

Still scratching the surface: how much of the 'black box' of soil ectomycorrhizal communities remains in the dark?

Article

Accepted Version

Pickles, B. J. and Pither, J. (2014) Still scratching the surface: how much of the 'black box' of soil ectomycorrhizal communities remains in the dark? New Phytologist, 201 (4). pp. 1101-1105. ISSN 0028-646X doi: https://doi.org/10.1111/nph.12616 Available at http://centaur.reading.ac.uk/48103/

It is advisable to refer to the publisher's version if you intend to cite from the work.

Published version at: http://dx.doi.org/10.1111/nph.12616

To link to this article DOI: http://dx.doi.org/10.1111/nph.12616

Publisher: Wiley

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the End User Agreement.

www.reading.ac.uk/centaur



CentAUR

Central Archive at the University of Reading Reading's research outputs online

- 1 Still scratching the surface: how much of the "black box" of soil
- 2 ectomycorrhizal communities remains in the dark?

4 Brian J. Pickles, Jason Pither

5

- 6 Irving K. Barber School of Arts and Sciences, Department of Biology, University of British
- 7 Columbia, Okanagan campus, 3333 University Way, Kelowna, BC V1V 1V7, Canada.

8

- 9 Author for correspondence:
- 10 Brian J Pickles
- 11 *Tel:* +1 250 807 9965
- 12 Fax: +1 250 807 8005
- 13 Email: brian.pickles@ubc.ca

15 Article type: Letter

16

- 17 **Key words**: Community ecology; diversity; ectomycorrhizas; meta-analysis; rooting depth;
- soil depth.

	rtı	ΛI	•
$\boldsymbol{\Gamma}$	rti	u	ĸ

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

Symbiotic soil organisms such as ectomycorrhizal fungi (EMF) were long thought of as an inscrutable "black-box", yet the advent of molecular technologies has driven rapid advances in identification and enumeration of their diversity (Horton & Bruns, 2001; Buée et al., 2009). For instance, one 20 cm soil core can impressively yield 100's of fungal OTUs (Taylor et al., 2013). Importantly root symbionts play functional roles in sequestration or breakdown of soil carbon pools (Trumbore & Czimczik, 2008; Harrison et al., 2011a,b; Clemmensen et al., 2013; Kramer et al., 2013), nutrient and water cycling (Virginia et al., 1986; Read & Perez-Moreno, 2003), alteration of soil porosity (Perry et al., 1990), and provision of sustenance for different trophic levels (Coleman & Whitman, 2005). Yet root symbionts occur and contribute to function far deeper in the soil than is usually sampled (Jenkins et al., 1988; Dalpé et al., 2000; Bornyasz et al., 2005). Soil properties vary considerably among ecosystems (Schenk, 2005; Dickie et al., 2013), hence so too does rooting depth (see below) – even within a single species (Stone & Kalisz, 1991; Canadell et al., 1996). However, in practice, we are (understandably) encouraged to employ uniform sampling strategies, even if these are known to only scratch the surface of potential symbiont habitat in some ecosystems (see below). Although the issue of limited-depth sampling has been raised before (Taylor, 2002), we are unaware of any efforts to quantify how much of the "black box" typically remains out of reach of standard sampling techniques. Such information would be extremely timely, due to the growing interest in accurately characterizing global patterns of EMF diversity and distribution (Dickie & Moyerson 2008; Vellinga et al., 2009; Tedersoo et al., 2012). To begin addressing this question, we gathered sampling depth data from recent field studies of EMF, and analysed these in relation to published data compiled by ecosystem ecologists regarding (i) maximum rooting depths of trees and shrubs, including 137 EMF host species distributed among 29 host genera, and (ii) estimates for 8 ecosystems of the mean depth above

