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Citation	Proceedings of the National Academy of Sciences of the United States of America, 112(14), 4304-4309 <a href="https://doi.org/10.1073/pnas.1423819112">https://doi.org/10.1073/pnas.1423819112</a>
Issue Date	2015-04-08
Doc URL	<a href="http://hdl.handle.net/2115/59917">http://hdl.handle.net/2115/59917</a>
Type	article (author version)
Note	This article contains supporting information online at <a href="http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1423819112/-/DCSupplemental">www.pnas.org/lookup/suppl/doi:10.1073/pnas.1423819112/-/DCSupplemental</a> .
Note(URL)	<a href="http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1423819112/-/DCSupplemental">http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1423819112/-/DCSupplemental</a>
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BIOLOGICAL SCIENCE: Biochemistry

**SWI/SNF chromatin-remodeling complexes function in noncoding RNA-dependent assembly of nuclear bodies**

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**Short title:** SWI/SNF complexes for nuclear RNP body assembly

## **Abstract**

Paraspeckles are subnuclear structures that form around nuclear paraspeckle assembly transcript 1 (*NEAT1*) long noncoding (lnc)RNA. Recently, paraspeckles were shown to be functional nuclear bodies involved in stress responses and the development of specific organs. Paraspeckle formation is initiated by transcription of the *NEAT1* chromosomal locus and proceeds in conjunction with *NEAT1* lncRNA biogenesis and a subsequent assembly step involving >40 paraspeckle proteins (PSPs). In this study, subunits of SWItch/Sucrose NonFermentable (SWI/SNF) chromatin-remodeling complexes were identified as paraspeckle components that interact with PSPs and *NEAT1* lncRNA. Electron microscopic observations revealed that SWI/SNF complexes were enriched in paraspeckle subdomains depleted of chromatin. Knockdown of SWI/SNF components resulted in paraspeckle disintegration, but mutation of the ATPase domain of the catalytic subunit BRG1 did not affect paraspeckle integrity, indicating that the essential role of SWI/SNF complexes in paraspeckle formation does not require their canonical activity. Knockdown of SWI/SNF complexes barely affected the levels of known essential paraspeckle components, but markedly diminished the interactions between essential PSPs, suggesting that SWI/SNF complexes facilitate organization of the PSP interaction network required for intact paraspeckle assembly. The interactions between SWI/SNF components and essential PSPs were maintained in *NEAT1*-depleted cells, suggesting that SWI/SNF complexes not only facilitate interactions between PSPs, but also recruit PSPs during paraspeckle assembly. SWI/SNF complexes were also required for Satellite III lncRNA-dependent formation of nuclear stress bodies under heat shock conditions. Our data suggest the existence of a

common mechanism underlying the formation of lncRNA-dependent nuclear body architectures in mammalian cells.

**Keywords:** chromatin-remodeling complex; long noncoding RNA; nuclear bodies; ribonucleoprotein assembly.

### **Significant Statement**

The postgenomic era has seen the identification of numerous lncRNAs in mammalian cells; however, their biological functions remain largely unknown. NEAT1 is a lncRNA that participates in the construction of the massive ribonucleoprotein subnuclear complex called the paraspeckle. Paraspeckle formation proceeds in conjunction with NEAT1 biogenesis and the assembly of >40 proteins. Here, we demonstrate that SWI/SNF chromatin-remodeling complexes play an essential role in organizing the protein-protein interaction network of paraspeckles, in a manner that does not require their canonical remodeling activity. We also show that SWI/SNF complexes are required for the assembly of the SatIII lncRNA-dependent nuclear stress body. These data suggest the presence of a common pathway for the assembly of lncRNA-dependent nuclear bodies in mammalian cells.

¥body

Paraspeckles are nuclear bodies that are typically detected as foci in close proximity to nuclear speckles (1, 2) and were initially defined as foci enriched in characteristic RNA-binding proteins, including PSPC1, NONO, and SFPQ (3, 4). Nuclear paraspeckle assembly transcript 1 (*NEAT1*), a long noncoding RNA (lncRNA), localizes exclusively to paraspeckles and acts as a core structural component of these ribonucleoprotein (RNP) bodies (5-7). Paraspeckles are approximately 360 nm in diameter (8); hence, they are considered huge among RNP particles. Paraspeckles regulate the expression of a number of genes via the sequestration of specific proteins and RNAs (4, 9, 10), and are physiologically involved in the development of the corpus luteum and the mammary gland in mice (11, 12).

Paraspeckle formation is initiated by *NEAT1* transcription at the *NEAT1* locus on human chromosome 11 (5, 13). The *NEAT1* gene generates two isoform transcripts, namely, 3.7 kb *NEAT1\_1* and 23 kb *NEAT1\_2*. Both isoforms are transcribed from the same promoter and can be processed at the 3'-end to produce a canonically polyadenylated *NEAT1\_1* isoform and a noncanonically processed *NEAT1\_2* isoform (6, 7, 14). Whereas *NEAT1\_2* is required for *de novo* paraspeckle construction, *NEAT1\_1* is not required for this process (6, 14, 15). Extensive RNA interference (RNAi) analyses of 40 paraspeckle proteins (PSPs) revealed that seven PSPs, namely, heterogeneous nuclear ribonucleoprotein K (HNRNPK), NONO, RNA-binding motif protein 14 (RBM14), SFPQ, DAZ-associated protein 1 (DAZAP1), fused in sarcoma (FUS), and HNRNPH3, are essential for paraspeckle formation (14). HNRNPK, NONO, RBM14, and SFPQ (category 1A proteins), but not

DAZAP1, FUS, or HNRNPH3 (category 1B proteins), are required for the accumulation of the essential *NEAT1\_2* isoform (14). Whereas HNRNPK facilitates *NEAT1\_2* synthesis by interfering with the 3'-end processing of *NEAT1\_1* (14), other proteins in category 1A stabilize the *NEAT1\_2* isoform. Although category 1B proteins barely affect *NEAT1\_2* accumulation, they are required for the assembly of intact paraspeckles (14). These data suggest that paraspeckle formation proceeds in conjunction with the biogenesis of *NEAT1* lncRNA and subsequent ribonucleoprotein assembly.

