

1 **Title:**

2 MR1: a multi-faceted metabolite sensor for T cell activation

3

4 **Author names and affiliations:**

5 Hamish E.G. McWilliam^{1-2#} & Jose A. Villadangos^{1-2#}

6

7 ¹Department of Microbiology and Immunology, Peter Doherty Institute for Infection and Immunity,
8 University of Melbourne, Parkville, Victoria 3010, Australia

9

10 ²Department of Biochemistry and Molecular Biology, Bio21 Molecular Science and Biotechnology
11 Institute, The University of Melbourne, Parkville, Victoria, Australia.

12

13 # joint corresponding authors. e-mail: hamish.mcwilliam@unimelb.edu.au,

14 j.villadangos@unimelb.edu.au

15 **Abstract:**

16 The major histocompatibility complex class I-related molecule MR1 captures and presents small
17 metabolites to MR1-restricted T cells including Mucosal Associated Invariant T (MAIT) cells. The
18 first MR1 ligands discovered were intermediates of microbial riboflavin synthesis, antigens presented
19 to alert inflammatory MAIT cells to bacterial infection. Recent advances have expanded the range of
20 MR1 ligands to include extracellular metabolites released by the commensal microbiome, and yet
21 undefined antigens presented by cancer cells to mediate MR1-dependent anti-tumor activity. MR1
22 thus exhibits a multifaceted ability to display a diverse range of ligands for immune surveillance in a
23 variety of contexts. The mechanisms of antigen presentation by MR1 are of central importance to
24 understanding metabolite-mediated immune homeostasis, immunity to infection and tumor
25 surveillance.

26

27

28 **Highlights:**

- 29 • MR1 captures a conserved metabolite signature of a diverse range of microbes and activates
30 MAIT cells for inflammation and immunity.
- 31 • Commensal microbes at barrier tissues secrete metabolites that are captured and presented by
32 MR1 in distant tissues such as the thymus.
- 33 • MR1 displayed by a various cancer cells elicit activation of MR1-restricted T cells for immune
34 control of cancers.
- 35 • The trafficking of MR1 to sample its varied metabolite cargo requires a unique pathway
36 among antigen-presenting molecules.

37

38 **Introduction**

39 The presentation of foreign antigen (Ag) by Major Histocompatibility Complex (MHC) molecules is
40 a pivotal step in adaptive immunity, which allows the activation, expansion and effector functions of
41 T lymphocytes. These cells possess a highly specific T cell receptor (TCR) that recognizes diverse
42 classes of Ag in the context of specialized MHC molecules. Conventional T cells recognize peptides
43 presented by classical MHC molecules, whereas innate-like T cells recognize conserved non-peptidic
44 Ag presented by non-classical MHC molecules, such as the monomorphic MHC class I-related
45 protein 1 (MR1). MR1 is highly conserved among mammals[1], and captures and presents small
46 metabolites to MR1-restricted T cells[2], the largest population of which are Mucosal Associated
47 Invariant T (MAIT) cells. The best characterized Ag presented by MR1 are transient metabolites
48 derived from the microbial synthesis of vitamin B2, riboflavin, which are produced by many microbes
49 but not mammals[3]. The riboflavin-precursor 5-amino-6-D-ribitylaminouracil (5-A-RU) combines
50 with methylglyoxal to form the MR1-binding derivative 5-(2-oxopropylideneamino)-6-D-
51 ribitylaminouracil (5-OP-RU)[2]. Unlike protein Ag which can mutate and drift, metabolites are
52 highly conserved and are essential bacterial ‘building blocks’ produced by a diverse range of
53 microbes. Therefore, MR1 presents a metabolite signature to allow the detection of pathogenic
54 microbes by the immune system for their subsequent clearance.

55

56 Recent discoveries that have expanded the roles attributed to MR1 metabolite Ag presentation (Figure
57 1): (i) surveillance of not only pathogenic but also commensal microbes; (ii) presentation of a cancer
58 metabolite signature for the immune recognition of tumors. The mechanism by which MR1 presents
59 this unique class of Ag is of central importance to understanding the whole range of functions of the
60 conserved MR1-MAIT cell axis. This review outlines these areas and proposes future directions for
61 the field.

62

63 **MR1 presents microbial metabolites for immunity and barrier homeostasis**

64 MAIT cells are classed as innate-like T cells. They are abundant in mucosal and barrier tissues and
65 are considered antigen-experienced, tissue-resident cells [4,5]. MAIT cells rapidly secrete pro-
66 inflammatory cytokines upon recognition of their MHC-presented cognate antigen, specifically
67 interferon- γ , tumor necrosis factor- α and interleukin-17A [5-8], and this inflammatory potential
68 somewhat defines MAIT cells. During infection they have also been shown to recruit conventional T
69 cells and secrete granulocyte-macrophage colony-stimulating factor which differentiates monocytes
70 into dendritic cells, together arming the adaptive response [9,10]. Furthermore, MAIT cells can
71 directly kill infected cells presenting MR1-metabolite complexes [11,12]. Many bacterial pathogens
72 produce the riboflavin-related metabolites presented by MR1, and much of the early work that

73 addressed the function of the MR1-MAIT cell axis focused on its role in immunity against
74 intracellular bacteria [13]. For example, in mice MAIT cells have been shown to be active and/or
75 protective against a range of bacterial infections including tuberculosis [9,10,14-18] whereas in
76 humans MAIT cells are stimulated and enriched in the airways of tuberculosis-infected patients
77 [19,20] and activated in many other bacterial diseases [11,21].

