

Final

14 November 2014

Title

Emerging concepts in the pathogenesis of ANCA-associated vasculitis.

Authors

Shaun M Flint^{1,2}, Eoin F McKinney^{1,2}, Kenneth GC Smith^{1,2}

Affiliations

¹ Department of Medicine, University of Cambridge

² Cambridge Institute of Medical Research.

Corresponding Author details.

Professor Kenneth GC Smith

Department of Medicine, The University of Cambridge,

Box 139, Addenbrooke's Hospital, Hills Road, Cambridge CB2 0XY, United Kingdom

e-mail: kgcs2@cam.ac.uk phone: +44 1223 762 639

Disclosure of specific funding.

This work was supported by the Wellcome Trust through a Translational Medicine and Therapeutics PhD Studentship, the National Institute of Health Research Cambridge Biomedical Research Centre and an MRC Programme grant. The Cambridge Institute for Medical Research is in receipt of Wellcome Trust Strategic Award 079895.

Abstract***Purpose of review:***

Anti-neutrophil cytoplasmic antibodies (ANCA) remain central to our current understanding of the pathogenesis of ANCA-associated vasculitis (AAV), and this review considers recent developments in the context of four key questions: (1) are there targets for ANCA beyond myeloperoxidase (MPO) and proteinase 3 (PR3); (2) are all ANCA pathogenic; (3) how are ANCA generated; and (4) how do ANCA cause disease?

Recent findings:

B-cell epitope mapping raises the possibility that only a subset of ANCA may be pathogenic. Anti-LAMP-2 autoantibodies have recently emerged as novel form of ANCA, and can be found in anti-MPO and anti-PR3-negative disease. These also provide recent evidence for molecular mimicry in the pathogenesis of AAV, but definitive proof in human AAV remains elusive. Neutrophil extracellular traps may represent an important mechanism by which MPO and PR3 are taken up by dendritic cells for presentation to the adaptive immune system, and the role of the alternative pathway of complement in AAV has recently been emphasized, with therapeutic implications.

Summary:

Our current understanding of the pathogenesis of AAV reinforces the central role of neutrophils, but also provides a sound rationale for B-cell and complement-directed therapies.

Keywords: (3-5)

Anti-Neutrophil Cytoplasmic Antibody-Associated Vasculitis;

Antibodies, Antineutrophil Cytoplasmic;

Neutrophils;

Pathogenesis

Introduction

Anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis (AAV) is a life-threatening, small vessel vasculitis characterized by the presence of pathogenic anti-proteinase 3 (PR3) and anti-myeloperoxidase (MPO) autoantibodies and renal, lung and upper respiratory tract involvement. Clinically and pathologically, three forms are traditionally recognized: granulomatosis with polyangiitis (GPA, formerly Wegener's granulomatosis), which is distinguished by necrotizing granulomas on biopsy and frequent upper respiratory tract involvement; microscopic polyangiitis (MPA); and eosinophilic granulomatosis with polyangiitis (EGPA, formerly Churg-Strauss Syndrome), which is much less common and distinguished by a prominent eosinophilia and allergic features.[1] There is significant clinical and pathological overlap between these diseases, and evidence is accumulating that sub-classification of AAV using ANCA antigen specificity may better reflect both prognosis and disease pathogenesis than traditional clinical definitions, reinforcing that fundamentally AAV remains an ANCA-centric disease.[2-4] This review, therefore, considers current research in the context of four key questions: (1) are there targets for ANCA beyond MPO and PR3; (2) are all ANCA pathogenic; (3) how are ANCA generated; and (4) how do ANCA cause disease?

Are there targets for ANCA beyond myeloperoxidase and proteinase-3, and are all ANCA pathogenic?

ANCA were first recognized for their ability to label neutrophils by indirect immunofluorescence.[5] The staining conformed to two patterns: perinuclear, and cytoplasmic, and MPO and PR3 were soon identified as their predominant target autoantigens, respectively. Multiple additional targets of ANCA were identified during the 1990s, including cathepsin-G and neutrophil elastase,[6] but these lack the specificity and the associated evidence of pathogenicity that has since accumulated for anti-PR3 and anti-MPO ANCA. The latter ranges from a case report of neonatal vasculitis in the context of exposure to maternal anti-MPO ANCA to evidence of glomerulonephritis after passive transfer of anti-MPO and anti-PR3 ANCA in murine models of disease.[7–9] On the other hand, there are also numerous examples of detectable anti-MPO and anti-PR3 ANCA in otherwise healthy individuals or individuals with a range of non-vasculitic diseases (e.g. systemic lupus erythematosus (SLE), ulcerative colitis), which demonstrates that ANCA are not always pathogenic.[10,11] In addition, up to a third of patients with a clinical diagnosis of AAV are ANCA-negative by conventional assays,[6] suggesting there may be additional targets for autoantibodies.