SWItch/Sucrose NonFermentable (SWI/SNF) chromatin-remodeling complexes are known to disrupt nucleosome architecture and elicit changes in gene expression. They play roles in a number of biological processes and are implicated in several diseases (16). The presence of either the BRG1 or the BRM subunit of SWI/SNF, which are both ATPases, is sufficient for nucleosome remodeling *in vitro*; however, maximal activity requires additional subunits (17). Here, we report the identification of SWI/SNF complexes as novel factors required for paraspeckle formation and demonstrate that this function does not require their ATPase activity. We demonstrate that several SWI/SNF subunits interact with known PSPs. The predominant localization of SWI/SNF components within paraspeckles was also confirmed. The results suggest a novel function of SWI/SNF complexes in the architectural lncRNA-dependent assembly of nuclear bodies.

## **Results**

### **Subunits of SWI/SNF complexes are major paraspeckle components**

A search for proteins that interact with the 40 known PSPs in information publically available

in the STRING database (ver. 9.05) (<http://string905.embl.de>) revealed core subunits of the SWI/SNF chromatin-remodeling complex, particularly essential category 1 proteins (Table 1). Immunocytochemical analyses of human HeLa cells showed that three SWI/SNF subunits (BRG1, BRM, and BAF155) exhibited broad nucleoplasmic distributions with several prominent foci that overlapped with the locations of *NEAT1* lncRNA detected by RNA fluorescence *in situ* hybridization (RNA-FISH) (Fig. 1A). The specific detection of BRG1 was confirmed by using a distinct BRG1 antibody with different fixation conditions (Fig. S1A), and by observing signal disappearance upon BRG1 knockdown (Fig. S1B). Paraspeckle localization of BRG1 and BRM was also observed in other human cell lines (A549 and HEK293) and in a mouse cell line (NIH3T3) (Fig. 1B). These results indicate that a subpopulation of the SWI/SNF subunits is localized to paraspeckles.

Transcriptional inhibition by actinomycin D leads to paraspeckle disintegration and relocation of PSPs, but not *NEAT1*, to perinucleolar cap structures (2, 6, 14). Similarly, in our hands, treatment of HeLa cells with actinomycin D resulted in the disappearance of SWI/SNF foci and relocation of BRG1 and BRM to perinucleolar cap structures (Fig. 1C). BRG1 signals overlapped with those of other PSPs such as NONO, but not with COIL, which is known to relocate to distinct perinucleolar caps (Fig. 1C) (2, 6, 14). Furthermore, knockdown of *NEAT1* with antisense oligonucleotides (Fig. S2A) resulted in the disappearance of SWI/SNF foci without affecting the levels of BRG1 and BRM (Fig. S2B and C), indicating that SWI/SNF foci require *NEAT1* for their integrity.

Immunogold electron microscopic (I-EM) detection revealed a patchy pattern of distribution of SWI/SNF, indicating that it was concentrated within paraspeckle subdomains

(Fig. 1D, upper left panel). This patchy pattern of localization, which was not observed for the other PSPs (Fig. S1C), was even more evident in enlarged paraspeckles generated by treatment with the proteasome inhibitor MG132 (9) (Fig. 1D, upper right panel). The use of an antibody against histone H3 revealed that the paraspeckle interior region, where the majority of the BRG1 protein was localized, contained very little chromatin (Fig. 1D, lower panels). Taken together, these results suggest that SWI/SNF subunits are *bona fide* paraspeckle components that do not overlap with distinct subnuclear structures or chromosome loci.

In UV crosslinking immunoprecipitation experiments, *NEAT1* coimmunoprecipitated with BRG1 and BRM, but only after UV irradiation (Fig. 2A), indicating a direct interaction between *NEAT1* and these SWI/SNF subunits. In addition, PSPs essential for paraspeckle formation were efficiently coimmunoprecipitated with BRG1 even after treatment with RNase A or ethidium bromide (18) (Fig. 2B), indicating the absence of an RNA or DNA bridge between the SWI/SNF subunits and PSPs. These observations suggest that SWI/SNF subunits are stable paraspeckle components.

### **SWI/SNF complexes are required for paraspeckle formation**

The results described above raised the possibility that SWI/SNF complexes are involved in paraspeckle formation. To test this hypothesis, the expression of six SWI/SNF core subunits was impaired by RNAi and the integrity of the paraspeckle structure was examined. Single knockdown of either BRG1 or BRM had no effect on paraspeckle appearance in HeLa cells (Fig. S1B and S3); however, double knockdown of BRG1 and BRM using a combination of



two siRNAs (Fig. S4A) resulted in obvious paraspeckle disintegration (Fig. 3A and B), suggesting that these SWI/SNF subunits are functionally redundant for paraspeckle formation. Single knockdowns of four BAF proteins (BAF170, BAF155, BAF57, and BAF47) (Fig. S4A) also resulted in marked paraspeckle disintegration in approximately 70% of cells (Fig. 3A and B). Based on these data, we propose that components of intact paraspeckle-localized SWI/SNF complexes are involved in the formation of paraspeckles.

In SW13 cells, SWI/SNF complexes are undetectable due to silencing of both BRG1 and BRM (Fig. S5A) (19). When we examined these cells, we observed normal nuclear speckles but no paraspeckle-like nuclear foci (Fig. 3C). In addition, immunoblot analyses and RNase protection assays (RPAs) of SW13 cells revealed the accumulation of essential PSPs and *NEAT1*, respectively (Fig. S5A and B). This result supports the hypothesis that paraspeckle formation specifically requires SWI/SNF complexes.

To investigate the requirement of the remodeling activity of SWI/SNF complexes for paraspeckle formation, the CRISPR/Cas9 system was used to create small deletions of the catalytic subunit BRG1 that spanned the most classically characterized essential lysine residue (K785) in the ATPase domain (20) (M1 and M2 in Fig. 3D, left panel). To accelerate genomic engineering, we employed the near-haploid human HAP1 cell line, in which we detected marginal expression of BRM (Fig. S6A), suggesting that paraspeckle formation depends on BRG1 without the functional redundancy of BRM. Paraspeckle formation was abolished by RNAi-mediated knockdown of BRG1 in HAP1 cells, but was barely affected by knockdown of BRM (Fig. S6B). Although the ATPase M1 and M2 mutations moderately diminished the expression level of BRG1 in HAP1 cells (Fig. S6C), both mutant proteins

were properly localized and sustained the paraspeckle structure (Fig. 3D, right panel). The size and number of paraspeckles were not altered in the M1 and M2 mutant cells (Fig. 3D, right panel). We also confirmed that the ATPase mutations abrogated the canonical chromatin-remodeling function of the SWI/SNF complexes in the interferon- $\gamma$  dependent transcriptional activation of the class II transactivator (*CIITA*) gene (21) (Fig. S6D). Taken together, these data indicate that the canonical ATP-dependent nucleosome remodeling activity of BRG1 is not required for paraspeckle formation.