78
79 Collectively this has suggested that MR1 evolved as a sensor of microbial, intracellular pathogens
80 [22]. However, two key recent studies have shown that *in vivo*, MR1 can capture soluble metabolites,
81 and not only from pathogenic but also from commensal bacteria. This suggests that MR1 may play a
82 key pivotal role in the crosstalk between microbiota and the mammalian immune system to maintain
83 homeostasis [23,24]. Interestingly, the commensal metabolites presented by MR1 in this scenario do
84 not need to originate within infected cells or in intracellular compartments of phagocytic cells
85 harboring microbes; instead they are bacterial metabolites released to the extracellular environment
86 in peripheral tissues [23,24]. The fate of these soluble compounds is remarkable. It was already
87 known that MAIT cells require commensal flora to colonize peripheral tissues [25], but a recent and
88 elegant study by Legoux et al [24] discovered that the first location where MAIT cells detect that
89 flora is actually the thymus. They showed that 5-OP-RU secreted by commensal microbes in the
90 periphery reaches the thymus, where it is presented to drive positive selection of developing MAIT
91 cells. Purified 5-OP-RU injected intraperitoneally, or administered on the skin or by oral gavage,
92 entered circulation and was likewise captured by thymocytes and presented on MR1. In another
93 unexpected development, Constantinides et al. [23] showed that 5-OP-RU administered on mouse
94 skin induced local accumulation of MAIT cells, another demonstration of presentation of soluble,
95 extracellular 5-OP-RU, in this case in a peripheral tissue. These studies provide direct evidence that
96 cells can be physically separated from commensals that synthesize 5-OP-RU and still capture and
97 present the metabolite on MR1 (Figure 1). There have been numerous studies showing cells readily
98 acquire and present extracellular metabolites *in vitro* [2,3,18,26-29], and Chen et al. [18] showed that
99 extracellular 5-OP-RU along with inflammatory signals can activate MAIT cells *in vivo*, but the
100 studies by Legoux et al [24] and Constantinides et al [23] represent the first evidence of a homeostatic
101 function for this source of MR1 ligands.

102
103 Other recent studies also implicate the MR1-MAIT cell axis in responding to and shaping the
104 microbiome; MR1-deficient mice harbored distinct microbiota to wildtype mice [30,31], and
105 antibiotic treatment depleted MAIT cell numbers [32]. Conversely, detection of changes in the
106 microbiome, or invasion of normally sterile tissues by commensal bacteria, may trigger reactions in
107 the host initiated by MAIT cells to restore homeostasis. Constantinides showed that MAIT cells had

108 wound-healing properties, and other studies have shown that MAIT cells are associated with gut
109 barrier integrity [31,33]. Together, these studies widen the scope of the roles played by the MR1-
110 MAIT cell axis and implicate multi-faceted mechanism for MR1 presentation of microbial
111 metabolites, catering for Ag from intra- and extracellular sources, from both commensal and infecting
112 pathogens (Figure 1), as will be discussed later.

113

114 **Presenting a metabolite signature of tumors?**

115 The metabolite 5-OP-RU is the canonical microbial MR1 ligand, a conserved antigen produced by a
116 range of microbes spanning different biological kingdoms; yet MR1 is capable of presenting a more
117 diverse range of metabolites. The structural flexibility of its Ag-binding cleft enables MR1 to
118 accommodate a diverse range ligands [34], including drug-like molecules [35] and other microbial
119 metabolites that have not been identified yet [36-38]. Recently, several studies have provided
120 compelling evidence that MR1 can present Ag from tumors for immune surveillance of cancers. Two
121 separate studies found MR1-restricted T cell clones that seem to recognize undefined cancer-specific
122 Ag [39,40]. Crowther et al [39] identified a T cell clone could recognize MR1 complexes presented
123 by cancer cells, but not healthy cells. This clone was highly cytolytic for a diverse range of human
124 tumor cells and it controlled a mouse model of leukemia. Furthermore, primary T cells transfected to
125 express the TCR of this clone could kill both autologous and non-autologous melanomas. These
126 results suggest that the TCR of this clone recognizes an MR1-presented Ag shared by human and
127 murine tumor cells, so T cells expressing this TCR have promise for use as a pan-cancer therapy [41].
128 Lepore et al. [40] also found MR1-restricted T cell clones that recognized MR1 likely presenting
129 tumor-derived ligands; these clones were stimulated by hydrophilic fractions from the lysates of THP-
130 1 leukemia cell line and murine breast tumors. Interestingly, the T cell clone discovered by Crowther
131 et al [39] did not recognize the canonical ligands 5-OP-RU or the vitamin B9-related metabolite, Ac-
132 6-FP. Further, it did not recognize MR1 with the residue lysine-43 mutated to alanine (K43A). This
133 lysine contains a positively charged side chain that needs to be neutralized by forming a Schiff base
134 bond with ligands lodged into the antigen binding site of MR1 to enable transport of the MR1-ligand
135 complexes from the endoplasmic reticulum (ER) to the cell surface [29]. MR1-K43A molecules
136 cannot form this covalent bond with metabolites although they can still bind non-Schiff base ligands
137 [42]. The fact that the anti-tumor clone described by Crowther et al requires MR1 molecules with the
138 lysine 43 present suggests it does not recognize MR1 alone but in complex with a Schiff base-forming
139 ligand.

