantly lower than 2010 (Watson et al., 2014). This provided the
106 opportunity to examine the effects of the natal environment on telomere length and dynamics
107 during early life and relate these to nestling survival. The study site and design also enabled
108 us to investigate whether exposure to an additional source of developmental stress, arising
109 from human recreational disturbance, has consequences for early-life telomere dynamics.
110 Parent-offspring relationships in TL were examined within a single cohort.

111 **Results**

112 The within-individual change in nestling TL was significantly different between the two
113 cohorts raised under different environmental conditions: in 2010, there was little change in
114 TL during the nestling period, whereas TL declined significantly with age in nestlings reared
115 in 2011 (Table 1, model 1; Fig. 1A; $F_{1,55.2} = 13.91$, $P < 0.001$). However, TL in early
116 postnatal development was significantly longer in individuals from the 2011 cohort,
117 compared with the 2010 cohort (Table 1, model 1; $F_{1,94.3} = 6.25$, $P = 0.014$). Removal of the
118 main effect of cohort, while retaining it in the interaction with Δ age, showed that, even after
119 adjusting for early differences in TL, the rate of change in TL was still significantly different
120 between the two cohorts ($\beta = -0.005 \pm 0.002$, $F_{1,95.5} = 5.82$, $P = 0.018$) and therefore the
121 cohort effect was above and beyond any effect due to regression to the mean. Furthermore,
122 there was considerable variation between individuals in the degree of change in TL, with
123 some individuals even exhibiting an apparent increase in TL during postnatal development
124 (Fig. 1B). The between-individual differences in TL were not quite significant (age at first
125 measurement: $F_{1,72.6} = 1.95$, $P = 0.055$). Neither nestling TL nor the within-individual change
126 in TL (tested by the respective interactions with Δ age) were affected by any of the other
127 variables considered, including sex, hatching date or visitor disturbance (all $P > 0.2$).
128 Subsequent analysis of TL in late postnatal development demonstrated that, although there
129 was a tendency for late TL to be shorter in 2011, the estimated cohort effect was not quite
130 significant at this stage (Table 1, model 2; $t_{47} = -1.73$, $P = 0.089$).

131 Of the 61 nestlings sampled, 6 died prior to fledging (3 individuals from each cohort).
132 When examining whether early TL was a good predictor of survival of the nestling phase, we
133 found that the probability of surviving to fledging increased significantly with TL (Table 1;
134 model 3; Fig. 2A; $z_{39} = 2.12$, $P = 0.034$). The mean TL of chicks that did not survive to
135 fledging was 22% shorter than that of chicks that successfully fledged (Fig. 2B). The model
136 fit was significantly improved by the inclusion of the rate of mass gain, though this variable

137 did not explain a significant portion of the variation in probability of fledging (Table 1, model
138 3; $z_{39} = 1.76$, $P = 0.078$). One of the chicks that did not fledge had very short telomeres (T/S
139 ratio = 0.68) and so the analysis was re-run excluding this nestling. The outlier had no
140 influence on the relationship, its removal having no effect on the magnitude or significance of
141 the effect of TL ($\beta = 0.99$, 95% CI: 0.64-1.0, $z_{38} = 2.08$, $P = 0.038$). The probability of
142 fledging was not affected by sex, hatching date, cohort, or visitor disturbance (all $P > 0.1$).
143 The relationship between nestling TL and probability of fledging was not affected by rate of
144 mass growth, as demonstrated by the respective interaction with TL (all $P > 0.1$). The AUC of
145 the minimum adequate model was 0.90, indicating the model performed very well in terms of
146 accuracy.

147 Analysis of parent-offspring relationships in the 2011 cohort revealed that nestling TL
148 was not significantly correlated with either maternal TL (Table 1, model 4; $t_{19} = 0.63$, $P =$
149 0.538) or paternal TL (Table 1, model 4; $t_{19} = 1.0$, $P = 0.330$). Parent-offspring TL
150 relationships were unaffected by nestling sex, as shown by testing the respective two-way
151 interactions (all $P > 0.5$).

