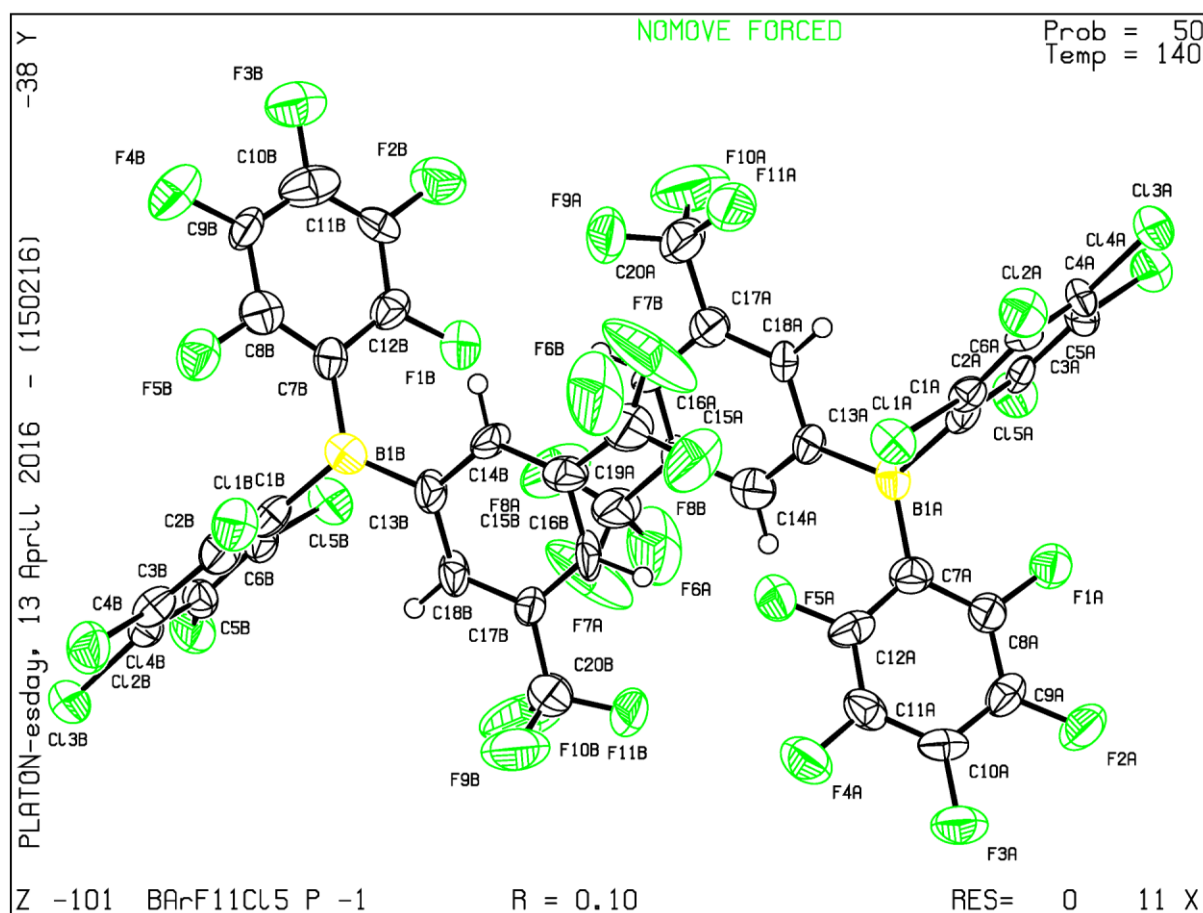


## SUPPLEMENTARY INFORMATION

H<sub>2</sub> activation using the first 1:1:1 hetero-tri(aryl)borane

Robin J. Blagg and Gregory G. Wildgoose

School of Chemistry, University of East Anglia, Norwich Research Park, Norwich, NR4 7TJ,  
U.K.

**Figure S1** X-ray crystallographic structure of B(C<sub>6</sub>F<sub>5</sub>){3,5-(CF<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>}(C<sub>6</sub>Cl<sub>5</sub>)<sub>3</sub>

## SUPPLEMENTARY INFORMATION

### “Gutmann-Beckett method” for measurement of Lewis acidity

**3** (Lewis acid) is combined with a three-fold excess of  $\text{OPEt}_3$  (Lewis base) in *ca.*  $0.8 \text{ cm}^3$   $\text{CD}_2\text{Cl}_2$  in a NMR tube, rapidly generating the Lewis acid-base adduct  $\text{Et}_3\text{PO-3}$ , and  $^1\text{H}$ ,  $^{11}\text{B}$ ,  $^{19}\text{F}$  and  $^{31}\text{P}\{^1\text{H}\}$  NMR spectra obtained.

