# Peer

# Climate, soil or both? Which variables are better predictors of the distributions of Australian shrub species?

Yasmin Hageer, Manuel Esperón-Rodríguez, John B. Baumgartner and Linda J. Beaumont

Department of Biological Sciences, Macquarie University, Sydney, New South Wales, Australia

## ABSTRACT

**Background**. Shrubs play a key role in biogeochemical cycles, prevent soil and water erosion, provide forage for livestock, and are a source of food, wood and non-wood products. However, despite their ecological and societal importance, the influence of different environmental variables on shrub distributions remains unclear. We evaluated the influence of climate and soil characteristics, and whether including soil variables improved the performance of a species distribution model (SDM), Maxent.

**Methods**. This study assessed variation in predictions of environmental suitability for 29 Australian shrub species (representing dominant members of six shrubland classes) due to the use of alternative sets of predictor variables. Models were calibrated with (1) climate variables only, (2) climate and soil variables, and (3) soil variables only.

**Results**. The predictive power of SDMs differed substantially across species, but generally models calibrated with both climate and soil data performed better than those calibrated only with climate variables. Models calibrated solely with soil variables were the least accurate. We found regional differences in potential shrub species richness across Australia due to the use of different sets of variables.

**Conclusions.** Our study provides evidence that predicted patterns of species richness may be sensitive to the choice of predictor set when multiple, plausible alternatives exist, and demonstrates the importance of considering soil properties when modeling availability of habitat for plants.

**Subjects** Biodiversity, Biogeography, Ecology, Environmental Sciences, Plant Science **Keywords** Australia, Climate, Growth form, Habitat suitability, Maxent, Predictor choice, Shrubs, Species distribution modelling, Species richness, Soil

## INTRODUCTION

Species distribution models (SDMs) are tools used to assess the spatial distribution of potentially suitable habitat for species, and to hypothesise how suitability is affected by environmental change (*Guisan & Thuiller, 2005*). These tools generally correlate species' occurrence patterns with environmental variables, which are frequently selected from a set of 19 'bioclimatic' indices (*Nix, 1986*) available in WorldClim (*Hijmans et al., 2005*).

Although climate is recognized as a major factor controlling species' distributions (*Brown & Gibson, 1983; Woodward, 1987*), climate variables are unlikely to be the only relevant predictors of habitat availability (*Chatfield et al., 2010; Austin & Van Niel, 2011*), as plant survival and reproduction also depends on light, nutrients, water, and CO<sub>2</sub>,

Submitted 3 February 2017 Accepted 19 May 2017 Published 22 June 2017

Corresponding author Yasmin Hageer, yasmin.hageer@gmail.com

Academic editor Budiman Minasny

Additional Information and Declarations can be found on page 14

DOI 10.7717/peerj.3446

Copyright 2017 Hageer et al.

Distributed under Creative Commons CC-BY 4.0

#### OPEN ACCESS

as well as disturbances and biotic interactions (*Hibbard et al.*, 2001; *Neher et al.*, 2004; *Jackson*, 2009).

However, comparatively a few SDM studies directly incorporate non-climatic environmental variables such as soil properties (but see Fitzpatrick et al., 2008; Martinson et al., 2011; Zhou et al., 2012; Condit et al., 2013; Taylor & Kumar, 2013), even though the soil properties are known to considerably impacts the distribution of plant species (*Elmendorf* & Moore, 2008; Dubuis et al., 2013) for its importance to the plants as source of water and nutrients (Aerts & Chapin, 2000) and physically for supporting root growth (Martre et al., 2002). Furthermore, some studies incorporated other factors in SDMs such as irradiance (see Franklin, 1998; Summers et al., 2012) as a light source for the plants, topography (Franklin, 1998; Hosseini et al., 2013), and landuse (Meier et al., 2012; Stanton et al., 2012; Titeux et al., 2016) in which degradation in plant habitats and loss of plant biodiversity is strongly influenced by changes in landuse and increase of urbanization is considered (Lawler et al., 2014). This poorly integration of non-climatic factors in modelling studies may partly reflect difficulties with obtaining appropriate data sets at relevant spatial scales, particularly with regards to soil variables that are related to plant functionality. It is highly recommended by all these modelling studies that SDMs should be calibrated with physiologically-relevant environmental variables, as this should lead to SDMs with greater predictive power (Austin, 2002; Austin, 2007; Williams et al., 2012).

Whilst not a strict botanical category, shrubs are generally regarded as low height, woody perennial plants with several base-stems (Zeng, Zeng & Barlage, 2008; Meng, Ni & Harrison, 2009). As the dominant flora in arid and semi-arid regions, shrubs play a key role in enhancing soil fertility, reducing runoff, soil loss (*Pressland, 1973; Xu et al., 2008; Song et al., 2013*) and dust emissions (*Engelstaedter et al., 2003*), and sequestering carbon in grassland ecosystems (*Yashiro et al., 2010*). By providing fodder for livestock (*Lefroy et al., 1992*), shrubs can enhance economic returns for dryland farms by providing an 'out-of-season' food source (*Monjardino, Revell & Pannell, 2010*).

The distribution of shrub species is strongly influenced by environmental conditions, such as climate (*Pedley, 1979*; *Westman, 1991*; *Kienast, Wildi & Brzeziecki, 1998*). Plant species occurring in arid to semi-arid regions have evolved several traits enabling them to tolerate extended periods of low precipitation and high temperature. These include small leaves (*Smith, Monson & Anderson, 1997*), slower growth rates, and more horizontal, rather than vertical, growth (*Zeng, Zeng & Barlage, 2008*). During the hot and dry season, stomata may be partly closed, reducing transpiration and water loss, leaves may be shed (*Smith, Monson & Anderson, 1997*), and physiological activity is restricted (*Reynolds, 1999*). Following rainfall events, leaves expand and stomata fully open (*Zeng, Zeng & Barlage, 2008*), and the negative impacts of the dry season may be compensated for via enhanced physiology and growth (*Reynolds, 1999*). Shrubs also have a deeper, wider rooting system than grasses, enabling the efficient extraction of water in low moisture environments (*Burgess, 1995*).

Physical and chemical soil properties, and biotic interactions play a major role in controlling the distribution of shrub species (*Pedley*, 1979). Shrubs usually occur on shallow, coarse and infertile soils (*Groves*, 1994), and are adapted to live on sandy soils with

limited moisture. Shrubs often accumulate their organic matter beneath their canopies, thereby enriching the nutrient pool horizontally, enabling these species to grow in infertile soils (*Zinke*, 1962; *Jackson & Caldwell*, 1993; *Schlesinger et al.*, 1996; *Burke et al.*, 1998) and providing microclimatic conditions that stimulate microbial biomass and activity (*Sandoval Pérez et al.*, 2016).

In this study, we assessed the extent to which soil variables, in conjunction with climate, may increase the predictive power of habitat suitability models of Australian shrub species. We hypothesised that models calibrated with both climate variables and soil properties will have greater predictive power compared with models that incorporate only climate or soil parameters. To test this hypothesis, we selected as a case study 29 shrub species that together span the distribution of major shrubland vegetation types across the continent.

#### **METHODS**

#### **Species data**

Shrubs are recognized as plants that are woody, "multi-stemmed at the base (or within 200 mm from ground level) or single stemmed, and less than 2 m" in height (*ESCAVI, 2003*, p. 87). Species are grouped into five growth forms: acacia, mallee (*Eucalypt* species), heath (which typically belong to Epacridaceae, Myrtaceae, Fabaceae and Proteaceae), chenopods, and samphire. Combined, these five shrub growth forms occupy a substantial part of the Australian landmass, mainly in semi-arid and arid regions, which form ca. 70% of the continent.

We identified 29 shrub species for inclusion in this study (Table 1), based on their dominance and endemism. Combined, these species represent the variety of shrub growth forms present on the continent. We obtained occurrence records from the Atlas of Living Australia (ALA, see http://www.ala.org.au/). Prior to downloading records, we applied filters to exclude records that did not contain coordinates (an average of 2% of records per species), were collected before 1960, or were identified by ALA as environmental outliers given the climatic envelope of the species. This resulted in an average of 3,523 (SD = 3,214) records per species.