The question of ANCA specificities and pathogenicity has been addressed by recent studies. Using a mass spectrometry-based approach, Roth and colleagues identified 25 epitopes bound by anti-MPO autoantibodies.[12] Autoantibodies binding one of the linear epitopes, MPO⁴⁴⁷⁻⁴⁵⁹, were exclusively found in patients with active

disease, suggesting that these autoantibodies may be pathogenic. This was supported by evidence that ANCA binding this epitope activated neutrophils *in vitro* and by the observation that DR2 transgenic mice immunized with the corresponding murine epitope developed a pauci-immune glomerulonephritis. However, complicating this, these immunized mice developed a polyclonal anti-MPO ANCA that bound a range of epitopes including but not limited to the MPO⁴⁴⁷⁻⁴⁵⁹. The remaining MPO epitopes were classified according to whether they bound ANCA from patients with active disease, remission or healthy volunteers, and this was used as a basis to infer potential pathogenicity. However, where ANCA were detectable in healthy volunteers, it was at extremely low levels. It remains unclear, therefore, whether ANCA targeting these epitopes are truly non-pathogenic at the higher concentrations observed in disease.

The same study also found that purified immunoglobulin from patients with ANCA-negative disease was able to bind the pathogenic MPO⁴⁴⁷⁻⁴⁵⁹ epitope, implying that detection of ANCA in a proportion of ANCA-negative patients may be masked by a serum factor (perhaps a fragment of ceruloplasmin) interfering with standard assays. Larger studies are needed to validate these findings and determine the proportion of ANCA negative patients accounted for by this proposed mechanism.

There are other potential mechanisms for ANCA-negative vasculitis. Espéli and colleagues have demonstrated that most auto-antibody producing plasma cells in mice with active SLE are found in inflammatory lesions, suggesting local production

of autoantibodies might help drive disease[13]; this potential mechanism has not been investigated in ANCA-negative AAV. Studies of anti-LAMP-2 autoantibodies performed by Kain and colleagues also suggest that these may represent a novel form of ANCA present in anti-PR3 and anti-MPO ANCA-negative disease. LAMP-2, a lysosomal membrane protein, was first identified as an antigenic target of ANCA in 1995 as part of a broader screen.[14] Follow-up publications in 2008 and 2012 reported that anti-LAMP-2 autoantibodies were widely present in patients with ANCA-positive vasculitis (14/16 patients in the 2008 study and 78/84 patients in 2012), coexisting with anti-MPO or anti-PR3 in most cases. These studies were not without controversy: the complex, highly glycosylated nature of native LAMP-2 meant that assays for anti-LAMP-2 antibodies were difficult to replicate,[15,16] however Kain argues plausibly that this is due to differences in patient selection (anti-LAMP-2 antibodies are sensitive to immunosuppression and disease activity) and the source of LAMP-2 antigen used.[17] More recently, anti-LAMP2 antibodies were found in eight of eleven ANCA-negative patients presenting with a pauci-immune glomerulonephritis.[18*] These autoantibodies were notable for binding the less-glycosylated glomerular form of LAMP-2, not the highly-glycosylated neutrophil form. Evidence for the pathogenicity of anti-LAMP-2 autoantibodies comes from their association with disease in animal models and their correlation with active disease in the human studies discussed above. Animal models include a 2013 Japanese study in which transgenic rats overexpressing the human T-cell lymphocyte virus type 1 gene env-pX develop a cutaneous vasculitis and spontaneous anti-LAMP-2 autoantibodies (without anti-PR3 or anti-MPO

ANCA).[19] When these antibodies were transferred into disease-free env-pX rats they progressed to develop an accelerated neutrophil-rich cutaneous vasculitis in comparison to non-treated control rats. The same study also identified anti-LAMP-2 autoantibodies in a subset of patients with cutaneous polyarteritis nodosa, and with Henoch-Schonlein purpura, diseases not classically associated with ANCA. This implies that, as with anti-MPO and anti-PR3 ANCA, anti-LAMP-2 autoantibodies may not wholly specific for AAV.