#### **$\text{Et}_3\text{POB}(\text{C}_6\text{F}_5)\{\text{3,5-(CF}_3)_2\text{C}_6\text{H}_3\}(\text{C}_6\text{Cl}_5)$ $\text{Et}_3\text{PO-3}$**

$^1\text{H}$  NMR (500.2 MHz,  $\text{CD}_2\text{Cl}_2$ , 25 °C,  $\delta$ ): +7.81 (s, 2H,  $\text{Ar}^{\text{F6}}$  2,6-H), +7.68 (s, 1H,  $\text{Ar}^{\text{F6}}$  4-H), +1.89 (br.m, 6H, Et  $\text{CH}_2$ ), +1.10 (br.m, 9H, Et  $\text{CH}_3$ );  $^{11}\text{B}$  NMR (160.5 MHz,  $\text{CD}_2\text{Cl}_2$ , 25 °C,  $\delta$ ): +2.51 (br.s);  $^{19}\text{F}$  NMR (470.7 MHz,  $\text{CD}_2\text{Cl}_2$ , 25 °C,  $\delta$ ): -62.9 (s, 6F,  $\text{Ar}^{\text{F6}}$  3,5- $\text{CF}_3$ ), -131.3 (m, 2F,  $\text{Ar}^{\text{F5}}$  2,6-F), -158.8 (t,  $^3J_{\text{FF}} = 19.9$  Hz, 1F,  $\text{Ar}^{\text{F5}}$  4-F), -163.8 (m, 2F,  $\text{Ar}^{\text{F5}}$  3,5-F);  $^{31}\text{P}\{^1\text{H}\}$  NMR (202.5 MHz,  $\text{CD}_2\text{Cl}_2$ , 25 °C,  $\delta$ ): +76.49 (s).

## SUPPLEMENTARY INFORMATION

### H<sub>2</sub> cleavage by FLPs

Equimolar quantities (*ca.* 30  $\mu\text{mol}$ ) of Lewis acid (**3**) and Lewis base {either P(<sup>t</sup>Bu)<sub>3</sub>, 2,2,6,6-tetramethylpiperidine (tmp), or 2,6-lutidine} are combined in *ca.* 0.8 cm<sup>3</sup> CD<sub>2</sub>Cl<sub>2</sub> in a NMR tube fitted with a J.Young valve. <sup>1</sup>H, <sup>11</sup>B, <sup>19</sup>F and <sup>31</sup>P{<sup>1</sup>H} NMR spectra are obtained. The solution is degassed in the NMR tube by three freeze-pump-thaw cycles, before being frozen and the head-space of the NMR tube filled with 1 bar H<sub>2</sub> (dried by passing through a P<sub>2</sub>O<sub>5</sub> column). The NMR tube is allowed to warm to room temperature, shaken, and the resulting reaction monitored by <sup>1</sup>H and <sup>11</sup>B NMR spectroscopy. A final set of <sup>1</sup>H, <sup>11</sup>B, <sup>19</sup>F and <sup>31</sup>P{<sup>1</sup>H} NMR spectra are then obtained.

#### **[(<sup>t</sup>Bu)<sub>3</sub>PH][HB(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>,5-(CF<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>](C<sub>6</sub>Cl<sub>5</sub>) [<sup>t</sup>Bu<sub>3</sub>PH][H-3]**

Spectral data at 57% conversion (164 hours reaction time).

<sup>1</sup>H NMR (500.2 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): +7.68 (s, 2H, Ar<sup>F6</sup> 2,6-H), +7.47 (s, 1H, Ar<sup>F6</sup> 4-H), +5.10 (d, <sup>1</sup>J<sub>HP</sub> = 430 Hz, 1H), +4.08 (br.q, <sup>1</sup>J<sub>HB</sub> = 88 Hz, 1H), +1.61 (d, <sup>3</sup>J<sub>HP</sub> = 15.7 Hz, 27H); <sup>11</sup>B NMR (160.5 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): -14.3 (d, <sup>1</sup>J<sub>BH</sub> = 88 Hz); <sup>19</sup>F NMR (470.7 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): -62.3 (s, 6F, Ar<sup>F6</sup> 3,5-CF<sub>3</sub>), -130.8 (br.m, 2F, Ar<sup>F5</sup> 2,6-F), -160.4 (t, <sup>3</sup>J<sub>FF</sub> = 20.3 Hz, 1F, Ar<sup>F5</sup> 4-F), -167.2 (m, 2F, Ar<sup>F5</sup> 3,5-F); <sup>31</sup>P{<sup>1</sup>H} NMR (202.5 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): +59.9 (s).