#### Selection of predictor variables

We considered annual, seasonal and monthly climate variables known to influence the distribution of shrubs (e.g., *Xin-Rong, 2001; Li et al., 2009; Gherardi & Sala, 2015*). Gridded data for nineteen climate variables, developed by the Wallace Initiative (http: //wallaceinitiative.org), were downloaded at a resolution of 0.05 × 0.05 arc-minutes. These data were derived from spatially interpolated monthly precipitation and temperature observations (baseline period 1976–2005) obtained from the Australian Water Availability Project (AWAP, *Raupach et al., 2009; Raupach et al., 2012;* http://www.bom.gov.au/jsp/ awap/) (for more details see *Vanderwal et al., 2011*). Multicollinearity of variables can result in over-fitting of SDMs and complicate interpretation of variables' contributions (*Elith, Kearney & Phillips, 2010; Williams et al., 2012; Zhou et al., 2012*), therefore we assessed pair-wise correlations among variables. When Pearson's correlation coefficients were greater than 0.85 we removed one of the variables. This reduced the number of climate **Table 1** Distribution changes in shrubs suitable habitats using the models of climate only variables vs. climate with-soil-variables. The projected area (km<sup>2</sup>) of suitable habitat for 29 Australian shrub species, based on models using climate-only variables ( $V_C$ ), and using climate-with-soil variables ( $V_{C+S}$ ). Also shown is the percentage of  $V_C$  habitat that is also suitable according to  $V_{C+S}$  models (Overlap), the percentage of  $V_{C+S}$  habitat that is not suitable in the corresponding  $V_C$  model (Gain), and the percentage of  $V_C$  habitat that is not suitable in the corresponding  $V_{C+S}$  model (Loss).

Family	Scientific authority	Species	<i>V<sub>C</sub></i> (km <sup>2</sup> )	$V_{C+S}$ (km <sup>2</sup> )	Overlap (%)	Gain (%)	Loss (%)
Asteraceae	DC	Ozothamnus turbinatus	170,433	135,372	73.1	7.9	26.9
Casuarinaceae	(Diels) LAS Johnson	Allocasuarina campestris	853,151	479,167	50.5	10.2	49.5
	Benth.	Atriplex angulate	2,308,110	1,051,859	42.4	6.9	57.6
	Aellen	Atriplex eardleyae	1,800,784	710,007	27.8	29.6	72.2
Chenopodiaceae	F.Muell.	Atriplex holocarpa	1,843,472	1,239,199	59.1	12.0	40.9
onenopounceae	Lindl.	Atriplex nummularia	2,895,215	1,633,454	52.0	7.8	48.0
	Heward ex Benth.	Atriplex vesicaria	1,969,767	1,762,939	82.4	7.9	17.6
	(R.Br.) Paul G. Wilson	Maireana aphylla	1,899,906	801,821	31.4	25.7	68.6
	Labill.	Epacris impressa	271,063	242,672	84.2	5.9	15.8
	F.Muell. ex Benth.	Acacia aneura	3,556,328	3,223,147	87.3	3.7	12.7
	F.Muell.	Acacia sclerosperma	1,346,499	1,535,144	89.9	21.1	10.1
	F.Muell.	Acacia tetragonophylla	3,124,692	2,161,370	66.4	4.1	33.6
	Benth.	Acacia victoriae	3,514,249	2,194,372	56.1	10.2	43.9
	Bonpl.	Eucalyptus diversifolia	231,681	180,003	74.8	3.8	25.2
	A. Cunn. ex J. Oxley	Eucalyptus dumosa	856,660	493,203	53.2	7.6	46.8
	F.Muell.	Eucalyptus gracilis	737,006	393,675	49.6	7.1	50.4
	Labill.	Eucalyptus incrassata	476,528	225,939	44.0	7.3	56.0
Myrtaceae	F.Muell. ex Miq.	Eucalyptus oleosa	847,148	544,794	59.8	7.0	40.2
	F.Muell. ex Miq.	Eucalyptus socialis	1,941,927	1,285,483	62.2	6.1	37.8
	Joy Thomps.	Leptospermum continentale	328,309	201,695	54.1	12.0	45.9
	S.Schauer	Leptospermum glaucescens	1,892,279	1,507,507	69.8	12.4	30.2
	(Gaertn.) F.Muell.	Leptospermum laevigatum	423,574	263,262	58.5	5.8	41.5
	(Sol. ex Ait.) Sm.	Leptospermum lanigerum	179,713	141,520	76.2	3.3	23.8
	J.R.Forst. & G.Forst.	Leptospermum scoparium	73,399	72,442	95.0	3.7	5.0
	Sm.	Melaleuca ericifolia	340,373	234,349	60.0	12.9	40.0
	Labill.	Melaleuca squamea	91,872	100,253	94.0	13.9	6.0
	Donn. ex Sm.	Melaleuca squarrosa	177,074	150,365	78.5	7.5	21.5
Sapindaceae	(F.Muell.) F.Muell. ex Benth.	Atalaya hemiglauca	5,463,020	3,736,940	65.6	4.0	34.4
Scrophulariaceae	F.Muell.	Eremophila freelingii	1,663,875	647,367	31.4	19.2	68.6

variables to a set of five: Mean Annual Temperature (T), Maximum Temperature of the Warmest Month (TM warm), Total Annual Precipitation (P), Precipitation of the Warmest Quarter (PQ warm), and Precipitation of the Coldest Quarter (PQ cold). See correlation matrix of the variables in Table S1.

Spatial data describing four soil variables that capture soil functionality (*Sauer*, *Cambardella & Meek*, 2006; *Fisher*, *Whittaker & Malhi*, 2011; *Meier et al.*, 2012) were obtained from the CSIRO Data Access Portal at a resolution of 3 × 3 arc-seconds (~90 × 90 m) (http://www.clw.csiro.au/aclep/soilandlandscapegrid/index.html). This data include: percent clay content (CLAY; *Viscarra Rossel et al.*, 2014*a*); bulk density (BD; *Viscarra Rossel* 

Abbreviation	Environment variable	$V_C$	$V_{C+S}$	$V_S$
Т	Annual mean temperature	•	•	
TM warm	Maximum temperature of warmest month	•	•	
Р	Mean annual precipitation	•	•	
PQ warm	Precipitation of warmest quarter	•	•	
PQ cold	Precipitation of coldest quarter	•	•	
BD	Bulk soil density (g/cm <sup>3</sup> )		•	•
CLAY	Clay content percentage		•	•
pН	pH CaCl <sub>2</sub>		•	•
OC	Organic carbon percentage		•	•

 Table 2
 The environmental predictor sets used in the different models.
 Alternative predictor sets used in models.

Notes.

 $V_C$ , climate variables only;  $V_{C+S}$ , climate and soil variables;  $V_S$ , soil variables only.

*et al.*, 2014*b*), which reflects soil porosity; pH CaCl<sub>2</sub> (pH; *Viscarra Rossel et al.*, 2014*c*), which reflects soil acidity; and organic carbon (OC; *Viscarra Rossel et al.*, 2014*d*). These variables describe the corresponding physical and chemical soil characteristics that are known to influence vascular plant growth and distribution (e.g., *Jarvis, 1974; Crawley, 1997*), and do not correlate highly to each other (Table S1). For each soil variable used in this study, the three soil depth layers (0–5 cm; 5–15 cm; 15–30 cm) were highly correlated (|r| > 0.98), thus, we used measurements from the first layer only. Using ArcGIS v10.4 (ESRI Inc., 2010), all soil data were resampled to a resolution of 0.05 × 0.05 arc-minutes using bilinear interpolation, thereby matching the resolution of the climate data. Finally, each predictor variable was projected to a 5 × 5 km equal area grid (Australian Albers, EPSG: 3577).

#### Generating Maxent models of shrub species' distributions

Three sets of models were calibrated for each species. Models referred to as  $V_{C+S}$  were calibrated with climate and soil variables,  $V_C$  models were calibrated with climate variables only, and  $V_S$  models used only soil variables (Table 2).

We used Maxent (version 3.3.3k; *Phillips, Dudík & Schapire, 2004; Phillips, Anderson & Schapire, 2006*) to develop SDMs for all 29 species. Maxent is a presence-only modelling approach that produces a continuous probability field that can be interpreted as a relative index of environmental suitability. Higher values represent greater suitability of a region for the target species (*Phillips, Dudík & Schapire, 2004; Phillips, Anderson & Schapire, 2006*). A more detailed description of Maxent can be found in *Elith et al. (2011)*.