Antibodies targeting moesin have also recently been described in a Japanese cohort of patients with anti-MPO AAV with some capacity to stimulate monocytes and neutrophils *in vitro*. [20]

How are ANCA generated?

Multiple hypotheses have been presented for the formation of ANCA, with molecular mimicry one of the most discussed. However, a pathogen may contribute to the risk of autoimmunity without necessarily being present at the onset of that disease (the so-called 'fertile field' hypothesis), [21] which makes establishing its role particularly challenging. Yet there is increasing evidence of heterologous immune responses to multiple pathogens making molecular mimicry a plausible hypothesis. [22] The most convincing evidence for its role in AAV comes from homology between FimH, a component of type 1 fimbriae found on a range of gram-negative pathogens, and LAMP-2. [23] Here, rats immunized with FimH progressed

to develop cross-reactive anti-LAMP-2 antibodies and a pauci-immune glomerulonephritis.

Another hypothesis is that homology between certain *Staphylococcus aureus* proteins and a peptide encoded by the reverse DNA strand to PR3 ('complementary' PR3, cPR3) results in an immune response to *S. aureus* generating antibodies that are also cross-reactive with cPR3.[24] These anti-cPR3 autoantibodies are then said to seed the development of idiotypic, PR3-reactive antibodies, ultimately leading to the development of clinical GPA. However, there is no direct evidence that anti-cPR3 antibodies cross-react with bacterial proteins, and others have not been able to confirm the presence of elevated anti-cPR3 antibody titres in GPA, nor establish a relationship between the presence of anti-cPR3 antibodies and *S. aureus* carriage.[25]

Nonetheless, an association between *S. aureus* carriage and AAV is well described,[26] and a role for *S. aureus* in the pathogenesis of AAV continues to be sought. Potential mechanisms include the ability of *S. aureus* to potently induce the formation of neutrophil extracellular traps (NETs),[27] discussed below or, more generally, bacterial activation of neutrophils through the activation of toll-like receptors. TLR4 and TLR9, whose ligands are gram-positive bacterial components and bacterial DNA respectively, have been shown to trigger MPO release and increased surface expression of PR3.[28] A recent candidate gene study has also identified polymorphisms at the *TLR9* locus as modulating susceptibility to GPA.[29]

This circumstantial evidence suggests a plausible mechanism by which *S. aureus* could drive GPA, but it remains far from conclusive.

Non-infectious triggers for ANCA formation have been described. Medications are chief among these and include levamisole (used as a 'cutting' agent for cocaine), one of the few instances in which both anti-PR3 and anti-MPO autoantibodies coexist, and propylthiouracil as recently reviewed in [30].

A role for T-cells in the generation of ANCA is suggested by their high affinity, class-switched nature and underscored by long-reported associations between MHC class II variants and AAV.[31,32] Recent GWAS studies have begun to define the latter more closely. Anti-MPO AAV and anti-PR3 AAV were strongly associated with HLA-DQ and HLA-DP variants, respectively.[3] The HLA-DP association was replicated in a second AAV GWAS, likely driven by the high anti-PR3 prevalence in this predominantly GPA cohort.[32] Precisely how a specific HLA risk allele predisposes to autoimmunity in AAV has not been determined, but in other diseases with a Class II HLA association it has been possible to map the genetic association onto key amino acid residues in the peptide-binding groove.[33] The ability of these HLA risk alleles to specifically bind peptides that have undergone post-translational modification seems to be an important property that may predispose to autoimmunity.[34]

What has also become clear is that the generation of ANCA may be facilitated by positive feedback loops within the innate immune system whereby, for example, ANCA stimulates the formation of NETs and which then facilitate the further generation of ANCA. These are discussed further in the next section, which examines recent developments in some of the effector pathways downstream of ANCA.

How do ANCA cause disease?

Recent research into the role of NETs and complement in AAV has added significantly to our understanding of how ANCA cause disease.