140

141 These studies suggest a new role for MR1; not only to present microbial Ag but to display a metabolic
142 signature of cancer, which is potentially a unique metabolite from within transformed cells. The
143 identity of such ligands is thus a crucial question.

144

145 **How does MR1 present these diverse metabolites signatures?**

146 The picture that has emerged from studies of MR1 ligands is that this molecule can sample metabolite
147 Ag from within cells, derived from intracellular bacterial infections or produced by cancer cells
148 themselves. In addition, MR1 can present Ag derived from extracellular sources, produced by
149 commensal or pathogenic organisms. This versatility sets MR1 apart from other antigen presenting
150 molecules. MHC class II molecules utilize a presentation pathway that enables sampling the proteome
151 that reaches the lumen of endosomal compartments, including extracellular proteins. The pathway
152 used by MHC class I molecules is adapted to present mostly the cytosolic proteome, although in some
153 cells MHC class I can also present Ag sampled from endosomes via cross-presentation [43,44]. The
154 various human CD1 subclasses employ distinct trafficking routes to survey the full range of lipids
155 that occur in different compartments [45]. What are the mechanisms that enable MR1, the product of
156 a single, monomorphic gene [46], to achieve its multifaceted display of so many different metabolite
157 ligands?

158 This is an ongoing area of research and we still do not understand all the nuances of MR1 presentation.
159 In all cell types examined, there is a pool of pre-synthesized MR1 that is poised to capture Ag from
160 either extracellular sources or intracellular infection [28,29]. The majority of this pool is located
161 inside the ER in a ligand-receptive conformation, with a small portion at the cell surface or in
162 endosomes [29,47-50]. Metabolite Ag binding to MR1 causes it to traffic to the cell surface for
163 presentation [2,3,27,29,51]. Logic dictates that the location where the majority of ligand-receptive
164 MR1 resides is the compartment where the Ag is captured. In support of this, we found that the
165 binding of ligands to MR1, accompanied by covalent binding to the K43 side chain, causes the release
166 from the ER. The mechanism is hypothesized to be the neutralization of the K43 charge, allowing the
167 stable folding of MR1 [29]. We and others support an ER-binding model as the primary mode of
168 presentation of extracellular metabolites [22,28,29,52,53]. But is this the only way that MR1 acquires
169 its cargo?

170

171 There is substantial evidence that MR1 molecules already expressed on the cell surface can be used
172 to present Ag from exogenous or intracellular Ag. The first evidence comes from the recycling of
173 MR1 molecules that acquired ligands in the ER and then trafficked to the cell surface. Most of these
174 molecules are endocytosed, delivered to lysosomes and degraded [29], but a small percentage (~5%)
175 returns to the surface after internalization [29]. We showed that MR1 pre-loaded with the ligand, 6-

176 formylpterin (6-FP) and expressed on the cell surface could replace this ligand with 5-OP-RU during
177 transit through endosomes and recycle back to the surface [29]. Karamooz et al [28] extended this
178 further and showed that cell lines incubated overnight with high concentrations of 6-FP enabled more
179 efficient presentation of subsequently added exogenous ligands, but not of ligands derived from
180 intracellular infections. The implication was that the recycling pathway could be used by ligands
181 derived from extracellular sources but not from intracellular bacteria, the latter relying on the ER-
182 loading pathway. Further indications of the ability of MR1 to capture endosomal ligands through a
183 recycling pathway comes from a recent study describing a modified form of the ligand, 5-A-RU that
184 requires processing in endosomes [54]. This 5-A-RU *prodrug* could not be presented by a mutant
185 MR1 that could not recycle, indicating this was its preferred presentation route.

186 The second evidence that MR1 employs more than one trafficking pathway comes from the
187 knockdown of several proteins in the secretory pathway. The silencing of VAMP4 and Rab6 inhibited
188 MR1 presentation of intracellular Ag without affecting extracellular 6-FP presentation [27], while
189 Syntaxin 4 silencing reduced the presentation of intracellular but not extracellular Ag [28]. One
190 interpretation of these experiments is that either MR1 employs different pathways in the same cell to
191 survey various compartments, hence the interruption of specific endosomal pathways can block one
192 mode of presentation over another. However, an alternative explanation is that the metabolite Ag
193 itself is captured and trafficked by different pathways, depending on its source; extracellular or from
194 within a phagosome. Future studies are needed to conclusively decide on the answer.