### **SWI/SNF complexes facilitate the formation of a PSP interaction network in paraspeckles**

To determine what is defective in SWI/SNF-depleted cells ( $\Delta$ SWI/SNF cells) for paraspeckle formation, we measured the expression of known essential paraspeckle components, including *NEAT1\_2* and category 1 PSPs. RPAs were used to determine whether the paraspeckle disintegration observed in  $\Delta$ SWI/SNF cells was caused by diminished accumulation of *NEAT1\_2*. These analyses, which were performed using an antisense riboprobe that discriminated between the two *NEAT1* isoforms, revealed no loss in the expression of *NEAT1* isoforms in  $\Delta$ SWI/SNF cells (Fig. S4B). It is noteworthy that the *NEAT1\_1* level was higher in  $\Delta$ SWI/SNF cells, and consequently the ratio between *NEAT1* isoforms was altered (Fig. S4B). Furthermore, immunoblot analyses showed that seven essential PSPs (14) were constantly accumulated in  $\Delta$ SWI/SNF cells (Fig. S4C). These results indicate that paraspeckle disintegration in  $\Delta$ SWI/SNF cells was not caused by the down-regulation of known essential paraspeckle components.

In the absence of SWI/SNF complexes, paraspeckle formation was arrested despite the continued accumulation of *NEAT1\_2*. To examine the defective process of paraspeckle formation in  $\Delta$ SWI/SNF cells further, we examined the intermolecular interactions between essential paraspeckle components. RNA immunoprecipitation experiments showed that the interactions between *NEAT1* and category 1 PSPs were unaffected in  $\Delta$ SWI/SNF cells. An exception was the interaction between *NEAT1* and HNRNPK, which was approximately 50% lower in  $\Delta$ SWI/SNF cells than in control cells (Fig. 4A). The lower interaction between *NEAT1* and HNRNPK may be a consequence of the change in the *NEAT1* isoform ratio shown in Fig. S4B (14). However, coimmunoprecipitation analyses of category 1A proteins showed that the interactions between category 1A proteins (HNRNPK-SFPQ, RBM14-HNRNPK, RBM14-NONO, and SFPQ-RBM14) were significantly diminished in  $\Delta$ SWI/SNF cells (Fig. 4B and C). To determine whether the decreased interactions between PSPs in  $\Delta$ SWI/SNF cells were a consequence of paraspeckle structural disintegration, the interactions between PSPs were monitored in cells in which paraspeckle formation was blocked by depletion of FUS (a category 1B protein) (14). Knockdown of FUS had no effect on the interactions between RBM14 and other category 1A proteins (Fig. 4D). Taken together, these results suggest that the decreased interactions between PSPs in  $\Delta$ SWI/SNF cells are due to the loss of specific SWI/SNF complex functions required for paraspeckle formation. Next, we monitored interactions between SWI/SNF components and essential PSPs in  $\Delta$ *NEAT1* cells that lacked paraspeckles. The interactions between BRG1 and three essential PSPs were not affected by *NEAT1* depletion (Fig. 4E), indicating that the interactions between proteins in SWI/SNF complexes and PSPs is not dependent on

paraspeckle integrity.

### **SWI/SNF complexes are required for the formation of another lncRNA-dependent nuclear body**

Nuclear stress bodies (nSBs), another type of RNA-dependent nuclear body, occur in response to heat shock in a process initiated by the synthesis of lncRNA derived from pericentric tandem repeats of Satellite III (Sat III) sequences (22). Sat III lncRNA sequesters several splicing-related factors, such as serine/arginine-rich splicing factor 1 and scaffold attachment factor B (SAFB) (22). To determine whether SWI/SNF complexes are required for Sat III lncRNA-dependent formation of nSBs, the numbers of nSB-positive control HeLa cells,  $\Delta$ BRG1/ $\Delta$ BRM HeLa cells, and SW13 cells lacking functional SWI/SNF complexes were determined before and after heat shock at 42°C for 1 h. The formation of nSBs in response to heat shock was impaired in both  $\Delta$ BRG1/ $\Delta$ BRM HeLa cells and SW13 cells (Fig. 5A, 5B, and S5C). In addition, an I-EM analysis revealed localization of BRG1 in the electron dense granules of nSBs (Fig. 5C), supporting the involvement of SWI/SNF complexes in nSB formation. RNA immunoprecipitation with an  $\alpha$ BRG1 antibody revealed the association of BRG1 with heat shock-induced Sat III lncRNA (Fig. 5D); however, the expression of Sat III lncRNA was still detected in  $\Delta$ BRG1/ $\Delta$ BRM HeLa cells and SW13 cells (Fig. 5E). These data indicate that SWI/SNF complexes contribute to the assembly of nSBs by interacting with Sat III lncRNA, a role that is remarkably similar to their role in paraspeckle formation. Taken together, these results raise the possibility that SWI/SNF complexes are part of a common mechanism for the lncRNA-dependent formation of two

distinct nuclear bodies (Fig. 5F).

## Discussion

In this paper, we demonstrated that subunits of SWI/SNF complexes are localized to paraspeckles, where they play an essential role in the formation of these cellular structures. The canonical ATP-dependent nucleosome remodeling activity of SWI/SNF complexes is not required for this role. In a number of previous studies, immunofluorescent analyses revealed a broad distribution of human SWI/SNF components (BRG1 and BRM) throughout the nucleoplasm (e.g., 23, 24), which is in contrast to our observations. We suggest that the different fixation conditions (use of ethanol and acetone) used in this study (Fig. S1A) or the use of RNA-FISH (Fig. 1 and 3) improved the detection of BRG1 in paraspeckles by immunostaining. We can argue for the paraspeckle localization of SWI/SNF complexes based on the following data: 1) immunofluorescent signals of BRG1 were detected in paraspeckles and were lost upon knockdown of BRG but not BRM, 2) the paraspeckle localization of BRG1 was confirmed by using two distinct antibodies against BRG1, 3) five other SWI/SNF subunits were localized to paraspeckles, and 4) SWI/SNF components physically interacted with *NEAT1* and PSPs.

