

## HOKKAIDO UNIVERSITY

Title	Draft Genome Sequence of Chlamydia trachomatis Strain 54, Isolated from the Urogenital Tract of a Male in Japan
Author(s)	Yamazaki, Tomohiro; Matsuo, Junji; Kikuchi, Momoka; Miyamoto, Kentaro; Oka, Kentaro; Takahashi, Motomichi; Takahashi, Satoshi; Okubo, Torahiko; Yamaguchi, Hiroyuki
Citation	Genome Announcements, 3(5), e01242-15 https://doi.org/10.1128/genomeA.01242-15
Issue Date	2015-10-22
Doc URL	http://hdl.handle.net/2115/60141
Rights(URL)	http://creativecommons.org/licenses/by/3.0/
Туре	article
File Information	Genome Announc_3(5)_e01242-15.pdf







## Draft Genome Sequence of *Chlamydia trachomatis* Strain 54, Isolated from the Urogenital Tract of a Male in Japan

Tomohiro Yamazaki,<sup>a,b</sup> Junji Matsuo,<sup>a</sup> Momoka Kikuchi,<sup>a</sup> Kentaro Miyamoto,<sup>c</sup> Kentaro Oka,<sup>c</sup> Motomichi Takahashi,<sup>c</sup> Satoshi Takahashi,<sup>d</sup> Torahiko Okubo,<sup>a</sup> Hiroyuki Yamaguchi<sup>a</sup>

Department of Medical Laboratory Science, Faculty of Health Sciences, Hokkaido University, Sapporo, Japan<sup>a</sup>; Research Fellow of Japan Society for the Promotion of Science, Tokyo, Japan<sup>b</sup>; Miyarisan Pharmaceutical Co., Ltd., Tokyo, Japan<sup>c</sup>; Department of Infection Control and Laboratory Medicine, Sapporo Medical University School of Medicine, Sapporo, Japan<sup>d</sup>

We report the draft genome sequence of *Chlamydia trachomatis* strain 54, isolated from the urogenital tract of a male in Japan, with unique polymorphic membrane proteins. Detailed genomic analysis will aid our understanding of the selective pressures that lead to sexual differentiation in chlamydial adaptive evolution.

Received 9 September 2015 Accepted 15 September 2015 Published 22 October 2015

Citation Yamazaki T, Matsuo J, Kikuchi M, Miyamoto K, Oka K, Takahashi M, Takahashi S, Okubo T, Yamaguchi H. 2015. Draft genome sequence of *Chlamydia trachomatis* strain 54, isolated from the urogenital tract of a male in Japan. Genome Announc 3(5):e01242-15. doi:10.1128/genomeA.01242-15.

Copyright © 2015 Yamazaki et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Hiroyuki Yamaguchi, hiroyuki@med.hokudai.ac.jp.

Obligate intracellular bacterium *Chlamydia trachomatis*, the causative agent of a sexually transmitted disease, can lead to serious medical complications such as infertility through ductal obstruction or pelvic inflammatory disease in females and testicular atrophy or epididymitis in males (1–6). Although chlamydial infection is reported in both men and women, the male urogenital tract lacks the microbial flora present in the female genital tract. This fact suggests that *C. trachomatis* strains would face different selective pressures, leading to specific genetic polymorphisms depending on sexual differentiation. However, *C. trachomatis* research has been limited to strains isolated from males because of difficulties in the isolation of organisms.

It is well established that the polymorphic membrane protein (Pmp) family generally possesses the properties of type V secretion systems (7). Interestingly, accumulating evidence has shown that among the nine Pmp families (A through I) expressed by *C. trachomatis*, PmpF is a high-definition phylogenetic marker that is strongly affected by host-selective pressures, such as those exerted by the microbial flora (7–9). In fact, we have recently demonstrated that PmpF of *C. trachomatis* isolated from the urogenital tracts of Japanese males showed less genetic diversity than the same protein isolated from the genital tracts of Japanese females, raising the idea that genetic diversity among *C. trachomatis* strains may be dependent on sexual differentiation (10). We therefore sequenced *C. trachomatis* strain 54 (*ompA* genotype B, accession number LC031846) isolated from the urogenital tract of a Japanese male.

The draft genome of *C. trachomatis* was obtained using an Illumina Miseq sequencer (Illumina, San Diego, CA, USA), with sequencing runs for paired-end sequences. The bacterial DNA libraries were prepared using an NEBNext DNA Library Prep master mix set for Illumina (New England Biolabs, Ipswich, Massachusetts, USA). The genome was assembled using *de novo* sequence assembler software (Platanus 1.2.1) (11), and 14 contigs were obtained with sizes ranging from 226 to 253,058 bp. Rapid Annotation using Subsystem Technology (RAST: http://rast .nmpdr.org/) was used for gene annotation (12). Functional annotation was performed using the Kyoto Encyclopedia of Genes and Genomes (KEGG) (http://www.genome.jp/kegg/) (13).