Models were initially fit using default settings. We then explored how different mathematical transformations of predictor variables ("features") influenced model predictions, and concluded that superior models were obtained when linear, quadratic, and product features were used. For each species, we generated a unique set of background points by identifying the subset of biogeographic subregion polygons (*IBRA*, 2015 version 7.0) that contained occurrence points for the species, and randomly selecting up to 10,000 occurrence records from all plant records that fell within that subset of polygons (i.e., a spatially-constrained target-group background sample). This approach aims to balance environmental sampling biases between the modelled species and the background records required by Maxent for model calibration (*Warren, Glor & Turelli, 2008; Phillips et al., 2009; Merow, Smith & Silander, 2013*).

To reduce bias caused by randomly selecting occurrence records for model training, we generated five cross-validated replicate models for each species, using a different subset of 20% of occurrence records to test each model. For each species, the five replicate models were projected to the model-fitting predictor grids, and these five projections were then averaged to produce the final projection.

#### Model performance

Currently, there is no ideal approach for evaluating model performance, although the area under the receiver-operating characteristic curve (AUC) is the most common measure of the performance of Maxent (Merow, Smith & Silander, 2013). A high AUC score indicates that the model can distinguish between presence and background points, with model performance generally considered good when AUC >0.75 (Pearce & Ferrier, 2000; Elith et al., 2006). Here, we used AUC as an indicator of model performance when models were calibrated with different sets of environmental factors. For each species we calculated the average test AUC over cross-validation replicates for each candidate model ( $V_C$ ,  $V_S$ ,  $V_{C+S}$ ). We then identified which of the models led to the highest AUC. To test whether increasing number of variables in the  $V_{C+S}$  would affect the value of AUC compared to the other models  $(V_C, V_S)$ , we computed the number of parameters used in each fitted model using lambdas file corresponds to the number of features in the model and correlated the number of parameters with the AUC score for each species in the model. In addition, for each model we calculated the maximum value of the True Skill Statistic (TSS) on test data. Unlike AUC, which is threshold-independent, TSS reflects a model's sensitivity and specificity at a particular threshold. As such, it was proposed as an appropriate measure of model performance when the modelling application requires dichotomous maps of habitat suitability (Allouche, Tsoar & Kadmon, 2006). TSS values range from -1 to 1, where 1 implies perfect sensitivity and specificity, and values of zero or less indicate considerable commission and/or omission errors.

#### **Statistical analyses**

Maxent suitability scores were converted to binary suitability (suitable/unsuitable) using the maximum training sensitivity plus specificity threshold (as recommended by *Liu*, *White* & *Newell*, 2013), which is numerically equivalent to the threshold corresponding to the maximum TSS. We then calculated pair-wise differences in the total area of suitable habitat predicted by the three sets of Maxent models using the ArcGIS extension SDMtoolbox (*Brown*, 2014). We also calculated potential shrub species 'richness', i.e., the number of species for which a grid cell was classified as suitable. These are not maps of species richness per se; rather, they are estimates of how many of the 29 species a given grid cell may be suitable for.





## RESULTS

#### Model performance

Of the 29 species,  $V_{C+S}$  models had the highest mean AUC and TSS for 27 and 25 species, respectively (Fig. 1). Across all species, AUC was, on average, 0.848 (SD = 0.0139), 0.823 (0.0137), and 0.773 (0.0187) for  $V_{C+S}$ ,  $V_C$  and  $V_S$  models, respectively, while the equivalent



**Figure 2** Habitat prediction maps of some species used in different models. Maxent predictions of habitat suitability for five Australian shrub species: *Acacia victoriae, Atriplex eardleyae, Eucalyptus gracilis, Leptospermum continentale,* and *Maireana aphylla.* Occurrence records for each species are shown in maps in the first column. Habitat suitability was modelled with different sets of environmental predictors: Climate-only ( $V_C$ ) (column A), Climate-with-soil ( $V_{C+S}$ ) (column B), and Soil-only ( $V_S$ ) (column C). Warmer colours (red) show areas predicted to have higher suitability. Bright blue represents unsuitable areas.

values for TSS were 0.583 (0.100), 0.546 (0.111), and 0.458 (0.010). Among all species, AUC showed a weak relationship with number of environmental variables used in each model, correlation coefficients |r| < 0.26 for each of  $V_{C+S}$ ,  $V_C$  and <0.20 for  $V_S$ . Visual inspection of maps generated by Maxent indicated that  $V_{C+S}$  and  $V_C$  models resulted in more realistic projections of habitat suitability than those calibrated with only soil variables (Fig. 2). However,  $V_C$  models over-predicted the realized distribution for some



**Figure 3** Species richness based in different used models. Maps of potential richness of 29 Australian shrub species, based on Maxent models calibrated with (A) climate variables only ( $V_C$ ), (B) climate and soil variables ( $V_{C+S}$ ), and (C) soil variables only ( $V_S$ ). Warmer colours (red) show areas with higher potential richness.

species, whereas  $V_{C+S}$  models provided a closer approximation (e.g., *Acacia victoriae* and *Eucalyptus* spp., Fig. 2).

The area of suitable habitat projected by  $V_{C+S}$  models ranged from 72,442 km<sup>2</sup> (*Leptospermum scoparium*) to 3,736,940 km<sup>2</sup> (*Atalaya hemiglauca*).  $V_C$  models projected suitable habitat ranging in area from 73,399 km<sup>2</sup> (*L. scoparium*) to 5,463,020 km<sup>2</sup> (*A. hemiglauca*) (Table 1). Maps from  $V_{C+S}$  and  $V_C$  models were similar for most species. Exceptions were *A. victoriae*, *Atriplex eardleyae*, *Eucalyptus gracilis*, *Leptospermum continentale*, and *Maireana aphylla* (Fig. 2). In contrast, projections from  $V_S$  models tended to cover a smaller spatial extent and had greater fragmentation of suitable habitat.

At a continental scale, the three alternate sets of variables used for model calibration resulted in different patterns of potential richness for the 29 species (Fig. 3). Highest potential richness was associated with  $V_S$  models, which predicted a total area of ~135,000 km<sup>2</sup> to be suitable for at least 11 (maximum = 18) of the 29 species. However, this map also showed substantial spatial discontinuities (i.e., contiguous areas of high potential richness were smaller than when using other predictor sets). In contrast,  $V_{C+S}$  models projected only ~50,000 km<sup>2</sup> to be suitable for at least 11 (maximum = 15) species. Although broadly similar patterns were projected by both  $V_C$  and  $V_{C+S}$  models, the potential richness of shrub species was higher based on  $V_C$  models, particularly in central South Australia (Fig. 3).

# Contribution of climate and soil variables to models of shrub distributions

In total, climate variables contributed more to  $V_{C+S}$  models than did soil variables for 20 of the 29 species (Table 3). The total contribution of climate variables in  $V_{C+S}$  models exceeded 80% for nine species (maximum = 98%, *Leptospermum lanigerum*) and was <20% for one species (18.7%, *E. gracilis*). In contrast, the total contribution of the soil variables in  $V_{C+S}$  models exceeded 80% for one species (*E. gracilis*) and was <20% for nine species. Of the individual climate and soil variables, TMwarm and pH were the most influential for ten and eight species, respectively; however, OC contributed the least for 11 species including *Atriplex* and *Eucalyptus* species (Table 3).

For  $V_C$  models, TMwarm and T were the most important variables for 14 and 10 species respectively, whereas P contributed the most for five species (Table 3). For models calibrated with soil variables only ( $V_S$ ), pH and BD were the most important variables for

**Table 3 Percent contribution of environmental variables used in the different models.** Percent contribution of environmental variables used in the climate-only predictor set  $(V_C)$ , in the climate-with-soil set  $(V_{C+S})$ , and in the soil-only predictor set  $(V_S)$  to model 29 Australian shrub species. Mean Annual Temperature (T), Maximum Temperature of the Warmest Month (TM warm), Mean Annual Precipitation (P), Precipitation of the Warmest Quarter (PQ warm), Precipitation of the Coldest Quarter (PQ cold), bulk density (BD), clay content percentage (CLAY), pH CaCl<sub>2</sub> (pH), and organic carbon (OC). For each species and predictor set, the highest value is shown in bold. For family names of species, see Table 2.