#### **[Me<sub>4</sub>H<sub>6</sub>C<sub>5</sub>NH<sub>2</sub>][HB(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>,5-(CF<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>](C<sub>6</sub>Cl<sub>5</sub>) [tmp-H][H-3]**

Spectral data at 38% conversion (164 hours reaction time); resonances for tmp correspond to a rapid equilibrium between [tmp-H]<sup>+</sup> and free tmp.

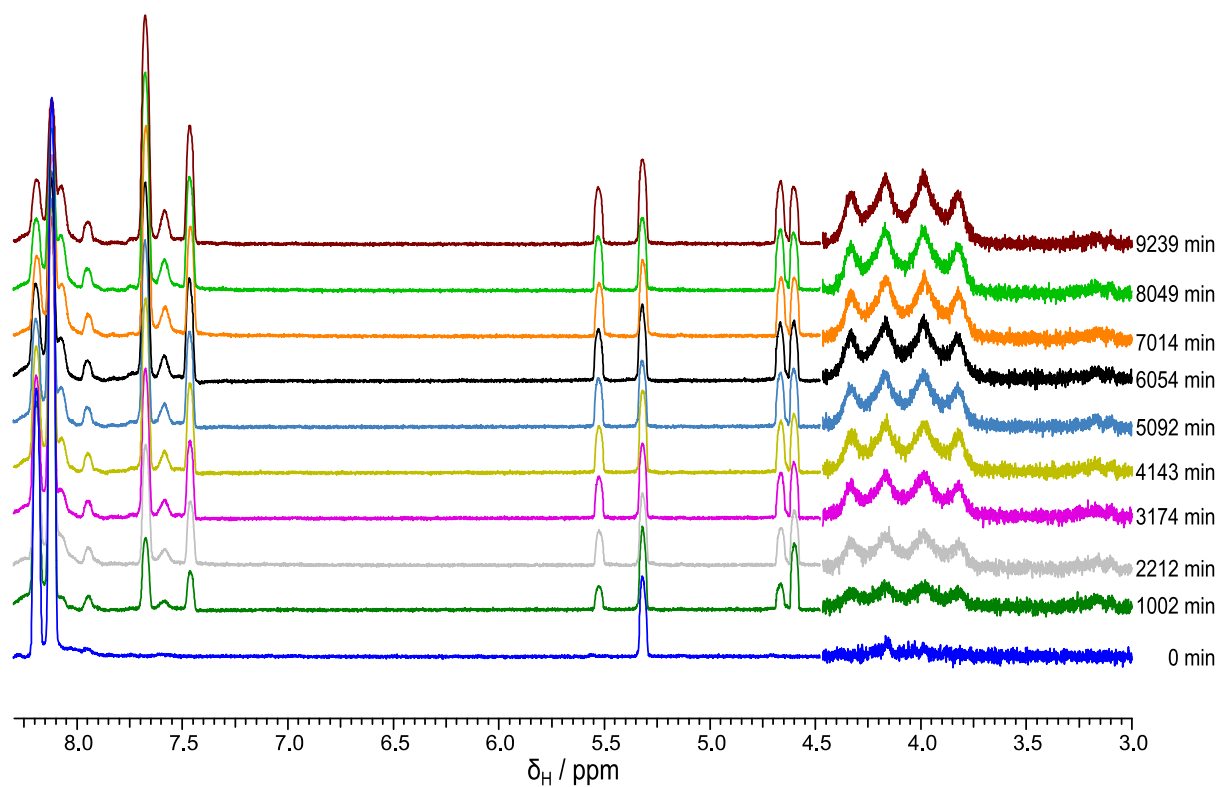
<sup>1</sup>H NMR (500.2 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): +7.63 (s, 2H, Ar<sup>F6</sup> 2,6-H), +7.52 (s, 1H, Ar<sup>F6</sup> 4-H), +3.98 (br.q, <sup>1</sup>J<sub>HB</sub> = 84 Hz, 1H), +2.90 (vbr.s, tmp NH<sub>2</sub>), +1.67 (m, tmp 4-H), +1.42 (m, tmp 3,5-H), +1.17 (s, tmp 2,6-CH<sub>3</sub>); <sup>11</sup>B NMR (160.5 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): -13.9 (d, <sup>1</sup>J<sub>BH</sub> = 84 Hz); <sup>19</sup>F NMR (470.7 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): -62.5 (s, 6F, Ar<sup>F6</sup> 3,5-CF<sub>3</sub>), -130.9 (br.m, 2F, Ar<sup>F5</sup> 2,6-F), -162.9 (t, <sup>3</sup>J<sub>FF</sub> = 20.3 Hz, 1F, Ar<sup>F5</sup> 4-F), -165.9 (m, 2F, Ar<sup>F5</sup> 3,5-F).