The results of the I-EM study also showed a marked enrichment of SWI/SNF components within paraspeckles; in particular, a characteristic patchy pattern of distribution in the interior region was observed. The 5' and 3' terminal regions and the middle region of *NEAT1\_2* are located mainly in the periphery and interior of paraspeckles, respectively (8). No other PSPs exhibited a similar patchy localization pattern, indicating that it was unique to

SWI/SNF components.

The I-EM study revealed the depletion of chromatin in SWI/SNF-located paraspeckle subdomains, suggesting that SWI/SNF complexes are localized to paraspeckles because they interact with *NEAT1* and PSPs, and not with the chromosomes. Similarly, SWI/SNF complexes, not chromosomes, may dictate the proper locations of *NEAT1* and other PSPs within the paraspeckle structure. This proposal is consistent with the finding that the nucleosome remodeling activity of SWI/SNF complexes is not required for their function in paraspeckle formation, and suggests that these complexes act as a part of the structural foundation of paraspeckles.

The evidence that SWI/SNF components (BRG1 and BRM) interact directly with *NEAT1*, as well as with a number of the essential PSPs, supports the pivotal functionality of the SWI/SNF components in sustaining paraspeckle structure. A recent study demonstrated that BRG1 binds to a specific lncRNA, termed Myheart, which acts to antagonize BRG1 function (25). The SWI/SNF component hSNF5 interacts with a specific lncRNA termed SChLAP1, which arrests the chromosome association of SWI/SNF complexes (26). Taken together, these findings suggest that some components of SWI/SNF complexes have affinity for lncRNAs.

According to our previous reports, paraspeckle formation involves at least two distinct steps: 1) *NEAT1\_2* expression by category 1A proteins, and 2) paraspeckle assembly by category 1B proteins that do not affect the *NEAT1\_2* expression level (14). In  $\Delta$ SWI/SNF cells, paraspeckle formation was defective despite *NEAT1\_2* accumulation, confirming that SWI/SNF complexes are involved in the second assembly step. SWI/SNF

depletion resulted in a marked loss of protein-protein interactions between essential PSPs, but these interactions were not affected by knockdown of FUS, a category 1B protein. Taken together, these results suggest that SWI/SNF complexes are involved in a novel essential step of paraspeckle formation.

In mammalian cells, additional nuclear bodies constructed around specific lncRNAs have been reported (27), and nSB foci form on stress-induced Sat III lncRNA in response to heat shock. SWI/SNF complexes are localized in nSBs, where they interact with SatIII lncRNA. Depletion of SWI/SNF complexes impaired the formation of nSBs in response to heat shock without affecting the level of Sat III lncRNA, suggesting that these complexes are involved in the assembly of Sat III sub-RNP complexes. The mechanism involved may be similar to that of SWI/SNF-dependent paraspeckle assembly. Paraspeckles and nSBs sequester specific sets of RNA-binding proteins whose specificity is defined by RNA sequence elements that reside in the lncRNAs. Similar to paraspeckles, nSBs form on the locus where Sat III lncRNA is transcribed; therefore, it is intriguing to hypothesize that SWI/SNF complexes bridge the architectural lncRNA to the sequestered RNA-binding protein to construct huge RNP complexes at the cognate chromosomal locus. Further characterization of the modes of action of SWI/SNF complexes will help unveil the common mechanisms underlying the lncRNA-dependent formation of nuclear bodies.

## **Materials and Methods**

Basic methods for techniques such as RPA, qRT-PCR, and immunoblotting are described in SI.

## **Cell culture**

HeLa, A549, HEK293, NIH3T3, and SW13 cells were grown and maintained as described previously (14). HAP1 cells were purchased from Haplogen GmbH and cultured in Iscove's Modified Dulbecco's Medium (IMDM). For heat shock experiments, the cells were incubated for 1 h at 42°C and then allowed to recover for 1 h at 37°C. Some cells were treated with actinomycin D (0.3 µg/ml) for 4 h or MG132 (5 µM) for 17 h.

## **RNA-FISH and immunocytochemistry**

The RNA-FISH probes were synthesized using SP6 RNA polymerase and a DIG/FITC RNA labeling kit (Roche Diagnostic). Linearized plasmids (1 µg) containing a *NEAT1* fragment (+1 to +1,000) or a *MALAT1* fragment (+5,114 to +5,712) were used as templates for transcription. RNA-FISH and immunocytochemical analyses were performed as described previously (14). The detailed conditions are described in the SI.

## **Immunoprecipitation of RNP complexes**

HeLa cells were lysed with IP lysis buffer [50 mM Tris-HCl (pH 7.5), 150 mM NaCl, 1% Triton X-100, 20 U/ml SUPERase•In (Ambion), cOmplete EDTA-free protease inhibitor, and PhoSTOP phosphatase inhibitor] and then disrupted by three pulses of sonication for 5 s. The cell extracts were incubated with or without RNase A (1 µg/ml) or ethidium bromide (50 µg/ml) on ice for 30 min, and then cleared by centrifugation at 10,000 × g for 10 min. The antibodies were incubated with Dynabeads Protein-G, Dynabeads anti-rabbit IgG, or



Dynabeads anti-mouse IgG (Life Technologies) for 1 h and then washed five times with IP lysis buffer. The remaining supernatants were mixed with the antibody-bead conjugates and rotated at 4°C for 3 h or overnight, after which the beads were washed three times with IP lysis buffer. For crosslinking immunoprecipitation, 1.8 J/cm<sup>2</sup> UV light was used for irradiation, as described previously (6). The antibodies used are listed in Table S1.

### **CRISPR/Cas9-mediated genome engineering.**

The guide RNA in the CRISPR/Cas9 system was designed using the CRISPR design tool (<http://crispr.mit.edu/>) and then cloned into the BbsI site of the PX459 vector (Addgene plasmid ID: 48141). HAP1 cells were transfected with the guide RNA plasmid using Nucleofection reagent (Ronza). For clonal selection of the mutants, the cells were selected in IMDM medium containing 167 ng/ml of puromycin for 2 days, and then diluted and incubated in IMDM medium without puromycin in 96-well plates. The selected clones were lysed, and the genomic region flanking the CRISPR target site was amplified by PCR to check for the presence of small deletions, which were confirmed by sequencing. The primers used are listed in Table S2.