The draft genome sequence of *C. trachomatis* strain 54 was 1,046,714 bp in length (G+C content, 46.37%; coverage, 300-fold). The genome sequence contained 976 coding sequences with 38 tRNAs and 2 ribosomal RNAs. Using KEGG analysis and comparisons with *C. trachomatis* strain D/UW-3/CX (female strain) (NC\_000117.1), the male strain was confirmed to possess conserved central carbon metabolic pathways and type III general secretion gene clusters. BLAST pairwise alignments interestingly revealed that the *pmpB* sequence of *C. trachomatis* strain 54 was unique, as well as the *pmpF*, a characteristic not previously reported for pathogenic *Chlamydiae*. A more detailed analysis of the genome will help us to understand the selective pressures that lead to sexual differentiation in chlamydial adaptive evolution.

**Nucleotide sequence accession numbers.** The draft genome sequence of *C. trachomatis* strain 54 has been deposited in the DDBJ database under accession numbers BCAM01000001 through BCAM01000014 (14 entries). The version described in this paper is the first version.

## ACKNOWLEDGMENTS

This study was supported by grants-in-aid for the Japan Society for the Promotion of Science fellows (13J04219) and scientific research from KAKENHI, grant numbers 21590474, 24659194, and 24117501 (Innovation Areas, Matryoshka-type evolution). The funders had no role in study design, data collection and analysis, the decision to publish, or preparation of the manuscript. In addition, the strain was provided from the Japanese surveillance committee, consisting of the Japanese Society of Chemotherapy, the Japanese Associations for Infectious Diseases, and the Japanese Society for Clinical Microbiology (Tokyo, Japan).

## REFERENCES

- 1. Horner P. 2006. The case for further treatment studies of uncomplicated genital *Chlamydia trachomatis* infection. Sex Transm Infect 82:340–343. http://dx.doi.org/10.1136/sti.2005.019158.
- 2. Paavonen J, Vesterinen E. 1982. *Chlamydia trachomatis* in cervicitis and urethritis in women. Scand J Infect Dis **32**(Suppl):45–54.
- Westergaard L, Philipsen T, Scheibel J. 1982. Significance of cervical *Chlamydia trachomatis* infection in postabortal pelvic inflammatory disease. Obstet Gynecol 60:322–325.
- Faxelid E, Krantz I. 1993. Experiences of disease and treatment among *Chlamydia* patients. Scand J Caring Sci 7:169–173. http://dx.doi.org/ 10.1111/j.1471-6712.1993.tb00194.x.
- Gimenes F, Souza RP, Bento JC, Teixeira JJV, Maria-Engler SS, Bonini MG, Consolaro MEL. 2014. Male infertility: a public health issue caused by sexually transmitted pathogens. Nat Rev Urol 11:672–687. http:// dx.doi.org/10.1038/nrurol.2014.285.
- Redgrove KA, McLaughlin EA. 2014. The role of the immune response in *Chlamydia trachomatis* infection of the male genital tract: a double-edged sword. Front Immunol 5:534. http://dx.doi.org/10.3389/ fimmu.2014.00534.
- 7. Henderson IR, Lam AC. 2001. Polymorphic proteins of *Chlamydia* spp.–autotransporters beyond the *Proteobacteria*. Trends Microbiol 9:573–578. http://dx.doi.org/10.1016/S0966-842X(01)02234-X.
- Tan C, Hsia R-, Shou H, Haggerty CL, Ness RB, Gaydos CA, Dean D, Scurlock AM, Wilson DP, Bavoil PM. 2009. *Chlamydia trachomatis*infected patients display variable antibody profiles against the nine-

member polymorphic membrane protein family. Infect Immun 77: 3218–3226. http://dx.doi.org/10.1128/IAI.01566-08.

- Gomes JP, Nunes A, Bruno WJ, Borrego MJ, Florindo C, Dean D. 2006. Polymorphisms in the nine polymorphic membrane proteins of *Chlamydia trachomatis* across all serovars: evidence for serovar da recombination and correlation with tissue tropism. J Bacteriol 188:275–286. http://dx.doi.org/10.1128/JB.188.1.275-286.2006.
- Yamazaki T, Matsuo J, Takahashi S, Kumagai S, Shimoda T, Abe K, Minami K, Yamaguchi H. 2015. A characteristic of polymorphic membrane protein F of *Chlamydia trachomatis* isolated from male urogenital tracts in Japan. J Infect Chemother, in press.
- 11. Kajitani R, Toshimoto K, Noguchi H, Toyoda A, Ogura Y, Okuno M, Yabana M, Harada M, Nagayasu E, Maruyama H, Kohara Y, Fujiyama A, Hayashi T, Itoh T. 2014. Efficient de novo assembly of highly heterozygous genomes from whole-genome shotgun short reads. Genome Res 24: 1384–1395. http://dx.doi.org/10.1101/gr.170720.113.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/ 1471-2164-9-75.
- Ogata H, Goto S, Sato K, Fujibuchi W, Bono H, Kanehisa M. 1999. KEGG: Kyoto encyclopedia of genes and genomes. Nucleic Acids Res 27: 29–34. http://dx.doi.org/10.1093/nar/27.1.29.