Neutrophil extracellular traps, and their role in AAV

A unique feature of neutrophils, which are key players in AAV, is their ability to generate NETs, a 'sticky mesh' of extruded chromatin, nucleic acid and a range of cytoplasmic proteins. These serve antibacterial functions, but also represent a means by which cytoplasmic proteins may be exposed to the immune system.[35] In 2009, AAV was one of the first autoimmune diseases to be linked to excessive NET formation.[36] This initial study demonstrated that NETs contain MPO and PR3 and are induced by ANCA. Moreover, extracellular DNA, histones and neutrophil granule components including PR3 and MPO co-localised in the glomeruli of patients with an active ANCA-associated glomerulonephritis, placing NETs at the site of tissue damage *in vivo*. Since then, increasing circumstantial evidence suggests that NETs represent an important means for autoantigen uptake by antigen presenting cells (APCs) in AAV. First, Sangaletti and colleagues showed in an elegant study that

murine NETs specifically interact with myeloid dendritic cells (mDCs) to facilitate the uptake of MPO and PR3. Moreover, when these NET-cultured mDCs were then transferred into healthy mice both new ANCA formation and an active glomerulonephritis were observed,[37] although exactly how an interaction with NETs licenses APCs to provide enhanced costimulation to B- and T-cells in subsequent interactions is unclear. Recent human studies provide additional evidence for the importance of NETs in AAV, showing that that the serum of patients with MPO+ vasculitis potently induces NET formation and retards their degradation, the latter potentially through an inhibitory action of anti-NET antibodies.[38*] It is hypothesized, but remains to be proven, that excessive or inappropriate NET formation or delayed NET degradation represents an important mechanism by which the key AAV autoantigens MPO and PR3 are taken up by APCs in a pro-inflammatory context for presentation to B- and T-cells (figure 1).

Complement

Our understanding of the role of complement in tissue damage in AAV has also undergone a substantial shift. The pauci-immune nature of the characteristic lesions had given rise to an initial belief that complement did not play a significant role in AAV pathogenesis.[39] Since then, a number of studies have convincingly shown that activation of the alternative pathway of complement and an intact C5a receptor (C5aR) are necessary for the development of glomerulonephritis in murine models of vasculitis, prompting a re-assessment of the role of complement in human AAV.[40,41] Experiments *in vitro* showed that ANCA-stimulated healthy human neutrophils are able to activate complement and that complement acting through

the C5aR is able to prime neutrophils and upregulate PR3 surface expression.[42] Moreover, C3F, a common coding genetic variant of C3, was found to be overrepresented in patients with AAV in small candidate gene studies.[43–45] Despite the pauci-immune tag, a number of studies have also demonstrated low level glomerular immune-complex deposition, including complement components.[46,47]

A Chinese group reported that complement deposition in renal biopsies of AAV patients correlated with worse renal injury, and that glomerular deposition and urine levels of the alternative pathway complement fragment Bb, in particular, correlate well with AAV activity. [48*] Other complement components including C3a, C5a and soluble C5b-9 were also found to be elevated and, separately, they reported increased plasma levels of activated complement components C3a, C5a and Bb in patients with active AAV.[49] C5aR (CD88) expression in renal biopsy specimens was predominantly tubular, was reduced in active vasculitis and absent on infiltrating glomerular neutrophils, raising questions about the importance of this receptor in AAV.[50] However, given that C5aR on neutrophils is internalized upon activation, and cleaved by neutrophil serine proteases from the surface of other cell types, this is not necessarily inconsistent with a role for C5aR in AAV.[51] Taken together these observations support a role for the alternative pathway of complement in human AAV, consistent with the murine data. An inappropriate positive feedback loop in which ANCA-activated neutrophils themselves activate complement, further increasing the amount of C5a present, may be important in

sustaining the inflammatory process (figure 2). These observations, especially if replicated, provide a rationale for the use of eculizumab, a C5a inhibitor, in AAV, with a preliminary report providing an early suggestion of clinical efficacy.[52]