195
196 Since MR1 can capture ligands in endosomal compartments via recycling, it is reasonable to assume
197 that any MR1 molecules already expressed on the plasma membrane prior to infection might be used
198 to present extracellular Ag. Indeed, a small amount of MR1 is found on the surface of cell lines or
199 primary cells in the absence of ligands [29]. To explain the origin of these molecules, we proposed
200 that empty ER-resident MR1 is maintained in a folding equilibrium between a partially misfolded,
201 predominant conformer that cannot leave the ER, and a minor one that can traffic to the cell surface
202 [53]. However, we believe this cohort of MR1 molecules is unlikely to play a prominent role in Ag
203 presentation because they represent a very small proportion compared to the ER-resident pool, and
204 of these only a small number (~5%) would be able to reach endosomes, escape transfer to lysosomes,
205 and return to the cell surface bound to ligands. It cannot be discarded that empty MR1 molecules may
206 recycle more efficiently than MR1-ligand complexes. However, it appears more likely that the
207 recycling pathway supplements the ER-binding route to enable presentation of ligands that cannot
208 reach the ER by employing the recycling pathway, but only once a sufficient number of MR1-Ag
209 complexes have been recruited from the ER to the plasma membrane (Figure 1). This would add

210 another layer of versatility to MR1 presentation pathway, enabling display of diverse ligands with
211 distinct intracellular trafficking properties.

212

213 **Future directions**

214 The MR1 presentation pathway is multifaceted; it can display metabolites from commensal and
215 pathogenic microbes and cancer, either distant to the presenting cell or from within. Understanding
216 how it manages this will require to answer numerous questions. Are there specialized mechanisms
217 for preservation and transport of extracellular MR1 ligands in blood or tissues? Their description
218 might improve vaccination protocols. What is the contribution of MR1 outside the ER to T cell
219 activation? To date this has not been addressed in a physiological setting and obtaining the answer
220 may need novel tools. Which cells present MR1-Ag complexes *in vivo*? MR1 is expressed by a
221 diverse range of cell types [53], but their relative roles in different scenarios of homeostasis or
222 infection *in vivo* remain unknown. Another complication is that the cells that capture and present for
223 commensal monitoring may be different from those presenting during acute infection. Recently Wang
224 et al. [15] used bone marrow chimeras to show that the cell type required to express MR1 for MAIT
225 cell expansion was dependent on the type of infection: *Salmonella* infection required MR1 on non-
226 bone marrow-derived cells, whereas for *Legionella longbeachae* infection MR1 expression was
227 required on bone-marrow-derived cells. This reinforces the idea that a range of professional and non-
228 professional APCs can perform MR1 presentation and their relative contribution depends on the
229 context. Understanding this is important to develop therapeutic strategies to arm MAIT cells in the
230 right setting and location without inducing side-effects. Finally, what is the identity of the cancer
231 ligands presented by MR1? The identity of these ligands would allow a novel approach for cancer
232 therapy and allow the creation of tetramers to study cancer-specific MR1-restricted T cells in healthy
233 donors or cancer patients.

234

235 **Acknowledgments**

236 J.A.V. is supported by an NHMRC Principal Research Fellowship (1154502) and Program Grant
237 (1113293). H.E.G.M. is supported by an Australian Research Council (ARC) Discovery Early Career
238 Researcher Award (DE170100575) and the AMP Tomorrow Fund. The authors declare that they have
239 no competing financial interests.

240

241 **Figure legends**

242

243 **Figure 1. The multifaceted presentation of metabolites by MR1 *in vivo*.**

244 Commensal microbes secrete MR1 ligands (5-OP-RU) at barrier tissues such as the skin (1). These
245 extracellular ligands are transported to the thymus by an unknown mechanism where they are
246 acquired by thymocytes (2) and presented for positive selection of MAIT cells. The commensal-
247 derived 5-OP-RU can also be acquired locally by an antigen presenting cell (APC; 3) for loading onto
248 MR1 molecules in the APC's endoplasmic reticulum (ER) and then presented at the cell surface.
249 During disease, such as the breach of a barrier (4), APCs may phagocytose microbes or be infected
250 by pathogens (5). The ligands in this situation may be sampled by MR1 in the ER or in endosomes
251 (5). Finally, MR1 expressed by tumor cells may present cancer metabolite ligands to T cells for cancer
252 surveillance (6).