152 Discussion

153 Although it has been widely suggested that the most rapid telomere loss occurs early in life in
154 a range of vertebrates (Zeichner et al., 1999; Baerlocher et al., 2007; Salomons et al., 2009),
155 only a few studies have investigated telomere dynamics within the timeframe of postnatal
156 development itself (see Foote et al., 2011; Geiger et al., 2012; Boonekamp et al., 2014;
157 Herborn et al., 2014). We found that TL and dynamics in growing nestlings of a long-lived
158 species were strongly influenced by the quality of the natal environment. Favourable
159 environmental conditions during development, as reflected by moderate colony productivity
160 in 2010, were associated with a negligible change in TL during early life, whereas conditions
161 resulting in significantly lower chick survival among the 2011 cohort were associated with
162 significant telomere attrition. While human recreational disturbance was previously shown to
163 reduce survival probabilities of nestlings (Watson et al., 2014), there was no effect of visitor
164 activity on telomere length or dynamics.

165 While some of the cohort effect might be explained by regression to the mean, the
166 difference in rate of change in TL remained even after adjusting for the differences in early
167 TL between the two cohorts. Accelerated attrition of longer telomeres, having controlled for
168 regression to the mean, has previously been reported (Salomons et al. 2009, Aviv et al. 2009)
169 and could be explained if longer telomeres offer a larger area for attack by free radicals (Aviv

170 et al. 2009). The high early TL in the 2011 cohort could potentially be explained if the high
171 early mortality observed within the 2011 cohort removed individuals with short telomeres
172 from the population or if poor-quality and/or inexperienced birds did not breed. Early
173 selective mortality of individuals with short telomeres was previously predicted to occur in
174 storm petrels (Hausmann and Mauck, 2008). Environmental conditions could be driving the
175 accelerated telomere attrition observed in the 2011 cohort either directly, for example via
176 elevated oxidative stress, and/or indirectly, if selective mortality or breeding by only high-
177 quality individuals were to result in a higher mean early TL, which is then subject to
178 increased attrition. The functional links between the environment and telomere dynamics
179 could be mediated by changes in parental care. Using a larger dataset from the same
180 population, we found that, in 2011, parents spent less time brooding their young and maximal
181 growth rate (in respect of mass) occurred later (Watson et al, unpubl.). An increased
182 frequency of extreme weather events in 2011 (Watson, 2014) might have been a key factor
183 influencing breeding success and parental care. Irrespective of the mechanism(s), this study
184 presents good evidence for environmental conditions influencing the rate of telomere loss and
185 shaping variation in early-life telomere dynamics between cohorts.

186 It is well known that cohort effects can arise in response to natal conditions. Survival
187 and recruitment among cohorts have previously been shown to be positively correlated with
188 the natal environment in mammals (Albon et al., 1987; Rose et al., 1998) and birds (Sedinger
189 et al., 1995; Reid et al., 2003). An increasing body of evidence links short telomeres and
190 accelerated rate of telomere loss to a shorter lifespan (Cawthon et al., 2003; Bize et al., 2009;
191 Olsson et al., 2011; Heidinger et al., 2012) and lower reproductive success (Pauliny et al.,
192 2006). The results of this study suggest that early-life telomere dynamics may contribute to
193 the marked differences in life-history traits that can arise among cohorts reared under
194 different environmental conditions. Indeed, TL in early life has been shown to be a strong
195 predictor of lifespan and a better predictor than TL in adulthood (Heidinger et al., 2012). In
196 addition to the increase in chick mortality observed in the cohort reared under unfavourable
197 natal conditions, the population-level effects might be exacerbated by a reduction in the
198 fitness of entire cohorts compared with those fledging under more favourable conditions.
199 Such cohort-wide effects can subsequently destabilise population dynamics (Lindström,
200 1999). Although our understanding will benefit from examination of longer-term data of
201 multiple cohorts, this study supports the idea that telomere dynamics may be a mechanism
202 linking early-life conditions with later-life performance.