Other notable recent findings – in brief

Under certain conditions it is clear that B cells can have a regulatory function, driven at least in part by the production of IL-10. These “regulatory B-cells” can be hard to define phenotypically, and there is only limited evidence that they comprise a distinct “lineage”. Four small recent clinical studies suggest that in AAV, especially GPA, a sub-population of B cells retains an ability to produce IL-10 and suppress T-cell differentiation and proliferation, but is reduced in number.[53–56] Further work is needed to clarify the role of B-cell regulation of AAV, however, as these changes may be secondary to changes in effector populations, and none of the studies established a convincing dose relationship between AAV disease activity and these cells. Regulatory T-cells (Tregs) also have the capacity to suppress immune responses. Studies of Treg populations in AAV are conflicting, and there is no consensus regarding changes in their suppressive ability or the size of the T-reg population in active disease.[57–59]. The most recent study to examine their role in AAV enrolled 62 patients in a cross-sectional fashion, finding that during active disease this population was expanded relative to other CD4+ T-cell populations but, again, it is difficult to know whether this is secondary to changes in the effector compartment given that the controls were paired remission samples and healthy volunteers.[60]

Conclusions

In the last few years, some controversies in the pathogenesis of AAV have moved closer to resolution but others remain hotly debated. NETs have emerged as a likely mechanism by which MPO and PR3 are taken up by APCs for presentation to the adaptive immune system, but the precise mechanisms by which tolerance to these self-antigens is broken remains unclear. Molecular mimicry remains an attractive hypothesis, and the most convincing evidence for this comes from controversial studies into the development of anti-LAMP-2 autoantibodies. Recent publications suggest an increasing acceptance of their role in AAV, but further research is needed. Large-scale genetics studies have identified a number of loci, including MHC Class II, that confer genetic susceptibility to AAV, and an important topic for future research will be determining how polymorphisms at these loci contribute to disease pathogenesis. An increasing appreciation of the importance of complement in generating tissue inflammation in AAV underscores the exciting potential of the complement pathway inhibitor eculizumab as a novel therapeutic for active disease. This demonstrates that the difficult process of teasing out the key pathogenic features driving AAV can provide real patient benefit.

Key Points: (3-5 bullet points)

- Emerging evidence suggests that epitope specificity may contribute to the pathogenicity of anti-neutrophil cytoplasmic antibodies (ANCA), and that anti-LAMP-2 antibodies represent a novel form of ANCA.
- The alternative pathway of complement is increasingly recognized as being

important in mediating the pathogenicity of ANCA.

- Neutrophils remain central to the pathogenesis of ANCA-associated vasculitis, with neutrophil extracellular traps playing a potentially important role in initiating the immune response.

Acknowledgements:

The authors acknowledge support from the Cambridge Biomedical Research Centre.

Financial support and sponsorship:

This work was supported by the Wellcome Trust [083650/Z/07/Z, 094227/Z/10/Z], MRC Programme grant [MR/L019027/1]. S.M.F holds a Translational Medicine and Therapeutics PhD Studentship part-funded by the Wellcome Trust and GlaxoSmithKline. The Cambridge Institute for Medical Research is in receipt of Wellcome Trust Strategic Award 079895.

Conflicts of interest:

The authors declare that there are no conflicts of interest relevant to this publication.

References:

1. Jayne D. The diagnosis of vasculitis. *Best Pract. Res. Clin. Rheumatol.* 2009;23:445–53.
2. Lionaki S, Blyth ER, Hogan SL, et al. Classification of antineutrophil cytoplasmic autoantibody vasculitides: the role of antineutrophil cytoplasmic autoantibody specificity for myeloperoxidase or proteinase 3 in disease recognition and prognosis. *Arthritis Rheum.* 2012;64:3452–62.
3. Lyons PA, Rayner TF, Trivedi S, et al. Genetically distinct subsets within ANCA-associated vasculitis. *N. Engl. J. Med.* 2012;367:214–23.