253

254

References

- 255 1. Reinink P, Van Rhijn I: **Mammalian CD1 and MR1 genes.** *Immunogenetics* 2016, **68**:515-523.
- 256 2. Corbett AJ, Eckle SB, Birkinshaw RW, Liu L, Patel O, Mahony J, Chen Z, Reantragoon R,
257 Meehan B, Cao H, et al.: **T-cell activation by transitory neo-antigens derived from**
258 **distinct microbial pathways.** *Nature* 2014, **509**:361-365.
- 259 3. Kjer-Nielsen L, Patel O, Corbett AJ, Le Nours J, Meehan B, Liu L, Bhati M, Chen Z, Kostenko
260 L, Reantragoon R, et al.: **MR1 presents microbial vitamin B metabolites to MAIT cells.**
261 *Nature* 2012, **491**:717-723.
- 262 4. Fan X, Rudensky Alexander Y: **Hallmarks of Tissue-Resident Lymphocytes.** *Cell* 2016,
263 **164**:1198-1211.
- 264 5. Dusseaux M, Martin E, Serriari N, Peguillet I, Premel V, Louis D, Milder M, Le Bourhis L,
265 Soudais C, Treiner E, et al.: **Human MAIT cells are xenobiotic-resistant, tissue-targeted,**
266 **CD161hi IL-17-secreting T cells.** *Blood* 2011, **117**:1250-1259.
- 267 6. Le Bourhis L, Martin E, Peguillet I, Guihot A, Froux N, Core M, Levy E, Dusseaux M,
268 Meyssonier V, Premel V, et al.: **Antimicrobial activity of mucosal-associated invariant**
269 **T cells.** *Nat Immunol* 2010, **11**:701-708.
- 270 7. Gold MC, Cerri S, Smyk-Pearson S, Cansler ME, Vogt TM, Delepine J, Winata E, Swarbrick
271 GM, Chua WJ, Yu YY, et al.: **Human mucosal associated invariant T cells detect**
272 **bacterially infected cells.** *PLoS Biol* 2010, **8**:e1000407.
- 273 8. Walker LJ, Kang YH, Smith MO, Tharmalingham H, Ramamurthy N, Fleming VM, Sahgal N,
274 Leslie A, Oo Y, Geremia A, et al.: **Human MAIT and CD8⁺ cells develop from a pool of**
275 **type-17 precommitted CD8⁺ T cells.** 2012, **119**:422-433.
- 276 9. Meierovics AI, Cowley SC: **MAIT cells promote inflammatory monocyte differentiation into**
277 **dendritic cells during pulmonary intracellular infection.** *J Exp Med* 2016, **213**:2793-
278 2809.
- 279 10. Meierovics A, Yankelevich WJ, Cowley SC: **MAIT cells are critical for optimal mucosal**
280 **immune responses during in vivo pulmonary bacterial infection.** *Proc Natl Acad Sci U S*
281 *A* 2013, **110**:E3119-3128.
- 282 11. Le Bourhis L, Dusseaux M, Bohineust A, Bessoles S, Martin E, Premel V, Core M, Sleurs D,
283 Serriari NE, Treiner E, et al.: **MAIT Cells Detect and Efficiently Lyse Bacterially-**
284 **Infected Epithelial Cells.** *PLoS Pathog* 2013, **9**:e1003681.
- 285 12. Kurioka A, Ussher JE, Cosgrove C, Clough C, Fergusson JR, Smith K, Kang YH, Walker LJ,
286 Hansen TH, Willberg CB, et al.: **MAIT cells are licensed through granzyme exchange to**
287 **kill bacterially sensitized targets.** 2015, **8**:429-440.
- 288 13. Salou M, Franciszkiewicz K, Lantz O: **MAIT cells in infectious diseases.** *Curr Opin Immunol*
289 2017, **48**:7-14.
- 290 14. Wang H, D'Souza C, Lim XY, Kostenko L, Pediongco TJ, Eckle SBG, Meehan BS, Shi M,
291 Wang N, Li S, et al.: **MAIT cells protect against pulmonary Legionella longbeachae**
292 **infection.** *Nat Commun* 2018, **9**:3350.
- 293 15. Wang H, Kjer-Nielsen L, Shi M, D'Souza C, Pediongco TJ, Cao H, Kostenko L, Lim XY,
294 Eckle SBG, Meehan BS, et al.: **IL-23 costimulates antigen-specific MAIT cell activation**
295 **and enables vaccination against bacterial infection.** *Science Immunology* 2019,
296 **4**:eaaw0402.
- 297 16. Cui Y, Franciszkiewicz K, Mburu YK, Mondot S, Le Bourhis L, Premel V, Martin E, Kachaner
298 A, Duban L, Ingersoll MA, et al.: **Mucosal-associated invariant T cell-rich congenic**
299 **mouse strain allows functional evaluation.** *J Clin Invest* 2015, **125**:4171-4185.
- 300 17. Georgel P, Radosavljevic M, Macquin C, Bahram S: **The non-conventional MHC class I MR1**
301 **molecule controls infection by Klebsiella pneumoniae in mice.** *Mol Immunol* 2011,
302 **48**:769-775.
- 303 18. Chen Z, Wang H, D'Souza C, Sun S, Kostenko L, Eckle SB, Meehan BS, Jackson DC, Strugnell
304 RA, Cao H, et al.: **Mucosal-associated invariant T-cell activation and accumulation**