Species			$V_C$							$V_{C+S}$						V	's	
	Р	Т	TM warm	PQ cold	PQ warm	Р	Т	TM warm	PQ cold	PQ warm	BD	CLAY	pН	OC	BD	CLAY	pН	ос
Ozothamnus turbinatus	< 0.01	35.3	42.9	14.9	6.9	< 0.01	28.3	46.8	6.7	2.5	1.4	4.4	8.6	1.3	62.8	12.2	19.8	5.2
Allocasuarina campestris	6.7	11.0	25.3	28.1	28.9	6.4	11.9	19.0	17.2	28.7	3.4	2.7	2.4	8.2	8.6	16.1	0.9	74.3
Atriplex angulata	45.6	45.9	5.1	0.9	2.6	37.2	33.5	6.4	0.5	1.5	7.3	6.8	6.5	0.5	24.0	20.4	37.6	18.1
Atriplex eardleyae	24.3	25.1	23.5	19.8	7.3	5.9	7.5	6.5	2.0	2.3	5.0	7.2	62.9	0.7	5.9	11.2	80.5	2.4
Atriplex holocarpa	47.0	46.8	2.4	3.3	0.4	34.8	32.3	9.0	1.7	1.7	3.6	2.3	12.6	2.1	27.9	2.6	42.3	27.2
Atriplex nummularia	41.1	7.6	41.1	6.8	3.4	28.8	4.8	1.3	2.5	6.8	33.9	14.2	7.2	0.4	15.8	10.0	57.7	16.5
Atriplex vesicaria	36.9	1.6	49.8	3.7	7.9	29.5	2.1	32.2	1.7	9.2	9.0	2.0	14.0	0.2	8.6	7.7	77.4	6.4
Maireana aphylla	37.7	52.0	5.9	1.2	3.1	22.5	23.8	1.8	2.4	3.3	15.5	29.0	1.6	0.2	23.1	33.4	35.6	7.9
Epacris impressa	1.5	2.5	85.8	2.9	7.3	0.9	3.2	83.0	3.2	7.4	0.5	0.4	0.4	1.0	58.9	4.2	3.0	33.9
Acacia aneura	30.9	15.2	29.6	21.5	2.8	27.8	10.8	4.2	7.2	1.0	26.0	4.8	10.2	8.1	18.8	18.9	26.0	36.4
Acacia sclerosperma	27.7	39.3	12.1	8.2	12.7	18.4	40.0	10.1	10.3	2.6	1.5	1.5	15.4	0.3	18.6	21.1	36.6	23.7
Acacia tetragonophylla	70.9	6.1	2.7	3.0	17.4	26.6	6.9	4.0	0.7	10.1	13.6	4.7	22.6	10.7	34.7	30.9	10.8	23.6
Acacia victoriae	33.4	15.8	30.4	19.4	1.0	9.5	7.6	7.2	3.6	2.7	12.4	4.0	44.7	8.3	17.2	7.3	59.7	15.7
Eucalyptus diversifolia	0.3	25.8	24.3	23.7	25.9	0.3	2.9	17.8	23.9	19.4	7.7	0.8	25.7	1.4	9.0	14.3	50.9	25.8
Eucalyptus dumosa	8.2	35.4	3.4	20.2	32.9	2.2	16.1	3.5	7.9	10.2	2.6	18.0	38.6	0.9	4.4	30.7	63.9	1.0
Eucalyptus gracilis	15.5	20.1	37.9	7.9	18.6	1.9	4.5	4.2	4.4	3.7	0.2	8.3	72.3	0.5	0.3	12.6	86.7	0.5
Eucalyptus incrassata	3.5	25.4	33.0	15.4	22.8	0.2	18.0	10.1	2.9	4.6	10.5	17.3	35.7	0.8	16.5	25.2	56.0	2.2
Eucalyptus oleosa	14.2	15.3	43.4	4.2	23.0	1.7	1.7	11.3	2.8	6.3	2.5	9.2	64.3	0.2	0.1	19.1	80.5	0.3
Eucalyptus socialis	8.4	34.4	18.8	11.5	27.0	1.7	10.8	4.1	4.6	3.7	2.8	13.8	<b>58</b> .0	0.4	2.9	19.6	76.6	0.9
Leptospermum continentale	4.1	10.4	62.5	19.0	3.9	5.2	3.6	46.2	16.6	1.8	11.6	3.1	10.0	2.0	1.7	39.2	25.5	33.7
Leptospermum glaucescens	2.9	42.4	13.4	38.7	2.6	7.3	39.6	4.4	19.7	2.1	7.2	4.4	11.1	4.2	27.7	27.2	37.4	7.6
Leptospermum laevigatum	9.8	31.3	28.0	26.8	4.1	1.4	26.1	23.1	20.4	1.7	0.9	12.1	8.9	5.5	14.3	19.8	38.4	27.5
Leptospermum lanigerum	0.7	1.4	92.2	2.7	3.0	2.2	1.4	90.7	1.2	2.5	0.5	0.3	0.1	1.1	69.7	10.0	2.2	18.2
Leptospermum scoparium	0.7	12.6	83.6	1.1	2.0	0.5	12.7	80.1	1.5	2.6	0.1	1.8	0.6	0.2	35.9	30.6	17.9	15.6
Melaleuca ericifolia	5.5	24.1	58.0	1.1	11.3	1.1	20.4	51.0	1.8	7.8	2.8	4.7	5.8	4.4	32.1	3.0	46.1	18.8
Melaleuca squamea	3.9	5.4	83.2	6.6	0.9	0.9	8.0	73.3	7.5	2.2	2.5	1.4	1.9	2.1	30.7	20.5	7.6	41.2
Melaleuca squarrosa	3.1	17.6	65.9	12.3	1.0	2.7	20.9	60.7	7.5	2.2	1.7	0.3	3.3	0.8	43.1	15.3	29.1	12.4
Atalaya hemiglauca	18.4	10.2	66.5	4.8	0.1	12.1	16.3	34.6	3.7	0.3	3.9	5.0	22.0	2.1	42.8	14.9	2.4	39.8
Eremophila freelingii	9.0	59.6	4.9	13.0	13.5	3.0	38.1	5.0	2.4	6.2	20.3	3.7	3.4	18.0	41.2	20.2	20.5	18.1

17 and eight species (maximum 86.7% *E. gracilis* and 69.7% *L. lanigerum*), respectively, while OC contributed > 74% to the model for *Allocasuarina campestris* and BD contributed 39.2% to the model for *L. continentale*.

### DISCUSSION

Species distribution models are frequently calibrated only with climate variables, but for plant species, does the addition of soil properties as predictors improve model performance? For 29 Australian shrub species, we found that: (a) on average, models calibrated with both climate and soil variables  $(V_{C+S})$  performed better than those calibrated solely with climate variables  $(V_C)$  (Fig. 1); (b) maximum temperature of the warmest month and pH were the most important contributors to  $V_{C+S}$  models for ten and eight species, respectively (Table 3); and (c) models calibrated with only soil variables  $(V_S)$  had lower AUC and TSS scores, indicating lower classification accuracy than  $V_C$  models (Fig. 1), and frequently generated unrealistic predictions (Figs. 2 and 3). For some species the inclusion of soil properties along with climate variables resulted in projections of current habitat that more closely approximated the realized distribution, compared to models calibrated with climate variables only. As a consequence, although broadly similar patterns of potential species richness occurred at the regional level, at finer spatial scales these patterns diverged substantially, particularly in central South Australia (Fig. 3). To date, few studies have explicitly assessed whether the inclusion of soil variables increases the predictive power of SDMs, although a number of studies have included these as variables in model calibration (e.g., Condit et al., 2013; Fitzpatrick et al., 2008; Martinson et al., 2011; Taylor & Kumar, 2013; Zhou et al., 2012). By themselves, the soil variables included in this study did not result in biologically realistic maps of the realised distribution of the 29 shrub species. The distributions of suitable habitat predicted by these models were frequently fragmented or had abrupt boundaries inconsistent with the known distributions of populations (Fig. 3). Topography and soil type, for instance, are important in determining the suitability of habitat for fire-prone chaparral species in California (Franklin, McCullough & Gray, 2000; Meentemeyer, Moody & Franklin, 2001). In Australia, Bui et al. (2014) found that although climate has higher impact on controlling the distribution of Acacia species at a continental scale, physical and chemical properties of soil were more useful in explaining the distribution of shrub species in southern Australia.