203 The maintenance of TL during postnatal development under favourable natal
204 conditions in storm petrels contrasts with studies in other birds suggesting that rapid telomere
205 loss occurs early in life (Hall et al., 2004; Baerlocher et al., 2007; Salomons et al., 2009;
206 Heidinger et al., 2012). A negligible change in early-life TL was also demonstrated in chicks
207 of the long-lived king penguin experiencing favourable growth conditions (Geiger et al.,
208 2012). A slow rate of telomere shortening could be causally linked to a higher resistance to
209 oxidative stress (Ogburn et al., 2001) and/or elevated telomerase activity (Verdun and
210 Karlseder, 2007; Haussmann et al., 2007). Stressful conditions, however, have been shown to
211 lead to accelerated telomere loss via downregulation of antioxidants in birds (Stier et al.,
212 2009) and telomerase in humans (Epel et al., 2004). When faced with a costly reproductive
213 event, adult Adélie penguins *Pygoscelis adeliae* are able to increase antioxidant defences and
214 avoid accelerated telomere attrition (Beaulieu et al., 2011). The results of this study suggest
215 that storm petrel nestlings, when faced with unfavourable natal conditions, are unable to
216 modulate or activate such regulatory mechanisms. If poor early conditions were to lead to
217 irreversible downregulation of antioxidants or telomerase, accelerated telomere loss may
218 even persist beyond the nestling period, exacerbating the effects of early conditions on later-
219 life senescence. An area for future research is to understand the links between telomere loss,
220 oxidative stress and telomerase activity during early life.

221 While recent studies have investigated the relationship between TL in early life and
222 survival in captive (Heidinger et al., 2012) and wild (Caprioli et al., 2013; Boonekamp et al.,
223 2014) birds, we are aware of only one study that has directly examined the association
224 between early-life TL and mortality within the development phase itself. Just as king penguin
225 chicks that died prior to fledging displayed shorter telomeres (Geiger et al., 2012), we also
226 found that post-hatching TL of storm petrel nestlings is directly related to imminent
227 mortality. Despite a small sample size, the model performed well (as measured by AUC) and
228 the results were not influenced by an outlying observation. Although confidence intervals are
229 wide at low TLs, they do not overlap with confidence intervals when the probability of
230 fledging is equal to one. TL was a much better predictor of fledging success than the rate of
231 mass gain. While food load and rate of mass gain are likely to be of importance in
232 determining survival within the first few days of hatching, storm petrel nestlings are well-
233 buffered against periodic food shortages and mortality beyond the brooding stage is unlikely
234 to be strongly influenced by environmental factors affecting rate of provisioning and
235 subsequent mass gain. Additionally, since they nest in cavities, the risk of nest predation is
236 very low. Consequently, it seems most likely that mortality was linked to intrinsic factors.

237 Accelerated telomere shortening at this stage is likely to be a biomarker rather than a cause of
238 poor survival prospects and may be indicative of exposure to high levels of glucocorticoids
239 and/or oxidative stress. Early TL is partly determined by genetic factors (Njajou et al., 2007),
240 but the rate of attrition is also known to be affected by environmental influences (Epel et al.,
241 2004; Tarry-Adkins et al., 2009; Geiger et al., 2012). The results demonstrate that early-life
242 TL is a good predictor of imminent mortality.

243 Recent evidence suggests that TL in birds is maternally inherited (an example of
244 heterogametic inheritance; Horn et al., 2011; Reichert et al., 2015; Asghar et al., 2015).
245 However, Reichert et al. (2014) also showed that environmental factors exerted a strong
246 influence on nestling TL and the maternal link disappeared with chick age. The absence of
247 any observable heritable component to early TL in this study may therefore be a consequence
248 of environmental effects obscuring any initial genetic effect on nestling TL. Since we only
249 have data on parent-offspring relationships for the cohort reared under unfavourable
250 conditions, Furthermore, parent-offspring relationships in this study could have been
251 obscured by parental age, which was not known; although telomere length was found to be
252 unrelated to age in two other long-lived avian species (Hall et al., 2004). Future research
253 should seek to disentangle the relative contributions from genetic, parental and environmental
254 effects on TL and dynamics. This will help to develop our understanding of the role of
255 telomere dynamics in driving the evolution of life histories.