Page 4 of 14

46	which 95% of all roots are located. Rooting depth data were derived from the following
47	sources: (i) EMF host species/genera from Stone & Kalisz (1991) and Canadell et al. (1996),
48	(ii) ecosystem data from Schenk & Jackson (2002). Sampling depth data were obtained from
49	EMF studies published in the last 5 years of <i>New Phytologist</i> (Table S1). While the concepts
50	discussed here are equally applicable to all soil-borne root symbionts, for the sake of brevity
51	we focus our attention specifically on EMF and their hosts.
52	Based on 27 articles that reported sampling depth, the average was 13.4 cm (\pm 1.59 s.e.m.),
53	with a median value of 10 cm. This sampling depth was approximately doubled in boreal and
54	semi-arid ecosystems, and halved in semi-arid and tropical evergreen ecosystems. In
55	comparison, none of the 29 ectomycorrhizal host genera for which data was available exhibited
56	maximum rooting depths shallower than 50 cm (Fig. 1a), and on average maximum rooting
57	depth among the 137 host species is 530 cm (± 44 cm s.e.m.) (Fig. 1b). Correspondingly, the
58	average proportion of maximum rooting depth assessed is estimated to be $0.068 \ (\pm \ 0.0071$
59	s.e.m.) across all host genera. If we consider maximum rooting depth as a proxy for the
60	amount of habitat available to symbionts, then an enormous amount of potential habitat
61	remains under-sampled, even within the Pinaceae (Fig. 1), which, according to a 2008
62	literature survey (Dickie & Moyerson, 2008), represented the focal family in 62% of all EMF
63	studies.
54	Although maximum rooting depth is a crucial variable in research examining ecosystem
65	function (Canadell et al., 1996; Jackson et al., 1996; Schenk, 2005), it could be argued that for
66	our purposes it provides an overly pessimistic outlook on the completeness of current sampling
67	efforts. We therefore also considered the EMF sampling depth data in relation to estimates of
68	ecosystem-specific mean rooting depths calculated using 16 to 59 observations per ecosystem
69	type, spanning all tree and shrub species for which rooting depth data existed (Schenk &
70	Jackson, 2002). Using an average sampling depth of 13.4 cm, the proportion of the mean
71	ecosystem rooting depth sampled varied from a high of 0.47 for tundra, to a low of 0.08 for

72	Mediterranean ecosystems (Fig. 2a), with a mean of 0.178 ± 0.0442 s.e.m.). Thus, even in
73	tundra ecosystems where rooting depths are comparatively shallow (Fig. 2b), typical sampling
74	methods are likely to access less than 50% of the mean depth of host roots.
75	Although striking, our findings do not necessarily mean that standard sampling methods are
76	always doomed to miss an important or sizeable component of the symbiont assemblage
77	associated with any given host. Indeed, it is likely that some studies - especially those
78	occurring in shallow rooting regions (e.g. tundra ecosystems) - could yield reasonable
79	estimates of the actual number of symbiont species associated with the host (using appropriate
80	analytical techniques; cf. Gotelli & Colwell, 2001). Nevertheless, it has long been
81	acknowledged that important characteristics of EMF communities vary with depth, but only in
82	recent years have studies begun to clarify these details. For example, fungal hyphae show
83	vertical niche differentiation (Dickie et al., 2002), EMF community composition changes
84	between soil horizons (Rosling et al., 2003), ECM root tips and EMF extramatrical mycelium
85	differ in their vertical structure (Genney et al., 2006), and other depth-associated patterns
86	continue to emerge (e.g. Egerton-Warburton et al., 2003; Landeweert et al., 2003; Baier et al.,
87	2006; Lindahl et al., 2007; Courty et al., 2008; Scattolin et al., 2008; Beiler et al., 2010;
88	Clemmensen et al., 2013; Taylor et al,. 2013). These observations, combined with our
89	findings, substantiate earlier statements that current sampling methods provide a limited view
90	of EMF assemblages (Taylor, 2002). Until more effort is spent sampling and characterising
91	symbiont diversity and function at depth, we cannot know the true extent of these limitations.
92	Since deep roots are features of most ecosystems worldwide (Schenk & Jackson, 2005), the
93	discoveries that could come with deeper sampling have the potential to profoundly change our
94	outlook on patterns of EMF diversity and function. To illustrate, consider a recent and
95	enlightening global-extent meta-analysis of local EMF diversity (Tedersoo et al., 2012). Based
96	on data from 55 published studies, total species richness (representing site-level species
97	richness) was significantly associated with a number of climate-based predictor variables (e.g.