4. Mahr A, Katsahian S, Varet H, et al. Revisiting the classification of clinical phenotypes of anti-neutrophil cytoplasmic antibody-associated vasculitis: a cluster analysis. *Ann. Rheum. Dis.* 2013;72:1003–10.
5. Davies DJ, Moran JE, Niall JF, Ryan GB. Segmental necrotising glomerulonephritis with antineutrophil antibody: possible arbovirus aetiology? *Br. Med. J. (Clin. Res. Ed).* 1982;285:606.
6. Hoffman GS, Specks U. Antineutrophil cytoplasmic antibodies. *Arthritis Rheum.* 1998;41:1521–37.
7. Schlieben DJ, Korbet SM, Kimura RE, et al. Pulmonary-renal syndrome in a newborn with placental transmission of ANCA. *Am. J. Kidney Dis.* 2005;45:758–61.
8. Xiao H, Heeringa P, Hu P, et al. Antineutrophil cytoplasmic autoantibodies specific for myeloperoxidase cause glomerulonephritis and vasculitis in mice. *J. Clin. Invest.* 2002;110:955–63.
9. Little M a, Al-Ani B, Ren S, et al. Anti-proteinase 3 anti-neutrophil cytoplasm autoantibodies recapitulate systemic vasculitis in mice with a humanized immune system. *PLoS One* 2012;7:e28626.
10. Hermann J, Demel U, Stünzner D, et al. Clinical interpretation of antineutrophil cytoplasmic antibodies: parvovirus B19 infection as a pitfall. *Ann. Rheum. Dis.* 2005;64:641–3.
11. Noh JY, Asari T, Hamada N, et al. Frequency of appearance of myeloperoxidase-antineutrophil cytoplasmic antibody (MPO-ANCA) in Graves' disease patients treated with propylthiouracil and the relationship between MPO-ANCA and clinical manifestations. *Clin. Endocrinol. (Oxf).* 2001;54:651–4.
12. Roth AJ, Ooi JD, Hess JJ, et al. Epitope specificity determines pathogenicity and detectability in ANCA-associated vasculitis. *J. Clin. Invest.* 2013;123:1773–83.
13. Espeli M, Bökers S, Giannico G, et al. Local renal autoantibody production in lupus nephritis. *J. Am. Soc. Nephrol.* 2011;22:296–305.
14. Kain R, Matsui K, Exner M, et al. A novel class of autoantigens of anti-neutrophil cytoplasmic antibodies in necrotizing and crescentic glomerulonephritis: the lysosomal membrane glycoprotein h-lamp-2 in neutrophil granulocytes and a related membrane protein in glomerular endothelial cell. *J. Exp. Med.* 1995;181:585–97.

15. Roth AJ, Brown MC, Smith RN, et al. Anti-LAMP-2 antibodies are not prevalent in patients with antineutrophil cytoplasmic autoantibody glomerulonephritis. *J. Am. Soc. Nephrol.* 2012;23:545–55.
16. Flint SM, Savage CO. Anti-LAMP-2 autoantibodies in ANCA-associated pauci-immune glomerulonephritis. *J. Am. Soc. Nephrol.* 2012;23:378–80.
17. Kain R. L29. Relevance of anti-LAMP-2 in vasculitis: why the controversy. *Presse Med.* 2013;42:584–8.
- *18. Peschel A, Basu N, Benharkou A, et al. Autoantibodies to hLAMP-2 in ANCA-negative pauci-immune focal necrotizing GN. *J. Am. Soc. Nephrol.* 2014;25:455–63.
Reports anti-LAMP-2 autoantibodies in patients with ANCA-negative vasculitis, and adds further information regarding LAMP-2 glycosylation patterns and detection.
19. Takeuchi S, Kimura S, Soma Y, et al. Lysosomal-associated membrane protein-2 plays an important role in the pathogenesis of primary cutaneous vasculitis. *Rheumatology (Oxford).* 2013;52:1592–8.
20. Suzuki K, Nagao T, Itabashi M, et al. A novel autoantibody against moesin in the serum of patients with MPO-ANCA-associated vasculitis. *Nephrol. Dial. Transplant* 2014;29:1168–77.
21. Fujinami RS, von Herrath MG, Christen U, Whitton JL. Molecular mimicry, bystander activation, or viral persistence: infections and autoimmune disease. *Clin. Microbiol. Rev.* 2006;19:80–94.
22. Welsh RM, Che JW, Brehm MA, Selin LK. Heterologous immunity between viruses. *Immunol. Rev.* 2010;235:244–66.
23. Kain R, Exner M, Brandes R, et al. Molecular mimicry in pauci-immune focal necrotizing glomerulonephritis. *Nat. Med.* 2008;14:1088–96.
24. Pendergraft WF, Preston G a, Shah RR, et al. Autoimmunity is triggered by cPR-3(105-201), a protein complementary to human autoantigen proteinase-3. *Nat. Med.* 2004;10:72–9.
25. Tadema H, Kallenberg CGM, Stegeman C a, Heeringa P. Reactivity against complementary proteinase-3 is not increased in patients with PR3-ANCA-associated vasculitis. *PLoS One* 2011;6:e17972.
26. Stegeman CA, Tervaert JW, Sluiter WJ, et al. Association of chronic nasal carriage of *Staphylococcus aureus* and higher relapse rates in Wegener granulomatosis. *Ann. Intern. Med.* 1994;120:12–7.