### **Acknowledgments**

The authors thank Y. Ikeuchi and the members of the T.H. laboratory for useful discussions. This research was supported by the Funding Program for the Ministry of Education, Culture, Sports, Science, and Technology of Japan (MEXT), as well as by grants from the New Energy and Industrial Technology Development Organization (NEDO), the Mitsubishi

Foundation, the Takeda Science Foundation, the Suminoto Foundation, the Naito Foundation, and the Mochida Memorial Foundation (to T.H.).

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## Figure Legends

### Fig. 1. Paraspeckle localization of SWI/SNF complex components.

(A, B) Localization of BRG1, BRM, and BAF155 to paraspeckles in human and mouse cells. The SWI/SNF components were detected by immunocytochemical analyses and the paraspeckles were visualized by RNA fluorescence *in situ* hybridization analyses of *NEAT1* lncRNA. The experiments were performed in HeLa cells (A) and other human cell lines (A549 and HEK293), as well as a mouse cell line (NIH3T3) (B). Scale bars: 10  $\mu$ m. (C) Relocalization of BRG1 and BRM to perinucleolar caps upon transcriptional arrest caused by treatment of HeLa cells with actinomycin D (+Act D). The indicated paraspeckle proteins and SWI/SNF components were detected by immunocytochemical analyses, and the paraspeckles were visualized by RNA-FISH analyses of *NEAT1* lncRNA. NONO, which is known to relocalize to perinucleolar caps, was used as a positive control. COIL relocates to another cap structure. Scale bars: 10  $\mu$ m. (D). Immunogold electron microscopic detection of BRG1 (upper panels) in normal (control) paraspeckles and enlarged paraspeckles induced by the treatment of HeLa cells with MG132. Chromosome localization was detected

using an anti-histone H3 antibody (lower photos). The arrows indicate paraspeckles. Cyt, cytoplasm; Nu, nucleus; IG, interchromatin granule clusters. Scale bars: 0.5  $\mu\text{m}$ .

**Fig. 2. SWI/SNF subunits interact directly with essential paraspeckle components.**

(A) Detection of direct interactions between SWI/SNF components (BRG1 and BRM) and *NEAT1* lncRNA by UV crosslinking immunoprecipitation. After UV crosslinking, the coimmunoprecipitated RNAs were quantified by qRT-PCR, and the immunoprecipitation (IP) efficiencies (%) were determined (n = 3). The UV- lanes are negative controls without UV crosslinking. IgG was used as a negative control for immunoprecipitation. (B) Interactions between BRG1 and five essential paraspeckle proteins (shown on the left), detected by coimmunoprecipitation with an anti-BRG1 antibody ( $\alpha$ BRG1) in the presence (+) or absence (-) of RNase A (left panel) or ethidium bromide (EtBr) (right panel). GAPDH was detected as a negative control.

**Fig. 3. SWI/SNF complexes are essential for paraspeckle formation.**

(A) The effects of RNAi-mediated knockdown of the indicated SWI/SNF components on paraspeckle formation. The effects were monitored by analyzing paraspeckle integrity using RNA fluorescence *in situ* hybridization detection of *NEAT1* and immunofluorescent detection of PSPC1. The siRNAs used in these experiments are listed in Table S3. Because BRG1 and BRM are functionally redundant, these proteins were concomitantly knocked down with a combination of two siRNAs (BRG1#2/BRM#1 and BRG1#4/BRM#2). The results of single RNAi-mediated knockdown of BRG1 or BRM are shown in Fig. S3. Scale bars: 10  $\mu\text{m}$ . (B)

Quantification of the number of paraspeckle-positive cells after RNAi-mediated knockdown of the indicated SWI/SNF components (as shown in A). Data are represented as the mean  $\pm$  SD of  $n = 3$  replicates. The cell numbers for the control, BRG1#2/BRM#1, BRG1#4/BRM#2, BAF170, BAF155, BAF57, and BAF47 groups were 435, 229, 266, 165, 149, 198, and 227, respectively.  $*P < 0.05$  and  $**P < 0.02$  by Student's t-tests. (C) The lack of paraspeckles in adrenal cortex adenocarcinoma SW13 cells (upper panels). RNA-FISH was used to detect *NEAT1* and immunocytochemistry was used to detect RBM14 in SW13 and HeLa (control) cells. Scale bars: 10  $\mu\text{m}$ . Nuclear speckles in SW13 cells as detected by RNA-FISH analyses of *MALAT1* and immunofluorescent analyses of SRSF2 (lower panels). Scale bars: 10  $\mu\text{m}$ . (D) Paraspeckle formation does not require the canonical nucleosome remodeling activity of SWI/SNF complexes. Schematic representations of wild type (WT) and two ATPase mutants (M1 and M2) of BRG1 are shown on the left. HAP1 cells were treated with siBRM to abolish the background expression of BRM. The essential lysine residue (K) in WT BRG1 and the artificially inserted proline residue (P) in the M2 mutant are shown in red and blue, respectively. The paraspeckles detected in WT, M1, and M2 HAP1 cells are shown on the right. Scale bar: 10  $\mu\text{m}$ .

**Fig. 4. SWI/SNF complexes are required for proper interactions between paraspeckle proteins (PSPs).**

(A) RNA immunoprecipitation of *NEAT1* with six essential PSPs from control and  $\Delta$ SWI/SNF cells. The amounts of immunoprecipitated RNAs were quantified by qRT-PCR using the *NEAT1* and *NEAT1\_2* primer pairs shown in Table S2. The ratio of immunoprecipitated RNA

from  $\Delta$ SWI/SNF cells to that from the control cells was calculated. The expression level of *GAPDH* was measured as a control. Data are represented as the mean  $\pm$  SD of  $n = 3$  replicates. (B, C) Interactions between the essential PSPs in control (-) and  $\Delta$ SWI/SNF (+) cells. Coimmunoprecipitation (coIP) was performed using antibodies against HNRNPK, RBM14, and SFPQ, and the expression levels of the indicated PSPs were monitored by immunoblotting (B). The interactions that were diminished by  $<40\%$  in  $\Delta$ SWI/SNF cells are indicated by I–IV, and the quantitated coIP ratios ( $\Delta$ SWI/SNF/control) in I–IV are plotted on the graph shown in (C). (D) Interactions between RBM14 and essential PSPs in control (-) and paraspeckle-depleted  $\Delta$ FUS (+) cells. CoIP was performed using an anti-RBM14 antibody and the expression levels of the indicated PSPs were monitored by immunoblotting. (E) Interactions between BRG1 and the essential PSPs in control (-) and *NEAT1*-depleted (+) cells.