mean annual temperature, mean annual precipitation), and not surprisingly, number of samples
and total sample volume. However, the meta-analysis included data gathered from a variety of
host genera and ecosystem types, meaning that the rooting depths of hosts also varied
substantially (see above). It would be interesting to determine if and how their findings would
change if sampling was deeper, or was adjusted to account for site-specific rooting depths.
Because different communities arise with increasing depth, we predict that deeper sampling
will increase estimates of total richness and reveal significant changes in community
composition. Perhaps every additional 50 cm of depth explored could provide as much
richness again as that found in the organic horizon (as per Rosling et al., 2003 & Landeweert et
al., 2003)? Based upon our findings, we speculate that the magnitude of this total increase will
vary significantly with ecosystem type due to the differences in host rooting depth and density.
Variation in the rooting depth of a given host species is related to multiple factors including
age, depth to bedrock, mean annual precipitation, mean annual potential evapotranspiration,
and depth to the water table (Schenk 2005), all co-varying with ecosystem type. Thus, a
Douglas-fir growing in seasonally dry evergreen forest is more likely to develop deep roots
than one growing in a cool-temperate to sub-boreal region (cf. Schenk & Jackson, 2002). This
has implications for sampling strategies (see below), and suggests that host species distributed
across multiple ecosystem types, like Douglas-fir, may be associated with a much more diverse
pool of EMF symbionts than current estimates indicate. This combination of varied rooting
depths and soil environments provides a greater diversity of habitat to symbionts than do hosts
whose distributions are predominantly restricted to a single ecosystem type (e.g. black spruce).
Another important finding concerns the thoroughness with which sampling methods are
described in published articles. Of the 30 EMF studies published in the past 5 years in New
Phytologist, 3 (10%) failed to report details about sampling depth. More generally, whereas
some authors give detailed descriptions of the soil environment in relation to sampling strategy
(e.g. Smith et al., 2005; Ryberg et al., 2009), depth information occasionally has to be derived

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

Columbia, Canada.

New Phytologist

Pickles & Pither 2013

or is missing entirely. We suggest that where possible, details about sampling should be accompanied by estimates of average rooting depths at the site, for the host species of interest, even if these estimates are speculative. This would provide for better and more consistent estimates of realized sampling effort across studies. Lastly, future research may not only require deeper sampling to minimise bias (depending upon the research objectives), but may also need to stratify sampling geographically according to potential rooting depth. Combining global estimates of soil depth (http://www.fao.org/nr/land/soils/harmonized-world-soil-database/en/) with global estimates of deep root distributions (Schenk & Jackson, 2005) and species' ranges (e.g. http://esp.cr.usgs.gov/data/little/) could help hone in on potential sampling regions, and ground penetrating radar technology (Sucre et al., 2011) could be used to identify final sample locations. The logistical impediments associated with deep soil sampling (including cost; Harrison et al., 2011) are daunting, but other research areas point to possible solutions, such as using drilling equipment to acquire ice or sediment cores (Nogué et al., 2013), or using excavation machinery such as a backhoe (Bornyasz et al., 2005). These challenges are worth tackling given the potentially crucial roles that symbionts at depth may play in ecosystem function (e.g. Clemmensen et al. 2013; Kramer et al. 2013). Acknowledgements We thank Colin Scherer and Emma Walker for assistance with data mining, and Ian Dickie and two anonymous reviewers for their insightful comments on this manuscript. BJP acknowledges financial support from the Simard and Mohn labs at the University of British Columbia, Canada. JP acknowledges financial support from the Natural Sciences and Engineering Research Council of Canada (Discovery Grants program), the Canada Foundation for Innovation, and the I.K. Barber School of Arts and Sciences at the Okanagan campus of the University of British

1	. 5	0

- References
- Baier R, Ingenhaag J, Blaschke H, Gottlein A, Agerer R. 2006. Vertical distribution of an
- ectomycorrhizal community in upper soil horizons of a young Norway spruce (*Picea abies* [L.]
- Karst.) stand of the Bavarian Limestone Alps. *Mycorrhiza* **16**: 197-206.
- Beiler KJ, Durall DM, Simard SW, Maxwell SA, Kretzer AM. 2010. Architecture of the
- wood-wide web: *Rhizopogon* spp. genets link multiple Douglas-fir cohorts. *New Phytologist*
- 157 **185**: 543-553.
- Bornyasz MA, Graham RC, Allen MF. 2005. Ectomycorrhizae in a soil-weathered granitic
- bedrock regolith: Linking matrix resources to plants. *Geoderma* **126**: 141-160.
- Buée M, Reich M, Murat C, Morin E, Nilsson RH, Uroz S, Martin F. 2009. 454
- pyrosequencing analyses of forest soils reveal an unexpectedly high fungal diversity. New
- 162 Phytologist 184: 449-456.
- 163 Canadell J, Jackson RB, Ehleringer JR, Mooney HA, Sala OE, Schulze E-D. 1996. Maximum
- rooting depth of vegetation types at the global scale. *Oecologia* **108**: 583-595.
- 165 Clemmensen KE, Bahr A, Ovaskainen O, Dahlberg A, Ekblad A, Wallander H, Stenlid J,
- Finlay RD, Wardle DA, Lindahl BD, 2013. Roots and associated fungi drive long-term carbon
- sequestration in Boreal forest. *Science* **339**: 1615-1618.
- 168 Coleman DC, Whitman WB. 2005. Linking species richness, biodiversity and ecosystem
- function in soil systems. *Pedobiologia* **49**: 479-497.
- 170 Courty P-E, Franc A, Pierrat J-C, Garbaye J. 2008. Temporal changes in the ectomycorrhizal
- 171 community in two soil horizons of a temperate oak forest. Applied and Environmental
- 172 *Microbiology* **79**: 5792-5801.
- Dalpé Y, Diop TA, Plenchette C, Gueye M. 2000. Glomales species associated with surface
- and deep rhizosphere of *Faidherbia albida* in Senegal. *Mycorrhiza* **10**: 125-129.