256 **Materials and methods**

257 The study was conducted at the island of Mousa, located in the Shetland archipelago, UK
258 (60°0'N, 1°10'W). Storm petrel nestlings are brooded for c. 7 d and do not leave the
259 underground nest cavity until fledging at c. 65-70 d (Davis, 1957). Fledging success (of eggs
260 hatched) was significantly lower in 2011 (0.60), compared with 2010 (0.78; see Watson et al.,
261 2014). This was probably in part linked to an increased frequency of extreme weather events
262 in 2011 (Watson, 2014). It is not always clear what environmental correlates are relevant,
263 however, and overall breeding performance of a population is widely used as a measure of
264 the natal environment (e.g. Reid et al. 2003). Blood was collected from 32 nestlings in 2010
265 and 29 nestlings in 2011 and from both parents at 21 nests in 2011 only. Whole blood was
266 obtained by venepuncture of the brachial vein under licence from the UK Home Office.
267 Nestlings were sampled on two occasions (where possible) in the first (hereafter, 'early';
268 median age: 11 d, range: 4-33 d, n = 59) and second (hereafter, 'late'; median: 45 d, range:
269 34-63 d, n = 52) half of postnatal development. Fifty nestlings were sampled twice and the

270 median number of days elapsed between repeated measurements was 31 d (range: 22-47 d).
271 Only one sample was available for some nestlings either because nestlings died ($n = 6$), were
272 not accessible in the nest or samples were accidentally destroyed. Of the 50 nestlings sampled
273 twice, 49 survived to fledging. The overall proportion of chicks that died was greatly
274 underrepresented in the data and the difference in mortality between the two cohorts was not
275 reflected within the dataset; this is because the majority of chick mortality occurred within a
276 few days of hatching and before initial blood sampling was carried out. Tarsus length
277 (nestlings only), mass and wing length (maximum flattened chord; adults only) were
278 recorded. Nestling mass was corrected to a standardised time (18:00 h) to account for the age-
279 related rate of proportional weight loss that chicks undergo during diurnal fasting (Bolton
280 1995).

281 **Determination of TL**

282 Blood samples were stored at $<5^{\circ}\text{C}$ for up to 8 hr prior to being separated by centrifugation.
283 Plasma and red blood cells were stored at $<5^{\circ}\text{C}$ in the field for a maximum of 3 d before
284 being transferred to -20°C for up to 3 months; following this, samples were stored at -80°C
285 until laboratory analyses were performed. DNA was extracted from red blood cells using
286 Machery-Nagel NucleoSpin Blood kits following the manufacturer's protocol. The quantity
287 and purity of DNA was assessed using a NanoDrop 8000 spectrophotometer (Thermo
288 Scientific); there were no differences in yield or purity between cohorts. Sex was determined
289 following the molecular method described by Griffiths et al. (1998). TL was measured by
290 quantitative PCR (qPCR) using an Mx3005P (Stratagene) as described by Cawthon (2002)
291 and adapted by Criscuolo et al. (2009) for birds. The method gives a relative value for TL by
292 determining the ratio of number of telomere repeats (T) to that of a single-copy control gene
293 (S; or more precisely, is non-variable in copy number) relative to a reference sample.
294 Consequently, the measure will include any interstitial repeats of the telomeric sequence
295 (Foote et al., 2013). However, good correlations have been found between TL measured
296 including and excluding the interstitial repeats using telomere restriction fragment (TRF)
297 analysis (Foote et al., 2013) and between TRF and qPCR analyses (Criscuolo et al., 2009;
298 Aviv et al., 2011). Since the extent of interstitial repeats is not expected to change with age,
299 qPCR is well suited to detecting within-individual changes. Furthermore, measurement of
300 telomeres from Leach's storm petrels, using TRF methods, suggests that interstitial repeats do
301 not occur to the extent that they would influence telomere measurements (Mark Haussmann,
302 pers. comm).