**Fig. 5. SWI/SNF complexes are required for the formation of nuclear stress bodies.**

(A) Immunostaining of scaffold attachment factor B (magenta) to detect the formation of nSBs before and after heat shock (HS) of control HeLa cells, SW13 cells, and BRG1-/BRM-specific siRNA-treated HeLa cells. Scale bars: 10  $\mu$ m. (B) Quantification of the numbers of nSB-positive cells shown in (A). Data are represented as the mean  $\pm$  SD of  $n = 3$  replicates. The cell numbers for the HeLa(HS-), SW13(HS-), HeLa(HS+), SW13(HS+), C(HS-),  $\Delta$ BRG1/ $\Delta$ BRM(HS-), C(HS+), and  $\Delta$ BRG1/ $\Delta$ BRM(HS+) groups were 276, 228, 303, 202, 341, 356, 332 and 403, respectively. **\*\* $P < 0.02$**  by a Student's t-test. (C) Immunogold electron microscopic detection of BRG1 in nSBs. In the left panel, the dashed line indicates



a single nSB. Scale bars: 0.5  $\mu\text{m}$  (left) and 100 nm (right). (D) RNA immunoprecipitation of Sat III lncRNA from heat shock-treated HeLa cells using the  $\alpha\text{BRG1}$  antibody. The expression level of  $\beta$ -actin was used as a control. RT, reverse transcription. The sequences of the PCR primers used are shown in Table S2. (E) RT-PCR analyses of Sat III lncRNA expression in control HeLa cells, SW13 cells, and BRG1-/BRM-specific siRNA-treated HeLa cells. HS, heat shock at 42°C for 1 h. (F) A model of lncRNA-dependent nuclear body assembly. SWI/SNF complexes may be common assembly factors in mammalian cells. Nuclear bodies act to sequester specific regulatory proteins to control the expression of specific genes (9).