At the scale of this study, models indicate that climate plays a greater role than soil characteristics in defining the distribution of most of the 29 shrub species, although soil pH was the key determinant for *Eucalyptus* species. *Bui et al.* (2017) found that incorporating soil variables with climate was efficient for defining the distribution of *Eucalyptus* species and strongly influenced some specific species in taxonomic sections (e.g., Aromatica and Dumaria), although that climate was more important factor. These results are similar to *Martinson et al.* (2011), who used Maxent to model the distributions of 30 species, including shrubs, across arid areas of North America using climate and soil variables. Temperature variables, mainly T, contributed the most to their models, and none of the three soil variables (pH, clay concentration, and total organic carbon) was the most important for

any species. Variables, such as cation exchange capacity and texture, also contributed little to models predicting the distributions of European trees (*Meier et al., 2012*).

However, soil variables made a substantial contribution to  $V_{C+S}$  models for some species. For instance, among *Eucalyptus* species, pH was the highest contributing variable in  $V_{C+S}$  models. *Eucalypts* are adapted to live in acidic soils ranging from pH 3.5 to 6 (*Evans, 1992*), through a symbiotic relationship with ectomycorrhizal fungi (*Malajczuk, McComb & Loneragan, 1975*). Conversely, growth of *Acacia* shrubs is restricted in high pH soils due to reductions in nutrient availability (*Nano & Clarke, 2008*). Again, pH contributed highly to  $V_{C+S}$  models for several *Acacia* species. Soil bulk density contributed the most to the  $V_{C+S}$  model for the saltbush *Atriplex nummularia*, which is known to favour heavy clay soils (*Cunningham & Cunningham, 2011*). Clay soils are found to have lower cation exchange from the organic matter than sandy soil (*McDonald et al., 2017*). This may explain the low contribution of soil organic carbon in the models for *Atriplex* species. These findings highlight the importance of knowledge about specific characteristic and biological idiosyncrasies that species possess to include them as variables predictors.

*González-Orozco et al. (2013)* found annual precipitation and percentage of sand in the topsoil to be key environmental factors influencing the distribution of Australian *Acacia* species. Our results support this, with precipitation contributing the most to  $V_{C+S}$  model for *A. aneura* and *A. tetragonophylla*, while bulk density was also important.

#### CAVEATS

The accuracy of SDMs is influenced by a number of factors, including accuracy and availability of environmental data used to calibrate models (*Buisson et al., 2010*), biases in occurrence records (*Liu, White & Newell, 2009*), and the selection of model parameters (*Beaumont, Hughes & Pitman, 2008*).

Environmental data frequently require manipulation before use in SDMs, and this often involves resampling data to different resolutions. Aggregation or interpolation to a coarser or finer resolution, respectively, can alter the accuracy of data. In order to match the spatial resolution of the climate data, the soil dataset used in this study was aggregated from 90 m to 5 km. Inconsistencies may have been magnified when the soil data were aggregated to a coarser resolution, and are apparent in some of the maps of suitable habitat. Interpolation and accuracy issues may also arise with climate data. For instance, although new high-resolution climate data (1 km) have recently become available (i.e., eMAST data products; http://www.emast.org.au/), precipitation-related variables may suffer accuracy problems when interpolating to areas with complex topography (*Hutchinson, 1995*). We point out that SDMs calibrated with climate and soil datasets that more closely align in spatial resolution may have different findings to our study. This remains an area requiring further exploration.

It is also likely that patterns in climate and soil do not influence species' distributions at the same spatial scale. For example, different mallee species (*Eucalyptus*) in Western Australia broadly occupy the same hot, dry climatic conditions. Within these climate zones, soil varies at a finer scale. As such, *E. diversifolia* is restricted to the limestone coastal dunes

and cliffs, while *E. incrassata* occurs on sand plains such as in South Australia (*Specht*, 1966). Therefore, trade-offs will occur when selecting the most appropriate spatial scale and environmental variables for modelling studies (*Guisan & Zimmermann*, 2000).

An additional hindrance might be the availability of environmental variables that used as predictors in the models. For example, soil temperature and available water capacity of soil that can be absorbed by roots, are suggested to be an important factors that potentially influence plant growth (*Dunne, 1996*; *Reddell, Bowen & Robson, 1985*) and could be incorporated in the SDMs. However, lack of data of these variables at continental scale can be a challenge for using such variables in modelling studies. Furthermore, for some studies that predictions of a variable under alternative scenarios, such as climate change, may be required. Yet these can be difficult to obtain. For instance, while climate change projections for the standard 19 bioclimatic variables included in WorldClim (*Hijmans et al., 2005*) and similar products are readily available, they may not be available for other climate variables.

Accuracy of occurrence records and sampling biases associated with them may affect SDM performance (*Hefley et al., 2013*). Sampling across arid and semi-arid zones of Australia has typically been poor and much clumped in space and time (*Haque et al., in press*). Thus, to reduce the likelihood of errors we applied filters to ALA records to exclude outliers. We decrease sampling bias by removing duplicate records in grid cells and adopted a target-group background approach. Additionally, although we selected dominant, easily-identified species for this study, it is not possible to determine whether their entire realized distribution (and hence, climate envelope) has been sampled.

Regarding species richness, this is a convenient way to describe and compare the biodiversity of different areas; however, there are concerns about over-estimating species richness using combined or so-called stacked SDMs (*Guisan & Rahbek, 2011*; *Hortal et al., 2012*). It is suggested that bias may be corrected by linking stacked SDMs to macroecological models; nevertheless, early comparisons indicate that this approach has not yielded much improvement in reducing overestimates of richness (*Calabrese et al., 2014*). The issue of how best to estimate richness from stacked SDMs will undoubtedly be a key area of research over the next few years.

#### CONCLUSIONS

We demonstrate that for most of the shrub species modelled in this study, the inclusion of soil properties, along with climate, resulted in more realistic predictions of the distribution of current habitat. We also demonstrate how maps of suitable habitat can differ substantially depending on whether models are calibrated with only climate variables or with climate and soil variables (e.g., *A. eardleyae* and *E. gracilis*), even when AUC and TSS scores are very similar. This emphasises the importance of interactive validation of model predictions, rather than relying solely on popular metrics for assessing their accuracy. Furthermore, a promising recent application of SDMs is to connect them with stochastic population models to estimate extinction risk (*Keith et al., 2008; Stanton et al., 2012*). Such estimates are dependent on reasonable predictions of suitable habitat as a function of climate conditions

and other parameters such as soil type. This is an important consideration when applying SDM methods to scenarios of temporal environmental change (e.g., climate change), whereby projections may continue to diverge. Our analysis validates the approach of incorporating soil variables into SDMs, and we recommend that future studies explore the contribution of soil variables when modelling the distributions of plant species.

#### Nomenclature: Australian Plant Census

AUC	Area under the receiver operating characteristic curve			
BD	bulk density			
CLAY	percent clay content			
Р	total annual precipitation (mm)			
OC	organic carbon			
PQ cold	total precipitation of the coldest quarter (mm)			
PQ warm	total precipitation of the warmest quarter (mm)			
SD	standard deviation			
SDM	species distribution model			
Т	mean annual temperature (°C)			
TSS	True Skill Statistic			
TM warm	maximum temperature of the warmest month (°C)			
V <sub>C</sub>	climate-only variable set			
$V_{C+S}$	climate-plus-soil variable set			
$V_S$	soil-only variable set			

## ACKNOWLEDGEMENTS

We thank Jeremy VanDerWal for assistance with climate data.

## **ADDITIONAL INFORMATION AND DECLARATIONS**

#### Funding

The authors received no funding for this work.

#### **Competing Interests**

The authors declare there are no competing interests.

#### **Author Contributions**

- Yasmin Hageer conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Manuel Esperón-Rodríguez contributed reagents/materials/analysis tools, prepared figures and/or tables, reviewed drafts of the paper.
- John B. Baumgartner analyzed the data, prepared figures and/or tables, reviewed drafts of the paper.
- Linda J. Beaumont conceived and designed the experiments, contributed reagents/materials/analysis tools, reviewed drafts of the paper, interpretaion of results.

#### **Data Availability**

The following information was supplied regarding data availability: The raw data has been supplied as Data S1.

#### **Supplemental Information**

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/ peerj.3446#supplemental-information.