Pickles & Pither 2013

- Dickie IA, Moyerson B. 2008. Towards a global view of ectomycorrhizas. *New Phytologist*
- 176 **180**: 263-265.
- Dickie IA, Xu B, Koide RT. 2002. Vertical niche differentiation of ectomycorrhizal hyphae in
- soil as shown by T-RFLP analysis. *New Phytologist* **156**: 527-535.
- Dickie IA, Martinez-Garcia LB, Koele N, Grelet G-A, Tylianakis JM, Peltzer DA, Richardson
- 180 SJ. 2013. Mycorrhizas and mycorrhizal fungal communities throughout ecosystem
- 181 development. *Plant & Soil* **367**: 11-39.
- 182 Egerton-Warburton LM, Graham RC, Hubbert KR. 2003. Spatial variability in mycorrhizal
- hyphae and nutrient and water availability in a soil-weathered bedrock profile. *Plant and Soil*
- **249**: 331-342.
- 185 Genney DR, Anderson IC, Alexander IA. 2006. Fine-scale distribution of pine
- ectomycorrhizas and their extramatrical mycelium. *New Phytologist* **170**: 381-390.
- Gotelli NJ, Coldwell RK. 2001. Quantifying biodiversity: procedures and pitfalls in the
- measurement and comparison of species richness. *Ecology Letters* **4**: 379-391.
- Harrison RB, Footen PW, Strahm BD. 2011a. Deep soil horizons: contribution and importance
- to soil carbon pools and in assessing whole-ecosystem response to management and global
- 191 change. Forest Science 57: 67-76.
- Harrison RB, Richter D, Fox T. 2011b. Deep soils. Forest Science 57: 1-2.
- Horton TR, Bruns TD. 2001. The molecular revolution in ectomycorrhizal ecology: peeking
- into the black-box. *Molecular Ecology* **10**: 1855-1871.
- 195 Jackson RB, Canadell J, Ehleringer JR, Mooney HA, Sala OE, Schulze ED. 1996. A global
- analysis of root distributions for terrestrial biomes. *Oecologia* **108**: 389-411.
- 197 Jenkins MB, Virginia RA, Jarrell WM. 1988. Depth distribution and seasonal populations of
- 198 Mesquite-nodulating Rhizobia in warm desert ecosystems. Soil Science Society of America
- 199 *Journal* **52**: 1644-1650.