Fig. 1

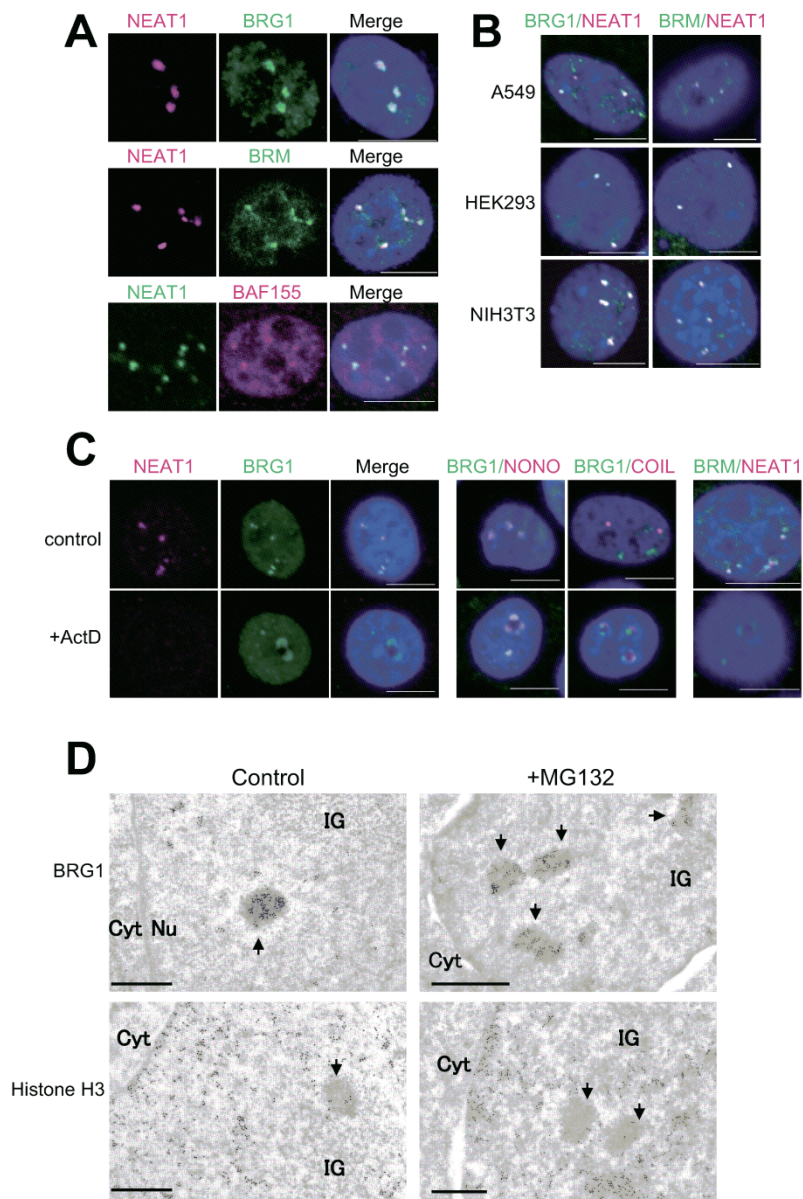


Fig. 2

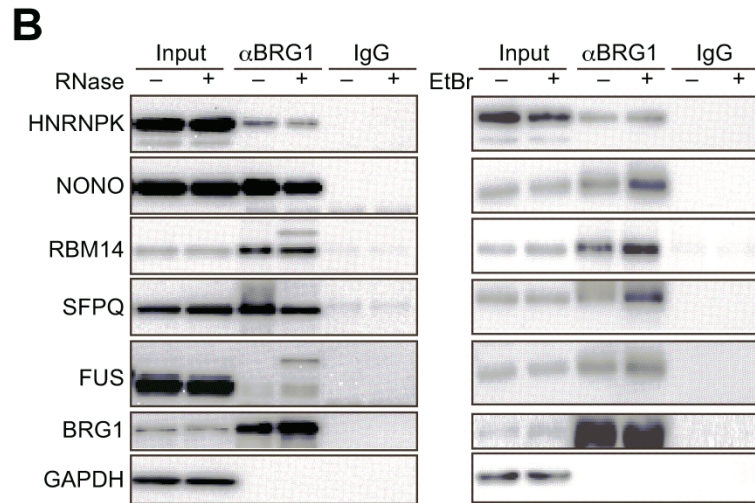
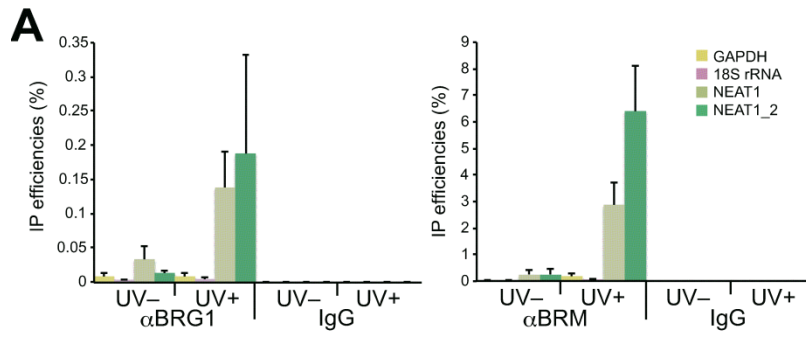