## REFERENCES

- Aerts R, Chapin FS. 2000. The mineral nutrition of wild plants revisited: a reevaluation of processes and patterns. *Advances in Ecological Research* **30**:1–67 DOI 10.1016/S0065-2504(08)60016-1.
- Allouche O, Tsoar A, Kadmon R. 2006. Assessing the accuracy of species distribution models: prevalence, kappa and the true skill statistic (TSS). *Journal of Applied Ecology* 43:1223–1232 DOI 10.1111/j.1365-2664.2006.01214.x.
- Austin M. 2002. Spatial prediction of species distribution: an interface between ecological theory and statistical modelling. *Ecological Modelling* 157:101–118 DOI 10.1016/S0304-3800(02)00205-3.
- Austin M. 2007. Species distribution models and ecological theory: a critical assessment and some possible new approaches. *Ecological Modelling* 200:1–19 DOI 10.1016/j.ecolmodel.2006.07.005.
- Austin MP, Van Niel KP. 2011. Impact of landscape predictors on climate change modelling of species distributions: a case study with Eucalyptus fastigata in southern New South Wales, Australia. *Journal of Biogeography* 38:9–19 DOI 10.1111/j.1365-2699.2010.02415.x.
- Beaumont LJ, Hughes L, Pitman A. 2008. Why is the choice of future climate scenarios for species distribution modelling important? *Ecology Letters* 11:1135–1146 DOI 10.1111/j.1461-0248.2008.01231.x.
- **Brown JL. 2014.** SDMtoolbox: a python-based GIS toolkit for landscape genetic, biogeographic and species distribution model analyses. *Methods in Ecology and Evolution* **5**:694–700 DOI 10.1111/2041-210X.12200.
- Brown JH, Gibson AC. 1983. Biogeography: Mosby. Missouri: St. Louis.
- Bui EN, Carlos E, González-Orozco CE, Miller JT. 2014. Acacia, climate, and geochemistry in Australia. *Plant and Soil* 381:161–175 DOI 10.1007/s11104-014-2113-x.
- Bui EN, Thornhill AH, González-Orozco CE, Knerr N, Miller JT. 2017. Climate and geochemistry as drivers of eucalypt diversification in Australia. *Geobiology* 15:427–440 DOI 10.1111/gbi.12235.
- Buisson L, Thuiller W, Casajus N, Lek S, Grenouillet G. 2010. Uncertainty in ensemble forecasting of species distribution. *Global Change Biology* 16:1145–1157 DOI 10.1111/j.1365-2486.2009.02000.x.
- **Burgess TL. 1995.** Desert Grassland, mixed shrub savanna, shrub steppe, or semidesert scrub? The dilemma of coexisting growth forms. In: McClaran MP, VanDevender TR, eds. *The Desert Grassland*. Tuscon: University of Arizona Press, 31–67.

- Burke IC, Lauenroth WK, Vinton MA, Hook PB, Kelly RH, Epstein HE, Aguiar MR, Robles MD, Aguilera MO, Murphy KL, Richard AG. 1998. Plant-soil interactions in temperate grasslands. *Biogeochemistry* **42**:121–143 DOI 10.1023/A:1005987807596.
- **Calabrese JM, Certain G, Kraan C, Dormann CF. 2014.** Stacking species distribution models and adjusting bias by linking them to macroecological models. *Global Ecology and Biogeography* **23**:99–112 DOI 10.1111/geb.12102.
- **Chatfield BS, Van Niel KP, Kendrick GA, Harvey ES. 2010.** Combining environmental gradients to explain and predict the structure of demersal fish distributions. *Journal of Biogeography* **37**:593–605 DOI 10.1111/j.1365-2699.2009.02246.x.
- **Condit R, Engelbrecht BM, Pino D, Pérez R, Turner BL. 2013.** Species distributions in response to individual soil nutrients and seasonal drought across a community of tropical trees. *Proceedings of the National Academy of Sciences of the United States of America* **110**:5064–5068 DOI 10.1073/pnas.1218042110.
- Crawley MJ. 1997. Plant ecology. 70. Cambridge: Blackwell Science, 357.
- **Cunningham GM (Geoffrey McIver), Cunningham GM. 2011.** *Plants of western New South Wales.* Collingwood: CSIRO Publishing.
- **Dubuis A, Giovanettina S, Pellissier L, Pottier J, Vittoz P, Guisan A. 2013.** Improving the prediction of plant species distribution and community composition by adding edaphic to topo-climatic variables. *Journal of Vegetation Science* **24**:593–606 DOI 10.1111/jvs.12002.
- Dunne KA. 1996. Global distribution of plant extractable-water capacity of soil. International Journal of Climatology 16:841–859 DOI 10.1002/(SICI)1097-0088(199608)16:8<841::AID-JOC60>3.0.CO;2-8.
- Elith J, Graham CH, Anderson RP, Dudik M, Ferrier S, Guisan A, Hijmans RJ, Huettmann F, Leathwick JR, Lehmann A, Li J, Lohmann LG, Loiselle BA, Manion G, Moritz C, Nakamura M, Nakazawa Y, Overton J McC, Peterson AT, Phillips SJ, Richardson K, Scachetti-Pereira R, Schapire RE, Soberon J, Williams S, Wisz MS, Zimmermann NE. 2006. Novel methods improve prediction of species' distributions from occurrence data. *Ecography* 29:129–151 DOI 10.1111/j.2006.0906-7590.04596.x.
- Elith J, Kearney M, Phillips S. 2010. The art of modelling range-shifting species. *Methods in Ecology and Evolution* 1:330–342 DOI 10.1111/j.2041-210X.2010.00036.x.
- Elith J, Phillips SJ, Hastie T, Dudik M, Chee YE, Yates CJ. 2011. A statistical explanation for MaxEnt for ecologists. *Diversity and Distributions* 17:43–57 DOI 10.1111/j.1472-4642.2010.00725.x.
- Elmendorf SC, Moore KA. 2008. Use of community-composition data to predict the fecundity and abundance of species. *Conservation Biology* 22:1523–1532 DOI 10.1111/j.1523-1739.2008.01051.x.
- **Engelstaedter S, Kohfeld K, Tegen I, Harrison S. 2003.** Controls of dust emissions by vegetation and topographic depressions: an evaluation using dust storm frequency data. *Geophysical Research Letters* **30**:Article 1294 DOI 10.1029/2002GL016471.

- **Evans J. 1992.** *Plantation forestry in the tropics: tree planting for industrial, social, environmental, and agroforestry purposes.* Second edition. Oxford: Oxford University Press, 403.
- **Executive Steering Committee for Australian Vegetation Information (ESCAVI). 2003.** Australian vegetation attribute manual: National vegetation information system. Version 6.0. Canberra: Department of the Environment and Heritage.
- **Fisher JB, Whittaker RJ, Malhi Y. 2011.** ET come home: potential evapotranspiration in geographical ecology. *Global Ecology and Biogeography* **20**:1–18 DOI 10.1111/j.1466-8238.2010.00578.x.
- Fitzpatrick MC, Gove AD, Sanders NJ, Dunn RR. 2008. Climate change, plant migration, and range collapse in a global biodiversity hotspot: the Banksia (Proteaceae) of Western Australia. *Global Change Biology* 14:1337–1352 DOI 10.1111/j.1365-2486.2008.01559.x.
- **Franklin J. 1998.** Predicting the distribution of shrub species in southern California from climate and terrain-derived variables. *Journal of Vegetation Science* **9**:733–748 DOI 10.2307/3237291.
- **Franklin J, McCullough P, Gray C. 2000.** Terrain variables used for predictive mapping of vegetation communities in Southern California. In: Gallant JP, Wilson JC, eds. *Terrain analysis: principles and applications.* New York: Wiley, 331–353.
- **Gherardi LA, Sala OE. 2015.** Enhanced precipitation variability decreases grass-and increases shrub-productivity. *Proceedings of the National Academy of Sciences of the United States of America* **112**:12735–12740 DOI 10.1073/pnas.1506433112.
- **González-Orozco CE, Shawn FW, Knerr N, Miller J. 2013.** A biogeographical regionalization of Australian Acacia species. *Journal of Biogeography* **40**:2156–2166 DOI 10.1111/jbi.12153.
- **Groves RH. 1994.** *Australian vegetation.* 2nd edition. Cambridge: Cambridge University Press.
- Guisan A, Rahbek C. 2011. SESAM–a new framework integrating macroecological and species distribution models for predicting spatio-temporal patterns of species assemblages. *Journal of Biogeography* 38:1433–1444 DOI 10.1111/j.1365-2699.2011.02550.x.
- Guisan A, Thuiller W. 2005. Predicting species distribution: offering more than simple habitat models. *Ecology Letters* 8:993–1009 DOI 10.1111/j.1461-0248.2005.00792.x.
- Guisan A, Zimmermann NE. 2000. Predictive habitat distribution models in ecology. *Ecological Modelling* 135:147–186 DOI 10.1016/S0304-3800(00)00354-9.
- Haque M, Nipperess D, Gallagher R, Beaumont L. A Legacy of sampling: exploring spatial patterns among occurrence records in Australia's virtual herbarium. *Austral Ecology* In press.
- Hefley TJ, Baasch DM, Tyre AJ, Blankenship EE. 2013. Correction of location errors for presence-only species distribution models. *Methods in Ecology and Evolution* 5:207–214.