303 Amplification of telomere sequences was achieved using the forward and reverse
304 primers: Tel1b (5'-CGGTTTGGTTTGGGTTTGGGTTTGGGTTTGGGTTTGGG TT-3') and
305 Tel2b (5'-GGCTTGCCTTACCCTTACCCTTACCCTTACCCTTACC CT-3'). The non-
306 variable copy control gene used was ornithine decarboxylase (OCD), isolated from the
307 European storm petrel (GenBank: DQ881744.1), which was amplified using the primers:
308 OCD Fwd1 (5'- GACCTTGCCATCATTGGAGTTAG-3') and OCD Rev1 (5'-
309 AAGGCATCCCTATTGTTAGGTAGA -3') sourced from Integrated DNA Technologies
310 (Leuven, Belgium). qPCR was performed using 10 ng of DNA per reaction. The
311 concentrations of primers used were 500 nM for telomere and 70 nM for OCD reactions.
312 Telomere qPCR reaction conditions started with 15 min at 95°C, followed by 27 cycles of 15
313 s at 95°C, 30 s annealing at 58°C and 30 s extending at 72°C. OCD reaction conditions started
314 with 15 min at 95°C, followed by 40 cycles of 30 s at 95°C and 30 s at 60°C. Melting curves
315 were assessed to confirm there was no non-specific amplification or primer-dimer formation.
316 For both telomere and OCD reactions, the number of PCR cycles required for accumulation
317 of sufficient products to exceed a threshold of fluorescent signal (C_t) was determined. A
318 standard curve, run on each plate, consisted of a serial dilution of a reference sample ranging
319 from 40 ng to 2.5 ng. The C_t threshold for each reaction was determined from the reference
320 sample. A negative control was also included on each plate. All samples, including the
321 standard curve, were run in triplicate. Amplification efficiencies were within an acceptable
322 range (Mean \pm SD: Telomere: $106.8 \pm 3.7\%$; OCD: $114.3 \pm 3.9\%$) and all samples fell within
323 the bounds of the standard curve. Inter- and intra-assay variations were 3.6% and 1.4%
324 respectively for telomere reactions and 0.82% and 0.33% respectively for OCD reactions.
325 Inter-assay variation in T/S ratio was 10.4%. The T/S ratio for each sample was calculated
326 from mean C_t values according to Pfaffl (2001). This method accounts for the variation in
327 amplification efficiencies. All repeated samples from nestlings were run on the same plate
328 along with parental samples.

329 **Statistical analysis**

330 All statistical analyses were performed in R 3.0.0 (R Core Team, 2013). Linear mixed models
331 (LMMs) with a normal error structure were fitted to data on nestling telomere length to
332 examine change in TL in surviving nestlings ($n = 100$; number of individuals = 55). Outlying
333 observations were removed to normalise the data and improve model fits. Since we were
334 primarily interested in the within-individual change in nestling TL, we employed within-
335 subject centring (van de Pol and Verhulst, 2006) to separate between-individual (cross-

336 sectional) and within-individual (longitudinal) effects. The age variable was split into two
337 covariates: age at first measurement (between-individual effects) and Δ age (the change in
338 age between measurements; within-individual effects). LMMs were fitted in the lme4
339 package (Bates et al., 2013). First, the optimal random effects structure was found by
340 comparing nested models, fitted by restricted maximum likelihood (REML), using likelihood
341 ratio tests (LRTs). A random intercept for nestling identity was included. We also considered
342 the inclusion of a random intercept for nest identity, to account for potential non-
343 independence of nestlings from different cohorts reared at the same nest-site. 30% of nests
344 were sampled in both years, though we do not know whether nests were occupied by the
345 same breeding pair in both years. However, the variances associated with nest identity were
346 estimated to be zero and inclusion of the random effect did not significantly improve model
347 fits. With nestling TL as the dependent variable, candidate explanatory variables included
348 cohort (year: 2010 or 2011), nestling sex and human disturbance (high or low, defined as ≤ 10
349 m or >150 m from the visitor path, respectively) as two-level fixed factors, and age at first
350 measurement, Δ age and hatching date as covariates. We tested for effects on within-
351 individual change in TL by considering all respective interactions with Δ age. To examine
352 variation in nestling TL in late development, linear models (LMs) with a normal error
353 structure were fitted to log-transformed late nestling TL. Candidate explanatory variables
354 included cohort, nestling sex and human disturbance as fixed factors and age and hatching
355 date as covariates.