- 305 **after in vivo infection depends on microbial riboflavin synthesis and co-stimulatory**
306 **signals.** *Mucosal Immunol* 2017, **10**:58-68.
- 307 19. Jiang J, Wang X, An H, Yang B, Cao Z, Liu Y, Su J, Zhai F, Wang R, Zhang G, et al.: **MAIT**
308 **Cell Function is Modulated by PD-1 Signaling in Patients with Active Tuberculosis.**
309 *American Journal of Respiratory and Critical Care Medicine* 2014:140630082918004.
- 310 20. Wong EB, Gold MC, Meermeier EW, Xulu BZ, Khuzwayo S, Sullivan ZA, Mahyari E, Rogers
311 Z, Kløverpris H, Sharma PK, et al.: **TRAV1-2+ CD8+ T-cells including oligoconal**
312 **expansions of MAIT cells are enriched in the airways in human tuberculosis.**
313 *Communications Biology* 2019, **2**.
- 314 21. Booth JS, Salerno-Goncalves R, Blanchard TG, Patil SA, Kader HA, Safta AM, Morningstar
315 LM, Czinn SJ, Greenwald BD, Sztein MB: **Mucosal-Associated Invariant T Cells in the**
316 **Human Gastric Mucosa and Blood: Role in Helicobacter pylori Infection.** 2015, **6**.
- 317 22. Karamooz E, Harriff MJ, Lewinsohn DM: **MR1-dependent antigen presentation.** *Semin Cell*
318 *Dev Biol* 2018, **84**:58-64.
- 319 23. Constantinides MG, Link VM, Tamoutounour S, Wong AC, Perez-Chaparro PJ, Han S-J, Chen
320 YE, Li K, Farhat S, Weckel A, et al.: **MAIT cells are imprinted by the microbiota in**
321 **early life and promote tissue repair.** *Science* 2019, **366**:eaax6624.
- 322 24. Legoux F, Bellet D, Daviaud C, El Morr Y, Darbois A, Niort K, Procopio E, Salou M, Gilet J,
323 Ryffel B, et al.: **Microbial metabolites control the thymic development of mucosal-**
324 **associated invariant T cells.** *Science* 2019, **366**:494-499.
- 325 25. Treiner E, Duban L, Bahram S, Radosavljevic M, Wanner V, Tilloy F, Affaticati P, Gilfillan S,
326 Lantz O: **Selection of evolutionarily conserved mucosal-associated invariant T cells by**
327 **MR1.** *Nature* 2003, **422**:164-169.
- 328 26. Eckle SB, Corbett AJ, Keller A, Chen Z, Godfrey DI, Liu L, Mak JY, Fairlie DP, Rossjohn J,
329 McCluskey J: **Recognition of Vitamin B precursors and byproducts by Mucosal**
330 **Associated Invariant T cells.** *J Biol Chem* 2015, **290**:30204-30211.
- 331 27. Harriff MJ, Karamooz E, Burr A, Grant WF, Canfield ET, Sorensen ML, Moita LF, Lewinsohn
332 DM: **Endosomal MR1 Trafficking Plays a Key Role in Presentation of Mycobacterium**
333 **tuberculosis Ligands to MAIT Cells.** *PLoS Pathog* 2016, **12**:e1005524.
- 334 28. Karamooz E, Harriff MJ, Narayanan GA, Worley A, Lewinsohn DM: **MR1 recycling and**
335 **blockade of endosomal trafficking reveal distinguishable antigen presentation**
336 **pathways between Mycobacterium tuberculosis infection and exogenously delivered**
337 **antigens.** *Scientific Reports* 2019, **9**.
- 338 29. McWilliam HE, Eckle SB, Theodossis A, Liu L, Chen Z, Wubben JM, Fairlie DP, Strugnell
339 RA, Mintern JD, McCluskey J, et al.: **The intracellular pathway for the presentation of**
340 **vitamin B-related antigens by the antigen-presenting molecule MR1.** *Nat Immunol*
341 2016, **17**:531-537.
- 342 30. Smith AD, Foss ED, Zhang I, Hastie JL, Giordano NP, Gasparyan L, Vinhnguyen LP, Schubert
343 AM, Prasad D, McMichael HL, et al.: **Microbiota of MR1 deficient mice confer**
344 **resistance against Clostridium difficile infection.** *PLOS ONE* 2019, **14**:e0223025.
- 345 31. Varelias A, Bunting MD, Ormerod KL, Koyama M, Olver SD, Straube J, Kuns RD, Robb RJ,
346 Henden AS, Cooper L, et al.: **Recipient mucosal-associated invariant T cells control**
347 **GVHD within the colon.** *Journal of Clinical Investigation* 2018, **128**:1919-1936.
- 348 32. Dumas A, Corral D, Colom A, Levillain F, Peixoto A, Hudrisier D, Poquet Y, Neyrolles O: **The**
349 **Host Microbiota Contributes to Early Protection Against Lung Colonization by**
350 **Mycobacterium tuberculosis.** *Frontiers in Immunology* 2018, **9**.
- 351 33. Rouxel O, Da Silva J, Beaudoin L, Nel I, Tard C, Cagninacci L, Kiaf B, Oshima M, Diedisheim
352 M, Salou M, et al.: **Cytotoxic and regulatory roles of mucosal-associated invariant T**
353 **cells in type 1 diabetes.** *Nat Immunol* 2017, **18**:1321-1331.
- 354 34. McWilliam HE, Birkinshaw RW, Villadangos JA, McCluskey J, Rossjohn J: **MR1**
355 **presentation of vitamin B-based metabolite ligands.** *Curr Opin Immunol* 2015, **34C**:28-
356 34.