- Hibbard K, Archer S, Schimel D, Valentine D. 2001. Biogeochemical changes accompanying woody plant encroachment in a subtropical savanna. *Ecology* **82**:1999–2011 DOI 10.1890/0012-9658(2001)082[1999:BCAWPE]2.0.CO;2.
- Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A. 2005. Very high resolution interpolated climate surfaces for global land areas. *International Journal of Climatology* 25:1965–1978 DOI 10.1002/joc.1276.
- Hortal J, De Marco Jr P, Santos A, Diniz-Filho JAF. 2012. Integrating biogeographical processes and local community assembly. *Journal of Biogeography* **39**:627–628 DOI 10.1111/j.1365-2699.2012.02684.x.
- Hosseini S, Kappas M, Chahouki MZ, Gerold G, Erasmi S, Emam AR. 2013. Modelling potential habitats for Artemisia sieberi and Artemisia aucheri in Poshtkouh area, central Iran using the maximum entropy model and geostatistics. *Ecological Informatics* 18:61–68 DOI 10.1016/j.ecoinf.2013.05.002.
- Hutchinson MF. 1995. Interpolation of mean rainfall using thin plate smoothing splines. *International Journal of Geographical Information Systems* 9:385–403 DOI 10.1080/02693799508902045.
- **IBRA. 2015.** Interim biogeographic regionalisation for Australia. *Bioregional Assessment Source Dataset.* IBRA Subregion Australia Version 7.0-PED. Adelaide: South Australian Department of Environment, Water and Natural Resources.
- Jackson R, Caldwell M. 1993. Geostatistical patterns of soil heterogeneity around individual perennial plants. *Journal of Ecology* 81:683–692 DOI 10.2307/2261666.
- Jackson SD. 2009. Plant responses to photoperiod. *New Phytologist* 181:517–531 DOI 10.1111/j.1469-8137.2008.02681.x.
- Jarvis SC. 1974. Soil factors affecting the distribution of plant communities on the cliffs of craig breidden, montgomeryshire. *Journal of Ecology* 62:721–733 DOI 10.2307/2258952.
- Keith DA, Akçakaya HR, Thuiller W, Midgley GF, Pearson RG, Phillips SJ, Regan HM, Araújo MB, Rebelo TG. 2008. Predicting extinction risks under climate change: coupling stochastic population models with dynamic bioclimatic habitat models. *Biology Letters* 4:560–563 DOI 10.1098/rsbl.2008.0049.
- Kienast F, Wildi O, Brzeziecki B. 1998. Potential impacts of climate change on species richness in mountain forests–an ecological risk assessment. *Biological Conservation* 83:291–305 DOI 10.1016/S0006-3207(97)00085-2.
- Lawler JJ, Lewisb DJ, Nelsonc E, Plantingad AJ, Polaskye S, Withey JC, Helmersg DP, Martinuzzig S, Penningtonh D, Radeloffg VC. 2014. Projected land-use change impacts on ecosystem services in the United States. *Proceedings of the National Academy of Sciences of the United States of America* 111:7492–7497 DOI 10.1073/pnas.1405557111.
- Lefroy E, Dann P, Wildin J, Wesley-Smith R, McGowan A. 1992. Trees and shrubs as sources of fodder in Australia. *Agroforestry Systems* 20:117–139.
- Li X, Tan H, He M, Wang X, Li X. 2009. Patterns of shrub species richness and abundance in relation to environmental factors on the Alxa Plateau: prerequisites for

conserving shrub diversity in extreme arid desert regions. *Science in China Series D* **52**:669–680 DOI 10.1007/s11430-009-0054-7.

- Liu C, White M, Newell G. 2009. Assessing the accuracy of species distribution models more thoroughly. 18th World IMACS/MODSIM Congress. Cairns: Citeseer, 4234–4240.
- Liu C, White M, Newell G. 2013. Selecting thresholds for the prediction of species occurrence with presence-only data. *Journal of Biogeography* 40:778–789 DOI 10.1111/jbi.12058.
- Malajczuk N, McComb A, Loneragan J. 1975. Phosphorus uptake and growth of mycorrhizal and uninfected seedlings of Eucalyptus calophylla R. Br. *Australian Journal of Botany* 23:231–238 DOI 10.1071/BT9750231.
- Martinson EJ, Eddy ZB, Commerford JL, Blevins E, Rolfsmeier SJ, McLauchlan KK. 2011. Biogeographic distributions of selected North American grassland plant species. *Physical Geography* 32:583–602 DOI 10.2747/0272-3646.32.6.583.
- Martre P, North GB, Bobich EG, Nobel PS. 2002. Root deployment and shoot growth for two desert species in response to soil rockiness. *American Journal of Botany* 89:1933–1939 DOI 10.3732/ajb.89.12.1933.
- McDonald GK, Tavakkoli E, Cozzolino D, Banas K, Derrien M, Rengasamy P. 2017. A survey of total and dissolved organic carbon in alkaline soils of southern Australia. *Australian Journal of Soil Research* Epub ahead of print Feb 9 2017 DOI 10.1071/SR16237.
- Meentemeyer RK, Moody A, Franklin J. 2001. Landscape scale patterns of shrub-species abundance in California chaparral: the role of topographically mediated resource gradients. *Plant Ecology* **156**:19–41 DOI 10.1023/A:1011944805738.
- Meier ES, Lischke H, Schmatz DR, Zimmermann NE. 2012. Climate, competition and connectivity affect future migration and ranges of European trees. *Global Ecology and Biogeography* 21:164–178 DOI 10.1111/j.1466-8238.2011.00669.x.
- Meng T-T, Ni J, Harrison SP. 2009. Plant morphometric traits and climate gradients in northern China: a meta-analysis using quadrat and flora data. *Annals of Botany* 104:1217–1229 DOI 10.1093/aob/mcp230.
- Merow C, Smith MJ, Silander JA. 2013. A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and settings matter. *Ecography* 36:1058–1069 DOI 10.1111/j.1600-0587.2013.07872.x.
- Monjardino M, Revell D, Pannell DJ. 2010. The potential contribution of forage shrubs to economic returns and environmental management in Australian dryland agricultural systems. *Agricultural Systems* 103:187–197 DOI 10.1016/j.agsy.2009.12.007.
- Nano CEM, Clarke PJ. 2008. Variegated desert vegetation: covariation of edaphic and fire variables provides a framework for understanding mulga-spinifex coexistence. *Austral Ecology* 33:848–862 DOI 10.1111/j.1442-9993.2008.01855.x.
- Neher D, Weicht T, Moorhead D, Sinsabaugh R. 2004. Elevated CO2 alters functional attributes of nematode communities in forest soils. *Functional Ecology* 18:584–591 DOI 10.1111/j.0269-8463.2004.00866.x.
- Nix HA. 1986. A biogeographic analysis of Australian elapid snakes. *Atlas of Elapid Snakes of Australia* 7:4–15.