356 Generalised linear models (GLMs) with a binomial error structure and logit link
357 function were fitted to data on survival (0/1; $n = 42$) to examine whether early TL (age: ≤ 16
358 d, median = 11 d) was a good predictor of survival of the nestling phase. The threshold of 16
359 d was adopted since this was the upper bound of the 95% confidence interval of age at first
360 measurement. These data did not include any repeated measures at the nest-level. In addition
361 to the fixed effect of nestling TL at ≤ 16 d, candidate explanatory variables included cohort,
362 sex and visitor disturbance as fixed factors and body size, body mass, hatching date, growth
363 rate (average daily incremental growth in mass) as covariates. We also tested for two-way
364 interactions between TL and mass growth rate. The minimum adequate model was evaluated
365 using Receiver Operating Characteristic plots (Sing et al., 2005). The resulting area under the
366 curve (AUC) offers a measure of predictive performance for a binomial model; a value of 1.0
367 indicates a perfect model, while a value of 0.5 indicates that a model performs no better than
368 random.

369 Finally, parent-offspring relationships in the 2011 cohort were examined in a linear
 370 model (LM) analysis with early nestling TL as the dependent variable ($n = 21$). Candidate
 371 fixed effects included the covariates of maternal TL, paternal TL and nestling sex as a fixed
 372 factor. We also tested for potential effects of nestling sex on parent-offspring relationships
 373 with the respective two-way interactions between parental TL and nestling sex and age.
 374 Having shown, in previous analyses, that neither age, hatching date, nor visitor disturbance
 375 affected nestling TL, these variables were not included in the parent-offspring analysis.

376 Taking into account the sample sizes and number of variables to potentially control
 377 for, a conservative approach to model fitting was adopted; starting from a null model, fixed
 378 effects that significantly differed from zero ($P < 0.05$) were added sequentially in a forward
 379 stepwise regression. Each time a new variable was added to the model, the significance of
 380 existing variables was re-examined. Nested models were compared using LRTs and the
 381 criterion for entry of a variable was a log-likelihood ratio P -value of < 0.05 . For LMMs,
 382 model selection was performed using models fitted by maximum likelihood; parameter
 383 estimates with standard errors are quoted from the minimum adequate LMM fitted by REML.
 384 The significance of parameter estimates was estimated using conditional F -tests based on
 385 Satterthwaite approximation for the denominator degrees of freedom. For LMs and GLMs,
 386 parameter estimates were estimated based on the t - and z -distributions, respectively; again
 387 estimates with standard errors are quoted from minimum adequate models.

388 **List of abbreviations**

| | | |
|-----|--------------|--|
| 389 | TL | telomere length |
| 390 | qPCR | quantitative PCR |
| 391 | C_t | the number of PCR cycles required for accumulation of sufficient products to |
| 392 | | exceed a set threshold of fluorescent signal |
| 393 | T/S ratio | ratio of telomeric sequence to that of a single copy gene, relative to the |
| 394 | | reference sample |
| 395 | OCD | a single copy gene isolated from the European storm petrel encoding for the |
| 396 | | protein, ornithine decarboxylase |
| 397 | Δ age | change in age (d) between first and second telomere length measurements |
| 398 | LM | linear model |
| 399 | GLM | generalised linear model |
| 400 | LMM | linear mixed-effects model |
| 401 | REML | restricted maximum likelihood estimation |

402 LRT likelihood ratio test
403 AUC area under the curve

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411 **Competing interests statement**