- 357 35. Keller AN, Eckle SB, Xu W, Liu L, Hughes VA, Mak JY, Meehan BS, Pediongco T,
358 Birkinshaw RW, Chen Z, et al.: **Drugs and drug-like molecules can modulate the**
359 **function of mucosal-associated invariant T cells.** *Nat Immunol* 2017, **18**:402-411.
- 360 36. Meermeier EW, Laugel BF, Sewell AK, Corbett AJ, Rossjohn J, McCluskey J, Harriff MJ,
361 Franks T, Gold MC, Lewinsohn DM: **Human TRAV1-2-negative MR1-restricted T cells**
362 **detect *S. pyogenes* and alternatives to MAIT riboflavin-based antigens.** *Nat Commun*
363 2016, **7**:12506.
- 364 37. Harriff MJ, McMurtrey C, Froyd CA, Jin H, Cansler M, Null M, Worley A, Meermeier EW,
365 Swarbrick G, Nilsen A, et al.: **MR1 displays the microbial metabolome driving selective**
366 **MR1-restricted T cell receptor usage.** *Sci Immunol* 2018, **3**.
- 367 38. Greene JM, Dash P, Roy S, McMurtrey C, Awad W, Reed JS, Hammond KB, Abdulhaqq S,
368 Wu HL, Burwitz BJ, et al.: **MR1-restricted mucosal-associated invariant T (MAIT) cells**
369 **respond to mycobacterial vaccination and infection in nonhuman primates.** *Mucosal*
370 *Immunol* 2017, **10**:802-813.
- 371 39. Crowther MD, Dolton G, Legut M, Caillaud ME, Lloyd A, Attaf M, Galloway SAE, Rius C,
372 Farrell CP, Szomolay B, et al.: **Genome-wide CRISPR–Cas9 screening reveals**
373 **ubiquitous T cell cancer targeting via the monomorphic MHC class I-related protein**
374 **MR1.** *Nature Immunology* 2020.
- 375 40. Lepore M, Kalinichenko A, Calogero S, Kumar P, Paleja B, Schmalzer M, Narang V, Zolezzi F,
376 Poidinger M, Mori L, et al.: **Functionally diverse human T cells recognize non-microbial**
377 **antigens presented by MR1.** *Elife* 2017, **6**:e24476.
- 378 41. Bird L: **MR1-restricted pan-cancer T cells.** *Nature Reviews Immunology* 2020.
- 379 42. Reantragoon R, Corbett AJ, Sakala IG, Gherardin NA, Furness JB, Chen Z, Eckle SBG, Uldrich
380 AP, Birkinshaw RW, Patel O, et al.: **Antigen-loaded MR1 tetramers define T cell**
381 **receptor heterogeneity in mucosal-associated invariant T cells.** *Journal of Experimental*
382 *Medicine* 2013, **210**:2305-2320.
- 383 43. Rock KL, Reits E, Neefjes J: **Present Yourself! By MHC Class I and MHC Class II**
384 **Molecules.** *Trends Immunol* 2016, **37**:724-737.
- 385 44. Heath WR, Belz GT, Behrens GM, Smith CM, Forehan SP, Parish IA, Davey GM, Wilson NS,
386 Carbone FR, Villadangos JA: **Cross-presentation, dendritic cell subsets, and the**
387 **generation of immunity to cellular antigens.** *Immunol Rev* 2004, **199**:9-26.
- 388 45. Barral DC, Brenner MB: **CD1 antigen presentation: How it works.** *Nature Reviews*
389 *Immunology* 2007, **7**:929-941.
- 390 46. Hashimoto K, Hirai M, Kurosawa Y: **A gene outside the human MHC related to classical**
391 **HLA class I genes.** *Science* 1995, **269**:693-695.
- 392 47. Abós B, Gómez del Moral M, Gozalbo-López B, López-Relaño J, Viana V, Martínez-Naves E:
393 **Human MR1 expression on the cell surface is acid sensitive, proteasome independent**
394 **and increases after culturing at 26°C.** *Biochem Biophys Res Commun* 2011, **411**:632-636.
- 395 48. Yamaguchi H, Hashimoto K: **Association of MR1 protein, an MHC class I-related molecule,**
396 **with β 2-microglobulin.** *Biochem Biophys Res Commun* 2002, **290**:722-729.
- 397 49. Miley MJ, Truscott SM, Yu YY, Gilfillan S, Fremont DH, Hansen TH, Lybarger L:
398 **Biochemical features of the MHC-related protein 1 consistent with an immunological**
399 **function.** *J Immunol* 2003, **170**:6090-6098.
- 400 50. Aldemir H: **Novel MHC class I-related molecule MR1 affects MHC class I expression in**
401 **293T cells.** *Biochemical and Biophysical Research Communications* 2008, **366**:328-334.
- 402 51. Eckle SB, Birkinshaw RW, Kostenko L, Corbett AJ, McWilliam HE, Reantragoon R, Chen Z,
403 Gherardin NA, Beddoe T, Liu L, et al.: **A molecular basis underpinning the T cell**
404 **receptor heterogeneity of mucosal-associated invariant T cells.** *J Exp Med* 2014,
405 **211**:1585-1600.
- 406 52. McWilliam HE, Villadangos JA: **MR1 antigen presentation to MAIT cells: new ligands,**
407 **diverse pathways?** *Curr Opin Immunol* 2018, **52**:108-113.

- 408 53. McWilliam HEG, Villadangos JA: **How MR1 presents a pathogen metabolic signature to**
409 **mucosal-associated invariant T (MAIT) cells.** *Trends Immunol* 2017, **38**:679-689.
- 410 54. Lange J, Anderson RJ, Marshall AJ, Chan STS, Bilbrough TS, Gasser O, Gonzalez-Lopez C,
411 Salio M, Cerundolo V, Hermans IF, et al.: **The Chemical Synthesis, Stability, and**
412 **Activity of MAIT Cell Prodrug Agonists That Access MR1 in Recycling Endosomes.**
413 *ACS Chemical Biology* 2020.
414

415 **Papers of interest:**

416 **Outstanding interest**

417 ** Legoux et al. *Science* 2019

418 *Discovered that 5-OP-RU at barrier tissues was presented by thymocytes to enable the selection of*
419 *MAIT cells. This is compelling evidence that MR1 can survey commensal bacteria at distant sites for*
420 *training immune system.*

421

422 ** Constatinidies et al *Science* 2019

423 *Showed that MAIT cells expand in skin by recognising soluble 5-OP-RU from commensal bacteria,*
424 *implying that MR1 captures extracellular ligands for monitoring commensal organisms.*

425

426 ** Crowther et al. *Nature Immunology* 2020.