- Pearce J, Ferrier S. 2000. Evaluating the predictive performance of habitat models developed using logistic regression. *Ecological Modelling* 133:225–245 DOI 10.1016/S0304-3800(00)00322-7.
- Pedley L. 1979. A revision of Acacia Mill. in Queensland. Austrobaileya 1:235–337.
- Phillips S, Anderson R, Schapire R. 2006. Maximum entropy modeling of species geographic distributions. *Ecological Modelling* 190:231–259 DOI 10.1016/j.ecolmodel.2005.03.026.
- Phillips SJ, Dudík M, Elith J, Graham CH, Lehmann A, Leathwick J, Ferrier S.
   2009. Sample selection bias and presence-only distribution models: implications for background and pseudo-absence data. *Ecological Applications* 19:181–197
   DOI 10.1890/07-2153.1.
- **Phillips SJ, Dudík M, Schapire RE. 2004.** A maximum entropy approach to species distribution modeling. In: *Proceedings of the twenty-first international conference on Machine learning*. ACM, 83.
- Pressland A. 1973. Rainfall partitioning by an arid woodland (Acacia aneura F. Muell.) in south-western Queensland. *Australian Journal of Botany* 21:235–245 DOI 10.1071/BT9730235.
- Raupach M, Briggs P, Haverd V, King E, Paget M, Trudinger C. 2009. *Australian water availability project (AWAP): CSIRO marine and atmospheric research component: final report for phase 3.* 67. Melbourne: Centre for Australian weather and climate research (bureau of meteorology and CSIRO).
- Raupach M, Briggs P, Haverd V, King E, Paget M, Trudinger C. 2012. *Australian water availability project*. Canberra: CSIRO Marine and Atmospheric Research.
- Reddell P, Bowen GD, Robson AD. 1985. The effects of soil temperature on plant growth, nodulation and nitrogen fixation in casuarina cunninghamiana miq. *The New Phytologist* 101:441–450 DOI 10.1111/j.1469-8137.1985.tb02850.x.
- **Reynolds MP. 1999.** Residual leaf area as a measure of shrub use. MSc Thesis, Oregon State University.
- Sandoval Pérez AL, Camargo-Ricalde SL, Montaño NM, García-Oliva F, Alarcón A, Montaño Arias SA, Esperón-Rodríguez M. 2016. Biocrusts, inside and outside resource islands of Mimosa luisana (Leguminosae), improve soil carbon and nitrogen dynamics in a tropical semiarid ecosystem. *European Journal of Soil Biology* 74:93–103 DOI 10.1016/j.ejsobi.2016.03.006.
- Sauer TJ, Cambardella CA, Meek DW. 2006. Spatial variation of soil properties relating to vegetation changes. *Plant and Soil* 280:1–5 DOI 10.1007/s11104-005-1545-8.
- Schlesinger WH, Raikes JA, Hartley AE, Cross AF. 1996. On the spatial pattern of soil nutrients in desert ecosystems. *Ecology* 77:364–374 DOI 10.2307/2265615.
- Smith SD, Monson R, Anderson JE. 1997. *Physiological ecology of North American desert plants*. Berlin: University of Nevada.
- Song W-K, Cui Y-J, Tang AM, Ding W-Q, Tran TD. 2013. Experimental study on water evaporation from sand using environmental chamber. *Canadian Geotechnical Journal* 51:115–128 DOI 10.1139/cgj-2013-0155.

- Specht R. 1966. The growth and distribution of mallee-broombush (Eucalyptus incrassata-Melaleuca uncinata association) and heath vegetation near Dark Island Soak, Ninety-Mile Plain, South Australia. *Australian Journal of Botany* 14:361–371 DOI 10.1071/BT9660361.
- Stanton JC, Pearson RG, Horning N, Ersts P, Akçakaya H. 2012. Combining static and dynamic variables in species distribution models under climate change. *Methods in Ecology and Evolution* 3:349–357 DOI 10.1111/j.2041-210X.2011.00157.x.
- Summers DM, Bryan BA, Crossman ND, Meyer WS. 2012. Species vulnerability to climate change: impacts on spatial conservation priorities and species representation. *Global Change Biology* 18:2335–2348 DOI 10.1111/j.1365-2486.2012.02700.x.
- Taylor S, Kumar L. 2013. Potential distribution of an invasive species under climate change scenarios using CLIMEX and soil drainage: a case study of Lantana camara L. in Queensland, Australia. *Journal of Environmental Management* 114:414–422 DOI 10.1016/j.jenvman.2012.10.039.
- Titeux N, Henle K, Mihoub JB, Regos A, Geijzendorffer IR, Cramer W, Verburg PH, Brotons L. 2016. Biodiversity scenarios neglect future land-use changes. *Global Change Biology* 22:2505–2515 DOI 10.1111/gcb.13272.
- Vanderwal J, Beaumont L, Zimmermann N, Lorch P. 2011. Package 'climates': methods for working with weather and climate. *Available at www rforge*.
- **Viscarra Rossel R, Chen C, Grundy M, Searle R, Clifford D, Odgers N. 2014a.** Soil and landscape grid national soil attribute maps-total phosphorus (3<sup>"</sup> resolution)-release 1. v4. *CSIRO Data Collection* DOI 10.4225/08/546F617719CAF.
- Viscarra Rossel R, Chen C, Grundy M, Searle R, Clifford D, Odgers N, Holmes K, Griffin T, Liddicoat C, Kidd D. 2014b. Soil and landscape grid national soil attribute maps—bulk density—whole earth (3<sup>"</sup> resolution). Release 1. v4. *CSIRO Data Collection* DOI 10.4225/08/546EE212B0048.
- Viscarra Rossel R, Chen C, Grundy M, Searle R, Clifford D, Odgers N, Holmes K, Griffin T, Liddicoat C, Kidd D. 2014c. Soil and landscape grid national soil attribute maps—organic carbon (3<sup>"</sup> resolution) Release 1. v1. *CSIRO Data Collection* DOI 10.4225/08/547523BB0801A.
- Viscarra Rossel R, Chen C, Grundy M, Searle R, Clifford D, Odgers N, Holmes K, Griffin T, Liddicoat C, Kidd D. 2014d. Soil and landscape grid national soil attribute maps - ph - cacl2 (3<sup>"</sup> resolution). Release 1. v2. *CSIRO Data Collection* DOI 10.4225/08/546F17EC6AB6E.
- Warren DL, Glor RE, Turelli M. 2008. Environmental niche equivalency versus conservatism: quantitative approaches to niche evolution. *Evolution* 62:2868–2883 DOI 10.1111/j.1558-5646.2008.00482.x.
- Westman WE. 1991. Measuring realized niche spaces: climatic response of chaparral and coastal sage scrub. *Ecology* 72:1678–1684 DOI 10.2307/1940967.
- Williams KJ, Belbin L, Austin MP, Stein JL, Ferrier S. 2012. Which environmental variables should I use in my biodiversity model? *International Journal of Geographical Information Science* 26:2009–2047 DOI 10.1080/13658816.2012.698015.

- Woodward FI. 1987. *Climate and plant distribution*. Cambridge: Cambridge University Press.
- Xin-Rong L. 2001. Study on shrub community diversity of Ordos Plateau, Inner Mongolia, northern China. *Journal of Arid Environments* **47**:271–279 DOI 10.1006/jare.2000.0707.
- Xu Z, Ward S, Chen C, Blumfield T, Prasolova N, Liu J. 2008. Soil carbon and nutrient pools, microbial properties and gross nitrogen transformations in adjacent natural forest and hoop pine plantations of subtropical Australia. *Journal of Soils and Sediments* 8:99–105 DOI 10.1065/jss2008.02.276.
- Yashiro Y, Shizu Y, Hirota M, Shimono A, Ohtsuka T. 2010. The role of shrub (Potentilla fruticosa) on ecosystem CO2 fluxes in an alpine shrub meadow. *Journal of Plant Ecology* 3:89–97 DOI 10.1093/jpe/rtq011.
- Zeng X, Zeng X, Barlage M. 2008. Growing temperate shrubs over arid and semiarid regions in the community land model–dynamic global vegetation model. *Global Biogeochemical Cycles* Epub ahead of print July 3 2008.
- Zhou Y, Pei Z, Su J, Zhang J, Zheng Y, Ni J, Xiao C, Wang R. 2012. Comparing soil organic carbon dynamics in perennial grasses and shrubs in a saline-alkaline arid region, northwestern China. *PLOS ONE* 7:e42927 DOI 10.1371/journal.pone.0042927.
- **Zinke PJ. 1962.** The pattern of influence of individual forest trees on soil properties. *Ecology* **43**:130–133 DOI 10.2307/1932049.

## **University Library**



# MINERVA A gateway to Melbourne's research publications

### Minerva Access is the Institutional Repository of The University of Melbourne

#### Author/s:

Hageer, Y; Esperon-Rodriguez, M; Baumgartner, JB; Beaumont, LJ

## Title:

Climate, soil or both? Which variables are better predictors of the distributions of Australian shrub species?

## Date:

2017-06-22

## Citation:

Hageer, Y., Esperon-Rodriguez, M., Baumgartner, J. B. & Beaumont, L. J. (2017). Climate, soil or both? Which variables are better predictors of the distributions of Australian shrub species?. PEERJ, 5 (6), https://doi.org/10.7717/peerj.3446.

## Persistent Link:

http://hdl.handle.net/11343/257310

File Description: published version License: CC BY