27. Pilszczek FH, Salina D, Poon KKH, et al. A novel mechanism of rapid nuclear neutrophil extracellular trap formation in response to *Staphylococcus aureus*. *J. Immunol.* 2010;185:7413–25.
28. Holle JU, Windmüller M, Lange C, et al. Toll-like receptor TLR2 and TLR9 ligation triggers neutrophil activation in granulomatosis with polyangiitis. *Rheumatology (Oxford)*. 2013;52:1183–9.
29. Husmann C a, Holle JU, Moosig F, et al. Genetics of toll like receptor 9 in ANCA associated vasculitides. *Ann. Rheum. Dis.* 2014;73:890–6.
30. Pendergraft WF, Niles JL. Trojan horses: drug culprits associated with antineutrophil cytoplasmic autoantibody (ANCA) vasculitis. *Curr. Opin. Rheumatol.* 2014;26:42–9.
31. Alberici F, Martorana D, Bonatti F, et al. Genetics of ANCA-associated vasculitides: HLA and beyond. *Clin. Exp. Rheumatol.* 2014;32 Suppl 8:90–7.
32. Xie G, Roshandel D, Sherva R, et al. Association of granulomatosis with polyangiitis (Wegener’s) with HLA-DPB1*04 and SEMA6A gene variants: evidence from genome-wide analysis. *Arthritis Rheum.* 2013;65:2457–68.
33. Raychaudhuri S, Sandor C, Stahl E a, et al. Five amino acids in three HLA proteins explain most of the association between MHC and seropositive rheumatoid arthritis. *Nat. Genet.* 2012;44:291–6.
34. Sollid LM, Pos W, Wucherpfennig KW. Molecular mechanisms for contribution of MHC molecules to autoimmune diseases. *Curr. Opin. Immunol.* 2014;31C:24–30.
35. Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. *Science* 2004;303:1532–5.
36. Kessenbrock K, Krumbholz M, Schönemärck U, et al. Netting neutrophils in autoimmune small-vessel vasculitis. *Nat. Med.* 2009;15:623–5.
37. Sangaletti S, Tripodo C, Chiodoni C, et al. Neutrophil extracellular traps mediate transfer of cytoplasmic neutrophil antigens to myeloid dendritic cells toward ANCA induction and associated autoimmunity. *Blood* 2012;120:3007–18.
- *38. Nakazawa D, Shida H, Tomaru U, et al. Enhanced formation and disordered regulation of NETs in myeloperoxidase-ANCA-associated microscopic polyangiitis. *J. Am. Soc. Nephrol.* 2014;25:990–7.
Demonstrates that anti-MPO ANCA can induce NET formation.

39. Ronco P, Verroust P, Mignon F, et al. Immunopathological studies of polyarteritis nodosa and Wegener's granulomatosis: a report of 43 patients with 51 renal biopsies. *Q. J. Med.* 1983;52:212–23.
40. Xiao H, Schreiber A, Heeringa P, et al. Alternative complement pathway in the pathogenesis of disease mediated by anti-neutrophil cytoplasmic autoantibodies. *Am. J. Pathol.* 2007;170:52–64.
41. Xiao H, Dairaghi DJ, Powers JP, et al. C5a receptor (CD88) blockade protects against MPO-ANCA GN. *J. Am. Soc. Nephrol.* 2014;25:225–31.
42. Schreiber A, Xiao H, Jennette JC, et al. C5a receptor mediates neutrophil activation and ANCA-induced glomerulonephritis. *J. Am. Soc. Nephrol.* 2009;20:289–98.
43. Persson U, Gullstrand B, Pettersson A, et al. A candidate gene approach to ANCA-associated vasculitis reveals links to the C3 and CTLA-4 genes but not to the IL1-Ra and Fcy-RIIa genes. *Kidney Blood Press. Res.* 2013;37:641–8.
44. Finn JE, Zhang L, Agrawal S, et al. Molecular analysis of C3 allotypes in patients with systemic vasculitis. *Nephrol. Dial. Transplant* 1994;9:1564–7.
45. Persson U, Truedsson L, Westman KW, Segelmark M. C3 and C4 allotypes in anti-neutrophil cytoplasmic autoantibody (ANCA)-positive vasculitis. *Clin. Exp. Immunol.* 1999;116:379–82.
46. Haas M, Eustace JA. Immune complex deposits in ANCA-associated crescentic glomerulonephritis: a study of 126 cases. *Kidney Int.* 2004;65:2145–52.
47. Chen M, Xing G-Q, Yu F, et al. Complement deposition in renal histopathology of patients with ANCA-associated pauci-immune glomerulonephritis. *Nephrol. Dial. Transplant* 2009;24:1247–52.
- *48. Gou S-J, Yuan J, Wang C, et al. Alternative complement pathway activation products in urine and kidneys of patients with ANCA-associated GN. *Clin. J. Am. Soc. Nephrol.* 2013;8:1884–91.
Provides evidence of activation of the alternative pathway for complement in human ANCA-associated vasculitis.
49. Gou S-J, Yuan J, Chen M, et al. Circulating complement activation in patients with anti-neutrophil cytoplasmic antibody-associated vasculitis. *Kidney Int.* 2013;83:129–37.
50. Yuan J, Gou S-J, Huang J, et al. C5a and its receptors in human anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis. *Arthritis Res. Ther.* 2012;14:R140.