- 200 Kraft NJB, Comita LS, Chase JM, Sanders NJ, Swenson NG, Crist TO, Stegan JC, Vellend M,
- Boyle B, Anderson MJ et al. 2011. Disentangling the drivers of β diversity along latitudinal
- and elevational gradients. *Science* **333**: 1755-1758.
- 203 Kramer S, Marhan S, Haslwimmer H, Ruess L, Kandeler E. 2013. Temporal variation in
- surface and subsoil abundance and function of the soil microbial community in an arable soil.
- 205 Soil Biology and Biochemistry **61**: 76-85.
- Landeweert R, Leeflang P, Kuyper TW, Hoffland E, Rosling A, Wernars K, Smit E. 2003.
- 207 Molecular identification of ectomycorrhizal mycelium in soil horizons. *Applied Environmental*
- 208 *Microbiology* **69**: 327-333.
- Lindahl BD, Ihrmark K, Boberg J, Trumbore SE, Högberg P, Stenlid J, Finlay RD. 2007.
- 210 Spatial separation of litter decomposition and mycorrhizal nitrogen uptake in a boreal forest.
- 211 *New Phytologist* **173**: 611-620.
- Nogué S, de Nascimento L, Fernández-Palacios JM, Whittaker RJ, Willis KJ. 2013. The
- ancient forests of La Gomera, Canary Islands, and their sensitivity to environmental change.
- 214 *Journal of Ecology* **101:** 368-377.
- 215 Perry DA, Borchers JG, Borchers SL, Amaranthus MP. 1990. Species migrations and
- 216 ecosystem stability during climate change: the belowground connection. Conservation Biology
- **4**: 266-274.
- 218 Read DJ, Perez-Moreno J. 2003. Mycorrhizas and nutrient cycling in ecosystems a journey
- towards relevance? *New Phytologist* **157**: 475-492.
- 220 Rosling A, Landeweert R, Lindahl BD, Larsson K-H, Kuyper TW, Taylor AFS Findlay RD.
- 221 2003. Vertical distribution of ectomycorrhizal fungal taxa in a podzol soil profile. New
- 222 *Phytologist* **159**: 775-783.
- 223 Ryberg M, Andreasen M, Björk RG. 2010. Weak habitat specificity in ectomycorrhizal
- communities associated with Salix herbacea and Salix polaris in alpine tundra. *Mycorrhiza* 21:
- 225 289-296.

Pickles & Pither 2013

- Scattolin L, Montecchio L, Mosca E, Agerer R. 2008. Vertical distribution of the
- 227 ectomycorrhizal community in the top soil of Norway spruce stands. European Journal of
- 228 Forest Research 127: 347-357.
- Schenk HJ. 2005. Vertical vegetation structure below ground: Scaling from root to globe.
- 230 *Progress in Botany* **66**: 341-373.
- Schenk HJ, Jackson RB. 2002. The global biogeography of roots. *Ecological Monographs*
- **72**: 311-328.
- Schenk HJ, Jackson RB. 2005 Mapping the global distribution of deep roots in relation to
- climate and soil characteristics. *Geoderma* **126:** 129-140.
- Smith JE, McKay D, Brenner G, McIver J, Spatafora JW. 2005. Early impacts of forest
- restoration treatments on the ectomycorrhizal fungal community and fine root biomass in a
- mixed conifer forest. *Journal of Applied Ecology* **42**: 526-535.
- 238 Stone EL, Kalisz PJ. 1991. On the maximum extent of tree roots. Forest Ecology &
- 239 Management **46**: 59 –102.
- 240 Sucre EB, Tuttle JW, Fox TR. 2011. The use of ground-penetrating radar to accurately
- estimate soil depth in rocky forest soils. Forest Science 57: 59-66.
- 242 Taylor AFS. 2002. Fungal diversity in ectomycorrhizal communities: sampling effort and
- species detection. Plant & Soil 244: 19-28.
- 244 Taylor DL, Hollingsworth TN, McFarland, Lennon NJ, Nusbaum C, Ruess RW. 2013. A first
- comprehensive census of fungi in soil reveals both hyperdiversity and fine-scale niche
- partitioning. *Ecology*, in press: http://dx.doi.org/10.1890/12-1693.1
- Trumbore SE, Czimczik CI. 2008. An uncertain future for soil carbon. *Science* **321**: 1455-1456.
- Vellinga EC, Wolfe BE, Pringle A. 2009. Global patterns of ectomycorrhizal introductions.
- 249 *New Phytologist* **181**: 960-973.
- Virginia RA, Jenkins MB, Jarell WM. 1986. Depth of root symbiont occurrence in soil. *Biology*
- 251 *and Fertility of Soils* **2**: 127-130.

Figure legends
Figure 1. a. Proportion of maximum recorded rooting depth examined (± s.e.m.) across general
using mean sampling depth derived from values reported in the literature (Table S1). Note
maximum y-axis value is a proportion of 0.15. b. Maximum rooting depths of selected host
species, with multiple bars (records) per species. Dashed red line represents average sampling
depth of EMF studies. In both panels, green bars indicate genera or species in the Pinaceae.
Figure 2. a. Sampled proportion of the mean depth at which 95% of ecosystem roots are
located, calculated using mean of sampling values reported in literature (Table S1). b.
Estimated mean depth (\pm s.e.m.) at which 95% of ecosystem roots are located using the
interpolated values of Schenk & Jackson (2002).
Supporting Information
Table S1. Citation, sample depth and host species for all ectomycorrhizal articles from the last
5 years of New Phytologist in which sampling depth was provided.