#### **[Me<sub>2</sub>H<sub>3</sub>C<sub>5</sub>NH][HB(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>,5-(CF<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>](C<sub>6</sub>Cl<sub>5</sub>) [lutidine-H][H-3]**

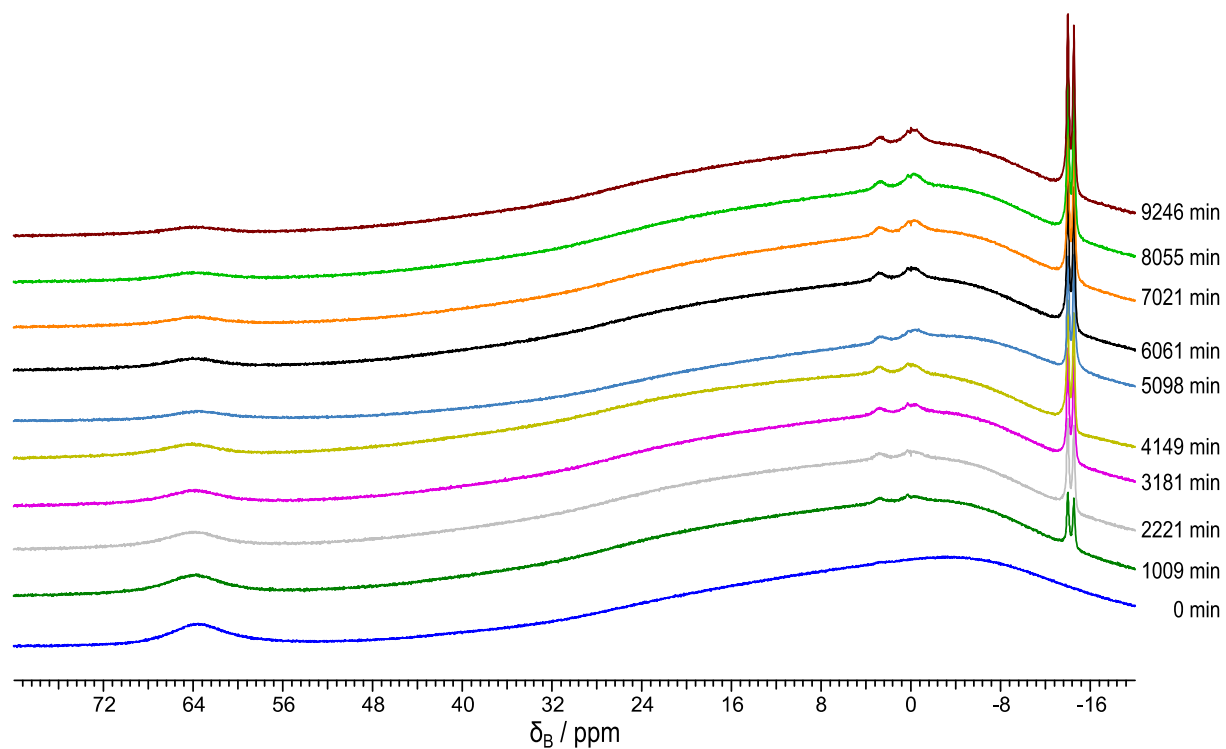
Spectral data at 64% conversion (164 hours reaction time); resonances for lutidine correspond to a rapid equilibrium between [lutidine-H]<sup>+</sup> and free lutidine.