51. Van den Berg CW, Tambourgi D V, Clark HW, et al. Mechanism of neutrophil dysfunction: neutrophil serine proteases cleave and inactivate the C5a receptor. *J. Immunol.* 2014;192:1787–95.
52. Niles J, Laliberte KA, Murphy AP, et al. Anti-C5 Monoclonal Antibody - Eculizumab: Use in the Treatment of Life Threatening ANCA Associated Vasculitis with Pulmonary Hemorrhage and Profound Anemia [Abstract]. *J. Am. Soc. Nephrol.* 2014;25:950A.
53. Wilde B, Thewissen M, Damoiseaux J, et al. Regulatory B cells in ANCA-associated vasculitis. *Ann. Rheum. Dis.* 2013;72:1416–9.
54. Todd SK, Pepper RJ, Draibe J, et al. Regulatory B cells are numerically but not functionally deficient in anti-neutrophil cytoplasm antibody-associated vasculitis. *Rheumatology (Oxford).* 2014;53:1693–703.
55. Lapse N, Abdulahad WH, Rutgers A, et al. Altered B cell balance, but unaffected B cell capacity to limit monocyte activation in anti-neutrophil cytoplasmic antibody-associated vasculitis in remission. *Rheumatology (Oxford).* 2014;53:1683–92.
56. Bunch DO, McGregor JG, Khandoobhai NB, et al. Decreased CD5⁺ B cells in active ANCA vasculitis and relapse after rituximab. *Clin. J. Am. Soc. Nephrol.* 2013;8:382–91.
57. Abdulahad WH, Stegeman C a, van der Geld YM, et al. Functional defect of circulating regulatory CD4⁺ T cells in patients with Wegener’s granulomatosis in remission. *Arthritis Rheum.* 2007;56:2080–91.
58. Rimbart M, Hamidou M, Braudeau C, et al. Decreased numbers of blood dendritic cells and defective function of regulatory T cells in antineutrophil cytoplasmic antibody-associated vasculitis. *PLoS One* 2011;6:e18734.
59. Chavele K-M, Shukla D, Ketepe-Arachi T, et al. Regulation of myeloperoxidase-specific T cell responses during disease remission in antineutrophil cytoplasmic antibody-associated vasculitis: the role of Treg cells and tryptophan degradation. *Arthritis Rheum.* 2010;62:1539–48.
60. Free ME, Bunch DO, McGregor JA, et al. Patients with antineutrophil cytoplasmic antibody-associated vasculitis have defective Treg cell function exacerbated by the presence of a suppression-resistant effector cell population. *Arthritis Rheum.* 2013;65:1922–33.

Figure titles and legends:**Figure 1.**

Excessive neutrophil extracellular trap (NET) formation promotes the development of ANCA by allowing uptake of MPO and PR3 by antigen presenting cells (APC) in a stimulatory context. ANCA further stimulates neutrophils to generate NETs through NETosis.

Figure 2.

Activation of the alternative complement pathway results in the generation of C5a, which exerts a broad range of downstream effects on neutrophils through the ligation of its receptor, C5aR, including the further generation of complement in a positive feedback loop.