412 No competing interests declared.

413 **Author contributions**

414 Conception and study design, and interpretation of the findings: H.W., M.B., P.M. Data
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Table 1. Effects of variation in natal conditions on telomere length (TL) and rate of telomere attrition during postnatal development and links to survival and parental TL. TL of nestlings was measured in early and late development in two successive cohorts reared under favourable (2010) and unfavourable (2011) conditions. Parental TL was only measured in the 2011 cohort. Parameter estimates and *P*-values are shown for fixed effects retained in minimum adequate models and for key terms that were rejected during forward stepwise regression. Models 1 and 2 also tested for the fixed effects of sex, hatching date and visitor disturbance; the effects were not significant in any model and, for simplicity, are not shown in the table. Model type is specified as either linear mixed model (LMM), linear model (LM) or generalised linear model (GLM).

| Model and dependent variable | Fixed effect(s) | Rejected term(s) | Parameter estimate (SE or 95% CI) | <i>P</i> -value |
|---|-----------------------------|--|---|-----------------|
| Within-individual change in telomere length during postnatal development (LMM^a) | | | | |
| (1) All TL (n = 100) | Δ age ^b | | 3.11e ⁻⁴ (1.90e ⁻³) | <0.001*** |
| | cohort(2011) | | 0.147 (0.059) | 0.014* |
| | Δ age x cohort(2011) | | -9.05e ⁻³ (2.59e ⁻³) | <0.001*** |
| | | age at first measurement | 6.09e ⁻³ (3.12e ⁻³) | 0.055 |
| | | all 2-way interactions with Δ age | | >0.2 |
| Telomere length in late postnatal development (LM) | | | | |
| (2) Late TL (n = 49) | | cohort(2011) | -0.092 (0.053) | 0.089 |
| | | age | 0.002 (0.004) | 0.640 |
| Nestling survival and telomere length (binomial GLM) | | | | |
| (3) Survival (0/1) (n = 42) | early TL | | 0.99 (0.68-1.0) | 0.034* |
| | growth rate | | 0.94 (0.42-1.0) | 0.078 |
| | | cohort(2011) | 0.648 (0.15-0.95) | 0.614 |
| | | TL x growth rate | 5.2e ⁻⁷ (1.5e ⁻¹⁸ -1.0) | 0.286 |
| Parent-offspring relationship in telomere length in 2011 cohort (LM) | | | | |
| (4) Early TL (n = 21) | | maternal TL | 0.148 (0.237) | 0.538 |
| | | paternal TL | 0.194 (0.194) | 0.330 |
| | | maternal TL x nestling sex(male) | -0.301 (0.506) | 0.560 |
| | | paternal TL x nestling sex(male) | -0.139 (0.593) | 0.818 |
| | | | | |

^aIncluded random intercept for nestling identity; mean variance \pm SD: 0.005 \pm 0.07.

^b Δ age is the change in age (d) between TL measurements.

Significance levels denoted as * *P* < 0.05, ** *P* < 0.01, *** *P* < 0.001.

Figure 1. Change in telomere length (TL) during growth in storm petrel nestlings from two consecutive cohorts reared under different natal conditions: (A) Mean within-individual change in TL in 2010 (closed circles; solid line) and 2011 (open circles; dashed line). Lines represent model predictions from the minimum adequate LMM (Δ age \times cohort(2011): $F_{1,55,2} = 12.17$, $P = <0.001$, $n = 100$) fitted within the range of observed values. Δ age is the change in age (d) between the first and second measurement. (B) Individual change in telomere length during postnatal development ($n = 98$) in 2010 (closed circles; solid lines) and 2011 (open circles; dashed lines). Lines link TL measurements in early and late postnatal development for each individual.

Figure 2. Relationship between early nestling telomere length (TL) and survival during postnatal development: (A) Predicted probability of fledging (solid line) in relation to early TL (≤ 16 d; GLM: $z_{39} = 2.12$, $P = 0.034$) with 95% confidence intervals (dashed lines). Line is fitted within the range of observed values (open circles; $n = 42$) and based on a mean rate of mass gain of 1.03 g d^{-1} ($z_{39} = 1.76$, $P = 0.078$). Removal of the outlier at the lower end of the TL scale did not affect model predictions. (B) Mean \pm SE early TL of nestlings that died ($n = 6$) during the nestling phase and those that survived to fledging ($n = 36$).