<sup>1</sup>H NMR (500.2 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): +7.67 (s, 2H, Ar<sup>F6</sup> 2,6-H), +7.62 (t, <sup>3</sup>J<sub>HH</sub> = 7.7 Hz, lutidine 4-H), +7.45 (s, 1H, Ar<sup>F6</sup> 4-H), +7.09 (d, <sup>3</sup>J<sub>HH</sub> = 7.7 Hz, lutidine 3,5-H), +4.08 (br.q, <sup>1</sup>J<sub>HB</sub> = 88 Hz, 1H), +2.50 (s, lutidine 2,6-CH<sub>3</sub>); <sup>11</sup>B NMR (160.5 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): -14.3 (d, <sup>1</sup>J<sub>BH</sub> = 88 Hz); <sup>19</sup>F NMR (470.7 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 25 °C,  $\delta$ ): -62.4 (s, 6F, Ar<sup>F6</sup> 3,5-CF<sub>3</sub>), -131.0 (br.m, 2F, Ar<sup>F5</sup> 2,6-F), -160.5 (t, <sup>3</sup>J<sub>FF</sub> = 21.2 Hz, 1F, Ar<sup>F5</sup> 4-F), -167.3 (m, 2F, Ar<sup>F5</sup> 3,5-F).

## SUPPLEMENTARY INFORMATION

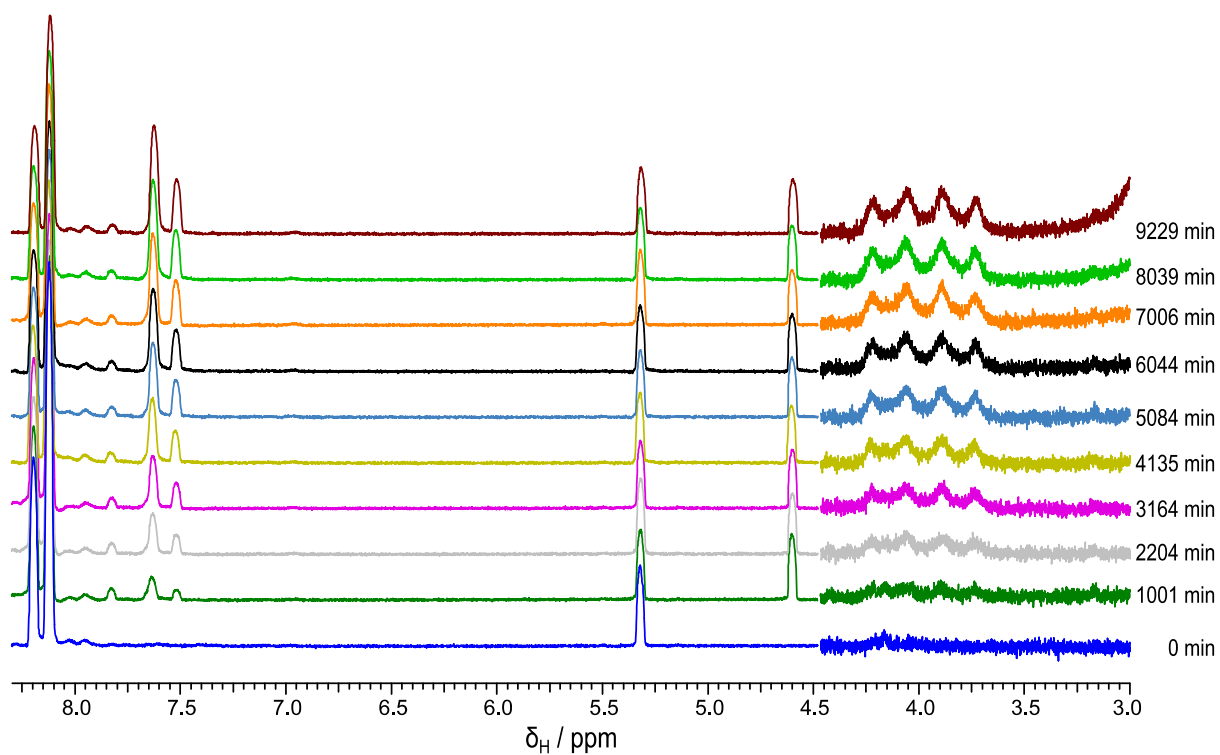


**Figure S2a**  $^1\text{H}$  NMR spectra showing the progress of  $\text{H}_2$  cleavage by the  $3/\text{P}(\text{tBu})_3$  FLP