Fig. 3

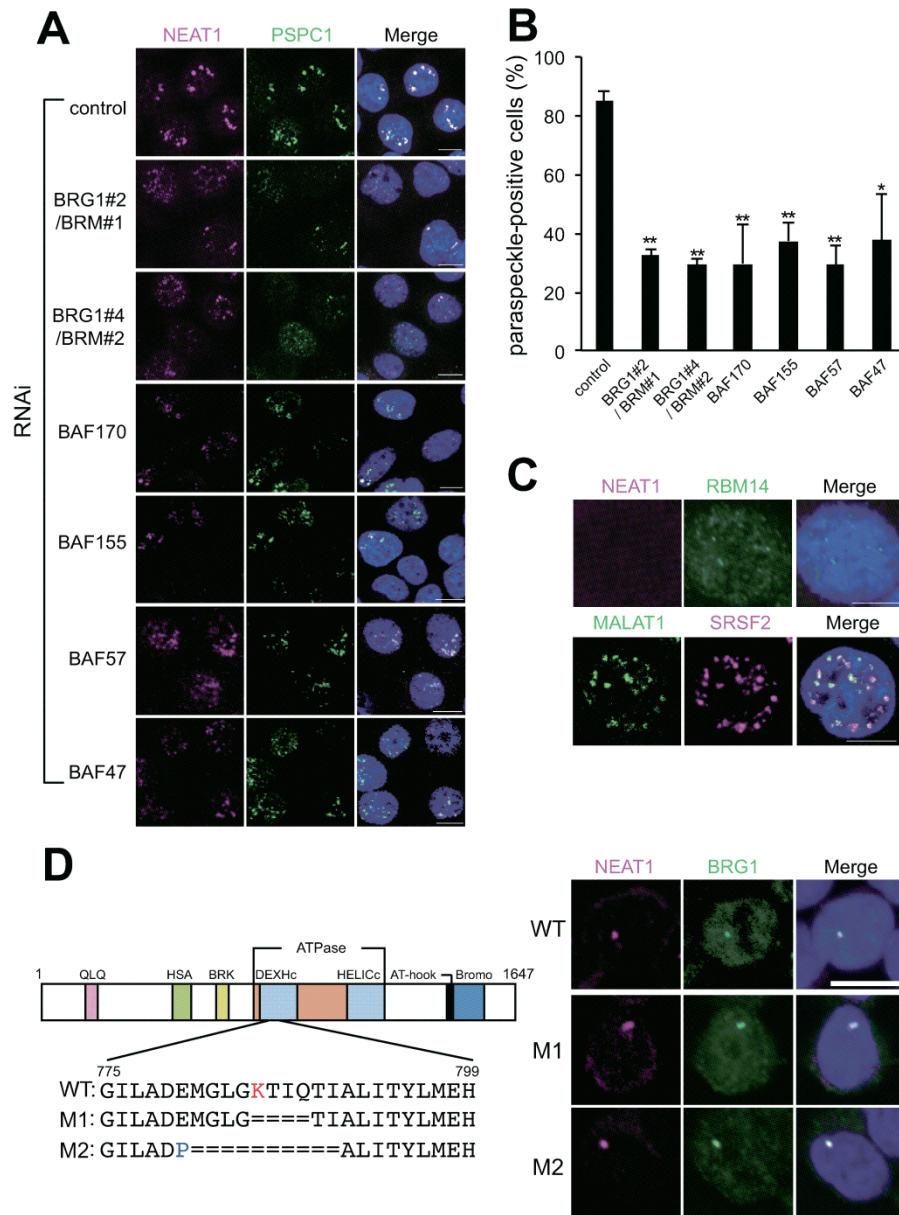


Fig. 4

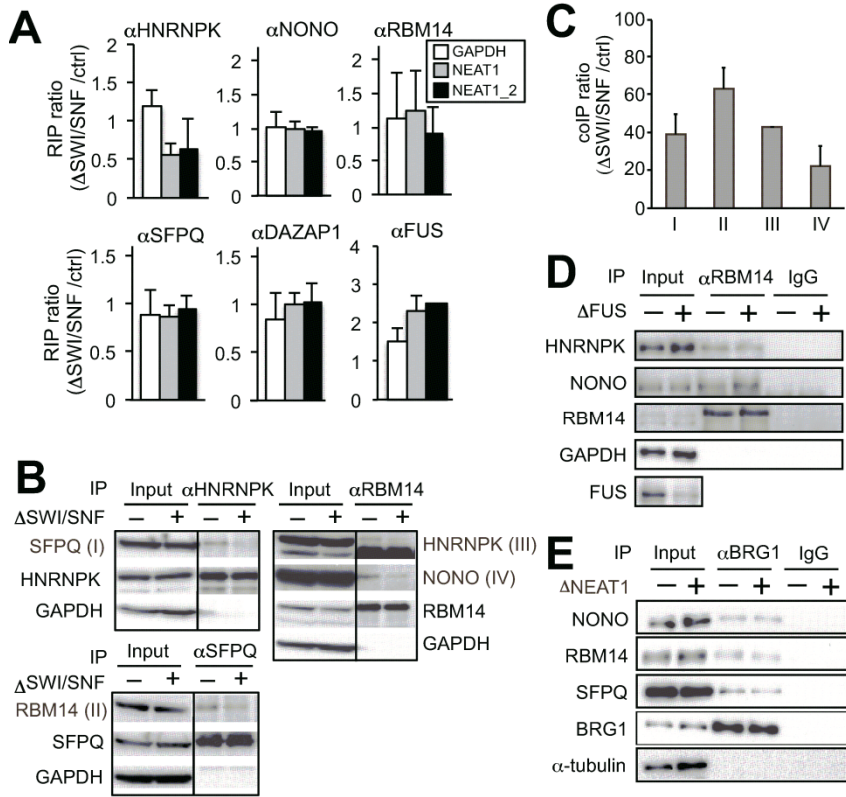
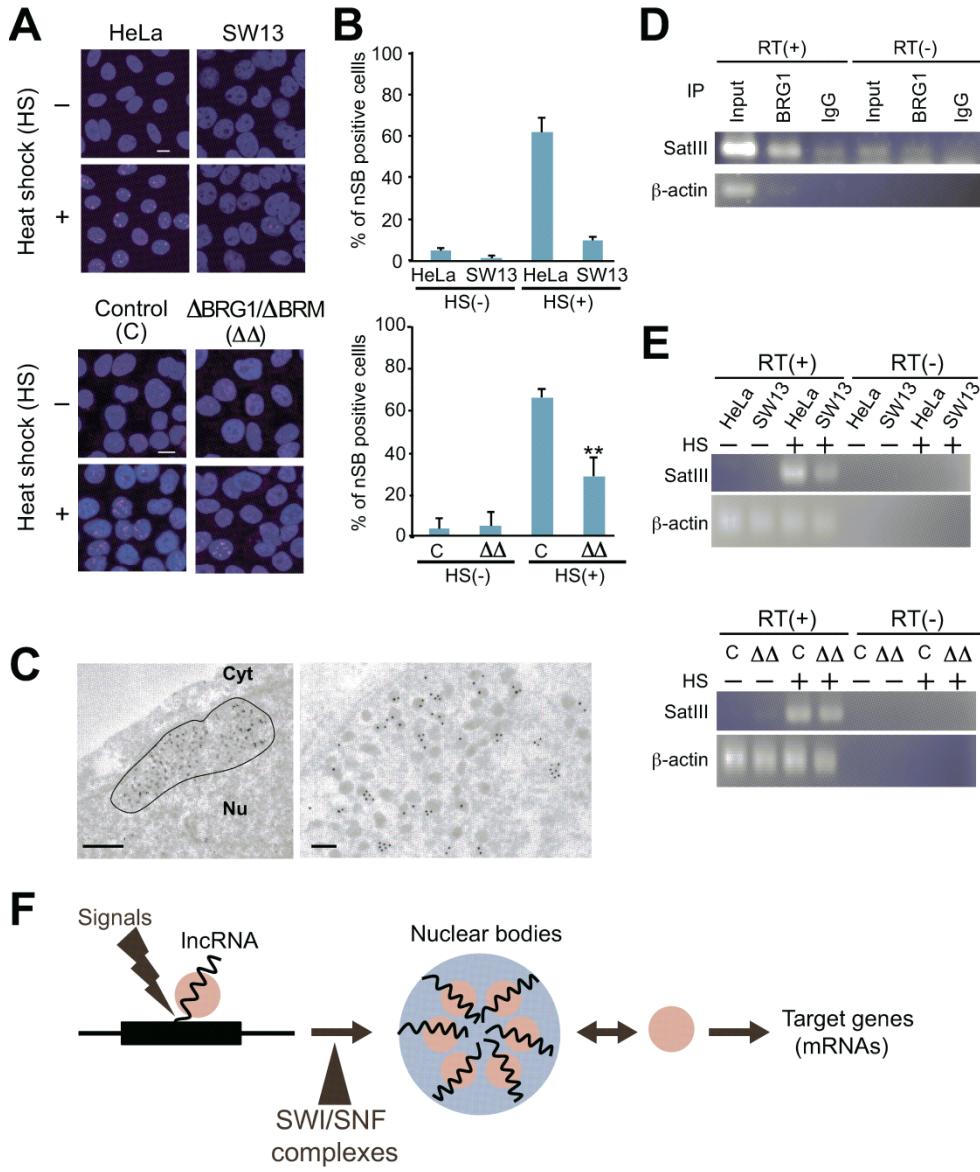


Fig. 5



**Table 1.** The paraspeckle proteins interacted with SW/SNF subunits

SWI/SNF	Paraspeckle protein
BRG1	NONO*, SFPQ*, CPSF7, CPSF6, SS18L1
BRM	NONO*, SFPQ*, RBM14*, HNRNPK*, FUS*, HNRNPA1, HNRNPR, TARDBP1, CPSF6, RBMX, HNRNPF, HNRNPH1, SS18L1
BAF170	NONO*, SFPQ*, RBM14*, HNRNPK*, FUS*, CPSF7, HNRNPA1, HNRNPR, TARDBP1, CPSF6, NUDT21, EWSR1, RBM7, RBMX, TAF15, HNRNPF, HNRNPH1
BAF155	NONO*, SFPQ*, RBM14*, HNRNPK*, FUS*, CPSF7, HNRNPA1, HNRNPR, TARDBP1, CPSF6, NUDT21, EWSR1, RBM7, RBMX, TAF15, HNRNPF, HNRNPH1
BAF57	NONO*, SFPQ*, RBM14*, HNRNPK*, FUS*, HNRNPA1, HNRNPR, TARDBP1, NUDT21, EWSR1, RBM7, RBMX, TAF15, HNRNPF, HNRNPH1
BAF47	NONO*, SFPQ*, RBM14*, HNRNPK*, FUS*, HNRNPA1, HNRNPR, TARDBP1, NUDT21, EWSR1, RBM7, RBMX, TAF15, HNRNPF, HNRNPH1

\*The essential paraspeckle proteins (Naganuma et al. 2012)