427 *Identified a T cell clone that recognises MR1 presented by cancerous cells, not healthy cells, and is*
428 *cytolytic for a range of diverse tumors. This is solid evidence that MR1 presents tumor metabolites.*

429

430

431 **Special interest**

432 * Karamooz et al. *Scientific Reports* 2019

433 *Showed that cell-surface MR1-ligand complexes can be re-used to present extracellular ligands but*
434 *not those from infecting intracellular bacteria.*

435

436 * Wang et al. *Science Immunology* 2019

437 *Found that the type of cells that present MR1 for efficient MAIT cell activation and proliferation was*
438 *dependent on the type of infection.*

439

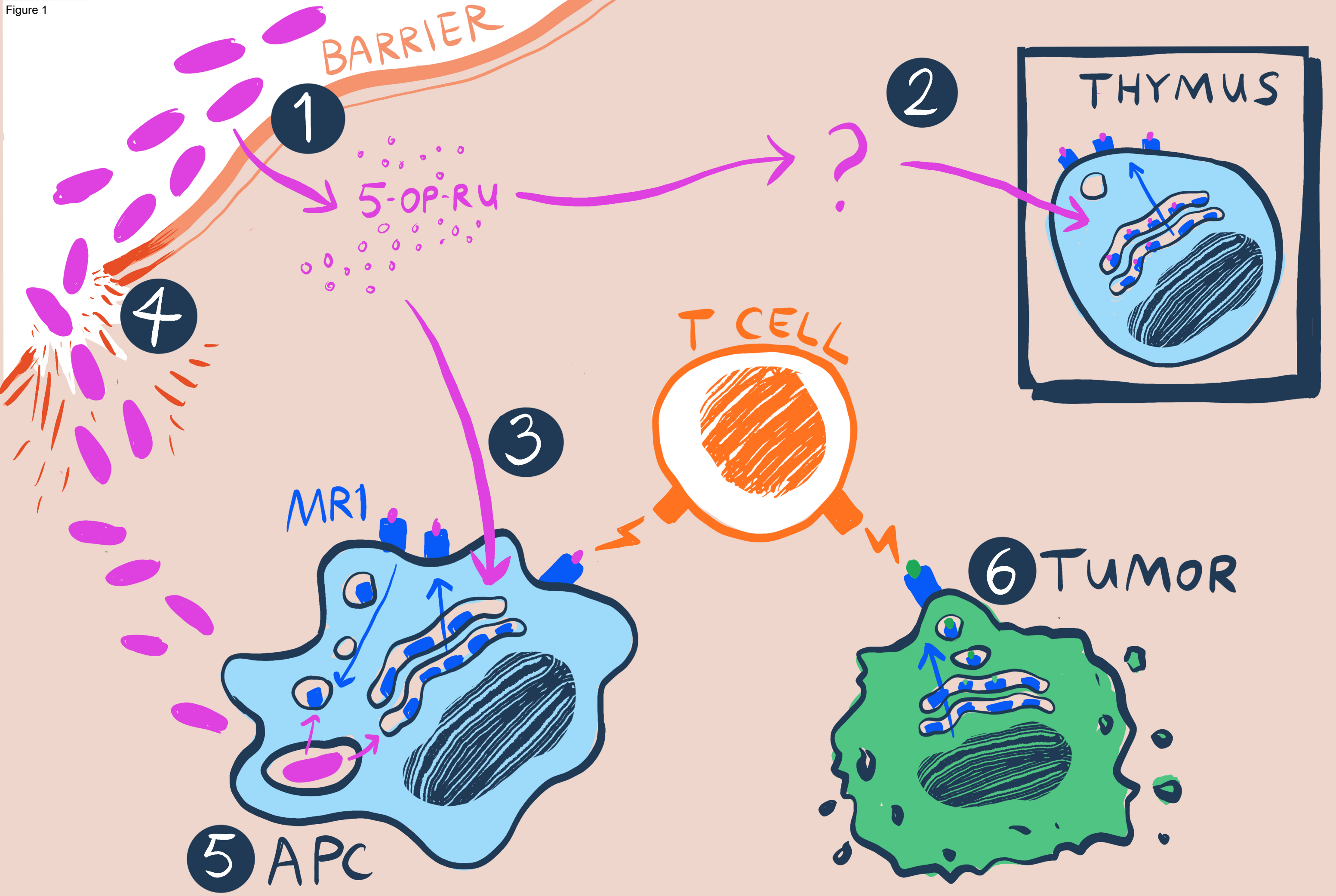
440

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Figure 1





Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

McWilliam, HEG; Villadangos, JA

Title:

MR1: a multi-faceted metabolite sensor for T cell activation

Date:

2020-06-01

Citation:

McWilliam, H. E. G. & Villadangos, J. A. (2020). MR1: a multi-faceted metabolite sensor for T cell activation. *Current Opinion in Immunology*, 64, pp.124-129.
<https://doi.org/10.1016/j.coi.2020.05.006>.

Persistent Link:

<http://hdl.handle.net/11343/267503>

File Description:

Accepted version