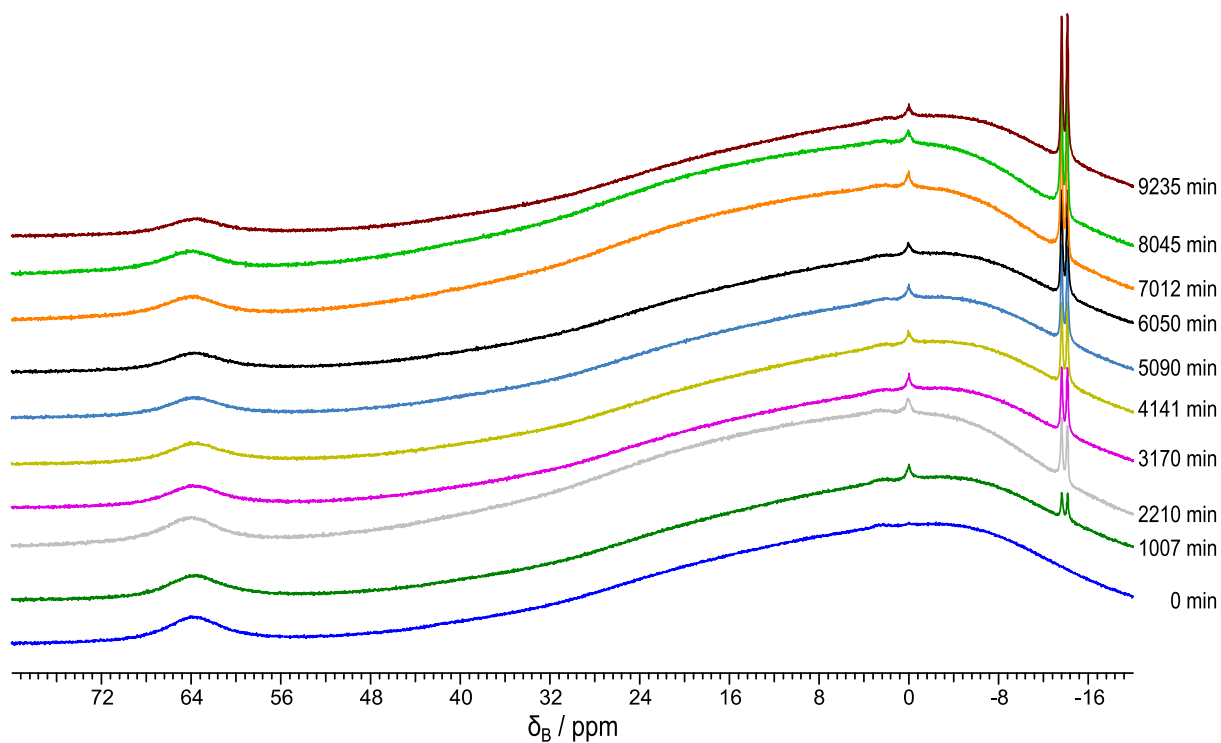


**Figure S2b**  $^{11}\text{B}$  NMR spectra showing the progress of  $\text{H}_2$  cleavage by the  $3/\text{P}(\text{tBu})_3$  FLP

## SUPPLEMENTARY INFORMATION

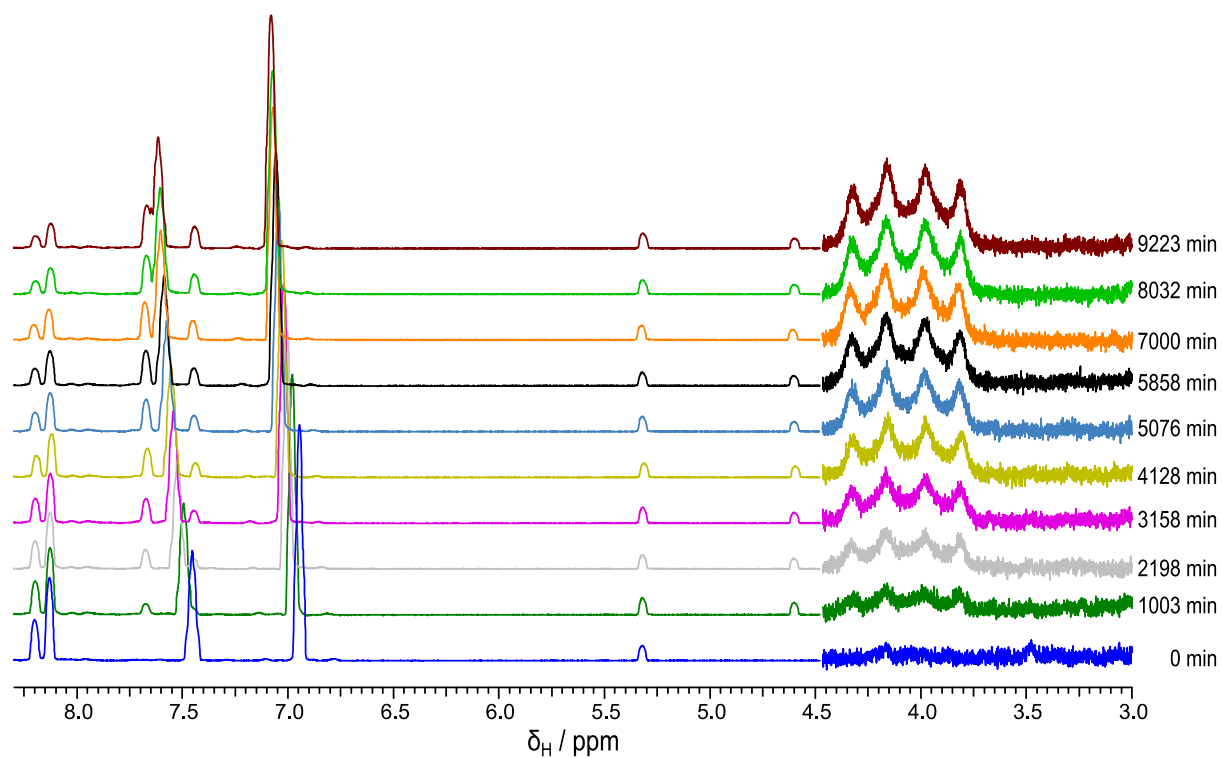


**Figure S3a**  $^1\text{H}$  NMR spectra showing the progress of  $\text{H}_2$  cleavage by the 3/tmp FLP

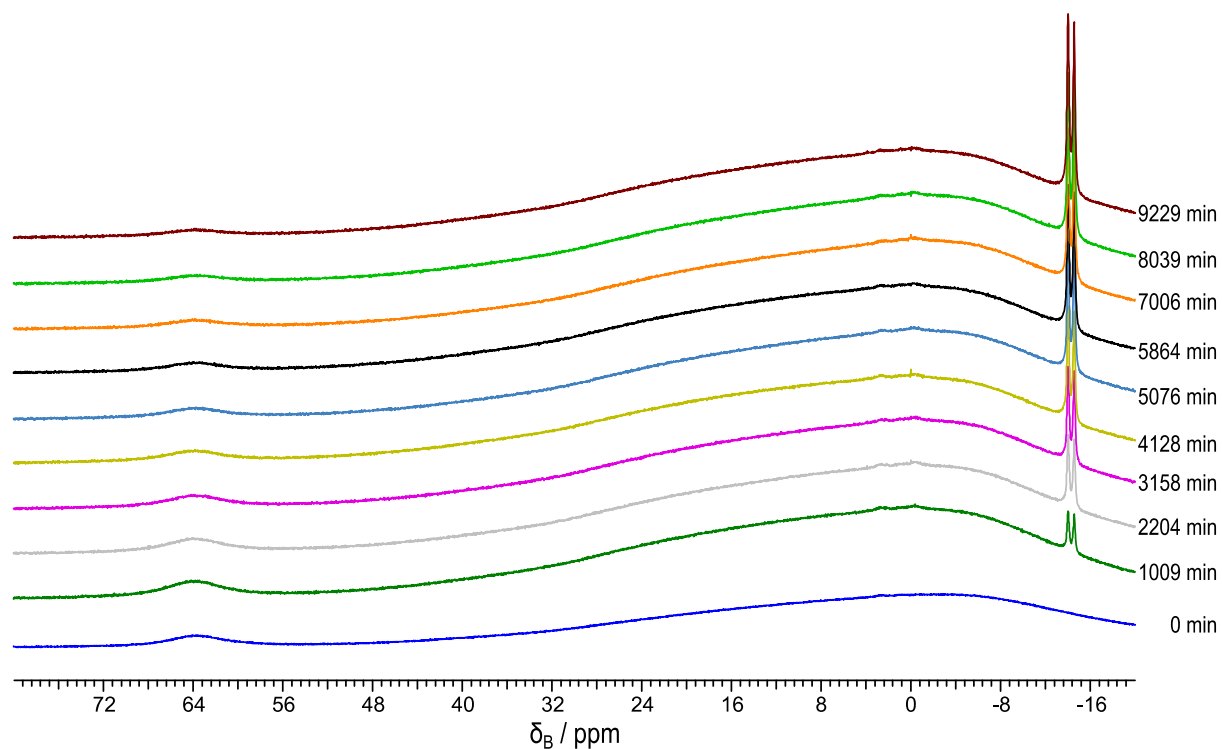


**Figure S3b**  $^{11}\text{B}$  NMR spectra showing the progress of  $\text{H}_2$  cleavage by the 3/tmp FLP

## SUPPLEMENTARY INFORMATION



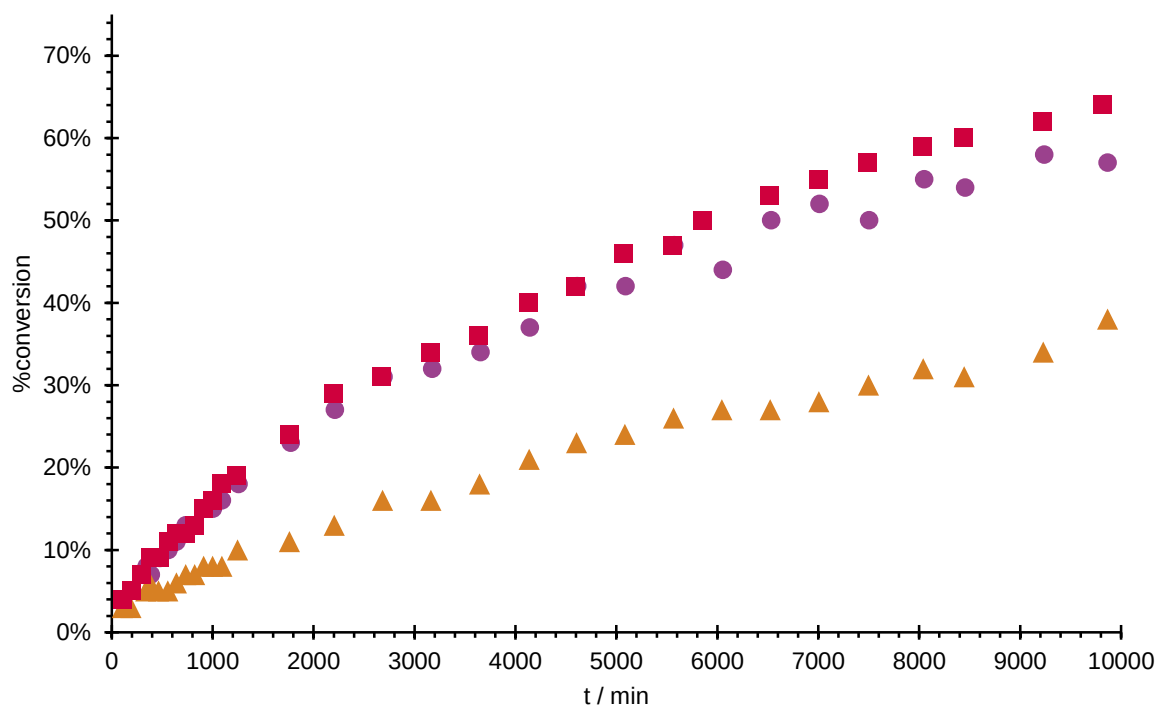
**Figure S4a**  $^1\text{H}$  NMR spectra showing the progress of  $\text{H}_2$  cleavage by the 3/lutidine FLP



**Figure S4b**  $^{11}\text{B}$  NMR spectra showing the progress of  $\text{H}_2$  cleavage by the 3/lutidine FLP

## SUPPLEMENTARY INFORMATION

>90% consumed **3**, is converted to the target H<sub>2</sub> cleavage product [H-**3**]<sup>-</sup>; non-negligible by-products are however observed in reactions where the Lewis base is P(<sup>t</sup>Bu)<sub>3</sub> (Figure S2) or tmp (Figure S3). The signals in the range δ<sub>B</sub> -4 - +4 ppm are indicative of tetrahedral boron and it is speculated that these are the water adduct **3**-OH<sub>2</sub>, or the hydroxide [**3**-OH]<sup>-</sup>. Similarly, this explains the by-product resonances observable in aromatic region of the <sup>1</sup>H spectra.



**Figure S5** Percentage conversion of **3** to [H-**3**]<sup>-</sup> (monitored by <sup>1</sup>H NMR spectroscopy of Ar<sup>F6</sup> 2,6-/4-H resonances), by reaction of a FLP with H<sub>2</sub> in CD<sub>2</sub>Cl<sub>2</sub> at 20 °C, with varying Lewis base:

● P(<sup>t</sup>Bu)<sub>3</sub>, ▲ tmp, ■ lutidine.